

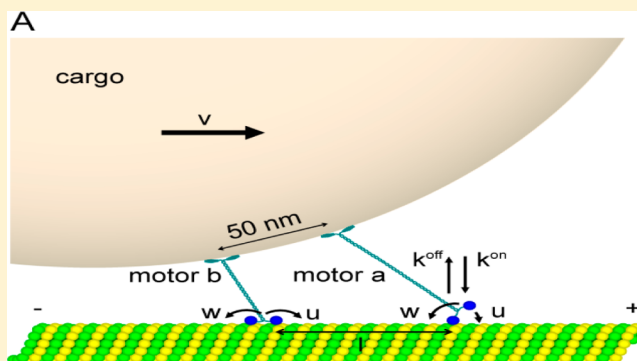
How the Interplay between Mechanical and Nonmechanical Interactions Affects Multiple Kinesin Dynamics

Karthik Uppulury,[‡] Artem K. Efremov,[†] Jonathan W. Driver,[†] D. Kenneth Jamison,[†] Michael R. Diehl,^{*,†,‡} and Anatoly B. Kolomeisky^{*,‡}

[†]Department of Bioengineering, Rice University, Houston, Texas 77005, United States

[‡]Department of Chemistry, Rice University, Houston, Texas 77005, United States

ABSTRACT: Intracellular transport is supported by enzymes called motor proteins that are often coupled to the same cargo and function collectively. Recent experiments and theoretical advances have been able to explain certain behaviors of multiple motor systems by elucidating how unequal load sharing between coupled motors changes how they bind, step, and detach. However, nonmechanical interactions are typically overlooked despite several studies suggesting that microtubule-bound kinesins interact locally via short-range nonmechanical potentials. This work develops a new stochastic model to explore how these types of interactions influence multiple kinesin functions in addition to mechanical coupling. Nonmechanical interactions are assumed to affect kinesin mechanochemistry only when the motors are separated by less than three microtubule lattice sites, and it is shown that relatively weak interaction energies ($\sim 2 k_B T$) can have an appreciable influence over collective motor velocities and detachment rates. In agreement with optical trapping experiments on structurally defined kinesin complexes, the model predicts that these effects primarily occur when cargos are transported against loads exceeding single-kinesin stalling forces. Overall, these results highlight the interdependent nature of factors influencing collective motor functions, namely, that the way the bound configuration of a multiple motor system evolves under load determines how local nonmechanical interactions influence motor cooperation.



1. INTRODUCTION

The transport of organelles and many other subcellular materials is strongly dependent on enzymes called motor proteins that use the chemical energy released from ATP hydrolysis to drive cargo transport along periodic cytoskeletal filaments.^{1,2} Many motor proteins can move processively along their filament tracks and can produce forces that should allow them to transport their cargos independently as single-motor molecules in many circumstances.^{3–8} Nevertheless, a cargo's motion is often driven by teams of similar and/or dissimilar motors that function either in a concerted fashion or antagonistically.^{9–11} Collective motor functions are important to various intracellular transport and trafficking processes, since they can determine how fast and far cargos are transported in the cytoplasm, the net directionality of cargo motions, and even how cargos switch between microtubule- and actin-dependent transport modes. Although these aspects of intracellular transport have received increased attention, understanding the impact of collective motor behaviors still requires improved knowledge of how the mechanochemistry of motor proteins is affected by motor cooperation.

Recent experimental and theoretical advances have provided important insights into mechanisms underlying the collective dynamics of processive motor molecules, particularly for

multiple kinesins.^{12–17} Several methods have been developed to characterize how cargo run lengths, velocities, detachment forces, and step sizes are influenced by the number of motors responsible for cargo transport.^{18–21} Among these studies, our group has examined the dynamic properties of structurally organized complexes composed of two interacting kinesin molecules using precision particle tracking and optical trapping procedures.^{12,13} These studies demonstrate that small collections of kinesins can transport cargos against larger forces and over longer distances than single kinesin molecules can produce on their own. However, analyses of two-kinesin run lengths, instantaneous velocities, average force–velocity relationships, and, perhaps most importantly, transition rates between different conformations of the two-motor system where either one of both kinesins within a complex bear the applied load imposed on a cargo all suggest that cargo transport by two kinesins will primarily be driven by a single-motor molecule. Given this behavior, the average transport behaviors of these two-kinesin complexes are best characterized as net negative cooperative, since they fall short of expectations from analytical

Received: April 25, 2012

Revised: June 21, 2012

Published: June 23, 2012



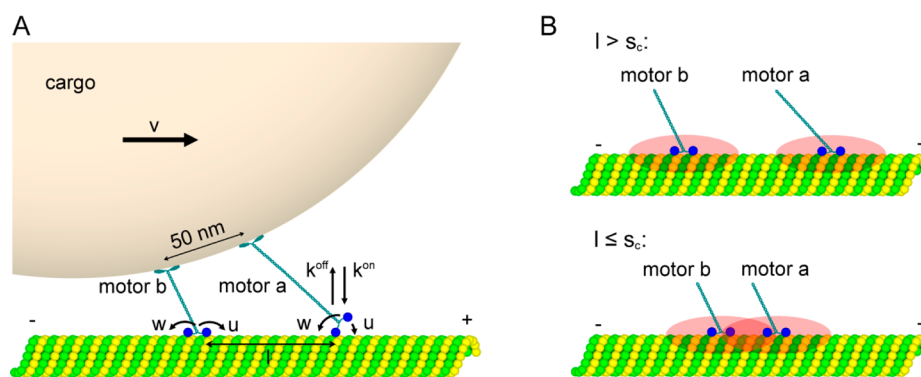


Figure 1. (A) A schematic picture of a model for transportation of cargo by a complex of two coupled kinesin motor proteins along the microtubule. Arrows indicate microscopic transition associated with kinesin binding, stepping, and detachment. (B) Local areas on microtubules that are affected by interactions with kinesins: (top panel) motor proteins are far away from each other; (bottom panel) closely spaced motor species that interact with each other.

treatments where two motors are assumed to function noncooperatively, meaning that they simply share their applied load equally when both kinesins are engaged in transport and do not interact in other ways.

The ability to construct organized motor systems has also facilitated comparisons of multiple kinesins under the different loading conditions provided by a static optical trap and an optical force clamp.^{13,22} As recognized by earlier theoretical studies,¹⁸ these analyses illustrate that the spatial and temporal dependencies of the applied loads experienced by multiple motor complexes can influence their ability to bind to microtubules in configurations where loads are shared equally between the motors.^{23,24} For example, loads in the static trap are found to increase rapidly compared to the rates at which the complexes evolve from single-motor-bound states into load-sharing configurations when cargo transport is driven by multiple kinesins. Consequently, a cargo's primary load-bearing kinesin will most often experience a high force and detach from the filament before its nonparticipating motor partner is able to assist in cargo motion. Two-kinesin complexes are only able to transport cargos to positions in the trap where the load exceeds kinesin's stalling force if they generate load-sharing states before they arrive at this load. This spatial filtering of multiple motor configurations therefore results in behaviors where motors cooperate negatively at low applied loads but function more productively via load sharing at high applied loads. Moreover, this behavior is further reinforced by the fact that partial cargo-filament detachment due to the release of one of the complex's motors results in rearward bead displacement, which only increases the probability that the transport will occur by a single kinesin molecule at low applied loads.

Key experimental signatures reflecting the relatively weak dependence of cargo transport on kinesin copy number can be reproduced by a discrete-state stochastic model that has been developed to explore the load-dependent dynamics of multiple kinesins.²⁵ This model is unique since it (i) accounts for chemical transitions between single-motor-bound and a spectrum of two-motor-bound states where motors are spaced by different distances on the microtubule and assume different portions of the applied load imposed on a cargo, (ii) parametrizes microscopic transition rates using fits to single-kinesin optical trapping data exclusively, and (iii) examines multiple motor dynamics via numerical calculations that facilitate more accurate analyses of the spatiotemporal dynamics of a multiple motor system. While illustrating that there are

significant kinetic barriers that limit a complex's ability to bind the filament in configurations where both motors share their applied load, this model also reproduces specific experimental observations including nonmonotonic dependencies of cargo velocities and motor-filament detachment rates on the magnitude of the applied load, and it even provides insight into how these unique behaviors result from the spatial and temporal dependence of the load experienced by a cargo.^{13,25} Finally, this theoretical framework predicts that the collective behavior of multiple motor proteins depends on their stepping mechanism. It is argued that efficient and strong motors like kinesins have greater difficulties cooperating productively compared to weaker and less efficient motors due to the relatively low susceptibility of kinesin's velocity to applied loads.²⁵

While the above theoretical approach is capable of capturing various important and unique experimental observations, this treatment assumes that multiple kinesins only interact mechanically through the elastic linkages connecting them to their cargo. Using this approach, the rates at which a motor complex will transition between configurations via motor binding, detachment, and stepping are therefore only dependent on the corresponding change in a complex's strain energy during these transitions. There are several lines of evidence suggesting that other forms of interactions between filament-bound motor proteins could also play an important role in multiple kinesin dynamics.^{26,27} Kinesins are found to phase segregate into patches of dense and sparsely decorated filaments in electron microscopy studies,²⁷ and to pause transiently on a microtubule when they are in close proximity to neighboring motors that are bound in a rigor state.²⁸ This behavior clearly indicates some form of short-range interaction potentials acting between microtubule-bound motor proteins. Similarly, local interactions among kinesins are evidenced by the clustering of kinesins along the microtubule filament even though the tails of these motors are not coupled mechanically. Importantly, the energy scale of these interactions was estimated to be relatively small ($\sim 1.6 \pm 0.5 k_B T$),²⁶ and they are found to affect both motor-filament binding and detachment in the absence of an applied load.

Outside of molecular crowding effects, the impact of such interactions on motor stepping has largely been overlooked.²⁸ Nevertheless, measured velocities in the recent optical trapping studies at loads exceeding kinesin's stalling force are much higher than those predicted by the models that only address

mechanical coupling between motors, even if more than two motors are assumed to be present on the cargo surface.²⁵ In fact, these velocities even appear to exceed predictions that assume motors share their applied load perfectly when transporting a cargo, implying some form of local interaction may potentially allow the motors to function more synergistically in this load regime.

Herein, we extend our previous discrete-state stochastic approach and develop a new theoretical model to examine the influence of local, nonmechanical, interactions between kinesins on their transport dynamics under applied loads. In particular, we explore how these interactions affect two-kinesin velocities and detachment rates when loads vary spatially and temporally. The present analyses suggest that local interactions can alter multiple motor detachment and stepping rates appreciably at high applied loads, even if the energies of these interactions are small. However, the overall impact of these effects is generally superseded by other kinetic constraints that determine how the bound geometries of a multiple motor complex evolve under load, as well as other factors influencing the spatiotemporal “filtering” of multiple kinesin states. From this basis, we discuss how this behavior stems from kinesin’s efficient stepping mechanism, and propose how similar interactions might influence other classes of motors possessing different mechano-chemical properties.

2. THEORETICAL METHODS

2.1. Discrete-State Stochastic Model. The present model builds upon a theoretical method that has been developed in refs 25 and 29. It was designed to examine the dynamics of the two-kinesin complexes as experimentally studied in refs 12 and 13. The geometry of the two-kinesin complexes bound to cargo and moving along microtubules as well as relevant kinetic transitions are illustrated in Figure 1A. To describe the structural organization of the complexes, we consider that two kinesins are bound 50 nm apart from one another on the surface of a 500 nm (diameter) bead. The motor–protein assemblies can associate with the microtubule via a single kinesin or via a range of two-motor-bound configurations where the separation distance l between motor-filament binding sites varies.

Each microtubule-bound kinesin molecule at the lattice site i can step forward (backward) with a rate u_i (w_i), and dissociates with a rate k_i^{off} . Unbound motors bind to the filament with a rate $k_{i \rightarrow (i,j)}^{\text{on}}$ (the meaning of labels here is explained below); see Figure 1A. These rates are specified by free-energy differences for conformations that are connected by these transitions following the requirement from the detailed balance conditions. The free energy of each state is calculated explicitly assuming that a mechanical (but not a chemical) equilibrium is achieved in each state after individual structural transitions in the complex are taking place.²⁵ This energy, which we refer to as a complex’s configurational energy (E_{config}), is computed from stretched motor lengths and bead-trap displacements obtained using a mechanical modeling and energy minimization procedure that balances the forces on the bead from the motors and trap to within 0.1 fN.²⁵ During this procedure, the elasticity of each motor-bead linkage is parametrized using measurements of single kinesin stiffnesses.¹³ A complex’s configurational energy can then be determined using the following expression:

$$E_{\text{config}} = \frac{1}{2} \kappa_T (x_T - x_b)^2 + \sum_M \int_{l_0}^{l_{\text{ax}}} \|\vec{F}_{\text{ax}}\| dl \quad (1)$$

where κ_T is the trap’s spring constant, $(x_T - x_b)$ is the distance by which a bead is displaced from the trap center, l_{ax} is the length of a motor when it experiences a force along its stalk axis (F_{ax}), and l_0 is the length of an unloaded motor. The rates at which complexes transition between configurations can then be determined using the corresponding difference in configurational energies (ΔE_{config}) as explained below.

The temporal evolution of the motor protein complexes can be described by the following set of master equations:

$$\frac{d\psi^0(t)}{dt} = \sum_i k_i^{\text{off}} \psi_i^a(t) + \sum_j k_j^{\text{off}} \psi_j^b(t) \quad (2)$$

$$\begin{aligned} \frac{d\psi_i^a(t)}{dt} = & u_{(i-1) \rightarrow i} \psi_{i-1}^a(t) + w_{(i+1) \rightarrow i} \psi_{i+1}^a(t) \\ & - (k_i^{\text{off}} + \sum_j k_{i \rightarrow (i,j)}^{\text{on}} + u_{i \rightarrow (i+1)} + w_{i \rightarrow (i-1)}) \\ & \psi_i^a(t) + \sum_j k_{(i,j) \rightarrow i}^{\text{off}} \psi_{(i,j)}^{a,b}(t) \end{aligned} \quad (3)$$

$$\begin{aligned} \frac{d\psi_j^b(t)}{dt} = & u_{(j-1) \rightarrow j} \psi_{j-1}^b(t) + w_{(j+1) \rightarrow j} \psi_{j+1}^b(t) \\ & - (k_j^{\text{off}} + \sum_i k_{j \rightarrow (i,j)}^{\text{on}} + u_{j \rightarrow (j+1)} + w_{j \rightarrow (j-1)}) \\ & \psi_j^b(t) + \sum_i k_{(i,j) \rightarrow j}^{\text{off}} \psi_{(i,j)}^{a,b}(t) \end{aligned} \quad (4)$$

$$\begin{aligned} \frac{d\psi_{(i,j)}^{a,b}(t)}{dt} = & u_{(i-1,j) \rightarrow (i,j)} \psi_{(i-1,j)}^{a,b}(t) + u_{(i,j-1) \rightarrow (i,j)} \psi_{(i,j-1)}^{a,b}(t) \\ & + w_{(i+1,j) \rightarrow (i,j)} \psi_{(i+1,j)}^{a,b}(t) \\ & + w_{(i,j+1) \rightarrow (i,j)} \psi_{(i,j+1)}^{a,b}(t) + k_{i \rightarrow (i,j)}^{\text{on}} \psi_i^a(t) \\ & + k_{j \rightarrow (i,j)}^{\text{on}} \psi_j^b(t) - (k_{(i,j) \rightarrow i}^{\text{off}} + k_{(i,j) \rightarrow j}^{\text{off}} \\ & + u_{(i,j) \rightarrow (i+1,j)} + u_{(i,j) \rightarrow (i,j+1)} + w_{(i,j) \rightarrow (i-1,j)} \\ & + w_{(i,j) \rightarrow (i,j-1)}) \psi_{(i,j)}^{a,b}(t) \end{aligned} \quad (5)$$

In these equations, $\psi^0(t)$ is the probability that the system will be found in a state where the motor protein complex is completely detached from the microtubule at time t . Similarly, the quantities $\psi_i^a(t)$ and $\psi_j^b(t)$ correspond to probabilities of the single-motor-bound states where the labels a and b specify different kinesin molecules and the labels i and j are used to specify positions of the motors on the microtubule lattice. The probabilities for two-motor-bound states are given by functions $\psi_{(i,j)}^{a,b}(t)$ for kinesin molecules a and b connected to the microtubule at positions i and j .

Transition rates in the master equations are estimated as follows. The detachment rate of a motor in the absence of load ($k_0^{\text{off}} = 0.312 \text{ s}^{-1}$) is taken from experiments,²⁵ and the unloaded binding (k_0^{on}) rate assumes a value of 4.7 s^{-1} from other experiments.³⁰ The ratio of binding to detachment rates under applied loads is related to the unloaded rates by the detailed balance condition:

$$\frac{k_{i \rightarrow (ij)}^{\text{on}}}{k_{(ij) \rightarrow i}^{\text{off}}} = \frac{k_0^{\text{on}}}{k_0^{\text{off}}} e^{-\beta \Delta E_{\text{config}}} \quad (6)$$

where $\Delta E_{\text{config}} = E_{\text{config}}(ij) - E_{\text{config}}(i)$ and β is equal to $1/k_B T$; $k_{i \rightarrow (ij)}^{\text{on}}$ ($k_{j \rightarrow (ij)}^{\text{on}}$) and $k_{(ij) \rightarrow i}^{\text{off}}$ ($k_{(ij) \rightarrow j}^{\text{off}}$) are the rates for transitioning between specified single- and two-motor-bound configurations via binding and detachment rates, respectively. The detachment rate $k_{(ij) \rightarrow i}^{\text{off}}$ of a motor from a two-motor-bound state to a single-motor state is estimated by considering motor detachment as a two-state process (where the positions and energies of the intermediate and transition states are determined from single-motor detachment rate data).²⁵ The detachment rates can be calculated for different loading conditions in the optical trap. Further, the binding rate of a motor $k_{i \rightarrow (ij)}^{\text{on}}$ can be obtained from eq 6.

The stepping rates of the motor under applied loads are calculated using the formalism of Fisher and Kim.³¹ Here, kinesin's reaction coordinate is approximated by a two-state model that accounts for intermediate biochemical states and for the work done by a motor as it transitions between spatially separated ground and intermediate states. The corresponding rates for these substeps can be written as

$$u_+ = u_+^0 e^{-\beta(E_{i,\text{TS1}} - E_i)} \quad (7)$$

$$u_{++} = u_{++}^0 e^{-\beta(E_{i,\text{TS2}} - E_{i,\text{IS}})} \quad (8)$$

$$w_- = w_-^0 e^{-\beta(E_{i,\text{TS1}} - E_{i,\text{IS}})} \quad (9)$$

$$w_{--} = w_{--}^0 e^{-\beta(E_{i,\text{TS2}} - E_{i+1})} \quad (10)$$

The unloaded stepping rates u_+^0 , u_{++}^0 , w_-^0 , and w_{--}^0 are obtained from fits to single kinesin optical trapping data.²⁵ E_i is the energy of the complex with the transitioning motor bound to the i th lattice site. $E_{i,\text{TS1}}$, $E_{i,\text{IS}}$, and $E_{i,\text{TS2}}$ are transition state energies and the energy of the intermediate biochemical state of the transitioning motor. These energies can be estimated by taking into account the positions of the transition states and the intermediate state along a well-defined motor stepping reaction coordinate.³¹ The composite stepping rates used in the master equations are given by³²

$$u_{(ij) \rightarrow (i+1,j)} = \frac{u_+ u_{++}}{u_+ + u_{++} + w_- + w_{--}} \quad (11)$$

$$w_{(i+1,j) \rightarrow (i,j)} = \frac{w_- w_{--}}{u_+ + u_{++} + w_- + w_{--}} \quad (12)$$

The above master equations are solved numerically for probabilities of different chemical states of the two-motor assembly using the forward Euler approximation.²⁵ For analyses of two-kinesin dynamics in a static optical trap, bead transport is assumed to begin with the binding of only one of the complex's motors to a microtubule lattice site where the bead is unloaded. This motor can then step forward, and the second motor can bind to the filament depending on the rates of these individual transitions. Bead runs are terminated when both motors detach from the filament. This procedure is modified for predictions of two kinesin behaviors in the force clamp in order to mimic the loading conditions of these experiments.²² In the force clamp assays, beads are first allowed to be transported in a static trap until the applied load reaches a threshold force (F_{Trig}). The load is then changed rapidly to a specified value and held constant via a force feedback algorithm.

To emulate these conditions, numerical calculations were performed to determine the probabilities the complexes would be bound in a particular configuration when the force clamp was first activated. The resulting probabilities are then used as a starting point for a second numerical calculation where the applied load is held constant. For all calculations, average bead velocities and detachment rates are weighted by the probabilities the complexes adopt different bound configurations as described in ref 25.

Note also that our model does not take into account the hard-core exclusion interactions between two motors. The kinesin molecules are allowed to occupy the same site on the microtubule during transport, since it does not affect many dynamic properties of the system and it also simplifies calculations significantly.²⁵ In addition, in our calculations, there are no states where motor-cargo linkers have crossed because the force equilibration assumption allows enough time for these states to relax to untangled conformations.

2.2. Introducing Local Motor Interactions. Both our experimental and theoretical analyses suggest that the two-kinesin complexes will primarily occupy configurations where both motors assume a substantial portion of the applied load when the load exceeds the kinesin's stalling force (7–8 pN), especially in the static trap.²⁵ This behavior is signified by observations of attenuated bead displacement sizes (3–5 nm) in this load regime as compared to kinesin's 8.2 nm step.¹³ Furthermore, the mechanical modeling shows that this behavior can only be produced if the kinesins are bound to closely spaced microtubule lattice sites, since, otherwise, only one kinesin in the complex will bear the load and the bead motion will resemble that of a single-kinesin molecule. Consequently, we assume that kinesins will interact in a way that could affect the transport dynamics when they are bound to microtubule lattice sites that are closer than some critical separation distance (s_c). Specifically, we choose $s_c = 16.4$ nm, which is equal to twice the length of tubulin subunits ($d = 8.2$ nm) from which the microtubules are assembled, since the strongest interaction is expected for kinesins with tubulin subunits to which they are connected; see Figure 1B. Local motor interactions were therefore introduced by assuming that the microscopic transition rates into and out of the states where the motors are positioned at spacing less than or equal to s_c would be altered according to the methods described in the following sections.

2.3. Local Modulation of Motor-Filament Affinities. The influence of local interactions on motor-filament binding and detachment rates was introduced using the treatment that is similar to one implemented by Roos et al.²⁶ to describe the local clustering of kinesins on microtubule filaments, where (attractive) interactions were found to enhance motor-filament binding rate in the absence of load by a factor of γ and attenuate motor detachment rates by a factor of δ when the motors are bound to closely spaced lattice sites on the microtubule.²⁶ Previous estimates of these parameters were $\gamma = 1.5$ – 2.5 and $\delta = 0.3$ – 0.6 , which corresponds to a total local interaction energy (E_{int}) of approximately $1.6 \pm 0.5 k_B T$. An important distinction of our treatment is that the microscopic binding rates ($k_{i \rightarrow (ij)}^{\text{on}}$) and detachment rates ($k_{(ij) \rightarrow i}^{\text{off}}$) in eq 6 are load-dependent, since motor proteins attached to the filament are coupled mechanically.

In the presence of mutual kinesin interactions (E_{int}), the ratio of binding to detachment rates should be modified

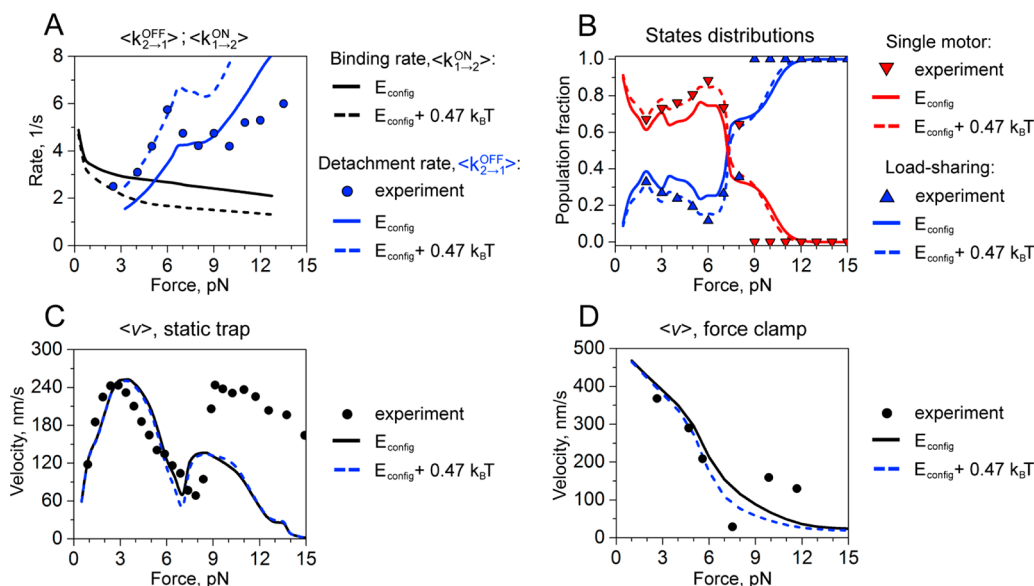


Figure 2. Dynamic properties of two-kinesin complexes in the model without local interactions. (A) Detachment and binding rates: experimentally measured detachment rates are shown by circles, while blue and black curves correspond to calculated values of detachment and binding rates, respectively. (B) Distribution of two-motor load-sharing and single-motor-bound states as a function of the load. The blue and red triangles represent experimentally measured fractions of load-sharing and single-motor states, respectively. Calculated two-motor load-sharing and non-load-sharing states are shown in blue and red, respectively. (C) Force–velocity relationships under static-trap conditions. Symbols correspond to experimental measurements. Black and blue curves describe calculated values for original configurational energies and for modified configurational energies (see text for details), respectively. (D) Force–velocity relationships for force-clamp conditions. Symbols correspond to experimental measurements. Black and blue curves describe calculated values for original configurational energies and for modified configurational energies (see text for details), respectively.

$$\frac{\tilde{k}_{i \rightarrow (i,j)}^{\text{on}}}{\tilde{k}_{(i,j) \rightarrow i}^{\text{off}}} = \frac{k_0^{\text{on}}}{k_0^{\text{off}}} e^{-\beta(\Delta E_{\text{config}} + E_{\text{int}})} \quad (13)$$

where the tilde (\sim) labels the binding and detachment rates affected by these interactions and which yields after taking into account eq 6

$$\frac{\tilde{k}_{i \rightarrow (i,j)}^{\text{on}}}{\tilde{k}_{(i,j) \rightarrow i}^{\text{off}}} = \frac{k_{i \rightarrow (i,j)}^{\text{on}}}{k_{(i,j) \rightarrow i}^{\text{off}}} e^{-\beta E_{\text{int}}} \quad (14)$$

In these expressions, E_{int} is the local nonmechanical motor/motor interaction energy that can be effectively split between binding and detachment as follows:

$$\tilde{k}_{i \rightarrow (i,j)}^{\text{on}} = \gamma k_{i \rightarrow (i,j)}^{\text{on}} \quad (15)$$

$$\tilde{k}_{(i,j) \rightarrow i}^{\text{off}} = \delta k_{(i,j) \rightarrow i}^{\text{off}} \quad (16)$$

Thus,

$$E_{\text{int}} = -k_B T \ln \left(\frac{\gamma}{\delta} \right) \quad (17)$$

2.4. The Effects of Interactions on Collective Motor Stepping. The present model also explicitly takes into account how local interactions between kinesins affect their individual stepping rates along the microtubule. Local interactions induce changes in the free-energy profiles and modify the individual substep transition rates u_{+} , u_{++} , w_{-} , and w_{--} . The substep transitions depend on the difference in the configurational energies of the initial state and the final state for each substep transition. The new substep transition rates \tilde{u}_{+} , \tilde{u}_{++} , \tilde{w}_{-} , and \tilde{w}_{--} , which account for these modifications in the free energy landscapes, are related to the original substep rates as follows:

$$\tilde{u}_{+} = u_{+} e^{-\beta \Delta E_{i, \text{TS1}}} \quad (18)$$

$$\tilde{u}_{++} = u_{++} e^{-\beta(\Delta E_{i, \text{TS2}} - \Delta E_{i, \text{IS}})} \quad (19)$$

$$\tilde{w}_{-} = w_{-} e^{-\beta(\Delta E_{i, \text{TS1}} - \Delta E_{i, \text{IS}})} \quad (20)$$

$$\tilde{w}_{--} = w_{--} e^{-\beta \Delta E_{i, \text{TS2}}} \quad (21)$$

where $\Delta E_{i, \text{TS1}} = E'_{i, \text{TS1}} - E_{i, \text{TS1}}$, $\Delta E_{i, \text{TS2}} = E'_{i, \text{TS2}} - E_{i, \text{TS2}}$, and $\Delta E_{i, \text{IS}} = E'_{i, \text{IS}} - E_{i, \text{IS}}$. The energies of the complex with the motor in the two transition states and the intermediate state are denoted as $E_{i, \text{TS1}}$, $E_{i, \text{TS2}}$, and $E_{i, \text{IS}}$, respectively, and the prime labels indicate the modified energies of these states with stepping interactions. The composite forward and backward transition rates depicted in Figure 1A are then computed similarly as in eqs 11 and 12 and then used in the numerical solutions of master equations.

3. RESULTS AND DISCUSSION

The theoretical framework described above has been used to test how local, nonmechanical interactions between elastically coupled kinesins affect their dynamics in the presence of applied loads. Specifically, the master eqs 2–5 were solved numerically with and without interactions in order to compute (i) average detachment and binding rates, (ii) the probabilities that both motors share their applied load, and (iii) force–velocity (F – V) relationships in both the static trap and the force clamp. Theoretical predictions were then compared with experimental observations to test whether including local interactions improves the agreement between the model predictions and experiments.^{13,25}

Characterizing the impact of local interactions on multiple-kinesin functions ultimately requires reasonable estimates of

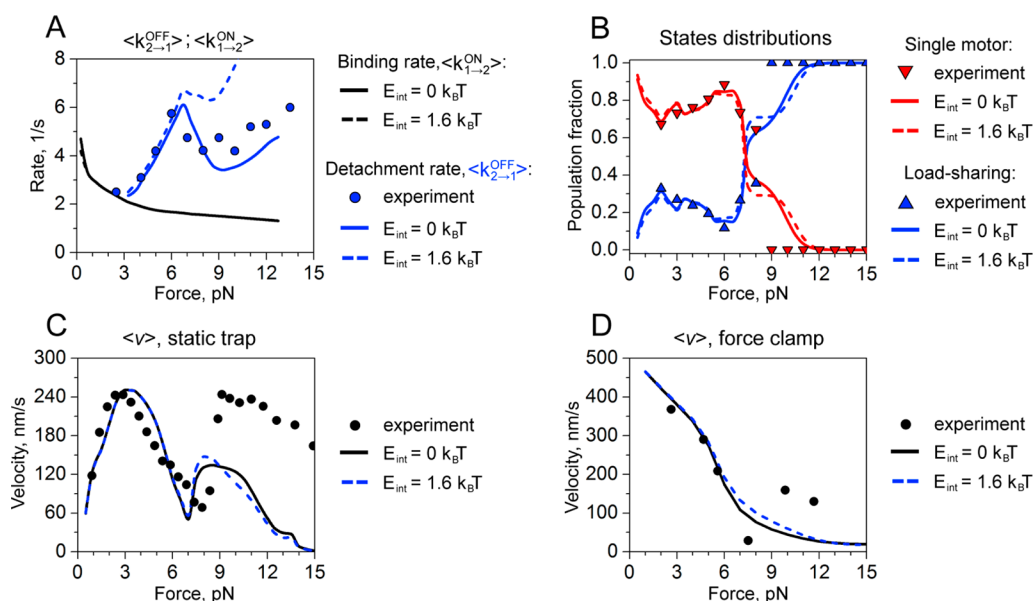


Figure 3. Dynamic properties of two-kinesin complexes in the model where local interactions affect only motor-filament affinities. (A) Detachment and binding rates: experimentally measured detachment rates are shown by circles, while blue and black curves correspond to calculated values of detachment and binding rates, respectively. (B) Distribution of two-motor load-sharing and single-motor-bound states as a function of the load. The blue and red triangles represent experimentally measured fractions of load-sharing and single-motor states, respectively. Calculated two-motor load-sharing and non-load-sharing states are shown in blue and red, respectively. (C) Force–velocity relationships under static-trap conditions. Symbols correspond to experimental measurements. Black and blue curves describe calculated values for original configurational energies and for modified by interactions configurational energies (see text for details), respectively. (D) Force–velocity relationships for force-clamp conditions. Symbols correspond to experimental measurements. Black and blue curves describe calculated values for original configurational energies and for modified configurational energies (see text for details), respectively.

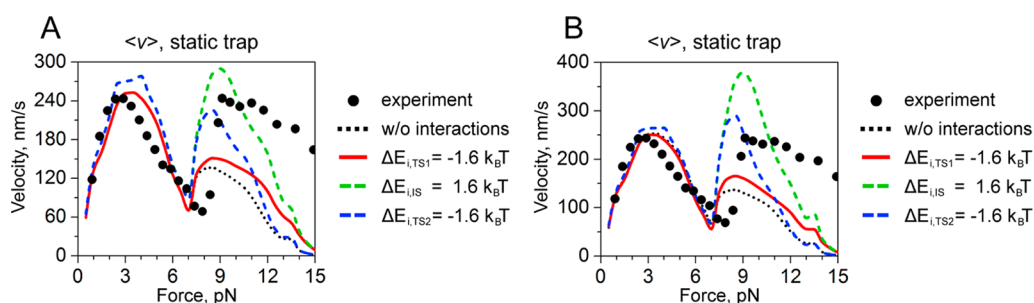


Figure 4. The effect of free-energy profile changes on force–velocity relationships in two-kinesin complexes. (A) The case of no interactions ($E_{\text{int}} = 0$ and the configurational energies of two-motor-bound states are unchanged). (B) The case with local interactions ($1.6 \text{ k}_B T$) and with the configurational energies of two-motor-bound states increased by $0.47 \text{ k}_B T$. Symbols correspond to experimental measurements; calculated curves describe lowering the free energy of the first transition state (red); or second transition state (blue), or increasing the energy of intermediate state (green).

how the free energy (E_{config}) of a complex changes when it transitions between different filament-bound conformations due to the binding, detachment, and stepping of individual motors within a complex. These energies were previously parametrized via analyses of single-motor elasticity measurements and a mechanical modeling procedure that computes a complex's configurational energy according to eq 1. While this treatment provides a reasonable approximation of the configuration-dependent balance of forces between the motors (see the Supporting Information in ref 25), E_{config} is purely determined by the axial force–extension properties of the kinesin-bead linkages, and it could potentially neglect the role of other energetic terms in eq 1 that arise due to the binding geometries of the complexes (e.g., rotational and torsional strain contributions to E_{config}). Consequently, we first re-evaluated this treatment in order to characterize the relative

significance of these factors (Figure 2). From this basis, the impact of local, nonmechanical interactions on multiple kinesin dynamics was then characterized using the model framework described in the Theoretical Methods section (Figures 3 and 4).

3.1. Evaluation of the Model without Local Interactions. Comparisons between predictions of the model without local interactions and experimental optical trapping data are displayed in Figure 2. As discussed in ref 25, this form of the model is able to capture key features found in the optical trapping data including the nonmonotonic force dependence of the detachment rate, $\langle k_{2 \rightarrow 1}^{\text{off}} \rangle$, describing how rapidly complexes detach partially from the filament (Figure 2A), and the analogous rapid increase in average two-kinesin velocities that are observed in the static trap when the load increases above kinesin's stalling force (7.6 pN in our experiments¹³); see

Figure 2C. Moreover, the model also reproduces experimentally measured trends in the static trap describing the load-dependent probability that a complex will associate with the microtubule in configurations where both kinesins share their applied load (Figure 2B). Here, load-sharing configurations are assumed to correspond to those where a complex's secondary load-bearing motor (the trailing motor) bears at least 25% of the total applied load imposed on the bead. With this criterion, the probability that both kinesins share their load increases initially until the load exceeds 2 pN but then slowly decreases with increasing load until the load approaches 7 pN, as is found in the data. Importantly, despite the liberal definition of load-sharing states, this probability never exceeds 35% throughout all force regimes. However, the complexes are found experimentally and predicted theoretically to primarily transport bead via load-sharing configurations above kinesin's stalling force (see Figure 2B). Overall, these results are significant, since they illustrate that the unique transport behaviors that arise in the static optical trap due to the spatiotemporal properties of applied load in these experiments¹³ can be recapitulated by the model that only accounts for mechanical interactions associated with elastic motor coupling.

Despite the ability to reproduce dynamic trends, there are still some significant differences between the model predictions and the optical trapping data. Two-kinesin velocities in both the static trap and force clamp are clearly underestimated by the model at high applied loads, again, even when three motors are assumed to be present on the bead's surface.²⁵ Thus, it is important to consider a possible role of local motor interactions in these responses. Measured $\langle k_{2 \rightarrow 1}^{\text{off}} \rangle$ transition rates (Figure 2A), which are average dissociation rates for two-motor-bound complexes, also exceed the model predictions at low applied loads (<7 pN) but fall below their predicted values at high applied loads, yielding a more pronounced nonmonotonic force dependence. While the latter result indicates that local interactions may enhance cargo-filament affinities at high applied loads (as discussed below), the difference between measured and calculated $\langle k_{2 \rightarrow 1}^{\text{off}} \rangle$ rates at low applied loads suggests that the original form of the model overestimates the free energy that a complex gains when both of its motors are anchored to the filament. For example, the agreement between theory and experiments can be improved greatly at low applied loads by simply raising E_{config} for each two-motor-bound state enumerated by $0.47 k_{\text{B}}T$ (Figure 2A). We suggest that this result signifies that mechanical factors other than the elastic coupling between motors can influence multiple-kinesin-filament affinities by altering the free energy difference between two-motor-bound configurations and single-motor-bound configurations. While these changes could potentially stem from various mechanical constraints (e.g., rotational and torsional strains) as well as nonmechanical factors that are not included in eq 1, the small energetic contributions of such factors to E_{config} can have an appreciable influence on $\langle k_{2 \rightarrow 1}^{\text{off}} \rangle$ transition rates. Nevertheless, including these energy terms does not modify motor stepping rates in the model, and it has little effect on calculated two-kinesin velocities in both the static optical trap and the force clamp, since these properties already remain dominated by the absence of load-sharing between the two motors. Finally, since this new treatment of E_{config} provides an overall improved fit to the two-kinesin data at low applied loads, this form of the model was employed for all subsequent analyses of local interactions between the kinesins.

3.2. Interactions Affecting Motor-Filament Affinities.

We next examined the impact of local, nonmechanical interactions between kinesins on their binding and detachment transition rates, load-sharing probabilities, and average velocities in the optical trap using the framework developed by Roos et al.²⁶ to analyze the clustering of kinesins on microtubules in motor-filament binding assays. As described in the Theoretical Methods section, microscopic binding ($k_{i \rightarrow (ij)}^{\text{on}}$) and detachment rates ($k_{(ij) \rightarrow i}^{\text{off}}$) in the master equations were assumed to be enhanced by the factors γ and δ only when the motors were bound to microtubule lattice sites that are spaced by less than $s_c = 16.4$ nm. With this treatment, average partial detachment rates $\langle k_{2 \rightarrow 1}^{\text{off}} \rangle$ are predicted to follow the experimental trend much more closely compared to the model without interactions when $\gamma = 2$ and $\delta = 0.4$ (Figure 3A). Importantly, these values fall in the range of microscopic rate enhancements reported by Roos et al.,²⁶ and according to eq 17 yield a small net interaction energy corresponding to $1.6 k_{\text{B}}T$.

Interestingly, plots of the average binding transition rates ($k_{1 \rightarrow 2}^{\text{on}}$) as a function of the applied loads show that motor-filament attachment transitions are largely unaffected by the introduction of local interactions into the model (Figure 3A). This insensitivity can be explained by the fact that the changes in the complex's strain energy are much larger than the amplitude of interaction energies specified by eq 17 when the motors transition from a single-motor-bound configuration to load-sharing configurations where the motors are spaced closely on the filament and have the potential to interact.²⁵ For example, when the total applied load is 4 pN, calculated values for ΔE_{config} associated with such transitions range between 15 and $26 k_{\text{B}}T$, since they require forward cargo displacements against the applied load of the trap in order for the newly bound motor to take its portion of the load. The strong energetic preference for a free motor to bind to a microtubule lattice site far behind its load-bearing partner described in refs 25 and 22 is therefore retained even when local interactions are included in the model. Accordingly, predictions of load-sharing probabilities (Figure 3B) and average two-kinesin F - V relationships (Figure 3C and D) are largely unchanged at all applied loads. As a result, although this treatment results in improved predictions of the complex's load-dependent detachment behavior, this form of local interactions has little effect on the ability for the motors to share their load and is insufficient to capture the apparent synergistic behaviors indicated by the large two-kinesin velocities that are found above 7 pN in the static trap and the force clamp.

3.3. Combined Affinity and Stepping Rate Enhancements. The above results imply that local interactions may affect both kinesin-microtubule-filament affinities and stepping rates at high applied loads. This is also more consistent with the idea that local changes in free energy profiles should modify all related chemical transitions. Consequently, we explored whether combining the treatment of Roos et al.²⁶ with analogous modifications to the free energy profile along kinesin's stepping pathway would be sufficient to capture the high velocities produced by the two-kinesin complexes above 7 pN in both optical trapping experiments. To do so, we first examined the sensitivities of the two-kinesin velocities to reductions of the free energy of the transition states associated with the first (TS1) or the second (TS2) substep transition enumerated in the Fisher-Kim model by $1.6 k_{\text{B}}T$, as specified by eqs 18–21. The effect of increasing the energy of the

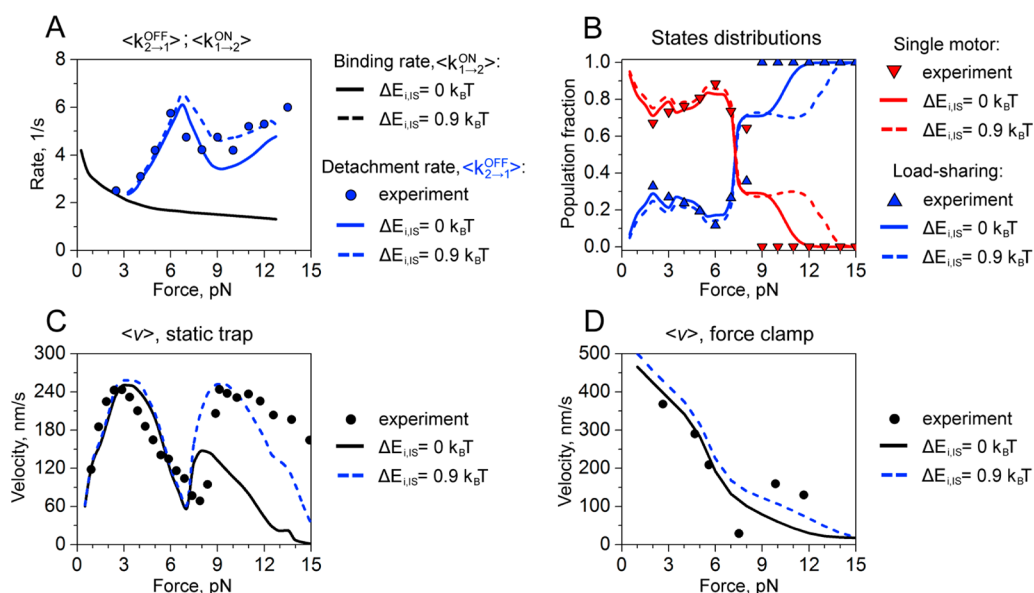


Figure 5. Dynamic properties of two-kinesin complexes in the model where local interactions affect motor-filament affinities and stepping rates. (A) Detachment and binding rates: experimentally measured detachment rates are shown by circles, while blue and black curves correspond to calculated values of detachment and binding rates, respectively. (B) Distribution of two-motor load-sharing and single-motor-bound states as a function of the load. The blue and red triangles represent experimentally measured fractions of load-sharing and single-motor states, respectively. Blue and red curves show calculated two-motor load-sharing and single-motor state probabilities, respectively. (C) Force–velocity relationships under static-trap conditions. Symbols correspond to experimental measurements. Black and blue curves describe calculated values for original configurational energies and for modified by interactions configurational energies (see text for details), respectively. (D) Force–velocity relationships for force-clamp conditions. Symbols correspond to experimental measurements. Black and blue curves describe calculated values for original configurational energies and for modified configurational energies (see text for details), respectively.

intermediate state (IS) along the kinesin stepping pathway by $1.6 k_B T$ was also examined in a separate calculation. Overall, each of these treatments produces substantial increases in two-kinesin velocities at high applied loads, particularly when the energies of TS2 and IS are altered, since the u_{++} substep transition is rate limiting at most loads for kinesin using the Fisher–Kim model (Figure 4). In fact, average two-kinesin velocities are even overapproximated when local interactions are assumed to increase both the free energy of IS and motor–filament affinities as described above (Figure 4B). Lowering TS1 by $1.6 k_B T$ generally produces more modest velocity enhancements but tends to enhance two-kinesin velocities at very large loads (>12 pN), since the u_+ transition becomes rate limiting in this load regime. In sum, these results generally show that small scale interactions of a few $k_B T$ of energy can indeed have a large effect on two-kinesin velocities, specifically at high applied loads.

Overall, we find, empirically, that the best agreement between the optical trapping data and the composite form of the model where local interactions are assumed to affect both motor–filament affinities and stepping rates is achieved when $s_c = 16.4$ nm, $\gamma = 2$, and $\delta = 0.4$, as in Figure 3, and when local interactions are assumed to modify the free energy profile of kinesin’s stepping pathway using: $\Delta E_{i,TS1} = -2.2 k_B T$, $\Delta E_{i,IS} = 0.9 k_B T$, and $\Delta E_{i,TS2} = 0.25 k_B T$ (Figure 5). These alterations have little effect on two-kinesin velocities at the low applied load, as well as on $\langle k_{2 \rightarrow 1}^{\text{OFF}} \rangle$ transition rates and on load-sharing state distributions at all loads. However, the rapid increase in two-kinesin velocities at 7 pN that is observed in the static trap is much more pronounced in the composite model, yielding F – V relationships that are much closer to the experimental trends. Moreover, the model retains key differences between the two-kinesin velocities measured in the static trap and in the force

clamp. The applied load of the static trap varies spatially and temporally depending on the trap’s spring constant and how the complexes move under load. In addition, the applied load decreases after detachment events in the static trap where only one of the two kinesins is released from the filament, since these events are accompanied by rearward bead displacements.¹³ Both our experimental and prior theoretical analyses suggest that this behavior reduces the probability that a two-kinesin complex will remain bound via a single-motor linkage significantly at high applied loads ($F_{ap} > 7$ pN). This spatiotemporal “filtering” of two-kinesin configurations also constitutes one of the main reasons the two-kinesin F – V relationship in the static trap is found to exhibit an unusual nonmonotonic dependence with or without interactions. The position of the trapping laser is updated to maintain a constant applied load in the force clamp. Bead velocities in the force clamp are therefore influenced by mixtures of single- and two-motor-bound states at all applied loads. Consequently, interactions result in larger velocity enhancements in the static trap compared to the force clamp when loads exceed kinesin’s stalling force. However, the velocities below 7 pN are quite similar in both experiments, and tend to closely follow the single-kinesin F – V curves in each case. This behavior occurs due to the preference for the complexes to transition from single-motor-bound states to two-motor states where both motors are positioned far apart on the microtubule under both loading conditions, since ΔE_{config} for transitions into load-sharing states is prohibitively large. These effects tend to dominate two-kinesin dynamics, since they determine whether a multiple motor complex is capable of generating configurations where the motors are able to interact.

Table 1. Model Parameters and Their Estimated Values

parameter	estimated/ measured values	reference
π_0 , unloaded binding rate	4.7 s^{-1}	ref 30
ϵ_0 , unloaded detachment rate	0.312 s^{-1}	estimated from ref 25
u_{++}^0 , unloaded stepping rate	$1.59 \times 10^{14} \text{ s}^{-1}$	estimated from ref 25, see also explanations in the text
u_{+-}^0 , unloaded stepping rate	61.7 s^{-1}	estimated from ref 25, see also explanations in the text
w_{-+}^0 , unloaded stepping rate	0.654 s^{-1}	estimated from ref 25, see also explanations in the text
w_{--}^0 , unloaded stepping rate	$1.69 \times 10^9 \text{ s}^{-1}$	estimated from ref 25, see also explanations in the text
γ , binding enhancement factor	2.0	ref 26
δ , affinity factor	0.4	ref 26

4. SUMMARY AND CONCLUSIONS

We have developed a discrete-state stochastic model of multiple kinesin dynamics that accounts for both (i) configuration-dependent mechanical interactions between motors that influence how forces are distributed between motors that are bound to the same cargo and (ii) local, nonmechanical interactions that have been found previously to enhance kinesin–filament affinities when the motors are bound to neighboring microtubule lattice sites. Although the mechanical coupling between motors still dominates most multiple kinesin behaviors, including the effects of weak local interactions ($<2 k_B T$) in transition rate expressions, describing how multiple kinesins bind to, detach from, and step along microtubules is shown to improve the model's agreement with experimentally determined force–velocity relationships produced by structurally defined kinesin complexes. In particular, this adaptation provides much better agreement between theoretical and the large experimental velocities observed in an optical trap at loads that exceed kinesin's stalling force, implying that the stepping rates of colocalized kinesins can be enhanced significantly by local, nonmechanical interactions even if their interaction strength is relatively small.

Although the interaction energy scales identified in this work are very similar to those determined from analyses of kinesin binding and detachment kinetics in the absence of load,²⁶ the origin of local interactions between kinesins is still unknown. The mutual attraction suggested by both studies could potentially stem from various sources including local electrostatic interactions that are more or less contact dependent, yielding an interaction distance that is comparable to the size of the motor itself ($d = 8.2 \text{ nm}$). The microtubule lattice structure has been found to be altered locally by kinesin associations,³³ and such effects indicate that local interactions could also be mediated through the microtubule filament. However, despite their source, there is now increasing evidence that mutual interactions between kinesins can enhance motor–filament affinities. To our knowledge, this study provides the first evidence that these effects can alter collective kinesin stepping rates, yielding larger velocities in the presence of superstalling loads.

One may expect that local interactions would result in more synergistic multiple motor functions due to improved coordination between the motors. The present analyses suggest, however, that other factors dominate multiple kinesin

dynamics by determining whether a complex can adopt bound geometries where the motors can experience nonmechanical interactions. The free energy changes associated with binding transitions into states where the motors are positioned closely and interact mechanically via load sharing are much higher than their nonmechanical interaction energies. Consequently, multiple kinesins still tend to transport their cargos via configurations where only one motor bears the applied load. Furthermore, the loads on cargos driven by multiple kinesins increase rapidly compared to the rate that the geometry evolves from single-motor to load-sharing transport modes.²⁵ Given this property and other spatiotemporal “filtering” effects, motor detachment will generally occur before the motors are given a chance to interact mechanically or nonmechanically. We finally note that this circumstance could change for different types of motors that have lower stalling forces and whose velocities change more sensitively to applied loads. In this case, a team of motors could potentially adopt load-sharing states much more readily, creating conditions where local interactions influence multiple motor force production and stepping dynamics over a larger range of applied loads. In this way, the microscopic details of stepping mechanisms for each motor can potentially determine the overall impact of local interactions and further distinguish how groups of different motor types function collectively.

AUTHOR INFORMATION

Corresponding Author

*E-mail: diehl@rice.edu (M.R.D.); tolya@rice.edu (A.B.K.).

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was supported by grants from the National Science Foundation (MCB-0643832), the National Institute of Health (1R01GM094489-01), and the Welch Foundation (C-1559 to A.B.K. and C-1625 to M.R.D.).

REFERENCES

- (1) Hirokawa, N. *Science* **1998**, *279*, 519–526.
- (2) Vale, R. D. *Cell* **2003**, *112*, 467–480.
- (3) Ashkin, A.; Schutze, K.; Dziedzic, J. M.; Euteneuer, U.; Schliwa, M. *Nature* **1990**, *348*, 346–348.
- (4) Wang, Z.; Khan, S.; Sheetz, M. P. *Biophys. J.* **1995**, *69*, 2011–2023.
- (5) Kojima, H.; Muto, E.; Higuchi, H.; Yanagida, T. *Biophys. J.* **1997**, *73*, 2012–2022.
- (6) Coy, D. L.; Wagenbach, M.; Howard, J. J. *Biol. Chem.* **1999**, *274*, 3667–3671.
- (7) Visscher, K.; Schnitzer, M. J.; Block, S. M. *Nature* **1999**, *400*, 184–189.
- (8) Toba, S.; Watanabe, T. M.; Yamaguchi-Okimoto, L.; Toyoshima, Y. Y.; Higuchi, H. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 5741–5745.
- (9) Kulić, I. M.; Brown, A. E. X.; Kim, H.; Kural, C.; Blehm, B.; Selvin, P. R.; Nelson, P. C.; Gelfand, V. I. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 10011–10016.
- (10) Ally, S.; Larson, A. G.; Barlan, K.; Rice, S. E.; Gelfand, V. I. *J. Cell Biol.* **2009**, *187*, 1071–1082.
- (11) Holzbaur, E. L. F.; Goldman, Y. E. *Curr. Opin. Cell Biol.* **2010**, *22*, 4–13.
- (12) Rogers, A. R.; Driver, J. W.; Constantinou, P. E.; Jamison, K. D.; Diehl, M. R. *Phys. Chem. Chem. Phys.* **2009**, *11*, 4882–4889.
- (13) Jamison, K. D.; Driver, J. W.; Rogers, A. R.; Constantinou, P. E.; Diehl, M. R. *Biophys. J.* **2010**, *99*, 2967–2977.

- (14) Leduc, C.; Ruhnnow, F.; Howard, J.; Diez, S. *Proc. Natl. Acad. Sci. U.S.A.* **2007**, *104*, 10847–10852.
- (15) Ali, Y. M.; Lu, H.; Bookwalter, C. S.; Warshaw, D. M.; Trybus, K. M. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 4691–4696.
- (16) Leduc, C.; Pavin, N.; Julicher, F.; Diez, S. *Phys. Rev. Lett.* **2010**, *105*, 128103.
- (17) Ali, Y. M.; Kennedy, G. G.; Safer, D.; Trybus, K. M.; Sweeney, L. H.; Warshaw, D. M. *Proc. Natl. Acad. Sci. U.S.A.* **2011**, *108*, E535–E541.
- (18) Klumpp, S.; Lipowsky, R. *Proc. Natl. Acad. Sci. U.S.A.* **2005**, *102*, 17284–17289.
- (19) Campas, O.; Kafri, Y.; Zeldovich, K. B.; Casademunt, J.; Joanny, J.-F. *Phys. Rev. Lett.* **2006**, *97*, 038101.
- (20) Muller, M. J. I.; Klumpp, S.; Lipowsky, R. *Proc. Natl. Acad. Sci. U.S.A.* **2008**, *105*, 4609–4614.
- (21) Kunwar, A.; Mogilner, A. *Phys. Biol.* **2010**, *7*, 016012.
- (22) Jamison, K. D.; Driver, J. W.; Diehl, M. R. *J. Biol. Chem.* **2012**, *287*, 3357–3365.
- (23) Beeg, J.; Klumpp, S.; Dimova, R.; Gracia, R. S.; Unger, E.; Lipowsky, R. *Biophys. J.* **2008**, *94*, 532–541.
- (24) Hendricks, A. G.; Epureanu, B. I.; Meyhöfer, E. *Physica D* **2009**, *238*, 677–686.
- (25) Driver, J. W.; Jamison, K. D.; Uppulury, K.; Rogers, A. R.; Kolomeisky, A. B.; Diehl, M. R. *Biophys. J.* **2011**, *101*, 386–395.
- (26) Roos, W. H.; Campàs, O.; Montel, F.; Woehlke, G.; Spatz, J. P.; Bassereau, P.; Cappello, G. *Phys. Biol.* **2008**, *5*, 046004.
- (27) Vilfan, A.; Frey, E.; Schwabl, F.; Thormählen, M.; Song, Y.-H.; Mandelkow, E. *J. Mol. Biol.* **2001**, *312*, 1011–1026.
- (28) Seitz, A.; Surrey, T. *EMBO J.* **2006**, *25*, 267–277.
- (29) Driver, J. W.; Rogers, A. R.; Jamison, K. D.; Das, R. K.; Kolomeisky, A. B.; Diehl, M. R. *Phys. Chem. Chem. Phys.* **2010**, *12*, 10398–10405.
- (30) Leduc, C.; Campas, O.; Zeldovich, K. B.; Roux, A.; Jolimaître, P.; Bourel-Bonnet, L.; Goud, B.; Joanny, J.-F.; Bassereau, P.; Prost, J. *Proc. Natl. Acad. Sci. U.S.A.* **2004**, *101*, 17096–17101.
- (31) Fisher, M. E.; Kim, Y. C. *Proc. Natl. Acad. Sci. U.S.A.* **2005**, *102*, 16209–16214.
- (32) Kolomeisky, A. B.; Fisher, M. E. *Annu. Rev. Phys. Chem.* **2007**, *58*, 675–695.
- (33) Krebs, A.; Goldie, K. N.; Hoenger, A. *J. Mol. Biol.* **2004**, *335*, 139–153.