ESSESA: An Expert System for Structure Elucidation from Spectra. 5. Substructure Constraints from Analysis of First-Order ¹H-NMR Spectra

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This paper describes the knowledge base of first-order ¹H-NMR spectral analysis and the interpretation program for analysis of ¹H-NMR spectral data in ESSESA. Logical representation and the production system rules concerning analysis of ¹H-NMR spectra are discussed as well as inferential models useful in firstorder ¹H-NMR spectral analysis. The unsaturation and the atomic composition of an unknown compound as well as the substructure constraints from the analysis of infrared spectrum, are passed to the interpretation program that develops the substructure constraints from its analysis of first-order ¹H-NMR spectral data by inference from the knowledge base of spectral analysis. The knowledge base contains 531 substructures.

INTRODUCTION

The elucidation of the structure of organic compounds based on spectroscopic methods, like ¹H-NMR, ¹³C-NMR, IR, mass spectrometry, and UV/vis spectroscopy, is still essentially an empirical procedure. This is due to the fact that the derivation of constitution from the spectra^{1,2} contains steps whose complexity prohibits formalization. Therefore the practical application of automatic interpretation programs³ is far more limited than sometimes postulated in the literature.4 Artificial intelligence, especially the expert systems method deriving rules out of a knowledge base, is generally applicable. ESSESA is an expert system for structure elucidation from spectral analysis.⁵⁻⁸

One of the most powerful tools for the determination of an unknown organic structure is the ¹H-NMR spectroscopy. From the first-order ¹H-NMR spectrum the types of hydrogen and structural environment in a structure can be derived. In some cases, the information from ¹H-NMR spectrum can lead to a unique structure, but in many cases there remains a large number of structural possibilities. In ESSESA the complete structure of a molecule is obtained by computer when data from several spectroscopic techniques are used simultaneously. The analysis of first-order ¹H-NMR spectral data gives the secondary substructure constrains which will be used in the generation of the complete structure.

KNOWLEDGE BASE OF FIRST-ORDER 1H-NMR SPECTRA ANALYSIS

The conventional approach taken by chemists to first-order ¹H-NMR spectral interpretation is based upon models, often simplified, of the physical processes underlying resonance and the resulting spectral absorption. The physical models can be used to relate specific spectral signals to particular structural components of the molecule. Usually, there are several factors that together determine the detailed characteristics (such as chemical shift, multiplicity, and area of peak) of a first-order ¹H-NMR spectral signal and which are often in some sort of a hierarchical relationship that defines their relative importance. The initial analysis of a spectral signal may identify the presence of a specific type of hydrogen in the unknown molecule, and more detailed analysis of the form of the signal may determine aspects of the larger environment of that type of hydrogen.

The chemist's knowledge of first-order ¹H-NMR spectral analysis that has been incorporated into ESSESA is encoded in the form of spectral feature-substructure relationship rules written in PROLOG, which comprise the knowledge base for ¹H-NMR spectral analysis.

If a set of specific ¹H-NMR absorption peaks given by an unknown compound is Wh, such that

$$\mathbf{W}_{h} = [\mathbf{W}_{hi}(\mathbf{c}, \mathbf{m}, r)] \quad i = 1, 2, 3, ..., n$$
 (1)

here c is the chemical shift, m is the multiplicity, and r is the area of the peak. The set of substructures in the knowledge base of first-order ¹H-NMR spectra analysis is

$$S_h = [S_{hi}]$$
 $j = 1, 2, 3, ..., k$ (2)

The set of specific absorption peaks that correspond to a substructure S_{hi} is W_{shi} that is expected to be

$$\mathbf{W}_{shi} = [W_h(c1, m, r), ..., W_h(c2, m, r)]$$
 (3)

And if the set of substructures from IR spectral analysis is S_{ir}

$$S_{ir} = [S_1, S_2, ...Sp]$$
 (4)

In order to analyze a first-order ¹H-NMR spectrum of an unknown compound, it is necessary to pick out the subset $S_{\rm hh}$ from the set $S_{\rm h}$, such that

$$\exists \ W_{\mathrm{h}i} \ \exists \ W_{\mathrm{sh}j} \ \exists \ \mathbf{S}_{ir} \{ W_{\mathrm{h}i} \subseteq W_{\mathrm{sh}j} \wedge S_j \subseteq \mathbf{S}_{ir} \} \longrightarrow \mathbf{S}_{\mathrm{h}\mathrm{h}} \supseteq S_j \ \ (5)$$

According to this procedure, the construction of the knowledge base for first-order ¹H-NMR spectral analysis requires that the logical representation formula (eq 3) of the set of substructures Sh be found and encoded in PROLOG rules. In ESSESA, the information in eq 3 is expressed by the production system rule—i.e., the spectral feature substructure relationships. Given a sufficiently large number of spectra, or alternatively the appropriate information derived from published observations, it is possible to develop the ability to associate certain absorption peaks with the corresponding hydrogen types and their structural environment. Thus a set of rules can be developed to derive the

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substructure	chem. shift mul	multiplicity a	area IR con	IR constraint no.		substructure	chem. shift	multiplicity	area	IR constraint
CH ₃ C ←	0.1-2.2	_	3	5.	S >CHC	>CH CH ₂ C(=0)0-	2.2-3.0	2	7	-C(=0)0-
CH ₂ CH ₂ -	0.6 - 1.6	3	3	53	_	CH ₃ CH ₂ C(=0)0-	2.3 - 3.1	4	7	-C(=0)0-
CH,CH <	Ĥ	2	3	54	/10	>CCH,C(=0)0-	2.5 - 3.0	_	2	-0(0=0)
CH ₂ C=C-	- 1	_	3	55	CH,CH,N <	\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	2.4 - 3.4	4	2	> N -
$CH_{\alpha}C(=0)$	- 1	_	3 -C(=0)-	56		$-CH_2CH_2N <$	2.4 - 3.8	3	7	> N -
CH ₂ S-	1.8-2.8	-	3 -S-	57		H ₂ N <	2.4 - 2.9	7	7	-N<
CH ₂ C(=0)0-		_	3 –C(=0)0-	- 58	,,,	× Z	2.4 - 3.5	_	7	> N -
CHAR		1	3 Ar	59	CH ₃ CH ₂ NH-	-HN-	2.4 - 3.4	5	7	-MH-
CH,N<	1	1	3 -N-	09		-CH ₂ CH ₂ NH-	2.4 - 3.8	4	7	-NH-
CH,OC(-)=0	1	_	3 -OC(=0)-	- 19	,	H,NH-	2.4-2.9	3	7	-NH-
CHO-		. —	3 -0-	62		-HN	2.4-3.5	7	7	-HN-
CH,C=C		. –		69	·	-CH,CH,NH,	24-38	ı v	. 6	HN-
CH.CH	1.6 2.0	, () 	25		>CHCH,NH,	- 1	4	0	-NH2
CH,NH-	2.5-2.7	2 (3 -NH-	65	,,,	NH.	2.4 - 3.5	· m	7	-NH,
N=U-HU	1.8-2.0	ı —	3 C=N	99		LAr	2.4 - 3.0	4	2	Ār
CH,CH=N	1.8-1.9	2	3 C=N	<i>L</i> 9		H,Ar	2.4 - 3.0	3	7	Ar
CH,CH,-	0.9-2.1	1 0		89		H,Ar		5	2	Ar
>CHCH.CH <	1.0-1.5	۳ (10	69	///	Ār	2.9-3.7	-	2	Ar
TOTAL CHI	11-17	4	1 C	02		V Z	29-33	_	. 6	× Z -
· »ZTTO	1.1-7.1	. ,	, c	71		-NH-	29-33	. 6	2	-NNH-
· CITATION · CHUITA	1.7 - 7.1	1 V	1 (CL		HZ	29-33	۳ ،	· C	۱ ۷
	1.2 -2.1	v	1 C	77	-	NHCH, NH-		, cr	, c	- 1
	4.2 2.1 1.2—2.1	٥	4 C			-NHCH-NH-	2.0 -3.3	7	1 C	-HN-
-CH2CH3CA	4.7—2.1	n -	7 (- 1		#12 112 C!	2.1-3.7	† <	1 C	1111
VCH2C*	6.1–7.1		7 (C/ 2F		12C1	$\frac{3.1-3.7}{2.1-4.0}$	† ~	4 6	
CHCH2CH2-	7.1 4.1	t -	ر ا	5, 5,			ŀ	n c	1 C	
	3.1-4.1	1 (81			ł	ı –	1 C	
C-CCH2MI	3.1-4.1	1 "		9/	. !	O(0=)C(H.S	3.0-3.0) C	-U(= 0)0-
ZIN ZINO HO		4		08	'	HS'HJJ(=0)0-	3.0-3.9	5	1 6	-C(=0)0
O OZIOSIO	19-28	۰, ۲۰		ã ∞	١	O(0=)CCH3N <	1	-	2	-0(0=0)
>HJ*HJU=U	ſ	, c		. 8	,	-O(O=)CCH3NH-	1	2	2	-C(=0)0
・ロングロントン	1.9 3.0	1 -		8.48	,	-O(O=)CCH;NH;	3.1-4.0) (r	1 %	-0(0=))-
NO HO	2.0-3.0	. 4		× ×	1	O(O=)CCH3C(=O)O-	3.2-3.5	· -	2	-C(=0)0-
CHJ:CHJ	2.0-3.0	۰, ۲۰	Z Z	98	⋖	S	3.2-4.0	-	7	Ar
NJ.HJ.AJ.	2.0-3.0	o 6		87		SH	3.2-4.0	2	2	Ar
VCH2CN	2.0-3.0	. —		88		(H ₂ O –	3.3-4.3	3	7	-0-
CH ₂ CH ₂ S-	2.1-3.0	4	2 -S-	68	^	H,0-	3.4-4.4	2	7	-0-
-CH,CH,S-	2.4-2.9	ω,	2 -S-	06		-0-	3.4-4.2	-	7	-0-
>CHCH.S-	2.4 - 3.1	2	2 -S-	91		-СН, СН ,ОН	3.4-4.4	4	7	НО-
>CH3C	2.4 - 3.1	-	2S-	92		Н ,ОН	3.4-4.2	3	7	Н0-
-CH,CH,SH	2.4-2.9	. 4	2 SH	93	,,,	HO	3.4-4.2	2	2	Н0-
>CHCH3H	2.4-3.1	· (r)		76		-CH ₂ CH ₃ N <	2.4 - 3.8	3	2	-N-
>CCH,SH	2.4-3.1	2	2 SH	95	1	-CH2CH2NH-	2.4 - 3.8	4	2	-NH-
$-CH_2CH_2C(=0)-$	2.4-3.5	3		96	1	-CH ₂ CH ₂ NH ₂	2.4 - 3.8	5	7	$-NH_2$
>CCH ₂ C(=0)-	2.2-3.2	-	2 -C(=0)-	16		0=CCH ₂ C=0	3.3 - 3.9	-	2	0=0-
$CH_2C(=0)$	2.3 - 3.0	4	2 - C(=0)	86	1	-(0=)CCH2CH=0	3.3-3.9	2	7	0=0-
>CH CH ₂ C(=0)-	2.2-2.8	2	2C(=0)-	66	1	-0(0=)C-CH2CN	3.4 - 3.6	-	7	CN, -C(=0)0-
-CH2CH2CH=0	2.2-3.5	4	2 -CH=0	001		×	3.4-4.0	1	7	Ar
>CH CH 2CH=0	2.2-2.8	3	2 -CH=0	101	1 ArCH2NH-	NH-	3.4 - 4.0	2	7	Ar, -NH-
>C CH 2CH=0	2.2-3.2	2	2CH=0	102	•	NH_2	3.4 - 4.0	3	7	Ar, $-NH_2$

Table 1 (Continued)

9	substructure	chem shift	multiplicity	area	IR constraint	٤	substructure	chem shift	multiplicity area	ea IR constraint
			Cuandama.		1					
212	>CH(-CH ₂)CHCN		4 (<u>S</u> 8	462	_ '	2.8-3.4	۰.	HS-
213	→C(-CH ₂)CHCN		.n. (٠.	Z	263		2.8-3.4	4 r	-S-
214	(-CH)CHCN	l	n (٠,	-CN	907		2.8-5.4	n c	HS-
212	≯C(>CH)CHCN	ļ	7 [٠ .		797	ز م	2.8-3.4	· · ·	-S-
216	(CH ₃)CHCN	2.6-2.8	- <			897	, ,	2.8-3.4	4 6	HS-
210		ن د ا	v t			607 07.0		2.6-3.4	0 z	181
210	/CH(-CH2)CHCH-O		7		-CII-	27.7		2.6-3.4	† ¢	-S
220	-CH:(CH:)(HC(=0)0-	- 1	۷ ع	-	C(=0)0- -((=0)0-	277	>CH(>C)CHS	2.8-3.4	J (r	HS-
22.	-0(0=)CHC(H3)KHC/<	- 1	o vr	-	-0(0=))-	273	>CH(>C)CHO!!	37-47	00	- d
222	>C(CH3)CHC(=0)0-	- 1	, 4	-	-0(0 =))-	274		3.2-4.2	ım	HO- I
223	(-CH ₂) CH C(=0)0-	- 1	. 5	-	-0(0=)) -0(0=)	275	//(3.3-4.5	4	-
224	(>CH)CHC(=0)0-		m	_	-C(=0)0-	276		3.3-4.5	٠,	HO-
225	>CH(>C)CHC(=0)0-	1	2 2	-	-0(0=))-	277		3.2-4.2	'n	-
226	>C(-CH ₂)CHC(=0)0-	3	'n	-	-0(0=) 2	278	_	3.2-4.2	9	HO- I
227	$(CH_3)CHC(=0)0-$		7	_	-0(0=)J-	279	_	3.3-4.5	ε.	-0-
228	$\rightarrow C(-CH_2)CHC(=0)-$	2.6 - 3.4	3	1	-C(=0)-	280	_	3.3-4.5	4	H0- 1
229	$\rightarrow C(-CH_2)CHCH=0$	2.6 - 3.4	4	-	O=H⊃−	281	/ I	3.3-4.1	4	-C(=0)0-
230	(CH ₃) CH Ar		7	_	Ar	282	1	3.3 - 4.1	33	0=0-
231	$-CH_2(CH_3)CHN <$		9	-	_ N _	283	ٺ	3.3-4.4	7	0=0-
232	$-CH_2(CH_3)CHNH-$	ξį.	7	_	-HN-	284		3.3 - 4.1	_	-00 - 0-
233	$-CH_2(CH_3)CHNH_2$	-	∞	_	$-\mathbf{NH}_2$	285	ت	2.4 - 3.6	7	-0 - 0-
234	>CH(CH ₃)CHN<	2.7 - 3.8	5	_	-N-	286	_	3.4 - 3.9	9	-0- 1
235	>CHCH3CHNH		9	-	-HN-	287	ٺ	3.4 - 3.9	7	H0 I
236	>CH(CH ₃)CHNH ₂		7	_	$-NH_2$	288	ı	4.1 - 5.1	æ	-0(0=)D- 1
237	≯C(CH ₃)CHN<		4	_	Z	289	1	4.2-5.2	_	
238	>C(CH ₃)CHNH-	ı,	5	_	-HN-	290	ı	3.4 - 3.8	4	1
239	►C(CH ₃)CHNH ₂		9	_	-NH ₂	291	-0(0=C)C(-CH ₃)HSH	3.4 - 3.8	vo (0
240	(-CH ₂)CHN <		s,	- .	_N_	292	-0(0=C)C(-CH ₂)HS-	3.4 - 3.8	m •	^
241	(-CH ₂)CHNH-		01		HN	567	-0(0=C)C(-CH ₂)HSH	Ļ	4 (-C(=0)0-, -SH
747	(-CH ₂)CHNH ₂	2.7-3.8	- <		NH ₂	467	-0(0-C)C(>CH)HS-	ļ I	7 6	C(=)), -3-
247 748	VCH(-CH ₂)CHN/-	1 1	t v	- -	L HN	267	1	3.4-3.0	n —	-C(=)0-, -sn -C(=0)0S-
245	>CH(-CH ₂)CHNH ₃		o ve	-	-NH,	297	1	3.4-3.8	. 7	٠,
246	≯C(−CH ₂)CHN <	ı	3	_	-N-	298	·	I.	m	Z
247	→C(-CH ₂)CHNH-	- 1	4	_	-NH-	299	.	3.4 - 50	4	C=0, -NH-
248	\rightarrow C($-$ CH ₂)CHNH ₂		S	_	$-NH_2$	300	ı	3.4 - 50	S	C=0, −NH ₂
249	(>CH)CHN<	Į.	с п.	_	-Z-	301	-0(0=C)C(>CH)HN<		7	C=0, -N-
250	(>CH)CHNH-	1	4 1		-HN-	302	Ö 0 0	3.4 - 5.0	m -	C=0, NH-
251	(>CH)CHNH ₂	1	'n		-NH ₂	303	-0(0=C)C(>CH)HNH2	1	4 -	C=0, -NH ₂
252	/CH(PC/CHND	7.7-2.8	7 6		-N-	200		2.4-5.0	- c	
254	>CH(>C)CHNH >CH(>C)CHNH	2.7-3.8	4		NH,	300	ı	3.4-5.0	4 cr	
255	(CH ₂)CHS-	- 1	7	. –	-S-	307	=	2.8-4.5		
256	>CH(-CH ₂)CHAr	- 1	4	_	Ar	308		2.8-4.3	∞	-HN- I
257	-CH2(CH3)CHS-	- 1	9	_	-S-	309	_	3.5-4.9	7	-0- 1
258	$-\mathrm{CH}_2(\mathrm{CH}_3)$ CH SH	2.8 - 3.4	7	_	HS-	310	, .	3.5 - 4.1	2	-0-
259	>CH(CH ₃ CHS-		vo /	- ·	-S-	311	Λ	3.5 - 4.1	9	HO- 1
7,00	>CH(CH ₃)CHSH	بر در د	۰ و	 -	HS-	312	ı	3.5-4.5	4 r	-0C=0, -N-
261 253	♦C(CH ₃)CHS−	2.8-3.4	4 4	- -	-S-	313	-0(0=C)C(CH ₃)HNH-	3.5-4.5	ο v	-0C=0,-NH-
707	CH ₂)CH ₂ —	2.8-3.4 2.8-3.4	n v		-S-	315	٨	3.5-4.0	0 4	
3	(11)	;	,	•	נ	,		:)

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で4で24で54-24で21-20で4452で12で422であるよう12で12で51-22で12で12で12で12で12で12で12で12で12で12で12で12で1	4
$\begin{array}{c} 4 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\$	4.8-2.7
>C(-CH ₂)CH-O- >C(-CH ₂)CHOH HO(>CH)CHC=C- -O(>CH)CHC=C- -O(CH ₃)CHC=C- HO(CH ₃)CHC=C- HO(CH ₃)CHC=C- HO(CH ₃)CHC=C- HO(-CH ₃)CHC=C- HO(-CH ₃)CHC=C- HO(-CH ₃)CHC=C- HO(-CH ₃)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=C- HO(>C)CHC=O- CI(CH ₃)CHC(=O)- CI(CH ₃)CHO(- HO(>C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CHO(HO)- -O(C)CH	HO(>CH)CHOH
372 373 373 373 373 373 373 373 373 373	_
-0H -0C(=0)-, Ar -0C(=0)-, Ar -0C(=0)-, Ar -0C(=0)-, Ar -0C(=0)-, Ar -0-, CH=0 -0-, CH	
	0
2.8.2.2.2.4.4.4.4.4.4.4.4.4.4.4.4.4.4.4.	4.0-4.3
	3/1 -CH ₂ CH ₃ CHCl

	no cubefricture	chem shift	multiplicity	эгез	TR constraint	9	substructure	chem chift	multiplicity	area	IR constraint
5 5		40.67	fundamen	-			TO HO	30 00			
428 429	Ar(>CH)CHO-	4.8-5.6	7 8	- -	Ar,0- Ar0H	480 481	VCH-SH WC-SH	0.9-2.5	7 -		HS-I
430	(Ar)CHC(=0)-		· —	-	-OH	482		3.0-4.0		1 1	ArSH
431	(Ar)CHCH=0	4.8 - 5.3	2	_	-CH=0, Ar	483		1.7-2.6	_		C=C
432	(Ar)CHC(=0)O-	4.8 - 5.3	-	_	-C(=0)0-, Ar	484		1.7 - 2.6		ပ -	C≡C
433	(Ar)CHN <	4.8-5.4	- (-N<, Ar	485		1.7-2.6		ن 	C=C
434 435	(Ar)CHNH (Ar)CHNH-	4.8-5.4 4.8-5.3	77 8		-NH-, Ar Ar -NH	486 487	HC=C-Ar HC=C-C=C<	2.7 - 3.4		₹ Ċ 	Ar, C=C C=C C=C
436	-0(Ar)CHC(=0)0-	4.9–5.5) -		-0C(=0)0 Ar			2.1 - 3.3		آ ر 	-C(=0) C=C
437	HO(Ar)CHC(=0)0-	4.9-5.5	5	-	-0H, C(=0)0- Ar		_	1.7-2.4	-	Ö -	•
438	$CH_3(-CH_2)CHOC(=O)-$	5.0-5.6	9	_	-(0=)J-O-	490		1.1 - 1.5	-	- -	-0-, C≡C
439	Cl(CH ₃)CHAr	5.0-5.6	4 (Ar	491		3.5-5.5	m (Ī —,	HO-
4 5	C(-CH ₂)CHAr > CHYCHA:	5.0-5.6	<i>.</i> 0 c		Ar Ar	492	#O-CH<	3.5-5.5	7 -	Ī Ī	# F F
4 4	(/Ch)CHAI Cl(*C)CHAI		v —		Ar	494		4.5-7.5		- I	-on ArOH
443	$-0(-CH_2)CHC(=0)-$	- 1	· κ	-	-0-, $-C(=0)-$	495		2.9-4.8	-	1 A	Ar, -NH-
44 4	$-0(-CH_2)$ CH CH=0	5.0-5.9	4	_	-0-, -CH=0	496		2.9-4.8	4	1 A	Ar, -NH-
445	$HO(-CH_2)CHC(=0)-$	- 1	4 '			497		2.9-4.8	κ,	Į.	Ar, -NH-
446	HO(-CH ₂)CHCH=0	1	vo c		-0H,CH=0	4 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5	1	2.9-4.8	- 5	₹ • 	Ar, NH-
4 5	CI(>CH)CHCI	5.0-0.5	7 -		٠,	5 5	AT-INH-CA	2.9-4.8 0.4-3.5	- "		Af,INH NIU
4	(A) CALCHO	50-61	- 4		7	3 5	` '''	0.4 - 3.5	0.0		-NH-
450	CI(>C)(CHC)	5.0-6.1	-	-		502	,,,	0.4 - 3.5	ı —	. –	-HN-
451	$CI(-CH_2)$ CH OH	5.2 - 6.0	4	-	Н0-	503	$>$ CH $-$ NH $-$ CH $_2-$	0.4 - 3.5	4	-	-NH-
452	CI(-CH ₂)CHO-	5.2-6.0	3	_	-0-	504	٨	0.4 - 3.5	3	-	NH
453	CI(CH ₃)CHO-	5.2 - 6.0	4 1	<u> </u>	-0-	505	j	0.4 - 3.5	ر ،	 	-HN-
454	CI(CH ₃)CHOH	5.2-6.0	v c		HO-	206	-CH ₂ -NH-CH ₃	0.4 - 3.5	9 4	 -	-HN-
455 456	CI(>CH)CHOH	5.2 - 6.0	4 m		HO-	50%	. //\	0.4 - 3.5	o 4		HZ
457	CI(≯C) CH 0−	5.2-6.0	. —	. –	-0-	208		7.8-9.4	· _	1 A	Ar, -NH-
458	CI(≯C)CHOH	5.2-6.0	2	_	Н0-	510	_	6.0 - 8.2	4		¦•
459	CI(-CH ₂)CHCI		m ,		i i	511		6.0 - 8.2	т (
94 60	-0(0=)CCH-C(=0)0- NC/-N/CHC(=0)0-	4.9-5.7		- -	-C(=0)0- -NC=00-	512	>CH~NH~C(=0)- *C-NH~C(=)-	6.0-8.2	2 -		-NH-,C(=0)- -NH-,C(=0)-
1.64	NC(~IN)CHC(=0)0-	- 1	- 6		-NHC=00-	514		6.0 8.2	- 4	-	٠,
463	NC(NH ₂)CH-C(=0)0-		ı m	· —	. •	515	,,	6.0-8.2	· ĸ	· -	• ¦ •
464	-0(-0) CH 0-	5.4-5.8	1	_	-0-	516	///	6.0 - 8.2	2	_	-NH-, -CH=0
465	-0(-0) CH OH	5.4-5.8	7 -	— -	-0H, -0-	517	-CH ₂ -NH ₂	0.4 - 3.5	m c	7 6	$-NH_2$
460	Ar(=0)CHC=0	5.4-6.1	٠ ,	- -	Ar, -0-, -((-0)- Ar -0H -((=0)-	510	. 71	0.4-3.5	7 -	4 C	-NH2
468	Ar(-0) CH CH=0	5.6-6.0	1 6		,	520		2.9-4.8		7 7	Ar, –NH,
469	Ar(HO)CHCH=0	5.6 - 6.0	3	_	Ar, -0H, -CH=0	521		5.0-6.5	_	2	$-C=0, -NH_2$
470	Ar(-O)CHAr	5.6 - 6.0	-	_	Ar, -0-	522	0	7.7-8.3	-	Ö _	0=CH-, -0-
471	Ar(HO)CHAr	5.6 - 6.0	7	- -	Ar, –0H	523		9.3 - 10.6	m (Ö (0=CH-
472	CI(CH ₃)CHNO ₂	5.6 - 6.0	4 "		-NO ₂	524	O=CH-CH <	9.3 - 10.6	7		O=CH- O=CH-
474	CI(>CH2/CHNO2	5.7-6.1	5 2	-	-NO ₂	526		10.0-13.5) i	-(0=)C-Oh
475	CI(>C)CHNO ₂	5.7-6.1	ı 	· 	$-N0_2$	527	4	6.0-8.5	· 	1 Ar	`
476	(CI)CHC(=0)0-	5.7 - 6.1	_	_	-C(=0)0-	528	_	4.8 - 8.0	_	5 C	$CH_2 = C$
477	-(0=)COC(Ar)HC(=0)O-	5.7-6.1			Ar, $-C(=0)0^{-}$	529		1	(ī (
4/8 479	(AI)CHCI -CH ₂ SH	0.9 - 2.5	3 -		ASH —SH	531	-CH=CH-	4.8—6.0 4.8—9.8	7 7	7 T	CH=C

subset S_{hh} of substructures that are indicated by a specific first-order ¹H-NMR spectrum.

In ESSESA, the first-order ¹H-NMR spectrum is not the only one to derive the substructure constraints, IR and ¹³C-NMR spectra are used also. The IR spectrum is the first one to be analyzed, and the result (eq 4)⁵ from the IR spectral analysis is used as the constraint in the first-order ¹H-NMR spectral analysis.

On the basis of eq 5, to derive the subset S_{hh} of substructures that are consistent with the IR and the first-order ¹H-NMR spectral data and other chemical information is to confirm that the expecting absorption peaks of S_{hh} exist in the spectral data. Equation 5 can be written in production system rules; for example, the production system rule that is derived the identification of the substructure CH_3 -Ar is as follows:

in the first-order ¹H-NMR spectrum of an unknown compound there are peak with chemical shift 1.9—
 2.8 ppm, multiplicity 1, and area equivalent to three hydrogen atoms, and there exists the aromatic substructure from the IR spectral analysis

Then the substructure CH₃-Ar may be present in the structure of the unknown compound

In ESSESA this rule can be written in PROLOG such as follows:

There are 531 substructures in the knowledge base used by ESSESA for the first-order ¹H-NMR spectrum analysis. It exceeds the databases used by Sasaki⁹ and Miller.¹⁰ These 531 substructures are shown in Table 1. There exists overlap within this table, for example substructures 17, 21, and 19 overlap substructure 2 and so on. This problem will be solved before the complete structure candidates are generated on the basis of information derived from the IR, ¹H-NMR, and ¹³C-NMR spectral analysis. The mutually consistent set of substructures that will be used by the structure generator can be achieved by means of a procedure that finds those combinations of permitted substructures that are compatible with the overall composition of the molecule and with the constraints derived from all of the spectral data. Each such combination of substructures then may be used to define a distinct problem that can be referred to a subsequent structure generation program.

INTERPRETER PROGRAM

After the ring and double-bond characteristics of the entered molecular formula and the digitized first-order ¹H-NMR spectrum as well as the analysis result of IR spectrum are passed to the interpreter program of first-order ¹H-NMR spectral analysis, the interpretation of the first-order ¹H-NMR spectrum is started. Using eq 5 and the knowledge base the interpreter program begins to identify the various substructural fragments that may be present in the structure. The result of interpretation is a substructure list that is used in the structure generation. The structure of this interpreter program is shown in Figure 1.

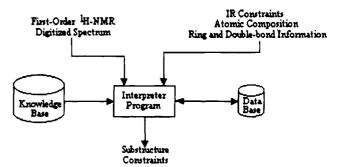


Figure 1. Overview of the structure of the interpreter program.

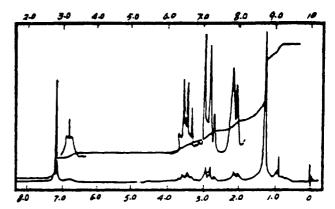


Figure 2. The ¹H-NMR spectrum of C₁₉H₃₁NO.

The interpreter program works with the set of 531 substructures. Each of these substructures is correlated with a defined pattern of first-order ¹H-NMR spectral absorption. The spectral features that characterize substructural fragments normally consist of spectral ranges within which specific types of peaks are expected. The initial set of substructures is, in effect, screened against the entered spectral data, and any substructure whose requisite spectral pattern is absent is discarded. The result of this analysis is a subset of the 531 substructures. Each of the members of the subset is related to absorption data, which is consistent with the first-order ¹H-NMR spectrum of the unknown compound.

In the interpretation, goal-driving inference tactics were used. In such an inference model, substructures from the knowledge base are used as the goals, and the program will seek spectral patterns that fulfill the premises of the goals, as defined by the production rules. If any single premise of a goal is not satisfied, that goal is determined to be false, that is to say, that a particular substructure cannot be contained in the unknown structure. If all the premises of a goal are satisfied, then the goal is considered to be true, and the substructure corresponding to the goal may be embedded within the complete structure of the unknown compound.

EXAMPLE

N-Phenethylundecanamide C₁₉H₃₁NO, structure 1, is taken as an example to illustrate the procedure in which the constraints from first-order ¹H-NMR spectral analysis are obtained in ESSESA. Figure 2 is the first-order ¹H-NMR spectrum of *N*-phenethyundecanamide.

At first, the first-order ¹H-NMR is entered. The spectral data must be presented in digital form, but the means of

Table 2. The Digitized ¹H-NMR Spectrum of C₁₉H₃₁NO

chemical shift	multiplicity	area	chemical shift	multiplicity	area
0.89	3	3	1.23	100 ^a	16
2.13	3	2	2.80	4	2
3.47	4	2	6.79	100^{a}	1
7.19	100^{a}	5			

^a The integer 100 indicates the multiplet structure.

Table 3. The Analysis Result of ¹H-NMR Spectrum of C₁₉H₃₁NO

Table 5.	The Amarysis Result of 11-1400	to opeculari of Cigrisii to
no.	LNSCS code	substructure
1	CC.	CH ₃ CH ₂ -
2	CCC.	CH ₃ CH ₂ CH ₂ -
3	C.2CC.2	>CHCH ₂ CH<
4	CCC.3	CH ₃ CH ₂ C€
5	C.CC.3	-CH ₂ CH ₂ -C€
6	CCC.2	CH ₃ CH ₂ CH <
7	C.CC.	-CH ₂ CH ₂ CH ₂ -
8	C.2CC.3	>CHCH ₂ C€
9	C.3CC.3	>CH ₂ C<
10	C.CC.2	-CH ₂ CH ₂ CH<
11	C.CC.=O	$-CH_2CH_2C(=O)-$
12	C.CN.	-CH ₂ CH ₂ NH-
13	C.CC1=CC=CC=C1	011 011
		-CH ₂ CH ₂
14	C.2CN.	>CHCH ₂ NH-
15	C.2C(C.2)2	>CHC(>CH)HCH<
16	C.2C(C.2)C.3	>CHC > CHCH<
17	C.C(C.)C.2	-CH2C(>CH)HCH2-
18	C.C(C.)C.3	-CH2C(>CH)HCH2-
19	C.C(C.2)C.3	-CH2C(>CH)HCH<
20	C.2C(C.2)2	>CHO(>CH)HCH<
21	C.C(C.3)C1=CC=CC=C1	-CH2C(>C)H-
		01120(>C)H
22	C.2C(C.2)Cl=CC=CC=C1	
		>CHC(>CH)H-()
23	C.2C(C.3)C.=O	>CHC>CHC(= O)
24	O=C.C(C.2)2	>CHC(>CH)HC(=O)
25	C.2C(C.3)N.	>CHC(>CHNH-
26	N.C(C.2)2	>CHC(CH)HNH-
27	C.C(C.3)N.	-CH₂C>CHNH-
28	C.C(C.2)Cl=CC=Cl	>CH ₂ C(>CH)H
29	C.NC.=O	$-CH_2NHC(=O)-$
30	C.2NC.=O	>CHNHC(=O)-
31	C.3NC.=O	>CNHC(=0)-
32	Cl=CC=CC=Cl.	

digitization is unimportant, as long as it is accurate. ESSESA accepts the following ranges for the digitized data: chemical shifts of 0.0-11.0 ppm; multiplicity of 0-10, the integer 100 is input if the peak is a broad multiplet group of absorption peaks; relative area of peaks is an integer of 1-20. The digital data from the spectrum in Figure 2 are given in Table 2. The constraints from IR spectral analysis and the ring and double-bond characteristics and the atomic composition, which are obtained in the IR spectral analysis, of N-phenethylundecanamide are passed to the interpreter program to help the analysis of first-order 1 H-NMR.

As the interpreter program acquires these data, it makes use of the rules in the knowledge base to compare the stored spectral patterns with the digital input data to identify the substructures that might be contributing to the *N*-phenethyl-undecanamide structure. The resulting constraints from the first-order ¹H-NMR spectral analysis of *N*-phenethylundecanamide are listed in Table 3. The structure of *N*-phenethylundecanamide can be compared with the substruc-

tures shown in Table 3, which ESSESA decided should be present in the structure of *N*-phenethylundecanamide, based upon the first-order ¹H-NMR spectrum. The substructures 1, 2, 7, 12, 13, 29, and 32 exist really in the structure of *N*-phenethylundecanamide. Most of the substructures from the analysis of the first-order ¹H-NMR spectra do not exist in the structure of *N*-phenethylundecanamide. These substructures are generated from multiplet explanation of the peaks, especially the multiplet peaks. They can be deleted from the analysis of ¹³C-NMR spectrum and substructural consistent analysis.

DISCUSSION

Chemical shift in NMR spectra is the physical property related with structural environment. The chemical shifts of two peaks in a ¹H-NMR spectrum will be very near if the structural environments of the two hydrogen atoms in the structure are very similar. The peaks of their chemical shifts are very near and will generate a complicated shape peak. The assignment of such peaks is very hard for first-order ¹H-NMR spectra. In ESSESA all possible explanations of such peaks are used as the substructure constraints in order to generate all possible candidate structures for an unknown compound. The correct substructures that really exist in the structure of an unknown compound will be obtained by analysis of all substructure constraints from IR, ¹H-NMR, and ¹³C-NMR spectral data.

Some substructures will generate peaks that have similar chemical shifts and the same multiplet as well as the areas of peaks. These peaks can be assigned to some different substructures, but not all of these different substructures are really included in the unknown structure. The incorrect substructure will be deleted by subsequent ¹³C-NMR spectral analysis and substructures consistent analysis. The details about this process will be presented in a future paper.

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