

# FITC-Conjugated Cyclic RGD Peptides as Fluorescent Probes for Staining Integrin $\alpha_v\beta_3/\alpha_v\beta_5$ in Tumor Tissues

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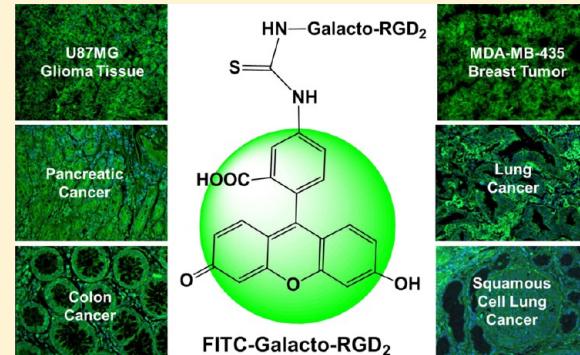
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## Supporting Information

**ABSTRACT:** This study sought to evaluate FITC-conjugated cyclic RGD peptides (FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub>) as fluorescent probes for in vitro assays of integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression in tumor tissues. FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub> were prepared, and their integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity was determined using the displacement assay against <sup>125</sup>I-echistatin bound to U87MG glioma cells. IC<sub>50</sub> values of FITC-Galacto-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-RGD<sub>2</sub> were calculated to be 28 ± 8, 32 ± 7, and 89 ± 17 nM, respectively. The integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity followed a general trend: FITC-Galacto-RGD<sub>2</sub> ~ FITC-3P-RGD<sub>2</sub> > FITC-RGD<sub>2</sub>. The xenografted tumor-bearing models were established by subcutaneous injection of 5 × 10<sup>6</sup> tumor cells into shoulder flank (U87MG, A549, HT29, and PC-3) or mammary fat pad (MDA-MB-435) of each athymic nude mouse. Three to six weeks after inoculation, the tumor size was 0.1–0.3 g. Tumors were harvested for integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  staining, as well as hematoxylin and eosin (H&E) staining. Six human carcinoma tissues (colon cancer, pancreatic cancer, lung adenocarcinoma, squamous cell lung cancer, gastric cancer, and esophageal cancer) were obtained from recently diagnosed cancer patients. Human carcinoma slides were deparaffinized in xylene, rehydrated with ethanol, and then used for integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  staining, as well as H&E staining. It was found that the tumor staining procedures with FITC-conjugated cyclic RGD peptides were much simpler than those with the fluorescence-labeled integrin  $\alpha_v\beta_3$  antibodies. Since FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub> were able to co-localize with the fluorescence-labeled integrin  $\beta_3$  antibody, their tumor localization and tumor cell binding are integrin  $\alpha_v\beta_3$ -specific. Quantification of the fluorescent intensity in five xenografted tumors (U87MG, MDA-MB-435, A549, HT29, and PC-3) and six human carcinoma tissues revealed an excellent linear relationship between the relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels determined with FITC-Galacto-RGD<sub>2</sub> and those obtained with the fluorescence-labeled anti-human integrin  $\beta_3$  antibody. There was also an excellent linear relationship between the tumor uptake (%ID/g) of <sup>99m</sup>Tc-3P-RGD<sub>2</sub> (an integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -targeted radiotracer) and the relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels from the quantification of fluorescent intensity in the tumor tissues stained with FITC-Galacto-RGD<sub>2</sub>. These results suggest that FITC-conjugated cyclic RGD peptides might be useful to correlate the in vitro findings with the in vivo imaging data from an integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -targeted radiotracer. The results from this study clearly showed that the FITC-conjugated cyclic RGD peptides (particularly FITC-3P-RGD<sub>2</sub> and FITC-Galacto-RGD<sub>2</sub>) are useful fluorescent probes for assaying relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels in tumor tissues.



## INTRODUCTION

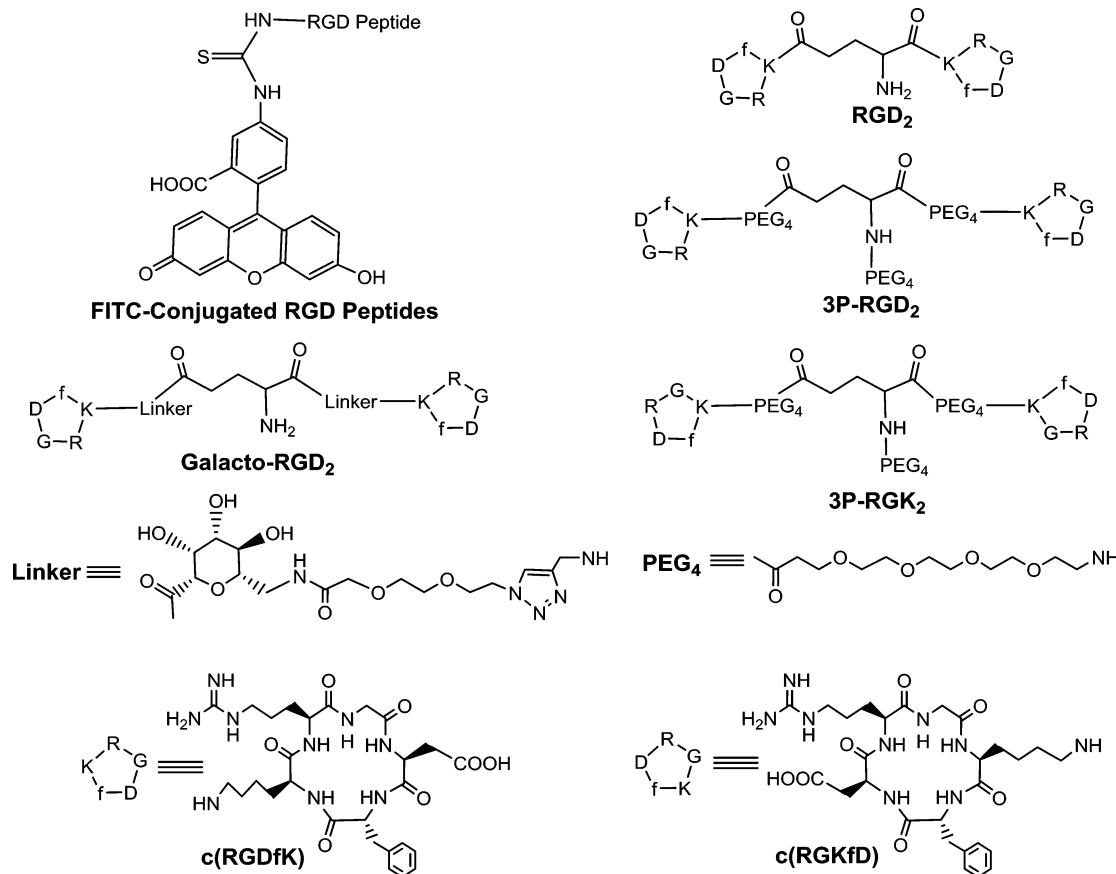
The integrin family is a group of transmembrane glycoproteins comprised of 19  $\alpha$ - and 8  $\beta$ -subunits that are expressed in 25 different  $\alpha/\beta$  heterodimeric combinations on the cell surface.<sup>1–4</sup> Integrins are critically important in many physiological processes, including cell attachment, proliferation, bone remodeling, and wound healing.<sup>3,4</sup> Integrins also contribute to pathological events such as thrombosis, atherosclerosis, tumor invasion, angiogenesis, and metastasis, infection by pathogenic microorganisms, and immune dysfunction.<sup>3–10</sup> Among 25 members of the integrin family, integrin  $\alpha_v\beta_3$  is

studied most extensively for its role in tumor growth, progression, and angiogenesis.

Integrin  $\alpha_v\beta_3$  is a receptor for extracellular matrix proteins (vitronectin, fibronectin, fibrinogen, laminin, collagen, Von Willebrand's factor, and osteoponin) with the exposed arginine-glycine-aspartic (RGD) tripeptide sequence.<sup>1,2</sup> Changes in the integrin  $\alpha_v\beta_3$  expression levels and activation state have been well documented during tumor growth and metastasis.<sup>5,7,10</sup>

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**Figure 1.** ChemDraw structures of FITC-conjugated cyclic peptide dimers (RGD<sub>2</sub>, 3P-RGD<sub>2</sub>, 3P-RGK<sub>2</sub>, and Galacto-RGD<sub>2</sub>).

Integrin  $\alpha_v\beta_3$  is expressed in low levels on the epithelial cells and mature endothelial cells, but it is highly expressed in many tumors, including osteosarcomas, glioblastomas, melanomas, and carcinomas of lung and breast.<sup>11–24</sup> Studies show that integrin  $\alpha_v\beta_3$  is overexpressed on both tumor cells and activated endothelial cells of neovasculature.<sup>11</sup>

Integrin  $\alpha_v\beta_3$  on endothelial cells modulate cellular adhesion during angiogenesis, while the integrin  $\alpha_v\beta_3$  on tumor cells potentiate metastasis by facilitating invasion of tumor cells across the blood vessels.<sup>19–36</sup> It has been shown that integrin  $\alpha_v\beta_3$  expression levels correlate well with the potential for metastasis and aggressiveness of tumors, including glioma, melanoma, and carcinomas of the breast and lungs.<sup>19–25</sup> Integrin  $\alpha_v\beta_3$  has been considered an interesting biological target for development of therapeutic pharmaceuticals for cancer treatment,<sup>13,36–40</sup> and molecular imaging probes for diagnosis of rapidly growing and highly metastatic tumors.<sup>41–52</sup>

Only two integrin family members ( $\alpha_v\beta_3$  and  $\alpha_{IIb}\beta_3$ ) contain the  $\beta_3$  chain. Since integrin  $\alpha_{IIb}\beta_3$  is expressed exclusively on the activated platelets,<sup>26,29,30</sup> the integrin  $\beta_3$  expression level on tumor cells or in tumor tissues should be the same as that of integrin  $\alpha_v\beta_3$ . Western blotting has been used to determine the integrin  $\alpha_v\beta_3$  concentration in tumor tissues,<sup>53–59</sup> but the percentage of integrin  $\alpha_v\beta_3$  recovery from tumor tissues and its activation state remained unknown. We have been using immunohistochemical (IHC) staining with anti-integrin  $\alpha_v\beta_3$  monoclonal antibodies to determine the integrin  $\alpha_v\beta_3$  expression levels on tumor cells (acetone-fixed or living) and in tumor tissues (acetone or methanol-fixed).<sup>60,61</sup> It was found that the IHC staining is an excellent technique to reflect the

activation state of integrin  $\alpha_v\beta_3$  because only the activated endothelial cells and some tumor cells are able to bind to the integrin  $\alpha_v\beta_3$  monoclonal antibody. However, the procedures for cellular and tissue staining with fluorescence-labeled anti-integrin  $\alpha_v\beta_3$  antibodies are often complicated and time-consuming because they involve the blocking of nonspecific binding and the use of secondary polyclonal antibody for the fluorescence detection. In addition, integrin  $\alpha_v\beta_3$  antibodies have limited stability in aqueous solution.

To overcome these shortcomings, we prepared three FITC conjugated dimeric cyclic RGD peptides (Figure 1): FITC-RGD<sub>2</sub> (RGD<sub>2</sub> = Glu[cyclo(Arg-Gly-Asp-d-Phe-Lys)]<sub>2</sub>), FITC-3P-RGD<sub>2</sub> (3P-RGD<sub>2</sub> = PEG<sub>4</sub>-Glu[cyclo[Arg-Gly-Asp-d-Phe-Lys(PEG<sub>4</sub>)]]<sub>2</sub> (PEG<sub>4</sub> = 15-amino-4,7,10,13-tetraoxapentadecanoic acid) and FITC-Galacto-RGD<sub>2</sub> (Galacto-RGD<sub>2</sub> = Glu[cyclo[Arg-Gly-Asp-d-Phe-Lys(SAA-PEG<sub>2</sub>-(1,2,3-triazole)-1-yl-4-methylamide)]]<sub>2</sub>, SAA = 7-amino-L-glycero-L-galacto-2,6-anhydro-7-deoxyheptanamide, and PEG<sub>2</sub> = 3,6-dioxaoctanoic acid). Since the cyclic RGD peptides, such as c(RGDFK), are antagonists for  $\alpha_v\beta_3$  and  $\alpha_v\beta_5$  integrins,<sup>4,36–39,62</sup> it is reasonable to believe that bioconjugates FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub> will target both integrin  $\alpha_v\beta_3$  and integrin  $\alpha_v\beta_5$ . We were interested in dimeric cyclic RGD peptides because they have significantly higher integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity than their monomeric analogs.<sup>44</sup>

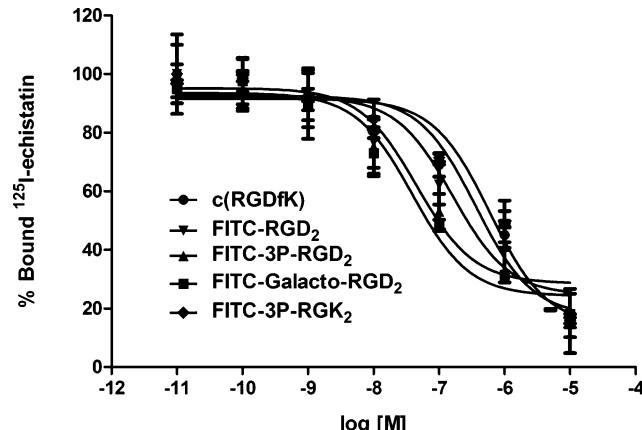
In this report, we present the evaluations of FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub> as fluorescent probes to assay integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels in tumor tissues. Their integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity was determined in a displacement assay against <sup>125</sup>I-echistatin

bound to U87MG glioma cells. To demonstrate their RGD-specificity, we have also prepared FITC-3P-RGK<sub>2</sub> (3P-RGK<sub>2</sub> = PEG<sub>4</sub>-E[PEG<sub>4</sub>-c(RGKfD)]<sub>2</sub> = PEG<sub>4</sub>-Glu[cyclo[Arg-Gly-Lys-(PEG<sub>4</sub>-D-Phe-Asp)]<sub>2</sub>]), which has the identical chemical composition to that of FITC-3P-RGD<sub>2</sub> but with low integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity due to the RGK tripeptide sequence. Even though the fluorescence-labeled cyclic RGD peptides have been reported as optical imaging probes,<sup>63</sup> very little information is available on their validity for in vitro assays of integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression. The objective of this study is to validate the utility of FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub> for determination of relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels in both the xenografted tumors and human carcinoma tissues in comparison with commercially available integrin  $\alpha_v\beta_3$  monoclonal antibodies.

## RESULTS

**Synthesis of FITC–Peptide Conjugates.** FITC-conjugated peptides (FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, FITC-3P-RGK<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub>) were prepared by reacting FITC with the corresponding cyclic peptides under basic conditions in the presence of excess DIEA. All new FITC–peptide conjugates were purified by HPLC. MALDI-MS data were completely consistent with the composition proposed for FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, FITC-3P-RGK<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub>. Their HPLC purity was >95% before being used for the integrin  $\alpha_v\beta_3$  binding assays and staining studies.

**Integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  Binding Affinity.** The whole-cell assay was used to determine integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity of FITC-conjugated cyclic RGD peptides. Figure 2 shows the



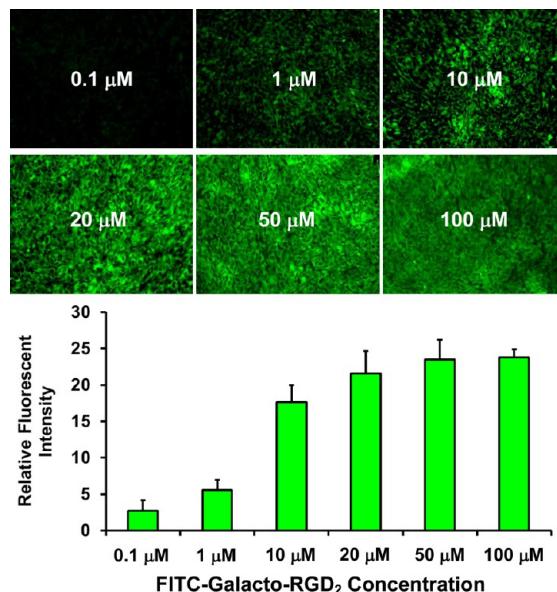
**Figure 2.** Competitive displacement curves of <sup>125</sup>I-echistatin bound to U87MG human glioma cells in the presence of FITC-conjugated cyclic RGD peptides. Their IC<sub>50</sub> values were obtained from curve fitting and were calculated to be 28 ± 8, 32 ± 7, 89 ± 17, 589 ± 73, and 414 ± 36 nM for FITC-Galacto-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, FITC-RGD<sub>2</sub>, FITC-3P-RGK<sub>2</sub> and c(RGDFK), respectively. c(RGDFK) was used as a standard. 3P-RGK<sub>2</sub> was the dimeric nonsense peptide to demonstrate the RGD-specificity of FITC-conjugated cyclic RGD peptides.

displacement curves of <sup>125</sup>I-echistatin bound to U87MG glioma cells in the presence of FITC-conjugated peptides. IC<sub>50</sub> values were calculated to be 28 ± 8, 32 ± 7, 89 ± 17, and 589 ± 73 nM for FITC-Galacto-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, FITC-RGD<sub>2</sub>, and FITC-3P-RGK<sub>2</sub>, respectively. The IC<sub>50</sub> value of c(RGDFK) (414 ± 36 nM) was close to that from our previous report.<sup>64–66</sup> The integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity followed a general trend: FITC-3P-RGD<sub>2</sub> ~ FITC-Galacto-RGD<sub>2</sub> > FITC-RGD<sub>2</sub>

» c(RGDFK) > FITC-3P-RGK<sub>2</sub>, which was consistent with the results for their DOTA (1,4,7,10-tetraazacyclododecane-1,4,7,10-tetracetic acid) and HYNIC (6-(2-(2-sulfonatobenzaldehyde)hydrazono)nicotinyl) derivatives.<sup>44,60–71</sup> The lower binding affinity of FITC-3P-RGK<sub>2</sub> (IC<sub>50</sub> = 589 ± 73 nM) than that of FITC-3P-RGD<sub>2</sub> (IC<sub>50</sub> = 32 ± 7 nM) clearly demonstrated the RGD-specificity of the FITC-conjugated cyclic RGD peptides.

### Optimal Concentration for Tumor Tissue Staining.

The concentration-dependence experiments were performed using the xenografted U87MG glioma and human carcinoma tissues. Figure 3 displays microscopic images of the xenografted

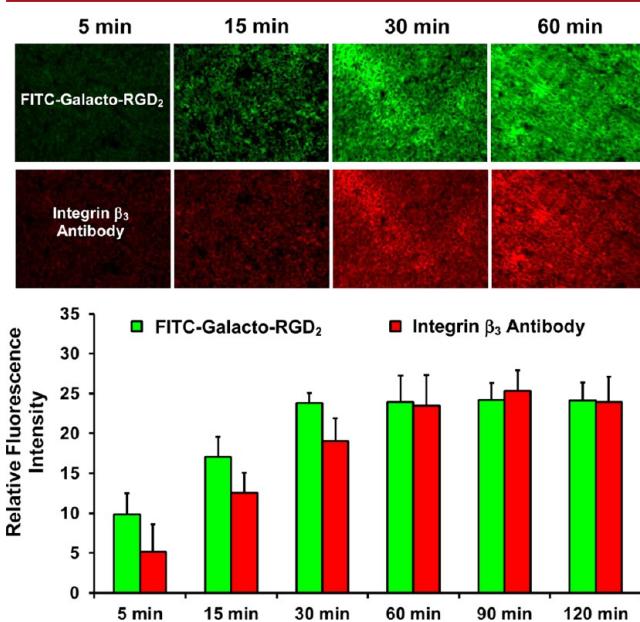


**Figure 3.** Microscopic images (Magnification: 200×) of the xenografted U87MG glioma tissues, and quantification of fluorescence intensity on tumor slice stained with FITC-Galacto-RGD<sub>2</sub> at 0.1, 1, 10, 20, 50, and 100 μM. Tumor tissues could be well-stained in the range of 10–100 μM.

U87MG glioma tissues stained with FITC-Galacto-RGD<sub>2</sub> at 0.1, 1, 10, 20, 50, and 100 μM, along with semiquantification of fluorescence intensity. The relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression in the U87MG glioma tumor was quantified as the percentage of green-colored (FITC-Galacto-RGD<sub>2</sub>) or red-colored (anti-integrin  $\beta_3$  antibody) area over the total area of each image. We found that all U87MG glioma tissues, which have high integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression on both tumor cells and neovasculature,<sup>22–33</sup> were well stained with FITC-Galacto-RGD<sub>2</sub> in the range of 10–100 μM. Very similar results were obtained with human carcinoma tissues (colon cancer, squamous cell lung cancer, and gastric cancer) stained with FITC-Galacto-RGD<sub>2</sub> (Supporting Information Figure S11). The minimal concentration for positive staining of colon carcinoma was ~5 μM. For squamous cell lung cancer tissues, a higher concentration (>25 μM) was needed in order to achieve adequate fluorescence staining. There was little staining in gastric carcinoma tissue, likely due to its low integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression. Since the higher concentrations were required for the tumor tissues with lower integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression, we used 100 μM of FITC-conjugated cyclic RGD peptides for most of the staining studies in xenografted tumor tissues, and

50  $\mu$ M of FITC-labeled cyclic RGD peptides for the staining of human carcinoma tissues.

**Tissue Staining Kinetics for Tumor Tissues.** We explored the impact of incubation time on the fluorescent intensity of U87MG glioma and human carcinoma tissues stained with FITC-Galacto-RGD<sub>2</sub> (100  $\mu$ M). We also performed IHC staining with anti-integrin  $\beta_3$  antibody using the same tumor slice in order to compare their staining kinetics (time-dependence) under identical experimental conditions. It was found that U87MG glioma tissues could achieve maximal fluorescent intensity with FITC-Galacto-RGD<sub>2</sub> within 30 min while it took >60 min for the same tumor tissue using the anti-integrin  $\beta_3$  antibody (Figure 4). Similar results were obtained



**Figure 4.** Microscopic images (Magnification: 200 $\times$ ) of the xenografted U87MG glioma tissues stained with FITC-Galacto-RGD<sub>2</sub> (green) and rabbit anti-human integrin  $\beta_3$  antibody (red), and tissue staining kinetics as indicated by fluorescence intensity at different incubation times (5–120 min).

with human carcinoma tissues stained with FITC-Galacto-RGD<sub>2</sub> (Supporting Information Figure SI2). The colon cancer tissues were positively stained within 15 min of incubation. The faster staining kinetics of FITC-Galacto-RGD<sub>2</sub> might explain the fact that the same tumor slice could be labeled with both FITC-Galacto-RGD<sub>2</sub> and anti-integrin  $\beta_3$  monoclonal antibody. Considering the tumor tissues with lower integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression than that in U87MG glioma and colon cancer, the 60 min incubation time was used for most of the tumor tissue staining studies.

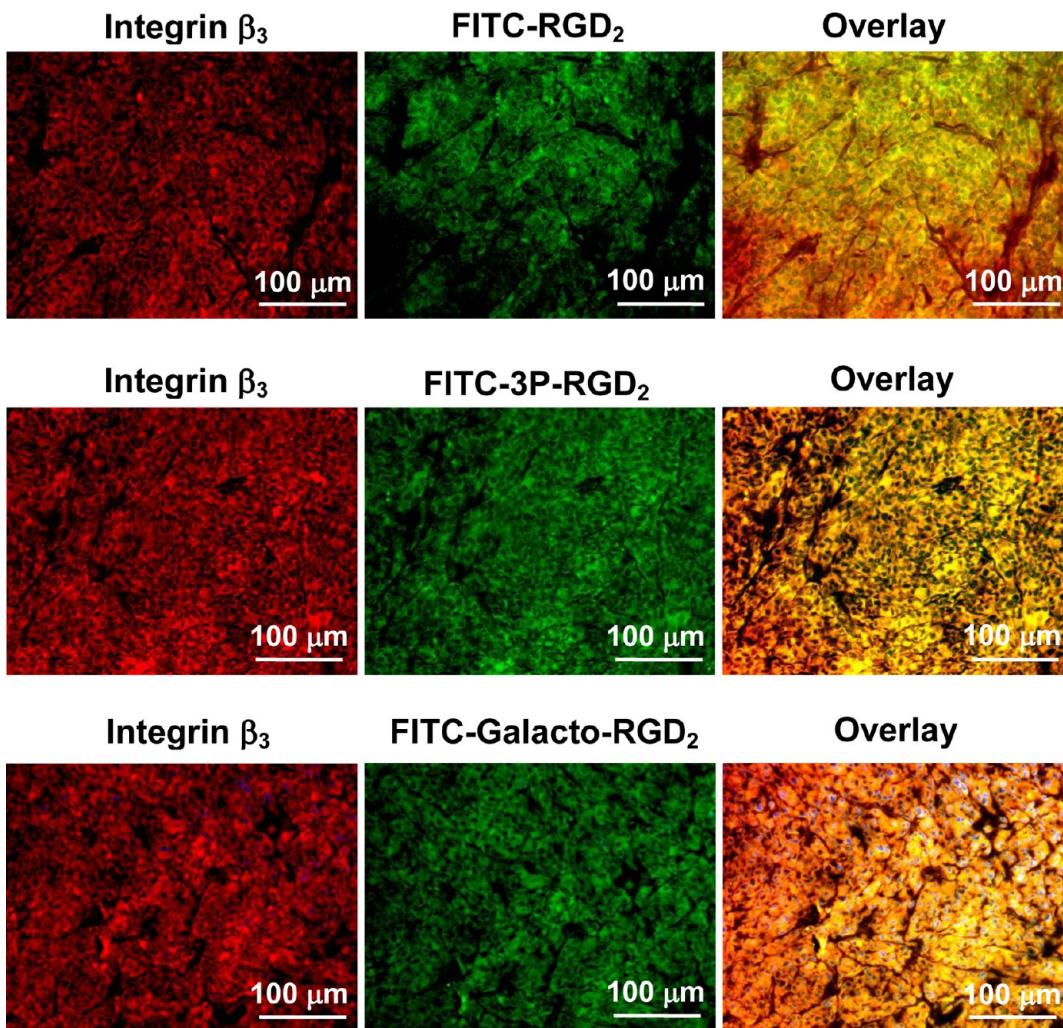
**Co-Localization of FITC-Conjugated Cyclic RGD Peptides with Integrin  $\alpha_v\beta_3$  Antibody in Tumor Tissues.** The overlay experiments were performed using the xenografted U87MG glioma (Figure 5) and human colon cancer (Figure 6) tissues to show the co-localization of FITC-conjugated cyclic RGD peptides and rabbit anti-integrin  $\beta_3$  antibody. We found that the tumor tissues stained with FITC-3P-RGD<sub>2</sub> or FITC-Galacto-RGD<sub>2</sub> showed more fluorescence intensity than that with FITC-RGD<sub>2</sub>. In all cases, they were able to co-localize with anti-integrin  $\beta_3$  antibody, as evidenced by the orange and yellow colors (red integrin  $\beta_3$  antibody merged with green FITC-labeled cyclic RGD peptide) in overlay images. FITC-3P-

RGD<sub>2</sub> and FITC-Galacto-RGD<sub>2</sub> were as effective for staining integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression as the anti-integrin  $\beta_3$  antibody for integrin  $\alpha_v\beta_3$  in U87MG glioma and human carcinoma tissues. The same conclusion could be made for cellular staining integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  (Supporting Information Figures SI3–SI5).

**Integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  Specificity.** Blocking experiments were performed using the xenografted U87MG glioma and human colon cancer to demonstrate the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  specificity of the FITC-conjugated cyclic RGD peptides. It was found that the xenografted U87MG glioma tissues were successfully stained with FITC-Galacto-RGD<sub>2</sub>, but not in the presence of excess RGD<sub>2</sub> (Figure 7A) due to blockage of integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ . The same results were obtained with the human colon carcinoma tissues (Figure 7B). Figure 7C shows microscopic images of the tumor slice obtained from a U87MG-bearing mouse injected with ~300  $\mu$ g of FITC-Galacto-RGD<sub>2</sub>. The fluorescence intensity and distribution patterns in the xenografted U87MG glioma tissue (Figure 7C: left) were almost identical to those stained with FITC-Galacto-RGD<sub>2</sub> (Figure 7B: left). Further staining with the hamster anti-mouse integrin  $\beta_3$  antibody (Figure 7C: right) was not successful since all integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding sites on tumor tissues had already been occupied by FITC-Galacto-RGD<sub>2</sub>. The results from these blocking studies clearly showed that the tumor localization of FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub> are integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -specific.

**RGD Specificity.** To demonstrate the RGD specificity of FITC-labeled cyclic RGD peptides, we used 3P-RGK<sub>2</sub> as a “nonsense” peptide. Due to the RGK tripeptide sequence, FITC-3P-RGK<sub>2</sub> had much lower integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity than FITC-3P-RGD<sub>2</sub> (Figure 2). Figure 8 shows microscopic images of the xenografted U87MG glioma tissues (top four panels) and the colon cancer tissues (bottom four panels) stained with FITC-RGD<sub>2</sub>, FITC-Galacto-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, or FITC-3P-RGK<sub>2</sub>. Apparently, the xenografted U87MG glioma and human colon cancer tissues were all positively stained with FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub>, but not with FITC-3P-RGK<sub>2</sub> under the same conditions. Thus, the tumor-binding of FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub> is RGD-specific.

**Quantification of Integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  Expression Xenografted Tumor Tissues.** Quantification of the relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels was performed using five xenografted tumors (U87MG, MDA-MB-435, A549, HT29, and PC-3). Figure 9 compares microscopic images of the xenografted tumors stained with FITC-Galacto-RGD<sub>2</sub> and anti-integrin  $\beta_3$  antibody. In overlay images, orange and yellow colors indicate co-localization of FITC-Galacto-RGD<sub>2</sub> and anti-integrin  $\beta_3$  antibody. Figure 10 summarizes the quantification data of relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels in the xenografted U87MG, MDA-MB-435, A549, HT29, and PC-3 tumor tissues. The fluorescent intensity (integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  density) on the tumor cells and tumor neovasculature was represented by the percentage of green (FITC-Galacto-RGD<sub>2</sub>) or red (anti-integrin  $\beta_3$  antibody) area over the total area in each image. We found that the integrin  $\beta_3$  (anti-integrin  $\beta_3$  antibody) and integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  (FITC-Galacto-RGD<sub>2</sub>) expression levels followed a general order of U87MG > HT29  $\approx$  MDA-MB-435  $\approx$  A549  $\gg$  PC-3 (Figure 10A,B). There was an excellent linear relationship (Figure 10C,D) between the relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels determined with FITC-Galacto-RGD<sub>2</sub> and the relative integrin



**Figure 5.** Microscopic images (200 $\times$ ) of the xenografted U87MG glioma tissues stained with FITC-conjugated cyclic RGD peptide (green) and rabbit anti-human integrin  $\beta_3$  antibody detected with Cy3 conjugated goat anti-rabbit antibody (red). Blue color indicates the presence of nuclei stained with DAPI. In overlay images, the orange and/or yellow color indicates co-localization of FITC-conjugated cyclic RGD peptides (FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub>) with rabbit anti-human integrin  $\beta_3$  antibody.

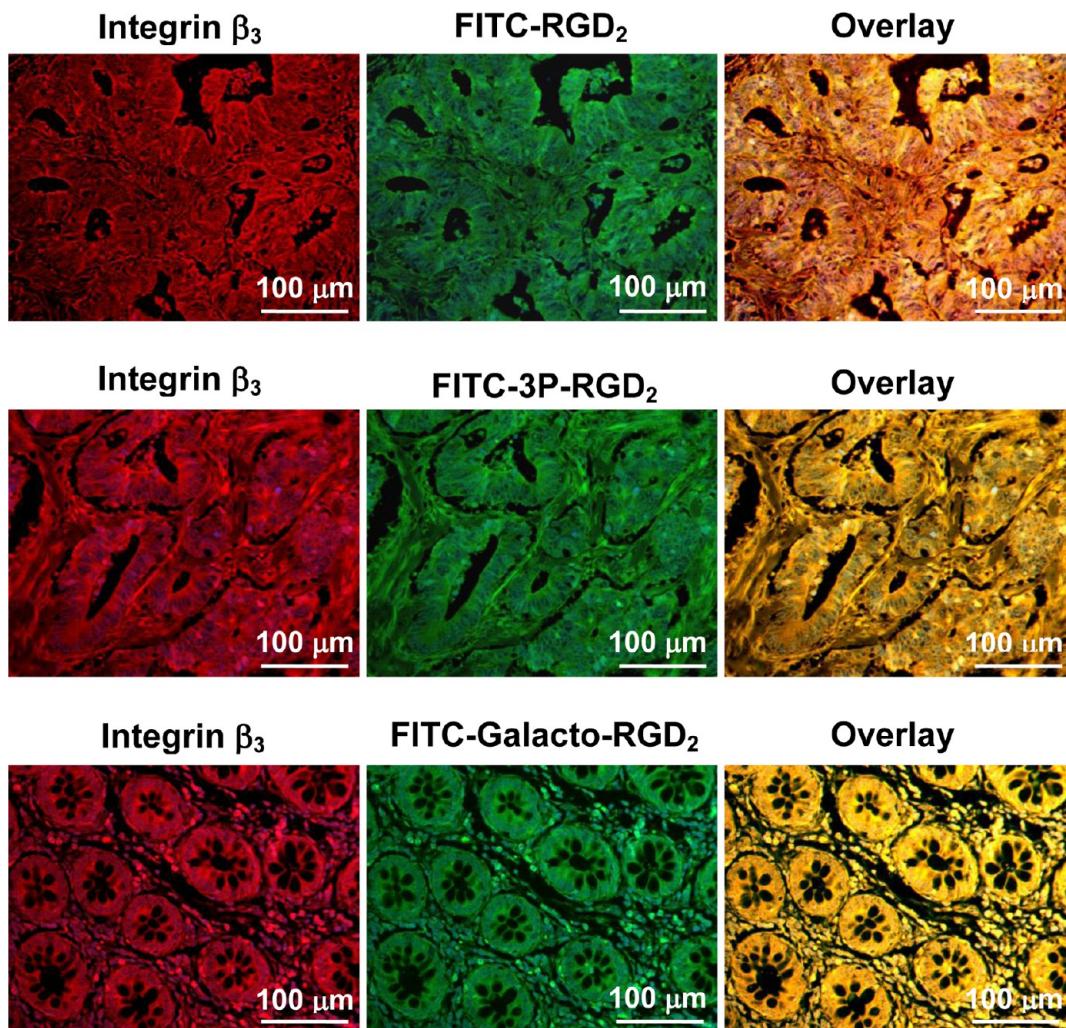
$\alpha_v\beta_3$  expression levels obtained from the anti-integrin  $\beta_3$  antibody.

**Linear Relationship between Tumor Uptake of  $^{99m}\text{Tc}$ -3P-RGD<sub>2</sub> and Relative Integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  Expression Level.**  $^{99m}\text{Tc}$ -3P-RGD<sub>2</sub> is an integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -targeted SPECT radiotracer under clinical evaluations for tumor imaging.<sup>72,73</sup> Figure 10E,F shows the plots of its %ID/g tumor uptake (radioactivity density) and the relative integrin  $\beta_3$  expression levels (fluorescence density) determined with FITC-Galacto-RGD<sub>2</sub> or the integrin  $\beta_3$  antibody in five different xenografted tumors (U87MG, MDA-MB-435, A549, HT29, and PC-3). It was found that there was an excellent linear relationship between the %ID/g tumor uptake of  $^{99m}\text{Tc}$ -3P-RGD<sub>2</sub> and relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels with  $R^2$  being 0.89 for FITC-Galacto-RGD<sub>2</sub>, and the relative integrin  $\alpha_v\beta_3$  expression levels with  $R^2$  being 0.86 for the anti-integrin  $\beta_3$  antibody. Therefore, the relative tumor integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels can be reflected by both %ID/g tumor uptake of  $^{99m}\text{Tc}$ -3P-RGD<sub>2</sub> and the fluorescence density determined by tissue staining with FITC-Galacto-RGD<sub>2</sub>.

**Integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  Expression on Tumor Neovasculature.** It is well-established that the integrins  $\alpha_v\beta_3/\alpha_v\beta_5$  are overexpressed on tumor cells and tumor neovascula-

ture.<sup>36–39,60–62</sup> The overlay experiments between FITC-Galacto-RGD<sub>2</sub> (green color) and CD31 antibody (red color) were performed using five different xenografted tumors (U87MG, MDA-MB-435, A549, HT29, and PC-3). CD31 is a biomarker for endothelial cells on blood vessels. In the overlay images (Figure 11), orange and yellow colors indicated the presence of integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  on neovasculature. Even though it was difficult to quantify the relative tumor vasculature integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels, the results from the overlay images clearly showed that the FITC-conjugated cyclic RGD peptides (e.g., FITC-Galacto-RGD<sub>2</sub>) were indeed able to bind to integrins  $\alpha_v\beta_3/\alpha_v\beta_5$  on the tumor neovasculature. Similar results were reported in the overlay experiments using fluorescence-labeled integrin  $\alpha_v\beta_3$  and CD31 antibodies.<sup>60,61</sup> If there were no yellow or orange color (as in the case of the PC3 prostate model), there would be very little integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression on the tumor vasculature.

**Quantification of Integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  Expression Levels in Human Carcinoma Tissues.** Because of the vasculature difference between the xenografted tumors and human carcinoma tissues, we also evaluated FITC-Galacto-RGD<sub>2</sub> for its utility as a fluorescent probe to quantify the relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels in human carcinoma tissues. Figure

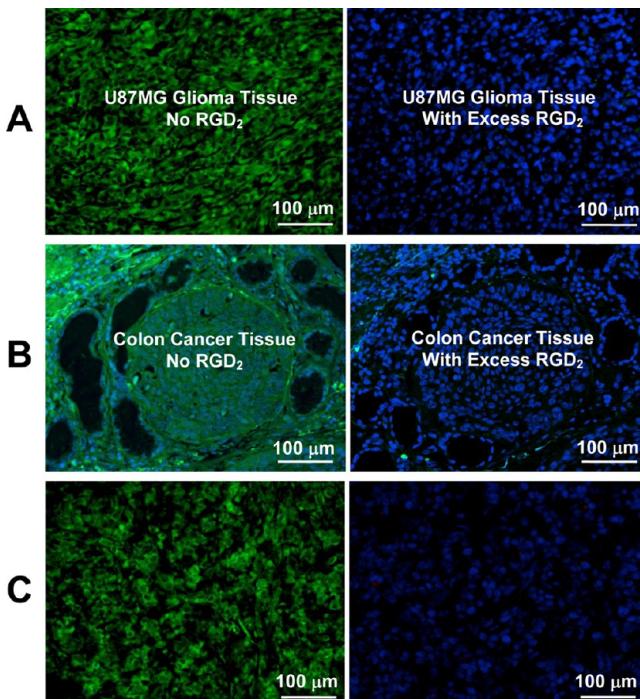


**Figure 6.** Selected microscopic images (200 $\times$  magnification) of colon carcinoma tissues stained with FITC-conjugated cyclic RGD peptide (green) and rabbit anti-human integrin  $\beta_3$  antibody detected with TR conjugated goat anti-rabbit antibody (red). Blue color indicates the presence of nuclei stained with DAPI. In overlay images, the orange and yellow colors (red integrin  $\beta_3$  merged with green cyclic RGD peptide) indicate co-localization of FITC-labeled cyclic RGD peptides (FITC-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-Galacto-RGD<sub>2</sub>) with anti-integrin  $\beta_3$  antibody.

12 compares macroscopic images of six human carcinoma tissues (colon cancer, pancreatic cancer, lung adenocarcinoma, squamous cell lung cancer, gastric cancer, and esophageal cancer) stained with both FITC-Galacto-RGD<sub>2</sub> and rabbit anti-human integrin  $\beta_3$  antibody. The quantitative analysis data were summarized in Figure 13 for human cancer tissues stained with FITC-Galacto-RGD<sub>2</sub> (green for integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ ) and rabbit anti-integrin  $\beta_3$  antibody (red for integrin  $\alpha_v\beta_3$ ). It was found that the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels followed a general order: colon cancer > pancreatic cancer > lung adenocarcinoma  $\approx$  squamous cell lung cancer  $\gg$  gastric cancer  $\approx$  esophageal cancer. There was an excellent linear relationship between the fluorescence density determined with FITC-Galacto-RGD<sub>2</sub> (relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels) and that with anti-human integrin  $\beta_3$  antibody (integron  $\alpha_v\beta_3$  expression levels). The FITC-conjugated cyclic RGD peptides (e.g., FITC-3P-RGD<sub>2</sub> and FITC-Galacto-RGD<sub>2</sub>) are useful for semi-quantification of relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels in human carcinoma tissues.

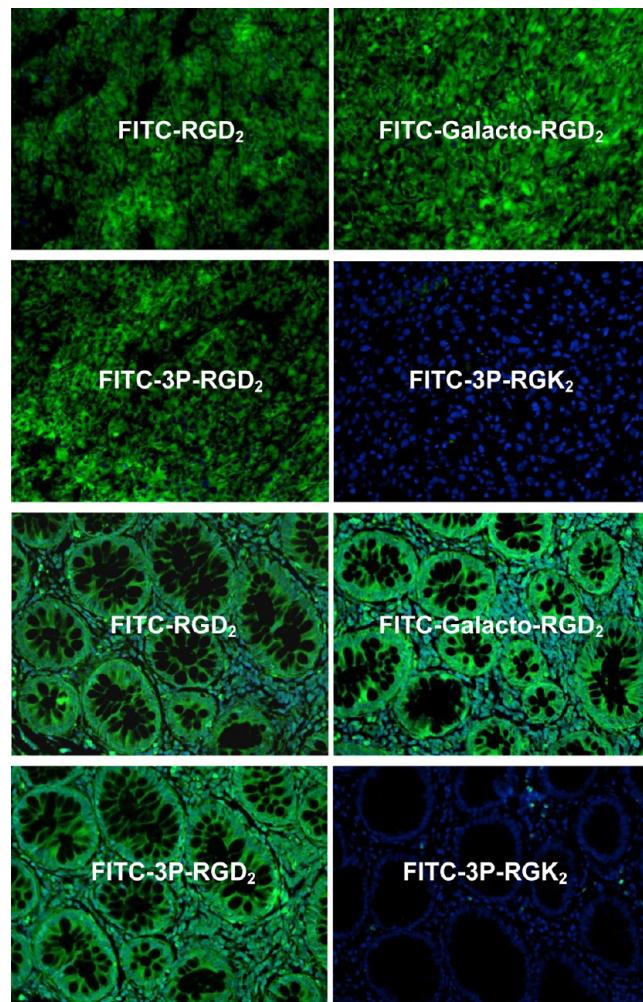
## ■ DISCUSSION

In this study, we found that FITC-3P-RGD<sub>2</sub> and FITC-Galacto-RGD<sub>2</sub> are excellent fluorescent probes for staining integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  in tumor tissues. The FITC-conjugated cyclic RGD peptides are as effective as the fluorescence-labeled anti-integrin  $\beta_3$  antibody despite the fact that cyclic RGD peptides target both integrin  $\alpha_v\beta_3$  and integrin  $\alpha_v\beta_5$ . The integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  specificity was demonstrated by the blocking experiments (Figure 7) and the RGD-specificity with the use of “nonsense” peptide conjugate FITC-3P-RGK<sub>2</sub> (Figure 8). There is also an excellent linear relationship between the relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels determined with FITC-Galacto-RGD<sub>2</sub> and the relative integrin  $\alpha_v\beta_3$  expression levels obtained with anti-integrin  $\beta_3$  antibody in the xenografted tumor tissues (U87MG, MDA-MB-435, A549, HT29, and PC-3) (Figure 10C,D) and in the human carcinoma tissues (Figure 13C,D). There are three possible explanations for this linear relationship. First, the anti-human integrin  $\beta_3$  antibody can bind to both  $\alpha_v\beta_3$  and  $\alpha_v\beta_5$  integrins. This explanation seems unlikely because the rabbit anti-human integrin  $\beta_3$  antibody (sc-14009) has been validated to be specific to integrin  $\beta_3$ . The second possibility is that the integrin  $\alpha_v\beta_5$  expression correlates



**Figure 7.** (A) Selected microscopic images (Magnification: 400×) of living U87MG glioma cells stained with FITC-Galacto-RGD<sub>2</sub> in the absence (left) and presence (right) of excess RGD<sub>2</sub>. (B) Microscopic images (Magnification: 200×) of a tumor slice stained with FITC-Galacto-RGD<sub>2</sub> in the absence (left) and presence (right) of excess RGD<sub>2</sub>. (C) Microscopic images (Magnification: 200×) of the tumor slice (left), which was obtained from a tumor-bearing mouse administered with FITC-Galacto-RGD<sub>2</sub> at a dose of 300 μg. Staining with hamster anti-mouse integrin β<sub>3</sub> antibody (right) detected with Cy3 conjugated goat anti-hamster antibody (red) was not successful due to blockage of integrin α<sub>v</sub>β<sub>3</sub> by administration of excess FITC-Galacto-RGD<sub>2</sub>. (D) Microscopic images (Magnification: 200×) of human colon cancer slice stained with FITC-Galacto-RGD<sub>2</sub> in the absence (left) and presence (right) of excess RGD<sub>2</sub>. Blue color indicates the nuclei.

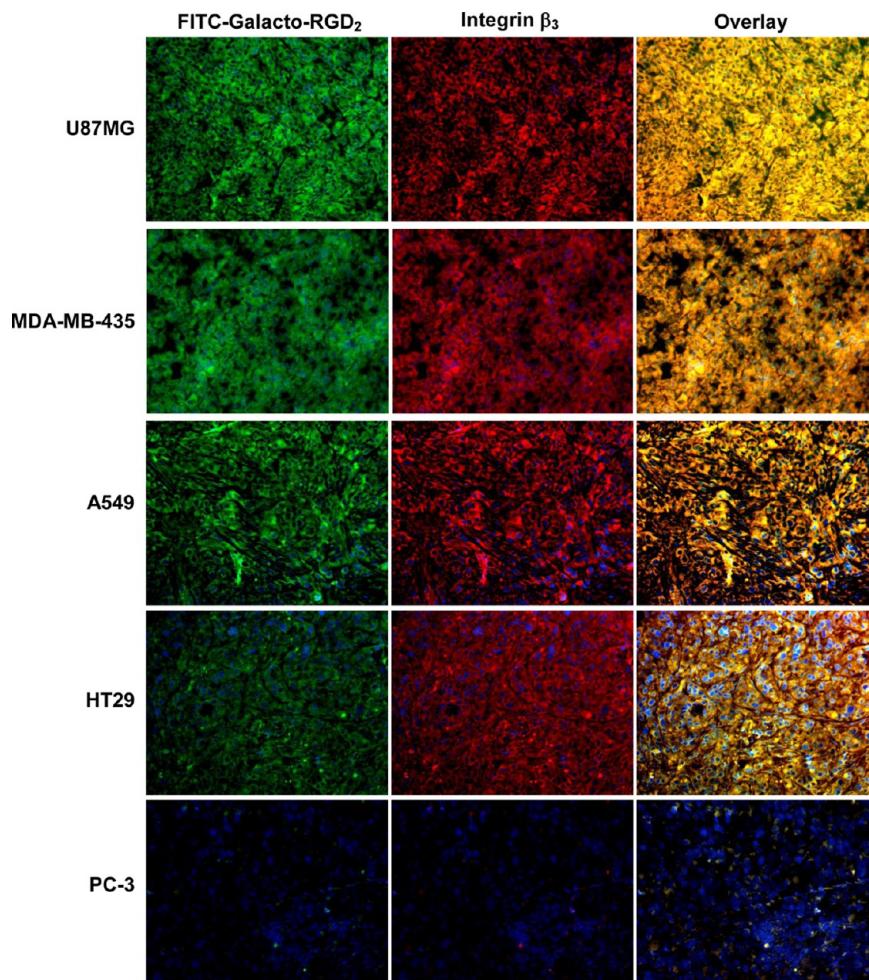
linearly with the integrin α<sub>v</sub>β<sub>3</sub> expression levels. This explanation is problematic since integrins α<sub>v</sub>β<sub>3</sub> and α<sub>v</sub>β<sub>5</sub> are expressed at different levels on various tumor cells and tumor vasculature. Finally, the contribution from integrin α<sub>v</sub>β<sub>5</sub> is much smaller as compared to that of integrin α<sub>v</sub>β<sub>3</sub>. This explanation is supported by careful examination of the slopes for the lines in Figure 10D (slope = 0.96 for five different xenografted tumor tissues) and Figure 13D (slope = 0.97 for six human carcinoma tissues). If both FITC-Galacto-RGD<sub>2</sub> and the anti-human integrin β<sub>3</sub> antibody (sc-14009) target only integrin α<sub>v</sub>β<sub>3</sub>, the slope of these two lines is expected to be 1.0. Obviously, the binding of FITC-Galacto-RGD<sub>2</sub> to integrin α<sub>v</sub>β<sub>5</sub> makes the slopes smaller than 1.0. The slight deviation suggests that only 3–4% of the fluorescent signals are from the integrin α<sub>v</sub>β<sub>5</sub>. If the slope of these lines were to be 1.0, there would have been no contribution from integrin α<sub>v</sub>β<sub>3</sub> and other integrins. One might argue that integrin α<sub>v</sub>β<sub>5</sub> is highly expressed on HT29 cells,<sup>74</sup> and U87MG human glioma cells have high expression of both integrin α<sub>v</sub>β<sub>3</sub> and integrin α<sub>v</sub>β<sub>5</sub>.<sup>75–79</sup> It is important to note that fluorescent intensity represents the total contribution from both tumor cells and neovasculature. It has been estimated that the % contribution from the tumor neovasculature to the total integrin α<sub>v</sub>β<sub>3</sub>/α<sub>v</sub>β<sub>5</sub> expression and tumor



**Figure 8.** (Top four images) Representative microscopic images of human colon cancer tissues (magnification: 200×) stained with FITC-RGD<sub>2</sub>, FITC-Galacto-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-3P-RGK<sub>2</sub>. (Bottom four images) Selected microscopic images of U87MG glioma tumor tissues (magnification: 200×) stained with FITC-RGD<sub>2</sub>, FITC-Galacto-RGD<sub>2</sub>, FITC-3P-RGD<sub>2</sub>, and FITC-3P-RGK<sub>2</sub>. Blue color indicates the presence of nuclei.

uptake of <sup>99m</sup>Tc-3P-RGD<sub>2</sub> is ~60% in the xenografted U87MG glioma model.<sup>80</sup> In the case of xenografted HT29 tumors, the main contribution to the total fluorescent intensity or the tumor uptake of <sup>99m</sup>Tc-3P-RGD<sub>2</sub> is actually from the integrin α<sub>v</sub>β<sub>3</sub> on new blood vessels.<sup>60,62</sup>

There is always a debate regarding whether one should separate α<sub>v</sub>β<sub>3</sub> from α<sub>v</sub>β<sub>5</sub>. Integrins α<sub>v</sub>β<sub>3</sub> and α<sub>v</sub>β<sub>5</sub> both play a significant role in tumor angiogenesis,<sup>3,4,6–10</sup> and often co-localize despite their different biological functions.<sup>16,30,39</sup> Several integrin family members (including, but not limited to, α<sub>v</sub>β<sub>3</sub>, α<sub>v</sub>β<sub>5</sub>, α<sub>v</sub>β<sub>6</sub>, and α<sub>v</sub>β<sub>1</sub>) are receptors of the RGD-containing extracellular matrix proteins (e.g., vitronectin, fibronectin, fibrinogen, laminin, collagen, and osteoponin).<sup>1–12</sup> As long as the biomolecule contains one or more RGD tripeptide sequences, it will target integrins α<sub>v</sub>β<sub>3</sub>, α<sub>v</sub>β<sub>5</sub>, α<sub>v</sub>β<sub>6</sub>, and α<sub>v</sub>β<sub>1</sub> regardless of its multiplicity. From this point of view, there is no need to differentiate them for the development of radiotracers and fluorescent probes. This is the exact reason we use the whole-cell displacement assay with U87MG glioma cells (high expression of integrin α<sub>v</sub>β<sub>3</sub> and integrin α<sub>v</sub>β<sub>5</sub>) as the host



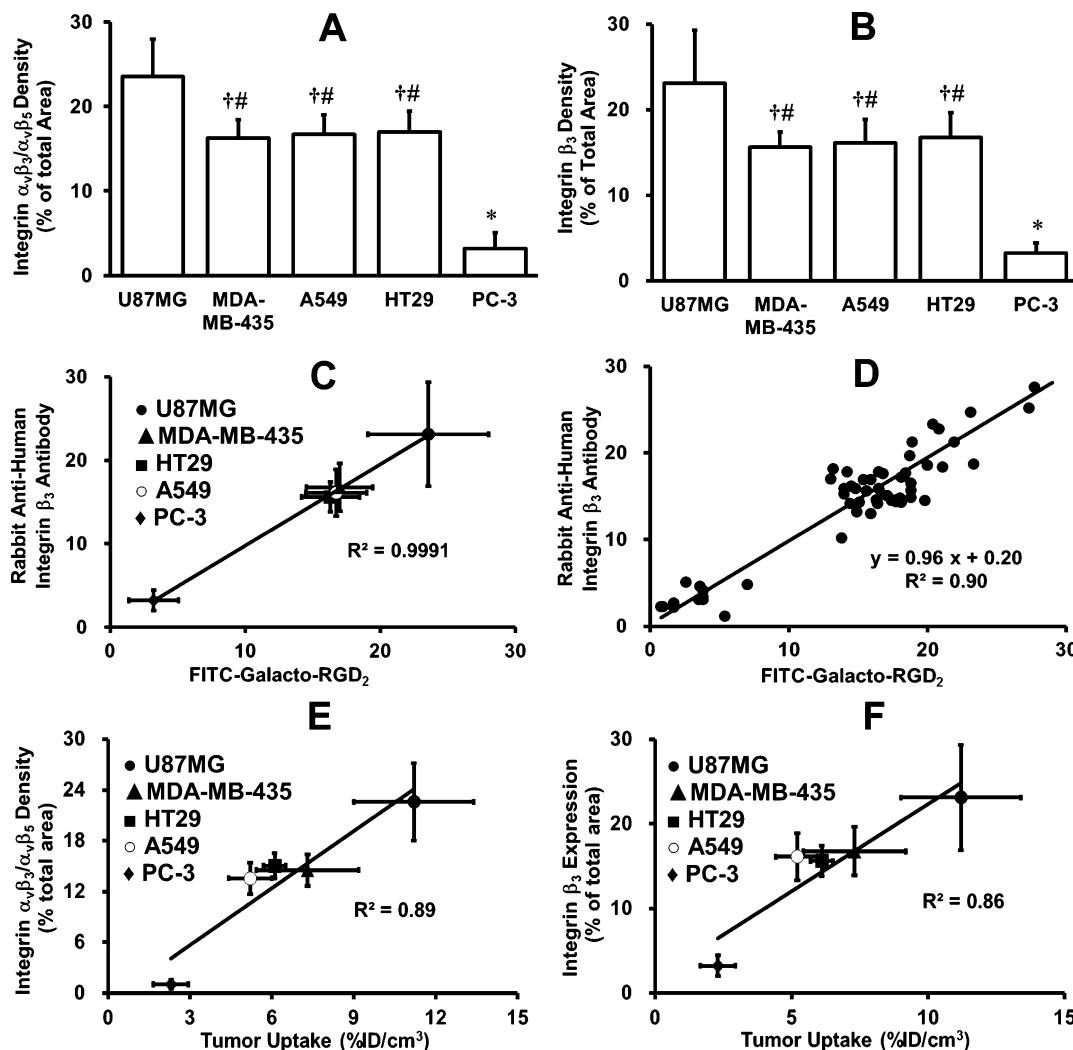
**Figure 9.** Representative fluorescence microscopic images (Magnification: 200 $\times$ ) of selected tumor slices from five xenografted tumors stained with FITC-Galacto-RGD<sub>2</sub> (green) and rabbit anti-human integrin  $\beta_3$  antibody detected with Cy3 conjugated goat anti-rabbit antibody (red). Orange or yellow in overlay image indicates co-localization of FITC-Galacto-RGD<sub>2</sub> (green) and integrin  $\beta_3$  antibody for tumor tissue staining of integrin  $\alpha_v\beta_3$ .

cells and <sup>125</sup>I-echistatin, which is a <sup>125</sup>I-labeled family member of disintegrins, as the radioligand in this study. Another important finding from this study is the linear relationship (Figure 10E,F) between the %ID/g tumor uptake of <sup>99m</sup>Tc-3P-RGD<sub>2</sub> and the fluorescence density from the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  staining with FITC-Galacto-RGD<sub>2</sub>. Visualization of integrins  $\alpha_v\beta_3/\alpha_v\beta_5$  with FITC-Galacto-RGD<sub>2</sub> makes it easier to correlate the in vitro tumor tissue staining data with the in vivo findings with the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -targeted radiotracers, such as <sup>99m</sup>Tc-3P-RGD<sub>2</sub> and [<sup>18</sup>F]Galacto-RGD (2-[<sup>18</sup>F]-fluoropropanamide-c(RGDFK(SAA); SAA = 7-amino-L-glyero-L-galacto-2,6-anhydro-7-deoxyheptanamide).<sup>55–59</sup> Such a combination of differently labeled analogs of the same cyclic peptide will improve the integration of both in vitro and in vivo diagnostics.

The tissue staining kinetics of the xenografted U87MG tumor and human colon cancer tissues with FITC-Galacto-RGD<sub>2</sub> (<30 min) is faster than that with anti-human integrin  $\beta_3$  antibody (>90 min required for blocking nonspecific binding, tissue staining, and attachment of secondary polyclonal antibody). This difference might be caused by the smaller size of FITC-Galacto-RGD<sub>2</sub>. Alternatively, this difference may also be caused by higher concentration of FITC-Galacto-RGD<sub>2</sub> (>5  $\mu$ M) than that of integrin  $\beta_3$  antibody (2.5  $\mu$ g/mL or ~12 nM), as illustrated by the concentration dependence of staining in

kinetics for human cancer tissues (Supporting Information Figure SI1). It is important to note that the staining kinetics of colon cancer tissue is faster than that of the xenografted U87MG tissues using the same fluorescent probe FITC-Galacto-RGD<sub>2</sub> under similar conditions. We believe that this difference might be related to the total integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels because the tumor tissues with higher integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression tend to have a significantly faster tissue staining kinetics (Supporting Information Figure SI2) at a much lower concentration (Supporting Information Figure SI1).

There are several advantages in using FITC-conjugated cyclic RGD peptides over the fluorescence-labeled integrin  $\alpha_v\beta_3$  antibodies. The staining procedures with FITC-conjugated cyclic RGD peptides are much simpler than those with fluorescence-labeled integrin  $\alpha_v\beta_3$  antibodies since the latter involves the blocking of nonspecific binding and the use of secondary antibody. There is a significant variability in the performance of fluorescence-labeled integrin  $\alpha_v\beta_3$  antibodies, depending on their origin (human, mouse, hamster, rabbit, or goat). In some cases, the results from tumor tissue staining may vary using different batches of antibodies from the same commercial source. In contrast, FITC-conjugated cyclic RGD peptides have high solution stability, and can be stored for a long time. They are useful for the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  staining in

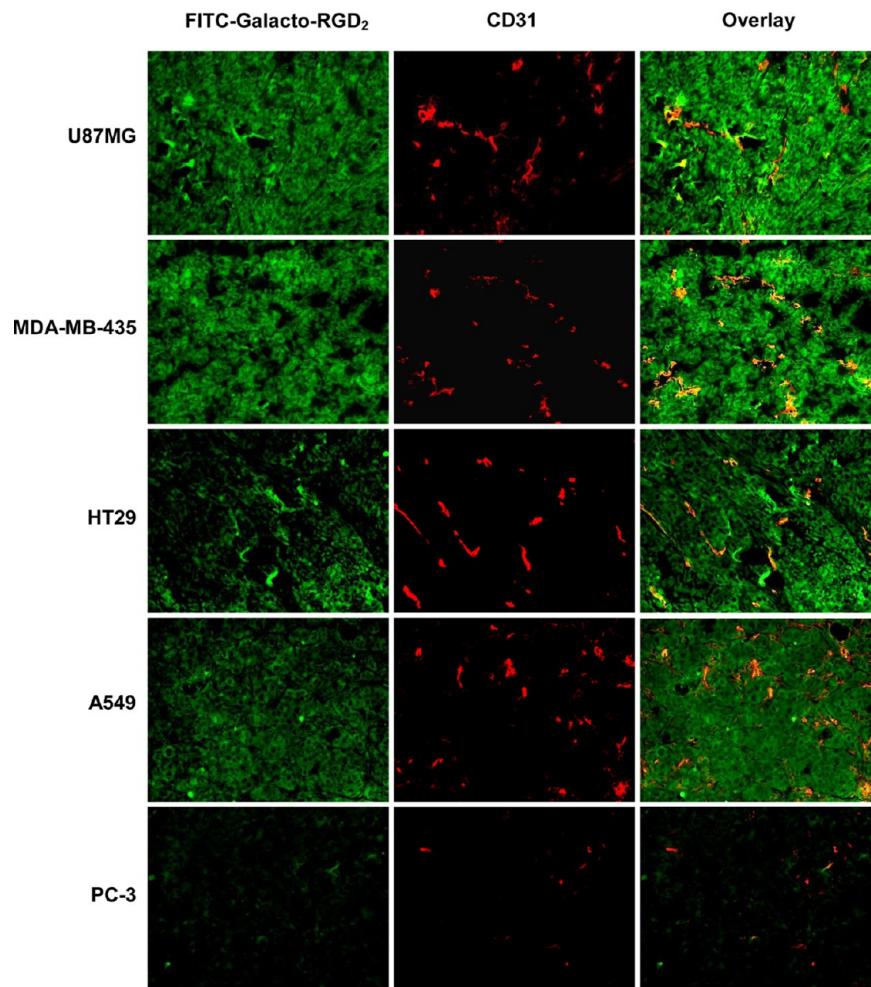


**Figure 10.** (A) Quantitative analysis of integrin  $\alpha_v\beta_3$  density for five xenografted tumor tissues (U87MG, MDAMB-435, A549, HT29, and PC-3) stained with FITC-Galacto-RGD<sub>2</sub> (green). Integrin  $\alpha_v\beta_3$  density on tumor cells and neovasculature is represented by the percentage of green area over the total area in each tumor slice. (B) Quantitative analysis of integrin  $\beta_3$  density for five different xenografted tumor tissues (U87MG, MDAMB-435, A549, HT29, and PC-3) stained with rabbit anti-human integrin  $\beta_3$  antibody detected with the Cy3-conjugated goat anti-rat antibody (red). Integrin  $\alpha_v\beta_3$  density on tumor cells and neovasculature is represented by the percentage of red area over the total area in each slice of tumor tissue. (C) (average data) and (D) (single data): Linear relationship between the relative integrin  $\alpha_v\beta_3$  expression levels (fluorescence density) determined with FITC-Galacto-RGD<sub>2</sub> and those obtained with rabbit anti-human integrin  $\beta_3$  antibody in five xenografted tumors (U87MG, MDA-MB-435, A549, HT29, and PC-3). Each data point was derived from 15 different areas of the same tumor slice. Experiments were repeated three times independently with very similar results. All experimental values are reported as the average plus/minus standard deviation. \*:  $p < 0.01$ , significantly different from all other groups; †:  $p < 0.01$ , significantly different from the U87MG group; #:  $p < 0.01$ , significantly different from the PC-3 group. (E) (FITC-Galacto-RGD<sub>2</sub>) and (F) (rabbit anti-human integrin  $\beta_3$  antibody): Linear relationship between the relative integrin  $\alpha_v\beta_3$  expression levels (fluorescence density) determined with FITC-Galacto-RGD<sub>2</sub> or rabbit anti-human integrin  $\beta_3$  antibody and the tumor uptake values of <sup>99m</sup>Tc-3P-RGD<sub>2</sub> in five xenografted tumors (U87MG, MDA-MB-435, A549, HT29, and PC-3). The tumor uptake values of <sup>99</sup>mTc-3P-RGD<sub>2</sub> were obtained from our previous studies.<sup>63</sup>

tumor tissues regardless of their origin (human vs murine). There is no need to block the nonspecific binding because they are integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  and RGD-specific. There is no need to use secondary antibody as fluorescence label because FITC can provide sufficient fluorescence signal to obtain excellent macroscopic images of integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -positive tumor tissues. There is also a significant disadvantage associated with FITC-conjugated cyclic RGD peptides since they bind several integrin family members ( $\alpha_v\beta_3$ ,  $\alpha_v\beta_5$ ,  $\alpha_v\beta_6$ , and  $\alpha_5\beta_1$ ). However, the tumor staining data with FITC-Galacto-RGD<sub>2</sub> can be used to reflect the relative integrin  $\alpha_v\beta_3$  expression levels because of the linear relationship between the fluorescent intensity with FITC-Galacto-RGD<sub>2</sub> and the integrin  $\alpha_v\beta_3$

expression levels determined with anti-integrin  $\beta_3$  antibody in the xenografted tumor tissues (Figure 10C,D) and in the human carcinoma tissues (Figure 13C,D).

The heterogeneity in fluorescence distribution is characteristic of the xenografted tumors (U87MG, MDA-MB-435, A549, and HT29). In general, larger tumors (>0.5 g) tend to have more necrosis. In necrotic regions, there is little integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression, as indicated by lack of fluorescence in the area.<sup>60,61</sup> It is not surprising that the human carcinoma tissues showed little integrin  $\alpha_v\beta_3$  staining with the fluorescence-labeled integrin  $\alpha_v\beta_3$  monoclonal antibodies even though the PET imaging data clearly showed high tumor uptake of [<sup>18</sup>F]Galacto-RGD in cancer patients.<sup>55–59</sup> Thus, the selection



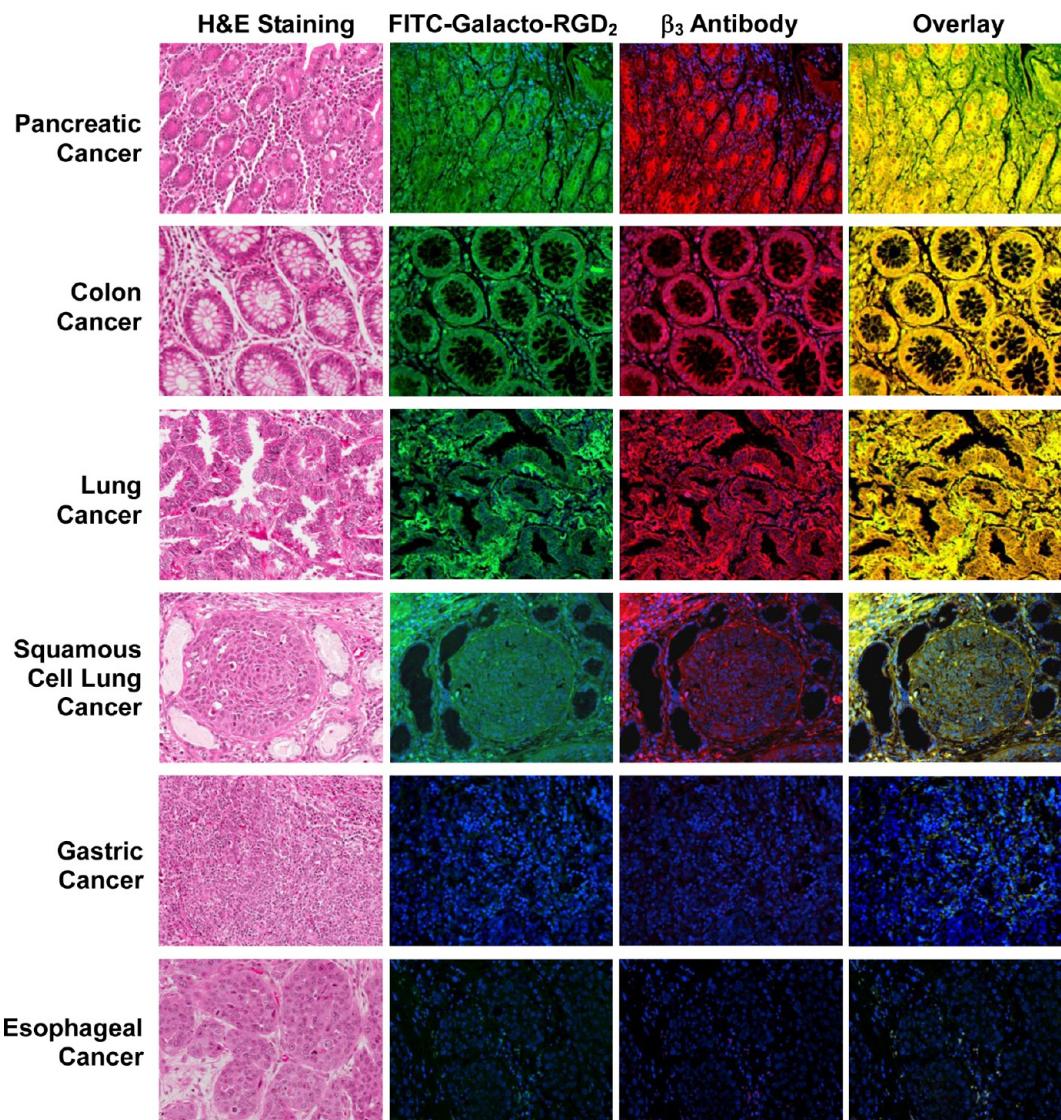
**Figure 11.** Representative fluorescence microscopic images (Magnification: 200 $\times$ ) of selected tumor slices stained with FITC-Galacto-RGD<sub>2</sub> (green) and rat anti-mouse CD31 antibody detected with Cy3 conjugated goat anti-rat antibody (red). Orange or yellow color in overlay images indicates co-localization of FITC-Galacto-RGD<sub>2</sub> (green) and CD31 antibody in tumor tissues.

of tumor biopsy samples becomes important for IHC staining of human carcinoma tissues with the FITC-conjugated cyclic RGD peptides or fluorescence-labeled integrin  $\alpha_v\beta_3$  antibodies. Since the human carcinoma tissues were only small parts of the whole tumor mass from cancer patients, caution must be taken when interpreting the quantification data in human carcinoma tissues stained with a FITC-conjugated cyclic RGD peptide or fluorescence-labeled integrin  $\alpha_v\beta_3$  antibody. The integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels only represent the status of that specific tissue (not whole tumor mass). In contrast, the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -targeted PET and SPECT radiotracers (e.g., <sup>99m</sup>Tc-3P-RGD<sub>2</sub> and [<sup>18</sup>F]Galacto-RGD) are better suited for determination and visualization of the tumor integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  heterogeneity due to the capability of PET and SPECT for in vivo imaging of entire tumor in cancer patients. Such a combination use of differently labeled (FITC vs <sup>99m</sup>Tc or <sup>18</sup>F) derivatives of the same cyclic RGD peptide would improve the integration in assessment of integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels. This statement is completely consistent with the linear relationship (Figure 10E) between the %ID/g tumor uptake of <sup>99m</sup>Tc-3P-RGD<sub>2</sub> in the xenografted tumors (U87MG, MDA-MB-435, A549, HT29, and PC-3) and the fluorescent intensity obtained from semiquantification of the same five xenografted tumor tissues stained with FITC-Galacto-RGD<sub>2</sub>. It must be noted that quantification of absolute fluorescence intensity is

operator-dependent. The relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels only represent the status under specific experimental conditions (animal species and sexes, inoculation location, tumor cell types, tumor growth time, and tumor sizes).

Theoretically, FITC-Galacto-RGD<sub>2</sub> would predominantly localize in the areas rich in blood vessels when it is injected intravenously. However, our results show that FITC-Galacto-RGD<sub>2</sub> distributes in the whole tumor tissue in a relatively homogeneous fashion (Figure 7C). This might be caused by the leaky nature of microvessels and its long tumor retention time, during which FITC-Galacto-RGD<sub>2</sub> could easily diffuse through the vessel walls and bind to all integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  sites in the whole tumor. This finding is important because it shows that FITC-conjugated cyclic RGD peptides might be useful as fluorescent probes to correlate the in vitro findings with those in vivo data using the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -targeted PET or SPECT radiotracers. This statement is supported by the linear relationship (Figure 10E) between the radioactivity density (% ID/g tumor uptake of <sup>99m</sup>Tc-3P-RGD<sub>2</sub>) and the integrin  $\alpha_v\beta_3$  density obtained from fluorescent signal quantification of tumor tissues stained with FITC-Galacto-RGD<sub>2</sub>.

The results from this study also show that FITC-Galacto-RGD<sub>2</sub> is able to stain integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  on tumor neovasculature despite the difficulty for quantitative analysis. It is important to note that the quantification of fluorescent signals



**Figure 12.** Representative fluorescence microscopic images (Magnification: 200 $\times$ ) of selected tumor slices from six different human tumors (colon cancer, pancreatic cancer, lung cancer, squamous cell lung cancer, gastric cancer, and esophageal cancer) stained with FITC-Galacto-RGD<sub>2</sub> (green) and rabbit anti-human integrin  $\beta_3$  antibody detected with Cy3 conjugated goat anti-rabbit antibody (red). Orange or yellow in overlay image indicates co-localization of FITC-Galacto-RGD<sub>2</sub> (green) and integrin  $\beta_3$  antibody for tumor tissue staining of integrin  $\alpha_v\beta_3$ . H&E staining data were used for pathological characterization of human carcinoma tissues.

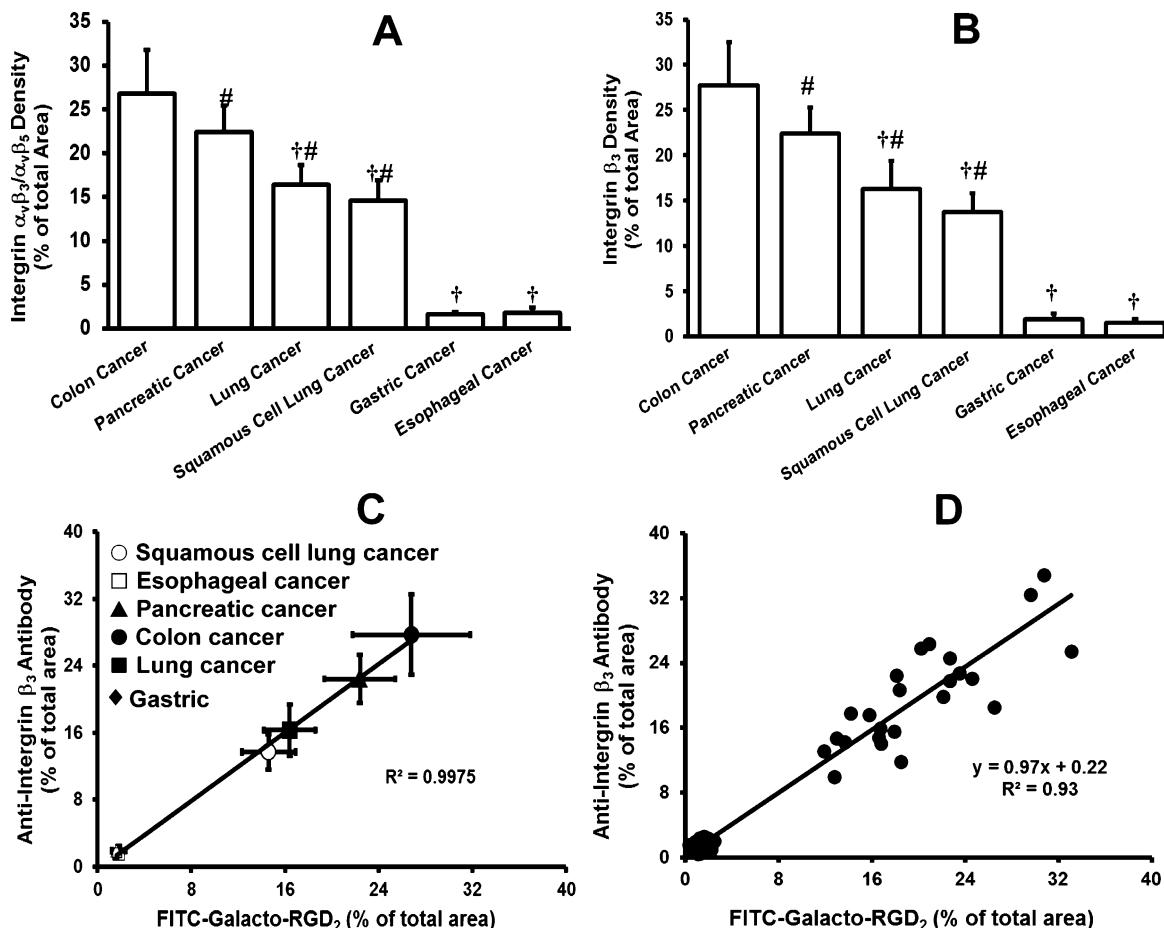
in each tumor slice reflects the total integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression on both tumor cells and neovasculature, and not all tumor blood vessels express integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ . In fact, some tumor blood vessels have little integrin  $\beta_3$  expression in the xenografted U87MG glioma tissues as indicated by the red-color in the overlay images (Figure 11). Since the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  is overexpressed only on tumor microvessels (not mature vessels), caution must be taken when correlating the radiotracer tumor uptake and blood vessel density (CD31 as a biomarker).

In this study, the FITC-conjugated cyclic RGD peptides (e.g., FITC-Galacto-RGD<sub>2</sub>) are used only as examples to demonstrate proof-of-concept for the general approach to stain tumor tissues with small peptide-based fluorescent probes. Theoretically, this approach applies to any receptor-based fluorescent probe, as long as the targeting biomolecule has sufficient receptor binding affinity and specificity. Many radiolabeled peptides have been evaluated as target-specific

radiotracers for molecular imaging of tumors by SPECT or PET.<sup>40–52</sup> These peptides could be readily conjugated to a fluorescent label to produce the corresponding fluorescent probes. In addition, FITC could be easily replaced by other fluorescent dyes (such as red-colored cyanine derivatives) if needed.

## ■ CONCLUSION

The results from this study clearly illustrate the value of the FITC-conjugated cyclic RGD peptides (particularly FITC-3P-RGD<sub>2</sub> and FITC-Galacto-RGD<sub>2</sub>) for the in vitro assays of integrins  $\alpha_v\beta_3/\alpha_v\beta_5$ . They have very high solution stability and faster integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding kinetics. FITC-3P-RGD<sub>2</sub> and FITC-Galacto-RGD<sub>2</sub> are excellent fluorescent probes for assaying relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression levels in tumor tissues.



**Figure 13.** (A) Quantitative analysis of integrin  $\alpha_v\beta_3$  density for six different human cancer tissues (colon cancer, pancreatic cancer, lung adenocarcinoma, squamous cell lung cancer, gastric cancer, and esophageal cancer) stained with FITC-Galacto-RGD<sub>2</sub> (green). Integrin  $\alpha_v\beta_3$  density on tumor cells and neovasculature is represented by the percentage of green area over the total area in each tumor slice. (B) Quantitative analysis of integrin  $\beta_3$  density for six human cancer tissues (colon cancer, pancreatic cancer, lung adenocarcinoma, squamous cell lung cancer, gastric cancer, and esophageal cancer) stained with rabbit anti-human integrin  $\beta_3$  antibody detected with the TR-conjugated goat anti-rat antibody (red). Integrin  $\alpha_v\beta_3$  density on tumor cells and neovasculature is represented by the percentage of red area over the total area in each slice of tumor tissue. (C) (average data) and (D) (single data): Linear relationship between the relative integrin  $\alpha_v\beta_3$  expression levels (fluorescence density) determined with FITC-Galacto-RGD<sub>2</sub> and those obtained with rabbit anti-human integrin  $\beta_3$  antibody in six human tumors (colon cancer, pancreatic cancer, lung adenocarcinoma, squamous cell lung cancer, gastric cancer, and esophageal cancer). Each data point was derived from 15 different areas of the same tumor slice. Experiments were repeated three times independently with very similar results. All values were reported as the average plus/minus standard deviation. †:  $p < 0.01$ , significantly different from the colon cancer group; #:  $p < 0.01$ , significantly different from the gastric cancer group.

## EXPERIMENTAL SECTION

**Materials and Instruments.** Common chemicals and solvents were purchased from Sigma/Aldrich (St. Louis, MO), and were used without further purification. Fluorescein isothiocyanate isomer I (FITC) was purchased from AnaSpec, Inc. (Fremont, CA). Cyclic peptides E[c(RGDFK)]<sub>2</sub> (RGD<sub>2</sub>), PEG<sub>4</sub>-E[PEG<sub>4</sub>-c(RGDFK)]<sub>2</sub> (3P-RGD<sub>2</sub>), and PEG<sub>4</sub>-E[PEG<sub>4</sub>-c(RGKfK)]<sub>2</sub> (3P-RGK<sub>2</sub>) were purchased from Peptides International, Inc. (Louisville, KY). Glu[cyclo[Arg-Gly-Asp-D-Phe-Lys(SAA-PEG<sub>2</sub>-(1,2,3-triazole)-1-yl-4-methylamide)]]<sub>2</sub> (Galacto-RGD<sub>2</sub>) was prepared according to the procedures described in our previous report.<sup>64</sup> The MALDI (matrix-assisted laser desorption ionization) mass spectral data were collected on an Applied Biosystems Voyager DE PRO mass spectrometer (Framingham, MA), in the Department of Chemistry, Purdue University.

**HPLC Methods.** The HPLC method used a LabAlliance HPLC system (Scientific Systems, Inc., State College, PA) equipped with a UV-vis detector ( $\lambda = 220$  nm) and Zorbax C<sub>18</sub>

column (9.4 mm  $\times$  250 mm, 100 Å pore size; Agilent Technologies, Santa Clara, CA). The flow rate was 2.5 mL/min with a mobile phase being 90% A (0.05% TFA in water) and 10% B (0.05% TFA in acetonitrile) at 0 min to 85% A and 15% B at 5 min, and to 75% A and 25% B at 20 min.

**FITC-E[c(RGDFK)]<sub>2</sub> (FITC-RGD<sub>2</sub>).** RGD<sub>2</sub> (3.1 mg, 2.35  $\mu$ mol) and FITC (1.2 mg, 3.08  $\mu$ mol) were dissolved in anhydrous DMF (1.5 mL). Upon addition of triethylamine (10  $\mu$ L, 71  $\mu$ mol), the reaction mixture was stirred for 12 h at room temperature. After completion of conjugation, 2 mL of water was added to the reaction mixture. The pH value was then adjusted to 3–4 using neat TFA. The resulting solution was subjected to HPLC purification. Fractions at 16 min were collected, combined, and lyophilized to yield FITC-RGD<sub>2</sub> as a yellow powder (2.5 mg, ~60%). MALDI-MS:  $m/z = 1707.4$  for  $[M + H]^+$  (MW = 1706.69 calcd. for  $[C_{80}H_{98}N_{20}O_{21}S]$ ).

**FITC-PEG<sub>4</sub>-E[PEG<sub>4</sub>-c(RGDFK)]<sub>2</sub> (FITC-3P-RGD<sub>2</sub>).** 3P-RGD<sub>2</sub> (4.15 mg, 2  $\mu$ mol) and FITC (2.5 mg, 6.4  $\mu$ mol) were dissolved in 2 mL of DMF. After addition of excess diisopropylethylamine (DIEA: 50  $\mu$ mol), the reaction mixture

was stirred for 2 days at room temperature. Upon completion of conjugation, 2 mL of water was added to the mixture above. The pH value was adjusted to 3–4 using neat TFA. The resulting solution was subjected to HPLC-purification. The fraction at ~18 min was collected. Lyophilization of collected fractions afforded FITC-3P-RGD<sub>2</sub> as a yellow powder (3.1 mg, ~63%). MALDI-MS: *m/z* = 2450.55 for [M + H]<sup>+</sup> (MW = 2449.68 calcd. for [C<sub>113</sub>H<sub>161</sub>N<sub>23</sub>O<sub>36</sub>S]).

**PEG<sub>4</sub>-E[PEG<sub>4</sub>-c(RGKfD)]<sub>2</sub> (FITC-3P-RGK<sub>2</sub>).** FITC-3P-RGK<sub>2</sub> (4.15 mg, 2  $\mu$ mol) was prepared and purified according to the same procedures for used for FITC-3P-RGD<sub>2</sub>. The fractions at ~18 min were collected, combined, and lyophilized to afford FITC-3P-RGK<sub>2</sub> as a yellow powder (2.3 mg, ~47%). MALDI-MS: *m/z* = 2450.35 for [M + H]<sup>+</sup> (MW = 2449.68 calcd. for [C<sub>113</sub>H<sub>161</sub>N<sub>23</sub>O<sub>36</sub>S]).

**FITC-Glu[cyclo[Arg-Gly-Asp-D-Phe-Lys(SAA-PEG<sub>2</sub>(1,2,3-triazole)-1-yl-4-methylamide)]]<sub>2</sub> (FITC-Galacto-RGD<sub>2</sub>).** Galacto-RGD<sub>2</sub> (6.5 mg, 3.02  $\mu$ mol) and FITC (2.4 mg, 6.15  $\mu$ mol) were dissolved in 2 mL of anhydrous DMF. After addition of excess DIEA (50  $\mu$ mol), the reaction mixture was stirred for 5 days at room temperature. The reaction was terminated by adding 0.5 mL of 25 mM NH<sub>4</sub>OAC solution. The pH value was then adjusted to 3–4 using neat TFA. The resulting solution was subjected to HPLC-purification. The fraction at ~16 min was collected. Lyophilization of the collected fractions afforded FITC-Galacto-RGD<sub>2</sub> as a yellow powder (4.1 mg, ~54%). MALDI-MS: *m/z* = 2538.85 for [M + H]<sup>+</sup> (MW = 2538.62 calcd. for [C<sub>112</sub>H<sub>148</sub>N<sub>30</sub>O<sub>37</sub>S]).

**Cellular Culture.** All human tumor cell lines (U87MG, MDA-MB-435, A549, HT29, and PC-3) were obtained from American Type Culture Collection (ATCC, Manassas, VA). U87MG glioma cells were cultured in the Minimum Essential Medium, Eagle with Earle's Balanced Salt Solution (non-essential amino acids sodium pyruvate, Invitrogen, Carlsbad, CA). PC-3 and A549 cells were cultured in the F-12 medium (GIBCO, Grand Island, NY). MDA-MB-435 and HT29 cells were grown in the RPMI Medium 1640 with L-Glutamine (GIBCO, Grand Island, NY). All human tumor cell lines were supplemented with 10% fetal bovine serum (FBS from Sigma/Aldrich, St. Louis, MO) and 1% penicillin and streptomycin solution (GIBCO, Grand Island, NY), and grown at 37 °C in a humidified atmosphere of 5% CO<sub>2</sub> in air. All tumor cells were grown as monolayers and were harvested or split when they reached 90% confluence to maintain exponential growth.

**Whole-Cell Integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  Binding Assay.** The integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  binding affinity was assessed via a cellular competitive displacement assay using <sup>125</sup>I-echistatin (PerkinElmer, Branford, CT) as integrin-specific radioligand. Experiments were performed using the integrin  $\alpha_v\beta_3/\alpha_v\beta_3$ -positive U87MG glioma cells by slight modification of the literature method.<sup>64–75</sup> Briefly, the filter multiscreen DV plates (Millipore, Billerica, MA) were seeded with 1 × 10<sup>5</sup> U87MG cells in the binding buffer (20 mM Tris, 150 mM NaCl, 2 mM CaCl<sub>2</sub>, 1 mM MnCl<sub>2</sub>, 1 mM MgCl<sub>2</sub>, 0.1% bovine serum albumin (BSA); and pH 7.4) and were incubated with <sup>125</sup>I-echistatin (0.75–1.0 kBq) for 2 h at room temperature in the presence of increasing concentrations of the FITC-conjugated peptides. After removing the unbound <sup>125</sup>I-echistatin, the hydrophilic PVDF filters were washed three times with the binding buffer, and were then collected. Radioactivity was determined using a PerkinElmer Wizard –1480  $\gamma$ -counter (Shelton, CT). All experiments were carried out at least twice with triplicate samples. IC<sub>50</sub> values were calculated by fitting

experimental data with nonlinear regression using GraphPad Prim 5.0 (GraphPad Software, Inc., San Diego, CA), and were reported as an average plus/minus standard deviation. Comparison between two FITC-conjugated cyclic RGD peptides was made using the one-way ANOVA test (GraphPad Prim 5.0, San Diego, CA). The level of significance was set at *p* < 0.05.

**Animal Model.** Animal studies were performed in compliance with the NIH animal experiment guidelines (Principles of Laboratory Animal Care, NIH Publication No. 86–23, revised 1985). The protocol for animal studies was approved by the Purdue University Animal Care and Use Committee (PACUC). Female athymic *nu/nu* mice (4–5 weeks) were purchased from Harlan (Indianapolis, IN), and were implanted subcutaneously with 5 × 10<sup>6</sup> human tumor cells in 0.1 mL of saline into shoulder flanks (U87MG, A549, HT29, and PC-3) or mammary fat pads (MDA-MB-435). Four to six weeks after inoculation, the tumor size was 0.1–0.3 g. The tumors were then harvested for IHC and hematoxylin and eosin (H&E) staining.

**Human Carcinoma Tissues.** Six human carcinoma tissues (colon cancer, pancreatic cancer, lung adenocarcinoma, squamous cell lung cancer, gastric cancer, and esophageal cancer) were obtained from the cancer patients with consent. Paraffin blocks of tumor tissues were obtained from the Department of Pathology, the China-Japan Friendship Hospital (Beijing, P. R. China). Original diagnoses were made between January 2011 and October 2012. All histological specimens were fixed for 12–24 h in neutrally buffered formaldehyde. The use of anonymous or coded leftover material for scientific purposes was part of the standard treatment contract with patients in the China-Japan Friendship Hospital (Beijing, P. R. China). For each case, hematoxylin-eosin (H&E) stained slides of the paraffin blocks were reviewed by pathologists to confirm the malignancy in tumor tissues. Histological type was assessed according to the WHO classification of tumors. The protocol for use of human cancer tissues was approved by the Purdue University Animal Care and Use Committee (PACUC).

**Tumor Tissue Staining.** U87MG, MDA-MB-435, A549, HT29, and PC-3 tumors were harvested from the tumor-bearing mice, and were immediately snap-frozen in OCT (optical cutting temperature) solution, and then cut into slices (5  $\mu$ m). After thorough drying at room temperature, slices were fixed with ice-cold acetone for 10 min, and dried in the air at the room temperature for 20 min. The tumor sections were incubated with 10% goat serum for 30 min at 37 °C to block nonspecific binding. In overlay experiments, the tumor sections were incubated with a FITC-conjugated cyclic RGD peptide (100  $\mu$ M) and the rabbit antihuman  $\beta_3$  antibody (sc-14009, 2.5  $\mu$ g/mL, BD Biosciences, San Jose, CA) for 1 h at room temperature. After washing with PBS buffer, the tumor slides were incubated for 1 h with the Cy3-conjugated goat anti-rabbit antibodies (1:100, V/V Jackson Immuno-Research Inc., West Grove, PA). Negative controls were incubated only with secondary antibody. In blocking experiments, tumor slides were incubated with 10% goat serum, and then with 100  $\mu$ M FITC-Galacto-RGD<sub>2</sub> in the presence of 10 mM c(RGDfK)<sub>2</sub> for 1 h at room temperature. After washing with PBS three times, the tumor slides were mounted with Dapi Fluormount G and cover glass. To demonstrate the RGD specificity, FITC-3P-RGK<sub>2</sub> (100  $\mu$ M) was used as the nonsense peptide. For human carcinoma tissues (colon cancer, pancreatic cancer, lung adenocarcinoma, squamous cell lung cancer, gastric cancer,

and esophageal cancer), the tumor slides were deparaffinized in xylene, rehydrated with alcohols, and then stained for integrin  $\alpha_v\beta_3$  using the same procedure. All pictures were taken under 200 $\times$  magnification with the same exposure time. Brightness and contrast adjustments were made equally to all images. The overlay images were obtained using Olympus MetaMorph software. Quantitative analysis of integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression was performed using ImageJ Software (the National Institutes of Health, Bethesda, MD). The fluorescence intensity within images was quantified by assigning every pixel a gray scale intensity value, which ranged from 0 (black) to 255 (white). For quantification, the green (FITC-Galacto-RGD<sub>2</sub>) and red (rabbit anti-human integrin  $\beta_3$  antibody) channels in each image was exported into the software program. After calibrating optical density and adjusting threshold, the area of interest was measured as the percentage of total area. The area percentage of nucleus defined by DAPI staining was subtracted from the area percentage of cytoplasmic to compare the intensity of FITC-Galacto-RGD<sub>2</sub> and anti-human integrin  $\beta_3$  antibody. At least 15 randomly selected fields of every section were used to assess relative fluorescent intensity. The fluorescence density was expressed as a percentage (%) of total area, and presented as an average plus/minus standard deviation.

**Tumor Tissue Staining Kinetics: Concentration Dependence.** U87MG tumor slices were fixed with ice-cold acetone for 10 min, and dried in the air for 20 min. The tumor sections were incubated for 1 h with FITC-Galacto-RGD<sub>2</sub> (0.1, 1.0, 10, 20, 50, and 100  $\mu$ M). After washing with PBS buffer three times, tumor slides were mounted with Dapi Fluormount G and cover glass. The same procedure was used for human carcinoma tissues (colon cancer with high integrin  $\alpha_v\beta_3$  expression, squamous cell lung cancer with moderate integrin  $\alpha_v\beta_3$  expression, and gastric cancer with low integrin  $\alpha_v\beta_3$  expression) after being cut into slices (5  $\mu$ m), deparaffinized in xylene, and rehydrated with degraded alcohols. Pictures were taken under 200 $\times$  magnification with the same exposure time. Brightness and contrast adjustments were made equally to all images. The intensity was analyzed for the integrin  $\alpha_v\beta_3/\alpha_v\beta_5$ -positive staining areas as a percentage of the total area. The concentration-dependence histogram was generated by plotting fluorescent density against the concentration of FITC-Galacto-RGD<sub>2</sub>.

**Tumor Tissue Staining Kinetics: Time Dependence.** U87MG tumor slices (5  $\mu$ m) were fixed with ice-cold acetone for 10 min, and dried in the air at room temperature. The tumor slices were then incubated with FITC-Galacto-RGD<sub>2</sub> (100  $\mu$ M) at room temperature for different times (5, 15, 30, 60, 90, and 120 min). After washing with PBS buffer three times, the tumor slices were mounted with Dapi Fluormount G and cover glass. For tissue staining with integrin  $\beta_3$  antibody, tumor slices were first incubated with 10% goat serum for 30 min at 37 °C to block the nonspecific binding, and then incubated with rabbit anti-integrin  $\beta_3$  antibody (sc-14009, 2.5  $\mu$ g/mL) for different incubation times (5, 15, 30, 60, 90, and 120 min). The slides were then incubated with Cy3-conjugated goat anti-rabbit secondary antibodies (1:100, V/V) for another 1 h before being mounted with Dapi Fluormount G and cover glass. Human carcinoma tissues (colon cancer, lung squamous cell cancer, and gastric cancer) were cut into slices (5  $\mu$ m), deparaffinized in xylene, and rehydrated with degraded alcohols. The same procedure was performed for integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  staining. All pictures were taken under 200 $\times$  magnification with the same exposure time. Brightness and

contrast adjustments were made equally to all images. The fluorescent intensity was analyzed for the areas with integrin  $\beta_3$ -positive staining after staining of tumor tissues according to the procedure above. The time-dependence histogram was generated by plotting fluorescent density against incubation time.

**Blocking Experiment with Intravenous Injection of FITC-Galacto-RGD<sub>2</sub>.** The athymic nude mice ( $n = 3$ ) bearing U87MG glioma xenografts were intravenously injected with FITC-Galacto-RGD<sub>2</sub> (300  $\mu$ g per animal). The animals were sacrificed at 24 h postinjection. Tumors were excised, immediately snap-frozen in the OCT solution, and cut into slices (5  $\mu$ m). The tumor slices were dried at room temperature, fixed with ice-cold acetone for 15 min, and then dried in the air for 20 min at room temperature. After washing with PBS buffer three times, microscopic images were obtained using the same procedure above. For the overlay experiments, the tumor sections were incubated for 1 h at room temperature with the hamster anti-mouse integrin  $\beta_3$  antibody (2.5  $\mu$ g/mL, BD Biosciences, San Jose, CA). After washing with PBS three times, the tumor slides were incubated with the Cy3-conjugated goat anti-hamster secondary antibody (1:100, V/V, Jackson Immuno-Research Inc., West Grove, PA) for another 1 h at room temperature. Upon washing with PBS buffer, the tumor slides were then mounted with Dapi Fluormount G and cover glass.

**Overlay Experiment with FITC-Galacto-RGD<sub>2</sub> and CD31 Monoclonal Antibody.** The tumor slices (5  $\mu$ m) were fixed with ice-cold acetone for 10 min, and dried in air for 20 min at room temperature. The tumor sections were blocked with 10% goat serum for 30 min, and then were incubated with 100  $\mu$ M FITC-Galacto-RGD<sub>2</sub> and the rat anti-mouse CD31 antibody (1:100, BD Biosciences, San Jose, CA) for 1 h at room temperature. After incubating with the Cy3-conjugated goat anti-rat secondary antibody (1:100, Jackson ImmunoResearch Inc., West Grove, PA) and washing with PBS buffer three times, the tumor slides were mounted with Dapi Fluormount G and cover glass. The fluorescence was visualized with an Olympic BX51 microscope (Olympus America Inc., Center Valley, PA).

**Histopathological H&E Staining.** Histological analysis of the tumor tissues was performed according to literature methods.<sup>64,65</sup> All tumor tissues were fixed in 10% neutrally buffered formalin and embedded in paraffin. The 4  $\mu$ m sections were deparaffinized and rehydrated through degraded ethanol. Tumor sections were stained for H&E to evaluate the morphology. Aperio's ImageScope Viewer (Vista, CA) was used to visualize the whole-slide digital scans and capture images.

**Data and Statistical Analysis.** The relative integrin  $\alpha_v\beta_3/\alpha_v\beta_5$  expression level was derived from at least 15 different regions, and expressed as means plus/minus standard deviation. Statistical analysis was performed by one-way analysis of variance (ANOVA) followed by the Newman-Keuls test for multiple comparisons. The level of significance was set at  $p < 0.05$ .

## ■ ASSOCIATED CONTENT

### S Supporting Information

Detailed protocols for cellular staining of tumor cells, the concentration-dependence curve (Figure SI1), the incubation time-dependence curve (Figure SI2), microscopic images (Figure SI3) of U87MG human glioma cells stained with FITC-conjugated cyclic RGD peptides (Green) and mouse

anti-human integrin  $\alpha_v\beta_3$  antibody (sc-7312), microscopic images (Figure S14) of acetone-fixed human cancer cells stained with FITC-Galacto-RGD<sub>2</sub> (green) and mouse anti-human  $\alpha_v\beta_3$  monoclonal antibody (sc-7312) detected with Cy3-conjugated goat anti-mouse antibody (red), and microscopic images (Figure S15) of living tumor cells stained with FITC-Galacto-RGD<sub>2</sub> (green) and mouse anti-human  $\alpha_v\beta_3$  monoclonal antibody (sc-7312) detected with Cy3-conjugated goat anti-mouse antibody (red). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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### Notes

The authors declare the following competing financial interest(s): Andrzej Czerwinski, Francisco Valenzuela and Michael Pennington are from Peptides International, Inc. (Louisville, KY 40299), a commercial provider of biologically active peptides. There are no known conflicts of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome. Yumin Zheng, Shundong Ji, and Shuang Liu declare that they have no conflicts of interest.

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## ABBREVIATIONS

DAPI, 4',6-diamidino-2-phenylindole; FITC, Fluorescein isothiocyanate isomer I; Galacto-RGD<sub>2</sub>, Glu[cyclo[Arg-Gly-Asp-D-Phe-Lys(SAA-PEG<sub>2</sub>-(1,2,3-triazole)-1-yl-4-methylamide)]]<sub>2</sub> (SAA = 7-amino-L-glycero-L-galacto-2,6-anhydro-7-deoxyheptanamide, and PEG<sub>2</sub> = 3,6-dioxaoctanoic acid); IHC, immunohistochemistry; MALDI, matrix-assisted laser desorption ionization; 3P-RGD<sub>2</sub>, PEG<sub>4</sub>-E[PEG<sub>4</sub>-c(RGKfD)]<sub>2</sub> = PEG<sub>4</sub>-Glu[cyclo[Arg-Gly-Asp-D-Phe-Lys(PEG<sub>4</sub>)]]<sub>2</sub> (PEG<sub>4</sub> = 15-amino-4,7,10,13-tetraoxapentadecanoic acid); 3P-RGK<sub>2</sub>, PEG<sub>4</sub>-E[PEG<sub>4</sub>-c(RGKfD)]<sub>2</sub> = PEG<sub>4</sub>-Glu[cyclo[Arg-Gly-Lys(PEG<sub>4</sub>)-D-Phe-Asp]]<sub>2</sub>; RGD<sub>2</sub>, E[c(RGDFK)]<sub>2</sub> = Glu[cyclo(Arg-Gly-Asp-D-Phe-Lys)]<sub>2</sub>; <sup>99m</sup>Tc-3P-RGD<sub>2</sub>, [<sup>99m</sup>Tc(HYNIC-3P-RGD<sub>2</sub>)(tricine)(TPPTS)] (HYNIC = 6-hydrazinonicotinyl, and TPPTS = trisodium triphenylphosphine-3,3',3"-trisulfonate); WHO, World Health Organization

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