



## Structural Changes in the Oxygen-Evolving Complex of Photosystem II Induced by the $S_1$ to $S_2$ Transition: A Combined XRD and QM/MM Study

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### Supporting Information

**ABSTRACT:** The  $S_1 \rightarrow S_2$  transition of the oxygen-evolving complex (OEC) of photosystem II does not involve the transfer of a proton to the lumen and occurs at cryogenic temperatures. Therefore, it is commonly thought to involve only Mn oxidation without any significant change in the structure of the OEC. Here, we analyze structural changes upon the  $S_1 \rightarrow S_2$  transition, as revealed by quantum mechanics/molecular mechanics methods and the isomorphous difference Fourier method applied to serial femtosecond X-ray diffraction data. We find that the main structural change in the OEC is in the position of the dangling Mn and its coordination environment.

Atmospheric oxygen is produced during the light reactions of photosynthesis in photosystem II (PSII), a complex of proteins and cofactors found in thylakoid membranes of green plant chloroplasts and internal membranes of cyanobacteria.<sup>1,2</sup> Evolution of oxygen occurs because of the light-driven water oxidation reaction that is catalyzed by the oxygen-evolving complex (OEC), a cuboidal  $\text{CaMn}_3$  cluster with a dangling Mn held together by five putative  $\mu$ -oxo bridges along with ligands that include four terminal water molecules and amino acid side chains of the D1 and CP43 protein subunits of PSII. The OEC often represents a model for biomimetic water oxidation catalysts because of its low overpotential (20 mV),<sup>3</sup> its high turnover numbers (50 oxygen molecules per second),<sup>4</sup> and the natural abundance of its constituent metal ions (Ca and Mn). A detailed investigation of the transformations of the OEC along the catalytic cycle is, therefore, a subject of great interest.

The catalytic cycle of PSII is initiated by absorption of photons by light-harvesting pigments and transfer of energy to the reaction center chlorophylls where charge separation and formation of a chlorophyll radical cation called  $\text{P}_{680}^{+\bullet}$  occur. The OEC is oxidized by transfer of an electron to  $\text{P}_{680}^{+\bullet}$  via the tyrosine residue ( $\text{Y}_Z$ ). With each charge separation, the OEC stores an oxidation equivalent and advances through the storage states  $S_i$  ( $i = 0-4$ ) according to the so-called Kok cycle.<sup>5,6</sup> Transformation of the dark-stable  $S_1$  state into the  $S_2$  state is the first step in the cycle and the only one that does not include the transfer of a proton to the lumen. It is, thus, commonly thought that the  $S_1 \rightarrow S_2$  transition involves only oxidation of a

Mn center without any significant change in the structure of the OEC or its ligation scheme.<sup>7</sup>

X-ray diffraction (XRD) experiments have determined the structure and ligation scheme of the OEC,<sup>8-11</sup> although the large doses of X-rays necessary for data collection typically induce radiation damage and alter the oxidation states of the Mn ions in the  $\text{CaMn}_4\text{O}_5$  cluster.<sup>12,13</sup> X-ray absorption measurements require much smaller X-ray doses and, therefore, are less affected by radiation damage.<sup>14,15</sup> Recently, a new approach to protein crystallography, based on ultrashort X-ray pulses of high intensity, has allowed the collection of PSII diffraction data before the onset of radiation damage, although currently at a low resolution of 5.9 Å. While the method has been applied to microcrystals of PSII in the  $S_1$  and  $S_2$  states, no significant changes in the structure of the OEC have been detected upon the  $S_1 \rightarrow S_2$  transition.<sup>7</sup>

The S-state transitions have also been extensively studied by a variety of other experimental techniques, including time-resolved mass spectrometry,<sup>16,17</sup> electron paramagnetic resonance (EPR) spectroscopy,<sup>18</sup> and Fourier transform infrared (FTIR) spectroscopy.<sup>19-21</sup> In particular, FTIR has been instrumental in detecting changes induced by the  $S_1 \rightarrow S_2$  transition in the properties of the carboxylate ligands. For example, the downshift of the  $\nu_{\text{sym}}(\text{COO}^-)$  mode of the  $\alpha\text{-COO}^-$  group of D1-Ala344 was evidence of weakening of the C=O bond.<sup>22</sup> In addition, the perturbation of a  $\nu_{\text{sym}}(\text{COO}^-)$  mode<sup>19</sup> was attributed to either D1-Glu333 directly ligated to the OEC cluster or D1-Asp61. Both findings were consistent with an increase in the charge of the  $\text{CaMn}_4\text{O}_5$  cluster, induced by its oxidation without deprotonation during the  $S_1 \rightarrow S_2$  transition, and the resulting perturbation of the surrounding hydrogen-bonding network.

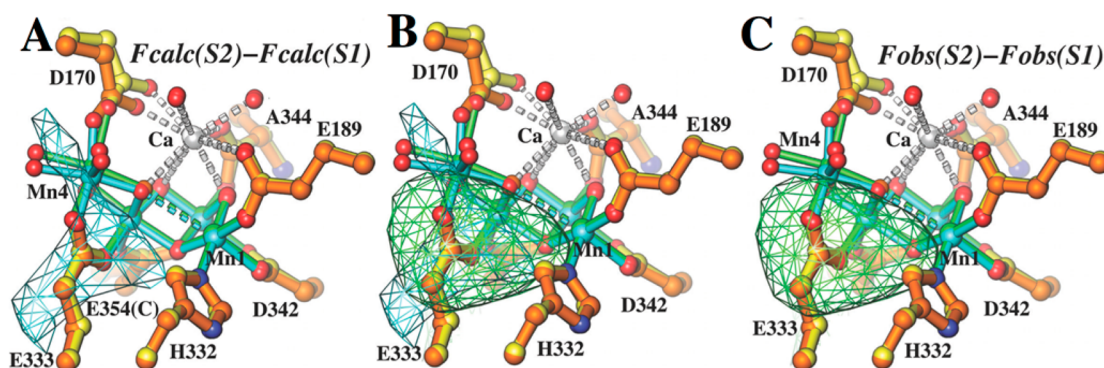
In this work, we re-examined the isomorphous difference Fourier maps using our newly improved phases and compare the experimental density difference maps to calculated density difference Fourier maps derived from quantum mechanics/molecular mechanics (QM/MM) models. The analysis shows subtle but significant structural differences, including changes in the position of the dangling Mn (denoted here Mn4) and its coordination environment, induced by the  $S_1 \rightarrow S_2$  transition (Figure 1).

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**Figure 1.** QM/MM  $S_1$  and  $S_2$  models and difference Fourier maps. (A) Simulated  $S_2$ -minus- $S_1$  difference Fourier maps calculated using the QM/MM  $S_1$  and  $S_2$  models and phases derived from multicrystal noncrystallographic symmetry averaging (see the text for computational procedures and contour levels). The highest peak near the OEC results from the displacement of Mn4. (B) Comparison of the simulated  $S_2$ -minus- $S_1$  (from panel A) and X-ray-observed  $S_2$ -minus- $S_1$  (from panel C) difference Fourier maps with color codes according to panels A and C. (C) Observed  $S_2$ -minus- $S_1$  difference Fourier maps calculated from ref 7.

The QM/MM model of the  $S_2$  state was prepared by oxidation of the previously reported  $S_1$  QM/MM model in the  $Mn_4[III,IV,IV,III]$  state.<sup>23,24</sup> We consider the spin isomer of the  $S_2$  state that is formed under native conditions,<sup>25</sup> corresponding to a doublet state ( $s = 1/2$ ) that gives rise to the  $g = 2$  multiline EPR signal. The oxidation-state pattern,  $Mn_4[III,IV,IV,IV]$ , is consistent with previous theoretical findings.<sup>26</sup>

Figure 1A shows the superposition of the QM/MM models for the  $S_1$  (yellow) and  $S_2$  (orange) states, with subtle structural rearrangements induced by the  $III \rightarrow IV$  oxidation of Mn4. These include displacement of Mn4 from the membrane and symmetrization of the Mn4 coordination environment because of the loss of Jahn–Teller distortions. In addition, Figure 1A shows the  $S_2$ -minus- $S_1$  density difference (green mesh) calculated at 5.9 Å resolution by using the QM/MM  $S_1$  and  $S_2$  models and phases derived from multicrystal noncrystallographic symmetry averaging. We also compare the  $F_{calc}(S_2)$ -minus- $F_{calc}(S_1)$  difference Fourier maps using phases initially derived from the partially refined  $S_1$  model and then from an averaging procedure (see the Supporting Information). Here,  $F_{calc}(S_2)$  and  $F_{calc}(S_1)$  denote the calculated structure factors from our hybrid  $S_2$  and  $S_1$  models, respectively.

We find that the underlying small structural displacements give rise to clear electron density differences, even at 5.9 Å resolution, because of the relatively high electronic density of Mn. In front of the displaced fragment that includes Mn4 and its ligands, we observe a positive density difference (green mesh) while the negative feature behind it is partially canceled out by other displacements in the same direction. Because the sizes of the density difference peaks are proportional to the magnitudes of the corresponding displacements, weighted by the absolute electronic density of the moving atoms, displacements of Mn centers are easier to detect than displacements of protein ligands. In fact, the peak height was  $\sim 2.9\sigma$  when difference densities of the entire unit cell were used to calculate the standard deviation (see the Supporting Information). This implies that such small differences should be detectable given that the amplitude differences of 22.9% upon comparison of the observed X-ray data for the  $S_1$  and  $S_2$  states.<sup>7</sup>

Guided by an expectation of well-defined features in the  $S_2$ -minus- $S_1$  difference Fourier map generated from our QM/MM  $S_1$  and  $S_2$  models, we asked if similar features would be present in the observed  $S_2$ -minus- $S_1$  difference Fourier maps calculated from experimental X-ray data.<sup>7</sup> Indeed, we find that the second

highest peak in the entire unit cell was next the OEC of monomer A and overlapped with similar features in the simulated  $S_2$ -minus- $S_1$  difference Fourier maps (Figure 1). The peak was approximately  $4.4$ – $5.9\sigma$ ; the height varied slightly, depending on the specific sets of phases used for the calculation (see the Supporting Information). In addition, there was a large negative peak near the putative electron extraction path (Figure S3C of the Supporting Information). There are two possible interpretations for the pair of difference density features. (i) The structure moiety between the positive and negative features is displaced as one rigid body, and (ii) the negative feature corresponds to the increased mobility of the aromatic residues nearby. It is noted that the two copies of PSII in the dimeric unit do not appear to behave the same during the  $S_1$  to  $S_2$  transition. Unfortunately, our QM/MM models cannot reliably address any structural changes away from the OEC.

In summary, we conclude that the structural changes of the OEC upon the  $S_1 \rightarrow S_2$  transition are subtle but can be addressed by an isomorphous difference Fourier method combined with QM/MM modeling. The most significant of those changes is a displacement of Mn4 from the protein membrane and a change in the coordination environment of Mn4 toward an ideal octahedron.

## ■ ASSOCIATED CONTENT

### ● Supporting Information

Description of computational methods, QM/MM coordinates of the newly reported  $S_2$  state, and discussion and analysis of electron density maps. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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## Notes

The authors declare no competing financial interest.

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## REFERENCES

- (1) McEvoy, J. P., and Brudvig, G. W. (2006) *Chem. Rev.* 106, 4455–4483.
- (2) Nelson, N., and Yocum, C. F. (2006) *Annu. Rev. Plant Biol.* 57, 521–565.
- (3) Grabolle, M., and Dau, H. (2005) *Biochim. Biophys. Acta* 1708, 209–218.
- (4) Blankenship, R. E. (2008) Frontmatter. In *Molecular Mechanisms of Photosynthesis*, pp i–vii, Blackwell Science Ltd., Oxford, U.K.
- (5) Kok, B., Forbush, B., and McGloin, M. (1970) *Photochem. Photobiol.* 11, 457–475.
- (6) Joliot, P., Barbieri, G., and Chabaud, R. (1969) *Photochem. Photobiol.* 10, 309–329.
- (7) Kern, J., Alonso-Mori, R., Tran, R., Hattne, J., Gildea, R. J., Echols, N., Glockner, C., Hellmich, J., Laksmono, H., Sierra, R. G., Lassalle-Kaiser, B., Koroidov, S., Lampe, A., Han, G., Gul, S., Difiore, D., Milathianaki, D., Fry, A. R., Miahnahri, A., Schafer, D. W., Messerschmidt, M., Seibert, M. M., Koglin, J. E., Sokaras, D., Weng, T. C., Sellberg, J., Latimer, M. J., Grosse-Kunstleve, R. W., Zwart, P. H., White, W. E., Glatzel, P., Adams, P. D., Bogan, M. J., Williams, G. J., Boutet, S., Messinger, J., Zouni, A., Sauter, N. K., Yachandra, V. K., Bergmann, U., and Yano, J. (2013) *Science* 340, 491–495.
- (8) Zouni, A., Witt, H.-T., Kern, J., Fromme, P., Krauss, N., Saenger, W., and Orth, P. (2001) *Nature* 409, 739–743.
- (9) Ferreira, K. N., Iverson, T. M., Maghlaoui, K., Barber, J., and Iwata, S. (2004) *Science* 303, 1831–1838.
- (10) Guskov, A., Kern, J., Gabdulkhakov, A., Broser, M., Zouni, A., and Saenger, W. (2009) *Nat. Struct. Mol. Biol.* 16, 334–342.
- (11) Umena, Y., Kawakami, K., Shen, J. R., and Kamiya, N. (2011) *Nature* 473, 55–60.
- (12) Yano, J., Kern, J., Irrgang, K. D., Latimer, M. J., Bergmann, U., Glatzel, P., Pushkar, Y., Biesiadka, J., Loll, B., Sauer, K., Messinger, J., Zouni, A., and Yachandra, V. K. (2005) *Proc. Natl. Acad. Sci. U.S.A.* 102, 12047–12052.
- (13) Grabolle, M., Haumann, M., Muller, C., Liebisch, P., and Dau, H. (2006) *J. Biol. Chem.* 281, 4580–4588.
- (14) Yachandra, V. K., and Yano, J. (2011) *J. Photochem. Photobiol. B* 104, 51–59.
- (15) Dau, H., and Haumann, M. (2008) *Coord. Chem. Rev.* 252, 273–295.
- (16) Messinger, J., Badger, M., and Wydrzynski, T. (1995) *Proc. Natl. Acad. Sci. U.S.A.* 92, 3209–3213.
- (17) Hillier, W., and Wydrzynski, T. (2004) *Phys. Chem. Chem. Phys.* 6, 4882–4889.
- (18) Miller, A. F., and Brudvig, G. W. (1991) *Biochim. Biophys. Acta* 1056, 1–18.
- (19) Lizasa, M., Suzuki, H., and Noguchi, T. (2010) *Biochemistry* 49, 3074–3082.
- (20) Debus, R. J. (1992) *Biochim. Biophys. Acta* 1102, 269–352.
- (21) Debus, R. J. (2008) *Coord. Chem. Rev.* 252, 244–258.
- (22) Chu, H. A., Hillier, W., and Debus, R. J. (2004) *Biochemistry* 43, 3152–3166.
- (23) Luber, S., Rivalta, I., Umena, Y., Kawakami, K., Shen, J.-R., Kamiya, N., Brudvig, G. W., and Batista, V. S. (2011) *Biochemistry* 50, 6308–6311.

- (24) Pal, R., Negre, C. F. A., Vogt, L., Pokhrel, R., Ertem, M. Z., Brudvig, G. W., and Batista, V. S. (2013) *Biochemistry* 52, 7703–7706.
- (25) Britt, R. D., Lorigan, G. A., Sauer, K., Klein, M. P., and Zimmermann, J. L. (1992) *Biochim. Biophys. Acta* 1140, 95–101.
- (26) Pantazis, D. A., Ames, W., Cox, N., Lubitz, W., and Neese, F. (2012) *Angew. Chem., Int. Ed.* 51, 9935–9940.