

# Changes in Serum Concentrations of Maternal Poly- and Perfluoroalkyl Substances over the Course of Pregnancy and Predictors of Exposure in a Multiethnic Cohort of Cincinnati, Ohio Pregnant Women during 2003–2006

Kayoko Kato,<sup>†</sup> Lee-Yang Wong,<sup>†</sup> Aimin Chen,<sup>‡</sup> Carmen Dunbar,<sup>†</sup> Glenys M. Webster,<sup>§</sup> Bruce P. Lanphear,<sup>§</sup> and Antonia M. Calafat<sup>\*,†</sup>

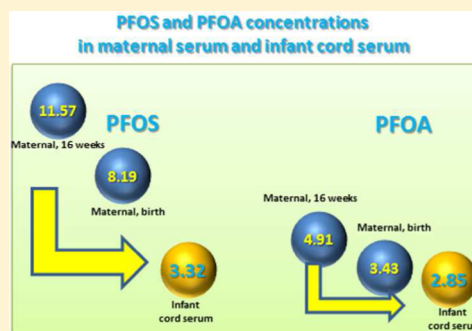
<sup>†</sup>Division of Laboratory Sciences, National Center for Environmental Health, Centers for Disease Control and Prevention, Atlanta, Georgia 30333, United States

<sup>‡</sup>Cincinnati Children's Hospital Medical Center, Cincinnati, Ohio 45229, United States

<sup>§</sup>Child & Family Research Institute, BC Children's Hospital and Simon Fraser University, Vancouver, British Columbia V6H 3V4, Canada

## S Supporting Information

**ABSTRACT:** Data on predictors of gestational exposure to poly- and perfluoroalkyl substances (PFASs) in the United States are limited. To fill in this gap, in a multiethnic cohort of Ohio pregnant women recruited in 2003–2006, we measured perfluorooctanesulfonate (PFOS), perfluorooctanoate (PFOA), and six additional PFASs in maternal serum at ~16 weeks gestation ( $N = 182$ ) and delivery ( $N = 78$ ), and in umbilical cord serum ( $N = 202$ ). We used linear regression to examine associations between maternal serum PFASs concentrations and demographic, perinatal, and lifestyle factors. PFASs concentrations in maternal sera and in their infants' cord sera were highly correlated (Spearman rank correlation coefficients = 0.73–0.95). In 71 maternal-infant dyads, unadjusted geometric mean (GM) concentrations (95% confidence interval) (in  $\mu\text{g/L}$ ) in maternal serum at delivery of PFOS [8.50 (7.01–9.58)] and PFOA [3.43 (3.01–3.90)] were significantly lower than at 16 weeks gestation [11.57 (9.90–13.53), 4.91 (4.32–5.59), respectively], but higher than in infants' cord serum [3.32 (2.84–3.89), 2.85 (2.51–3.24), respectively] ( $P < 0.001$ ). Women who were parous, with a history of previous breastfeeding, black, or in the lowest income category had significantly lower PFOS and PFOA GM concentrations than other women. These data suggest transplacental transfer of PFASs during pregnancy and nursing for the first time in a U.S. birth cohort.



## INTRODUCTION

Poly- and perfluoroalkyl substances (PFASs) are synthetic fluorinated chemicals that have been used in a wide range of consumer products and industrial applications over six decades.<sup>1</sup> Because the remarkable strength of the fluorine–carbon covalent bond confers thermal and chemical stability, some PFASs are highly resistant to both chemical and biological degradation under normal environmental conditions.<sup>1</sup>

The global occurrence of certain PFASs, persistence in the environment and bioaccumulation in biota have raised concerns about human exposures to PFASs,<sup>2</sup> and human biomonitoring has gained importance for exposure assessment of PFASs in the general population of several countries.<sup>3–9</sup> While human exposure to PFASs is well recognized, pregnant women, infants, and young children are often poorly represented in general population biomonitoring surveys. Biomonitoring studies among subpopulations susceptible to the effects of potentially harmful environmental chemicals, such as PFASs, are of interest because exposures during these critical windows of susceptibility may impact health later in life.

Pregnant women worldwide are exposed to PFASs<sup>10–24</sup>—even though data on pregnant women in the United States are limited<sup>14,15,25</sup>—and several studies have identified previous pregnancies, breastfeeding duration, and diet as important determinants of exposure to PFASs among pregnant women.<sup>12,14,15,23,26</sup> Of interest, PFASs can be transported across the placenta; several PFASs have been detected in cord serum.<sup>13,17,18,20,21,24,27–43</sup> Furthermore, the transfer of PFASs across the placenta may differ depending on the compound. For example, available scientific evidence suggest that PFOA may pass through the placenta more easily than PFOS.<sup>17,20,21,24,28,30,32,34,35,39,41</sup> Data on paired maternal and cord blood PFASs concentrations also exist for

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populations around the world, but not in the United States.<sup>13,17,18,20,21,24,28,30,32,34,35,37,39,41,43</sup>

To improve our understanding of in utero exposure to PFASs among U.S. women, we quantified the concentrations of PFASs in maternal serum during pregnancy and at delivery, and in cord sera, and evaluated the trends of PFASs concentrations and their correlations during gestation in a cohort of Ohio women. We also determined the correlations of select PFASs for paired maternal–cord sera, and examined associations between maternal serum PFASs concentrations and demographic and lifestyle factors.

## ■ EXPERIMENTAL SECTION

**Study Population.** Data were collected from mothers and their children participating in the Health Outcomes and Measures of the Environment (HOME) Study, an ongoing prospective birth cohort in the Cincinnati, Ohio (U.S.A.) metropolitan area, designed to examine low-level exposure to environmental toxicants and the efficacy of injury and lead hazard controls in the home. Between March 2003 and January 2006, pregnant women were recruited from seven prenatal clinics associated with three hospitals. Eligibility criteria for the study included  $\leq 19$  weeks of gestation, age  $\geq 18$  years, living in a house built before 1978, negative HIV status, and not taking medications for seizure or thyroid disorders. HOME Study staff mailed letters to 5184 women who were  $\geq 18$  years of age and living in a house built before 1978. Of the 1263 eligible women, 468 provided informed consent and enrolled in the study. The Institutional review boards of all involved research institutions, hospitals, and laboratories approved the study protocol.

Women provided three serum samples collected at approximately 16 and 26 weeks of gestation and within 24 h of parturition. For this project, we used the concentrations of PFASs in the 16 week samples, collected at the women's prenatal care appointment visit, to estimate the woman's gestational exposure to PFASs. Umbilical cord blood was collected at the time of infant delivery. We analyzed a total of 182 (16-week), 78 (delivery), and 202 (cord) serum samples, including 71 maternal–infant dyads with mothers' PFASs concentrations available at both 16 week of gestation and delivery.

**Quantification of PFASs Concentrations in Serum.** By using a modification of our analytical method,<sup>44</sup> we measured eight PFASs in maternal and umbilical cord sera: 2-(*N*-methyl-perfluorooctane sulfonamido) acetate (Me-PFOSA-AcOH), 2-(*N*-ethyl-perfluorooctane sulfonamido) acetate (Et-PFOSA-AcOH), perfluorohexanesulfonate (PFHxS), perfluorooctanesulfonate (PFOS), perfluorooctanoate (PFOA), perfluorononanoate (PFNA), perfluorodecanoate (PFDeA), and perfluorooctane sulfonamide (PFOSA). We used the following isotope-labeled internal standards for quantification:  $^{13}\text{C}_4$ -PFOS,  $^{18}\text{O}_2$ -PFHxS,  $^{13}\text{C}_2$ -PFOA,  $^{13}\text{C}_5$ -PFNA,  $^{13}\text{C}_2$ -PFDeA,  $\text{D}_3$ -Me-PFOSA-AcOH, and  $\text{D}_5$ -Et-PFOSA-AcOH,  $^{18}\text{O}_2$ -PFOSA. To account for potential matrix effects, we spiked the calibration standards into calf serum (Gibco, Grand Island, NY). Briefly, we added 325  $\mu\text{L}$  of 0.1 M formic acid and 25  $\mu\text{L}$  of internal standard solution to 50  $\mu\text{L}$  of serum in a 1.5 mL polypropylene autosampler vial, and the spiked serum was vortex-mixed. The sera vials were placed on a Symbiosis online SPE system (Spark Holland, Plainsboro, NJ) for the preconcentration of the analytes on a Polaris C18 cartridge (7  $\mu\text{m}$ ,  $10 \times 1$  mm; Spark Holland). The analytes were transferred onto a Betasil C8 HPLC column (3  $\times$  50 mm, 5  $\mu\text{m}$ ; ThermoHypersil Keystone, Bellefonte, PA), separated by HPLC (mobile phase A: 20 mM ammonium acetate in water, pH = 4;

mobile phase B: acetonitrile), and detected by negative-ion TurboIonSpray-tandem mass spectrometry on an API 4000 mass spectrometer (Applied Biosystems, Foster City, CA). The limits of detection (LODs) were 0.087  $\mu\text{g/L}$  for Me-PFOSA-AcOH, 0.082  $\mu\text{g/L}$  for PFNA, 0.1  $\mu\text{g/L}$  for PFHxS, PFOA, PFDeA, PFOSA, and Et-PFOSA-AcOH, and 0.2  $\mu\text{g/L}$  for PFOS. The reported concentrations are for the sum of linear and branched isomers of PFOS and PFOA. Low-concentration quality control materials (QCs) and high-concentration QCs, prepared from a calf serum pool, were analyzed with the study samples and with reagent and serum blanks to ensure the accuracy and reliability of the data.<sup>44</sup> The coefficients of variation of repeated measurements of the QCs within a period of almost one year are around 6% ([http://www.cdc.gov/nchs/data/nhanes/nhanes\\_09\\_10/PFAS\\_F\\_Polyfluorinated\\_Compounds\\_met.pdf](http://www.cdc.gov/nchs/data/nhanes/nhanes_09_10/PFAS_F_Polyfluorinated_Compounds_met.pdf)).

**Predictors of PFASs Concentrations.** We examined the association between maternal serum PFASs concentrations and demographic, perinatal, and environmental variables collected from questionnaires and biological samples. These variables have been used as covariates in PFASs-related epidemiological studies. Demographic factors included maternal age, education, race, and household income. Perinatal and maternal factors included gestational age, parity, history of breastfeeding, prepregnancy body mass index (BMI), and serum cotinine. Serum cotinine concentrations were measured in maternal samples collected at 16 weeks of gestation and within 24 h of birth, as well as in cord serum at delivery.

**Statistical Analysis.** All analyses were conducted using SAS version 9.1 (SAS Institute, Cary, NC). For statistical analysis, PFAS concentrations were  $\log_{10}$  transformed. For concentrations below the LOD, we used LOD divided by the square root of 2.<sup>45</sup> We calculated the median concentration for maternal and cord sera, and for the 71 maternal–infant dyads, we used the non-parametric Wilcoxon signed rank test to compare each woman's medians at different time points over the course of pregnancy and in their infant's cord sera. Moreover, for the 71 pairs with available maternal serum concentrations at 16 weeks and birth, and cord serum concentrations at delivery, we calculated the unadjusted geometric mean (GM) PFASs concentrations, and used a paired *t* test to compare the unadjusted GM for each pair of time points. We also determined the Spearman rank correlations among the  $\log_{10}$  transformed concentrations of PFASs in maternal and cord sera at birth, and in maternal sera at birth and at 16 weeks gestation. Statistical significance was set at  $P < 0.05$  for all analyses.

We used linear regression to model separately the  $\log_{10}$  transformed PFAS concentrations of cord serum or mothers' serum at 16 weeks gestation for each predictor variable (covariate). We included the following covariates one at a time: gestational age (term [ $\geq 37$  weeks], preterm [ $< 37$  weeks]), maternal age, parity, prepregnancy BMI (obese [ $\geq 30$  kg/m<sup>2</sup>], overweight [25–29.9 kg/m<sup>2</sup>], normal weight [18.5–24.9 kg/m<sup>2</sup>], underweight [ $< 18.5$  kg/m<sup>2</sup>]), history of previous breast feeding (yes/no), household income ( $< \$20\text{K}$ ,  $\$20\text{K}$ – $\$40\text{K}$ ,  $\$40\text{K}$ – $< \$80\text{K}$ ,  $> \$80\text{K}$ ), maternal education, race (non-Hispanic white, non-Hispanic black, other), and serum cotinine [active ( $> 3$   $\mu\text{g/L}$ ), secondhand (0.015–3  $\mu\text{g/L}$ ), or no ( $< 0.015$   $\mu\text{g/L}$ ) environmental tobacco smoke exposure]. Cotinine in serum was measured using HPLC-tandem mass spectrometry (LOD = 0.015  $\mu\text{g/L}$ ) as described before.<sup>46</sup> We calculated the GM concentrations and their 95% confidence intervals (CI) by categories of each predictor. Beta coefficients were exponentiated from the regression models to produce the ratio of PFAS GM

concentrations between categories of predictor variables. Thus, numeric estimates indicate that the GM concentrations were higher ( $>1$ ) or lower ( $<1$ ) for the predictor in that category compared with the reference category.

## RESULTS

The frequency of detection and concentrations of 8 PFASs in maternal sera during pregnancy and delivery and in cord sera are shown in Table 1 for the 71 paired maternal/infant samples

**Table 1. Median Concentrations (in  $\mu\text{g/L}$ ) of PFASs and Frequency of Detection for 71 Paired Maternal–Infant Serum Samples Collected during Gestation and at Delivery<sup>a</sup>**

	maternal, 16 weeks [frequency of detection, %]	maternal, delivery [frequency of detection, %]	infant's cord serum [frequency of detection, %]
Et-PFOA-AcOH	<LOD [23.9]	<LOD [9.9]	<LOD [11.3]
Me-PFOA-AcOH	0.50 [100]	0.20 [88.7]	0.30 [93.0]
PFOSA	<LOD [1.4]	<LOD [0]	<LOD [0]
PFHxS	1.20 [98.6]	1.20 [93.0]	0.60 [97.2]
PFOS	12.70 [100]	8.50 [100]	3.50 [98.6]
PFOA	4.80 [100]	3.30 [100]	3.10 [100]
PFNA	0.82 [100]	0.66 [100]	0.41 [98.6]
PFDeA	0.20 [97.2]	0.20 [90.1]	<LOD [16.9]

<sup>a</sup>The limits of detection (LODs) were 0.087  $\mu\text{g/L}$  (Me-PFOA-AcOH), 0.082  $\mu\text{g/L}$  (PFNA), 0.1  $\mu\text{g/L}$  (PFHxS, PFOA, PFDeA, PFOSA, Et-PFOA-AcOH), and 0.2  $\mu\text{g/L}$  (PFOS).

and in the Supporting Information, SI (Tables S1 and S2). The geometric means of concentration ratios for the 71 dyads are also shown in the SI (Table S3). We detected PFOS, PFOA, PFNA, and PFHxS in more than 90% of the paired samples, and Me-PFOA-AcOH in about 90%. For the other analytes, we did not perform additional analyses because the frequency of detection was  $<30\%$  at all of the time points examined (i.e., Et-PFOA-AcOH, PFOSA) or the median was close to the LOD (i.e., PFDeA). The median concentrations in cord serum were lower than the corresponding paired maternal concentrations (Table 1).

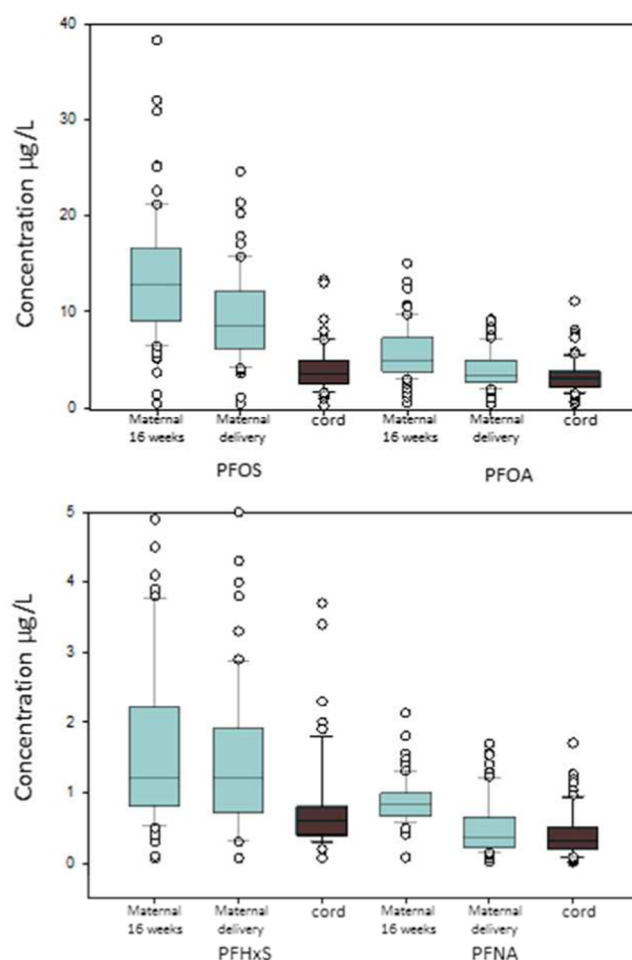
We determined the Spearman rank correlations between the  $\log_{10}$  transformed maternal concentrations of Me-PFOA-AcOH, PFHxS, PFOS, PFOA, and PFNA at 16 weeks and at delivery and their infants' cord serum concentrations (Table 2). Correlations between the concentrations in maternal and infants' samples collected at delivery (Spearman rank correlation coefficient ( $r$ ) = 0.79–0.92) and between the maternal samples at 16 weeks gestation and the infants' cord serum ( $r$  = 0.73–0.95) were high. The PFAS concentrations in maternal sera at 16 weeks gestation and at delivery were also highly correlated ( $r$  = 0.84–0.94).

We observed a statistically significant decrease in the unadjusted GM maternal serum concentrations of most PFASs from 16 weeks gestation to delivery (all paired  $t$  test  $P$  values  $<0.001$ ) (Figure 1). Still, maternal concentrations of PFOS, PFOA, PFNA, and PFHxS at birth were significantly higher than cord concentrations (all paired  $t$  test  $P$  values  $<0.001$ ), but were lower for Me-PFOA-AcOH ( $P$  = 0.01). Specifically, the maternal geometric mean concentrations of PFOS, PFOA, PFNA, and PFHxS decreased 25–43%, depending on the compound, between 16 weeks gestation and delivery (SI Table S4). The percent change of geometric mean concentrations between

**Table 2. Spearman Rank Correlation Coefficients between  $\log_{10}$  Concentrations of Select PFASs in Maternal Serum at 16 Weeks Gestation, At Birth, And in Infant Cord Serum ( $n$  = 71 Mother–Infant Pairs)<sup>a,b</sup>**

analyte	maternal serum at 16 weeks and at birth ( $N$ = 71)	maternal serum at 16 weeks and infant cord serum ( $N$ = 182)	maternal serum at birth and infant cord serum ( $N$ = 78)
Me-PFOA-AcOH	0.84	0.73	0.92
PFHxS	0.92	0.95	0.89
PFOS	0.87	0.83	0.82
PFOA	0.94	0.92	0.88
PFNA	0.88	0.77	0.79

<sup>a</sup>Selected PFASs were those detected in  $>60\%$  of samples at all time points. <sup>b</sup> $P$  values for all the comparisons were  $<0.05$ .



**Figure 1.** Concentrations (in  $\mu\text{g/L}$ ) for select PFASs in maternal serum at 16 weeks gestation and delivery, and in cord serum ( $n$  = 71 paired samples). Whiskers represent the 5<sup>th</sup> and 95<sup>th</sup> percentiles. The top and bottom edges of each box represent the 75<sup>th</sup> and 25<sup>th</sup> percentiles, respectively. The horizontal line in each box represents the median. Circles represent observations outside the 10<sup>th</sup> and 90<sup>th</sup> percentiles.

maternal and cord sera during gestation and delivery for these five PFASs are shown in the SI (Table S4).

The Wilcoxon signed rank test showed the same pattern for the medians: maternal serum median concentrations of PFOS, PFOA, PFNA, and Me-PFOA-AcOH decreased significantly from 16 weeks gestation to birth (all  $P$  values  $<0.001$ ).



Also, maternal median concentrations of PFOS, PFOA, PFNA, and PFHxS at delivery were significantly higher than the infants' cord serum concentrations ( $P$  values  $<0.001$ ). The maternal median concentrations of Me-PFOSA-AcOH were lower than in cord serum, but they did not reach statistical significance ( $P = 0.46$ ). Because concentrations of Me-PFOSA-AcOH in the mothers' serum at birth and their infants' cord serum were close to the LOD, these data should be interpreted cautiously.

Maternal serum concentrations of PFOS, PFOA, PFNA, and PFHxS at 16 weeks gestation were lower in women younger than 25 years of age compared with older women, but only reached statistical significance for PFOS and PFHxS (Table 3). Non-Hispanic black women and women with the lowest household income had significantly lower concentrations of PFOS, PFOA, PFNA, and PFHxS than non-Hispanic white women and women in the most affluent households, respectively. Obese women had significantly lower PFOS and PFNA concentrations than other women; active smokers had significantly lower PFOS concentrations than nonsmokers or second-hand smokers; women with a high school education or below had significantly lower PFHxS concentrations than women having more than a high school education. Also, a history of previous breastfeeding was inversely and significantly associated with PFOS and PFOA serum concentrations, and nulliparous women had significantly higher concentrations of PFOS and PFOA than parous women (Table 3). Breastfeeding and parity were significantly associated with each other ( $\chi^2$  test  $P$ -value =  $<0.001$ ), but parous women's GM concentrations were essentially the same when we included parity and breastfeeding history together (data not shown) or separately (Table 3) in the model. For example, GM concentrations of PFOS were 13.19  $\mu\text{g/L}$  (parity = 1) and 11.3  $\mu\text{g/L}$  (parity  $>1$ ) compared to 13.06  $\mu\text{g/L}$  and 11.37  $\mu\text{g/L}$ , respectively. Gestational age (preterm vs term) was not significantly associated with the concentrations of any of the PFASs examined. Cord serum concentrations varied similarly by demographic, perinatal, and lifestyle factors (SI Table S5) and will not be discussed further.

## DISCUSSION

We found that serum concentrations of PFOS, PFOA, PFNA, and PFHxS among a group of U.S. pregnant women in 2003–2006 were within the concentration ranges of these compounds in the U.S. general population.<sup>5</sup> These results suggest that PFAS exposure is ubiquitous in U.S. pregnant women. This information is important because, to date, published data on background exposure to PFASs among pregnant women in the United States are limited to 180 pregnant participants of the 2003–2008 National Health and Nutrition Examination Survey (NHANES).<sup>14,25</sup>

Of interest, the PFOS median concentrations in these Cincinnati women recruited during early gestation in 2003–2006 (12.7  $\mu\text{g/L}$ ) are quite close to those reported for NHANES 2003–2004 pregnant participants (12.0  $\mu\text{g/L}$ ), while the PFOA median concentrations (4.8  $\mu\text{g/L}$ ) are about two times higher (2.6  $\mu\text{g/L}$ ).<sup>14,25</sup> These findings suggest similar exposure to PFOS but higher exposure to PFOA for this cohort of Cincinnati pregnant women compared to the U.S. general population. Consistent with our findings, previous investigations reported higher serum concentrations of PFOA, but not of other PFASs, in a group of 6–8 year-old Cincinnati girls recruited starting in the spring of 2004 from a school district in northern Kentucky<sup>47,48</sup> and in more than 65 000 residents of six water

districts in two states near Parkersburg, West Virginia enrolled in 2005–2006.<sup>49</sup> The source of the exposure to PFOA in these populations was identified to be drinking water from the Ohio River that had been contaminated with PFOA.<sup>50</sup> Therefore, we speculate that the higher than background concentrations of PFOA in our study population may also be related to the consumption of PFOA contaminated drinking water as reported recently for Cincinnati girls recruited around the same time<sup>48</sup> as the HOME Study participants.

The observed PFAS concentration sequence PFOS  $\gg$  PFOA  $>$  PFNA  $\sim$  PFHxS in maternal and cord sera and the strong correlations between the PFAS serum concentrations between all of the maternal and infant samples are consistent with previous reports.<sup>13,17,18,20,21,24,28,30,32,34,35,37,39,41,43</sup> The significantly higher maternal serum concentrations of these PFASs at 16 weeks than both the maternal and infant's concentrations at birth suggest transplacental transfer of PFASs. Of interest, the ratio of concentrations between maternal and infant's samples varied depending on the compound. For example, at delivery, the ratio between the median maternal and cord serum concentration was about 1 for PFOA but  $\sim 2$  for PFOS. These ratios, which are similar to those previously reported, indicate that there are differences in the transplacental transfer of these compounds.<sup>17,20,21,24,28,30,32,34,35,39,41</sup>

Our results are consistent with other studies that examined PFAS concentrations in serial maternal blood samples collected during pregnancy, and compared maternal and infant's concentrations at birth.<sup>18,21</sup> Glynn et al. reported a significant decline in mean serum concentrations between the first and third trimester of pregnancy for PFOS, PFOA, and PFNA among 19 primiparous Swedish women.<sup>18</sup> Fromme et al. did not observe significant concentration differences of PFOS, PFOA, PFHxS, and PFNA between maternal samples collected at 34–37 weeks of pregnancy and at delivery from 27 German women,<sup>21</sup> but the time interval between the two maternal samples was much shorter than the interval Glynn et al. examined. Consistent with our findings, both research groups reported lower concentrations of PFASs in the cord serum compared with maternal samples. Thus, our results, which show strong correlations between the concentrations of all five PFASs in the maternal sera early during pregnancy and at birth, are consistent with these two previous studies,<sup>18,21</sup> while also providing information for PFHxS and Me-PFOSA-AcOH, and with a larger sample size.

We found that maternal age was positively associated with PFAS concentrations during pregnancy. PFOS, PFOA, PFNA, and PFHxS concentrations were lower in women younger than 25 years of age compared with older women. In the early 2000s, 3M, the main manufacturer of PFOS worldwide, discontinued the production of PFOS precursors and related compounds (including PFHxS and PFOA) in the United States. This fact may explain why younger women had lower concentrations of these compounds than older women, who may have had experienced increased exposures when production of these PFASs peaked during the 1980s–1990s time period.<sup>51</sup> NHANES data during 1999–2008 also suggest that PFOS, PFOA, PFHxS, and PFNA concentrations in females increased with age.<sup>52</sup>

Non-Hispanic black women and women who reported the lowest household income had lower concentrations of PFOS, PFOA, PFNA, and PFHxS than non-Hispanic white women and women in the more affluent households, respectively. These results are in agreement with findings from the U.S. general population. Specifically, 1999–2008 NHANES data also suggest that non-Hispanic black females had lower concentrations of

Table 3. Geometric Mean Maternal Prenatal (16 weeks) Concentrations of PFOS, PFOA, PFHxS, and Me-PFOA-AcOH ( $\mu\text{g/L}$ ) According to Demographic, Perinatal and Lifestyle Factors<sup>a</sup>

variables	N (%)	PFOS			PFOA			PFHxS			Me-PFOA-AcOH		
		GM (95% CI)	GM ratio (95% CI)	GM ratio (95% CI)	GM (95% CI)	GM ratio (95% CI)	GM ratio (95% CI)	GM (95% CI)	GM ratio (95% CI)	GM ratio (95% CI)	GM (95% CI)	GM ratio (95% CI)	GM ratio (95% CI)
maternal age (years)													
<25	40 (22)	10.87 (9.16–12.9)	0.8 (0.65–0.97) <sup>b</sup>	0.94 (0.77–1.14)	4.89 (4.12–5.81)	0.73 (0.64–0.83)	0.91 (0.78–1.05)	1.02 (0.8–1.29)	0.63 (0.48–0.83) <sup>b</sup>	1.12 (0.88–1.41)	0.47 (0.38–0.57)	1.12 (0.88–1.41)	
>35	30 (16)	14.94 (12.26–18.21)	1.09 (0.87–1.37)	1.3 (1.04–1.62) <sup>b</sup>	6.77 (5.55–8.26)	0.93 (0.8–1.07)	1.15 (0.98–1.36)	1.65 (1.26–2.16)	1.02 (0.75–1.38)	1.02 (0.78–1.32)	0.43 (0.34–0.54)	1.02 (0.78–1.32)	
25–34	112 (62)	13.67 (12.34–15.14)	ref	ref	5.22 (4.71–5.79)	0.8 (0.75–0.87)	ref	1.62 (1.41–1.87)	ref	ref	0.42 (0.37–0.47)	ref	
race													
non-hispanic black	56 (31.1)	10.85 (9.4–12.53)	0.76 (0.64–0.9) <sup>b</sup>	0.75 (0.63–0.9) <sup>b</sup>	4.41 (3.82–5.1)	0.73 (0.65–0.81)	0.86 (0.76–0.98) <sup>b</sup>	0.86 (0.72–1.03)	0.47 (0.38–0.59) <sup>b</sup>	1.12 (0.91–1.38)	0.46 (0.39–0.54)	1.12 (0.91–1.38)	
other	10 (5.6)	16.07 (11.43–22.59)	1.12 (0.79–1.6)	1.03 (0.72–1.47)	6.05 (4.3–8.53)	0.98 (0.76–1.26)	1.17 (0.9–1.52)	2.43 (1.58–3.74)	1.34 (0.85–2.09)	1.25 (0.83–1.89)	0.51 (0.34–0.76)	1.25 (0.83–1.89)	
non-hispanic white	114 (63.3)	14.3 (12.93–15.82)	ref	ref	5.87 (5.3–6.5)	0.84 (0.78–0.9)	ref	1.82 (1.6–2.07)	ref	ref	0.41 (0.36–0.46)	ref	
household income													
<\$20 000	29 (16)	9.44 (7.73–11.51)	0.63 (0.49–0.81) <sup>b</sup>	0.7 (0.54–0.9) <sup>b</sup>	4.1 (3.35–5.03)	0.64 (0.55–0.74)	0.77 (0.64–0.93) <sup>b</sup>	0.84 (0.64–1.1)	0.49 (0.35–0.69) <sup>b</sup>	1.02 (0.76–1.36)	0.48 (0.38–0.6)	1.02 (0.76–1.36)	
\$20 000–\$40 000	37 (20.6)	13.29 (11.15–15.85)	0.89 (0.71–1.13)	0.91 (0.72–1.15)	5.35 (4.47–6.4)	0.84 (0.74–0.96)	1.02 (0.86–1.21)	1.4 (1.1–1.77)	0.82 (0.59–1.12)	0.92 (0.7–1.21)	0.43 (0.35–0.53)	0.92 (0.7–1.21)	
\$40 000–\$80 000	64 (35.6)	13.98 (12.23–15.98)	0.94 (0.77–1.15)	0.97 (0.79–1.19)	5.69 (4.96–6.52)	0.86 (0.78–0.95)	1.04 (0.9–1.21)	1.72 (1.43–2.06)	1 (0.76–1.32)	0.81 (0.64–1.03)	0.38 (0.33–0.45)	0.81 (0.64–1.03)	
>\$80 000	50 (27.8)	14.87 (12.78–17.3)	ref	ref	5.89 (5.05–6.87)	0.83 (0.74–0.92)	ref	1.71 (1.39–2.1)	ref	ref	0.47 (0.39–0.56)	ref	
BMI <sup>c</sup>													
obese ( $\geq 30 \text{ kg/m}^2$ )	67 (36.8)	11.45 (9.77–13.43)	0.81 (0.66–1) <sup>b</sup>	0.88 (0.71–1.08)	4.95 (4.22–5.82)	0.7 (0.63–0.79)	0.81 (0.7–0.94) <sup>b</sup>	1.22 (0.98–1.52)	0.84 (0.63–1.11)	0.93 (0.73–1.17)	0.44 (0.36–0.52)	0.93 (0.73–1.17)	
overweight (25–29.9 $\text{kg/m}^2$ )	47 (25.8)	13.59 (11.89–15.54)	0.97 (0.8–1.17)	0.96 (0.8–1.17)	5.45 (4.75–6.24)	0.84 (0.76–0.92)	0.96 (0.84–1.1)	1.71 (1.42–2.06)	1.17 (0.9–1.52)	0.83 (0.67–1.04)	0.39 (0.34–0.46)	0.83 (0.67–1.04)	
normal ( $<24.9 \text{ kg/m}^2$ )	66 (36.4)	14.08 (12.32–16.08)	ref	ref	5.65 (4.93–6.46)	0.87 (0.79–0.96)	ref	1.46 (1.21–1.76)	ref	ref	0.47 (0.4–0.55)	ref	
breast feeding history													
prev breast feed	74 (41)	11.85 (10.44–13.45)	0.83 (0.71–0.98) <sup>b</sup>	0.73 (0.62–0.86) <sup>b</sup>	4.47 (3.95–5.06)	0.77 (0.7–0.85)	0.93 (0.82–1.05)	1.34 (1.13–1.61)	0.86 (0.69–1.09)	0.99 (0.82–1.2)	0.43 (0.37–0.5)	0.99 (0.82–1.2)	
not prev breast feed	108 (59)	14.2 (12.78–15.76)	ref	ref	6.1 (5.5–6.75)	0.83 (0.77–0.9)	ref	1.56 (1.35–1.8)	ref	ref	0.43 (0.38–0.49)	ref	
education													
<12 years	12 (6.7)	11.08 (8.07–15.23)	0.81 (0.58–1.13)	0.92 (0.66–1.28)	4.99 (3.62–6.88)	0.69 (0.55–0.87)	0.84 (0.66–1.07)	0.94 (0.61–1.45)	0.59 (0.38–0.93) <sup>b</sup>	1.24 (0.85–1.8)	0.51 (0.36–0.73)	1.24 (0.85–1.8)	
12 years	25 (13.9)	11.77 (9.45–14.67)	0.86 (0.68–1.09)	0.96 (0.76–1.23)	5.24 (4.19–6.54)	0.8 (0.68–0.94)	0.97 (0.82–1.16)	1.14 (0.84–1.54)	0.72 (0.52–1.00) <sup>b</sup>	1.2 (0.91–1.58)	0.5 (0.39–0.64)	1.2 (0.91–1.58)	
>12 years	143 (79.4)	13.68 (12.48–15)	ref	ref	5.44 (4.96–5.97)	0.82 (0.77–0.88)	ref	1.59 (1.4–1.8)	ref	ref	0.41 (0.37–0.46)	ref	
gestational age													
preterm ( $<37$ weeks)	14 (7.7)	11.25 (8.39–15.09)	0.84 (0.62–1.14)	0.96 (0.71–1.31)	5.18 (3.85–6.96)	0.73 (0.59–0.91)	0.9 (0.72–1.13)	1.31 (0.87–1.97)	0.89 (0.58–1.36)	1.18 (0.83–1.68)	0.5 (0.36–0.7)	1.18 (0.83–1.68)	
term ( $\geq 37$ weeks)	168 (92.3)	13.37 (12.28–14.55)	ref	ref	5.39 (4.95–5.87)	0.81 (0.76–0.86)	ref	1.48 (1.32–1.67)	ref	ref	0.43 (0.39–0.47)	ref	
serum cotinine ( $\mu\text{g/L}$ )													
active smoker ( $\geq 3$ )	14 (7.7)	9.04 (6.76–12.07)	0.68 (0.49–0.94) <sup>b</sup>	0.95 (0.68–1.32)	4.76 (3.55–6.39)	0.66 (0.53–0.81)	0.82 (0.64–1.04)	1.06 (0.7–1.59)	0.67 (0.42–1.06)	1.12 (0.76–1.63)	0.43 (0.31–0.61)	1.12 (0.76–1.63)	
secondhand smoker (0.015–3)	117 (64.3)	13.76 (12.45–15.21)	1.04 (0.86–1.24)	1.12 (0.93–1.34)	5.61 (5.07–6.22)	0.83 (0.77–0.89)	1.03 (0.9–1.18)	1.48 (1.29–1.7)	0.94 (0.73–1.22)	1.16 (0.94–1.43)	0.45 (0.4–0.51)	1.16 (0.94–1.43)	
non smoker ( $<0.015$ )	51 (28)	13.28 (11.41–15.45)	ref	ref	5.03 (4.31–5.87)	0.8 (0.72–0.9)	ref	1.57 (1.27–1.95)	ref	ref	0.39 (0.33–0.46)	ref	
parity													
>1	47 (25.8)	11.37 (9.7–13.33)	0.78 (0.64–0.96) <sup>b</sup>	0.74 (0.61–0.91) <sup>b</sup>	4.83 (4.14–5.64)	0.75 (0.67–0.85)	0.87 (0.75–1.01)	1.31 (1.05–1.64)	0.79 (0.6–1.05)	1.08 (0.85–1.36)	0.52 (0.43–0.62)	1.08 (0.85–1.36)	
1	57 (31.3)	13.06 (11.31–15.08)	0.9 (0.74–1.09)	0.7 (0.58–0.84) <sup>b</sup>	4.53 (3.94–5.21)	0.77 (0.7–0.86)	0.89 (0.78–1.03)	1.36 (1.11–1.67)	0.82 (0.63–1.07)	1.04 (0.84–1.3)	0.5 (0.42–0.59)	1.04 (0.84–1.3)	
0	78 (42.9)	14.53 (12.85–16.43)	ref	ref	6.49 (5.76–7.32)	0.86 (0.79–0.95)	ref	1.65 (1.39–1.97)	ref	ref	0.48 (0.42–0.55)	ref	

<sup>a</sup>Ratios are the exponentiated beta coefficients from a linear regression model with the maternal concentrations as the outcome. Ratios represent the multiplicative difference in PFAS concentrations from the reference category. Thus, numeric estimates indicate that GM concentrations were higher ( $>1$ ) or lower ( $<1$ ) for the predictor (variable) in that category compared with the reference category. Each predictor was run in a separate model. <sup>b</sup> $P < 0.05$ . <sup>c</sup>We excluded two underweight participants with BMI  $< 18 \text{ kg/m}^2$ .

PFOA and PFHxS than non-Hispanic white females.<sup>52</sup> Similarly, in an analysis of 2003–2008 NHANES data, PFOA and PFOS concentrations were higher in more affluent populations.<sup>53</sup> The reason for these differences is unknown, but may reflect differences related to lifestyle (e.g., use of PFASs-containing products), diet, physiology (e.g., elimination), or a combination of the above.<sup>52</sup>

Active smokers had significantly lower PFOS concentrations than nonsmokers or second-hand smokers. Of interest, the associations between smoking and PFASs exposure are inconsistent among studies. In agreement with our findings, current Danish male smokers had lower PFOA and PFOS plasma concentrations than never smokers.<sup>54</sup> By contrast, smoking was positively associated with serum concentrations of PFNA and PFOA in 17–39 year old 2003–2008 NHANES female participants.<sup>14</sup> In another study, smoking was not associated with concentrations of PFASs in French women's breast milk.<sup>55</sup>

Obese women had significantly lower PFOS and PFNA concentrations than other women. Like for smoking, there is no consistency in the literature about the potential associations between exposure to PFASs and BMI. Similar to our findings, plasma concentrations of PFOS and PFOA were inversely associated with BMI in Danish men.<sup>54</sup> However, in Korean adolescents and adults,<sup>7,56</sup> serum concentrations of several PFASs, including PFHxS, PFOS, PFOA, and PFNA, showed a positive association with BMI. Finally, others have reported no associations between plasma concentrations of PFASs (e.g., PFOS, PFOA, PFHxS, and PFNA) in a group of Vietnamese pregnant women<sup>57</sup> or middle-aged Norwegian women.<sup>58</sup> Interpreting these results is difficult because of the differences in study populations and inconsistent findings among studies.

Nulliparous women had significantly higher concentrations of PFOS and PFOA than parous women, in agreement with previous studies.<sup>12,26,28,40,59,60</sup> We also observed a significant decrease in maternal concentrations of these compounds from 16 weeks gestation to delivery. Together, these findings suggest gestational transfer of PFASs to the fetus.<sup>17,20,28,30,32,34,40</sup> However, we cannot rule out that physiologic changes occurring during pregnancy (e.g., increased maternal blood volume, fat and total body water; decreased plasma protein concentrations, especially albumin) may also be related to the observed decrease in maternal PFAS concentrations.<sup>61,62</sup>

Finally, we observed that a history of previous breastfeeding was inversely associated with PFOS and PFOA serum concentrations, suggesting that breastfeeding may further decrease the mother's body burden of these compounds and contribute to infants' exposure to PFASs.<sup>14,21,35,37,43,55,63–72</sup> Consistent with our findings, the duration of being breast-fed was also positively associated with serum PFOA concentrations among 6–8 year-old girls in Greater Cincinnati (N = 353) and the San Francisco Bay Area (N = 351).<sup>48</sup>

Our study has several limitations. We lacked detailed information on some potentially important sources of PFAS exposure including paper/cardboard use, packaged food consumption, or use of fabric or carpet treatment products during pregnancy. Furthermore, we did not examine sources of dietary exposure as dietary variables collected in the larger HOME Study were designed to assess exposures to heavy metals, pesticides and other persistent pollutants rather than PFASs. Lastly, results from this study population may not be generalizable to the general population of U.S. pregnant women because our participants resided in an area in the United States

where the nature of the exposure to at least some of the PFASs (i.e., PFOA) may have differed from the rest of the country. Nonetheless, our data support the hypothesis that PFASs transfer from the mother to her child, and suggests that nursing may decrease a woman's body burden of PFASs. Our findings also show for the first time in a U.S. longitudinal birth cohort that exposure to PFASs among a population of pregnant women is a function of age, race, household income, parity, and previous history of breastfeeding. Future research should build on these results to examine the impact of prenatal and postnatal exposure to these persistent organic pollutants among the children in the HOME Study and other birth cohorts.

## ■ ASSOCIATED CONTENT

### § Supporting Information

Median concentrations ( $\mu\text{g/L}$ ) of PFASs and frequency of detection for maternal sera (at each collection point) and cord sera (Table S1); geometric mean serum concentrations (in  $\mu\text{g/L}$ ) for select PFASs for 71 paired samples (Table S2); geometric means of serum concentration ratios for select PFASs for 71 paired samples (Table S3); percent change of geometric mean serum concentrations for select PFASs for 71 paired samples (Table S4); geometric mean cord serum concentrations of PFOS, PFOA, PFNA, PFHxS, and Me-PFOA-AcOH ( $\mu\text{g/L}$ ) according to demographic, perinatal, and lifestyle factors (Table S5). This material is available free of charge via the Internet at <http://pubs.acs.org>.

## ■ AUTHOR INFORMATION

### Corresponding Author

\*Phone: (770) 488-7891; e-mail: [acalafat@cdc.gov](mailto:acalafat@cdc.gov).

### Notes

The use of trade names is for identification only and does not constitute endorsement by the U.S. Department of Health and Human Services or the CDC. The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the NIEHS or the CDC.

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