

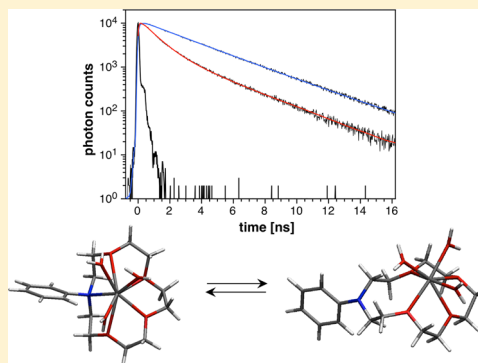
Probing Ternary Complex Equilibria of Crown Ether Ligands by Time-Resolved Fluorescence Spectroscopy

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S Supporting Information

ABSTRACT: Ternary complex formation with solvent molecules and other adventitious ligands may compromise the performance of metal-ion-selective fluorescent probes. As Ca(II) can accommodate more than 6 donors in the first coordination sphere, commonly used crown ether ligands are prone to ternary complex formation with this cation. The steric strain imposed by auxiliary ligands, however, may result in an ensemble of rapidly equilibrating coordination species with varying degrees of interaction between the cation and the specific donor atoms mediating the fluorescence response, thus diminishing the change in fluorescence properties upon Ca(II) binding. To explore the influence of ligand architecture on these equilibria, we tethered two structurally distinct aza-15-crown-5 ligands to pyrazoline fluorophores as reporters. Due to ultrafast photoinduced electron-transfer (PET) quenching of the fluorophore by the ligand moiety, the fluorescence decay profile directly reflects the species composition in the ground state. By adjusting the PET driving force through electronic tuning of the pyrazoline fluorophores, we were able to differentiate between species with only subtle variations in PET donor abilities. Concluding from a global analysis of the corresponding fluorescence decay profiles, the coordination species composition was indeed strongly dependent on the ligand architecture. Altogether, the combination of time-resolved fluorescence spectroscopy with selective tuning of the PET driving force represents an effective analytical tool to study dynamic coordination equilibria and thus to optimize ligand architectures for the design of high-contrast cation-responsive fluorescence switches.



1. INTRODUCTION

Macrocyclic ligands, particularly aza-substituted crown and thiocrown ethers, are commonly utilized in the design of cation-selective fluorescent probes responding with either a spectral shift or enhancement in fluorescence emission.^{1,2} If an *N*-aryl azacrown moiety serves as the electron-donor in fluorophore platforms with donor–acceptor (D–A) architecture, the degree of excited-state intramolecular charge transfer (ICT) decreases upon metal cation binding to produce a hypsochromic shift in the fluorescence emission. Alternatively, if the crown ether-based electron donor is electronically decoupled from the fluorophore π -system, it can quench fluorescence via photoinduced electron transfer (PET), yielding an increased emission intensity upon coordination of the cation via suppression of PET. The overall fluorescence contrast, defined as the ratio of emission intensities between the metal-bound and free forms, is dependent on the change in PET driving force, which in turn is governed by the increase in donor potential upon metal binding.³ Thus, the fluorescence contrast and final quantum yield are dependent on the ability of the metal cation to reduce the PET donor strength. In previous work with *N*-aryl thiazacrown-based Cu(I)-responsive fluorescent probes, in which the arylamine moiety serves as the PET donor, we observed that the fluorescence contrast ratios

and quantum yields are significantly compromised by formation of ternary complexes with solvent molecules, which allow for dissociation of the Cu(I)–N bond and thus significant residual PET.⁴ We suspected that such effects are not confined to Cu(I)–thiazacrown systems but might extend to other ligands and metal ions as well. In fact, Freidzon et al. had previously conducted computational studies of ternary complex formation in the Ca(II)-*N*-phenylaza-15-crown-5 system in an effort to explain a long wavelength shoulder observed in the absorption spectrum of Ca(II) responsive styryl dyes containing this ligand motif.^{5–7} Furthermore, X-ray structural analysis of the Ca(II)-*N*-phenylaza-15-crown-5 complex revealed the presence of three water molecules in the first coordination sphere.⁸ As illustrated with Scheme 1A, ternary complex formation with solvent molecules may increase the steric strain to push the equilibrium toward coordination species with weakened Ca–N interaction,⁶ thus increasing the electron donating ability of the aniline moiety. In the case of styryl dyes, this results in

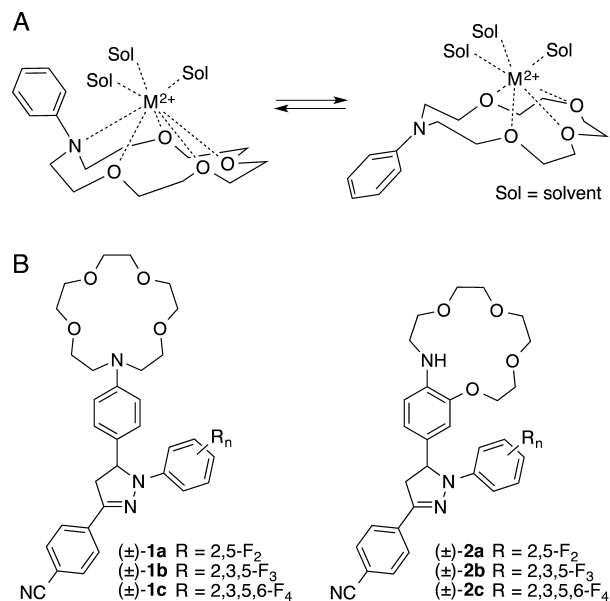
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Scheme 1



increased ICT and thus a longer absorption and emission wavelength. As the equilibration between different coordination species likely occurs with rapid kinetics, individual coordination species may not be distinguishable by solution NMR methods.^{4,9} Therefore, we decided to study this system by picosecond-resolved fluorescence spectroscopy as an extension of our previous work on the design of Cu(I)-responsive fluorescent probes.^{4,10–13}

For this purpose, we combined the *N*-phenylaza-15-crown-5 ligand, as well as a modified version with the aryl ring fused to the ligand backbone, with electronically decoupled pyrazoline fluorophores as PET-sensitive fluorescence reporters (Scheme 1B). Compared to probe **1**, the ring-fused ligand of **2** was expected to reduce steric strain between the ortho-hydrogen atoms of the aniline ring and the ligand backbone in the ternary complex,^{9,14} thus suppressing dissociation of the metal–nitrogen bond for improved contrast and quantum yield.¹⁵

Given the different donor strengths for PET quenching, each species is expected to exhibit a characteristic fluorescence lifetime, thus yielding a multiexponential decay profile if more than one coordination species is present in solution. Due to the correlative relationship between the pre-exponential terms and fluorescence lifetimes, least-squares fitting of multiexponential decay data can yield more than a single solution with satisfactory statistics.¹⁶ For this reason, we thought to measure the fluorescence decay profiles at three different PET driving forces and perform a global analysis based on a single set of pre-exponential components reflecting the fractional contribution of each species. Taking advantage of the electronic tunability of 1,3,5-triarylpyrazoline fluorophores,^{3,17} we synthesized a set of three derivatives for each ligand, **1a–c** and **2a–c**, respectively, with an increasing number of fluoro-substituents on the 1-aryl ring (Scheme 1B, Table 1). As previously demonstrated, the increasing electron withdrawing character of the 1-aryl ring enhances the excited-state energy while leaving the fluorophore acceptor potential mostly unchanged.^{3,17} As a consequence, the PET driving force steadily increases with the number of fluoro substituents. Although the radical cation formed by PET may promote a significant reorganization of the metal–ligand interaction and even lead to metal ejection,^{18–20} PET

quenching of the electronically decoupled fluorophore precedes subsequent rearrangements, and therefore the fluorescence decay profile is a direct reflection of the ensemble composition in the ground state at the time of excitation. Furthermore, unlike a directly coupled ICT-based fluorescence reporter, the attachment of the pyrazoline fluorophore via an sp³-hybridized carbon is not expected to substantially perturb the donor strength of the arylamine nitrogen compared to the unsubstituted ligand.

2. EXPERIMENTAL METHODS

Synthesis. Procedures for the synthesis of compounds **1a–c**, **2a–c**, and all intermediates are provided in the Supporting Information.

Steady-State Absorption and Fluorescence Spectroscopy. UV–vis absorption spectra were acquired at 25 °C with a Varian Cary Bio50 spectrophotometer with constant temperature accessory. Emission spectra were recorded with a PTI fluorometer and corrected for the spectral response of the detection system and the spectral irradiance of the excitation source (through a calibrated photodiode). To convert the emission spectra to wavenumber scale, the measured emission intensities were corrected by multiplication with λ^2 .²¹ All measurements were conducted in a quartz cuvette with 1 cm path length and a cell volume of 3.0 mL with HPLC grade acetonitrile as the solvent. Quantum yields were determined in air-equilibrated solutions using norharmane in 0.1 N H₂SO₄ as fluorescence standard ($\Phi_f = 0.58$).²²

Picosecond Time-Resolved Fluorescence Spectroscopy. The fluorescence decay profiles were acquired in air-equilibrated HPLC grade acetonitrile in a 1 cm path length quartz cuvette at a fluorophore concentration of 5 μ M using a single photon counting spectrometer (Edinburgh Instruments, LifeSpec Series) equipped with a pulsed laser diode as the excitation source (372 nm, FMHM = 80 ps, 20 MHz repetition rate, 1024 channel resolution). The wavelength of the emission monochromator was adjusted to the emission maximum of the respective fluorophore. The decay data were analyzed by nonlinear least-squares fitting with deconvolution of the instrument response function using the DecayFit software package.²³

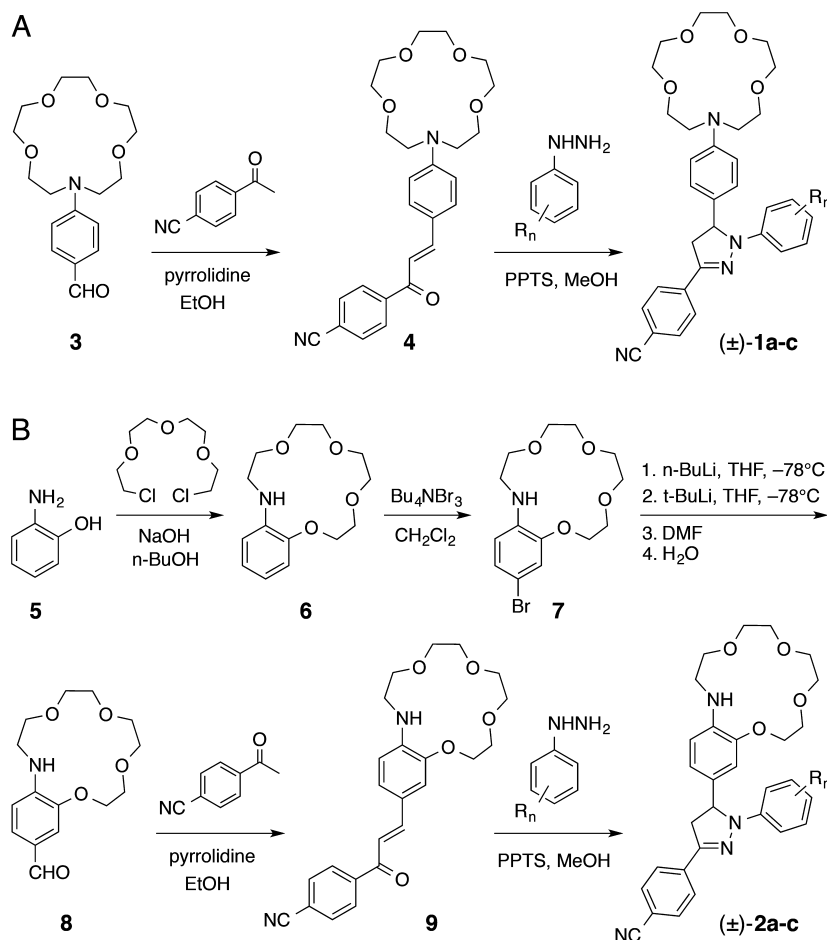
Cyclic Voltammetry. The donor and acceptor potentials of each probe were determined in deoxygenated acetonitrile containing 0.1 M Bu₄NPF₆ as the electrolyte using a CH-Instruments potentiostat equipped with a single compartment cell with a glassy carbon working electrode, a Pt counter electrode, and a nonaqueous Ag/AgNO₃ reference electrode (10 mM). The measured potentials were referenced to ferrocene as external standard. All measurements were performed at a scan rate of 100 mV s^{−1}.

Determination of Stability Constants. Titrations were carried out in acetonitrile at a probe concentration of 5 μ M by addition of Ca(ClO₄)₂·4H₂O up to a total concentration of 10 mM. After addition of each aliquot, the solution was equilibrated for 5 min and a fluorescence spectrum was acquired over the entire emission range. The data were analyzed by nonlinear least-squares fitting based on equation S5 as described in the Supporting Information.

3. RESULTS AND DISCUSSION

Synthesis. The two sets of fluorescent probes **1** and **2** were synthesized from the corresponding aldehyde precursors **3** and

Scheme 2. Synthesis of Pyrazoline Derivatives 1a–c (A) and 2a–c (B)



6, respectively, by aldol condensation with *para*-cyano acetophenone followed by cyclization of the chalcone intermediates with the respective fluoro-substituted arylhydrazines (Scheme 2). Aldehyde 3 was obtained through Vilsmeier formylation of commercially available *N*-phenylaza-15-crown-5,²⁴ whereas 8 was accessed through lithium halogen exchange of bromide 7 followed by quenching with dimethylformamide. Ligand 6 was synthesized from 2-aminophenol by cyclization with tetraethylene glycol dichloride according to a literature procedure,⁹ and converted to 7 by bromination with tetrabutylammonium tribromide. Each probe was isolated by column chromatography and further purified by recrystallization from MTBE–cyclohexane.

Steady-State Absorption and Emission Spectroscopy.

As the stability of crown ether complexes is significantly reduced in protic compared to nonprotic solvents, all complexation studies were conducted in acetonitrile. We selected Ca(II) as the metal cation to probe ternary complex formation, as it has been frequently used in complexation studies of *N*-phenylaza-15-crown-5 derivatives.^{8,24–32} Ca(II) was supplied as Ca(ClO₄)₂·4H₂O, yielding four water molecules per Ca(II) center, a number that satisfies the stoichiometry of ternary complexes according to experimental and computational studies.^{6–8} The pertinent photophysical properties of the pyrazoline probes 1 and 2 are compiled in Table 1. Due to the low quantum yield of the free probes, the emission maxima were determined in the presence of a large excess of Ca(ClO₄)₂·4H₂O. The PET driving force for each derivative

Table 1. Photophysical Data of Pyrazoline Derivatives 1a–c and 2a–c in Acetonitrile at 298 K

compd	R	abs λ_{max} / nm	em λ_{max} / nm ^a	ΔE_{00} / eV ^b	$\Delta G_{\text{et}}^{\text{c}}$ / eV ^c	$\Phi_{\text{F}}^{\text{d}}$
1a	2,5-F ₂	377	489	2.90	–0.27	0.68
1b	2,3,5-F ₃	371	475	2.97	–0.35	0.42
1c	2,3,5,6-F ₄	357	450	3.09	–0.42	0.32
2a	2,5-F ₂	377	489	2.90	–0.35	0.71
2b	2,3,5-F ₃	370	471	2.98	–0.45	0.73
2c	2,3,5,6-F ₄	354	451	3.11	–0.51	0.79

^aMeasured in the presence of 0.1 M Ca(ClO₄)₂·4H₂O. ^bZero–zero transition energy; estimated as $\bar{\nu}_{00} \approx 1/2(\bar{\nu}_{\text{abs}}^{\text{max}} + \bar{\nu}_{\text{em}}^{\text{max}})$.³⁴ ^cPhotoinduced electron-transfer free energy estimated on the basis of the Rehm–Weller formalism³³ using experimental ground-state donor and acceptor potentials (Supporting Information). ^dApparent fluorescence quantum yield; norharmane in 0.1 N H₂SO₄ as reference ($\Phi_{\text{F}} = 0.58$).²²

was estimated on the basis of the Rehm–Weller formalism³³ using the experimentally determined ground-state donor and acceptor potentials (Supporting Information).

Consistent with previous observations on 1,3,5-triarylpyrazoline fluorophores, the absorption and emission bands of all derivatives are broad and unstructured, indicating a lowest excited state with charge-transfer character (Figure 1).^{4,27,35} With increasing number of fluoro-substituents on the 1-aryl ring, the absorption and emission maxima are shifted to higher energy while maintaining similar Stokes shifts (Figure 1),^{3,17}

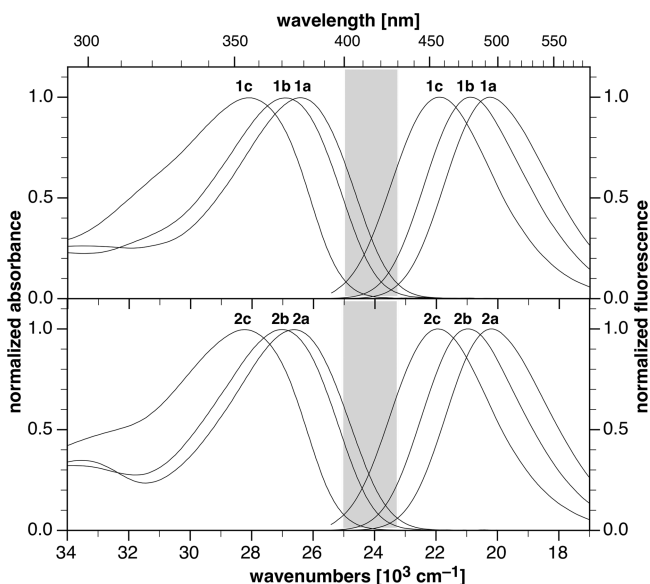


Figure 1. Normalized absorption (left traces) and emission spectra (right traces) of pyrazoline derivatives **1a–c** and **2a–c** in acetonitrile in the presence of 0.1 M $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$ at 298 K. The shaded area indicates the tunable range of the 0–0 transition energy to the lowest excited state.

and consequently the zero–zero excited-state energies ΔE_{00} increase from 2.90 to 3.11 eV with approximately 0.1 eV increments (Table 1). Because the crown ether moiety is electronically decoupled from the fluorophore π -system, both sets of probes exhibit very similar absorption and emission trends; however, the PET driving force differs by approximately 80–100 mV between derivatives with identical numbers of fluoro substituents. This difference can be attributed to the lower donor potential of the crown ether moiety in series **2**, which features an additional oxygen π -donor for improved stabilization of the radical cation. Despite the higher PET driving force, the apparent fluorescence quantum yields of the $\text{Ca}(\text{II})$ -saturated probes consistently remain above 70% in the **2** series but decrease from 0.68 to 0.32 for **1**, indicating an incomplete fluorescence recovery.

Proton NMR Analysis. In previous work with $\text{Cu}(\text{I})$ –thiazacrown systems,⁴ including the direct thio-analogue of *N*-phenylaza-15-crown-5,³⁶ we observed a single set of NMR resonances at all $\text{Cu}(\text{I})$ –ligand ratios, indicating that the binary complex, free ligand, and any ternary complexes present all equilibrate rapidly on the NMR time scale. To determine whether this is also the case for the $\text{Ca}(\text{II})$ –*N*-phenylaza-15-crown-5 system, we recorded ^1H NMR spectra of this ligand in the absence and presence of 0.1 M $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$. As shown in Figure 2, addition of excess $\text{Ca}(\text{II})$ to a solution of the ligand in CD_3CN resulted in substantial deshielding of all aryl protons; however, only a single resonance for each set of equivalent protons in the ligand was observed, indicating that all coordination species present equilibrate too rapidly to be distinguished by proton NMR.

Coordination Equilibria with $\text{Ca}(\text{II})$. To determine the apparent binding affinity and complex stoichiometry of the fluorescent probes, we performed a series of fluorescence titrations with $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$ in acetonitrile. Because the metal ion binding sites are electronically decoupled from the fluorophore moieties, we anticipated only minor affinity variations across derivatives of the same series. For this reason

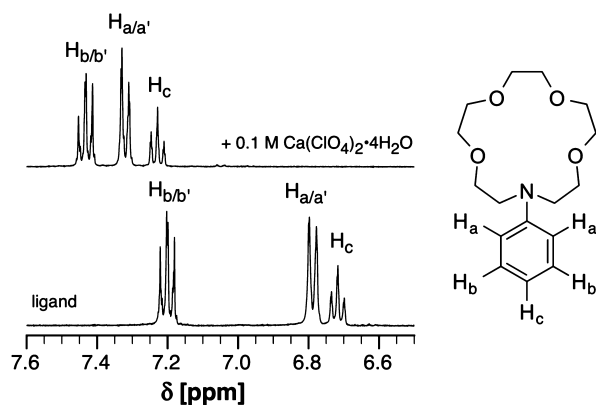


Figure 2. Aromatic region of the ^1H NMR spectrum of *N*-phenylaza-15-crown-5 (25 mM) in the presence (top trace) and absence (bottom trace) of 0.1 M $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$ in acetonitrile- d_3 at 298 K.

we restricted the $\text{Ca}(\text{II})$ binding affinity studies to derivatives **1c** and **2c** as representative examples. Both compounds displayed negligible fluorescence emission in their free form but responded with strong fluorescence enhancements upon saturation with $\text{Ca}(\text{II})$ (Figure 3). Due to the low quantum yield of the free forms, it was not possible to extract reliable fluorescence enhancement factors; nevertheless, the data indicate that the fluorescence contrast exceeds 1:1000 for both probes. Given the rapid equilibration between different

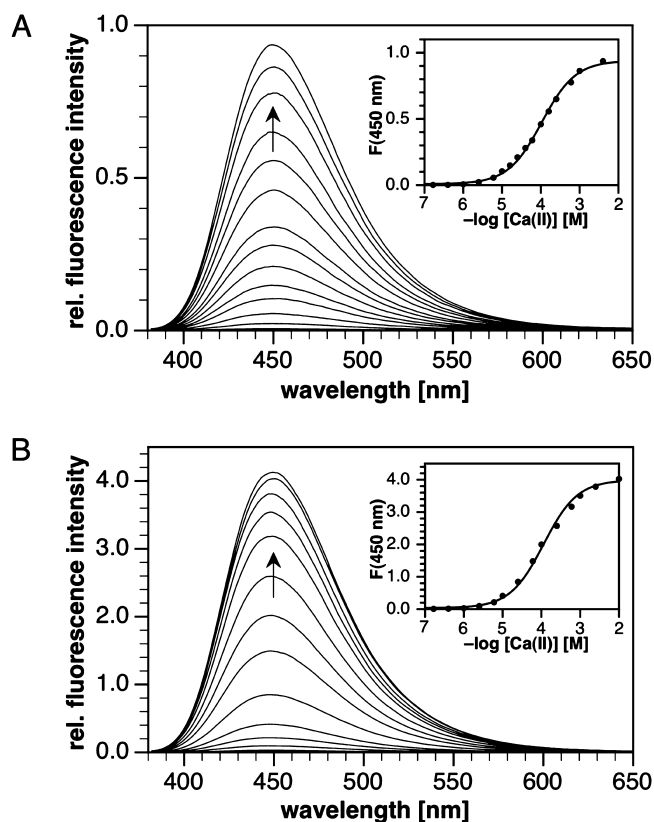


Figure 3. Fluorescence emission spectra of pyrazoline derivative **1c** (A) and **2c** (B) in acetonitrile as a function of the total concentration of $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$. The inset shows the fluorescence intensity at 450 nm and the corresponding $\text{Ca}(\text{II})$ concentrations for each trace, as well as the curve-fit for a binding isotherm with 1:1 metal-probe stoichiometry using equation S5 (Supporting Information).

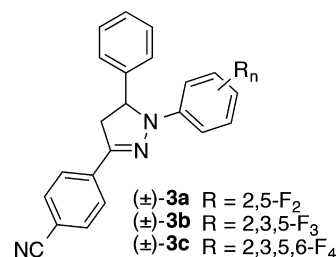
Table 2. Time-Resolved Fluorescence Decay Data for Pyrazoline Derivatives **1a–c** and **2a–c** and Reference Compounds **3a–c** in Acetonitrile Containing 0.1 M $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$ at 25 °C

compd	R	τ /ns	χ^2 ^a
1a	2,5-F ₂	3.22	1.578
		4.06 (0.26); 2.89 (0.74) ^b	1.212
1b	2,3,5-F ₃	3.61 (0.26); 1.88 (0.74) ^b	1.104
1c	2,3,5,6-F ₄	2.99 (0.26); 0.91 (0.74) ^b	1.220
2a	2,5-F ₂	3.95	1.318
2b	2,3,5-F ₃	3.78	1.166
2c	2,3,5,6-F ₄	3.01	2.412
		3.19 (0.88); 1.62 (0.12) ^c	1.408
3a	2,5-F ₂	3.99 ^d	
3b	2,3,5-F ₃	3.74	1.248
3c	2,3,5,6-F ₄	3.09 ^d	

^aGoodness of fit parameter. ^bGlobal least-squares fit with biexponential decay model (global $\chi^2 = 1.179$). ^cLeast-squares fit with unconstrained biexponential decay model. ^dData from ref 3.

coordination species, the steady-state fluorescence data reflect an ensemble-averaged response of the probes toward $\text{Ca}(\text{II})$ binding. Nonlinear least-squares fitting of the fluorescence intensity at 450 nm as a function of $\text{Ca}(\text{II})$ concentrations fit well to an equilibrium model with 1:1 metal–ligand stoichiometry and yielded the apparent stability constants of $\log K = 3.98 \pm 0.02$ and 3.94 ± 0.04 for derivatives **1c** and **2c**, respectively (Supporting Information). These values are comparable to the $\text{Ca}(\text{II})$ stability constants of other fluorescent probes featuring the same parent ligand *N*-phenylaza-15-crown-5.^{8,25,27} In the presence of 1% (0.56 M) additional water, the apparent stability constant of **1c** decreased to $\log K = 2.12 \pm 0.02$, and even further to $\log K = 1.71 \pm 0.02$ in the case of derivative **2c** (Figures S1 and S2, Supporting Information). The somewhat lower affinity of **2c** is possibly due to additional stabilization of the N–H functionality in the free ligand through hydrogen bonding with water molecules, which must be reorganized upon binding of $\text{Ca}(\text{II})$.

Time-Resolved Fluorescence Decay Data. The lower recovery quantum yields observed for **1a–c** compared to **2a–c** upon saturation with $\text{Ca}(\text{II})$ is indicative of a weakened interaction between the metal ion and the aniline nitrogen donor, possibly involving the formation of various equilibrating coordination species. To investigate whether multiple coordination species contribute to the observed fluorescence recovery, we performed picosecond time-resolved fluorescence decay measurements. Depending on the interaction strength between the metal ion and PET donor, the different coordination species are expected to exhibit distinct fluorescence decay lifetimes. Each probe was dissolved in acetonitrile containing 0.1 M $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$, a concentration at which complex formation is complete by >99.9% based on the measured $\text{Ca}(\text{II})$ stability constants. All compounds were excited at 372 nm with a pulsed laser, and the decay signal was collected at the maximum emission wavelength by single photon counting. The fluorescence lifetimes were determined from nonlinear least-squares fitting of the decay profiles with mono- or biexponential decay models (Table 2). We also include in Table 2 data from reference compounds **3a–c** carrying a simple phenyl ring in the 5-position in lieu of the arylazacrown moiety to gauge the intrinsic lifetime of the fluorophore in the absence of PET.³



As evident from the multiexponential fluorescence decay profiles displayed in Figure 4A, the pyrazoline derivatives **1a–c** produce more than a single coordination species with $\text{Ca}(\text{II})$. Although the time trace of **1a** can be fit to a single lifetime of 3.22 ns, the quality of the fit improves significantly with a biexponential decay model (Table 2). Because the metal binding site is electronically decoupled from the fluorophore π -system, the species composition is expected to remain uniform across all three derivatives. In agreement with this notion, it was possible to extract a single pair of pre-exponential factors through global least-squares curve fitting²³ of the three decay profiles using a biexponential decay model (Table 2). Despite the fitting constraints, each decay profile fit to the same pair of pre-exponential factors with an excellent global χ^2 value of 1.179 (Table 2). The longer lifetime component, which contributes 26% to the decay signal, matches within ~ 0.1 ns the lifetime of the respective reference compounds **3a–c**, indicating effective inhibition of PET and complete fluores-

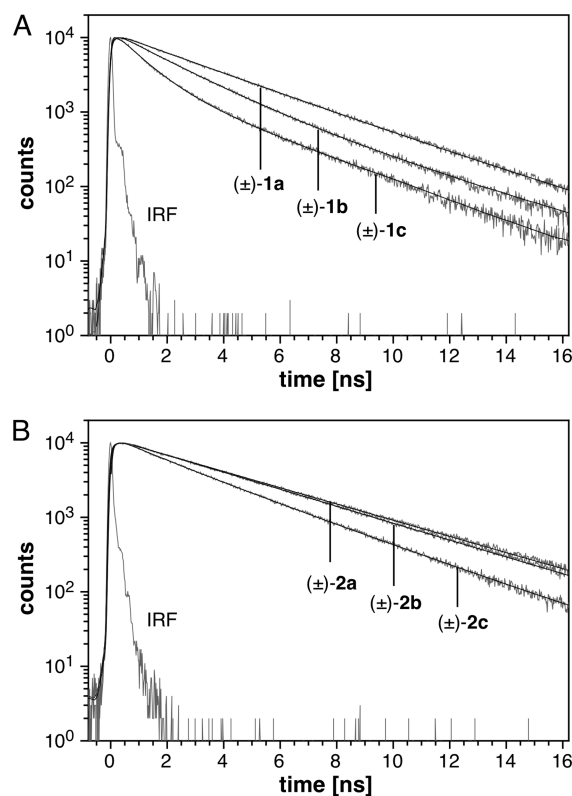


Figure 4. Fluorescence decay profiles of pyrazoline derivatives **1a–c** (A) and **2a–c** (B) in acetonitrile in the presence of 0.1 M $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$. Each sample was excited at 372 nm (80 ps fwhm), and the decay signal was detected at the maximum emission wavelength by single photon counting. Nonlinear least-squares fitted traces are shown as solid lines (IRF = instrument response function; see Table 2 for curve fitting data).

cence recovery. In contrast, the lifetime of the more prevalent component, which contributes 74% to the decay signal, is consistently shorter and decreases significantly from 2.89 to 0.91 ns with increasing PET driving force (Table 2). Consistent with the observed decrease of the ensemble-averaged quantum yields (Table 1), the differences in fluorescence decay lifetimes between the two coordination species is more pronounced at higher PET driving force.

To test whether the species with shorter fluorescence lifetimes might arise from direct Ca(II) interactions with the pyrazoline fluorophore, we measured the decay profile under acidic conditions in the presence of 0.1 M $\text{Ca}(\text{ClO}_4)_2 \cdot 4\text{H}_2\text{O}$ and 1 mM methanesulfonic acid. Concluding from the monoexponential decay (Figure S5B, Supporting Information) and fluorescence lifetime of 3.13 ns, which agrees well with that of the unquenched reference compound 3c (Table 1), the emission is completely restored through protonation of the aniline nitrogen and thus Ca(II)–pyrazoline interactions do not appear to constitute a significant nonradiative deactivation pathway. Hence, the fluorescence lifetime data are consistent with the presence of a coordination species in which the metal–ligand donor interaction is weakened to alleviate steric strain imposed by the auxiliary solvent ligands. Interestingly, the fluorescence decay profile of 1c acquired under anhydrous conditions using rigorously dried $\text{Ca}(\text{ClO}_4)_2$ remain unchanged (Figure S5C, Supporting Information), suggesting that acetonitrile rather than water is engaged in ternary complex formation.

In contrast to 1a–c, the fluorescence decay profiles of 2a–c appear to involve only a single component (Figure 4B). In the case of 2a and 2b, deconvolution of the decay profiles with a monoexponential decay model produced excellent least-squares curve fits with lifetimes closely matching those of the reference compounds 3a and 3b, respectively. Although the decay profile of 2c can be fit to a single lifetime of 3.01 ns matching reference compound 3c, the fit is significantly improved with a biexponential decay model. However, unlike in the case of 1c, the longer lifetime component is now the dominant contributor (88%), presumably indicating a strengthened Ca(II)–N interaction. At the same time, the shorter component (1.62 ns) is significantly longer-lived than that of 1c (0.94 ns), despite the greater PET driving force of the free probe. This effect may be due to coordination of Ca(II) to the anisole oxygen of the PET donor even in the absence of a strong Ca–N interaction. Altogether, these data demonstrate that the modified azacrown ether ligand of probes 2a–c engages Ca(II) more effectively in coordination to the actual electron donor moiety, which in turn results in more effective inhibition of PET quenching and full fluorescence recovery with a greater than 2-fold improvement in quantum yield for the tetrafluoro derivative 2c.

4. CONCLUSIONS

Taking advantage of the tunable PET framework offered by 1,3,5-triarylpyrazoline fluorophores, we were able to specifically probe for rapidly equilibrating coordination species of two arylazacrown ether derivatives by picosecond time-resolved fluorescence spectroscopy. Similar to previous observations on Cu(I)-selective thiaza crowns,⁴ the Ca(II)-solution chemistry of the *N*-phenylaza-15-crown-5 parent ligand of 1a–c is characterized by the presence of at least two coordination species with distinct fluorescence decay lifetimes. As the difference in lifetimes not only is a function of the PET donor

strength of each species but also depends on the contribution of the fluorophore to the PET driving force, the latter can be used as a tool to amplify lifetime differences and thus to differentiate between species with only subtle variations in PET donor abilities. As demonstrated for 1a–c, the lifetime difference between the two components was small at the lowest PET driving force, resulting in an apparent monoexponential decay profile but substantially increased at more favorable driving forces to yield a biexponential decay with distinct lifetimes. In contrast, the fused-ring azacrown ligand of 2a and 2b produced uniform decay profiles with lifetimes that mirrored those of the reference compounds 3a and 3b, suggesting full fluorescence recovery upon saturation with Ca(II). Even the tetrafluoro derivative 2c, which offers an initial PET driving force 90 mV higher than that of 1c, achieved nearly complete fluorescence recovery with only 12% of a shorter-lived component in the decay lifetime. These differences are probably due to the more favorable ligand architecture of 2a–c toward Ca(II) coordination, where the aryl ring is fully integrated into the ligand backbone and thus able to alleviate steric strain imposed by the ortho-hydrogens and solvent auxiliary ligands as in the case of 1a–c. In summary, ternary complex formation with solvent molecules plays an important but often overlooked role in the design of high-contrast cation-selective fluorescent probes. In addition to the solvent, other adventitious ligands may contribute to ternary complex formation, e.g. if fluorescent probes are operating in a biological environment, as recently demonstrated by Bal and co-workers for the ZnAF family of zinc sensors.³⁷ The combination of time-resolved fluorescence spectroscopy with selective tuning of the PET driving force offers an effective analytical tool to probe ternary complex formation and thus to optimize ligand architectures for the design of high-contrast cation-responsive fluorescence switches.

■ ASSOCIATED CONTENT

Supporting Information

Synthetic procedures, ^1H -NMR and ^{13}C -NMR spectra, estimation of the photoinduced electron-transfer driving force (including a table of reduction potentials), data fitting for the determination of stability constants (including fluorescence titration plots), and fluorescence decay profiles. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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