See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/24250872

ChemInform Abstract: Synthesis of Novel Thiourea, Thiazolidinedione and Thioparabanic Acid Derivatives of 4-Aminoquinoline as Potent Antimalarials

ARTICLE in BIOORGANIC & MEDICINAL CHEMISTRY LETTERS · APRIL 2009

Impact Factor: 2.42 · DOI: 10.1016/j.bmcl.2009.03.026 · Source: PubMed

CITATIONS

34

READS

58

6 AUTHORS, INCLUDING:



Naresh Sunduru

9 PUBLICATIONS 223 CITATIONS

SEE PROFILE



Kumkum Srivastava

Central Drug Research Institute

88 PUBLICATIONS 1,419 CITATIONS

SEE PROFILE



Jitendra Saxena

Central Drug Research Institute

83 PUBLICATIONS 776 CITATIONS

SEE PROFILE



Prem M.S. Chauhan

Central Drug Research Institute

85 PUBLICATIONS **1,496** CITATIONS

SEE PROFILE

Synthesis of novel thiourea, thiazolidinedione and thioparabanic acid derivatives of 4-aminoquinoline as potent antimalarials

Naresh Sunduru,^a Kumkum Srivastava,^b S. Rajakumar,^b S. K. Puri,^b J. K. Saxena^c and Prem M. S. Chauhan ^a*

^aDivision of Medicinal and Process Chemistry, Central Drug Research Institute, Lucknow 226001, India

^bDivision of Parasitology, Central Drug Research Institute, Lucknow 226001, India

^cDivision of Biochemistry, Central Drug Research Institute, Lucknow 226001, India

Abstract- In search of new 4-aminoquinolines which are not recognized by CQR mechanism, thiourea, thiazolidinedione and thioparabanic acid derivatives of 4-aminoquinoline were synthesized and screened for their antimalarial activity. Thiourea derivative 3 found to be the most active against CQ sensitive strain 3D7 of *P. falciparum* in an *in vitro* model with an IC₅₀ of 6.07 ng/mL and also showed *in vivo* suppression of 99.27% on day 4 against CQ resistant strain N-67 of *P. yoelii*.

Malaria is a devastating disease caused by four species of genus plasmodium which afflicts more than 40% of the world population, causing an estimated mortality of 1.5-2.7 million people annually. The current epidemic is fueled about the parasite Plasmodium falciparum, responsible for the most deadly cases of malaria and its resistance to the antimalarial drugs. Chloroquine (CQ) has historically been mainstay of malaria treatment, particularly with P. falciparum in pregnant women and children under the age of five.² It acts by binding to heme molecules released from the hemoglobin that is digested by malaria parasites as they grow within their host red blood cells. This binding interferes with the process by which heme is normally incorporated into inert crystals (β-hematin) and detoxified, thereby accumulation of toxic levels of heme leads to the death of parasite.

Chloroquine resistance (CQR) in *P. falciparum* is primarily conferred by complex point mutations in *P. falciparum* resistant transporter (PfCRT), a putative transporter involved in drug flux and proton equilibrium

Keywords: Antimalarial; *P. falciparum*; 4-Aminoquinoline; Thiourea; Thiazolidinedione; Thioparabanic acid.

across the digestive vacuole membrane, which is preventing the parasite by non-accumulating lethal concentration of the chloroquine.^{3,4} Thus, developing resistant strains of chloroquine and other drugs are forcing & alarming the researchers for the development of new effective antimalarial agents.

$$\begin{array}{c|c} & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & &$$

Figure 1. Structure of Chloroquine (CQ) and synthesised compounds

Among old and new drug targets of malaria, host heme molecule remains one of the most attractive target and 7-chloroquinoline compounds are very selective towards heme binding. 5-7 So, rather than identifying the new molecules for efficacy, 7-chloroquinolines having many advantages and efficiency are now in priority for antimalarial chemotherapy. Based on this observation, modifications of 1,4-diaminoalkyl chain of chloroquine has been done and promising results against chloroquine sensitive and resistant strains of plasmodium were obtained, due to non-recognization of these molecules by chloroquine resistant (CQR) mechanism. 8-12 Keeping this in view we hypothesized and synthesized new prototypes by incorporating new entities like thiourea, thiazolidinedione and thioparabanic acid on 4-aminoalkyl chain of 7-chloroquinoline. In this

^{*} Corresponding author. Tel.: +91 522 221 2411x4332; fax: +91 522 262 3405; e-mail addresses: prem_chauhan_2000@yahoo.com; premsc58@hotmail.com

manuscript, we would like to report the antimalarial activity of these new prototypes against both sensitive, resistant strains of chloroquine and also the inhibition of β -hematin formation.

Scheme 1. Reagents and conditions: (a) α,γ-diaminoalkanes, reflux, 4h; (b) isothiocyanates, CH₃CN, rt, 30-60 min; (c) (COCl)₂, DCM, 0 °C, 30 min; (d) ClCOCO₂Et, DCM, 0 °C, 30 min.

Our synthesis approach toward the targeted compounds (2-25), involved less reaction time and good yielding steps with commercially available 4,7-dichloroquinoline as outlined in **Scheme 1**. Amination of 4,7dichloroquinoline with α, γ -diaminoalkanes gave N-(7chloro-4-quinolyl)-diaminoalkanes 1 in 80-87% yield.8 Followed by reacting 1 with different isothiocyanate (Table 1) in acetonitrile for 30-60 minutes furnished respective thioureas (2-9) in 76-80% yield. Finally, to obtain the structural isomeric derivatives thiazolidinedione (10-17) and thioparabanic acid (18-25), thiourea derivatives (2-9) were cyclised with oxalyl chloride and chlorooxoethylacetate respectively, 13 at 0 °C in DCM for 30 minutes with the yield in the range of new 72-78%. These compounds were characterized by spectroscopic means and their purity is established by elemental analysis.¹⁴

The compounds were evaluated for their in vitro

antimalarial activity against CQ sensitive 3D7 strain of P.falciparum BY SYBER Green I-based fluorescence (MSF) assay. 15 The compounds were dissolved in DMSO at 5mg/ml. For the assays, fresh dilutions of all compounds in screening medium were prepared and 50 μl of highest starting concentration (500 ng/ml) was dispensed in duplicate wells in row B of 96 well tissue culture plate. The highest concentration for chloroquine was 25 ng/ml. Subsequently two fold serial dilutions were prepared up to row H (seven concentrations). Finally 50 µl of 2.5% parasitized cell suspension containing 0.5% parasitaemia was added to each well except 4 wells in row A which received non infected cell suspension. These wells containing non infected erythrocytes in the absence of drugs served as negative controls, while parasitized erythrocytes in the presence of CQ served as positive control. After 72 hours of incubation, 100 µl of lysis buffer [20 mM tris (Ph 7.5), 5mM EDTA, 0.008% (wt/vol) saponin, and 0.08% (vol/vol) Triton X - 100] containing 1 x concentration of SYBER Green I (Invitrogen) was added to each cell. The plates were re-incubated for one hour at room temperature and examined for the relative fluoroscence units (RFUs) per well using the FLUOstar, BMG lab technologies. The 50% inhibitory concentration (IC₅₀) was determined using non-linear regression analysis dose-response curves.

The compounds inhibitory activity of β-hematin formation was measured according to the described protocol. 16 Male swiss mice, weighing 15-20 gm were inoculated with 1 X 10⁵ P. yoelii infected RBCs. Blood of infected animal at ~50% parasitemia was collected by cardiac puncture in 2.0% citrate buffer and centrifuged at 5000 rpm for 10 min at 4 °C. The plasma was used in assay of β-hematin formation. The assay mixture contained 100 mM sodium acetate buffer pH (5.1), 50 μl plasma, 100 μM hemin as the substrate and 1-20 μg compound/drug in a total volume of 1.0 ml. The control tube contained all reagents except compound. The reaction mixture in triplicate was incubated at 37°C for 16h in a rotary shaker. The reaction was stopped by centrifugation at 10,000 rpm for 10 min at 30°C. The pellet was suspended in 100 mM Tris-HCl buffer pH (7.4) containing 2.5% SDS. The pellet obtained after centrifugation was washed thrice with distilled water (TDW) to remove free hemin attached to β -hematin. The pellet was solubilized in 50 µl of 2N NaOH and volume was made up to 1.0 ml with TDW. Absorbance was measured at 400 nm.

Table 1. Biological activity of the synthesized compounds

In vitro Inhibition of In vivo

Compound	n	R	antimalarial activity $IC_{50} (ng/mL)^{a}$	SI ^b	β-hematin formation IC_{50} (µg/mL) ^c	% suppression on day 4 ^d
2	0	Phenyl	9.22	996.75	9.86	•
3	0	Butyl	6.07	749.61	7.11	99.27
4	0	Allyl	26.11	668.32	7.31	,, <u>,</u> ,
5	0	o-Chlorophenyl	20.51	332.06	5.56	
6	1	Phenyl	10.01	968.11	5.67	
7	1	Butyl	11.82	599.81	6.17	
8	1	Allyl	42.02	581.63	9.23	
9	1	o-Chlorophenyl	10.16	1113.27	6.46	16.82
10	0	Phenyl	29.49	605.14	9.76	
11	0	Butyl	17.44	480.27	9.54	
12	0	Allyl	32.44	602.71	8.38	
13	0	o-Chlorophenyl	12.11	542.58	8.79	5.45
14	1	Phenyl	11.05	637.13	8.42	
15	1	Butyl	11.03	255.76	8.28	
16	1	Allyl	111.61	475.22	6.45	
17	1	o-Chlorophenyl	17.48	433.64	9.72	
18	0	Phenyl	119.10	118.22	10.71	
19	0	Butyl	33.08	405.98	9.33	
20	0	Allyl	104.09	140.64	9.41	
21	0	o-Chlorophenyl	199.31	3.36	13.01	
22	1	Phenyl	150.55	65.49	7.78	
23	1	Butyĺ	69.70	27.25	8.65	
24	1	Allyl	54.85	80.41	12.16	
25	1	o-Chlorophenyl	39.71	122.38	9.02	
CQ		1 2	5.2	8983	4.87	99.9

^a IC_{50} : concentration corresponding to 50% growth inhibition of chloroquine sensitive strain 3D7 of *P. falciparum*; ^b $SI=IC_{50}$ values of toxicity against VERO cell line/ IC_{50} values of antimlarial activity; ^C The 50% inhibitory concentration (IC_{50}) was determined using nonlinear regression analysis dose-response curves; ^d In vivo antimalarial activity against chloroquine resistant strain N-67 of *P. yoelii* in swiss mice at dose 50 mg/Kg/day by intraperitoneal route.

Comparing the antimalarial activity, based on entities incorporated on 4-aminoalkyl chain of 7chloroquinoline, thiourea derivatives showed good antimalarial profile of IC₅₀ ranging from 6.07 to 42.02 ng/mL and thiazolidinedione showed moderate activity with IC₅₀ in the range of 11.03-111.61 ng/mL, while thioparabanic acid derivatives showed below moderate activity of 33.08-199.31 ng/mL in comparison with chloroquine (Table 1). The most active compound is of thiourea derivative (3) consisting ethyl chain, n-butyl group as **R** showed IC₅₀ of 6.07 ng/mL and β-hematin formation inhibition of IC_{50} 7.11 µg/mL. On replacing ethyl with propyl chain (7) activity decreases to 11.82 ng/mL, though its βhematin inhibitory activity is increased to 6.17 µg/mL. Whereas, compounds (2, 6) having phenyl as \mathbf{R} , ethyl and propyl chains showed equal potency with IC50 of 9.22, 10.01 ng/mL respectively. While compound (9) has shown increase in antimalarial activity of IC50 10.16 ng/mL by replacing ethyl (5) with propyl chain

and *o*-chlorophenyl group as **R** even though β -hematin inhibitory activity is decreased to 6.46 from 5.56 μ g/mL. Compounds having allyl group as **R**, ethyl (4)

and propyl (8) chains have shown below moderate activity of IC₅₀ 26.11, 42.02 ng/mL respectively.

Cyclisation of thiourea derivatives lead to drop off in antimalarial profile of thiazolidinedione thioparabanic acid compounds, due to decreased inhibitory activity of β-hematin formation. Compound (11) consisting of ethyl chain and n-butyl group as R shown moderate activity of IC₅₀ 17.44 ng/mL, while by replacing ethyl with propyl chain (15) showed increase in activity of IC₅₀ 12.11 ng/mL due to increase of inhibitory activity of β-hematin formation to 8.28 from 9.54 μ g/mL, similarly compound (14) having phenyl as **R** showed IC₅₀ of 11.05 ng/mL while its ethyl chain analogue (10) showed IC₅₀ of 29.49 ng/mL. Whereas compound (13) having ethyl chain and o-chlorophenyl group as R showed IC50 of 12.11 ng/mL, while its propyl chain derivative (17) showed decrease in activity of 17.48 ng/mL. All the thioparabanic acid derivatives showed less activity in comparison with thiourea and thiazolidinedione due to poor inhibitory activity toward β-hematin formation (Table 1).

These compounds were also tested for their

cytotoxicity against VERO cells using MTT assay. 17 three thiourea Among prototypes, thiazolidinedione derivatives have good selectivity index in comparison with thioparabanic acid derivatives (Table 1). Compound (9) having an IC₅₀ 10.16 ng/mL showed highest selectivity index of 1113.27, while most potent compound (3) having an IC₅₀ 6.07 ng/mL showed selectivity index of 749.61, thus illustrating the good activity profile. Based on this selectivity index, compounds (3, 9 and 13) were also screened in an in vivo model against chloroquine resistant N-67 strain of *P.yoelii* in swiss mice¹⁸ at 50mg/Kg/day for 4 days by intraperitoneal route (ip) (Table 1). Out of three evaluated compounds, thiourea derivative (3) found to be the most active against chloroquine resistant strain with 99.27% suppression on day 4. Thus confirming that the thiourea entity on 4-aminoalkyl chain of 7-chloroquinoline is useful for generating new effective antimalarials for both chloroquine sensitive and resistant strains.

It has become necessary to identify the new 4-aminoquinolines which are not identified by CQR mechanism to overcome the parasite resistance as well as to eradicate global malaria problem. Among all synthesized molecules, thiourea derivative of 4-aminoquinoline 3 showed promising activity against both chloroquine sensitive and resistant strains. Thus, optimization of this new prototype may be useful for the generation of effective antimalarial agents.

Acknowledgements

N. Sunduru thanks the Council of Scientific and Industrial Research, India, for the award of Senior Research Fellowship. We are also thankful to S.A.I.F. Division, CDRI, Lucknow, for providing spectroscopic data. CDRI Communication no.7684

References and notes

- WHO Expert Committee on Malaria. Technical Report Series. Twentieth Report, World Health Organization, Geneva 2000.
- Dominguez, J. N. Curr. Top. Med. Chem. 2002, 2, 1173.
- 3. Wellems, T. E. Science 2002, 298, 124.
- Sidhu, A. B. S.; Verdier-Pinard, D.; Fidock, D. A. Science 2002, 298, 210.
- De, D.; Krogstad, F. M.; Cogswell, F. B.; Krogstad, D. J. Am. J. Trop. Med. Hyg. 1996, 55, 579.
- Ridley, R. G.; Hofheinz, W.; Matile, H.; Jaquet, C.; Dorn, A.; Masciadri, R.; Jolidon, S.; Richter, W. F.; Guenzi, A.; Girometta, M. A.; Urwyler, H.; Huber, W.; Thaithong, S.; Peters, W. Antimicrob. Agents Chemother. 1996, 40, 1846.

- 7. Vippagunta, S. R.; Dorn, A.; Matile, H.; Bhattacharjee, A. K.; Karle, J. M.; Ellis, W. Y.; Ridely, R. G.; Vennerstrom, J. L. *J. Med. Chem.* **1999**, *42*, 4630.
- 8. Madrid, P. B.; Liou, A. P.; DeRisi, J. L.; Guy, R. K. *J. Med. Chem.* **2006**, *49*, 4535.
- Burgess, S. J.; Selzer, A.; Kelly, J. X.; Smilkstein, M. J.; Riscoe, M. K.; Peyton. D. H. J. Med. Chem. 2006, 49, 5623.
- 10. Gupta, L.; Srivastava, K.; Singh, S.; Puri, S. K.; Chauhan, P. M. S. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 3306.
- Yearick, K.; Ekoue-Kovi, K.; Iwaniuk, D. P.; Natarajan, J. K.; Alumasa, J.; de Dios, A. C.; Roepe, P. D.; Wolf, C. *J. Med. Chem.* **2008**, *51*, 1995.
- 12. Natarajan, J. K.; Alumasa, J. N.; Yearick, K.; Ekoue-Kovi, K. A.; Leah B.; Casabianca, L. B.; de Dios, A. C.; Wolf, C.; Roepe, P. D. *J. Med. Chem.* **2008**, *51*, 3466.
- (a) Elokdah, H.; Sulkowski, T. S.; Abou-Gharbia, M.; Butera, J. A.; Chai, S.; McFarlane, G. R.; McKean, M.; Babiak, J. L.; Adelman, S. J.; Quinet, E. M. J. Med. Chem. 2004, 47, 681.
 (b) Mahajan, A.; Yeh, S.; Nell, M.; van Rensburg, C. E. J.; Chibale, K. Bioorg. Med. Chem. Lett. 2007, 17, 5683.
- 14. Spectroscopic data for 3: yield: 78%; mp 158-160 °C; FAB-MS: 337 (M+1); IR(KBr) 3419 (NH), 1216 (C=S) cm⁻¹; ¹H NMR (200 MHz, DMSO-d₆): δ (ppm) 8.38 (d, 1H, J = 5.41 Hz), 8.22 (d, 1H, J = 9.23 Hz), 7.77 (d, 1H, J = 2.07Hz), 7.52 (br-s, 3H), 7.44 (dd, 1H, J = 2.02, 8.89Hz), 6.62 (d, 1H, J = 5.42 Hz), 3.71 (t, 2H, J =5.48 Hz), 3.42 (t, 4H, J = 5.53 Hz), 1.49-1.33(m, 2H), 1.30-1.15 (m, 2H), 0.84 (t, 3H, J = 7.36)Hz); ¹³C NMR (50 MHz, CDCl₃+CD₃OD): 182.05, 155.46, 155.21, 152.33, 139.67, 130.99, 129.65, 127.54, 121.53, 102.37, 49.34, 48.21, 46.61, 35.25, 24.25, 17.84; Anal. Calcd for C₁₆H₂₁CIN₄S: C, 57.04; H, 6.28; N, 16.63. Found: C, 57.12; H, 6.23; N, 16.57. Compound **11:** yield: 73%; mp 180-182 °C; FAB-MS: 391 (M+1); IR(KBr) 3426 (NH), 1773 (C=O), 1615 (C=N) cm⁻¹; 1 H NMR (200 MHz, DMSO-d₆): δ (ppm) 8.81 (br-s, 1H), 8.56 (dd, 1H, J = 2.86, 6.42 Hz), 8.32-8.24 (m, 1H), 7.95 (d, 1H, J =2.25 Hz), 7.62 (dd, 1H, J = 2.02, 9.06 Hz), 6.88(t, 1H, J = 6.24 Hz), 4.08 (t, 2H, J = 5.56 Hz),3.74 (t, 4H, J = 6.93 Hz), 1.55-1.40 (m, 2H), 1.36-1.17 (m, 2H), 0.84 (t, 3H, J = 7.33 Hz); ¹³C NMR (50 MHz, DMSO-d₆): 181.57, 156.68, 154.90, 152.68, 145.71, 141.87, 135.34, 124.95, 124.27, 121.56, 115.66, 97.80, 41.62, 40.02, 39.81, 29.61, 19.75, 13.98; Anal. Calcd for $C_{18}H_{19}CIN_4O_2S$: C, 55.31; H, 4.90; N, 14.33. Found: C, 55.29; H, 4.96; N, 14.26. Compound 19: yield: 75%; mp 154-156 °C; FAB-MS: 391

- (M+1); IR(KBr) 3429 (NH), 1766 (C=O), 1223 (C=S) cm⁻¹; ¹H NMR (200 MHz, DMSO-d₆): δ (ppm) 8.47 (d, 1H, J = 5.39 Hz), 8.04 (d, 1H, J = 9.03 Hz), 7.82 (d, 1H, J = 1.98 Hz), 7.48 (d, 1H, J = 1.88 Hz), 7.44 (br-s, 1H), 6.67 (d, 1H, J = 5.43 Hz), 4.07 (t, 2H, J = 5.86 Hz), 3.79 (t, 2H, J = 7.03 Hz), 3.63 (t, 2H, J = 5.98 Hz) 1.57-1.42 (m, 2H), 1.33-1.21 (m, 2H), 0.91 (t, 3H, J = 7.16 Hz); ¹³C NMR (50 MHz, CDCl₃+CD₃OD): 184.40, 158.04, 157.66, 149.05, 145.60, 141.89, 130.59, 127.54, 125.66, 121.56, 119.78, 101.66, 45.47, 44.08, 43.14, 32.97, 23.24, 16.76; Anal. Calcd for C₁₈H₁₉ClN₄O₂S: C, 55.31; H, 4.90; N, 14.33. Found: C, 55.35; H, 4.83; N, 14.31.
- 15. Smilkstein, M.; Sriwilaijaroen, N.; Kelly, J. X.; Wilairat, P.; Riscoe, M. *Antimicrob. agents Chemother.* **2004**, *48*, 1803.
- Pandey, A. V.; Singh, N.; Tekwani, B. L.; Puri,
 S. K.; Chauhan, V. S. J. pharm. Biomed. Anal. 1999, 20, 203.
- 17. Ctotoxicity of the compounds was determined against VERO cell lines (C-1008; Monkey kidney fibroblast cells) using MTT assay. A total of 1x10⁴ cells/well were incubated with varying concentrations of compound for 72 hours. The highest concentration of compound was 100 μg/ml. The 50% inhibitory concentration (IC₅₀) was determined using nonlinear regression analysis dose-response curves and represented the concentration of compound required to kill 50% of the fibroblast cells. Mosmann T. *J. Immunol. Methods.* **1983**, 65, 55.
- 18. The in vivo drug response was evaluated in Swiss mice infected with P. yoelii (N-67 strain) which is innately resistant to CQ. The mice $(22\pm2g)$ were inoculated with $1x10^6$ parasitized RBC on day 0 and treatment was administered to a group of five mice from day 0 to 3, once daily. The aqueous suspensions of compounds were prepared with a few drops of Tween 80. The efficacy of test compounds was evaluated at 50 mg/kg/day and required daily dose was administered in 0.2 шL volume intraperitoneal route. Parasitaemia levels were recorded from thin blood smears on days 4. The mean value determined for a group of 5 mice was used to calculate the percent suppression of parasitaemia with respect to the untreated control group. Mice treated with CQ served as reference controls. Puri, S. K.; Singh, N. Expl. Parasit. 2000, 94, 8.