

See discussions, stats, and author profiles for this publication at:
<https://www.researchgate.net/publication/51524725>

Recent advances in the multitarget-directed ligands approach for the treatment of Alzheimer's disease

ARTICLE *in* MEDICINAL RESEARCH REVIEWS · JANUARY 2013

Impact Factor: 8.43 · DOI: 10.1002/med.20248 · Source: PubMed

CITATIONS

78

READS

127

3 AUTHORS, INCLUDING:



Rafael Leon

Universidad Autónoma de Madrid

48 PUBLICATIONS 690 CITATIONS

SEE PROFILE



José Marco-Contelles

Spanish National Research Council

347 PUBLICATIONS 5,178 CITATIONS

SEE PROFILE

Recent Advances in the Multitarget-Directed Ligands Approach for the Treatment of Alzheimer's Disease

Rafael León,¹ Antonio G. Garcia,^{2,3} and José Marco-Contelles⁴

¹Department of Chemistry, University of Cambridge, Cambridge, Lensfield road, Cambridge CB2 1EW, United Kingdom

²Instituto Teófilo Hernando, Departamento de Farmacología y Terapéutica, Facultad de Medicina, Universidad Autónoma de Madrid, 28029 Madrid, Spain

³Servicio de Farmacología Clínica, Instituto de Investigación Sanitaria, Hospital Universitario de la Princesa, Madrid, Spain

⁴Laboratorio de Radicales Libres (IQOG, CSIC), C/Juan de la Cierva 3, 28006-Madrid, Spain

Published online 26 July 2011 in Wiley Online Library (wileyonlinelibrary.com).
DOI 10.1002/med.20248



Abstract: With 27 million cases worldwide documented in 2006, Alzheimer's disease (AD) constitutes an overwhelming health, social, economic, and political problem to nations. Unless a new medicine capable to delay disease progression is found, the number of cases will reach 107 million in 2050. So far, the therapeutic paradigm one-compound-one-target has failed. This could be due to the multiple pathogenic mechanisms involved in AD including amyloid β ($A\beta$) aggregation to form plaques, τ hyperphosphorylation to disrupt microtubule to form neurofibrillary tangles, calcium imbalance, enhanced oxidative stress, impaired mitochondrial function, apoptotic neuronal death, and deterioration of synaptic transmission, particularly at cholinergic neurons. Approximately 100 compounds are presently been investigated directed to single targets, namely inhibitors of β and γ secretase, vaccines or antibodies that clear $A\beta$, metal chelators to inhibit $A\beta$ aggregation, blockers of glycogen synthase kinase 3β , enhancers of mitochondrial function, antioxidants, modulators of calcium-permeable channels such as voltage-dependent calcium channels, *N*-methyl-D-aspartate receptors for glutamate, or enhancers of cholinergic neurotransmission such as inhibitors of acetylcholinesterase or butyrylcholinesterase. In view of this complex pathogenic mechanisms, and the successful treatment of chronic diseases such as HIV or cancer, with multiple drugs having complementary mechanisms of action, the concern is growing that AD could better be treated with a single compound targeting two or more of the pathogenic mechanisms leading to neuronal death. This review summarizes the current therapeutic

Contract grant sponsor: Fundación CIEN; Contract grant number: PI 016/09; Contract grant sponsor: Agencia Lain Entralgo; Contract grant number: NDG07/9; Contract grant sponsor: MICINN; Contract grant numbers: SAF2006-08764-C02-01; SAF2009-07271; Contract grant sponsor: CSIC-GRICES; Contract grant number: 2007PT-13; Contract grant sponsors: ISCIII; MICINN; Contract grant number: RD06/0026/1002 Retic "RENEVAS"; Contract grant sponsor: Comunidad de Madrid; Contract grant number: S/SAL-0275-2006.

Correspondence to: Rafael León, Department of Chemistry, University of Cambridge, Cambridge, Lensfield road, Cambridge CB2 1EW, UK, E-mail: rl389@cam.ac.uk

strategies based on the paradigm one-compound-various targets to treat AD. A treatment that delays disease onset and/or progression by 5 years could halve the number of people requiring institutionalization and/or dying from AD. © 2011 Wiley Periodicals, Inc. *Med Res Rev.*, 33, No. 1, 139–189, 2013

Key words: Alzheimer's disease; multitarget drugs; dual AChE inhibitors; AChE peripheral anionic site; multiactive compounds; Ca^{2+} dyshomeostasis; 1,4-dihydropyridines; voltage-dependent calcium channels; amyloid- β antiaggregating agents; antioxidant drugs; anti-inflammatory drugs; NSAIDs; neuroprotection; Ca^{2+} overload; oxidative stress; metal chelators; BACE-1 inhibitors; GSK-3 β inhibitors; ERK2-inhibitors; CDK5 inhibitors; CK-1 inhibitors

1. INTRODUCTION

When Alois Alzheimer treated his patient Auguste D. more than 100 years ago, he did not realize that he was about to describe one of the most complex and challenging diseases in the history of humanity.¹ Since then, a great research effort has been devoted to increase the knowledge and understanding of this disease. Many scientists are currently investigating the pathogenic mechanism of Alzheimer's disease (AD) and millions are invested every year; however, the molecular basis of the disease is yet to be determined. The estimated number of patients needing treatment is 7–8 million in Europe, 4–5 million in the USA contributing to a total of 24 million worldwide. The number is expected to be doubled to 42 million in 2020 due to aging population.² Owing to the sheer number of patients, AD is considered the most common neurodegenerative disorder and a major health concern to societies worldwide.

AD is characterized by progressive memory loss, language skills decline, and other cognitive impairments.³ The etiology of AD is not completely known; however, there are diverse factors such as amyloid- β ($\text{A}\beta$) deposits,⁴ τ -protein (τ) aggregation,⁵ oxidative stress,⁶ and decreased levels of acetylcholine (ACh)⁷, which are thought to play significant roles in the pathophysiology of the disease.⁸ Because of its complexity, AD has been described as a multifactorial disease.⁹ AD develops as a complex network of events, which are all involved and interconnected with the symptoms to induce the subsequent evolution of the disease.

Amyloid plaques are formed, mainly, of $\text{A}\beta$ peptides that aggregate after structural modifications. The interaction of $\text{A}\beta$ peptides with acetylcholinesterase (AChE) increases the rate of $\text{A}\beta$ aggregation and also its toxicity.¹⁰ The τ -protein acts as a stabilizer of the neuron's cytoskeleton. After being hyperphosphorylated, it becomes detached of the microtubules to form intracellular aggregates called neurofibrillary tangles (NFTs). NFTs are aberrant structures linked to the $\text{A}\beta$ -induced neurotoxicity.¹¹ Among the molecular factors linked to AD, the apolipoprotein E (APOE) genotype is implicated in AD pathogenesis. Hence, APOE4 mutations are a high risk factor that can lead to early onset AD.¹² Although a number of mechanisms have been proposed to link APOE4 to $\text{A}\beta$,^{13,14} the general consensus is that APOE4 isoform activity increases $\text{A}\beta$ peptide concentration by decreasing its clearance.^{15–17} Simultaneously, APOE increases the activity of glycogen synthase kinase 3 β (GSK3 β),¹⁸ (kinase that specifically hyperphosphorylates τ protein). On the other hand, it has been established that GSK3 β is essential for $\text{A}\beta$ -induced neurotoxicity.¹⁹ $\text{A}\beta$ not only activates GSK3 β but also cyclin-dependent kinase 5 (CDK5) and extracellular signal-regulated kinase 2 (ERK2), leading to τ hyperphosphorylation and, ultimately, to apoptosis.^{20–22}

The Ca^{2+} ion is one of the most important second messengers in the brain. Tight control of Ca^{2+} homeostasis is crucial for cell survival. In AD, Ca^{2+} imbalance is likely to be one of the main causes of neurodegeneration.^{23,24} Since Khachaturian proposed the pathological role of Ca^{2+} in AD,²⁵ much evidence has substantiated this hypothesis.^{26,27} $\text{A}\beta$ is able to affect neuronal membranes by making them unable to regulate their internal concentration of ions, particularly

Ca^{2+} .^{25,28,29} $\text{A}\beta$ increases the Ca^{2+} influx that occurs when neurotransmitter glutamate activates the *N*-methyl *D*-aspartate receptors (NMDAr).³⁰ This finding is supported by the fact that memantine³¹ (a NMDAr antagonist, the first non cholinergic drug approved for AD) blocks $\text{A}\beta$ -induced Ca^{2+} influx, indicating that drugs restoring the Ca^{2+} balance in neurons might indeed be therapeutic options for the disease.³² The fact that Ca^{2+} imbalance does cause neuron death during the disease evolution suggests that restoration of Ca^{2+} homeostasis may become a new therapeutic strategy.

Recent drug development projects were based on the emergence of new potential targets in different genomic and proteomic studies. Despite all efforts of drug development undertaken, the number of successful drugs and novel targets have been lower than expected during the past few decades.^{33–35} The “one-target one-compound” paradigm was highly successful in the past, thanks to biochemical studies and discovery of the molecular mechanisms that underly common diseases. Biologists were able to define a key target for a particular disease, leading to medicinal chemists strategically designing a molecule that interacts with this target selectively, with a potential drug as the outcome. After 20 years, it is evident that this target-based approach does not always guarantee success. Drugs directed to a single target might not always modify complex systems, even if they act in the way they are expected to proceed. It is very common in the cell to have “back-up” systems yielding the same effect such as gene expression, protein synthesis, receptors response, and protein degradation. Proteins and intermediates involved in this back-up systems can be completely different and therefore, drugs targeting primary pathways will have no effect over this back-up pathway, an effect known as redundancy.³⁶ Moreover, many cellular networks are buffered to prevent major changes in their outputs, despite dramatic changes in the constituents.^{37–40} Complex disorders, such as cancer, cardiovascular disease, depression, and neurodegenerative diseases, tend to result from multiple molecular abnormalities and not from a single defect. By using target-directed drugs it is not always likely that the effect will modify the evolution of the illness.

In the last decade, new strategies are emerging to overcome the lack of efficiency of the “one-target one-compound” development based on the complexity of the disease and the relationship between different pathways in the pathological evolution of the illness. Many of these drug development programs aim to influence multiple targets in a parallel fashion. One of the most widespread multiple target approaches, the so-called combination therapy, is increasingly used to treat many types of diseases, including AIDS, atherosclerosis, cancer, and depression. As one of the newly developed combination therapies, “multitarget lead discovery” is a promising strategy for the identification of unexpectedly novel effects of drug combinations.^{41–45}

A similar approach that might have inherent advantages is the development of ligands including multiple targets, or the so-called “multitarget-directed ligands” (MTDLs).⁴⁶ Multitarget therapeutics can be more efficacious making the biological system more sensitive to the action of a drug with two or more targets simultaneously, thereby, mitigating the redundancy effect. The idea that multitarget drugs might be more effective than those directed to a single target has emerged from efforts made in understanding the action of different treatments such as antipsychotic drugs. One example is Clorazil® (clozapine), which targets a large number of proteins and has moderate side effects. In an effort to reduce its side effects, a range of analogs were developed, attempting to bind fewer targets. Surprisingly, many of these analogs had reduced activity, but still showed similar side effects.⁴⁷ Another example is amitriptyline, a dual inhibitor of serotonin reuptake transporter (SERT) and norepinephrine transporter (NET). It appears to combine a faster onset of action with increased efficacy.^{48,49} This effect results from the demonstration of antagonism of 5-hydroxytryptamine receptors (5-HT) subtypes namely 5-HT_{2A}, 5-HT_{2C}, 5-HT₆, 5-HT₇, α_1 -adrenergic, histamine H₁, muscarinic ACh receptors, and σ_1 receptor agonist properties.^{50–56}

Some examples of marketed multitarget drugs are based on cancer drug development.⁵⁷ Sutent[®] (sunitinib malate), marketed by Pfizer, is a multitargeted inhibitor of tyrosine kinase (RTC), which blocks epidermal growth factor receptor and two other similar kinases.⁵⁸ On the other hand, sorafenib (Nexavar[®]; Bayer, Leverkusen, North Rhine-Westphalia, Germany), used in the treatment of various types of cancer, is an inhibitor of several protein kinases, vascular endothelial growth factor (VEGF) receptor 2 and 3 kinases and c kit. Sorafenib was the first molecule targeting the Raf/Mek/Erk pathway (within the MAP kinase pathway).⁵⁹ Furthermore, Gleevec[®] (imatinib mesilate),⁶⁰ which represents one of the first developments approved in 2001 by the FDA, is a multikinase inhibitor.⁶¹ Several efficient drugs such as salicylate, nonsteroidal anti-inflammatory drugs (NSAIDs), metformin, or antidepressants⁴⁴ also affect many targets simultaneously.^{44,60,62–66}

The concept of the multitarget approach is particularly applicable to AD. Efficacious and durable responses in AD require multitarget drugs, as the pathogenic mechanisms leading to neurodegeneration are known to be multifactorial.⁶⁷ The molecular basis of AD can be considered as a complex network with multiple pathological crossroads in the system, which must be modified simultaneously in order to induce a positive effect. The above-mentioned complexity of AD expressed as a linked network of several pathological events lead to the final conclusion that the best approach to treat this illness will involve the multitarget approach.⁶⁸ Not surprisingly, some of the most important nodes in the pathological network of AD are the most relevant targets currently under development such as amyloid precursor protein⁶⁹ (APP), β -secretase-1⁷⁰ (BACE-1), GSK3 β ,⁷¹ Ca²⁺,⁷² NFTs, and oxidative stress.⁷³ The multitarget approach is the starting point for the development of ligands capable of modifying the pathological system at several nodes simultaneously, resulting in a major action over the network. This strategy is proposed to avoid redundant mechanisms that could escape to a single-target attack, facilitating the recovery of the cells to a healthy state. This review will focus on the recent efforts devoted to the development of multitarget modifying molecules (Fig. 1).

2. CURRENT TREATMENT OF AD

The most widely used concept for AD drug development is the cholinergic hypothesis,⁷⁴ used for the discovery of palliative drugs during the last 20 years. The cholinergic hypothesis is based on the observation of a deficiency of ACh in the central nervous system. Subsequently, Whitehouse et al.⁷⁵ discovered the degeneration of neurons from the nucleus basalis of Meynert, mainly constituted by cholinergic innervation, in patients with AD. The loss of cholinergic function is closely related to cognitive dysfunction.⁷⁶ Also, in early onset patients observation showed abnormalities such as diminished activity of choline acetyl transferase (ChAT),⁷⁷ enzyme responsible for the synthesis of ACh. Activities of AChE and butyrylcholinesterase (BuChE) were increased and concentrations of nicotinic ACh receptors (nAChRs) were diminished.⁷⁷ As a result of these findings, the primary therapeutic approach to address cognitive loss associated with AD has been the cholinergic replacement strategy. This approach was attempted using muscarinic and nicotinic cholinergic ligands and AChE inhibitors (AChEIs).⁷⁸

Current treatments for AD are mainly based on the inhibition of AChE; however, new hypotheses are emerging. In 1993, tacrine (**1**, Fig. 2), an AChEI, was approved by the FDA for clinical use against AD. After tacrine, three more AChEIs were approved for the treatment of AD namely, donepezil⁷⁹ (**2**, Fig. 2), galantamine⁸⁰ (**3**, Fig. 2), and rivastigmine⁸¹ (**4**, Fig. 2). These three AChEIs are currently the most common treatments for mild to moderate stages of AD sufferers. However, their clinical usefulness is limited, largely because

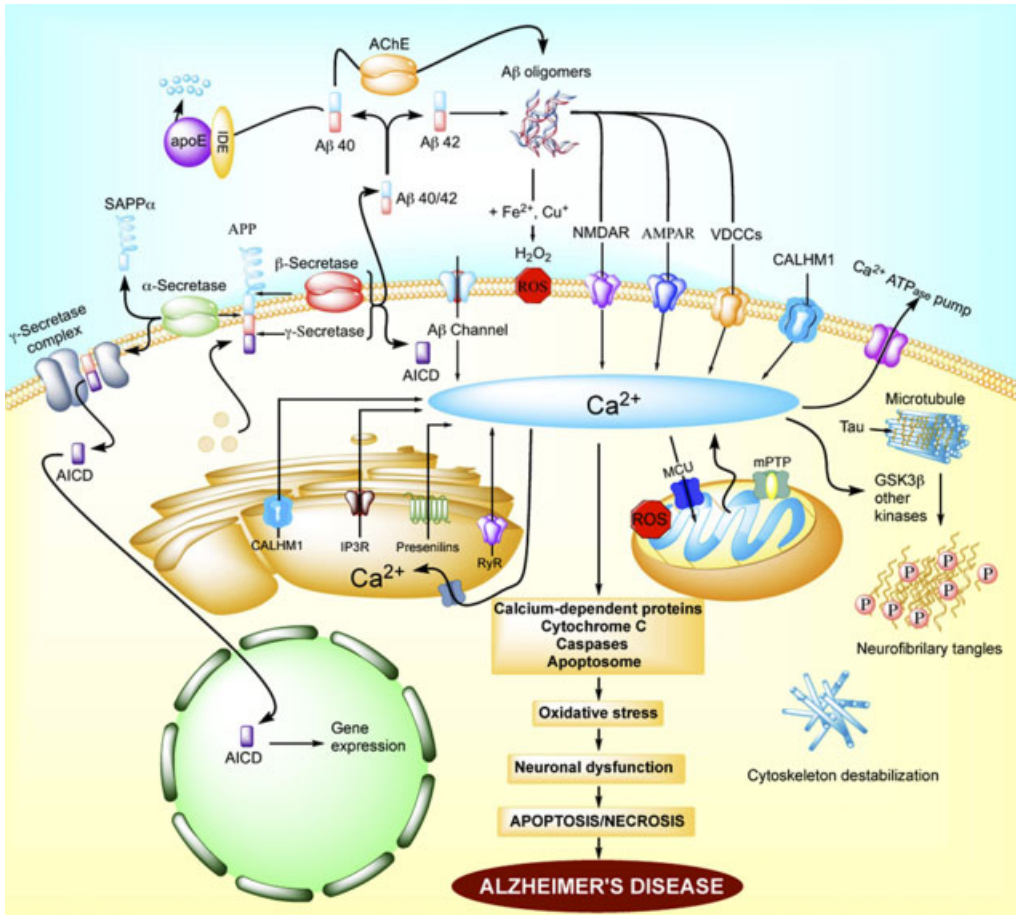


Figure 1. Interconnected pathways in Alzheimer's disease. The nonamyloidogenic pathway starts with the processing of the amyloid precursor protein (APP) by α -secretase to generate the soluble APP fragment α (SAPP α) and a second fragment that is further processed by the γ -secretase complex. In the amyloid pathway, APP is cleaved by β -secretase followed by γ -secretase complex to generate amyloid β peptide (A β 40/42), soluble APP β (sAPP β), and amyloid intracellular domain (AICD). The A β monomers that are released outside neurons can be removed by microglia, which release insulin-degrading enzyme (IDE) that destroys them. This aberrant peptide increases the influx of Ca^{2+} by forming Ca^{2+} permeable channels in the plasma membrane. A β_{42} has high potential to aggregate to form A β oligomers, which rate of aggregation is also increased by the interaction of A β with the peripheral anionic site of AChE. A β oligomers can interact with metal ions Fe^{2+} or Cu^{+} activating the generation of reactive oxygen species (ROS). These species damage the plasma membrane facilitating the influx of different ions resulting in membrane depolarization. Membrane depolarization and subproducts of lipid peroxidation affect the function of different receptor and channels such as *N*-methyl *D*-aspartate receptors (NMDAR), α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPA), voltage-dependent calcium channels (VDCCs). A β oligomers can affect also the activity of these receptors and channels directly. Presenilins (PS) function as Ca^{2+} leak channels at the endoplasmic reticulum (ER), mutated PS are highly related to familial AD. PS have been found at ER membranes. Mutations in PS modify their ability to regulate Ca^{2+} in the ER enhancing Ca^{2+} release through ryanodine receptors (RyR) and inositol triphosphate receptors (InsP3R) channels. There is also evidence that PS can interact directly with InsP3Rs, RyRs, and the SERCA pump to alter ER Ca^{2+} release and uptake. AICD migrates to the nucleus, interacts with transcription regulators, and modifies gene transcription to disrupt Ca^{2+} homeostasis. Elevated Ca^{2+} can also affect the attachment/detachment equilibrium of protein Tau (τ) to tubulin to form the cytoskeleton. Glycogen synthase 3 β (GSK3 β) and other kinases are involved in this equilibrium; Ca^{2+} can modify their activity inducing τ hyperphosphorylation to generate, after its aggregation, the neurofibrillary tangles. A β also affect mitochondria inducing oxidative stress and Ca^{2+} dysregulation resulting in an increase in the production of radicals and decreased production of ATP. High concentrations of Ca^{2+} can be stored by mitochondria through mitochondrial Ca^{2+} uniporter (MCU), resulting in Ca^{2+} overload of mitochondria leading to opening of mitochondrial permeability-transition pore (mtPTP) and apoptosis.

of adverse peripheral effects arising from excessive activation of cholinergic systems, with side effects such as confusion, hallucinations, extreme or sudden changes in behavior, nausea, or stomach pain.⁸² Additionally, the hepatotoxic effects⁸³ of tacrine led to its drop out from

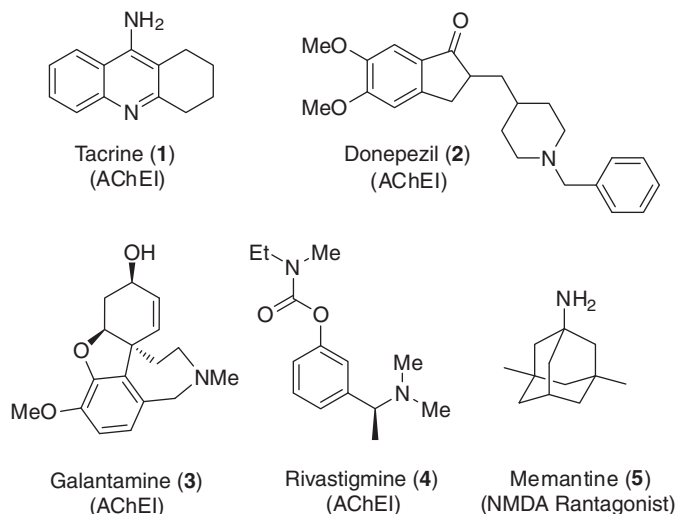


Figure 2. Tacrine and other drugs currently available to treat AD patients.

clinical use. Later on, memantine (**5**, Fig. 2)³¹ was approved for the treatment of AD. Memantine is an NMDA receptor antagonist developed considering the central role of Ca^{2+} in the pathogenesis of AD.⁸⁴

Although the cholinergic hypothesis initiated palliative treatment, increasing evidence is accumulating suggesting that AChEIs have only minor, if any efficacy on AD.⁸⁵ Furthermore, these drugs do not address the underlying causes of the disease.^{85–88} Recently, a new demonstrated effect of AChE, the so-called “non-classical” effect,¹⁰ has arisen increased interest in developing dual interacting inhibitors of this enzyme. New developments under clinical trials include $\text{A}\beta$ antiaggregating or plaque-dissolving agents like pheneserine (an AChE inhibitor and $\text{A}\beta$ synthesis inhibitor, phase III unsuccessful)⁸⁹ and ELND005 (anti-aggregating agent, phase II ongoing).^{90,91}

Inflammation and glia activation observed in AD patients suggested that NSAIDs might be effective for the treatment and more importantly, for the prevention of AD.⁹² Although phase III clinical trials with various NSAIDs were unsuccessful, new evidence has emerged supporting this theory, resulting in the development of new molecules.^{93–98} Other targets such as GSK3 β , BACE-1,⁷⁰ γ -secretase,⁹⁹ nAChRs,¹⁰⁰ $\text{A}\beta$ -immunization, histamine-3 (H_3) receptors, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPA), γ -aminobutyric acid (GABA) receptors, as well as neuroprotective and neurotrophic agents are now being addressed. In 2009, Sabbagh reviewed selected molecules and antibodies that are currently in clinical trials with diverse and interesting results.¹⁰¹

Many approaches and targets have been addressed in the treatment of AD.^{23,46,102–107} However, important physiopathological aspects of AD remain unclear. Growing evidence suggests an underlying complex network that comprises genetics, enzyme activities, receptor expression, protein interactions, alteration of metal concentrations, cell cycle survival disruption, ion homeostasis dysregulation, protein misfolding, among others. Medicinal chemists were prompted by this evidence to accept the multifactorial hypothesis, and the idea of the “MTDLs” that Melchiorre and coworkers proposed.⁴⁶

In this review, we will summarize the most recent approaches on basic developments that will bring the next generation of multitarget AD drugs during the next years. Our summary will reflect the recent understanding of new targets in the design process that leads to new multitarget ligand development.

3. DUAL BINDING SITE AChE INHIBITORS

The development of dual binding site AChEIs is based on the cholinergic hypothesis. As referred to above, the cholinergic system is the most affected in AD patients with almost 80% of cholinergic neuron loss.⁷⁴ Early observations have demonstrated a decrease in ACh concentration within the synaptic cleft. At the same time, a decrease in the activity of ChAT, and hyperactivity of AChE and BuChE is also observed.⁷⁴ These findings led to the rational development of the first generation of AD drugs, AChEIs. Inhibition of AChE increases the concentration of ACh at the synaptic cleft, thereby enhancing central cholinergic neurotransmission. The increased bioavailability of ACh was supposed to be the treatment of the cognitive and behavioral symptoms of the patients. More than a decade since the approval of the first AChEI, some concerns about their benefits are emerging.¹⁰⁸

Nevertheless, in 1996,¹⁰⁹ the interest in this kind of drugs was renewed due to the first demonstration of the so-called “non cholinergic action of AChE.”¹¹⁰ Inestrosa et al. proved for the first time the relationship between AChE and the aggregation of A β .¹⁰⁹ Interaction of A β at the peripheral anionic site (PAS) of AChE greatly accelerates the aggregation of this toxic peptide.¹¹¹ Interactions at the PAS of AChE catalyze some conformational changes in A β fibrils to form the β -sheet with increased aggregating potential.¹¹² Thus, inhibitors targeting the PAS of the enzyme will decrease the aggregation rate of A β keeping it in solution, therefore facilitating its clearance. This strategy could represent a new area of research for the AD treatment based on both, the cholinergic and the amyloid hypothesis.

Early developments on dual inhibitors of AChE were based on the elucidation by Sussman et al. of the 3D structure of AChE¹¹³ and the reported structure of the donepezil-AChE complex.¹¹⁴ Donepezil (**2**, Fig. 2) is able to interact in both, catalytic and PAS binding sites of AChE via extended solvent organization.¹¹⁵ Several studies demonstrated the ability of donepezil (22% inhibition)¹¹² and other AChEIs to decrease the AChE-induced A β aggregation. A large antiaggregating effect was observed for PAS ligands such as propidium (**9**, Fig. 3) (82% inhibition), giving further insight into the connection between PAS and amyloid plaques.¹¹² Subsequent developments of dual binding sites inhibitors were directed toward the modification of donepezil structure, in order to achieve a better pharmacological profile while keeping both interactions. A wide range of *N*-benzylpiperidine analogs were developed with different properties; a key review on those derivatives was reported by Muñoz-Torrero et al.¹¹⁶ Following this hypothesis, new families of compounds were reported

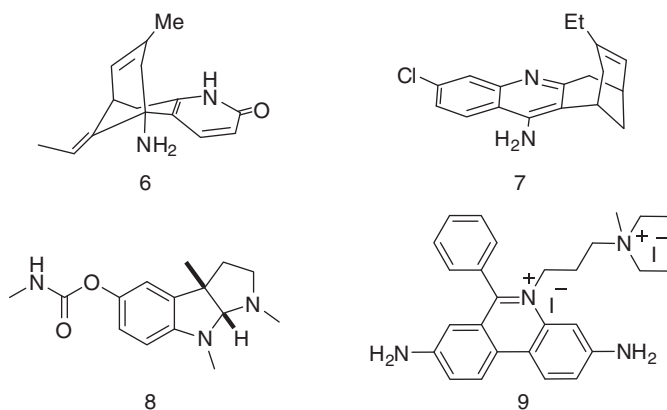


Figure 3. Chemical structure of known AChE inhibitors (–)-huperzine A (**6**), huprine X (**7**), physostigmine (**8**), and propidium iodide (**9**).

as homodimers or heterodimers of known inhibitors such as tacrine (**1**, Fig. 2), donepezil (**2**, Fig. 2),¹¹⁷ galantamine (**3**, Fig. 2),⁸⁰ (–)-huperzine A¹¹⁸ (**6**, Fig. 3), huprine X (**7**, Fig. 3),¹¹⁹ physostigmine (**8**, Fig. 3),¹²⁰ and propidium (**9**, Fig. 3)¹²¹ among others.

Bis(7)-tacrine (**10**, Fig. 4) was one of the first homodimers reported in the literature with increased potency as AChEI ($IC_{50} = 0.4 \text{ nM}$), and showing good selectivity toward BuChE.¹²² Bis(7)-tacrine also improved memory-enhancement capabilities.¹²³ Compound **10** increased the spontaneous quantal ACh release from peripheral cholinergic terminals in the electric organ of *Torpedo marmorata*, at lower concentrations than tacrine. It also had some effect on nAChRs.¹²⁴ Neuroprotective properties were tested in different models where

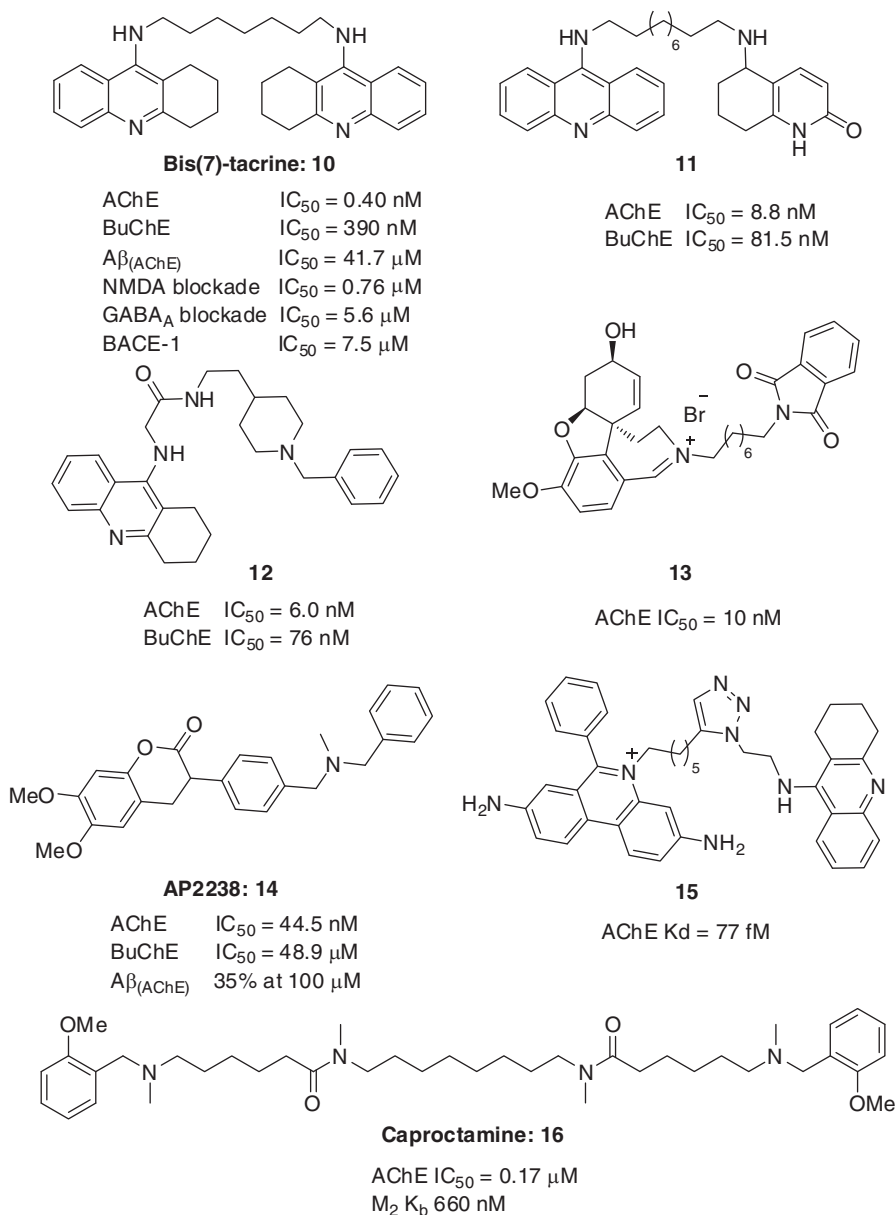


Figure 4. Multitarget drug developments as dual binding site AChE inhibitors.

Bis(7)-tacrine revealed a NMDAR antagonist character, with an IC_{50} of $0.76\ \mu\text{M}$ preventing glutamate-induced neuronal apoptosis.¹²⁵ Interestingly, a few years later a regulatory effect of L-type calcium channels was reported;¹²⁶ such an effect has several neuroprotective implications discussed in following paragraphs. Compound **10** also showed neuroprotection against hydrogen peroxide-induced oxidative stress.¹²⁷ Its large pharmacological profile did not include any $A\beta$ implications until 2007 when Bolognesi et al.¹²⁸ reported for the first time such activity. Bis(7)-tacrine inhibited the AChE-induced $A\beta$ aggregation with an IC_{50} of $41.7\ \mu\text{M}$. A year later, Fu et al. reported a decrease by **10** in the generation of both, secreted and intracellular $A\beta_{42}$ (48.5% reduction at $3\ \mu\text{M}$) and $A\beta_{40}$ (37.7% reduction at $3\ \mu\text{M}$).¹²⁹ An initial explanation might be the interaction of this drug at the PAS of AChE; however, surprisingly, it was able to increase the amount of soluble amyloid protein precursor α fragment ($APP\alpha$) and decrease the aberrant insoluble β -fragment ($APP\beta$). This indicates the activation of a different processing pathway. Fu et al.¹²⁹ demonstrated the possible activation of α -secretase, although it has a minor role (less than 10% activation). More importantly, Bis(7)-tacrine behaved as a BACE-1 inhibitor with an IC_{50} of $7.5\ \mu\text{M}$.¹²⁹

In addition to the homodimers of tacrine, other AChEIs have been used to form heterodimers; this is the case of compound **11**¹³⁰ (Fig. 4). Designed as heterodimer of tacrine and the 5-amino-5,6,7,8-tetrahydro-2(1*H*)-quinolone fragment from (–)-huperzine A (**6**, Fig. 3),¹¹⁸ it is a selective and potent AChEI. Compound **11** ($IC_{50} = 8.8\ \text{nM}$) is 13-fold more potent than (–)-huperzine A, 25-fold more potent than tacrine, and 10-fold more selective toward BuChE than tacrine. Several heterodimers were reported with improved activities and selectivity over the monomers.^{130–132} The crystal structure was solved for this type of heterodimers.¹³² The tacrine moiety binds to the PAS of AChE. To the best of our knowledge, no further studies were performed in order to elucidate their potential $A\beta$ -antiaggregating effect. Donepezil was also dimerized with tacrine, resulting in a new family of inhibitors yielding compounds type **12** (Fig. 4), with similar potency as donepezil.¹³³ In the case of galantamine, homodimers led to better inhibitors than the parent compound.¹³⁴ Galantamine hetero-hybrid **13** (Fig. 4) was the lead compound with an IC_{50} of $10\ \text{nM}$.¹³⁵

In 2003, Piazzini et al.¹³⁶ reported the synthesis and biological activity of AP2238 (**14**, Fig. 4), an interesting molecule based on the study of crystal structures of different AChE–ligand complexes and molecular modeling studies. This compound was designed as a merger of two optimal binders for both catalytic and PAS sites. A benzylamino group and a coumarin (2*H*-2-chromene) were fused with a linker able to interact with the residues at the gorge of the enzyme. AP2238 is a good inhibitor of AChE ($IC_{50} = 44.5\ \text{nM}$), including good antiaggregating properties (35% inhibition at $100\ \mu\text{M}$, 1.6-fold more potent than donepezil).

An inspired approach for the design of dual binding site inhibitors of AChE was reported by Sharpless et al. in 2002.¹³⁷ Using click chemistry as a tool for lead optimization, they reported one of the most potent inhibitors known to date. Compound **15** (Fig. 4) inhibited AChE with a dissociation constant as low as $77\ \text{fM}$. Co-crystallization of **15** with AChE revealed a multivalent interaction, the expected interaction of the separated monomeric moieties and further hydrogen bonding of the triazole ring.¹³⁸ Another example of early developments of dual binding site AChEIs is caproctamine¹³⁹ (**16**, Fig. 4) a polyamine-based dual inhibitor of AChE designed from benextramine,¹⁴⁰ a muscarinic receptor (M_2) antagonist. Caproctamine was able to inhibit AChE ($IC_{50} = 0.17\ \mu\text{M}$) and block the M_2 muscarinic receptor ($M_2\ K_b = 0.66\ \mu\text{M}$).¹⁴¹ Recently, new approaches to dual AChEIs with improved potencies and better $A\beta$ anti-aggregating effects have been reported.^{142–145} Some of these new structures are summarized in Figure 4.

In 2007, Kwon et al.¹⁴² reported a new family of compounds based on the structure of donepezil with the core structure of *N*-substituted piperidines. These have the ability to bind on both sites of the AChE, including effective inhibition of AChE-induced $A\beta_{1-42}$

aggregation and on self-aggregation. Compound **17** (Fig. 5), bearing a *p*-Cl substituent, was selected as the best inhibitor of the family. Compound **17** has an IC_{50} of $0.32\ \mu\text{M}$ to inhibit AChE and a good selectivity (BuChE $IC_{50} = 34.4\ \mu\text{M}$) being 120-fold more active toward AChE. Molecular modeling studies showed an interesting interaction with both catalytic and PAS sites. This suggests that the 4-chlorobenzene moiety in **17** might be placed at the bottom of the gorge, bound by π - π stacking to the aromatic indole-ring of Trp84. The aliphatic alkyl chain of **17** is placed in the middle of the gorge surrounded with phenyl rings of Tyr121, Phe330, and Tyr334. Compound **17** was approximately 37-fold less potent inhibiting AChE than donepezil; however, **17** was able to inhibit AChE-induced A β aggregation by 42.4% at $1\ \mu\text{M}$ and by 55.5% at $100\ \mu\text{M}$. It is more effective than tacrine and donepezil, which were unable to inhibit the aggregation at these concentrations. Furthermore, compound **17** was able to inhibit A β self-aggregation with an IC_{50} of $28.2\ \mu\text{M}$, 3-fold more potent than donepezil.

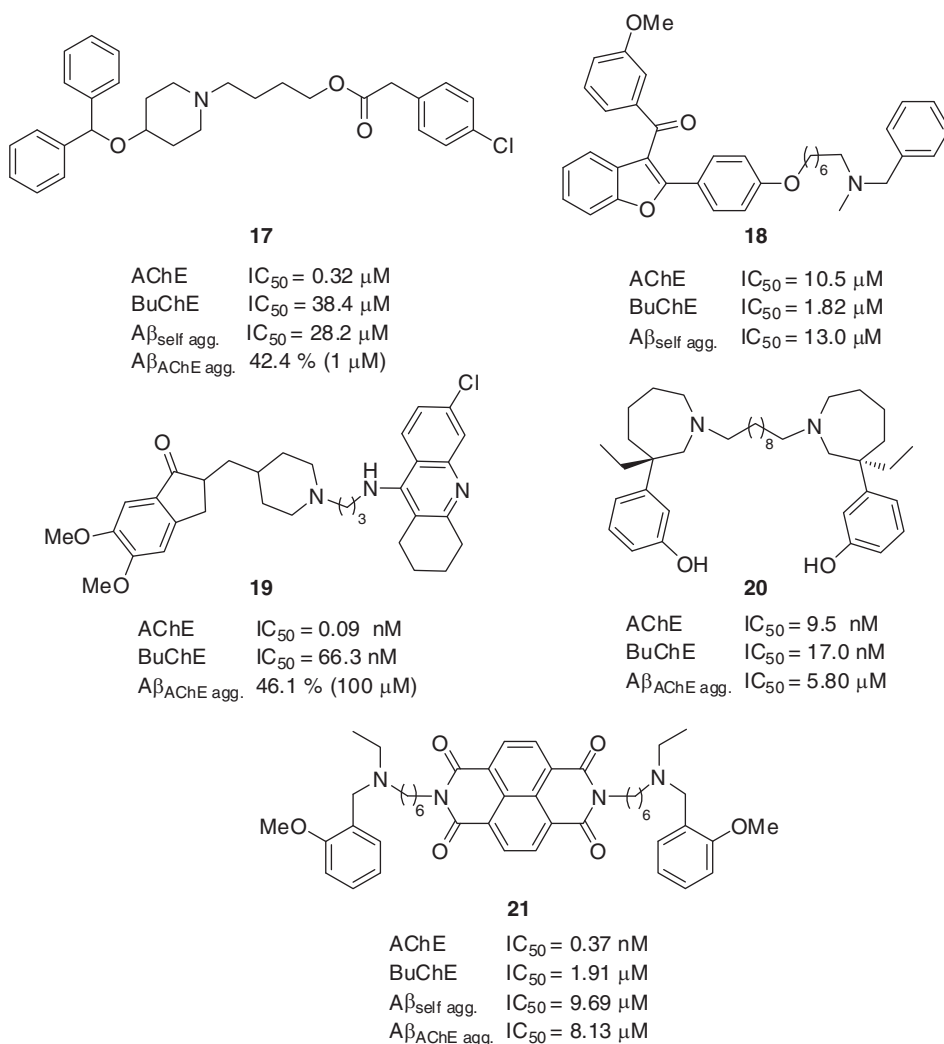


Figure 5. Dual binding site AChE inhibitors with improved anti-A β aggregating properties.

Evidence indicates that BuChE might be a co-regulator of the activity of the neurotransmitter ACh.¹⁴⁶ Remarkably, cortical levels of BuChE show a significant increase in AD patients. This observation prompted the development of selective inhibitors to determine the role of this enzyme and the therapeutic feasibility of its inhibition.^{147,148} Compound **18** (Fig. 5)¹⁴³ was designed as a new selective BuChE inhibitor, maintaining the ability to bind to the PAS of AChE. Compound **18** was based on two molecules, *N*-Methyl-*N*-benzylamine,¹⁴⁹ an AChEI and compound SKF-64346, a benzofuran derivative with good properties as an inhibitor of A β -fibril formation.¹⁵⁰ *N*-methyl-*N*-benzylamine moiety was linked to the benzofuran residue with a heptyloxy chain. Compound **18** showed an IC₅₀ of 1.82 μ M to inhibit BuChE and was 5.7-fold more selective toward BuChE than AChE (IC₅₀ = 10.5 μ M). Some derivatives are even more selective, up to 100-fold difference in inhibition. Furthermore, compound **18** was able to inhibit the A β self-aggregation with an IC₅₀ of 13 μ M and showed a marked neuroprotective effect against A β _{25–35} peptide-induced neurotoxicity with a maximum neuroprotective effect of 63% at 30 μ M.

Compound **19** (Fig. 5) was developed as a dual binding site inhibitor based on donepezil and tacrine, reported by Camps et al. in 2008.¹⁴⁴ It was designed on the basis of the binding modes of donepezil¹¹⁴ and tacrine¹⁵¹ within TcAChE, by combining the 5,6-dimethoxy-2[(4-piperidinyl)-methyl-1-indanone] moiety of donepezil with tacrine. Most of the donepezil structure was maintained in order to allow favorable interactions of the indanone ring at the PAS, piperidine moiety at the centre of the gorge, and tacrine at the bottom of the cavity. This family were potent inhibitors of AChE, where compound **19** was a subnanomolar inhibitor with an IC₅₀ of 0.09 nM, more potent than all parent compounds (tacrine, 6-chlorotacrine and donepezil). Derivative **19** was also a potent inhibitor of BuChE (IC₅₀ = 66.3 nM). The higher inhibitory activity against AChE in comparison with BuChE of tacrine-based homo- and heterodimers is ascribed to the lack of PAS of BuChE,^{122,152} however, recent studies have suggested that Phe278 would be responsible for π – π interactions with aromatic moieties of tacrine-based heterodimers, thus explaining the higher inhibitory potencies in this case.¹⁵³ Furthermore, compound **19** inhibits the AChE-induced A β aggregation up to 46.1% (at 100 μ M) presumably by interaction at the PAS of the enzyme. In this line, **19** decreased thioflavin T (an A β -aggregate¹⁵⁴ and AChE-PAS¹⁵⁵ specific binder) fluorescence by 57% at 100 μ M, an indirect proof of direct interaction of **19** with the PAS.

A new approach to the development of dual binding site AChE inhibitors was reported by Xie et al. in 2008.¹⁴⁵ Their approach was focused on meptazinol,¹⁵⁶ a racemic analgesic opioid with low addiction liability. Its minus enantiomer is a moderate AChE inhibitor.¹⁵⁷ Based on molecular modeling and pharmacological data, they synthesized a new series of homobivalent (–)-*N*-demethylmeptazinols; compound **20**¹⁴⁵ was found to be the family lead with high potency to inhibit AChE (IC₅₀ = 9.5 nM). It was 1.8-fold more potent against AChE than BuChE. The crystal structure of compound **20**¹⁵⁸ demonstrates a dual interaction simultaneously with both the catalytic site and PAS of AChE. Key interactions in both sites are π -interaction, cation π -interaction and two hydrogen bonds at the catalytic site, explaining the activity on BuChE. Further testing with thioflavin T-based assay resulted in a high activity of compound **20**, preventing the AChE-induced A β -aggregation with an IC₅₀ value of 5.8 μ M (95% aggregation inhibition at 100 μ M). Continuing an extensive program in the development of MTDLs performed by Melchiorre and co-workers, Tumiaiti et al.¹⁵⁹ reported the synthesis and pharmacological profile of polyamine-based dual inhibitors with special emphasis on the investigation of the inner spacer. Caproctamine¹⁶⁰ (**16**, Fig. 4) and related derivatives were unable to inhibit the AChE-induced A β aggregation, although they made contact with both PAS and active AChE binding sites.¹⁶⁰ New molecules, such as compound **21**,¹⁵⁹ were designed aiming to test if this failure in the inhibition of A β aggregation was due to their high structural flexibility. Results revealed that the insertion of

constrained moieties strongly influenced the ability to inhibit AChE and BuChE. Compound **21** having a 1,4,5,8-naphthalenetetracarboxylic diimide moiety was the best inhibitor ($IC_{50} = 0.37 \text{ nM}$), with a 9-fold improvement in potency in comparison with its parent analog (bis-piperidin derivative).¹⁵⁹ Compound **21** was also the most selective and potent of the series with an AChE/BuChE ratio greater than 5,000. Looking at its potential properties as A β aggregating inhibitor agent, kinetic studies reveal the dual ability of compound **21** to interact with both binding sites of AChE. Thioflavin-T-based assay determined an IC_{50} of $8.13 \mu\text{M}$ (90% inhibition at $100 \mu\text{M}$) to inhibit AChE-induced A β aggregation. Finally, **21** was able to inhibit the A β -self-aggregation with an IC_{50} of $9.69 \mu\text{M}$. In this case, the modification of the 1,4,5,8-naphthalenetetracarboxylic diimide structural motif results in a more potent activity and better pharmacological profile that deserves further in vivo development.

Proof of the interest of the dual binding site AChEIs type of compounds is the fact that NP-61 a member of the dual AChEIs type of compounds and is the first one to be in phase II clinical trials for AD.¹⁶¹

4. DUAL BINDING AChE AND BACE-1 INHIBITORS

A β is proposed to play a key role in the pathogenesis of AD.^{69,162} A β is formed through the amyloidogenic pathway, in which APP is sequentially cleaved by BACE-1 and γ -secretase, rather than through nonamyloidogenic processing by α -secretase.¹⁶³ BACE-1, an aspartyl protease, cleaves APP in a first instance at the extracellular domain of APP.^{99,164,165} Subsequent cleavage by γ -secretase produces A β fibrils in their soluble form.⁹⁹ Further structural modifications of soluble A β fibrils form high β -sheet content peptides, inducing their aggregation to form senile plaques. As before, AChE can catalyze these conformational changes accelerating the aggregation process. Activation of γ -secretase and the inhibition of BACE-1 and/or γ -secretase reduce the production of A β . Targeting these three secretase activities is currently an objective in the development of new treatments for AD.^{166,167} According to the amyloid hypothesis, a drug able to reduce brain A β levels should act as an effective drug against the disease. Both enzymes, AChE and BACE-1 are connected by the production-aggregation process; therefore, development of agents targeting at both enzymes should be a good approach to design effective pharmaceutical agents.

Bis(7)-tacrine (**10**, Fig. 4) has been recently reported as a selective BACE-1 inhibitor.¹²⁹ In experiments performed in N2a APPswe cells, bis(7)-tacrine reduced the generation of both secreted and intracellular A β_{42} and A β_{40} in a concentration-dependent manner up to 48.5% (A β_{42} , $3 \mu\text{M}$) and 37.7% (A β_{40} , $3 \mu\text{M}$) without any toxic effect for cell viability at those concentrations. In order to rationalize this result, the authors measured the amounts of APP cleavage products in presence and in absence of bis(7)-tacrine. A reduction in the amount of C-terminal fragments β , and increased amount of C-terminal fragments α , was observed without affecting the overall expression of APP. This could indicate a significant inhibition of BACE-1 or an activation of α -secretase. Bis(7)-tacrine was found to be a moderate activator of α -secretase and a selective potent inhibitor of BACE-1 with an IC_{50} of $7.5 \mu\text{M}$. Furthermore, bis(7)-tacrine mitigates A β -induced neuronal apoptosis,¹²⁶ corroborating the multiple-target activities in the amyloid pathological cascade of AD.

Dual inhibition of AChE and BACE-1 is not an easy task and only a few privileged structures are able to effectively inhibit both enzymes. In 2008, Piazza et al. reported the design and evaluation of the first dual inhibitors, exemplified by compound **22**.¹⁶⁸ Its design was based on compound **14** (Fig. 4) (AP2238,¹⁶⁹ AChE dual binding inhibitor) and BACE-1 inhibitors bearing a dihalophenyl acid motif was reported in the literature.^{170,171} Compound **22** contains the core structure of AP2243; the methoxy groups of the coumarin were

alternatively substituted by the amidic chain to extend the activity to BACE-1. Amidic-coumarin derivatives were found to be among the poorest inhibitors of AChE compared with the dimethoxy-derivatives; however, they were successful inhibitors of BACE-1. Compound **22** was the best balanced derivative able to inhibit AChE with an IC_{50} of 181 nM and BACE-1 with an IC_{50} of 150 nM. Compound **22** was designed to be a dual inhibitor of AChE and BACE-1. To the best of our knowledge, no data on its ability to modify A β aggregation or production have been reported.

Similar design methodology of hybridizing different molecules with known activity in one single entity was used to obtain derivative **23**.¹⁷² This was reported in 2009 as dual AChE/BACE-1 inhibitor aiming to include both pharmacological profiles. Development was based on the reported isophthalamide, a widely used pharmacophore for BACE-1 inhibitors^{173–177} and donepezil. It was envisaged that interactions may occur between the *N*-benzylpiperidine group and the catalytic site of AChE, and the isophthalamide moiety at the PAS. The best balanced inhibitor of the series was **23**. Moderate potencies against AChE and BACE-1 (IC_{50} = 1.83 and 0.567 μ M respectively) were observed. In order to elucidate the activity of this compound on the A β production/aggregation by intracellular inhibition of endogenous BACE-1, compound **23** was tested in a cell-based assay using HEK293 cells transfected with human β APP695wt. Compound **23** displayed an excellent inhibitory effect in the cell-based assay on A β production (IC_{50} = 98.7 nM). With this encouraging result, its potential neuroprotective profile of **23** against free radicals (relevancy of oxidative stress in AD will be discussed in following sections) was investigated. Measuring the ability of derivative **23** to protect against H₂O₂ showed a mild protective effect in PC12 cells. Owing to its favorable overall profile, in vivo efficacy was also investigated. Intracerebroventricular administration of compound **23** to APP transgenic mice induced a 29% decrease in endogenous A β_{1-40} production compared with the vehicle-treated control mice.

Propidium (**9**, Fig. 3) binds to the PAS of AChE via π – π stacking interactions with the indole moiety of Trp286.¹⁷⁸ This is reinforced by concomitant cation– π interactions between a quaternary aromatic nitrogen atom and the same residue.¹⁷⁹ After development of a synthetic route to obtain pyrano[3,2-*c*]quinoline scaffold,¹⁸⁰ Camps et al. reported a new series of compounds, structurally similar to **24** (Fig. 6).¹⁸¹ These were designed to interact at the PAS, without being protonated. This property can implement a better profile to cross the blood–brain barrier (BBB). Derivative **24** was a potent inhibitor of AChE (IC_{50} = 14.0 nM) and a mild inhibitor of BuChE (IC_{50} = 1.07 μ M). Molecular modeling predicted interactions between **24** and both catalytic and PAS sites of AChE. With this dual binding site character, **24** was tested as potential A β antiaggregating agent, either AChE-induced or self-induced aggregation, showing good results in both assays (45.7% inhibition at 100 μ M and 47.3% at 50 μ M respectively). Based on the assumption that some dual AChEIs are able to inhibit BACE-1.^{129,168,182–185} Compound **24** was also a potent BACE-1 inhibitor, exhibiting a 78% inhibition at 2.5 μ M and emerging as a promising compound to treat AD. Finally, compound **24** proved to be able to cross the BBB and reach its pharmacological targets located in the central nervous system (CNS).

Finally, Cavalli et al.¹⁸⁵ reported the synthesis and pharmacological profile of the highly promising compound **25** (Fig. 6), called memoquin. Memoquin constitutes the natural evolution of the polyamine-core-based derivatives previously studied in the same group.^{136,141,186–188} Design was based on the existing polyamine core, an anticholinesterasic agent with other interesting properties including muscarinic antagonism and A β anti-aggregating effects. The authors aimed at adding these properties to the ability to counteract oxidative stress introducing a 1,4-benzoquinone fragment from coenzyme Q10 (CoQ10), a fragment with antioxidant properties.^{189,190} Initial evaluation of compound **25** as an anticholinesterasic agent showed promising results with improved ability to inhibit AChE

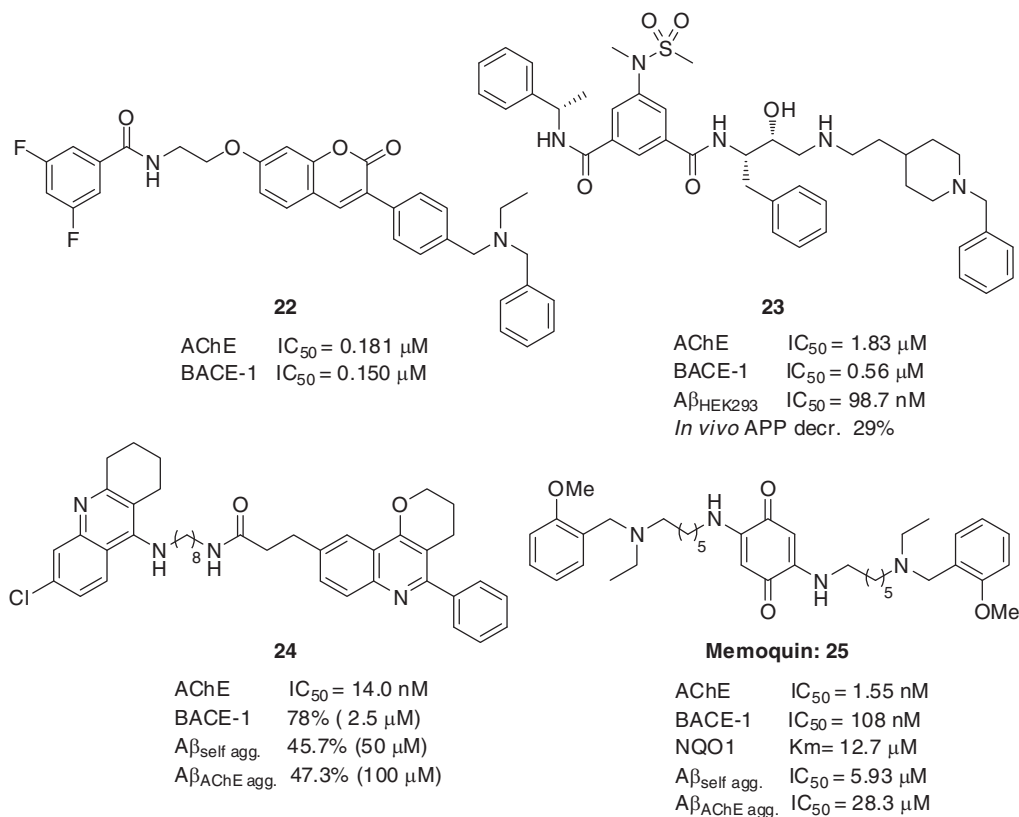


Figure 6. Multitarget-directed ligands as dual inhibitors of AChE and BACE-1.

($IC_{50} = 1.55 nM$), almost 15-fold more potent than the reference compound.¹⁸⁵ Related derivatives proved to be both AChE and self-induced $A\beta$ antiaggregating agents. Memoquin was tested with encouraging results. Memoquin was able to inhibit AChE-induced $A\beta_{1-40}$ aggregation with an IC_{50} of 28.3 μM and the self-induced $A\beta_{1-42}$ aggregation with an IC_{50} of 5.93 μM . This revealed the ability of memoquin to interact with $A\beta$ directly. Furthermore, memoquin was found to inhibit BACE-1 activity at submicromolar concentrations with an IC_{50} value of 108 nM, more potent than other molecules designed to have the same activity. Finally, memoquin's antioxidant properties were investigated. Memoquin was able to reduce the formation of free radicals by 44.1%, a value slightly lower to that for trolox;¹⁹¹ however, this activity was tested only in the oxidized form. In vivo, the 1,4-benzoquinone moiety can be reduced to the 1,4-dihydroquinone form, increasing its antioxidant potential and scavenging properties. The enzyme NAD(P)H:quinine oxidoreductase 1 (NQO1) was shown to be responsible for the reduction of CoQ10 oxidized state increasing its scavenging properties. The capability of this enzyme to reduce memoquin to the 1,4-dihydroquinone form was tested showing that memoquin is a good substrate for NQO1 with K_m and V_{max} values of 12.7 μM and 3,480 ($\mu mol/min$)/mg, respectively. On in vitro cell-based experiments, these authors corroborated the expected increased antioxidant properties of the reduced form by improving its ability to reduce reactive oxygen species (ROS) formation in the presence or absence of sulforaphane (a potent inducer of NQO1).¹⁹² The most potent antioxidant form of memoquin is directly generated in situ in the affected cells, avoiding possible secondary effects. Further in vivo characterization was recently reported,¹⁹¹ demonstrating the

capabilities of memoquin to restore cholinergic deficit, by reverting the neuronal death observed in a mouse model. Memoquin was also able to successfully reduce A β expression and accumulation at the same time as reducing τ hyperphosphorylation. Finally, memoquin proved to ameliorate the behavioral deficits in scopolamine-based object recognition test. The wide pharmacological profile of memoquin gives evidence of the excellent potential of this derivative, endowed with a plethora of activities that could become a disease modifying drug.

5. AChE INHIBITORS AND ANTIOXIDANTS

Oxidative stress is one of the main causes of neuronal death in AD. It refers to the production of high concentrations of ROS in AD patients.¹⁹³ Abnormal pathological mitochondria produce much more O $_2^-$, increasing the concentration of H $_2$ O $_2$ in the cytoplasm. Interaction of this hydrogen peroxide with iron can induce the production of free radicals via Fenton chemistry.¹⁹⁴ This interaction is possible due to the increased concentrations of free iron arising from decreased concentrations of ferritin observed in AD.¹⁹⁵ ROS oxidize lipids and damage membranes in the brains of AD patients.¹⁹⁶ Lipid peroxidation products, for example, aldehydes, arising from polyunsaturated fatty acids oxidation, have much longer half-lives than the radicals. Thus, aldehydes can diffuse to other sites within the cell and react there.^{197,198} Free radicals can oxidize proteins to form protein carbonyl species, which are observed in increased levels in several brain regions in AD patients. Build up of these oxidized proteins results in the loss of their activity leading to the destabilization of several systems or equilibriums inside neurons.¹⁹⁶ Free radicals can also oxidize DNA and RNA and their oxidation products were found to be elevated in vulnerable neurons in the brains of AD patients. Oxidation would impact on ribosomal functioning and reduce protein synthesis.¹⁹⁹ In addition to iron homeostasis disturbance, modifications in the concentration of zinc and copper are also important in AD pathology.²⁰⁰ A β binds copper ions with high affinity²⁰¹ due to three histidine residues and a tyrosine,²⁰² after some redox changes, Cu $^{2+}$ is able to reduce oxygen to generate H $_2$ O $_2$.²⁰³

By considering this evidence, lipocrine (**26**, Fig. 7)¹⁸⁷ was designed to have the capability to reduce free radical concentrations and their toxic effects. Rational design was based on the structure of lipoic acid, a potent antioxidant with multiple neuroprotective effects,²⁰⁴ and tacrine. Several series of derivatives yield lipocrine as one of the most potent AChEIs (IC $_{50}$ = 0.25 nM).¹⁸⁷ Owing to the linked lipoic acid moiety, a linear extension of the molecule was argued that lipocrine can interact with the PAS of AChE with the ability to decrease AChE-induced A β aggregation. Lipocrine reduced A β aggregation with an IC $_{50}$ of 45 μ M. Considering its antioxidant effect, lipocrine decreased by 51% the production of ROS species at 10 μ M. Moreover, compound **26** demonstrated a good profile as a neuroprotectant against oxidative stress to a larger extent than its parent compound lipoic acid.⁴⁶

Compound **27**²⁰⁵ (Fig. 7) was the lead compound of a series of hybrids of tacrine and feluric acid, a potent phenolic natural antioxidant, which concomitantly appears as a secondary metabolite with antioxidant properties.^{206,207} It has been proven to reduce the toxicity in the brain of rats exposed to A β_{1-42} .²⁰⁸ Compound **27** was able to inhibit AChE with high potency (IC $_{50}$ = 4.4 nM), 10-fold more effective than tacrine. Interestingly, this family of derivatives kept antioxidant properties of the parent compound. In experiments performed by the oxygen radical absorbance capacity (ORAC) method,²⁰⁹ compound **27** showed a value of 1.5 trolox equiv. in its ability to decrease ROS species.

Compound **28** (Fig. 7) was developed by Rodríguez-Franco et al. in 2006,²¹⁰ as multi-targeted drug for AD. The design of **28** was based on the structure of tacrine and melatonin,

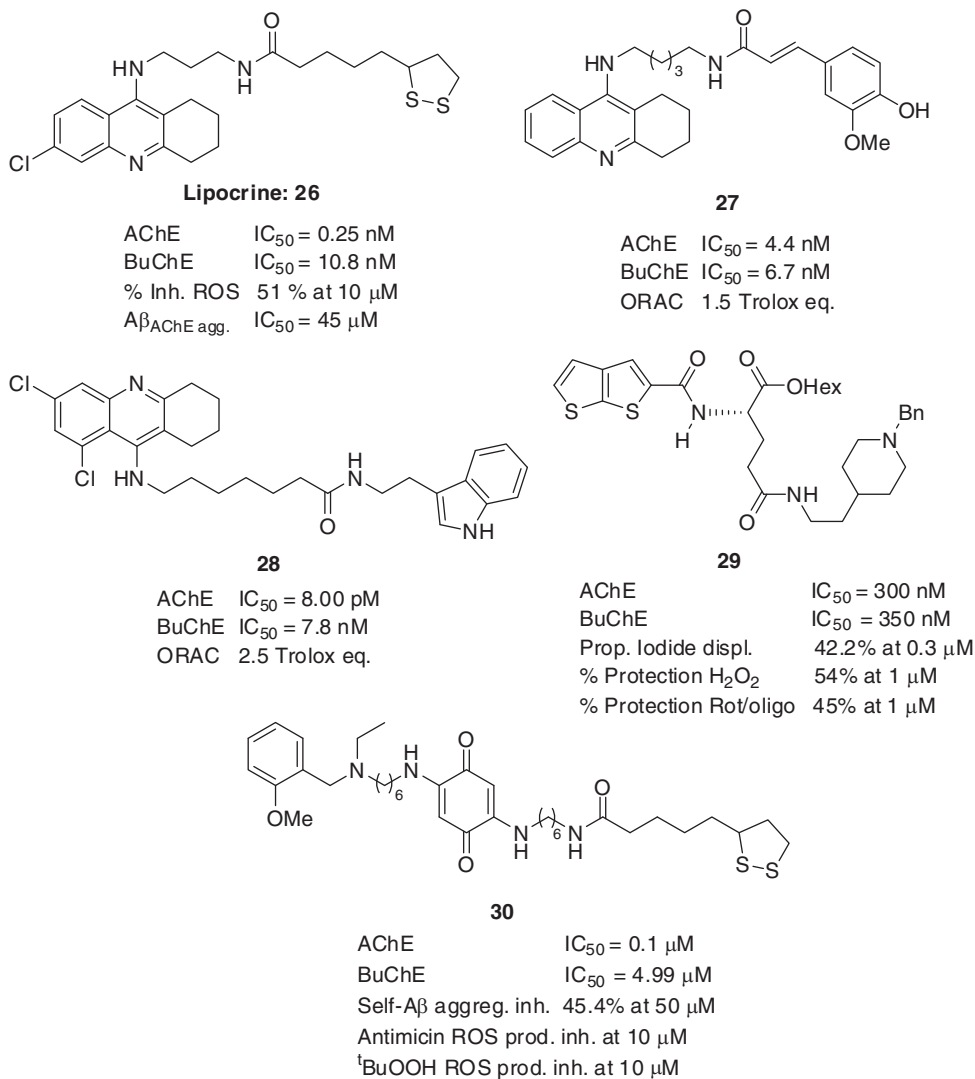


Figure 7. Chemical structure of dual AChE inhibitors and antioxidants.

a neurohormone that possesses strong antioxidant properties, and also acts as scavenger for some ROS species.²¹¹ Melatonin has also proved to exert neuroprotective effect against Aβ toxic insult in microglial cells.²¹² Compound **28** was a high potent inhibitor of human AChE (IC₅₀ = 8 pM). Interesting selectivity between AChE and BuChE was observed, compound **28** was a 1,000-fold more active toward AChE than toward BuChE, and 40,000 times more effective AChEI than tacrine. Interestingly, compound **28** was 2.5 times more potent than trolox in its antioxidant properties. Some member of this chemical family had even more potency, up to 4-fold higher. To complete its exciting profile, compound **28** was found to cross the BBB with a permeability of 7.610⁻⁶ cm s⁻¹.²¹³ Recently, derivatives of this family were evaluated as potential Aβ₁₋₄₀ self-aggregation inhibitors.²¹³ An analog compound of **28** reduced Aβ self-aggregation up to 63% at 100 μM. Members of this family were neuroprotectant agents against Ca²⁺ overload and oxidative stress models, showing a moderate neuroprotective profile. Further developments of this class of compounds have been recently

achieved with a tacrine-melatonin hybrid *N*-(2-(1*H*-indol-3-yl)ethyl-7-(1,2,3,4-tetrahydroacridin-9-ylamino)heptanamide.²¹⁴ This molecule successfully decreased the A β deposits in organotypic brain slices of APP/Ps1 mouse.²¹⁵ This compound induced a significant decrease in brain A β levels; moreover, it reduced the A β -cytotoxicity in primary neuronal cultures as well as in the APP/Ps1 mouse model. Evidence indicates that the observed neuroprotective activity may involve caspase-3 and -9 expression modifications. Finally, this derivative alleviated behavioral deficits in an APP/Ps1 mice model, with cognition deficits; the compound exerted an effective action only after 6 weeks of treatment.

In 2009, a new family of derivatives inspired in compound **29** was investigated.²¹⁶ Design was based in dicarboxylic amino acids as a biocompatible scaffold, allowing the attachment of different bioactive moieties. The authors reported L-glutamic acid as a linker for three groups: (a) A ω -situated *N*-benzylpiperidine moiety able to bind to the catalytic active site of AChE, based on the structure of donepezil (**2**, Fig. 2). (b) An *N*-protecting group able to interact with the PAS of AChE proposed to inhibit A β -aggregation. Finally, (c) a lipophilic α -hexyl ester that could favor the crossing of the BBB.²¹⁷ Overall properties displayed by exemplar compound **29** fits the hypothesis and therefore, it was selected as the lead compound of the family. Compound **29** showed high inhibitory activity of both cholinesterases in the nanomolar range. Furthermore, it decreased in a significant manner (42.2% at 0.3 μ M) the fluorescence elicited by propidium, demonstrating the interaction at the PAS of AChE. This result is an indirect test of the potential A β anti-aggregating effect of these molecules. Neuroprotective potential was assessed against two different oxidative stress models. Generation of ROS from H₂O₂, and incubation with a combination of rotenone/ oligomycin-A, which blocks complexes I and V of the electron transport chain in the mitochondrion, leading to the generation of ROS. Compound **29** showed good properties as a neuroprotectant agent rescuing 54.5% of cell death originated by 60 μ M H₂O₂ and 45.4% of cell death induced by rotenone/oligomycin-A cocktail. Finally, in vitro permeability experiments demonstrated the potential of **29** to cross the BBB.

Based on the structure of memoquin¹⁹¹ (**25**, Fig. 6) and lipocrine¹⁸⁷ (**26**, Fig. 7), compound **30**⁶⁸ (Fig. 7) was designed trying to summarize the excellent multitarget profile of memoquin, and the inherent antioxidant activity of lipoic acid.¹⁸⁸ Bolognesi et al.⁶⁸ reported **30**, a new hybrid with good AChE inhibitory activity (IC₅₀ = 0.1 μ M) and selectivity (50-fold more potent against AChE). Furthermore, **30** successfully reduced A β self-aggregation up to 45.4% at 10 μ M. Insertion of the benzoquinone (from CoQ10) and lipoyl moieties (from lipoic acid) into **30**, direct its antioxidant action to mitochondria. Thus, the ability of compound **30** to reduce the production of ROS was tested. Compound **30** reduced almost to 50% ROS production at 10 μ M in the oxidized form. As observed with memoquin, antioxidant character of **30** enhance when quinone core is reduced by NQO1¹⁸⁵ (confirmed by experiments in cells treated with sulforaphane⁶⁸). Reduced form of compound **30** exerted an increased antioxidant activity, decreasing ROS production to basal levels.

Considering oxidative damage as a key process in AD pathogenesis,¹⁹³ several targets have been defined with potential interest for further therapeutic developments. Monoamino oxidase (MAO) enzymatic family releases ROS during its catalytic deamination of neurotransmitters (noradrenaline, dopamine, and serotonin).²¹⁸ Inhibition of MAO will reduce ROS production, decreasing oxidative stress in AD patients. Several compounds have been designed and synthesized to include AChE and MAO inhibition in one molecule inserting propargylamine moieties.²¹⁹ Several families with dual activity over AChE and MAO have been reported; key compounds have been reported in recent reviews.⁴⁶ A prominent example of this type of compounds is ladostigil,²²⁰ a novel multifunctional AChEI/MAO-AB inhibitor for the treatment of AD that currently is in phase II clinical trial.²²¹

6. AChE INHIBITORS AND CALCIUM CHANNEL BLOCKERS

As stated before, Ca^{2+} overload is a major pathway initiating the processes leading to cell death.²²² Much evidence has shown that Ca^{2+} dysfunction is involved in the pathogenesis of AD,²²³ increasing A β formation,²²⁴ and τ hyperphosphorylation.²²⁵ In pathological conditions, Ca^{2+} entry through L-channels causes calcium overload and mitochondrial disruption, leading to the activation of the apoptotic cascade and cell death.²²⁶ Modulation of Ca^{2+} entry through this specific Ca^{2+} channel subtype could be an effective strategy to prevent cell death.

Compounds **31**,²²⁷ **32**,²²⁸ and **33**^{229,230} (Fig. 8) were developed in an extensive program targeting Ca^{2+} dyshomeostasis produced in the pathological progression of AD.^{227–240} Families were designed as hybrids of tacrine and 1,4-dihydropyridines, which are voltage-dependent Ca^{2+} channel (VDCC) modulators. 1,4-dihydropyridines demonstrated both antagonist (nimodipine)²⁴¹ or agonist (Bay K 8644)²⁴² activities due to their affinity for L channels. Combining both structures in one molecule, the new compounds were expected to have both activities. Derivative **31**, bearing the 4*H*-pyrano[2,3-*b*]quinoline core was reported as part of a new family of AChEIs with Ca^{2+} antagonism activities.²²⁷ Compound **31** selectively inhibited AChE at the micromolar range (IC_{50} = 1.86 μM) with almost no activity over BuChE. More importantly, it blocked up to 43.4% the increase in cytosolic Ca^{2+} evoked by K^+ in SH-SY5Y neuroblastoma cells, thus providing evidence for the dual activity exerted by this hybrid. Furthermore, compound **31** showed neuroprotective effect in two AD in vitro models, Ca^{2+} overload induced by 70 mM K^+ , and oxidative stress induced by 60 μM H_2O_2 . Compound **31** rescued 42% of the cell death induced by K^+ and 52% of the toxicity induced in the oxidative stress model. Nevertheless, no evidence of the neuroprotection mechanism was given, and there was no correlation between neuroprotection effect and VDCC blockade. This lack of correlation implies a different effect by this family of compounds.

Neuroprotective effect may be explained by different studies performed with compound **32**, named as ITH4012.²²⁸ ITH4012 was reported in 2004 as the best AChE inhibitor of the 1,8-naphthiridine family, with an IC_{50} of 820 nM.²³⁶ This compound blocked the Ca^{2+} influx induced by 70 mM K^+ up to 20% at 3 μM . Even more interesting was its Ca^{2+} promoting activity, demonstrated in the chromaffin cell model. ITH4012 increased the cytosolic basal Ca^{2+} concentration of the cells from 47 to 250 nM at 10 nM. ITH4012 was found to have neuroprotective properties against thapsigargin (calcium overload model), H_2O_2 -induced

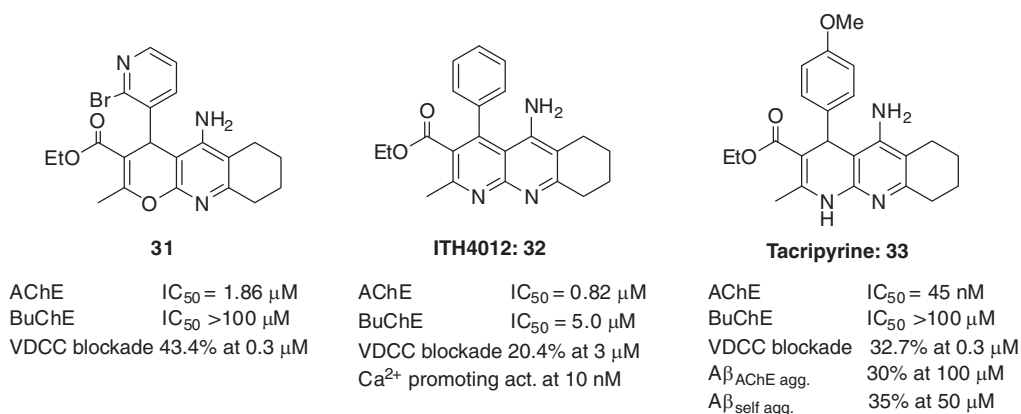


Figure 8. Multipotent AChE inhibitors with antagonist voltage-dependent Ca^{2+} channel activity.

oxidative stress, A β -induced toxicity, and veratridine.²³⁶ Further studies showed that protein synthesis inhibition by cycloheximide reverted ITH4012 cytoprotective effect. Investigations revealed overexpression of Bcl-2 (antiapoptotic protein), which possibly mediates the ITH4012 antiapoptotic effect.²²⁸

Continuing this line of research, compound **33** (Fig. 8), named as tacripyrine, was developed as the third generation of AChEIs–VDCC antagonists. In this new family, the 1,4-dihydropyridine moiety was integrated into the tetrahydroacridine core of tacrine.²³⁰ Compound **33** was the most potent AChEI with an IC₅₀ of 45 nM (4-fold more potent than tacrine) and a highly selective, 2,000-fold more active against AChE compared with BuChE. Kinetic studies and molecular modeling data proposed that this compound may interact with the PAS of the AChE. Thus, compound **33** was tested in both AChE-induced and A β -self aggregation models, and was found to be moderately active in both models (30 and 35% inhibition respectively). Compound **33** also exhibits Ca²⁺ blocking activity, showing a 32% blockade of the Ca²⁺ signal in the same model. As previous generations, tacripyrine **33** included neuroprotective activity decreasing mortality induced by Ca²⁺ overload and oxidative stress. Finally, tacripyrine **33** successfully penetrated the BBB. These multiple activities and properties have made this family one of the most interesting candidates for further development in the search of future treatment for AD.

7. AChE AND OTHER SYSTEM MODIFYING TARGETS

The links between cognitive impairment, neuronal death, A β production–aggregation, oxidative stress, metal accumulation, and Ca²⁺ dyshomeostasis are widely accepted.⁹ All the above are mostly related with the cholinergic system. Nevertheless, in the last decade advances in the understanding of the molecular basis of the disease have led to the description of new important targets related with several stages of AD.¹⁰² As an example, mitochondrial abnormalities are known markers for AD. Brains of AD patients show decreased activity of important enzymes such as cytochrome oxidase (COX, enzyme responsible for reducing molecular oxygen), pyruvate dehydrogenase complex, and the α -ketoglutarate dehydrogenase complex.¹⁰² However, similar mitochondria abnormalities are found in neurons that lack pathological neurofibrils, indicating that these abnormalities occur at very early stages of the disease.²⁴³ Compromised mitochondria activity may induce Ca²⁺ dyshomeostasis as mitochondria can buffer large quantities of this key ion. Mitochondria are involved in the tight control of intracellular Ca²⁺ homeostasis. Thus, mitochondrial damage leads to high basal cytosolic Ca²⁺ concentrations. In these pathological conditions, any stimulation can lead to Ca²⁺ overload toxicity for example, activation of NMDARs by glutamate cause greater neurotoxicity when mitochondria are damaged.²⁴³ This effect has been proved in rats cholinergic neurons after compromising mitochondrial activity.¹⁰² In fact, memantine (**5**, Fig. 2) (the only non AChEI treatment for AD) is an NMDAR antagonist, which is able to block the toxicity elicited by soluble A β oligomers. This observation suggests a direct link between soluble A β oligomers and NMDAR at early stages of the disease.

With these observations in mind, Melchiorre et al.³² designed a new multitarget ligand based on carvedilol. Carvedilol²⁴⁴ is a vasodilating β -blocker and antioxidant approved for the treatment of mild-to-moderate hypertension, which shows low-affinity NMDAR antagonism.²⁴⁴ In this project, the tetrahydroacridine moiety from tacrine and the carbazole core from carvedilol were linked. The resulting dimeric molecules were expected to inhibit A β fibril formation, since carbazoles are efficient inhibitors of A β aggregation.²⁴⁵ Carbacrine (**34**, Fig. 9),³² had effective anticholinesterase activity at the nanomolar range (IC₅₀ = 2.15 nM),

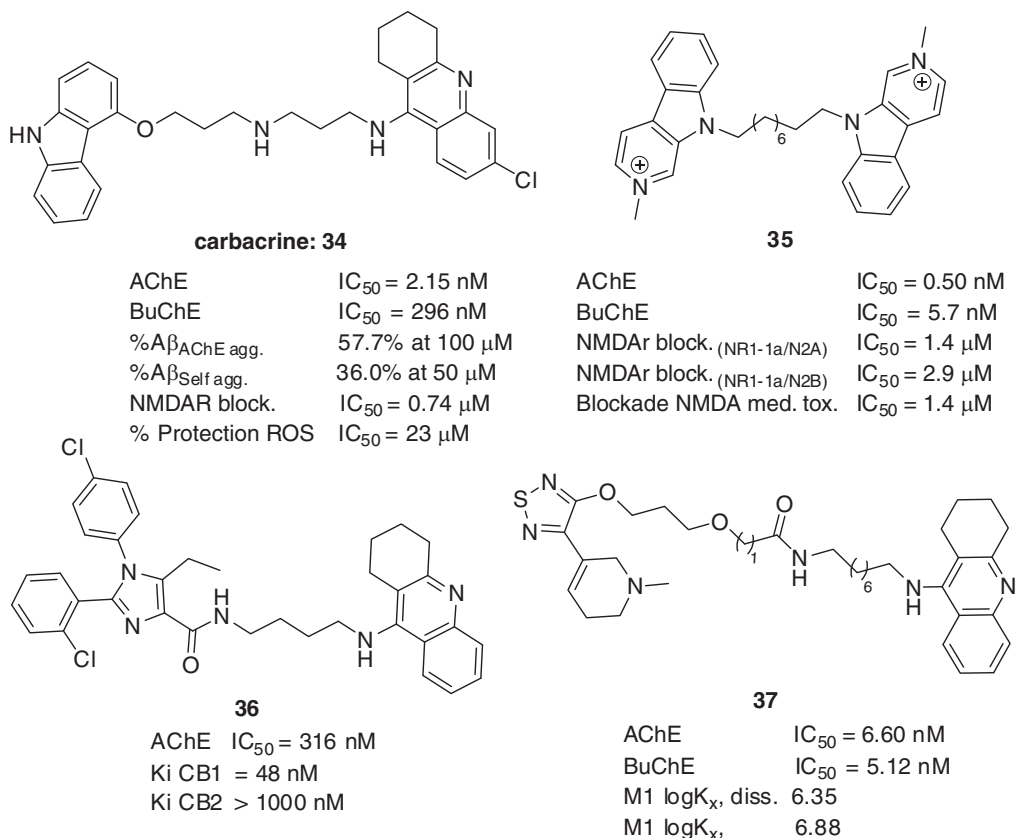


Figure 9. Chemical structure of AChE inhibitors including activity over NMDA, cannabinoid, or muscarinic receptors as potential targets for AD treatment.

more potent than tacrine and with good selectivity against AChE. Docking of carbacrine predicted interactions in both catalytic and PAS sites of AChE. Potential antiaggregating effect was therefore investigated. A good profile was observed, inhibiting both AChE and self-induced Aβ aggregation. As this family was designed as NMDAR modulators, their potential activity was tested. Interestingly, **34** NMDAR antagonistic effect (IC₅₀ = 0.74 μM) was more potent than the reference compound carvedilol. Further experiments with carbacrine suggested that it can be a noncompetitive open-channel blocker. This ability may imply a well-tolerated antagonism that preferentially blocks excessive NMDAR activity without affecting normal neuronal function. Finally, and as an effect of all the intrinsic activity included in this molecule, neuroprotective potential was investigated. This potential effect was tested in ROS-generating models where carbacrine showed neuroprotective effects against ROS formation in a significant manner (IC₅₀ = 23 μM). In the ORAC text, compound **34** was a good antioxidant (IC₅₀ = 0.07 μM), more potent than trolox.

Based on the structure of monovalent β-carbolines (potent AChEI),^{246,247} a new family for bivalent β-carbolines (pyrido[3,4-*b*]indoles) has recently been developed. Rook et al.²⁴⁸ found that several bivalent β-carboline compounds were also potent NMDAR blockers, providing a possible multitarget approach to the treatment of AD. Compound **35** was the most potent anticholinesterasic agent of the series with IC₅₀ to inhibit AChE as low as 0.5 nM. Also, compound **35** proved to be a potent inhibitor of the Ca²⁺ transient induced by glutamate, with an IC₅₀ of 1.4 μM, 4-fold more potent than memantine (IC₅₀ = 5.6 μM in the

same assay conditions).²⁴⁸ No experiments were performed in order to elucidate its possible ability to inhibit A β -aggregation and to determine its potential antioxidant properties. However, the presence of a quaternary pyridine moiety may imply these activities and further exploration will be interesting in this area.

Cannabinoid receptors (CBR) are also interesting targets for AD treatment.^{249,250} Cognitive disorders constitute a potential therapeutic area for cannabinoid CB₁ receptor antagonists.^{250,251} CB₁ receptor antagonists increased ACh release in certain brain areas including cortical regions and the hippocampus.²⁴⁹ Co-application of subthreshold doses of rimonabant (a CB₁ receptor antagonist) and donepezil (**2**, Fig. 2), in noneffective doses, induce memory enhancement.²⁵² Lange et al.²⁵³ have recently targeted this type of receptors including two activities, AChE inhibition and CB₁ receptor antagonism. Design was based on tacrine and two known CB₁ antagonists, 3,4-diarylpyrazoline moiety²⁵⁴ and 1,2-diarylimidazol.²⁵⁵ Thus, imidazol **36** (Fig. 9) was a potent inhibitor of AChE (IC₅₀ = 316 nM), while it was able to block CB₁ receptors with high affinity (K_i = 48.0 nM). Furthermore, a high CB₁/CB₂ receptor subtype selectivity (>20-fold) was observed. Molecular modeling with both (AChE and CB₁ receptor) pharmacophores explained both activities; the *p*-Cl-phenyl moiety is known to be important in the interaction with CB₁R, while the pyrazoline core is surprisingly well accommodated at the PAS of AChE. Furthermore, tacrine-like core is proposed to interacting at the bottom of the gorge, at the catalytic site of AChE.

In the search of new targets of potential interest for the pathogenesis of AD, muscarinic receptors have gained great importance in the development of AD drugs.²⁵⁶ Neurotransmission via M₁ receptors can be stimulated either directly by muscarinic agonists or indirectly by allosteric agents acting as ACh enhancers.²⁵⁶ M₁ stimulation has been targeted as a symptomatic treatment. In addition, M₁ stimulation can reduce A β ₄₂ and τ pathologies via activation of protein kinase C (PKC), which led to the production of soluble APP among other pathways.^{257,258} Heterodimer **37** (Fig. 9) was developed in a program launched by Fang et al.²⁵⁹ linking the tetrahydroacridine moiety from tacrine, and xanomeline, a functionally selective M₁ muscarinic agonist with promising in vivo antidemential properties.²⁶⁰ The lead compound **37** (AChE IC₅₀ = 6.6 nM) was significantly more potent than tacrine, and inhibited BuChE with a similar potency. Its M₁ muscarinic potential affinity was investigated using the affinity of the compound for unliganded receptors labeling them with the orthosteric radioligand [³H]-N-methylscopolamine. Compound **37** showed enhanced M₁ allosteric affinity (logK_x = 6.35) exceeding the binding affinity of xanomeline by a factor of 3. Structure–activity relationships suggest that compound **37** prefers a purely allosteric binding topography even in orthosterically free receptors, inhibiting binding of the endogenous neurotransmitter ACh. In vivo studies of scopolamine-induced memory impairments in rats showed a significant reduction in scopolamine impairment activity in the presence of **37**.

Another target widely used for drug development is the SERT,^{261,262} expected to bestow antidepressant efficacy, as a symptomatic treatment of the psychiatric behavior of AD patients. Balanced inhibition of AChE and SERT is considered to be a promising dual-target for the treatment of AD, due to a potential improvement in cognitive deficits. After years of development, Kogen et al.²⁶³ summarize results in this area. Compound (**R**)-**38** (Fig. 10) was characterized as a potent AChE inhibitor (IC₅₀ = 14 nM) and SERT antagonist (IC₅₀ = 6 nM). (**R**)-**38** was 155 times more potent antagonist of SERT than the correspondent (**S**)-**38** enantiomer. However, no further experiments with (**R**)-**38** were performed due to a poor penetration across the BBB. Related compounds were evaluated showing interesting properties, leading to compound BGC20-1259,²⁶³ the most promising inhibitor of AChE and SERT that has completed phase I clinical studies in healthy volunteers.

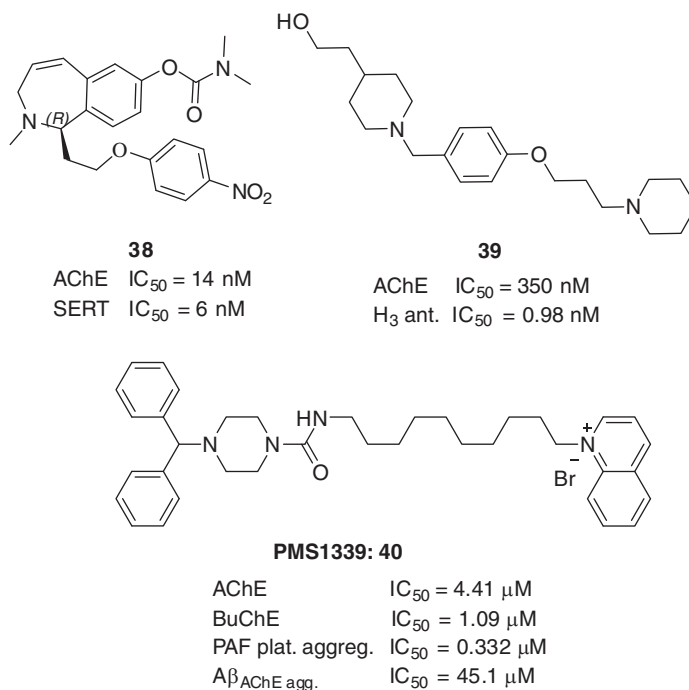


Figure 10. Chemical structure of AChE inhibitors including activity over SERT or histaminic or PAF receptors as potential target for AD treatment.

The H₃ receptor (H₃R) is an attractive G protein-coupled receptor drug target that regulates neurotransmission in the CNS and plays a role in cognitive and homeostatic functions.²⁶⁴ H₃R antagonists improve cholinergic neurotransmission in the cortex via different mechanisms. When an H₃R antagonist is present, ACh release is improved. A combination of AChE inhibition with H₃R antagonism may induce an increased memory enhancing effect. Combination of both activities in one molecule has been achieved by Bembenek et al.²⁶⁵ Based on virtual screening toward both (AChE and H₃) targets,²⁶⁶ led to the discovery of family type **39** (Fig. 10). Compound **39** was a potent anti-cholinesterase agent ($IC_{50} = 350$ nM) including an interesting H₃R antagonist profile with an IC_{50} of 0.98 nM. QM/MM calculations of the interaction mode of **39** with AChE suggest a dual-inhibition mode interacting at the PAS including a possible A β antiaggregating effect.

Converging lines of evidence suggests that platelet-activating factor (PAF), a potent pro-inflammatory mediator, is implicated in the inflammatory events,^{267,268} promoting neuronal death in demential disorders.²⁶⁹ Previous developments in dual PAF and AChE targets include series of 2,5-disubstituted tetrahydrofuran derivatives.²⁷⁰ Among them PMS777 was shown to inhibit AChE, reverse scopolamine-induced dementia in mouse models, prevent PAF-induced neurotoxicity,²⁷¹ and LPS-induced oxidative/inflammatory disturbances in human neuroblastoma cell lines.²⁷² Recently, Ezoulin et al. reported a new improved candidate, **40** (Fig. 10) (PMS1339),²⁷³ a piperazine derivative with improved AChE inhibitory activity ($IC_{50} = 4.41$ μ M) and inhibition of PAF-induced platelet aggregation ($IC_{50} = 332$ nM). Compound **40** reduced the AChE-induced A β -aggregation ($IC_{50} = 45.1$ μ M) by direct interaction at the PAS of AChE. Finally, compound **40** reversed the scopolamine-induced memory impairment, a proof of the benefits provided by the biological target included in this molecule.

8. NON-ACH-E-DIRECTED MULTITARGET DRUG DEVELOPMENTS

Non cholinergic-based drug discovery has been extensively investigated due to the implication of different mechanisms and pathways in AD pathology.⁹ Great efforts have been devoted to the investigation of the A β processing targets,⁶⁹ τ hyperphosphorylation and aggregation,⁵ and metal chemistry²⁷⁴ related to AD. APP function is not completely understood; the extracellular domain has been suggested to serve as neurotrophic factor, while the intracellular domain may have a role as gene transcription regulator.^{275,276} APP has been also linked to axonal transport inside neurons, helping the transport of different vesicles through kinesin-1.²⁷⁷ APP can be proteolytically processed by three aspartic acid proteases, named α -, β -, and γ -secretase.²⁷⁸ Subsequent cleavage by β - and γ -secretase generates a group of peptides which differ in length at their C terminus, A β ₄₀ being the dominant species and A β ₄₂ being the second major peptide. A β ₄₂ readily aggregates to form the seed for larger oligomers and fibrils, eventually generating macroscopic amyloid plaques.^{279,280} As referred before, this process can be catalyzed by the interaction of A β peptides at the PAS of AChE.¹⁰ Thus, BACE-1 inhibition and γ -secretase modulation are two areas of great interest for drug development in AD. Aspartic acid protease inhibitors have been discovered in the past and some of them have entered in early clinical trials, for example, CTS-21166.²⁸¹ On the other hand, it should be noted that the proportion of APP that is processed by α -, β - and γ -secretase depends on the equilibrium processes inside the cell. Activation of G-protein-coupled-type receptors such as M₁ ACh receptor or 5-HT₄ serotonin receptor can induce an increase in the α -secretase–APP processing pathway, thus reducing the production of A β peptides. In this sense, it is worth to note that two partial agonists of these receptors, SL-65.0155²⁸² (Sanofi-Aventis, Paris, France), and PRX-03140²⁸³ (EPIX Pharmaceuticals, Lexington, MA) have reached phase II clinical trials for the treatment of AD,^{284,285} although none of them have progressed to more advanced phases. Their mechanism may be mediated by modulation of PKC, since activators of PKC have similar effects. Surprisingly, shifting between APP processing pathways was observed with some NSAIDs such as ibuprofen and indometacin.²⁸⁶ This action is unrelated to their inhibition of COX-1 and -2 enzymes. The shifting effect is related to the peroxisome proliferator-activated receptor γ (PPAR γ) agonist effect, included in NSAIDs.^{287,288} Evidence indicates that PPAR γ agonists may have multiple beneficial effects in AD. These benefits can be both on core pathological processes in brain and on peripheral factors, such as serum glucose levels and insulin sensitivity.²⁸⁹

In 2010, a new molecule trying to integrate α -secretase modulation activity and PPAR γ activity was reported by Hieke et al.²⁹⁰ Its development was based on the 2-(bis-(phenoxy)pyrimidine-2-ylthio)-hexanoic acid that displayed γ -secretase modulation and effective PPAR γ modulation. SAR studies and lead optimization yielded compound **41** (Fig. 11). Optimized activity over γ -secretase (inhibition of A β ₄₂ production: 6 μ M; activation of A β ₃₈: 1.8 μ M) and improved PPAR γ activation (EC₅₀ of 11.0 μ M) was observed. Most NSAID type PPAR γ -modulators are inhibitors of COX enzymes at the same time, resulting in side effects in their long-term clinical use. In order to minimize potential side effects, a decrease in the activity over these enzymes was needed. Thus, compound **41** was a weaker inhibitor of both enzymes providing a promising strategy to address the increased dementia risk in AD patients.

As pointed out before, 5-HT₄ agonists are of special interest in AD drug development; their benefits are inherent to the pathways of this G-protein-coupled receptor. The structure of ML10302,²⁹¹ a partial agonist with a benzoate base structure, was found to selectively enhance soluble APP α release in the hippocampus and cortex of mouse. This was the starting point for the development of derivative **51** (Fig. 11).²⁹² This compound is a potent 5-HT₄ receptor agonist with an EC₅₀ of 9 nM and, as expected, in vivo studies showed an increased

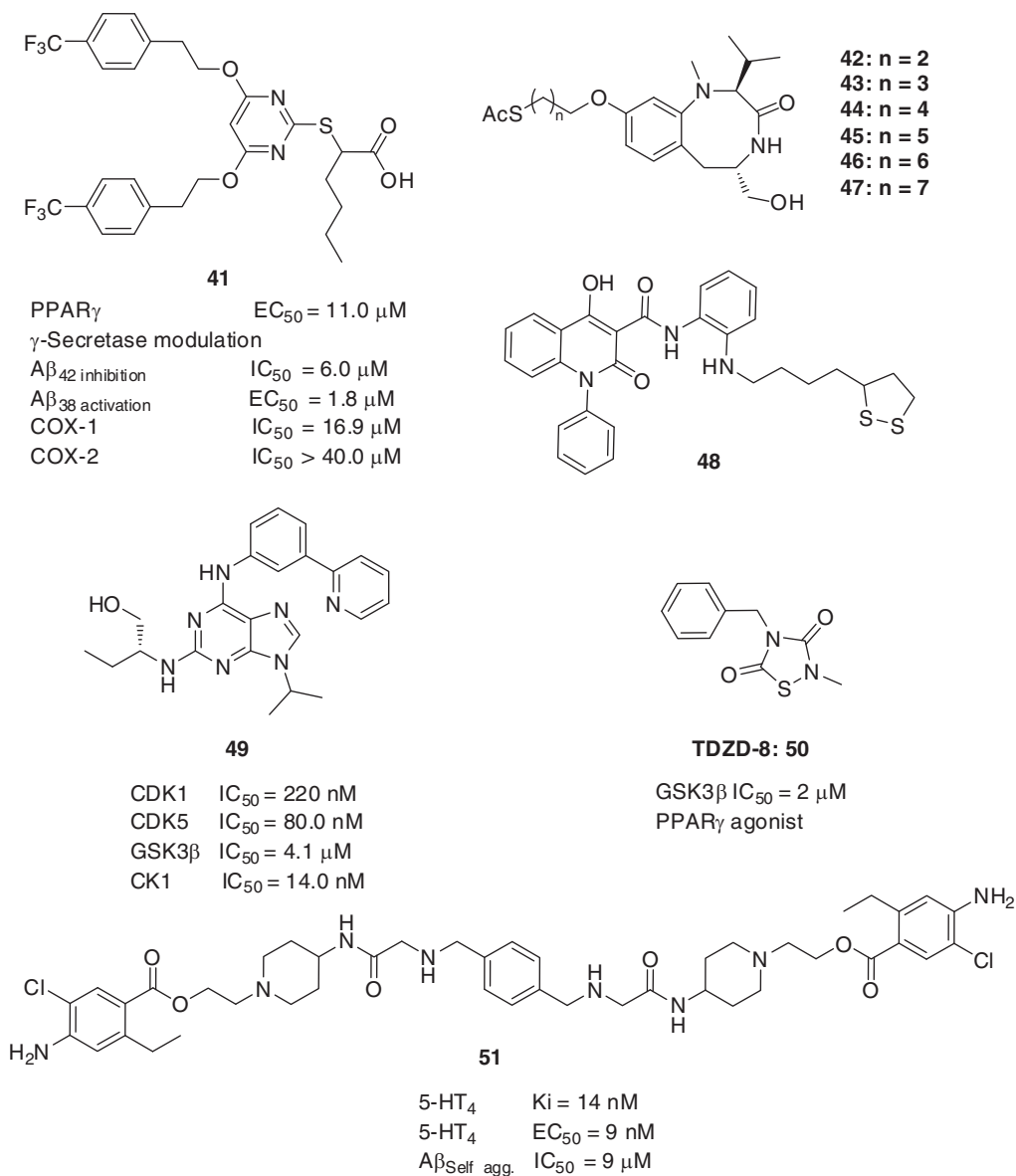


Figure 11. Chemical structure of noncholinergic multitarget-directed ligands developments.

concentration of soluble APP α in hippocampus and cortex in mouse. Moreover, **51** displayed an inhibitory effect on A β fibril formation ($IC_{50} = 9 \mu M$) being, to the best of our knowledge, the first designed molecule exhibiting these two activities together.

Similar effects have been described for PKC activators, due to their ability to modulate the α -secretase activity.²⁹³ This modulation increases the nonamyloidogenic pathway to produce soluble APP α . Another emerging approach which could affect multiple pathways to ameliorate AD pathophysiology and cognitive impairment is epigenetic remodeling through the inhibition of histone deacetylase (HDAC).²⁹⁴ In 2009, Kozikowski et al.²⁹⁵ included both targets in one molecule generating a small chemical library of benzolactam analogs. This scaffold has previously been found to have high-binding affinity for PKC.²⁹⁶ Among all the

benzolactam derivatives generated, compound **43** (Fig. 11) induced a statistically significant elevation of soluble APP α at a concentration as low as 1 nM. This compound, however, was not the best ligand for PKC (K_i = 15.8 nM) with a medium potency among the library. Compound **47** (Fig. 11) was the most potent with a K_i of 2.8 nM. Compounds **42–47** (Fig. 11) induced an increase in histone H4 acetylation, confirming HDAC inhibition activity at 10 μ M. These compounds were found to activate PKC and inhibit HDAC-inducing histone H4 acetylation. HDAC inhibition blocks the deacetylation process, increasing the global acetylation level of histones. Consequently, profound changes in gene expression are induced activating survival pathways. Compounds **42–47** protected cortical neurons upon exposure to homocysteate (oxidative stress model) with high potency.²⁹⁵

As discussed previously, oxidative stress is one of the most important pathological pathways of AD.¹⁹⁶ Oxidative stress has been widely investigated leading to the discovery of antioxidant molecules or MTDLs with antioxidant properties (discussed above).⁷³ In this line, a new antioxidant molecule was designed to include anti-inflammatory activity. Inflammation is a well-known process related to AD pathology.⁷³ In the past, several NSAIDs have been investigated as potential drugs targeting glia inflammation in AD.⁷³ In this line of research, several unsuccessful clinical trials have been reported;⁹⁸ nevertheless, inflammation continues to be a key target in AD drug development. The quinolinone structure includes motifs that are characteristic of numerous natural products and synthetic analogs, exhibiting a wide variety of biological activities.^{297–300} Linomide (*N*-phenyl-methylpz-1,2-dihydro-4-hydroxy-1-methyl-2-oxo-3-quinoline carboxamide) is a synthetic immunomodulator used as a starting point for lead optimization of several analogs.^{297,301–306} Rebamipide,³⁰⁷ another example of quinolinone derivative, has effective antioxidant effect due to its ROS scavenging ability and over-expression induction of endogenous prostaglandin. Scavenger properties are due to the presence of the 3,4 double bond together with the 2-oxo functionality in the quinoline moiety.³⁰⁷ On the basis of these observations, Detsi et al.³⁰⁸ used this quinoline structure to generate hybrid molecules. This class of compounds included α -lipoic acid and the quinoline core. α -Lipoic acid is a very well-known antioxidant with anti-inflammatory activity.¹⁸⁸ Compound **48** (Fig. 11) as lead compound in this series showed good anti-inflammatory properties; thus, it reduced the carragenin-induced inflammation by 65% at low dosages in rat paw. It also proved to be a good antioxidant.

Formation of τ -protein aggregates has been mentioned throughout the text giving some hints about the importance of these aberrant structures in AD pathology. Much attention has been devoted to the destabilization of the neuron cytoskeleton; however, no drug has yet reached the clinic to directly interrupt this pathological process. The cytoskeleton consists of microtubules, polymers of tubulin. Microtubules bind microtubule-associated proteins, which regulate their stability. The most important stabilizing protein in the axon is τ -protein, a very hydrophilic protein which contains at least 25 potential phosphorylation sites. τ -Protein is phosphorylated by kinases CDK5, GSK3 β , ERK2, and CK1 (casein kinase 1). Phosphorylation can detach τ from microtubules, inducing neuronal damage in AD.³⁰⁹ Phosphorylation of τ under normal conditions induces equilibria between bound and unbound protein. This dynamic equilibrium helps the axonal transport of vacuoles and integrity of the cell. Thus, hyperphosphorylation of τ , as a result of an imbalance in the kinase and phosphatase activities, generates aberrant hyperphosphorylated τ . These aberrant structures induce toxicity and axonal transport dyshomeostasis previously mentioned. In AD brains, total expression of τ is around 8-fold higher than in controls, and it is abnormally hyperphosphorylated.³¹⁰ Implication of τ protein in AD pathology is supported by the existence of tauopathies such as amyotrophic lateral sclerosis, Pick's disease, progressive supranuclear palsy, and frontotemporal dementia with Parkinsonism.³¹¹ Drug development based on the τ hypothesis has been focused on the screening of protein kinase inhibitors.

GSK3 β is the most widely investigated, believed to be the most important τ -phosphorylating agent.³¹² Early developments on kinase inhibitors pursued high selectivity against only one of the kinases. These inhibitors were used to study the, not yet very well understood, phosphorylation–dephosphorylation equilibria of τ -protein.³¹³ Recently, kinase inhibitors developments pursue mainly multitarget kinase inhibitors. These new inhibitors target several protein kinases simultaneously, with the right balance of inhibition of different kinases, to stop the pathological phosphorylation of τ -protein. Oumata et al. reported the synthesis and biological evaluation of dual-specificity inhibitors of CDKs and CK-1³¹⁴ based on roscovitine³¹⁵ (CYC202 or seliciclib), a purine-based inhibitor.³¹⁶ Roscovitine has finished phase IIb of clinical trials against nonsmall-cell lung cancer,³¹⁷ and starting phase I against advanced solid tumors,³¹⁸ and several other diseases. Frequently considered as highly selective for CDKs, roscovitine has been shown to interact with several other kinases (DYRK1A, CK1, pyridoxal kinase).³¹⁹ Purines are a large family of biologically active molecules, constituting a scaffold of a wide variety of promising drugs, including kinase inhibitors as 2,6,9-trisubstituted purines.³²⁰ Tri-substituted purine **49** (Fig. 11) was one of the most potent inhibitors of CK1 (IC_{50} = 14 nM).³²¹ Compound **49** inhibited also with high potency CDK5 (IC_{50} = 80 nM), GSK3 β (IC_{50} = 4.1 μ M), and CDK1 (IC_{50} = 0.22 μ M).³²⁰ On the other hand, proliferation assays were performed in order to elucidate its possible toxic effect; in general, biarylamine type compounds were highly potent CDK/CK1 inhibitors but displayed low antiproliferative effect. As a general rule, CK1 inhibition inversely correlated with antiproliferative activity; thus, compound **49** can be described as a protective agent. Some evidence suggests a regulatory effect of CK1 on the production of A β as CK1 inhibitors were able to inhibit the production of A β .³²² Compound **49** was the most effective agent reducing the production of A β in the N2A-APP₆₉₅ cellular model.

Noscira (Madrid, Spain), a pharmaceutical company, has developed an interesting family of thiadiazolidindinones exemplified by compound **50**³²³ (Fig. 11). Noscira has finished a safety study with NP031112 for AD treatment.³²⁴ NP031112 is a potent GSK3 β inhibitor developed as selective inhibitor. More importantly, it is one of the first non-ATP competitive inhibitor molecule of this important kinase. Compound **50** was developed as a selective kinase inhibitor; however, NP031112 was found to be a nuclear receptor PPAR γ agonist,³²⁵ showing effective anti-inflammatory and neuroprotective properties. Both properties could constitute a significant advance in further developments of multitarget GSK3 β inhibitors.

9. MULTITARGET BIOAVAILABLE METAL CHELATORS

As mentioned above, oxidative stress is considered to play a central role in AD pathogenesis.¹⁹³ Recently, different drug development programs have pursued different strategies in order to combat increased levels of ROS species in AD patients. Strategies included interference in ROS production,²⁴³ development of antioxidants,¹⁰⁷ metal chelators,²⁷⁴ and others. Multitarget drug development programs have implemented some of these targets in one molecule. Cholinergic neurotransmission, A β antiaggregating effect and/or clearance facilitation, and neuroprotection activity against ROS species have been included. Several studies have indicated that cerebral biometal (Fe, Cu, and Zn) dyshomeostasis and oxidative stress are intimately associated with the formation of A β plaques and NFTs. For instance, iron increases production and translation of APP via activation of APP mRNA iron-responsive element. Consequently, it induces A β plaque formation.³²⁶ Dyshomeostasis of biometals and their interactions with A β cause A β aggregation and deposition.^{274,327} Metal chelators have the ability to attenuate a broad spectrum of oxidative stress as well as APP translation, A β production/aggregation, and NFTs formation. Metal chelators as desferrioxamine[®]³²⁸ and

clioquinol³²⁹ were under investigation as potential drugs for AD. However, they are not brain specific chelators, and possibly will display undesirable interactions with beneficial biometals, affecting the normal physiological function of essential metal-requiring metalloenzymes. With the aim to overcome these problems, some chelators or pro-chelators with improved target specificity have been developed. Franz et al. reported in 2006 one of the first molecules based on the prodrug approach.³³⁰ Compound **52** (Fig. 12) is a pro-chelator that has almost no affinity for metal ions until the boronic pinacol ester mask is removed by ROS species. In the absence of oxidative stress, the masked molecule is a poor ligand and thus cannot alter healthy metal ion distribution. When ROS are elevated in pathogenic conditions, compound **52** is unmasked and is revealed as a potent iron chelator, inhibiting the iron ability to generate OH^\bullet species. It includes a boronic ester as a latent phenolic oxygen, a key donor of the aroylhydrazone class chelators.³³¹ As expected, experiments showed the ability of compound **53** to inhibit the formation of ROS via effective iron chelation. Although **52–53** pair is not considered as a multitarget drug, we felt it was interesting to mention in this review. It was the first example of a pro-chelating drug that open a new line of research to new target-directed drugs aiming to diminish nondiscriminating biometal chelation.

On the basis of the same approach, Schugar et al. developed the **54–55** pair.³³² It is also a prodrug; however in this case, the metal chelator moiety is based on a bidentate

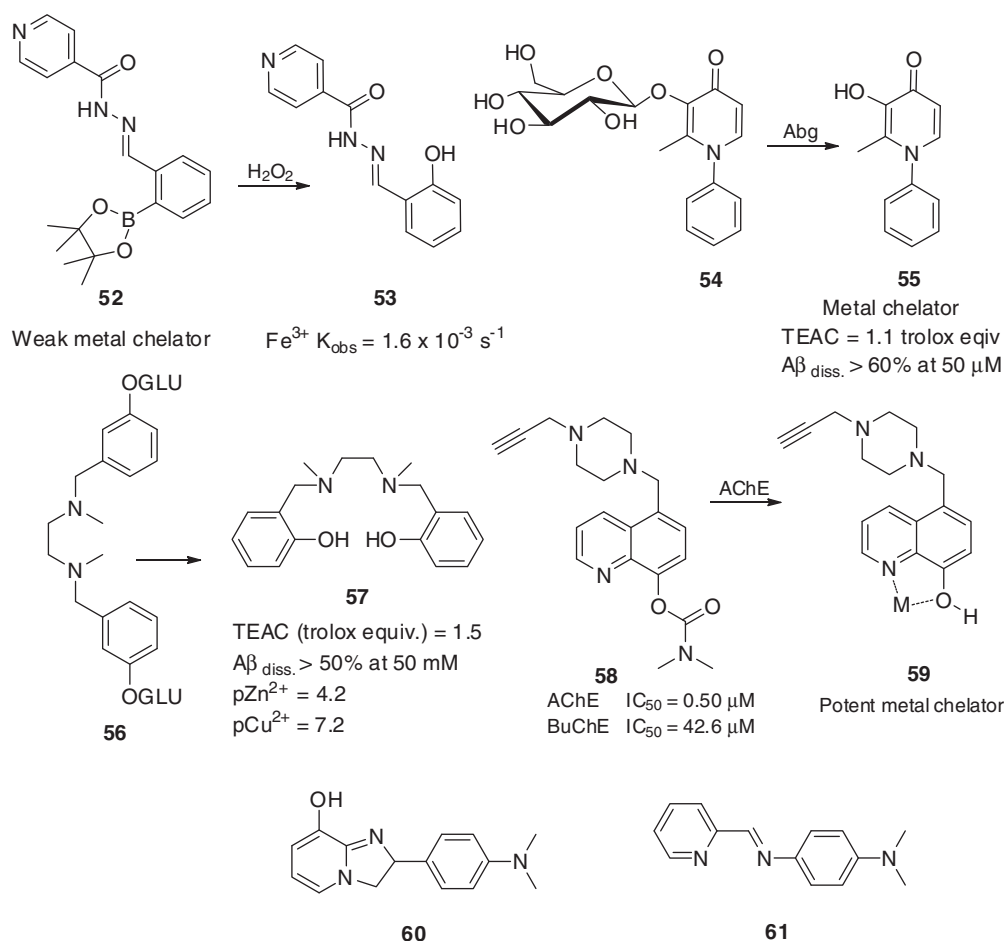


Figure 12. Metal chelator-based ligands with multiple activities included.

hydroxypyridinone. The presence of this moiety addresses both the metal ion and the oxidative imbalances linked to a glucose receptor targeting moiety. The carbohydrate moiety is expected to interact with the glucose receptor in the BBB. This receptor is present in the BBB to facilitate the entry of glucose across it. The trifunctionalized compound **54** successfully penetrated the BBB demonstrated by brain uptake of a radiolabeled hydroxypyridinone glucoconjugate. The presence of the carbohydrate moiety serves at the same time to inhibit systemic metal binding, thereby reducing side effects. Within the brain, compound **54** is hydrolyzed by glycoside hydrolysis. The authors used β -glucosidase (Abg), which successfully cleaves glucose from hydroxypyridinone. Hydroxypyridinones are known binders of di- and trivalent metal ions like Cu^{2+} and Fe^{3+} .^{333,334} It is also known that free alcohols are scavengers of free radicals; thus, compound **55** was assayed in the TEAC test, proven to be a more potent antioxidant than vitamin E, known antioxidant, equivalent to trolox. The potential of compound **55** to dissolve A β plaque was assayed by turbidimetry; it reduced the preaggregated A β_{1-40} by 60% at 50 μM . This trifunctional ligand combines metal sequestering and antioxidant properties with glucose conjugation, helping BBB crossing and minimizing systemic complexation of metal ions. The same hypothesis was used in the development of the pair **56–57** which demonstrated improved chelating properties toward Zn^{2+} and Cu^{2+} .³³⁵ The **56–57** pair reduced the preaggregated A β by 50% at 50 μM . Furthermore, **57** proved to be a good antioxidant being, 1.5-fold more potent than trolox in the TEAC assay.

A further advance in the pro-chelator approach was made by Zheng et al. including several new ideas in its design.³³⁶ As described before, by using masked chelators, the systemic chelation of healthy metal homeostasis is avoided. Masking the chelator as an AChE inhibitor included a new activity in the molecule. The pair pro-chelator **58**–chelator **59** was based on the structure of the bifunctional chelator 5-(4-propargylpiperazin-1-yl-methyl)-8-hydroxyquinol (HLA20)³³⁷ and the AChEI rivastigmine (**4**, Fig. 2), and donepezil (**2**, Fig. 2). Compound **58** included moieties from these three structures, where the carbamoyl moiety of rivastigmine also acts as a protective group of the quinolinol OH. Its biological evaluation revealed the AChE inhibition properties. Thus, compound **58** was a time-dependent AChEI ($\text{IC}_{50} = 500 \text{ nM}$) slightly more potent than rivastigmine and 85-fold more active against AChE than BuChE. After the anticholinesterasic properties were described, the potential chelating effect was investigated. Although compound **58** was unable to bind Cu^{2+} or Zn^{2+} ions, AChE successfully cleaves its carbamyl moiety, releasing the unprotected form **59**. Complexation experiments of Fe^{3+} and Cu^{2+} with chelator **59**, using absorbance spectroscopy, showed the formation of the different complexes. Complexation was confirmed by mass spectrometry studies. Among the benefits of the masked pro-chelators, activity against AChE can induce an antiaggregating effect. This together with the potential antioxidative properties makes these derivatives highly interesting for further preclinical³³⁸ and clinical development.

A slightly different approach was reported by Rodríguez-Rodríguez et al.³³⁹ using the structure of thioflavin-T as substructure for recognition/interaction with A β aggregates. In this proposal, instead of masking the metal chelator as performed in previous developments, they used molecular recognition molecules targeting A β aggregates. Based on thioflavin-T which exerts specific affinity for amyloid plaques, and clioquinol,³²⁹ known metal chelator, the idea was to insert metal chelator properties into these models without important modifications of the core structure. In an extensive and exhaustive study, the authors described three new hybrids with highly encouraging capabilities as intercalation properties in amyloid fibrils and their potential use for radioisotopic detection of A β_{1-42} deposits in the human brain.³³⁹

A similar approach has been described by Hindo et al. in 2009.³⁴⁰ Structures of IMPY³⁴¹ and *p*-¹²⁵I-stilbene (two A β aggregate-imaging probes),³⁴² were used to provide the core structures. Derivatives **60** (Fig. 12) and **61** are the result of this development and they were

able to bind copper forming different stoichiometry complexes, 1:2 and 1:1 for **60** and 1:1 for **61**. NMR studies indicated the ability of both metal chelators to directly interact with A β -aggregates. Interaction is probably in positions close to where metal ions bind to A β -aggregate structures. These compounds exert a bifunctional ability, binding directly A β -aggregates and chelating metals as independent activities. Their potential to inhibit Cu²⁺-induced A β -aggregation was therefore tested. Co-incubation experiments resulted in the inhibition of the high-molecular-weight aggregates in the presence of compounds **60** and **61**, an effect that was not observed in the presence of other metal chelators (EDTA) or the model substructures MPY and stilbene. This suggests the need for both activities, metal chelating and A β direct interaction, to be able to inhibit the aggregation effect of Cu²⁺. Metal chelators are known to inhibit H₂O₂ production by Cu-bound A β .²⁷⁴ Interestingly, compounds **60** and **61** are reduced by 70% H₂O₂ production. Finally, toxicity experiments proved compound **61** to be non toxic at concentrations as high as 200 μ M, suggesting that it is a good candidate to be further studied *in vitro* and *in vivo*. These small compounds are neutral, lipophilic and able to cross the BBB. By mimicking them and inserting oxygen and/or nitrogen donors, these molecules should have low toxicity, and selective chelating properties.

10. CONCLUSIONS

During the last decade, substantial research efforts have been devoted to the development of multitarget drugs for different diseases. This change in the emphasis has been driven by the increase in the understanding of the pathogenesis of complex chronic diseases, and the fact that the paradigm one-target-one-molecule has not been as successful as expected. Complex diseases, such as cancer and neurodegenerative diseases, particularly AD, are composed of many different cross-linked pathological pathways, affecting a continuously increasing number of metabolic routes. AD pathogenesis is composed of many different mechanisms, interacting between them to generate a high complex network.⁶⁷ Relationships between different routes generate key nodes implicated in disease progression. Even now, this network is not well understood; however, there are some hints about important nodes that are known targets for disease modifying drugs.¹⁰⁷ Examples of these targets include APP pathogenic cleavage,³⁴³ cytoskeletal destabilization,³⁴⁴ axonal transport impairment, neurotransmitter and Ca²⁺ dyshomeostasis,²⁷ metal ion accumulation,³⁴⁵ protein misfolding,³⁴³ oxidative stress,⁷³ neuronal death, and gene mutations.¹⁴ There are drugs effective against almost all of these targets; however, none of them are likely to provide an efficient treatment for AD. On the other hand, only a few marketed drugs are available. These drugs constitute only a symptomatic treatment, effective for a few months; afterwards, disease progression is unavoidable. Furthermore, compelling evidence is questioning the effectiveness of the treatment with AChEIs,¹⁰⁸ while other treatments are emerging such as memantine, the first non-AChEI marketed for AD.

Initial evidence for the multitarget approach was the discovery of multiple activities in some natural products. These have been used as starting points for further developments. Compounds such as curcumin,³⁴⁶ resveratrol,³⁴⁶ and some flavonoids³⁴⁷ were studied, because of their interesting antioxidant and anti-inflammatory properties. Further studies demonstrated their ability to modify A β aggregation and metal dyshomeostasis. Although natural products are not discussed in this review, however, there is a plethora of compounds which may be important in multitarget drug design. These can also serve as templates to achieve new biological activities, which medicinal chemists should have in mind, during the discovery process.^{348–350} Recent multitarget drugs have mainly been designed by studying the

3D structure of previous molecules with known activities and crystal structures of target proteins. This information is focused to the virtual design of new chemical entities that include more than one activity in a single molecule.³⁵¹

Multitarget approach has been used to describe the benefits of the use of combination of drugs. It is now widely accepted in the treatment of complex diseases such as cancer, where the use of several one-target specific drugs in combination leads to better results, than the treatment with each one separately. This effect can be explained by the alteration of several interconnected pathological pathways, modifying the progression of the disease. This type of therapy has opened a broad line of research for scientist in different fields. Defining correlations and links among the pathological network, and the use of this information to design specific molecules. These molecules will include several targets which will be tested as new potential drugs, among many others.⁴⁶ Nevertheless, feedback currents of information between all different fields will be crucial for success in the search of a real disease modifying drug for AD. Future advances in AD treatment will summarize all knowledge about the physiopathological progress of the disease and the different relations among them, in order to achieve the right balance of biological activity in the optimal multitarget drug.

ABBREVIATIONS

A β	amyloid- β
ACh	acetylcholine
AD	Alzheimer's disease
AChEI	AChE inhibitor
AChEIs	AChE inhibitors
AICD	amyloid intracellular domain
AMPA	α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid
APOE	apolipoprotein E
APOE4	apolipoprotein E 4
APP	amyloid precursor protein
APP α	amyloid protein precursor α fragment
BACE-1	β -secretase
BBB	blood-brain barrier
BuChE	butyrylcholinesterase
ChAT	choline acetyl transferase
CoQ10	coenzyme Q10
CDK5	cyclin-dependent kinase 5
EDTA	ethylene-diamino tetracetic acid
EGFR	epidermal growth factor receptor
ER	endoplasmic reticulum
ERK2	extracellular signal-regulated kinase 2
GABA	γ -aminobutyric acid
GSK3 β	glycogen synthase kinase 3 β
H ₃	Histamine 3
5-HT	5-hydroxytryptamine receptors
IDE	insulin-degrading enzyme
InsP3R	inositol triphosphate receptors
MCU	mitochondrial Ca ²⁺ uniporter
mtPTP	mitochondrial permeability-transition pore
MTDLs	multitarget-directed ligands

nAChRs	nicotinic acetylcholine receptors
NET	norepinephrine transporter
NFTs	neurofibrillary tangles
NMDAr	<i>N</i> -methyl <i>D</i> -aspartate receptors
NQO1	NAD(P)H:quinone oxidoreductase 1
NSAIDs	nonsteroidal anti-inflammatory drugs
ORAC	oxygen radical absorbance capacity
PAS	peripheral anionic site
PS	presenilins
ROS	reactive oxygen species
RTC	tyrosine kinase
RyR	ryanodine receptors
sAPP α	APP fragment α
sAPP β	soluble APP β
SERT	serotonin reuptake transporter
τ	tau-protein
VDCCs	voltage-dependent calcium channels
VEGF	vascular endothelial growth factor

ACKNOWLEDGMENTS

R.L. thanks Ministerio de educación y ciencia of the Spanish government and Marie Curie Foundation for fellowships. The authors gratefully acknowledge to Peter Wathen and Rhiannon Beard for helpful discussion. A.G.G. thanks Fundación CIEN (PI 016/09) and Agencia Laín Entralgo (NDG07/9) for financial support. J.M.C. thanks MICINN (SAF2006-08764-C02-01; SAF2009-07271), CSIC-GRICES (2007PT-13), ISCIII, MICINN (RD06/0026/1002 Retic “RENEVAS”), and Comunidad de Madrid (S/SAL-0275-2006) for financial support.

REFERENCES

1. Alzheimer A. Über einen eigenartigen schweren Erkrankungsprozeß der Hirnrinde. *Neurologisches Centralblatt* 1906;23:1129–1136.
2. Jellinger KA. Alzheimer 100—Highlights in the history of Alzheimer research. *J Neural Transm* 2006;113:1603–1623.
3. Goedert M, Spillantini MG. A century of Alzheimer’s disease. *Science* 2006;314:777–781.
4. Castro A, Martinez A. Targeting beta-amyloid pathogenesis through acetylcholinesterase inhibitors. *Curr Pharm Des* 2006;12:4377–4387.
5. Alonso AC, Grundke-Iqbal I, Iqbal K. Alzheimer’s disease hyperphosphorylated tau sequesters normal tau into tangles of filaments and disassembles microtubules. *Nat Med* 1996;2:783–787.
6. Andersen JK. Oxidative stress in neurodegeneration: Cause or consequence? *Nat Med* 2004;10:S18–S25.
7. Cummings JL. Treatment of Alzheimer’s disease: Current and future therapeutic approaches. *Rev Neurol Dis* 2004;1:60–69.
8. Scarpini E, Scheltens P, Feldman H. Treatment of Alzheimer’s disease: Current status and new perspectives. *Lancet Neurol* 2003;2:539–547.

9. Iqbal K, Grundke-Iqbal I. Alzheimer disease is multifactorial and heterogeneous. *Neurobiol Aging* 2000;21:901–902; discussion 903–904.
10. Inestrosa NC, Alvarez A, Perez CA, Moreno RD, Vicente M, Linker C, Casanueva OI, Soto C, Garrido J. Acetylcholinesterase accelerates assembly of amyloid-beta-peptides into Alzheimer's fibrils: Possible role of the peripheral site of the enzyme. *Neuron* 1996;16:881–891.
11. Grimes CA, Jope RS. The multifaceted roles of glycogen synthase kinase 3beta in cellular signaling. *Prog Neurobiol* 2001;65:391–426.
12. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 1993;261:921–923.
13. Tiraboschi P, Hansen LA, Masliah E, Alford M, Thal LJ, Corey-Bloom J. Impact of APOE genotype on neuropathologic and neurochemical markers of Alzheimer disease. *Neurology* 2004;62:1977–1983.
14. Ye S, Huang Y, Mullendorff K, Dong L, Giedt G, Meng EC, Cohen FE, Kuntz ID, Weisgraber KH, Mahley RW. Apolipoprotein (apo) E4 enhances amyloid beta peptide production in cultured neuronal cells: apoE structure as a potential therapeutic target. *Proc Natl Acad Sci USA* 2005;102:18700–18705.
15. Bales KR, Dodart JC, DeMattos RB, Holtzman DM, Paul SM. Apolipoprotein E, amyloid, and Alzheimer disease. *Mol Interv* 2002;2:363–375, 339.
16. DeMattos RB, Cirrito JR, Parsadanian M, May PC, O'Dell MA, Taylor JW, Harmony JA, Aronow BJ, Bales KR, Paul SM, Holtzman DM. ApoE and clusterin cooperatively suppress Abeta levels and deposition: Evidence that ApoE regulates extracellular Abeta metabolism in vivo. *Neuron* 2004;41:193–202.
17. Fagan AM, Watson M, Parsadanian M, Bales KR, Paul SM, Holtzman DM. Human and murine ApoE markedly alters A beta metabolism before and after plaque formation in a mouse model of Alzheimer's disease. *Neurobiol Dis* 2002;9:305–318.
18. Cedazo-Minguez A, Popescu BO, Blanco-Millan JM, Akterin S, Pei JJ, Winblad B, Cowburn RF. Apolipoprotein E and beta-amyloid (1–42) regulation of glycogen synthase kinase-3beta. *J Neurochem* 2003;87:1152–1164.
19. Takashima A, Noguchi K, Sato K, Hoshino T, Imahori K. Tau protein kinase I is essential for amyloid beta-protein-induced neurotoxicity. *Proc Natl Acad Sci USA* 1993;90:7789–7793.
20. Takashima A, Noguchi K, Michel G, Mercken M, Hoshi M, Ishiguro K, Imahori K. Exposure of rat hippocampal neurons to amyloid beta peptide (25–35) induces the inactivation of phosphatidylinositol-3 kinase and the activation of tau protein kinase I/glycogen synthase kinase-3 beta. *Neurosci Lett* 1996;203:33–36.
21. Takashima A, Honda T, Yasutake K, Michel G, Murayama O, Murayama M, Ishiguro K, Yamaguchi H. Activation of tau protein kinase I/glycogen synthase kinase-3beta by amyloid beta peptide (25–35) enhances phosphorylation of tau in hippocampal neurons. *Neurosci Res* 1998;31:317–323.
22. Ferreira A, Lu Q, Orecchio L, Kosik KS. Selective phosphorylation of adult tau isoforms in mature hippocampal neurons exposed to fibrillar A beta. *Mol Cell Neurosci* 1997;9:220–234.
23. Marx J. Alzheimer's disease. Fresh evidence points to an old suspect: calcium. *Science* 2007;318:384–385.
24. Green KN, LaFerla FM. Linking calcium to Abeta and Alzheimer's disease. *Neuron* 2008;59:190–194.
25. Khachaturian ZS. Calcium, membranes, aging, and Alzheimer's disease. Introduction and overview. *Ann N Y Acad Sci* 1989;568:1–4.
26. Mattson MP. ER calcium and Alzheimer's disease: in a state of flux. *Sci Signal* 2010;3:pe10.
27. Demuro A, Parker I, Stutzmann GE. Calcium signaling and amyloid toxicity in Alzheimer disease. *J Biol Chem* 2010;285:12463–12468.

28. Khachaturian ZS. Calcium hypothesis of Alzheimer's disease and brain aging. *Ann N Y Acad Sci* 1994;747:1–11.
29. Pollard HB, Rojas E, Arispe N. A new hypothesis for the mechanism of amyloid toxicity, based on the calcium channel activity of amyloid beta protein (A beta P) in phospholipid bilayer membranes. *Ann N Y Acad Sci* 1993;695:165–168.
30. Li S, Hong S, Shepardson NE, Walsh DM, Shankar GM, Selkoe D. Soluble oligomers of amyloid Beta protein facilitate hippocampal long-term depression by disrupting neuronal glutamate uptake. *Neuron* 2009;62:788–801.
31. Parsons CG, Stoffler A, Danysz W. Memantine: A NMDA receptor antagonist that improves memory by restoration of homeostasis in the glutamatergic system—Too little activation is bad, too much is even worse. *Neuropharmacology* 2007;53:699–723.
32. Rosini M, Simoni E, Bartolini M, Cavalli A, Ceccarini L, Pascu N, McClymont DW, Tarozzi A, Bolognesi ML, Minarini A, Tumiatti V, Andrisano V, Mellor IR, Melchiorre C. Inhibition of acetylcholinesterase, beta-amyloid aggregation, and NMDA receptors in Alzheimer's disease: a promising direction for the multi-target-directed ligands gold rush. *J Med Chem* 2008;51:4381–4384.
33. Brown D, Superti-Furga G. Rediscovering the sweet spot in drug discovery. *Drug Discov Today* 2003;8:1067–1077.
34. Overington JP, Al-Lazikani B, Hopkins AL. How many drug targets are there? *Nat Rev Drug Discov* 2006;5:993–996.
35. Szuromi P, Vinson V, Marshal E. Rethinking drug discovery. *Science* 2004;303:1795.
36. Hartman JLt, Garvik B, Hartwell L. Principles for the buffering of genetic variation. *Science* 2001;291:1001–1004.
37. Papp B, Pal C, Hurst LD. Metabolic network analysis of the causes and evolution of enzyme dispensability in yeast. *Nature* 2004;429:661–664.
38. Pal C, Papp B, Lercher MJ, Csermely P, Oliver SG, Hurst LD. Chance and necessity in the evolution of minimal metabolic networks. *Nature* 2006;440:667–670.
39. Kitano H. A robustness-based approach to systems-oriented drug design. *Nat Rev Drug Discov* 2007;6:202–210.
40. Sharom JR, Bellows DS, Tyers M. From large networks to small molecules. *Curr Opin Chem Biol* 2004;8:81–90.
41. Borisy AA, Elliott PJ, Hurst NW, Lee MS, Lehar J, Price ER, Serbedzija G, Zimmermann GR, Foley MA, Stockwell BR, Keith CT. Systematic discovery of multicomponent therapeutics. *Proc Natl Acad Sci USA* 2003;100:7977–7982.
42. Dancey JE, Chen HX. Strategies for optimizing combinations of molecularly targeted anticancer agents. *Nat Rev Drug Discov* 2006;5:649–659.
43. Keith CT, Borisy AA, Stockwell BR. Multicomponent therapeutics for networked systems. *Nat Rev Drug Discov* 2005;4:71–78.
44. Millan MJ. Multi-target strategies for the improved treatment of depressive states: Conceptual foundations and neuronal substrates, drug discovery and therapeutic application. *Pharmacol Ther* 2006;110:135–370.
45. Zimmermann GR, Lehar J, Keith CT. Multi-target therapeutics: when the whole is greater than the sum of the parts. *Drug Discov Today* 2007;12:34–42.
46. Cavalli A, Bolognesi ML, Minarini A, Rosini M, Tumiatti V, Recanatini M, Melchiorre C. Multi-target-directed ligands to combat neurodegenerative diseases. *J Med Chem* 2008;51:347–372.
47. Roth BL, Sheffler DJ, Kroeze WK. Magic shotguns versus magic bullets: selectively non-selective drugs for mood disorders and schizophrenia. *Nat Rev Drug Discov* 2004;3:353–359.
48. Stahl SM, Entsuah R, Rudolph RL. Comparative efficacy between venlafaxine and SSRIs: a pooled analysis of patients with depression. *Biol Psychiatry* 2002;52:1166–1174.

49. Tatsumi M, Groshan K, Blakely RD, Richelson E. Pharmacological profile of antidepressants and related compounds at human monoamine transporters. *Eur J Pharmacol* 1997;340:249–258.
50. Owens MJ, Morgan WN, Plott SJ, Nemeroff CB. Neurotransmitter receptor and transporter binding profile of antidepressants and their metabolites. *J Pharmacol Exp Ther* 1997;283:1305–1322.
51. Rauser L, Savage JE, Meltzer HY, Roth BL. Inverse agonist actions of typical and atypical antipsychotic drugs at the human 5-hydroxytryptamine(2C) receptor. *J Pharmacol Exp Ther* 2001;299:83–89.
52. Werling LL, Keller A, Frank JG, Nuwayhid SJ. A comparison of the binding profiles of dextromethorphan, memantine, fluoxetine and amitriptyline: treatment of involuntary emotional expression disorder. *Exp Neurol* 2007;207:248–257.
53. Sills MA, Loo PS. Tricyclic antidepressants and dextromethorphan bind with higher affinity to the phencyclidine receptor in the absence of magnesium and L-glutamate. *Mol Pharmacol* 1989;36:160–165.
54. Zahradnik I, Minarovic I, Zahradnikova A. Inhibition of the cardiac L-type calcium channel current by antidepressant drugs. *J Pharmacol Exp Ther* 2008;324:977–984.
55. Punke MA, Friederich P. Amitriptyline is a potent blocker of human Kv1.1 and Kv7.2/7.3 channels. *Anesth Analg* 2007;104:1256–1264, tables of contents.
56. Jang SW, Liu X, Chan CB, Weinshenker D, Hall RA, Xiao G, Ye K. Amitriptyline is a TrkA and TrkB receptor agonist that promotes TrkA/TrkB heterodimerization and has potent neurotrophic activity. *Chem Biol* 2009;16:644–656.
57. Mencher SK, Wang LG. Promiscuous drugs compared to selective drugs (promiscuity can be a virtue). *BMC Clin Pharmacol* 2005;5:3.
58. Mendel DB, Laird AD, Xin X, Louie SG, Christensen JG, Li G, Schreck RE, Abrams TJ, Ngai TJ, Lee LB, Murray LJ, Carver J, Chan E, Moss KG, Haznedar JO, Sukbuntherng J, Blake RA, Sun L, Tang C, Miller T, Shirazian S, McMahon G, Cherrington JM. In vivo antitumor activity of SU11248, a novel tyrosine kinase inhibitor targeting vascular endothelial growth factor and platelet-derived growth factor receptors: determination of a pharmacokinetic/pharmacodynamic relationship. *Clin Cancer Res* 2003;9:327–337.
59. Mitsui H, Takuwa N, Maruyama T, Maekawa H, Hirayama M, Sawatari T, Hashimoto N, Takuwa Y, Kimura S. The MEK1-ERK map kinase pathway and the PI 3-kinase-Akt pathway independently mediate anti-apoptotic signals in HepG2 liver cancer cells. *Int J Cancer* 2001;92:55–62.
60. Kaelin WG, Jr. Gleevec: prototype or outlier? *Sci STKE* 2004;2004:pe12.
61. Druker BJ, Lydon NB. Lessons learned from the development of an abl tyrosine kinase inhibitor for chronic myelogenous leukemia. *J Clin Invest* 2000;105:3–7.
62. Frantz S. Drug discovery: Playing dirty. *Nature* 2005;437:942–943.
63. Shashkin PN, Huang LC, Larner J, Vandenhoff GE, Katz A. Fasting decreases the content of D-chiroinositol in human skeletal muscle. *Int J Exp Diabetes Res* 2002;3:163–169.
64. Morphy R, Kay C, Rankovic Z. From magic bullets to designed multiple ligands. *Drug Discov Today* 2004;9:641–651.
65. Rogawski MA. Low affinity channel blocking (uncompetitive) NMDA receptor antagonists as therapeutic agents—Toward an understanding of their favorable tolerability. *Amino Acids* 2000;19:133–149.
66. Youdim MB, Buccafusco JJ. Multi-functional drugs for various CNS targets in the treatment of neurodegenerative disorders. *Trends Pharmacol Sci* 2005;26:27–35.
67. De Strooper B. Proteases and proteolysis in Alzheimer disease: a multifactorial view on the disease process. *Physiol Rev* 2010;90:465–494.
68. Bolognesi ML, Cavalli A, Bergamini C, Fato R, Lenaz G, Rosini M, Bartolini M, Andrisano V, Melchiorre C. Toward a rational design of multitarget-directed antioxidants: merging melatonin and lipoic acid molecular frameworks. *J Med Chem* 2009;52:7883–7886.

69. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002;297:353–356.
70. Venugopal C, Demos CM, Rao KS, Pappolla MA, Sambamurti K. Beta-secretase: structure, function, and evolution. *CNS Neurol Disord Drug Targets* 2008;7:278–294.
71. Hernandez F, Gomez de Barreda E, Fuster-Matanzo A, Lucas JJ, Avila J. GSK3: A possible link between beta amyloid peptide and tau protein. *Exp Neurol* 2010;223:322–325.
72. Berridge MJ. Calcium signalling and Alzheimer's disease. *Neurochem Res* 2011. DOI: 10.1007/s11064-010-0371-4.
73. Di Bona D, Scapagnini G, Candore G, Castiglia L, Colonna-Romano G, Duro G, Nuzzo D, Iemolo F, Lio D, Pellicano M, Scafidi V, Caruso C, Vasto S. Immune-inflammatory responses and oxidative stress in Alzheimer' disease: Therapeutic implications. *Curr Pharm Des* 2010; 16:684–691.
74. Francis PT, Palmer AM, Snape M, Wilcock GK. The cholinergic hypothesis of Alzheimer's disease: A review of progress. *J Neurol Neurosurg Psychiatry* 1999;66:137–147.
75. Whitehouse PJ, Price DL, Struble RG, Clark AW, Coyle JT, Delon MR. Alzheimer's disease and senile dementia: Loss of neurons in the basal forebrain. *Science* 1982;215:1237–1239.
76. Farlow MR, Evans RM. Pharmacologic treatment of cognition in Alzheimer's dementia. *Neurology* 1998;51:S36–S44; discussion S65–S37.
77. Perry EK, Tomlinson BE, Blessed G, Bergmann K, Gibson PH, Perry RH. Correlation of cholinergic abnormalities with senile plaques and mental test scores in senile dementia. *Br Med J* 1978;2:1457–1459.
78. Buccafusco JJ, Terry AV, Jr. Multiple central nervous system targets for eliciting beneficial effects on memory and cognition. *J Pharmacol Exp Ther* 2000;295:438–446.
79. Bryson HM, Benfield P. Donepezil. *Drugs Aging* 1997;10:234–239; discussion 240–231.
80. Scott LJ, Goa KL. Galantamine: A review of its use in Alzheimer's disease. *Drugs* 2000;60: 1095–1122.
81. Polinsky RJ. Clinical pharmacology of rivastigmine: A new-generation acetylcholinesterase inhibitor for the treatment of Alzheimer's disease. *Clin Ther* 1998;20:634–647.
82. Nordberg A, Svensson AL. Cholinesterase inhibitors in the treatment of Alzheimer's disease: A comparison of tolerability and pharmacology. *Drug Saf* 1998;19:465–480.
83. Fariss MW, Mumaw VR, Walton LP. Tetrahydroaminoacridine-induced apoptosis in rat hepatocytes. *Toxicol In Vitro* 1996;10:383–393.
84. Parsons CG, Danysz W, Quack G. Memantine is a clinically well tolerated N-methyl-D-aspartate (NMDA) receptor antagonist—A review of preclinical data. *Neuropharmacology* 1999;38:735–767.
85. Takeda A, Loveman E, Clegg A, Kirby J, Picot J, Payne E, Green C. A systematic review of the clinical effectiveness of donepezil, rivastigmine and galantamine on cognition, quality of life and adverse events in Alzheimer's disease. *Int J Geriatr Psychiatry* 2006;21:17–28.
86. Birks J, Flicker L. Donepezil for mild cognitive impairment. *Cochrane Database Syst Rev* 2006;3: CD006104.
87. Birks J. Cholinesterase inhibitors for Alzheimer's disease. *Cochrane Database Syst Rev* 2006;25: CD005593.
88. Farlow MR, Miller ML, Pejovic V. Treatment options in Alzheimer's disease: maximizing benefit, managing expectations. *Dement Geriatr Cogn Disord* 2008;25:408–422.
89. Thatte U. Phenserine Axonyx. *Curr Opin Investig Drugs* 2005;6:729–739.
90. <http://clinicaltrials.gov/ct2/show/NCT00568776>. Accessed May 14, 2010.
91. Reger MA, Watson GS, Green PS, Wilkinson CW, Baker LD, Cholerton B, Fishel MA, Plymate SR, Breitner JC, DeGroodt W, Mehta P, Craft S. Intranasal insulin improves cognition and modulates beta-amyloid in early AD. *Neurology* 2008;70:440–448.
92. Szekely CA, Zandi PP. Non-steroidal anti-inflammatory drugs and Alzheimer's disease: the epidemiological evidence. *CNS Neurol Disord Drug Targets* 2010;9:132–139.

93. Aisen PS, Schafer KA, Grundman M, Pfeiffer E, Sano M, Davis KL, Farlow MR, Jin S, Thomas RG, Thal LJ. Effects of rofecoxib or naproxen vs placebo on Alzheimer disease progression: A randomized controlled trial. *JAMA* 2003;289:2819–2826.
94. Rogers J, Kirby LC, Hempelman SR, Berry DL, McGeer PL, Kaszniak AW, Zilinski J, Cofield M, Mansukhani L, Willson P, Kogan F. Clinical trial of indomethacin in Alzheimer's disease. *Neurology* 1993;43:1609–1611.
95. Scharf S, Mander A, Ugoni A, Vajda F, Christophidis N. A double-blind, placebo-controlled trial of diclofenac/misoprostol in Alzheimer's disease. *Neurology* 1999;53:197–201.
96. Thal LJ, Ferris SH, Kirby L, Block GA, Lines CR, Yuen E, Assaid C, Nessly ML, Norman BA, Baranak CC, Reines SA. A randomized, double-blind, study of rofecoxib in patients with mild cognitive impairment. *Neuropsychopharmacology* 2005;30:1204–1215.
97. Group AR, Lyketsos CG, Breitner JC, Green RC, Martin BK, Meinert C, Piantadosi S, Sabbagh M. Naproxen and celecoxib do not prevent AD in early results from a randomized controlled trial. *Neurology* 2007;68:1800–1808.
98. Martin BK, Szekely C, Brandt J, Piantadosi S, Breitner JC, Craft S, Evans D, Green R, Mullan M. Cognitive function over time in the Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT): Results of a randomized, controlled trial of naproxen and celecoxib. *Arch Neurol* 2008;65:896–905.
99. Lundkvist J, Naslund J. Gamma-secretase: a complex target for Alzheimer's disease. *Curr Opin Pharmacol* 2007;7:112–118.
100. Belluardo N, Mudo G, Blum M, Fuxe K. Central nicotinic receptors, neurotrophic factors and neuroprotection. *Behav Brain Res* 2000;113:21–34.
101. Sabbagh MN. Drug development for Alzheimer's disease: Where are we now and where are we headed? *Am J Geriatr Pharmacother* 2009;7:167–185.
102. Jakob-Roetne R, Jacobsen H. Alzheimer's disease: From pathology to therapeutic approaches. *Angew Chem Int Ed Engl* 2009;48:3030–3059.
103. Hooper C, Killick R, Lovestone S. The GSK3 hypothesis of Alzheimer's disease. *J Neurochem* 2008;104:1433–1439.
104. Brunden KR, Trojanowski JQ, Lee VM. Advances in tau-focused drug discovery for Alzheimer's disease and related tauopathies. *Nat Rev Drug Discov* 2009;8:783–793.
105. Kim J, Basak JM, Holtzman DM. The role of apolipoprotein E in Alzheimer's disease. *Neuron* 2009;63:287–303.
106. Shah RS, Lee HG, Xiongwei Z, Perry G, Smith MA, Castellani RJ. Current approaches in the treatment of Alzheimer's disease. *Biomed Pharmacother* 2008;62:199–207.
107. Biran Y, Masters CL, Barnham KJ, Bush AI, Adlard PA. Pharmacotherapeutic targets in Alzheimer's disease. *J Cell Mol Med* 2009;13:61–86.
108. Shanks M, Kivipelto M, Bullock R, Lane R. Cholinesterase inhibition: Is there evidence for disease-modifying effects? *Curr Med Res Opin* 2009;25:2439–2446.
109. Inestrosa NC, Alvarez A, Calderon F. Acetylcholinesterase is a senile plaque component that promotes assembly of amyloid beta-peptide into Alzheimer's filaments. *Mol Psychiatry* 1996;1:359–361.
110. Selkoe DJ. Translating cell biology into therapeutic advances in Alzheimer's disease. *Nature* 1999;399:A23–A31.
111. De Ferrari GV, Canales MA, Shin I, Weiner LM, Silman I, Inestrosa NC. A structural motif of acetylcholinesterase that promotes amyloid beta-peptide fibril formation. *Biochemistry* 2001;40:10447–10457.
112. Bartolini M, Bertucci C, Cavrini V, Andrisano V. beta-Amyloid aggregation induced by human acetylcholinesterase: Inhibition studies. *Biochem Pharmacol* 2003;65:407–416.
113. Sussman JL, Harel M, Frolow F, Oefner C, Goldman A, Toker L, Silman I. Atomic structure of acetylcholinesterase from *Torpedo californica*: A prototypic acetylcholine-binding protein. *Science* 1991;253:872–879.

114. Kryger G, Silman I, Sussman JL. Structure of acetylcholinesterase complexed with E2020 (Aricept): Implications for the design of new anti-Alzheimer drugs. *Structure* 1999;7:297–307.
115. Greenblatt HM, Sillman I, Sussman JL. Structural studies on vertebrate and invertebrate acetylcholinesterases and their complexes with functional ligands. *Drug Dev Res* 2000;50:573.
116. Munoz-Torrero D, Camps P. Dimeric and hybrid anti-Alzheimer drug candidates. *Curr Med Chem* 2006;13:399–422.
117. Pang YP, Hong F, Quiram P, Jelacic T, Brimijoin S. Synthesis of alkylene linked bis-THA and alkylene linked benzyl-THA as highly potent and selective inhibitors and molecular probes of acetylcholinesterase. *J Chem Soc Perkin Trans* 1997;1:171.
118. Tang XC, De Sarno P, Sugaya K, Giacobini E. Effect of huperzine A, a new cholinesterase inhibitor, on the central cholinergic system of the rat. *J Neurosci Res* 1989;24:276–285.
119. Camps P, Cusack B, Mallender WD, El Achab RE, Morral J, Munoz-Torrero D, Rosenberry TL. Huprine X is a novel high-affinity inhibitor of acetylcholinesterase that is of interest for treatment of Alzheimer's disease. *Mol Pharmacol* 2000;57:409–417.
120. Nilsson L, Nordberg A, Hardy J, Wester P, Winblad B. Physostigmine restores 3H-acetylcholine efflux from Alzheimer brain slices to normal level. *J Neural Transm* 1986;67:275–285.
121. Cavalli A, Bottegoni G, Raco C, De Vivo M, Recanatini M. A computational study of the binding of propidium to the peripheral anionic site of human acetylcholinesterase. *J Med Chem* 2004;47:3991–3999.
122. Pang YP, Quiram P, Jelacic T, Hong F, Brimijoin S. Highly potent, selective, and low cost bis-tetrahydroaminacrine inhibitors of acetylcholinesterase. Steps toward novel drugs for treating Alzheimer's disease. *J Biol Chem* 1996;271:23646–23649.
123. Liu J, Ho W, Lee NT, Carlier PR, Pang Y, Han Y. Bis(7)-tacrine, a novel acetylcholinesterase inhibitor, reverses AF64A-induced deficits in navigational memory in rats. *Neurosci Lett* 2000;282:165–168.
124. Ros E, Aleu J, Gomez de Aranda I, Canti C, Pang YP, Marsal J, Solsona C. Effects of bis(7)-tacrine on spontaneous synaptic activity and on the nicotinic ACh receptor of Torpedo electric organ. *J Neurophysiol* 2001;86:183–189.
125. Li W, Pi R, Chan HH, Fu H, Lee NT, Tsang HW, Pu Y, Chang DC, Li C, Luo J, Xiong K, Li Z, Xue H, Carlier PR, Pang Y, Tsim KW, Li M, Han Y. Novel dimeric acetylcholinesterase inhibitor bis(7)-tacrine, but not donepezil, prevents glutamate-induced neuronal apoptosis by blocking N-methyl-D-aspartate receptors. *J Biol Chem* 2005;280:18179–18188.
126. Fu H, Li W, Luo J, Lee NT, Kan KK, Tsang HW, Tsim KW, Pang Y, Li Z, Chang DC, Li M, Han Y. Bis(7)-tacrine attenuates beta amyloid-induced neuronal apoptosis by regulating L-type calcium channels. *J Neurochem* 2006;98:1400–1410.
127. Xiao XQ, Lee NT, Carlier PR, Pang Y, Han YF. Bis(7)-tacrine, a promising anti-Alzheimer's agent, reduces hydrogen peroxide-induced injury in rat pheochromocytoma cells: comparison with tacrine. *Neurosci Lett* 2000;290:197–200.
128. Bolognesi ML, Cavalli A, Valgimigli L, Bartolini M, Rosini M, Andrisano V, Recanatini M, Melchiorre C. Multi-target-directed drug design strategy: From a dual binding site acetylcholinesterase inhibitor to a trifunctional compound against Alzheimer's disease. *J Med Chem* 2007;50:6446–6449.
129. Fu H, Li W, Luo J, Lee NT, Li M, Tsim KW, Pang Y, Youdim MB, Han Y. Promising anti-Alzheimer's dimer bis(7)-tacrine reduces beta-amyloid generation by directly inhibiting BACE-1 activity. *Biochem Biophys Res Commun* 2008;366:631–636.
130. Carlier PR, Du DM, Han Y, Liu J, Pang YP. Potent, easily synthesized huperzine A-tacrine hybrid acetylcholinesterase inhibitors. *Bioorg Med Chem Lett* 1999;9:2335–2338.
131. Carlier PR, Du DM, Han YF, Liu J, Perola E, Williams ID, Pang YP. Dimerization of an inactive fragment of huperzine A produces a drug with twice the potency of the natural product. *Angew Chem Int Ed Engl* 2000;39:1775–1777.

132. Wong DM, Greenblatt HM, Dvir H, Carlier PR, Han YF, Pang YP, Silman I, Sussman JL. Acetylcholinesterase complexed with bivalent ligands related to huperzine A: Experimental evidence for species-dependent protein-ligand complementarity. *J Am Chem Soc* 2003;125:363–373.
133. Shao D, Zou C, Luo C, Tang X, Li Y. Synthesis and evaluation of tacrine-E2020 hybrids as acetylcholinesterase inhibitors for the treatment of Alzheimer's disease. *Bioorg Med Chem Lett* 2004;14:4639–4642.
134. Guillou C, Mary A, Renko DZ, Gras E, Thal C. Potent acetylcholinesterase inhibitors: design, synthesis and structure-activity relationships of alkylene linked bis-galanthamine and galanthamine-galanthaminium salts. *Bioorg Med Chem Lett* 2000;10:637–639.
135. Mary A, Renko DZ, Guillou C, Thal C. Potent acetylcholinesterase inhibitors: design, synthesis, and structure-activity relationships of bis-interacting ligands in the galanthamine series. *Bioorg Med Chem* 1998;6:1835–1850.
136. Piazzzi L, Rampa A, Bisi A, Gobbi S, Belluti F, Cavalli A, Bartolini M, Andrisano V, Valenti P, Recanatini M. 3-(4-[[Benzyl(methyl)amino]methyl]phenyl)-6,7-dimethoxy-2H-2-chromenone (AP2238) inhibits both acetylcholinesterase and acetylcholinesterase-induced beta-amyloid aggregation: a dual function lead for Alzheimer's disease therapy. *J Med Chem* 2003;46:2279–2282.
137. Lewis WG, Green LG, Grynszpan F, Radic Z, Carlier PR, Taylor P, Finn MG, Sharpless KB. Click chemistry in situ: acetylcholinesterase as a reaction vessel for the selective assembly of a femtomolar inhibitor from an array of building blocks. *Angew Chem Int Ed Engl* 2002;41:1053–1057.
138. Manetsch R, Krasinski A, Radic Z, Raushel J, Taylor P, Sharpless KB, Kolb HC. In situ click chemistry: enzyme inhibitors made to their own specifications. *J Am Chem Soc* 2004;126:12809–12818.
139. Melchiorre C, Antonello A, Banzi R, Bolognesi ML, Minarini A, Rosini M, Tumiatti V. Polymethylene tetraamine backbone as template for the development of biologically active polyamines. *Med Res Rev* 2003;23:200–233.
140. Melchiorre C, Romualdi P, Bolognesi ML, Donatini A, Ferri S. Binding profile of benextramine at neuropeptide Y receptor subtypes in rat brain areas. *Eur J Pharmacol* 1994;265:93–98.
141. Melchiorre C, Andrisano V, Bolognesi ML, Budriesi R, Cavalli A, Cavrini V, Rosini M, Tumiatti V, Recanatini M. Acetylcholinesterase noncovalent inhibitors based on a polyamine backbone for potential use against Alzheimer's disease. *J Med Chem* 1998;41:4186–4189.
142. Kwon YE, Park JY, No KT, Shin JH, Lee SK, Eun JS, Yang JH, Shin TY, Kim DK, Chae BS, Leem JY, Kim KH. Synthesis, in vitro assay, and molecular modeling of new piperidine derivatives having dual inhibitory potency against acetylcholinesterase and Abeta1-42 aggregation for Alzheimer's disease therapeutics. *Bioorg Med Chem* 2007;15:6596–6607.
143. Rizzo S, Riviere C, Piazzzi L, Bisi A, Gobbi S, Bartolini M, Andrisano V, Morroni F, Tarozzi A, Monti JP, Rampa A. Benzofuran-based hybrid compounds for the inhibition of cholinesterase activity, beta amyloid aggregation, and abeta neurotoxicity. *J Med Chem* 2008;51:2883–2886.
144. Camps P, Formosa X, Galdeano C, Gomez T, Munoz-Torrero D, Scarpellini M, Viayna E, Badia A, Clos MV, Camins A, Pallas M, Bartolini M, Mancini F, Andrisano V, Estelrich J, Lizondo M, Bidon-Chanal A, Luque FJ. Novel donepezil-based inhibitors of acetyl- and butyrylcholinesterase and acetylcholinesterase-induced beta-amyloid aggregation. *J Med Chem* 2008;51:3588–3598.
145. Xie Q, Wang H, Xia Z, Lu M, Zhang W, Wang X, Fu W, Tang Y, Sheng W, Li W, Zhou W, Zhu X, Qiu Z, Chen H. Bis-(–)-nor-meptazinols as novel nanomolar cholinesterase inhibitors with high inhibitory potency on amyloid-beta aggregation. *J Med Chem* 2008;51:2027–2036.
146. Darvesh S, Hopkins DA, Geula C. Neurobiology of butyrylcholinesterase. *Nat Rev Neurosci* 2003;4:131–138.
147. Giacobini E. Cholinesterase inhibitors: New roles and therapeutic alternatives. *Pharmacol Res* 2004;50:433–440.

148. Greig NH, Utsuki T, Ingram DK, Wang Y, Pepeu G, Scali C, Yu QS, Mamczarz J, Holloway HW, Giordano T, Chen D, Furukawa K, Sambamurti K, Brossi A, Lahiri DK. Selective butyrylcholinesterase inhibition elevates brain acetylcholine, augments learning and lowers Alzheimer beta-amyloid peptide in rodent. *Proc Natl Acad Sci USA* 2005;102:17213–17218.
149. Rampa A, Piazzi L, Belluti F, Gobbi S, Bisi A, Bartolini M, Andrisano V, Cavrini V, Cavalli A, Recanatini M, Valenti P. Acetylcholinesterase inhibitors: SAR and kinetic studies on omega-[N-methyl-N-(3-alkylcarbamoyloxyphenyl)methyl]aminoalkoxyaryl derivatives. *J Med Chem* 2001;44:3810–3820.
150. Howlett DR, Perry AE, Godfrey F, Swatton JE, Jennings KH, Spitzfaden C, Wadsworth H, Wood SJ, Markwell RE. Inhibition of fibril formation in beta-amyloid peptide by a novel series of benzofurans. *Biochem J* 1999;340:283–289.
151. Harel M, Schalk I, Ehret-Sabatier L, Bouet F, Goeldner M, Hirth C, Axelsen PH, Silman I, Sussman JL. Quaternary ligand binding to aromatic residues in the active-site gorge of acetylcholinesterase. *Proc Natl Acad Sci USA* 1993;90:9031–9035.
152. Carlier PR, Chow ES, Han Y, Liu J, El Yazal J, Pang YP. Heterodimeric tacrine-based acetylcholinesterase inhibitors: Investigating ligand-peripheral site interactions. *J Med Chem* 1999;42:4225–4231.
153. Savini L, Gaeta A, Fattorusso C, Catalanotti B, Campiani G, Chiasserini L, Pellerano C, Novellino E, McKissic D, Saxena A. Specific targeting of acetylcholinesterase and butyrylcholinesterase recognition sites. Rational design of novel, selective, and highly potent cholinesterase inhibitors. *J Med Chem* 2003;46:1–4.
154. LeVine H, III. Quantification of beta-sheet amyloid fibril structures with thioflavin T. *Methods Enzymol* 1999;309:274–284.
155. De Ferrari GV, Mallender WD, Inestrosa NC, Rosenberry TL. Thioflavin T is a fluorescent probe of the acetylcholinesterase peripheral site that reveals conformational interactions between the peripheral and acylation sites. *J Biol Chem* 2001;276:23282–23287.
156. Li W, Hao JL, Tang Y, Chen Y, Qiu ZB. Structural comparisons of meptazinol with opioid analgesics. *Acta Pharmacol Sin* 2005;26:334–338.
157. Ennis C, Haroun F, Lattimer N. Can the effects of meptazinol on the guinea-pig isolated ileum be explained by inhibition of acetylcholinesterase? *J Pharm Pharmacol* 1986;38:24–27.
158. Paz A, Xie Q, Greenblatt HM, Fu W, Tang Y, Silman I, Qiu Z, Sussman JL. The crystal structure of a complex of acetylcholinesterase with a bis-(-)-nor-meptazinol derivative reveals disruption of the catalytic triad. *J Med Chem* 2009;52:2543–2549.
159. Tumiatti V, Milelli A, Minarini A, Rosini M, Bolognesi ML, Micco M, Andrisano V, Bartolini M, Mancini F, Recanatini M, Cavalli A, Melchiorre C. Structure-activity relationships of acetylcholinesterase noncovalent inhibitors based on a polyamine backbone. 4. Further investigation on the inner spacer. *J Med Chem* 2008;51:7308–7312.
160. Tumiatti V, Rosini M, Bartolini M, Cavalli A, Marucci G, Andrisano V, Angeli P, Banzi R, Minarini A, Recanatini M, Melchiorre C. Structure-activity relationships of acetylcholinesterase noncovalent inhibitors based on a polyamine backbone. 2. Role of the substituents on the phenyl ring and nitrogen atoms of caproctamine. *J Med Chem* 2003;46:954–966.
161. Available from: <http://www.noscira.com>
162. Mattson MP. Pathways towards and away from Alzheimer's disease. *Nature* 2004;430:631–639.
163. Pereira C, Agostinho P, Moreira PI, Cardoso SM, Oliveira CR. Alzheimer's disease-associated neurotoxic mechanisms and neuroprotective strategies. *Curr Drug Targets CNS Neurol Disord* 2005;4:383–403.
164. Sinha S, Lieberburg I. Cellular mechanisms of beta-amyloid production and secretion. *Proc Natl Acad Sci USA* 1999;96:11049–11053.
165. Tang BL, Liou YC. Novel modulators of amyloid-beta precursor protein processing. *J Neurochem* 2007;100:314–323.

166. Golde TE. Disease modifying therapy for AD? *J Neurochem* 2006;99:689–707.
167. Hull M, Berger M, Heneka M. Disease-modifying therapies in Alzheimer's disease: How far have we come? *Drugs* 2006;66:2075–2093.
168. Piazzzi L, Cavalli A, Colizzi F, Belluti F, Bartolini M, Mancini F, Recanatini M, Andrisano V, Rampa A. Multi-target-directed coumarin derivatives: hAChE and BACE1 inhibitors as potential anti-Alzheimer compounds. *Bioorg Med Chem Lett* 2008;18:423–426.
169. Piazzzi L, Cavalli A, Belluti F, Bisi A, Gobbi S, Rizzo S, Bartolini M, Andrisano V, Recanatini M, Rampa A. Extensive SAR and computational studies of 3-{4-[(benzylmethylamino)methyl]phenyl}-6,7-dimethoxy-2H-2-chromenone (AP2238) derivatives. *J Med Chem* 2007;50:4250–4254.
170. Drug Data Report 2002;24:119, 309, 597, 885.
171. Guo T, Hobbs DW. Development of BACE1 inhibitors for Alzheimer's disease. *Curr Med Chem* 2006;13:1811–1829.
172. Zhu Y, Xiao K, Ma L, Xiong B, Fu Y, Yu H, Wang W, Wang X, Hu D, Peng H, Li J, Gong Q, Chai Q, Tang X, Zhang H, Li J, Shen J. Design, synthesis and biological evaluation of novel dual inhibitors of acetylcholinesterase and beta-secretase. *Bioorg Med Chem* 2009;17:1600–1613.
173. Cumming JN, Le TX, Babu S, Carroll C, Chen X, Favreau L, Gaspari P, Guo T, Hobbs DW, Huang Y, Iserloh U, Kennedy ME, Kuvelkar R, Li G, Lowrie J, McHugh NA, Ozgur L, Pan J, Parker EM, Saionz K, Stamford AW, Strickland C, Tadesse D, Voigt J, Wang L, Wu Y, Zhang L, Zhang Q. Rational design of novel, potent piperazinone and imidazolidinone BACE1 inhibitors. *Bioorg Med Chem Lett* 2008;18:3236–3241.
174. Yang W, Lu W, Lu Y, Zhong M, Sun J, Thomas AE, Wilkinson JM, Fucini RV, Lam M, Randal M, Shi XP, Jacobs JW, McDowell RS, Gordon EM, Ballinger MD. Aminoethylenes: A tetrahedral intermediate isostere yielding potent inhibitors of the aspartyl protease BACE-1. *J Med Chem* 2006;49:839–842.
175. Rajapakse HA, Nantermet PG, Selnick HG, Munshi S, McGaughey GB, Lindsley SR, Young MB, Lai MT, Espeseth AS, Shi XP, Colussi D, Pietrak B, Crouthamel MC, Tugusheva K, Huang Q, Xu M, Simon AJ, Kuo L, Hazuda DJ, Graham S, Vacca JP. Discovery of oxadiazoyl tertiary carbinamine inhibitors of beta-secretase (BACE-1). *J Med Chem* 2006;49:7270–7273.
176. Ghosh AK, Kumaragurubaran N, Hong L, Kulkarni S, Xu X, Miller HB, Reddy DS, Weerasena V, Turner R, Chang W, Koelsch G, Tang J. Potent memapsin 2 (beta-secretase) inhibitors: Design, synthesis, protein-ligand X-ray structure, and in vivo evaluation. *Bioorg Med Chem Lett* 2008;18:1031–1036.
177. Maillard MC, Hom RK, Benson TE, Moon JB, Mamo S, Bienkowski M, Tomasselli AG, Woods DD, Prince DB, Paddock DJ, Emmons TL, Tucker JA, Dappen MS, Brogley L, Thorsett ED, Jewett N, Sinha S, John V. Design, synthesis, and crystal structure of hydroxyethyl secondary amine-based peptidomimetic inhibitors of human beta-secretase. *J Med Chem* 2007;50:776–781.
178. Radic Z, Reiner E, Taylor P. Role of the peripheral anionic site on acetylcholinesterase: Inhibition by substrates and coumarin derivatives. *Mol Pharmacol* 1991;39:98–104.
179. Bourne Y, Taylor P, Radic Z, Marchot P. Structural insights into ligand interactions at the acetylcholinesterase peripheral anionic site. *Embo J* 2003;22:1–12.
180. Jimenez O, de la Rosa G, Lavilla R. Straightforward access to a structurally diverse set of oxacyclic scaffolds through a four-component reaction. *Angew Chem Int Ed Engl* 2005;44:6521–6525.
181. Camps P, Formosa X, Galdeano C, Gomez T, Munoz-Torrero D, Ramirez L, Viayna E, Gomez E, Isambert N, Lavilla R, Badia A, Clos MV, Bartolini M, Mancini F, Andrisano V, Bidon-Chanal A, Huertas O, Dafni T, Luque FJ. Tacrine-based dual binding site acetylcholinesterase inhibitors as potential disease-modifying anti-Alzheimer drug candidates. *Chem Biol Interact* 2010;187:411–415.
182. Mancini F, Naldi M, Cavrini V, Andrisano V. Multiwell fluorometric and colorimetric microassays for the evaluation of beta-secretase (BACE-1) inhibitors. *Anal Bioanal Chem* 2007;388:1175–1183.

183. Rosini M, Andrisano V, Bartolini M, Melchiorre C. Organic compounds useful for the treatment of alzheimer's disease; their use and method of preparation. 2006.
184. Hanessian S, Yun H, Hou Y, Yang G, Bayrakdarian M, Therrien E, Moitessier N, Roggo S, Veenstra S, Tintelnot-Blomley M, Rondeau JM, Ostermeier C, Strauss A, Ramage P, Paganetti P, Neumann U, Betschart C. Structure-based design, synthesis, and memapsin 2 (BACE) inhibitory activity of carbocyclic and heterocyclic peptidomimetics. *J Med Chem* 2005;48:5175–5190.
185. Cavalli A, Bolognesi ML, Capsoni S, Andrisano V, Bartolini M, Margotti E, Cattaneo A, Recanatini M, Melchiorre C. A small molecule targeting the multifactorial nature of Alzheimer's disease. *Angew Chem Int Ed Engl* 2007;46:3689–3692.
186. Bolognesi ML, Andrisano V, Bartolini M, Banzi R, Melchiorre C. Propidium-based polyamine ligands as potent inhibitors of acetylcholinesterase and acetylcholinesterase-induced amyloid-beta aggregation. *J Med Chem* 2005;48:24–27.
187. Rosini M, Andrisano V, Bartolini M, Bolognesi ML, Hrelia P, Minarini A, Tarozzi A, Melchiorre C. Rational approach to discover multipotent anti-Alzheimer drugs. *J Med Chem* 2005;48:360–363.
188. Bolognesi ML, Minarini A, Tumiatti V, Melchiorre C. Lipoic acid, a lead structure for multi-target-directed drugs for neurodegeneration. *Mini Rev Med Chem* 2006;6:1269–1274.
189. Beal MF. Mitochondrial dysfunction and oxidative damage in Alzheimer's and Parkinson's diseases and coenzyme Q10 as a potential treatment. *J Bioenerg Biomembr* 2004;36:381–386.
190. Bragin V, Chemodanova M, Dzhaferova N, Bragin I, Czerniawski JL, Aliev G. Integrated treatment approach improves cognitive function in demented and clinically depressed patients. *Am J Alzheimers Dis Other Dement* 2005;20:21–26.
191. Bolognesi ML, Cavalli A, Melchiorre C. Memoquin: a multi-target-directed ligand as an innovative therapeutic opportunity for Alzheimer's disease. *Neurotherapeutics* 2009;6:152–162.
192. Kang YH, Pezzuto JM. Induction of quinone reductase as a primary screen for natural product anticarcinogens. *Methods Enzymol* 2004;382:380–414.
193. Sultana R, Butterfield DA. Role of oxidative stress in the progression of Alzheimer's disease. *J Alzheimers Dis* 2010;19:341–353.
194. Sutton HC, Winterbourn CC. On the participation of higher oxidation states of iron and copper in Fenton reactions. *Free Radic Biol Med* 1989;6:53–60.
195. Petersen RB, Nunomura A, Lee HG, Casadesus G, Perry G, Smith MA, Zhu X. Signal transduction cascades associated with oxidative stress in Alzheimer's disease. *J Alzheimers Dis* 2007;11:143–152.
196. Butterfield DA, Drake J, Pocernich C, Castegna A. Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid beta-peptide. *Trends Mol Med* 2001;7:548–554.
197. Mattson MP, Chan SL. Neuronal and glial calcium signaling in Alzheimer's disease. *Cell Calcium* 2003;34:385–397.
198. Bieschke J, Zhang Q, Powers ET, Lerner RA, Kelly JW. Oxidative metabolites accelerate Alzheimer's amyloidogenesis by a two-step mechanism, eliminating the requirement for nucleation. *Biochemistry* 2005;44:4977–4983.
199. Honda K, Smith MA, Zhu X, Baus D, Merrick WC, Tartakoff AM, Hattier T, Harris PL, Siedlak SL, Fujioka H, Liu Q, Moreira PI, Miller FP, Nunomura A, Shimohama S, Perry G. Ribosomal RNA in Alzheimer disease is oxidized by bound redox-active iron. *J Biol Chem* 2005;280:20978–20986.
200. Crouch PJ, White AR, Bush AI. The modulation of metal bio-availability as a therapeutic strategy for the treatment of Alzheimer's disease. *FEBS J* 2007;274:3775–3783.
201. Atwood CS, Scarpa RC, Huang X, Moir RD, Jones WD, Fairlie DP, Tanzi RE, Bush AI. Characterization of copper interactions with alzheimer amyloid beta peptides: Identification of an attomolar-affinity copper binding site on amyloid beta1-42. *J Neurochem* 2000;75:1219–1233.
202. Curtain CC, Ali F, Volitakis I, Cherny RA, Norton RS, Beyreuther K, Barrow CJ, Masters CL, Bush AI, Barnham KJ. Alzheimer's disease amyloid-beta binds copper and zinc to generate an

- allosterically ordered membrane-penetrating structure containing superoxide dismutase-like subunits. *J Biol Chem* 2001;276:20466–20473.
203. Jiang D, Men L, Wang J, Zhang Y, Chickenyen S, Wang Y, Zhou F. Redox reactions of copper complexes formed with different beta-amyloid peptides and their neuropathological [correction of neuropathological] relevance. *Biochemistry* 2007;46:9270–9282.
 204. Holmquist L, Stuchbury G, Berbaum K, Muscat S, Young S, Hager K, Engel J, Munch G. Lipoic acid as a novel treatment for Alzheimer's disease and related dementias. *Pharmacol Ther* 2007; 113:154–164.
 205. Fang L, Kraus B, Lehmann J, Heilmann J, Zhang Y, Decker M. Design and synthesis of tacrine-ferulic acid hybrids as multi-potent anti-Alzheimer drug candidates. *Bioorg Med Chem Lett* 2008; 18:2905–2909.
 206. Heilmann J, Calis I, Kirmizibekmez H, Schuhly W, Harput S, Sticher O. Radical scavenger activity of phenylethanoid glycosides in FMLP stimulated human polymorphonuclear leukocytes: Structure-activity relationships. *Planta Med* 2000;66:746–748.
 207. Trombino S, Serini S, Di Nicuolo F, Celleno L, Ando S, Picci N, Calviello G, Palozza P. Antioxidant effect of ferulic acid in isolated membranes and intact cells: Synergistic interactions with alpha-tocopherol, beta-carotene, and ascorbic acid. *J Agric Food Chem* 2004;52:2411–2420.
 208. Yan JJ, Cho JY, Kim HS, Kim KL, Jung JS, Huh SO, Suh HW, Kim YH, Song DK. Protection against beta-amyloid peptide toxicity in vivo with long-term administration of ferulic acid. *Br J Pharmacol* 2001;133:89–96.
 209. Davalos A, Gomez-Cordoves C, Bartolome B. Extending applicability of the oxygen radical absorbance capacity (ORAC-fluorescein) assay. *J Agric Food Chem* 2004;52:48–54.
 210. Rodriguez-Franco MI, Fernandez-Bachiller MI, Perez C, Hernandez-Ledesma B, Bartolome B. Novel tacrine-melatonin hybrids as dual-acting drugs for Alzheimer disease, with improved acetylcholinesterase inhibitory and antioxidant properties. *J Med Chem* 2006;49:459–462.
 211. Reiter RJ. Oxidative damage in the central nervous system: protection by melatonin. *Prog Neurobiol* 1998;56:359–384.
 212. Jang MH, Jung SB, Lee MH, Kim CJ, Oh YT, Kang I, Kim J, Kim EH. Melatonin attenuates amyloid beta25–35-induced apoptosis in mouse microglial BV2 cells. *Neurosci Lett* 2005;380: 26–31.
 213. Fernandez-Bachiller MI, Perez C, Campillo NE, Paez JA, Gonzalez-Munoz GC, Usan P, Garcia-Palomero E, Lopez MG, Villarroya M, Garcia AG, Martinez A, Rodriguez-Franco MI. Tacrine-melatonin hybrids as multifunctional agents for Alzheimer's disease, with cholinergic, antioxidant, and neuroprotective properties. *Chem Med Chem* 2009;4:828–841.
 214. Spuch C, Antequera D, Isabel Fernandez-Bachiller M, Isabel Rodriguez-Franco M, Carro E. A new tacrine-melatonin hybrid reduces amyloid burden and behavioral deficits in a mouse model of Alzheimer's disease. *Neurotox Res* 2010;17:421–431.
 215. Kurt MA, Davies DC, Kidd M, Duff K, Rolph SC, Jennings KH, Howlett DR. Neurodegenerative changes associated with beta-amyloid deposition in the brains of mice carrying mutant amyloid precursor protein and mutant presenilin-1 transgenes. *Exp Neurol* 2001;171:59–71.
 216. Arce MP, Rodriguez-Franco MI, Gonzalez-Munoz GC, Perez C, Lopez B, Villarroya M, Lopez MG, Garcia AG, Conde S. Neuroprotective and cholinergic properties of multifunctional glutamic acid derivatives for the treatment of Alzheimer's disease. *J Med Chem* 2009;52:7249–7257.
 217. Prokai-Tatrai K, Nguyen V, Zharikova AD, Braddy AC, Stevens SM, Prokai L. Prodrugs to enhance central nervous system effects of the TRH-like peptide pGlu-Glu-Pro-NH₂. *Bioorg Med Chem Lett* 2003;13:1011–1014.
 218. Alper G, Girgin FK, Ozgonul M, Montes G, Ersoz B. MAO inhibitors and oxidant stress in aging brain tissue. *Eur Neuropsychopharmacol* 1999;9:247–252.
 219. Fink DM, Palermo MG, Bores GM, Huger FP, Kurys BE, Merriman MC, Olsen GE, Petko W, O'mMalley GJ. Imino 1,2,3,4-tetrahydrocyclopent[b]indole carbamates as dual inhibitors of acetylcholinesterase and monoamine oxidase. *Bioorg Med Chem Lett* 1996;6:625–630.

220. Bar-Am O, Weinreb O, Amit T, Youdim MB. The novel cholinesterase-monoamine oxidase inhibitor and antioxidant, ladostigil, confers neuroprotection in neuroblastoma cells and aged rats. *J Mol Neurosci* 2009;37:135–145.
221. Available from: <http://www.businesswire.com/news/home/20100414005702/en/Yissum-Pontifax-Clal-Biotechnology-Industries-Invest-9>. 2010.
222. Mattson MP, Chan SL. Dysregulation of cellular calcium homeostasis in Alzheimer's disease: Bad genes and bad habits. *J Mol Neurosci* 2001;17:205–224.
223. Selkoe DJ. Biochemistry of altered brain proteins in Alzheimer's disease. *Annu Rev Neurosci* 1989;12:463–490.
224. Kruman I, Guo Q, Mattson MP. Calcium and reactive oxygen species mediate staurosporine-induced mitochondrial dysfunction and apoptosis in PC12 cells. *J Neurosci Res* 1998;51:293–308.
225. Mattson MP, Cheng B, Davis D, Bryant K, Lieberburg I, Rydel RE. beta-Amyloid peptides destabilize calcium homeostasis and render human cortical neurons vulnerable to excitotoxicity. *J Neurosci* 1992;12:376–389.
226. Cano-Abad MF, Villarroya M, Garcia AG, Gabilan NH, Lopez MG. Calcium entry through L-type calcium channels causes mitochondrial disruption and chromaffin cell death. *J Biol Chem* 2001;276:39695–39704.
227. Leon R, Marco-Contelles J, Garcia AG, Villarroya M. Synthesis, acetylcholinesterase inhibition and neuroprotective activity of new tacrine analogues. *Bioorg Med Chem* 2005;13:1167–1175.
228. Orozco C, de Los Rios C, Arias E, Leon R, Garcia AG, Marco JL, Villarroya M, Lopez MG. ITH4012 (ethyl 5-amino-6,7,8,9-tetrahydro-2-methyl-4-phenylbenzol[1,8]naphthyridine-3-carboxylate), a novel acetylcholinesterase inhibitor with “calcium promotor” and neuroprotective properties. *J Pharmacol Exp Ther* 2004;310:987–994.
229. Marco-Contelles J, Leon R, de Los Rios C, Guglietta A, Terencio J, Lopez MG, Garcia AG, Villarroya M. Novel multipotent tacrine-dihydropyridine hybrids with improved acetylcholinesterase inhibitory and neuroprotective activities as potential drugs for the treatment of Alzheimer's disease. *J Med Chem* 2006;49:7607–7610.
230. Marco-Contelles J, Leon R, de los Rios C, Samadi A, Bartolini M, Andrisano V, Huertas O, Barril X, Luque FJ, Rodriguez-Franco MI, Lopez B, Lopez MG, Garcia AG, Carreiras Mdo C, Villarroya M. Tacripyrines, the first tacrine-dihydropyridine hybrids, as multitarget-directed ligands for the treatment of Alzheimer's disease. *J Med Chem* 2009;52:2724–2732.
231. Martinez-Grau MA, Marco JL. Friedländer reaction on 2-amino-3-cyano-4h-pyrans: synthesis of derivatives of 4h-pyran [2,3-b] quinoline, new tacrine analogues. *Bioorg Med Chem Lett* 1997;7:3165–3170.
232. de los Rios C, Marco JL, Carreiras MD, Chinchon PM, Garcia AG, Villarroya M. Novel tacrine derivatives that block neuronal calcium channels. *Bioorg Med Chem* 2002;10:2077–2088.
233. Marco JL, de los Rios C, Carreiras MC, Banos JE, Badia A, Vivas NM. Synthesis and acetylcholinesterase/butyrylcholinesterase inhibition activity of new tacrine-like analogues. *Bioorg Med Chem* 2001;9:727–732.
234. Marco JL, De Los Rios C, Carreiras MC, Banos JE, Badia A, Vivas NM. Synthesis and acetylcholinesterase/butyrylcholinesterase inhibition activity of 4-amino-2, 3-diaryl-5, 6, 7, 8-tetrahydrofuro(and thieno)[2, 3-b]-quinolines, and 4-amino-5, 6, 7, 8, 9-pentahydro-2, 3-diphenylcyclohepta[e]furo(and thieno)-[2, 3-b]pyridines. *Arch Pharm (Weinheim)* 2002;335:347–353.
235. Marco-Contelles JL, León R, Morales E, Villarroya M, García AG. Synthesis, electrochemical and biological studies onpolyfunctionalized 4-ferrocenyl-4*H*-pyran and 4-ferrocenyl-1,4-dihydropyridine derivatives. *Tetrahedron Lett* 2004;45:5203–5205.
236. Marco JL, de los Rios C, Garcia AG, Villarroya M, Carreiras MC, Martins C, Eleuterio A, Morreale A, Orozco M, Luque FJ. Synthesis, biological evaluation and molecular modelling of diversely functionalized heterocyclic derivatives as inhibitors of acetylcholinesterase/butyrylcholinesterase and modulators of Ca²⁺ channels and nicotinic receptors. *Bioorg Med Chem* 2004;12:2199–2218.

237. Marco-Contelles J, Leon R, Lopez MG, Garcia AG, Villarroya M. Synthesis and biological evaluation of new 4H-pyrano[2,3-b]quinoline derivatives that block acetylcholinesterase and cell calcium signals, and cause neuroprotection against calcium overload and free radicals. *Eur J Med Chem* 2006;41:1464–1469.
238. Leon R, de los Rios C, Marco-Contelles J, Huertas O, Barril X, Luque FJ, Lopez MG, Garcia AG, Villarroya M. New tacrine-dihydropyridine hybrids that inhibit acetylcholinesterase, calcium entry, and exhibit neuroprotection properties. *Bioorg Med Chem* 2008;16:7759–7769.
239. Leon R, de Los Rios C, Marco-Contelles J, Lopez MG, Garcia AG, Villarroya M. Synthesis of 6-amino-1,4-dihydropyridines that prevent calcium overload and neuronal death. *Eur J Med Chem* 2008;43:668–674.
240. Marco-Contelles J, Leon R, de los Rios C, Garcia AG, Lopez MG, Villarroya M. New multi-potent tetracyclic tacrines with neuroprotective activity. *Bioorg Med Chem* 2006;14:8176–8185.
241. Towart R, Kazda S. The cellular mechanism of action of nimodipine (BAY e 9736), a new calcium antagonist [proceedings]. *Br J Pharmacol* 1979;67:409P–410P.
242. Garcia AG, Sala F, Reig JA, Viniegra S, Frias J, Fonteriz R, Gandia L. Dihydropyridine BAY-K-8644 activates chromaffin cell calcium channels. *Nature* 1984;309:69–71.
243. Aliev G, Palacios HH, Walrafen B, Lipsitt AE, Obrenovich ME, Morales L. Brain mitochondria as a primary target in the development of treatment strategies for Alzheimer disease. *Int J Biochem Cell Biol* 2009;41:1989–2004.
244. Lysko PG, Lysko KA, Webb CL, Feuerstein G, Mason PE, Walter MF, Mason RP. Neuroprotective activities of carvedilol and a hydroxylated derivative: Role of membrane biophysical interactions. *Biochem Pharmacol* 1998;56:1645–1656.
245. Howlett DR, George AR, Owen DE, Ward RV, Markwell RE. Common structural features determine the effectiveness of carvedilol, daunomycin and rolitetracycline as inhibitors of Alzheimer beta-amyloid fibril formation. *Biochem J* 1999;343:419–423.
246. Becher PG, Beuchat J, Gademann K, Juttner F. Nostocarboline: isolation and synthesis of a new cholinesterase inhibitor from Nostoc 78-12A. *J Nat Prod* 2005;68:1793–1795.
247. Schott Y, Decker M, Rommelspacher H, Lehmann J. 6-Hydroxy- and 6-methoxy-beta-carbolines as acetyl- and butyrylcholinesterase inhibitors. *Bioorg Med Chem Lett* 2006;16:5840–5843.
248. Rook Y, Schmidtke KU, Gaube F, Schepmann D, Wunsch B, Heilmann J, Lehmann J, Winckler T. Bivalent beta-carbolines as potential multitarget anti-Alzheimer agents. *J Med Chem* 2010;53:3611–3617.
249. Degroot A, Kofalvi A, Wade MR, Davis RJ, Rodrigues RJ, Rebola N, Cunha RA, Nomikos GG. CB1 receptor antagonism increases hippocampal acetylcholine release: Site and mechanism of action. *Mol Pharmacol* 2006;70:1236–1245.
250. Castellano C, Rossi-Arnaud C, Cestari V, Costanzi M. Cannabinoids and memory: Animal studies. *Curr Drug Targets CNS Neurol Disord* 2003;2:389–402.
251. Wolff MC, Leander JD. SR141716A, a cannabinoid CB1 receptor antagonist, improves memory in a delayed radial maze task. *Eur J Pharmacol* 2003;477:213–217.
252. Hikida T, Kitabatake Y, Pastan I, Nakanishi S. Acetylcholine enhancement in the nucleus accumbens prevents addictive behaviors of cocaine and morphine. *Proc Natl Acad Sci USA* 2003;100:6169–6173.
253. Lange JH, Coolen HK, van der Neut MA, Borst AJ, Stork B, Verveer PC, Kruse CG. Design, synthesis, biological properties, and molecular modeling investigations of novel tacrine derivatives with a combination of acetylcholinesterase inhibition and cannabinoid CB1 receptor antagonism. *J Med Chem* 2010;53:1338–1346.
254. Lange JH, Coolen HK, van Stuijvenberg HH, Dijkman JA, Herremans AH, Ronken E, Keizer HG, Tipker K, McCreary AC, Veerman W, Wals HC, Stork B, Verveer PC, den Hartog AP, de Jong NM, Adolfs TJ, Hoogendoorn J, Kruse CG. Synthesis, biological properties, and molecular modeling investigations of novel 3,4-diarylpyrazolines as potent and selective CB(1) cannabinoid receptor antagonists. *J Med Chem* 2004;47:627–643.

255. Lange JH, van Stuivenberg HH, Coolen HK, Adolfs TJ, McCreary AC, Keizer HG, Wals HC, Veerman W, Borst AJ, de Looft W, Verveer PC, Kruse CG. Bioisosteric replacements of the pyrazole moiety of rimonabant: synthesis, biological properties, and molecular modeling investigations of thiazoles, triazoles, and imidazoles as potent and selective CB1 cannabinoid receptor antagonists. *J Med Chem* 2005;48:1823–1838.
256. Fisher A. M1 muscarinic agonists target major hallmarks of Alzheimer's disease—the pivotal role of brain M1 receptors. *Neurodegener Dis* 2008;5:237–240.
257. Fisher A, Brandeis R, Bar-Ner RH, Kliger-Spatz M, Natan N, Sonogo H, Marcovitch I, Pittel Z. AF150(S) and AF267B: M1 muscarinic agonists as innovative therapies for Alzheimer's disease. *J Mol Neurosci* 2002;19:145–153.
258. Sadot E, Gurwitz D, Barg J, Behar L, Ginzburg I, Fisher A. Activation of m1 muscarinic acetylcholine receptor regulates tau phosphorylation in transfected PC12 cells. *J Neurochem* 1996; 66:877–880.
259. Fang L, Jumpertz S, Zhang Y, Appenroth D, Fleck C, Mohr K, Trankle C, Decker M. Hybrid molecules from xanomeline and tacrine: enhanced tacrine actions on cholinesterases and muscarinic M1 receptors. *J Med Chem* 2010;53:2094–2103.
260. Veroff AE, Bodick NC, Offen WW, Sramek JJ, Cutler NR. Efficacy of xanomeline in Alzheimer disease: Cognitive improvement measured using the Computerized Neuropsychological Test Battery (CNTB). *Alzheimer Dis Assoc Disord* 1998;12:304–312.
261. Kogen H, Toda N, Tago K, Marumoto S, Takami K, Ori M, Yamada N, Koyama K, Naruto S, Abe K, Yamazaki R, Hara T, Aoyagi A, Abe Y, Kaneko T. Design and synthesis of dual inhibitors of acetylcholinesterase and serotonin transporter targeting potential agents for Alzheimer's disease. *Org Lett* 2002;4:3359–3362.
262. Toda N, Tago K, Marumoto S, Takami K, Ori M, Yamada N, Koyama K, Naruto S, Abe K, Yamazaki R, Hara T, Aoyagi A, Abe Y, Kaneko T, Kogen H. Design, synthesis and structure-activity relationships of dual inhibitors of acetylcholinesterase and serotonin transporter as potential agents for Alzheimer's disease. *Bioorg Med Chem* 2003;11:1935–1955.
263. Toda N, Kaneko T, Kogen H. Development of an efficient therapeutic agent for Alzheimer's disease: design and synthesis of dual inhibitors of acetylcholinesterase and serotonin transporter. *Chem Pharm Bull (Tokyo)* 2010;58:273–287.
264. Alguacil LF, Perez-Garcia C. Histamine H3 receptor: a potential drug target for the treatment of central nervous system disorders. *Curr Drug Targets CNS Neurol Disord* 2003;2:303–313.
265. Bembenek SD, Keith JM, Letavic MA, Apodaca R, Barbier AJ, Dvorak L, Aluisio L, Miller KL, Lovenberg TW, Carruthers NI. Lead identification of acetylcholinesterase inhibitors-histamine H3 receptor antagonists from molecular modeling. *Bioorg Med Chem* 2008;16:2968–2973.
266. Axe FU, Bembenek SD, Szalma S. Three-dimensional models of histamine H3 receptor antagonist complexes and their pharmacophore. *J Mol Graph Model* 2006;24:456–464.
267. Engelberts I, von Asmuth EJ, van der Linden CJ, Buurman WA. The interrelation between TNF, IL-6, and PAF secretion induced by LPS in an in vivo and in vitro murine model. *Lymphokine Cytokine Res* 1991;10:127–131.
268. Lo CJ, Cryer HG, Fu M, Kim B. Endotoxin-induced macrophage gene expression depends on platelet-activating factor. *Arch Surg* 1997;132:1342–1347.
269. Bate C, Kempster S, Williams A. Platelet-activating factor antagonists protect amyloid-beta damaged neurons from microglia-mediated death. *Neuropharmacology* 2006;51:173–181.
270. Le Texier L, Favre E, Ronzani N, Massicot F, Blavet N, Pirotzky E, Godfroid JJ. Structure-activity relationships in platelet-activating factor (PAF). 8. Tetrahydrofuran derivatives as dual PAF antagonists and acetylcholinesterase inhibitors: anti-acetylcholinesterase activity and comparative SAR. *J Lipid Mediat Cell Signal* 1996;13:207–222.
271. Li J, Huang H, Miezian Ezoulin JM, Gao XL, Massicot F, Dong CZ, Heymans F, Chen HZ. Pharmacological profile of PMS777, a new AChE inhibitor with PAF antagonistic activity. *Int J Neuropsychopharmacol* 2007;10:21–29.

272. Ezoulin MJ, Li J, Wu G, Dong CZ, Ombetta JE, Chen HZ, Massicot F, Heymans F. Differential effect of PMS777, a new type of acetylcholinesterase inhibitor, and galanthamine on oxidative injury induced in human neuroblastoma SK-N-SH cells. *Neurosci Lett* 2005;389: 61–65.
273. Miezian Ezoulin JM, Shao BY, Xia Z, Xie Q, Li J, Cui YY, Wang H, Dong CZ, Zhao YX, Massicot F, Qiu ZB, Heymans F, Chen HZ. Novel piperazine derivative PMS1339 exhibits tri-functional properties and cognitive improvement in mice. *Int J Neuropsychopharmacol* 2009; 12:1409–1419.
274. Bush AI. Drug development based on the metals hypothesis of Alzheimer's disease. *J Alzheimers Dis* 2008;15:223–240.
275. Turner PR, O'Connor K, Tate WP, Abraham WC. Roles of amyloid precursor protein and its fragments in regulating neural activity, plasticity and memory. *Prog Neurobiol* 2003;70:1–32.
276. Cao X, Sudhof TC. Dissection of amyloid-beta precursor protein-dependent transcriptional transactivation. *J Biol Chem* 2004;279:24601–24611.
277. Satpute-Krishnan P, DeGiorgis JA, Bearer EL. Fast anterograde transport of herpes simplex virus: Role for the amyloid precursor protein of alzheimer's disease. *Aging Cell* 2003;2: 305–318.
278. Dillen K, Annaert W. A two decade contribution of molecular cell biology to the centennial of Alzheimer's disease: Are we progressing toward therapy? *Int Rev Cytol* 2006;254:215–300.
279. Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, Wolfe MS, Rowan MJ, Selkoe DJ. Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature* 2002;416:535–539.
280. Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, Choi EY, Nairn AC, Salter MW, Lombroso PJ, Gouras GK, Greengard P. Regulation of NMDA receptor trafficking by amyloid-beta. *Nat Neurosci* 2005;8:1051–1058.
281. <http://clinicaltrials.gov/ct2/show/NCT00621010?term=ZPQ-21166&rank=1>. Accessed on May 27, 2010.
282. Moser PC, Bergis OE, Jegham S, Lochead A, Duconseille E, Terranova JP, Caille D, Berque-Bestel I, Lezoualc'h F, Fischmeister R, Dumuis A, Bockaert J, George P, Soubrie P, Scatton B. SL65.0155, a novel 5-hydroxytryptamine(4) receptor partial agonist with potent cognition-enhancing properties. *J Pharmacol Exp Ther* 2002;302:731–741.
283. Mohler EG, Shacham S, Noiman S, Lezoualc'h F, Robert S, Gastineau M, Rutkowski J, Marantz Y, Dumuis A, Bockaert J, Gold PE, Ragozzino ME. VRX-03011, a novel 5-HT4 agonist, enhances memory and hippocampal acetylcholine efflux. *Neuropharmacology* 2007;53:563–573.
284. http://www.neuronetrix.com/investors_files/Neuronetrix%20Business%20Plan.pdf. 2011.
285. Short Term Effects of PRX-03140 in Patients With Mild Alzheimer's Disease Being Treated With Aricept; <http://clinicaltrials.gov/ct2/show/NCT00384423?term=PRX-03140&rank=1>. Accessed on February 15, 2011. 2011.
286. Weggen S, Eriksen JL, Das P, Sagi SA, Wang R, Pietrzik CU, Findlay KA, Smith TE, Murphy MP, Bulter T, Kang DE, Marquez-Sterling N, Golde TE, Koo EH. A subset of NSAIDs lower amyloidogenic Abeta42 independently of cyclooxygenase activity. *Nature* 2001;414: 212–216.
287. Jaradat MS, Wongsud B, Phornchirasilp S, Rangwala SM, Shams G, Sutton M, Romstedt KJ, Noonan DJ, Feller DR. Activation of peroxisome proliferator-activated receptor isoforms and inhibition of prostaglandin H(2) synthases by ibuprofen, naproxen, and indomethacin. *Biochem Pharmacol* 2001;62:1587–1595.
288. Lehmann JM, Lenhard JM, Oliver BB, Ringold GM, Kliewer SA. Peroxisome proliferator-activated receptors alpha and gamma are activated by indomethacin and other non-steroidal anti-inflammatory drugs. *J Biol Chem* 1997;272:3406–3410.
289. Landreth G, Jiang Q, Mandrekar S, Heneka M. PPARgamma agonists as therapeutics for the treatment of Alzheimer's disease. *Neurotherapeutics* 2008;5:481–489.

290. Hieke M, Ness J, Steri R, Dittrich M, Greiner C, Werz O, Baumann K, Schubert-Zsilavecz M, Weggen S, Zettl H. Design, synthesis, and biological evaluation of a novel class of gamma-secretase modulators with PPARgamma activity. *J Med Chem* 2010;53:4691–4700.
291. Yang D, Soulier JL, Sicsic S, Mathe-Allainmat M, Bremont B, Croci T, Cardamone R, Aureggi G, Langlois M. New esters of 4-amino-5-chloro-2-methoxybenzoic acid as potent agonists and antagonists for 5-HT4 receptors. *J Med Chem* 1997;40:608–621.
292. Russo O, Cachard-Chastel M, Riviere C, Giner M, Soulier JL, Berthouze M, Richard T, Monti JP, Sicsic S, Lezoualc'h F, Berque-Bestel I. Design, synthesis, and biological evaluation of new 5-HT4 receptor agonists: Application as amyloid cascade modulators and potential therapeutic utility in Alzheimer's disease. *J Med Chem* 2009;52:2214–2225.
293. Gandy S, Greengard P. Processing of Alzheimer A beta-amyloid precursor protein: Cell biology, regulation, and role in Alzheimer disease. *Int Rev Neurobiol* 1994;36:29–50.
294. Chuang DM, Leng Y, Marinova Z, Kim HJ, Chiu CT. Multiple roles of HDAC inhibition in neurodegenerative conditions. *Trends Neurosci* 2009;32:591–601.
295. Kozikowski AP, Chen Y, Subhasish T, Lewin NE, Blumberg PM, Zhong Z, D'Annibale MA, Wang WL, Shen Y, Langley B. Searching for disease modifiers-PKC activation and HDAC inhibition—a dual drug approach to Alzheimer's disease that decreases Abeta production while blocking oxidative stress. *Chem Med Chem* 2009;4:1095–1105.
296. Kozikowski AP, Nowak I, Petukhov PA, Etcheberrigaray R, Mohamed A, Tan M, Lewin N, Hennings H, Pearce LL, Blumberg PM. New amide-bearing benzolactam-based protein kinase C modulators induce enhanced secretion of the amyloid precursor protein metabolite sAPPalpha. *J Med Chem* 2003;46:364–373.
297. Tsuji K, Spears GW, Nakamura K, Tojo T, Seki N, Sugiyama A, Matsuo M. Synthesis and antinephritic activities of quinoline-3-carboxamides and related compounds. *Bioorg Med Chem Lett* 2002;12:85–88.
298. Rezai-Zadeh K, Shytle D, Sun N, Mori T, Hou H, Jeanniton D, Ehrhart J, Townsend K, Zeng J, Morgan D, Hardy J, Town T, Tan J. Green tea epigallocatechin-3-gallate (EGCG) modulates amyloid precursor protein cleavage and reduces cerebral amyloidosis in Alzheimer transgenic mice. *J Neurosci* 2005;25:8807–8814.
299. Ono K, Condrón MM, Ho L, Wang J, Zhao W, Pasinetti GM, Teplow DB. Effects of grape seed-derived polyphenols on amyloid beta-protein self-assembly and cytotoxicity. *J Biol Chem* 2008;283:32176–32187.
300. Ehrnhoefer DE, Bieschke J, Boeddrich A, Herbst M, Masino L, Lurz R, Engemann S, Pastore A, Wanker EE. EGCG redirects amyloidogenic polypeptides into unstructured, off-pathway oligomers. *Nat Struct Mol Biol* 2008;15:558–566.
301. Jonsson S, Andersson G, Fex T, Fristedt T, Hedlund G, Jansson K, Abramo L, Fritzson I, Pekarski O, Runstrom A, Sandin H, Thuvesson I, Bjork A. Synthesis and biological evaluation of new 1,2-dihydro-4-hydroxy-2-oxo-3-quinolinecarboxamides for treatment of autoimmune disorders: Structure-activity relationship. *J Med Chem* 2004;47:2075–2088.
302. Kojima Y, Hashiguchi H, Hashimoto T, Tsuji S, Shoji H, Kazuyama Y. Recurrent herpes simplex virus type 2 meningitis: A case report of Mollaret's meningitis. *Jpn J Infect Dis* 2002;55:85–88.
303. Tsuji A, Ishiko A, Hirose M, Takasaki T, Ikeda N. Maternity testing using mitochondrial DNA analysis. *Fukuoka Igaku Zasshi* 2002;93:85–90.
304. Tsuji K. Effect of (+/-)-pindolol on the central 5-HT1A receptor by the use of in vivo microdialysis and hippocampal slice preparations. *Nihon Shinkei Seishin Yakurigaku Zasshi* 2002;22:85–95.
305. Yamauchi J, Hirasawa A, Miyamoto Y, Kokubu H, Nishii H, Okamoto M, Sugawara Y, Tsujimoto G, Itoh H. Role of Dbp's big sister in the anti-mitogenic pathway from alpha1B-adrenergic receptor to c-Jun N-terminal kinase. *Biochem Biophys Res Commun* 2002;296:85–92.
306. Jansson K, Fristedt T, Olsson A, Svensson B, Jonsson S. Synthesis and reactivity of laquinimod, a quinoline-3-carboxamide: intramolecular transfer of the enol proton to a nitrogen atom as a plausible mechanism for ketene formation. *J Org Chem* 2006;71:1658–1667.

307. Naito Y, Yoshikawa T, Tanigawa T, Sakurai K, Yamasaki K, Uchida M, Kondo M. Hydroxyl radical scavenging by rebamipide and related compounds: electron paramagnetic resonance study. *Free Radic Biol Med* 1995;18:117–123.
308. Detsi A, Bouloubasi D, Prousis KC, Koufaki M, Athanasellis G, Melagraki G, Afantitis A, Igglessi-Markopoulou O, Kontogiorgis C, Hadjipavlou-Litina DJ. Design and synthesis of novel quinolinone-3-aminoamides and their alpha-lipoic acid adducts as antioxidant and anti-inflammatory agents. *J Med Chem* 2007;50:2450–2458.
309. Alonso AC, Li B, Grundke-Iqbal I, Iqbal K. Mechanism of tau-induced neurodegeneration in Alzheimer disease and related tauopathies. *Curr Alzheimer Res* 2008;5:375–384.
310. Khatoon S, Grundke-Iqbal I, Iqbal K. Brain levels of microtubule-associated protein tau are elevated in Alzheimer's disease: A radioimmuno-slot-blot assay for nanograms of the protein. *J Neurochem* 1992;59:750–753.
311. Ballatore C, Lee VM, Trojanowski JQ. Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. *Nat Rev Neurosci* 2007;8:663–672.
312. Takashima A. Drug development targeting the glycogen synthase kinase-3beta (GSK-3beta)-mediated signal transduction pathway: Role of GSK-3beta in adult brain. *J Pharmacol Sci* 2009;109:174–178.
313. Bartolini M, Andrisano V. Strategies for the inhibition of protein aggregation in human diseases. *Chembiochem* 2010;11:1018–1035.
314. Oumata N, Bettayeb K, Ferandin Y, Demange L, Lopez-Giral A, Goddard ML, Myrianthopoulos V, Mikros E, Flajolet M, Greengard P, Meijer L, Galons H. Roscovitine-derived, dual-specificity inhibitors of cyclin-dependent kinases and casein kinases 1. *J Med Chem* 2008;51:5229–5242.
315. Bukanov NO, Smith LA, Klinger KW, Ledbetter SR, Ibraghimov-Beskrovnaya O. Long-lasting arrest of murine polycystic kidney disease with CDK inhibitor roscovitine. *Nature* 2006;444:949–952.
316. Meijer L, Raymond E. Roscovitine and other purines as kinase inhibitors. From starfish oocytes to clinical trials. *Acc Chem Res* 2003;36:417–425.
317. Efficacy study of oral seliciclib to treat non-small cell lung cancer. <http://clinicaltrials.gov/ct2/show/NCT00372073?term=Seliciclib&rank=2> Accessed on December 06, 2010.
318. A study of oral sapacitabine and oral seliciclib in patients with advanced solid tumors. <http://clinicaltrials.gov/ct2/show/NCT00999401?term=roscovitine&rank=1> Accessed on December 06, 2010.
319. Bain J, McLauchlan H, Elliott M, Cohen P. The specificities of protein kinase inhibitors: An update. *Biochem J* 2003;371:199–204.
320. Haesslein JL, Jullian N. Recent advances in cyclin-dependent kinase inhibition. Purine-based derivatives as anti-cancer agents. Roles and perspectives for the future. *Curr Top Med Chem* 2002;2:1037–1050.
321. Reinhardt J, Ferandin Y, Meijer L. Purification of CK1 by affinity chromatography on immobilised axin. *Protein Expr Purif* 2007;54:101–109.
322. Flajolet M, He G, Heiman M, Lin A, Nairn AC, Greengard P. Regulation of Alzheimer's disease amyloid-beta formation by casein kinase I. *Proc Natl Acad Sci USA* 2007;104:4159–4164.
323. Martinez A. Preclinical efficacy on GSK-3 inhibitors: towards a future generation of powerful drugs. *Med Res Rev* 2008;28:773–796.
324. <http://clinicaltrials.gov/ct2/results?term=Noscira>; Accessed on May 30, 2010.
325. Luna-Medina R, Cortes-Canteli M, Sanchez-Galiano S, Morales-Garcia JA, Martinez A, Santos A, Perez-Castillo A. NP031112, a thiadiazolidinone compound, prevents inflammation and neurodegeneration under excitotoxic conditions: Potential therapeutic role in brain disorders. *J Neurosci* 2007;27:5766–5776.
326. Rogers JT, Randall JD, Cahill CM, Eder PS, Huang X, Gunshin H, Leiter L, McPhee J, Sarang SS, Utsuki T, Greig NH, Lahiri DK, Tanzi RE, Bush AI, Giordano T, Gullans SR. An

- iron-responsive element type II in the 5'-untranslated region of the Alzheimer's amyloid precursor protein transcript. *J Biol Chem* 2002;277:45518–45528.
327. Barnham KJ, Bush AI. Metals in Alzheimer's and Parkinson's diseases. *Curr Opin Chem Biol* 2008;12:222–228.
 328. Bannerman RM, Callender ST, Williams DL. Effect of desferrioxamine and D.T.P.A. in iron overload. *Br Med J* 1962;2:1573–1577.
 329. Ritchie CW, Bush AI, Mackinnon A, Macfarlane S, Mastwyk M, MacGregor L, Kiers L, Cherny R, Li QX, Tammer A, Carrington D, Mavros C, Volitakis I, Xilinas M, Ames D, Davis S, Beyreuther K, Tanzi RE, Masters CL. Metal-protein attenuation with iodochlorhydroxyquin (clioquinol) targeting Abeta amyloid deposition and toxicity in Alzheimer disease: a pilot phase 2 clinical trial. *Arch Neurol* 2003;60:1685–1691.
 330. Charkoudian LK, Pham DM, Franz KJ. A pro-chelator triggered by hydrogen peroxide inhibits iron-promoted hydroxyl radical formation. *J Am Chem Soc* 2006;128:12424–12425.
 331. Kalinowski DS, Richardson DR. The evolution of iron chelators for the treatment of iron overload disease and cancer. *Pharmacol Rev* 2005;57:547–583.
 332. Schugar H, Green DE, Bowen ML, Scott LE, Storr T, Bohmerle K, Thomas F, Allen DD, Lockman PR, Merkel M, Thompson KH, Orvig C. Combating Alzheimer's disease with multifunctional molecules designed for metal passivation. *Angew Chem Int Ed Engl* 2007;46:1716–1718.
 333. el-Jammal A, Howell PL, Turner MA, Li N, Templeton DM. Copper complexation by 3-hydroxypyridin-4-one iron chelators: structural and iron competition studies. *J Med Chem* 1994;37:461–466.
 334. Kontoghiorghes GJ. New concepts of iron and aluminium chelation therapy with oral L1 (deferiprone) and other chelators. A review. *Analyst* 1995;120:845–851.
 335. Storr T, Scott LE, Bowen ML, Green DE, Thompson KH, Schugar HJ, Orvig C. Glycosylated tetrahydrosalens as multifunctional molecules for Alzheimer's therapy. *Dalton Trans* 2009;16:3034–3043.
 336. Zheng H, Youdim MB, Fridkin M. Site-activated multifunctional chelator with acetylcholinesterase and neuroprotective-neurorestorative moieties for Alzheimer's therapy. *J Med Chem* 2009;52:4095–4098.
 337. Amit T, Avramovich-Tirosh Y, Youdim MB, Mandel S. Targeting multiple Alzheimer's disease etiologies with multimodal neuroprotective and neurorestorative iron chelators. *FASEB J* 2008;22:1296–1305.
 338. Zheng H, Youdim MBH, Fridkin M. Selective acetylcholinesterase inhibitor activated by acetylcholinesterase releases an active chelator with neurorescuing and anti-amyloid activities. *ACS Chem Neurosci* 2010;1:737–746.
 339. Rodriguez-Rodriguez C, Sanchez de Groot N, Rimola A, Alvarez-Larena A, Lloveras V, Vidal-Gancedo J, Ventura S, Vendrell J, Sodupe M, Gonzalez-Duarte P. Design, selection, and characterization of thioflavin-based intercalation compounds with metal chelating properties for application in Alzheimer's disease. *J Am Chem Soc* 2009;131:1436–1451.
 340. Hinds SS, Mancino AM, Braymer JJ, Liu Y, Vivekanandan S, Ramamoorthy A, Lim MH. Small molecule modulators of copper-induced Abeta aggregation. *J Am Chem Soc* 2009;131:16663–16665.
 341. Newberg AB, Wintering NA, Plossl K, Hochold J, Stabin MG, Watson M, Skovronsky D, Clark CM, Kung MP, Kung HF. Safety, biodistribution, and dosimetry of 123I-IMPY: a novel amyloid plaque-imaging agent for the diagnosis of Alzheimer's disease. *J Nucl Med* 2006;47:748–754.
 342. Kung HF, Kung MP, Zhuang ZP, Hou C, Lee CW, Plossl K, Zhuang B, Skovronsky DM, Lee VM, Trojanowski JQ. Iodinated tracers for imaging amyloid plaques in the brain. *Mol Imaging Biol* 2003;5:418–426.
 343. Lee MS, Kao SC, Lemere CA, Xia W, Tseng HC, Zhou Y, Neve R, Ahljianian MK, Tsai LH. APP processing is regulated by cytoplasmic phosphorylation. *J Cell Biol* 2003;163:83–95.

344. Phiel CJ, Wilson CA, Lee VM, Klein PS. GSK-3 α regulates production of Alzheimer's disease amyloid-beta peptides. *Nature* 2003;423:435–439.
345. Scott LE, Orvig C. Medicinal inorganic chemistry approaches to passivation and removal of aberrant metal ions in disease. *Chem Rev* 2009;109:4885–4910.
346. Kim J, Lee HJ, Lee KW. Naturally occurring phytochemicals for the prevention of Alzheimer's disease. *J Neurochem* 2010;112:1415–1430.
347. Choi RC, Zhu JT, Leung KW, Chu GK, Xie HQ, Chen VP, Zheng KY, Lau DT, Dong TT, Chow PC, Han YF, Wang ZT, Tsim KW. A flavonol glycoside, isolated from roots of *Panax notoginseng*, reduces amyloid-beta-induced neurotoxicity in cultured neurons: Signaling transduction and drug development for Alzheimer's disease. *J Alzheimers Dis* 2010;19:795–811.
348. Zhao B. Natural antioxidants protect neurons in Alzheimer's disease and Parkinson's disease. *Neurochem Res* 2009;34:630–638.
349. Mandel SA, Amit T, Kalfon L, Reznichenko L, Weinreb O, Youdim MB. Cell signaling pathways and iron chelation in the neurorestorative activity of green tea polyphenols: Special reference to epigallocatechin gallate (EGCG). *J Alzheimers Dis* 2008;15:211–222.
350. Man SC, Durairajan SS, Kum WF, Lu JH, Huang JD, Cheng CF, Chung V, Xu M, Li M. Systematic review on the efficacy and safety of herbal medicines for Alzheimer's disease. *J Alzheimers Dis* 2008;14:209–223.
351. Keller TH, Pichota A, Yin Z. A practical view of “druggability”. *Curr Opin Chem Biol* 2006;10:357–361.

Rafael León graduated in chemistry from the University of Castilla la Mancha in 2001. In 2002, he joined the groups of Prof. Antonio G. García at the Department of Pharmacology, Autónoma University of Madrid and Prof. José Marco-Contelles at CSIC. He obtained his Ph.D. in Organic Chemistry in 2006 with honors. His main research focused on drug development and medicinal chemistry related to neurodegenerative diseases. After a year in industry, he joined UAM as a Postdoctoral Fellow, where he was involved in the pharmacological study of the neuronal nicotinic receptor and in an Alzheimer's disease drug development project. Shortly after he moved to the University of Victoria, Canada, as a Postdoctoral Fellow in the group of Dr. Fraser Hof where he increased his knowledge in medicinal chemistry. Currently, he is a Marie Curie Fellow at the Department of chemistry of the Cambridge University, under the supervision of Dr. Matthew J. Gaunt. He is developing new chemical strategies for enantioselective organic chemistry. He has been related in 13 research projects and has published more than 20 research articles.

Antonio G. García holds an M.D. and Ph.D. from Universidad Complutense de Madrid (Spain) 1970. He received postdoctoral training at the State University of New York, Health Science Center, 1971–1974. He is a Professor at the Pharmacology & Therapeutics department, Medicine School, Universidad Autónoma de Madrid (Spain) and an author of 304 papers in international journals. He founded the foundations “Fundación Teófilo Hernando” and “Fundación de Estudios Médicos de Molina de Segura,” and also four journals on medical education (in Spanish): “SNC Pharmacology,” “Pharmacotherapy,” “Drug Prescription,” and “Topics in Pharmacology and Therapeutics.” He is also a founder and first chairman of Institute “Teófilo Hernando” for Drug Research and Development, Autónoma University of Madrid, Spain. He was honored with the price Severo Ochoa, Spain, the Gold Medal of the Government of Region de Murcia, Spain, and is Doctor Honoris Causa by Universidad La Laguna, Spain.

José Marco Contelles studied chemistry at the Universidad Complutense de Madrid (UCM) (graduating with honors), where he obtained his Ph.D. under Professor Benjamin Rodríguez's supervision (Instituto de Química Orgánica, Consejo Superior de Investigaciones Científicas, CSIC) in 1984. After working as a post-doctoral fellow for 2 years under Dr. H.-P. Husson (Institut de Chimie de Substance Naturelles, CNRS, Gif-sur-Yvette, France) (CNRS methods in asymmetric synthesis) (1984–1985), he was an Associate Researcher under Professor Wolfgang Oppolzer (Département de Chimie Organique, Genève, Suisse) (aldol reaction) (1986), and a visiting Professor at the Department of Chemistry, Duke University, North Carolina working with Professor Fraser-Reid (free radical chemistry; annulated furanoses; formal total synthesis of phyllathocin) (1988–1989). In 1986, he was appointed as an Associate Researcher, and in 1992 he was promoted to Research Scientist in the CSIC (Spain). Since 2004, Dr. Marco-Contelles is Profesor de Investigación (Senior Research Scientist) at CSIC. He has been a Invited Professor at the Université Pierre et Marie Curie, Paris VI (2000), at the Université Jules Verne-Picardie (Amiens, France) (2003–2005), and at the Okayama University (Faculty of Engineering) (2003). In 2002, he was awarded with the French-Spanish award of the French Chemical Society. His main scientific interests include carbohydrate, free radical and heterocyclic chemistry (synthesis and biological evaluation of molecules for neurodegenerative diseases, and antiviral compounds), as well as transition metal promoted cycloisomerization of polyunsaturated precursors.