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Four-Photon Excitation of 2,2'-Dimethyl-*p*-terphenyl†

Ignacy Gryczynski, Grzegorz Piszczek, Zygmunt Gryczynski, and Joseph R. Lakowicz*

University of Maryland Baltimore, Center for Fluorescence Spectroscopy, Department of Biochemistry and Molecular Biology, 725 West Lombard Street, Baltimore, Maryland 21201

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We report the emission spectra, intensity decays, and anisotropy decay of 2,2'-dimethyl-*p*-terphenyl (DMT) with four-photon excitation. When excited with a fs Ti:Sapphire laser the emission intensity of DMT was found to depend on the third power of the incident intensity for excitation of 783 nm, and on the fourth power of the incident intensity for excitation at 882 nm. Surprisingly, at the highest value incident power, the emission intensity for four-photon excitation was about 10-fold less than with three-photon excitation. The emission spectra, intensity decays and correlation times were found to be identical for three- and four-photon excitation. However, the fundamental anisotropy (r_0) of DMT depended on the mode of excitation. To the best of our knowledge, the r_0 value of 0.70 is the highest ever observed for an isotropic solution. These results suggest that four-photon excitation can be used with red-NIR lasers to obtain excitation of UV-absorbing chromophores.

Introduction

Multiphoton absorption processes can be used to characterize the excited states of organic molecules.^{1–2} The interest in multiphoton excitation increased dramatically after it was shown that two-photon imaging was possible with laser scanning fluorescence microscopy.^{3–4} Since this time there has been intense activity in using multiphoton excitation in fluorescence spectroscopy.⁵ Two-photon excitation has been used to characterize biological macromolecules^{6–9} and tissues,^{10–12} and in biochemical assays.^{13–14} Simultaneously, the use of two-photon imaging was implemented by many other laboratories.^{15–17} Subsequently, three-photon excitation was shown to be useful in fluorescence spectroscopy^{18–20} and imaging.^{21–23} Additionally, there is an ongoing effort to design and synthesize fluorophores with improved two-photon cross sections for absorption.^{24–28}

In the present report we describe the first observation of four-photon excitation. We examine the UV-absorbing chromophore 2,2'-dimethyl-*p*-terphenyl (DMT) which display a maximum absorbance near 250 nm, with the absorption ending near 300 nm. When excited with the 882 nm output of a fs Ti:Sapphire laser the emissive intensity depended on the fourth power of the incident intensity. The emission was demonstrated to be due to a direct four-photon process based on the steady state and time-resolved anisotropies. These results suggest that four-photon excitation can be useful for the study of UV-absorbing species.

Theory

It is useful to briefly review the theory for anisotropies with multiphoton excitation. The emission anisotropy depends on the extent of excitation photoselection and the angle between the excitation and emission transition moments (β). For z -axis polarized one-photon excitation the anisotropy is given by^{29–30}

$$r_0(\theta, \beta) = \left(\frac{3}{2} \langle \cos^2 \theta \rangle - \frac{1}{2}\right) \left(\frac{3}{2} \cos^2 \beta - \frac{1}{2}\right) \quad (1)$$

where θ is the angle from the z -axis, and β is the angle between the absorption and emission transition moments. The subscript zero indicates the absence of rotational diffusion during the excited-state lifetime or the time-zero anisotropy. The average value of $\cos^2 \theta$ depends on the type of photoselection and is given by excitation this distribution is given by

$$\langle \cos^2 \theta \rangle = \frac{\int_0^{\pi/2} \cos^2 \theta f_i(\theta) d\theta}{\int_0^{\pi/2} f_i(\theta) d\theta} \quad (2)$$

The term $f_i(\theta)$ is the directional distribution of the excited state.²⁹ For one-photon excitation this distribution is given by

$$f_1(\theta) = \cos^2 \theta \sin \theta \quad (3)$$

and for one-photon excitation eq 1 becomes

$$r_{01}(\beta) = \frac{2}{5} \left(\frac{3}{2} \cos^2 \beta - \frac{1}{2}\right) \quad (4)$$

The factor of 2/5 originates with $\cos^2 \theta$ or one-photon photoselection (eq 3). For collinear transitions ($\beta = 0$) the fundamental anisotropy (r_{01}) without rotational diffusion is 0.40. For nonzero values of β the anisotropy ranges from 0.40 to -0.20 (Table 1).

The anisotropy expected for multi-photon excitation can be calculated using

$$f_i(\theta) = \cos^{2i} \theta \sin \theta \quad (5)$$

where i indicated the number of simultaneous absorbed photons. Substitution into eq 1 yields

$$r_{0i}(\beta) = \frac{4}{7} \left(\frac{3}{2} \cos^2 \beta - \frac{1}{2}\right) \quad (6)$$

* Corresponding author.

† Abbreviations: DMT, 2,2'-dimethyl-*p*-terphenyl; FD, frequency-domain; 1-, 2-, 3-, and 4-PE, one-, two-, three-, or four-photon excitation; PPO, 2,5-diphenyloxazole.

TABLE 1: Fundamental Anisotropies for One-, Two-, and Three-Photon Excitation

| β (degrees) | one-photon r_{01} | two-photon r_{02} | three-photon r_{03} | four-photon r_{04} |
|----------------------|------------------------|------------------------|--------------------------|-------------------------|
| 0 | 0.40 | 0.57 | 0.66 ^a | 0.73 |
| 45 | 0.10 | 0.14 | 0.17 | 0.18 |
| 54.7 | 0.00 | 0.00 | 0.00 | 0.00 |
| 90 | -0.20 | -0.29 | -0.33 | -0.36 |

$$r_{03}(\beta) = \frac{2}{3} \left(\frac{3}{2} \cos^2 \beta - \frac{1}{2} \right) \quad (7)$$

$$r_{04}(\beta) = \frac{8}{11} \left(\frac{3}{2} \cos^2 \beta - \frac{1}{2} \right) \quad (8)$$

For any number (i) of simultaneous absorption events the value of $\langle \cos^2 \theta \rangle$ is given by

$$\langle \cos^2 \theta \rangle = \frac{2i + 1}{2i + 3} \quad (9)$$

so that for $\beta = 0$

$$r_{0i} = \frac{2i}{2i + 3} \quad (10)$$

The anisotropy values for multiphoton excitation and for various values of β are given in Table 1. For collinear transitions multiphoton excitation is expected to result in a more highly oriented excited-state population than for one-photon excitation. Observation of a larger anisotropy than possible for three- or two-photon excitation provides strong evidence for four-photon excitation. The description in eqs 1–10 assumes that all transitions involve matrix elements with a single transition dipole orientation. For completeness we note that the anisotropy for two-photon excitation can be as high as 0.61.^{31–32} However, the simple theory described above is adequate for interpretation of these initial results.

Materials and Methods

2,2'-Dimethyl-*p*-terphenyl (DMT) was from Lambda Physics and was used without further purification. The solvents cyclohexane (Aldrich), hexadecane (Aldrich), and triacetin (Becker) were of spectroscopic grade. The concentration of DMT was 5×10^{-4} M. Emission spectra were recorded on a SLM 8000 spectrofluorometer with 10 nm band-pass. Solutions were in equilibrium with air and stirred during the measurements with Teflon-coated magnets.

Multiphoton excitation was accomplished with the fundamental output of Tsunami Ti:Sapphire laser from Spectra Physics, Inc. The pulse width was near 80 fs. The repetition rate of 80 MHz was held fixed by the lock-to-clock accessory. The Ti:Sapphire output (770 to 900 nm) was focused on the sample with a best form 2 cm confocal length lens. The beam waist was estimated to have a diameter of about 20 μ m. The average excitation power was 0.5 W. This condition results in a pulse peak intensity of 2×10^{28} photon $\text{cm}^{-2} \text{s}^{-1}$. For time-resolved measurements the emission was isolated with a combination of BG-39 and UG-11 filters transmitting from 320 to 370 (CVI, Inc.). For all measurements, the background signals from solvent references were less than 1.0%. All intensity and intensity decay measurements were performed using magic angle conditions, which is excitation with vertically polarized light and measurement of the emission through a polarizer oriented 54.7° from the vertical. Under these conditions the intensity and intensity decays are not affected by Brownian motion or

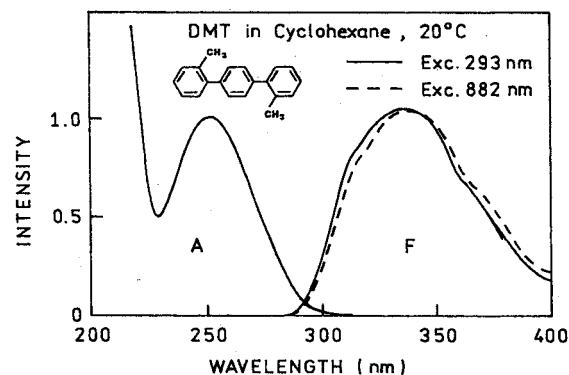


Figure 1. Absorption and emission spectra of DMT in cyclohexane. Also shown (— —) is the emission spectrum with 884 nm excitation, which revealed a four-photon process.

the anisotropy decay of the sample. For measurements of the dependence of the emission on laser intensity, the peak power was attenuated with neutral density filters. To avoid any effect of widening the laser pulses by the filters, a single filter of the same design and thickness but varying optical density was used for the intensity measurements at various peak powers.

Frequency-domain intensity and anisotropy decays were obtained using the instrumentation described previously.^{33–34} The data were analyzed in terms of single exponential models

$$I(t) = I_0 \exp(-t/\tau_i) \quad (11)$$

$$r(t) = r_{0i} \exp(-t/\theta_i) \quad (12)$$

where τ_i is the decay time and θ_i is the rotational correlation time. The subscript i refers to the number of photons in the excitation process. The FD data were fit as described previously.^{35–36}

Results

Emission spectra of DMT are shown in Figure 1 for one-photon excitation at 293 nm and for excitation at 882 nm. These emission spectra were obtained from the same solution of DMT, using the same instrument, the only difference being the excitation wavelength. The emission spectra are essentially identical for both modes of excitation. We believe the small difference in the emission spectra for one- and four-photon excitation is due to a different spatial distribution for the excited volume. Since the absorption of DMT in cyclohexane is essentially zero above 300 nm, it seems clear that this emission must result from three- or four-photon excitation.

To clarify the nature of the excitation process we examined the effect of attenuating the incident power at 882 nm. A 2-fold decrease in the incident power resulted in an over 15-fold decrease in the emission intensity of DMT (Figure 2). The extent of decrease is in close agreement with that expected for the 2.03-fold attenuation, that is a decrease by a factor of 17 due to the fourth-order dependence of the 2.03-fold attenuation. The dependence on incident power is obviously not in agreement with that expected for 1, 2, or 3-PE. For excitation at 793 nm the emission depended on the incident power cubed (Figure 3). For excitation at 882 nm the emission depended on the fourth power of the incident intensity. We further examined how the power dependence varied with excitation wavelength (Figure 4). As the excitation wavelength changes from 810 to 870 nm the power dependence shifted from the cubed dependence to a fourth-power dependence (Figure 4).

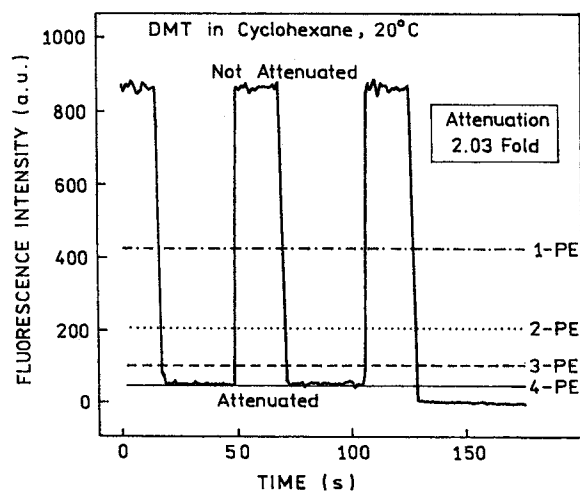


Figure 2. Fluorescence intensity of DMT in cyclohexane with and without a 2.03-fold attenuation of the 882 nm excitation intensity. The horizontal lines are the expectations for 1-PE (---), 2-PE (···), 3-PE (- · -), and 4-PE (—) excitation.

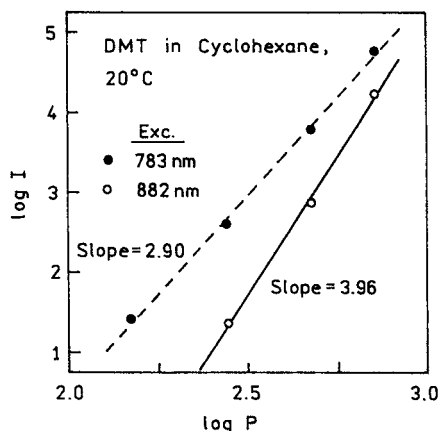


Figure 3. Dependence of the emission intensity of DMT on the excitation intensities at 783 and 882 nm. The power is in mW.

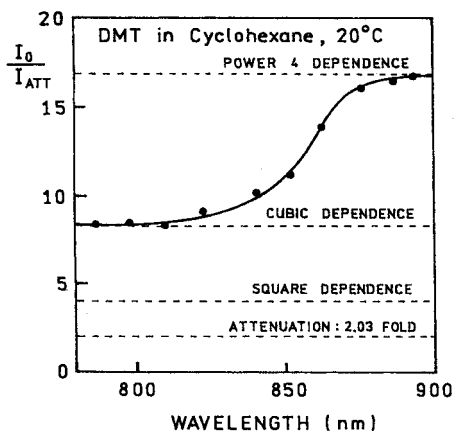


Figure 4. Effect of a 2.03-fold attenuation of the excitation light on the emission intensity of DMT in cyclohexane at 20 °C. The horizontal dashed lines show the expected values of I_0/I_{att} for 1-, 2-, 3-, and 4-photon excitation.

We questioned whether the DMT sample was being damaged by the intense 882 nm excitation needed for 4-PE. Hence we examined the intensity decays measured in the frequency domain (Figure 5). The same single-exponential decay was found for 3-PE at 783 nm and 4-PE at 882 nm. Additionally, the same single exponential decays were measured for 3-PE and 4-PE of DMT in hexadecane and triacetin (Table 2). These results

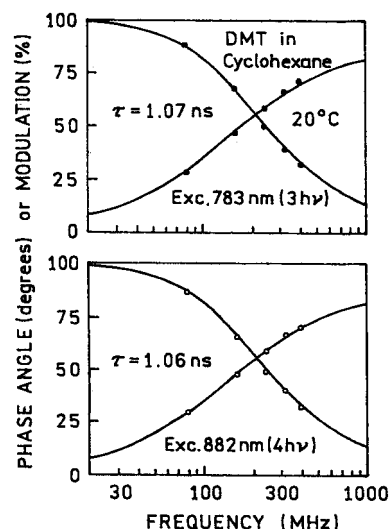


Figure 5. Frequency-domain intensity decays of DMT in cyclohexane with 3-PE at 783 nm and 4-PE at 882 nm.

TABLE 2: Intensity and Anisotropy Decay Parameters of DMT in Cyclohexane, Hexadecane, and Triacetin for Three- and Four-Photon Excitation

| solvent | excitation (nm) | intensity decays τ (ns) | χ_R^2 | anisotropy decays | | |
|-------------|-----------------|------------------------------|------------|-------------------|---------------|------------|
| | | | | r_0 | θ (ns) | χ_R^2 |
| cyclohexane | 783 | 1.07 (0.01) ^a | 1.7 | 0.53 (0.05) | 0.06 (0.03) | 0.9 |
| | 882 | 1.06 (0.01) | 1.8 | 0.57 (0.03) | 0.10 (0.01) | 1.0 |
| hexadecane | 783 | 1.12 (0.01) | 1.6 | 0.61 (0.03) | 0.11 (0.01) | 1.5 |
| | 882 | 1.11 (0.01) | 2.8 | 0.69 (0.03) | 0.11 (0.01) | 1.5 |
| triacetin | 783 | 1.15 (0.01) | 2.5 | 0.61 (0.02) | 0.79 (0.03) | 2.6 |
| | 882 | 1.14 (0.01) | 1.5 | 0.70 (0.02) | 0.78 (0.03) | 1.5 |
| triacetin | 783 | | | 0.61 (0.01) | | |
| | 882 | | | 0.70 (0.01) | 0.78 (0.02) | 2.4 |

^a The numbers in parentheses are the estimated uncertainties.

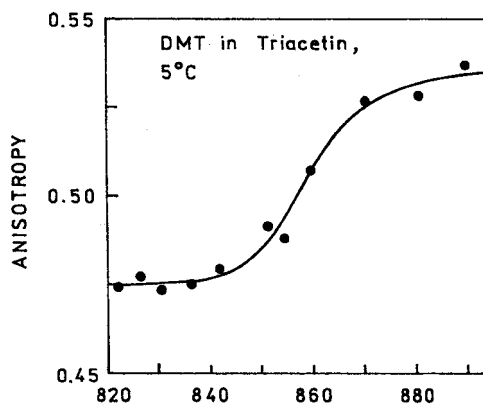


Figure 6. Excitation wavelength-dependent anisotropies of DMT in triacetin at 5 °C.

suggest the DMT is not undergoing photochemical transformation to a different emitting species.

We examined the steady-state anisotropy of DMT in triacetin at 5° where rotational diffusion occurs (Figure 6) and at -50 °C where triacetin forms a rigid glass (Figure 6). Higher anisotropies were observed at lower temperature. The anisotropies of DMT at -55 °C for excitation near 830 and 870 nm are in good agreement with those expected for three- and four-photon excitation with collinear absorption and emission transition moments (Table 1). One can use the anisotropies in Figures 6 and 7 to calculate the rotational correlation time of DMT using the Perrin equation:

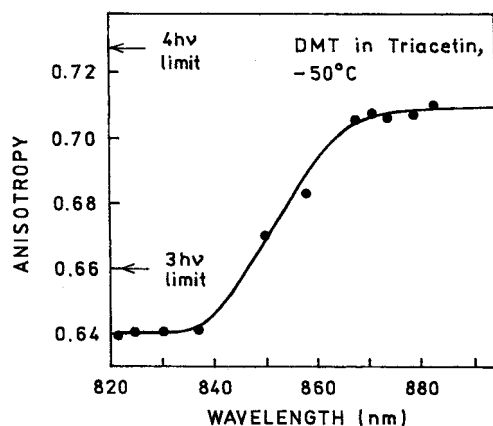


Figure 7. Excitation wavelength-dependent anisotropies of DMT in triacetin at $-50\text{ }^{\circ}\text{C}$.

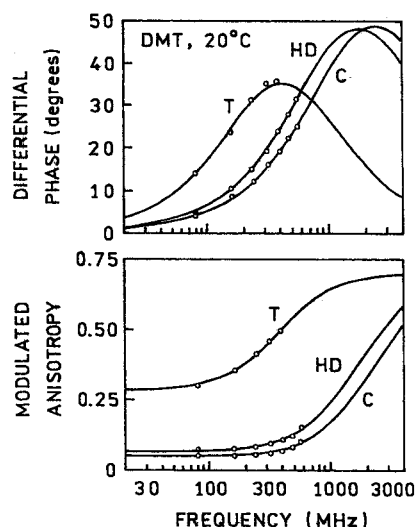


Figure 8. Frequency-domain anisotropy decays of DMT at $20\text{ }^{\circ}\text{C}$ in cyclohexane (C), hexadecane (HD), and triacetin (T) at $20\text{ }^{\circ}\text{C}$.

$$\frac{r_0}{r} = 1 + \tau/\theta \quad (13)$$

where r_0 is the fundamental anisotropy observed at $-50\text{ }^{\circ}\text{C}$, r is the anisotropy at $5\text{ }^{\circ}\text{C}$, and $\tau = 1.81\text{ ns}$ the lifetime at $5\text{ }^{\circ}\text{C}$. This calculation yields a correlation time of 3.3 and 3.4 ns for excitation at 830 and 870 nm, respectively. The closeness of these values suggests that the DMT solution does not undergo local heating at the intensities needed for four-photon excitation. The change in excitation from a three-photon to a four-photon process resulted in a less than 10-fold difference in emission intensities at the highest incident power. Incident powers about 1 W should result in higher emission intensity for 882 nm illumination than for 793 nm illumination (Figure 4).

To further determine if the DMT samples were heated by illumination we measured the frequency-domain anisotropy decay in cyclohexane and triacetin (Figure 8). In all cases nearly the same time-zero anisotropies (r_0) and correlation times were observed for three- and four-photon excitation (Table 2). The slightly different values of r_0 and θ found for DMT in cyclohexane can be due to the shorter correlation times which are more difficult to measure. The fact that the anisotropy decays were the same for 3-PE and 4-PE is demonstrated by the ability to globally fit the FD anisotropy data for 783 and 882 nm excitation (Figure 9) to a single correlation time (Table 2). The closeness of the correlation times for different excitation

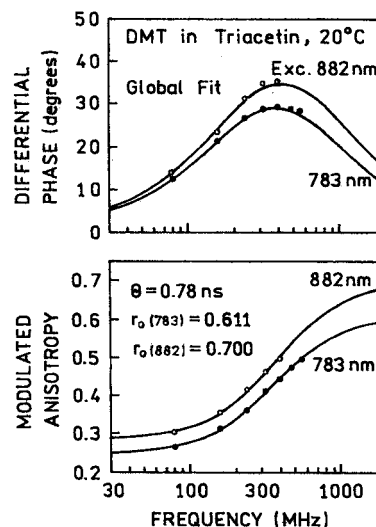


Figure 9. Global analysis of the frequency-domain anisotropy decay of DMT in triacetin with 3-PE at 783 nm and 4-PE at 882 nm.

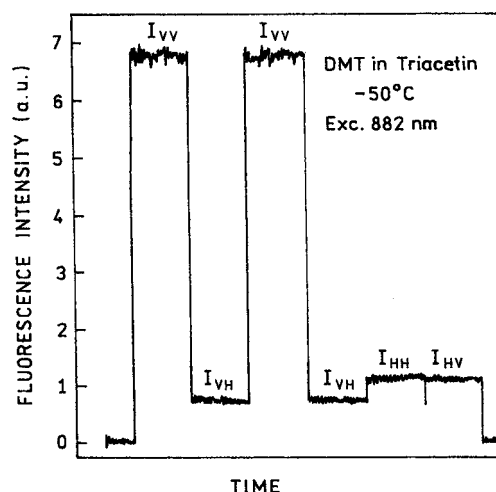


Figure 10. Emission intensities of the four polarized components of DMT with 882 nm excitation, in triacetin at $-50\text{ }^{\circ}\text{C}$.

wavelengths demonstrates that 4-PE can be accomplished without significant heating of the samples.

The r_0 values found for DMT with 4-PE are the largest yet observed in an isotropic solution. For one-photon excitation the maximum ratio of intensities of the parallel and perpendicularly polarized component of the emission is 3.0. It is interesting to see the relative magnitude of these components with four-photon excitation (Figure 10). The G -factor for our instrument was near unity, as can be seen from the vertical (I_{HV}) and horizontal (I_{HH}) components with horizontally polarized excitation (Figure 10, right side). For vertically polarized excitation the vertical (I_{VV}) and horizontal (I_{VH}) polarized components of the emission differ by 8-fold. Such high ratios for the vertical and horizontal components is an indication of the high degree of orientation of the excited-state transition moment distribution (Figure 11). As the mode of the excitation changes from 1-PE to 4-PE the fundamental anisotropy increases from 0.40 to 0.727.

An important advantage of two-photon excitation microscopy is localized excitation at the focal point of the lens.³⁻⁴ This effect is due to the quadratic dependence on the illumination intensity. As a result one expects still greater excitation localization with higher modes of excitation. Figure 12 shows the calculated distribution of the excited-state population along the excitation beam. As the mode of absorption increases the

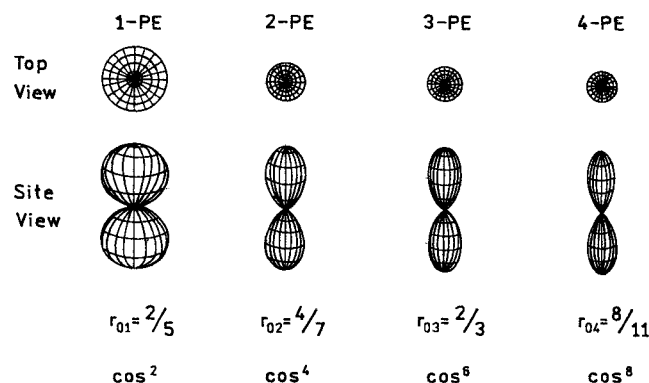


Figure 11. Theoretical distribution of the excited-state transition moments for 1-, 2-, 3-, and 4-photon excitation.

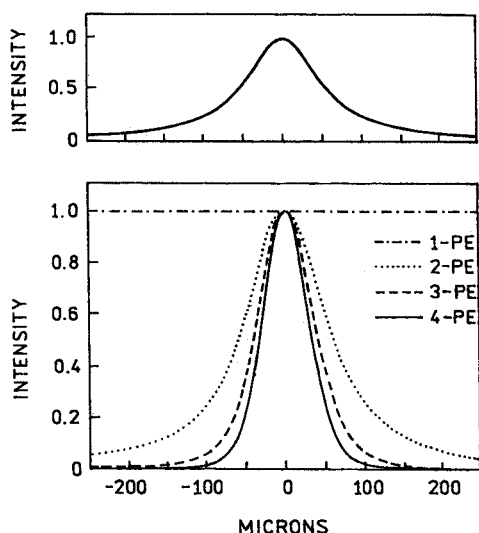


Figure 12. Theoretical predictions for the spatial distribution of excited fluorophores along the direction of the beam for 1-, 2-, 3-, and 4-photon excitation. The assumed Gaussian intensity distribution across the beam is shown in the upper panel.

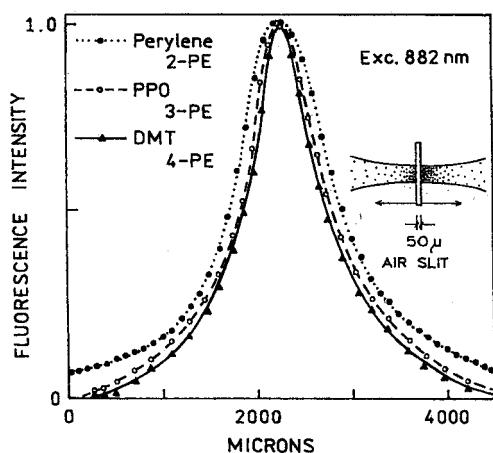


Figure 13. Measured spatial intensity distribution along the laser beam for 882 nm excitation. This wavelength provides 2-PE of perylene, 3-PE of PPO, and 4-PE of DMT.

excited-state population is expected to become more localized along the z -axis.

We examined the excited state distribution along the excitation beam (Figure 13) and across the excitation beam (Figure 14). We scanned images of the fluorescence spots using a 50 μ precision air slit along the beam and a 10 μ pinhole across the beam. These measurements were all performed with the same

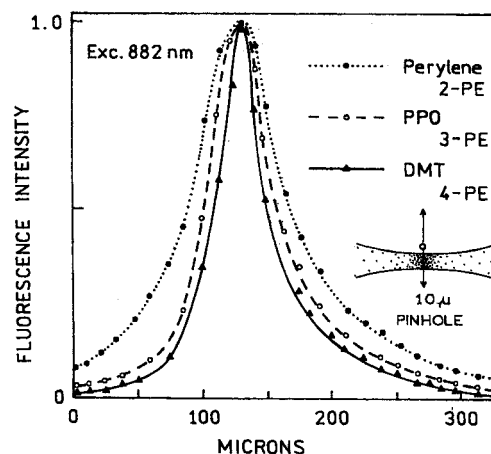


Figure 14. Measured distribution of intensities across the laser beam with 882 nm excitation. This wavelength provides 2-PE of perylene, 3-PE of PPO, and 4-PE of DMT.

excitation wavelength of 882 nm. We obtained 2-PE using perylene, 3-PE using PPO, and 4-PE using DMT. These data demonstrate that at the same excitation wavelength, more localized excitation can be obtained with a higher mode of excitation. Importantly, the intensities rapidly decrease to zero at short distances from the center of the laser beam.

Discussion

Our observation of four-photon excitation has implications for the use of multiphoton excitation in spectroscopy and imaging. For example, we have shown the emission from saturated alkanes can be observed with excitation near 300 nm via two-photon excitation.^{37–39} It would be difficult to obtain good transmission of 300 nm through a microscope objective. However, higher-order three- or four-photon excitation of saturated hydrocarbons may allow detection of deep UV emission from cellular membranes in an optical microscope.

The high anisotropies possible with 4-PE may also be useful in cellular imaging. It is possible to accomplish anisotropy imaging in laser scanning microscopy.⁴⁰ As shown in Figure 10 there is an approximate 8-fold intensity difference between the parallel (V) and perpendicular (H) components of the polarized emission. Such large intensity differences should facilitate the use of anisotropy measurements in cell imaging.

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References and Notes

- (1) Fredrich, D. M. Two-photon molecular spectroscopy. *J. Chem. Educ.* **1982**, *59*, 472–481.
- (2) Callis, P. E. Two-photon-induced fluorescence. *Annu. Rev. Phys. Chem.* **1997**, *48*, 271–297.
- (3) Denk, W.; Strickler, J. H.; Webb, W. W. Two-photon laser scanning fluorescence microscopy. *Science* **1990**, *248*, 73–76.
- (4) Xu, C.; Webb, W. W. Multiphoton excitation of molecular fluorophores and nonlinear laser microscopy. In *Topics in Fluorescence Spectroscopy*; Vol. 5: *Nonlinear and Two-Photon-Induced Fluorescence*; Lakowicz, J. R., Ed.; Plenum Press: New York, 1997; pp 471–540.
- (5) *Topics in Fluorescence Spectroscopy*; Vol. 5: *Nonlinear and Two-Photon-Induced Fluorescence*; Lakowicz, J. R., Ed.; Plenum Press: New York, 1997; 544 pp.
- (6) Walla, P. J.; Yom, J.; Krueger, B. P.; Fleming, G. R. Two-photon excitation spectrum of light-harvesting complex II and fluorescence upconversion after one- and two-photon excitation of the carotenoids. *J. Phys. Chem. B* **2000**, *104*, 4799–4806.

- (7) Bagatolli, L.; Gratton, E.; Khan, T. K.; Chong, P. L.-G. Two-photon fluorescence microscopy studies of bipolar tetraether giant liposomes from thermoacidophilic archaeobacteria *Sulfolobus acidocaldarius*. *Biophys. J.* **2000**, *79*, 416–425.
- (8) Bagtoli, L. A.; Gratton, E. Two-photon fluorescence microscopy of coexisting lipid domains in giant unilamellar vesicles of binary phospholipid mixtures. *Biophys. J.* **2000**, *78*, 290–305.
- (9) Brown, E. B.; Wu, E. S.; Zipfel, W.; Webb, W. W. Measurement of molecular diffusion in solution by multiphoton fluorescence photobleaching recovery. *Biophys. J.* **1999**, *77*, 2837–2849.
- (10) Teuchner, K.; Freyer, W.; Leupold, D.; Volkmer, A.; Birch, D. J. S.; Altmeyer, P.; Stücker, M.; Hoffman, K. Femtosecond two-photon excited fluorescence of melanin. *Photochem. Photobiol.* **1999**, *70* (2), 146–151.
- (11) Masters, B. R.; So, P. T. C.; Gratton, E. Multiphoton excitation fluorescence microscopy and spectroscopy of in vivo human skin. *Biophys. J.* **1997**, *72*, 2405–2412.
- (12) Guo, Y.; Wang, Q. Z.; Zhadin, N.; Liu, F.; Demos, S.; Calistru, D.; Tirsklunas, A.; Katz, A.; Budansky, Y.; Ho, P. P.; Alfano, R. R. Two-photon excitation of fluorescence from chicken tissue. *Appl. Opt.* **1997**, *36*, 968–970.
- (13) Baker, G. A.; Pandey, S.; Bright, F. V. Extending the reach of immunoassays to optically dense specimens by using two-photon excited fluorescence polarization. *Anal. Chem.* **2000**, *72*, 5748–5752.
- (14) Okerberg, E.; Shear, J. B. Attomole-level protein fingerprinting based on intrinsic peptide fluorescence. *Anal. Chem.* **2001**, *73*, 1610–1613.
- (15) Fan, G. Y.; Fujisaki, H.; Miyawaki, A.; Tsay, R.-K.; Tsien, R. Y.; Ellisman, M. H. Video-rate scanning two-photon excitation fluorescence microscopy and ratio imaging with cameleons. *Biophys. J.* **1999**, *76*, 2412–2420.
- (16) Diaspro, A. Introduction to two-photon microscopy. *Microscopy Res. Technique* **1999**, *47*, 163–164.
- (17) So, P. T. C.; Dong, C. Y.; Masters, B. R.; Berland, K. M. Two-photon excitation fluorescence microscopy. *Annu. Rev. Biomed. Eng.* **2000**, *02*, 399–429.
- (18) Lakowicz, J. R.; Gryczynski, I.; Malak, H.; Schrader, M.; Engelhardt, P.; Kano, H.; Hell, S. W. Time-resolved fluorescence spectroscopy and imaging of DNA labeled with DAPI and Hoechst 33342 using three-photon excitation. *Biophys. J.* **1997**, *72*, 567–578.
- (19) Szmajnski, H.; Gryczynski, I.; Lakowicz, J. R. Three-photon induced fluorescence of the calcium probe Indo-1. *Biophys. J.* **1996**, *70*, 547–555.
- (20) Gryczynski, I.; Malak, H.; Lakowicz, J. R.; Cheung, H. C.; Robinson, J.; Umeda, P. K. Fluorescence spectral properties of troponin C mutant F22W with one-, two-, and three-photon excitation. *Biophys. J.* **1996**, *71*, 3448–3453.
- (21) Gryczynski, I.; Malak, H.; Lakowicz, J. R. Multiphoton excitation of the DNA stains DAPI and Hoechst. *Bioimaging* **1996**, *4*, 138–148.
- (22) Wokosin, D. L.; Centoze, V. E.; Crittenden, S.; White, J. Three-photon excitation fluorescence imaging of biological specimens using an all-solid-state laser. *Bioimaging* **1996**, *4*, 208–214.
- (23) Maiti, S.; Shear, J. B.; Williams, R. M.; Zipfel, W. R.; Webb, W. W. Measuring serotonin distribution in live cells with three-photon excitation. *Science* **1997**, *275*, 530–532.
- (24) Belfield, K. D.; Hagan, D. J.; Van Stryland, E. W.; Schafer, K. J.; Negres, R. A. New two-photon absorbing fluorene derivatives: synthesis and nonlinear optical characterization. *Org. Lett.* **1999**, *1* (10), 1557–1578.
- (25) Rumi, M.; Ehrlich, J. E.; Heikai, A. A.; Perry, J. W.; Barlow, S.; Hu, Z.; McCord-Maughon, D.; Parker, T. C.; Röckel, H.; Thayumanavan, S.; Marder, S. R.; Beljonne, D.; Brédas, J.-L. Structure–property relationships for two-photon absorbing chromophores: bis-donor diphenylpolyene and bis(styryl)benzene derivatives. *J. Am. Chem. Soc.* **2000**, *122*, 9500–9510.
- (26) Chung, S.-J.; Kim, K.-S.; Lin, T.-C.; He, G. S.; Swiatkiewicz, J.; Prasad, P. N. Cooperative enhancement of two-photon absorption in multi-branched structures. *J. Phys. Chem. B* **1999**, *103*, 10741–10745.
- (27) Belfield, K. D.; Schafer, K. D.; Mourad, W.; Reinhardt, B. A. Synthesis of new two-photon absorbing fluorene derivatives via Cu-mediated Ullmann condensations. *J. Org. Chem.* **2000**, *65* (15), 4475–4481.
- (28) Adronov, A.; Fréchet, J. M. J. Novel two-photon absorbing dendritic structures. *Chem. Mater.* **2000**, *12*, 2838–2841.
- (29) Lakowicz, J. R.; Gryczynski, I.; Gryczynski, Z.; Danielsen, E.; Wirth, M. J. Time-resolved fluorescence intensity and anisotropy decays of 2,5 diphenylloxazole by two-photon excitation and frequency-domain fluorometry. *J. Phys. Chem.* **1992**, *98*, 3000–3006.
- (30) Weber, G. Polarization of the fluorescence of solutions. In *Fluorescence and Phosphorescence Analysis: Principles and Applications*; Interscience Publishers: New York, 1966; pp 217–240.
- (31) Callis, P. R. On the theory of two-photon induced fluorescence anisotropy with application to indoles. *J. Chem. Phys.* **1993**, *21*, 61–78.
- (32) Chen, S.-Y.; Van Der Meer, B. W. Theory of two-photon induced fluorescence anisotropy decay in membranes. *Biophys. J.* **1993**, *64*, 1567–1575.
- (33) Lakowicz, J. R.; Maliwal, B. P. Construction and performance of a variable-frequency phase-modulation fluorometer. *Biophys. Chem.* **1985**, *21*, 61–78.
- (34) Laczkó, G.; Lakowicz, J. R.; Gryczynski, I.; Gryczynski, Z.; Malak, H. A 10 GHz frequency-domain fluorometer. *Rev. Sci. Instrum.* **1990**, *61*, 2331–2337.
- (35) Gratton, E.; Lakowicz, J. R.; Maliwal, B. P.; Cherek, H.; Laczkó, G.; Limkeman, M. Resolution of mixtures of fluorophores using variable-frequency phase shift and modulation data. *Biophys. J.* **1984**, *46*, 479–486.
- (36) Lakowicz, J. R.; Cherek, H.; Kusba, J.; Gryczynski, I.; Johnson, M. L. Review of fluorescence anisotropy decay analysis by frequency-domain fluorescence spectroscopy. *J. Fluoresc.* **1993**, *3*, 103–116.
- (37) Lakowicz, J. R.; Gryczynski, I. Fluorescence intensity decays of cyclohexane and methylcyclohexane with two-photon excitation from a high repetition rate frequency-doubled dye laser. *Biospectrosc.* **1995**, *1*, 3–8.
- (38) Gryczynski, I.; Lakowicz, J. R. Quenching of methylcyclohexane fluorescence by methanol. *Photochem. Photobiol.* **1995**, *3*, 426–432.
- (39) Gryczynski, I.; Razyńska, A.; Lakowicz, J. R. Two-photon induced fluorescence of linear alkanes: A possible intrinsic lipid probe. *Biophys. Chem.* **1996**, *57*, 291–295.
- (40) Toyonaga, S.; Nakano, Y. Imaging of rotational Brownian motion of bimolecules in living cells using fluorescence depolarization microscopy. *Proc. SPIE-Int. Soc. Opt. Eng.* **1999**, *3260*, 84–95.