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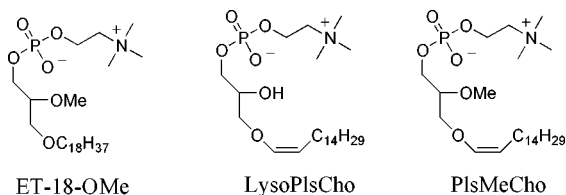
# An Efficient New Route to Plasmenyl-type Lipids: Synthesis and Cytotoxicity of a Plasmenylcholine Analogue of the Antitumor Ether Lipid ET-18-OMe

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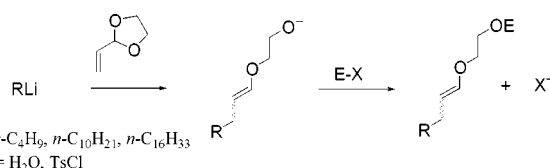
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Plasmalogens<sup>1-3</sup> (i.e. plasmenylcholines or plasmenylethanolamines) are an arachidonate-rich family of mammalian phospholipids containing *sn*-1-*Z*-1'-*O*-alkenyl chains of varying length and degrees of unsaturation. They are members of a broad class of ether-linked phospholipids which also includes the cytotoxic antitumor ether lipids (ATL): single chain ether lipids that interfere with phospholipid metabolism and signal transduction pathways. One such ATL, 1-*O*-octadecyl-2-*O*-methyl-*rac*-glycero-3-phosphocholine (ET-18-OMe, edelfosine), induces apoptosis in a variety of Fas-positive tumor cell lines<sup>4</sup> and has shown promising activity in Phase II clinical trials for bone marrow purging autologous transplantation treatment of acute leukemia.<sup>5</sup>

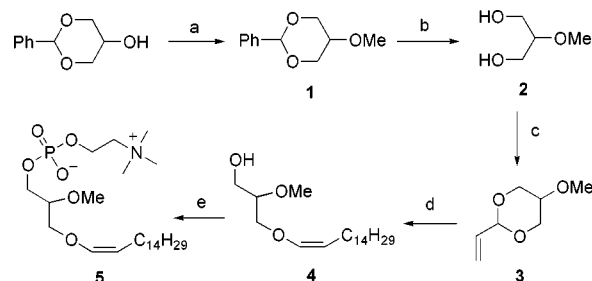


The involvement of plasmalogens in cell signaling pathways, combined with their utility in drug and gene delivery applications,<sup>6-8</sup> has prompted the development of a more direct route to this important class of phospholipids. We now report the development of a facile new pathway to glyceryl vinyl ether lipids based on the addition of alkylolithium reagents to 2-vinyl-1,3-dioxolane (Figure 1) or 5-methoxy-2-vinyl-1,3-dioxane intermediates (Figure 2). This method has been applied to the synthesis of 1-*O*-1'-(*Z*)-hexadecenyl-2-*O*-methyl-*rac*-glycero-3-phosphocholine (PlsMeCho), a *Z*-vinyl ether analogue of ET-18-OMe, that shows significant antitumor activity in MIAPACA-2 pancreatic tumor cells.

Our initial investigations focused on Micheal-type additions of preformed alkylolithium reagents to 2-vinyl-1,3-dioxolane<sup>9</sup> as a model substrate (Figure 1) using lithium-iodide exchange,<sup>10,11</sup>



**Figure 1.** Generalized pathway for the preparation of vinyl ether lipids from 2-vinyl-1,3-dioxolane.



**Figure 2.** Pathway for the synthesis of 1-*O*-1'-(*Z*)-hexadecenyl-2-*O*-methyl-*rac*-glycero-3-phosphocholine (PlsMeCho): (a) NaH, MeI, THF, 25 °C, 3 h (92%); (b) AG 50W-X2 resin, THF, H<sub>2</sub>O, reflux, 5 h (100%); (c) acrolein, *n*-BuSnCl<sub>3</sub>, 25 °C, 1 h (28% *trans*, 36% *cis*); (d) Li, DBB, C<sub>13</sub>H<sub>27</sub>I, THF, 0 °C, 0.5 h (53%); (e) (i) 2-oxo-2-chloro-1,3,2-dioxaphospholane, Et<sub>3</sub>N, C<sub>6</sub>H<sub>6</sub>, 5 °C, 20 h; (ii) Me<sub>3</sub>N, MeCN/C<sub>6</sub>H<sub>6</sub>, 70 °C, 26 h (62% for steps i and ii, combined).

direct lithiation,<sup>12</sup> and lithiation catalyzed by 4,4-di-*tert*-butylbiphenyl (LiDBB).<sup>13,14</sup> Conditions for the reaction between 2-vinyl-1,3-dioxolane and the alkylolithium species were varied to determine the best conditions for *Z*-vinyl ether product formation (Table 1). Although couplings resulting from lithium-iodide exchange gave a modest excess of *Z*-vinyl ether product (60:40 *Z:E* ratio) in 51% yield, LiDBB-mediated reactions under Barbier-type conditions<sup>15,16</sup> were much more efficient, giving the corresponding vinyl ether adduct in 61% yield with *Z:E* ratios as high as 87:13.

On the basis of these observations, a related synthetic pathway for PlsMeCho was explored (Figure 2). This sequence begins with the preparation of 5-methoxy-2-vinyl-1,3-dioxane (**3**) via methylation of 2-phenyl-1,3-dioxane-5-ol, cleavage of acetal **1** under acidic conditions, and condensation of **2** with acrolein in the presence of *n*-BuSnCl<sub>3</sub>.<sup>17</sup> The *cis:trans* ratio of **3** prepared in this manner was 1.3:1, as determined by <sup>1</sup>H NMR comparisons with *cis*- and *trans*-5-methyl-2-phenyl-1,3-dioxane.<sup>18</sup> The key step in this strategy, coupling of **3** under Barbier-type reaction conditions with 1-chloroalkanes, was then conducted in the presence of lithium 4,4-di-*tert*-butylbiphenyl radical anion (LiDBB). Reaction of *cis*-**3** and 1-chlorotridecane under Barbier conditions successfully gave the penultimate alcohol **4** in 49% yield and 88:12 *Z:E* ratio (Table 2).<sup>19</sup> Similar results were obtained by using *trans*-**3** as substrate. As the data in Table 2 also show, the highest yields of the desired 4-(*Z*) product were obtained by using a 2-fold excess of 1-iodotridecane relative to **3** under Barbier conditions (53% yield, 95:5 *Z:E*). Introduction of the phosphocholine headgroup in the final step was accomplished by treating alcohol **4**-(*Z*) with

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**Table 1.** Alkylolithium Additions to 2-Vinyl-1,3-dioxane

RLi	E-X	% yield	Z:E
<i>n</i> -BuLi	H <sub>2</sub> O	43	40:60
<i>n</i> -C <sub>10</sub> H <sub>21</sub> I/ <i>t</i> -BuLi	TsCl	51	60:40
<i>n</i> -C <sub>13</sub> H <sub>27</sub> Cl/Li	TsCl	36	31:69
<i>n</i> -C <sub>13</sub> H <sub>27</sub> Cl/LiDBB	TsCl	27	40:60
<i>n</i> -C <sub>13</sub> H <sub>27</sub> Cl/LiDBB (Barbier cond.)	TsCl	61	87:13

**Table 2.** Barbier-type Reactions of 2-Vinyl-1,3-dioxanes

3: R = Me; 6: R = TBDMS					
Substrate	Electrophiles	equiv. E	% Yield	Z:E	Comments
trans-3	<i>n</i> -C <sub>13</sub> H <sub>27</sub> I	2	53	95:5	—
cis-3	<i>n</i> -C <sub>13</sub> H <sub>27</sub> Cl	0.5	49	88:12	—
trans-6	<i>n</i> -C <sub>13</sub> H <sub>27</sub> Cl	1	33	24:76	—
trans-6	<i>n</i> -C <sub>13</sub> H <sub>27</sub> Cl	2	57	30:70	—
trans-6	<i>n</i> -C <sub>13</sub> H <sub>27</sub> Cl	5	53	30:70	—
cis-6	<i>n</i> -C <sub>13</sub> H <sub>27</sub> Br	2	32	67:33	—
cis-6	<i>n</i> -C <sub>13</sub> H <sub>27</sub> I	2	47	98:2	—
cis-6	<i>n</i> -C <sub>13</sub> H <sub>27</sub> Ms <sup>a</sup>	2	47	0	>98% cis
cis-6	ONBS <sup>b</sup>	2	0	0	>98% cis
cis-6	Cl	2	41	0	>98% cis

<sup>a</sup> Ms ≡ methanesulfonate. <sup>b</sup> NBS ≡ *m*-nitrobenzenesulfonate.

2-oxo-2-chloro-1,3,2-dioxaphospholane, followed by excess trimethylamine,<sup>2,3</sup> to give racemic PlsMeCho (**5**) in 62% yield.

The cytotoxicity of PlsMeCho was evaluated in MIAPACA-2 cells and compared to the cytotoxicities of ET-18-OMe, ET-16-OMe, and the corresponding semisynthetic lysoplasmenechole (LysoPlsCho)<sup>20</sup> (Figure 3) using the MTT assay. Cells were exposed to 10 μM lipid in Dulbecco's modified media for 24 h then examined for viability after incubation with MTT for 3–4 h. Our results indicate that PlsMeCho exhibits similar cytotoxicity within experimental error as the antitumor ether lipids ET-18-OMe and ET-16-OMe in MIAPACA-2 cells. These observations are consistent with the apoptosis of cells that actively accumulate ATL.<sup>4,21–23</sup> Originally, this sensitivity was thought to arise from the absence of *O*-alkyl cleavage enzyme activity;<sup>24</sup> however, it is now recognized that the cytotoxic and cytostatic properties of ATL are attributable to their interferences with both CTP:

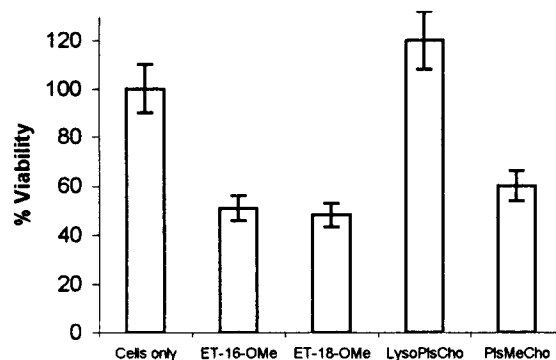
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**Figure 3.** Viability of MIAPACA-2 cells after a 24 h exposure to 10 μM lipid concentrations as determined by MTT assay. Cells were plated at 15 000 cells/well (96 well plates) in Dulbecco's modified Eagle medium supplemented with 10% heat-inactivated fetal calf serum, penicillin (100 units/mL), and streptomycin (100 μg/mL). The reported data represent the mean and standard deviation of quadruplicate determinations.

phosphocholine cytidyltransferase-mediated phosphatidylcholine synthesis<sup>25,26</sup> and cell cycling processes leading from G<sub>0</sub>/G<sub>1</sub> to S phase.<sup>27</sup> They further suggest that the insensitivity of MIAPACA-2 cells to LysoPlsCho may be due to the rapid remodeling of this lysolipid precursor, via reacylation, after uptake. This would have the net effect of maintaining phospholipid homeostasis<sup>28</sup> while augmenting the cellular plasmalogen pool.

In conclusion, PlsMeCho has been prepared via a facile reaction sequence with use of an acrolein acetal derivative and 1-iodotridecane as precursors under Barbier-type reaction conditions. The observed regioselectivities and high Z:E ratios of the LiDBB-mediated coupling reaction are likely due to a lithiated allylic anion that undergoes stereoselective γ-coupling with electrophiles<sup>29</sup> via lithium ion–acetal chelation. The resulting racemic PlsMeCho shows cytotoxic properties similar to the clinically relevant ATL analogue ET-18-OMe; however, it is not presently known whether the cytotoxic mechanism for PlsMeCho is apoptotic in nature as it is for ET-18-OMe. Experiments designed to adapt this method for the preparation of plasmalogens and the pure *R*-5/*S*-5 stereoisomers are in progress. Additional experiments aimed at probing the relative cytotoxicity of *R*-5, *S*-5, and racemic **5**, as well as the mechanism of cell death upon exposure to these ATL, are also planned.

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**Supporting Information Available:** <sup>1</sup>H and <sup>13</sup>C NMR spectral data for compounds **1–5** (PDF). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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