QSAR of Chemical Polarizability and Nerve Toxicity. 2

Corwin Hansch* and Alka Kurup

Department of Chemistry, Pomona College, Claremont, California 91711

Received April 18, 2003

Polarizability is a property of molecules that has long been of interest to scientists from a variety of viewpoints. However, in the area of the QSAR of chemical—biological interactions, it has received little attention. Recently we have shown that one can use the simple summation of the valence electrons (H = 1, C = 4, O = 6, etc.) in a molecule as a measure of its polarizability. We have found this parameter to correlate nerve toxicity of a wide variety of chemicals acting on nerves of frogs, rabbits, cockroaches, and humans.

INTRODUCTION

Among the electronic factors that must be considered to understand how chemicals effect living systems or their parts is polarizability. Over the years, there has been much discussion of the subject;^{1–5} however, taking into consideration this molecular property in attempts to formulate QSAR has been largely neglected except for the work in our laboratory. For many years we have used the Lorentz–Lorenz equation:

$$MR = n^2 - 1/n^2 + 2\left(\frac{MW}{d}\right)$$
 (1)

In this expression, molar refractivity is defined by n (refractive index), MW (molecular weight), and d (density). Thus, MR is composed of two components, volume and the ability of the electrons to interact with light. It is of interest that this equation was defined in 1880 before anyone had thought of electrons. Leo developed an algorithm somewhat like what he formulated for calculating log P for octanol/ water partitions.6 We have found 1450 QSAR where calculated CMR plays an important role in chemical-biological interactions. However, we have realized that CMR is ambivalent, but that it could not be replaced by molar volume. We have now gained a deeper understanding of polarizabiliy⁵ from the work of Agin et al.4 In an unusual study of a very miscellaneous set of compounds that blocked the action of frog sartorius muscle, they showed that a plot of log 1/C vs αI_p yielded a very straight line. From their data we formulated eq 2.

$$\log 1/C = 0.010(\pm 0.00)\alpha I_p - 0.983(\pm 0.174)$$
 (2)

$$n = 39, r^2 = 0.987, s = 0.240, q^2 = 0.984$$

The polarizability α was calculated as was the ionization potential I_p .

Deriving QSAR 2 for α alone, we obtained QSAR 3:

log 1/C =
$$0.082(\pm 0.005)\alpha$$
 - $0.664(\pm 0.23)$ (3)
 $n = 39, r^2 = 0.973, s = 0.344, a^2 = 0.969$

Thus we see that polarizability is the main factor in rationalizing their data. Using CMR calculated via the

BioByte Clog P Program⁶ we obtained QSAR 4.

$$\log 1/C = 0.82(\pm 0.05)CMR - 0.64(\pm 0.25)$$
 (4)

$$n=37, r^2=0.969, s=0.375, q^2=0.964$$

outliers: antipyrine; methyl anthranilate

Next we showed that simply by adding up the number of valence electrons of the elements in the compounds (H = 1, C = 4, O = 6, N = 5, halogens = 7, S = 6, P = 5) to define NVE, we obtained QSAR 5.

$$\log 1/C = 0.064(\pm 0.005)$$
NVE $-0.779(\pm 0.301)$ (5)

$$n = 37$$
, $r^2 = 0.958$, $s = 0.458$, $q^2 = 0.953$
outliers: antipyrine; octanol

QSAR 5 is not quite as good as QSAR 4. It does not contain the large volume component contained in the Lorentz—Lorenz equation and CMR, although volume is involved to some degree. That is the more electrons the greater the volume. We assumed that elements beyond F would be more easily polarized and hence would not fit QSAR 5 as well. However, this is not true; all of the halogens are well fit in many examples. We now have 373 QSAR based on NVE. In fact, CHCl₃ is not an outlier in QSAR 5.

In our earlier report⁵ we found that NVE QSAR were often associated with nerve toxicity. Examples were found for the inhibition of cockroach, rabbit, and frog nerves. It is of special interest that we concluded that the toxic effect occurs at the synapse of the nerve axon. This appears to be a polar region, because most often we find no dependence on log P. No doubt, nerves could be blocked by hydrophobic compounds, but this is not our current interest. We believe that understanding this type of toxicity will have an important role in drug development in medicinal chemistry. However, it is now abundantly clear that NVE is important for other types of chemical—biological interactions.

RESULTS

A. Barbiturates. A logical place to look for nerve toxicity would be with barbiturates. At present, we have many QSAR on all aspects of this class of drugs. However, only a few are suitable for assaying nerve toxicity. Most of these are correlated by log P. In part, this is due to the fact that NVE and log P are perfectly collinear for saturated hydrocarbons. Many studies with barbiturates have been made with varia-

^{*} Corresponding author phone: (909)621-8445; fax: (909)607-7726; e-mail: atessier@pomona.edu.

Table 1. LD₅₀ of Barbiturates for Female White Mice

		log 1/C			Clog	
no.	substituents	obsd	pred (eq 6)	Δ	P	NVE
1	5-Me, 5-C(Me)=CHC ₂ H ₅	2.62	2.60	0.02	1.50	82
2	$5-C_2H_5$, $5-CH(Me)=CHC_2H_5^a$	3.09	2.82	0.27	2.03	88
3	$5-C_3H_7$, $5-CH(Me)=CHC_2H_5$	2.95	3.05	-0.10	2.56	94
4	5-CHMe ₂ , 5-C(Me)=CHC ₂ H ₅ ^{a}	3.38	3.23	0.15	2.43	94
5	5-Me, 5-C(C_2H_5)=CHMe	2.56	2.60	-0.04	1.50	82
6	$5-C_2H_5$, $5-C(C_2H_5)=CHMe$	2.90	2.82	0.08	2.03	88
7	$5-C_3H_7$, $5-C(C_2H_5)=CHMe$	2.95	3.05	-0.10	2.56	94
8	5-CHMe ₂ , 5-C(C_2H_5)=CHMe	3.19	3.23	-0.04	2.43	94
9	5-Me, 5-C(Me)=CHC ₃ H ₇	2.78	2.82	-0.04	2.03	88
10	$5-C_2H_5$, $5-C(Me)=CHC_3H_7$	3.17	3.05	0.13	2.56	94
11	5-Me, 5-C(Me)=CHCHMe ₂	3.05	3.01	0.04	1.90	88
12	5-Me, 5-C(Me)=CHC ₄ H ₉ a	2.82	3.05	-0.23	2.56	94
13	$5-C_2H_5$, $5-C(Me)=CHC_4H_9$	3.29	3.27	0.02	3.09	100
14	$5-C_2H5$, $5-C(C_3H_7)=CHC_2H_5$	3.29	3.27	0.02	3.09	100

a Outliers.

Table 2. Elimination Rate Constant of Barbiturates for Rabbits¹⁰

			log k		
no.	substituents	obsd	pred (eq 7)	Δ	NVE
1	hexobarbital	0.17	0.01	0.17	92
2	pentobarbital	-0.01	-0.14	0.13	90
3	cyclobarbital	-0.05	0.01	-0.06	92
4	amobarbital	-0.35	-0.14	-0.22	90
5	allobarbaital	-0.92	-0.84	-0.09	80
6	phenobarbital ^a	-1.60	-0.28	-1.33	88
7	barbital	-1.34	-1.40	0.06	72
^a O	utlier.				

tion at the 5-position, using alkyl groups. There are a few illustrative examples that contain unsaturated alkyl groups. For these we can distinguish NVE from log P correlation. The SAR of barbiturates has been discussed.^{7,8}

 LD_{50} for Female White Mice. Data from Cope and Hancock⁹ (Table 1).

$$\log 1/C = -1.44(\pm 1.07) \operatorname{Clog} P + 0.16(\pm 0.095) \operatorname{NVE} - 8.70(\pm 6.25)$$
 (6)

$$n = 11$$
, $r^2 = 0.924$, $s = 0.077$, $q^2 = 0.879$
outliers: 5-C₂H₅, 5-C(Me)=CHC₂H₅; 5-CH
(Me)₂, 5-CH(Me)=CHC₂H₅; 5-Me, 5-C(Me)=CHC₄H₉

The negative C log P term is what we would expect if the synapse is polar in character.

Elimination Rate Constant for Rabbits. Data from Watari et al. ¹⁰ (Table 2).

$$\log k = 0.070(\pm 0.025) \text{ NVE} - 6.45(\pm 2.13)$$
 (7)

$$n = 6$$
, $r^2 = 0.940$, $s = 0.163$, $q^2 = 0.866$
outlier: phenobarbital

Possibly, nerves are involved in controlling elimination.

Inhibition of Acid Secretion from Gastric Mucosa of Frog.

Data from Dinno et al. 11 (Table 3).

$$\log k = 0.028(\pm 0.003) \text{ NVE} - 0.886(\pm 0.276)$$
 (8)

$$n = 5$$
, $r^2 = 0.996$, $s = 0.018$, $q^2 = 0.993$

outlier: thiamylal

Table 3. Inhibition of Acid Secretion from Gastric Mucosa of Frog by Barbiturates

			log k					
no.	substituents	obsd	pred (eq 8)	Δ	NVE			
1	barbital	1.15	1.15	0.00	72			
2	diallylbarbituric acid	1.38	1.38	0.00	80			
3	phenobarbital	1.58	1.60	-0.02	88			
4	pentobarbital	1.68	1.66	0.02	90			
5	secobarbital	1.77	1.77	0.00	94			
6	thiamylal ^a	1.89	1.77	0.12	94			
a C	^a Outlier.							

Table 4. Narcosis of Mice by Barbiturates¹²

			log 1/C		
			pred		
no.	substituents	obsd	(eq 9)	Δ	NVE
1	5-Me, 5-C(C_3H_7)=CH C_2H_5				
2	$5-C_2H_5$, $5-C(Me)=CHC_4H_9$	3.75	3.72	0.03	100
3	5-CHMe ₂ , 5-C(C ₂ H ₅)=CHMe ^a	3.72	3.52	0.21	94
4	$5-C_2H_5$, $5-C(Me)=CHC_2H_5^a$	3.65	3.31	0.34	88
5	$5-C_2H_5$, $5-C(Me)=CHC_3H_7$	3.64	3.52	0.13	94
6	$5-C_3H_7$, $5-C(Me)=CHC_2H_5$	3.56	3.52	0.05	94
7	$5-C_3H_7$, $5-C(C_2H_5)=CHMe$	3.42	3.52	-0.10	94
8	$5-C_2H_5$, $5-C(C_2H_5)=CHMe$	3.40	3.31	0.09	88
9	5-Me, 5-C(Me)= CHC_4H_9	3.38	3.52	-0.14	94
10	5-Me, 5-C(Me)=CHC ₃ H ₇	3.27	3.31	-0.04	88
11	5-Me, 5-C(Me)= CHC_2H_5	3.21	3.11	0.11	82
12	5-Me, 5-C(Me)=CHCHMe ₂	3.20	3.31	-0.11	88
13	5-Me, 5-C(C_2H_5)=CHMe	3.06	3.11	0.05	82
14	5-CHMe ₂ , 5-C(Me)=CHC ₂ H ₅ a	3.98	3.52	0.47	94

a Outliers.

Table 5. LD₁₀₀ of Miscellaneous Drugs for Humans¹³

			log 1/C				
no.		obsd	pred (eq 10)	Δ	log P	NVE	
1	chlorpromazine	5.20	5.38	-0.18	3.22	110	
2	propoxyphene	5.08	4.98	0.10	2.36	134	
3	amitriptyline	4.92	4.76	0.16	2.50	108	
4	dothiepin	4.75	4.97	-0.22	2.76	108	
5	secobarbital	4.19	4.13	0.06	1.97	94	
6	phenobarbital	3.71	3.37	0.34	1.14	88	
7	chloroform ^a	3.60	3.24	0.36	1.97	26	
8	chlormethiazole	3.51	3.68	-0.17	2.12	50	
9	paraldehyde	2.88	2.53	0.35	0.67	54	
10	ether	2.17	2.42	-0.25	0.89	32	
11	ethanol	1.06	1.27	-0.21	-0.31	20	
^a Outlier.							

Narcosis of Mice. Data from Hansch et al. 12 (Table 4).

$$\log 1/C = 0.034(\pm 0.011) \text{ NVE} + 0.303(\pm 1.00)$$
 (9)

$$n = 11, r^2 = 0.847, s = 0.095, q^2 = 0.782$$

outliers: 5-CH(Me)₂, 5-C
(C₂H₅)=CHMe; 5-C₂H₅, 5-C(Me)=CHC₂H₅; 5-CH
(Me)₂, 5-CH(Me)=CHC₂H₅

B. Drug Toxicity to Humans. Of course, toxicity to humans is of greatest interest and the studies of King are surprising.

 LD_{100} of Miscellaneous Drugs to Humans. Data from King¹³ (Table 5).

$$n = 10, r^2 = 0.970, s = 0.270, q^2 = 0.922$$

outlier: phenobarbital

In a previous study based only on log P, $r^2 = 0.876$. These data come from studies in England where, whenever an individual commits suicide or dies from an overdose of a drug, the drug concentration in their blood is immediately determined. From average values of many examples, King established log 1/C.

In a second, larger study¹⁴ we were unable to formulate a QSAR, now using NVE we have derived QSAR 11 (Table 6).

$$\log 1/C = 0.61(\pm 0.17) \log P + 0.017(\pm 0.004) \text{NVE} + 1.44(\pm 0.37) (11)$$

$$n=36, r^2=0.850, s=0.438, q^2=0.817$$
 outliers: morphine; theophylline; CF₂ (Cl)₂; halothane; paraldehyde

It is noteworthy that a number of drugs containing Cl and Br are reasonably well fit. Two of the drugs having these elements are outliers. The terms in QSAR 11 are similar to those of QSAR 10. In QSAR 10 and 11, there are log P terms that account for the movement of the drugs from the stomach to the nerves of the central nervous system.

Antiinflammatory Potency of Glucocorticoid Derivatives in Man. Data from Ahmad and Mellors¹⁵ (Table 7).

log AE =
$$-0.78(\pm 0.35)$$
 Clog P + $0.091(\pm 0.03)$ NVE $-11.6(\pm 4.6)$ (12)

$$n = 16$$
, $r^2 = 0.897$, $s = 0.278$, $q^2 = 0.836$
outliers: 6-Me-16-OH-prednisolone; 6,9-di-F-16-OH-prednisolone

AE is esonipenic potency of glucorticoids.

Obviously nerves are involved in inflammation so that one would anticipate that NVE would be important.

Paralysis of Rabbit Diaphragm by Injection of into Phrenic

$$R_3N$$
 (CH₂)_nNR₃

Nerve. Data from Wein and Mason¹⁶ (Table 8)

$$\log RBR = 0.034(\pm 0.005) \text{ NVE} - 4.22(\pm 0.70)$$
 (13)

$$n = 16$$
, $r^2 = 0.933$, $s = 0.211$, $q^2 = 0.916$
outliers: $R = Me$, $n = 3$; $R = Me$, $n = 5$

RBR = relative biological response.

The same authors tested a similar set of congeners for which we found a similar QSAR.⁵

Inhibition of Cockroach Nerve Conduction by Miscellaneous Chemicals. Data from Uchida et al. 18 (Table 9).

$$\log 1/C = 0.084(\pm 0.009) \text{ NVE} - 1.68(\pm 0.41)$$
 (14)

$$n = 11, r^2 = 0.981, s = 0.198, q^2 = 0.969$$

outlier: L-menthol

Table 6. LD₁₀₀ of Miscellaneous Drugs to Humans¹⁴

			log 1/C			
no.		obsd	pred (eq 11)	Δ	log P	NVE
1	morphine ^a	5.45	3.45	2.00	0.15	110
2	chlorpromazine	5.24	5.32	-0.08	3.22	110
3	propoxyphene	5.08	5.22	-0.14	2.36	134
4	strychnine	4.57	4.09	0.48	0.68	128
5	quinine	4.45	4.93	-0.48	2.11	126
6	maprotiline	4.74	4.19	0.55	1.42	108
7	pentazocine	4.50	4.68	-0.18	2.04	114
8	dothiepin	4.75	5.01	-0.26	2.76	108
9	flurazepam	4.86	5.35	-0.49	2.35	142
10	amitriptyline	4.92	4.85	0.07	2.50	108
11	nortriptyline	4.24	4.27	-0.03	1.71	102
12	cocaine	4.70	4.14	0.56	1.05	118
13	secobarbital	4.14	4.28	-0.14	1.97	94
14	desipramine	4.15	4.14	0.01	1.45	104
15	propranolol	4.46	3.94	0.52	1.18	102
16	diazepam	4.20	4.89	-0.69	2.80	100
17	phenobarbital	3.71	3.67	0.04	1.14	88
18	chlormethiazole	3.51	3.61	-0.10	2.12	50
19	theophylline ^a	3.51	2.62	0.89	-0.02	68
20	caffeine	3.23	2.69	0.54	-0.07	74
21	tetrachloroethylene	4.56	4.14	0.42	3.40	36
22	$CF_2(Cl)_2^a$	4.61	3.32	1.29	2.16	32
23	toluene	4.34	3.73	0.61	2.73	36
24	$C(Cl)_4$	3.14	3.73	-0.59	2.83	32
25	$CH_3C(Cl)_3$	3.22	3.52	-0.30	2.49	32
26	trichloroethylene	3.91	3.44	0.47	2.42	30
27	C(Cl) ₃ F	4.06	3.54	0.52	2.53	32
28	nitrous oxide	2.39	1.99	0.41	0.43	16
29	benzene	3.99	3.26	0.73	2.13	30
30	CH(F) ₂ Cl	2.37	2.56	-0.19	1.08	26
31	halothane ^a	2.68	3.61	-0.93	2.30	44
32	ClCH ₂ CH ₂ Cl	2.49	2.80	-0.31	1.48	26
33	chloroform	3.60	3.10	0.50	1.97	26
34	CH ₂ BrCl	2.81	2.65	0.16	1.41	20
35	CH_2Cl_2	2.37	2.55	-0.18	1.25	20
36	CH ₃ CH ₂ Cl	2.21	2.66	-0.45	1.43	20
37	paraldehyde ^a	1.52	2.79	-1.27	0.67	54
38	EtOEt	2.17	2.54	-0.37	0.89	32
39	butanol	2.19	2.54	-0.35	0.88	32
40	2-propanol	1.26	1.93	-0.67	0.05	26
41	ethanol	1.26	1.82	-0.56	0.05	20

Table 7. Antiinflammatory Potency of Glucocorticoids in Humans¹⁵

			log AE	Clog		
no.		obsd	pred (eq 12)	Δ	P	NVE
1	cortisol	0.00	0.24	-0.24	1.70	144
2	coticosterone	-1.22	-0.79	-0.43	2.32	138
3	prednisolone	0.60	0.30	0.30	1.38	142
4	6-Me-11-OH-progesterone	-1.30	-1.45	0.15	3.17	138
5	6-Me-9-F-21-deoxycortisol	0.30	0.25	0.05	2.39	150
6	6-Me-prednisolone	0.70	0.60	0.10	1.70	148
7	6-Me-0-F-prednisolone	1.00	1.27	-0.27	1.55	154
8	6-Me-9-F-21-deoxyprednisolone	0.30	0.31	-0.01	2.07	148
9	9-Me-16-OH-prednisolone ^a	0.00	1.58	-1.58	1.14	154
10	6-F-cortisol	0.60	0.75	-0.15	1.74	150
11	6-F-prednisolone	0.95	0.81	0.14	1.43	148
12	6,9-di-F-16-OH-prednisolone ^a	0.70	2.46	-1.77	0.71	160
13	9-F-16-Me-prednisolone	1.08	1.11	-0.04	1.75	154
14	6,9-di-F-16-Me-prednisolone	1.48	1.63	-0.15	1.79	160
15	9-F-cortisol	0.90	0.91	-0.01	1.54	150
16	9-F-prednisolone	1.30	0.97	0.33	1.23	148
17	9-F-21-deoxy-prednisolone	-0.30	0.01	-0.32	1.75	142
18	9-F-16-OH-prednisolone ^a	0.70	1.95	-1.25	0.67	154
19	9-F-16-Me-21-deoxy-prednisolone	0.70	0.16	0.54	2.27	148

EC₅₀ To Induce a Repetitive Train of Impulses and Conduction Blockage by Benzyl Chrysanthemates in Cockroach Nerves. Data from Nishimura et al.¹⁹ (Table 10).

Table 8. Paralysis of Rabbit Diaphragm into Phrenic Nerve by Injection of 16

$$R_3N(CH_2)_n$$
 NR_3

	substituent	S				
no.	R	\overline{n}	obsd pred (eq 13) Δ		Δ	NVE
1	Me, Me, Me	0	-1.00	-1.00	0.00	94
2	Me, Me, Me	1	-0.70	-0.79	0.06	100
3	Me, Me, Me	2	-0.70	-0.59	0.11	106
4	Et, Me, Me	2	-0.22	-0.17	-0.05	118
5	Et, Et, Me	2	0.40	0.24	0.16	130
6	Et, Et, Et	2	0.48	0.65	-0.17	142
7	Me, Me, Me	3	-0.70	-0.38	-0.32	112
8	Et, Me, Me	3	0.00	0.03	-0.03	124
9	Et, Et, Me	3	0.60	0.44	-0.16	136
10	Et, Et, Et	3	0.78	0.86	-0.08	148
11	Me, Me, Me	4^a	0.48	-0.17	0.65	118
12	Et, Me, Me	4	0.18	0.24	-0.06	130
13	Et, Et, Me	4	0.70	0.65	0.05	142
14	Et, Et, Et	4	1.30	1.06	0.24	154
15	Me, Me, Me	5^a	0.78	0.03	0.75	124
16	Et, Et, Et	5	1.00	1.27	-0.27	160
17	Me, Me, Me	6	0.70	0.24	0.46	130
18	Et, Et, Et	6	1.40	1.47	-0.07	166

Table 9. Inhibition of Cockroach Nerve Conduction by Miscellaneous Chemicals¹⁸

a Outlier.

			pred			
no.		log 1/C	(eq 14)	Dev	NVE	
1	C ₂ H ₅ OH	-0.01	-0.01	0.00	20	
2	C_3H_7OH	0.41	0.49	-0.08	26	
3	C ₄ H ₉ OH	1.05	0.99	0.06	32	
4	$C_5H_{11}OH$	1.55	1.49	0.06	38	
5	$C_6H_{13}OH$	2.08	2.00	0.09	44	
6	$C_7H_{15}OH$	2.71	2.50	0.21	50	
7	$C_8H_{17}OH$	2.92	3.00	-0.08	56	
8	L-menthol ^a	2.80	3.83	-1.03	66	
9	thymol	3.52	3.33	0.19	60	
10	gamma benzenehexachloride	3.40	3.83	-0.43	66	
11	delta-benzenehexachloride	4.00	3.83	0.17	66	
12	ether	0.82	0.99	-0.17	32	
a O 41'						
	Outlier.					

$$\log 1/C = 0.97(\pm 0.31) \text{ Mlog P} + 0.070(\pm 0.053) \text{ NVE} - 0.85(\pm 4.63)$$
 (15)

$$n = 15$$
, $r^2 = 0.829$, $s = 0.369$, $q^2 = 0.720$
outliers: $X = 6$ -Cl-3-pyridyl, $Y = CHNO_2$,
 $Z = NH$; $X = CH_2CH_2$ -(6-Cl-3-pyridyl), $Y = CHNO_2$, $Z = NH$; $X = CH_2$ -(6-Cl-3-pyridyl), $Y = NNO_2$, $Z = NHCH_2$; $X = CH_2$ -(6-Cl-3-pyridyl), $Y = CHNO_2$, $Z = O$

Minimum Blocking Concentration To Suppress Action Potential in Excised Central Nerve Cord of Cockroach Below 1 MV by Pyrethroids. Data from Nishimura et al.²⁰ (Table 11).

$$\log 1/C = 0.25(\pm 0.10) \operatorname{Clog} P - 0.36(\pm 0.32) \operatorname{B1}_3 + 0.020(\pm 0.008) \operatorname{NVE} + 1.47(\pm 0.95)$$
 (16)

$$n = 12$$
, $r^2 = 0.938$, $s = 0.120$, $q^2 = 0.892$
outliers: CH₂C₆H₅; NO₂

Table 10. EC50 To Induce a Repetitive Train of Impulses and Conduction Blockage by

				log 1/C				
	substitue	nts			pred		Mlog	
no.	X	Y	Z	obsd	(eq 15)	Δ	P	NVE
1	CH ₂ -3-Pyridyl	CHNO ₂	NH	4.54	4.07	0.47	-1.02	84
2	CH ₂ -(6-Cl-3-pyridyl)	CHNO ₂	NH	5.65	5.30	0.35	-0.19	90
3	CH2-(6-Me-3- pyridyl)	CHNO ₂	NH	4.94	4.91	0.03	-0.59	90
4	6-Cl-3-pyridyl	$CHNO_2$	NH^a	6.10	4.84	1.26	-0.23	84
5	CH ₂ CH ₂ -(6-Cl-3- pyridyl)	CHNO ₂	NH ^a	4.12	5.68	-1.56	-0.23	96
6	CH ₂ -2-pyridyl	$CHNO_2$	NH	3.70	4.22	-0.52	-0.87	84
7	CH ₂ -4-pyridyl	$CHNO_2$	NH	4.09	4.05	0.05	-1.05	84
8	benzyl	$CHNO_2$	NH	4.66	5.38	-0.72	0.33	84
9	4-Cl-benzyl	$CHNO_2$	NH	6.04	6.44	-0.40	0.99	90
10	CH ₂ -[5-(2-Cl)- thiazolyl]	CHNO ₂	NH	5.35	5.16	0.19	-0.04	86
11	CH ₂ -3-pyridyl	NNO_2	NH	4.80	4.88	-0.08	-0.19	84
12	CH ₂ -(6-Cl-3-pyridyl)	NNO_2	NH	6.30	6.05	0.25	0.59	90
13	CH ₂ -(6-Cl-3-pyridyl)	NNO_2	$ \begin{array}{c} \text{NH-}\\ \text{CH}_2{}^a \end{array} $	3.86	6.15	-2.29	0.26	96
14	CH ₂ -(6-Cl-3-pyridyl)	NCN	NH	5.82	5.66	0.16	0.77	82
15	CH ₂ -(6-Cl-3-pyridyl)	CHCN	NH	5.34	5.23	0.11	0.32	82
16	CH ₂ -(6-Cl-3-pyridyl)	$CHNO_2$	CH_2	6.25	6.14	0.11	0.68	90
17	CH ₂ -(6-Cl-3-pyridyl)	$CHNO_2$	O^a	5.22	6.00	-0.78	0.54	90
18	CH ₂ -(6-Cl-3-pyridyl)	$CHNO_2$	S	6.42	5.11	0.31	0.65	90
19	CH ₂ -(6-Cl-3-pyridyl)	CHNO ₂	NH	4.99	5.30	-0.31	-0.62	96

a Outlier.

Table 11. Minimum Blocking Concentration To Suppress Action Potential of Cockroach Nerve below 1 MV by Pyrethroids²⁰

$$CH_2$$
- OC Me Me C = $CCOOMe$ Me

			log 1/C				
			pred				
no.	substituents X	obsd	(eq 16)	Δ	Clog P	B1-3	NVE
1	Н	4.61	4.57	0.04	4.24	1.00	118
2	F	4.39	4.60	-0.21	4.38	1.35	124
3	Br	4.56	4.57	-0.01	5.11	1.95	124
4	Me	4.78	4.63	0.15	4.74	1.52	124
5	C_2H_5	4.80	4.89	-0.09	5.27	1.52	130
6	$CH_2C_6H_5$	5.27	5.59	-0.32	6.31	1.52	152
7	OMe	4.83	4.67	0.16	4.16	1.35	130
8	OC_2H_5	4.84	4.92	-0.08	4.69	1.35	136
9	$OCHMe_2$	5.11	5.12	-0.01	5.00	1.35	142
10	OC_6H_5	5.67	5.66	0.01	6.34	1.35	152
11	COC_6H_5	5.32	5.27	0.05	5.28	1.92	156
12	NO_2^a	5.00	4.58	0.42	3.99	1.70	134
13	CN	4.42	4.38	0.05	3.68	1.60	126
14	SO_2Me	4.23	4.28	-0.05	2.60	2.06	142
a	Outlier.						

B1 is the Verloop sterimol parameter (ref 2, pp 76–78). *Inhibition of* [³H]Quinuclidinyl Benzylate Binding in Rat Forebrain Membrane by (Y-Pyrrole) Amino-pyridin-4-yls. Data from Kushwana et al.¹⁷ (Table 12).

$$\log 1/C = -0.53(\pm 0.15) \ \sigma^*$$
-sum + $0.018(\pm 0.011) \ \text{NVE} + 3.65(\pm 0.95) \ (17)$

$$n = 12, r^2 = 0.912, s = 0.183, q^2 = 0.799$$

outliers: X = Me, Y = 2-CHO; X = C₂H₅, Y = H; X = Me, Y = 2-CN

Table 12. IC₅₀ Inhibition of [³H]Quinuclidkinyl Benzilate Binding to Rate Forebrain Membrane by

	subs	tituents		S'-		
no.	X	Y	obsd	pred (eq 17)	Δ	sum
1	C ₄ H ₉	Н	4.90	4.60	0.30	0.85
2	Me	2-CH=CHC ₆ H ₅	4.77	5.01	0.24	0.90
3	Me	2-CH=CH ₂	4.74	4.52	0.22	0.89
4	C_3H_7	H	4.67	4.57	0.10	0.86
5	Me	$2-C_2H_5$	4.66	4.82	-0.16	0.39
6	Me	3-CH(OH)Me	4.62	4.63	0.01	0.95
7	$CH_2CH=CH_2$	Н	4.57	4.41	0.16	1.10
8	Me	2-CHO^a	4.28	3.59	0.70	2.64
9	Me	H	4.15	4.29	-0.14	0.98
10	C_2H_5	H^a	3.92	4.45	-0.53	0.88
11	C_3H_7	3-CHO	3.86	3.87	0.01	2.52
12	Н	H	3.73	3.93	-0.26	1.47
13	Me	3-CHO	3.73	3.59	0.15	2.64
14	Me	$2-CN^a$	3.53	2.77	0.76	4.13
15	H	2-CHO	3.14	3.23	-0.09	3.13
a (Outliers.					
и	Outners.					

 σ^* -sum is the σ^* for all substituents. This is Taft's definition of an inductive effect.² This is an interesting study in that the compounds studied possessed anti-Alzheimer activity. The most active compound was $X = C_4H_9$, Y = H. It would be interesting to test compounds with higher NVE and electron releasing ability such as Y = OC₄H₉, X = 2-CH=CHOMe.

SUMMARY

Polarizability is a property of chemicals that has received almost no attention, outside of our laboratory, by those studying chemical-biological interactions. This despite the fact that an algorithm for calculating molar refractivity has been available for many years on the widely used Clog P program.⁶ As we have pointed out⁵ there is surprisingly little interest in those developing new software for drug design in considering specific electronic interactions of any kind between simple chemicals and the more complex chemicals that make up living systems.

Our database contains 2400 biological QSAR that are based on the Hammett electronic parameters, 373 based on NVE and 2311 based on MR or CMR. Where these are a measure of polarizability based on the Lorentz-Lorenz equation. Thus, there is strong support for consideration of electronic effects in chemical biological interactions. Another surprising aspect of our findings is that good correlations can be obtained with whole animals, including humans! This highlights the unique character of the synaptic site associated with NVE. Recent studies of this region reveal a very complex mode of its operation.^{21,22}

Another important observation of the present study as well as unpublished results is that our fear of elements beyond F (e.g., Cl, Br, I, S, P) would be more easily polarizable and hence would not fit QSAR based on the smaller elements is not justified. An important aspect of having a database of over 10 000 QSAR for chemical-biological interactions is that one can almost instantly retrieve long forgotten QSAR to support new ideas. Traditional QSAR is far from being a static subject.

REFERENCES AND NOTES

- (1) Pauling, L. The Nature of the Chemical Bond, 3rd ed.; Cornell University Press: 1969; p 607.
- (2) Hansch, C.; Leo, A. Exploring QSAR. Fundamentals and Applications in Chemistry and Biology; American Chemical Society: 1995; p 78.
- (3) Zissimos, A. M.; Abraham, M. H.; Barker, M. C.; Box, K. J.; Tam, K. Y. Calculation of Abraham Descriptors from Solvent-Water Partition Coefficients in Four Different Systems; Evaluation of Different Methods of Calculation. J. Chem. Soc., Perkin Trans. 2 2002, 470 - 477.
- (4) Agin, D.; Hersh, L.; Holtzman, D. The Action of Anesthetics on Excitable Membranes; A Quantum-chemical Analysis. Proc. Natl. Acad. Sci. U.S.A. 1965, 53, 952-958.
- (5) Hansch, C.; Steinmetz, W. E.; Leo, A. J.; Mekapati, S. B.; Kurup, A.; Hoekman, D. On the Role of Polarizability in Chemical-Biological Interactions. J. Chem. Inf. Comput. Sci. 2003, 43, 120-125.
- (6) BioByte Corporation. 201 E. Fourth Street, Suite 204, Claremont, CA 91711 U.S.A.
- Principles of Medicinal Chemistry; Foye, W. O., Ed.; Lea and Febiger: Philadelphia, PA, 1974; pp 165-170.
- (8) Textbook of Organic and Pharmaceutical Chemistry, 6th ed.; Wilson, C. O., Gisvold, O., Doerge, R. E., Eds.; Lippencott: Philadelphia, PA, 1971; p 405.
- (9) Cope, A. C.; Hancock, E. M. Substituted Vinyl Barbituric Acids III. Derivatives Containing a Dialkylvinyl Group having Five or More Carbon Atoms. J. Am. Chem. Soc. 1939, 61, 776-779.
- (10) Watari, N.; Sugiyama, Y.; Kaneniwa, N.; Hiura, M. Prediction of Hepatic First-Pass Metabolism and Plasma Levels Following Intravenous and Oral Administration of Barbiturates in the Rabbit Based on Quantitative Structure-Pharmacokinetic Relationships. J. Pharmacok. Biopharm. 1988, 16, 279-301.
- (11) Dinno, M. A.; Holloman, T. L.; Schwartz, M. Potency of Barbiturates in Inhibition of Frog Gastric Secretion. Proc. Soc. Exptl. Biol. Med. **1972**, 141, 397-399.
- (12) Hansch, C.; Steward, A. R.; Anderson, S. M.; Bently, D. The Parabolic Dependence of Drug Action upon Lipophilic Character as Revealed by a Study of Hypnotics. *J. Med. Chem.* **1967**, *11*, 1–11.
- (13) King, L. A. Fergusons Principle and the Prediction of Fatal Drug Levels in Blood. Human Toxicol. 1985, 4, 273-278.
- (14) King, L. A. The Use of Drug Solubility Data in Forensic Toxicology. Meeting of the International Association of forensic Toxicologists. August 25-30 1985, Rigi Kalbad, Switzerland. R. Bandenberger,
- (15) Ahmad, P.; Mellors, A. Glucocorticoid Potency and Parachor. J. Steroid Biochem. 1976, 7, 19-28.
- (16) Wien, R.; Mason, D. F. J. The Pharmacological Actions of a Series of Phenyl Alkane p- ω -Bistrialkylammonium Compounds. Brit. J. Pharmacol. 1953, 8, 306-314.
- (17) Kushwaha, P. S.; Shukla, M. K.; Mishra, P. C. A Geometry Optimization and Molecular Electrostatic Potential Mapping Study of Structure-Activity Relationship for some Anti-Alzheimer Agents. Indian J. Biochem. Biophys. 1999, 36, 101-106.
- (18) Uchida, M.; Kurihara, M.; Fujita, T.; Nakajin, J. M. Inhibitory Effects of BHC Isomers on Na+-K+-ATP-Ase, Yeast Growth, and Nerve Conduction. Pestic. Biochem. Physiol. 1974, 4, 260-265.
- (19) Nishimura, K.; Kanda, Y.; Okazawa, A.; Ueno, T. Relationship Between Insecticidal Neurophysiological Activities of Imidacloprid and Related Compounds. Pestic. Biochem. Physiol. 1994, 50, 51-59.
- (20) Nishimura, K.; Ohoka, M.; Fujita, T. Quantitative Structure—Activity Studies of Pyrethroids. Pestic. Biochem. Physiol. 1987, 28, 257-270.
- (21) Schoch, S.; Castillo, P. E.; Jo, T.; Mukherjee, K.; Geptert, M.; Wang, Y.; Schmitz, F.; Malenka, R. C.; Südhof, T. C. R1M1α Forms a Protein Scafford for the Regulating Neurotransmitter Release at the Active Zone. Nature 2002, 415, 321-326.
- (22) Castillo, P. E.; Shoch, S.; Schmitz, F.; Südhof, T. C.; Melenka, R. C. R1M1a is Required for Presynaptic Long-term Potentiation. Nature **2002**, 415, 327-330.

CI030289E