See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/283576233

Betulinic acid derived hydroxamates and betulin derived carbamates are interesting scaffolds for the synthesis of novel cytotoxic compounds

ARTICLE in EUROPEAN JOURNAL OF MEDICINAL CHEMISTRY · NOVEMBER 2015

Impact Factor: 3.45 · DOI: 10.1016/j.ejmech.2015.10.043

READS

30

6 AUTHORS, INCLUDING:



Ralph Kluge

Martin Luther University Halle-Wittenberg

86 PUBLICATIONS **680** CITATIONS

SEE PROFILE



Dieter Ströhl

Martin Luther University Halle-Wittenberg

77 PUBLICATIONS 521 CITATIONS

SEE PROFILE



René Csuk

Martin Luther University Halle-Wittenberg

212 PUBLICATIONS 1,919 CITATIONS

SEE PROFILE

FISEVIER

Contents lists available at ScienceDirect

European Journal of Medicinal Chemistry

journal homepage: http://www.elsevier.com/locate/ejmech



Research paper

Betulinic acid derived hydroxamates and betulin derived carbamates are interesting scaffolds for the synthesis of novel cytotoxic compounds



Jana Wiemann, Lucie Heller, Vincent Perl, Ralph Kluge, Dieter Ströhl, René Csuk*

Martin-Luther-University Halle-Wittenberg, Organic Chemistry, Kurt-Mothes-Str. 2, D-06120 Halle (Saale), Germany

ARTICLE INFO

Article history:
Received 27 August 2015
Received in revised form
20 October 2015
Accepted 26 October 2015
Available online 5 November 2015

Keywords: Triterpenes Betulinic acid Betulin Hydroxamates Carbamates Tumor cells

ABSTRACT

The betulinic acid-derived hydroxamates **5–18**, the amides **19–24**, and betulin-derived bis-carbamates **25–28** as well as the carbamates **31–40** and **44–48** were prepared and evaluated for their antiproliferative activity in a photometric sulforhodamine B (SRB) assay against several human cancer cell lines and nonmalignant mouse fibroblasts (NIH 3T3). While for 3-*O*-acetyl hydroxamic acid **5** EC₅₀ values as low as EC₅₀ = 1.3 μ M were found, *N*,*O*-bis-alkyl substituted hydroxamates showed lowered cytotoxicity (EC₅₀ = 16–20 μ M). In general, hydroxamic acid derivatives showed only reduced selectivity for tumor cells, except for allyl substituted compound **13** (EC₅₀ = 5.9 μ M for A2780 human ovarian carcinoma cells and EC₅₀ > 30 μ M for nonmalignant mouse fibroblasts). The cytotoxicity of betulinic acid derived amides **19–24** and of betulin derived bis-carbamates **25–28** was low, except for *N*-ethyl substituted **25**. Hexyl substituted **39** showed EC₅₀ = 5.6 μ M (518A2 cells) while for mouse fibroblasts EC₅₀ > 30 was determined.

© 2015 Elsevier Masson SAS. All rights reserved.

1. Introduction

Hydroxamic acids have been known since Lossen's discovery of oxalohydroxamic acid (Fig. 1) in 1869 [1,2]. Then and for many years to follow they were regarded as rather exotic compounds. Nowadays, they have been recognized [3] as a unique family of compounds that hold a wide spectrum of biological activities. They act as selective inhibitors of many enzymes, such as matrix metalloproteinases [4,5], hydrolases [6], ureases [7], lipoxygenase [8], tumor necrosis factor- α converting enzyme [9,10], carbonic anhydrase [11,12], ribonucleotide reductase [13,14] and many others. Their acidity is much weaker than that of structurally related carboxylic acids but the hydroxamic acid moiety may act as a bidentate ligand to chelate with several metal ions. It also controls multiple sites for potential hydrogen bond interactions with enzymes and recentors [15]

Nonspecific hydroxamic acid derivatives have been prepared starting inter alia from benzodiazepines [16], α -amino-suberic acids [17], N-alkylated amino acids [18] as well as from arylsulfonamides [19]. Although triterpenoic acids represent an important

E-mail address: rene.csuk@chemie.uni-halle.de (R. Csuk).

class of compounds bearing high potential as antitumor-active compounds [20,21], to our knowledge, there are only two reports describing a hydroxamic acid derived from glycyrrhetinic acid. These compounds (Fig. 1) acted as selective inhibitors of 11 β -hydroxysteroid dehydrogenase 2; there were no data provided whether these compounds were cytotoxic.

During our continuing search for antitumor active compounds from natural products [20–22] we became interested in the synthesis of betulinic acid (**BA**, **1**, Fig. 1) derived hydroxamic acid and derivatives and their cytotoxicity. Previous studies suggested introducing a sulfamate [23,24] or carbamate [25,26] to the pentacyclic skeleton of triterpenes can significantly improve the cytotoxicity of these compounds. Thus, we decided to prepare several mono- and bis-carbamates derived from betulin (**2**, Fig. 1). Betulin as well as betulinic acid have been shown to be interesting scaffolds for developing analogs displaying various biological and medicinal properties especially potent anticancer effects [27–37].

2. Results and discussion

2.1. Chemistry

Our synthetic approach started from **BA** whose acetylation gave 3-O-acetyl-**BA** (3, Scheme 1) while from the Jones oxidation of **BA**

^{*} Corresponding author. Bereich Organische Chemie, Martin-Luther Universität Halle-Wittenberg, Kurt-Mothes-Str. 2, D-06120 Halle (Saale), Germany.

Fig. 1. Structure of Lossen's oxalohydroxamic acid (A) and a representative glycyrrhetinic acid derived hydroxamic acid (B), betulinic acid (BA, 1) and betulin (2).

betulonic acid (4) was obtained [38,39]. Treatment of **3** with oxalyl chloride in DCM for 2 h at 25 °C followed by adding hydroxylammonium chloride in the presence of trimethylamine [40] provided a 68% yield of hydroxamic acid **5**. Deacetylation of **5** with potassium hydroxide in methanol gave 63% of **6** [41].

While the reaction of $\bf 1$ with oxalyl chloride/hydroxylammonium chloride gave target compound $\bf 5$ nicely, the reaction of $\bf 1$ with propylphosphonic anhydride (T3P) or with $\bf 1,1'$ -carbonyldiimidazole failed to give high yields under a broad variety of different conditions [42–44].

Hydroxamic acid **5** is characterized in its ¹³C NMR spectrum by a signal at $\delta = 175.1$ ppm being assigned to the **CONHOH** moiety (C-28) of **5**. For comparison, C-28 was found in **3** at $\delta = 182.5$ ppm. This shifting of the resonance signal of carbonyl carbon C-28 to higher fields is typical for hydroxamic acids [45,46]. In the IR spectrum the C-O stretch vibration was detected at $\nu = 1643$ cm⁻¹.

Reaction of **3** with oxalyl chloride followed by reaction with *N*,*O*-dimethylhydroxylammonium chloride, or *N*-

Scheme 1. Synthesis of betulinic acid derived hydroxamic acids **5–18**: a) Ac_2O , NEt_3 , pyridine, DMAP, 12 h, 25 °C, 75%; b) Jones oxidation (4 h, 25 °C), 81%; c) oxalyl chloride, DCM, 2 h, 25 °C; then NHR¹ OR^2 , NEt_3 , DCM, 2-12 h, 25 °C; **5** (from NH $_2OH$ -HCI, 68%), **7** (from HNMeOM-HCI, 77%), **9** (from HNMeOH-HCI, 66%), **11** (from NH $_2OM$ -HCI, 63%), **13** (from NH $_2OM$ -HCI, 90%), **15** (from HNMeOM-HCI, 52%), **16** (from HNMeOH-HCI, 81%), **17** (from NH $_2OM$ -HCI, 45%), **18** (from NH $_2OM$ -HCI, 68%); d) NOH in MeOH, 25 °C: **6** (4 d, 63%), **8** (5 d, 89%), **10** (5 d, 60%), **12** (5 d, 95%), **14** (7 d, 94%).

methylhydroxylammonium chloride or *O*-methylhydroxylammonium chloride or *O*-allylhydroxylammonium chloride in the presence of triethylamine furnished products **7**, **9**, **11** and **13**; their deacetylation yielded compounds **8**, **10**, **12** or **14**, respectively.

Under similar conditions betulonic acid (**4**) gave substituted hydroxamic acids **15–18**. Yields dropped slightly for these reactions because of the accompanying formation of C-3-oximes.

For comparison, we prepared several betulinic acid derived amides as well as betulin derived carbamates. Reaction of 3-0acetyl-betulinic acid (3) with oxalyl chloride (Scheme 2) followed by a reaction with dry ammonia in DCM furnished amide 19 in 95% yield. Deacetylation of 19 with potassium hydroxide in methanol gave amide **20** [47,48]. In a similar way, from the reaction of **3** with oxalyl chloride and benzylamine, benzylamide 21 was obtained whose deacetylation yielded 22; the Jones oxidation of 21 gave 3oxo compound 23. Following the procedure given for the synthesis of 12, from betulonic acid (4) with dry ammonia in DCM 3-oxoamide **24** [49–52] was obtained. Compound **19** is characterized in its ¹H NMR spectrum by the presence of a signal at $\delta = 5.55$ ppm that was assigned to the CONH2 moiety. In addition, the signal for H-19 was shifted to lower fields (compared to parent compound 3 showing $\Delta \delta = 0.08$ ppm). In the ¹³C NMR spectrum the **C**ONH₂ moiety was detected at $\delta = 179.3$ ppm.

Previous studies suggested the introduction of a sulfamate [23] or carbamate [25] to the skeleton of a pentacyclic triterpenoid can significantly improve its cytotoxicity. Thus, we decided to prepare several bis-carbamates. The synthesis of these bis-carbamates 25–29 started from betulin (2, Scheme 3). While the reaction of betulin with ethyl isocyanate in refluxing chloroform for 48 h gave only low yields, the microwave assisted reaction of 2 with ethyl isocyanate in dry THF worked nicely, and 3,28-bis-*N*-ethyl-carbamate 25 was obtained in 81% isolated yield. Similarly biscarbamates 26–28 were prepared.

For the synthesis of 3-*O*-acetyl-28-*N*-alkyl-carbamates **31**—**35**, 3-*O*-acetyl-betulin (**30**) was used as a starting material. Compound **30** is easily accessible from betulin; thus, diacetylation of betulin gave diacetate **29** whose selective deacetylation with KOH in MeOH/THF at 0 °C yielded 57% of monoacetate **30**.

The microwave assisted reaction of **30** with alkyl isocyanates or phenyl isocyanate allowed a quick and reliable synthesis of 3-*O*-acetylated 28-*N*-substituted carbamates **31–35**. From their deacetylation with potassium hydroxide in MeOH compounds **36–40** were obtained.

The synthesis of 3-oxo-28-*N*-alkyl-carbamates **44**—**48** (Scheme 4) started from 3-oxo-betulin **43**, and the microwave assisted reaction of **43** with alkyl isocyanates in THF gave target compounds **44**—**48** in good yields. The starting material for these reactions, **43**, was obtained from a deacetylation reaction of **42** (KOH in MeOH) in 90% isolated yield. Compound **42** was easily prepared by Jones oxidation of **41** (72% yield); the latter was made by a selective acetylation of betulin in 60% isolated yield.

2.2. Biology

The betulinic acid-derived hydroxamates **5–18**, the amides **19–24**, and betulin-derived bis-carbamates **25–28** as well as the carbamates **31–40** and **44–48** were evaluated for their antiproliferative activity in a photometric sulforhodamine B (SRB) assay [53–56] against several human cancer cell lines and nonmalignant mouse fibroblasts (NIH 3T3). For comparison, betulinic acid (**1**), betulin (**2**), betulonic acid (**4**) and acetates **3**, **29** and **30** were included into this screening (Table 1).

Betulin (2) and its diacetate 29 displayed no (EC₅₀ > 30 μ M; cut-off of the assay) cytotoxicity; low activity was found for the betulin-monoacetate 30. Betulinic acid (1) is well-known for its

Scheme 2. Synthesis of betulinic or betulonic acid derived amides **19–24**: a) oxalyl chloride, DCM 2 h 25 °C, then NH₃, 1 h, 25 °C, 95% (for **19**) and 84% (for **24**); b) oxalyl chloride, DCM, 2 h, 25 °C, then BnNH₂, 1 h, reflux, 59%; c) KOH, MeOH, THF, 1 d, 25 °C, 97% (for **20**) and 92% (for **22**); d) Jones oxidation, 20 min, 25 °C, 74%; f) a) oxalyl chloride, DCM, 2 h, 25 °C, then NH₃, 1 h, 25 °C, 84%.

2
$$\frac{1}{R}$$
 $\frac{1}{H}$ $\frac{$

Scheme 3. Synthesis of betulin derived 3,28-bis-carbamates **25–28** and 28-mono-carbamates **31–40**: a) microwave assisted (7 h, 120 °C, THF, RNCO): **25** (from EtNCO, 81%), **26** (from PropNCO, 83%), **27** (from ButNCO, 81%), **28** (from HexNCO, 73%); b) Ac₂O, NEt₃, DMAP, DCM, 12 h, 25 °C, 90%; c) KOH, MeOH, THF, 0 °C, 30 min, 57%; d) KOH, MeOH, THF, 25 °C: **36** (2 d, 95%), **37** (1 d, 93%), **38** (2 d, 94%), **39** (1 d, 88%, **40** (2 d, 89%).

Scheme 4. Synthesis of 3-oxo-betulin derived 28-mono-carbamates 44—48: a) Ac₂O, NEt₃, DMAP, 12 h, 25 °C, 60%; b) Jones oxidation, 40 min, 25 °C, 72%; c) KOH, MeOH, THF, 12 h, 25 °C, 90%; d) miocrowave assisted (7 h, 120 °C, THF, RNCO): 44 (from EtNCO, 68%), 45 (from PropNCO, 62%), 46 (from ButNCO, 72%), 47 (from HexNCO, 55%), 48 (from PhNCO, 85%).

cytotoxicity, and EC_{50} values between 8.8 and 17.1 μ M were determined while for betulonic acid (**4**) EC_{50} values between $EC_{50} = 13.1-29.0$ μ M were found. Betulinic acid 3-acetate **3** showed higher cytotoxicity, especially for 518A2 human melanoma cells ($EC_{50} = 4.5$ μ M). Betulinic acid derived 3-*O*-acetyl hydroxamic acid **5** showed significantly higher cytotoxicity, and

 EC_{50} values as low as $EC_{50}=1.3~\mu M$ were found. Bis-alkyl substitution (as in 7) in the hydroxamate moiety lowered cytotoxicity ($EC_{50}=16-20~\mu M$) significantly. The presence of at least one acidic hydrogen substituent in the hydroxamic acid part seems mandatory to obtain cytotoxic compounds: Compounds 9 and 11 gave EC_{50} values between $EC_{50}=2.0-13.5~\mu M$. For allyl-

Table 1 Cytotoxicity of betulinic acid-derived hydroxamates **5–18** and starting materials **1–4, 29, 30** (EC₅₀ values in μM from SRB assays after 96 h of treatment; the values are averaged from three independent experiments performed each in triplicate; confidence interval CI = 95%). The cell lines are human cancer cell lines: 518A2 (melanoma), A2780 (ovarian carcinoma), A549 (alveolar basal epithelial adenocarcinoma), MCF7 (breast adenocarcinoma) and non-malignant mouse fibroblasts (NIH 3T3).

•		,			` ,
	518A2	A2780	A549	MCF7	NIH 3T3
1	9.4 ± 0.7	8.8 ± 0.9	17.1 ± 1.1	10.2 ± 1.2	16.1 ± 1.4
2	>30	>30	>30	>30	>30
3	4.5 ± 0.1	18.3 ± 0.5	6.2 ± 0.4	11.0 ± 0.5	>30
4	29.0 ± 2.0	20.4 ± 1.9	25.9 ± 2.5	25.9 ± 2.0	13.1 ± 0.9
29	>30	>30	>30	>30	>30
30	15.2 ± 1.6	11.0 ± 1.3	14.0 ± 2.4	11.2 ± 1.4	>30
5	1.6 ± 0.1	1.3 ± 0.1	1.3 ± 0.1	1.4 ± 0.1	1.1 ± 0.1
6	2.4 ± 0.2	3.7 ± 0.5	5.3 ± 0.6	3.9 ± 0.6	2.0 ± 0.5
7	17.5 ± 0.6	19.3 ± 1.1	20.1 ± 1.4	16.4 ± 0.9	17.4 ± 1.1
8	7.3 ± 0.5	7.6 ± 0.3	19.0 ± 3.0	10.2 ± 1.4	7.7 ± 1.3
9	3.5 ± 0.3	2.0 ± 0.1	4.3 ± 0.7	2.3 ± 0.3	2.9 ± 0.6
10	Not soluble				
11	8.0 ± 0.6	8.9 ± 0.5	11.7 ± 2.0	13.5 ± 0.9	9.5 ± 0.5
12	5.8 ± 1.2	12.3 ± 0.9	6.9 ± 0.2	21.9 ± 0.7	15.2 ± 0.5
13	7.6 ± 0.1	5.9 ± 0.6	6.6 ± 1.5	12.2 ± 2.0	>30
14	6.7 ± 0.1	7.3 ± 0.7	4.1 ± 1.7	8.6 ± 1.0	6.1 ± 0.4
15	>30	20.7 ± 1.6	14.5 ± 1.3	18.7 ± 1.2	26.7 ± 2.0
16	6.3 ± 0.2	3.0 ± 0.2	6.0 ± 0.3	10.2 ± 0.9	5.0 ± 0.4
17	17.2 ± 1.6	12.0 ± 1.5	6.1 ± 1.1	13.8 ± 1.6	12.7 ± 1.1
18	22.4 ± 1.5	19.7 ± 2.0	25.0 ± 2.1	18.4 ± 1.9	19.7 ± 1.7

substituted **14** EC_{50} values of the same magnitude as for **11** were found (Table 2).

Deacetylated compound **6** also gave low EC₅₀ values, but this compound was less active than parent **5**. Substituted hydroxamic acids **8**, **12** and **14** were of similar cytotoxicity. Despite the low EC₅₀ values, hydroxamic acid derivatives showed only reduced selectivity for tumor cells, except for allyl substituted compound **13** (EC₅₀ = 5.9 μ M for A2780 human ovarian carcinoma cells and EC₅₀ > 30 μ M for nonmalignant mouse fibroblasts) (Table 3).

The cytotoxicity of amides **19–24**, however, was moderate; the lowest EC₅₀ value was determined for 3-*O*-acetylated compounds **19** and **21**. The cytotoxicity of betulin derived bis-carbamates **25–28** was also low, except for *N*-ethyl substituted **25**; the same was observed for their 3-*O*-acetylated analogues **31–35** and their 3-oxo-analogues **44–48**.

Significantly improved cytotoxicity was found for carbamates **36–40** holding a free hydroxyl group at position C-3. While almost all of these compounds showed more or less the same cytotoxicity for tumor cell lines as well as for mouse fibroblasts, hexyl substituted **39** showed EC₅₀ = 5.5 μ M (518A2 cells) while for mouse fibroblasts EC₅₀ > 30 was determined.

3. Conclusion

The betulinic acid-derived hydroxamates 5–18, the amides

19-24, and betulin-derived bis-carbamates 25-28 as well as the carbamates 31-40 and 44-48 were prepared and evaluated for their antiproliferative activity in a photometric sulforhodamine B (SRB) assay against several human cancer cell lines and nonmalignant mouse fibroblasts (NIH 3T3). While betulin (2) and its diacetate 29 exhibited no cytotoxicity, low activity was found for the betulin-monoacetate **30**. Betulinic acid 3-acetate **3** showed higher cytotoxicity than its parent compound betulinic acid, but significantly lower EC₅₀ values were determined for 3-0-acetyl hydroxamic acid **5**, and EC₅₀ values as low as EC₅₀ = 1.3 μ M were found. Bis-alkyl substitution in the hydroxamate moiety lowered cytotoxicity (EC₅₀ = $16-20 \mu M$). The presence of at least one acidic hydrogen substituent in the hydroxamic acid part seems mandatory to obtain cytotoxic compounds. Despite their low EC₅₀ values, hydroxamic acid derivatives showed only reduced selectivity for tumor cells, except for allyl substituted compound 13 $(EC_{50} = 5.9 \mu M \text{ for A2780 human ovarian carcinoma cells and}$ $EC_{50} > 30 \,\mu\text{M}$ for nonmalignant mouse fibroblasts). The cytotoxicity of betulinic acid derived amides 19-24 as well as of betulin derived bis-carbamates 25–28 was low, except for N-ethyl substituted 25. While almost all of these compounds showed more or less the same cytotoxicity for tumor cell lines as well as for mouse fibroblasts, hexyl substituted **39** showed $EC_{50} = 5.6 \mu M$ (518A2 cells) while for mouse fibroblasts $EC_{50} > 30$ was determined.

4. Experimental section

4.1. General - chemistry

Melting points are uncorrected (Leica hot stage microscope), NMR spectra were recorded using the Varian spectrometers Gemini 2000 or Unity 500 (δ given in ppm, J in Hz, internal Me4Si, MS spectra were taken on a Finnigan MAT LCQ 7000 (electrospray, voltage 4.1 kV, sheath gas nitrogen) instrument. The optical rotation was measured on a Perkin–Elmer polarimeter at 20 °C; TLC was performed on silica gel (Merck 5554); elemental analyses were performed on a Vario EL (CHNS). The solvents were dried according to usual procedures. The purity of the compounds was determined by HPLC and found to be >98%. Betulinic acid (1) and betulin (2) and were obtained from different commercial suppliers in bulk quantities. Microwave assisted reactions were performed in a Monowave 300 apparatus (Anton Parr, Austria).

4.2. General – biological screening

The SRB assay was performed as previously described [53–56].

4.3. Syntheses

4.3.1. (3β) 3-Acetyloxy-lup-20(29)-en-28-acid (**3**)
Prepared by reaction (12 h, 25 °C) of **1** (4.0 g, 8.78 mmol) with

Table 2 Cytotoxicity of betulinic acid derived amides 19–24 (EC₅₀ values in μM from SRB assays after 96 h of treatment; the values are averaged from three independent experiments performed each in triplicate; confidence interval CI = 95%). The cell lines are human cancer cell lines: 518A2 (melanoma), A2780 (ovarian carcinoma), HT29 (colorectal adenocarcinoma), MCF7 (breast adenocarcinoma), A549 (alveolar basal epithelial adenocarcinoma), HeLa (epitheloid cervix carcinoma) and non-malignant mouse fibroblasts (NIH 3T3).

	518A2	A2780	HT-29	MCF7	A549	HeLa	NIH 3T3
19	13.4 ± 1.1	5.7 ± 0.6	11.5 ± 0.7	11.1 ± 0.6	12.4 ± 1.4	12.0 ± 1.1	20.0 ± 1.3
20	9.9 ± 0.7	15.7 ± 1.4	10.2 ± 1.3	10.7 ± 0.5	8.2 ± 0.9	11.2 ± 1.2	10.7 ± 0.6
21	6.2 ± 1.3	6.2 ± 1.1	>30	>30	10.3 ± 0.8	16.9 ± 1.8	>30
22	>30	11.8 ± 1.4	_	18.0 ± 2.1	>30	>30	11.1 ± 0.9
23	>30	7.4 ± 1.1	>30	>30	>30	>30	>30
24	21.5 ± 1.9	15.6 ± 1.4	17.9 ± 1.4	10.3 ± 1.1	14.7 ± 0.9	22.9 ± 1.7	17.7 ± 1.1

Table 3
Cytotoxicity of betulin derived bis-carbamates 25–28 and mono-carbamates 31–40 and 44–48 (EC₅₀ values in μM from SRB assays after 96 h of treatment; the values are averaged from three independent experiments performed each in triplicate; confidence interval CI = 95%). The cell lines are human cancer cell lines: 518A2 (melanoma), A2780 (ovarian carcinoma), HT29 (colorectal adenocarcinoma), MCF7 (breast adenocarcinoma), A549 (alveolar basal epithelial adenocarcinoma), HeLa (epitheloid cervix carcinoma) and non-malignant mouse fibroblasts (NIH 3T3).

	518A2	A2780	HT-29	MCF7	A549	HeLa	NIH 3T3
25	12.6 ± 1.3	5.8 ± 0.6	8.3 ± 0.9	23.3 ± 2.0	10.7 ± 1.3	17.8 ± 0.9	>30
26	>30	19.4 ± 2.0	>30	>30	>30	>30	>30
27, 28	>30	>30	>30	>30	>30	>30	>30
31	10.2 ± 1.3	11.1 ± 1.4	>30	20.4 ± 2.5	14.1 ± 1.2	17.0 ± 1.5	>30
32-35	>30	>30	>30	>30	>30	>30	>30
36	12.5 ± 1.6	4.6 ± 0.7	>30	12.1 ± 1.1	12.0 ± 1.6	>30	8.8 ± 1.0
37	15.9 ± 2.0	10.6 ± 1.5	16.3 ± 1.1	15.5 ± 1.4	14.2 ± 2.1	9.4 ± 0.9	7.3 ± 0.8
38	8.0 ± 1.1	7.0 ± 0.9	7.5 ± 1.0	12.5 ± 0.8	8.6 ± 0.5	10.1 ± 1.2	5.7 ± 0.7
39	5.5 ± 0.7	7.7 ± 0.9	>30	>30	12.7 ± 1.4	7.4 ± 1.1	>30
40	23.9 ± 1.4	7.2 ± 1.4	16.3 ± 2.5	24.6 ± 1.9	>30	11.5 ± 1.3	14.3 ± 2.0
44	16.6 ± 1.1	8.3 ± 0.9	11.5 ± 0.9	19.7 ± 2.5	20.3 ± 1.9	13.4 ± 1.2	4.6 ± 0.7
45-48	>30	>30	>30	>30	>30	>30	>30

NEt₃ (2.2 mL, 15.90 mmol), acetic anhydride (1.6 mL, 16.94 mmol) and catalytic amounts of DMAP in dry pyridine (80 mL), and **3** (3.28 g, 75%) was obtained as a colorless solid; $R_F = 0.60$ (silica gel, toluene/ethyl acetate/heptane/formic acid, 80:20:10:3); m.p.: 275–277 °C (lit.: [57] 277–278 °C; [α]_D = +20.54° (c = 0.58, CHCl₃) (lit.: [57] +22° (c = 0.49, CHCl₃); MS (ESI, MeOH): m/z = 497.4 (25%, [M – H]⁻), 543.1 (100%, [M + HCO₂]⁻), 995.3 (100%, [2M – H]⁻), 1017.8 (8%, [2M – 2H + Na]⁻).

4.3.2. Betulonic acid (4)

Prepared from **1** (13.1 g, 28.7 mmol) by Jones oxidation (4 h, 25 °C) as previously reported [38], followed by chromatographic workup (silica gel, hexanes/ethyl acetate, 9:1), and **4** (10.6 g, 81%) was obtained as a colorless solid; $R_F = 0.79$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 242–244 °C (lit.: [38] 245–247 °C; $[\alpha]_D = +39.42^\circ$ (c = 0.64, CHCl₃) (lit.: [57] $[\alpha]_D = +32^\circ$ (c = 0.37, CHCl₃), $[\alpha]_D = +40^\circ$ (c = 0.86, CHCl₃) [58]; MS (ESI, MeOH): m/z = 455.2 (100%, $[M+H]^+$), 508.9 (24%, $[M+Na+MeOH]^+$), 909.3 (16%, $[2M+H]^+$), 931.3 (46%, $[2M+Na]^+$).

4.3.3. (3β) 3-Acetyloxy-N-hydroxy-lup-20(29)-en-28-amide (5)

To an ice cold solution of 3 (0.39 g, 0.78 mmol) in dry DCM (25 mL), oxalyl chloride (0.40 mL, 4.70 mmol) was slowly added, and the mixture was allowed to warm to 25 °C and stirred for 2 h. The solvent was removed under diminished pressure, the residue dissolved in dry THF (20 mL), and the solvent was again removed. The residue was dissolved in dry DCM (25 mL) containing NEt₃ (0.70 mL, 5.05 mmol), and hydroxylammonium chloride (0.20 g, 2.90 mmol) was added. Stirring at 25 °C was continued for another 2 h. Usual aqueous workup gave a residue that was subjected to chromatography (silica gel, hexanes/ethyl acetate, 7:3), and 5 (0.27 g, 68%) was obtained as a colorless solid; $R_F = 0.36$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 207–210 °C; $[\alpha]_D = +9.02^\circ$ $(c = 0.58, CHCl_3)$; IR (KBr): $\nu = 3355br$, 3075m, 2943s, 2870s, 1736s, 1716s, 1643s, 1451s, 1377s, 1317w, 1265s, 1196m, 1152w, 1107m, 1066w, 1030s, 979s, 917m, 885m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.93 - 7.70$ (m, 2H, NH + OH), 4.73 (s, 1H, CH_a (29)), 4.60 (s, 1H, $CH_b(29)$, 4.50–4.42 (m, 1H, CH (3)), 3.03 (ddd, J = 11.1, 11.1, 4.2 Hz, 1H, CH (19)), 2.33 (ddd, J = 12.6, 12.6, 3.2 Hz, 1H, CH (13)), 2.03 (s, 3H, CH_3 (32)), 2.02–1.87 (m, 2H, CH_a (21) + CH_a (16)), 1.83–1.73 (m, 1H, CH_a (22)), 1.74–1.68 (m, 2H, CH_a (12) + CH_a (1)), 1.68 (s, 3H, CH_3 (30)), 1.66–1.28 (m, 12H, CH (18) + CH₂ (2) + CH_b (22) + CH₂ $(7) + CH_b (16) + CH_b (21) + CH_a (15) + CH_a (11) + CH_2 (6)$, 1.29-1.21 (m, 2H, CH_b (11) + CH (9)), 1.20-1.09 (m, 1H, CH_b (15)), 1.09-0.96 (m, 2H, CH_b (12) + CH_b (1)), 0.95 (s, 3H, CH_3 (27)), 0.93 (s, 3H, CH₃ (24)), 0.84 (s, 3H, CH₃ (26)), 0.83 (s, 3H, CH₃ (23)), 0.82 (s,

3H, CH_3 (25)), 0.80-0.74 (m, 1H, CH (5)) ppm; ^{13}C NMR (100 MHz, CDCl₃): $\delta = 175.1$ (C=0, C28), 171.1 (C=0, C31), 150.2 (C_q , C20), 109.8 (CH_2 =C, C29), 81.0 (CH, C3), 55.4 (CH, C5), 54.3 (C_q , C17), 50.4 (CH, C9), 50.4 (CH, C18), 46.9 (CH, C19), 42.4 (C_q , C14), 40.8 (C_q , C8), 38.4 (CH_2 , C1), 38.3 (CH_2 , C22), 38.0 (CH, C13), 37.8 (C_q , C4), 37.1 (C_q , C10), 34.3 (CH_2 , C7), 32.8 (CH_2 , C16), 30.7 (CH_2 , C21), 29.4 (CH_2 , C15), 27.9 (CH_3 , C23), 25.5 (CH_2 , C12), 23.7 (CH_2 , C7), 21.3 (CH_3 , C32), 20.8 (CH_2 , C11), 19.3 (CH_3 , C30), 18.2 (CH_2 , C6), 16.5 (CH_3 , C26), 16.2 (CH_3 , C25), 16.1 (CH_3 , C24), 14.6 (CH_3 , C27) ppm; MS (ESI, MeOH): m/z = 514.4 (100%, $[M+H]^+$), 536.3 (30%, $[M+H]^+$), 1027.3 (50%, $[2M+H]^+$), 1049.4 (90%, $[2M+Na]^+$); analysis calcd for $C_{32}H_{51}NO_4$ (513.75); C 74.81, H 10.01, N 2.73; found: C 74.63, H 10.17, N 2.51.

4.3.4. (3β) 3-Hydroxy-N-hydroxy-lup-20(29)-en-28-amide (**6**)

To a solution of potassium hydroxide (0.28 g, 5.00 mmol) in methanol (25 mL) compound 5 (0.20 g, 0.39 mmol) was added, and stirring at 25 °C was continued for 4 days. Usual aqueous work-up followed by chromatography (silica gel, hexanes/ethyl acetate, 7:3, then 8:2, then CHCl₃/MeOH 9:1 followed by 10:0) gave 6 [41] (0.12 g, 63%) as an off-white solid; $R_F = 0.27$ (silica gel, hexanes/ ethyl acetate, 7:3); m.p.: 188–190 °C; $[\alpha]_D = -5.30^\circ$ (c = 0.67, CHCl₃/MeOH 5:1); IR (KBr): $\nu = 3252br$, 2946s, 2868m, 1628m, 1466m, 1450m, 1388m, 1376m, 1256w, 1156w, 1110w, 1042m, 1030m, 1010*m*, 886*m* cm⁻¹; ¹H NMR (400 MHz, CDCl₃/CD₃OD): $\delta = 4.60$ (*s*, 1H, CH_a (29)), 4.47 (s, 1H, CH_b (29)), 3.07–3.00 (m, 1H, CH (3)), 2.96 (ddd, J = 10.9, 10.9, 3.8 Hz, 1H, CH (19)), 2.35-2.25 (m, 1H, CH (13)),1.94-1.76 (m, 2H, CH_a (16) + CH_a (21)), 1.73-1.62 (m, 1H, CH_a (22)), 1.62-1.57 (m, 2H, CH_a (12) + CH_a (1)), 1.56 (s, 3H, CH₃ (30)), 1.51-1.42 (m, 3H, CH_2 (2) + CH (18)), 1.42-1.35 (m, 3H, CH_b $(16) + CH_a (15) + CH_a (6)$, 1.35–1.18 (m, 6H, CH₂ (7) + CH_b $(21) + CH_b (22) + CH_b (6) + CH_a (11)$, 1.19-1.10 (m, 2H, CH_b (11) + CH(9), 1.07 - 0.96 (m, 1H, CH_b (15)), 0.96 - 0.86 (m, 1H, CH_b (12)), 0.84 (s, 3H, CH₃ (27)), 0.82 (s, 6H, CH₃ (23) + CH₃ (25)), 0.80-0.72 (m, 1H, CH_b (1)), 0.70 (s, 3H, CH₃ (26)), 0.62 (s, 3H, CH₃ (24)), 0.57–0.52 (*m*, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃/CD₃OD): δ = 174.8 (*C*=0, C28), 150.7 (*C*_q, C20), 109.3 (*C*H₂=C, C29), 78.7 (CH, C9), 55.4 (CH, C5), 54.2 (C_q, C17), 50.5 (CH, C9), 50.4 (CH, C18), 46.7 (CH, C19), 42.3 (C_q , C14), 40.7 (C_q , C8), 38.7 (CH₂, C1), 38.2 $(CH_2, C22)$, 37.8 (CH, C13), 37.1 $(C_q, C4)$, 36.9 $(C_q, C10)$, 34.3 $(CH_2, C7)$, 32.6 (CH₂, C16), 30.7 (CH₂, C21), 29.3 (CH₂, C15), 27.8 (CH₃, C23), 26.9 (CH₂, C2), 25.5 (CH₂, C12), 20.8 (CH₂, C11), 19.2 (CH₃, C30), 18.2 (CH₂, C6), 16.0 (CH₃, C26), 15.9 (CH₃, C25), 15.2 (CH₃, C24), 14.5 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 472.4 (100\%, [M+H]^+), 494.3 (6\%, M)$ $[M+Na]^+$), 943.4 [20%, [2M + H]⁺), 965.5 (60%, [2M + Na]⁺); analysis calcd for C₃₀H₄₉NO₃ (471.71): C 76.39, H 10.47, N 2.97; found: C 76.21, H 10.90, N 2.76.

4.3.5. (3β) 3-Acetyloxy-N-methoxy-N-methyl-lup-20(29)-en-28-amide (7)

Following the procedure given for the synthesis of 5, from the reaction of 3 (0.30 g, 0.60 mmol) with oxalyl chloride (0.30 mL, 3.52 mmol) in dry DCM (25 mL) for 2 h, then with NEt₃ (0.70 mL, 5.05 mmol) and N,O-dimethylhydroxylammonium chloride (0.12 g, 1.24 mmol) in dry DCM (25 mL) for 1 day at 25 °C followed by chromatography (silica gel. hexanes/ethyl acetate, 9:1) 7 (0.25 g. 77%) [59] was obtained as a colorless solid; $R_F = 0.72$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 260–261 °C; $[\alpha]_D = +8.28^\circ$ $(c = 0.66, CHCl_3)$; IR (KBr): v = 3441br, 3077w, 2936s, 2868s, 1737s, 1651s, 1446m, 1370m, 1327w, 1249s, 1165w, 1105w, 1029m, 994m, 976*m*, 887*m* cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.71$ (*d*, J = 2.4 Hz, 1H, $CH_a(29)$), 4.56 (d, J = 2.3 Hz, 1H, $CH_b(29)$), 4.48–4.44 (m, 1H, CH (3)), 3.64 (s, 3H, OCH₃ (34)), 3.15 (s, 3H, NCH₃ (33)), 2.97 (ddd, J = 10.8, 10.8, 4.3 Hz, 1H, CH (19)), 2.71 (ddd, J = 12.0, 13.6 Hz, 1H, CH (13)), 2.34–2.29 (m, 1H, CH_a (16)), 2.11–2.05 (m, 1H, CH_a (22)), 2.03 (s, 3H, CH₃ (32)), 1.84-1.75 (m, 1H, CH_b (16)), 1.73-1.56 (m, 4H, CH_a (1) + CH_a (12) + CH₂ (2)), 1.67 (s, 3H, CH₃ (30)), 1.56–1.50 (m, 1H, CH (9)), 1.50–1.42 $(m, 1H, CH_a (6))$, 1.42-1.20 (m, 10H, CH_b (6) + CH₂ (11) + CH_a (15) + CH₂ (7) + CH_b $(22) + CH_2(21) + CH(18)$, 1.17–1.12 (m, $CH_a(15)$), 1.01–0.92 (m, 2H, $CH_b(12) + CH_b(1)$, 0.95 (s, 3H, $CH_3(27)$), 0.93 (s, 3H, $CH_3(26)$), 0.84 (s, 3H, CH₃ (25)), 0.83 (s, 3H, CH₃ (23)), 0.82 (s, 3H, CH₃ (24)), 0.80-0.76 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 177.1 (C=0, C28), 171.1 (C=0, C31), 151.5 (C_q , C20), 109.2 (CH₂= C, C29), 81.1 (CH, C3), 60.4 (OCH₃, C34), 56.2 (C_q, C17), 55.7 (CH, C5), 52.2 (CH, C9), 50.9 (CH, C18), 46.5 (CH, C19), 42.4 (C_q, C14), 40.9 (C_q, C8), 38.6 (CH₂, C1), 38.0 (C_q, C4), 37.3 (C_q, C10), 37.2 (CH, C13), 35.2 (CH₂, C22), 34.5 (CH₂, C7), 34.1 (NCH₃, C33), 31.2 (CH₂, C21), 31.0 (CH₂, C16), 30.1 (CH₂, C15), 28.1 (CH₃, C23), 25.8 (CH₂, C12), 23.9 (CH₂, C2), 21.4 (CH₃, C32), 21.3 (CH₂, C11), 19.7 (CH₃, C30), 18.4 (CH₂, C6), 16.6 (CH₃, C24), 16.4 (CH₃, C26), 16.3 (CH₃, C25), 14.9 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 542.5 (100\%, [M+H]^+), 564.3 (38\%, 100\%)$ $[M+Na]^+$), 580.1 (24%, $[M+Li+MeOH]^+$), 1089.6 (40%, $[2M+Li]^+$), 1105.5 [2M + Na]⁺); analysis calcd for $C_{34}H_{55}NO_4$ (541.80): C 75.37, H 10.37, N 2.59; found: C 75.34, H 10.51, N 2.38.

4.3.6. (3β) 3-Hydroxy-N-methoxy-N-methyl-lup-20(29)-en-28-amide (**8**)

Deacetylation of 7 (0.10 g, 0.18 mmol) with potassium hydroxide (0.10 g, 1.79 mmol) in MeOH/CHCl₃ (13 mL/3.5 mL) for 5 days at 25 °C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 8 (0.80 g, 89%) as a colorless solid; $R_F = 0.59$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 219–220 °C; $[\alpha]_D = +4.66^{\circ} \ (c = 0.64, \text{CHCl}_3); \ \text{IR (KBr): } \nu = 3456s, 3078w, 2944s,$ 2866s, 1655s, 1621s, 1447s, 1415m, 1390m, 1357m, 1259w, 1190m, 1166m, 1109m, 1081w, 1044m, 1032m, 995s, 885m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.71$ (d, J = 2.3 Hz, 1H, CH_a (29)), 4.56 (dd, $I = 2.3, 1.4 \text{ Hz}, 1H, CH_b (29), 3.65 (s, 3H, OCH_3 (32)), 3.17 (dd,$ I = 11.8, 5.4 Hz, 1H, CH(3), 3.15 (s, 3H, NCH₃(31)), 2.98 (ddd, <math>I = 11.1, I11.1, 4.0 Hz, 1H, CH (19)), 2.71 (ddd, J = 13.0, 11.8, 3.6 Hz, 1H, CH (13)), 2.34-2.30 (m, 1H, CH_a (16)), 2.12-2.06 (m, 1H, CH_a (22)), 1.85-1.74 $(m, 1H, CH_b(16)), 1.73-1.64 (m, 2H, CH_a(12) + CH_a(1)), 1.67 (s, 3H, CH_a(12) + CH_a(1)), 1.67 (s, 3H, CH_a(12) + CH_a(12))$ CH_3 (30)), 1.64–1.43 (m, 4H, CH_2 (2) + CH (18) + CH_a (6)), 1.43–1.19 $(m, 10H, CH_2(11) + CH_b(6) + CH_2(21) + CH_a(15) + CH_2(7) + CH_b$ (22) + CH(9), 1.18-1.12 (m, 1H, CH_b (15)), 1.00-0.94 (m, 1H, CH_b (12)), 0.96 (s, 3H, CH₃ (27)), 0.95 (s, 3H, CH₃ (23)), 0.94 (s, 3H, CH₃ (25)), 0.93-0.85 (ddd, J = 13.2, 13.2, 3.6 Hz, 1H, $CH_b(1)$), 0.82 (s, 3H, CH₃ (26)), 0.75 (s, 3H, CH₃ (24)), 0.69–0.66 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 177.1 (*C*=0, C28), 151.6 (C_q , C20), 109.2 (CH₂=C, C29), 79.2 (CH, C3), 60.4 (OCH₃, C32), 56.2 (C_q, C17), 55.6 (CH, C5), 52.3 (CH, C18), 51.0 (CH, C9), 46.5 (CH, C19), 42.4 (C₀, C14), $40.9 (C_q, C8), 39.0 (CH_2, C1), 38.9 (C_q, C4), 37.4 (C_q, C10), 37.2 (CH, C10), 37.2 (CH,$ C13), 35.2 (CH₂, C22), 34.6 (CH₂, C7), 34.1 (NCH₃, C31), 31.2 (CH₂, C21), 31.1 (CH₂, C16), 30.1 (CH₂, C15), 28.1 (CH₃, C23), 27.6 (CH₂, C2), 25.9 (CH₂, C12), 21.3 (CH₂, C11), 19.8 (CH₃, C30), 18.5 (CH₂, C6), 16.4 (CH₃, C26), 16.3 (CH₃, C25), 15.5 (CH₃, C24), 14.9 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 500.5 (100\%, [M+H]^+), 1005.7 (28\%, [2M+Li]^+), 1021.4 (55\%, [2M+Na]^+); analysis calcd for C₃₂H₅₃NO₃ (499.77): C 76.90, H 10.69, N 2.80; found: C 76.77, H 10.84, N 2.69.$

4.3.7. (3β) 3-Acetyloxy-N-hydroxy-N-methyl-lup-20(29)-en-28-amide (**9**)

Following the procedure for the synthesis of 5, reaction of 3 (0.32 g, 0.64 mmol) with oxalyl chloride (0.30 mL, 3.52 mmol), then with NEt₃ (0.70 mL, 5.05 mmol) and N-methylhydroxylammonium chloride (0.15 g, 1.81 mmol) in dry DCM (25 mL) for 12 h at 25 °C, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave **9** (0.22 g, 0.42 mmol, 66%) as a colorless solid; $R_F = 0.57$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 226–228 °C; $[\alpha]_D = +3.52^\circ$ (c 0.84, CHCl₃); IR (KBr): $\nu = 3302br$, 3073w, 2947s, 2875m, 1736m, 1701s, 1630s, 1451m, 1390m, 1375s, 1334w, 1259s, 1271s, 1193w, 1109w, 1073w, 1029m, 980m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.56$ (br, 1H, OH), 4.74 (d, J = 1.9 Hz, 1H, CH_a (29)), 4.60 (dd, $J = 1.9, 1.5 \text{ Hz}, 1\text{H}, CH_b(29), 4.49-4.44 (m, 1\text{H}, CH(3)), 3.41 (s, 3\text{H}, 1.9)$ NCH_3 (33)), 2.98 (ddd, J = 11.1, 11.1, 3.9 Hz, 1H, CH (19)), 2.81 (ddd, J = 13.0, 13.0, 3.5 Hz, 1H, CH (13)), 2.11 (ddd, J = 13.2, 3.1, 3.1 Hz, 1H, 1H, 1H) CH_a (16)), 2.03 (s, 3H, CH_3 (32)), 1.96 (dd, J = 11.0, 7.8 Hz, 1H, CH_a (22)), 1.91–1.80 (m, 1H, CH_a (15)), 1.76–1.71 (m, 1H, CH_a (12)), 1.69 (s, 3H, CH_3 (30)), 1.67–1.57 (m, 4H, CH (18) + CH_2 (2) + CH_a (1)), 1.57-1.22 (m, 10H, CH_b (16) + CH_2 (6) + CH_2 (11) + CH_b (22) + CH_2 $(7) + CH_b(15) + CH_a(21)$, 1.16 (ddd, J = 13.5, 3.2, 3.2 Hz, 1H, CH_b (21)), 1.03-0.94 (m, 2H, CH_b (1) + CH_b (12)), 0.96 (s, 3H, CH_3 (27)), 0.94 (s, 3H, CH₃ (24)), 0.85 (s, 3H, CH₃ (26)), 0.83 (s, 3H, CH₃ (23)), 0.83 (s, 3H, CH₃ (25)), 0.81–0.75 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 171.5 (C=0, C28), 171.2 (C=0, C31), 150.9 (C_q, C20), 109.7 (CH_2 =C, C29), 81.1 (CH, C3), 55.7 (CH, C5), 53.3 (C_0 , C17), 52.4 (CH, C18), 50.9 (CH, C9), 46.1 (CH, C19), 42.2 (Cq, C14), 40.9 (Cq, C8), 38.6 (CH₂, C1), 38.4 (NCH₃, C33), 38.0 (C_q, C4), 37.3 (CH, C13), 37.3 (C₀, C10), 35.9 (CH₂, C22), 34.5 (CH₂, C7), 32.4 (CH₂, C16), 31.2 (CH₂, C15), 29.8 (CH₂, C21), 28.1 (CH₃, C23), 25.7 (CH₂, C12), 23.9 (CH₂, C2), 21.5 (CH₃, C32), 21.3 (CH₂, C11), 19.7 (CH₃, C30), 18.4 (CH₂, C6), 16.6 (CH₃, C26), 16.4 (CH₃, C25), 16.3 (CH₃, C24), 14.8 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 528.4 (70\%, [M+H]^+), 550.2 (20\%,$ $[M+Na]^+$), 1077.4 (100%, $[2M+Na]^+$); analysis calcd for $C_{33}H_{53}NO_4$ (527.78): C 75.10, H 10.12, N 2.65; found: C 75.00, H 10.21, N 2.47.

4.3.8. (3β) 3-Hydroxy-N-hydroxy-N-methyl-lup-20(29)-en-28-amide (**10**)

Deacetylation of **9** (0.22 g, 0.42 mmol) with potassium hydroxide (0.28 g, 5.00 mmol) in MeOH for 5 days at 25 °C, followed by chromatography gave **10** (0.12 g, 60%) as a colorless solid (only scarcely soluble in usual solvents); $R_F = 0.25$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 258–260 °C; IR (KBr): $\nu = 3385s$, 3198m, 3084m, 2956s, 2927s, 2868s, 1653m, 1607s, 1550w, 1452m, 1419m, 1384s, 1294w, 1256w, 1196m, 1107w, 1092w, 1023m, 982m, 884m cm⁻¹; MS (ESI, MeOH): m/z = 486.5 (100%, [M+H]⁺), 508.4 (10%, [M+Na]⁺), 993.6 (50%, [2M + Na]⁺); analysis calcd for C₃₁H₅₁NO₃ (485.74): C 76.65, H 10.58, N 2.88; found: C 76.55, H 10.51, N 2.67.

4.3.9. (3β) 3-Acetyloxy-N-methoxy-lup-20(29)-en-28-amide (11)

Following the procedure for the synthesis of **5**, reaction of **3** (0.20 g, 0.40 mmol) with oxalyl chloride (0.20 mL, 2.35 mmol) followed by reaction with NEt₃ (0.70 mL, 5.05 mmol) and *O*-methylhydroxylammonium chloride (0.15 g, 1.81 mmol) in dry DCM (25 mL) for 5 h at 25 °C followed by chromatography (silica gel, hexanes/ethyl acetate, 7:3) gave **11** (0.13 g, 63%) as a colorless solid; $R_F = 0.43$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.:

217–220 °C; $[\alpha]_D = +14.92^\circ$ (c = 0.54, CHCl₃); IR (KBr): $\nu = 3639m$, 3385m, 3226m, 3071w, 2952s, 2866m, 1740s, 1654s, 1638m, 1511w, 1466m, 1448m, 1392m, 1377m, 1350w, 1317w, 1250s, 1151w, 1108w, 1084w, 1032m, 980m, 890m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.46$ (s, 1H, NH), 4.73 (s, 1H, CH_a (29)), 4.60 (s, 1H, CH_b (29)), 4.50-4.42 (m, 1H, CH(3)), 3.75 (s, 3H, OCH₃(33)), 3.11 (ddd, J = 11.0, 11.0, 4.2 Hz, 1H, CH (19)), 2.42 (ddd, I = 12.6, 12.6, 3.2 Hz, 1H, CH (13)), 2.03 (s, 3H, CH₃ (32)), 2.02–1.93 (m, 1H, CH_a (16)), 1.93–1.80 $(m, 1H, CH_a(21)), 1.80-1.73$ $(m, 1H, CH_a(22)), 1.73-1.58$ $(m, 4H, CH_a(21)), 1.80-1.73$ $(12) + CH_a(1) + CH_2(2)$, 1.67 (s, 3H, CH₃(30)), 1.57–1.51 (m, 3H, CH $(9) + CH_b(21) + CH_a(15), 1.50 - 1.44(m, 1H, CH_a(6)), 1.44 - 1.29(m, 1.44 - 1.29(m, 1.44 - 1.29(m, 1.44 - 1.29(m, 1.44 - 1.49(m, 1.44 - 1$ 6H, $CH_b(6) + CH_a(11) + CH_b(16) + CH_2(7) + CH_b(22)$, 1.29-1.20 $(m, 2H, CH_b (11) + CH (18)), 1.20-1.11 (m, 1H, CH_b (15)), 1.01-0.90$ $(m, 2H, CH_b(12) + CH_b(1)), 0.96 (s, 3H, CH_3(27)), 0.95 (s, 3H, CH_3(27)),$ (26)), 0.84 (s, 3H, CH₃ (25)), 0.83 (s, 3H, CH₃ (23)), 0.82 (s, 3H, CH₃ (24)), 0.80–0.75 (*m*, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.7$ (C=0, C28), 171.2 (C=0, C31), 150.7 (C₀, C20), 109.7 (CH₂=C, C29), 81.1 (CH, C3), 64.3 (OCH₃, C33), 55.6 (CH, C5), 54.8 (C_q, C17), 50.7 (CH, C9), 50.7 (CH, C18), 46.9 (CH, C19), 42.5 (C_q, C14), $41.0 (C_q, C8), 38.6 (CH_2, C1), 38.2 (CH_2, C22), 37.9 (C_q, C4), 37.9 (CH, C1)$ C13), 37.3 (C₀, C10), 34.5 (CH₂ C7), 33.2 (CH₂, C21), 30.9 (CH₂, C16), 29.6 (CH₂, C15), 28.1 (CH₃, C23), 25.7 (CH₂, C12), 23.8 (CH₂, C2), 21.4 (CH₃, C32), 21.0 (CH₂, C11), 19.6 (CH₃, C30), 18.3 (CH₂, C6), 16.6 (CH₃, C24), 16.3 (CH₃, C26), 16.3 (CH₃, C25), 14.8 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 528.5 (100\%, [M+H]^+), 550.3 (24\%, [M+Na]^+), 1055.3$ $(24\%, [2M + H]^+)$, 1077.40 (80%, $[2M + Na]^+$); analysis calcd for C₃₃H₅₃NO₄ (527.78): C 75.10, H 10.12, N 2.65; found: C 74.88, H 10.34, N 2.60.

4.3.10. (3β) 3-Hydroxy-N-methoxy-lup-20(29)-en-28-amide (**12**)

Deacetylation of 11 (0.12 g, 0.22 mmol) with potassium hydroxide (0.18 g, 3.21 mmol) in MeOH/THF (18 mL/6 mL) for 5 days at 25 °C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 7:3) gave 12 (0.10 g, 95%) as a colorless solid; $R_F = 0.36$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 222–224 °C; $[\alpha]_D = +3.22^{\circ} (c = 0.62, CHCl_3); IR (KBr): \nu = 3421m, 3243m, 3082w,$ 2940s, 2869m, 1655s, 1464m, 1438m, 1390m, 1377w, 1256w, 1193w, 1108w, 1039m, 888m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.57$ (s, 1H, NH), $4.72 (d, J = 1.9 \text{ Hz}, 1\text{H}, CH_a(29)), 4.58 (dd, J = 2.0, 1.4 \text{ Hz}, 1\text{H},$ CH_b (29)), 3.74 (s, 3H, OCH₃ (31)), 3.16 (dd, J = 11.3, 4.9 Hz, 1H, CH (3)), 3.10 (ddd, J = 11.1, 11.1, 4.3 Hz, 1H, CH (19)), 2.41 (ddd, J = 12.8, 12.8, 3.6 Hz, 1H, CH (13)), 2.02–1.94 (m, 1H, CH_a (16)), 1.92–1.87 (m, 1H, $CH_a(21)$), 1.76 (dd, J = 11.8, 7.8 Hz, 1H, $CH_a(22)$), 1.72–1.67 (m, 1H, CH_a (12)), 1.66 (s, 3H, CH₃ (30)), 1.66–1.63 (m, 1H, CH_a (1)), $1.63 - 1.47 \; (\textit{m}, \, 6H, \, CH_2 \, (2) \, + \, CH_a \, (15) \, + \, CH_b \, (21) \, + \, CH \, (18) \, + \, CH_a \, (11) \, + \, CH$ (6)), 1.47–1.30 (m, 6H, CH_b (6) + CH_a (11) + CH_b (16) + CH_b $(22) + CH_2(7)$), 1.28-1.21 (m, 2H, $CH(9) + CH_b(11)$), 1.20-1.14 (m, 1H, CH_b (15)), 1.03-0.96 (m, 1H, CH_b (12)), 0.95 (s, 9H, CH₃ $(27) + CH_3(25) + CH_3(25)$, 0.88 (ddd, J = 12.9, 12.9, 4.1 Hz, 1H, CH_b (1)), 0.80 (s, 3H, CH₃ (26)), 0.74 (s, 3H, CH₃ (24)), 0.69–0.63 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.7$ (C=0, C28), 150.7 (C_q, C20), 109.6 (CH₂=C, C29), 79.1 (CH, C3), 64.3 (OCH₃, C31), 55.5 (CH, C5), 54.8 (C_q, C17), 50.7 (CH, C9), 50.7 (CH, C18), 46.8 (CH, C19), 42.5 (C_q , C14), 40.9 (C_q , C8), 39.0 (CH_2 , C1), 38.9 (C_q , C4), 38.2 (CH₂, C22), 37.9 (CH, C13), 37.3 (C_q, C10), 34.5 (CH₂, C7), 33.2 (CH₂, C21), 30.9 (CH₂, C16), 29.6 (CH₂, C15), 28.1 (CH₃, C23), 27.5 (CH₂, C2), 25.7 (CH₂, C12), 21.0 (CH₂, C11), 19.6 (CH₃, C30), 18.4 (CH₂, C6), 16.3 (CH₃, C26), 16.3 (CH₃, C25), 15.5 (CH₃, C25), 14.8 (CH₃, C24) ppm; MS (ESI, MeOH): $m/z = 486.5 (60\%, [M+H]^+), 508.4 (5\%, [M+Na]^+),$ 971.4 (20%, $[2M + H]^+$), 993.5 (100%, $[2M + Na]^+$); analysis calcd for $C_{31}H_{51}NO_3$ (485.74): C 76.65, H 10.58, N 2.88; found: C 76.44, H 10.67, N 2.80.

4.3.11. (3β) -3-Acetyloxy-N-allyloxy-lup-20(29)-en-28-amide (**13**) Following the procedure for the synthesis of **5**, reaction of **3**

(0.40 g, 0.80 mmol) with oxalyl chloride (0.40 mL, 4.60 mmol), followed by reaction with NEt3 (0.70 mL, 5.05 mmol) and O-allylhydroxylammonium chloride (0.30 g, 2.75 mmol) in dry DCM (25 mL) for 2 h at 25 °C, followed by chromatography (silica gel, hexanes/ethyl acetate, 9:1) gave **13** (0.40 g, 90%) as a colorless solid; $R_F = 0.65$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 128–131 °C; $[\alpha]_D = +22.16^{\circ} (c = 0.59, CHCl_3); IR (KBr): \nu = 3260br, 2947s, 1737s,$ 1644s, 1455s, 1376s, 1317m, 1245s, 1154m, 1108m, 1030s, 979s, 927s, 883s, 754m, 544m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.42$ (s, 1H, NH), 5.99 (ddt, I = 16.8, 10.3, 6.4 Hz, 1H, CH(34)), 5.36-5.25 (m, 2H, 2H)CH₂, C35)), 4.72 (s, 1H, CH_a (29)), 4.58 (s, 1H, CH_b (29)), 4.50–4.43 (m, 1H, CH(3)), 4.42-4.32 (m, 2H, CH₂(33)), 3.10 (ddd, J = 11.0, 11.0, 11.0)3.9 Hz, 1H, CH (19)), 2.42 (ddd, J = 12.7, 12.7, 3.3 Hz, 1H, CH (13)), 2.02 (s, 3H, CH₃ (32)), 2.02-1.92 (m, 1H, CH₃ (16)), 1.92-1.84 (m, 1H, $CH_a(21)$), 1.79–1.72 (m, 1H), 1.66 (s, 3H, $CH_3(30)$), 1.71–1.63 (m, 2H, $CH_a(12) + CH_a(1)$, 1.62–1.56 (m, 2H, $CH_2(2)$), 1.55–1.49 (m, 3H, $CH_2(2)$) $(18) + CH_b (21) + CH_a (15), 1.49 - 1.28 (m, 7H, CH_2 (6) + CH_a)$ $(11) + CH_b (16) + CH_2 (7) + CH_b (22)$, 1.27–1.20 (m, 2H, CH $(9) + CH_b(11)$, 1.19-1.10 (m, 1H, $CH_b(15)$), 1.05-0.93 (m, 2H, CH_b $(12) + CH_b(1)$, 0.95 (s, 3H, CH₃ (27)), 0.94 (s, 3H, CH₃ (26)), 0.83 (s, 3H, CH₃ (25)), 0.82 (s, 3H, CH₃ (23)), 0.81 (s, 3H, CH₃ (24)), 0.79–0.72 (*m*, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.6$ (C=0, C28), 171.2 (C=0, C31), 150.7 (C₀, C20), 132.7 (CH= CH₂, C34), 120.4 (CH₂=CH, C35), 109.7 (CH₂=C, C29), 81.1 (CH, C3), 77.4 (CH₂, C33), 55.6 (CH, C5), 54.8 (C_q, C17), 50.7 (CH, C18), 50.7 (CH, C9), 46.8 (CH, C19), 42.5 (C_q, C14), 40.9 (C_q, C8), 38.5 (CH₂, C1), 38.3 (CH₂, C22), 37.9 (C_q, C4), 37.8 (CH, C13), 37.3 (C_q, C10), 34.5 (CH₂, C7), 33.3 (CH₂, C21), 30.9 (CH₂, C16), 29.6 (CH₂, C15), 28.1 (CH₃, C23), 25.7 (CH₂, C12), 23.8 (CH₂, C2), 21.4 (CH₃, C32), 21.0 (CH₂, C11), 19.6 (CH₃, C30), 18.3 (CH₂, C6), 16.6 (CH₃, C24), 16.4 (CH₃, C25), 16.3 (CH₃, C26), 14.8 (CH₃, C27) ppm; MS (ESI, MeOH): m/ $z = 554.4 (96\%, [M+H]^+), 576.4 (20\%, [M+Na]^+), 1129.5 (100\%,$ $[2M + Na]^+$); analysis calcd for $C_{35}H_{55}NO_4$ (553.82): C 75.91, H 10.01, N 11.56; found: C 75.79, H 10.14, N 11.49.

4.3.12. (3β) 3-Hydroxy-N-allyloxy-lup-20(29)-en-28-amide (**14**)

Deacetylation of 13 (0.17 g, 0.31 mmol) with potassium hydroxide (0.17 g, 3.03 mmol) in MeOH (18 mL) and THF (7 mL) for 7 days at 25 °C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 14 (0.15 g, 94%) as a colorless solid; $R_F = 0.44$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 129–131 °C; $[\alpha]_D = +3.36^\circ$ (c = 0.78, CHCl₃); IR (KBr): $\nu = 3423br$, 3075w, 2944s, 2869s, 1655m, 1453m, 1389m, 1376m, 1279w, 1247w, 1189w, 1138w, 1107w, 1034m, 1008m, 983m, 924m, 883*m* cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.31$ (s, 1H, N*H*), 5.99 (ddt, J = 16.8, 10.3, 6.4 Hz, 1H, CH(32)), 5.37-5.27 (m, 2H, CH₂(33)),4.73 (s, 1H, CH_a (29)), 4.59 (s, 1H, CH_b (29)), 4.43-4.33 (m, 2H, CH₂ (31)), 3.17 (dd, J = 11.3, 4.8 Hz, 1H, CH (3)), 3.11 (ddd, J = 11.1, 11.1, 4.3 Hz, 1H, CH (19)), 2.42 (ddd, J = 12.7, 12.7, 3.5 Hz, 1H, CH (13)), 2.03-1.93 (m, 1H, CH_a (16)), 1.86 (m, 1H, CH_a (21)), 1.75 (dd, J=11.8, 7.9 Hz, 1H, CH_a (22)), 1.72–1.63 (m, 2H, CH_a (12) + CH_a (1)), 1.67 (s, 3H, CH_3 (30)), 1.63-1.46 (m, 6H, CH_2 (2) + CH_a (15) + CH_b (21) + CH $(18) + CH_a$ (6)), 1.45–1.30 (m, 6H, CH_b (6) + CH_a (11) + CH_b (16) + CH₂(7) + CH_b(22)), 1.29-1.20 (m, 2H, CH (9) + CH_b (11)),1.19-1.12 (m, 1H, CH_b (15)), 1.03-0.96 (m, 1H, CH_b (12)), 0.96 (s, 3H, CH_3 (23)), 0.95 (s, 6H, CH_3 (25) + CH_3 (27)), 0.92–0.84 (m, 1H, CH_b (1)), 0.81 (s, 3H, CH₃ (26)), 0.75 (s, 3H, CH₃ (24)), 0.70–0.64 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 174.6$ (C=0, C28), 150.7 (C₀, C20), 132.7 (CH=CH₂, C32), 120.5 (CH₂=CH, C33), 109.7 $(CH_2=C, C29)$, 79.1 (CH, C3), 77.4 $(CH_2, C31)$, 55.5 (CH, C5), 54.9 $(C_q, C31)$ C17), 50.8 (CH, C18), 50.7 (CH, C9), 46.8 (CH, C19), 42.5 (Cq, C14), 41.0 (C_q, C8), 39.0 (CH₂, C1), 38.9 (C_q, C4), 38.4 (CH₂, C22), 37.9 (CH, C13), 37.4 (C_q, C10), 34.6 (CH₂, C7), 33.4 (CH₂, C21), 30.9 (CH₂, C16), 29.6 (CH₂, C15), 28.1 (CH₃, C23), 27.6 (CH₂, C2), 25.8 (CH₂, C12), 21.0 (CH₂, C11), 19.6 (CH₃, C30), 18.4 (CH₂, C6), 16.4 (CH₃, C26), 16.3 (CH₃, C25), 15.5 (CH₃, C24), 14.8 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 512.5 (70%, $[M+H]^+$), 534.4 (12%, $[M+Na]^+$), 1023.3 (18%, $[2M + H]^+$), 1045.6 (100%, $[2M + Na]^+$); analysis calcd for C₃₃H₅₃NO₃ (511.78): C 77.45, H 10.44, N 2.74; found: C 77.39, H 10.51, N 2.62.

4.3.13. 3-Oxo-N-methoxy-N-methyl-lup-20(29)-en-28-amide (**15**)

Following the procedure for the synthesis of 5, reaction of 4 (0.25 g, 0.50 mmol) in DCM (20 mL) with oxalyl chloride (0.30 mL, 3.5 mmol), then with NEt₃ (0.70 mL, 5.06 mmol) and N,O-dimethylhydroxylammonium chloride (0.15 g, 1.55 mmol) for 12 h at 25 °C followed by chromatography (silica gel, hexanes/ethyl acetate, 9:1) gave **15** (0.13 g, 52%) as a colorless solid; $R_F = 0.70$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 174–176 °C; $[\alpha]_D = +33.12^\circ$ $(c = 0.60, CHCl_3)$; IR (KBr): $\nu = 3398br$, 3077w, 2942s, 2867s, 1709s, 1651s, 1456s, 1376m, 1356s, 1345m, 1283w, 1243w, 1168m, 1137w, 1110*m*, 1082*w*, 1022*w*, 993*s*, 881*m* cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.70$ (s, 1H, CH_a (29)), 4.56 (s, 1H, CH_b (29)), 3.65 (s, 3H, OCH₃ (32)), 3.15 (s, 3H, NCH₃ (31)), 2.97 (ddd, J = 11.0, 11.0, 3.5 Hz, 1H, CH (19)), 2.75 (ddd, J = 12.9, 12.9, 3.3 Hz, 1H, CH (13)), 2.47 (ddd, $J = 15.8, 8.2, 7.6 \text{ Hz}, 1H, CH_a(2), 2.42-2.27 (m, 2H, CH_b(2) + CH_a(2))$ (16)), 2.15-2.05 (m, 1H, CH_a (22)), 1.88 (ddd, J = 13.1, 8.1, 4.6 Hz, 2H, $CH_a(1)$), 1.79 (m, 1H, $CH_b(16)$), 1.75–1.69 (m, 1H, $CH_a(12)$), 1.67 (s, 3H, CH_3 (30)), 1.54 (m, 1H, CH (18)), 1.50–1.21 (m, 13H, CH_2 (6) + CH_2 $(11) + CH_{2}(21) + CH_{2}(7) + CH_{a}(15) + CH_{b}(1) + CH_{b}(22) + CH_{a}(15) + CH_{b}(15) + CH_{b}(15)$ (5) + CH(9), 1.20–1.13 (m, 1H, $CH_b(15)$), 1.05 (s, 3H, $CH_3(23)$), 1.00 (s, 3H, CH₃ (24)), 0.97 (s, 3H, CH₃ (25)), 0.96 (s, 3H, CH₃ (27)), 0.96–0.93 (m, 1H, CH_b (12)), 0.91 (s, 3H, CH₃ (26)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 218.3$ (C=0, C3), 177.0 (C=0, C28), 151.4 (C_0 , C20), 109.2 (CH₂=C, C29), 60.4 (OCH₃, C32), 56.1 (C_q, C17), 55.2 (CH, C5), 52.1 (CH, C18), 50.3 (CH, C9), 47.5 (Cq, C4), 46.4 (CH, C19), 42.4 $(C_q, C14), 40.8 (C_q, C8), 39.8 (CH_2, C1), 37.3 (CH, C13), 37.1 (C_q, C10),$ 35.1 (CH₂, C22), 34.3 (CH₂, C2), 34.1 (NCH₃, C31), 33.8 (CH₂, C7), 31.1 (CH₂, C16), 31.0 (CH₂, C21), 30.0 (CH₂, C15), 26.7 (CH₃, C23), 25.8 (CH₂, C12), 21.8 (CH₂, C11), 21.2 (CH₃, C24), 19.8 (CH₂, C6), 19.7 (CH₃, C30), 16.1 (CH₃, C26), 16.1 (CH₃, C25), 14.8 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 498.5 (100\%, [M+H]^+), 1017.6 (40\%, [2M + Na]^+);$ analysis calcd for C₃₂H₅₁NO₃ (497.75): C 77.22, H 10.33, N 2.81; found: C 76.98, H 10.54, N 2.77.

4.3.14. 3-Oxo-N-hydroxy-N-methyl-lup-20(29)-en-28-amide (16)

Following the procedure for the synthesis of 5, reaction of 4 (0.24 g, 0.53 mmol) with oxalyl chloride (0.30 mL, 3.52 mmol) followed by reaction with NEt₃ (0.70 mL, 5.06 mmol) and Nmethylhydroxylammonium chloride (0.10 g, 1.20 mmol) in dry DCM (25 mL) for 12 h at 25 °C, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 16 (0.21 g, 81%) as a colorless solid; $R_F = 0.25$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 214–216 °C; $[\alpha]_D = +21.54^\circ$ (c = 0.70, CHCl₃); IR (KBr): $\nu = 3405br$, 2942s, 2868s, 1704m, 1640m, 1595m, 1455m, 1376m, 1184w, 1104w, 1076w, 1022w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.53$ (*br*, 1H, OH), 4.73 (s, 1H, CH_a (29)), 4.59 (s, 1H, CH_b (29)), 3.40 (s, 1H, NCH₃ (31), 2.98 (ddd, J = 11.1, 11.1, 3.6 Hz, 1H, CH (19)), 2.85 <math>(ddd, J = 12.9, 11.12.9, 3.4 Hz, 1H, CH (13)), 2.53-2.44 (m, 1H, CH_a (2)), 2.38 (ddd, $J = 15.6, 7.5, 4.3 \text{ Hz}, 1\text{H}, CH_b(2), 2.14 (ddd, J = 13.4, 2.8, 2.8 \text{ Hz}, 1\text{H},$ $CH_a(16)$), 1.98 (dd, J = 10.6, 7.9 Hz, 1H, $CH_a(22)$), 1.94–1.80 (m, 2H, $CH_a(1) + CH_a(21)$, 1.79–1.69 (m, 1H, $CH_a(12)$), 1.68 (s, 3H, CH_3) (30)), 1.62 (dd, J = 11.3 Hz, 1H, CH (18)), 1.54 (ddd, J = 13.9, 13.9, 3.5 Hz, 1H, CH_b (16)), 1.48–1.22 (m, 12H, CH_2 (6) + CH_2 (11) + CH_2 $(7) + CH_b(1) + CH_b(22) + CH_b(21) + CH_a(15) + CH(9) + CH(5),$ $1.17 \ (ddd, J = 13.5, 3.2, 3.2 \ Hz, 1H, CH_b (15)), 1.05 \ (s, 3H, CH_3 (23)),$ 1.01 (s, 3H, CH₃ (24)), 0.97–0.89 (m, 1H, CH_b (12)), 0.97 (s, 3H, CH₃ (27)), 0.97 (s, 3H, CH₃ (25)), 0.92 (s, 3H, CH₃ (26)), ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 218.4 (C=0, C3), 171.5 (C=0, C28), 150.9 (C_q , C20), 109.7 (CH₂=C, C29), 55.2 (CH, C5), 53.4 (C_q, C17), 52.3 (CH, C18), 50.3 (CH, C9), 47.5 (C_q , C4), 46.1 (CH, C19), 42.2 (C_q , C14), 40.8 (C_q , C8), 39.8 (CH₂, C1), 38.3 (NCH₃, C31), 37.4 (CH, C13), 37.1 (C_q , C10), 35.9 (CH₂, C22), 34.3 (CH₂, C2), 33.9 (CH₂, C7), 32.2 (CH₂, C16), 31.2 (CH₂, C21), 29.8 (CH₂, C15), 26.7 (CH₃, C23), 25.8 (CH₂, C12), 21.8 (CH₂, C11), 21.2 (CH₃, C24), 19.8 (CH₂, C6), 19.7 (CH₃, C30), 16.1 (CH₃, C26), 16.1 (CH₃, C25), 14.2 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 484.5 (100%, [M+H]⁺), 507.5 (5%, [M+Na]⁺), 537.8 (12%, [M + Na + MeOH]⁺), 748.5 (80%, [3M+2Na]²⁺), 989.5 (50%, [2M + Na]⁺); analysis calcd for $C_{31}H_{49}NO_3$ (483.73): C 76.97, H 10.21, N 2.90; found: C 76.76, H 10.34, N 2.74.

4.3.15. 3-Oxo-N-methoxy-lup-20(29)-en-28-amide (17)

Following the procedure for the synthesis of 5, reaction of 4 (0.25 g, 0.55 mmol) dry DCM (20 mL) with oxalyl chloride (0.4 mL, 4.70 mmol), followed by reaction with NEt₃ (0.75 mL, 5.42 mmol) and O-methylhydroxylammonium chloride (0.15 g, 1.81 mmol) in dry DCM (20 mL) for 12 h at 25 °C, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 17 (0.12 g, 45%) as a colorless solid; $R_F = 0.43$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 159–162 °C; $[\alpha]_D = +24.32^\circ$ (c = 0.62, CHCl₃); IR (KBr): $\nu = 3173br$, 2944s, 2866s, 1708s, 1637s, 1515m, 1464m, 1376m, 1318w, 1281w, 1243w, 1195w, 1140w, 1117w, 1081w, 1046m, 938m, 886m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.46$ (s, 1H, NH), 4.73 (s, 1H, CH_a (29)), 4.60 (s, 1H, CH_b (29)), 3.75 (s, 3H, OCH₃ (31)), 3.11 (ddd, J = 10.9, 10.9, 3.9 Hz, 1H, CH (19)), 2.54-2.43 (m, 2H, CH) $(13) + CH_a(2)$, 2.38 (ddd, J = 15.6, 7.4, 4.3 Hz, 1H, $CH_b(2)$), 2.06-1.94 (m, 1H, CH_a (16)), 1.94-1.83 (m, 2H, CH_a (21) + CH_a (1)), 1.81-1.68 (m, 2H, CH_a (22) + CH_a (21)), 1.67 (s, 3H, CH_3 (30)), 1.63-1.52 (m, 3H, $CH_a(15) + CH_b(21) + CH(18)$), 1.52-1.22 (m, 11H, $CH_2(6) + CH_2(11) + CH_b(16) + CH_2(7) + CH_b(22) + CH_b(1) + CH_b(11) + C$ (9) + CH(5), 1.21–1.15 (m, 1H, CH_b (15)), 1.05 (s, 3H, CH₃ (23)), 1.06-1.02 (m, 1H, CH_b (12)), 1.01 (s, 3H, CH₃ (24)), 1.00 (s, 3H, CH₃ (27)), 0.97 (s, 3H, CH₃ (25)), 0.92 (s, 3H, CH₃ (26)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 218.3$ (C=0, C3), 174.7 (C=0, C28), 150.6 (C_0 , C20), 109.7 (CH₂=C, C29), 64.3 (OCH₃, C31), 55.2 (CH, C5), 54.7 (C_q, C17), 50.7 (CH, C18), 50.1 (CH, C9), 47.5 (C₀, C4), 46.8 (CH, C19), 42.6 $(C_0, C14), 40.9 (C_0, C8), 39.8 (CH_2, C1), 38.2 (CH_2, C22), 38.0 (CH_2, C14), 38.2 (CH_2, C14), 38.2 (CH_2, C15), 38.0 (CH_2, C15), 38.2 (CH_2, C15), 3$ C13), 37.1 (C₀, C10), 34.3 (CH₂, C2), 33.8 (CH₂, C7), 33.2 (CH₂, C21), 30.9 (CH₂, C16), 29.6 (CH₂, C15), 26.7 (CH₃, C23), 25.8 (CH₂, C12), 21.6 (CH₂, C11), 21.2 (CH₃, C24), 19.8 (CH₂, C6), 19.6 (CH₃, C30), 16.1 (CH₃, C26), 16.1 (CH₃, C25), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 484.5 (100\%, [M+H]^+), 506.3 (4\%, [M+Na]^+), 967.3 (20\%, M)$ $[2M + H]^+$), 989.6 (82%, $[2M + Na]^+$); analysis calcd for $C_{31}H_{49}NO_3$ (483.37): C 76.97, H 10.21, N 2.90; found: C 76.88, H 10.50, N 2.81.

4.3.16. 3-Oxo-N-allyloxy-lup-20(29)-en-28-amide (18)

Following the procedure for the synthesis of 5, reaction of 4 (0.26 g, 0.57 mmol) with oxalyl chloride (0.3 mL, 3.52 mmol), followed by reaction with NEt₃ (0.75 mL, 5.42 mmol) and O-allylhydroxylammonium chloride (0.25 g, 2.29 mmol) in dry DCM (25 mL) for 5 h at 25 °C, followed by chromatography (silica gel, hexanes/ ethyl acetate, 8:2) gave 18 (0.20 g, 68%) as a colorless solid; $R_F = 0.33$ (silica gel, toluene/ethyl acetate/heptane/formic acid, 80:20:30:4); m.p.: 113–115 °C; $[\alpha]_D = +22.88^\circ$ (c = 0.89, CHCl₃); IR (KBr): $\nu = 3277br$, 3074w, 2948s, 2868s, 1705s, 1642s, 1458m, 1376m, 1241w, 1203w, 1140w, 1115w, 1039m, 924m, 883m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.45$ (s, 1H, NH), 5.99 (ddt, J = 16.8, 10.3, 6.4 Hz, 1H, CH (32)), 5.37–5.24 (m, 2H, CH₂ (33)), 4.71 (s, 1H, CH_a (29)), 4.58 (s, 1H, CH_b (29)), 4.43–4.31 (m, 2H, CH₂ (31)), 3.10 (ddd, $J = 11.0, 11.0, 3.9 \text{ Hz}, 1H, CH (19)), 2.53-2.42 (m, 2H, CH (13) + CH_a)$ (2)), 2.37 (ddd, J = 15.6, 7.4, 4.3 Hz, 1H, $CH_b(2)$), 2.01–1.92 (m, 1H, CH_a (16)), 1.92–1.83 (m, 2H, CH_a (21) + CH_a (1)), 1.80–1.67 (m, 2H, CH_a (22) + CH_a (12)), 1.66 (s, 3H, CH_3 (30)), 1.60–1.50 (m, 3H, CH_3 $(18) + CH_b (21) + CH_a (15), 1.49-1.21 (m, 11H, CH_2 (6) + CH_2)$ $(11) + CH_b(16) + CH_2(7) + CH_b(22) + CH_b(1) + CH(9) + CH(5),$

1.19–1.14 (m, 1H, CH_b (15)), 1.04 (s, 3H, CH_3 (23)), 1.00 (s, 3H, CH_3 (24)), 0.99 (s, 3H, CH_3 (27)), 0.95 (s, 3H, CH_3 (25)), 0.91 (s, 3H, CH_3 (26)) ppm; ¹³C NMR (100 MHz, CDCl₃): δ = 218.3 (C=0, C3), 174.6 (C=0, C28), 150.6 (C_q , C20), 132.7 (CH=CH₂, C32), 120.5 (CH₂=C, C33), 109.7 (CH₂=C, C29), 77.4 (CH₂, C31), 55.1 (CH, C5), 54.8 (C_q , C17), 50.7 (CH, C18), 50.1 (CH, C9), 47.5 (C_q , C4), 46.7 (CH, C19), 42.6 (C_q , C14), 40.9 (C_q , C8), 39.8 (CH₂, C1), 38.3 (CH₂, C22), 37.9 (CH, C13), 37.1 (C_q , C10), 34.3 (CH₂, C2), 33.8 (CH₂, C7), 33.3 (CH₂, C21), 30.9 (CH₂, C16), 29.6 (CH₂, C15), 26.7 (CH₃, C23), 25.7 (CH₂, C12), 21.6 (CH₂, C11), 21.1 (CH₃, C24), 19.8 (CH₂, C6), 19.6 (CH₃, C30), 16.2 (CH₃, C26), 16.1 (CH₃, C25), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 510.5 (100%, [M+H]⁺), 532.4 (14%, [M+Na]⁺), 784.9 (20%, [3M+K+H]²⁺), 1019.3 (24%, [2M+H]⁺), 1041.6 (84%, [2M+Na]⁺); analysis calcd for C_{33} H₅₁NO₃ (509.76): C 77.75, H 10.08, N 2.75; found: C 77.59, H 10.19, N 2.63.

4.3.17. (3 β) 3-Acetyloxylup-20(29)-en-28-amide (19)

Compound 3 (4.0 g, 8.0 mmol) was allowed to react with oxalyl chloride (3.7 g, 2.5 mL, 29.2 mmol) in dry DCM (50 mL) as described above. The volatiles were removed, and the residue was dissolved in dry DCM (50 mL). A solution of dry ammonia in DCM (satd., 50 mL) was added, and the mixture was stirred at 25 °C for 1 h. Water (50 mL) and diluted aqueous hydrochloric acid (1 N) were added until the aqueous layer showed pH = 7. The layers were separated, the aqueous layer was extracted with DCM (3 \times 50 mL), the combined organic phases were dried (Na₂SO₄), and the solvent was distilled off under reduced pressure. The crude product was purified by chromatography (silica gel, hexanes/ethyl acetate, 7:3) to yield **19** (3.8 g, 95%) as a colorless solid; $R_F = 0.12$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 336-338 °C (lit.: [60] 340-341 °C); $[\alpha]_D = +17.0^\circ$ (c = 0.48, CHCl₃); IR (KBr): $\nu = 3471m$, 3325m, 3202w, 3074w, 2940s, 2868m, 1742s, 1686s, 1642s, 1610m, 1449w, 1392m, 1367s, 1301w, 1242s, 1139w, 1108w, 1082w, 1025m, 1009m, 979m, 946w, 899w, 875m, 738w, 655w, 608w, 579w, 558w, 541w, 516w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 5.53$ (br, 2H, NH₂), 4.73 (s, 1H, CH_a (29)), 4.60 (s, 1H, CH_b (29)), 4.46 (dd, J = 9.7, 6.6 Hz, 1H, CH (3)), 3.07 (dt, J = 11.1, 4.4 Hz, 1H, CH)(19)), 2.44 (dt, J = 12.8, 3.5 Hz, 1H, CH (13)), 2.03 (s, 3H, CH₃ (32)), 2.02-1.89 (m, 2H, CH_a (16) + CH_a (21)), 1.81 (dd, J = 11.9, 7.8 Hz, 1H, CH_a (22)), 1.73–1.64 (m, 2H, CH_a (1) + CH_a (12)), 1.68 (s, 3H, CH_3 (30)), 1.63–1.52 (m, 5H, CH_2 (2) + CH (18) + CH_a (15) + CH_b (16)), 1.52-1.44 (m, 2H, CH_a (6) + CH_a (11)), 1.44-1.34 (m, 5H, CH_2 $(7) + CH_b(6) + CH_b(21) + CH_b(22)$, 1.30–1.21 (m, 2H, CH(9) + CH_b (11)), 1.21–1.14 (m, 1H, CH_b (15)), 1.03–0.92 (m, 2H, CH_b (1) + CH_b (12)), 0.96 (s, 3H, CH₃ (27)), 0.95 (s, 3H, CH₃ (25)), 0.84 (s, 3H, CH₃ (23)), 0.83 (s, 3H, CH₃ (26)), 0.82 (s, 3H, CH₃ (24)), 0.78 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 179.3$ (C=0, C28), 171.0 (C=0, C31), 150.7 (C=CH₂, C20), 109.5 (H₂C=C, C29), 80.9 (CH, C3),55.8 (*C*₀, C17), 55.5 (*C*H, C5), 50.5 (*C*H, C9), 49.9 (*C*H, C18), 46.6 (*C*H, C19), 42.5 (Cq, C14), 40.8 (Cq, C8), 38.4 (CH, C13), 38.4 (CH₂, C1), 37.8 (Cq, C4), 37.7 (CH₂, C22), 37.1 (Cq, C10), 34.3 (CH₂, C7), 33.9 (CH₂, C16), 30.7 (CH₂, C21), 29.5 (CH₂, C15), 27.9 (CH₃, C23), 25.6 (CH₂, C12), 23.7 (CH₂, C2), 21.3 (CH₂, C32), 20.9 (CH₂, C11), 19.4 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.2 (CH₃, C26), 16.2 (CH₃, C25), 14.6 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 498.5 (34%, [M + H]⁺), 520.4 (11%, [M + Na]⁺), 552.0 (5%, [M + Na + MeOH]⁺), 995.4 $(100\%, [2M + H]^+), 1017.4 (46\%, [2M + Na]^+).$

4.3.18. (3 β) 3-Hydroxylup-20(29)-en-28-amide (**20**)

Deacetylation of **19** (1.78 g, 3.58 mmol) with potassium hydroxide (0.4 g, 7.16 mmol) in THF (30 mL)/MeOH (20 mL) for 1 day at 25 °C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 7:3) gave **20** (1.57 g, 96.5%) as a colorless solid; $R_F = 0.1$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 256-257 °C (lit.: [47] 254-257 °C); [α]_D = +2.0° (c = 0.3, CHCl₃),

(lit.: [61] $[\alpha]_D = +6.0^\circ$, CHCl₃); IR (KBr): $\nu = 3422br$, 2943s, 2869m, 1656m, 1451m, 1376m, 1044w, 883m, 543w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 5.53$ (br, 1H, NH_a), 5.43 (br, 1H, NH_b), 4.73 (s, 1H, CH_a (29)), 4.59 (s, 1H, CH_b (29)), 3.17 (dd, J = 11.1, 5.1 Hz, 1H, CH(3)), 3.07 (dt, J = 11.0, 4.2 Hz, 1H, CH (19)), 2.45 (dt, J = 12.7, 3.3 Hz, 1H, CH (13)), 2.03-1.89 (m, 2H, CH_a (16) + CH_a (21)), 1.80 (dd, $J = 11.9, 7.8 \text{ Hz}, 1H, CH_a(22), 1.72 - 1.63 (m, 2H, CH_a(1) + CH_a(12)),$ 1.68 (s, 3H, CH_3 (30)), 1.63–1.52 (m, 5H, CH_2 (2) + CH (18) + CH_3 $(15) + CH_b$ (16)), 1.52–1.49 (m, 1H, CH_a (6)), 1.45–1.32 (m, 6H, CH₂) $(7) + CH_b(6) + CH_a(11) + CH_b(21) + CH_b(22)$, 1.30–1.21 (m, 2H, $CH(9) + CH_b(11)$, 1.20–1.13 (m, 1H, $CH_b(15)$), 1.05–0.94 (m, 1H, CH_b (12)), 0.96 (s, 3H, CH₃ (27)), 0.95 (s, 3H, CH₃ (23)), 0.95 (s, 3H, CH_3 (25)), 0.93–0.84 (m, 1H, CH_b (1)), 0.82 (s, 3H, CH_3 (26)), 0.75 (s, 3H, CH₃ (24)), 0.67 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 178.9$ (C=0, C28), 150.8 (C=CH₂, C20), 109.4 (H₂C=C, C29), 79.0 (CH, C3), 55.8 (C₀, C17), 55.4 (CH, C5), 50.6 (CH, C9), 49.9 (CH, C18), 46.6 (CH, C19), 42.5 (C_q, C14), 40.7 (C_q, C8), 38.8 (CH₂, C1), 38.7 (CH₂, C22), 38.4 (*C*_q, C4), 37.7 (*C*H, C13), 37.2 (*C*_q, C10), 34.4 (*C*H₂, C7), 34.0 (CH₂, C16), 30.7 (CH₂, C21), 29.5 (CH₂, C15), 28.0 (CH₃, C23), 27.4 (CH₂, C2), 25.6 (CH₂, C12), 20.9 (CH₂, C11), 19.5 (CH₃, C30), 18.3 (CH₂, C6), 16.2 (CH₃, C25), 16.1 (CH₃, C26), 15.3 (CH₃, C24), 14.6 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 456.3 (84\%, [M + H]^+), 911.4 (78\%, M)$ $[2M + H]^+$), 933.4 (100%, $[2M + Na]^+$).

4.3.19. (3 β) 3-Acetyloxylup-N-benzyl-20(29)-en-28-amide (**21**)

Following the procedure given for the synthesis of 19, the reaction of 3 (0.35 g, 0.7 mmol) with oxalyl chloride (0.22 g, 0.15 mL, 1.8 mmol) in dry DCM (15 mL), followed by the reaction with benzylamine (0.15 g, 0.15 mL) (1 h, reflux) and chromatographic work-up (silica gel, hexanes/ethyl acetate, 8:2) gave **21** (0.24 g, 59%) as a colorless solid; $R_F = 0.87$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 124–127 °C; $[\alpha]_D = +23.2^\circ$ (c = 0.35, CHCl₃); IR (KBr): $\nu = 3425 br, 2946 s, 1736 m, 1638 m, 1517 w, 1454 w, 1384 m, 1246 m,$ 1028w, 979w, 882m, 698w cm⁻¹; UV-vis (CHCl₃): λ_{max} $(\log \varepsilon) = 253.8 (3.49) \text{ nm}; {}^{1}\text{H NMR } (400 \text{ MHz}, \text{CDCl}_{3}); \delta = 7.35 - 7.29$ (m, 2H, CH(36/36')), 7.29-7.22(m, 3H, CH(35/35') + CH(37)), 5.89(br, 1H, NH)), 4.74 (s, 1H, CH_a (29)), 4.59 (s, 1H, CH_b (29)), 4.53–4.42 $(m, 2H, CH(3) + CH_a(33)), 4.36 (dd, J = 14.7, 5.6 Hz, 1H, CH_b(33)),$ 3.17 (dt, J = 11.0, 4.0 Hz, 1H, CH (19)), 2.49 (dt, J = 12.8, 3.5 Hz, 1H, CH)(13)), 2.03 (s, 3H, CH₃ (32)), 2.02–1.94 (m, 2H, CH₂ (21)), 1.94–1.87 $(m, 2H, CH_2(16)), 1.80-1.70 (m, 2H, CH_a(22) + CH_a(12)), 1.68 (s, 3H, CH_a(12)), 1.68 (s, 3H, CH_a(12)), 1.80-1.70 (m, 2H, CH_a($ CH₃ (30)), 1.67-1.62 (m, 1H, CH_a (1)), 1.62-1.51 (m, 3H, CH $(18) + CH_2(2)$, 1.50-1.41 (m, 3H, $CH_b(22) + CH_a(15) + CH_a(6)$), 1.40-1.28 (m, 7H, CH_2 (7) + CH_a (11) + CH_b (6)), 1.28-1.21 (m, 2H, $CH(9) + CH_b(11)$, 1.15–1.08 (m, 1H, $CH_b(15)$), 1.04–0.93 (m, 2H, $CH_b(1) + CH_b(12)$), 0.95 (s, 3H, $CH_3(27)$), 0.91 (s, 3H, $CH_3(25)$), 0.84 (s, 3H, CH₃ (26)), 0.83 (s, 3H, CH₃ (23)), 0.83 (s, 3H, CH₃ (24)), 0.77 (*m*, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 175.9$ (C=O, C28), 171.0 (C=O, C31), 150.8 (C=CH₂, C20), 139.2 (C_q, C34), 128.6 (CH, C36/36'), 127.8 (CH, C35/35'), 127.3 (CH, C37), 109.4 (H₂C=C, C29), 81.0 (CH, C3), 55.7 (Cq, C17), 55.5 (CH, C5), 50.6 (CH, C9), 50.2 (CH, C18), 46.7 (CH, C19), 43.3 (CH₂, C33), 42.5 (C_q, C14), 40.8 (C_q, C8), 38.4 (CH₂, C1), 38.4 (CH₂, C22), 37.8 (C_q, C4), 37.7 (CH, C13), 37.1 (C_q, C10), 34.4 (CH₂, C7), 33.8 (CH₂, C16), 30.9 (CH₂, C21), 29.4 (CH₂, C15), 27.9 (CH₃, C23), 25.6 (CH₂, C12), 23.7 (CH₂, C2), 21.3 (CH₃, C32), 21.0 (CH₂, C11), 19.5 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.2 (CH₃, C26), 16.1 (CH₃, C25), 14.6 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 588.5 (58\%, [M + H]^+), 610.5 (26\%, [M + Na]^+), 1175.3 (90\%,$ $[2M + H]^+$), 1197.4 (100%, $[2M + Na]^+$); analysis calcd for $C_{39}H_{57}NO_3$ (587.87): C 79.68, H 9.77, N 2.38; found: C 79.52, H 9.85, N 2.21.

4.3.20. (3 β) N-Benzyl-3-hydroxylup-20(29)-en-28-amide (**22**)

Deacetylation of **21** (210 mg, 0.36 mmol) with potassium hydroxide (0.1 g, 1.8 mmol) in THF (15 mL)/MeOH (5 mL) for 1 day at

25 °C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 7:3) gave 22 (180 mg, 91.8%) as a colorless solid; $R_F = 0.6$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 246–248 °C; $[\alpha]_D = +14.3^\circ$ (c = 0.31, CHCl₃); IR (KBr): $\nu = 3496$ s, 3448s, 2941s, 2868m, 1634s, 1496s, 1458m, 1376w, 1319w, 1280w, 1223w, 1190w, 1105w, 1077w, 1045w, 892w, 761w, 702w, 596w, 496w cm⁻¹; UV-vis (CHCl₃): λ_{max} (log ε) = 258.8 (3.43) nm; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.35-7.29$ (m, 2H, CH (34/34')), 7.29-7.23 (m, 3H, CH (33/33') + CH (35)), 5.88 (br, 1H, NH)), 4.74 (s, 1H, $CH_a(29)$), 4.59 (s, 1H, $CH_b(29)$), 4.48 (dd, J = 14.7, 5.8 Hz, 1H, CH_a (31)), 4.36 (dd, I = 14.7, 5.6 Hz, 1H, CH_b (31)), 3.17 (m, 2H, CH (19) + CH(3), 2.49 (dt, I = 12.8, 3.5 Hz, 1H, CH(13)), 2.03–1.94 (m, 2H, CH₂ (21)), 1.94–1.87 (m, 2H, CH₂ (16)), 1.80–1.69 (m, 2H, CH_a $(22) + CH_a(12)$, 1.68 (s, 3H, $CH_3(30)$), 1.68–1.63 (m, 1H, $CH_a(1)$), 1.62-1.54 (m, 3H, CH (18) + CH₂ (2)), 1.53-1.44 (m, 2H, CH_a $(15) + CH_a(6)$, 1.43–1.36 (m, 2H, $CH_b(22) + CH_a(11)$), 1.36–1.28 (m, 3H, CH₂(7) + CH_b(6)), 1.29-1.20 (m, 2H, CH(9) + CH_b(11)),1.16-1.08 (m, 1H, CH_b (15)), 1.05-0.97 (m, 1H, CH_b (12)), 0.96 (s, 3H, CH₃ (27)), 0.96 (s, 3H, CH₃ (23)), 0.91 (s, 3H, CH₃ (25)), 0.91–0.87 (m, 1H, CH_b (1)), 0.82 (s, 3H, CH₃ (26)), 0.75 (s, 3H, CH₃ (24)), 0.66 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 175.9$ (C=0, C28), 151.0 (C=CH₂, C20), 139.2 (C_q, C32), 128.6 (CH, C34/34'), 127.8 (CH, C33/33'), 127.3 (CH, C35), 109.4 (H₂C=C, C29), 79.0 (CH, C3), 55.7 (C_a, C17), 55.4 (CH, C5), 50.7 (CH, C9), 50.2 (CH, C18), 46.7 (CH, C19), 43.3 (CH₂, C31), 42.5 (C_q, C14), 40.8 (C_q, C8), 38.9 (CH₂, C1), 38.7 (CH₂, C22), 38.4 (C_q, C4), 37.7 (CH, C13), 37.2 (C_q, C10), 34.5 (CH₂, C7), 33.8 (CH₂, C16), 30.9 (CH₂, C21), 29.5 (CH₂, C15), 28.0 (CH₃, C23), 27.4 (CH₂, C2), 25.7 (CH₂, C12), 21.0 (CH₂, C11), 19.5 (CH₃, C30), 18.3 (CH₂, C6), 16.2 (CH₃, C26), 16.2 (CH₃, C25), 15.4 (CH₃, C24), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 546.5 (100%, $[M + H]^+$), 568.4 (9%, $[M + Na]^+$), 1091.4 (28%, $[2M + H]^+$), 1113.4 (52%, $[2M + Na]^{+}$); analysis calcd for $C_{37}H_{55}NO_{2}$ (545.84): C 81.42, H 10.16, N 2.57; found: C 81.39, H 10.29, N 2.43.

4.3.21. N-Benzyl-3-oxolup-20(29)-en-28-amide (23)

Jones oxidation of 22 (137 mg, 0.25 mmol) in acetone (25 mL) using freshly prepared reagent [from CrO₃ (63 mg, 0.63 mmol), sulfuric acid (62 mg, 34 μ L, 0.63 mmol, 96%) and water (0.5 mL)] for 20 min, followed by usual aqueous work-up and chromatography (silica gel, hexanes/ethyl acetate, 7:3) gave 23 (100 mg, 73.5%) as a colorless solid; $R_F = 0.68$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 108-111 °C; $[\alpha]_D = +38.3^\circ$ (c = 0.37, CHCl₃), (lit.: [61] $[\alpha]_D = +43.0^\circ$, CHCl₃); IR (KBr): $\nu = 3396br$, 2947s, 2867m, 1705s, 1640s, 1519m, 1454m, 1384m, 1241w, 1185w, 1080w, 986w, 882w, 724w, 698w, 583w cm⁻¹; UV-vis (CHCl₃): λ_{max} (log ε) = 259.0 (3.42) nm; ¹H NMR $(500 \text{ MHz}, \text{CDCl}_3)$: $\delta = 7.35 - 7.30$ (m, 2H, CH (34))34')), 7.30-7.24 (m, 3H, CH (33/33') + CH (35)), 5.89 (br, 1H, NH)), 4.74 (s, 1H, CH_a (29)), 4.60 (s, 1H, CH_b (29)), 4.49 (dd, J = 14.7, 5.8 Hz, 1H, CH_a (31)), 4.36 (dd, J = 14.7, 5.6 Hz, 1H, CH_b (31)), 3.18 (dt, J = 11.1, 4.3 Hz, 1H, CH (19), 2.54 (dt, J = 12.8, 3.5 Hz, 1H, CH (13)),2.51-2.44 (m, 1H, CH_a (2)), 2.48 (ddd, I = 14.7, 7.5, 2.2 Hz, 1H, CH_a (2)), 2.38 (ddd, J = 15.6, 7.5, 4.3 Hz, 1H, $CH_b(2)$), 2.03–1.95 (m, 1H, CH_a (21)), 1.94–1.85 (m, 2H, CH_a (1) + CH_a (16)), 1.80–1.70 (m, 2H, CH_a (22) + CH_a (12)), 1.68 (s, 3H, CH_3 (30)), 1.64–1.52 (m, 2H, CH_3 $(18) + CH_b (16)$, $1.51-1.39 (m, 5H, CH_b (22) + CH_a (15) + CH_a$ (11) + CH₂(6), 1.39 - 1.33 (m, 5H, CH(9) + CH_b(1) + CH₂(7) + CH_b(21)), 1.32-1.23 (m, 1H, CH_b (15) + CH_b (11)), 1.18-1.11 (m, 1H, CH(5)), 1.06 (s, 3H, CH₃ (23)), 1.02 (s, 3H, CH₃ (24)), 1.02–0.98 (m, 1H, CH_b (12)), 0.97 (s, 3H, CH₃ (27)), 0.94 (s, 3H, CH₃ (25)), 0.92 (s, 3H, CH₃ (26)) ppm; 13 C NMR (125 MHz, CDCl₃): $\delta = 218.1$ (C=0, C3), 175.8 (C=O, C28), 150.8 (C=CH₂, C20), 139.2 (C_q, C32), 128.6 (CH, C34/34'), 127.8 (CH, C33/33'), 127.3 (CH, C35), 109.4 (H₂C=C, C29), 55.6 (C_q, C17), 55.0 (CH, C5), 50.1 (CH, C9), 50.1 (CH, C18), 47.3 (C_q, C4), 46.6 (CH, C19), 43.3 (CH₂, C31), 42.6 (C_q , C14), 40.7 (C_q , C8), 39.7(CH₂, C1), 38.4 (CH₂, C22), 37.8 (CH, C13), 36.9 (C_q, C10), 34.1 (CH₂, C2), 33.8 (CH₂, C16), 33.7 (CH₂, C7), 30.9 (CH₂, C21), 29.4 (CH₂, C15), 26.6 (CH₃, C23), 25.7 (CH₂, C12), 21.5 (CH₂, C11), 21.0 (CH₃, C24), 19.7 (CH₂, C6), 19.5 (CH₃, C30), 16.0 (CH₃, C26), 16.0 (CH₃, C25), 14.6 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 544.5 (66%, [M + H]⁺), 566.2 (9%, [M + Na]⁺), 1087.3 (70%, [2M + H]⁺), 1109.4 (100%, [2M + Na]⁺); analysis calcd for C₃₇H₅₃NO₂ (543.82): C 81.72, H 9.82, N 2.58; found: C 81.54, H 9.70, N 2.64.

4.3.22. 3-Oxolup-20(29)-en-28-amide (**24**)

Reaction of 4 (3.0 g, 6.61 mmol) with oxalyl chloride (3.0 g, 2 mL, 23.3 mmol) in abs. DCM (50 mL) followed by a reaction with ammonia in DCM as described above and chromatographic workup (silica gel, hexanes/ethyl acetate, 7:3) gave 24 (2.52, 84.1%) as a colorless solid; $R_F = 0.21$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 155–1157 °C (lit.: [50] 156–157 °C); $[\alpha]_D = +32.9^\circ$ (c = 0.37, CHCl₃); IR (KBr): $\nu = 34\overline{46b}r$, 2947s, 2868m, 1703s, 1670s, 1458m, 1375*m*, 1204*w*, 1116*w*, 882*w*, 755*w*, 543*w* cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 5.55$ (br, 1H, NH_a), 5.32 (br, 1H, NH_b), 4.74 (s, 1H, CH_a (29)), 4.59 (s, 1H, CH_b (29)), 3.08 (dt, J = 11.1, 4.4 Hz, 1H, CH (19)), 2.56-2.43 (m, 2H, CH (13) + CH_a (2)), 2.42-2.35 (m, 1H, CH_b (2)), 2.03-1.94 (m, 1H, CH_a (21)), 1.93-1.85 (m, 2H, CH_a (16) + CH_a (1)), 1.84-1.77 (m, 1H, CH_a (22)), 1.76-1.69 (m, 1H, CH_a (12)), 1.68 (s, 3H, $CH_3(30)$), 1.62–1.53 (m, 3H, $CH(9) + CH_b(16) + CH_a(15)$), 1.53–1.47 $(m, 2H, CH_a (6) + CH_b (22)), 1.46-1.34 (m, 7H, CH (18) + CH_b)$ (1) + CH₂(7) + CH_b(21) + CH_a(11) + CH_b(6), 1.34–1.28 (m, 2H, CH $(5) + CH_b(11)$, 1.23–1.17 (m, 1H, $CH_b(15)$), 1.06 (s, 3H, $CH_3(23)$), 1.03-0.99 (m, 1H, CH_b (12)), 1.01 (s, 3H, CH₃ (24)), 1.00 (s, 3H, CH₃ (25)), 0.98 (s, 3H, CH₃ (27)), 0.92 (s, 3H, CH₃ (26)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 218.1$ (C=0, C3), 178.9 (C=0, C28), 150.7 (C= CH₂, C20), 109.4 (H₂C=C, C29), 55.8 (C_q, C17), 55.0 (CH, C5), 50.0 (CH, C18), 49.8 (CH, C9), 47.3 (C_q, C4), 46.5 (CH, C19), 42.6 (C_q, C14), 40.7 (C₀, C8), 39.6 (CH₂, C1), 38.3 (CH₂, C22), 37.7 (CH, C13), 36.9 (C₀, C10), 34.1 (CH₂, C2), 33.9 (CH₂, C16), 33.7 (CH₂, C7), 30.7 (CH₂, C21), 29.5 (CH₂, C15), 26.6 (CH₃, C23), 25.6 (CH₂, C12), 21.4 (CH₂, C11), 21.0 (CH₃, C24), 19.6 (CH₂, C6), 19.5 (CH₃, C30), 16.0 (CH₃, C26), 15.9 $(CH_3, C25)$, 14.5 $(CH_3, C27)$ ppm; MS (ESI, MeOH): m/z = 454.5 (59%, $[M + H]^+$), 907.3 (100%, $[2M + H]^+$), 929.5 (71%, $[2M + Na]^+$).

4.3.23. (3β) Lup-20(29)-en-3,28-bis-N-ethylcarbamate (**25**)

Microwave assisted reaction (7 h, 120 °C) of **2** (0.3 g, 0.68 mmol) in dry THF (5 mL) with ethyl isocyanate (0.29 g, 0.32 mL, 4.1 mmol), followed by chromatographic work-up (silica gel, hexanes/ethyl acetate, 8:2) gave **25** (0.32 g, 80.8%) as a colorless solid; $R_F = 0.29$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 233-235 °C; $[\alpha]_D = +24.9^{\circ} (c = 0.44, CHCl_3); IR (KBr): \nu = 3330br, 2945s, 2870m,$ 2361w, 1687s, 1536s, 1449m, 1376w, 1306m, 1264s, 1142w, 1107w, 1083w, 1020m, 986w, 894w, 781w, 636w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.65–4.51 (br, 2H, 2× NH), 4.58 (s, 1H, CH_b (29)), 4.34 (dd, J = 11.2, 4.0 Hz, 1H, CH (3)), 4.25 (d, $J = 10.8 \text{ Hz}, 1\text{H}, CH_a(28)), 3.83 (d, J = 10.3 \text{ Hz}, 1\text{H}, CH_b(28)), 3.21 (br,$ 4H, CH_2 (32) + CH_2 (32')), 2.4 (dt, I = 11.0, 5.8 Hz, 1H, CH (19)), 2.02-1.91 (m, 1H, CH_a (21)), 1.86-1.79 (m, 1H, CH_a (16)), 1.78-1.72 (m, 1H, CH_a (22)) 1.72–1.68 (m, 1H, CH_a (15)), 1.68 (s, 3H, CH₃ (30)), 1.68-1.64 (m, 1H, CH (13)), 1.63-1.53 (m, 4H, CH_a (2) + CH_a (1) + CH₂(12), 1.52–1.45 (m, 2H, CH_b(2) + CH(18)), 1.45–1.41 (m, 1H, $CH_a(6)$), 1.40–1.33 (m, 6H, $CH_b(6) + CH_2(11) + CH_b(21) + CH_2(11)$ (7)), 1.32–1.26 (m, 1H, CH (9)), 1.25–1.17 (m, 1H, CH_b (16)), 1.14 (t, J = 7.3 Hz, 3H, CH₃ (33)), 1.13 (t, J = 7.2 Hz, 3H, CH₃ (33')), 1.09–0.95 $(m, 3H, CH_b(15) + CH_b(22) + CH_b(1)), 1.03 (s, 3H, CH_3(25)), 0.96 (s, 3H, CH_b(15)), 0.96 (s, 3H_b(15)), 0.96 (s, 3H_b(15)),$ 3H, CH₃ (27)), 0.88 (s, 3H, CH₃ (23)), 0.83 (s, 3H, CH₃ (26)), 0.81–0.76 (m, 1H, CH (5)), 0.80 (s, 3H, CH₃ (24)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 157.2$ (C=0, C31), 156.9 (C=0, C31'), 150.4 (C=CH₂, C20), 109.7 (H₂C=C, C29), 80.9 (CH, C3), 62.8 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C₀, C17), 42.7 $(C_q, C14), 40.9 (C_q, C8), 38.4 (CH_2, C1), 38.0 (C_q, C4), 37.5 (CH, C13),$

37.0 (C_q , C10), 35.9 (CH₂, C32/C32'), 34.5 (CH₂, C22), 34.1 (CH₂, C7), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 24.1 (CH₂, C2), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.1 (CH₂, C6), 16.1 (CH₃, C24), 16.0 (CH₃, C26), 15.7 (CH₃, C25), 15.3 (CH₃, C33/C33'), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 607.4 (37%, [M + Na]⁺), 1169.5 (100%, [2M + H]⁺), 1191.5 (71%, [2M + Na]⁺); analysis calcd for $C_{36}H_{60}N_2O_4$ (584.87): C 73.93, H 10.34, N 4.79; found: C 73.77, H 10.51, N 4.69.

4.3.24. (3β) Lup-20(29)-en-3,28-bis-N-propylcarbamate (**26**)

Microwave assisted reaction (7 h, 120 °C) of 2 (0.3 g, 0.68 mmol) in dry THF (5 mL) with propyl isocyanate (0.35 g, 0.39 mL, 4.1 mmol) followed by chromatographic work-up (silica gel, hexanes/ethyl acetate, 8:2) gave **26** (0.362 g, 83.4%) as a colorless solid; $R_F = 0.4$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 123–126 °C $[\alpha]_D = +25.5^{\circ} (c = 0.32, CHCl_3); IR (KBr): \nu = 3358s, 2963s, 2873s,$ 1717s, 1520s, 1458s, 1389m, 1265s, 1138m, 1106m, 1042m, 1008m, 976m, 882w, 774w, 638w cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 4.64 - 4.45$ (br, 2H, 2× NH), 4.61 (s, 1H, CH_a (29)), 4.57 (s, 1H, CH_b (29)), 4.27 (dd, J = 7.7, 2.9 Hz, 1H, CH(3)), 4.19 (d, J = 10.6 Hz, 1H, CH_a (28)), 3.76 (d, J = 10.1 Hz, 1H, CH_b (28)), 3.07 (br, 4H, CH_2 (32) + CH_2 (32')), 2.38 (dt, J = 11.1, 5.7 Hz, 1H, CH (19)), 1.92 <math>(dq, J = 13.8, 1.0)10.5 Hz, 1H, CH_a (21)), 1.80–1.72 (*m*, 1H, CH_a (16)), 1.72–1.63 (*m*, 2H, CH_a (22) + CH_a (15)), 1.61 (s, 3H, CH_3 (30)), 1.62–1.54 (m, 4H, CH_a $(2) + CH(13) + CH_{a}(1) + CH_{a}(12)), 1.54 - 1.47 (m, 2H, CH_{b}(2) + CH_{b}(2)), 1.54 - 1.47 (m, 2H, CH_{b}(2) + CH_{$ (18)), 1.47-1.40 (m, 5H, CH_2 (33) + CH_2 (33') + CH_a (6)), 1.37-1.28 $(m, 5H, CH_2(7) + CH_b(21) + CH_a(11) + CH_b(6)), 1.25 - 1.20 (m, 1H, 1H)$ CH(9)), 1.20–1.11 (m, 2H, $CH_b(16) + CH_b(11)$), 1.03–0.92 (m, 3H, $CH_b(22) + CH_b(15) + CH_b(12)$, 0.97 (s, 3H, $CH_3(25)$), 0.92–0.88 $(m, 1H, CH_b(1)), 0.90 (s, 3H, CH_3(27)), 0.86 (t, I = 7.7 Hz, 3H, CH_3)$ (34)), 0.85 (t, J = 7.7 Hz, 3H, CH_3 (34')), 0.81 (s, 3H, CH_3 (23)), 0.77 (s, 3H, CH₃ (26)), 0.74–0.70 (*m*, 1H, CH (5)), 0.73 (*s*, 3H, CH₃ (24)) ppm; ¹³C NMR (125 MHz, CDCl₃): $\delta = 157.2$ (C=0, C31), 156.9 (C=0, C31'), 150.3 (C=CH₂, C20), 109.8 (H₂C=C, C29), 81.0 (CH, C3), 62.9 (CH₂, C28), 55.4 (CH, C5), 50.3(CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C_q , C17), 42.8 (CH_2 , C32/C32'), 42.7 (C_q , C14), 40.9 (C_q , C8), 38.4 (CH₂, C1), 38.0 (C_q, C4), 37.5 (CH, C13), 37.0 (C_q, C10), 34.6 (CH₂, C22), 34.1 (CH₂, C7), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 24.1 (CH₂, C2), 23.3 (CH₂, C33/C33'), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 14.7 (CH₃, C27), 11.2 (CH₃, C34/ C34') ppm; MS (ESI, MeOH): m/z = 612.8 (6%, $[M + H]^+$), 635.5 (26%, $[M + Na]^+$), 1125.5 (100%, $[2M + H]^+$), 1247.6 (32%, $[2M + Na]^+$); analysis calcd for C₃₈H₆₄N₂O₄ (612.93): C 74.46, H 10.52, N 4.57; found: C 74.39, H 10.69, N 4.50.

4.3.25. (3 β) Lup-20(29)-en-3,28-bis-N-butylcarbamate (**27**)

Microwave assisted reaction (7 h, 120 °C) of 2 (0.3 g, 0.68 mmol) in dry THF (5 mL) with butyl isocyanate (0.4 g, 0.45 mL, 4.1 mmol) followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave **27** (0.35 g, 80.7%) as a colorless solid; $R_F = 0.41$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 106-109 °C; $[\alpha]_D = +31.8$ ° $(c = 0.32, CHCl_3)$; IR (KBr): v = 3364br, 2957s, 2872m, 2361w, 1709s, 1528*m*, 1465*m*, 1376*w*, 1247*m*, 1138*w*, 1018*w*, 883*w*, 775*w* cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.66–4.50 (br, 2H, $2 \times NH$), 4.58 (s, 1H, CH_b (29)), 4.34 (dd, J = 11.0, 4.0 Hz, 1H, CH (3)), $4.26 (d, J = 10.9 \text{ Hz}, 1\text{H}, CH_a(28)), 3.83 (d, J = 10.2 \text{ Hz}, 1\text{H}, CH_b(28)),$ 3.17 (br, 4H, CH_2 (32) + CH_2 (32')), 2.45 (dt, J = 10.9, 5.6 Hz, 1H, CH(19)), 2.04–1.93 (m, 1H, CH_a (21)), 1.86–1.79 (m, 1H, CH_a (16)), 1.78-1.70 (m, 2H, CH_a (22) + CH_a (15)), 1.69-1.61 (m, 4H, CH_a $(1) + CH(13) + CH_a(12) + CH_a(2)$, 1.68 (s, 3H, CH₃(30)), 1.61–1.54 $(m, 2H, CH (18) + CH_b (2)), 1.52-1.44 (m, 5H, CH_2 (33) + CH_2)$ $(33') + CH_a$ (6)), 1.44–1.38 (m, 6H, CH₂ (7) + CH₂ (11) + CH_b $(21) + CH_b(6)$, 1.38-1.32 (m, 4H, $CH_2(34) + CH_2(34')$), 1.29-1.24(m, 1H, CH (9)), 1.23–1.17 (m, 1H, CH_b (16)), 1.10–0.99 (m, 3H, CH_b $(15) + CH_b(12) + CH_b(22)$, 1.04 (s, 3H, CH₃(25)), 0.99–0.94 (m, 1H, $CH_b(1)$, 0.97 (s, 3H, $CH_3(27)$), 0.92 (t, J = 7.3 Hz, 3H, $CH_3(35)$), 0.92 $(t, J = 7.3 \text{ Hz}, 3H, CH_3 (35')), 0.88 (s, 3H, CH_3 (23)), 0.84 (s$ (26)), 0.81–0.76 (*m*, 1H, CH(5)), 0.80 (*s*, 3H, CH₃ (24)) ppm; ¹³C NMR (125 MHz, CDCl₃): $\delta = 157.2$ (C=0, C31), 156.8 (C=0, C31'), 150.3 (C=CH₂, C20), 109.7 (H₂C=C, C29), 81.0 (CH, C3), 62.9 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C₀, C17), 42.7 (C_q, C14), 40.9 (C_q, C8), 40.7 (CH₂, C32/C32'), 38.4 (CH₂, C1), 38.0 (Cq, C4), 37.5 (CH, C13), 37.0 (Cq, C10), 34.6 (CH₂, C22), 34.1 (CH₂, C7), 32.1 (CH₂, C33/C33'), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 24.1 (CH₂, C2), 20.8 (CH₂, C11), 19.9 (C34/C34'), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 14.7 (CH₃, C27), 13.7 (CH₃, C35/35') ppm; MS (ESI, MeOH): $m/z = 641.1 (7\%, [M + H]^+), 663.3 (30\%, M)$ $[M + Na]^+$), 1281.4 (100%, $[2M + H]^+$), 1303.5 (24%, $[2M + Na]^+$); analysis calcd for C₄₀H₆₈N₂O₄ (640.98): C 74.95, H 10.69, N 4.37; found: C 74.87, H 10.82, N 4.19.

4.3.26. (3 β) Lup-20(29)-en-3,28-bis-N-hexylcarbamate (**28**)

Microwave assisted reaction (7 h, 120 °C) of **2** (0.3 g, 0.68 mmol) in dry THF (5 mL) with hexyl isocyanate (0.52 g, 0.58 mL, 4.1 mmol) followed by chromatography (silica gel, hexanes/ethyl acetate, 9:1) gave 28 (0.35 g, 73.4%) as a colorless solid; $R_F = 0.49$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 85–88 °C; $[\alpha]_D = +20.5^{\circ}$ $(c = 0.53, CHCl_3)$; IR (KBr): $\nu = 3347br$, 2954s, 2870m, 2361w, 1700s, 1540m, 1458m, 1376w, 1247s, 1140w, 1040w, 1012w, 982w, 883w, 775w, 668w cm⁻¹; ¹H NMR (500 MHz, CDCl₃): $\delta = 4.67$ (s, 1H, CH_a (29)), 4.64–4.59 (br, 2H, $2 \times NH$), 4.57 (s, 1H, CH_b (29)), 4.34 (dd, I = 10.9, 3.0 Hz, 1H, CH(3), 4.25 (d, I = 10.7 Hz, 1H, CH_a(28)), 3.82 $(d, I = 10.4 \text{ Hz}, 1\text{H}, CH_b(28)), 3.15 (br, 4\text{H}, CH_2(32) + CH_2(32')), 2.44$ (dt, J = 11.0, 5.9 Hz, 1H, CH (19)), 2.04-1.92 (m, 1H, CH_a (21)),1.85-1.78 (m, 1H, CH_a (16)), 1.77-1.69 (m, 2H, CH_a (15) + CH_a (22)), 1.69-1.60 (m, 4H, CH (13) + CH_a (1) + CH_a (2) + CH_a (12)), 1.67 (s, 3H, CH_3 (30)), 1.60–1.53 (m, 2H, CH (18) + CH_b (2)), 1.52–1.43 (m, $5H, CH_2(33) + CH_2(33') + CH_a(6), 1.43 - 1.34(m, 5H, CH_2(7) + CH_b)$ $(6) + CH_a (11) + CH_b (21), 1.34-1.25 (m, 13H, CH_2 (34) + CH_2)$ $(34') + CH_2(35) + CH_2(35') + CH_2(36) + CH_2(36') + CH(9)$ 1.23-1.13 (m, 2H, CH_b (11) + CH_b (16)), 1.10-0.99 (m, 3H, CH_b $(12) + CH_b(15) + CH_b(22)$, 1.03 (s, 3H, CH₃(25)), 0.98–0.92 (m, 1H, CH_b (1)), 0.96 (s, 3H, CH_3 (27)), 0.91–0.85 (m, 9H, CH_3 (23) + CH_3 (37) + CH₃ (37')), 0.83 (s, 3H, CH₃ (26)), 0.79 (s, 3H, CH₃ (24)), 0.78-0.75 (m, 1H, CH (5)) ppm; ¹³C NMR (125 MHz, CDCl₃): $\delta = 157.1$ (C=0, C31), 156.8 (C=0, C31'), 150.3 (C=CH₂, C20), 109.7 (H₂C=C, C29), 81.0 (CH, C3), 62.9 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C_a, C17), 42.7 (C_a, C14), 41.0 $(CH_2, C32/C32')$, $40.9(C_q, C8)$, $38.4(CH_2, C1)$, $38.0(C_q, C4)$, 37.5(CH, C1)C13), 37.0 (C_q, C10), 34.6 (CH₂, C22), 34.1 (CH₂, C7), 31.5 (CH₂, C35/ 35'), 30.0 (CH₂, C33/C33'), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 26.4 (CH₂, C34/34'), 25.2 (CH₂, C12), 24.1 (CH₂, C2), 22.5 (CH₂, C36/36'), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 14.7 (CH₃, C27), 14.0 (CH₃, C37/37') ppm; MS (ESI, MeOH): m/z = 697.0 (12%, $[M + H]^+$), 719.5 (34%, $[M + Na]^+$), 1394.6 (100%, $[2M + H]^+$), 1416.4 $(20\%, [2M + Na]^+)$; analysis calcd for $C_{44}H_{76}N_2O_4$ (697.09): C 75.81, H 10.99, N 4.02; found: C 75.69, H 11.14, N 3.94.

4.3.27. 3,28-Di-O-acetyl-betulin (29)

This compound was prepared from **1** (4.5 g, 10.16 mmol) by acetylation (12 h, 25 °C) with acetic anhydride (7.5 mL, 79.2 mmol), NEt₃ (15 mL, 108.2 mmol) and DMAP (0.12 g, 0.95 mmol) as previously described, followed by one re-crystallization from EtOH (30 mL) to yield **29** (4.78 g, 89.5%) as a colorless solid; $R_F = 0.73$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 219–222 °C (lit.: [62] 216–218 °C); $[\alpha]_D = +20.7^\circ$ (c = 0.36, CHCl₃), (lit.: [62] $[\alpha]_D = +19.7^\circ$ (CHCl₃)); MS (ESI, MeOH): m/z = 467.3 (91%,

 $[M + H-HOAc]^+$), 544.2 (16%, $[M + NH4]^+$), 549.4 (12%, $[M+Na]^+$), 1075.6 (10%, $[2M + Na]^+$).

4.3.28. 3-O-Acetyl-betulin (30)

Deacetylation of **29** (8.2 g, 15.6 mmol) with KOH (0.87 g, 15.6 mmol) in THF (50 mL) and MeOH (20 mL) for 30 min at 0 °C followed by usual aqueous workup and chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave **30** (4.33 g, 57.4%) as a colorless solid; $R_F = 0.40$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 255-258 °C (lit.: [62] 258-260 °C); [α]_D = +34.1° (c = 0.37, CHCl₃), (lit.: [62] [α]_D = +25.7° (CHCl₃)); MS (ESI, MeOH): m/z = 485.3 (8%, [M+H]⁺), 538.9 (25%, [M + Na + MeOH]⁺), 969.5 (18%, [2M + H]⁺), 991.4 (100%, [2M + Na]⁺).

4.3.29. (3 β) 3-Acetyloxylup-20(29)-en-28-N-ethylcarbamate (31)

Microwave assisted reaction (7 h, 120 °C) of **30** (1.0 g, 2.1 mmol) in dry THF (5 mL) with ethyl isocyanate (0.44 g, 0.49 mL, 6.2 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 9:1) gave 31 (1.05 g, 91.2%) as a colorless solid; $R_F = 0.44$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 116–119 °C; $[\alpha]_D = +21.0^{\circ} (c = 0.32, \text{CHCl}_3); \text{IR (KBr): } \nu = 3424br, 2946s, 2872m,$ 1731s, 1641w, 1521m, 1456m, 1369m, 1245s, 1146w, 1106w, 1081w, 1026m, 980w, 883w, 775w, 548w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.61 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), 4.46 (dd, J = 9.9, 6.2 Hz, 1H, CH (3)), 4.25 (d, J = 10.7 Hz, 1H, CH_a (28)), 3.82 (d, J = 10.7 Hz, 1H, CH_b (28)), 3.22 (br, 2H, CH_2 (34)), 2.44 (dt, J = 11.0, 5.7 Hz, 1H, CH(19)), 2.03 (s, 3H, CH₃(32)), 2.02-1.92 (m, CH₃(32)), 2.02 (m, C1H, CH_a (21)), 1.86–1.76 (m, 1H, CH_a (16)), 1.75–1.69 (m, 2H, CH_a $(22) + CH_a$ (15), 1.68 (s, 3H, CH₃ (30)), 1.67–1.61 (m, 4H, CH_a (1) + CH(13) + CH₂(12), 1.60-1.52 (m, 3H, CH(18) + CH₂(2)), 1.52-1.47 (m, 1H, CH_a (6)), 1.44-1.33 (m, 6H, CH₂ (7) + CH_b (21) + CH₂ (11) + CH_b (6), 1.32-1.24 (m, 1H, CH (9)), 1.20 (dd, 1.32-1.24) $J = 12.6, 3.6 \text{ Hz}, 1H, CH_b (16), 1.14 (t, J = 7.2 \text{ Hz}, 3H, CH_3 (35)),$ 1.10-0.99 (m, 2H, CH_b (22) + CH_b (15)), 1.03 (s, 3H, CH₃ (25)), 0.98–0.92 (m, 1H, CH_b (1)), 0.96 (s, 3H, CH₃ (27)), 0.84 (s, 3H, CH₃ (23)), 0.84 (s, 3H, CH₃ (26)), 0.83 (s, 3H, CH₃ (24)), 0.78 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.0$ (*C*=0, C31), 157.0 (C=O, C33), 150.3 (C=CH₂, C20), 109.8 (H₂C=C, C29), 80.9 (CH, C3), 63.0 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C_q, C17), 42.7 (C_q, C14), 40.9 (C_q, C8), 38.4 (CH₂, C1), 37.8 (Cq, C4), 37.5 (CH, C13), 37.1 (Cq, C10), 35.1 (CH₂, C34), 34.5 (CH₂, C22), 34.1 (CH₂, C7), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 23.7 (CH₂, C2), 21.3 (CH₃, C32), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 15.3 (CH₃, C35), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 555.9 (18\%, [M + H]^+), 578.3 (15\%, [M + Na]^+),$ 1111.3 (100%, [2M + H]⁺), 1133.3 (53%, [2M + Na]⁺); analysis calcd for C₃₅H₅₇NO₄ (555.83): C 75.63, H 10.34, N 2.52; found: C 75.41, H 10.47, N 2.39.

4.3.30. (3 β) 3-Acetyloxylup-20(29)-en-28-N-propylcarbamate (**32**)

Microwave assisted reaction (7 h, 120 °C) of **30** (0.78 g, 1.6 mmol) in dry THF (5 mL) with propyl isocyanate (0.41 g, 0.45 mL, 4.8 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 9:1) gave **32** (0.68 g, 73.6%) as a colorless solid; $R_F = 0.52$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 108-111 °C; $[\alpha]_D = +29.9$ ° (c = 0.34, CHCl₃); IR (KBr): v = 3418br, 2949s, 2872m, 2362w, 1732m, 1642w, 1530w, 1461w, 1372w, 1246m, 1136w, 1106w, 1031w, 980w, 882w, 776w, 668w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.65 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), 4.46 (dd, J = 9.9, 6.1 Hz, 1H, CH (3)), 4.26 (d, J = 10.6 Hz, 1H, CH_a (28)), 3.14 (br, 2H, CH_2 (34)), 2.44 (dt, J = 11.1, 5.8 Hz, 1H, CH (19)), 2.03 (s, 3H, CH_3 (32)), 2.02–1.92 (m, 1H, CH_a (21)), 1.85–1.76 (m, 1H, CH_a (16)), 1.76–1.69 (m, 2H, CH_a (22) + CH_a (15)), 1.68 (s, 3H, CH_3 (30)),

1.67-1.58 (m, 4H, $CH_a(1) + CH(13) + CH_2(12)$), 1.58-1.48 (m, 3H, $CH_2(2) + CH(18)$, 1.44–1.34 (m, 3H, $CH_a(6) + CH_2(35)$), 1.32–1.20 (m, 6H, CH₂(7) + CH_b(21) + CH₂(11) + CH_b(6)), 1.25-1.16 (m, 2H, 2H, 2H) CH_{b} (16) + CH (9)), 1.10–0.99 (m, 2H, CH_{b} (22) + CH_{b} (15)), 1.04 (s, 3H, CH_3 (25)), 0.99–0.95 (m, 1H, CH_b (1)), 0.96 (s, 3H, CH_3 (27)), 0.92 $(t, I = 4.0 \text{ Hz}, 3H, CH_3 (36)), 0.84 (s, 3H, CH_3 (23)), 0.84 (s, 3H, CH_3 (36)), 0.84 (s,$ (26)), 0.83 (s, 3H, CH₃ (24)), 0.79–0.76 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.0$ (C=0, C31), 157.2 (C=0, C33). 150.3 (C=CH₂, C20), 109.8 (H₂C=C, C29), 80.9 (CH, C3), 62.9 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 $(C_q, C17)$, 42.8 $(CH_2, C34)$, 42.7 $(C_q, C14)$, 40.9 $(C_q, C8)$, 38.4 $(CH_2, C1)$, 37.8 (C_q, C4), 37.5 (CH, C13), 37.1 (C_q, C10), 34.6 (CH₂, C22), 34.1 (CH₂, C7), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 23.7 (CH₂, C2), 23.2 (CH₂, C35), 21.3 (CH₃, C32), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 14.7 (CH₃, C27), 11.2 (CH₃, C36) ppm; MS (ESI, MeOH): m/z = 570.1 (8%, $[M + H]^+$), 592.4 (9%, $[M + Na]^+$), 643.3 $(13\%, [M + H + DMF]^+), 665.2 (14\%, [M + Na + DMF]^+), 1139.5$ $(100\%, [2M + H]^+)$, 1161.6 (66%, $[2M + Na]^+$); analysis calcd for C₃₆H₅₉NO₄ (569.86): C 75.88, H 10.44, N 2.46; found: C 75.69, H 10.53, N 2.31.

4.3.31. (3β) 3-Acetyloxylup-20(29)-en-28-N-butylcarbamate (33)

Microwave assisted reaction (7 h, 120 °C) of **30** (1 g, 2.1 mmol) in dry THF (5 mL) with butyl isocyanate (0.61 g, 0.7 mL, 6.2 mol) as described above, followed by chromatography (silica gel, hexanes/ ethyl acetate, 9:1) gave 33 (1.0 g, 83.3%) as a colorless solid; $R_F = 0.57$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 94–97 °C; $[\alpha]_D = +18.6^{\circ} (c = 0.29, CHCl_3); IR (KBr): \nu = 3421br, 2951s, 2872m,$ 2366w, 1734m, 1636w, 1534w, 1458w, 1374w, 1245m, 1134w, 1106w, 1025w, 980w, 882w, 776w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.62 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), $4.46 (dd, J = 10.3, 5.8 \text{ Hz}, 1H, CH (3)), 4.26 (d, J = 10.7 \text{ Hz}, 1H, CH_a)$ (28)), 3.82 (d, J = 10.5 Hz, 1H, CH_b (28)), 3.17 (br, 2H, CH_2 (34)), 2.44 $(dt, J = 11.1, 5.8 \text{ Hz}, 1\text{H. CH} (19)), 2.03 (s, 3\text{H, CH}_3 (32)), 2.02 - 1.92 (m, 3.03)$ 1H, CH_a (21)), 1.87–1.76 (m, 1H, CH_a (16)), 1.75–1.69 (m, 2H, CH_a $(22) + CH_a$ (15)), 1.68 (s, 3H, CH₃ (30)), 1.67–1.61 (m, 4H, CH_a (1) + CH(13) + CH₂(12), 1.61-1.53 (m, 3H, CH₂(2) + CH(18)),1.52-1.44 (m, 3H, CH_a (6) + CH_2 (35)), 1.43-1.29 (m, 6H, CH_2 $(7) + CH_b(21) + CH_2(11) + CH_b(6)$, 1.29–1.24 (m, 2H, CH₂(36)), 1.23-1.15 (m, 2H, CH_b (16) + CH (9)), 1.10-0.99 (m, 2H, CH_b $(22) + CH_b(15)$, 1.04 (s, 3H, CH₃ (25)), 0.98–0.95 (m, 1H, CH_b (1)), 0.96 (s, 3H, CH_3 (27)), 0.92 (t, J = 4.0 Hz, 3H, CH_3 (37)), 0.84 (s, 3H, $CH_3(23)$), $0.84(s, 3H, CH_3(26))$, $0.83(s, 3H, CH_3(24))$, $0.79-0.76(m, CH_3(23))$ 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.0$ (C=0, C31), 157.1 (C=O, C33), 150.3 (C=CH₂, C20), 109.8 (H₂C=C, C29), 80.9 (CH, C3), 62.9 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C_q , C17), 42.7 (C_q , C14), 40.9 (C_q , C8), 40.8 (CH₂, C34), 38.4 (CH₂, C1), 37.8 (C_q, C4), 37.5 (CH, C13), 37.1 (C_q, C10), 34.6 (CH₂, C22), 34.1 (CH₂, C7), 32.1 (CH₂, C35), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 23.7 (CH₂, C2), 21.3 (CH₃, C32), 20.8 (CH₂, C11), 19.9 (CH₂, C36), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 14.7 (CH₃, C27), 13.7 (CH₃, C37) ppm; MS (ESI, MeOH): m/z = 584.1 $(25\%, [M + H]^+)$, 606.5 $(10\%, [M + Na]^+)$, 1167.5 $(100\%, [2M + H]^+)$, 1189.5 (22%, $[2M + Na]^+$); analysis calcd for $C_{37}H_{61}NO_4$ (583.88): C 76.11, H 10.53, N 2.40; found: C 76.00, H 10.63, N 2.29.

4.3.32. (3 β) 3-Acetyloxylup-20(29)-en-28-N-hexylcarbamate (**34**)

Microwave assisted reaction (7 h, 120 °C) of **30** (1 g, 2.1 mmol) in dry THF (5 mL) with hexyl isocyanate (0.79 g, 0.9 mL, 6.2 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 9:1) gave **34** (1.09 g, 86.5%) as a colorless solid; $R_F = 0.82$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 80-82 °C; $[\alpha]_D = +19.6$ ° (c = 0.43, CHCl₃); IR (KBr): $\nu = 3420br$, 2950s, 2870m,

2362w, 1734m, 1646w, 1534w, 1457w, 1347w, 1245m, 1134w, 1106w, 1030w, 980w, 882w, 776w, 668w, 610w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.62 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), 4.46 (dd, J = 9.4, 6.5 Hz, 1H, CH (3)), 4.26 (d, J = 10.8 Hz, 1H, CH_a (28)), 3.82 (d, J = 10.3 Hz, 1H, CH_b (28)), 3.16 (br, 2H, CH_2 (34)), 2.44 (dt, J = 11.2, 5.9 Hz, 1H, CH (19)), 2.03 (s, 3H, CH₃ (32)),2.02–1.92 (m, 1H, CH_a (21)), 1.85–1.76 (m, 1H, CH_a (16)), 1.76–1.70 $CH_a(1) + CH(13)$, 1.61–1.53 (m, 5H, $CH(18) + CH_2(12) + CH_2(2)$), 1.53-1.44 (m, 3H, CH₂ (35) + CH_a (6)), 1.44-1.35 (m, 6H, CH₂ $(7) + CH_b(21) + CH_2(11) + CH_b(6), 1.33 - 1.24(m, 7H, CH(9) + CH_2(11) +$ (36) + CH₂(37) + CH₂(38), 1.24-1.15 (m, 1H, CH_b(16)), 1.08-0.99 $(m, 2H, CH_b (22) + CH_b (15)), 1.04 (s, 3H, CH_3 (25)), 0.98-0.93 (m,$ 1H, $CH_b(1)$, 0.96 (s, 3H, $CH_3(27)$), 0.88 (t, J = 6.4 Hz, 3H, $CH_3(39)$), 0.84 (s, 3H, CH₃ (23)), 0.84 (s, 3H, CH₃ (26)), 0.83 (s, 3H, CH₃ (24)), 0.79–0.76 (*m*, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.0$ (C=0, C31), 157.2 (C=0, C33), 150.3 (C=CH₂, C20), 109.8 (H₂C=C, C29), 80.9 (CH, C3), 62.9 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C_q, C17), 42.7 (C_q, C14), 41.0 (CH₂, C34), 40.9 (C_q, C8), 38.4 (CH₂, C1), 37.8 (C_q, C4), 37.5 (CH, C13), 37.1 (C₀, C10), 34.6 (CH₂, C22), 34.1 (CH₂, C7), 31.5 (CH₂, C37), 30.0 (CH₂, C35), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 26.4 (CH₂, C36), 25.2 (CH₂, C12), 23.7 (CH₂, C2), 22.5 (CH₂, C38), 21.3 (CH₃, C32), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 14.7 (CH₃, C27), 14.0 (CH₃, C39) ppm; MS (ESI, MeOH): m/z = 612.2 (60%, $[M + H]^+$), 634.5 (19%, $[M + Na]^+$), 1223.6 (100%, $[2M + H]^+$), 1245.6 $(26\%, [2M + Na]^+)$; analysis calcd for $C_{39}H_{65}NO_4$ (611.94): C 76.55, H 10.71, N 2.29; found: C 76.37, H 10.84, N 2.19.

4.3.33. (3β) 3-Acetyloxylup-20(29)-en-28-N-phenylcarbamate (**35**)

Microwave assisted reaction (7 h, 120 °C) of 30 (1 g, 2.1 mmol) in dry THF (5 mL) with phenyl isocyanate (0.74 g, 0.67 mL, 6.2 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 9:1) gave **35** (1.08 g, 87.1%) as a colorless solid; $R_F = 0.84$ (silica gel, hexanes/ethyl acetate, 7:3); m.p.: 221–223 °C; $[\alpha]_D = +13.2^{\circ} (c = 0.44, CHCl_3); IR (KBr): \nu = 3431m, 3080w, 2948s,$ 2873m, 1737s, 1647w, 1602m, 1523s, 1501w, 1442s, 1391w, 1374m, 1311m, 1246m, 1204s, 1178w, 1105w, 1084w, 1054m, 1030m, 978m, 898w, 872w, 848w, 757m, 693w, 545w, 510w cm⁻¹; UV-vis (CHCl₃): $\lambda_{\text{max}} (\log_{e}) = 236.1 (4.17) \text{ nm; }^{1}\text{H NMR } (400 \text{ MHz, CDCl}_{3}): \delta = 7.39$ (d, J = 7.9 Hz, 2H, HC = C (35/35')), 7.31 (t, J = 7.9 Hz, 2H, HC = C (36/35'))36'), 7.06(t, J = 7.3 Hz, 1H, HC = C(37)), 6.61(br, 1H, NH), 4.70(s, 1H, NH)) $CH_a(29)$), 4.60 (s, 1H, $CH_b(29)$), 4.47 (dd, J = 10.2, 5.9 Hz, 1H, CH(3)), 4.37 (d, J = 10.8 Hz, 1H, $CH_a(28)$), 3.95 (d, J = 10.8 Hz, 1H, $CH_b(28)$), 2.47 (dt, J = 11.0, 5.8 Hz, 1H, CH (19)), 2.04 (s, 3H, CH₃ (32)),2.03-1.96 (m, 1H, CH_a (21)), 1.92-1.82 (m, 1H, CH_a (16)), 1.81-1.71 $(m, 2H, CH_a(22) + CH_a(15)), 1.69 (s, 3H, CH_3(30)), 1.68 - 1.66 (m, 1H, 1H)$ CH(13)), 1.66-1.59 (m, 3H, $CH_a(1) + CH_2(12)$), 1.60-1.55 (m, 3H, CH(18) + CH₂(2), 1.53–1.48 (m, 1H, CH_a(6)), 1.46–1.36 (m, 6H, CH₂ $(7) + CH_b(21) + CH_2(11) + CH_b(6)$, 1.33–1.19 (m, 2H, CH (9) + CH_b (16)), 1.15-1.02 (m, 2H, CH_b (22) + CH_b (15)), 1.05 (s, 3H, CH_3 (25)), 1.02-0.93 (m, 1H, CH_b (1)), 0.98 (s, 3H, CH₃ (27)), 0.85 (s, 3H, CH₃ (23)), 0.84 (s, 3H, CH_3 (26)), 0.84 (s, 3H, CH_3 (24)), 0.79-0.76 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.0$ (C=0, C31), 154.0 (C=0, C33), 150.1 (C=CH₂, C20), 137.9 (C₀, C34), 129.0 (CH, C36/36'), 123.4 (CH, C37), 118.6 (CH, C35/35'), 109.9 (H₂C=C, C29), 80.9 (CH, C3), 63.6 (CH₂, C28), 55.4 (CH, C5), 50.3 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C_0 , C17), 42.7 (C_0 , C14), 40.9 (C_0 , C8), 38.4 (CH₂, C1), 37.8 (C_q, C4), 37.6 (CH, C13), 37.1 (C_q, C10), 34.5 (CH₂, C22), 34.1 (CH₂, C7), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 27.9 (CH₃, C23), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 23.7 (CH₂, C2), 21.3 (CH₃, C32), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.2 (CH₂, C6), 16.5 (CH₃, C24), 16.1 (CH₃, C26), 16.1 (CH₃, C25), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): m/ $z = 604.1 (6\%, [M + H]^+), 621.0 (10\%, [M + NH_4]^+), 626.3 (24\%,$

 $[M + Na]^+$), 1207.3 (48%, $[2M + H]^+$), 1229.3 (100%, $[2M + Na]^+$); analysis calcd for $C_{39}H_{57}NO_4$ (603.87): C 77.57, H 9.51, N 2.32; found: C 77.36, H 9.69, N 2.18.

4.3.34. (3 β) 3-Hydroxylup-20(29)-en-28-N-ethylcarbamate (**36**)

Deacetylation of 31 (0.98 g, 1.77 mmol) with potassium hydroxide (0.25 g, 5.3 mmol) in THF (30 mL) and MeOH (10 mL) for 2 days at 25 °C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 7:3) gave 36 (0.86 g, 94.5%) as a colorless solid; $R_F = 0.13$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 223–225 °C; $[\alpha]_D = +10.1^\circ$ (c = 0.29, CHCl₃); IR (KBr): $\nu = 3432br$, 2938s, 1689m, 1640w, 1536w, 1458w, 1375w, 1248m, 1024w, 883m, 543w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.67$ (s, 1H, CH_a (29)), 4.62 (br, 1H, NH), 4.57 (s, 1H, CH_b (29)), 4.24 (d, $J = 10.8 \text{ Hz}, 1\text{H}, CH_a(28), 3.83 (d, J = 10.5 \text{ Hz}, 1\text{H}, CH_b(28)), 3.21 (br,$ 2H, $CH_2(32)$), 3.17 (dd, J = 11.2, 5.0 Hz, 1H, CH(3)), 2.44 (dt, J = 11.1, 5.8 Hz, 1H, CH (19)), 1.98 (ddd, J = 19.5, 11.4, 7.0 Hz, 1H, CH_a (21)), 1.86-1.72 (m, 3H, CH_a (15) + CH_a (16) + CH_a (22)), 1.72-1.54 (m, 5H, $CH(18) + CH(13) + CH_a(1) + CH_a(2) + CH_a(12)$, 1.67 (s, 3H, CH_3) (30)), 1.54–1.48 (m, 2H, CH_b (2) + CH_a (6)), 1.44–1.33 (m, 5H, CH₂ $(7) + CH_b(6) + CH_a(11) + CH_b(21), 1.29 - 1.17 (m, 3H, CH(9) + CH_b(11))$ $(11) + CH_b(16)$, 1.13 (t, J = 7.2 Hz, 3H, $CH_3(33)$), 1.10–0.99 (m, 3H, $CH_b(12) + CH_b(15) + CH_b(22)$, 1.03 (s, 3H, $CH_3(25)$), 0.96 (s, 3H, CH_3 (27)), 0.96 (s, 3H, CH_3 (23)), 0.89 (dt, J = 12.8, 4.4 Hz, 1H, CH_b (1)), 0.81 (s, 3H, CH₃ (26)), 0.75 (s, 3H, CH₃ (24)), 0.67–0.65 (m, 1H, CH (5)) ppm; 13 C NMR (100 MHz, CDCl₃): $\delta = 157.0$ (C=0, C31), 150.3 (C=CH₂, C20), 109.7 (H₂C=C, C29), 79.0 (CH, C3), 62.9 (CH₂, C28), 55.3 (CH, C5), 50.4 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 $(C_q, C17), 42.7 (C_q, C14), 40.9 (C_q, C8), 38.8 (C_q, C4), 38.7 (CH_2, C1),$ 37.5 (CH, C13), 37.1 (C_q, C10), 35.9 (CH₂, C32), 34.6 (CH₂, C22), 34.2 (CH₂, C7), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 28.0 (CH₃, C23), 27.4 (CH₂, C2), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.3 (CH₂, C6), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 15.3 (CH₃, C24), 15.3 (CH₃, C33), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 514.1 $(14\%, [M + H]^+), 536.4 (6\%, [M + Na]^+), 1027.5 (100\%, [2M + H]^+),$ $1049.4 (96\%, [2M + Na]^+)$; analysis calcd for $C_{33}H_{55}NO_3 (513.79)$: C 77.14, H 10.79, N 2.73; found: C 77.04, H 10.90, N 2.59.

4.3.35. (3 β) 3-Hydroxylup-20(29)-en-28-N-propylcarbamate (37)

Deacetylation of 32 (0.53 g, 0.93 mmol) with potassium hydroxide (0.10 g, 1.8 mmol) in THF (5 mL)/MeOH (5 mL) for 1 day as decribed above, followed by chromatography (silica gel, hexanes/ ethyl acetate, 8:2) gave 37 (0.46 g, 93.0%) as a colorless solid; $R_F = 0.26$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 111–114 °C; $[\alpha]_D = +10.5^{\circ} (c = 0.31, CHCl_3); IR (KBr): \nu = 3456br, 2941s, 2869s,$ 2362w, 1702s, 1528m, 1460m, 1375w, 1244m, 1106w, 1045m, 883w, 668w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.64 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), 4.25 (d, J = 10.7 Hz, 1H, CH_a (28)), $3.84 (d, J = 10.6 \text{ Hz}, 1\text{H}, CH_b (28)), 3.18 (dd, J = 11.2, 5.0 \text{ Hz}, 1\text{H},$ CH(3)), 3.14 (*br*, 2H, $CH_2(32)$), 2.44 (*dt*, J = 11.1, 5.7 Hz, 1H, CH(19)), 2.03–1.92 (m, 1H, CH_a (21)), 1.87–1.79 (m, 1H, CH_a (16)), 1.79–1.69 $(m, 2H, CH_a(22) + CH_a(15)), 1.67 (s, 3H, CH_3(30)), 1.66-1.61 (m, 3H, CH_a(22) + CH_a(15)), 1.67 (s, 3H, CH_3(30)), 1.66-1.61 (m, 3H, CH_a(22) + CH_a(15)), 1.67 (s, 3H, CH_3(30)), 1.66-1.61 (m, 3H, CH_3(30)), 1.60 (m, 3H, CH_3(30)), 1.60 (m, 3H, CH_3(30)), 1.60 (m, 3H, CH_$ $CH_a(1) + CH(13) + CH_a(12)$, 1.61–1.55 (m, 3H, $CH(18) + CH_2(2)$), 1.55-1.47 (m, 3H, CH_2 (33) + CH_a (6)), 1.44-1.33 (m, 5H, CH_2 $(7) + CH_b(21) + CH_a(11) + CH_b(6)$, 1.29–1.17 (m, 3H, CH (19) + CH_b $(16) + CH_b(11), 1.10 - 0.99 (m, 3H, CH_b(22) + CH_b(15) + CH_b(12)),$ 1.04 (s, 3H, CH₃ (25)), 0.97 (s, 3H, CH₃ (23)), 0.96 (s, 3H, CH₃ (27)), $0.92 (t, J = 7.3 \text{ Hz}, 3H, CH_3 (34)), 0.89 - 0.84 (m, 1H, CH_b (1)), 0.82 (s, 1.85)$ 3H, CH_3 (26)), 0.75 (s, 3H, CH_3 (24)), 0.69–0.66 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 157.2$ (C=0, C31), 150.3 (C=CH₂, C20), 109.7 (H₂C=C, C29), 79.0 (CH, C3), 63.0 (CH₂, C28), 55.3 (CH, C5), 50.4 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (Cq, C17), 42.8 $(CH_2, C32), 42.7 (C_q, C14), 40.9 (C_q, C8), 38.8 (C_q, C4), 38.7 (CH_2, C1),$ 37.5 (CH, C13), 37.1 (C_q, C10), 34.6 (CH₂, C22), 34.2 (CH₂, C7), 29.9 (CH₂, C16), 29.7 (CH₂, C21), 28.0 (CH₃, C23), 27.4 (CH₂, C2), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 23.3 (CH₂, C33), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.3 (CH₂, C6), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 15.3 (CH₃, C24), 14.7 (CH₃, C27), 11.2 (CH₃, C34) ppm; MS (ESI, MeOH): m/z = 528.1 (14%, $[M + H]^+$), 1055.4 (100%, $[2M + H]^+$), 1077.5 (72%, $[2M + Na]^+$); analysis calcd for $C_{34}H_{57}NO_3$ (527.82): C 77.37, H 10.88, N 2.65; found: C 77.15, H 10.91, N 2.52.

4.3.36. (3 β) 3-Hydroxylup-20(29)-en-28-N-butylcarbamate (**38**)

Deacetylation of 33 (0.23 g, 0.39 mmol) with potassium hydroxide (0.11 g, 2.0 mmol) in THF (10 mL)/MeOH (5 mL) for 2 days at 25 °C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 38 (0.2 g, 94.0%) as a colorless solid; $R_F = 0.39$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 100–102 °C; $[\alpha]_D = +11.6^\circ$ (c = 0.43, CHCl₃); IR (KBr): $\nu = 3456br$, 2941s, 2870s, 2362w, 1700m, 1638w, 1522w, 1458m, 1374w, 1248w, 1136w, 1106w, 1047w, 984w, 882w, 774w, 638w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.62 (br, 1H, NH), 4.58 $(s, 1H, CH_b (29)), 4.25 (d, J = 10.9 Hz, 1H, CH_a (28)), 3.82 (d, J)$ $J = 10.5 \text{ Hz}, 1\text{H}, CH_b(28), 3.18 (m, 3\text{H}, CH_2(32) + CH(3)), 2.45 (dt, 3)$ $J = 11.0, 5.7 \text{ Hz}, 1\text{H}, CH(19), 2.03-1.92 (m, 1\text{H}, CH_a(21)), 1.86-1.76$ $(m, 1H, CH_a(16)), 1.76-1.69 (m, 2H, CH_a(22) + CH_a(15)), 1.67 (s, 3H, CH_a(16)), 1.76-1.69 (m, 2H, CH_a(22) + CH_a(15)), 1.67 (s, 3H, CH_a(16)), 1.76-1.69 (m, 2H, CH_a(22) + CH_a(15)), 1.67 (s, 3H, CH_a(16)), 1.76-1.69 (m, 2H, CH_a(22) + CH_a(15)), 1.67 (s, 3H, CH_a(16)), 1.76-1.69 (m, 2H, CH_a(16)),$ $CH_3(30)$), 1.66–1.61 (m, 3H, $CH_a(1) + CH(13) + CH_a(12)$), 1.61–1.55 $(m, 3H, CH_2(2) + CH(18)), 1.54-1.44 (m, 3H, CH_a(6) + CH_2(33)),$ 1.43-1.36 (m, 5H, CH_2 (7) + CH_b (21) + CH_a (11) + CH_b (6)), 1.36-1.30 (m, 2H, CH₂ (34)), 1.29-1.17 (m, 3H, CH_b (16) + CH $(9) + CH_b(11)$, 1.10-1.00 (m, 3H, $CH_b(22) + CH_b(15) + CH_b(12)$), 1.04 (s, 3H, CH₃ (25)), 0.96 (s, 3H, CH₃ (23)), 0.96 (s, 3H, CH₃ (27)), $0.92 (t, I = 7.3 \text{ Hz}, 3H, CH_3 (35)), 0.89 - 0.84 (m, 1H, CH_b (1)), 0.82 (s, 1.85)$ 3H, $CH_3(26)$), 0.76 (s, 3H, $CH_3(24)$), 0.69–0.66 (m, 1H, CH(5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 157.1$ (C=0, C31), 150.3 (C=CH₂, C20), 109.7 (H₂C=C, C29), 79.0 (CH, C3), 62.9 (CH₂, C28), 55.3 (CH, C5), 50.4 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (Cq, C17), 42.7 $(C_q, C14), 40.9 (C_q, C8), 40.8 (CH_2, C32), 38.8 (C_q, C4), 38.7 (CH_2, C1),$ 37.5 (CH, C13), 37.1 (C_q, C10), 34.6 (CH₂, C22), 34.2 (CH₂, C7), 32.1 (CH₂, C33), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 28.0 (CH₃, C23), 27.4 (CH₂, C2), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 20.8 (CH₂, C11), 19.9 (CH₂, C34), 19.1 (CH₃, C30), 18.3 (CH₂, C6), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 15.3 (CH₃, C24), 14.7 (CH₃, C27), 13.7 (CH₃, C35) ppm; MS (ESI, MeOH): $m/z = 542.1 (14\%, [M + H]^+), 1083.5 (100\%, [2M + H]^+),$ 1105.5 (22%, $[2M + Na]^+$); analysis calcd for $C_{35}H_{59}NO_3$ (541.85): C 77.58, H 10.98, N 2.58; found: C 77.40, H 11.14, N 2.41.

4.3.37. (3 β) 3-Hydroxylup-20(29)-en-28-N-hexylcarbamate (**39**)

Deacetylation of 34 (0.27 g, 0.56 mmol) with potassium hydroxide (0.1 g, 1.8 mmol) in THF (10 mL)/MeOH (5 mL) for 1 day as described above, followed by chromatography (silica gel, hexanes/ ethyl acetate, 8:2) gave 39 (0.22 g, 87.5%) as a colorless solid; $R_F = 0.32$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 95–97 °C; $[\alpha]_D = +11.7^{\circ}$ (c = 0.41, CHCl₃); IR (KBr): $\nu = 3456br$, 2940s, 2869s, 1702s, 1642w, 1523m, 1456m, 1375w, 1248m, 1106w, 1045m, 883w, 637w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.63 (br, 1H, NH), 4.57 (s, 1H, CH_b (29)), 4.25 (d, J = 10.4 Hz, 1H, CH_a (28)), 3.83 (d, J = 10.4 Hz, 1H, CH_b (28)), 3.17 (m, 3H, CH_2 (32) + CH_2 (3)), 2.44 (dt, J = 11.0, 5.7 Hz, 1H, CH (19)), 2.06–1.92 (m, 1H, CH_a (21)), 1.79 (dd, J = 17.5, 9.3 Hz, 1H, CH_a (16)), 1.76–1.69 (m, 2H, CH_a $(22) + CH_a$ (15)), 1.67 (s, 3H, CH₃ (30)), 1.66–1.61 (m, 3H, CH_a $(1) + CH(13) + CH_a(12), 1.61 - 1.51 (m, 4H, CH(18) + CH_2(2) + CH_a(12))$ (6)), 1.51-1.44 (m, 2H, CH_2 (33)), 1.44-1.36 (m, 5H, CH_2 (7) + CH_b $(21) + CH_a (11) + CH_b (6), 1.34-1.23 (m, 7H, CH (9) + CH_2)$ $(34) + CH_2(35) + CH_2(36)$, 1.23–1.14 (m, 2H, CH_b(11) + CH_b(16)), 1.09-1.01 (m, 3H, CH_b (22) + CH_b (15) + CH_b (12)), 1.03 (s, 3H, CH_3 (25)), 0.96 (s, 3H, CH₃ (23)), 0.96 (s, 3H, CH₃ (27)), 0.93–0.87 (m, 1H, $CH_b(1)$), 0.88 (t, J = 6.7 Hz, 3H, $CH_3(37)$), 0.82 ($s, 3H, CH_3(26)$), 0.75 (s, 3H, CH₃ (24)), 0.69–0.66 (m, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 157.2$ (C=0, C31), 150.3 (C=CH₂, C20), 109.7 (H₂C=C, C29), 79.0 (CH, C3), 62.9 (CH₂, C28), 55.3 (CH, C5), 50.4 (CH, C9), 48.8 (CH, C18), 47.7 (CH, C19), 46.6 (C_q , C17), 42.7 (C_q , C14), 41.1 (CH₂, C32), 40.9 (C_q , C8), 38.8 (C_q , C4), 38.7 (CH₂, C1), 37.5 (CH, C13), 37.1 (C_q , C10), 34.6 (CH₂, C22), 34.2 (CH₂, C7), 31.5 (CH₂, C35), 30.0 (CH₂, C33), 29.8 (CH₂, C16), 29.7 (CH₂, C21), 28.0 (CH₃, C23), 27.4 (CH₂, C2), 27.1 (CH₂, C15), 26.4 (CH₂, C34), 25.2 (CH₂, C12), 22.5 (CH₂, C36), 20.8 (CH₂, C11), 19.1 (CH₃, C30), 18.3 (CH₂, C6), 16.1 (CH₃, C26), 16.0 (CH₃, C25), 15.3 (CH₃, C24), 14.7 (CH₃, C27), 14.0 (CH₃, C37) ppm; MS (ESI, MeOH): m/z = 570.1 (12%, [M + H]⁺), 592.3 (4%, [M + Na]⁺), 1139.5 (100%, [2M + H]⁺), 1161.5 (60%, [2M + Na]⁺); analysis calcd for C₃₇H₆₃NO₃ (569.90): C 77.98, H 11.14, N 2.46; found: C 77.77, H 11.29, N 2.27.

4.3.38. (3 β) 3-Hydroxylup-20(29)-en-28-N-phenylcarbamate (**40**)

Deacetylation of **35** (1 g, 1.8 mmol) with potassium hydroxide (0.4 g, 7.16 mmol) in THF (30 mL)/MeOH (20 mL) for 2 days at 25 $^{\circ}$ C as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 7:3) gave 40 (0.83 g, 89.2%) as a colorless solid; $R_F = 0.28$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 233–234 °C; $[\alpha]_D = +7.2^{\circ} (c = 0.27, \text{CHCl}_3); \text{ IR (KBr): } \nu = 3511br, 2945s, 2867m,$ 1721s, 1602m, 1550s, 1503w, 1446s, 1385w, 1316w, 1228s, 1086m, 1070*m*, 1031*w*, 984*w*, 886*w*, 751*m*, 692*w* 506*w* cm⁻¹; UV–vis (CHCl₃): $\lambda_{\text{max}} (\log \varepsilon) = 235.8 (4.06) \text{ nm}; {}^{1}\text{H NMR} (400 \text{ MHz, CDCl}_{3}):$ $\delta = 7.39 (d, J = 7.9 \text{ Hz}, 2H, HC=C (33/33')), 7.31 (t, J = 7.9 \text{ Hz}, 2H, 2H, 2H, 3H)$ HC = C(34/34'), 7.06 (t, J = 7.3 Hz, 1H, HC = C(35)), 6.63 (br, 1H, NH), $4.70 (s, 1H, CH_a (29)), 4.60 (s, 1H, CH_b (29)), 4.37 (d, J = 10.7 Hz, 1H, 1H, 1H)$ $CH_a(28)$), 3.96 (d, J = 10.8 Hz, 1H, $CH_b(28)$), 3.18 (dd, J = 11.1, 5.0 Hz, 1H, CH (3)), 2.47 (dt, I = 11.0, 5.8 Hz, 1H, CH (19)), 2.02 (ddd, I = 19.2, 13.9, 10.5 Hz, 1H, CH_a (21)), 1.93–1.82 (m, 1H, CH_a (16)), 1.81–1.72 $(m, 2H, CH_a(22) + CH_a(15)), 1.69 (s, 3H, CH_3(30)), 1.68 - 1.55 (m, 4H, CH_a(30)), 1.68 (m, 4H, CH_a(30))$ $CH(18) + CH_a(1) + CH(13) + CH_a(12), 1.55-1.44 (m, 3H, CH₂)$ $(2) + CH_a(6), 1.44 - 1.36 (m, 5H, CH_2(7) + CH_b(21) + CH_a(11) + CH_b$ (6)), 1.32-1.22 (m, 3H, CH (9) + CH_b (16) + CH_b (11)), 1.15-1.02 (m, 2H, CH_b (22) + CH_b (15)), 1.05 (s, 3H, CH_3 (25)), 1.01–0.95 (m, 1H, CH_b (12)), 0.98 (s, 3H, CH₃ (27)), 0.97 (s, 3H, CH₃ (23)), 0.90 (dt, $J = 12.9, 4.3 \text{ Hz}, 1\text{H}, CH_b(1), 0.83 (s, 3\text{H}, CH_3(26)), 0.76 (s, 3\text{H}, CH_3(26))$ (24)), 0.69–0.66 (*m*, 1H, CH (5)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 154.1 (C=0, C31), 150.1 (C=CH₂, C20), 137.9 (C₀, C32), 129.0 (CH, CH₂, C20), 129.0 (CH₂, C20), 129.0 (CH, CH₂, C20), 129.0 (CH, CH_$ C34/34'), 123.4 (CH, C35), 118.6 (CH, C33/33'), 109.8 (H₂C=C, C29), 79.0 (CH, C3), 63.5 (CH₂, C28), 55.3 (CH, C5), 50.4 (CH, C9), 48.9 (CH, C18), 47.7 (CH, C19), 46.6 (C_q , C17), 42.7 (C_q , C14), 40.9 (C_q , C8), 38.9 (C_q, C4), 38.7 (CH₂, C1), 37.6 (CH, C13), 37.2 (C_q, C10), 34.6 (CH₂, C22), 34.2 (CH₂, C7), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 28.0 (CH₃, C23), 27.4 (CH₂, C2), 27.1 (CH₂, C15), 25.2 (CH₂, C12), 20.8 (CH₂, C11), 19.2 (CH₃, C30), 18.3 (CH₂, C6), 16.1 (CH₃, C26), 16.1 (CH₃, C25), 15.3 (CH₃, C24), 14.8 (CH₃, C27) ppm; MS (ESI, MeOH): m/z = 579.1 (6%, $[M + NH_4]^+$), 584.3 (6%, $[M + Na]^+$), 1123.4 (48%, $[2M + H]^+$), 1146.5 $(44\%, [2M + Na]^+)$; analysis calcd for $C_{37}H_{55}NO_3$ (561.84): C 79.10, H 9.87, N 2.49; found: C 79.03, H 10.02, N 2.33.

4.3.39. 28-O-Acetyl-betulin (41)

Acetylation of **2** (15.0 g, 33.88 mmol) in DCM (100 mL) with acetic anhydride (3.2 mL, 33.88 mmol) in the presence of NEt₃ (4.3 mL) and DMAP (0.24 g, 1.96 mmol) as previously described, followed by usual workup and chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave **41** (9.87 g, 60.4%) as a colorless solid; $R_F = 0.40$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 216–217 °C (lit.: [63] 214–215 °C); $[\alpha]_D = +11.1^\circ$ (c = 0.39, CHCl₃), (lit.: [64] $[\alpha]_D = +10.3^\circ$ (CHCl₃)); MS (ESI, MeOH): m/z = 467.3 (100%, $[M+H-H_2O]^+$), 485.1 (4%, $[M+H]^+$), 991.4 (16%, $[2M+Na]^+$).

4.3.40. 28-Acetyloxy-lup-20(29)-en-3-one (42)

Jones oxidation of **41** (3.0 g, 6.22 mmol) for 40 min at 25 °C as previously described, followed by usual aqueous workup and chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave **42**

(2.14 g, 71.6%) as a colorless solid; $R_F = 0.73$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 91–94 °C (lit.: [65] 77–79 °C); $[\alpha]_D = +37.1^\circ$ (c = 0.35, CHCl₃), (lit.: [66] $[\alpha]_D = +38.0^\circ$ (CHCl₃)); MS (ESI, MeOH): m/z = 483.2 (26%, $[M+H]^+$), 505.3 (8%, $[M+Na]^+$), 536.8 (10%, $[M+Na+MeOH]^+$), 965.1 (12%, $[2M+H]^+$), 987.3 (70%, $[2M+Na]^+$).

4.3.41. 28-Hydroxy-lup-20(29)en-3-one (43)

Deacetylation of 42 (2.1 g, 4.36 mmol) with KOH (0.61 g, 10.9 mmol) in THF (20 mL) and MeOH (10 mL) for 12 h at 25 °C as described above, followed by usual workup and chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 43 (1.73 g, 90.4%) as a colorless solid; $R_F=0.45$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 103-105 °C (lit.: [67] 94-96 °C); $[\alpha]_D=+45.0$ ° (c=0.40, CHCl₃), (lit.: [67] $[\alpha]_D=+52.7$ ° (CHCl₃)); MS (ESI, MeOH): m/z=423.1 (31%, $[M+H-H_2O]^+$), 441.2 (46%, $[M+H]^+$), 881.4 (29%, $[2M+H]^+$), 903.4 (100%, $[2M+Na]^+$).

4.3.42. 3-Oxo-lup-20(29)-en-28-N-ethylcarbamate (44)

Microwave assisted reaction (7 h, 120 °C) of 43 (0.3 g, 0.682 mmol) in dry THF (5 mL) with ethyl isocyanate (0.15 mg, 0.16 mL, 2.05 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 44 (0.237 g, 68.0%) as a colorless solid; $R_F = 0.32$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 83–86 °C; [α]_D = +34.7° (c = 0.32, CHCl₃); IR (KBr): v = 3441s, 2945s, 2870m, 1706s, 1639m, 1523w, 1458m, 1384w, 1243m, 1138w, 1112w, 1080w, 1023m, 882w, 774w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.60 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), 4.26 (d, I = 10.7 Hz, 1H, CH_a (28)), 3.83 (d, $I = 10.9 \text{ Hz}, 1\text{H}, CH_b(28), 3.22 (br, 2\text{H}, CH_2(32)), 2.53 - 2.43 (m, 2\text{H}, 2\text{H},$ $CH(19) + CH_a(2)$, 2.37 (ddd, I = 15.7, 7.4, 4.5 Hz, 1H, CH_b(2)),2.05-1.94 (m, 1H, CH_a (21)), 1.92-1.86 (m, 1H, CH_a (1)), 1.81-1.71 $(m, 3H, CH_a (22) + CH_a (16) + CH_a (15)), 1.68 (s, 3H, CH_3 (30)),$ 1.68-1.64 (m, 2H, CH (13) + CH_a (12)), 1.62-1.56 (m, 1H, CH (18)), 1.49-1.39 (m, 4H, CH₂ (6) + CH₂ (7)), 1.36-1.30 (m, 4H, CH $(9) + CH_b(1) + CH_b(21) + CH_a(11), 1.28-1.26$ (m, 1H, CH (5)), 1.25-1.20 (m, 2H, CH_b (11) + CH_b (16)), 1.14 (t, J = 7.2 Hz, 3H, CH₃ (33)), 1.08 (s, 3H, CH₃ (25)), 1.06 (s, 3H, CH₃, (23)), 1.05–1.00 (m, 3H, $CH_b(22) + CH_b(15) + CH_b(12)$, $1.02(s, 3H, CH_3(24))$, $0.98(s, 3H, CH_3(24))$ 3H, CH₃ (27)), 0.93 (s, 3H, CH₃ (26)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 218.0$ (C=0, C3), 157.0 (C=0, C31), 150.2 (C=CH₂, C20), 109.8 (H₂C=C, C29), 62.8 (CH₂, C28), 55.0 (CH, C5), 49.8 (CH, C9), 48.7 (CH, C18), 47.6 (CH, C19), 47.3 (C_q, C4), 46.6 (C_q, C17), 42.8 $(C_q, C14), 40.8 (C_q, C8), 39.6 (CH_2, C1), 37.6 (CH, C13), 36.9 (C_q, C10),$ 36.1 (CH₂, C32), 34.5 (CH₂, C22), 34.1 (CH₂, C2), 33.5 (CH₂, C7), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 27.1 (CH₂, C15), 26.5 (CH₃, C23), 25.2 (CH₂, C12), 21.3 (CH₂, C11), 21.0 (CH₃, C24), 19.6 (CH₂, C6), 19.1 (CH₃, C30), 15.9 (CH₃, 25), 15.8 (CH₃, C26), 15.3 (CH₃, C33), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 512.1 (100\%, [M + H]^+)$, 529.1 (7%, $[M + NH_4]^+$), 534.3 (17%, $[M + Na]^+$), 1023.5 (100%, $[2M + H]^+$), 1025.5 (64%, $[2M + Na]^+$); analysis calcd for C₃₃H₅₃NO₃ (511.78): C 77.45, H 10.44, N 2.74; found: C 77.39, H 10.51, N 2.57.

4.3.43. 3-Oxo-lup-20(29)-en-28-N-propylcarbamate (45)

Microwave assisted reaction (7 h, 120 °C) of **43** (0.29 g, 0.659 mmol) in dry THF (5 mL) with propyl isocyanate (0.17 g, 0.19 mL, 1.98 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave **45** (0.216 g, 62.0%) as a colorless solid; $R_F = 0.35$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 87–91 °C; [α]_D = +34.5° (c = 3.4, CHCl₃); IR (KBr): $\nu = 3406$ s, 2958s, 2870m, 1706s, 1642m, 1530m, 1459m, 1384m, 1236m, 1138m, 1112m, 1044m, 1004m, 882m, 776m cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.72-4.64$ (br, 1H, NH), 4.67 (s, 1H, CH_a (29)), 4.57 (s, 1H, CH_b (29)), 4.26 (d, J = 10.8 Hz, 1H, CH_a (28)), 3.82 (d, J = 10.7 Hz, 1H,

 CH_b (28)), 3.13 (br, 2H, CH_2 (32)), 2.53–2.42 (m, 2H, CH (19) + CH_a (2)), 2.37 (ddd, J = 15.6, 7.5, 4.4 Hz, 1H, $CH_b(2)$), 2.07–1.93 (m, 1H, CH_a (21)), 1.92-1.82 (m, 1H, CH_a (1)), 1.82-1.69 (m, 3H, CH_a $(16) + CH_a(22) + CH_a(15)$, 1.67 (s, 3H, CH₃(30)), 1.69–1.55 (m, 4H, CH (13) + CH_2 (12) + CH (18)), 1.54–1.47 (m, 2H, CH_2 (33)), 1.47-1.33 (m, 8H, CH_2 (6) + CH_2 (7) + CH_a (11) + CH_b (21) + CH_b (1) + CH(9), 1.25 (m, 3H, CH(5) + $CH_b(11) + CH_b(16)$), 1.10–0.99 $(m, 2H, CH_b(22) + CH_b(15)), 1.07(s, 3H, CH_3(25)) 1.05(s, 3H, CH_3, CH_3))$ (23)), 1.01 (s, 3H, CH₃ (24)), 0.97 (s, 3H, CH₃, (27)), 0.9 (s, 3H, CH₃ (26)), 0.91 (t, J = 2.0 Hz, 3H, CH_3 (34)) ppm; ¹³C NMR (100 MHz. CDCl₃): $\delta = 218.0$ (C=0, C3), 157.1 (C=0, C31), 150.2 (C=CH₂, C20), 109.8 (H₂C=C, C29), 62.8 (CH₂, C28), 55.0 (CH, C5), 49.7 (CH, C9), 48.7 (CH, C18), 47.6 (CH, C19), 47.3 (C_q, C4), 46.6 (C_q, C17), 43.3 (CH₂, C32), 42.7 (C_q, C14), 40.8 (C_q, C8), 39.6 (CH₂, C1), 37.6 (CH, C13), 36.9 (C₀, C10), 34.5 (CH₂, C22), 34.1 (CH₂, C2), 33.5 (CH₂, C7), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 27.1 (CH₂, C15), 26.5 (CH₃, C23), 25.2 (CH₂, C2), 23.2 (CH₂, C33), 21.3 (CH₂, C11), 21.0 (CH₃, C24), 19.6 (CH₂, C6), 19.1 (CH₃, C30), 15.9 (CH₃, 25), 15.8 (CH₃, C26), 14.7 (CH₃, C27), 11.2 (CH₃, C34) ppm; MS (ESI, MeOH): $m/z = 526.2 (40\%, [M + H]^+), 548.4 (6\%, M)$ $[M + Na]^+$), 1051.6 (100%, $[2M + H]^+$], 1073.6 (46%, $[2M + Na]^+$); analysis calcd for C₃₄H₅₅NO₃ (525.81): C 77.66, H 10.54, N 2.66; found: C 77.51, H 10.69, N 2.50.

4.3.44. 3-Oxo-lup-20(29)-en-28-N-butylcarbamate (46)

Microwave assisted reaction (7 h, 120 °C) of 43 (0.28 g, 0.682 mmol) in dry THF (5 mL) with butyl isocyanate (0.2 g, 0.23 mL, 2.05 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 46 (0.246 g, 72.0%) as a colorless solid; $R_F = 0.51$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 84–86 °C; $[\alpha]_D = +32.4^\circ$ (c = 3.65, CHCl₃); IR (KBr): $\nu = 3416s$, 2956s, 2870m, 1706s, 1642w, 1530m, 1460m, 1384w, 1245m, 1136w, 1112w, 1052w, 1021m, 882w, 776w cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.64 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), 4.26 (d, J = 10.5 Hz, 1H, CH_a (28)), 3.83 (d, $J = 10.5 \text{ Hz}, 1\text{H}, CH_b(28), 3.17 (br, 2\text{H}, CH_2(32)), 2.55-2.42 (m, 2\text{H}, 2\text{H})$ $CH(19) + CH_a(2)$, 2.38 (ddd, J = 15.5, 7.4, 4.3 Hz, 1H, $CH_b(2)$), 2.07–1.93 (m, 1H, CH_a (21)), 1.92–1.83 (m, 1H, CH_a (1)), 1.82–1.69 $(m, 3H, CH_a (16) + CH_a (22) + CH_a (15)), 1.69-1.54 (m, 4H, CH)$ $(13) + CH_2(12) + CH(18)$, 1.67 (s, 3H, CH₃(30)), 1.53–1.41 (m, 6H, $CH_2(33) + CH_2(6) + CH_2(7)$), 1.41–1.33 (m, 5H, $CH_2(11) + CH_b$ $(21) + CH_b(1) + CH(9), 1.32 - 1.17 (m, 4H, CH(5) + CH_b(16) + CH_2$ (34)), 1.11-1.05 (m, 2H, CH_b $(22) + CH_b$ (15)), 1.07 (s, 3H, CH_3 (25)), 1.06 (s, 3H, CH₃ (23)), 1.02 (s, 3H, CH₃ (24)), 0.97 (s, 3H, CH₃ (27)), 0.94 (s, 3H, CH₃ (26)), 0.92 (t, J = 7.7 Hz, 3H, CH₃ (35)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 218.0$ (C=0, C3), 157.2 (C=0, C31), 150.2 (C=CH₂, C20), 109.8 (H₂C=O, C29), 62.8 (CH₂, C28), 55.0 (CH, C5), 49.7 (CH, C9), 48.8 (CH, C18), 47.6 (CH, C19), 47.3 (C_q, C4), 46.6 (C_q, C17), 42.8 (C_q, C14), 40.8 (C_q, C8), 40.8 (CH₂, C32), 39.1 (CH₂, C1), 37.6 (CH, C13), 36.9 (C_q, C10), 34.5 (CH₂, C22), 34.1 (CH₂, C2), 33.5 (CH₂, C7), 32.1 (CH₂, C33), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 27.1 (CH₂, C15), 26.5 (CH₃, C23), 25.2 (CH₂, C12), 21.3 (CH₂, C11), 21.0 (CH₃, C24), 19.9 (CH₂, C34), 19.6 (CH₂, C6), 19.1 (CH₃, C30), 15.9 (CH₃, C25), 15.9 (CH₃, C26), 14.7 (CH₃, C27), 13.7 (CH₃, C35) ppm; MS (ESI, MeOH): m/z = 540.2 (72%, [M + H]⁺), 562.5 $(8\%, [M + Na]^+), 1079.5 (100\%, [2M + H]^+), 1101.5 (49\%,$ $[2M + Na]^+$); analysis calcd for $C_{35}H_{57}NO_3$ (539.83): C 77.87, H 10.64, N 2.59; found: C 77.61, H 10.83, N 2.41.

4.3.45. 3-Oxo-lup-20(29)-en-28-N-hexylcarbamate (47)

Microwave assisted reaction (7 h, 120 °C) of **43** (0.29 g, 0.66 mmol) in dry THF (5 mL) with hexyl isocyanate (0.25 g, 0.29 mL, 1.98 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave **47** (0.205 g, 55.1%) as a colorless solid; $R_F = 0.57$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 76–79 °C; $[\alpha]_D = +29.8^\circ$ (c = 0.52, CHCl₃); IR (KBr):

 $\nu = 3397m$, 2951s, 2869s, 1706s, 1640w, 1522m, 1458m, 1384w, 1243*m*, 1138*w*, 1112*w*, 1041*w*, 1018*w*, 882*w*, 776*w*, 580*w* cm⁻¹; ¹H NMR (400 MHz, CDCl₃): $\delta = 4.68$ (s, 1H, CH_a (29)), 4.63 (br, 1H, NH), 4.58 (s, 1H, CH_b (29)), 4.26 (d, J = 10.7 Hz, 1H, CH_a (28)), 3.83 (d, $J = 10.8 \text{ Hz}, 1\text{H}, CH_b(28), 3.17 (br, 2\text{H}, CH_2(32)), 2.54-2.42 (m, 2\text{H}, 2\text{H})$ $CH(19) + CH_a(2) \times 2.38 (ddd, I = 15.6, 7.5, 4.3 Hz, 1H, CH_b(2)), 2.00$ $(m, 1H, CH_a(21)), 1.93-1.83 (m, 1H, CH_a(1)), 1.83-1.69 (m, 3H, CH_a(1)), 1.83-1.69$ $(16) + CH_a(22) + CH_a(15)$, 1.67 (s, 3H, CH₃(30)), 1.69–1.54 (m, 4H, $CH(13) + CH_2(12) + CH(18)$, 1.54–1.40 (m, 6H, $CH_2(33) + CH_2$ (6) + CH₂(7), 1.39-1.31 (m, 4H, CH(9) + CH_b(1) + CH_b(21) + CH_a(21))(11), 1.30-1.26 (m, 7H, CH₂ (34) + CH₂ (36) + CH₂ (35) + CH (5)),1.26-1.16 (m, 2H, CH_b (11) + CH_b (16)), 1.07 (s, 3H, CH₃ (25)), 1.06 (s, 3H, CH_3 (23)) 1.12–0.95 (m, 2H, CH_b (22) + CH_b (15)), 1.02 (s, 3H, CH₃ (24)), 0.97 (s, 3H, CH₃ (27)), 0.92 (s, 3H, CH₃ (26)), 0.88 (t, I = 6.5 Hz, 3H, CH₃ (37)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 218.0$ (C=0, C3), 157.1 (C=0, C31), 150.2 $(C=CH_2, C20)$, 109.8 $(H_2C=C, C30)$ C29), 62.8 (CH₂, C28), 55.0 (CH, C5), 49.8 (CH, C9), 48.8 (CH, C18), 47.6 (CH, C19), 47.3 (C_q , C4), 46.6 (C_q , C17), 42.8 (C_q , C14), 41.1 (CH₂, C32), 40.8 (C_q, C8), 39.6 (CH₂, C1), 37.6 (CH, C13), 36.9 (C_q, C10), 34.5 (CH₂, C22), 34.1 (CH₂, C2), 33.5 (CH₂, C7), 31.5 (CH₂, C35), 30.0 (CH₂, C33), 29.8 (CH₂, C6), 29.7 (CH₂, C21), 27.1 (CH₂, C15), 26.5 (CH₃, C23), 26.4 (CH₂, C34), 25.2 (CH₂, C12), 22.5 (CH₂, C36), 21.3 (CH₂, C11), 21.0 (CH₃, C24), 19.6 (CH₂, C6), 19.1 (CH₃, C30), 15.9 (CH₃, C25), 15.8 (CH₃, C26), 14.7 (CH₃, C27), 14.0 (CH₃, C37) ppm; MS (ESI, MeOH): $m/z = 568.1 (30\%, [M + H]^+), 590.5 (4\%, [M + Na]^+), 1135.6$ $(100\%, [2M + H]^+)$, 1157.8 $(32\%, [2M + Na]^+)$; analysis calcd for C₃₇H₆₁NO₃ (567.89): C 78.25, H 10.83, N 2.47; found: C 78.01, H 10.97. N 2.38.

4.3.46. 3-Oxo-lup-20(29)-en-28-N-phenylcarbamate (48)

Microwave assisted reaction (7 h, 120 °C) of 43 (0.3 g, 0.682 mmol) in dry THF (5 mL) with phenyl isocyanate (0.24 g, 0.22 mL, 2.05 mmol) as described above, followed by chromatography (silica gel, hexanes/ethyl acetate, 8:2) gave 48 (0.325 g, 85.3%) as a colorless solid; $R_F = 0.53$ (silica gel, hexanes/ethyl acetate, 8:2); m.p.: 200–202 °C; $[\alpha]_D = +23.3^\circ$ (c = 0.46, CHCl₃); IR (KBr): $\nu = 3443s$, 2945s, 2868m, 2362w, 2344w, 1734s, 1700s, 1602m, 1540m, 1442m, 1386w, 1374w, 1216m, 1057w, 1030w, 882w, 752w, 692w cm⁻¹; UV–vis (CHCl₃): λ_{max} (log ε) = 234.4 (4.23) nm; ¹H NMR (400 MHz, CDCl₃): $\delta = 7.39$ (*d*, J = 7.9 Hz, 2H, HC = C (33/ 33')), 7.31 (t, J = 7.8 Hz, 2H, HC = C(34/34')), 7.06 (t, J = 7.3 Hz, 1H, HC=C(35)), 6.63 (br, 1H, NH), 4.70 (s, 1H, $CH_a(29)$), 4.60 (s, 1H, CH_b (29)), 4.38 (d, J = 10.8 Hz, 1H, CH_a (28)), 3.96 (d, J = 10.7 Hz, 1H, CH_b (28)), 2.55-2.43 (m, 2H, CH (19) + CH_a (2)), 2.39 (ddd, J = 15.6, 7.4, 4.2 Hz, 1H, CH_b (2)), 2.10–1.95 (m, 1H, CH_a (21)), 1.95–1.80 (m, 3H, $CH_{a}(1) + CH_{a}(22) + CH_{a}(16)$), 1.78–1.57 (m, 4H, $CH_{a}(15) + CH_{a}(16)$) (12) + CH 13 + CH (18), 1.69 (s, 3H, CH₃ (30)), 1.55–1.35 (m, 8H, CH₂ $(7) + CH_2(6) + CH_b(21) + CH_a(11) + CH_b(1) + CH(9), 1.35-1.22$ $(m, 3H, CH (5) + CH_b (11) + CH_b (16)), 1.14-1.04 (m, 3H, CH_b)$ $(22) + CH_b (15) + CH_b (12) + CH_b (12)$ (23)), 1.03 (s, 3H, CH₃ (24)), 0.99 (s, 3H, CH₃ (27)), 0.94 (s, 3H, CH₃ (26)) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 218.5$ (C=0, C3), 154.0 (C=0, C31), 150.0 (C=CH₂, C20), 137.9 (C_q, C32), 129.1 (CH, C34/ 34'), 123.4 (CH, C35), 118.6 (CH, C33/33'), 109.9 (H₂C=C, C29), 63.5 (CH₂, C28), 55.0 (CH, C5), 49.7 (CH, C9), 48.8 (CH, C18), 47.6 (CH, C19), 47.4 (*C*_q, C4), 46.6 (*C*_q, C17), 42.8 (*C*_q, C14), 40.8 (*C*_q, C8), 39.6 (CH₂, C1), 37.7 (CH, C13), 36.9 (C_q, C10), 34.5 (CH₂, C22), 34.2 (CH₂, C2), 33.5 (CH₂, C7), 29.8 (CH₂, C16), 29.6 (CH₂, C21), 27.1 (CH₂, C15), 26.6 (CH₃, C23), 25.2 (CH₂, C12), 21.3 (CH₂, C11), 21.1 (CH₃, C24), 19.6 (CH₂, C6), 19.2 (CH₃, C30), 15.9 (CH₃, C25), 15.9 (CH₃, C26), 14.7 (CH₃, C27) ppm; MS (ESI, MeOH): $m/z = 560.0 (12\%, [M + H]^+), 582.3$ $(34\%, [M + Na]^+)$, 613.3 $(10\%, [M + Na + MeOH]^+)$, 1119.3 (58%, MeOH) $[2M + H]^+$), 1141.5 (100%, $[2M + Na]^+$); analysis calcd for C₃₇H₅₃NO₃ (559.82): C 79.38, H 9.54, N 2.50; found: C 79.13, H 9.61, N 2.43.

Acknowledgments

Thanks are due to Mrs J. Wiese for measuring the IR spectra and optical rotations. The cell lines were kindly provided by Dr. Th. Müller (Dept. of Haematology/Oncology, Martin-Luther Universität Halle-Wittenberg). Support by the "Gründerwerkstatt—Biowissenschaften" is gratefully acknowledged.

Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.ejmech.2015.10.043.

References

- [1] H. Lossen, Über die Oxalohydroxamsäure, Liebigs Ann. Chem. 150 (1869) 314–316
- [2] A.S. Begum, V.K. Jain, C.L. Khetrapal, N.C. Shivaprakash, Structural studies of oxalohydroxamic acid by single-crystal X-ray-diffraction and spectroscopic methods, J. Cryst. Spectrosc. 17 (1987) 545–555.
- [3] S.P.E. Gupta, Hydroxamic Acids a Unique Family of Chemicals with Multiple Biological Activities, Springer, Berlin, 2013.
- [4] E.M.F. Muri, M.J. Nieto, R.D. Sindelar, J.S. Williamson, Hydroxamic acids as pharmacological agents, Curr. Med. Chem. 9 (2002) 1631–1653.
- [5] M. Sani, D. Belotti, R. Giavazzi, W. Panzeri, A. Volonterio, M. Zanda, Synthesis and evaluation of stereopure alpha-trifluoromethyl-malic hydroxamates as inhibitors of matrix metalloproteinases, Tetrahedron Lett. 45 (2004) 1611–1615.
- [6] D.A. Brown, W.K. Glass, N.J. Fitzpatrick, T.J. Kemp, W. Errington, G.J. Clarkson, W. Haase, F. Karsten, A.H. Mahdy, Structural variations in dinuclear model hydrolases and hydroxamate inhibitor models: synthetic, spectroscopic and structural studies, Inorg, Chim. Acta 357 (2004) 1411–1436.
- [7] Z. Amtul, Atta-ur-Rahman, R.A. Siddiqui, M.I. Choudhary, Chemistry and mechanism of urease inhibition, Curr. Med. Chem. 9 (2002) 1323–1348.
- [8] C. Pergola, O. Werz, 5-Lipoxygenase inhibitors: a review of recent developments and patents, Expert Opin. Ther. Pat. 20 (2010) 355–375.
- [9] G.R. Ott, N. Asakawa, R.Q. Liu, M.B. Covington, M.X. Qian, K. Vaddi, R.C. Newton, J.M. Trzaskos, D.D. Christ, L. Galya, T. Scholz, W. Marshall, J.J.W. Duan, alpha,beta-Cyclic-beta-benzamido hydroxamic acids: novel oxaspiro[4.4] nonane templates for the discovery of potent, selective, orally bioavailable inhibitors of tumor necrosis factor-alpha converting enzyme (TACE), Bioorg. Med. Chem. Lett. 18 (2008) 1288—1292.
- [10] G.R. Ott, N. Asakawa, Z. Lu, R.Q. Liu, M.B. Covington, K. Vaddi, M. Qian, R.C. Newton, D.D. Christ, J.M. Traskos, C.P.D. James, alpha,beta-Cyclic-beta-benzamido hydroxamic acids: novel templates for the design, synthesis, and evaluation of selective inhibitors of TNF-alpha converting enzyme (TACE), Bioorg, Med. Chem. Lett. 18 (2008) 694–699.
- [11] A. Scozzafava, A. Mastrolorenzo, C.T. Supuran, Modulation of carbonic anhydrase activity and its applications in therapy, Expert Opin. Ther. Pat. 14 (2004) 667–702.
- [12] J.Y. Winum, A. Scozzafava, J.L. Montero, C.T. Supuran, Therapeutic applications of sulfamates, Expert Opin. Ther. Pat. 14 (2004) 1273–1308.
- [13] B. Stearns, K.A. Losee, J. Bernstein, Hydroxyurea a new type of potential antitumor agent, J. Med. Chem. 6 (1963) 201.
- [14] B. Chetan, M. Bunha, M. Jagrat, B.N. Sinha, P. Saiko, G. Graser, T. Szekeres, G. Raman, P. Rajendran, D. Moorthy, A. Basu, V. Jayaprakash, Design, synthesis and anticancer activity of piperazine hydroxamates and their histone deacetylase (HDAC) inhibitory activity, Bioorg. Med. Chem. Lett. 20 (2010) 3906—3910
- [15] C.J. Marmion, D. Griffith, K.B. Nolan, Hydroxamic acids an intriguing family of enzyme inhibitors and biomedical ligands, Eur. J. Inorg. Chem. (2004) 3003—3016
- [16] L.P. Tardibono, M.J. Miller, Synthesis and anticancer activity of new hydroxamic acid containing 1,4-benzodiazepines, Org. Lett. 11 (2009) 1575–1578.
- [17] P. Kahnberg, A.J. Lucke, M.P. Glenn, G.M. Boyle, J.D.A. Tyndall, P.G. Parsons, D.P. Fairlie, Design, synthesis, potency, and cytoselectivity of anticancer agents derived by parallel synthesis from alpha-aminosuberic acid, J. Med. Chem. 49 (2006) 7611–7622.
- [18] J. Jiang, A. Thyagarajan-Sahu, V. Krchňák, A. Jedinak, G.E. Sandusky, D. Sliva, NAHA, a novel hydroxamic acid-derivative, inhibits growth and angiogenesis of breast cancer in vitro and in vivo, PLoS ONE 7 (2012) e34283.
- [19] S. Mochizuki, Y. Okada, ADAMs in cancer cell proliferation and progression, Cancer Sci. 98 (2007) 621–628.
- [20] R. Csuk, Recent developments in the synthesis of antitumor-active glycyrrhetinic acid derivatives, Mini-Rev. Org. Chem. 11 (2014) 253–261.
- [21] B. Siewert, E. Pianowski, A. Obernauer, R. Csuk, Towards cytotoxic and selective derivatives of maslinic acid, Bioorg. Med. Chem. 22 (2014) 594–615.
- [22] R. Csuk, Betulinic acid and its derivatives: a patent review (2008–2013), Expert Opin. Ther. Pat. 24 (2014) 913–923.

- [23] S. Sommerwerk, L. Heller, R. Csuk, Synthesis and cytotoxic activity of pentacyclic triterpenoid sulfamates, Arch. Pharm. 348 (2015) 46–54.
- [24] S. Schwarz, S. Sommerwerk, S.D. Lucas, L. Heller, R. Csuk, Sulfamates of methyl triterpenoates are effective and competitive inhibitors of carbonic anhydrase II, Eur. I. Med. Chem. 86 (2014) 95—102.
- [25] H. Kommera, G.N. Kaluderović, S. Dittrich, J. Kalbitz, B. Dräger, T. Müller, R. Paschke, Carbamate derivatives of betulinic acid and betulin with selective cytotoxic activity, Bioorg, Med. Chem. Lett. 20 (2010) 3409—3412.
- [26] D. Chaturvedi, Role of organic carbamates in anticancer drug design, Chem. Pharmacol. Nat. Occurr. Bioactive Comp. (2013) 117–139.
- [27] E.E. Rufino-Palomares, A. Perez-Jimenez, F.J. Reyes-Zurita, L. Garcia-Salguero, K. Mokhtari, A. Herrera-Merchan, P.P. Medina, J. Peragon, J.A. Lupianez, Anticancer and anti-angiogenic properties of various natural tri-terpenoids and some of their chemical derivatives, Curr. Org. Chem. 19 (2015) 919–947.
- [28] G. Periasamy, G. Teketelew, M. Gebrelibanos, B. Sintayehu, M. Gebrehiwot, A. Karim, G. Geremedhin, Betulinic acid and ist derivatives as anti-cancer agent: a review, Arch. Appl. Sci. Res. 6 (2014) 47–58.
- [29] D. Gheorgheosu, O. Duicu, C. Dehelean, C. Soica, D. Muntean, Betulinic acid as a potent and complex antitumor phytochemical: a minireview, Anti-Cancer Agents Med. Chem. 14 (2014) 936–945.
- [30] J.A.R. Salvador, A.S. Leal, D.P.S. Alho, B.M.F. Goncalves, A.S. Valdeira, V.I.S. Mendes, Y. Jing, Highlights of pentacyclic triterpenoids in the cancer settings, Stud. Nat. Prod. Chem. 41 (2014) 33-73.
- [31] S.C. Jonnalagadda, M.A. Corsello, C.E. Sleet, Betulin-betulinic acid natural product based analogs as anti-cancer agents, Anti-Cancer Agents Med. Chem. 13 (2013) 1477—1499.
- [32] M.G. Moghaddam, F.B.H. Ahmad, A. Samzadeh-Kermani, Biological activity of betulinic acid: a review, Pharmacol, Pharm. 3 (2012) 119—123.
- [33] F.B. Mullauer, J.H. Kessler, J.P. Medema, Betulinic acid, a natural compound with potent anticancer effects, Anti-Cancer Drugs 21 (2010) 215–227.
- [34] L. Tripathi, P. Kumar, R. Singh, A review on extraction, synthesis and anticancer activity of betulinic acid, Curr. Bioactive Compd. 5 (2009) 160–168.
- [35] S. Fulda, Betulinic acid: a natural product with anticancer activity, Mol. Nutr. Food Res. 53 (2009) 140–146.
- [36] S. Fulda, Betulinic acid for cancer treatment and prevention, Int. J. Mol. Sci. 9 (2008) 1096–1107.
- [37] R. Csuk, Betulinic acid and its derivatives: a patent review (2008–2013), Expert Opin. Ther. Pat. 24 (2014) 913–923.
- [38] A. Barthel, S. Stark, R. Csuk, Oxidative transformations of betulinol, Tetrahedron 64 (2008) 9225–9229.
- [39] R. Csuk, K. Schmuck, R. Schäfer, A practical synthesis of betulinic acid, Tetrahedron Lett. 47 (2006) 8769–8770.
- [40] C. Stanetty, L. Czollner, I. Koller, P. Shah, R. Gaware, T. Da Cunha, A. Odermatt, U. Jordis, P. Kosma, D. Claßen-Houben, Synthesis of novel 3-amino and 29-hydroxamic acid derivatives of glycyrrhetinic acid as selective 11 beta-hydroxysteroid dehydrogenase 2 inhibitors, Bioorg. Med. Chem. 18 (2010) 7522—7541.
- [41] H.H. Sun, P.V. Kaplita, D.R. Houck, M.B. Stawicki, R. McGarry, R.C. Wahl, A.M. Gillum, R. Cooper, A metalloproteinase inhibitor from Doliocarpus verruculosus, Phytother. Res. 10 (1996) 194–197.
- [42] B. Vasantha, H.P. Hemantha, V.V. Sureshbabu, 1-Propanephosphonic acid cyclic anhydride (T3P) as an efficient promoter for the lossen rearrangement: application to the synthesis of urea and carbamate derivatives, Synthesis (2010) 2990—2996.
- [43] A. Ech-Chahad, A. Minassi, L. Berton, G. Appendino, An expeditious hydroxyamidation of carboxylic acids, Tetrahedron Lett. 46 (2005) 5113–5115.
- [44] H.A. Staab, M. Lueking, F.H. Dürr, The preparation of imidazolides. Synthesis of amides, hydrazides, and hydroxamic acids by the imidazolide method, Chem. Ber. 95 (1962) 1275–1283.
- [45] R.C. Santos, J.A.R. Salvador, S. Marin, M. Cascante, J.N. Moreira, T.C.P. Dinis, Synthesis and structure-activity relationship study of novel cytotoxic carbamate and N-acylheterocyclic bearing derivatives of betulin and betulinic acid, Bioorg. Med. Chem. 18 (2010) 4385–4396.
- [46] R.C. Santos, J.A.R. Salvador, S. Marin, M. Cascante, Novel semisynthetic derivatives of betulin and betulinic acid with cytotoxic activity, Bioorg. Med. Chem. 17 (2009) 6241–6250.

- [47] H.L. Ziegler, H. Franzyk, M. Sairafianpour, M. Tabatabai, M.D. Tehrani, K. Bagherzadeh, H. Hagerstrand, D. Staerk, J.W. Jaroszewski, Erythrocyte membrane modifying agents and the inhibition of Plasmodium falciparum growth: structure—activity relationships for betulinic acid analogues, Bioorg. Med. Chem. 12 (2004) 119—127.
- [48] Shakeel-u-Rehman, S.N. Sofi, M.A. Khuroo, S.C. Taneja, K.A. Bhat, R. Vishwakarma, New natural compounds from Rhododendron lepidotum, Nat. Prod. Res. 27 (2013) 2033–2038.
- [49] L.R. Mikhailova, L.A. Baltina Jr., L.A. Baltina, R.M. Kondratenko, S.A. Nepogodiev, R.A. Field, O. Kunert, M.C. Yin, A simple method of synthesis of triterpene glycosides similar to glycyrrhizic acid and their hepatoprotective activity in vitro, Russ. J. Bioorg. Chem. 35 (2009) 619–627.
- [50] O.B. Flekhter, E.I. Boreko, L.R. Nigmatullina, E.V. Tret'yakova, N.I. Pavlova, L.A. Baltina, S.N. Nikolaeva, O.V. Savinova, V.F. Eremin, F.Z. Galin, G.A. Tolstikov, Synthesis and antiviral activity of betulonic acid amides and conjugates with amino acids, Russ. J. Bioorg. Chem. 30 (2004) 80–88.
- [51] L.A. Baltina, Chemical modification of glycyrrhizic acid as a route to new bioactive compounds for medicine, Curr. Med. Chem. 10 (2003) 155–171.
- [52] L.A. Baltina, O.B. Flekhter, L.R. Nigmatullina, E.I. Boreko, N.I. Pavlova, S.N. Nikolaeva, O.V. Savinova, G.A. Tolstikov, Lupane triterpenes and derivatives with antiviral activity, Bioorg. Med. Chem. Lett. 13 (2003) 3549–3552.
- [53] P. Skehan, R. Storeng, D. Scudiero, A. Monks, J. Mcmahon, D. Vistica, J.T. Warren, H. Bokesch, S. Kenney, M.R. Boyd, New colorimetric cytotoxicity assay for anticancer-drug screening, J. Natl. Cancer Inst. 82 (1990) 1107–1112.
- [54] B. Siewert, J. Wiemann, A. Köwitsch, R. Csuk, The chemical and biological potential of C ring modified triterpenoids, Eur. J. Med. Chem. 72 (2014) 84–101.
- [55] B. Siewert, E. Pianowski, R. Csuk, Esters and amides of maslinic acid trigger apoptosis in human tumor cells and alter their mode of action with respect to the substitution pattern at C-28, Eur. J. Med. Chem. 70 (2013) 259–272.
- [56] B. Siewert, E. Pianowski, A. Obernauer, R. Csuk, Towards cytotoxic and selective derivatives of maslinic acid, Bioorg. Med. Chem. 22 (2014) 594–615.
- [57] M. Urban, J. Sarek, J. Klinot, G. Korinkova, M. Hajduch, Synthesis of A-seco derivatives of betulinic acid with cytotoxic activity, J. Nat. Prod. 67 (2004) 1100—1105
- [58] J. Ito, F.R. Chang, H.K. Wang, Y.K. Park, M. Ikegaki, N. Kilgore, K.H. Lee, Anti-AIDS agents. 48. Anti-HIV activity of moronic acid derivatives and the new melliferone-related triterpenoid isolated from Brazilian propolis, J. Nat. Prod. 64 (2001) 1278—1281.
- [59] T.J. Nitz, C. Montalbetti, R. Mears, X. Gai, E. Glenn, Extended triterpene derivatives as antiretroviral agents, WO2008057420A2, 2008; Chem. Abstr. (2008) 585712.
- [60] C.R. Dorr, S. Yemets, O. Kolomitsyna, P. Krasutsky, L.M. Mansky, Triterpene derivatives that inhibit human immunodeficiency virus type 1 replication, Bioorg. Med. Chem. Lett. 21 (2011) 542–545.
- [61] A.N. Antimonova, N.V. Uzenkova, N.I. Petrenko, M.M. Shakirov, E.E. Shul'ts, G.A. Tolstikov, Synthesis of betulonic acid amides, Chem. Nat. Compd. 44 (2008) 327–333.
- [62] D. Thibeault, C. Gauthier, J. Legault, J. Bouchard, P. Dufour, A. Pichette, Synthesis and structure-activity relationship study of cytotoxic germanicane- and lupane-type 3 beta-O-monodesmosidic saponins starting from betulin, Bioorg. Med. Chem. 15 (2007) 6144–6157.
- [63] S. Boryczka, E. Bebenek, J. Wietrzyk, K. Kempinska, M. Jastrzebska, J. Kusz, M. Nowak, Synthesis, structure and cytotoxic activity of new acetylenic derivatives of betulin, Molecules 18 (2013) 4526–4543.
- [64] C. Horwedel, S.B. Tsogoeva, S.W. Wei, T. Efferth, Cytotoxicity of artesunic acid homo- and heterodimer molecules toward sensitive and multidrug-resistant CCRF-CEM leukemia cells, J. Med. Chem. 53 (2010) 4842–4848.
- [65] L. Pohjala, S. Alakurtti, T. Ahola, J. Yli-Kauhaluoma, P. Tammela, Betulin-derived compounds as inhibitors of alphavirus replication, J. Nat. Prod. 72 (2009) 1917–1926.
- [66] A.V. Korovin, A.V. Tkachev, Synthesis of quinoxalines fused with triterpenes, ursolic acid and betulin derivatives, Russ. Chem. B 50 (2001) 304–310.
- [67] K. Hata, K. Hori, S. Takahashi, Differentiation- and apoptosis-inducing activities by pentacyclic triterpenes on a mouse melanoma cell line, J. Nat. Prod. 65 (2002) 645–648.