

Chapter 5

Molecular Approaches to Explore Natural and Food-Compound Modulators in Cancer Epigenetics and Metabolism

Alberto Del Rio and Fernando B. Da Costa

5.1 Introduction

Let food be thy medicine and medicine be thy food (Hippocrates)

The biological activity of chemical constituents from natural sources and food is crucial in many cellular processes. Several clinical, physiopathological, and epidemiological studies highlight the detrimental or beneficial role of natural/food factors in conjunction with epigenetic and metabolic alterations. Chemical constituents isolated from various sources can interfere with many different biological targets and have been considered as possible starting points for therapeutic purposes. These agents include, for example, curcumin (turmeric), genistein (soybean), polyphenols (green tea, berries, and cocoa), resveratrol (grapes), and sulforaphane (cruciferous vegetables). Moreover, a wide variety of compounds from medicinal plants, spices, bees, or fish can also be mentioned as examples in this category. Among pathways and functions of cells that are notably modulated by these natural constituents, metabolism and epigenetics have emerged in the context of cancer prevention and therapy. Interestingly, epigenetic changes are tightly linked to metabolism, thus adding a higher level of complexity to elucidate the biological role of these compounds. A deeper understanding on how metabolism and epigenetics are influenced by compounds from natural sources and food can be achieved at molecular level by using a variety of chemoinformatic and computer-aided techniques. These include data mining, molecular databasing, and molecular design techniques such as

A. Del Rio (✉)

Institute of Organic Synthesis and Photoreactivity (ISOF), National Research Council (CNR),
Via P. Gobetti 101, 40129 Bologna, Italy
e-mail: alberto.delrio@gmail.com, alberto.delrio@isof.cnr.it

Department of Experimental, Diagnostic and Specialty Medicine (DIMES), Alma Mater
Studiorum, University of Bologna, Via S. Giacomo 14, 40126 Bologna, Italy

F. B. Da Costa

School of Pharmaceutical Sciences of Ribeirão Preto, University of São Paulo,
Avenida do Café s/n, Ribeirão Preto, SP 14040-903, Brazil

pharmacophore-based methods or molecular docking. An overview of these techniques will be described in this chapter in the view of using them as valuable tools to elucidate molecular determinants, mechanism of actions, and polypharmacological role of chemical constituents of food and natural sources.

5.2 Bioactivity of Natural and Food Compounds

The idea that nature is a rich source of bioactive constituents is a 4000-year-old concept. Indians, Egyptians, and Chinese have used natural sources as medicines in early periods of the human civilization. Hippocrates often described diet as a valuable way to treat diseases such as diabetes. Dioscorides, in his five-volume encyclopedia, described the medical uses of herbs, animals, and minerals, and this fantastic work remained alive for more than 15 centuries. Today, lifestyle modifications based on healthy diet, thus on the intake of food and natural compounds, is called lifestyle medicine. The perception that bioactivity of nutraceuticals may have causal relations with the cure or treatment of diseases and, therefore, influence the biological balance of our organism, was spurred starting from the early 1900s. A valuable example of this concept is the treatment of goiter, a disease caused, for over the 90% of cases, by an iodine deficiency, successfully carried out by the administration of iodine-rich foods or potassium iodine. Yet, the beneficial role of natural compounds has been progressively associated to specific food intake. For instance, it has been observed that consuming fish could contribute to keep in good health heart of healthy people as well as positively influence people who are affected by cardiovascular diseases and are exposed to correlated risks. Thanks to the progress in the analytical techniques of food chemicals, fish was identified to be a good source of omega-3 fatty acids (Fig. 5.1a). Indeed, this class of compounds has the capacity to decrease the risks of arrhythmia, triglycerides level, the rate of atherosclerotic plaque, and to lower blood pressure. Consequently, the beneficial effects of fish have been linked to omega-3 fatty acids.

The awareness that natural compounds and food have beneficial or detrimental effects on our life has been also fuelled by the growing epidemiological evidences that have been made possible by the effective exchange of scientific data, the growing availability of specific natural sources, and the effective number of scientists dedicated to the study of phytochemicals, e.g., in the field of pharmacognosy. This kind of research has also assumed in the past decades the “multidisciplinary” dimension involving not only pharmacists, chemists, and pharmacologists but also biochemists, cellular and molecular biologists, toxicologists, and clinicians, among others. Despite the growing information about natural and food components that would suggest their usage as valuable chemicals to prevent and/or treat diseases, contributing to people well-being, there are still several hurdles to clear in this field pervaded by misinformation, not only in the scientific literature but also in the common knowledge. For instance, a common misconception is the assumption that “natural

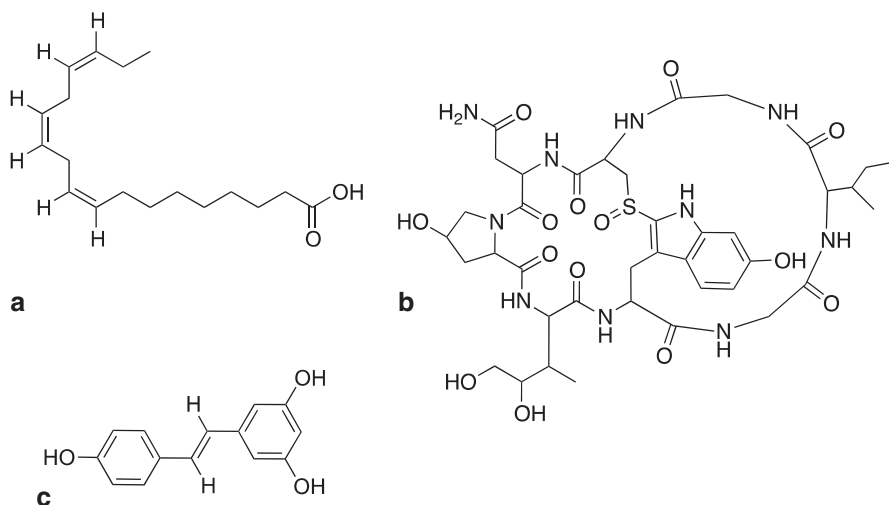


Fig. 5.1 Chemical structures reporting examples of natural compounds with different biological effects. **a** α -Linolenic acid, an essential omega-3 fatty acid. Omega-3 is known to decrease the risk of cardiovascular diseases; **b** α -amantine is a deadly natural compound found in the *Amanita phalloides* mushrooms; **c** resveratrol, a polyphenolic stilbenoid produced in plants with several reported pharmacological actions

is always good,” which is an easily falsifiable statement. Indeed, a large number of phytochemicals are known to be harmful for health and, in several cases, also lethal. For example, α -amantine (Fig. 5.1b) is a natural cyclic octapeptide contained in the *Amanita phalloides* fungus, which is widely distributed across Europe and resembles several species of edible mushrooms. α -Amantine is an example of highly poisonous and deadly natural compound which was proved to bind to the bridge helix in RNA polymerase II, interfering with the translocation of RNA and DNA, leading to a drastically reduced rate of synthesis of the RNA molecule [1]. There are numerous classical examples of natural constituents from plants or food which are dangerous to health, such as strychnine from *Strychnos* species, cyanogenic glucosides from cassava (*Manihot* species) or myristicin from nutmeg (*Myristica fragrans*).

To clarify the role of natural and dietary compounds, an elucidation of the interaction mechanisms of these molecules with the human biological network is required, especially at a molecular level. This includes the uncover of the biophysical mechanisms by which these compounds bind to receptors or enzymes (i.e., allosteric regulation and inhibition/activation profile) and their kinetics (i.e., reversible/irreversible, substrate and cofactors competition/non-competition) that could underlie to specific pharmacological actions. These studies are far to be accomplished because, in many cases, it is experimentally difficult to isolate large amounts of compounds from the natural source and, even when this is possible, it is complicated to dissect their intrinsically polypharmacological roles, rendering this area of research

extremely challenging. An exemplifying case of polypharmacology is resveratrol (Fig. 5.1c), a polyphenolic stilbenoid produced in plants and found in wine which possesses several reported pharmacological actions, including anti-inflammatory, anticarcinogenic, antimutagenic, antiaging, antioxidant, and anticoagulant. Many examples reporting bioactivity of resveratrol in different molecular pathways can be found in the literature [2–6].

5.2.1 *Pharmaceutical Development of Natural Compounds*

It was only after the advent of advanced technologies for isolation, purification, and structure elucidation of organic compounds that scientists could realize how natural sources were able to deliver an important amount of diverse chemical entities. Nowadays, it is well known that the natural product landscape constitute a very varied supply of building blocks and intermediates useful for the drug discovery process, which, in many cases, represent the starting point for generating lead compounds. The latter can be further synthetically modified in order to create and develop specific therapeutically relevant pharmaceuticals [7]. The impressive chemical diversity along with the structural complexity of natural compounds represents a source of inspiration for the generation of chemical libraries belonging, in most cases, to an unexplored and “intellectual property free” chemical space, allowing pharmaceutical companies to protect composition of matter together with medical uses [8]. In this sense, we assist to a conceptual shift, passing from the classical era of combinatorial chemistry, during which pharmaceutical companies essentially disregarded the development of natural products as potential drug candidates, to the development of targeted or focused compound libraries inspired by natural sources [9]. The accumulating evidence that the natural selection process represents a unique way to diversify the chemistry of natural compounds and the way in which the latter evolved in biological organisms has favored this process. For these reasons, the interactions of natural compounds with other biological macromolecules reflect, in different cases, high specificity and potency profiles. Since natural products can be considered the richest source of novel chemical scaffolds for biological studies, technologies and strategies to extract them from different sources have evolved rapidly in the past years [10]. A number of advanced separation and structure elucidation techniques are now available for chemists/pharmacists that can now have access to an increasing number of purified natural compounds [11]. Among the separation procedures, high-performance liquid chromatography (HPLC) is the technique of choice because it allows isolation of compounds from the analytical to the preparative scale level. In addition, HPLC can also be coupled to ultraviolet (UV), mass spectrometry (MS), or nuclear magnetic resonance (NMR), comprising the so-called hyphenated or tandem techniques (LC-MS or LC-NMR), which greatly increase the efficiency of compound identification [11].

However, despite the advance in purification techniques, natural products resources are still largely unexplored, mostly due to the technical obstacles to collect

samples, especially from the most concealed places on earth, e.g., deep sea level, arid or extremely cold regions. Historically, the most widely used natural compounds have been isolated from plants and animals by means of classical chromatographic techniques such as column or thin-layer chromatography. Subsequently, cultured soil microorganisms, or the direct access to the genome of soil organisms clonable into culturable organisms, provided a rich source of natural products [12]. In the last decade, compounds recovered from the marine environment have come into focus: Indeed, oceans harbor one of the widest variety of ecosystems on earth, a fact reasonably reflected by an unprecedented discovery of new chemical entities of marine origin.

Food compounds, most of them plant secondary metabolites, can be seen as a particular class of natural compounds since they have to be considered as materials designated as “generally recognized as safe” (GRAS) [13]. There is currently a great deal of interest in exploring benefits of bioactive food components and relate them to health and wellness. However, despite the efforts made by researchers to identify food-compounds, few studies report the systematic extraction and purification of a specific bioactive component from different food sources, with the notable exceptions of fruits, vegetables, beverages, and essential oils [14, 15].

5.2.2 *Anticancer Compounds from Natural and Food Sources*

Natural and dietary compounds present molecular scaffolds that are particularly attracting as sources of lead compounds for cancer therapy. Indeed, more than 60% of the anticancer drugs have natural origin or are the result of chemical optimizations of natural scaffolds. Accordingly, it is not surprising that the interest in natural products have gained momentum in the past years, as their application as lead compounds is source of novel chemical entities (NCEs) in different areas of anticancer drug design [16–18]. With their unique chemical diversity, the usage of natural compounds in cancer therapies is even more justified if considered the wide range of variability in terms of biochemical and biological pathways that are present in cancer pathologies. The result of the drift toward natural compounds and their derivatives is reflected by the wide range of chemical compounds from very different sources already associated to bioactivities of oncogenic targets.

Historically, this discovery resulted mainly in the development of anticancer agents from plants (e.g., vinca alkaloids like vincristine and vinblastine; *Podophyllum* lignans like podophyllotoxin; taxanes like paclitaxel and docetaxel; and quinoline alkaloid like camptothecin, topotecan, and irinotecan), marine organisms (i.e., toxins like latrunculins; didemnins like aplidine and trabectedin; and stronglylophorines) and microorganisms (e.g., anthracyclines like doxorubicin, daunorubicin, mitoxantrone and idarubicin; chromomycins like dactinomycin and plicamycin; and miscellaneous antibiotics like mitomycin and bleomycin). More recently, different types of terpenoids have been demonstrated to inhibit the NF- κ B signaling, to suppress inflammation processes and to reduce cancer progression

[19] while α -methylene- γ -lactones, in particular sesquiterpene lactones (especially found in Asteraceae species), have proven to be promising candidates for treatment of various types of cancer [20–22]. Salinosporamides, a class of marine natural compounds, present in *Salinispora tropica* bacterium, were identified to be potent inhibitors of proteasome [23].

Among natural sources, several food-component agents have also been identified as beneficial for anticancer therapy. Dietary sources including fruits, vegetables, and spices have drawn a great deal of attention from the scientific community due to their demonstrated ability to interfere with cancer mechanisms; nevertheless, speculations by the general public has fomented the idea that fabricated supplements can be a panacea [24]. Scientific literature provided evidence that the regular consumption of fruits, vegetables and spices lowers the incidence of cancers (i.e., stomach, esophagus, lung, oral, endometrium, pancreas, and colon) [25]. These agents include curcumin (turmeric), resveratrol (red grapes, peanuts and berries), genistein (soybean), diallylsulfide (allium), *S*-allyl cysteine (allium), allicin (garlic), lycopene (tomato), capsaicin (red chili), diosgenin (fenugreek), 6-gingerol (ginger), ellagic acid (pomegranate), ursolic acid (apple, pears, prunes), silymarin (milk thistle), anethol (anise, camphor, and fennel), catechins (green and white tea, berries and cocoa), eugenol (cloves), indole-3-carbinol (cruciferous vegetables), limonene (citrus fruits), beta-carotene (carrots), and several dietary fibers.

Many other examples of natural and dietary compounds that have a role in cancer-related diseases underline the importance of this topic in oncological research. In the following paragraphs, we provide an overview of these compounds that specifically modulate cell pathways and functions connected to epigenetic and metabolic changes in cancer diseases.

5.3 Epigenetic and Metabolic Pathophysiology of Cancer

Cancer is a complex set of diseases. Genetic aberrations, epigenetic alterations, and inflammations constitute some of the known mechanisms by which normal cells develop and progress towards neoplastic pathologies. While last decades marked a major understating in cancer genetics, it is now evident that the dissection of the mechanisms of this multifaceted set of diseases requires a deeper look in other paradigms of cancer biology in order to conceive new prevention or therapeutic approaches. This larger framework has evolved in the recent years on novel lines of research, for instance, toward the understanding of the immune system regulation [26, 27] and the epigenetic modifications, but also on the reinterpretation of old studies by means of new scientific awareness that marked a return to cancer metabolism [28–31]. In the next paragraphs, we will discuss cancer metabolism and epigenetics, focusing on the possibilities to interfere with the mechanism of pathogenesis and progression of cancer diseases by means of small molecules of natural and food origin.

5.3.1 *Natural Compounds Modulating Epigenetic and Metabolic Mechanisms*

Epigenetics is a general term that refers to modifications of genes expression through alteration of chromatin structure and/or DNA methylation occurring without changes in the DNA sequence, from which the term *epi*-(from greek: over, outside of, around)*genetics*. Global modifications of chromatin packaging and its influence in the transcription of associated genes fuelled the research on cancer epigenetics in the past years. The ensemble of known epigenetic mechanisms can be categorized into three classes: i) histone posttranslational modifications (PTMs) that represent one of the major way to arrange the different states of chromatin; ii) DNA methylation, i.e., the methylation of DNA cytosines to 5-methylcytosines; and iii) regulation of gene expression by non-coding RNA (ncRNA). The elucidation of epigenetic phenomena, representing nowadays an important topic of research, is necessary to understand the basis of several biological processes and is progressively translating into the development of new therapeutic *epi-compounds* or *epi-drugs* [32–34]. Different studies have highlighted how alterations in the epigenetic code contribute to the onset and growth of a variety of cancers [35–48]. Consequently, epigenetic modifications are constituting attractive therapeutic targets for the development of new cancer therapies [33, 49–52]. An increasing number of reports describe, in particular, new types of histone post-translational modifications (PTMs) associated with the characterization of the enzymes that are in charge of operating these chemical reactions [53]. Yet, other studies point on the validation of these PTMs in the context of chromatin remodeling and regulation, as well as their clinicopathological relevance in human diseases [54]. It is important to point out that the increasing evidences linking epigenetic targets and cancer pathologies have been boosted by the surge of structural data describing these proteins, thus creating the basis to develop specific probe compounds and start new drug discovery campaigns [54, 55]. However, although the ensemble of these data promises to shed light on cancer epigenetics, the way in which epigenetic modifications relate to cancer and, consequently, their therapeutic relevance in cancer diseases, is still largely unknown. Most of these targets, despite being linked to cancer pathologies, may not have causal role in specific malignant transformations. Some notable exceptions [56, 57] are the recent success stories documenting the potential to interfere with these mechanisms by means of small organic molecules [34]. In particular, the first clinical results have been obtained with histone deacetylases (class I, II and IV HDACs) inhibitors [58], DNA methyltransferases inhibitors (DNMTi) [59] and histone methyltransferases inhibitors [60]. Other classes of epigenetic enzymes are rapidly reaching the potential to become pharmaceutically validated biological targets. Among them are sirtuins, which are NAD⁺-dependent histone deacetylases also known as class III HDACs [6], and histone demethylases [61]. Apart from histones PTMs and DNA methylation, growing evidences indicate that modulating microRNAs expression might be useful to interfere with epigenetic mechanisms and develop novel RNA-based drugs for a wide range of diseases [62–65]. Indeed,

the deregulation of microRNAs expression and activity is frequently observed in a variety of human pathologies including cancer [66]. Therefore, in addition to the general strategy of increasing or decreasing miRNA abundance and activity by using oligonucleotides or plasmid- and virus-based constructs, a novel paradigm aims to target miRNA expression by means of specific compounds targeting miRNA transcription and processing. Clearly, the potential success of small molecules can be ascribed to their capacity to circumvent the issue of delivery into most tissues making them very attractive as a therapeutic tool.

Metabolic changes have been rediscovered in the context of cancer diseases after the initial observations of Otto Warburg in the early 1920s [30, 31, 67]. Warburg noticed that proliferating cancer cells consume glucose at a high rate, releasing lactate and not carbon dioxide. Indeed, one of the primary metabolic changes in cancer transformation is constituted by an increased catabolic glucose metabolism characterized by high rates of anaerobic glycolysis, regardless of oxygen concentration. While the underlying mechanisms that alter metabolic programs of cancer cells are still to be fully elucidated, it is known that several genetic alterations in cell pathways responsible for the regulation of cells metabolism contribute to cancer growth and progression. For instance, the conversion of glucose to glucose-6-phosphate (G6P) is critical to different cancer phenotypes, a process catalyzed by the enzyme hexokinase-II. Thus, intermediates of glycolysis like G6P can therefore accumulate, creating a highly advantageous environment for cancer survival and growth. On these bases, the pharmacological modulation of specific metabolic enzymes is currently under investigation by various research groups as a viable strategy to block cancer cell proliferation [68–72].

Several natural and dietary components have been already identified as capable to interfere with different epigenetic and metabolic mechanisms [29, 73, 74] (Fig. 5.2). Dietary components like phenolics from green tea, genistein from soybean, isothiocyanates from plant foods (e.g., from Brassicaceae species), diallylsulfide from garlic, curcumin from turmeric, resveratrol from grapes, and sulforaphane from cruciferous vegetables have been studied for their ability to target the epigenome, in relation, for instance, to breast cancer [73, 75–79]. While in most of the cases the mechanisms of action of natural compounds are still poorly understood, some of them have been identified. For instance, luteolin (Fig. 5.2), a common flavonoid found in parsley and celery has been demonstrated to inhibit DNMTs and sirtuins (SIRT), while retinoic acid, found in carrots, spinach and eggs, and used nowadays to treat leukemias, is an HDAC inhibitor. Among polyphenols, epigallocatechin-3-gallate, the major compound found in green tea, was reported to have a complex polypharmacology, as inhibitor of histone acetyltransferases (HATs), HDACs, SIRTs, DNMTs, retinoic acid receptor (RAR β), proteasome, 78 kDa glucose regulated protein (Grp78) and heat shock protein 90 (Hsp90). Similarly, curcumin (Fig. 5.2) and curcuminoids have also been widely studied for their anti-inflammatory, antiangiogenic, antioxidant, wound healing, and anticancer effects. Importantly, curcumin analogs, like dihydrocoumarins, have been demonstrated to inhibit sirtuins. Since the isoform SIRT1 has been shown to have a role in deacetylating p53, a master regulator of metabolic function in the cell, the inhibition of enzymes like SIRT1 likely contributes to the

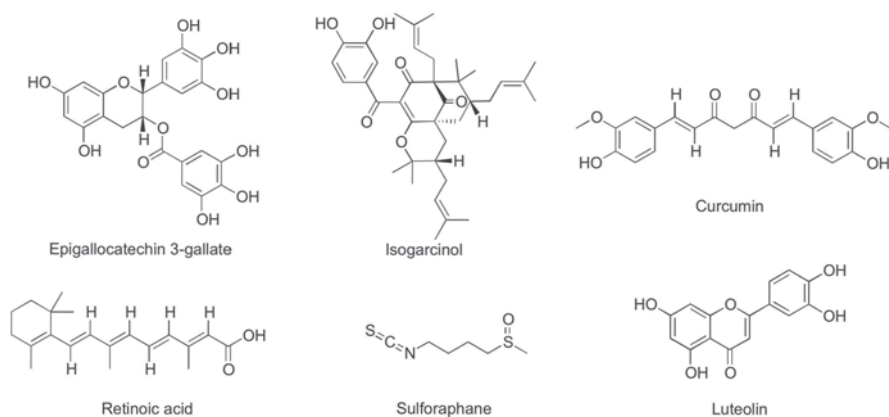


Fig. 5.2 Examples of the chemical diversity of natural compounds with a role in epigenetics and metabolic pathways

regulation of both epigenetic mechanisms and metabolic pathways like glycolysis. Other classes of natural compounds, such as anacardic acid and related compounds from cashew nut, alkaloids such as sanguinarine, quinone derivatives, peptides and peptide conjugates, and polyisoprenylated benzophenone derivatives (PBDs), have been demonstrated to have activities against HATs [80]. As previously pointed out, the discovery of natural scaffolds is allowing the development of focused libraries of compounds that are able to act on epigenetic enzymes with more potent and specific profiles. An example of this strategy is given by Kundu and co-workers, who could generate garcinol derivatives starting from isogarcinol (Fig. 5.2), in order to develop inhibitors for p300 and PCAF HATs [81]. Because of the tight connection with epigenetic and metabolic changes, it is known that specific cancer conditions are strongly influenced by lifestyle and environmental factors, including the intake of food and nutrients [82]. For instance, the absorption of compounds like flavonoids and folates through diet has been shown to alter DNA methylation and modify the risk of human colon cancer and cardiovascular diseases, even though their mechanisms of action have to be ascertained, yet [83–85]. Additional researches on the effects that nutraceuticals have on epigenetic and metabolic changes promise to be relevant for devising new preventive and therapeutic interventions.

5.3.2 Linking Metabolism and Epigenetic Mechanism

Growing evidences show how epigenetic changes are linked to cancer metabolism in different cancer pathologies [29]. It is meaningful to stress on how many enzymes, substrates, and co-factors are common in metabolic and epigenetic pathways/targets, as shown in Fig. 5.3. For example, sirtuins deacetylate histone proteins and have also a primary role in metabolic regulation which is dependent on the pool of intracellular NAD^+ , whose biosynthesis and signaling became an emerging area in

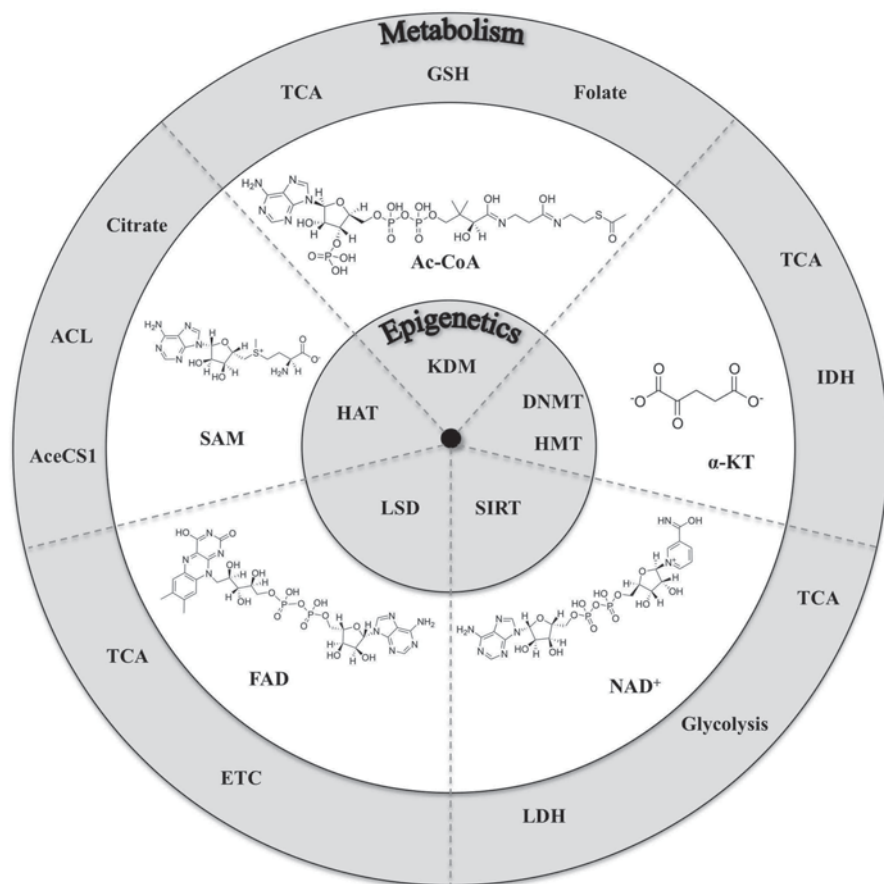


Fig. 5.3 Examples of connections between epigenetics and metabolic pathways. (Abbreviations: α -KT α -ketoglutarate; *AcCoA* acetyl coenzyme A; *AcCS1* acetyl-CoA synthase 1; *ACL* ATP-citrate lyase; *ETC* electron-transport chain; *FAD* flavin adenine dinucleotide; *GSH* glutathione; *IDH* isocitrate dehydrogenase; *LDH* lactate dehydrogenase; *NAD⁺* nicotinamide adenine dinucleotide; *SAM* S-adenosyl methionine; *TCA* tricarboxylic acid cycle)

medicinal chemistry [86]. Many cancer cells rely on glycolysis to satisfy their energy requirements, a process that leads to the production of lactate and not of acetyl-CoA (AcCoA), like for healthy cells. Since AcCoA is also a substrate of epigenetic enzymes, such as histone acetyltransferases (HATs), depletion of the AcCoA in cancer cells might contribute to epigenetic alterations. A similar consideration can be drawn for other metabolic co-substrate and co-factors like S-adenosylmethionine (SAM), flavin adenine dinucleotide (FAD), and α -ketoglutarate (Fig. 5.3), which are all involved in the epigenetic regulation through various enzymatic mechanisms [87]. Moreover, compounds of natural and food origin can be converted by cell

metabolites into chemical intermediates implicated in epigenetic and metabolic alterations [25, 29, 44, 75, 82, 88]. So, it is evident that a molecular-level knowledge of the connections between metabolism and epigenetic mechanisms is required in order to define the polypharmacological role of small molecules. It should be noted that the biological effect of many chemical scaffolds, especially of natural origin, is in most cases ascribable to a promiscuous activity towards biological targets that uses common substrates and cofactors like NAD⁺/NADH, FAD, SAM, AcCoA, α -ketoglutarate, and ATP. Therefore, in the framework of developing compound libraries from natural and food origin, it is essential to assess compounds against their impact on epigenome and metabolism by looking at their polypharmacological behavior. In particular, the screening of biological activities acquires importance if considered that the detrimental or beneficial effects of natural compounds for the treatment of a specific disease, is dependent on the physiopathological context [89].

5.4 Computer-Aided Molecular Design Approaches

Computer-aided molecular techniques are heavily used in academia and industrial settings to assist the selection of new compounds with predefined biological activity. Several examples testify their successful applications in the development of new chemical entities [90–92] and a wide range of disciplines nowadays revolve around computer-aided drug discovery (CADD), including chemoinformatics, computational chemistry, structural biology, biophysics, medicinal chemistry, organic chemistry, and pharmacology. Among the various computational techniques available, virtual screening is certainly the most popular to screen rapidly and cost-effectively new chemicals from large libraries of compounds [93–95]. In principle, this technique can be divided in ligand- and structure-based drug design techniques (LBDD and SBDD): the first category usually takes advantage of information from known bioactive compounds (ligand), while the second usually exploits three-dimensional structure of the biological target (protein) in order to identify putative modulators of the protein activity. In the past years, the growing availability of protein structures, resolved by structural biologists, progressively raised the possibility to deploy SBDD. Nevertheless, ligand-based techniques are still essential tools, for example when structural information of a biological target is missing or when the molecular design is not directed towards a target-centric approach, but point to modulate cellular pathways or phenotypic traits without a precise knowledge of the mechanism of action. In addition, it should be noted that, despite the apparent advantage and the success of the target-centric approach, which consist in the design of small molecules having high-selectivity profiles against a specific target, it has failed in many other cases [96].

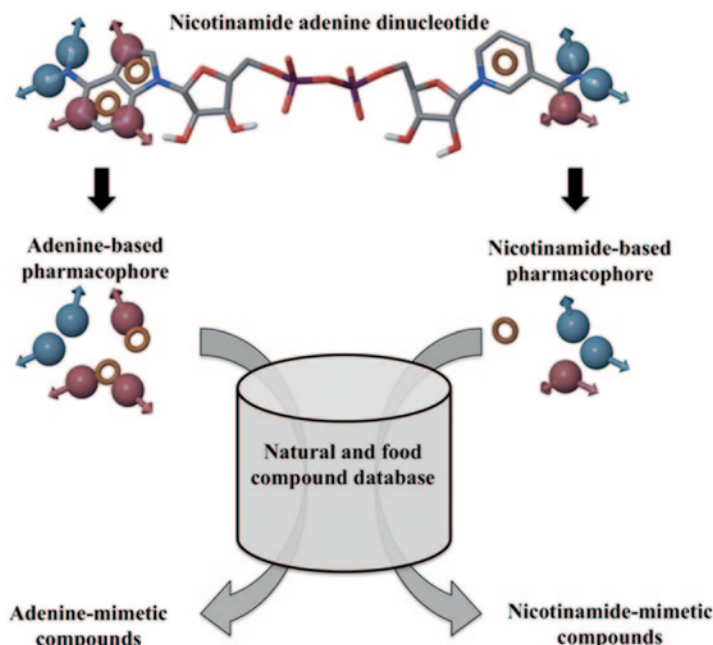


Fig. 5.4 Example of pharmacophore-based *in silico* approach to discover adenine and nicotinamide mimetic compound of NAD⁺ originating from natural or food sources

5.4.1 CADD Approaches on Epigenetic and Metabolic Targets

As seen in previous sections, the research aiming at developing new therapeutic anticancer strategies against epigenetic and metabolic targets has flourished in the past years. Several reports describe rationales, targets, new drugs, approaches, novel compounds, and methodologies [34, 35, 37, 41–48, 52, 53, 56, 97–107]. Computational techniques are being actively used in this field and several reviews and articles have been published recently on this topic [56, 57, 59, 105, 108, 109]. A valuable example is the extensive usage of computer-aided techniques for epigenetic enzymes like sirtuins [6, 108, 110–114]. A variety of computational tools like molecular docking and pharmacophore mapping have been used to identify novel modulating compounds while trying to explain the mechanism of actions of these small molecules. Equally, theoretical tools have also been applied to identify and elucidate pharmacological mechanisms of metabolic enzymes like lactate dehydrogenase and hexokinase-II [69, 114, 115]. Of note, many of these targets use NAD⁺ as a cofactor and several computational strategies were directed to find competitive compounds of either the adenine or the nicotinamide pocket, or both. As an example, Fig. 5.4 depicts a typical *in silico* screening workflow that uses pharmacophore techniques to identify NAD⁺ competitive inhibitors with natural or dietary-derived scaffolds mimicking the adenine or the nicotinamide moieties. In fact, three-dimen-

sional pharmacophore modeling techniques revealed to be useful for virtual screening and computational purposes to analyze diverse compound databases in order to define pharmaceutical values of new compounds [116, 117]. Interestingly, the use of less-sophisticated techniques based on topological-structural descriptors and subsequent statistical treatment, i.e., discriminant analysis, have also been demonstrated as very efficient methodologies for the selection of new natural compounds. Even in this case, the validated model could be readily applied for searching new chemicals of natural origin in large databases [118, 119]. It is expected that the usage and combination of various *in silico* approaches and the availability of compound databases of natural and dietary sources (see below) could constitute an effective step toward the identification, development, and pharmacological definition of natural and dietary-derived components in metabolic and epigenetic mechanism of cancer.

5.4.2 Chemical Space of Natural and Food Compounds

Since natural products and dietary components are known to represent a vast chemical diversity with underlying scaffold complexities and architectures, exploring the chemical space of these compounds it currently a major field of research for different groups [13, 120–125]. Most of the natural products are assorted by chemical groups reflecting novel molecular properties/features as compared to synthetic compounds and available drugs. Several chemoinformatic analyzes, in fact, highlight this behavior and, at the same time, recognize the adherence to drug- and lead-like rules purporting the idea that several classes can be considered as pharmaceutically relevant entities [13, 124]. In addition, despite this diversity, natural products insure the presence of privileged scaffolds that could offer the advantage to address the coverage of poorly explored chemical space [121, 126]. As previously indicated, this feature is particularly appealing for industrial settings to insure the appropriate intellectual property protection requested for the pharmaceutical development. In this direction, it should be noted also that natural products are providing line principles for novel library design in combinatorial chemistry and targeted compound libraries inspired by nature [126–128].

From the chemical point of view, the analysis of natural products databases available in the public domain shows a low-molecular overlap of compounds and highlight as the most representative molecular fragment benzene, acyclic compounds, flavones, coumarins, and flavanones [121, 122, 125]. A particular class of natural compounds that can be considered as dietary component are flavoring substances like menthol, camphor, and anethol, that are discrete chemical entities that usually are considered “generally recognized as safe” (GRAS) compounds. Interestingly, the comparison of collections of compounds including GRAS, natural products, approved drugs, and dataset from commercial molecules by means of chemoinformatic analysis demonstrated that GRAS products are an important source of bioactive compounds that possess all the characteristics for drug discovery and nutraceutical purposes [13].

Among computational approaches that can help driving the discovery of new bioactive compounds, a prominent workflow is the screening of large database of readily available molecules. It is with surprise that the scientific community has not developed yet a freely available and fully chemically annotated database of food components [8, 9]. Despite this lack, some examples are starting to appear in the literature and on the Internet. Among them, we can list the INFOODS of FAO [129], the USDA national nutrient database [130] and the FoodDB that has been recently released [131]. In the direction of the creation of a comprehensive and freely available collection of food chemicals, it should be noted also the necessity to include the possible procurement from commercial sources of purified samples of food components that, ideally, should complement the major efforts that have been done in the past years for other natural sources [123].

5.5 Conclusions

Many anticancer drugs have natural origin or are the result of chemical optimizations of natural scaffolds. Because the natural product landscape constitutes a varied supply of building blocks and intermediates, they can represent the starting point for generating lead compounds with bioactive relevance. A thriving topic in cancer research deals with metabolism and epigenetics mechanisms that lead to malignant transformation and the way to interfere pharmacologically with the pathogenesis and progression of cancer diseases by means of small molecules. Natural and food-derived compounds able to modulate epigenetic and metabolic mechanisms are of great interest because they promise to provide new therapeutic interventions, as they are capable to exert anti-inflammatory, antiangiogenic and antioxidant effects that could also be beneficial for anticancer purposes. In this framework, it is expected that advances in computational approaches, with emphasis on pharmacophore and docking-based techniques, together with the systematic cataloguing of natural and dietary-related components, would greatly help to track molecular mechanisms involved in nutriepigenomics and nutrimetabolomics, and therefore constitute a launching platform for new drug-discovery pipelines.

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References

1. Bushnell DA, Cramer P, Kornberg RD (2002) Structural basis of transcription: alpha-amanitin-RNA polymerase II cocystal at 2.8 Å resolution. *P Natl Acad Sci U S A* 99:1218–1222
2. Beher D, Wu J, Cumine S, Kim KW, Lu S-C, Atangan L, Wang M (2009) Resveratrol is not a direct activator of SIRT1 enzyme activity. *Chem Biol Drug Des* 74:619–624

3. Denu JM (2012) Fortifying the link between SIRT1, resveratrol, and mitochondrial function. *Cell Metab* 15:566–567
4. Price NL, Gomes AP, Ling AJY et al (2012) SIRT1 is required for AMPK activation and the beneficial effects of resveratrol on mitochondrial function. *Cell Metab* 15:675–690
5. Moniot S, Weyand M, Steegborn C (2012) Structures, substrates, and regulators of mammalian sirtuins—opportunities and challenges for drug development. *Front Pharmacol* 3:16
6. Bruzzone S, Parenti MD, Grozio A, Ballestrero A, Bauer I, Del Rio A, Nencioni A (2013) Rejuvenating sirtuins: the rise of a new family of cancer drug targets. *Curr Pharm Des* 19:614–623
7. Newman DJ, Cragg GM (2012) Natural products as sources of new drugs over the 30 years from 1981 to 2010. *J Nat Prod* 75:311–335
8. Hong J (2011) Role of natural product diversity in chemical biology. *Curr Opin Chem Biol* 15:350–354
9. Sheridan C (2012) Recasting natural product research. *Nat Biotechnol* 30:385–387
10. Paterson I, Anderson EA (2005) Chemistry. The renaissance of natural products as drug candidates. *Science* 310:451–453
11. Pauli GF, Chen S-N, Friesen JB, McAlpine JB, Jaki BU (2012) Analysis and purification of bioactive natural products: the AnaPurNa study. *J Nat Prod* 75:1243–1255
12. Handelsman J, Rondon MR, Brady SF, Clardy J, Goodman RM (1998) Molecular biological access to the chemistry of unknown soil microbes: a new frontier for natural products. *Chem Biol* 5:R245–R249
13. Medina-Franco JL, Martínez-Mayorga K, Peppard TL, Del Rio A (2012) Chemoinformatic analysis of GRAS (Generally Recognized as Safe) flavor chemicals and natural products. *PLoS One* 7:e50798
14. Barbosa-Pereira L, Pocheville A, Angulo I, Paseiro-Losada P, Cruz JM (2013) Fractionation and purification of bioactive compounds obtained from a brewery waste stream. *Biomed Res Int* 2013(2013):408491
15. Angela A, Meireles M (eds) (2008) Extracting bioactive compounds for food products. CRC, Boca Raton. doi:10.1201/9781420062397
16. Kinghorn AD, Chin Y-W, Swanson SM (2009) Discovery of natural product anticancer agents from biodiverse organisms. *Curr Opin Drug Discov Dev* 12:189–196
17. Newman DJ (2008) Natural products as leads to potential drugs: an old process or the new hope for drug discovery? *J Med Chem* 51:2589–2599
18. Gordaliza M (2008) Natural products as leads to anticancer drugs. *Clin Transl Oncol* 9:767–776
19. Salminen A, Lehtonen M, Suuronen T, Kaarniranta K, Huuskonen J (2008) Terpenoids: natural inhibitors of NF-kappaB signaling with anti-inflammatory and anticancer potential. *Cell Mol Life Sci* 65:2979–2999
20. Merfort I (2011) Perspectives on sesquiterpene lactones in inflammation and cancer. *Curr Drug Targets* 12:1560–1573
21. Ghantous A, Gali-Muhtasib H, Vuorela H, Saliba NA, Darwiche N (2010) What made sesquiterpene lactones reach cancer clinical trials? *Drug Discov Today* 15:668–678
22. Janecka A, Wyrębska A, Gach K, Fichna J, Janecki T (2012) Natural and synthetic α -methylene lactones and α -methylene lactams with anticancer potential. *Drug Discov Today* 17:561–572
23. Gulder TAM, Moore BS (2010) Salinosporamide natural products: Potent 20 S proteasome inhibitors as promising cancer chemotherapeutics. *Angew Chem Int Edit* 49:9346–9367
24. Szic KS, Palagani A, Hassannia B (2011) Phytochemicals and cancer chemoprevention: epigenetic friends or foe? In: Rasooli I (ed) *Phytochemicals—Bioactivities and Impact on Health*. InTech, Croatia, pp 159–198. doi:10.5772/28499
25. Szic KS, Ndlovu MN, Haegeman G, Vanden Berghe W (2010) Nature or nurture: let food be your epigenetic medicine in chronic inflammatory disorders. *Biochem Pharmacol* 80:1816–1832
26. Galluzzi L, Senovilla L, Zitvogel L, Kroemer G (2012) The secret ally: immunostimulation by anticancer drugs. *Nat Rev Drug Discov* 11:215–233

27. Cavallo F, De Giovanni C, Nanni P, Forni G, Lollini P-L (2011) 2011: the immune hallmarks of cancer. *Cell* 60:319–326
28. Cairns RA, Harris IS, Mak TW (2011) Regulation of cancer cell metabolism. *Nat Rev Cancer* 11:85–95
29. Gerhäuser C (2012) Cancer cell metabolism, epigenetics and the potential influence of dietary components—a perspective. *Biomed Res* 23:1–21
30. Semenza GL (2011) A return to cancer metabolism. *J Mol Med* 89:203–204
31. Koppenol WH, Bounds PL, Dang CV (2011) Otto Warburg's contributions to current concepts of cancer metabolism. *Nat Rev Cancer* 11:325–337
32. Best JD, Carey N (2010) Epigenetic therapies for non-oncology indications. *Drug Discov Today* 15:1008–1014
33. Best JD, Carey N (2010) Epigenetic opportunities and challenges in cancer. *Drug Discov Today* 15:65–70
34. Dhanak D (2012) Cracking the code: the promise of epigenetics. *ACS Med Chem Lett* 3(7):521–523. doi:10.1021/ml300141h
35. Ellis L, Atadja PW, Johnstone RW (2009) Epigenetics in cancer: targeting chromatin modifications. *Mol Cancer Ther* 8:1409–1420
36. Altucci L, Minucci S (2009) Epigenetic therapies in haematological malignancies: searching for true targets. *Eur J Cancer* 45:1137–1145
37. Chi P, Allis CD, Wang GG (2010) Covalent histone modifications—miswritten, misinterpreted and mis-erased in human cancers. *Nat Rev Cancer* 10:457–469
38. Herranz M, Esteller M (2006) New therapeutic targets in cancer: the epigenetic connection. *Clin Transl Oncol* 8:242–249
39. Graham JS, Kaye SB, Brown R (2009) The promises and pitfalls of epigenetic therapies in solid tumours. *Eur J Cancer* 45:1129–1136
40. Santos-Rosa H, Caldas C (2005) Chromatin modifier enzymes, the histone code and cancer. *Eur J Cancer* 41:2381–2402
41. Rodríguez-Paredes M, Esteller M (2011) Cancer epigenetics reaches mainstream oncology. *Nat Med* 17:330–339
42. Rius M, Lyko F (2012) Epigenetic cancer therapy: rationales, targets and drugs. *Oncogene* 31:4257–4265
43. Kulis M, Esteller M (2010) DNA methylation and cancer. *Adv Genet* 70:27–56
44. Meeran SM, Ahmed A, Tollefsbol TO (2010) Epigenetic targets of bioactive dietary components for cancer prevention and therapy. *Clin Epigenetics* 1:101–116
45. Ljungman M (2009) Targeting the DNA damage response in cancer. *Chem Rev* 109:2929–2950
46. Claes B, Buyschaert I, Lambrechts D (2010) Pharmaco-epigenomics: discovering therapeutic approaches and biomarkers for cancer therapy. *Heredity* 105:152–160
47. Pollock RM, Richon VM (2009) Epigenetic approaches to cancer therapy. *Drug Discov Today Ther Strategy* 6:71–79
48. Spannhoff A, Sippl W, Jung M (2009) Cancer treatment of the future: inhibitors of histone methyltransferases. *Int J Biochem Cell Biol* 41:4–11
49. Sala A, Corona DFV (2008) Epigenetics: More than genetics. *Fly* 2:165–168
50. Baylin SB (2008) Epigenetics and cancer. *Mol Basis Cancer*. 2:57–65
51. Lohrum M, Stunnenberg HG, Logie C (2007) The new frontier in cancer research: deciphering cancer epigenetics. *Int J Biochem Cell Biol* 39:1450–1461
52. Inche AG, La Thangue NB (2006) Chromatin control and cancer-drug discovery: realizing the promise. *Drug Discov Today* 11:97–109
53. Bannister AJ, Kouzarides T (2011) Regulation of chromatin by histone modifications. *Cell Res* 21:381–395
54. Andreoli F, Barbosa AJM, Parenti MD, Del Rio A (2013) Modulation of epigenetic targets for anticancer therapy: clinicopathological relevance, structural data and drug discovery perspectives. *Curr Pharm Des* 19:578–613
55. Hou H, Yu H (2010) Structural insights into histone lysine demethylation. *Curr Opin Struct Biol* 20:739–748

56. Sippl W, Jung M (2009) Epigenetic drug discovery special issue. *Bioorg Med Chem* 19:3603–3604
57. Sippl W, Jung M (2009) Epigenetic targets in drug discovery. Wiley, Weinheim
58. Lombardi PM, Cole KE, Dowling DP, Christianson DW (2011) Structure, mechanism, and inhibition of histone deacetylases and related metalloenzymes. *Curr Opin Struct Biol* 21:735–743
59. Yoo J, Medina-Franco JL (2012) Inhibitors of DNA methyltransferases: insights from computational studies. *Curr Med Chem* 19(21):3475–3487
60. Copeland RA, Solomon ME, Richon VM (2009) Protein methyltransferases as a target class for drug discovery. *Nat Rev Drug Discov* 8:724–732
61. Lohse B, Kristensen JL, Kristensen LH, Agger K, Helin K, Gajhede M, Clausen RP (2011) Inhibitors of histone demethylases. *Bioorg Med Chem* 19:3625–3636
62. Cho WC (2012) Exploiting the therapeutic potential of microRNAs in human cancer. *Expert Opin Ther Targets* 16:345–350
63. Esau CC, Monia BP (2007) Therapeutic potential for microRNAs. *Adv Drug Deliv Rev* 59:101–114
64. Bratkovič T, Glavan G, Strukelj B, Zivin M, Rogelj B (2012) Exploiting microRNAs for cell engineering and therapy. *Biotechnol Adv* 30:753–765
65. Ling H, Fabbri M, Calin GA (2013) MicroRNAs and other non-coding RNAs as targets for anticancer drug development. *Nat Rev Drug Discov* 12:847–865
66. De Santa F, Iosue I, Del Rio A, Fazi F (2013) microRNA biogenesis pathway as a therapeutic target for human disease and cancer. *Curr Pharm Des* 19:745–764
67. Bayley J-P, Devilee P (2012) The Warburg effect in 2012. *Curr Opin Oncol* 24:62–67
68. Le A, Cooper CR, Gouw AM, Dinavahi R, Maitra A, Deck LM, Royer RE, Vander Jagt DL, Semenza GL, Dang C V (2010) Inhibition of lactate dehydrogenase A induces oxidative stress and inhibits tumor progression. *Proc Natl Acad Sci U S A* 107:2037–2042
69. Salani B, Marini C, Del Rio A et al (2013) Metformin impairs glucose consumption and survival in Calu-1 cells by direct inhibition of hexokinase-II. *Sci Rep* 3:2070
70. Zhao Y, Butler EB, Tan M (2013) Targeting cellular metabolism to improve cancer therapeutics. *Cell Death Dis* 4:e532
71. Birsoy K, Sabatini DM, Possemato R (2012) Untuning the tumor metabolic machine: targeting cancer metabolism: a bedside lesson. *Nat Med* 18:1022–1023
72. Vander Heiden MG (2011) Targeting cancer metabolism: a therapeutic window opens. *Nat Rev Drug Discov* 10:671–684
73. Stefanska B, Karlic H, Varga F, Fabianowska-Majewska K, Haslberger A (2012) Epigenetic mechanisms in anti-cancer actions of bioactive food components—the implications in cancer prevention. *Br J Pharmacol* 167:279–297
74. Kirk H, Cefalu WT, Ribnicky D, Liu Z, Eilertsen KJ (2008) Botanicals as epigenetic modulators for mechanisms contributing to development of metabolic syndrome. *Metabolism* 57:16–23
75. Khan SI, Aumsuwan P, Khan IA, Walker LA, Dasmahapatra AK (2012) Epigenetic events associated with breast cancer and their prevention by dietary components targeting the epigenome. *Chem Res Toxicol* 25:61–73
76. Su Y, Shankar K, Rahal O, Simmen RCM (2011) Bidirectional signaling of mammary epithelium and stroma: implications for breast cancer—preventive actions of dietary factors. *J Nutr Biochem* 22:605–611
77. Lustberg MB, Ramaswamy B (2010) Epigenetic therapy in breast cancer. *Curr Breast Cancer Rep* 3:34–43
78. Ramaswamy B, Sparano JA (2010) Targeting epigenetic modifications for the treatment and prevention of breast cancer. *Curr Breast Cancer Rep* 2:198–207
79. Thornburg KL, Shannon J, Thuillier P, Turker MS (2010) In utero life and epigenetic predisposition for disease. *Adv Genet* 71:57–78
80. Piaz FD, Vassallo A, Rubio OC, Castellano S, Sbardella G, De Tommasi N (2011) Chemical biology of histone acetyltransferase natural compounds modulators. *Mol Divers* 15:401–416

81. Mantelingu K, Reddy BAA, Swaminathan V et al (2007) Specific inhibition of p300-HAT alters global gene expression and represses HIV replication. *Chem Biol* 14:645–657
82. Nyström M (2009) Diet and epigenetics in colon cancer. *World J Gastroenterol* 15:257
83. Duthie SJ (2011) Epigenetic modifications and human pathologies: cancer and CVD. *P Nutr Soc* 70:47–56
84. Van Engeland M, Herman JG (2010) Viewing the epigenetics of colorectal cancer through the window of folic acid effects. *Cancer Prev Res* 3:1509–1512
85. Garagnani P, Pirazzini C, Franceschi C (2013) Colorectal cancer microenvironment: among nutrition, gut microbiota, inflammation and epigenetics. *Curr Pharm Des* 19:765–778
86. Nencioni A, Bruzzone S, Del Rio A (2013) Editorial: NAD⁺ biosynthesis and signaling as an emerging area in medicinal chemistry. *Curr Top Med Chem* 13:2905–2906
87. Teperino R, Schoonjans K, Auwerx J (2010) Histone methyl transferases and demethylases; can they link metabolism and transcription? *Cell Metab* 12:321–327
88. Rajendran P, Williams DE, Ho E, Dashwood RH (2011) Metabolism as a key to histone deacetylase inhibition. *Crit Rev Biochem Mol Biol* 46:181–199
89. Petrelli A, Giordano S (2008) From single- to multi-target drugs in cancer therapy: when aspecificity becomes an advantage. *Curr Med Chem* 15:422–432
90. Talele TT, Khedkar SA, Rigby AC (2010) Successful applications of computer aided drug discovery: moving drugs from concept to the clinic. *Curr Top Med Chem* 10:127–141
91. Van Drie JH (2007) Computer-aided drug design: the next 20 years. *J Comput Aided Mol Des* 21:591–601
92. Borhani DW, Shaw DE (2012) The future of molecular dynamics simulations in drug discovery. *J Comput Aided Mol Des* 26:15–26
93. Moura Barbosa AJ, Del Rio A (2012) Freely accessible databases of commercial compounds for high- throughput virtual screenings. *Curr Top Med Chem* 12:866–877
94. Del Rio A, Barbosa AJM, Caporuscio F, Mangiatordi GF (2010) CoCoCo: a free suite of multiconformational chemical databases for high-throughput virtual screening purposes. *Mol Biosyst* 6:2122–2128
95. Del Rio A, Barbosa AJM, Caporuscio F (2011) Use of large multiconformational databases with structure-based pharmacophore models for fast screening of commercial compound collections. *J Cheminform* 3:P27
96. Bottegoni G, Favia AD, Recanatini M, Cavalli A (2012) The role of fragment-based and computational methods in polypharmacology. *Drug Discov Today* 17:23–34
97. Copeland RA, Olhava EJ, Scott MP (2010) Targeting epigenetic enzymes for drug discovery. *Curr Opin Chem Biol* 14:505–510
98. De Koning L, Corpet A, Haber JE, Almouzni G (2007) Histone chaperones: an escort network regulating histone traffic. *Nat Struct Mol Biol* 14:997–1007
99. Golbabapour S, Abdulla MA, Hajrezaei M (2011) A concise review on epigenetic regulation: insight into molecular mechanisms. *Int J Mol Sci* 12:8661–8694
100. Mai A, Cheng D, Bedford MT et al (2008) Epigenetic multiple ligands: mixed histone/protein methyltransferase, acetyltransferase, and class III deacetylase (sirtuin) inhibitors. *J Med Chem* 51:2279–2290
101. Suganuma T, Workman JL (2008) Crosstalk among histone modifications. *Cell* 135:604–607
102. Jones P (2012) Development of second generation epigenetic agents. *Med Chem Comm* 3:135
103. Mani S, Herceg Z (2010) DNA demethylating agents and epigenetic therapy of cancer. *Adv Genet* 70:327–340
104. Karberg S (2009) Switching on epigenetic therapy. *Cell* 139:1029–1031
105. Medina-Franco JL, Caulfield T (2011) Advances in the computational development of DNA methyltransferase inhibitors. *Drug Discov Today* 16:418–425
106. Kristensen LS, Nielsen HM, Hansen LL (2009) Epigenetics and cancer treatment. *Eur J Pharmacol* 625:131–142
107. Hamm CA, Costa FF (2011) The impact of epigenomics on future drug design and new therapies. *Drug Discov Today* 16:626–635

108. Heinke R, Carlino L, Kannan S, Jung M, Sippl W (2011) Computer- and structure-based lead design for epigenetic targets. *Bioorg Med Chem* 19:3605–3615
109. Yoo J, Medina-Franco JL (2011) Discovery and optimization of inhibitors of DNA methyl-transferase as novel drugs for cancer therapy. In: Rundefeldt C (ed) *Drug development—a case study based insight into modern strategies*. InTech, Croatia
110. Neugebauer RC, Uchieczowska U, Meier R, Hruby H, Valkov V, Verdin E, Sippl W, Jung M (2008) Structure-activity studies on splitomicin derivatives as sirtuin inhibitors and computational prediction of binding mode. *J Med Chem* 51:1203–1213
111. Costantini S, Sharma A, Raucci R, Costantini M, Autiero I, Colonna G (2013) Genealogy of an ancient protein family: the Sirtuins, a family of disordered members. *BMC Evol Biol* 13:60
112. Sakkiah S, Arooj M, Kumar MR, Eom SH, Lee KW (2013) Identification of inhibitor binding site in human sirtuin 2 using molecular docking and dynamics simulations. *PLoS One* 8:e51429
113. Chen L (2011) Medicinal chemistry of sirtuin inhibitors. *Curr Med Chem* 18:1936–1946
114. Mak L, Liggi S, Tan L, Kusonmano K, Rollinger JM, Glen RC, Kirchmair J, Koutsoukas A (2012) Anti-cancer drug development: computational strategies to identify and target proteins involved in cancer metabolism. *Curr Pharm Des* 19(4):532–577
115. Manerba M, Vettraiño M, Fiume L, Di Stefano G, Sartini A, Giacomini E, Buonfiglio R, Roberti M, Recanatini M (2012) Galloflavin (CAS 568–80-9): a novel inhibitor of lactate dehydrogenase. *Chem Med Chem* 7:311–317
116. Sanders MPA, Barbosa AJM, Zarzycka B, Nicolaes GAF, Klomp JPG, de Vlieg J, Del Rio A (2012) Comparative analysis of pharmacophore screening tools. *J Chem Inf Model* 52:1607–1620
117. Schuster D, Wolber G (2010) Identification of bioactive natural products by pharmacophore-based virtual screening. *Curr Pharm Des* 16:1666–1681
118. Galvez-Llompert M, Zanni R, García-Domenech R (2011) Modeling natural anti-inflammatory compounds by molecular topology. *Int J Mol Sci* 12:9481–9503
119. Gálvez-Llompert M, Recio MC, García-Domenech R (2011) Topological virtual screening: a way to find new compounds active in ulcerative colitis by inhibiting NF- κ B. *Mol Divers* 15:917–926
120. Harvey AL (2008) Natural products in drug discovery. *Drug Discov Today* 13:894–901
121. Rosén J, Gottfries J, Muresan S, Backlund A, Oprea TI (2009) Novel chemical space exploration via natural products. *J Med Chem* 52:1953–1962
122. Singh N, Guha R, Giulianotti MA, Pinilla C, Houghten RA, Medina-Franco JL (2009) Chemoinformatic analysis of combinatorial libraries, drugs, natural products, and molecular libraries small molecule repository. *J Chem Inf Model* 49:1010–1024
123. Füllbeck M, Michalsky E, Dunkel M, Preissner R (2006) Natural products: sources and databases. *Nat Prod Rep* 23:347–356
124. Lachance H, Wetzel S, Kumar K, Waldmann H (2012) Charting, navigating, and populating natural product chemical space for drug discovery. *J Med Chem* 55:5989–6001
125. Yongye AB, Waddell J, Medina-Franco JL (2012) Molecular scaffold analysis of natural products databases in the public domain. *Chem Biol Drug Des* 80:717–724
126. Bauer RA, Wurst JM, Tan DS (2010) Expanding the range of “druggable” targets with natural product-based libraries: an academic perspective. *Curr Opin Chem Biol* 14:308–314
127. Kumar K, Waldmann H (2009) Synthesis of natural product inspired compound collections. *Angew Chem Int Ed Engl* 48:3224–3242
128. Over B, Wetzel S, Grütter C, Nakai Y, Renner S, Rauh D, Waldmann H (2012) Natural-product-derived fragments for fragment-based ligand discovery. *Nat Chem* 5:21–28
129. International network of food data systems. <http://www.fao.org/infoods/infoods/en/>. Accessed 7 Sept 2014
130. USDA National Nutrient Database. <http://ndb.nal.usda.gov/>. Accessed 7 Sept 2014
131. FooDB. <http://www.foodb.ca/>. Accessed 7 Sept 2014