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Synthesis, spectral characterization, in vitro antibacterial, antifungal and cytotoxic activities of Co(II), Ni(II) and Cu(II) complexes with 1,2,4-triazole Schiff bases

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Abstract

A series of metal complexes of cobalt(II), nickel(II) and copper(II) have been synthesized with newly synthesized biologically active 1,2,4-triazole Schiff bases derived from the condensation of 3-substituted-4-amino-5-mercapto-1,2,4-triazole and 8-formyl-7-hydroxy-4-methylcoumarin, which have been characterized by elemental analyses, spectroscopic measurements (IR, UV–vis, fluorescence, ESR), magnetic measurements and thermal studies. Electrochemical study of the complexes is also reported. All the complexes are soluble to limited extent in common organic solvents but soluble to larger extent in DMF and DMSO and are non-electrolytes in DMF and DMSO. All these Schiff bases and their complexes have also been screened for their antibacterial (*Escherichia coli*, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Pseudomonas aeruginosa* and *Salmonella typhi*) and antifungal activities (*Aspergillus niger*, *Aspergillus flavus* and *Cladosporium*) by MIC method. The brine shrimp bioassay was also carried out to study their in vitro cytotoxic properties.

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Keywords: Synthesis; Biological activity; Electrochemical; 1,2,4-Triazole; Coumarin; Complexes

1. Introduction

Coumarins have long been recognized to possess anti-inflammatory [1], antioxidant [2], antithrombotic [3], antiallergic [4], hepatoprotective [4], antiviral [2] and anticarcinogenic [4] activities. The hydroxycoumarins are typical phenolic compounds and therefore act as potent metal chelators and free radical scavengers. They are powerful chain-breaking antioxidants [5]. The coumarins display a remarkable array of biochemical and pharmacological actions [1–4]; the antitumor effects of coumarin and its major metabolite, 7-hydroxycoumarin, were tested in several human tumor cell lines [6]. Furthermore, cytotoxic effects of complexes of coumarin derivatives were examined on several neuronal cell lines [7].

It is well known that N and S atoms play a key role in the coordination of metals at the active sites of numerous metallo-biomolecules. Metallo-organic chemistry is becoming an emerging area of research due to the demand for new metal-based antibacterial and antifungal compounds [8,9]. The serious medical problem [9–12] of bacterial and fungal resistance and the rate at which it develops have led to increasing levels of resistance to classical antibiotics. The discovery and development of effective antibacterial and antifungal drugs with novel mechanism of action have become an urgent task for infectious diseases research programs [13]. Many investigations have proved that binding of a drug to a metalloelement enhances its activity and in some cases, the complex possesses even more healing properties than the parent drug [14]. Triazole derivatives [15–18] are known to possess antibacterial, fungicidal, hypotensive and hypothermic activities. Metal complexes of 1,2,4-triazole derivatives have been extensively

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[illegible]

Table 2
Antibacterial and antifungal results of Co(II), Ni(II) and Cu(II) complexes (**1–12**) and standard

| Compound | Conc. ($\mu\text{g ml}^{-1}$) | Antibacterial activity (zone of inhibition in %) | | | | | Antifungal activity (zone of inhibition in %) | | |
|-----------|------------------------------------|--|------------------|--------------------|----------------------|-----------------|---|---------------------|-----------------|
| | | <i>E. coli</i> | <i>S. aureus</i> | <i>S. pyogenes</i> | <i>P. aeruginosa</i> | <i>S. typhi</i> | <i>A. flavus</i> | <i>Cladosporium</i> | <i>A. niger</i> |
| 1 | 100 | 58 | 55 | 61 | 58 | 70 | 58 | 67 | 59 |
| | 50 | 45 | 42 | 50 | 50 | 64 | 49 | 63 | 51 |
| | 25 | 48 | 39 | 47 | 49 | 66 | 47 | 63 | 48 |
| 2 | 100 | 67 | — | 69 | 63 | 75 | 60 | — | 69 |
| | 50 | 62 | — | 57 | 54 | 60 | 52 | — | 54 |
| | 25 | 60 | — | 52 | 55 | 62 | 50 | — | 48 |
| 3 | 100 | 68 | 60 | 71 | 77 | 79 | 77 | 84 | 72 |
| | 50 | 63 | 54 | 64 | 68 | 62 | 70 | 77 | 67 |
| | 25 | 58 | 49 | 60 | 70 | 61 | 69 | 80 | 65 |
| 4 | 100 | 68 | 61 | 73 | 80 | 80 | 80 | 95 | 78 |
| | 50 | 63 | 55 | 67 | 69 | 70 | 71 | 80 | 68 |
| | 25 | 60 | 47 | 66 | 72 | 74 | 67 | 83 | 57 |
| 5 | 100 | 64 | 63 | 50 | 52 | 71 | 52 | 78 | 73 |
| | 50 | 58 | 59 | — | — | 60 | — | 72 | 67 |
| | 25 | 52 | 53 | — | — | 59 | — | 71 | 59 |
| 6 | 100 | 67 | 69 | 55 | 55 | 78 | 58 | 85 | 75 |
| | 50 | 60 | 62 | 48 | 44 | 68 | 52 | 76 | 70 |
| | 25 | 59 | 62 | 41 | 39 | 68 | 47 | 73 | 61 |
| 7 | 100 | 71 | 70 | 59 | 58 | 80 | 64 | 81 | 70 |
| | 50 | 67 | 65 | 49 | 45 | 69 | 57 | 73 | 61 |
| | 25 | 68 | 66 | 43 | 46 | 70 | 51 | 71 | 52 |
| 8 | 100 | 74 | 71 | — | 60 | 83 | 66 | 80 | 66 |
| | 50 | 70 | 62 | — | 52 | 71 | 57 | 78 | 52 |
| | 25 | 67 | 60 | — | 50 | 70 | 52 | 73 | 48 |
| 9 | 100 | 58 | 59 | 52 | 49 | 68 | 51 | 60 | 68 |
| | 50 | 47 | 53 | — | — | 59 | — | 52 | 60 |
| | 25 | 46 | 52 | — | — | 54 | — | 51 | 59 |
| 10 | 100 | 59 | — | 59 | — | 72 | 54 | 58 | 70 |
| | 50 | 50 | — | 43 | — | 61 | 47 | 45 | 63 |
| | 25 | 49 | — | 41 | — | 57 | 41 | 42 | 60 |
| 11 | 100 | 68 | 70 | 63 | 62 | 76 | 58 | 69 | 68 |
| | 50 | 62 | 63 | 54 | 51 | 64 | 51 | 58 | 58 |
| | 25 | 59 | 61 | 52 | 46 | 60 | 48 | 52 | 51 |
| 12 | 100 | 71 | 73 | 64 | 65 | 80 | — | 79 | 62 |
| | 50 | 67 | 65 | 54 | 52 | 71 | — | 70 | 54 |
| | 25 | 65 | 64 | 53 | 48 | 65 | — | 75 | 50 |
| Standard | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 50 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| | 25 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |

suitably diluted with sterilized distilled water to get dilution of 100, 50 and $25 \mu\text{g ml}^{-1}$. Control for each dilution was prepared by diluting 10 ml of solvent instead of stock solution with sterilized distilled water.

The bacteria were subcultured in agar medium. The Petri dishes were incubated for 24 h at 37°C . Standard antibacterial drug (gentamycine) was also screened under similar conditions for comparison. The fungi were subcultured in potato dextrose agar medium. Standard antifungal drug (fluconazole) was used for comparison. The Petri dishes were incubated for 48 h at 37°C . The wells were dug in the agar media using a sterile metallic borer. Activity was determined by measuring the diameter of the zone showing complete inhibition (mm). Growth inhibition was compared with the standard drugs. In order to clarify any effect of DMF on the biological screening, separate studies were carried out with solutions alone of DMF and they showed no activity against any microbial strains.

3.1.1. Minimum inhibitory concentration (MIC)

Compounds showing promising antibacterial/antifungal activity were selected for minimum inhibitory concentration studies. The minimum inhibitory concentration was determined by assaying at 100, 50 and $25 \mu\text{g ml}^{-1}$ concentrations along with standards at the same concentrations.

3.1.2. Pharmacology results

The microbial results are systematized in Tables 1 and 2 and Figs. 2 and 3. The antibacterial and antifungal studies suggested that all the Schiff bases were found to be biologically active and their metal(II) complexes showed significantly enhanced antibacterial and antifungal activities. It is, however, known [30,31] that chelation tends to make the Schiff bases act as more powerful and potent bacteriostatic agents, thus inhibiting the growth of bacteria and fungi more than the parent Schiff bases. It is suspected that factors such as solubility, conductivity, dipole moment and cell permeability mechanism

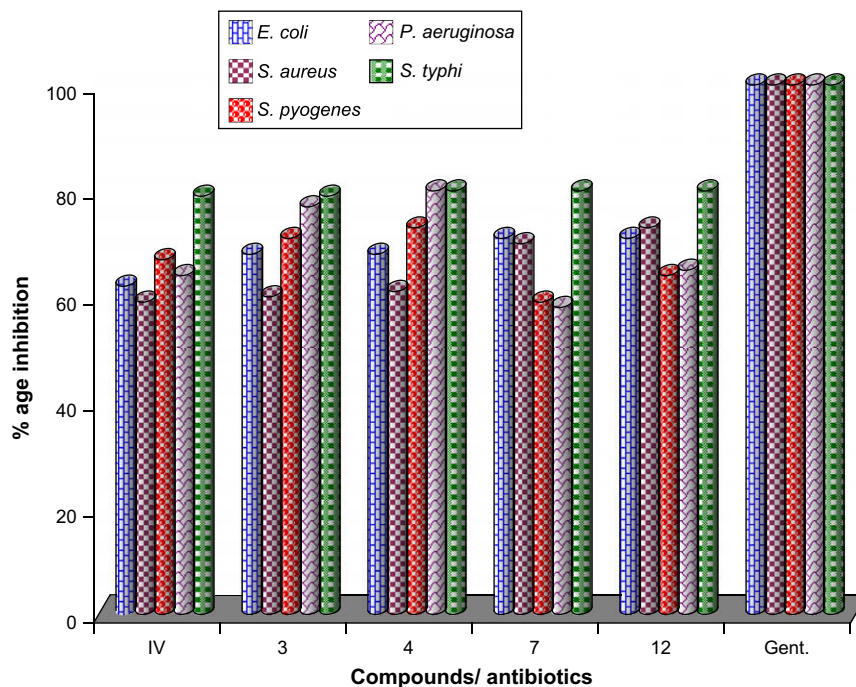


Fig. 2. In vitro antibacterial spectrum of compounds **IV**, **3**, **4**, **7**, **12**, and gentamycin (Std) at $100 \mu\text{g ml}^{-1}$ concentration.

(influenced by the presence of metal ions) may be the possible reasons for the increase in activity.

In the case of bacteriological studies it was observed that some of the Schiff bases were found potentially active against all bacterial strains. Metal(II) complexes (**1–12**) of these Schiff bases (**I–IV**) were also screened against the same bacterial strains. It was evident that overall potency of the uncoordinated compounds was enhanced on coordination with

metal ions, especially with *S. typhi*. Among these metal complexes compounds **3**, **4**, **7** and **12** show high activity against *S. typhi*.

In the case of antifungal activity, the results were compared with the standard drug (fluconazole). All Schiff bases showed activity against fungal species. However, the Co(II), Ni(II) and Cu(II) complexes (**1–12**) of these Schiff bases showed much enhanced activity as compared to the uncoordinated

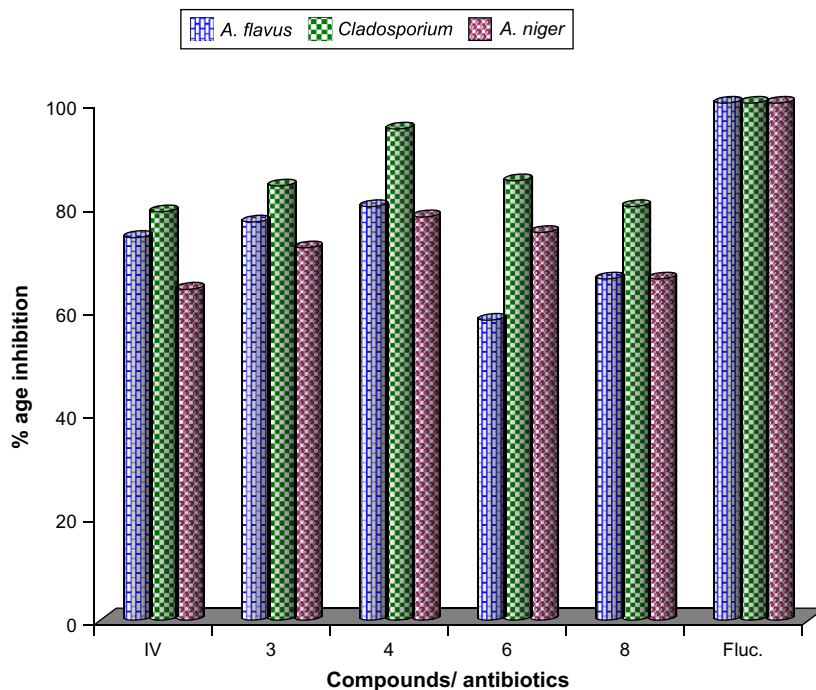


Fig. 3. In vitro antifungal spectrum of compounds **IV**, **3**, **4**, **6**, **8**, and fluconazole (Std) at $100 \mu\text{g ml}^{-1}$ concentration.

Table 3

Elemental analyses of Co(II), Ni(II) and Cu(II) complexes and their magnetic and molar conductance data

| Compound | Empirical formula | M% | | C% | | N% | | S% | | Molar conductance ($\text{Ohm}^{-1} \text{cm}^{-2} \text{mol}^{-1}$) | Mag. moments (μ_{eff} in BM) |
|----------|---|-------|-------|-------|-------|-------|-------|------|-------|---|---|
| | | Obsd | Calcd | Obsd | Calcd | Obsd | Calcd | Obsd | Calcd | | |
| 1 | $\text{Co}(\text{C}_{13}\text{H}_8\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 14.90 | 14.92 | 39.48 | 39.50 | 14.17 | 14.18 | 8.08 | 8.10 | 28 | 4.72 |
| 2 | $\text{Co}(\text{C}_{14}\text{H}_{10}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 14.39 | 14.41 | 41.05 | 41.08 | 13.68 | 13.69 | 7.81 | 7.83 | 26.12 | 4.84 |
| 3 | $\text{Co}(\text{C}_{15}\text{H}_{12}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 13.90 | 13.93 | 42.55 | 42.56 | 13.22 | 13.24 | 7.54 | 7.56 | 25.34 | 4.95 |
| 4 | $\text{Co}(\text{C}_{16}\text{H}_{14}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 13.45 | 13.48 | 43.92 | 43.94 | 12.80 | 12.82 | 7.30 | 7.32 | 22.28 | 4.86 |
| 5 | $\text{Ni}(\text{C}_{13}\text{H}_8\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 14.85 | 14.86 | 39.49 | 39.52 | 14.17 | 14.19 | 8.10 | 8.11 | 18.26 | 3.24 |
| 6 | $\text{Ni}(\text{C}_{14}\text{H}_{10}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 14.33 | 14.36 | 41.10 | 41.11 | 13.69 | 13.70 | 7.80 | 7.82 | 20.18 | 3.26 |
| 7 | $\text{Ni}(\text{C}_{15}\text{H}_{12}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 13.86 | 13.88 | 42.55 | 42.58 | 13.24 | 13.25 | 7.55 | 7.57 | 21.35 | 3.22 |
| 8 | $\text{Ni}(\text{C}_{16}\text{H}_{14}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 13.42 | 13.44 | 43.95 | 43.96 | 12.80 | 12.82 | 7.31 | 7.33 | 24.12 | 3.28 |
| 9 | $\text{Cu}(\text{C}_{13}\text{H}_8\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 15.88 | 15.90 | 39.02 | 39.04 | 14.01 | 14.02 | 8.00 | 8.01 | 25.14 | 1.75 |
| 10 | $\text{Cu}(\text{C}_{14}\text{H}_{10}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 15.33 | 15.36 | 40.60 | 40.62 | 13.51 | 13.54 | 7.71 | 7.74 | 26.45 | 1.83 |
| 11 | $\text{Cu}(\text{C}_{15}\text{H}_{12}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 14.85 | 14.86 | 42.09 | 42.10 | 13.07 | 13.09 | 7.45 | 7.48 | 24.56 | 1.87 |
| 12 | $\text{Cu}(\text{C}_{16}\text{H}_{14}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 14.37 | 14.39 | 43.46 | 43.48 | 12.65 | 12.68 | 7.23 | 7.25 | 23.27 | 1.90 |

compounds, especially with *Cladosporium*. All Schiff bases show high activity against *Cladosporium*. Schiff base (II) is inactive towards *A. flavus*. Compounds 3, 4, 6, 8 and 11 show promising results.

The biological activity of the ligands exhibited a marked enhancement on coordination with the metal ions against all fungal strains. However, the metal complexes showed good antifungal activity against *A. niger*, *A. flavus* and *Cladosporium*. It was evident from the data that this activity significantly increased on coordination. This enhancement in the activity may be rationalized on the basis that their structures mainly possess an additional C=N bond. It has been suggested that the ligands with nitrogen and oxygen donor systems inhibit enzyme activity, since the enzymes which require these groups for their activity appear to be especially more susceptible to deactivation by metal ions on coordination. Moreover, coordination reduces the polarity [32] of the metal ion mainly because of the partial sharing of its positive charge with the donor groups [30] within the chelate ring system formed during coordination. This process, in turn, increases the lipophilic nature of the central metal atom, which favors its permeation more efficiently through the lipid layer of the microorganism [31], thus destroying them more aggressively.

3.1.3. In vitro cytotoxicity

The synthesized Schiff bases and their Co(II), Ni(II) and Cu(II) complexes were screened for their cytotoxicity (brine shrimp bioassay) by using the protocol of Meyer et al. [32]. Brine shrimp (*Artemia salina* leach) eggs were hatched in a shallow rectangular plastic dish (22 × 32 cm) filled with

artificial seawater, which was prepared with a commercial salt mixture and double distilled water. An unequal partition was made in the plastic dish with the help of a perforated device. Approximately 50 mg of eggs were sprinkled into the large compartment, which was darkened while the minor compartment was open to ordinary light.

After two days nauplii were collected by a pipette from the lighted side. A sample of the test compound was prepared by dissolving 20 mg of each compound in 2 ml of DMF. From this stock solution 100, 50 and 10 $\mu\text{g ml}^{-1}$ were transferred to nine vials (three for each dilutions were used for each test sample and LD₅₀ is the mean of three values) and one vial was kept as control having 2 ml of DMF only. The solvent was allowed to evaporate overnight. After two days, when shrimp larvae were ready, 1 ml of seawater and 10 shrimps were added to each vial (30 shrimps/dilution) and the volume was adjusted with seawater to 5 ml per vial. After 24 h the number of survivors was counted. Data were analyzed by a Finney computer program to determine the LD₅₀ values [33].

In the case of cytotoxic activity it was observed that only Schiff bases III and IV and compounds 3, 7, 11, and 12 displayed weak cytotoxic activity against *A. salina*, while the other compounds gave values of LD₅₀ therefore can be considered non-cytotoxic.

Table 5

The important infrared frequencies (in cm^{-1}) of Co(II), Ni(II) and Cu(II) complexes of 3-substituted-4-amino(8-formyl-7-hydroxy-4-methylcoumarin)-5-mercapto-1,2,4-triazole Schiff bases

| Complex | $\nu(\text{OH})$ | $\nu(\text{C}=\text{O})$ | $\nu(\text{C}=\text{N})$ | Phenolic $\nu(\text{C}-\text{O})$ | $\nu(\text{M}-\text{N})$ | $\nu(\text{M}-\text{S})$ | $\nu(\text{M}-\text{O})$ |
|---------|------------------|--------------------------|--------------------------|--------------------------------------|--------------------------|--------------------------|--------------------------|
| 1 | 3415 | 1685 | 1618 | 1345 | 447 | 365 | 376 |
| 2 | 3435 | 1681 | 1622 | 1342 | 456 | 368 | 378 |
| 3 | 3430 | 1683 | 1620 | 1348 | 463 | 367 | 375 |
| 4 | 3418 | 1680 | 1621 | 1349 | 448 | 364 | 379 |
| 5 | 3429 | 1688 | 1617 | 1342 | 450 | 370 | 380 |
| 6 | 3431 | 1690 | 1620 | 1345 | 458 | 369 | 381 |
| 7 | 3430 | 1685 | 1615 | 1350 | 463 | 371 | 379 |
| 8 | 3426 | 1689 | 1618 | 1347 | 476 | 368 | 384 |
| 9 | 3420 | 1685 | 1615 | 1351 | 449 | 348 | 382 |
| 10 | 3434 | 1687 | 1612 | 1348 | 475 | 346 | 385 |
| 11 | 3432 | 1690 | 1622 | 1346 | 480 | 347 | 381 |
| 12 | 3431 | 1689 | 1620 | 1353 | 446 | 345 | 378 |

Table 4

The important infrared frequencies (in cm^{-1}) of 3-substituted-4-amino(8-formyl-7-hydroxy-4-methylcoumarin)-5-mercapto-1,2,4-triazole Schiff bases

| Ligand | $\nu(\text{NH})$ | Lactonil $\nu(\text{C}=\text{O})$ | $\nu(\text{C}=\text{N})$ | H-bonded —OH stretching | $\nu(\text{C}=\text{C})$ | $\nu(\text{SH})$ | Phenolic $\nu(\text{C}-\text{O})$ |
|--------|------------------|--------------------------------------|--------------------------|----------------------------|--------------------------|------------------|--------------------------------------|
| I | 3140 | 1700 | 1629 | 3260 | 1590 | 2730 | 1279 |
| II | 3147 | 1707 | 1631 | 3220 | 1600 | 2720 | 1285 |
| III | 3138 | 1705 | 1630 | 3250 | 1595 | 2725 | 1292 |
| IV | 3135 | 1710 | 1628 | 3265 | 1597 | 2720 | 1294 |

Table 6

Ligand field parameters of Ni(II) complex with 3-substituted-4-amino(8-formyl-7-hydroxy-4-methycoumarin)-5-mercapto-1,2,4-triazole Schiff bases (I–IV)

| Complex | Transitions (cm^{-1}) | | | ν_2 Calcd (cm^{-1}) | Dq (cm^{-1}) | B^1 (cm^{-1}) | Distortion (%) | ν_1/ν_2 | LSFE | μ_{eff} Calcd (BM) | β | β^0 (%) |
|---------|----------------------------------|---------|---------|------------------------------------|-------------------------|----------------------------|----------------|---------------|--------|-------------------------------|---------|---------------|
| | ν_1 | ν_2 | ν_3 | | | | | | | | | |
| 5 | 9794 | 15 838 | 26 297 | 15858.55 | 979.4 | 851.57 | 0.130 | 1.617 | 33.579 | 3.194 | 0.806 | 19.359 |
| 6 | 9854 | 15 842 | 26 261 | 15914.52 | 985.4 | 840.91 | 0.456 | 1.600 | 33.785 | 3.192 | 0.796 | 20.369 |
| 7 | 9577 | 15 597 | 26 421 | 15640.43 | 957.7 | 888.69 | 0.278 | 1.629 | 32.835 | 3.202 | 0.842 | 15.843 |
| 8 | 9642 | 15 682 | 26 140 | 15662.26 | 964.2 | 858.41 | 0.126 | 1.626 | 33.058 | 3.200 | 0.813 | 18.710 |

4. Results and discussion

The Schiff bases were soluble in some organic solvents. All the Co(II), Ni(II) and Cu(II) complexes were stable in room temperature, non-hygroscopic, insoluble in water and many common organic solvents, infusible at high temperature and all of them were polymeric in nature. The elemental analyses shown in Table 3 agree well with the formation of 1:1 stoichiometry of the type $\text{ML} \cdot 2\text{H}_2\text{O}$. All the complexes are sparingly soluble in common organic solvents but these complexes are soluble to a larger extent in DMF and DMSO. The molar conductance values are too low to account for any dissociation in DMF indicating that complexes are non-electrolytic in nature.

4.1. IR spectra

The selected IR spectra of the Schiff bases and their metal complexes along with their tentative assignments are reported in Tables 4 and 5.

The IR spectra of the Schiff bases show characteristic bands due to $\nu(\text{NH})$ and $\nu(\text{SH})$ at 3145 and 2700 cm^{-1} , respectively [34]. Another band at 1100 cm^{-1} is assigned to $\nu(\text{C}=\text{S})$ [34]. These observations suggest that the Schiff bases exhibit thiol–thione tautomerism (Fig. 1). The broad band at 3220–3270 cm^{-1} , a strong band at 1705–1715, 1630–1625 and 1285 cm^{-1} in the IR spectra of the Schiff bases are assigned to H-bonded –OH stretching, $\nu(\text{C}=\text{O})$ lactonic carbonyl [35],

$\nu(\text{C}=\text{N})$ and phenolic $\nu(\text{C}-\text{O})$ vibrations, respectively. The medium intensity band in the region 770–760 cm^{-1} has been attributed to $\nu(\text{C}=\text{S})$. A medium band around 1055 cm^{-1} is characterized for $\nu(\text{O}-\text{C}-\text{O})$. In comparison with the spectra of the Schiff bases, all the complexes exhibit downward shift (10–20 cm^{-1}) of $\nu(\text{C}=\text{N})$ indicating the participation of azomethine nitrogen in the coordination to the metal ion.

The high intensity band due to phenolic C–O appeared in the region at 1285 cm^{-1} in the Schiff bases appeared as a medium to high intensity band in the 1350 cm^{-1} region in the complexes. These observations support the formation of M–O bonds via deprotonation. So the H-bonded –OH groups have been replaced by the metal ion.

The deprotonation of the thiol group is indicated by the absence of a band in the metal complexes at 2700 cm^{-1} , which is due to $\nu(\text{S}-\text{H})$ of Schiff bases, indicating that the metal is coordinated through sulphur atom also. This is supported by the lower frequency shift which appears around 685–670 cm^{-1} in the metal complexes due to $\nu(\text{C}-\text{S})$.

The presence of coordinated water molecules in the complexes [34] is indicated by a broad band in the region 3200–3500 cm^{-1} and two weaker bands in the region 750–800 and 700–720 cm^{-1} due to $\nu(\text{OH})$ rocking and wagging mode of vibrations, respectively [36].

An interesting feature observed is the red shift in lactone $\nu(\text{C}=\text{O})$ to the extent of about 15–20 cm^{-1} , suggesting that the metal is coordinated to the lactone oxygen [37]. This is

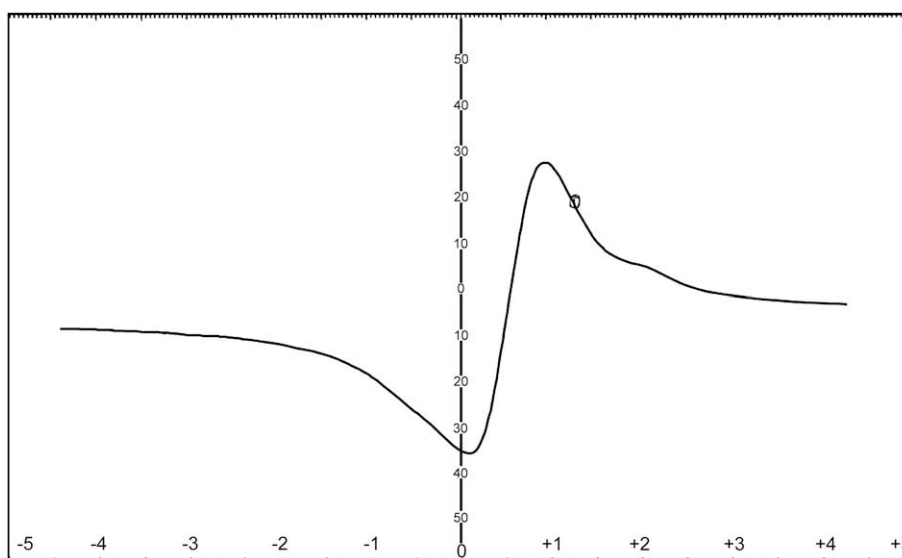


Fig. 4. ESR spectrum of Cu(II) complex (10).

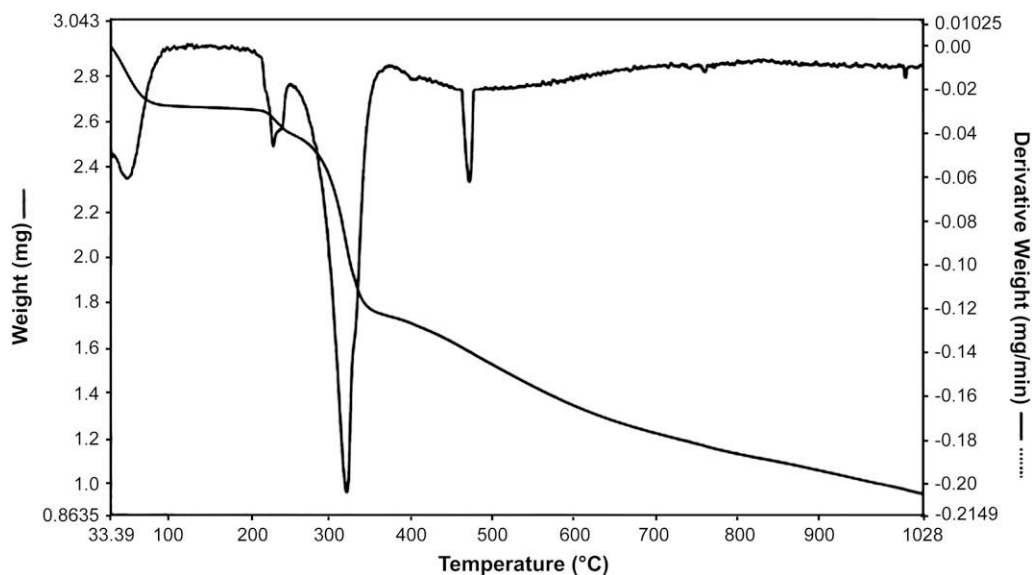


Fig. 5. Thermogravimetric (TGA/DTG) curves of Cu(II) complex (10).

further supported by downward shift in $\nu(\text{O}=\text{C}-\text{O})$ of the coumarin ring [38]. On the basis of IR data, it is concluded that all the metal ions are coordinated to the azomethine nitrogen, phenolic oxygen, sulphur atom and lactone oxygen.

The new bands in the region of 375–350 and 450–480 cm^{-1} in all the complexes are assigned to stretching frequencies of (M–O) and (M–N) bonds, respectively. The band in the region 333–379 cm^{-1} of far IR spectra is due to metal–sulphur bond formation.

Thus the IR spectral data results provide strong evidences for the complexation of the potentially tetradentate Schiff bases and also suggests that the complexes exist in the solid state as polymeric structure with bonding of metal(II) likely to both the deprotonated phenolic oxygen and lactose carbonyl oxygen.

4.2. ^1H NMR spectra

The ^1H NMR spectra of Schiff bases exhibit singlet at 13.58, 10.21, 8.62 ppm and multiplet at 7.2–7.5 ppm due to $-\text{NH}$

[39], phenolic OH, $-\text{CH}=\text{N}$ [40] and aromatic protons, respectively. In addition to these signals, a sharp signal at 3.5 ppm is attributed to SH protons [19]. These observations suggest that the Schiff bases exist in thiol–thione tautomerism. The singlet around 2.84 ppm is due to methyl protons [40].

4.3. Electronic spectra

The cobalt complexes exhibited two distinct bands in the region 9790–10 000 and 18 950–20 661 cm^{-1} which may be assigned to $^4\text{T}_{1g}(\text{F}) \rightarrow ^4\text{T}_{2g}(\text{F})$ (ν_1) and $^4\text{T}_{1g}(\text{F}) \rightarrow ^4\text{T}_{1g}(\text{P})$ (ν_3) transitions, respectively, and these are suggestive of octahedral geometry around the cobalt(II) ions [41,42]. The electronic spectra of nickel complexes showed d–d bands in the region 9570–10 000, 15 597–15 845 and 20 492–27 248 cm^{-1} . These are assigned [41] to the transitions $^3\text{A}_{2g}(\text{F}) \rightarrow ^3\text{T}_{2g}(\text{F})$ (ν_1), $^3\text{A}_{2g}(\text{F}) \rightarrow ^3\text{T}_{1g}(\text{F})$ (ν_2) and $^3\text{A}_{2g}(\text{F}) \rightarrow ^3\text{T}_{1g}(\text{P})$ (ν_3), respectively, consistent with their well-defined octahedral configuration [41,42]. The ligand field parameters [43] Dq , β , B' ,

Table 7

Thermogravimetric data of Co(II) (2), Ni(II) (6) and Cu(II) (10) complexes of 3-substituted-4-amino(8-formyl-7-hydroxy-4-methylcoumarin)-5-mercapto-1,2,4-triazole Schiff base (II)

| Empirical formula | Decomposition temperature (°C) | Weight loss (%) | | Metal oxide (%) | | Inference |
|---|--------------------------------|-----------------|-------|-----------------|-------|-------------------------------------|
| | | Obsd | Calcd | Obsd | Calcd | |
| $\text{Co}(\text{C}_{14}\text{H}_{10}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 205–235 | 8.77 | 8.80 | 18.30 | 18.32 | Loss of coordinated water molecules |
| | 310–325 | 31.05 | 31.06 | | | Loss of triazole moieties |
| | 490–485 | 45.71 | 45.73 | | | Loss of coumarin moieties |
| $\text{Ni}(\text{C}_{14}\text{H}_{10}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 210–228 | 8.79 | 8.81 | 18.25 | 18.28 | Loss of coordinated water molecules |
| | 320–330 | 31.04 | 31.07 | | | Loss of triazole moieties |
| | 485–495 | 45.74 | 45.76 | | | Loss of coumarin moieties |
| $\text{Cu}(\text{C}_{14}\text{H}_{10}\text{N}_4\text{O}_3\text{S}) \cdot 2\text{H}_2\text{O}$ | 200–230 | 8.70 | 8.71 | 19.20 | 19.23 | Loss of coordinated water molecules |
| | 290–330 | 30.68 | 30.71 | | | Loss of triazole moieties |
| | 480–510 | 45.20 | 45.22 | | | Loss of coumarin moieties |

ν_2/ν_1 and LFSE have been calculated (Table 6). The electronic spectra of Cu(II) complexes showed absorption band in the region 14 540–14 780 cm^{-1} attributed to ${}^2T_g \leftarrow {}^2E_g$ transition indicative of distorted octahedral geometry [44,45].

4.4. Magnetic studies

The magnetic moments obtained at room temperature are listed in Table 3. The magnetic measurements for Co(II) and Ni(II) complexes showed magnetic moment values of 4.3–5.2 and 2.8–3.5 BM, respectively, suggesting [46] consistency with their octahedral environment. The Cu(II) complexes show magnetic moments, 1.75–1.87 BM, slightly higher than the spin-only value 1.73 BM expected for one unpaired electron, which offers possibility of an octahedral geometry [47].

4.5. ESR spectra of copper(II) complex (10)

The ESR spectrum of copper(II) complex (10) with ligand (2) has been studied and depicted in Fig. 4. The g_{\parallel} and g_{\perp} values have been found to be 2.04351 and 2.15835, respectively. The g_{av} was calculated to be 2.12007. The Cu(II) complex shows reversed axial (compressed octahedral) with $g_{\parallel} < g_{\perp}$. The trend $g_{\parallel} < g_{\perp}$ showed that the electron is delocalised in d_z^2 orbital of the ground state of Cu(II). In this case ($g_{\parallel} < g_{\perp}$) distortion occurs by compression [48]. The parameter G , determined as $G = (g_{\parallel} - 2)/(g_{\perp} - 2)$, is found to be much less than 4 suggesting considerable interaction in the solid state [49].

4.6. Thermal studies

The thermal behavior of all the complexes is almost same. Hence, only Co(II) (2), Ni(II) (6) and Cu(II) (10) complexes were discussed.

The Cu(II) complex (10) has been reproduced in Fig. 5. Co(II) (2), Ni(II) (6) and Cu(II) (10) complexes decompose gradually with the formation of metal oxide above 530 °C. The nature of proposed chemical change with the temperature range and the percentage of metal oxide obtained are given in Table 7. The thermal decomposition of Co(II) (2), Ni(II) (6) and Cu(II) (10) complexes takes place in three steps as

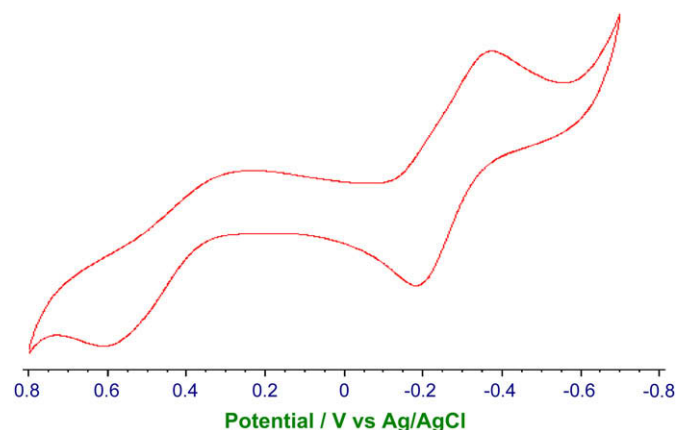


Fig. 6. Cyclic voltammogram of Cu(II) complex (10).

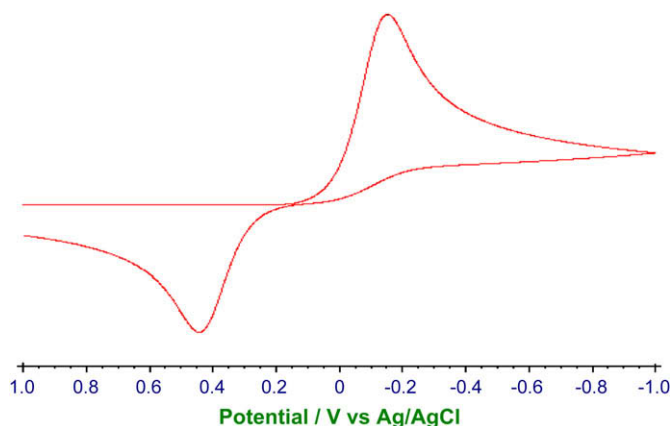


Fig. 7. Cyclic voltammogram of Ni(II) complex (6).

indicated by DTG peaks around 200–230, 290–330 and 480–510 °C corresponding to the mass loss of two coordinated water molecules, triazole moiety and coumarin moiety, respectively.

4.7. Electrochemistry

Electrochemical properties of the complexes were studied on a CHI1110A-Electrochemical analyzer in *N,N*-dimethyl formamide (DMF) containing 0.05 M *n*-Bu₄NClO₄ as the supporting electrolyte. A cyclic voltammogram of Cu(II) (10) (Fig. 6) radical displays a reduction peak at $E_{\text{pc}} = 0.2673$ V and again it reduced to Cu(I) and displays a reduction peak at $E_{\text{pc}} = -0.3733$ V, respectively, with a corresponding oxidation peak (Cu(I) radical) at $E_{\text{pa}} = -0.1822$ V and $E_{\text{pa}} = 0.6027$ V for Cu(II), respectively. The peak separation of this couple (ΔE_p) is 0.33 and 0.191 V at 0.05 V and increases with scan rate. The most significant feature of the Cu(II) complex is the Cu(II)/Cu(I) couple. The difference between forward and backward peak potentials can provide a rough evaluation of the degree of the reversibility of one electron transfer reaction. The analyses of cyclic voltammetric responses with the scan rate varying 50–250 mV/s give the evidence for quasi-reversible one electron oxidation state. The ratio of cathodic to anodic peak height was less than one. However, the peak current increases with the increase of the square root of the scan rates. This establishes the electrode process as diffusion controlled [50].

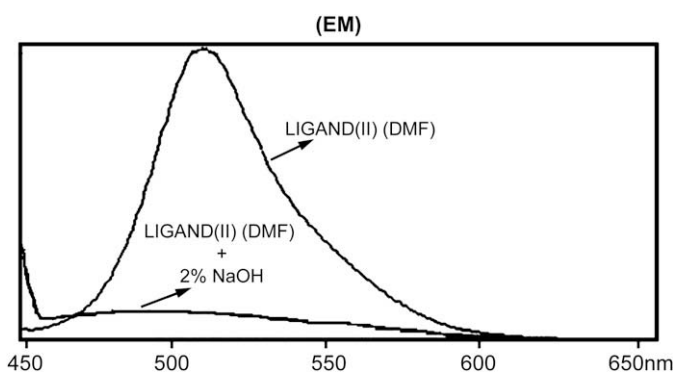


Fig. 8. Emission spectra of Schiff base (II) in DMF and red shift of Schiff base (II) in DMF with 2% NaOH.

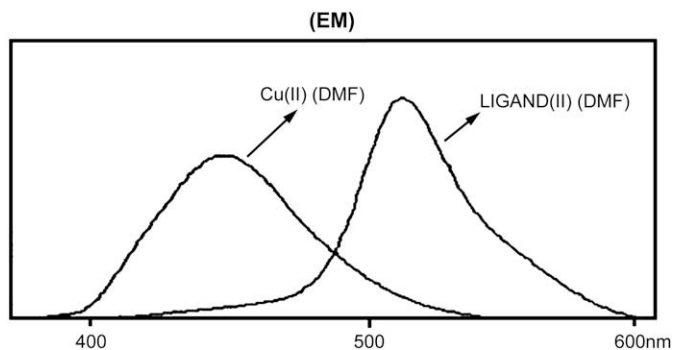


Fig. 9. Emission band of Schiff bases (II) and Cu(II) complex (10) in DMF solution.

The separation in peak potentials increases at higher scan rates. These characteristic features are consistent with the quasi-reversibility of Cu(II)/Cu(I) couple.

The Ni(II) (6) complex (Fig. 7) exhibits a reduction peak at $E_{pc} = 0.152$ V with a direct re-oxidation peak at $E_{pa} = 0.443$ V corresponding to the formation of Ni(II)/Ni(I) couple. The peak separation of this couple (ΔE_p) is 0.29 V. This Ni(II) complex also have a quasi-reversible character as the separation in peak potential is higher than 59 mV and the peak currents rise with increasing $\nu^{1/2}$. The difference between forward and backward peak potentials can provide a rough evaluation of the degree of the reversibility.

4.8. Fluorescence studies

The emission spectra of the Schiff bases derived from 3-substituted-4-amino-5-mercapto-1,2,4-triazole and 8-formyl-7-hydroxy-4-methylcoumarin and their complexes were investigated in various solvents such as DMF, DMSO, MeCN and dioxan.

4.8.1. Emission spectra

The Schiff bases characterized by an emission band around 511 nm in DMF, DMSO, MeCN and dioxan is due to the

formation of phenoxide anion and cleavage of the imine bond is observed in the Schiff bases. Upon addition of aqueous alkali (2% NaOH) to all the above-prepared solutions, we observed the band at 480 nm in DMF, DMSO, MeCN and dioxan solutions. The changes clearly indicate that proton transferred (H-bonded ion pair) species exist in equilibrium [51] and also we observed the λ_{max} of the Schiff bases undergoes red shift in DMF, DMSO, MeCN and dioxan solutions due to the hydrogen bond formation (Fig. 8) [52].

We have also studied the emission spectra of the Co(II), Ni(II) and Cu(II) complexes with 3-methyl-4-amino(8-formyl-7-hydroxy-4-methylcoumarin)-5-mercapto-1,2,4-triazole. The Co(II), Ni(II) and Cu(II) complexes were characterized by the emission band around 450 nm and it is observed that the emission band of Schiff bases around 510 nm disappeared because of the interaction of the phenolic oxygen with the metal ion (Fig. 9). There was decrease in intensity of fluorescence of Co(II), Ni(II) and Cu(II) complexes in all prepared solutions. In all other previous studies, it has been reported that transition metal ions decrease the fluorescence quite effectively [53,54]. Magnetic perturbation, redox activity, etc., have been invoked [54] in the past to rationalize fluorescence quenching by transition metal ions. But in the case of Cu(II) complexes we could observe the enhancement of fluorescence in MeCN solution.

5. Conclusion

The synthesized 3-substituted-4-amino(8-formyl-7-hydroxy-4-methylcoumarin)-5-mercapto-1,2,4-triazole Schiff bases act as tetradentate Schiff bases. The metals are coordinated to azomethine nitrogen, lactonyl oxygen, phenolic oxygen and sulphur atom. The analytical, IR, ESR, electronic, magnetic, and thermal studies confirm the bonding of Schiff bases to metal ions. Electrochemical study of Cu(II) and Ni(II) complexes can provide the degree of the reversibility of one electron transfer reaction and they have a quasi-reversible character. All Schiff bases were found potentially active

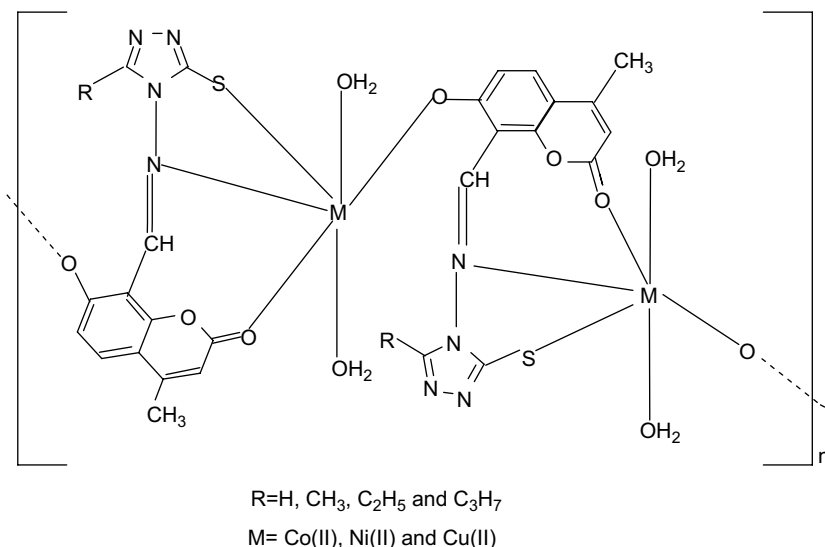


Fig. 10. Proposed structure of metal(II) complexes.

towards all microbial strains. Bacterial studies of the metal(II) complexes **3**, **4**, **7** and **12** and the fungal studies of complexes **3**, **4**, **6**, **8** and **11** show promising results.

The IR data, taken together with the insolubility of the complexes in water and as all the complexes are fusible at higher temperatures, suggest that they exist in the solid state as polymeric structures with bonding of metal(II) likely to both the deprotonated phenolic oxygen and lactone carbonyl oxygen [55–57].

All these observations put together lead us to propose the following structure (Fig. 10) in which the metal(II) complexes exhibit coordination number six on the basis of magnetic and electronic spectral data having the stoichiometry of the type $(ML \cdot 2H_2O)_2$.

6. Experimental protocols

6.1. Chemistry

All chemicals and solvents used were of AR grade. All metal(II) salts were used as their chlorides. Further remaining pure reagents were purchased from Ranbaxy Chemicals. The metal contents were determined gravimetrically by the known method [58]. The results of elemental analyses, molar conductance and magnetic data are listed in Table 1.

The IR spectra of the Schiff bases and their metal complexes were recorded on a HITACHI-270 IR spectrophotometer in the 4000–250 cm^{-1} region in KBr disks. The electronic spectra of the complexes were recorded in DMF on a VARIAN CARY 50-BIO UV-spectrophotometer in the region of 200–1100 nm. The 1H NMR spectra of Schiff bases were recorded in DMSO on a BRUKER 300 MHz spectrometer at room temperature using TMS as an internal reference. Thermogravimetric analyses data were measured from room temperature to 1000 °C at a heating rate of 10 °C/min. The data were obtained by using a PERKIN–ELMER DIAMOND TG/DTG instrument. Molar conductivity measurements were recorded on an ELICO-CM-82 T conductivity bridge with a cell having cell constant 0.51. Electrochemical studies were carried out using CHI1110A-Electrochemical analyzer and magnetic moments were carried out by Faraday balance.

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References

- [1] L. Pochet, C. Doucet, M. Schynts, N. Thierry, N. Boggeto, B. Pirotte, K.Y. Liang, B. Masereel, P. de Tulio, J. Delarge, M. Reboud-Ravaux, *J. Med. Chem.* 39 (1996) 2579.
- [2] X. Shao, D.H.L. Ekstrand, R. Bhikhabhai, C.F.R. Kallander, J.S. Gronowitz, *Antiviral Chem. Chemother.* 8 (1997) 149–159.
- [3] B. Rendenbach-Muller, R. Schelcker, M. Traut, H. Weifenbach, *Bioorg. Med. Chem. Lett.* 4 (1994) 1195.
- [4] S. Kostova, P. Raleva, Genova, R. Argirova, *Bioinorg. Chem. Appl.* (2006) 1–9.
- [5] Irena Kostova, *Curr. Med. Chem. — Anti-Cancer Agents* 5 (2005) 29–46.
- [6] U.S. Weber, B. Steffen, C. Sigers, *Res. Commun. Mol. Pathol. Pharmacol.* 99 (1998) 193–206.
- [7] I. Kostova, I. Manolov, M. Karaivanova, *Arch. Pharm.* 334 (2001) 157–162.
- [8] A. Scozzafava, C.T. Supuran, *J. Med. Chem.* 43 (2000) 3677.
- [9] S.A. Rice, M. Givskov, P. Steinberg, S. Kjelleberg, *J. Mol. Microbiol. Biotechnol.* 1 (1999) 23–31.
- [10] J.G. Lara, M. Masalha, S.J. Foster, *Drug Discov. Today* 10 (2005) 643–651.
- [11] G.D. Wright, *Chem. Biol.* 7 (2000) R127–R132.
- [12] J. Travis, J. Potempa, *Biochim. Biophys. Acta* 14 (2000) 35–50.
- [13] H.J. Smith, C. Simons, *Proteinase and Peptidase Inhibition: Recent Potential Targets for Drug Development*, Taylor and Francis, London, 2001.
- [14] S.J. Lippard, J.M. Berg, *Principles of Bioinorganic Chemistry*, University Science Books, Mill Valley, CA, 1999.
- [15] Ya Al-Sound, M.N. Al-Dweri, Na Al-Masoudi, *Il Farmaco* 59 (10) (2004) 775–783.
- [16] Hp Michael, C. Dines, *J. Mol. Struct.* 705 (2004) 177–187.
- [17] Z.H. Chohan, *Synth. React. Inorg. Met.-Org. Chem.* 34 (2004) 833.
- [18] Z.H. Chohan, A. Scozzafava, C.T. Supuran, *J. Enzyme Inhib. Med. Chem.* 17 (4) (2002) 261–266.
- [19] S.A. Patil, B.M. Badiger, S.M. Kudari, V.H. Kulkarni, *Transition Met. Chem.* 8 (1983) 238.
- [20] B.M. Badiger, S.A. Patil, S.M. Kudari, V.H. Kulkarni, *Rev. Roum. Chim.* 31 (1986) 849.
- [21] A.Y. Naik, S.D. Angadi, V.H. Kulkarni, *Oriental J. Chem.* 10 (1994) 23.
- [22] M.S. Yadawe, S.A. Patil, *Transition Met. Chem.* 22 (1997) 220.
- [23] P.G. Avaji, B.N. Reddy, P.S. Badami, S.A. Patil, *Transition Met. Chem.* 31 (2006) 842–848.
- [24] A. Kumar, G. Singh, R.N. Handa, S.N. Dubey, *Indian J. Chem.* 38A (1999) 613–617.
- [25] A.K. Sen, G. Singh, K. Singh, R.K. Noren, R.N. Handa, S.N. Dubey, *Indian J. Chem.* 36A (1997) 891–894.
- [26] D. Heng-Shan, Q. Bin, W. Kun, W. Qing-Lian, Z. Zi-Yi, *Magn. Reson. Chem.* 38 (2000) 210–212.
- [27] S.S. Papakonstantinou-Garuofalia, E. Tani, O. Todoulou, A. Papadaki-Valiraki, E. Filippatos, E.D. Clercq, P.N. Kourounakis, *J. Pharm. Pharmacol.* 50 (1) (1998) 117–124.
- [28] E. Spath, M. Pailer, *Chem. Ber.* 68 (1935) 940.
- [29] A.K. Sadana, Y. Miraza, K.R. Aneja, O. Prakash, *Eur. J. Med. Chem.* 38 (2003) 533–536.
- [30] Z.H. Chohan, C.T. Supuran, A. Scozzafava, *J. Enzyme Inhib. Med. Chem.* 19 (1) (2004) 79–84.
- [31] Z.H. Chohan, M. Praveen, *Appl. Organomet. Chem.* 15 (2001) 617–625.
- [32] B.N. Meyer, N.R. Ferrigni, J.E. Putnam, L.B. Jacobsen, D.E. Nichols, J.L. McLaughlin, *Planta Med.* 45 (1982) 31.
- [33] A.W. Bauer, W.M. Kirby, J.C. Sherris, M. Turck, *Am. J. Clin. Pathol.* 45 (1966) 493.
- [34] G. Singh, P.A. Singh, K. Singh, D.P. Singh, R.N. Handa, S.N. Dubey, *Proc. Natl. Acad. Sci. Ind.* 72A (2002) 87–94.
- [35] W.J. Geary, *Coord. Chem. Rev.* 7 (1971) 81.
- [36] P.R. Shukla, V.K. Singh, A.M. Jaiswal, J. Narain, *J. Indian Chem. Soc.* 60 (1983) 321–324.
- [37] V.J. Tyaga Raju, Vilas Ranbaore, Vasudha Atre, M.C. Ganorkar, *J. Indian Chem. Soc. Vol. (LIX) (Feb. 1982)* 199–203.
- [38] F.D. Lewis, S.V. Barancyk, *J. Am. Chem. Soc.* 111 (1989) 8653–8661.
- [39] M.S. Gunthkal, T.R. Goudar, S.A. Patil, *Oriental J. Chem.* 16 (2000) 151–154.
- [40] Sayaji Rao, *Asian J. Chem.* 17 (2005) 2663–2668.
- [41] A.B.P. Lever, *Inorganic Electronic Spectroscopy*, Elsevier, Amsterdam, 1984.
- [42] S.M.E. Khalil, *Chem. Pap.* 54 (1) (2000) 12–18.
- [43] R.S. Drago, *Physical Methods in Inorganic Chemistry*, Reinhold Publishing Corporation, New York, 1968.

- [44] C.H. Krishna, C.M. Mahapatra, K.A. Dash, *J. Inorg. Nucl. Chem.* 39 (1977) 1253.
- [45] B.K. Rai, Mukesh Kumar, *J. Indian Coun. Chem.* 20 (2003) 22.
- [46] C.J. Balhausen, *Introduction to Ligand Fields*, McGraw Hill, New York, 1962.
- [47] B.K. Patel, M.M. Patel, *Indian J. Chem.* 29 (1990) 90–92.
- [48] R.C. Rossenberg, A.C. Root, P.K. Bernstein, H.B. Gray, *J. Am. Chem. Soc.* 97 (1975) 2092.
- [49] B.T. Hathaway, *Struct. Bonding* 14 (1973) 60.
- [50] A.-J. Bard, L.-R. Izatt (Eds.), *Electrochemical Methods: Fundamentals and Applications*, second ed. Wiley, New York, 2001.
- [51] S. Mukherjee, *Indian J. Chem.* 26 (1987) 1002.
- [52] H. Beens, K.H. Grellmann, M. Gurr, A.H. Weller, *Discuss. Faraday Soc.* 39 (1965) 183.
- [53] A.W. Czarnik, *Chem. Biol.* 2 (1995) 423.
- [54] A.W. Varnes, R.B. Dodson, E.L. Wehry, *J. Am. Chem. Soc.* 94 (1972) 946.
- [55] T. Kaliyappan, S. Rajagopan, P. Kannan, *J. Appl. Polym. Sci.* 91 (2003) 494–500.
- [56] G.S.V. Kumar, B. Mathew, *J. Mac. Sci.* 41 (2004) 1037–1050.
- [57] S.C. Bernadette, A.E. Denise, Kevin Kavanagh, Malachy McCann, Andy Noble, Bhumika Thati, Maureen Walsh, *Inorg. Chim. Acta* 359 (2006) 3976–3984.
- [58] A.L. Vogel, *A Text Book of Quantitative Chemical Analysis*, fifth ed. Addison Wesley Longman, London, 1999.