See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/259108870

Stereoselective facile synthesis of 2 '-spiro pyrimidine pyranonucleosides via their key intermediate 2 '-C-cyano analogues. Evaluation of their bioactivity

ARTICLE in CARBOHYDRATE RESEARCH · NOVEMBER 2013

Impact Factor: 1.93 · DOI: 10.1016/j.carres.2013.11.001 · Source: PubMed

CITATION

1

READS

43

7 AUTHORS, INCLUDING:



Stella Manta

University of Thessaly

49 PUBLICATIONS 384 CITATIONS

SEE PROFILE



Petros Gkizis

Aristotle University of Thessaloniki

13 PUBLICATIONS 62 CITATIONS

SEE PROFILE



Dimitri Komiotis

University of Thessaly

89 PUBLICATIONS 788 CITATIONS

SEE PROFILE

ELSEVIER

Contents lists available at ScienceDirect

Carbohydrate Research

journal homepage: www.elsevier.com/locate/carres



Note

Stereoselective facile synthesis of 2'-spiro pyrimidine pyranonucleosides via their key intermediate 2'-C-cyano analogues. Evaluation of their bioactivity



Christos Kiritsis ^a, Stella Manta ^a, Athina Dimopoulou ^a, Vanessa Parmenopoulou ^a, Petros Gkizis ^b, Ian Balzarini ^c, Dimitri Komiotis ^{a,*}

- ^a Department of Biochemistry and Biotechnology, Laboratory of Bio-Organic Chemistry, University of Thessaly, 26 Ploutonos Str., 41221 Larissa, Greece
- ^b Department of Chemistry, Aristotle University of Thessaloniki, University Campus, 54124 Thessaloniki, Greece
- ^c Rega Institute for Medical Research, KU Leuven, B-3000 Leuven, Belgium

ARTICLE INFO

Article history: Received 27 August 2013 Received in revised form 31 October 2013 Accepted 3 November 2013 Available online 9 November 2013

Keywords: C-Cyano pyranonucleosides Spironucleosides Cytostatic agents 5-Fluorouracil

ABSTRACT

A novel series of 2'-spiro pyrimidine pyranonucleosides has been designed and synthesized. Their precursors, 2'-C-cyano nucleosides **5a,b** and **6a,b**, were obtained by subjecting **1a,b** to the sequence of selective protection of the primary hydroxyl group, acetalation, oxidation, and finally treatment with sodium cyanide. Deoxygenation at the 2'-position of cyanohydrins **5a,b** or **6a,b** led to the 2'-deoxy derivatives **9a,b**. Fully deprotection of **5a,b**, **6a,b**, and **9a,b** gave the desired 2'-C-cyano **7a,b**, **8a,b**, and 2'-C-cyano-2'-deoxy pyranonucleosides **10a,b**, respectively. Mesylation of the corresponding cyanohydrins **5a,b** and **6a,b** afforded compounds **11a,b** and **12a,b** which after base treatment and subsequent deprotection furnished the spiro nucleosides **15a,b** and **16a**. The new analogues were evaluated for their potential cytostatic activities in cell culture.

© 2013 Elsevier Ltd. All rights reserved.

Nucleoside analogues have long played a pivotal role in the treatment of viral infections and cancer.^{1–4} These therapeutic compounds mimic physiological nucleosides in terms of uptake and metabolism and are incorporated into newly synthesized DNA, resulting in synthesis inhibition and DNA chain termination.⁵ However, their clinical use is often limited by important side-effects and natural or acquired drug resistance, thus there is an ongoing medical need for more effective, selective, and non-toxic derivatives.

Nucleosides containing sugar-ring modifications^{6,7} are recognized as an important class of biologically active molecules. Particularly, branched-chain cyano furanonucleosides have drawn much attention from medicinal chemists due to their antiviral and anticancer properties^{8,9} and their potential to provide an access to the synthesis of branched-chain spiro derivatives endowed with potent anti-HIV properties.^{10,11} Spirobicyclic cores display an important role in the development of new bioactive substances,¹² while a number of spiro nucleosides, as conformationally restricted analogues, exhibited interesting antiviral activities.^{13–15}

Pyranosyl nucleosides are undoubtedly one of the most important modifications of natural nucleosides, offering promising avenues in the development of potential antitumor, ^{16,17} antiviral, ^{18,19} antioxidant, ²⁰ and antimicrobial ²¹ agents. In our previous investigations, we demonstrated that insertion of the cyano functional group at the 3′-C- or 4′-C-position of the pyranose ring, leads to derivatives endowed with significant cytostatic activity against different cancer cell lines. ^{22,23}

With the above applications in mind and in order to: (i) better understand the cyano substituent position effect on the properties of this class of analogues and (ii) promote an efficient way to pharmacologically active spiro branched-chain nucleosides, we found it of particular interest to further extend our studies toward the preparation of 2'-C-cyano pyrimidine pyranonucleosides as potential useful synthons for 2'-spiro pyranonucleosides bearing a 4-amino-1,2-oxathiole-2,2-dioxide ring. The chemical synthesis and biological activity of these compounds are presented herein.

Our synthetic approach was focused on the preparation of 2'-C-cyano pyranonucleosides of uracil **5a,6a** and 5-fluorouracil **5b,6b** as key intermediates (Scheme 1). Commercially available 1,2,3,4,6-penta-O-acetyl-D-galactopyranose was condensed with the proper pyrimidine base to give, after deacetylation, the β -galactopyranonucleosides of uracil **1a**²⁴ and 5-fluorouracil **1b**.²⁵ Selective protection of the primary hydroxyl group of **1a** and **1b** with t-butyldimethylsilyl chloride (TBDMSCl) followed by acetalation,

^{*} Corresponding author. Tel.: +30 2410 565285; fax: +30 2410 565290. E-mail address: dkom@bio.uth.gr (D. Komiotis).

Scheme 1. Reagents and conditions: (i) TBDMSCI, pyridine, DMAP, 2a: 85%, 2b: 83%; (ii) acetone, 2,2-dimethoxypropane, *p*-toluenesulfonic acid, 3a: 86%, 3b: 85%; (iii) PDC, Ac₂O, dry CH₂Cl₂, 4a: 80%, 4b: 83%; (iv) H₂O, Et₂O, NaCN, NaHCO₃, 5a: 28% and 6a: 52%, 5b: 25% and 6b: 56%; (v) TFA 90% in MeOH, 7a: 74%, 7b: 74%, 8a: 79%, 8b: 78%.

furnished the partially protected derivatives 3a,b, which upon oxidation²⁶ gave the 2'-ketonucleosides **4a**,**b**. Reaction of **4a**,**b** with sodium cyanide (NaCN),²³ afforded a chromatographically separable mixture of the two possible 2'-cyanohydrin epimers 5a,b and 6a,b in an approximate 1:2 ratio, respectively, contrary to previously published stereospecific preparation of 3'-C- and 4'-C-cyano pyranonucleosides.^{22,23} The proposed structures of compounds **5a,b** and **6a,b** were established by comparison of their ¹H NMR spectra and were assigned as talopyranosyl and galactopyranosyl analogues, respectively, based on the deshielding effect of the adjacent cyano group^{27–29} over H-1' which appeared at lower field in **5a,b** than in **6a.b** ($\Delta \delta = 0.42$ and 0.31 ppm, respectively). This suggests that H-1' and the cyano group in **5a.b** are on the same side of the plane. The stereochemistry of C-2' was further confirmed by NOE measurements performed on pyranosyl analogue 6b, as depicted in Figure 1. The mutual NOE enhancements observed between the free OH group of galactopyranosyl derivative 6b and H-1', show that these protons are on the same side of the ring system. In contrast, no effect was observed between the 2'-OH hydroxyl proton and the H-1' on talopyranosyl analogue **5b**, when either 2'-OH group or H-1' was irradiated. Finally, deprotection of analogues **5a,b** and **6a,b** using trifluoroacetic acid (TFA) gave the desired nucleosides 7a,b and 8a,b, in good yields (74-79%).

OTBDMS

CN O NH

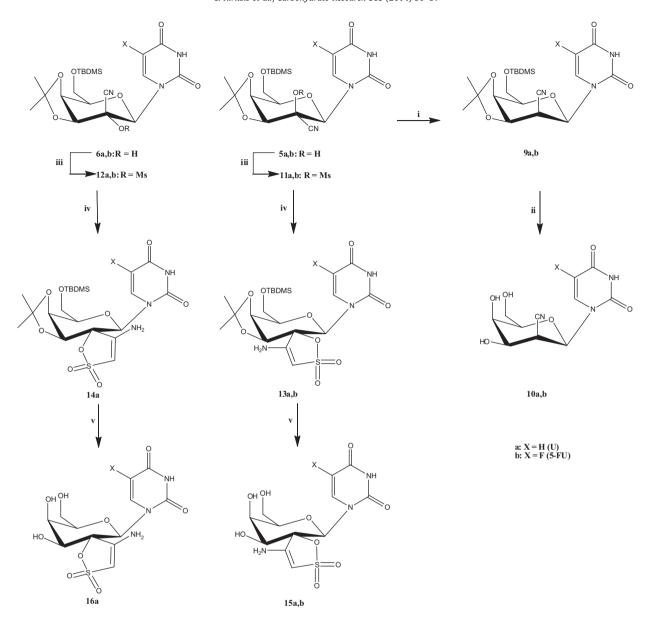
S.0 H 2% H 5.77

7%

Figure 1. NOEs observed for compound 6b.

In an effort to further explore structure–activity relationships of this series of compounds, the synthesis of the 2'-C-cyano-2'-deoxy derivatives **10a,b** was also accomplished (Scheme 2). Phenoxythiocarbonylation of **5a,b** or **6a,b**, followed by deoxygenation with tris(trimethylsilyl)silane [(Me₃Si)₃SiH]³⁰ in the presence of 2,2'-azobis(isobutyronitrile) (AIBN), and subsequent deprotection with TFA led to the formation of the 2'-C-cyano-2'-deoxy derivatives **10a,b**. ¹H NMR data obtained for the newly synthesized nucleosides **10a,b** ($J_{1',2'}$ = 2.6 and 1.7 Hz, respectively) revealed an axial/equatorial relationship between H-1' and H-2', and thus an axial-oriented cyano group. This demonstrated that deoxygenation of 2'-C-cyanohydrins occurs in a stereospecific manner, in accordance with the approach of the hydrogen atom from the less hindered side of the molecule.^{22,23}

Finally, the cyanohydrins 5a,b and 6a,b were used as starting materials for the synthesis of the desired 2'-spiro pyrimidine pyranonucleosides 13-16, as outlined in Scheme 2. Their reaction with mesyl chloride (MsCl) in pyridine furnished the corresponding 2'-C-cyano-2'-O-mesyl derivatives 11a,b and 12a,b. Subsequent treatment of the cyanomesyl nucleosides 11a,b and 12a with 8diazabicycloundec-7-ene (DBU)³¹ in the presence of acetonitrile (CH₃CN) furnished the 2'-spiro derivatives 13a,b and 14a, respectively, in fair to good yields (60-74%), while only decomposition products were recovered upon treatment of 5-fluorouracil analogue 12b either with DBU or cesium carbonate. The cyclization step of spiro derivatives 13a,b and 14a most likely involves abstraction of the acidic proton from the mesylate group, followed by nucleophilic attack of the resulting carbanion at the nitrile carbon atom.^{31,32} The formation of the spiroaminooxathiole dioxide ring in the aforementioned compounds was evident by the disappearance in their ¹H NMR spectra of the characteristic signal at 3.28-3.24 ppm assigned to the mesyl group and the presence of two new singlets at 6.02-5.35 ppm corresponding to NH₂-4" and at 6.04-5.43 ppm assigned to H-3". In agreement, the IR spectra of the newly synthesised compounds 13a,b and 14a, exhibited characteristic absorption bands at 3320-3455 cm⁻¹ assigned to the stretching vibrations of the NH₂ group. In the final step, deprotection of 13a,b and 14a under mild acidic conditions,



Scheme 2. Reagents and conditions: (i) phenyl chlorothionoformate, Et₃N, DMAP, CH₃CN, 0 °C; then (Me₃Si)₃SiH, AlBN, toluene, 100 °C, **9a**: 70% from **5a** or 67% from **6a**, **9b**: 64% from **5b** or 67% from **6b**, (two steps); (ii) TFA 90% in MeOH, **10a**: 74%, **10b**: 79%; (iii) MsCl, pyridine, **11a**: 69%, **11b**: 68%, **12a**: 69%, **12b**: 59%; (iv) DBU, CH₃CN, **13a**: 60%, **13b**: 74%, **14a**: 70%; (v) CH₂Cl₂, methanolic hydrogen chloride, **15a**: 58%, **15b**: 59%, **16a**: 67%.

performed by methanolic hydrogen chloride, ^{33,34} resulted in removal of both silyl and isopropylidene groups to deliver the target nucleosides **15a,b** and **16a**, respectively.

of 2'-C-cyano The cytostatic activity 7a,b, 8a.b. 2'-C-cyano-2'-deoxy 10a,b, 2'-spiro pyranonucleosides 13a,b, 14a and their unprotected derivatives 15a,b, 16a was determined against murine leukemia L1210, human CD4⁺ T-lymphocyte (CEM) and human cervix carcinoma HeLa cell cultures (Table 1). Whereas the uracil derivatives (a-series of compounds) showed rather poor if any cytostatic activity, several 5-fluorouracil derivatives (b-series) showed pronounced antiproliferative activity. In particular, the 2'-C-cyano (up or down)-2'-hydroxy derivatives **7b** and 8b showed a cytostatic activity profile identical to that of the free base 5-fluorouracil (5-FU). The three other 5-FU derivatives, that is, the 2'-C-cyano(up)-2'-deoxynucleoside analogue 10b and the 2'-spironucleoside analogues 13b and 15b were devoid of significant cytostatic activity (Table 1). The pronounced cytostatic activity of 7b and 8b can be due to intrinsic instability of the compounds (spontaneously releasing 5-FU upon cell culture exposure)

 Table 1

 Cytostatic activity of test compounds against tumor cell proliferation in cell culture

Compound	$IC_{50}^{a}(\mu M)$		
	L1210	CEM	HeLa
7a	>250	>250	>250
8a	>250	>250	>250
10a	>250	>250	>250
13a	30 ± 10	22 ± 1	28 ± 12
14a	111 ± 3	110 ± 2	87 ± 2
15a	>250	>250	>250
16a	>250	>250	≥250
7b	0.79 ± 0.45	16 ± 4	0.60 ± 0.04
8b	0.68 ± 0.11	20 ± 6	0.65 ± 0.23
10b	34 ± 1	112 ± 6	22 ± 2
13b	23 ± 2	24 ± 1	16 ± 5
15b	88 ± 13	>250	49 ± 6
F-Uracil	0.33 ± 0.17	18 ± 5	0.54 ± 0.12

^a 50% inhibitory concentration.

and/or to enzymatic cleavage by serum proteins and/or intracellular enzymes. The presence of a TBDMS group at the 6'-position of the molecules (i.e., **13a**, **13b**) seems to provoke an increased cytostatic activity of the compounds.

In conclusion, we have accomplished the stereoselective synthesis of 2'-C-cyano pyranonucleosides bearing 5-fluorouracil and uracil, along with the preparation of their 2'-deoxy derivatives. We further utilized the cyanohydrins as excellent templates for the synthesis of novel 2'-spiro nucleosides, via a convenient strategy, involving the intramolecular cyclocondensation of their corresponding cyanomesylates. The target nucleosides were tested for their inhibitory effects on the proliferation of murine leukemia (L1210), human lymphocyte (CEM), and human cervix carcinoma (HeLa) cell cultures. Several 5-fluorouracil derivatives showed a pronounced cytostatic activity, comparable to the parent compound. 5-FU.

1. Experimental

1.1. General procedure

Melting points were recorded in a Mel-Temp apparatus and are uncorrected. Thin layer chromatography (TLC) was performed on Merck precoated 60F₂₅₄ plates. Reactions were monitored by TLC on silica gel, with detection by UV light (254 nm) or by charring with sulfuric acid. Flash chromatography was performed using silica gel (240-400 mesh, Merck). Microanalyses were performed on a Perkin-Elmer 2400-II Analyzer. ¹H NMR spectra were recorded at 300 MHz on a Bruker AVANCE^{III} 300 spectrometer and ¹³C NMR spectra at 75.5 MHz on the same spectrometer, using chloroform-d (CDCl₃), dimethyl sulfoxide- d_6 (DMSO- d_6). Chemical shifts are reported in parts per million (δ) downfield from tetramethylsilane (TMS) as internal standard. UV-Vis spectra were recorded on a PG T70 UV-Vis spectrometer and mass spectra were obtained on a Thermo Quest Finnigan AQA Mass Spectrometer (electrospray ionization). Optical rotations were measured using an Autopol I polarimeter. Infrared spectra were obtained with a Thermo Scientific Nicolet IR100 FT-IR spectrometer. Dichloromethane (CH₂Cl₂) was distilled from phosphorus pentoxide and stored over 4 Å molecular sieves. CH₃CN and toluene were distilled from calcium hydride and stored over 3 Å molecular sieves. Pyridine was stored over potassium hydroxide pellets. All reactions sensitive to oxygen or moisture were carried out under nitrogen atmosphere using oven-dried glassware.

1.2. Synthesis of 1-(2'-C-cyano- β -D-talopyranosyl)uracil (7a) and 1-(2'-C-cyano- β -D-galactopyranosyl)uracil (8a)

1.2.1. 1-(6'-0-t-Butyldimethylsilyl- β -D-galactopyranosyl)uracil (2a)

To a stirred solution of **1a** (4.11 g, 15.0 mmol) in pyridine (75.0 mL) were added successively TBDMSCl (3.39 g, 22.5 mmol) and a catalytic amount of 4-dimethylaminopyridine (DMAP). The reaction mixture was stirred for 30 min at 0 °C under nitrogen and then at room temperature for 5 h. After, the reaction mixture was quenched with MeOH and evaporated under reduced pressure. The resulting residue was purified by flash chromatography (EtOAc–MeOH, 9:1), to give **2a** (4.97 g, 85%) as a colorless oil. $[\alpha]_D^{22}$ +2 (c 0.2, MeOH); R_f = 0.52 (EtOAc–MeOH, 8:2); UV (MeOH): $\lambda_{\rm max}$ 258 nm (ε 7286); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.07 (br s, 1H, NH), 7.63 (d, 1H, $J_{5,6}$ = 8.1 Hz, H-6), 5.68 (d, 1H, H-5), 5.34 (d, 1H, $J_{1'.2'}$ = 9.1 Hz, H-1'), 5.27 (d, 1H, J = 4.5 Hz, OH), 5.01 (s, 1H, OH), 4.58 (d, 1H, J = 6.1 Hz, OH), 3.76–3.49 (m, 6H, H-2', H-3', H-4', H-5', H-6a',b'), 0.83 (s, 9H, t-Bu), 0.02, 0.01 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 163.3, 151.1, 141.4,

102.2, 83.2, 78.2, 75.9, 74.0, 68.7, 62.7, 26.1, 18.3, -4.9. Anal. Calcd for $C_{16}H_{28}N_2O_7Si$: C, 49.47; H, 7.26; N, 7.21. Found: C, 49.71; H, 7.43; N, 7.02. ESIMS m/z 389.18 [M+H⁺].

1.2.2. 1-(6'-O-t-Butyldimethylsilyl-3',4'-O-isopropylidene- β -D-galactopyranosyl)uracil (3a)

To a stirred suspension of 2a (4.97 g, 12.8 mmol) in anhydrous acetone (226.0 mL) and 2,2-dimethoxypropane (12.7 mL) was added p-toluenesulfonic acid monohydrate (0.46 g, 2.4 mmol). After 3 h the resulting solution was neutralized with triethylamine (Et₃N) so pH did not exceed 7. The solution was concentrated and the residue was purified by flash chromatography (CH₂Cl₂-MeOH, 9.5:0.5), to give **3a** (4.72 g, 86%) as a white solid, mp 205-207 °C. $[\alpha]_D^{22}$ +42 (c 0.6, CDCl₃); R_f = 0.47 (CH₂Cl₂-MeOH, 9:1); UV (CDCl₃): $\lambda_{\rm max}$ 259 nm (ε 4831); ¹H NMR (CDCl₃, 300 MHz): δ 9.43 (br s, 1H, NH), 7.41 (d, 1H, $J_{5,6}$ = 8.2 Hz, H-6), 5.74 (d, 1H, $J_{1',2'}$ = 8.0 Hz, H-1'), 5.57 (d, 1H, H-5), 4.32-4.26 (m, 2H, H-3', H-4'), 4.07 (dt, 1H, $J_{4',5'} = 1.4 \text{ Hz}, \ J_{5',6a'} = J_{5',6b'} = 6.4 \text{ Hz}, \ \text{H--5'}), \ 3.89 - 3.78 \ (\text{m}, \ 2\text{H}, \ \text{H--})$ 6a',b'), 3.72 (dd, 1H, $J_{2',3'}$ = 5.8 Hz, H-2'), 1.53, 1.37 (2s, 6H, 2CH₃), 0.88 (s, 9H, t-Bu), 0.06, 0.05 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 163.1, 151.0, 139.9, 110.1, 102.9, 82.9, 78.9, 76.3, 72.9, 71.9, 61.8, 28.0, 25.8, 18.3, -5.3, -5.4. Anal. Calcd for $C_{19}H_{32}$ N₂O₇Si: C, 53.25; H, 7.53; N, 6.54. Found: C, 53.32; H, 7.43; N, 6.72. ESIMS m/z 429.22 [M+H⁺].

1.2.3. 1-(6'-O-t-Butyldimethylsilyl-3',4'-O-isopropylidene- β -D-lyxo-hexopyranosyl-2'-ulose)uracil (4a)

A mixture of **3a** (4.72 g, 11.0 mmol), pyridinium dichromate (PDC) (4.93 g, 13.1 mmol) and Ac₂O (3.10 mL, 32.7 mmol) was stirred in dry CH₂Cl₂ (110.0 mL) for 6 h, under nitrogen at room temperature. Purification by flash chromatography (EtOAc-hexane, 5:5) gave **4a** (3.75 g, 80%) as a white solid, mp 153–155 °C. [α]_D²² –80 (c 1.0, CDCl₃); R_f = 0.45 (EtOAc-hexane, 6:4); UV (CDCl₃): λ _{max} 257 nm (ε 11296); ¹H NMR (CDCl₃, 300 MHz): δ 8.72 (br s, 1H, NH), 7.10 (d, 1H, $J_{5,6}$ = 8.2 Hz, H-6), 6.21 (s, 1H, H-1'), 5.77 (d, 1H, H-5), 4.71–4.67 (m, 2H, H-3', H-4'), 4.32 (dt, 1H, $J_{4',5'}$ = 0.7 Hz, $J_{5',6a'}$ = $J_{5',6b'}$ = 6.7 Hz, H-5'), 3.89 (d, 2H, H-6a',b'), 1.45, 1.41 (2s, 6H, 2CH₃), 0.89 (s, 9H, t-Bu), 0.08, 0.07 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 198.5, 162.5, 150.2, 141.0, 111.2, 103.0, 82.2, 77.8, 77.1, 75.9, 61.3, 27.4, 25.8, 25.7, 18.3, –5.4, –5.5. Anal. Calcd for C₁₉H₃₀N₂O₇Si: C, 53.50; H, 7.09; N, 6.57. Found: C, 53.69; H, 7.13; N, 6.41. ESIMS m/z 427.21 [M+H⁺].

1.2.4. $1-(6'-O-t-Butyldimethylsilyl-2'-C-cyano-3',4'-O-isopropylidene-\beta-D-talopyranosyl)uracil (5a) and <math>1-(6'-O-t-butyldimethylsilyl-2'-C-cyano-3',4'-O-isopropylidene-\beta-D-galactopyranosyl)uracil (6a)$

A mixture of 4a (3.75 g, 8.80 mmol), H_2O (130.0 mL), Et_2O (260.0 mL), $NaHCO_3$ (1.48 g, 17.6 mmol) and NaCN (430 mg, 8.80 mmol) was stirred vigorously at room temperature for 72 h. The organic phase was separated, and the aqueous phase was washed with Et_2O . The combined ether phases were dried over Na_2SO_4 , filtered, and evaporated to dryness. The residue was purified by flash chromatography (CH_2Cl_2 -EtOAc, 7:3).

The fastest moving fractions afforded **5a** (1.12 g, 28%) as a white foam. $[\alpha]_D^{22} - 18$ (c 0.6, CDCl₃); $R_f = 0.30$ (CH₂Cl₂-EtOAc, 7:3); UV (CDCl₃): λ_{max} 259 nm (ϵ 9500); ^1H NMR (CDCl₃, 300 MHz): δ 9.13 (br s, 1H, NH), 7.52 (d, 1H, $J_{5,6} = 8.2$ Hz, H-6), 6.54 (br s, 1H, 2'-OH), 6.15 (s, 1H, H-1'), 5.72 (d, 1H, H-5), 4.57-4.53 (m, 2H, H-3', H-4'), 4.42 (t, 1H, $J_{5',6a'} = J_{5',6b'} = 6.5$ Hz, H-5'), 3.78 (d, 2H, H-6a',b'), 1.59, 1.38 (2s, 6H, 2CH₃), 0.89 (s, 9H, t-Bu), 0.07, 0.06 (2s, 6H, 2Si-CH₃). 13 C NMR (CDCl₃, 75.5 MHz): δ 162.6, 150.1, 141.5, 116.4, 110.7, 102.2, 79.4, 76.1, 74.0, 71.6, 71.2, 62.6, 26.1, 25.7, 24.8, 18.1, -5.4, -5.6. Anal. Calcd for $C_{20}H_{31}N_3O_7Si$: C, 52.96; H, 6.89; N, 9.26. Found: C, 52.88; H, 6.93; N, 9.17. ESIMS m/z 454.20 [M+H $^+$].

The slowest moving fractions afforded **6a** (2.08 g, 52%) as a white foam. $[\alpha]_D^{22}$ +24 (c 0.5, CDCl₃); R_f = 0.18 (CH₂Cl₂–EtOAc, 7:3); UV (CDCl₃): λ_{max} 258 nm (ε 3883); ¹H NMR (CDCl₃, 300 MHz): δ 10.06 (br s, 1H, NH), 7.45 (d, 1H, $J_{5,6}$ = 8.2 Hz, H-6), 6.15 (br s, 1H, 2′-OH), 5.84 (d, 1H, H-5), 5.73 (s, 1H, H-1′), 4.52 (d, 1H, $J_{3',4'}$ = 5.9 Hz, H-3′), 4.43 (dd, 1H, $J_{4',5'}$ = 2.8 Hz, H-4′), 4.28–4.24 (m, 1H, H-5′), 4.01–3.89 (m, 2H, H-6a′,b′), 1.74, 1.39 (2s, 6H, 2CH₃), 0.89 (s, 9H, t-Bu), 0.08, 0.07 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 162.0, 150.7, 141.6, 116.8, 111.2, 102.4, 80.1, 75.7, 73.7, 71.9, 69.8, 62.1, 26.4, 25.9, 24.3, 18.2, –5.4, –5.6. Anal. Calcd for C₂₀H₃₁N₃O₇Si: C, 52.96; H, 6.89; N, 9.26. Found: C, 52.80; H, 6.96; N, 9.32. ESIMS m/z 454.21 [M+H⁺].

1.2.5. 1-(2'-C-Cyano-β-D-talopyranosyl)uracil (7a)

The protected nucleoside **5a** (0.37 g, 0.82 mmol) was dissolved in 4.1 mL of 90% TFA in MeOH. The solution was stirred for 10 min at room temperature and then concentrated under reduced pressure, in order to remove traces of TFA. The residue was purified by flash chromatography (CH₂Cl₂-MeOH, 9:1), to give 7a (0.18 g, 74%) as a white foam. $[\alpha]_D^{22}$ –6 (*c* 0.1, MeOH); R_f = 0.24 (CH₂Cl₂– MeOH, 8.5:1.5); UV (MeOH): λ_{max} 257 nm (ϵ 5362); IR (KBr, cm⁻¹): 2225 (CN); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.40 (br s, 1H, NH), 7.66 (d, 1H, $I_{5.6}$ = 8.2 Hz, H-6), 7.58 (s, 1H, H-1'), 6.34 (d, 1H, I = 5.2 Hz, OH), 6.24 (s, 1H, OH), 5.63 (d, 1H, H-5), 5.26 (d, 1H, J = 4.3 Hz, OH), 4.71 (t, 1H, J = 4.3 Hz, OH), 4.18–4.93 (m, 4H, H-3', H-4', H-5', H-6a'), 4.42 (dd, 1H, $J_{5',6b'}$ = 5.3 Hz, $J_{6a',6b'}$ = 11.3 Hz, H-6b'); 13 C NMR (DMSO- d_6 , 75.5 MHz): δ 163.1, 150.6, 142.9, 118.9, 101.2, 80.6, 73.6, 73.3, 72.4, 63.4, 56.9. Anal. Calcd for C₁₁H₁₃N₃O₇: C, 44.15; H, 4.38; N, 14.04. Found: C, 44.29; H, 4.31; N, 14.20. ESIMS m/z 300.09 [M+H⁺].

1.2.6. 1-(2'-C-Cyano-β-D-galactopyranosyl)uracil (8a)

Uracil derivative **8a** was synthesized from **6a** by the similar procedure as described for **7a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9:1), to give **8a** (0.36 g, 79%) as a white foam. $|\alpha|_{1}^{22}$ +14 (c 0.3, MeOH); R_f = 0.23 (CH₂Cl₂–MeOH, 8.5:1.5); UV (MeOH): λ_{max} 257 nm (ε 4032); IR (KBr, cm⁻¹): 2250 (CN); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.19 (br s, 1H, NH), 7.69 (d, 1H, $J_{5,6}$ = 8.2 Hz, H-6), 7.08 (s, 1H, H-1'), 5.82 (d, 1H, J = 2.7 Hz, OH), 5.75 (d, 1H, H-5), 5.59 (s, 1H, OH), 4.93 (s, 1H, OH), 4.71 (t, 1H, J = 4.7 Hz, OH), 3.87–3.58 (m, 5H, H-3', H-4', H-5', H-6a',b'); ¹³C NMR (DMSO- d_6 , 75.5 MHz): δ 163.2, 149.9, 140.2, 117.6, 101.5, 80.1, 79.6, 67.3, 66.2, 60.9, 36.8. Anal. Calcd for C₁₁H₁₃N₃O₇: C, 44.15; H, 4.38; N, 14.04. Found: C, 44.49; H, 4.56; N, 13.96. ESIMS m/z 300.10 [M+H⁺].

1.3. Synthesis of 1-(2'-C-cyano-2'-deoxy- β -D-talopyranosyl) uracil (10a)

1.3.1. 1-(6'-O-t-Butyldimethylsilyl-2'-C-cyano-2'-deoxy-3',4'-O-isopropylidene-β-b-talopyranosyl)uracil (9a)

Phenyl chlorothionoformate (0.17 mL, 1.2 mmol) was added to a solution of **5a** or **6a** (0.37 g, 0.82 mmol), DMAP (0.04 g, 0.3 mmol), and Et₃N (0.17 mL, 1.2 mmol) in CH₃CN (12.0 mL) under nitrogen at 0 °C. The mixture was stirred for 1 h and then diluted with AcOEt. The whole was washed with H₂O and the separated organic phase was dried over anhydrous sodium sulfate, filtered and evaporated to dryness. The residue was coevaporated two times with toluene and then was dissolved in toluene (12.0 mL). (Me₃Si)₃SiH (0.34 mL, 1.1 mmol) was added to the above solution containing AIBN (0.08 g, 0.5 mmol) at 90 °C under nitrogen. After being heated for 50 min, the solvent was removed under reduced pressure. The residue was purified by flash chromatography (CH₂Cl₂–MeOH, 9.8:0.2) to give **9a** (0.25 g, 70% from **5a** or 0.24 g, 67% from **6a**) as a colorless oil. $[\alpha]_D^{22}$ +54 (c 0.3, CDCl₃); R_f = 0.33 (CH₂Cl₂–MeOH, 9.8:0.2); UV (CDCl₃): λ_{max} 257 nm (ε 4773); 1 H

NMR (CDCl₃, 300 MHz): δ 8.78 (br s, 1H, NH), 7.23 (d, 1H, $J_{5,6}$ = 8.3 Hz, H-6), 5.79 (d, 1H, H-5), 5.55 (d, 1H, $J_{1',2'}$ = 2.7 Hz, H-1'), 4.57 (dd, 1H, $J_{2',3'}$ = 7.1 Hz, $J_{3',4'}$ = 5.7 Hz, H-3'), 4.29 (dd, 1H, $J_{4',5'}$ = 2.9 Hz, H-4'), 4.11–3.89 (m, 3H, H-5',H-6a',b'), 3.65 (dd, 1H, H-2'), 1.74, 1.39 (2s, 6H, 2CH₃), 0.89 (s, 9H, t-Bu), 0.10, 0.09 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 161.6, 150.7, 141.0, 116.8, 110.9, 102.7, 80.1, 75.7, 73.4, 71.3, 62.6, 36.8, 26.1, 25.6, 24.8, 18.2, -5.4, -5.6. Anal. Calcd for $C_{20}H_{31}N_3O_6Si$: C, 54.90; H, 7.14; N, 9.60. Found: C, 54.78; H, 7.11; N, 9.82. ESIMS m/z 438.22 [M+H⁺].

1.3.2. 1-(2'-C-Cyano-2'-deoxy-β-D-talopyranosyl)uracil (10a)

Uracil derivative **10a** was synthesized from **9a** by the similar procedure as described for **7a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9:1), to give **10a** (0.12 g, 74%) as a white foam. $[\alpha]_D^{2D}$ +6 (c 0.2, MeOH); R_f = 0.27 (CH₂Cl₂–MeOH, 8.5:1.5); UV (MeOH): λ_{max} 258 nm (ε 6229); IR (KBr, cm⁻¹): 2250 (CN); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.48 (br s, 1H, NH), 7.70 (d, 1H, $J_{5,6}$ = 8.2 Hz, H-6), 5.74–5.69 (m, 2H, H-5, OH), 5.66 (d, 1H, $J_{1',2'}$ = 2.6 Hz, H-1'), 4.86 (d, 1H, J = 2.8 Hz, OH), 4.76 (t, 1H, J = 5.3 Hz, OH), 4.01–3.65 (m, 5H, H-3', H-4', H-5', H-6a',b'), 3.47 (dd, 1H, $J_{2',3'}$ = 5.3 Hz, H-2'); ¹³C NMR (DMSO- d_6 , 75.5 MHz): δ 163.2, 149.9, 140.2, 117.6, 101.5, 80.0, 79.6, 67.3, 66.2, 60.9, 36.9. Anal. Calcd for C₁₁H₁₃N₃O₆: C, 46.65; H, 4.63; N, 14.84. Found: C, 46.78; H, 4.58; N, 14.76. ESIMS m/z 284.09 [M+H⁺].

1.4. Synthesis of $[1-(\beta-D-talopyranosyl)uracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (15a) and <math>[1-(\beta-D-talopyranosyl)uracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (16a)$

1.4.1. 1-(6'-*O*-*t*-Butyldimethylsilyl-2'-*C*-cyano-3',4'-*O*-isopropylidene-2'-*O*-mesyl-β-D-talopyranosyl)uracil (11a)

The cyanohydrin 5a (0.37 g, 0.82 mmol) was dissolved in dry pyridine (4.0 mL) and mesyl chloride (0.32 mL, 4.1 mmol) was added. The mixture was stirred at 5-8 °C for 48 h. poured into ice and water, and extracted with chloroform $(2 \times 15 \text{ mL})$. The combined extracts were washed with 1 N HCl (15 mL), aqueous sodium hydrogen carbonate (15 mL), and brine (15 mL), dried over anhydrous sodium sulfate, filtered, and evaporated to dryness. The residue was purified by flash chromatography (CH₂Cl₂-MeOH, 9.8:0.2) to give **11a** (0.30 g, 69%) as a colorless oil. $[\alpha]_D^{22}$ -3 (c 0.2, CDCl₃); $R_f = 0.52$ (CH₂Cl₂-MeOH, 9.5:0.5); UV (CDCl₃): λ_{max} 256 nm (ε 9053); ¹H NMR (CDCl₃, 300 MHz): δ 8.47 (br s, 1H, NH), 7.49 (d, 1H, $J_{5.6}$ = 8.3 Hz, H-6), 6.65 (s, 1H, H-1'), 5.80 (d, 1H, H-5), 5.03 (d, 1H, $J_{3',4'}$ = 6.9 Hz, H-3'), 4.65 (dd, 1H, $J_{4',5'}$ = 1.4 Hz, H-4'), 4.33 (dt, 1H, $J_{5',6a'} = J_{5',6b'} = 6.5$ Hz, H-5'), 3.83-3.79 (m, 2H, H-6a',b'), 3.26 (s, 3H, CH₃SO₂), 1.69, 1.41 (2s, 6H, 2CH₃), 0.89 (s, 9H, t-Bu), 0.07, 0.06 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 161.6, 150.6, 138.6, 112.4, 112.3, 103.1, 81.5, 79.8, 79.0, 78.4, 72.6, 61.4, 40.3, 25.8, 25.7, 25.1, 18.3, -5.3, -5.4. Anal. Calcd for C₂₁H₃₃N₃O₉SSi: C, 47.44; H, 6.26; N, 7.90. Found: C, 47.32; H, 6.30; N, 7.97. ESIMS m/z 532.16 [M+H⁺].

1.4.2. 1-(6'-0-t-Butyldimethylsilyl-2'-C-cyano-3',4'-0-isopropylidene-2'-0-mesyl-β-p-galactopyranosyl)uracil (12a)

The cyanomesylate **12a** was synthesized from **6a** by the similar procedure as described for **11a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9.8:0.2) to give **12a** (0.56 g, 69%) as a colorless oil. $[\alpha]_D^{12} + 26$ (c 0.5, CDCl₃); $R_f = 0.45$ (CH₂Cl₂–MeOH, 9.5:0.5); UV (CDCl₃): λ_{max} 256 nm (ε 9647); ¹H NMR (CDCl₃, 300 MHz): δ 8.64 (br s, 1H, NH), 7.68 (d, 1H, $J_{5,6} = 8.3$ Hz, H-6), 5.92 (s, 1H, H-1'), 5.83 (d, 1H, H-5), 4.79 (d, 1H, $J_{3',4'} = 6.1$ Hz, H-3'), 4.50 (dd, 1H, $J_{4',5'} = 2.7$ Hz, H-4'), 4.24 (dt, 1H, $J_{5',6a'} = J_{5',6b'} = 6.4$ Hz, H-5'), 4.01–3.89 (m, 2H, H-6a',b'), 3.24 (s, 3H, CH₃SO₂), 1.77, 1.42 (2s,

6H, 2CH₃), 0.89 (s, 9H, *t*-Bu), 0.08, 0.07 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 161.7, 150.3, 138.9, 112.9, 112.4, 103.4, 81.8, 79.9, 78.7, 78.6, 72.5, 61.3, 40.2, 25.9, 25.8, 25.0, 18.3, -5.3, -5.4. Anal. Calcd for C₂₁H₃₃N₃O₉SSi: C, 47.44; H, 6.26; N, 7.90. Found: C, 47.51; H, 6.19; N, 7.82. ESIMS m/z 532.18 [M+H⁺].

1.4.3. [1-(6'-O-t-Butyldimethylsilyl-3',4'-O-isopropylidene-β-D-talopyranosyl)uracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (13a)

A solution of **11a** (0.3 g, 0.6 mmol) in dry acetonitrile (6.0 mL) was treated with DBU (0.09 mL, 0.6 mmol), and the mixture was stirred at room temperature for 36 h. The solution was neutralized with acetic acid and the solvent was evaporated to dryness. The residue was purified by flash chromatography (CH2Cl2-MeOH, 9.5:0.5) to give **13a** (0.18 g, 60%) as a white foam. $[\alpha]_D^{22}$ -6 (c 0.4, CDCl₃); $R_f = 0.38$ (CH₂Cl₂-MeOH, 9.5:0.5); UV (CDCl₃): λ_{max} 253 nm (ε 3429); IR (KBr, cm⁻¹): 3420, 3320 (NH₂), 1665 (C=C-N); ¹H NMR (CDCl₃, 300 MHz): δ 10.29 (br s, 1H, NH), 7.56 (d, 1H, $J_{5.6}$ = 8.2 Hz, H-6), 6.49 (s, 1H, H-1'), 6.02 (br s, 2H, NH₂-4"), 5.79 (d, 1H, H-5), 5.43 (s, 1H, H-3"), 4.65 (d, 1H, $J_{3',4'}$ = 7.5 Hz, H-3'), 4.48 (d, 1H, H-4'), 4.35 (t, 1H, $J_{5',6a'} = J_{5',6b'} = 4.4$ Hz, H-5'), 3.84 (d, 2H, H-6a',b'), 1.57, 1.37 (2s, 6H, 2CH₃), 0.92 (s, 9H, t-Bu), 0.10, 0.08 (2s, 6H, 2Si-CH₃); 13 C NMR (CDCl₃, 75.5 MHz): δ 163.9, 153.4, 151.8, 140.4, 112.5, 103.5, 89.1, 83.0, 79.9, 77.3, 75.7, 73.1, 62.5, 25.9, 25.6, 24.6, 17.9, -5.5, -5.8. Anal. Calcd for C₂₁H₃₃N₃O₉₋ SSi: C, 47.44; H, 6.26; N, 7.90. Found: C, 47.62; H, 6.34; N, 7.84. ESIMS m/z 532.20 [M+H⁺].

1.4.4. [1-(6'-O-t-Butyldimethylsilyl-3',4'-O-isopropylidene-β-D-galactopyranosyl)uracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (14a)

Spiro derivative **14a** was synthesized from **12a** by the similar procedure as described for **13a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9.5:0.5) to give **14a** (0.39 g, 70%) as a white foam. $[\alpha]_0^{22}$ +6 (c 0.5, CDCl₃); R_f = 0.37 (CH₂Cl₂–MeOH, 9.5:0.5); UV (CDCl₃): λ_{max} 257 nm (ε 8611); IR (KBr, cm⁻¹): 3455, 3360 (NH₂), 1650 (C=C-N); ¹H NMR (CDCl₃, 300 MHz): δ 10.94 (br s, 1H, NH), 7.88 (d, 1H, $J_{5,6}$ = 8.1 Hz, H-6), 6.27 (s, 1H, H-1′), 6.04 (s, 1H, H-3″), 5.43 (d, 1H, H-5), 5.35 (br s, 2H, NH₂–4″), 4.61–4.52 (m, 2H, H-3′,H-4′), 4.43 (t, 1H, $J_{5',6a'}$ = $J_{5',6b'}$ = 6.1 Hz, H-5′), 3.91–3.81 (m, 2H, H-6a′,b′), 1.59, 1.41 (2s, 6H, 2CH₃), 0.91 (s, 9H, t-Bu), 0.09 (s, 6H, 2Si-CH₃); ¹³C NMR (CDCl₃, 75.5 MHz): δ 164.7, 151.6, 150.3, 141.4, 110.6, 100.5, 92.9, 83.2, 79.3, 75.7, 73.3, 71.4, 61.8, 26.2, 25.7, 24.9, 18.1, -5.5, -5.6. Anal. Calcd for C₂₁H₃₃N₃O₉SSi: C, 47.44; H, 6.26; N, 7.90. Found: C, 47.52; H, 6.39; N, 7.78. ESIMS m/z 532.19 [M+H⁺].

1.4.5. $[1-(\beta-D-Talopyranosyl)uracil]-2'-spiro-5''-(4''-amino-1'',2''-oxathiole-2'',2''-dioxide)$ (15a)

To a solution of **13a** (0.17 g, 0.32 mmol) in dry CH₂Cl₂ (1.0 mL) was added methanolic hydrogen chloride (2.5 mL), which was obtained from acetyl chloride (0.2 mL) in dry MeOH (5.0 mL) at 0 °C. After completion of the reaction (6 h), the mixture was diluted with MeOH, and treated with an excess of NaHCO₃. The resulting mixture was filtered through Celite and the solvent was evaporated to dryness. The residue was purified by flash chromatography (CH₂Cl₂-MeOH, 8:2) to give **15a** (0.07 g, 58%) as a white foam. $[\alpha]_D^{22}$ –26 (c 0.1, DMSO); R_f = 0.22 (CH₂Cl₂-MeOH, 8:2); UV (DMSO): λ_{max} 260 nm (ε 10206); IR (KBr, cm⁻¹): 3420, 3330 (NH₂), 1655 (C=C-N); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.23 (br s, 1H, NH), 7.50 (d, 1H, $J_{5,6}$ = 8.2 Hz, H-6), 6.67 (br s, 2H, NH₂-4"), 6.34 (s, 1H, H-1'), 6.18 (br s, 2H, 2OH), 5.58 (d, 1H, H-5), 5.46 (s, 1H, H-3"), 4.72 (br s, 1H, OH), 4.13-3.59 (m, 5H, H-3', H-4', H-5', H-6a',b'); 13 C NMR (DMSO- d_6 , 75.5 MHz); δ 163.2, 154.4, 150.5, 142.4, 101.6, 91.3, 86.4, 79.5, 75.4, 73.3, 66.1, 58.4. Anal. Calcd for $C_{12}H_{15}N_3O_9S$: C, 38.20; H, 4.01; N, 11.14. Found: C, 38.29; H, 3.92; N, 11.19. ESIMS m/z 378.08 [M+H $^+$].

1.4.6. [1-(β -D-Galactopyranosyl)uracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (16a)

Spiro derivative **16a** was synthesized from **14a** by the similar procedure as described for **15a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 8:2) to give **16a** (0.18 g, 67%) as a white foam. $[\alpha]_2^{12}$ +36 (c 0.1, DMSO); R_f = 0.21 (CH₂Cl₂–MeOH, 8:2); UV (DMSO): λ_{max} 260 nm (ε 8426); IR (KBr, cm⁻¹): 3400, 3380 (NH₂), 1620 (C=C-N); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.19 (br s, 1H, NH), 7.65 (d, 1H, $J_{5,6}$ = 8.2 Hz, H-6), 7.26 (br s, 2H, NH₂–4″), 6.53 (br s, 1H, OH), 5.97 (s, 1H, H-1′), 5.92 (br s, 1H, OH), 5.58 (m, 2H, H-5, H-3″), 4.73 (br s, 1H, OH), 4.21–3.61 (m, 5H, H-3′, H-4′, H-5′, H-6a′,b′); ¹³C NMR (DMSO- d_6 , 75.5 MHz): δ 162.9, 152.8, 150.7, 140.8, 101.8, 92.2, 91.4, 82.2, 81.2, 72.3, 68.7, 60.6. Anal. Calcd for $C_{12}H_{15}N_3O_9S$: C, 38.20; H, 4.01; N, 11.14. Found: C, 38.46; H, 4.11; N, 11.25. ESIMS m/z 378.06 [M+H⁺].

1.5. Synthesis of 1-(2'-C-cyano- β -D-talopyranosyl)-5-fluorouracil (7b) and 1-(2'-C-cyano- β -D-galactopyranosyl)-5-fluorouracil (8b)

1.5.1. 1-(6'-O-t-Butyldimethylsilyl- β -D-galactopyranosyl)-5-fluorouracil (2b)

5-Fluorouracil derivative **2b** was synthesized from **1b** by the similar procedure as described for **1a**. It was purified by flash chromatography (EtOAc–MeOH, 9:1) to give **2b** (5.06 g, 83%) as a white solid, mp 136–138 °C. $[\alpha]_D^{22}$ +7 (c 0.5, MeOH); R_f = 0.60 (EtOAc–MeOH, 8:2); UV (MeOH): λ_{max} 267 nm (ε 4579); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.75 (br s, 1H, NH), 8.01 (d, 1H, $J_{6,F5}$ = 7.1 Hz, H-6), 5.33 (d, 1H, $J_{1',2'}$ = 9.0 Hz, H-1'), 5.18 (d, 1H, J = 2.9 Hz, OH), 4.79 (s, 1H, OH), 4.35 (d, 1H, J = 7.4 Hz, OH), 3.82–3.51 (m, 6H, H-2', H-3', H-4', H-5', H-6a',b'), 0.86 (s, 9H, t-Bu), 0.03, 0.02 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 157.4, 149.9, 140.5, 126.0, 83.6, 78.3, 73.7, 68.9, 68.8, 62.6, 26.2, 18.5, -4.8. Anal. Calcd for $C_{16}H_{27}FN_2O_7Si$: C, 47.28; H, 6.70; N, 6.89. Found: C, 47.34; H, 6.82; N, 6.96. ESIMS m/z 407.15 [M+H+].

1.5.2. 1-(6'-O-t-Butyldimethylsilyl-3',4'-O-isopropylidene- β -D-galactopyranosyl)-5-fluorouracil (3b)

5-Fluorouracil derivative **3b** was synthesized from **2b** by the similar procedure as described for **2a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9.5:0.5) to give **3b** (4.70 g, 85%) as a white solid, mp 94–96 °C. [α]_D²² +20 (c 0.5, CDCl₃); R_f = 0.50 (CH₂Cl₂–MeOH, 9:1); UV (CDCl₃): λ_{max} 267 nm (ε 4452); ¹H NMR (CDCl₃, 300 MHz): δ 9.15 (br s, 1H, NH), 7.51 (d, 1H, $J_{6,F5}$ = 5.9 Hz, H-6), 5.57 (d, 1H, $J_{1',2'}$ = 7.9 Hz, H-1'), 4.33–4.26 (m, 2H, H-3', H-4'), 4.07 (dt, 1H, $J_{4',5'}$ = 2.1 Hz, $J_{5',6a'}$ = $J_{5',6b'}$ = 6.3 Hz, H-5'), 3.92–3.80 (m, 2H, H-6a',b'), 3.74 (dd, 1H, $J_{2',3'}$ = 6.1 Hz, H-2'), 1.56, 1.38 (2s, 6H, 2CH₃), 0.90 (s, 9H, t-Bu), 0.08, 0.07 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 156.9, 149.7, 140.7, 124.3, 110.2, 83.2, 78.5, 76.2, 72.8, 71.8, 61.8, 27.9, 25.8, 18.3, –5.3, –5.4. Anal. Calcd for $C_{19}H_{31}FN_2O_7Si$: C, 51.10; H, 7.00; N, 6.27. Found: C, 50.92; H, 7.17; N, 6.21. ESIMS m/z 447.21 [M+H⁺].

1.5.3. 1-(6'-O-t-Butyldimethylsilyl-3',4'-O-isopropylidene-β-D-lyxo-hexopyranosyl-2'-ulose)-5-fluorouracil (4b)

5-Fluorouracil derivative **4b** was synthesized from **3b** by the similar procedure as described for **4a**. It was purified by flash chromatography (EtOAc–hexane, 5:5) to give **4b** (3.87 g, 83%) as a white solid, mp 155–157 °C. $[\alpha]_D^{22}$ –2 (c 0.3, CDCl₃); R_f = 0.50 (EtOAc–hexane, 6:4); UV (CDCl₃): λ_{max} 264 nm (ε 13813); ¹H NMR (CDCl₃, 300 MHz): δ 8.46 (br s, 1H, NH), 7.19 (d, 1H,

 $J_{6,F5}$ = 5.7 Hz, H-6), 6.15 (s, 1H, H-1'), 4.71–4.68 (m, 2H, H-3', H-4'), 4.31 (t, 1H, $J_{5',6a'}$ = $J_{5',6b'}$ = 6.7 Hz, H-5'), 3.89 (d, 2H, H-6a',b'), 1.48, 1.42 (2s, 6H, 2CH₃), 0.91 (s, 9H, *t*-Bu), 0.09 (s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 198.0, 156.2, 148.9, 140.6, 128.6, 111.4, 82.7, 77.7, 76.4, 76.3, 61.4, 27.3, 26.0, 25.8, 18.3, -5.4, -5.5. Anal. Calcd for C₁₉H₂₉FN₂O₇Si: C, 51.34; H, 6.58; N, 6.30. Found: C, 51.42; H, 6.72; N, 6.22. ESIMS m/z 445.20 [M+H⁺].

1.5.4. 1-(6'-O-t-Butyldimethylsilyl-2'-C-cyano-3',4'-O-isopropylidene- β -D-talopyranosyl)-5-fluorouracil (5b) and 1-(6'-O-t-butyldimethylsilyl-2'-C-cyano-3',4'-O-isopropylidene- β -D-galactopyranosyl)-5-fluorouracil (6b)

The cyanohydrins **5b** and **6b** were synthesized from **4b** by the similar procedure as described for **5a** and **6a**. The residue was purified by flash chromatography (CH₂Cl₂–EtOAc, 7:3).

The fastest moving fractions afforded **5b** (1.03 g, 25%) as a white foam. $[\alpha]_D^{22}$ +2 (c 0.6, CDCl₃); R_f = 0.41 (CH₂Cl₂-EtOAc, 7:3); UV (CDCl₃): λ_{max} 261 nm (ε 7775); ¹H NMR (CDCl₃, 300 MHz): δ 8.72 (br s, 1H, NH), 7.55 (d, 1H, $J_{6,F5}$ = 5.3 Hz, H-6), 6.08 (s, 1H, H-1'), 5.78 (br s, 1H, 2'-OH), 4.56-4.48 (m, 2H, H-3', H-4'), 4.42 (dt, 1H, $J_{4',5'}$ = 1.9 Hz, $J_{5',6a'}$ = $J_{5',6b'}$ = 6.5 Hz, H-5'), 3.81-3.79 (m, 2H, H-6a',b'), 1.62, 1.39 (2s, 6H, 2CH₃), 0.90 (s, 9H, t-Bu), 0.08, 0.07 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 157.4, 148.7, 139.8, 127.0, 116.5, 111.5, 79.2, 75.6, 73.6, 71.5, 70.6, 62.1, 26.4, 25.8, 24.1, 18.3, -5.4, -5.6. Anal. Calcd for C₂₀H₃₀FN₃O₇Si: C, 50.94; H, 6.41; N, 8.91. Found: C, 50.81; H, 6.52; N, 8.85. ESIMS m/z 472.19 [M+H⁺].

The slowest moving fractions afforded **6b** (2.29 g, 56%) as a white foam. $[\alpha]_D^{22}$ +7 (c 0.2, CDCl₃); R_f = 0.27 (CH₂Cl₂–EtOAc, 7:3); UV (CDCl₃): λ_{max} 263 nm (ε 13813); ¹H NMR (CDCl₃, 300 MHz): δ 9.47 (br s, 1H, NH), 7.85 (d, 1H, $J_{6,F5}$ = 6.1 Hz, H-6), 5.77 (s, 1H, H-1'), 5.00 (br s, 1H, 2'-OH), 4.52 (d, 1H, $J_{3',4'}$ = 6.3 Hz, H-3'), 4.43 (dd, 1H, $J_{4',5'}$ = 2.6 Hz, H-4'), 4.22–4.18 (m, 1H, H-5'), 4.02–3.88 (m, 2H, H-6a',b'), 1.73, 1.41 (2s, 6H, 2CH₃), 0.91 (s, 9H, t-Bu), 0.10, 0.09 (2s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 157.3, 149.0, 139.9, 126.3, 116.7, 111.6, 79.3, 75.3, 73.7, 71.2, 70.8, 62.3, 26.5, 25.4, 24.6, 18.2, -5.4, -5.6. Anal. Calcd for C₂₀H₃₀FN₃O₇Si: C, 50.94; H, 6.41; N, 8.91. Found: C, 51.03; H, 6.44; N, 8.97. ESIMS m/z 472.21 [M+H⁺].

1.5.5. 1-(2'-C-Cyano-β-D-talopyranosyl)-5-fluorouracil (7b)

5-Fluorouracil derivative 7b was synthesized from 5b by the similar procedure as described for 7a. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9:1) to give 7b (0.17 g, 74%) as a white foam. [α]₂²² -2 (c 0.4, MeOH); R_f = 0.29 (CH₂Cl₂–MeOH, 8.5:1.5); UV (MeOH): $\lambda_{\rm max}$ 266 nm (ε 3192); IR (KBr, cm⁻¹): 2240 (CN); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.98 (br s, 1H, NH), 7.80 (d, 1H, $J_{6,F5}$ = 7.2 Hz, H-6), 7.62 (s, 1H, H-1'), 6.36 (d, 1H, J = 5.6 Hz, OH), 6.21 (s, 1H, OH), 5.28 (d, 1H, J = 4.8 Hz, OH), 4.72 (t, 1H, J = 5.2 Hz, OH), 4.07–3.92 (m, 4H, H-3', H-4', H-5', H-6a'), 3.64 (dd, 1H, $J_{5',6b'}$ = 5.7 Hz, $J_{6a',6b'}$ = 12.8 Hz, H-6b'); ¹³C NMR (DMSO- d_6 , 75.5 MHz): δ 157.3, 149.6, 140.8, 124.7, 118.6, 82.7, 80.4, 73.9, 72.6, 68.2, 60.6. Anal. Calcd for C₁₁H₁₂FN₃O₇: C, 41.65; H, 3.81; N, 13.25. Found: C, 41.54; H, 3.76; N, 13.35. ESIMS m/z 318.08 [M+H⁺].

1.5.6. 1-(2'-C-Cyano-β-D-galactopyranosyl)-5-fluorouracil (8b)

5-Fluorouracil derivative **8b** was synthesized from **6b** by the similar procedure as described for **7a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9:1) to give **8b** (0.40 g, 78%) as a white foam. [α]_D²² +8 (c 0.2, MeOH); R_f = 0.29 (CH₂Cl₂–MeOH, 8.5:1.5); UV (MeOH): λ _{max} 266 nm (ε 6595); IR (KBr, cm⁻¹): 2235 (CN); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.89 (br s, 1H, NH), 7.81 (d, 1H, $J_{6,F5}$ = 7.0 Hz, H-6), 7.04 (s, 1H, H-1′), 5.74 (s, 1H, OH), 5.60 (s, 1H, OH), 4.85 (s, 1H, OH), 4.66 (s, 1H, OH), 3.91–3.61

(m, 5H, H-3', H-4', H-5', H-6a',b'); 13 C NMR (DMSO- d_6 , 75.5 MHz): δ 159.3, 150.4, 140.7, 128.8, 117.9, 101.2, 81.1, 76.4, 68.3, 66.8, 61.2. Anal. Calcd for $C_{11}H_{12}FN_3O_7$: C, 41.65; H, 3.81; N, 13.25. Found: C, 41.62; H, 3.90; N, 13.39. ESIMS m/z 318.10 [M+H $^+$].

1.6. Synthesis of 1-(2'-C-cyano-2'-deoxy- β -D-talopyranosyl)-5-fluorouracil (10b)

1.6.1. 1-(6'-O-t-Butyldimethylsilyl-2'-C-cyano-2'-deoxy-3',4'-O-isopropylidene-β-D-talopyranosyl)-5-fluorouracil (9b)

5-Fluorouracil derivative **9b** was synthesized from **5b** or **6b** by the similar procedure as described for **9a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9.8:0.2) to give **9b** (0.21 g, 64% from **5b** or 0.22 g, 67% from **6b**) as a colorless oil. $|\alpha|_D^{22} - 2$ (c 0.2, CDCl₃); $R_f = 0.36$ (CH₂Cl₂–MeOH, 9.8:0.2); UV (CDCl₃): λ_{max} 264 nm (ε 4259); ¹H NMR (CDCl₃, 300 MHz): δ 9.01 (br s, 1H, NH), 7.83 (d, 1H, $J_{6,F5} = 5.9$ Hz, H-6), 5.59 (d, 1H, $J_{1',2'} = 1.8$ Hz, H-1'), 4.59 (t, 1H, $J_{2',3'} = J_{3',4'} = 6.5$ Hz, H-3'), 4.30 (dd, 1H, $J_{4',5'} = 2.4$ Hz, H-4'), 4.10–3.89 (m, 3H, H-5',H-6a',b'), 3.67 (dd, 1H, H-2'), 1.74, 1.39 (2s, 6H, 2CH₃), 0.90 (s, 9H, t-Bu), 0.10 (s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 157.3, 148.9, 141.0, 127.2, 116.5, 111.3, 79.4, 75.8, 73.5, 71.6, 62.1, 36.9, 26.2, 25.3, 24.9, 18.2, -5.4, -5.6. Anal. Calcd for $C_{20}H_{30}FN_{30}GSi$: C, 52.73; H, 6.64; N, 9.22. Found: C, 52.60; H, 6.72; N, 9.33. ESIMS m/z 456.22 [M+H⁺].

1.6.2. 1-(2'-C-Cyano-2'-deoxy-β-D-talopyranosyl)-5-fluorouracil (10b)

5-Fluorouracil derivative **10b** was synthesized from **9b** by the similar procedure as described for **10a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9:1) to give **10b** (0.11 g, 79%) as a white foam. [α]_D²² –40 (c 0.2, MeOH); R_f = 0.33 (CH₂Cl₂–MeOH, 8.5:1.5); UV (MeOH): $\lambda_{\rm max}$ 264 nm (ϵ 7366); IR (KBr, cm⁻¹): 2220 (CN); ¹H NMR (DMSO- d_6 , 300 MHz): δ 12.15 (br s, 1H, NH), 7.92 (d, 1H, $J_{6,F5}$ = 7.0 Hz, H-6), 5.77 (br s, 1H, OH), 5.66 (d, 1H, $J_{1',2'}$ = 1.7 Hz, H-1'), 4.96 (d, 1H, J = 3.9 Hz, OH), 4.85 (t, 1H, J = 5.5 Hz, OH), 3.99–3.61 (m, 5H, H-3', H-4', H-5', H-6a',b'), 3.49 (dd, 1H, $J_{2',3'}$ = 5.3 Hz, H-2'); ¹³C NMR (DMSO- d_6 , 75.5 MHz): δ 158.4, 149.8, 140.3, 127.9, 118.9, 81.6, 76.7, 75.0, 73.2, 65.1, 37.8. Anal. Calcd for C₁₁H₁₂FN₃O₆: C, 43.86; H, 4.02; N, 13.95. Found: C, 43.94; H, 4.13; N, 13.73. ESIMS m/z 302.10 [M+H⁺].

1.7. Synthesis of [1-(β -D-talopyranosyl)-5-fluorouracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (15b)

1.7.1. 1-(6'-O-t-Butyldimethylsilyl-2'-C-cyano-3',4'-O-isopropylidene-2'-O-mesyl- β -D-talopyranosyl)-5-fluorouracil (11b)

The cyanomesylate **11b** was synthesized from **5b** by the similar procedure as described for **11a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9.8:0.2) to give **11b** (0.27 g, 68%) as a colorless oil. $[\alpha]_{2}^{12}$ +2 (c 0.2, CDCl₃); R_f = 0.54 (CH₂Cl₂–MeOH, 9.5:0.5); UV (CDCl₃): λ_{max} 263 nm (ϵ 8012); ¹H NMR (CDCl₃, 300 MHz): δ 8.16 (br s, 1H, NH), 7.53 (d, 1H, $J_{6,F5}$ = 6.2 Hz, H-6), 6.62 (s, 1H, H-1'), 5.05 (d, 1H, $J_{3',4'}$ = 6.8 Hz, H-3'), 4.66 (dd, 1H, $J_{4',5'}$ = 1.4 Hz, H-4'), 4.34 (dt, 1H, $J_{5',6a'}$ = $J_{5',6b'}$ = 6.2 Hz, H-5'), 3.84–3.82 (m, 2H, H-6a',b'), 3.28 (s, 3H, CH₃SO₂), 1.69, 1.42 (2s, 6H, 2CH₃), 0.91 (s, 9H, t-Bu), 0.09 (s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 156.4, 149.2, 140.7, 123.6, 112.9, 112.3, 81.1, 80.2, 78.8, 78.4, 72.7, 61.4, 39.9, 25.8, 25.7, 24.9, 18.4, -5.3, -5.4. Anal. Calcd for C₂₁H₃₂FN₃O₉SSi: C, 45.89; H, 5.87; N, 7.64. Found: C, 45.71; H, 5.96; N, 7.54. ESIMS m/z 550.17 [M+H⁺].

1.7.2. 1-(6'-O-t-Butyldimethylsilyl-2'-C-cyano-3',4'-O-isopropylidene-2'-O-mesyl- β -D-galactopyranosyl)-5-fluorouracil (12b)

The cyanomesylate **12b** was synthesized from **6b** by the similar procedure as described for **11a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9.8:0.2) to give **12b** (0.52 g, 59%) as a colorless oil. $[\alpha]_D^{12}$ +12 (c 0.3, CDCl₃); R_f = 0.48 (CH₂Cl₂–MeOH, 9.5:0.5); UV (CDCl₃): $\lambda_{\rm max}$ 262 nm (ϵ 3678); ¹H NMR (CDCl₃, 300 MHz): δ 8.59 (br s, 1H, NH), 7.75 (d, 1H, $J_{6,F5}$ = 5.9 Hz, H-6), 5.93 (s, 1H, H-1'), 4.81 (d, 1H, $J_{3',4'}$ = 6.1 Hz, H-3'), 4.50 (dd, 1H, $J_{4',5'}$ = 2.3 Hz, H-4'), 4.23–4.18 (m, 1H, H-5'), 4.02–3.89 (m, 2H, H-6a',b'), 3.26 (s, 3H, CH₃SO₂), 1.77, 1.43 (2s, 6H, 2CH₃), 0.90 (s, 9H, t-Bu), 0.09 (s, 6H, 2Si-CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 156.5, 149.2, 140.8, 123.6, 112.9, 112.4, 81.8, 79.8, 78.6, 78.5, 72.5, 61.3, 40.2, 25.9, 25.8, 24.9, 18.3, -5.4, -5.5. Anal. Calcd for C₂₁H₃₂FN₃O₉SSi: C, 45.89; H, 5.87; N, 7.64. Found: C, 45.82; H, 5.94; N, 7.73. ESIMS m/z 550.15 [M+H⁺1.

1.7.3. [1-(6'-*O*-*t*-Butyldimethylsilyl-3',4'-*O*-isopropylidene-β-D-talopyranosyl)-5-fluorouracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (13b)

Spiro derivative **13b** was synthesized from **11b** by the similar procedure as described for **13a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 9.5:0.5) to give **13b** (0.20 g, 74%) as a white foam. $[\alpha]_2^{D^2}$ –22 (c 0.5, CDCl₃); R_f = 0.40 (CH₂Cl₂–MeOH, 9.5:0.5); UV (CDCl₃): λ_{max} 265 nm (ε 7399); IR (KBr, cm⁻¹): 3410, 3360 (NH₂), 1620 (C=C-N); ¹H NMR (CDCl₃, 300 MHz): δ 10.61 (br s, 1H, NH), 7.68 (d, 1H, $J_{6,F5}$ = 6.0 Hz, H-6), 6.47 (s, 1H, H-1'), 5.86 (br s, 2H, NH₂–4"), 5.45 (s, 1H, H-3"), 4.65 (d, 1H, $J_{3',4'}$ = 7.5 Hz, H-3'), 4.48 (d, 1H, H-4'), 4.36 (t, 1H, $J_{5',6a'}$ = $J_{5',6b'}$ = 4.6 Hz, H-5'), 3.86 (d, 2H, H-6a',b'), 1.58, 1.38 (2s, 6H, 2CH₃), 0.93 (s, 9H, t-Bu), 0.12, 0.10 (2s, 6H, 2Si–CH₃). ¹³C NMR (CDCl₃, 75.5 MHz): δ 157.9, 153.2, 150.5, 141.3, 124.8, 112.6, 89.4, 82.9, 80.4, 77.3, 75.7, 72.9, 62.6, 25.9, 25.6, 24.6, 18.7, –5.6, –5.7. Anal. Calcd for C₂₁H₃₂FN₃O₉SSi: C, 45.89; H, 5.87; N, 7.64. Found: C, 45.99; H, 5.81; N, 7.70. ESIMS m/z 550.19 [M+H⁺].

1.7.4. [1-(β-D-Talopyranosyl)-5-fluorouracil]-2'-spiro-5"-(4"-amino-1",2"-oxathiole-2",2"-dioxide) (15b)

Spiro derivative **15b** was synthesized from **13b** by the similar procedure as described for **15a**. It was purified by flash chromatography (CH₂Cl₂–MeOH, 8:2) to give **15b** (0.08 g, 59%) as a white foam. $[\alpha]_D^{22}$ –12 (c 0.1, DMSO); R_f = 0.24 (CH₂Cl₂–MeOH, 8:2); UV (DMSO): λ_{max} 263 nm (ε 2532); IR (KBr, cm⁻¹): 3450, 3320 (NH₂), 1655 (C=C-N); ¹H NMR (DMSO- d_6 , 300 MHz): δ 11.61 (br s, 1H, NH), 7.69 (d, 1H, $J_{6,F5}$ = 7.3 Hz, H-6), 6.69 (s, 1H, H-1'), 6.32 (br s, 2H, NH₂–4"), 6.19 and 6.15 (2 br s, 2H, 2OH), 5.47 (s, 1H, H-3"), 4.66 (br s, 1H, OH), 4.16–3.59 (m, 5H, H-3', H-4', H-5', 6-a',b'); ¹³C NMR (DMSO- d_6 , 75.5 MHz): δ 162.3, 158.7, 150.4, 140.8, 126.5, 90.8, 86.2, 79.1, 75.2, 70.4, 66.5, 62.5. Anal. Calcd for C₁₂H₁₄FN₃O₉S: C, 36.46; H, 3.57; N, 10.63. Found: C, 36.52; H, 3.68; N, 10.78. ESIMS m/z 396.05 [M+H⁺].

1.8. Antiproliferative assays

The cytostatic effects of the test compounds on murine leukemia cells (L1210), human T-lymphocyte cells (CEM), and human cervix carcinoma cells (HeLa) were evaluated as follows: an appropriate number of cells suspended in growth medium were allowed to proliferate in 200 μ L-wells of 96-well-microtiter plates in the presence of variable amounts of test compounds at 37 °C in a humidified CO₂-controlled atmosphere. After 48 h (L1210), 72 h (CEM), or 96 h (HeLa), the number of cells was counted in a Coulter

counter. The IC_{50} value was defined as the compound concentration required to inhibit cell proliferation by 50%.

Acknowledgments

This work was supported in part by the Postgraduate Programmes 'Biotechnology-Quality assessment in Nutrition and the Environment', 'Application of Molecular Biology-Molecular Genetics-Molecular Markers', Department of Biochemistry and Biotechnology, University of Thessaly. The biological experiments were supported by the KU Leuven (GOA 10/014). We are grateful to Mrs. Lizette van Berckelaer, Mrs. Leentje Persoons and Mrs. Frieda De Meyer for excellent technical assistance.

References

- Jordheim, L. P.; Durantel, D.; Zoulim, F.; Dumontet, C. Nat. Rev. Drug Disc. 2013, 12, 447–464.
- 2. Galmarini, C. M.; Mackey, J. R.; Dumontet, C. Lancet Oncol. 2002, 3, 415-424.
- 3. De Clercq, E. J. Med. Chem. 2010, 53, 1438–1450.
- Manta, S.; Kiritsis, C.; Dimopoulou, A.; Parmenopoulou, V.; Kollatos, N.; Tsotinis, A.; Komiotis, D. Anti-Infect. Agents 2014, 12.
- 5. Galmarini, C. M.; Mackey, J. R.; Dumontet, C. Leukemia 2001, 15, 875-890.
- 6. Ichikawa, E.; Kato, K. Curr. Med. Chem. 2001, 8, 385-423.
- 7. Li, N.-S.; Lu, J.; Piccirilli, J. A. Org. Prep. Proced. Int. 2010, 42, 191–283.
- 8. Camarasa, M. J.; Diaz-Ortiz, A.; Calvo-Mateo, A.; De las Heras, F. G.; Balzarini, J.; De Clercq, E. *J. Med. Chem.* **1989**, 32, 1732–1738.
- Azuma, A.; Nakajima, Y.; Nishizono, N.; Minakawa, N.; Suzuki, M.; Hanaoka, K.; Kobayashi, T.; Tanaka, M.; Sasaki, T.; Matsuda, A. J. Med. Chem. 1993, 36, 4183–4189.
- Camarasa, M. J.; Pérez-Pérez, M. J.; San-Félix, A.; Balzarini, J.; De Clercq, E. J. Med. Chem. 1992. 35, 2721–2727.
- Bonache, M. C.; Chamorro, C.; Velázquez, S.; De Clercq, E.; Balzarini, J.; Barrios, F. R.; Gago, F.; Camarasa, M. J.; San-Félix, A. J. Med. Chem. 2005, 48, 6653–6660.
- 12. Boivin, T. L. B. Tetrahedron 1987, 43, 3309-3362.
- Tronchet, J. M. J.; Kovacs, I.; Seman, M.; Dilda, P.; De Clercq, E.; Balzarini, J. Nucleosides Nucleotides Nucleic Acids 2000, 19, 775–794.
- De Castro, S.; Peromingo, M. T.; Naesens, L.; Andrei, G.; Snoeck, R.; Balzarini, J.; Velázquez, S.; Camarasa, M. J. J. Med. Chem. 2008, 51, 5823–5832.
- Moura, M.; Josse, S.; Nguyen Van Nhien, A.; Fournier, C.; Duverlie, G.; Castelain, S.; Soriano, E.; Marco-Contelles, J.; Balzarini, J.; Postel, D. Eur. J. Med. Chem. 2011, 46, 5046–5056.
- Tsoukala, E.; Manta, S.; Kiritsis, C.; Komiotis, D. Mini-Rev. Med. Chem. 2012, 12, 255–275.
- Dimopoulou, A.; Manta, S.; Kiritsis, C.; Gkaragkouni, D. N.; Papasotiriou, I.; Balzarini, J.; Komiotis, D. Bioorg. Med. Chem. Lett. 2013, 23, 1330–1333.
- Verheggen, I.; Van Aerschot, A.; Toppet, S.; Snoeck, R.; Janssen, G.; Balzarini, J.; De Clercq, E.; Herdewijn, P. J. Med. Chem. 1993, 36, 2033–2040.
- Verheggen, I.; Van Aerschot, A.; Van Meervelt, L.; Rozenski, J.; Wiebe, L.; Snoeck, R.; Andrei, G.; Balzarini, J.; Claes, P.; De Clercq, E.; Herdewijn, P. J. Med. Chem. 1995, 38, 826–835.
- Spanou, C.; Manta, S.; Komiotis, D.; Dervishi, A.; Kouretas, D. Int. J. Mol. Sci. 2007, 8, 695–704.
- Haouz, A.; Vanheusden, V.; Munier-Lechman, H.; Froeyen, M.; Herdewijn, P.; Van Galenbergh, S.; Delarue, M. J. Biol. Chem. 2003, 278, 4963–4971.
- Kiritsis, C.; Manta, S.; Parmenopoulou, V.; Balzarini, J.; Komiotis, D. Eur. J. Med. Chem. 2011, 46, 5668–5674.
- Kiritsis, C.; Manta, S.; Parmenopoulou, V.; Dimopoulou, A.; Kollatos, N.; Papasotiriou, I.; Balzarini, J.; Komiotis, D. Carbohydr. Res. 2012, 364, 8–14.
- 24. Kondo, T.; Goto, T. Agric. Biol. Chem. 1971, 35, 625-628.
- 25. Haeckel, R.; Weber, K.; Germann, C.; Haberkorn, U.; Zeisler, S.; Eisenbarth, J.; Wiessler, M.; Oberdorfer, F. J. Labelled Comp. Rad. 1996, 38, 1061–1070.
- Agelis, G.; Tzioumaki, N.; Botić, T.; Cencič, A.; Komiotis, D. Bioorg. Med. Chem. 2007, 15, 5448–5456.
- 27. Abraham, R. J.; Reid, M. Magn. Reson. Chem. 2000, 38, 570-579.
- 28. Martin, N. H.; Nance, K. H. J. Mol. Graph. Model. 2002, 21, 51-56.
- Rouillard, M.; Geribaldi, S.; Khazarian, J.; Azzaro, M. Org. Magn. Reson. 1980, 13, 323–327
- 30. Chatgilialoglu, C.; Lalevée, J. *Molecules* **2012**, *17*, 527–555.
- Velázquez, S.; Jimeno, M. L.; Camarasa, M. J.; Balzarini, J. Tetrahedron 1994, 50, 11013–11022.
- **32.** Perez-Perez, M. J.; Camarasa, M. J.; Diaz-Ortiz, A.; San-Felix, A.; De las Heras, F. G. *Carbohydr. Res.* **1991**, *216*, 399–411.
- 33. Khan, A. T.; Mondal, E. Synlett 2003, 694-698.
- Tzioumaki, N.; Tsoukala, E.; Manta, S.; Kiritsis, C.; Balzarini, J.; Komiotis, D. Carbohydr. Res. 2011, 346, 328–333.