See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/263899287

Investigations of amide bond variation and biaryl modification in analogues of $\alpha7$ nAChR agonist SEN12333

ARTICLE in EUROPEAN JOURNAL OF MEDICINAL CHEMISTRY · JULY 2014

 $Impact\ Factor:\ 3.45\cdot DOI:\ 10.1016/j.ejmech.2014.07.029\cdot Source:\ PubMed$

READS

39

11 AUTHORS, INCLUDING:



Corinne Beinat

University of Sydney

18 PUBLICATIONS 63 CITATIONS

SEE PROFILE



Yingxian Xiao

Georgetown University

70 PUBLICATIONS 2,121 CITATIONS

SEE PROFILE



John Tsanaktsidis

The Commonwealth Scientific and Industri...

72 PUBLICATIONS 520 CITATIONS

SEE PROFILE



Michael Kassiou

University of Sydney

232 PUBLICATIONS 3,089 CITATIONS

SEE PROFILE

FISEVIER

Contents lists available at ScienceDirect

European Journal of Medicinal Chemistry

journal homepage: http://www.elsevier.com/locate/ejmech



Short communication

Investigations of amide bond variation and biaryl modification in analogues of $\alpha 7$ nAChR agonist SEN12333



Corinne Beinat ^a, Tristan Reekie ^a, David Hibbs ^b, Teresa Xie ^c, Thao T. Olson ^c, Yingxian Xiao ^c, Andrew Harvey ^d, Susan O'Connor ^d, Carolyn Coles ^d, John Tsanaktsidis ^e, Michael Kassiou ^{a, f, g, *}

- ^a School of Chemistry, The University of Sydney, Sydney, NSW 2006, Australia
- ^b School of Pharmacy, The University of Sydney, Sydney, NSW 2006, Australia
- ^c Department of Pharmacology and Physiology, Georgetown University, Washington, DC 20057, USA
- ^d Bionomics Limited, Thebarton, SA 5031, Australia
- e CSIRO Materials Science & Engineering, Ian Wark Laboratory, Bayview Avenue, Clayton Victoria 3168, Australia
- f Discipline of Medical Radiation Sciences, The University of Sydney, Sydney, NSW 2006, Australia
- g Brain and Mind Research Institute, Sydney, NSW 2050, Australia

ARTICLE INFO

Article history: Received 12 May 2014 Received in revised form 2 July 2014 Accepted 3 July 2014 Available online 9 July 2014

Keywords: α₇ nicotinic receptors Acetylcholine receptor AChR CNS Structure—activity relationships Tetrazole Isostere

ABSTRACT

Several lines of experimental evidence support the involvement of the α_7 nAChR in schizophrenia and Alzheimer's disease. Modulators of the α_7 nAChR have been extensively reviewed for the treatment of the cognitive deficits associated with these pathologies. SEN12333 represents a novel α_7 nAChR agonist chemotype with potential for reduced side effects but requiring further SAR exploration. The present work investigates the amide bond of SEN12333, specifically its connectivity and replacement with the tetrazole functionality, a known cis amide isostere. The results reveal the original amide bond connectivity of SEN12333 to be favorable for binding affinity and agonist activity at α_7 nAChRs. The use of a tetrazole isostere completely abolishes affinity and functional activity and suggests that SEN12333 binds in a linear conformation. Results reported herein also suggest the pyridine nitrogen within the terminal aromatic ring of SEN12333 is not essential for binding affinity or functional activity. Further SAR investigations involving manipulation of other moieties contained within SEN12333 are warranted.

© 2014 Elsevier Masson SAS. All rights reserved.

1. Introduction

Neuronal nicotinic acetylcholine receptors (nAChRs) belong to the superfamily of ligand-gated cation channels and have recently generated extensive interest as potential targets of therapeutics in the treatment of cognitive disorders [1–3]. These receptors exist as pentameric transmembrane ion channels where subtype is formed from either a homo- or heteropentameric combination of twelve possible subunits: $\alpha 2 - \alpha 10$ and $\beta 2 - \beta 4$. The homomeric $\alpha 7$ nAChR is one of the most commonly expressed nicotinic receptors in the human brain, found in high levels in regions associated with learning and memory, such as the cerebral cortex and the hippocampus [4]. Experimental evidence supports the involvement of

E-mail address: michael.kassiou@sydney.edu.au (M. Kassiou).

the α 7 nAChR in schizophrenia and Alzheimer's disease (AD) [5]. Modulators of α 7 nAChRs have been extensively reviewed for the treatment of the cognitive deficits associated with these pathologies [6–8].

Multiple $\alpha 7$ agonists have entered clinical trials. Unfortunately, due to unfavorable side effects and pharmacokinetic issues the majority of these studies have been discontinued [9]. The known $\alpha 7$ ligands currently and previously in clinical trials display relatively little structural diversity and are primarily centered on anabaseine, quinuclidine, or diazabicyclic scaffolds [10]. Consequently, the majority of them possess the same cross-reactivity profiles with other sites and similar unwanted side effects. There is a strong demand for structurally differentiated $\alpha 7$ nAChR agonists with more desirable drug-like properties and improved cross-reactivity profiles for research in this field to further progress.

High-throughput screening by Siena Biotech and Wyeth has identified piperazine **1** (Fig. 1) as a novel chemotype of weak, partial agonist activity at α 7 nAChRs. Preliminary SAR investigation

^{*} Corresponding author. School of Chemistry, The University of Sydney, Sydney, NSW 2006, Australia.

1:
$$EC_{50} = 2.84 \,\mu\text{M}$$
 2 (SEN12333): $EC_{50} = 1.65 \,\mu\text{M}$

Fig. 1. Agonists at α_7 nAChRs identified by Siena Biotech and Wyeth.

Fig. 2. The incorporation of the urea functionality with pharmacophoric elements: basic center (blue), carbon chain (black), central linker (green) and biaryl system (red). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

Fig. 3. Amido-varied SEN12333 analogues.

of the piperazine, biaryl and amide regions of compound **1** resulted in the development of SEN12333 (**2**) [11]. SEN12333 is an α7 nAChR agonist, with selectivity over other nAChR subtypes, 5-HT3 receptors, and hERG channels [11,12]. Additionally, it displayed favorable bioavailability and brain permeation *in vivo* [12]. Initial investigations of SEN12333 (**2**) in animal models of episodic memory confirmed its ability to reverse both scopolamine- and MK-801-induced amnesia [11,12].

Limited SAR data is reported for this unique class of $\alpha 7$ nAChR ligands, with brief studies conducted into modification of the morpholine ring and variation of the arylanilide groups [11]. Previous work within our laboratory has identified the 4-carbon chain linker of compound **2** to be optimal for binding and functional activity at $\alpha 7$ nAChRs [13]. Minimal SAR investigation is available for the amide bond of SEN12333 (**2**). Haydaar and co-workers have described replacement of the amide moiety with the urea functionality (Fig. 2) [14]. Though these compounds generally exhibited

enhanced potency at α7 nAChRs, they also demonstrated low selectivity, particularly against receptors containing α3 subunit, unfavorable hERG activity and substantial CYP450 inhibition [14].

The present investigations were aimed at exploring the connectivity of the amide bond. The transposition of amide to reverse-amide is a concept well-known to medicinal chemistry [15–17]. Additionally, the effect of the pyridine nitrogen within the terminal aromatic ring of SEN12333 (2) was simultaneously investigated. The lack of a clear rationale why the pyridine nitrogen was moved from the 2-position in original hit 1 to the 3-position in SEN12333 (2), prompted us to question if it was required at all. Accordingly, compounds 3–5 were designed to address this question (Fig. 3).

Aside from investigating the amide connectivity, the amide linker was also replaced with a tetrazole group. The selection of a tetrazole linker is based upon the knowledge that tetrazoles are *cis*-amide isosteres [18–21]. It was envisioned that this would provide us further mechanistic detail into the binding mode of SEN12333 (2)

Scheme 1. Reagents and conditions: (a) (COCl)₂, rt, 45 min; (b) 4-bromoaniline, Et₃N, CH₂Cl₂, -78 °C to rt, 4 h, 91% yield over 2 steps; (c) morpholine, NaI, Et₃N, DMF, reflux, 16 h, 85%; (d) Pd(PPh₃)₄, appropriate boronic acid, MeCN/sat. aq. Na₂CO₃ (1:1), reflux, 18 h, 68 or 83%.

Scheme 2. Reagents and conditions: (a) Morpholine, Nal, Et₃N, THF, reflux, 16 h, 66%; (b) NH₂NH₂·H₂O, EtOH, reflux, 1 h; (c) 4-bromobenzoyl chloride, Et₃N, CH₂CH₂, -78 °C to rt, 2 h, 64% over two steps; (d) Pd(PPh₃)₄, appropriate boronic acid, MeCN/sat. aq. Na₂CO₃ (1:1), reflux, 18 h, 70 or 91%.

Scheme 3. Reagents and conditions. (a) (i) PCl₅, CH₂Cl₂, rt, 2 h; (ii) TMSN₃, 0 °C, 16 h, 40–71%.

at α_7 nAChRs and potentially reduce the number of rotatable bonds within the molecule. Tetrazoles are resistant to certain metabolic pathways, and their incorporation can often result in increased bioavailability of a drug [18]. There are two regioisomeric tetrazoles to be studied. One series would be accessed from SEN12333 (2) and analogues containing this original amide bond connectivity (3); whereas the other tetrazole series could be accessed from the related 'reverse-amide' bond analogues (4 and 5).

2. Chemistry

The syntheses of analogues **2** and **3** are depicted in Scheme **1** and involved the initial conversion of 5-bromovaleric acid (**6**) to the corresponding acid chloride. Subsequent treatment with 4-bromoaniline in the presence of triethylamine gave desired anilide **7** (91%). Alkylation of morpholine with bromide **7** in the presence of catalytic iodide afford compound **8** in 85% yield. Subjecting bromoarene **8** to Suzuki cross-coupling with either 3-pyridylboronic acid or phenylboronic acid gave the desired ligands SEN12333 (**2**) and **3** (83 or 68%) respectively. SEN12333 (**2**) was synthesized for direct comparison with the new analogues.

An alternative synthesis was proposed for reverse-amide analogues **4** and **5** (Scheme 2). Alkylation of morpholine with bromide **9** gave amine **10** (66%), subsequent hydrazinolysis and amide bond formation yielded compound **12** (64% over two steps). Final Suzuki cross-coupling with either 3-pyridylboronic acid or phenylboronic

Fig. 4. ORTEP representation of SEN12333 (2).

acid gave the desired retro-amide ligands **4** and **5** (70 or 91%) respectively.

The corresponding tetrazole analogues **13–16** were readily synthesized from amides **2–5**, a representative example is shown in Scheme 3. The amides **2–5** were treated with phosphorus pentachloride to form the presumed corresponding imidoyl chloride. However this was not isolated as the addition of trimethylsilylazide furnished the desired tetrazoles **13–16** following basic workup (40–71%).

Single crystals of SEN12333 (2) and tetrazole 13 were grown by slow recrystallization in CH₂Cl₂ and *n*-hexane and subjected to X-Ray crystal analysis. In addition to structure confirmation, it was thought that by identifying their solid-state conformations it would help to further predict the mode of binding of the amide and tetrazole analogues. The Oak Ridge Thermal Ellipsoid Plots (ORTEP) of SEN12333 (2) and tetrazole 13 are shown in Figs. 4 and 5 respectively. SEN12333 (2, Fig. 4) has a large degree of conformational flexibility and it is immediately apparent that its lowest energy configuration state adopts a *trans* amide conformation to linearize the structure and minimize steric interactions. In contrast, the tetrazole ring present within compound 13 locks the amide bond

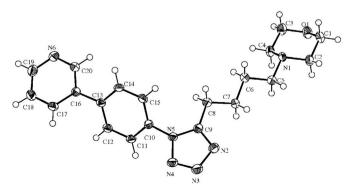


Fig. 5. ORTREP representation of tetrazole 13.

 Table 1

 Binding affinities of newly synthesized compounds.

Compound	$K_{i} (nM)^{a}$			Physicochemical properties		
	α7	α4β2	α3β4	CLogP ^b	PSA ^b	LLE ^c
	670 ± 119	>10,000	>10,000	2.25	53.93	3.92
	2300 ± 1000	>10,000	>10,000	3.58	41.57	2.06
	11,000 ± 4000	>10,000	>10,000	1.93	53.93	3.02
	1800 ± 200	>10,000	>10,000	3.28	41.57	2.46
N, N, N, N	216,000	DNC	DNC	2.08	65.15	1.59
N, N, N	236,000	DNC	>1,000,000	3.47	52.79	0.16
N, N, N	165,000	DNC	>1,000,000	2.81	65.15	0.97
N, N, N	156,000	DNC	>100,000	4.31	52.79	-0.5

^a Competition curves were constructed in which 10 concentrations of a sample were included. DNC = the fitting does not converge due to extremely low affinities. K_i values are the Mean \pm SEM of three to six independent measurements. For low binding affinity compounds, estimated K_i values or K_i value ranges were provided.

into a *cis* conformation and as illustrated in Fig. 5. It is evident that the compound **13** resides in a bent form. If the binding conformation of SEN12333 (2) is similar to the low energy, *trans* amide conformation depicted in Fig. 4, these studies suggest the mode of binding of SEN12333 (2) and tetrazole 13 would be vastly different. It was envisioned that this would additionally provide greater insight into the binding mode of SEN12333 (2), specifically, if the amide bond was residing in either a *cis* or *trans* conformation. It was thought that reverse-amide **4** would have a similar conformation to SEN12333 (**2**) while the tetrazole **15** would mimic analogue **13**, however, X-Ray crystal analysis was only obtained for a representative example of each class of compounds.

3. Pharmacology

All newly synthesized compounds and SEN12333 (2) as a reference were evaluated in competitive binding assays to

determine their affinity and selectivity for the $\alpha 7$ nAChR. The results are summarized in Table 1. All compounds were selective for the α7 nAChR subtype; however, a large range of affinities was evident. Removal of the pyridine nitrogen from reference SEN12333 (2) to give biphenyl analogue 3 resulted in a decrease in affinity. Reversal of the amide bond order of SEN12333 (2) to give compound 4 led to a marked decrease in affinity, however, this decrease in affinity was restored partially in retro-amide biphenyl analogue 5. These results suggest the original amide bond order of SEN12333 (2) to be favorable and that the pyridine nitrogen is not essential for binding or selectivity at α 7 receptors. Conversion of compounds to their corresponding tetrazole resulted in a loss of affinity at α 7 receptors with K_i values over 150 μ M for all tetrazole analogues. Considering the high concentrations required for the tetrazole series, a clear determination of the effect of the pyridine or biphenyl compounds in this series was not possible. In contrast to compounds **2–5**, tetrazoles synthesized from the original amide

^b Calculated using ChemDraw12.0.

^c Calculated according to equation LLE = plC_{50} (or pK_i) – cLogP.

SEN12333 (2) dose response on rat A7/GH4C1

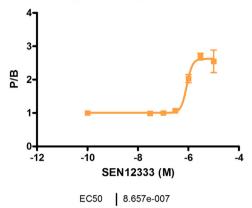


Fig. 6. Dose response curve of SEN12333 (2) on rat α_7 GH4C1 cells; EC₅₀ = 0.86 μ M.

bond order exhibited decreased affinity compared to tetrazoles synthesized from analogues containing the reversed amide bond order (K_i values **13** and **14** approximately 220 μ M vs. those of **15** and **16** approximately 130 μ M). These findings illustrate that the tetrazole motif is not tolerated as an amide isostere in this series of compounds. This could be attributed to one or more of a combination of effects including the electronics of the tetrazole ring, increased steric bulk, or the *cis* conformation of the tetrazole isostere as illustrated in Fig. 5.

Table 1 also describes the physicochemical properties cLogP, polar surface area (PSA) and ligand lipophilicity efficiency (LLE) for all newly synthesized compounds. It has been recognized for numerous years that the success of a lead in drug development is largely affected by its lipophilicity and the influence of LogP on drug potency, pharmacokinetics and toxicity [22], Lipophilic bases can potentially trigger cardiovascular toxicological effects through interaction with hERG channels and tissue toxicity through promotion of cellular phospholipidosis [23]. In terms of absorption and permeability, the 'rule of five' guidelines define the optimum for cLogP to be less than 5. In line with this, the average clogP value of marketed CNS drugs is 3.43 [24–27]. PSA is commonly utilized in drug design in the optimization of a drug's capability to permeate cell membranes such as the BBB [28]. PSA of approximately 120 Å² is deemed necessary to permeate cell membranes and $40-70 \text{ Å}^2$ is required to cross the BBB [28,29]. The development of a novel index known as ligand lipophilicity efficiency (LLE) has served as an

Compound 3 and 5 dose response on rat A7/GH4C1

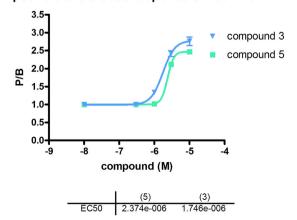


Fig. 7. Dose response curve of compound 3 and 5 on rat α_7 GH4C1 cells; 3: EC $_{50}=1.7~\mu M$, 5: EC $_{50}=2~\mu M$.

important guide in the process of lead optimization, and also as a flag for selection of preclinical candidates [30]. A typical oral drug with cLogP of 2.5 and potency within the range of 1–10 nM suggests a target LLE of 5–7 or greater, a value >5 is considered highly optimized for drug discovery. The advantages of high *in vivo* potency are low total doses in humans and decreased adventitious compound related toxicity [31].

All compounds display acceptable PSA values and cLogP < 5. Whilst biphenyl compound **5** displays a binding affinity similar to that of SEN12333 (**2**) (K_i values as 670 vs 1800 nM, respectively), its decreased LLE value of 2.46 suggests that it may not be as successful *in vivo*. While the cLogP and PSA values of compounds **4** and **5** are acceptable, their LLE values (3.02 and 2.46 respectively) also suggest their limited potential *in vivo*. Due to the extremely low binding affinities of tetrazole containing analogues **13**–**16**, they all display unacceptable LLE values, further confirming the limited use of tetrazoles as an amide isostere in SEN12333 (**2**) and related analogues.

To determine their agonistic activity at $\alpha 7$ nAChRs, SEN12333 and all amide-varied and tetrazole analogues were evaluated in a fluorescent calcium assay using a GH4C1 cell line expressing rat $\alpha 7$ receptors. Compounds were tested at concentrations of 0.3, 1, 3 and 10 μ M (n=3). Fluo4 Direct Dye from Molecular Probes was used and the assay was performed on a Perkin Elmer CellLux high throughput fluorescent reader. Peak/Base ratios were calculated to show activity where a peak/base values >1.00 was indicative of agonist activity.

Unfortunately, aside from SEN12333 (2), only compounds **3** and **5** showed agonism at α 7 nAChRs (Figs. 6 and 7). This was to be expected given the concentration range tested and the K_i values from the binding studies (Table 1). The EC₅₀ of SEN12333 (2) was observed to be 0.86 μ M. The biphenyl analogue **3**, retained agonistic activity at α 7 receptors with an EC₅₀ of 1.7 μ M. Retro-amide biphenyl analogue **5** similarly maintained agonistic activity with an EC₅₀ of 2.0 μ M. The lack of activity of the tetrazole containing compounds **13–16** highlights incorporation of this moiety is also detrimental for agonistic activity at α 7 nAChRs.

4. Conclusions

In summary, seven new analogues of SEN12333 (2) have been synthesized to examine the effects of amide bond orientation, the use of a tetrazole as an amide bond isostere and the importance of the pyridine nitrogen within the biaryl system. The original amide bond orientation of SEN12333 (2) was found to be favored; reversal of the amide bond resulted in a significant loss of binding and agonistic activity at α7 nAChRs (2 vs 4). Biphenyl analogues of the reverse amide series were found to be more potent than their corresponding phenyl-pyridine analogues (5 vs 4) and importantly maintained agonistic activity at $\alpha 7$ nAChRs. Furthermore, the biphenyl motif of compound 5 was found to restore the potency lost in the reversed amide pyridine compound 4. The use of a tetrazole functionality as an amide bond isostere resulted in a dramatic loss of potency and a complete loss of agonist activity for all types of analogues. The most potent compound of the series was still SEN12333 (2, $K_i = 670$ nM) where this amide bond orientation appears to be optimal for binding and activity at α 7 nAChRs.

Acknowledgments

CB acknowledges receipt of an Australian Postgraduate Award and a John A. Lamberton research scholarship.

Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.ejmech.2014.07.029.

References

- [1] LA. Dani, D. Bertrand, Annu. Rev. Pharmacol, Toxicol, 47 (2007) 699-729.
- [2] L.M. Broad, E. Sher, P.C. Astles, R. Zwart, M.J. O'Neill, Drug Future 32 (2007)
- [3] D. D'Hoedt, D. Bertrand, Expert Opin. Ther. Targets 13 (2009) 395–411.
 [4] P. Seguela, J. Wadiche, K. Dineley-Miller, J.A. Dani, J.W. Patrick, J. Neurosci. 13 (1993) 596-604.
- [5] A. Taly, P.-J. Corringer, D. Guedin, P. Lestage, J.-P. Changeux, Nat. Rev. Drug Discov. 8 (2009) 733-750.
- [6] P. Lippiello, M. Bencherif, T. Hauser, K. Jordan, S. Letchworth, A. Mazurov, Expert Opin. Drug Discov. 2 (2007) 1185-1203.
- R. Freedman, C.E. Adams, S. Leonard, J. Chem. Neuroanat. 20 (2000) 299-306.
- [8] W.R. Kem, Behav. Brain Res. 113 (2000) 169–181.
- [9] J. Toyohara, K. Hashimoto, Open Med. Chem. J. 4 (2010) 37–56.
- [10] A. Mazurov, T. Hauser, C.H. Miller, Curr. Med. Chem. 13 (2006) 1567–1584.
- [11] S.N. Haydar, C. Ghiron, L. Bettinetti, H. Bothmann, T.A. Comery, J. Dunlop, S. La Rosa, I. Micco, M. Pollastrini, J. Quinn, R. Roncarati, C. Scali, M. Valacchi, M. Varrone, R. Zanaletti, Bioorg. Med. Chem. 17 (2009) 5247-5258.
- [12] R. Roncarati, C. Scali, T.A. Comery, S.M. Grauer, S. Aschmi, H. Bothmann, B. Jow, D. Kowal, M. Gianfriddo, C. Kelley, U. Zanelli, C. Ghiron, S. Haydar, J. Dunlop, G.C. Terstappen, J. Pharmacol. Exp. Ther. 329 (2009) 459-468.
- [13] C. Beinat, S.D. Banister, P.S. van, M.R. Doddareddy, D. Hibbs, M. Sako, M. Chebib, T. Tran, N. Al-Muhtasib, Y. Xiao, M. Kassiou, Bioorg. Med. Chem. Lett. 22 (2012) 2380-2384.
- [14] C. Ghiron, S.N. Haydar, S. Aschmies, H. Bothmann, C. Castaldo, G. Cocconcelli, T.A. Comery, L. Di, J. Dunlop, T. Lock, A. Kramer, D. Kowal, F. Jow, S. Grauer, B. Harrison, S. La Rosa, L. Maccari, K.L. Marquis, I. Micco, A. Nencini, J. Quinn, A.J. Robichaud, R. Roncarati, C. Scali, G.C. Terstappen, E. Turlizzi, M. Valacchi, M. Varrone, R. Zanaletti, U. Zanelli, J. Med. Chem. 53 (2010) 4379-4389.

- [15] A. Reichelt, C. Gaul, R.R. Frey, A. Kennedy, S.F. Martin, J. Org. Chem. 67 (2002) 4062-4075.
- [16] B. Hu, J. Ellingboe, S. Han, E. Largis, R. Mulvey, A. Oliphant, F.-W. Sum, J. Tillett, J. Med. Chem. 44 (2001) 1456-1466.
- [17] S.L. Roderick, M.C. Fournie-Zaluski, B.P. Roques, B.W. Matthews, Biochemistry 28 (1989) 1493-1497.
- [18] R.J. Herr, Bioorg. Med. Chem. 10 (2002) 3379-3393.
- [19] K.L. Yu, R.L. Johnson, J. Org. Chem. 52 (1987) 2051–2059.
- [20] J. Zabrocki, J.B. Dunbar, K.W. Marshall, M.V. Toth, G.R. Marshall, J. Org. Chem. 57 (1992) 202–209.
- [21] J. Zabrocki, G.D. Smith, J.B. Dunbar, H. lijima, G.R. Marshall, J. Am. Chem. Soc. 110 (1988) 5875–5880.
- [22] H. van de Waterbeemd, D.A. Smith, K. Beaumont, D.K. Walker, J. Med. Chem. 44 (2001) 1313-1333.
- [23] M.J. Waring, C. Johnstone, Bioorg. Med. Chem. Lett. 17 (2007) 1759–1764.
- [24] C.A. Lipinski, F. Lombardo, B.W. Dominy, P.J. Feeney, Adv. Drug Deliv. Rev. 23 $(1997)^{\circ}3-25$
- [25] C.A. Lipinski, Drug Discov, Today Technol. 1 (2004) 337–341.
- [26] T. Fichert, M. Yazdanian, J.R. Proudfoot, Bioorg. Med. Chem. Lett. 13 (2003) 719-722.
- [27] H.L. Pajouhesh, G.R. Lenz, NeuroRx 2 (2005) 541–553.[28] J.J. Lu, K. Crimin, J.T. Goodwin, P. Crivori, C. Orrenius, L. Xing, P.J. Tandler, T.J. Vidmar, B.M. Amore, A.G.E. Wilson, P.F.W. Stouten, P.S. Burton, J. Med. Chem. 47 (2004) 6104-6107.
- [29] J. Kelder, P.J. Grootenhuis, D. Bayada, L.C. Delbressine, J.-P. Ploemen, Pharm. Res 16 (1999) 1514-1519
- [30] A.R. Leach, M.M. Hann, J.N. Burrows, E.J. Griffen, Mol. Biosyst. 2 (2006) 429-446.
- [31] P.D. Leeson, B. Springthorpe, Nat. Rev. Drug Discov. 6 (2007) 881-890.