

Targeting phosphatidylinositol 3 kinase (PI3K)-Akt beyond rapalogs

Shin Ogita · Patricia LoRusso

Received: 8 February 2011 / Accepted: 9 March 2011 / Published online: 6 May 2011
© Springer-Verlag 2011

Abstract The activation of the phosphatidylinositol 3 kinase (PI3K)-Akt pathway is a known causal mechanism of oncogenesis and resistance to cancer treatments. The process of PI3K-Akt pathway activation is complex and includes receptor tyrosine kinase (RTK) activation, PIK3CA mutations, loss of phosphatase and tensin homolog (PTEN), Akt mutations, tuberous sclerosis complex (TSC) mutations, and Ras homologue enriched in brain (RHEB) gene amplifications. The blockage of mammalian target of rapamycin (mTOR), the key downstream pathway protein, has been successful in selected cancer types, with mTOR-targeting agents available for clinical use. Other novel drugs blocking this pathway such as PI3K inhibitors, Akt inhibitors and PDK-1 inhibitors are currently only available for investigational use, but have shown promise as cancer therapies in both preclinical and early phase clinical studies. The newer generations of these inhibitors are more specific and have improved potency and safety. The combinations of targeted treatments against this pathway, blocking multiple different steps, are under preliminary investigation. Further research is needed to identify the biomarkers that predict treatment response and resistance in order to optimize personalized medicine.

Keywords PI3K · Akt · PDK-1 · Inhibitor · PTEN · mTOR

Introduction

The phosphatidylinositol 3 kinase (PI3K)-Akt pathway is one of the most popular areas for targeted cancer therapy. Both preclinical and clinical data indicate that alterations of

this pathway lead to oncogenesis. Mammalian target of rapamycin (mTOR) is one of the downstream enzymes in the pathway, and two of its inhibitors, everolimus and temsirolimus, currently approved by the United States Food and Drug Administration (FDA) for commercial use against renal cell cancer (RCC), have been found to be active against multiple tumor types [1–4]. Multiple other signal transduction proteins, both upstream and downstream of mTOR, are involved in the PI3K-Akt pathway and can potentially be targeted and “drugable.” mTOR as a target of inhibition has been reviewed extensively in recently published articles [5–7]. Our review will explain the PI3K-Akt pathway, including key proteins and enzymes and their relationships, followed by novel strategies other than rapamycin analogues to block the PI3K-Akt pathway.

PI3K-AKT pathway and oncogenesis

The PI3K-Akt pathway is a common intracellular signaling pathway that can be stimulated by the activation of various receptor tyrosine kinases (RTKs), including insulin like growth factor receptor (IGF-R), epithelial growth factor receptor (EGFR), and HER-2. Over-expression and/or gene mutations of the RTKs activate downstream pathways, causing increased proliferation and cell division. The RTKs can be inhibited by small molecule tyrosine kinase inhibitors (TKIs) or monoclonal antibodies (MoAbs) and are effective for cancer treatments. However, the treatment effect is often transient in the metastatic setting, and almost all the cancers eventually progress. The mechanisms of resistance include secondary mutations in RTKs and/or activation of downstream pathways such as the PI3K-Akt and RAS-RAF-MAPK/ERK pathways. Furthermore, some cancers have primary resistance to RTK inhibitors despite

S. Ogita · P. LoRusso (✉)
Karmanos Cancer Institute, Wayne State University,
4100 John R,
Detroit, MI 48201, USA
e-mail: lorussop@karmanos.org

known pathway activation, which suggests that activation of the downstream signaling cascades can be the primary oncogenic event. By blocking downstream proteins at select sites or at multiple locations, resistant cancers may potentially regain sensitivity to these therapies.

Figure 1 illustrates the PI3K-Akt pathway. We will begin by explaining the key enzymes involved in the pathway.

PI3K

PI3K converts phosphatidylinositol (4,5)-bisphosphate (PIP-2) to phosphatidylinositol (3,4,5)-trisphosphate (PIP-3) by phosphorylation. There are three classes of PI3K isoforms, namely class I, II and III [8]. Class I PI3K can be further subdivided to class IA and IB, and class IA is implicated in oncogenesis in human cancers. Class IA PI3Ks are heterodimeric, being comprised of one catalytic subunit (p110 α [also known as PIK3CA], p110 β [PIK3CB], and p110 δ [PIK3CD]), and one regulatory subunit (p85 α , p85 β , p55 α , p55 γ , and p50 α). The α catalytic subunit (PIK3CA) has been most extensively studied in human cancer, and its mutations can activate the PI3K-Akt pathway and are oncogenic [9]. PIK3CA mutations are frequently found in multiple malignancies including glioblastoma, gastric, breast, and colorectal cancers, and their prevalence varies not only among different cancer types (e.g., breast versus lung), but also among different subtypes of each cancer (e.g., estrogen receptor positive versus triple negative breast cancer) [10–12]. Well over 10 different PIK3CA mutations have been

discovered, and the most common mutations are H1047R, E545K and E542K [13]. Amplification of the PIK3CA gene occurs less commonly than mutations [14].

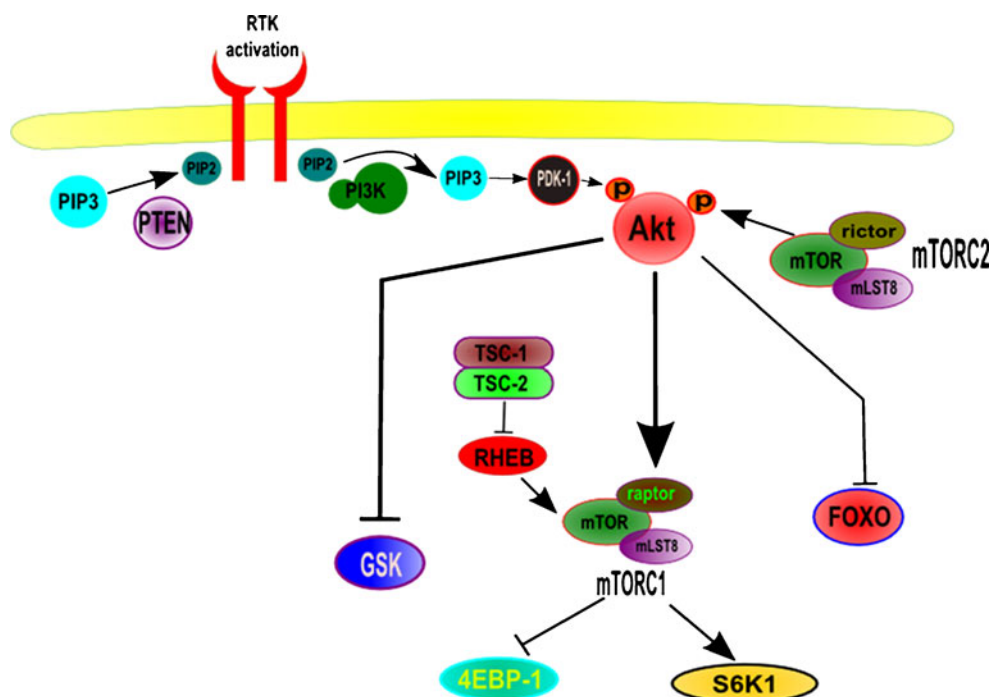
PTEN

Phosphatase and tensin homolog (PTEN), also referred to mutated multiple cancers 1 (MMAC 1) or TGF- β regulated and epithelial- cell-enriched phosphatase 1 (TEP1), dephosphorylates PIP-3 to PIP-2 counteracting PI3K. Inactivation or hypoactivation of PTEN causes overactivation of the PI3K-Akt pathway [10]. Germline mutations and loss of PTEN function are known to cause various syndromes including Bannayan-Zonana and Cowden syndromes [15, 16]. Patients with Cowden syndrome have up to a 90% increased risk of various malignancies, such as breast and thyroid cancers, during their lifetime [17]. Inactivation of PTEN by somatic mutations, deletions, promoter methylation and/or loss of heterozygosity (LOH) is commonly found in many cancers including prostate cancer, endometrial cancer, and glioblastoma and is associated with advanced stage of the diseases [18–23].

AKT

Akt, also known as protein kinase B (PKB), is a serine/threonine protein kinase that is activated by phosphorylation at Thr³⁰⁸ by phosphoinositide-dependent protein kinase 1 (PDK-1) and at Ser⁴⁷³ by mTOR complex 2 (mTORC2) [24]. There is some controversy if other kinases, or Akt alone, can phosphorylate Ser⁴⁷³. The activation of both

Fig. 1 PI3K-Akt pathway



kinase domains of Akt is required for its full activation [25]. The potential importance of Akt and its blockage in cancer is due to its pluri-functional role in intracellular signal transduction. Akt activates and/or inhibits multiple other proteins including BAD, FOXO and mTOR complex 1 (mTORC1). By acting on these multiple downstream pathways, Akt promotes cell survival, cell growth, cell proliferation, angiogenesis, cell migration, and invasion [26]. Akt activation is one of the important mechanisms in chemotherapy/radiotherapy resistance [27]. There are three isoforms of Akt, namely Akt1/PKB α , Akt2/PKB β , and Akt3/PKB γ . Although their functions overlap in some areas, each has its own distinct functional role. Akt1 is vital for normal growth, Akt2 for glucose regulation, and Akt3 for brain development [28–31].

The Akt1 mutation (E17K) at the pleckstrin homology (PH) domain has been observed in breast, colorectal, and ovarian carcinomas, and the mutation was found to enhance Akt activity [32]. Although the prevalence of this Akt mutation is generally low, it has been the target of novel Akt inhibitors [33–35]. Other alterations, including Akt 1 gene amplification, Akt1 and Akt2 deletion, and Akt3 amplifications have been found, and their clinical significance in human cancers is yet to be defined [36].

The degree of Akt phosphorylation of the tumor may reflect treatment effects from upstream pathway inhibition with such treatments as trastuzumab, cetuximab, and erlotinib. The comparison of pre- and post-treatment Akt phosphorylation can indicate if the pathway is adequately suppressed by these treatments, and select markers relative to this pathway are currently under investigation as pharmacodynamic (PD) markers in the setting of early phase clinical trials.

mTOR, S6K1 and 4EBP1

mTOR is a serine/threonine kinase and is structurally related to PI3K. mTOR binds to other intracellular proteins and forms two distinctive complexes, mTORC1 and mTORC2, respectively. mTORC1 is composed of mTOR, regulatory-associated protein of mTOR (raptor), and mLST8/G β L [37]. Activated mTORC1 phosphorylates ribosomal S6 kinase 1 (S6K1) and eukaryotic initiation factor 4E-binding protein (4E-BP1 or EIF4EBP-1). S6K1 promotes increased mRNA translation by phosphorylating the ribosomal protein S6. Hypophosphorylated 4E-BP protein binds to eIF4E and disrupts the interaction between ribosomes and mRNA. When hyperphosphorylated by mTOR, 4E-BP releases eIF4E and allows enhanced mRNA translation [38].

mTORC2 has been more recently discovered and consists of mTOR, rapamycin-insensitive companion of mTOR (ric-tor), and mLST8/G β L [39]. mTORC2 regulates cell morphology and the cytoskeleton [40]. Interestingly, mTORC2

plays a critical part in Akt activation [24]. Therefore, mTOR works both upstream and downstream of Akt in a different complex. The currently approved mTOR inhibitors—rapamycin (sirolimus), everolimus and temsirolimus—block mTORC1, only without affecting mTORC2 [41].

TSC-1/–2 and RHEB

Tuberous sclerosis complex (TSC) is a rare genetic disease caused by the germline mutation TSC-1 (hamartin) and TSC-2 (tuberin) genes. TSC patients develop variable clinical manifestations including benign tumors (hamartomas, angiofibromas, and rhabdomyoma) in the skin, kidney, lung, brain, eye, heart, and liver. Although they also have an increased risk of malignant tumors, especially renal cell carcinoma, there are still many TSC patients who do not develop malignancies [42]. This implies that TSC mutations alone are probably insufficient to cause cancer.

TSC-1 and TSC-2 form a heterodimer and function as tumor suppressors by inhibiting Ras homologue enriched in brain (RHEB), a guanosine triphosphate (GTP)-binding protein [43]. TSC has a GTPase-activating protein (GAP) domain, and exchange GTP with GDP [44]. This replacement inactivates RHEB. The functional loss of TSC by gene mutations or its promoter methylation results in RHEB overactivation [45–47].

The main role of RHEB is the activation of mTORC1, and its multiple downstream pathway proteins [48]. RHEB does not activate mTORC2 [49]. The two isoforms, RHEB 1 and RHEB 2, have been identified, and RHEB1 is most extensively studied [50]. RHEB gene amplifications can activate the pathway and have been found in prostate cancer [51]. RHEB overexpression was observed to varying degrees in human cancers including lymphoma, breast, and head/neck cancers. RHEB overexpression has been associated with aggressive disease and poor clinical outcome [52, 53]. RHEB overactivation is oncogenic and the process is mediated through mTORC1 and signal transducer and activator of transcription 3 (STAT3) [52]. In vitro studies suggest that an mTOR inhibitor can potentially induce chemotherapy sensitization in RHEB overexpressing lymphoma, but not in RHEB normoexpressing lymphoma [53].

PKD-1

PIP-3 recruits PDK-1 to the cell membrane where PDK-1 can exert its activity. PDK-1 is a serine/threonine kinase, and in the presence of PIP-3, PDK-1 phosphorylates and activates Akt [54]. Elevated phosphorylation of PDK-1 has been observed in select malignancies (i.e., breast) in patients. It is yet to be elucidated if PDK-1 overactivation is the primary oncogenic mechanism or just a consequence of upstream pathway activation [55].

GSK-3

Glycogen synthesis kinase-3 (GSK-3) has two isoforms: GSK-3 α and GSK-3 β . GSK-3 is known to both directly and indirectly regulate multiple intracellular proteins including transcription factors. GSK-3 not only affects glucose metabolism but also alters cell survival by inducing apoptosis [56]. Many proteins, including Akt, regulate GSK-3 by phosphorylation. As GSK-3 is involved in many pathways including Wnt, Notch and Hedgehog; it is beyond this review's scope to mention the detail of GSK-3 related oncogenesis. It is, however, constitutively active and suppresses pathways such as Wnt and Hedgehog. There was concern that GSK-3 inhibitors could induce malignancies. However, preclinical data paradoxically suggest GSK-3 inhibition is indeed effective as a cancer target [57, 58]. Further research in this area will eventually elucidate the role of GSK-3 in cancer pathophysiology and treatment.

FOXO

Forkhead box (FOX) is a family of more than 100 transcription factors, and the O subgroup (FOXO) plays a key role in cell cycle control and apoptosis [59]. FOXO has four subtypes, namely FOXO1, FOXO3, FOXO4 and FOXO6. FOXO is regulated by multiple mechanisms including phosphorylation, acetylation and ubiquitination. FOXO consists of four domains, namely a DNA-binding domain (DBD), a nuclear localization signal (NLS), a nuclear export sequence (NES), and a C-terminal transactivation domain. Akt phosphorylates FOXO at multiple domains and prevents FOXO from binding to DNA.

Rationale of blocking PI3K-AKT pathway

Although the PI3K-Akt signaling pathway is complex and blocking the pathway can potentially cause significant adverse effects on cell regulation, there are multiple scientific grounds to support this strategy in cancer therapy. As PI3K-Akt pathway activation is one of the important mechanisms of resistance to cytotoxic chemotherapies, hormonal therapies, biological treatments and radiotherapy, pathway inhibition can potentially overcome resistance [27].

We will first summarize the various topics of the pathway blockade, followed by class-specific compound discussions.

Upstream receptors. What receptors activate the PI3K-Akt pathway?

HER family (or ErbB family), platelet-derived growth factor receptor (PDGFR), and insulin-like growth factor (IGFR)

receptors are the transmembrane receptors, which are well known to activate the PI3K-Akt pathway. Many other receptors including fibroblast growth factor receptor (FGFR), vascular endothelial growth factor receptor (VEGFR), cMET/hepatocyte growth factor receptor (HGFR), T cell receptor and Notch have also been found to use the PI3K-Akt signaling cascade for cell proliferation, growth, and migration [60–64]. Overexpression of those receptors is one of the plausible oncogenic processes in many types of cancers. Direct receptor blockade by MoAb and/or TKI is the easiest method of inhibiting receptor-mediated tumor growth. However, the activation or overexpression of alternative pathways can occur when treatment resistance emerges. The blockade of common downstream intracellular pathways such as PI3K-Akt and/or RAS/RAF/MAPK can potentially overcome the treatment resistance.

Crosstalk with other pathways

Transmembrane receptors, upon activation, can stimulate multiple other intracellular pathways such as RAS/RAF/MAPK and JAK-STAT pathways. In some cancers the primary oncogenic event is the mutational activation of the pathway's protein enzymes.

The best-described example is the V600E BRAF mutation, which occurs in approximately 60% of melanoma patients [65]. BRAF inhibition, or its downstream protein MEK inhibition, can suppress the pathway. The proof of concept was observed in the phase I clinical study, where almost 70% of melanoma patients with such BRAF mutations responded to BRAF or MEK inhibitors [66, 67]. However, resistance can occur and one of the main causes is the crosstalk between RAS/RAF/MAPK and PI3K-Akt pathways. BRAF or MEK inhibitor-resistant melanoma was found to have Akt upregulation [68]. BRAF-resistant melanoma cells do not appear sensitive to downstream MEK inhibition, but potentially susceptible to PI3K inhibition. This is currently an area of intense research and multiple combination phase I studies are ongoing to prove this concept in the clinical setting.

mTOR inhibitors resistance and the role of PI3K or Akt inhibition

Currently mTOR inhibitors are the only commercially available agents for PI3K-Akt pathway blockade. Clinical response rates to mTOR inhibitor monotherapy in renal cell cancer are approximately 3–8% [1, 2]. mTOR inhibition can cause paradoxical activation of IGF-1 and its downstream pathway (i.e., PI3K-Akt pathway) by releasing physiological negative feedback inhibition [69, 70]. In a preclinical setting, the combination of mTOR inhibitors with IGF-1R inhibitors or Akt inhibitors has shown greater activity than either agent

used alone [70]. A more in-depth discussion of the relevance of mTOR inhibitor resistance is reviewed by Carew et al. [71].

What is the best combination?

It is clear that blockage of one pathway or target is not the ultimate solution. Even in very oncogene-addicted cancers, blockage of a single pathway can yield transient clinical benefit, but eventually results in resistance, often due to bypass mechanisms. What is the best combination? Is it mTOR+PI3K, MEK+PI3K, or BRAF+Akt inhibitors? The answer could be all or none of them. Clearly, the diverse biology of cancer cannot be generalizable even in the same histological type. Treating cancer patients based on traditional organ-specific strategies does work, but in the majority of cases of metastatic disease results in relatively ineffective treatment. Even if the primary oncogenic events are the same, the subsequent resistant mechanisms may be quite different. Various mechanisms of EGFR TKI resistance in lung adenocarcinoma—including T790M secondary mutations or MET protooncogene amplification—and dozens of different secondary mutations in chronic myeloid leukemia (CML) after imatinib failure clearly illustrate the diversity and complexity of resistance, and demonstrate great need for an individual approach to treatment [72–74]. Multiple tissue samplings at the time of diagnosis and at the time of disease progression will potentially be one of the tools needed to identify individual mechanisms of treatment resistance. Circulating tumor cells, if obtainable, may one day replace the need for invasive biopsies as technological advances improve their identification and sensitivity.

PI3K inhibitors

Dual PI3K and mTOR inhibitors

Like other kinase inhibitors, the mechanism of PI3K inhibition is mediated through the blockage of the adenosine triphosphate (ATP)-binding pocket of the catalytic subunit [75]. More than ten PI3K inhibitors have been developed and tested. It is not uncommon to observe one kinase inhibitor possessing inhibitory activities against multiple different kinases [76]. mTOR belongs to a family of PI3K-related kinases (PIKKs) and it shares structural similarity with PI3K [77]. More specifically, mTOR contains HEAT (Huntingtin, EF3, A subunit of PP2A and TOR) repeats, and its three dimensional structure resembles the PI3K catalytic subunit [77, 78]. Therefore, many but not all PI3K inhibitors have mTOR inhibitory potency.

There are potential advantages of dual mTOR and PI3K blocking agents over pure PI3K inhibitors. In some cancers, alterations of PDK-1, Akt, TSC, or RHEB are the oncogenic

processes. They are located downstream of PI3K and PI3K inhibition alone is unlikely to suppress the downstream pathway activation. However, as they sit upstream of mTOR, the mTOR inhibitory component of dual PI3K-mTOR inhibition can be effective for those select cancers. Additionally, the crosstalk between the RAS/RAF/MAPK and PI3K-Akt pathways occurs at the level of Akt. Potential disadvantages are side effects due to multi-blockade and the resultant inability to increase the dose high enough to get adequate tumor-drug exposure to cause cell death. Further basic and clinical research will elucidate the difference between dual blockade and pure PI3K inhibitors.

The PI3K inhibitor prototypes are **LY294002** and **wortmannin**. The naturally occurring flavonoid **quercetin** is found in many vegetables and fruits. It has potent PI3K inhibitory activity. LY294002 is a quercetin derivative and has higher potency for PI3K inhibition [79]. In both in vitro and in vivo studies, LY 294002 has reversed paclitaxel resistance in ovarian cancer, and the combination of paclitaxel with LY294002 has demonstrated synergy [80]. However, poor bioavailability has precluded further development of LY294002. **SF1126** is a prodrug of LY294002 that is linked with an integrin antagonist. This enhances delivery of LY294002 to targeted cells. After IV administration, SF1126 is hydrolyzed in a pH-dependent manner. In a phase I study, SF1126 was well tolerated and anticancer activity was observed [81].

Wortmannin is a fungal metabolite and is a potent irreversible PI3K inhibitor [75, 82]. Wortmannin is an unstable compound, less active in vivo than in vitro, and is presently not in clinical use [83]. To overcome its pharmacological problems, multiple derivatives have been developed. **PX-866**, the most potent wortmannin derivative currently available, recently completed phase I investigation in solid tumor malignancies and was well tolerated with a 20% stable disease rate [84]. The agent is currently under clinical investigation in combination with cytotoxic chemotherapies [85].

BEZ 235 is an oral potent dual PI3K and mTOR inhibitor. In addition to mTORC 1, BEZ 235 also inhibits mTORC2. In a phase I study, a partial response was observed in a lung cancer patient with Cowden syndrome and in an ER(+) breast cancer patient [86]. Additionally, 24% of patients had stable disease (SD) ≥4 months. Although these results are encouraging, once again it can be observed that monotherapy with many agents of this class will be insufficient for most metastatic patients. Unlike BKM 120 (see below), hyperglycemia was not observed.

More specific PI3K inhibitors

BKM 120 is a specific PI3K inhibitor [87]. Its antitumor activity was observed initially in a xenograft model of

PIK3CA mutated breast cancer [88]. In a phase I study, a partial response (PR) was observed in a triple-negative breast cancer patient whose tumor harbored the KRAS mutation [89]. KRAS is known to activate not only the Ras-RAF-MAPK pathway but also PI3K [90]. This KRAS-PI3K crosstalk is a known mechanism of resistance to MEK inhibitors and the preliminary positive response in this KRAS-mutated patient lends consideration for the possible use of PI3K inhibitors in combination with MEK inhibitors [91]. Only one colorectal patient in the study carried PIK3CA mutation and did not respond to the BKM120 compound. The dose-limiting toxicities included grade 4 hyperglycemia, grade 3 mood alteration, and rash. The mood alteration from PI3K inhibitors was somewhat unexpected, but was explainable. PI3K inhibition can cause insulin resistance. This resistance was associated with increased risk of neurocognitive dysfunction including Alzheimer's disease [92]. In an animal model study, PI3K inhibition caused depression and anxiety [93].

XL147 (SAR 245408) is another oral PI3K inhibitor. XL147 was investigated as a single agent and in combination with carboplatin plus paclitaxel, and with erlotinib [94–96]. The agent was well tolerated, and a partial response was observed in a lung cancer patient in a monotherapy study [94]. **XL765** (SAR245409) is an oral PI3K and TORC1/2 inhibitor. XL765 is well tolerated, both as a single agent as well as in combination with erlotinib or temozolomide (TMZ) [97–99]. **GDC-0941**, another PI3K inhibitor, has demonstrated improved bioavailability relative to its prototype compounds, PI-103, PI-540 and PI-620 [100]. In phase I studies, a partial response has been reported in a breast cancer patient and prolonged SD was achieved in multiple ovarian cancer patients [101, 102].

Other PI3K inhibitors

CAL-101 is a p110 δ specific PI3K inhibitor, while all the other PI3K inhibitors are p110 α inhibitors. Unlike p110 α , which is expressed ubiquitously in many tissues, p110 δ is exclusively found in leukocytes, making this agent potentially useful for the treatment of hematological malignancies [103]. Chronic lymphocytic leukemia (CLL) cells express higher p110 δ compared to normal B-cells. CAL-101 induced apoptosis in CLL cells without affecting normal B-cells [104]. In phase I studies, single agent CAL-101 showed high response rates of 62% in heavily pretreated indolent non-Hodgkin's lymphoma patients and in 33% of CLL patients [105, 106]. The clinical study results and main pharmacological actions of notable PI3K inhibitors are summarized in Table 1 [81, 85, 86, 89, 94, 97, 101, 102, 105–112].

AKT inhibitors

Alkyl-lysophospholipids (ALPs) are synthetic lipids and cannot be properly degraded by phospholipase, resulting in abnormal accumulation in the cell membrane, disruption of cell metabolism, and cell death [113]. ALP also has Akt inhibitory activity and anti-neoplastic properties [114]. **Miltefosine** is one of the ALPs and is very effective in the treatment of visceral leishmaniasis [115]. It is available as both a topical solution as well as an oral agent. Topical miltefosine can effectively treat cutaneous breast cancer metastasis [116]. Oral miltefosine was evaluated for cancer treatment and the dose was 3–4 times higher than the dose used for the treatment of leishmaniasis. Responses were observed in a fraction of patients, however toxicities including nausea, vomiting, and kidney impairment precluded further development of this medication as an anti-neoplastic agent [117]. **Perifosine** is structurally related to miltefosine, but has a superior safety profile. Perifosine does not inhibit Akt directly, but inhibits Akt translocation to the cell membrane, which is necessary for Akt phosphorylation and activation [118]. Perifosine has been studied in many phase I and phase II studies. Although the agent was well tolerated, perifosine did not exhibit antitumor activity in melanoma, squamous cell carcinoma of the head and neck, or breast cancer as a single agent [119–121]. In contrast, durable responses were observed in heavily pretreated Waldenström's macroglobulinemia (WM), chronic lymphocytic leukemia (CLL), and soft tissue sarcoma patients [122–124]. Multiple combination studies with various agents are ongoing.

Triciribine, also known as Akt/protein kinase B signaling inhibitor-2 (API-2), is a potent and specific Akt inhibitor [125]. Triciribine has been around for over 20 years and has been used in phase I and phase II studies. At high doses, it has clearly shown anti-neoplastic activity as a single agent, including one complete response in metastatic squamous cell carcinoma of cervix [126]. Due to its significant side effect profile, including hepatotoxicity, hyperglycemia, and hypocalcemia, its clinical use has been limited.

MK2206, an allosteric Akt inhibitor, has high inhibitory potency against both Akt1 and Akt2 with less activity toward Akt3. In a phase I study with single agent MK2206, SD was observed in 16% of refractory solid tumor patients and a minor response was observed in a patient who had chemo-refractory pancreatic cancer with PTEN loss [127]. In vitro studies suggest enhanced effects when MK2206 is used in combination with other cytotoxic agents [128]. Currently, multiple studies with various drug combinations, including both targeted and cytotoxic agents, are ongoing. The summary data of Akt inhibitors are presented in Table 1 [122–124, 126, 127, 129–131].

Table 1 Summary of PI3K inhibitors, Akt inhibitors and PDK-1 inhibitors

Drug Class	Drug [reference]	Other target action	Mode of admin.	Clinical Efficacy ^a	DLT (notable non-DLT toxicities)	Current phase of clinical studies and combination use
PI3K inhibitor	Quercetin [107]	PI4K	IV, oral	AFP decrease in HCC and CA-125 decrease in ovarian Ca	Nephrotoxicity	Chemoprevention No therapeutic trials
	wortmannin	mTOR, PI4K				Not in clinical use
	LY294002 [108]	mTOR, PI4K DNA-PK				Not in clinical use
	BKM120 [89]	PIM1, PLK1, CK2, ATM	oral	PR in breast Ca. 42% SD	hyperglycemia, rash, epigastralgia mood alteration, pruritus	Phase I/II AI, trastuzumab, MEK inhibitor
	BEZ 235 [86, 108]	mTOR DNA-PK	oral	PR in lung and breast Ca 24% SD	No DLT (fatigue, diarrhea)	Phase I/II AI, trastuzumab
	XL147 (SAR245408) [94]		oral	PR in lung 17% SD	Rash, Hypersensitivity (nausea, fatigue, diarrhea)	Phase I/II
	XL 765 (SAR245409) [97]	mTOR	oral	14% SD	nausea, vomiting, rash, fatigue, hypophosphatemia, abnormal EKG, transaminitis	Phase I AI, temozolomide, erlotinib
	SF1126 [81]	mTOR (TORC1/TORC2), DNA-PK PIM1, PLK1, CK2, ATM	IV	21% SD	Diarrhea (nausea, vomiting, fever, fatigue)	Phase I
	GDC-0941 [101, 102, 108]	mTOR DNA-PK	oral	PR in breast Ca 16-21% SD	Headache Pleural effusion Decreased DLCO (nausea, vomiting, diarrhea, dyspnea)	Phase I MEK inhibitor Trastuzumab
	GDC-0980 [109]	mTOR	oral	23% SD	No DLT (nausea, diarrhea, fatigue, flatulence)	Erlotinib Carbo/Pac/Bev Phase I
	PX 866 (wortmannin analogue) [85]	mTOR	oral	22% SD	diarrhea increased AST (nausea, vomiting, fatigue)	Pac/Bev/trastuzumab Phase I cetuximab, docetaxel
	CAL-101 (p110 δ inhibitor) [105, 106]		oral	33% PR in CLL 62% PR in indolent NHL	33% AST/ALT elevation (cytopenia)	Phase I Bendamustine ofatumumab
	PF-04691502 [110]	mTOR	oral			Phase I
	BGT226	mTOR	oral			Phase I
	GSK2126458 [111]	mTOR	oral			Phase I
	ZSTK474 [108, 112]	mTOR DNA-PK	oral			Phase I
	XL-499 IC486068 (p110 δ inhibitor)					preclinical

Table 1 (continued)

Drug Class	Drug [reference]	Other target action	Mode of admin.	Clinical Efficacy ^a	DLT (notable non-DLT toxicities)	Current phase of clinical studies and combination use
Akt inhibitor	Triciribine (APL-2) [126, 129]		IV	PR and CR in cervical SCC	Hepatotoxicity Hypocalcemia hyperglycemia	Phase I
	MK2206 [127]		oral	16% SD	Rash, hyperglycemia Pruritus, diarrhea (nausea, fatigue,)	Phase I Carbo/Pac Docetaxel Trastuzumab, lapatinib MEK inhibitor Gefitinib
PDK-1 inhibitor	Perifosine [122–124, 130]	Cell membrane	oral	6% PR in WM PR in CLL PR in sarcoma	Fatigue diarrhea	IGF-1R MoAb Phase I/II/III VEGF TKIs Bortezomib Various cytotoxic agents mTOR inhibitors imatinib Preclinical
	GSK690693 [131]	PAK6, PKC η PKC θ , PrkX S6K	IV			preclinical
	XL418					preclinical
	A443654					preclinical
	PX-316					preclinical
	OSU-03012					preclinical
	BX912 [142]					preclinical
	BX320 [143]	PKA				preclinical
	BX795 [144]					preclinical
	GSK2334470 [145]					preclinical
PDK-1 inhibitor	BAG956 [144]	P13K				preclinical
	dibenzo [c,f][2, 7] naphthyridines [145]					preclinical

^a Efficacy is single agent response. If cancer subtype is not specified, efficacy was observed in unselected solid malignancy patients.

AFP alpha fetoprotein, *AI* aromatase inhibitor, *Benda* bendamustine, *Bev* bevacizumab, *Ca* carcinoma, *Carbo* carboplatin, *CLL* chronic lymphocytic leukemia, *CR* complete response, *DLCO* Diffusing Capacity of the Lung for Carbon Monoxide, *DLT* dose-limiting toxicities, *HCC* hepatocellular carcinoma, *IGF-1R* insulin like growth factor 1 receptor, *IV* intravenous, *K* kinase, *MoAb* monoclonal antibody, *mTOR* mammalian target of rapamycin, *NHL* non-Hodgkin's lymphoma, *Pac* paclitaxel, *P1* Phosphatidylinositol, *PIM-1* Proto-oncogene serine/threonine-protein kinase pim-1, *PKA* protein kinase A, *PR* partial response, *SD* stable disease, *S6K* ribosomal S6 kinase, *TKIs* tyrosine kinase inhibitors, *TORC* mammalian target of rapamycin complex, *VEGF* vascular endothelial growth factor, *WM* Waldenström's macroglobulinemia

PDK-1 inhibitors

Table 1 summarizes PDK1 inhibitors. PDK1 inhibitors are still in the preclinical drug development stage, but are worth mentioning. **Celecoxib** is a cyclooxygenase –2 (COX-2) inhibitor and has anti-inflammatory activity. Celecoxib also has moderate PDK-1 inhibitory action, and can induce apoptosis in cancer cells [132]. However, because of its low potency, celecoxib at currently approved dosages for inflammation does not have clinically meaningful antitumor effects [133, 134]. **OSU-03012** is a celecoxib derivative and has a twenty-fold greater inhibitory potency against PDK-1 compared with celecoxib [135]. In vitro studies have demonstrated cytotoxicity against various cancer cell lines including those of CLL, glioblastoma, lymphoma, myeloma, breast, and pancreatic cancers [136–138].

UCN-01, 7-Hydroxystaurosporine, is a multi-kinase inhibitor whose targets include PKC α , PKC β , PKC γ , Chk1 as well as PDK-1 [139]. Multiple studies have confirmed its antitumor potency, and several trials are currently ongoing in different phases of clinical investigation [140]. However, more recent data has indicated its main mechanism of anticancer activity is through Chk1 inhibition, and its role as a PDK-1 inhibitor is not yet defined [141]. Currently investigated PDK-1 inhibitors are listed in Table 1 [142–145].

Unanswered questions and future directions

The obvious and yet most difficult question is which agent or agents will be the champion in each class and why. Most of the new agents currently under clinical investigation are safe and well tolerated. Their side effect profiles, which include hyperglycemia, pulmonary toxicity and mood alteration, may limit their clinical use in certain population groups such as those with diabetes, chronic obstructive pulmonary disease, and depression, respectively. Most of the agents demonstrated, at best, only cytostatic activity with an occasional response. Except for one case involving a KRAS-mutated breast cancer patient who responded to BKM120, there have been no relevant potential predictive biomarkers seen clinically. Like other targeted agents, these agents will most likely show activity in selected patient populations with specific molecular alterations, and in drug combinations. The importance of simultaneous development of predictive biomarkers cannot be overemphasized. Additionally, a more effective and efficient way to screen for appropriate patient selection is needed. Some pathway alterations, such as the Akt1 mutation, are generally uncommon. If the association of clinical characteristics with molecular characteristics can be identified, screening strategies will be more efficient.

Although our understanding of PI3K-Akt and its surrounding pathways has improved, further elucidation (especially its crosstalk with other signaling cascades and defining appropriate patient subsets), is critical if we are to more definitively define its role in cancer treatment. The early therapeutics team at MD Anderson has presented an initial and updated response profile of patients treated with inhibitors of the PI3K pathway. They looked at response primarily of monotherapy treatment in patients preselected by either a PIK3CA mutation or PTEN deletion [146]. Although preliminarily, extremely positive results relative to mutational status, the most recent presentation showed <25% response. Although still encouraging, it demonstrates to the cancer community the importance of the next steps of PIK3CA mutations. We have to think and act smarter but to do that we will also need to gather the facts, as difficult as it may be. Even when we harvest fresh tissue, it is unknown how reflective its expression profile is relative to other disease sites within the same patient. However, when we finally define the keys to therapeutic success, we must be prepared to act: we must continue to develop in parallel the necessary pharmacodynamic tools and clinical drug combinations so as to be able to tackle the results head on and hopefully make a difference in the treatment of this disease both collectively, as well as one patient at a time.

Conflict of interest statement Although the authors have not received and will not receive benefits for personal or professional use from a commercial party related directly or indirectly to the subject of this manuscript, benefits have been or will be received but are directed solely to a research fund, foundation, educational institution or other non-profit organization with which one or more of the authors is(are) associated.

References

1. Hudes G, Carducci M, Tomczak P, Dutcher J, Figlin R, Kapoor A, Staroslawska E, Sosman J, McDermott D, Bodrogi I, Kovacevic Z, Lesovoy V, Schmidt-Wolf IGH, Barbarash O, Gokmen E, O'Toole T, Lustgarten S, Moore L, Motzer RJ, the Global ARCC Trial, (2007) Temsirolimus, interferon alfa, or both for advanced renal-cell carcinoma. *N Engl J Med* 356 (22):2271–2281. doi:10.1056/NEJMoa066838
2. Motzer RJ, Escudier B, Oudard S, Hutson TE, Porta C, Bracarda S, Grünwald V, Thompson JA, Figlin RA, Hollaender N, Urbanowitz G, Berg WJ, Kay A, Lebwohl D, Ravaud A (2008) Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomised, placebo-controlled phase III trial. *Lancet* 372(9637):449–456
3. Witzig TE, Reeder CB, LaPlant BR, Gupta M, Johnston PB, Micallef IN, Porrata LF, Ansell SM, Colgan JP, Jacobsen ED, Ghobrial IM, Habermann TM (2010) A phase II trial of the oral mTOR inhibitor everolimus in relapsed aggressive lymphoma. *Leukemia* 25(2):341–347
4. Baselga J, Semiglazov V, van Dam P, Manikhas A, Bellet M, Mayordomo J, Campone M, Kubista E, Greil R, Bianchi G, Steinseifer J, Molloy B, Tokaji E, Gardner H, Phillips P, Stumm

- M, Lane HA, Dixon JM, Jonat W, Rugo HS (2009) Phase II randomized study of neoadjuvant everolimus plus letrozole compared with placebo plus letrozole in patients with estrogen receptor-positive breast cancer. *J Clin Oncol* 27(16):2630–2637. doi:10.1200/jco.2008.18.8391
5. Sabatini DM (2006) mTOR and cancer: insights into a complex relationship. *Nat Rev Cancer* 6(9):729–734
 6. Wulschleger S, Loewith R, Hall MN (2006) TOR signaling in growth and metabolism. *Cell* 124(3):471–484. doi:10.1016/j.cell.2006.01.016
 7. Sparks CA, Guertin DA (2010) Targeting mTOR: prospects for mTOR complex 2 inhibitors in cancer therapy. *Oncogene* 29(26):3733–3744
 8. Vanhaesebroeck B, Guillermet-Guibert J, Graupera M, Bilanges B (2010) The emerging mechanisms of isoform-specific PI3K signalling. *Nat Rev Mol Cell Biol* 11(5):329–341. doi:10.1038/nrm2882
 9. Kang S, Bader AG, Vogt PK (2005) Phosphatidylinositol 3-kinase mutations identified in human cancer are oncogenic. *Proc Natl Acad Sci USA* 102(3):802–807. doi:10.1073/pnas.0408864102
 10. Stemke-Hale K, Gonzalez-Angulo AM, Lluch A, Neve RM, Kuo WL, Davies M, Carey M, Hu Z, Guan Y, Sahin A, Symmans WF, Pusztai L, Nolden LK, Horlings H, Berns K, Hung MC, van de Vijver MJ, Valero V, Gray JW, Bernards R, Mills GB, Hennessy BT (2008) An integrative genomic and proteomic analysis of PIK3CA, PTEN, and AKT mutations in breast cancer. *Cancer Res* 68(15):6084–6091. doi:10.1158/0008-5472.CAN-07-6854
 11. Samuels Y, Wang Z, Bardelli A, Silliman N, Ptak J, Szabo S, Yan H, Gazdar A, Powell SM, Riggins GJ, Willson JK, Markowitz S, Kinzler KW, Vogelstein B, Velculescu VE (2004) High frequency of mutations of the PIK3CA gene in human cancers. *Science* 304(5670):554. doi:10.1126/science.1096502
 12. Saal LH, Holm K, Maurer M, Memeo L, Su T, Wang X, Yu JS, Malmström P-O, Mansukhani M, Enoksson J, Hibshoosh H, Borg Å, Parsons R (2005) PIK3CA Mutations Correlate with Hormone Receptors, Node Metastasis, and ERBB2, and Are Mutually Exclusive with PTEN Loss in Human Breast Carcinoma. *Cancer Res* 65(7):2554–2559
 13. Karakas B, Bachman KE, Park BH (2006) Mutation of the PIK3CA oncogene in human cancers. *Br J Cancer* 94(4):455–459
 14. Wu G, Xing M, Mambo E, Huang X, Liu J, Guo Z, Chatterjee A, Goldenberg D, Gollin SM, Sukumar S, Trink B, Sidransky D (2005) Somatic mutation and gain of copy number of PIK3CA in human breast cancer. *Breast Cancer Res* 7(5):R609–616. doi:10.1186/bcr1262
 15. Marsh DJ, Dahia PL, Zheng Z, Liaw D, Parsons R, Gorlin RJ, Eng C (1997) Germline mutations in PTEN are present in Bannayan-Zonana syndrome. *Nat Genet* 16(4):333–334. doi:10.1038/ng0897-333
 16. Liaw D, Marsh DJ, Li J, Dahia PL, Wang SI, Zheng Z, Bose S, Call KM, Tsou HC, Peacocke M, Eng C, Parsons R (1997) Germline mutations of the PTEN gene in Cowden disease, an inherited breast and thyroid cancer syndrome. *Nat Genet* 16(1):64–67. doi:10.1038/ng0597-64
 17. Riegert-Johnson DL, Gleeson FC, Roberts M, Tholen K, Youngborg L, Bullock M, Boardman LA (2010) Cancer and Lhermitte-Duclos disease are common in Cowden syndrome patients. *Hered Cancer Clin Pract* 8(1):6. doi:10.1186/1897-4287-8-6
 18. Wang SI, Puc J, Li J, Bruce JN, Cairns P, Sidransky D, Parsons R (1997) Somatic Mutations of PTEN in Glioblastoma Multiforme. *Cancer Res* 57(19):4183–4186
 19. Sano T, Lin H, Chen X, Langford LA, Koul D, Bondy ML, Hess KR, Myers JN, Hong Y-K, Yung WKA, Steck PA (1999) Differential Expression of MMAC/PTEN in Glioblastoma Multiforme. *Cancer Res* 59(8):1820–1824
 20. Cairns P, Okami K, Halachmi S, Halachmi N, Esteller M, Herman JG, Jen J, Isaacs WB, Bova GS, Sidransky D (1997) Frequent Inactivation of PTEN/MMAC1 in Primary Prostate Cancer. *Cancer Res* 57(22):4997–5000
 21. Kong D, Suzuki A, Zou TT, Sakurada A, Kemp LW, Wakatsuki S, Yokoyama T, Yamakawa H, Furukawa T, Sato M, Ohuchi N, Sato S, Yin J, Wang S, Abraham JM, Souza RF, Smolinski KN, Meltzer SJ, Horii A (1997) PTEN1 is frequently mutated in primary endometrial carcinomas. *Nat Genet* 17(2):143–144. doi:10.1038/ng1097-143
 22. Kang Y-H, Lee HS, Kim WH (2002) Promoter Methylation and Silencing of PTEN in Gastric Carcinoma. *Lab Invest* 82(3):285–291
 23. Salvesen HB, MacDonald N, Ryan A, Jacobs IJ, Lynch ED, Akslen LA, Das S (2001) PTEN methylation is associated with advanced stage and microsatellite instability in endometrial carcinoma. *Int J Cancer* 91(1):22–26. doi:10.1002/1097-0215
 24. Sarbassov DD, Guertin DA, Ali SM, Sabatini DM (2005) Phosphorylation and Regulation of Akt/PKB by the Rictor-mTOR Complex. *Science* 307(5712):1098–1101
 25. Bellacosa A, Chan TO, Ahmed NN, Datta K, Malstrom S, Stokoe D, McCormick F, Feng J, Tsichlis P (1998) Akt activation by growth factors is a multiple-step process: the role of the PH domain. *Oncogene* 17(3):313–325. doi:10.1038/sj.onc.1201947
 26. Manning BD, Cantley LC (2007) AKT/PKB Signaling: Navigating Downstream. *Cell* 129(7):1261–1274. doi:10.1016/j.cell.2007.06.009
 27. Clark AS, West K, Streicher S, Dennis PA (2002) Constitutive and Inducible Akt Activity Promotes Resistance to Chemotherapy, Trastuzumab, or Tamoxifen in Breast Cancer Cells. *Mol Cancer Ther* 1(9):707–717
 28. Chen WS, Xu P-Z, Gottlob K, Chen M-L, Sokol K, Shiyanova T, Roninson I, Weng W, Suzuki R, Tobe K, Kadowaki T, Hay N (2001) Growth retardation and increased apoptosis in mice with homozygous disruption of the akt1 gene. *Genes Dev* 15(17):2203–2208
 29. Cho H, Thorvaldsen JL, Chu Q, Feng F, Birnbaum MJ (2001) Akt1/PKB α Is Required for Normal Growth but Dispensable for Maintenance of Glucose Homeostasis in Mice. *J Biol Chem* 276(42):38349–38352
 30. Cho H, Mu J, Kim JK, Thorvaldsen JL, Chu Q, Crenshaw EB, Kaestner KH, Bartolomei MS, Shulman GI, Birnbaum MJ (2001) Insulin Resistance and a Diabetes Mellitus-Like Syndrome in Mice Lacking the Protein Kinase Akt2 (PKB β). *Science* 292(5522):1728–1731
 31. Tschopp O, Yang Z-Z, Brodbeck D, Dummler BA, Hemmings-Mieszczak M, Watanabe T, Michaelis T, Frahm J, Hemmings BA (2005) Essential role of protein kinase B γ (PKB γ /Akt3) in postnatal brain development but not in glucose homeostasis. *Development* 132(13):2943–2954
 32. Carpten JD, Faber AL, Horn C, Donoho GP, Briggs SL, Robbins CM, Hostetter G, Boguslawski S, Moses TY, Savage S, Uhlik M, Lin A, Du J, Qian Y-W, Zeckner DJ, Tucker-Kellogg G, Touchman J, Patel K, Mousses S, Bittner M, Schevitz R, Lai M-HT, Blanchard KL, Thomas JE (2007) A transforming mutation in the pleckstrin homology domain of AKT1 in cancer. *Nature* 448(7152):439–444
 33. Kim MS, Jeong EG, Yoo NJ, Lee SH (2008) Mutational analysis of oncogenic AKT E17K mutation in common solid cancers and acute leukaemias. *Br J Cancer* 98(9):1533–1535
 34. Bleeker FE, Lamba S, Zanon C, van Tilborg AA, Leenstra S, Troost D, Hulsebos T, Vandertop WP, Bardelli A (2009) Absence of *AKT1* Mutations in Glioblastoma. *PLoS ONE* 4(5):e5638

35. Askham JM, Platt F, Chambers PA, Snowden H, Taylor CF, Knowles MA (2009) AKT1 mutations in bladder cancer: identification of a novel oncogenic mutation that can co-operate with E17K. *Oncogene* 29(1):150–155
36. Kirkegaard T, Witton CJ, Edwards J, Nielsen KV, Jensen LB, Campbell FM, Cooke TG, Bartlett JMS (2010) Molecular alterations in AKT1, AKT2 and AKT3 detected in breast and prostatic cancer by FISH. *Histopathology* 56(2):203–211. doi:10.1111/j.1365-2559.2009.03467.x
37. Inoki K, Guan K-L (2006) Complexity of the TOR signaling network. *Trends Cell Biol* 16(4):206–212. doi:10.1016/j.tcb.2006.02.002
38. Hay N, Sonenberg N (2004) Upstream and downstream of mTOR. *Genes Dev* 18(16):1926–1945. doi:10.1101/gad.1212704
39. Sarbassov DD, Ali SM, Kim D-H, Guertin DA, Latek RR, Erdjument-Bromage H, Tempst P, Sabatini DM (2004) Rictor, a Novel Binding Partner of mTOR, Defines a Rapamycin-Insensitive and Raptor-Independent Pathway that Regulates the Cytoskeleton. *Curr Biol* 14(14):1296–1302
40. Jacinto E, Loewith R, Schmidt A, Lin S, Ruegg MA, Hall A, Hall MN (2004) Mammalian TOR complex 2 controls the actin cytoskeleton and is rapamycin insensitive. *Nat Cell Biol* 6(11):1122–1128
41. Jacinto E, Loewith R, Schmidt A, Lin S, Ruegg MA, Hall A, Hall MN (2004) Mammalian TOR complex 2 controls the actin cytoskeleton and is rapamycin insensitive. *Nat Cell Biol* 6(11):1122–1128. doi:10.1038/ncb1183
42. Bjornsson J, Short MP, Kwiatkowski DJ, Henske EP (1996) Tuberous sclerosis-associated renal cell carcinoma. Clinical, pathological, and genetic features. *Am J Pathol* 149(4):1201–1208
43. Garami A, Zwartkruis FJT, Nobukuni T, Joaquin M, Rocco M, Stocker H, Kozma SC, Hafen E, Bos JL, Thomas G (2003) Insulin Activation of Rheb, a Mediator of mTOR/S6K/4E-BP Signaling, Is Inhibited by TSC1 and 2. *Mol Cell* 11(6):1457–1466. doi:10.1016/s1097-2765(03)00220-x
44. Li Y, Corradetti MN, Inoki K, Guan KL (2004) TSC2: Filling the GAP in the mTOR signaling pathway. *Trends Biochem Sci* 29(1):32–38
45. Gao X, Zhang Y, Arrazola P, Hino O, Kobayashi T, Yeung RS, Ru B, Pan D (2002) Tsc tumour suppressor proteins antagonize amino-acid-TOR signalling. *Nat Cell Biol* 4(9):699–704
46. Zhang Y, Gao X, Saucedo LJ, Ru B, Edgar BA, Pan D (2003) Rheb is a direct target of the tuberous sclerosis tumour suppressor proteins. *Nat Cell Biol* 5(6):578–581
47. Jiang WG, Sampson J, Martin TA, Lee-Jones L, Watkins G, Douglas-Jones A, Mokbel K, Mansel RE (2005) Tuberlin and hamartin are aberrantly expressed and linked to clinical outcome in human breast cancer: the role of promoter methylation of TSC genes. *Eur J Cancer* 41(11):1628–1636. doi:10.1016/j.ejca.2005.03.023
48. Inoki K, Li Y, Xu T, Guan KL (2003) Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. *Genes Dev* 17(15):1829–1834. doi:10.1101/gad.1110003
49. Sato T, Nakashima A, Guo L, Tamanoi F (2009) Specific Activation of mTORC1 by Rheb G-protein in Vitro Involves Enhanced Recruitment of Its Substrate Protein. *J Biol Chem* 284(19):12783–12791
50. Aspuria P-J, Tamanoi F (2004) The Rheb family of GTP-binding proteins. *Cell Signal* 16(10):1105–1112. doi:10.1016/j.cellsig.2004.03.019
51. Nardella C, Chen Z, Salmena L, Carracedo A, Alimonti A, Egia A, Carver B, Gerald W, Cordon-Cardo C, Pandolfi PP (2008) Aberrant Rheb-mediated mTORC1 activation and Pten haploinsufficiency are cooperative oncogenic events. *Genes Dev* 22(16):2172–2177
52. Lu ZH, Shvartsman MB, Lee AY, Shao JM, Murray MM, Kladney RD, Fan D, Krajewski S, Chiang GG, Mills GB, Arbeit JM (2010) Mammalian target of rapamycin activator RHEB is frequently overexpressed in human carcinomas and is critical and sufficient for skin epithelial carcinogenesis. *Cancer Res* 70(8):3287–3298. doi:10.1158/0008-5472.CAN-09-3467
53. Mavrakis KJ, Zhu H, Silva RL, Mills JR, Teruya-Feldstein J, Lowe SW, Tam W, Pelletier J, Wendel HG (2008) Tumorigenic activity and therapeutic inhibition of Rheb GTPase. *Genes Dev* 22(16):2178–2188. doi:10.1101/gad.1690808
54. Mora A, Komander D, van Aalten DMF, Alessi DR (2004) PDK1, the master regulator of AGC kinase signal transduction. *Sem Cell Dev Biol* 15(2):161–170. doi:10.1016/j.semcdb.2003.12.022
55. Lin HJ, Hsieh FC, Song H, Lin J (2005) Elevated phosphorylation and activation of PDK-1/AKT pathway in human breast cancer. *Br J Cancer* 93(12):1372–1381
56. Jope RS, Johnson GVW (2004) The glamour and gloom of glycogen synthase kinase-3. *Trends Biochem Sci* 29(2):95–102. doi:10.1016/j.tibs.2003.12.004
57. Ougolkov AV, Fernandez-Zapico ME, Bilim VN, Smyrk TC, Chari ST, Billadeau DD (2006) Aberrant nuclear accumulation of glycogen synthase kinase-3 β in human pancreatic cancer: association with kinase activity and tumor dedifferentiation. *Clin Cancer Res* 12(17):5074–5081. doi:10.1158/1078-0432.CCR-06-0196
58. Wang Z, Smith KS, Murphy M, Piloto O, Somervaille TC, Cleary ML (2008) Glycogen synthase kinase 3 in MLL leukaemia maintenance and targeted therapy. *Nature* 455(7217):1205–1209. doi:10.1038/nature07284
59. Obsil T, Obsilova V (2008) Structure/function relationships underlying regulation of FOXO transcription factors. *Oncogene* 27(16):2263–2275
60. Abid MR, Guo S, Minami T, Spokes KC, Ueki K, Skurk C, Walsh K, Aird WC (2004) Vascular Endothelial Growth Factor Activates PI3K/Akt/Forkhead Signaling in Endothelial Cells. *Arterioscler Thromb Vasc Biol* 24(2):294–300. doi:10.1161/01.atv.0000110502.10593.06
61. Dey JH, Bianchi F, Voshol J, Bonenfant D, Oakeley EJ, Hynes NE (2010) Targeting Fibroblast Growth Factor Receptors Blocks PI3K/AKT Signaling, Induces Apoptosis, and Impairs Mammary Tumor Outgrowth and Metastasis. *Cancer Res* 70(10):4151–4162. doi:10.1158/0008-5472.can-09-4479
62. Palomero T, Sulis ML, Cortina M, Real PJ, Barnes K, Ciofani M, Caparros E, Buteau J, Brown K, Perkins SL, Bhagat G, Agarwal AM, Basso G, Castillo M, Nagase S, Cordon-Cardo C, Parsons R, Zuniga-Pflucker JC, Dominguez M, Ferrando AA (2007) Mutational loss of PTEN induces resistance to NOTCH1 inhibition in T-cell leukemia. *Nat Med* 13(10):1203–1210
63. Schnell CR, Stauffer F, Allegrini PR, O'Reilly T, McSheehy PMJ, Dartois C, Stumm M, Cozens R, Littlewood-Evans A, García-Echeverría C, Maira S-M (2008) Effects of the Dual Phosphatidylinositol 3-Kinase/Mammalian Target of Rapamycin Inhibitor NVP-BEZ235 on the Tumor Vasculature: Implications for Clinical Imaging. *Cancer Res* 68(16):6598–6607. doi:10.1158/0008-5472.can-08-1044
64. Ye M, Hu D, Tu L, Zhou X, Lu F, Wen B, Wu W, Lin Y, Zhou Z, Qu J (2008) Involvement of PI3K/Akt Signaling Pathway in Hepatocyte Growth Factor-Induced Migration of Uveal Melanoma Cells. *Invest Ophthalmol Vis Sci* 49(2):497–504. doi:10.1167/iovs.07-0975
65. Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, Teague J, Woffendin H, Garnett MJ, Bottomley W, Davis N, Dicks E, Ewing R, Floyd Y, Gray K, Hall S, Hawes R, Hughes J, Kosmidou V, Menzies A, Mould C, Parker A, Stevens C, Watt S, Hooper S, Wilson R, Jayatilake H, Gusterson BA, Cooper C,

- Shipley J, Hargrave D, Pritchard-Jones K, Maitland N, Chenevix-Trench G, Riggins GJ, Bigner DD, Palmieri G, Cossu A, Flanagan A, Nicholson A, Ho JW, Leung SY, Yuen ST, Weber BL, Seigler HF, Darrow TL, Paterson H, Marais R, Marshall CJ, Wooster R, Stratton MR, Futreal PA (2002) Mutations of the BRAF gene in human cancer. *Nature* 417 (6892):949–954. doi:10.1038/nature00766
66. Kefford R, Arkenau H, Brown MP, Millward M, Infante JR, Long GV, Ouellet D, Curtis M, Lebowitz PF, Falchook GS (2010) Phase I/II study of GSK2118436, a selective inhibitor of oncogenic mutant BRAF kinase, in patients with metastatic melanoma and other solid tumors. *ASCO Meet Abstr* 28 (15_suppl):8503
67. Patel SP, Lazar AJ, Mahoney S, Vaughn C, Gonzalez N, Papadopoulos NE, Liu P, Infante JR, LoRusso P, Kim KB (2010) Clinical responses to AZD6244 (ARRY-142886)-based combination therapy stratified by gene mutations in patients with metastatic melanoma. *ASCO Meet Abstr* 28(15_suppl):8501
68. Jiang CC, Lai F, Thorne RF, Yang F, Liu H, Hersey P, Zhang XD (2010) MEK-Independent Survival of B-RAFV600E Melanoma Cells Selected for Resistance to Apoptosis Induced by the RAF Inhibitor PLX4720. *Clin Cancer Res*. doi:10.1158/1078-0432.ccr-10-2225
69. Wan X, Harkavy B, Shen N, Grohar P, Helman LJ (2006) Rapamycin induces feedback activation of Akt signaling through an IGF-1R-dependent mechanism. *Oncogene* 26(13):1932–1940
70. O'Reilly KE, Rojo F, She Q-B, Solit D, Mills GB, Smith D, Lane H, Hofmann F, Hicklin DJ, Ludwig DL, Baselga J, Rosen N (2006) mTOR Inhibition Induces Upstream Receptor Tyrosine Kinase Signaling and Activates Akt. *Cancer Res* 66(3):1500–1508
71. Carew J et al (2011) Mechanisms of mTOR resistance in cancer therapy. *Targ Oncol*. doi:10.1007/s11523-011-0167-8
72. Engelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, Park JO, Lindeman N, Gale CM, Zhao X, Christensen J, Kosaka T, Holmes AJ, Rogers AM, Cappuzzo F, Mok T, Lee C, Johnson BE, Cantley LC, Janne PA (2007) MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Science* 316(5827):1039–1043. doi:10.1126/science.1141478
73. Kobayashi S, Boggon TJ, Dayaram T, Janne PA, Kocher O, Meyerson M, Johnson BE, Eck MJ, Tenen DG, Halmos B (2005) EGFR Mutation and Resistance of Non-Small-Cell Lung Cancer to Gefitinib. *New Eng J Med* 352(8):786–792. doi:10.1056/NEJMoa044238
74. Hughes T, Deininger M, Hochhaus A, Branford S, Radich J, Kaeda J, Baccarani M, Cortes J, Cross NCP, Druker BJ, Gabert J, Grimwade D, Hehlmann R, Kamel-Reid S, Lipton JH, Longtine J, Martinelli G, Saglio G, Soverini S, Stock W, Goldman JM (2006) Monitoring CML patients responding to treatment with tyrosine kinase inhibitors: review and recommendations for harmonizing current methodology for detecting BCR-ABL transcripts and kinase domain mutations and for expressing results. *Blood* 108(1):28–37. doi:10.1182/blood-2006-01-0092
75. Wymann MP, Bulgarelli-Leva G, Zvelebil MJ, Pirola L, Vanhaesebroeck B, Waterfield MD, Panayotou G (1996) Wortmannin inactivates phosphoinositide 3-kinase by covalent modification of Lys-802, a residue involved in the phosphate transfer reaction. *Mol Cell Biol* 16(4):1722–1733
76. Davies SP, Reddy H, Caivano M, Cohen P (2000) Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochem J* 351(Pt 1):95–105
77. Lempiainen H, Halazonetis TD (2009) Emerging common themes in regulation of PIKKs and PI3Ks. *EMBO J* 28 (20):3067–3073. doi:10.1038/emboj.2009.281
78. Dennis PB, Fumagalli S, Thomas G (1999) Target of rapamycin (TOR): balancing the opposing forces of protein synthesis and degradation. *Curr Opin Gen Dev* 9(1):49–54. doi:10.1016/S0959-437X(99)80007-0
79. Vlahos CJ, Matter WF, Hui KY, Brown RF (1994) A specific inhibitor of phosphatidylinositol 3-kinase, 2-(4-morpholinyl)-8-phenyl-4 H-1-benzopyran-4-one (LY294002). *J Biol Chem* 269 (7):5241–5248
80. Hu L, Hofmann J, Lu Y, Mills GB, Jaffe RB (2002) Inhibition of Phosphatidylinositol 3'-Kinase Increases Efficacy of Paclitaxel in In Vitro and in Vivo Ovarian Cancer Models. *Cancer Res* 62 (4):1087–1092
81. Garlich JR, Becker MD, Shelton CF, Qi W, Liu X, Cooke L, Mahadevan D (2010) Phase I Study of Novel Prodrug Dual PI3K/mTOR Inhibitor SF1126 In B-Cell Malignancies. *ASH Annu Meet Abstr* 116(21):1783
82. Arcaro A, Wymann MP (1993) Wortmannin is a potent phosphatidylinositol 3-kinase inhibitor: the role of phosphatidylinositol 3,4,5-trisphosphate in neutrophil responses. *Biochem J* 296(2):297–301
83. Schultz RM, Merriman RL, Andis SL, Bonjouklian R, Grindey GB, Rutherford PG, Gallegos A, Massey K, Powis G (1995) In vitro and in vivo antitumor activity of the phosphatidylinositol-3-kinase inhibitor, wortmannin. *Anticancer Res* 15(4):1135–1139
84. Ihle NT, Williams R, Chow S, Chew W, Berggren MI, Paine-Murrieta G, Minion DJ, Halter RJ, Wipf P, Abraham R, Kirkpatrick L, Powis G (2004) Molecular pharmacology and antitumor activity of PX-866, a novel inhibitor of phosphoinositide-3-kinase signaling. *Mol Cancer Ther* 3(7):763–772
85. Jimeno A, Herbst RS, Falchook GS, Messersmith WA, Hecker S, Peterson S, Hausman DF, Kurzrock R, Eckhardt SG, Hong DS (2010) Final results from a phase I, dose-escalation study of PX-866, an irreversible, pan-isoform inhibitor of PI3 kinase. *ASCO Meet Abstr* 28(15_suppl):3089
86. Burris H, Rodon J, Sharma S, Herbst RS, Tabernero J, Infante JR, Silva A, Demanase D, Hackl W, Baselga J (2010) First-in-human phase I study of the oral PI3K inhibitor BEZ235 in patients (pts) with advanced solid tumors. *ASCO Meet Abstr* 28 (15_suppl):3005
87. Voliva cF, Pecchi S, Burger M, Nagel T, Schnell C, Fritsch c, Brachmann s, Menezes D, Knapp M, Shoemaker K, Wiesmann M, Huh k, Zaror I, Dorsch M, Sellers WR, Garcia-Echeverria C, Maira M (2011) Abstract 4498: Biological characterization of NVP-BKM120, a novel inhibitor of phosphoinositide 3-kinase in Phase I/II clinical trials. *Cancer Res* 70(8 Supplement):4498. doi:10.1158/1538-7445.am10-4498
88. Maira M, Menezes D, Pecchi S, Shoemaker K, Burger M, Schnell c, Fritsch c, Brachmann S, Nagel T, Sellers WR, Garcia-Echeverria C, Wiesmann M, Voliva cF (2010) NVP-BKM120, a novel inhibitor of phosphoinositide 3-kinase in Phase I/II clinical trials, shows significant antitumor activity in xenograft and primary tumor models Paper presented at the AACR 101st Annual Meeting
89. Baselga J, De Jonge MJ, Rodon J, Burris HA, Birl DC, De Buck SS, Demanase D, Ru QC, Goldbrunner M, Bendell JC (2010) A first-in-human phase I study of BKM120, an oral pan-class I PI3K inhibitor, in patients (pts) with advanced solid tumors. *ASCO Meet Abstr* 28(15_suppl):3003
90. Repasky GA, Chenette EJ, Der CJ (2004) Renewing the conspiracy theory debate: does Raf function alone to mediate Ras oncogenesis? *Trends Cell Biol* 14(11):639–647. doi:10.1016/j.tcb.2004.09.014
91. Wee S, Jagani Z, Xiang KX, Loo A, Dorsch M, Yao Y-M, Sellers WR, Lengauer C, Stegmeier F (2009) PI3K Pathway Activation Mediates Resistance to MEK Inhibitors in KRAS Mutant Cancers. *Cancer Res* 69(10):4286–4293
92. Matsuzaki T, Sasaki K, Tanizaki Y, Hata J, Fujimi K, Matsui Y, Sekita A, Suzuki SO, Kanba S, Kiyohara Y, Iwaki T (2010)

- Insulin resistance is associated with the pathology of Alzheimer disease: the Hisayama study. *Neurology* 75(9):764–770. doi:10.1212/WNL.0b013e3181ee25f
93. Bandaru SS, Lin K, Roming SL, Vellipuram R, Harney JP (2010) Effects of PI3K inhibition and low docosahexaenoic acid on cognition and behavior. *Phys Beh* 100(3):239–244. doi:10.1016/j.physbeh.2009.10.019
 94. Edelman G, Bedell C, Shapiro G, Pandya SS, Kwak EL, Scheffold C, Nguyen LT, Laird A, Baselga J, Rodon J (2010) A phase I dose-escalation study of XL147 (SAR245408), a PI3K inhibitor administered orally to patients (pts) with advanced malignancies. *ASCO Meet Abstr* 28(15_suppl):3004
 95. Moldovan C, Soria J, LoRusso P, Guthrie T, Song C, Nguyen LT, Martini J, Infante JR, Burris HA (2010) A phase I safety and pharmacokinetic (PK) study of the PI3K inhibitor XL147 (SAR245408) in combination with erlotinib in patients (pts) with advanced solid tumors. *ASCO Meet Abstr* 28(15_suppl):3070
 96. Traynor AM, Kurzrock R, Bailey HH, Attia S, Scheffold C, van Leeuwen B, Wu B, Falchook GS, Moulder SL, Wheler J (2010) A phase I safety and pharmacokinetic (PK) study of the PI3K inhibitor XL147 (SAR245408) in combination with paclitaxel (P) and carboplatin (C) in patients (pts) with advanced solid tumors. *ASCO Meet Abstr* 28(15_suppl):3078
 97. Brana I, LoRusso P, Baselga J, Heath EI, Patnaik A, Gendreau S, Laird A, Papadopoulos K (2010) A phase I dose-escalation study of the safety, pharmacokinetics (PK), and pharmacodynamics of XL765 (SAR245409), a PI3K/TORC1/TORC2 inhibitor administered orally to patients (pts) with advanced malignancies. *ASCO Meet Abstr* 28(15_suppl):3030
 98. Cohen RB, Janne PA, Engelman JA, Martinez P, Nishida Y, Gendreau S, Wu B, Felipe E (2010) A phase I safety and pharmacokinetic (PK) study of PI3K/TORC1/TORC2 inhibitor XL765 (SAR245409) in combination with erlotinib (E) in patients (pts) with advanced solid tumors. *ASCO Meet Abstr* 28(15_suppl):3015
 99. Nghiemphu PL, Omuro AM, Cloughesy T, Mellingshoff IK, Norden AD, Nguyen LT, Rajangam K, Wen PY (2010) A phase I safety and pharmacokinetic study of XL765 (SAR245409), a novel PI3K/TORC1/TORC2 inhibitor, in combination with temozolomide (TMZ) in patients (pts) with newly diagnosed malignant glioma. *ASCO Meet Abstr* 28(15_suppl):3085
 100. Raynaud FI, Eccles SA, Patel S, Alix S, Box G, Chuckowree I, Folkes A, Gowan S, De Haven BA, Di Stefano F, Hayes A, Henley AT, Lensun L, Pergl-Wilson G, Robson A, Saghir N, Zhyvoloup A, McDonald E, Sheldrake P, Shuttleworth S, Valenti M, Wan NC, Clarke PA, Workman P (2009) Biological properties of potent inhibitors of class I phosphatidylinositol 3-kinases: from PI-103 through PI-540, PI-620 to the oral agent GDC-0941. *Mol Cancer Ther* 8(7):1725–1738
 101. Hoff DDV, LoRusso P, Tibes R, Shapiro G, Weiss GJ, Ware JA, Fredrickson J, Mazina KE, Levy GG, Wagner AJ (2010) A first-in-human phase I study to evaluate the pan-PI3K inhibitor GDC-0941 administered QD or BID in patients with advanced solid tumors. *ASCO Meet Abstr* 28(15_suppl):2541
 102. Baird RD, Kristeleit RS, Sarker D, Olmos D, Sandhu SK, Yan Y, Koeppen H, Levy GG, Jin J, Bono JSD (2010) A phase I study evaluating the pharmacokinetics (PK) and pharmacodynamics (PD) of the oral pan-phosphoinositide-3 kinase (PI3K) inhibitor GDC-0941. *ASCO Meet Abstr* 28(15_suppl):2613
 103. Vanhaesebroeck B, Welham MJ, Kotani K, Stein R, Warne PH, Zvelebil MJ, Higashi K, Volinia S, Downward J, Waterfield MD (1997) p110 δ , a novel phosphoinositide 3-kinase in leukocytes. *Proc Nat Acad Sci USA* 94(9):4330–4335
 104. Herman SEM, Gordon AL, Wagner AJ, Heerema NA, Zhao W, Flynn JM, Jones J, Andritsos L, Puri KD, Lannutti BJ, Giese NA, Zhang X, Wei L, Byrd JC, Johnson AJ (2010) Phosphatidylinositol 3-kinase- δ inhibitor CAL-101 shows promising preclinical activity in chronic lymphocytic leukemia by antagonizing intrinsic and extrinsic cellular survival signals. *Blood* 116(12):2078–2088
 105. Furman RR, Byrd JC, Brown JR, Coutre SE, Benson DM Jr, Wagner-Johnston ND, Flinn IW, Kahl BS, Spurgeon SE, Lannutti B, Giese NA, Webb HK, Ulrich RG, Peterman S, Holes LM, Yu AS (2010) CAL-101, An Isoform-Selective Inhibitor of Phosphatidylinositol 3-Kinase P110 δ , Demonstrates Clinical Activity and Pharmacodynamic Effects In Patients with Relapsed or Refractory Chronic Lymphocytic Leukemia. *ASH Annu Meet Abstr* 116(21):55
 106. Kahl B, Byrd JC, Flinn IW, Wagner-Johnston N, Spurgeon S, Benson DM Jr, Furman RR, Brown JR, Coutre S, Lannutti B, Giese NA, Ulrich RG, Webb HK, Peterman S, Holes L, Yu AS (2010) Clinical Safety and Activity In a Phase I Study of CAL-101, An Isoform-Selective Inhibitor of Phosphatidylinositol 3-Kinase P110 δ , In Patients with Relapsed or Refractory Non-Hodgkin Lymphoma. *ASH Annu Meet Abstr* 116(21):1777
 107. Ferry DR, Smith A, Malkhandi J, Fyfe DW, deTakats PG, Anderson D, Baker J, Kerr DJ (1996) Phase I clinical trial of the flavonoid quercetin: pharmacokinetics and evidence for in vivo tyrosine kinase inhibition. *Clin Cancer Res* 2(4):659–668
 108. Kong D, Dan S, Yamazaki K, Yamori T (2010) Inhibition profiles of phosphatidylinositol 3-kinase inhibitors against PI3K superfamily and human cancer cell line panel JFCR39. *Eur J Cancer* 46(6):1111–1121. doi:10.1016/j.ejca.2010.01.005
 109. Dolly S, Wagner AJ, Bendell JC, Yan Y, Ware JA, Mazina KE, Holden SN, Derynck MK, Bono JSD, H. A. Burris I (2010) A first-in-human, phase I study to evaluate the dual PI3K/mTOR inhibitor GDC-0980 administered QD in patients with advanced solid tumors or non-Hodgkin's lymphoma. *ASCO Meet Abstr* 28(15_suppl):3079
 110. Cheng H, Bagrodia S, Bailey S, Edwards M, Hoffman J, Hu Q, Kania R, Knighton DR, Marx MA, Ninkovic S, Sun S, Zhang E (2010) Discovery of the highly potent PI3K/mTOR dual inhibitor PF-04691502 through structure based drug design. *Med Chem Commun* 1(2):139–144
 111. Knight SD, Adams ND, Burgess JL, Chaudhari AM, Darcy MG, Donatelli CA, Luengo JI, Newlander KA, Parrish CA, Ridgers LH, Sarpong MA, Schmidt SJ, Van Aller GS, Carson JD, Diamond MA, Elkins PA, Gardiner CM, Garver E, Gilbert SA, Gontarek RR, Jackson JR, Kershner KL, Luo L, Raha K, Sherk CS, Sung C-M, Sutton D, Tummino PJ, Wegrzyn RJ, Auger KR, Dhanak D (2010) Discovery of GSK2126458, a Highly Potent Inhibitor of PI3K and the Mammalian Target of Rapamycin. *ACS Med Chem Lett* 1(1):39–43. doi:10.1021/ml900028r
 112. Kong D, S-i Y, Yamori T (2009) Effect of ZSTK474, a Novel Phosphatidylinositol 3-Kinase Inhibitor, on DNA-Dependent Protein Kinase. *Biol Pharm Bull* 32(2):297–300
 113. van Blitterswijk WJ, Hilkmann H, Storme GA (1987) Accumulation of an alkyl lysophospholipid in tumor cell membranes affects membrane fluidity and tumor cell invasion. *Lipids* 22(11):820–823
 114. Ruiter GA, Zerp SF, Bartelink H, van Blitterswijk WJ, Verheij M (2003) Anti-cancer alkyl-lysophospholipids inhibit the phosphatidylinositol 3-kinase-Akt/PKB survival pathway. *Anticancer Drugs* 14(2):167–173
 115. Sundar S, Jha TK, Thakur CP, Engel J, Sindermann H, Fischer C, Junge K, Bryceson A, Berman J (2002) Oral Miltefosine for Indian Visceral Leishmaniasis. *New Eng J Med* 347(22):1739–1746. doi:10.1056/NEJMoa021556
 116. Leonard R, Hardy J, van Tienhoven G, Houston S, Simmonds P, David M, Mansi J (2001) Randomized, Double-Blind, Placebo-Controlled, Multicenter Trial of 6% Miltefosine Solution, a

- Topical Chemotherapy in Cutaneous Metastases From Breast Cancer. *J Clin Oncol* 19(21):4150–4159
117. Planting AS, Stoter G, Verweij J (1993) Phase II study of daily oral miltefosine (hexadecylphosphocholine) in advanced colorectal cancer. *Eur J Cancer* 29A(4):518–519
 118. Kondapaka SB, Singh SS, Dasmahapatra GP, Sausville EA, Roy KK (2003) Perifosine, a novel alkylphospholipid, inhibits protein kinase B activation. *Mol Cancer Ther* 2(11):1093–1103
 119. Argriris A, Cohen E, Karrison T, Esparaz B, Mauer A, Ansari R, Wong S, Lu Y, Pins M, Dancey J, Vokes E (2006) A phase II trial of perifosine, an oral alkylphospholipid, in recurrent or metastatic head and neck cancer. *Cancer Biol Ther* 5(7):766–770
 120. Ernst D, Eisenhauer E, Wainman N, Davis M, Lohmann R, Baetz T, Belanger K, Smylie M (2005) Phase II Study of Perifosine in Previously Untreated Patients with Metastatic Melanoma. *Investig New Drugs* 23(6):569–575. doi:10.1007/s10637-005-1157-4
 121. Leighl N, Dent S, Clemons M, Vandenberg T, Tozer R, Warr D, Crump R, Hedley D, Pond G, Dancey J, Moore M (2008) A Phase 2 study of perifosine in advanced or metastatic breast cancer. *Breast Cancer Res Treat* 108(1):87–92. doi:10.1007/s10549-007-9584-x
 122. Friedman DR, Davis PH, Lanasa MC, Moore JO, Gockerman JP, Nelson T, Bond KM, Jiang N, Davis ED, Allgood SD, Chen Y, Sportelli P, Weinberg JB (2010) Pre-Clinical and Interim Results of a Phase II Trial of Perifosine In Patients with Relapsed or Refractory Chronic Lymphocytic Leukemia (CLL). *ASH Annu Meet Abstr* 116(21):1842
 123. Bailey HH, Mahoney MR, Ettinger DS, Maples WJ, Fracasso PM, Traynor AM, Erlichman C, Okuno SH (2006) Phase II study of daily oral perifosine in patients with advanced soft tissue sarcoma. *Cancer* 107(10):2462–2467. doi:10.1002/ncr.22308
 124. Ghobrial IM, Roccaro A, Hong F, Weller E, Rubin N, Leduc R, Rourke M, Chuma S, Sacco A, Jia X, Azab F, Azab AK, Rodig S, Warren D, Harris B, Varticovski L, Sportelli P, Leleu X, Anderson KC, Richardson PG (2010) Clinical and Translational Studies of a Phase II Trial of the Novel Oral Akt Inhibitor Perifosine in Relapsed or Relapsed/Refractory Waldenström's Macroglobulinemia. *Clin Cancer Res* 16(3):1033–1041
 125. Yang L, Dan HC, Sun M, Liu Q, Sun XM, Feldman RI, Hamilton AD, Polokoff M, Nicosia SV, Herlyn M, Sehti SM, Cheng JQ (2004) Akt/protein kinase B signaling inhibitor-2, a selective small molecule inhibitor of Akt signaling with antitumor activity in cancer cells overexpressing Akt. *Cancer Res* 64(13):4394–4399. doi:10.1158/0008-5472.CAN-04-0343
 126. Feun LG, Blessing JA, Barrett RJ, Hanjani P (1993) A phase II trial of tricyclic nucleoside phosphate in patients with advanced squamous cell carcinoma of the cervix. A Gynecologic Oncology Group Study. *Am J Clin Oncol* 16(6):506–508
 127. TA Yap AP, Fearon I, Olmos D, Papadopoulos K, Tunariu N, Sullivan D, Yan L, De Bono JS, Tolcher AW (2010) First-in-class phase I trial of a selective Akt inhibitor, MK2206 (MK), evaluating alternate day (QOD) and once weekly (QW) doses in advanced cancer patients (pts) with evidence of target modulation and antitumor activity. *ASCO Annu Meet* 28(15_suppl):3009
 128. Yan L (2009) MK-2206: A potent oral allosteric AKT inhibitor. *AACR Annual Meeting:DDT01-01*
 129. Mittelman A, Casper ES, Godwin TA, Cassidy C, Young CW (1983) Phase I study of tricyclic nucleoside phosphate. *Cancer Treat Rep* 67(2):159–162
 130. Van Ummersen L, Binger K, Volkman J, Marnocha R, Tutsch K, Kolesar J, Arzoumanian R, Alberti D, Wilding G (2004) A Phase I Trial of Perifosine (NSC 639966) on a Loading Dose/Maintenance Dose Schedule in Patients with Advanced Cancer. *Clin Cancer Res* 10(22):7450–7456
 131. Rhodes N, Heerding DA, Duckett DR, Eberwein DJ, Knick VB, Lansing TJ, McConnell RT, Gilmer TM, Zhang S-Y, Robell K, Kahana JA, Geske RS, Kleymanova EV, Choudhry AE, Lai Z, Leber JD, Minthorn EA, Strum SL, Wood ER, Huang PS, Copeland RA, Kumar R (2008) Characterization of an Akt Kinase Inhibitor with Potent Pharmacodynamic and Antitumor Activity. *Cancer Res* 68(7):2366–2374
 132. Arico S, Pattingre S, Bauvy C, Gane P, Barbat A, Codogno P, Ogier-Denis E (2002) Celecoxib Induces Apoptosis by Inhibiting 3-Phosphoinositide-dependent Protein Kinase-1 Activity in the Human Colon Cancer HT-29 Cell Line. *J Biol Chem* 277(31):27613–27621
 133. Gadgeel SM, Ruckdeschel JC, Heath EI, Heilbrun LK, Venkatramanamoorthy R, Wozniak A (2007) Phase II Study of Gefitinib, an Epidermal Growth Factor Receptor Tyrosine Kinase Inhibitor (EGFR-TKI), and Celecoxib, a Cyclooxygenase-2 (COX-2) Inhibitor, in Patients with Platinum Refractory Non-small Cell Lung Cancer (NSCLC). *J Thorac Oncol* 2(4):299–305
 134. Pierga J-Y, Delaloge S, Espié M, Brain E, Sigal-Zafrani B, Mathieu M-C, Bertheau P, Guinebretière J, Spielmann M, Savignoni A, Marty M (2010) A multicenter randomized phase II study of sequential epirubicin/cyclophosphamide followed by docetaxel with or without celecoxib or trastuzumab according to HER2 status, as primary chemotherapy for localized invasive breast cancer patients. *Breast Cancer Res Treat* 122(2):429–437. doi:10.1007/s10549-010-0939-3
 135. Zhu J, Huang JW, Tseng PH, Yang YT, Fowble J, Shiau CW, Shaw YJ, Kulp SK, Chen CS (2004) From the cyclooxygenase-2 inhibitor celecoxib to a novel class of 3-phosphoinositide-dependent protein kinase-1 inhibitors. *Cancer Res* 64(12):4309–4318. doi:10.1158/0008-5472.CAN-03-4063
 136. Zhang S, Suvannasankha A, Crean CD, White VL, Johnson A, Chen C-S, Farag SS (2007) OSU-03012, a Novel Celecoxib Derivative, Is Cytotoxic to Myeloma Cells and Acts through Multiple Mechanisms. *Clin Cancer Res* 13(16):4750–4758
 137. Johnson AJ, Smith LL, Zhu J, Heerema NA, Jefferson S, Mone A, Grever M, Chen CS, Byrd JC (2005) A novel celecoxib derivative, OSU03012, induces cytotoxicity in primary CLL cells and transformed B-cell lymphoma cell line via a caspase- and Bcl-2-independent mechanism. *Blood* 105(6):2504–2509. doi:10.1182/blood-2004-05-1957
 138. McCubrey JA, Lahair MM, Franklin RA (2006) OSU-03012 in the treatment of glioblastoma. *Mol Pharmacol* 70(2):437–439. doi:10.1124/mol.106.026252
 139. Sato S, Fujita N, Tsuruo T (2002) Interference with PDK1-Akt survival signaling pathway by UCN-01 (7-hydroxystaurosporine). *Oncogene* 21(11):1727–1738. doi:10.1038/sj.onc.1205225
 140. Sausville EA, Arbuck SG, Messmann R, Headlee D, Bauer KS, Lush RM, Murgo A, Figg WD, Lahusen T, Jaken S, X-x J, Roberge M, Fuse E, Kuwabara T, Senderowicz AM (2001) Phase I Trial of 72-Hour Continuous Infusion UCN-01 in Patients With Refractory Neoplasms. *J Clin Oncol* 19(8):2319–2333
 141. Graves PR, Yu L, Schwarz JK, Gales J, Sausville EA, O'Connor PM, Piwnicka-Worms H (2000) The Chk1 Protein Kinase and the Cdc25C Regulatory Pathways Are Targets of the Anticancer Agent UCN-01. *J Biol Chem* 275(8):5600–5605
 142. Feldman RI, Wu JM, Polokoff MA, Kochanny MJ, Dinter H, Zhu D, Biroc SL, Alicke B, Bryant J, Yuan S, Buckman BO, Lentz D, Ferrer M, Whitlow M, Adler M, Finster S, Chang Z, Arnaiz DO (2005) Novel Small Molecule Inhibitors of 3-Phosphoinositide-dependent Kinase-1. *J Biol Chem* 280(20):19867–19874. doi:10.1074/jbc.M501367200
 143. Najafav A, Sommer EM, Axten JM, Deyoung MP, Alessi DR (2010) Characterization of GSK2334470, a novel and highly specific inhibitor of PDK1. *Biochem J* 433(2):357–369. doi:10.1042/BJ20101732

144. Weisberg E, Banerji L, Wright RD, Barrett R, Ray A, Moreno D, Catley L, Jiang J, Hall-Meyers E, Sauveur-Michel M, Stone R, Galinsky I, Fox E, Kung AL, Griffin JD (2008) Potentiation of antileukemic therapies by the dual PI3K/PDK-1 inhibitor, BAG956: effects on BCR-ABL- and mutant FLT3-expressing cells. *Blood* 111(7):3723–3734. doi:[10.1182/blood-2007-09-114454](https://doi.org/10.1182/blood-2007-09-114454)
145. Gopalsamy A, Shi M, Boschelli DH, Williamson R, Olland A, Hu Y, Krishnamurthy G, Han X, Arndt K, Guo B (2007) Discovery of Dibenzo[c, f][2, 7]naphthyridines as Potent and Selective 3-Phosphoinositide-Dependent Kinase-1 Inhibitors. *J Med Chem* 50(23):5547–5549. doi:[10.1021/jm070851i](https://doi.org/10.1021/jm070851i)
146. Janku F, Tsimberidou AM, Garrido-Laguna I, Hong DS, Naing A, Falchook GS, Wheler JJ, Fu S, Piha-Paul SA, Kurzrock R (2010) PIK3CA, KRAS, and BRAF mutations in patients with advanced cancers treated with PI3K/AKT/mTOR axis inhibitors. *ASCO Meet Abstr* 28(15_suppl):2583