

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/10757131>

Substituted 2-Pyridinemethanol derivatives as potent and selective phosphodiesterase-4 inhibitors

ARTICLE in BIOORGANIC & MEDICINAL CHEMISTRY LETTERS · JULY 2003

Impact Factor: 2.42 · DOI: 10.1016/S0960-894X(03)00314-7 · Source: PubMed

CITATIONS

11

READS

29

13 AUTHORS, INCLUDING:



Diane Ethier

Merck Canada

50 PUBLICATIONS 2,790 CITATIONS

SEE PROFILE



Joseph A Mancini

Vertex Pharmaceuticals

105 PUBLICATIONS 6,542 CITATIONS

SEE PROFILE



Angela Styhler

McGill University

19 PUBLICATIONS 401 CITATIONS

SEE PROFILE



Robert Young

Simon Fraser University

124 PUBLICATIONS 3,370 CITATIONS

SEE PROFILE

Substituted 2-Pyridinemethanol Derivatives as Potent and Selective Phosphodiesterase-4 Inhibitors

Yves Ducharme,* Richard W. Friesen, Marc Blouin, Bernard Côté, Daniel Dubé, Diane Ethier, Richard Frenette, France Laliberté, Joseph A. Mancini, Paul Masson, Angela Styhler, Robert N. Young and Yves Girard

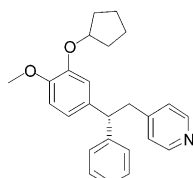
Merck Frosst Centre for Therapeutic Research, PO Box 1005, Pointe Claire-Dorval, Québec, Canada H9R 4P8

Received 29 October 2002; accepted 12 February 2003

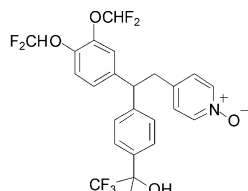
Abstract—The synthesis and the phosphodiesterase-4 (PDE4) inhibitory activity of 2-pyridinemethanol derivatives is described. The evaluation of the structure–activity relationship (SAR) in this series of novel PDE4 inhibitors led to the identification of compound **9** which exhibits excellent in vitro activity, desirable pharmacokinetic parameters and good efficacy in animal models of bronchoconstriction.

© 2003 Elsevier Science Ltd. All rights reserved.

Cyclic nucleotide phosphodiesterases (PDEs) constitute a broad family of enzymes responsible for the hydrolysis and consequent deactivation of the second messengers cAMP and cGMP.¹ The cAMP specific PDE4 isozymes,² encoded by four genes (A–D), are particularly abundant in inflammatory and immune cells and in airway smooth muscles.³ It is now well established that inhibition of PDE4 results in antiinflammatory and immunomodulatory activities, both in vitro and in animal models.⁴ A number of PDE4 inhibitors are under clinical evaluation for the treatment of asthma, chronic obstructive pulmonary disease (COPD) and atopic dermatitis.⁵ Despite promising results, the therapeutic potential of PDE4 inhibitors remains hampered by their dose-limiting side effects such as nausea and emesis.⁶ Therefore, the search for novel PDE4 inhibitors with superior therapeutic index remains a very active field of research.^{5,7}

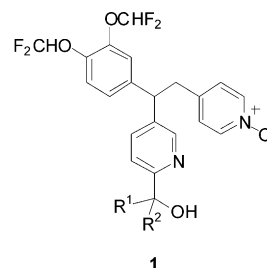


CDP840



L-791,943

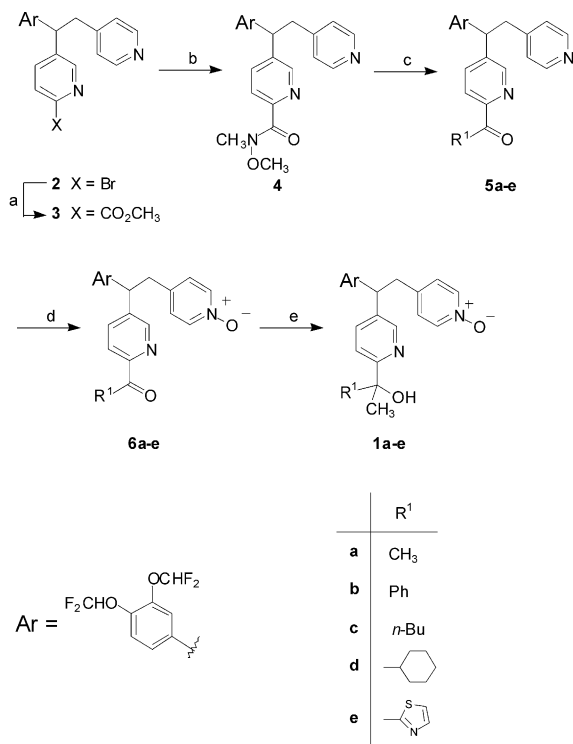
Recently, we reported how a SAR study directed toward the improvement of the metabolic stability of PDE4 inhibitor CDP840⁸ led to the discovery of compound L-791,943.⁹ The development of L-791,943 was precluded, despite an otherwise favorable pharmacological profile, by an excessively long half-life in a variety of animal species. The introduction of a soft metabolic site¹⁰ to the structure of L-791,943 and the substitution of the metabolically resistant bis(trifluoromethyl)carbinolbenzene moiety by an aminopyridine residue¹¹ both produced inhibitors with improved pharmacokinetic profiles. We now report on the optimization of a novel series of triarylethane derivatives of general structure **1**, bearing a 2-pyridinemethanol residue.¹²



The synthesis of PDE4 inhibitors **1** is presented in Scheme 1. Palladium catalyzed methoxycarbonylation¹³ of bromopyridine **2**¹¹ afforded in 95% yield the ester **3**, which was transformed to the Weinreb amide¹⁴ **4**.

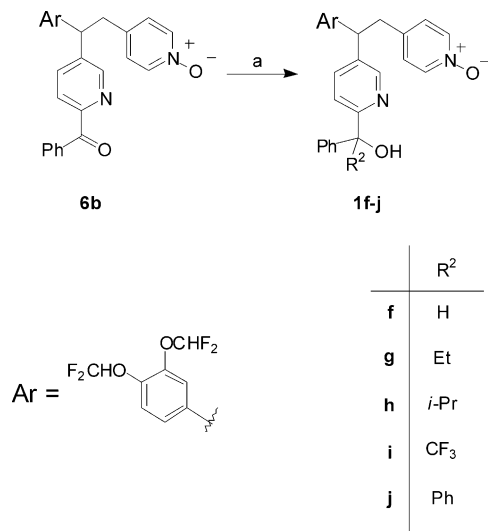
Addition of a variety of organometallic reagents to this intermediate gave access to ketones **5a–e**. Selective

*Corresponding author. Tel.: +1-514-428-8621; fax: +1-514-428-4900; e-mail: yves_ducharme@merck.com



Scheme 1. Reagents and conditions: (a) CO, CH₃OH, Pd(OAc)₂, dppf, Et₃N, DMF, 60 °C, 95%; (b) CH₃O(CH₃)NH·HCl, CH₃MgBr, THF, -78 °C, 91%; (c) **5a**: CH₃MgBr, THF, 0 °C, 100%; **5b**: PhMgCl, THF, -78 °C, 51%; **5c**: *n*-BuLi, THF, -78 °C, 51%; **5d**: cyclohexylmagnesium chloride, THF, 0 °C; **5e**: thiazole, *n*-BuLi, THF, -78 °C, 98%; (d) MMPP, CH₂Cl₂/CH₃OH (9/1), rt; (e) CH₃MgBr, CH₂Cl₂, -78 °C, 50–80%.

oxidation of the monosubstituted pyridine ring was then accomplished by treatment with magnesium monoperoxyphthalate (MMPP) to afford pyridine-*N*-oxides **6a–e**. Addition of methylmagnesium bromide to these ketones gave the desired tertiary alcohols **1a–e**. Analogues **1f–j** were obtained by the addition of the requisite organometallic reagents to phenylketone **6b** (Scheme 2). Use of ethylmagnesium bromide gave access to secondary



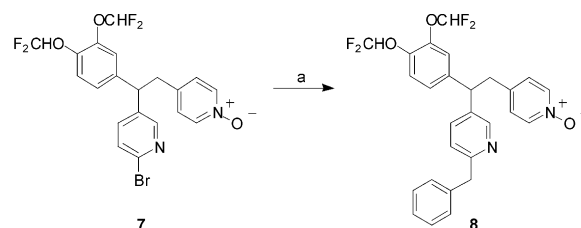
Scheme 2. Reagents and conditions: (a) **1f** and **1g**: EtMgBr, CH₂Cl₂, -78 °C, 35 and 50%; **1h**: *i*-PrMgBr, CH₂Cl₂, -78 °C, 47%; **1i**: TMSCF₃/TBAF, THF, 0 °C, 71%; **1j**: PhMgCl, CH₂Cl₂, -78 °C, 39%.

and tertiary alcohols **1f** and **1g** in yields of 35 and 50% respectively. Benzyl derivative **8** was obtained by palladium catalyzed coupling of bromopyridine **7**¹¹ with benzylmagnesium bromide in 73% yield (Scheme 3).

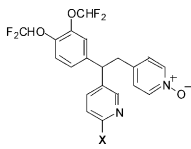
The compounds prepared were evaluated for their potency to inhibit the PDE4A enzyme *in vitro*¹⁵ and for their ability to inhibit LPS-induced TNF- α formation in human whole blood (HWB).¹⁶ Initially, the effect of a variety of substituents at position 2 of the pyridine ring was evaluated (Table 1). Replacement of the methyl group present in compound **6a** by a lipophilic phenyl group produces ketone **6b**, an inhibitor showing a 4-fold improvement in potency. The analogous secondary alcohol **1f** is even more potent, while the reduced benzyl analogue **8** shows a decreased ability to inhibit LPS-induced TNF- α formation in HWB. The favorable effect brought by a hydroxyl group at this position is confirmed by the fact that tertiary alcohol **1b** is the most potent inhibitor in this series with an IC₅₀ of 0.6 nM in the PDE4A assay and 0.35 μ M in the HWB assay.






Next, the effect of substitution at the carbinol position was investigated (Table 2). Reduction of lipophilicity (methyl derivative **1a**) has a negative effect on potency. Non-aromatic analogues **1c** and **1d** are equipotent to phenyl derivative **1b** in the HWB assay despite lower potencies in the enzyme assay. Replacement of the phenyl group by heterocycles (e.g., thiazole **1e**) did not improve the inhibitory profile. The phenyl analogue **1b** remains the best PDE4 inhibitor in this series. Substitution of the methyl group (R²), at the carbinol position, by larger alkyl (**1g** and **1h**), trifluoromethyl (**1i**) or phenyl (**1j**) units did not have significant effects on potency (Table 3). Tertiary alcohol **1b** was selected for further evaluation.

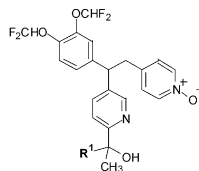
Pure diastereomers of inhibitor **1b** were obtained in the following way (Scheme 4). First, the enantiomers of ketone **6b** were resolved by chromatography, using a Chiralpak AD HPLC column (25% *i*-PrOH in hexanes). Under these conditions, the PDE4 inhibitory activity resides mainly in the fast-eluting enantiomer (–)-**6b** (data not shown). Enantiomer (–)-**6b** was then converted to **1b** diastereomers 1 and 2 by treatment with methylmagnesium bromide. A second chromatographic separation on the Chiralpak AD HPLC column (25% EtOH in hexanes) yielded the fast-eluting diastereomer 1 (**9**) and the slow-eluting diastereomer 2 (**10**). The same procedure applied to enantiomer (+)-**6b** afforded diastereomers 3 and 4 (**11**), which were not separated.






Scheme 3. Reagents and conditions: (a) BnMgBr, ZnBr₂, Pd(PPh₃)₄, THF, -78 °C to rt, 73%.

Table 1. SAR at position 2 of the pyridine ring


Compd	X	PDE 4A IC ₅₀ (nM) ^a	HWB (TNF-α) IC ₅₀ (μM) ^a
6a		102	2.2
6b		23	0.60
1f		6	0.36
8		3	0.77
1b		0.6	0.35
CDP840	—	4	16

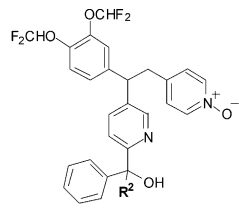
^aEach IC₅₀ value is an average of at least three experiments.**Table 2.** SAR on substituent R¹ of the carbinol moiety


Compd	R ¹	PDE 4A IC ₅₀ (nM) ^a	HWB (TNF-α) IC ₅₀ (μM) ^a
1b		0.6	0.35
1a	CH ₃	40	0.89
1c	<i>n</i> -Bu	6	0.34
1d		1.2	0.30
1e		6	0.49

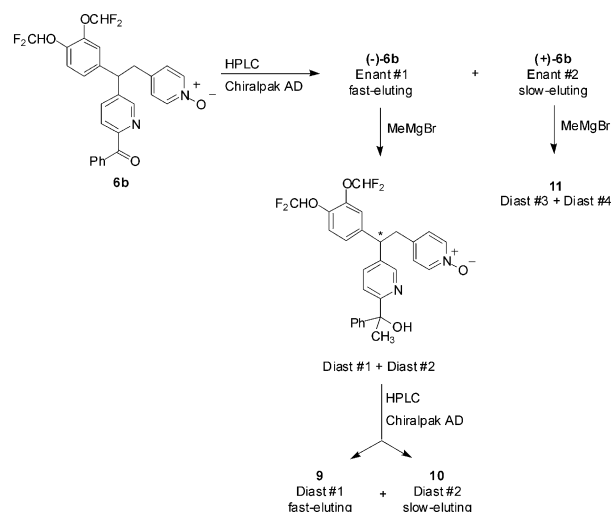
^aEach IC₅₀ value is an average of at least three experiments.

As observed previously in the triarylethane class of PDE4 inhibitors,^{8,10,11} a major stereochemical effect on inhibitory potency is observed at the chiral methine carbon. Diastereomers 3 and 4 (**11**) are significantly less potent than diastereomers 1 (**9**) and 2 (**10**) (Table 4). In contrast, the stereochemistry at the chiral carbinol center does not influence significantly the biological activity since both diastereomers **9** and **10** are essentially equipotent in vitro.

Despite similar intrinsic potencies, diastereomer **9** presents a slightly superior in vivo profile compared to **10**.

Table 3. SAR on substituent R² of the carbinol moiety


Compd	R ²	PDE 4A IC ₅₀ (nM) ^a	HWB (TNF-α) IC ₅₀ (μM) ^a
1b	CH ₃	0.6	0.35
1g	Et	1.0	0.16
1h	<i>i</i> -Pr	0.8	0.37
1i	CF ₃	2.4	0.46
1j	Ph	0.8	0.55

^aEach IC₅₀ value is an average of at least three experiments.**Scheme 4.** Resolution of tertiary alcohol **1b**.**Table 4.** Biological activity of diastereomers **9**, **10** and **11**

Compd		PDE 4A IC ₅₀ (nM) ^a	HWB (TNF-α) IC ₅₀ (μM) ^a
1b	Racemic Mixture	0.6	0.35
9	Diastereomer 1	2	0.16
10	Diastereomer 2	1	0.11
11	Diastereomers 3 and 4	48	3.5

^aEach IC₅₀ value is an average of at least three experiments.

For example, inhibitors **9** and **10** inhibit ovalbumin induced bronchoconstriction in conscious guinea pigs¹⁷ by 62 and 34% respectively when administered ip at 0.3 mg/kg (0.5 h pre-treatment). This behavior may correlate with the fact that diastereomer **9** presents a superior pharmacokinetic profile in various animal species. For this reason, compound **9** was selected for further evaluation. Inhibitor **9** is highly selective against the PDE4 enzyme (IC₅₀ = 2 nM); no inhibitory activity was detected against other phosphodiesterases at concentrations up to 5 μM. In contrast to its predecessor L-791,943,⁹ compound **9** presents suitable half-lives of 1.5 h in rats and 2.2 h in squirrel monkeys following intravenous administration at 5 mg/kg. The compound is also well

absorbed in both species, following oral dosage, with bioavailabilities of 57 and 73% respectively. Induction of emesis in squirrel monkeys is observed at an oral dose of 10 mg/kg. Under these conditions, the maximal concentration (C_{\max}) of **9** measured in plasma is 2.2 μM . Knowing that compound **9** inhibits LPS-induced TNF- α production in squirrel monkey whole blood with an IC_{50} of 0.11 μM , an estimation of the emetic window can be made by taking the ratio of these two concentrations. In the case of **9**, this ratio is 20, while for CDP840 and L-791,943 the same analysis provides ratios of 8 and >20 respectively. Inhibitor **9** is also effective for the inhibition of ascaris-induced bronchoconstriction in conscious sheep,¹⁸ exhibiting 33% and 91% inhibition of the early and late-phase responses respectively, at a dose of 0.5 mg/kg iv (4 days of dosing) given 2 h prior to challenge. On the whole, the data indicates that inhibitor **9** shows good in vivo efficacy in pulmonary function models while maintaining an improved window with respect to emesis.

In conclusion, we have developed a novel series of potent PDE4 inhibitors by replacing the metabolically resistant bis(trifluoromethyl)carbinolbenzene moiety found in L-791,943 by substituted 2-pyridinemethanol residues. The results of the SAR study in this series led to the identification of PDE4 inhibitor **9**, which exhibits excellent in vitro activity (HWB IC_{50} = 0.16 μM). Furthermore, compound **9** is well absorbed and presents a shorter half-life than L-791,943 in rats and squirrel monkeys. Compound **9** is active in the guinea pig model of ovalbumin-induced bronchoconstriction (0.3 mg/kg) and in the sheep model of ascaris-induced bronchoconstriction (0.5 mg/kg).

References and Notes

- (a) Conti, M.; Jin, S.-L. C. *Prog. Nucleic Acid Res. Mol. Biol.* **1999**, 63, 1. (b) Soderling, S. H.; Beavo, J. A. *Curr. Opin. Cell. Biol.* **2000**, 12, 174.
- Houslay, M. D.; Sullivan, M.; Bolger, G. B. *Adv. Pharmacol.* **1998**, 44, 225.
- Souness, J. E.; Aldous, D.; Sargent, C. *Immunopharmacol.* **2000**, 47, 127.
- Torphy, T. J. *Am. J. Respir. Crit. Care Med.* **1998**, 157, 351.
- (a) Huang, Z.; Ducharme, Y.; MacDonald, D.; Robichaud, A. *Curr. Opin. Chem. Biol.* **2001**, 5, 432. (b) Dyke, H. J.; Montana, J. G. *Exp. Opin. Investig. Drugs* **2002**, 11, 1.
- Robichaud, A.; Savoie, C.; Stamatiou, P. B.; Lachance, N.; Jolicoeur, P.; Rasori, R.; Chan, C. C. *Br. J. Pharmacol.* **2002**, 135, 113.
- (a) Burnouf, C.; Pruniaux, M.-P. *Curr. Pharm. Des.* **2002**, 8, 1255. (b) Martin, T. J. *IDrugs* **2001**, 4, 312.
- Alexander, R. P.; Warrelow, G. J.; Eaton, M. A. W.; Boyd, E. C.; Head, J. C.; Porter, J. R.; Brown, J. A.; Reuberson, J. T.; Hutchinson, B.; Turner, P.; Boyce, B.; Barnes, D.; Mason, B.; Cannell, A.; Taylor, R. J.; Zomaya, A.; Millican, A.; Leonard, J.; Morphy, R.; Wales, M.; Perry, M.; Allen, R. A.; Gozzard, N.; Hughes, B.; Higgs, G. *Bioorg. Med. Chem. Lett.* **2002**, 12, 1451.
- (a) Guay, D.; Hamel, P.; Blouin, M.; Brideau, C.; Chan, C. C.; Chauret, N.; Ducharme, Y.; Huang, Z.; Girard, M.; Jones, T. R.; Laliberté, F.; Masson, P.; McAuliffe, M.; Piechuta, H.; Silva, J.; Young, R. N.; Girard, Y. *Bioorg. Med. Chem. Lett.* **2002**, 12, 1457. (b) Chauret, N.; Guay, D.; Li, C.; Day, S.; Silva, J.; Blouin, M.; Ducharme, Y.; Yergey, J. A.; Nicoll-Griffith, D. A. *Bioorg. Med. Chem. Lett.* **2002**, 12, 2149.
- Frenette, R.; Blouin, M.; Brideau, C.; Chauret, N.; Ducharme, Y.; Friesen, R. W.; Hamel, P.; Jones, T. R.; Laliberté, F.; Li, C.; Masson, P.; McAuliffe, M.; Girard, Y. *Bioorg. Med. Chem. Lett.* **2002**, 12, 3009.
- Côté, B.; Frenette, R.; Prescott, S.; Blouin, M.; Brideau, C.; Ducharme, Y.; Friesen, R. W.; Laliberté, F.; Masson, P.; Styhler, A.; Girard, Y. *Bioorg. Med. Chem. Lett.* **2003**, 13, 741.
- A portion of this work was presented at the 17th International Symposium on Medicinal Chemistry, Barcelona, Spain, 1–5 September, 2002, Abstract P-225.
- Chambers, R. J.; Marfat, A. *Synth. Commun.* **1997**, 27, 515.
- (a) Nahm, S.; Weinreb, S. M. *Tetrahedron Lett.* **1981**, 22, 3815. (b) Williams, J. M.; Jobson, R. B.; Yasuda, N.; Marchesini, G.; Dolling, U.-H.; Grabowski, E. J. J. *Tetrahedron Lett.* **1995**, 36, 5461.
- Laliberté, F.; Han, Y.; Govindarajan, A.; Giroux, A.; Liu, S.; Bobechko, B.; Lario, P.; Bartlett, A.; Gorseth, E.; Gresser, M.; Huang, Z. *Biochemistry* **2000**, 39, 6449.
- Brideau, C.; Van Staden, C.; Styhler, A.; Rodger, I. W.; Chan, C.-C. *Br. J. Pharmacol.* **1999**, 126, 979.
- Muise, E. S.; Chute, I. C.; Claveau, D.; Masson, P.; Boulet, L.; Tkalec, L.; Pon, D. J.; Girard, Y.; Frenette, R.; Mancini, J. A. *Biochem. Pharmacol.* **2002**, 63, 1527.
- Abraham, W. M.; Ahmed, A.; Cortes, A.; Sielczak, M. W.; Hinz, W.; Bouska, J.; Lanni, C.; Bell, R. L. *Eur. J. Pharmacol.* **1992**, 217, 119.