

# Molecular Markers Assisted Identification of Intraspecific Hybrids in *Ziziphus mauritiana*

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**Abstract** Breeding in *Ziziphus mauritiana* through hybridization is limited by its small sized flowers, cross incompatibility, low fruit set and poor retention. Correct identification of a hybrid is often difficult when it resembles more with one parent or when new morphological combination of characters arise from recombination of distinct genotypes. Under present study three varieties of *Z. mauritiana* viz., Tikadi, Seb and Katha were found cross compatible and resulted in improved hybrids viz.,  $F_1$  (Seb  $\times$  Tikadi),  $F_1 \times$  Seb and Seb  $\times$  Katha. Ten RAPD primers revealed a total of 108 bands in the range of 150 bp to 3 kb. Among these, 69 bands were polymorphic and exhibited 33.33–90.9 % polymorphism in the banding patterns. The dendrogram clearly delineated variety Tikadi and Katha from Seb. All the three hybrids formed a single cluster with the variety Seb. The 5.8S gene region was found to be highly conserved (99.39 %) followed by ITS-2 (97.77 %). and ITS-1 (90.6–90.8 %). Heterozygous positions in ITS sequences clearly show their hybrid nature and could perform as highly polymorphic molecular markers. Despite SNPs, INDELS and total length polymorphism among parents and their hybrids, all were molecularly identified as *Z. mauritiana* and did not cause any phylogenetic error. The nucleotide polymorphism in ITS-1 region can serve as a bar code to detect and address genetic diversity within *Z. mauritiana*.

**Keywords** Indian jujube · Seb · Tikadi · Katha · Hybridization · ITS · RAPD

## Introduction

Indian jujube (*Ziziphus mauritiana* Lamk.) commonly known as Ber is mainly distributed in the tropic zone of southern Asia, Australia and Africa [1] including Taiwan and China. Endowed with a high degree of climatic adaptability, it is cultivated all over the arid parts of Indian subcontinent and plays a major role in preventing soil erosion and desertification [2]. The fruits of Indian jujube are rich in vitamins and are used as sedative, flavonoids, anticancer, tonic, wound healer and against asthma [2–6]. The extract from fruits [7], leaves [8], and seeds [9] have been reported to exhibit antioxidant activity.

Indian jujube germplasm characterization, evaluation and improvement are fundamentally based on morphophysiological traits. The lack of breakthrough has been due to underutilization of genetic variability for superior quality, resistance to biotic and abiotic stresses and high yield potential. Selection of promising plant types for widespread variability resulted in evolution of promising varieties. However, these cultivars lack one or more quality and/or productivity traits. Presently very little information is available in *Z. mauritiana* regarding genetic parameters to be helpful for the effective selection leading to an ideal genotype having high fruit yield. Saran et al. [10] assessed morphological diversity among *Z. mauritiana* genotypes and suggested that the most divergent genotypes could be selected for crop improvement.

Out of more than 90 cultivars, only 11 are commonly cultivated in different agro-climate regions of India [11] and show substantial variations in terms of their natural habitats, growth characteristics and aroma. The species demonstrates a rich genetic diversity mainly through natural cross pollination due to self incompatibility [12]. The elite plant types with desirable traits have been released as

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varieties, mass multiplied and propagated through standard vegetative multiplication method for this crop. However, the authenticity of cultivar identification in *Z. mauritiana* is unclear and a subject of some controversy [13].

The variety Tikadi, which is resistant to fruit fly and also exhibits frost tolerance was used as a parent in a crossing programme with the popular commercial cultivars Seb, which shows drought tolerance, market acceptability, mid early fruit maturity, apple shape of fruit and tolerance to powdery mildew [14–16]. The  $F_1$  hybrid (Seb  $\times$  Tikadi) showed resistance to fruit fly but inferior in fruit quality in respect of low pulp stone ratio and smaller sized fruits. The other crosses were incompatible. Subsequently, the back cross of  $F_1 \times$  Seb regained the desired fruit quality whilst retaining the fruit fly resistant trait. Katha fruit have better shelf life [17–19]. Therefore it was crossed with Seb to get hybrid with better shelf life. The  $F_1$  hybrid (Seb  $\times$  Katha) was found to be a promising precocious, prolific bearer and early maturing with good fruit set at high temperatures under arid conditions [20]. All these parental varieties and their hybrids are vegetatively propagated and are being maintained at Central Arid Zone Research Institute (CAZRI), Jodhpur.

Even if some morphological traits may help in hybrid recognition, a correct identification of a hybrid plant may often be difficult when it resembles one parent more than the other or when new morphological combination of characters arise from recombination of distinct genotypes or when only a few morphological traits are available for analysis/overlapping. It is in this context that the molecular approach may be useful tool for the assessment of hybrid status and for the correct identification of parental lineages [21].

Trees in general and *Z. mauritiana* in particular have a long juvenile phase when plants are raised through seeds (even up to 7 years). During this phase true elements of hybridism based on a few morphological descriptors cannot be validated due to the absence of fruiting stages. Molecular marker technologies circumvent this problem and hasten the breeding process by detecting coinheritance of both parental characters at very early juvenile stage.

Among the various molecular marker systems, the Randomly Amplified Polymorphic DNA (RAPD) is one of the most popular DNA based approaches [22]. It allows the analysis of individual and large number of markers in relatively short time, as only a few primers allow the generation of sufficient data to obtain a robust estimate of diversity index and have allowed the resolution of complex taxonomic relationships. This technique has recently been used in a number of fruit trees i.e., Citrus species [23] and *Punica granatum* [24]. The nuclear ribosomal Internal Transcribed Spacer (ITS) intron regions encompassing highly conserved 5.8S rRNA gene is considered as one of the most useful

phylogenetic markers due to less functional constraints and comparatively higher evolutionary changes [25]. The ITS polymorphism might occur at generic, specific or individual level, making it useful for phylogenetic, evolutionary and bio-geographical diversity studies [26, 27]. Because of the similarities between the methods, both ISSR [28] and RAPD markers have been successfully used to assess genetic diversity in *Z. mauritiana* but switching from RAPD to ISSR requires purchasing new primers and re-optimizing PCR parameters. Currently, ITS and RAPD techniques have been used together and have shown complimentary results for genetic diversity studies [24, 29, 30].

The authors report the extent of genetic diversity using nuclear ribosomal DNA and RAPD polymorphism to validate successful intra-specific hybridization within *Z. mauritiana*.

## Material and Methods

### Field Experiment

Morpho-physiological and molecular characterization of three putative *Z. mauritiana* varieties Tikadi, Seb and Katha and their crosses were carried out from ber orchard maintained in field gene bank at horticulture block, CR Farm of CAZRI, Jodhpur. Observations of fruit weight (g), fruit volume (CC), fruit width (cm) and fruit length (cm) were recorded from the composite samples collected from ten randomly selected trees of each of the parent and hybrid during January–February 2012 and 2013 (Table 1). Whereas the observations from five randomly selected plants were recorded for plant height (m), canopy area ( $m^2$ ), fruit yield (kg per plant), pulp: stone ratio, total soluble solids (TSS %) and specific gravity. The phenotypic correlation of all the morpho-physiological traits studied with fruit yield was calculated to identify possible phenotypic markers.

### DNA Isolation and Quantification

The genomic DNA was extracted from the leaves of three putative varieties viz., Tikadi, Seb, and Katha and three intraspecific hybrids Seb  $\times$  Tikadi ( $F_1$ ),  $F_1 \times$  Seb (BC) and Seb  $\times$  Katha of *Z. mauritiana*. To eliminate polyphenols, mucilages and polysaccharides as glucans and cuticular wax on leaves, a hybrid protocol for genomic DNA isolation developed using initial steps of CTAB method and subsequently columns and solutions of the Plant Genomic DNA Purification kit recently developed by Singh et al. [24] was followed. The genomic DNA was dissolved in 200  $\mu$ l of Tris–EDTA buffer and diluted to make a working concentration of 40–50 ng/ $\mu$ l for ITS amplification and RAPD analyses.

**Table 1** Morpho-physiological characterization of *Z. mauritiana* parents and their hybrids during 2012–2013

Variety	Plant height (m)	Canopy area (m <sup>2</sup> )	Fruit weight (g)	Fruit volume (cc)	Fruit breadth (cm)	Fruit length (cm)	Pulp: stone ratio	TSS (%)	Specific gravity	Fruit yield (kg plant <sup>-1</sup> )
Tikadi	4.55	32.52	3.48	3.10	1.50	2.90	4.28	26.33	0.990	31.80
Seb	2.74	16.90	15.8	15.6	2.86	3.13	9.69	19.20	1.012	45.20
Katha	2.61	28.20	25.60	25.2	3.30	3.80	14.33	24.80	1.016	23.60
F <sub>1</sub> (Seb × Tikadi)	4.20	23.74	5.50	9.42	2.30	2.5	6.76	24.20	0.988	18.23
BC <sub>1</sub> (F <sub>1</sub> × Seb)	2.71	22.25	15.80	16.50	2.90	3.51	10.98	22.40	0.988	43.80
F <sub>1</sub> (Seb × Katha)	2.54	18.08	17.20	17.50	3.12	2.80	12.25	23.80	0.980	32.50
C.D. (5 %)	0.22	3.13	1.22	1.70	0.26	0.32	0.99	2.82	0.042	3.51
Phenotypic correlation with fruit yield	−0.44	−0.47	0.16	0.05	0.16	0.29	0.10	−0.69	0.04	1.0

### RAPD Analysis

The random primers of OPA, OPB, OPP series (Operon Technologies) and a custom primer were used for initial screening of *Z. mauritiana* varieties and their hybrids. Based on the reproducibility of scorable bands, the RAPD was finally performed using 10 decamer arbitrary primers. Each amplification was performed in a total volume of 25 µl containing: decamer primer, 1 µl (50 pmol/µl); dNTP mix, 2 µl (2.5 mM/µl Bangalore Genei); *Taq* DNA polymerase, 0.4 µl (5 U/µl, Sigma Chem); MgCl<sub>2</sub>, 1 µl (25-mM, Sigma Chem); 10× PCR buffer, 2.5 µl (100 mM, Tris–HCl, pH 8.3, 15 mM MgCl<sub>2</sub>, 250 mM KCl), 14.1 µl of dH<sub>2</sub>O and 4 µl of genomic DNA (50 ng/µl). RAPD-PCR amplifications in a thermal cycler (Corbett Research, USA) with initial denaturation step of 94 °C for 3 min followed by 38 amplification cycles at 94 °C for 40 s, 50 °C for 40 s and 72 °C for 2 min and final elongation at 72 °C for 7 min. Amplicons were separated on a 1.6 % agarose gel and stained with ethidium bromide in 1× TAE buffer. The gels were run for 3 h at 45 V. The size of the amplified fragments was determined using 1 kb ladder. RAPD amplified DNA fragments were examined by repeating PCR reactions as well as running on gel for three times.

### PCR Amplification and Sequencing of ITS Region

The genomic DNA of the parents and hybrids of *Z. mauritiana* were used for amplification of 5.8S gene region. The Polymerase Chain Reaction (PCR) universal primer ITS-1 and ITS-4 developed by White et al. [31] were used to amplify ITS region of ribosomal DNA. Each PCR amplification was performed in a total volume of 50 µl containing: 1 U *Taq* DNA polymerase (Bangalore Genei), 2.5 mM MgCl<sub>2</sub>, 160 µM dNTP mix (MBI Fermentas), 50 pmol of each of ITS-1 and ITS-4 primers (Bangalore Genei), 50 ng genomic DNA in dH<sub>2</sub>O. The reactions were performed in a thermal cycler (Corbett Research, USA) with following

conditions: 1 min denaturation at 95 °C, 30 s annealing at 50 °C, 80 s elongation at 72 °C, for 35 cycles with a final elongation step of 72 °C for 10 min. Agarose gel was stained with ethidium bromide and photographed under UV light using Syngene gel documentation system.

PCR amplified products were purified using single step ExoSAP PCR clean up enzyme at 37 °C for 15 min followed by enzyme inactivation at 80 °C for a further 15 min to remove unconsumed dNTPs and ITS primers. The primers were diluted to 10 pM concentration for sequencing. Amplified ITS regions were then sequenced with an ABI Prism DNA sequencer (Applied Biosystems, Carlsbad, CA, USA) using ITS-1 and ITS-4 primers separately for labeling of each DNA by the BigDye terminator method (Applied Biosystems, Foster City, CA, USA). Comparison of nucleotide sequences was performed using the Basic Local Alignment Search Tool (BLAST) network services of the National Centre for Biotechnology Information (NCBI) database (<http://www.ncbi.nlm.nih.gov>). The phylogenetic relationships of parents and their hybrids were established by multiple alignment of sequences using ClustalX 2.0.11 and generating phylogram depicting bootstrap values using NJ plot software [32] based on Single Nucleotide Polymorphisms (SNPs), insertions/deletions (INDELs), and or length polymorphism in the ITS and 5.8S nuclear rDNA regions. A composite phylogenetic tree with bootstrap values showing grouping was generated to measure phylogenetic accuracy.

### Molecular Analysis of RAPD

Molecular data of RAPD profiles was entered into a binomial matrix and was used to determine Jaccard's similarity coefficient with NTSYS-pc software [33, 34]. The polymorphic information content (PIC) values for all the selected primers amplified by a particular primer pair was calculated for the RAPD markers to characterize the capacity of each primer to detect polymorphic loci using the formula derived by Smith et al. [35].

## Results

### Morpho-Physiological Characterization

The data on morpho-physiological characterization of 10 characters recorded during 2012–2013 are presented in Table 1. The plant height varied from 2.61 m (Katha) to 4.55 m (Tikadi). The canopy area varied from 16.9 m<sup>2</sup> (Seb) to 32.52 m<sup>2</sup> (Tikadi). The lowest fruit weight of 3.48 g was recorded from Tikadi and the highest of 25.6 g from Katha. Similarly the minimum fruit volume, fruit width, length and pulp stone ratio were recorded from the parent variety Tikadi and the maximum were recorded from the variety Katha. The gross fruit morphology with element of hybridity in parental varieties and hybrids are shown in Fig. 1(a–c). The minimum TSS% of 19.2 % was recorded from the variety Seb and the maximum of 26.33 % was recorded from the variety Tikadi. The specific gravity did not vary significantly among the parental varieties and their hybrids and were statistically at par with each other. The minimum fruit yield per plant varied from 18.23 kg [F<sub>1</sub> (Seb × Tikadi)] to 45.2 kg (Seb).

The analysis of phenotypic correlation vis-à-vis fruit yield revealed that plant height, canopy area and TSS% had negative and highly significant correlation, whereas, positive and significant phenotypic correlation was recorded in most of the fruit related traits for example, the highest correlation of fruit length (0.29) followed by fruit weight, fruit width (0.16) and pulp stone ratio (0.10) with fruit yield.

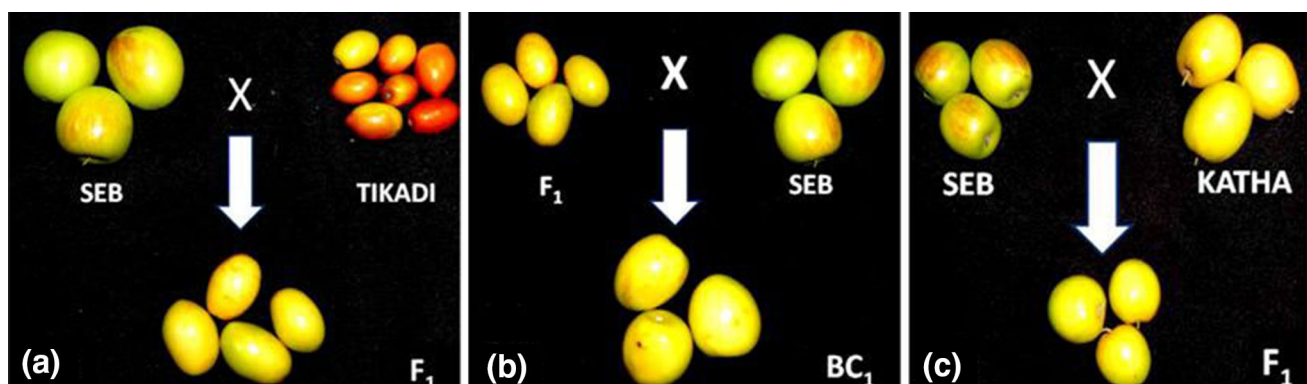
### RAPD Analysis

A set of 20 random primers of OPA and OPB series and one custom primer were used for initial screening of parents (Tikadi, Seb and Katha) of *Z. mauritiana* and their hybrids [F<sub>1</sub>(Seb × Tikadi), BC (F<sub>1</sub> × Seb) and F<sub>1</sub> (Seb × Katha)]. Based on reproducibility and intra-specific variation

patterns, 10 primers revealed a total of 108 bands in the range of 150 bp–3 kb (Table 2). Among these, 69 bands were polymorphic and exhibited 33.33–90.9 % polymorphism in the banding patterns. The number of PCR amplified products ranged from seven (OPA-03) to 14 (OPA-16). Primer OPA-17 was recorded the most informative primer and exhibited the maximum polymorphism amounting to 90.9 % in banding patterns. The RAPD profile generated by OPA-16 exhibited the maximum PIC value of 90 % (Fig. 2a). The dendrogram obtained from cumulative binary analysis of 10 RAPD primers scorable fragments clearly delineated Tikadi and Katha from Seb.

### Phylogenetic Analysis

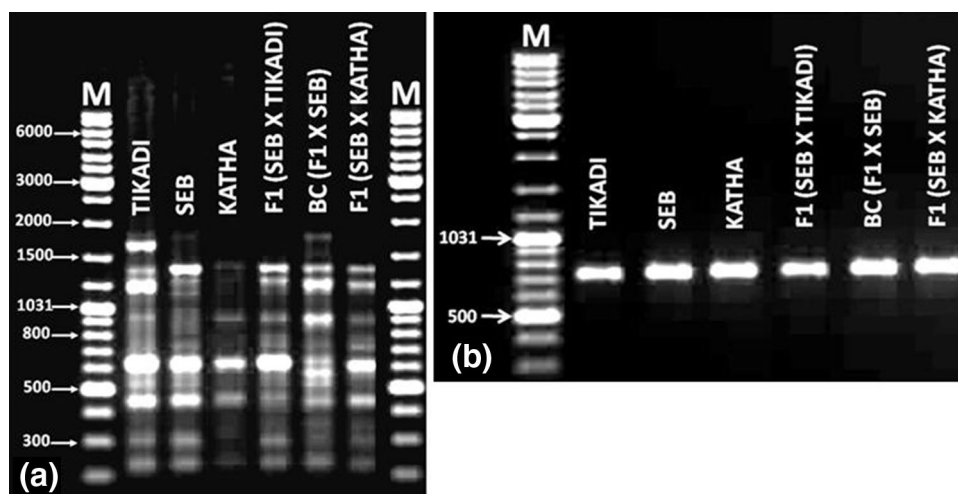
All the parental varieties of *Z. mauritiana* and their hybrids exhibited a single prominent band upon gel electrophoresis containing partial sequence of 18S gene complete sequence (ITS-1, 5.8S gene, ITS-2) and partial sequence of 28S gene upon direct sequencing using universal primer ITS-1 and ITS-4 (Fig. 2b). All the novel gene sequences have been assigned GenBank accession numbers by National Centre for Biotechnology Information (NCBI), USA (Table 3). The 5.8S rDNA region was recorded with a uniform nucleotide length of 164 bp and ITS-2 with 179 bp. The ITS-1 region exhibited base pair length diversity from 255 to 260 which eventually resulted in variation in the total length of the gene from 598 to 603 bp. The multiple sequence alignment of all the three parental varieties with their hybrids resulted in detection of Single Nucleotide Polymorphism (SNPs) at 20, Insertions at 2 and deletions at 2 positions in ITS 1 region, SNP at one position in 5.8S rDNA region and at 4 places in ITS2 region. All the three hybrids formed a single cluster with the variety Seb (Fig. 3a). The phylogram generated based on multiple sequence alignment further delineated all the parents and their hybrids with significant bootstrap values (Fig. 3b).



**Fig. 1** a F<sub>1</sub> (Seb × Tikadi), b BC (F<sub>1</sub> × Seb) and c F<sub>1</sub> (Seb × Katha)

**Table 2** Details of primer code, GC content, percent polymorphism and PIC values of RAPD primers used for molecular profiling of *Z. mauritiana* parents and hybrids

S. no.	Primer code	Primer sequence	GC (%)	No. of bands	No. of polymorphic bands	Polymorphism (%)	PIC values
1.	OPA-01	CAG GCC CTT C	70	12	8	66.66	88 %
2.	OPA-02	TGC CGA GCT G	70	12	6	50.00	89 %
3.	OPA-03	AGT CAG CCA C	60	7	5	71.43	83 %
4.	OPA-09	GGG TAA CGC C	70	8	4	50.00	84 %
5.	OPA-16	AGC CAG CGA A	60	14	9	64.29	90 %
6.	OPA-17	GAC CGC TTG T	60	11	10	90.90	87 %
7.	OPA-18	AGG TGA CCG T	60	13	11	84.61	89 %
8.	OPB-06	TGC TCT GCC C	70	9	3	33.33	88 %
9.	OPB-14	TCC GCT CTG G	70	10	6	60.00	87 %
10.	Custom primer	CGC ACC GCA G	80	12	7	58.33	89 %
	Total			108	69		
	Average					63.00	87.40

**Fig. 2** **a** RAPD profiles (OPA-16) and **b** ITS profiles of *Z. mauritiana* parents and hybrids

## Discussion

Hybrids offer an opportunity to recombine complementary quantitative and qualitative traits. Breeding in *Z. mauritiana* through hybridization is limited by its small sized flowers, cross incompatibility, low fruit set and poor retention [36]. Since nuclear markers are co-dominantly inherited, a putative hybrid plant must show the additive

profile of the two parental sequences [37], otherwise the resultant cross may not be a hybrid but simply a morphological variant of one parental species.

Under present study all the three varieties of *Z. mauritiana* viz., Tikadi, Seb and Katha were found cross compatible and resulted in improved hybrids. For instance, the hybrid F<sub>1</sub> (Seb × Tikadi) exhibited element of hybridity with inheritance of fruit fly resistance and improvement in

**Table 3** Nucleotide base pair lengths of nuclear ribosomal RNA gene of *Z. mauritiana* parents and hybrids

S. no.	Genotype	Gen accession number	ITS-1 (bp)	5.8 S (bp)	ITS-2 (bp)	Total (bp)
1.	Tikadi	JQ627029	255	164	179	598
2.	Seb	JQ627037	259	164	179	602
3.	Katha	JQ627045	257	164	179	600
4.	Tikadi × Seb (F <sub>1</sub> )	KC155273	260	164	179	603
5.	Tikadi × Seb (BC)	KC155274	259	164	179	602
6.	Seb × Katha	KC155275	259	164	179	602



the fruit size in  $F_1$  generation, back cross BC1 ( $F_1 \times \text{Seb}$ ) regained the desired fruit quality of variety Seb and  $\text{Seb} \times \text{Katha}$  acquired the characters of early maturing and improved shelf life that validate the co-inheritance of both the parental characters and a novel component of hybridity which is different from both the parents. The acquired desirable traits of the successful hybrids have been perpetuated through vegetative propagation.

The phenotypic correlation of different morpho-physiological characters with fruit yield suggests that TSS, plant height and canopy area are inversely proportional to fruit yield. The maximum positive and significant correlation of fruit length followed by fruit weight and breadth with fruit yield suggest that these morphological traits are of significance in large scale screening of wild and cultivated germplasm for improving fruit yield in *Z. mauritiana*. It can be inferred that these traits are under the control of additive gene action and phenotypic selection for their improvement to be effective.

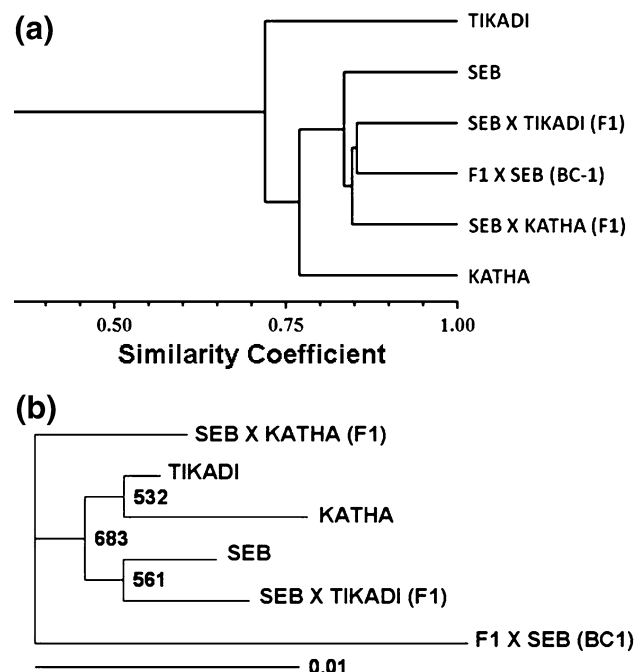
The methods of morphology or isozymes are insufficient to distinguish cultivars and the results are often liable to be influenced by the environment. It has been proved that ber genotypes earlier reported to be similar based on morphology are genetically different [13]. However the effect of hybridity on morphometric fruit characters and flavonoid spectrum facilitated characterization of varieties in *Z. mauritiana* to some extent [38–40] as well as in assessing the phylogenetic relationship between the parents and hybrids [41, 42]. By contrast, Tiwari et al. [43] attributed morphological variations within cultivar Banarsi of *Z. mauritiana* due to scion effect or clonal variation.

Molecular markers based on polymerase chain reaction (PCR) method offer several advantages over the conventional morphological markers. In general, the robustness of molecular marker technique depends on the amount of polymorphism it can detect. DNA markers have been often used for hybrid characterization in higher plants including *Ziziphus* [36, 44, 45]. Although genetic diversity analysis and cultivar identification by RAPD and other molecular markers have been performed in many fruit crops [24, 46], including *Ziziphus* [47], its application in identification of intraspecific hybrid within *Z. mauritiana* has not been carried out. AFLP and RAPD markers have been used to increase the diversity of the genetic linkage of *Z. jujube* [48, 49].

The cumulative analysis of all the ten informative RAPD primers detected an average of 63 % polymorphism in banding pattern with overall 87.4 % polymorphism indicating its efficacy for evaluating genetic diversity within species *Z. mauritiana*. All the three hybrid crosses clustered together with the parental variety Seb as all the three crosses had variety Seb as one of the parent. The test RAPD primers distinguished all the three parent DNA

profiles from each other and exhibited the presence of distinct parental bands in the individual. Patterns of all the three hybrids, each of separate origin confirmed their hybrid status. The extent of co-inherited RAPD markers was high from 72 to 86 % indicating a substantial element of co-inheritance of parental characteristics into progenies. Intraspecific classification within *Z. jujuba* has been addressed using RAPD [50, 51] to distinguish or to reduce the number of varieties. Based on the presence of male parents specific DNA fragments produced by RAPD and ISSR markers, true hybrids within *Z. mauritiana* have been identified by Khan et al. [36].

The genetic variations ranging from 14 to 28 % (Fig. 3a) indicate that all the parents and hybrids were distinguished from each other. Co-inherited RAPD markers have also been reported from other plants [52]. Comparison of the RAPD band patterns between an  $F_1$  hybrid, *Papaver bracteatum* and *Papaver pseudo-orientale* clearly showed that part of the bands of both parents were induced into the  $F_1$  hybrid [53]. Lee et al. [54] investigated the segregation ratio of morphologic characters (4 qualitative and 15 quantitative) and RAPD marker in 55 intra-specific hybrids and their parents to obtain complex inheritance information in the genus *Dianthus*. Shasany et al. [52] experimentally demonstrated that through RAPD, hybridization patterns can be revealed as distinctly as by AFLP, and therefore RAPD analysis may be applicable to various plant species where polypoidy has played a role in evolution and genomic constitution.



**Fig. 3** a RAPD dendrogram and b phylogram of *Z. mauritiana* parents and hybrids

Sharma et al. [55] suggested that the origin of new compounds through hybridity can be explained on the basis of new gene combinations leading to novel expression. The disappearance of spots from the hybrids can be explained on the basis of epistatic phenomenon i.e. the masking effect of one gene over the other. In addition some compounds present in parents were not found in the hybrids [56].

Both the forward and reverse ITS primers amplified a bigger fragment which also included partial 18S and 28S sequences and therefore measured approximately plus 700 bp on gel electrophoresis (Fig. 2b). Total ITS region (ITS1-5.8S-ITS2) length of parents and hybrids was approximately 600 bp (Table 3). A uniform base pair length of 164 and 169 bp of all the 3 parents and their hybrids validate the conserve nature of 5.8S gene and ITS-2 regions. The total length polymorphism in the intron ITS-1 region indicates its non-coding and variable nature. The 5.8S region was found to be highly conserved (99.39 %) followed by ITS-2 (97.77 %) and ITS-1 (90.6–90.8 %). The multiple sequence alignment of nucleotide sequence of parents *vis-a-vis* their hybrids exhibited that ITS-1 region contains the highest per cent of parsimony informative sites with high divergence mainly due to SNPs and can serve as a genetic bar code to reveal genetic diversity within *Z. mauritiana*. The delineation of all the parents and hybrids into distinct sub-clad with significant bootstrap values was mainly because of nucleotide polymorphism in ITS-1 region that can serve as a bar code to detect and address genetic diversity within *Z. mauritiana*. Heterozygous positions in ITS sequences clearly show their hybrid nature and could serve as highly polymorphic molecular markers. Ma et al. [57] reported that the sequence specific microsatellites could efficiently reveal intra-specific variations within *Z. jujuba*. The ITS length variants and polymorphism have been reported in several plant species. Raturi et al. [30] reported heterogeneity in nuclear rDNA ITS region of *Vigna radiata* which did not cause any phylogenetic errors at species level. Barkley et al. [58] observed SNPs and suggested Eco-TILLING as a powerful genetic analysis tool for rapid identification of naturally occurring variations in plants. Under the present study, despite SNPs, INDELS and total length polymorphism among parents and their hybrids, all were molecularly identified as *Z. mauritiana* and did not cause any phylogenetic error using BLAST search of NCBI.

The phylogram generated based on multiple sequence alignment further delineated all the parents and their hybrids with highly significant bootstrap values. Recent phylogenetic studies carried out by using the sequences of the internal transcribed spacer (ITS 1 and ITS 2) of nuclear ribosomal DNA have shown a higher correlation between ITS nucleotide and species within genus [21, 59] and at intra specific levels of crop plants [29, 30] and tree species [24].

Since it is co-dominantly inherited, a putative hybrid plant must show the additive profile of the two parental sequences.

From molecular point of view, heterozygous positions in ITS sequences clearly showed the hybrid nature of the crosses. Together with RAPD, ITS facilitated the detection of maternal lineages in hybrid progenies because of similarity coefficients and polymorphism. These specific RAPD markers can be utilized to develop SCAR markers [60–62], adding a comparable precision for the probing of improved genotypes through hybridization programme. The nucleotide polymorphism in ITS-1 region can serve as a bar code to detect and address genetic diversity within *Z. mauritiana*. Consequently, variety specific restriction sites in the nucleotide sequences may be used as marker for identification of the varieties and their hybrids.

## References

1. Qu ZZ, Wang YH (1991) China fruit's monograph—Chinese Jujube volume. China Forestry Publishing House, Beijing (in Chinese)
2. Pareek OP (2001) Fruits for the future 2: Ber. International Centre for Underutilized Crop. Redwood Books, Wiltshire, p 290
3. Morton J (1987) Indian jujube. In: Morton JF, Miami FL (eds) Fruits of warm climates. Center for New Crops & Plant Products, Purdue University, Lafayette, pp 272–275. <http://www.hort.purdue.edu/newcrop/morton/indianjujube.html>
4. Verheij EWM, Calabura M (1991) Plant resources of South-East Asia 2. In: Coronel RE (ed) Edible fruits and nuts. PROSEA, Pudoc, Wageningen, pp 223–225
5. Hudina M, Liu M, Veberic R, Stampar F, Colaric M (2008) Phenolic compounds in the fruit of different varieties of Chinese jujube (*Ziziphus jujuba* Mill.). J Hort Sci Biotechnol 83:305–308
6. Mishra T, Khullar M, Bhatia A (2011) Anticancer potential of aqueous ethanol seed extract of *Ziziphus mauritiana* against cancer cell lines and Ehrlich ascites carcinoma. Evid Based Complement Altern Med. doi:10.1155/2011/765029
7. Ndhalala AR, Mupure CH, Chitindingue KM, Benhura AN, Muchwet M (2006) Antioxidant potentials and degree of polymerization of six wild fruits. Life Sci Res Assays 1:87–92
8. Dahiru D, Obidoa O (2007) Pretreatment of albino rats with aqueous leaf extract of *Ziziphus mauritiana* protects against alcohol induced liver damage. Trop J Pharma Res 6:705–710
9. Bhatia A, Mishra T (2009) Free radical scavenging and antioxidant potential of *Ziziphus mauritiana* (Lamk.) seed extract. J. Complement. Integra Med 8:42–46
10. Saran PL, Godara AK, Sehrawat SK (2006) Characterization of ber (*Ziziphus mauritiana* Lamk.) genotypes. Haryana J Hort Sci 35(3/4):215–218
11. Morton JF (2005) Indian jujube. J Expt Bot 56(422):3082–3092
12. Godara NR (1980) Studies on floral biology and compatibility behavior in ber (*Ziziphus mauritiana* Lamk.) Ph.D. thesis, Haryana Agricultural University, Hisar, India
13. Devanshi, Singh AK, Sharma P, Singh B, Singh R, Singh NK (2007) Molecular profiling and genetic relationship among ber (*Ziziphus* sp.) genotypes using RAPD markers. Indian J Genet 67(2):121–127
14. Singh MP, Vashishtha BB (1984) Field screening of ber cultivars for resistance to ber fruitfly, *Carpomyia vesuviana* Costa. Indian J Plant Prot 12(1):55–56

15. Pareek OP, Nath V (1996) Ber. Coordinated fruit research in Indian Arid Zone—a two decades profiles (1976–1995). National Research Centre for Arid Horticulture, Bikaner, pp 9–30
16. Vashishtha BB, Singh MP, Singh GK, Pareek RA (1999) Breeding in Ber (*Ziziphus mauritiana* Lamk.) for the resistance of fruit fly. Abst. Fourth Agricultural Science Congress-Sustainable Agricultural Export, Jaipur, India
17. Dhingra RP, Singh JP, Chitkara SD (1973) Varietal variations in physico-chemical characters of ber (*Ziziphus mauritiana* Lamk.). Haryana J Hort Sci 2(3/4):61–65
18. Pareek OP, Vashishtha BB (1983) Delicious ber varieties of Rajasthan. Indian Hort 28(2):13–15
19. Pareek OP, Vashishtha BB (1986) Vegetative and fruit characteristics of ber (*Ziziphus mauritiana* Lamk.) cultivars of Rajasthan grown in the Thar Desert. Ann Arid Zone 25(3):214–221
20. Shukla AK, Dhandar DG, Vashishtha BB, Umamabeswari C (2004) New promising ber varieties. Indian Hort 49(3):36
21. Cennamo P, Cafasso D (2002) Molecular markers as a tool for the identification of hybrid plane trees. Delpinoa 44:89–94
22. Williams JGK, Kubelik AR, Livak KJ, Rafalski JA, Tingey SV (1990) DNA polymorphism amplified by arbitrary primers are useful as genetic markers. Nucleic Acids Res 18:6531–6535
23. El-Mouei R, Choumane W, Dway F (2011) Characterization and estimation of genetic diversity in Citrus rootstocks. Intl J Agric Biol 13:571–575
24. Singh SK, Meghwal PR, Pathak R, Gautam R, Kumar S (2013) Genetic diversity in *Punica granatum* revealed by nuclear rDNA, internal transcribed spacer and RAPD Polymorphism. Natl Acad Sci Lett 36(2):115–124
25. Alvarez I, Wendel JF (2003) Ribosomal ITS sequences and plant phylogenetic inference. Mol Phylogenet Evol 29:435–455
26. Carvalho A, Guedes-Pinto H, Lima BJ (2009) Genetic diversity among old Portuguese bread wheat cultivars and botanical varieties evaluated by ITS rDNA PCR-RFLP markers. J Genet 88(3):363–367
27. Xiao LQ, Moller M, Zhu H (2010) High nrDNA ITS polymorphism in the ancient extant seed plant cycas: incomplete concerted evolution and the origin of pseudogenes. Mol Phylogenet Evol 55:168–177
28. Singh AK, Devanshi S, Sharma P, Singh R, Singh B, Koundal KR, Singh NK (2007) Assessment of genetic diversity in *Ziziphus mauritiana* using inter-simple sequence repeat markers. J Plant Biochem Biotechnol 16:35–40
29. Kakani RK, Singh SK, Pancholy A, Meena RS, Pathak R, Raturi A (2011) Assessment of genetic diversity in *Trigonella foenum-graecum* based on nuclear ribosomal DNA, internal transcribed spacer and RAPD analysis. Plant Mol Bio Rep 29:315–323
30. Raturi A, Singh SK, Sharma V, Pathak R (2012) Molecular characterization of *Vigna radiata* (L.) Wilczek genotypes based on nuclear ribosomal DNA and RAPD polymorphism. Mol Biol Rep 39(3):2455–2465
31. White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (eds) PCR protocols: a guide to methods and applications. Academic Press, San Diego, pp 315–322
32. Perrière G, Gouy M (1996) WWW-Query: an on-line retrieval system for biological sequence banks. Biochimie 78:364–369
33. Sneath PHA, Sokal RR (1973) Numerical taxonomy. W. H. Freeman, San Francisco
34. Rohlf FJ (1997) NTSYS pc: numerical taxonomy and multivariate analysis system version 2.02h. Exeter software, New York
35. Smith JSC, Chin ECL, Shu H, Smith OS, Wall SJ, Senior ML, Mitchell SE, Kresovich S, Zeigler J (1997) An evaluation of the utility of SSR loci as molecular markers in maize (*Zea mays* L.): comparison with data from RFLPs and pedigree. Theor Appl Genet 95:163–173
36. Khan H, Sivalingam PN, Chauhan S, Awasthi OP, More TA (2013) Improved crossing technique and identification of true F1 hybrids of *Ziziphus mauritiana* Lam. by molecular markers. Sci Hortic 150:164–171
37. Rieserberg LH, Carted R, Zona S (1990) Molecular test of hypothesized hybrid origin of two diploid *Helianthus* species (Asteraceae). Evolution 44:1498–1511
38. Kanzaki S, Sugivra A, Yonemori K, Utsunomiya N, Subhadrabandhu S (1997) Chemotaxonomic studies on some Diospyros species by flavonoids compositions in leaves. In: Subhadrabandhu S (Ed) Proceedings of the first international persimmon symposium, Chiang mai, Thailand, 17–19 July 1996. Acta Hort 436:93–99
39. Williams CA, Harborne JB, Newman M, Greenham J, Eagles J (1997) Chrysin and other leaf exudates flavonoids in the genus Pelargonium. Phytochemistry 46:1349–1353
40. Robards K, Antolovich M, Boyd S (1997) Characterisation of Citrus by chromatographic analysis of flavonoids. J Sci Food Agric 75(1):87–101
41. Bhargava R (1983) Metabolites as indicators of polyploidy and hybridity in some bulbous angiosperms. Ph.D. Thesis, University of Jammu, Jammu
42. Bhargava R, Sharma MC, Koul AK (1986) Origin of variety Odorus Plenus of Narcissus in the light of flavonoids analysis. Nucleus 29(3):139–140
43. Tiwari R, Baghel BS, Gupta N (2003) Morphological characters of top-worked ber cv. Banarsi. JNKVV Res J 37(2):94–96
44. Harris SA, Ingram R (1992) Chloroplast DNA and biosystematics: the effect of intra-specific diversity and plastid transmission. Taxon 40:393–412
45. Dumolin S, Demesure B, Petit RJ (1995) Inheritance of chloroplast and mitochondrial genomes in pedunculate oak investigated with an efficient PCR-based method. Theor Appl Genet 91:1253–1256
46. Cervera MT, Cabezas JA, Sancha JC (1998) Application of AFLP's to the characterization of grapevine (*Vitis vinifera* L.) genetic resources: a case study with accessions from Rioja (Spain). Theor Appl Genet 97:51–59
47. Obeed RS, Harhash MM, Abdel-Mawgood AL (2008) Fruit properties and genetic diversity of five ber (*Ziziphus mauritiana* Lamk.) cultivars. Pak J Biol Sci 11(6):888–893
48. Fu JX, Liu CM, Xie JH (2007) Identification and classification of ber cultivars based on ISSR and RAPD analysis. Acta Hort 764:119–126
49. Jing Q, Zhen D, Min MY, Ying SL, Xing ZY, Jie L, Ling WX (2009) Construction of a dense genetic linkage map and QTL analysis of trunk diameter in Chinese jujube. Sci Silvae Sin 45(8):44–49
50. Peng JY, Shu HR, Peng SQ (2002) To address the problem of infraspecific classification of *Ziziphus jujube* Mill. using RAPD data. Acta Phytotaxon Sin 40:89–94
51. Liu P, Peng JY, Peng SQ, Zhou JY, Dai L (2005) Study on systematic relationships of *Ziziphus jujube* and *Ziziphus spinosa* using RAPD technique. Sci Silvae Sin 41:182–185
52. Shasany AK, Darokar MP, Dhawan S, Gupta AK, Patra NK, Shukla AK, Patra NK, Khanuja SPS (2005) Use of RAPD and AFLP markers to identify inter and intraspecific hybrids of Mentha. J Hered 96(5):542–549
53. Shoyama Y, Kawachi F, Tanaka H, Nakai R, Shibata T, Nishi K (1998) Genetic and alkaloid analysis of *Papaver* species and their F1 hybrid by RAPD, HPLC and ELISA. Forensic Sci Int 91:207–217
54. Lee SY, Yae BW, Kim KS (2005) Segregation patterns of several morphological characters and RAPD markers in interspecific



- hybrids between *Dianthus giganteus* and *D. Carthusianorum*. *Sci Hortic* 105:53–64
55. Sharma PK, Bhargava R, Koul AK (1989) Studies on the flavonoids spectrum of disomic. *Cytologia* 54:197–202
56. Bhargava R, Shukla AK, Chauhan NB, Vashishtha B, Dhandar DG (2005) Impact of hybridity on flavonoid spectrum of ber (*Ziziphus mauritiana* Lamk.). *Environ Expt Bot* 53:135–138
57. Ma QH, Wang GX, Liang LS (2011) Development and characterization of SSR markers in Chinese jujube (*Ziziphus jujuba* Mill.) and its related species. *Sci Hortic* 129:597–602
58. Barkley NA, Wang ML, Gillaspie AG, Dean RE, Pederson GA, Jenkins TM (2008) Discovering and verifying DNA polymorphisms in a mungbean [*V. radiata* (L.) R Wilczek] collection by Eco-Tilling and sequencing. *BMC Res Notes* 1:28
59. Schoch CL, Seifert KA, Huhndorf S, Robert V, Spouge JL, Levesque CA, Chen W, Fungal Barcoding Consortium (2013) Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proc Natl Aca Sci of USA* 109(16):6241–6246
60. Paran I, Michelmore RW (1993) Development of reliable PCR-based markers linked to downy mildew resistance genes in lettuce. *Theor Appl Gent* 85:985–993
61. Hernandez P, Martin A, Dorado G (1999) Development of SCARs by direct sequencing of RAPD products: a practical tool for the introgression and maker-assisted selection of wheat. *Mol Breed* 5:245–253
62. Lee GP, Lee CH, Kim CS (2004) Molecular markers derived from RAPD, SCAR and the conserved 18S rDNA sequences for classification and identification in *Pyrus pyrifolia* and *P. communis*. *Theor Appl Gent* 108:1487–1491