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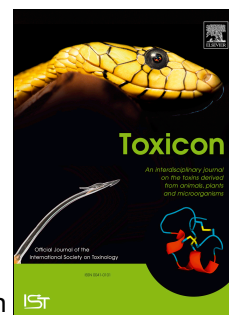
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Venomomics of the Australian eastern brown snake (*Pseudonaja textilis*): detection of new venom proteins and splicing variants.

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Abstract

The eastern brown snake is the predominant cause of snakebites in mainland Australia. Its venom induces defibrination coagulopathy, renal failure and microangiopathic hemolytic anemia. Cardiovascular collapse has been described as an early cause of death in patients, but, so far, the mechanisms involved have not been fully identified. In the present work, we analyzed the venom of *Pseudonaja textilis* by combining high throughput proteomics and

transcriptomics, aiming to further characterize the components of this venom. The combination of these techniques in the analysis and identification of toxins, venom proteins and putative toxins allowed the sequence description and the identification of the following: prothrombinase coagulation factors, neurotoxic textilotoxin phospholipase A₂ (PLA₂) subunits and “acidic PLA₂”, three-finger toxins (3FTx) and the Kunitz-type protease inhibitor textilinin, venom metalloproteinase, C-type lectins, cysteine rich secretory proteins, calreticulin, dipeptidase 2, as well as evidences of *Heloderma* lizard peptides. Deep data-mining analysis revealed the secretion of a new transcript variant of venom coagulation factor 5a and the existence of a splicing variant of PLA₂ modifying the UTR and signal peptide from a same mature protein. The transcriptome revealed the diversity of transcripts and mutations, and also indicates that splicing variants can be an important source of toxin variation.

Keywords: *Pseudonaja textilis*; transcriptome; proteome; toxin; splicing variant.

1. Introduction.

Snake bites are an important public health problem in many parts of the rural, developing world, mostly in poor, tropical and subtropical areas (Harrison *et al.*, 2009). There may be as many as 4.5 to 5 million snake bites/year globally, resulting in 2.5 million envenomings, 125,000 deaths and perhaps three times that number with permanent disabilities (Chippaux, 1998). However, an accurate estimation of snake bite envenoming is difficult to establish (WHO, 2007) and these numbers may be much higher, as many estimates rely on hospital returns, which are not available in many areas, and do not capture data for cases that do not present to public health services. The mainstay of treatment for snake bite envenoming is antivenom obtained and purified from hyper-immune plasma, mostly from horses. The value of antivenom is however determined by its ability to effectively neutralize all of the medically relevant toxins in a venom from which is raised, and some toxins may be poorly neutralized

(Gutiérrez *et al.*, 1981; Judge *et al.*, 2006). A better knowledge of venom components and their role in the onset of the pathophysiological effects might circumvent the lack of efficacy of available antivenoms against some components of snake venoms. The currently available cutting-edge proteomic and transcriptomic tools have made such studies possible, enabling the detection of new and/or rare toxins (Calvete, 2013; Paiva *et al.*, 2014) as well as intraspecific ontogenic and geographical variations of the venom (Castro *et al.*, 2013; Madrigal *et al.*, 2012). Such studies have improved our knowledge of venoms and how they vary within species and genera, providing tools to better understand the molecular evolution of toxins, as well as to identify potential targets for the design of more effective antisera. The combined proteomic/transcriptomics approach is also relevant to affirm new transcripts as real toxins, as common tissues can express toxin transcripts, even in non-venomous snakes such as pythons (Reyes-Velasco *et al.*, 2014).

In the present work, our aim was to characterize the venom gland products from the eastern brown snake *Pseudonaja textilis* by combining transcriptomic and proteomic studies of the gland and its venom. The eastern brown snake is the predominant cause of accidents in mainland Australia (White, 2009), due to its abundance and adaptation to peridomicile, even in urban environments. Its venom has been described as containing toxins that induce defibrination coagulopathy, renal failure and microangiopathic hemolytic anemia (White, 2009). Although neurotoxicity is considered to be rare (Barber *et al.*, 2012; White, 2009), a potent presynaptic neurotoxin named textilotoxin has been described (Aquilina, 2009; Pearson *et al.*, 1993) as well as short and long three-finger postsynaptic toxins (3FTx) (Gong *et al.*, 2001, 2000; St Pierre *et al.*, 2007b). The low *P. textilis* venom yield (Mirtschin *et al.*, 2002) associated to low neurotoxins concentrations (Barber *et al.*, 2012) and the usual low venom/victim weight ratio (Mirtschin *et al.*, 1998) can be considered as an explanation for this apparent paradox, although specificity of the toxin towards nerve terminals in a specific prey type should not be discounted as a possible explanation. According to Judge *et al.* (2006), there is an accumulating body of evidence to suggest that the efficacy of the brown snake antivenom is limited. These authors report that the antivenom does not recognize the low molecular mass protein components of the venoms of *P. textilis*, *P. affinis affinis* and *P. nuchalis* when assayed by western blot, nor was the antiserum able to neutralize the contractile response of tracheal nerve/muscle preparations. These observations suggest that either these toxins are poorly immunogenic or that they might not be present in the venom pool used to produce the antiserum. Indeed, according to the World Health Organization

(2007) “Ineffective antivenoms may also be prepared because of an inappropriate selection of the venoms used as immunizing mixtures. This illustrates a lack of information on the snake fauna of the area or region as well as on the composition and immunochemistry of venoms”. This highlights the need of further investigation of venom components and how they correlate with clinical observations.

2. Material and Methods.

2.1. Biological samples.

The venom gland was extracted from an adult *Pseudonaja textilis* male individual captured at Barossa Valley, near Adelaide, South Australia. The venom gland was extracted three days after milking to obtain a tissue with a high level of toxin transcript expression, and stored in RNA-Later® (QIAGEN N.V., Netherlands) at -80 °C until RNA extraction. The venom of *P. textilis* used for proteomics and toxin purification was a pool from five captive individuals from the same region. The crude venom was lyophilized and stored at -20°C until use. The animal was euthanized for tissue collection in accordance with Euthanasia of Animals Used for Scientific Purposes guidelines (2001), Australian and New Zealand Council for the Care of Animals in Research and Teaching, under the monitoring of the SA Pathology/ CHN Animal Ethics Committee, Project Approval 93/12.

2.2. Venom gland *de novo* transcriptome.

The venom gland *de novo* transcriptome was obtained by shotgun pyrosequencing (GS-FLX, Roche) of a normalized cDNA library (GATC Biotech, Konstanz, Germany). After trimming, the resulting reads were aligned and assembled with Newbler™ (Roche). Resulting isotigs and singletons were identified and annotated with BLAST2GO (Conesa *et al.*, 2005; Götz *et al.*, 2008). All steps were manually eye-checked and fixed when necessary.

2.3. Venom 2D-PAGE, in-gel digestion and MS² analysis.

Prior to use, the venom sample was dissolved to 170 µg/ml in 9 M urea, 2 % ampholytes and 70 mM DTT. After 30 min room temperature incubation and centrifugation (45 min, 15000 g) the supernatant was removed and frozen at -80°C. The protein mixture was decomplexed by 2D-PAGE using a slightly adapted method from previous works (Georgieva *et al.*, 2011; Meganathan *et al.*, 2012). The selected spots were collected and in-gel digested with Trypsin (Promega, USA). Peptides were analyzed by liquid chromatography (LC) followed by electrospray ionization (ESI) and detected in an ion trap mass spectrometry system (Agilent

1100 LC/MSD-trap XCT series system) (Viala *et al.*, 2014). The most intense ions were fragmented by collision-induced dissociation (CID) and MS² spectra were acquired. The protein identification was performed based on the public protein database enriched with our in house *P. textilis* transcriptome, using the InChorus multi-algorithmic tool from PEAKS (Bioinformatics Solutions Inc., Canada) that integrates PEAKS and MASCOT (Matrix Science Inc., USA) identification results. Identity was considered when significant score was achieved. All MS/MS assignments and automatic *de novo* sequencing results were manually revised for correctness as well as the quality of the mass spectra of peptides from near-threshold identification.

2.4. Anti-jararhagin western blot.

Crude lyophilized *P. textilis* venom was redissolved (2 mg/mL in PBS pH 7), centrifuged and 30 µL of the supernatant were diluted in 10 µL of non-reducing buffer and submitted to 15% SDS-PAGE (Laemmli, 1970). The gel was placed in the electroblot apparatus and transferred to nitrocellulose paper in transfer buffer for 90 min at 0.85 mA/cm² (Towbin *et al.*, 1979). The nitrocellulose paper was then incubated with polyclonal anti-jararhagin antibodies (diluted 1:5000). Jararhagin is a P-III metalloproteinase from *Bothrops jararaca* and the antibodies were gently provided by Dr. Máisa Splendore Della Casa (Instituto Butantan, São Paulo, Brazil). The immunoreactive proteins were detected using peroxidase-labeled anti-rabbit IgG and the blot was developed with orthophenyldiamine in the presence of 0.03% H₂O₂ (v/v).

2.5. Metalloproteinase cDNA cloning and sequencing.

The metalloprotease transcript was cloned from a *P. textilis* venom gland cDNA library, built using In-Fusion SMARTer cDNA library construction kit (Clontech Laboratories Inc., USA). RNA was extracted with Trizol® reagent (Life Technologies, USA) in an RNase free environment. The 20-mers primers (5'UTR: 5'-TTGGAAGCAGAAAGAGATTC-3' and 3'UTR: 5'-GTAGGATAAAGACAGATGGG-3') were designed based on conserved regions found by aligning metalloproteases UTR sequences from Elapidae, Colubridae and Viperidae species, available in public databases (GenBank, NCBI). The 5'UTR and 3'UTR sequences were first separated from the open reading frame (ORF), then the UTRs were aligned independently (Hall, 1999; Lassmann *et al.*, 2009). PCR reaction was performed using Taq DNA polymerase (Biotools B&M Labs S.A., Spain) in the following conditions: 1X 94°C 5 min + 40X 94 °C 30s, 51°C 30s and 72°C 2min + 1X 72°C 5min (Thermal cycler CG1-96, Corbett Research, QIAGEN N.V., Netherlands). The resulting amplification bands were

excised from the agarose gel, extracted with DNA gel extraction kit (Norgenbiotek, Canada) and cloned into pGEM-Teasy® (Promega, USA). *E. coli* DH5α electrocompetent cells (New England Biolabs Inc., USA) were transformed with the ligated plasmids. After ampicillin and white/blue selection of the recombinant colonies, plasmids were extracted by miniprep (QIAGEN N.V., Netherlands) and clones sequencing was performed on an ABI 3730 DNA Analyser with BigDye (Applied Biosystems) and universal forward and reverse M13 primers. As the forward and reverse sequences did not overlap, another forward primer Ptint1F (5'-ACTTCGGAGTCAGATGAGCC-3') was designed to obtain the missing overlap sequence. Resulting sequences were aligned and the final consensus sequence was generated.

3. Results.

By combining transcriptomics and proteomics high-throughput techniques in the analysis and identification of toxins, venom proteins and putative toxins, a big volume of data was generated. The data-mining results were evaluated and discussed in the light of toxinology, biochemistry, genetics, biology, and evolutionary knowledge. A previous proteomic analysis of this venom was performed by Birrell *et al.* (2006) in which the following toxins were identified: Pseutarin-C (catalytic and non-catalytic subunits), neurotoxic phospholipase A₂ (PLA₂) textilotoxin subunits and other PLA₂s, 3FTxs, textilinin, the pseudochetoxin-like cysteine rich secretory proteins (CRiSP) and an additional unknown protein in venoms, the Glucose Regulated Protein 78. Our combined analysis identified the following toxins in the *P. textilis* venome: prothrombinase complex coagulation factors, textilotoxin subunits, procoagulant PLA₂, short and long 3FTxs, the Kunitz-type protease inhibitor textilinin (bovine protease trypsin inhibitors family – BPTI), CRiSP and for the first time, a new splicing variant of the snake venom coagulation factor V_a (VF5_a), a yet undescribed long 3FTx (LNTX-2), C-type lectins (CLect), as well as evidences of lizard toxins from *Heloderma* genus and other toxin candidates such as calreticulin and dipeptidase 2. Some of those toxins were identified in a pool of venoms at the isoform level, supported by the observation of unique peptides. A venom metalloproteinase (SVMP), a class of toxins poorly investigated in elapidae, was detected by western blot, cloned and sequenced. Also, we identified at the transcript level a PLA₂ signal peptide switching mechanism driven by alternative splicing.

3.1. Venom gland transcriptome.

Next Generation Sequencing of the transcriptome generated 104,878 reads (c.a. 37 Mb) with 34 to 944 base pairs (bp) (N50 = 466 bp) and 42.25% GC. The assembly generated 1,518 isotigs (N50 = 916 bp, longer isotig = 6,179 bp), clustered into 1266 isogroups, and 41,514 reads were considered singletons, totalizing 43,032 sequences. The BLASTx resulted in transcripts of interest annotated as toxins, related to venom or considered putative new toxins. Coding and truncated toxins transcript/sequences were classified into 3FTx, BPTIs, VF5_a and snake venom coagulation factor X_a (VF10_a), PLA₂, CLect, hyaluronidase, natriuretic peptides (NP), wey acidic proteins (WAP), veficolin, CRiSP, SVMP, calreticulin, transferrin, cobra venom factor (CVF), dipeptidylpeptidase IV and VIP/glucagon-like (vasoactive intestinal peptides). These sequences were individually checked for quality and untranslated regions (UTR), open reading frames (ORF) and their translated amino acid sequences, signal peptides, mutations and alternative splicing were identified. An in house transcriptomic annotated database was compiled and used for the next steps of venomomics identification. This Transcriptome Shotgun Assembly project has been deposited at DDBJ/EMBL/GenBank under the accession GDAD00000000. The version described in this paper is the first version, GDAD01000000.

3.2. Venom proteome.

The 2D gel decomplexed the crude venom into spots with a wide range of molecular mass (from 10 to > 100 kDa) and a wide range of pI (3 to 10) (Fig. 1). 120 spots were collected manually from different regions of the 2D gel and protein clusters. Venom protein identification was successful in 86 spots. After data mining they were classified into eight toxin families (VF5_a; VF10_a; PLA₂: A and B chains from the multimeric PLA₂ textilotoxin and “acidic PLA₂”; 3FTx; BPTI textilinin; CLect; CRiSP and *Heloderma* peptides). The InChorus multi-algorithmic identification results are listed in the Supplementary Material. Nineteen sequenced transcripts encoding for VF5_a, VF10_a, Acidic PLA₂, PLA₂ textilotoxin A, CRiSP, LNTx-1, LNTx-2, SNTx, NXS7, textilinin, CLect and calreticulin were identified in 67 out of the 85 positive spots (Supplementary Material). The long 3FTx (LNTX-2) and the VF5a-3 are new variants that were identified at the protein level.

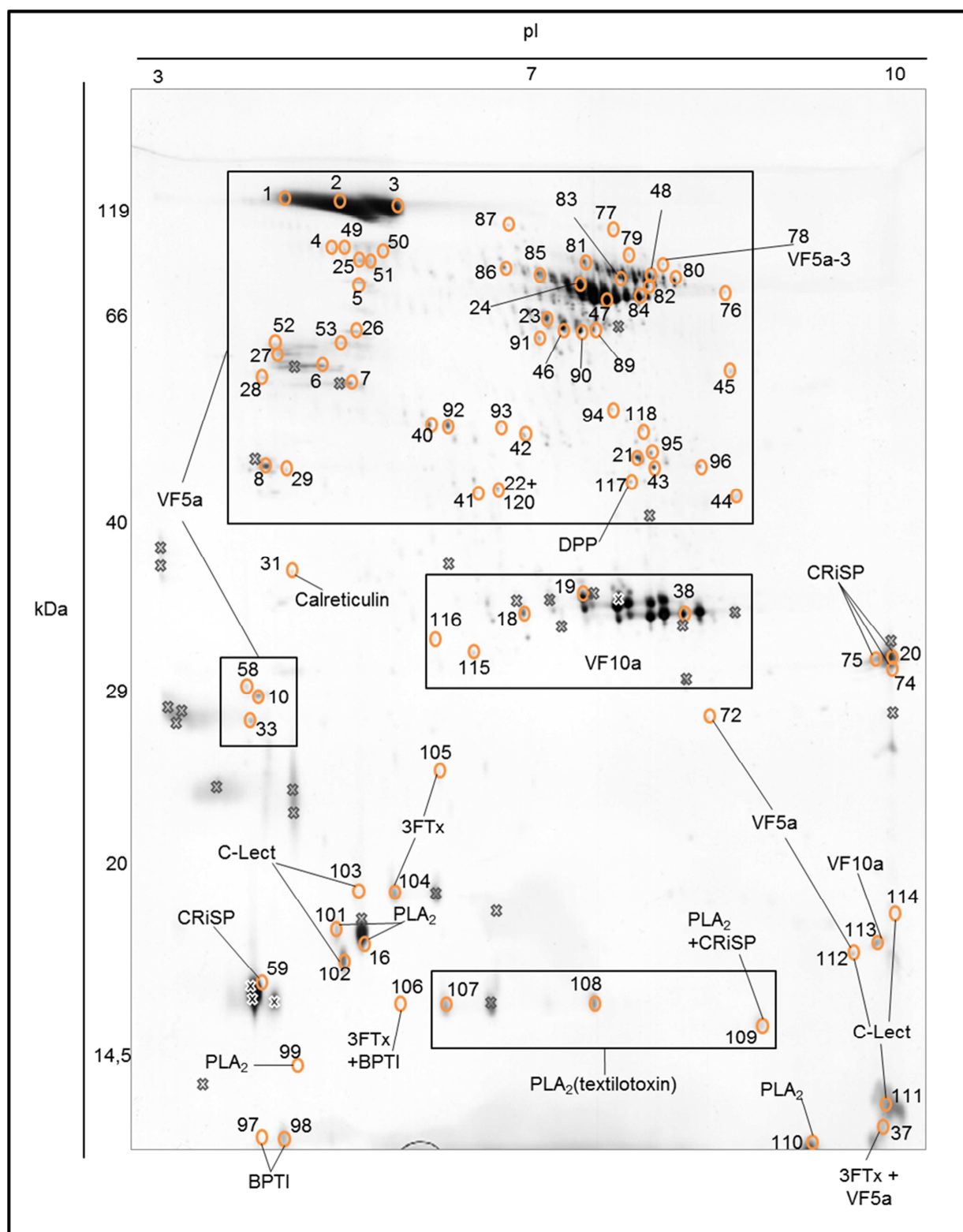


Fig. 1. 2D-PAGE of *Pseudonaja textilis* and the assigned spots (circles). Collected spots without positive match are crossed. Clusters of toxins are highlighted by labeled rectangles. VF5a: Coagulation venom factor 5a; VF10a: Coagulation venom factor 10a; CRiSP: Cysteine rich secretory protein; C-Lect: C type lectins; PLA₂: Venom phospholipase A₂; BPTI: Bovine protease trypsin inhibitor (textilinin); 3FTx: Three-finger toxin; DPP: Dipeptidylpeptidase-2.

3.3. Venom metalloproteinase detection, cDNA cloning and sequencing.

When the crude venom was analyzed by western blot using a polyclonal antibody raised against jararhagin, a single reactive band was detected around 45 kDa (data not shown), indicating the presence of a metalloproteinase. A full transcript was cloned and the sequence revealed to encode a P-III SVMP. The sequence was translated and the protein domains were identified by homology with similar sequences and a DCD motif was observed in the disintegrin domain (Fig. 2). The translated protein has 612 residues and calculated molecular mass (MM) of 68.5 kDa. After trimming out the signal peptide and the propeptide, the putative secreted protein has a predicted mass around 45 kDa and pI around 5.

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10      20      30      40      50      60      70      80
-----|-----|-----|-----|-----|-----|-----|-----|
TGGAAGCAGAAAGAGATTCTATCCACCAGTCCAATCCAGGCTCCAAAATGATCCAAGCTCTTGGTAAGTATATGCTT
M I Q A L L V T I C F
|-----|
signal peptide

90      100     110     120     130     140     150     160
-----|-----|-----|-----|-----|-----|-----|-----|
CGCGGTTTTCATATCAAGGGAGCTCTATAATCCTGGAATCCGGGAATGTTAATGATTATGAAGTAGTATCCACAAA
A V F P Y Q G S S I I L E S G N V N D Y E V V Y P Q K
|-----|
signal peptide      propeptide

170     180     190     200     210     220     230     240
-----|-----|-----|-----|-----|-----|-----|-----|
AAATGCCTGCATTGCCCAAAGGAGGAGTTCGGAATCCTCAGCCAGAGACCAAGTATGAAGATACAATGCAATATGAATTT
M P A L P K G G V R N P Q P E T K Y E D T M Q Y E F
|-----|
propeptide

250     260     270     280     290     300     310     320
-----|-----|-----|-----|-----|-----|-----|-----|
CAAGTGAATGGAGAGCCAGTGGTCTTCACTTAGAAAGAAATAAAGAACTTTTTCAGAGAGATTACACTGAAATTCATTA
Q V N G E P V V L H L E R N K E L F S E D Y T E I H Y
|-----|
propeptide

330     340     350     360     370     380     390     400
-----|-----|-----|-----|-----|-----|-----|-----|
TTCCTCTGATGACACAGAAATTATAACAAGCCCTCCGGTTCAGATCATTGCTATTATCATGGCTACATTGAGAATGAAG
S S D D T E I I T S P P V Q D H C Y Y H G Y I Q N E A
|-----|
propeptide

410     420     430     440     450     460     470     480
-----|-----|-----|-----|-----|-----|-----|-----|
CTAATTCAGTGCAATCATCAGTGCATGCGACGCTTGAAAGGACATTTCAAGCATCAAGGGGAGACATACTTTATTGAG
N S S A V I S A C D G L K G H F K H Q G E T Y F I E
|-----|
propeptide

490     500     510     520     530     540     550     560
-----|-----|-----|-----|-----|-----|-----|-----|
CCCTTGAAGCTTTCCGACAGTAAATCCCATGCAATCTACAAGATGAAAATGTAGAAGAAGAGGAAAAGACCCCAACTG
P L K L S D S K S H A I Y K D E N V E E E E K T P N C

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282 propeptide
283
284
285       570       580       590       600       610       620       630       640
286 -----|-----|-----|-----|-----|-----|-----|-----|
287 TGGGATAACCCAGACTACTTCGGAGTCAGATGAGCCCATCGAAAAGATCTCTCAATTAACATAATTTCTGAACAAGAAA
288   G I T Q T T S E S D E P I E K I S Q L T N I S E Q E R
289 -----|
290 propeptide
291
292       650       660       670       680       690       700       710       720
293 -----|-----|-----|-----|-----|-----|-----|-----|
294 GGTACTTGAAGGTCAAAAATACATTGAGTTGTATGTAGTTGTGGACAACAAAATGTACAAGAATTACGACAGCARGAGA
295   Y L K V K K Y I E L Y V V V D N K M Y K N Y D S X R
296 -----|-----|
297 metalloproteinase
298
299       730       740       750       760       770       780       790       800
300 -----|-----|-----|-----|-----|-----|-----|-----|
301 CATGCTATAAAAAAGAAAGTATATGAAACAATCAACCCCTTAAACATGATGTACAGACCTTTGAATTTTCTCATTGCACT
302   H A I K R K V Y E T I N P L N M M Y R P L N F L I A L
303 -----|-----|
304 metalloproteinase
305
306       810       820       830       840       850       860       870       880
307 -----|-----|-----|-----|-----|-----|-----|-----|
308 GATTGGCCTAGAAATTTGGTCCAACCGAGATAAGATTAAACATTGAACAGAGGTGGCTGTCACTTTGAAATCATTGGAA
309   I G L E I W S N R D K I N I E P E V A V T L K S F G K
310 -----|-----|
311 metalloproteinase
312
313       890       900       910       920       930       940       950       960
314 -----|-----|-----|-----|-----|-----|-----|-----|
315 AATGGAGAGAAACAGTTTACTGCCACGCAAAAGGAATGATAATGCTCAGTTACTTACGCAGATTGAGTTCAGCGGAAC
316   W R E T V L L P R K R N D N A Q L L T Q I E F S G T
317 -----|-----|
318 metalloproteinase
319
320       970       980       990       1000       1010       1020       1030       1040
321 -----|-----|-----|-----|-----|-----|-----|-----|
322 ACTGTAGACTTGTCTTATGTGGGAGCATCTGCAGTCCGAGGAATCTGTAGCAGTTATGGAGGTTTATAGCAGAAKAWC
323   T V G L A Y V G S I C S P E E S V A V M E V Y S R X X
324 -----|-----|
325 metalloproteinase
326
327       1050       1060       1070       1080       1090       1100       1110       1120
328 -----|-----|-----|-----|-----|-----|-----|-----|
329 AAATATAATGGCATCTGGAATGGCCCATGAGTTGGGTCAATCTGGGCATTACTCATGACCACGCTTCTGTAAATTGCA
330   N I M A S G M A H E L G H N L G I T H D H A S C N C N
331 -----|-----|
332 metalloproteinase
333
334       1130       1140       1150       1160       1170       1180       1190       1200
335 -----|-----|-----|-----|-----|-----|-----|-----|
336 ATGCTGAACATATGCATTATGTCTGCGATAATAAGTTTGAACCTCTCTCTGAGTTCAGCTCTTGTAGTATCCAGGAACAT
337   A E L C I M S A I I S F E P L S E F S S C S I Q E H
338 -----|-----|
339 metalloproteinase
340
341       1210       1220       1230       1240       1250       1260       1270       1280
342 -----|-----|-----|-----|-----|-----|-----|-----|
343 CAGAGGTGTCTTCTTAGAGAGAGACCACAATGCATTCTCAACAGACCCTTGAGCACAGATATTGTTACAMCTCCAGTTTG
344   Q R C L L R E R P Q C I L N R P L S T D I V T X P V C
345 -----|-----|
346 metalloproteinase disintegrin
347
348       1290       1300       1310       1320       1330       1340       1350       1360
349 -----|-----|-----|-----|-----|-----|-----|-----|
350 TGGAAATTAACCTAGTGGAGGTGGGAGAAGAATGTGACTGTGGCTTTCTTATGGATTGTCAAAGTGCTGCTGCAACGCTA
351   G N Y L V E V G E E C D C G F P M D C Q S A C C N A T
352 -----|-----|
353 disintegrin
354
355       1370       1380       1390       1400       1410       1420       1430       1440
356 -----|-----|-----|-----|-----|-----|-----|-----|
357 CAACTTGTAAACTGCAACATGAGGCACAGTGTGACTCTGAAGAGTGTGTGAGAAATGCAAACTTAAGAAAGCAGGAGCA
358   T C K L Q H E A Q C D S E E C C E K C K L K K A G A

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disintegrin
-----
          1450      1460      1470      1480      1490      1500      1510      1520
-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|
GAATGCCGGGCGAGCAAAGGATGACTGTGACTTGCCTGAAATCTGCACTGGCCAATCTGCTGAGTGTCCCATGGACAGCTT
E  C  R  A  A  K  D  D  C  D  L  P  E  I  C  T  G  Q  S  A  E  C  P  M  D  S  F
-----
disintegrin
-----
          1530      1540      1550      1560      1570      1580      1590      1600
-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|
CCAGAGGAATGGACATCCATGCCAAAACAACCAAGGTTACTGCTACAATGGGAAATGCCCATCATGACAAACCAATGTA
Q  R  N  G  H  P  C  Q  N  N  Q  G  Y  C  Y  N  G  K  C  P  I  M  T  N  Q  C  I
-----|-----|-----|-----|-----|-----|-----|-----|
          Cysteine rich
-----
          1610      1620      1630      1640      1650      1660      1670      1680
-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|
TCGATCTCTGGGGGCCAGGTGTAATGTGTCTCCAGATATATGTTTACGTTGAACCAAGTATAGCCAAGGTTGTGGCTTC
D  L  W  G  P  G  V  N  V  S  P  D  I  C  F  T  L  N  Q  Y  S  Q  G  C  G  F
-----
Cysteine rich
-----
          1690      1700      1710      1720      1730      1740      1750      1760
-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|
TGCAGAAATGGAAATGATACAAAGATTCCGTGTGCAGCAAAGGATAAAATGTGTGGCAAGTTAATATGCCGAAAGGGAAA
C  R  M  E  N  D  T  K  I  P  C  A  A  K  D  K  M  C  G  K  L  I  C  E  K  G  N
-----
Cysteine rich
-----
          1770      1780      1790      1800      1810      1820      1830      1840
-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|
CTCGACATGCACCTGCTTTCCTACAACAGATGACCCAGATGATGGAATGGTTGAACCTGGAACAAAATGTGGAGATGGAA
S  T  C  T  C  F  P  T  T  D  D  P  D  D  G  M  V  E  P  G  T  K  C  G  D  G  M
-----
Cysteine rich
-----
          1850      1860      1870      1880      1890      1900      1910      1920
-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|
TGGTGTGCAGCAACAGGCAGTGTGTTGATGTGCAGACAGCCTACTGATCAAGCACTGGCTTCTCTCAATTGATTTTGAA
V  C  S  N  R  Q  C  V  D  V  Q  T  A  Y  *
-----|-----|
          Cysteine rich
-----
          1930      1940      1950      1960      1970      1980      1990
-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|-----:-----|
GATCCTCATCCAGAAAGCTTCCCTCAAGTCCAAGAGACCCATCTGTCTTTATCCTACAATCACTAGTGAATTC

```

Fig. 2. cDNA sequence and translation of the cloned *P. textilis* P-III SVMP. Domains were identified by homology. The highlighted gray box indicates the DCD disintegrin tripeptide motif.

4. Discussion.

4.1. *P. textilis* toxins.

4.1.1. Prothrombinase complex: Coagulations factors Va and Xa.

The venom coagulation factors VF5_a and VF10_a are present in large amounts in some Australian Elapidae venom such as *Oxyuranus microlepidotus*, *O. scutellatus* and *P. textilis*

(Bos and Camire, 2010; Masci *et al.*, 1988). In *P. textilis* venom, VF5_a and VF10_a form the prothrombinase complex named Pseutarin-C (Kini *et al.*, 2001), with similar characteristics to the blood plasma prothrombinase complex, essential for blood clot formation by converting prothrombin into thrombin (Mann *et al.*, 1990; Rao and Kini, 2002). The venom prothrombinase complex was recruited to the venom gland after successive gene duplication and rapid mutation accumulation (Minh Le *et al.*, 2005), which enabled the toxins to be secreted already activated, modified to escape the protein C inactivation system and with no need for membrane interaction (Bos *et al.*, 2009; Rao *et al.*, 2003). Pseutarin-C is an effective *in vitro* procoagulant, but has the opposite effect *in vivo* by quickly exhausting the prey's blood plasma prothrombin, resulting in coagulopathy and spontaneous bleeding due to exhaustion of clotting factors (Tibballs *et al.*, 1992, 1991).

VF5_a

Two full transcripts encoding for VF5_a were identified (isotigs 199 and 200). The isotig 200 encodes a new isoform, named here VF5a2, similar to the venom prothrombin activator pseutarin-C non-catalytic subunit FA5V_PSETE (UniProt accession number) (Rao *et al.*, 2003), the only one described so far in *P. textilis*. It contains minor mutations in the nucleotide and amino acid sequences, when compared to the previously described toxin FA5V_PSETE, but none at cleavage and known interaction sites (Camire and Bos, 2009). The isotig 199, named here VF5a3, encodes for the isoform VF5a2 increased by a 102 bp insertion at the ORF position 2528. Both VF5_a2 and VF5_a3 sequence were aligned (BLAST) against the *Ophiophagus hannah* genome (Vonk *et al.*, 2013), as no *P. textilis* genomic sequence coding for VF5_a is available in public databases, and both matched entirely and uniquely into the assembly scaffold 988.1 (Accession gb|AZIM01000987.1). The alignment shows that the VF5_a3's 102 bp insertion is an extra exon, named here exon 15 (Fig. 3). Consequently, VF5_a3 might be a splicing variant. More interestingly, this unique additional amino acid sequence, translated from exon 15, was identified in the proteome analysis as described below.

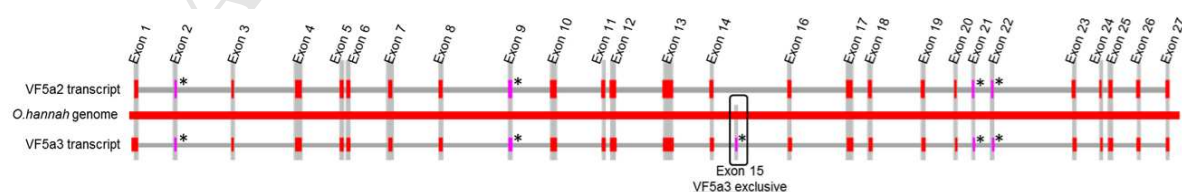


Fig. 3: Graphical overview of *Pseudonaja textilis* VF5a2 and VF5a3 transcripts alignment (BLAST) with the king cobra (*Ophiophagus hannah*) genome scaffold 988.1 (Accession gb|AZIM01000987.1), described by Vonk *et al.* (2013). An alternative splicing of this gene is demonstrated by the existence of exclusive exon 15 on the VF5a3 transcript (box). Alignments are coded by BLAST score. Exons labeled with * have a similarity score ranging from 80 to 200, unlabeled exons have similarity scores above 200. Horizontal lines connect exon sequences.

In general, the proteome analysis detected VF5_a tryptic peptides in 57 spots (1-8, 10, 21-29, 33, 37, 40-53, 58, 72, 76-87, 90-96, 112 and 120). A wide range of mass and pI distribution was observed in the 2D gel, with evident clusters, indicating the diversity of post-translational modifications (PTMs), and possibly proteolysis (Fig. 1).

The VF5a isoforms identified were: the *Oxyuranus scutellatus* (FA5V_OXYSU) (Welton and Burnell, 2005), *P. textilis* venom and blood coagulation factor (FA5V_PSETE and FA5_PSETE) (Rao *et al.*, 2003) and the two new isoforms described in this transcriptome (VF5a2 and VF5a3). The splicing variant VF5a3 contains an extra sequence of 34 amino acid residues (exon 15), after the Gln846 (KSQKLFWKIEESELESRKRIEKDKYIYSEENIKE). A set of two unique peptides IESELESR and DKYIYSEENIK identified in spot 78 confirm that this product of alternative splicing is indeed translated and secreted in the venom. (Supplementary Material). To our knowledge, this is the first time that an alternative splicing variant of snake venom toxin is detected at the protein level.

VF10_a.

Two isotigs (188 and 189) encoding for VF10_a were identified, with a slight difference only between their 3'UTR regions (17 pb gap). The ORF have 99% of identity with the only one described sequence (Accession: FAXC_PSETE) (Filippovich *et al.*, 2005), but no mutations were found in known active sites. VF10_a was identified in five spots (18, 38, 113, 115 e 116) from a single cluster with c.a. 35 kDa and pI between 6 and 8.5 (Fig. 1).

4.1.2. Phospholipases A₂.

PLA₂s are ubiquitous enzymes in nature (Kini, 2003). Snake venom PLA₂s were recruited to the venom gland and lost their original function (Ohno *et al.*, 1998). The great diversity of

venom PLA₂s is best illustrated by the many different neurotoxic, myotoxic, cardiotoxic, haemolytic, haemorrhagic, hypotensive, anticoagulant and other functions (Kini, 2003). The most studied *P. textilis* PLA₂s are the textilotoxin subunits (A to D), which form a potent heterohexameric presynaptic neurotoxin (Aquilina, 2009; Pearson *et al.*, 1993). Another PLA₂ with apparent procoagulant activity has also been described (Armugam *et al.*, 2004).

Many transcripts were identified as snake venom PLA₂ (Table 1). The “acidic PLA₂” was found in isotigs 824, 263 and 264. Isotig 824 ORF is identical to the “acidic PLA₂ 1” (Accession: PA2A1_PSETE) (Armugam *et al.*, 2004). The isotigs 263 and 264 encode for the same mature protein, with 99% identity to the “acidic PLA₂ isoform 2” (Accession: PA2A2_PSETE) (Armugam *et al.*, 2004), and were named here as “PLA₂ isoform 3” and “PLA₂ isoform 4”, respectively. Isotig 264 5’UTR and signal peptide nucleotide sequences are different from the 263’s (and consequently the signal peptide first 9 aa residues).

Table 1: Venomic PLA₂ identification details. The numbering of the spots (proteome) and the isotigs (transcriptome) are cited in columns 2 and 3 respectively. ND: not detected. Hit description: name obtained from SwissProt database. IA and IB are subclasses of class I PLA₂, depending on the presence or absence of the pancreatic loop.

Hit description	Spot	Istotig	Hit's activity	Pancreatic loop
<i>Basic PLA₂ CM-II</i>	90, 91, 99	ND	Myonecrotic and neurotoxic	No (IA)
<i>textilotoxin A chain</i>	107, 109	814	Hexameric neurotoxin	
<i>textilotoxin B chain</i>		883		
<i>textilotoxin C chain</i>	ND	819		
<i>textilotoxin D chain</i>		800		
<i>Acidic A2 1</i>	ND	824	pro-coagulant	Yes (IB)
<i>Acidic A2 2</i>	16, 101, 110	263		
<i>Acidic A2 2</i>		264		

When carefully analyzed, aligning the nucleotide sequences of isotigs 263 and 264 with the available gene sequences of this PLA₂ (GenBank Accession: AY027495.1) (Armugam *et al.*, 2004), the isotig 264 was identified as a new splicing variant, with a different 5’UTR and a new start codon inside the usual first intron (Fig. 4). Interestingly, this new signal peptide matches with *Micrurus fulvius* (e.g. Accession: U3F5A1_MICFL) and *Bungarus multicinctus* (e.g. Accession: Q8AXW0_BUNMU) PLA₂ signal peptide. We identified unique peptides from this acidic PLA₂ in spots 16, 101 and 110.

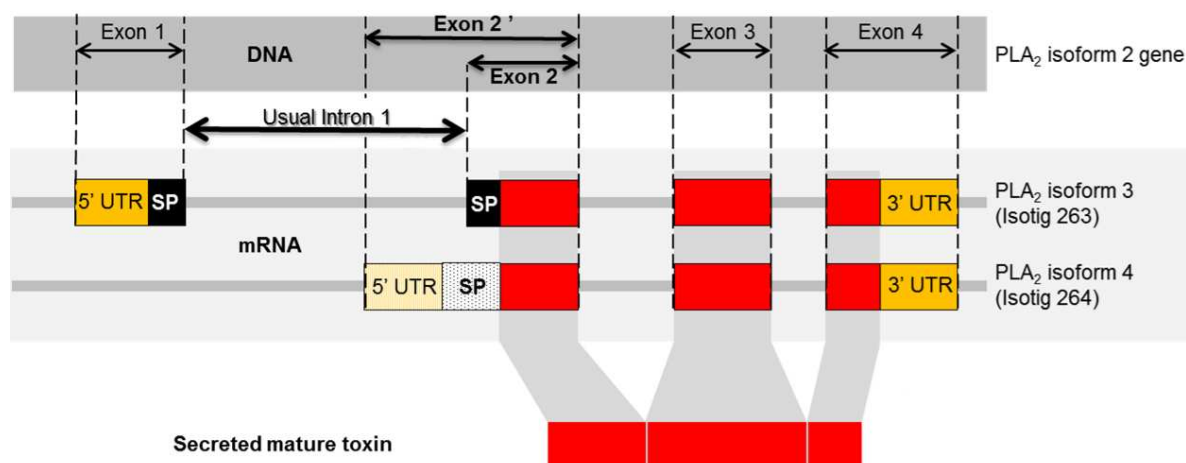


Fig. 4: Graphical overview of the alternative splicing occurring between two PLA₂ transcripts. The alignment with the gene, described in *Pseudonaja textilis* (Armugam *et al.*, 2004), highlights an alternative splicing occurring at exon 2. Although the alternative splicing generates two different signal peptides (SP), the secreted mature toxin is the same. UTR: untranslated region.

Textilotoxin was also identified in the venom. Isotigs 814, 883, 819 and 800 encode for subunits A, B, C and D respectively, which translated amino acid sequences are all 100% identical to those already described in the literature, except for the isotig 883 (named here as subunit B isoform 2) with two substitutions out of the 121 residues. These data indicate a strong positive selection upon those toxins. The textilotoxin B subunit nucleotide sequence is being described for the first time. The proteomics identified textilotoxin A and B subunits in spots 107 to 109.

A tryptic peptide with a sequence only found in the “basic PLA₂ CM-II” (UniProt Accession number: PA2B2_NAJMO) described in *Naja mossambica* (Joubert, 1977) was identified in the spots 90, 91 and 99. CM-II causes myonecrosis when injected intramuscularly, causes neuromuscular blockade with a gradual contracture and a decreased sensitivity to ACh and KCl (Lin *et al.*, 1987).

4.1.3. Three-finger toxins:

The 3FTx are non-enzymatic polypeptides with 60 to 74 aa residues. They are commonly known as 3FTx due to the three finger-like loops emerging from a hydrophobic core and linked together by four conserved disulfide bonds. Despite this similar structure, 3FTx can

have different biological activities including the blockade of nicotinic acetylcholine receptors, distinct muscarinic acetylcholine receptor subtypes, L type Ca^{2+} channels, inhibition of acetylcholinesterase, platelet aggregation, or as analgesic and membrane pore formation agents (Hegde *et al.*, 2009; Kini and Doley, 2010; Utkin, 2013). 3FTx are common component of elapid snake venoms and are often responsible for their lethal effects. The α -neurotoxin antagonists of postsynaptic nicotinic acetylcholine receptors are the most studied 3FTx. They promote paralysis, respiratory failure and death (Servent *et al.*, 1997). The molecular accelerated evolution, classification and nomenclature of these toxins are subject to debate (Doley *et al.*, 2009; Sunagar *et al.*, 2013). Although a huge diversity of genes and transcripts have been described (Gong *et al.*, 1999; Jackson *et al.*, 2013; St Pierre *et al.*, 2007b; Tyler *et al.*, 1987), the usual simplified short-chain neurotoxin (SNTx) or long-chain neurotoxin (LNTx) classification is used here (Hegde *et al.*, 2009).

This toxin family presented the greatest number of isotigs and isogroups in the transcriptome, after assembly. A closer analysis into the sequences ORFs and UTRs enabled the subclassification of isogroup 2 into three more subgroups (Table 2).

Table 2: Diversity of 3FTx transcripts of the *P. textilis* venom gland transcriptome. Isogroups are derived from assembly. Subgroups were assigned based on sequence similarity. Blast2Go annotations are based on public NCBI protein database annotations. NA: Not Available.

Isogroup	Subgroup	Istotig	Blast2Go annotation	Note
2	1	67, 68, 72, 74, 76 to 79	Short neurotoxin precursor	SNTx isoforms
		75	---NA---	Deletion nonsense
	2	60 to 62	Short neurotoxin precursor	Premature stop codon
	3	57 to 59	Uncharacterized transposon-derived protein	Loss of signal peptide by insertion
		63 to 66, 69 to 71, 73	Short neurotoxin precursor	
27	4	185 and 187	Short neurotoxin precursor	New SNTx
		186	---NA---	Deletion nonsense
23	5	174 and 176	Long chain neurotoxin precursor	LNTx-1 (100% similar)
		175		LNTx-1 new isoform
83	6	295 and 296	LNTx-1 precursor	LNTx Pseudonajatoxin b homologue (100% similar)
1076	7	1328	Long neurotoxin	LNTx-2 (new)
6	8	112 to 118	---NA---	Non-coding, with gene introns.

The isogroups 2 and 27 are clusters of transcripts encoding for SNTx. Isotig 74 is the exact NXS7 variant (Gong *et al.*, 2000) and isotigs 67, 68, 72 and 76 to 79 encode for new SNTx isoforms. Isotigs 185 and 187 encode for new SNTxs similar to 3FTxs from *Pseudonaja modesta* “Pse-290” (Jackson *et al.*, 2013) (~90%) and *Oxyuranus microlepidotus* “3FTx-Oxy6” (Fry *et al.*, 2008) (84%). Other transcripts had nonsense mutations (*e.g.*: insertions, deletions, intron insertions) suggesting that the 3FTx gene family undergoes wide-ranging alternative splicing and deserves further investigation.

The previously described “LNTx-1” (St Pierre *et al.*, 2007b) and an isoform were identified in isogroup 23. The isogroup 83 encodes for “Pseudonajatoxin b homologue” (Gong *et al.*, 2001) and isotig 1328 is a new LNTx transcript, named here “LNTx-2”, with 93% translation similarity to a *Pseudonaja modesta* LNTx (Accession: R4G319_9SAUR) (Jackson *et al.*, 2013), 85% to α -neurotoxin “LNTx 20” from *Drysdalia coronoides* (Accession: 3FL20_DRYCN) (Chatrath *et al.*, 2011) and only 78% similar to “Pseudonajatoxin b homologue”. Some singletons showed truncated sequences of diverse LNTxs.

The proteome analysis identified SNTx variants in spots 37, 105 and 106, namely “NXS2”, “NXS3” and “NXS7”. The “LNTx-1” was identified in spot 37 and the new “LNTx-2” (isotig 1328) was identified in spot 104 (Supplementary Material).

4.1.4. Serine protease inhibitors peptides: Kunitz-like and WAP.

Kunitz-like serine protease inhibitors (I2 family), also known as Bovine Protease Trypsin Inhibitors (BPTI), are present in a great variety of tissues in almost all organisms (except for Archaeae and Fungi). A variety of BPTIs with different activities are found in venoms of different creatures, ranging from anemones (*e.g.* Schweitz *et al.*, 1995), wasps (*e.g.* Hisada *et al.*, 2005), spiders (*e.g.* Chung *et al.*, 2002), shrews (*e.g.* Kita *et al.*, 2004), scorpions (*e.g.* Chen *et al.*, 2012), viperid (*e.g.* Cheng *et al.*, 2012) and elapid (*e.g.* Masci *et al.*, 2000) snakes. Textilinin, a selective and reversible inhibitor of plasmin was previously described in *P. textilis* venom, and has applications in the control of blood clotting (Earl *et al.*, 2012; Masci *et al.*, 2000).

Among the transcripts, isotig 173 was the only one with a functional venom BPTI sequence, identical to “textilinin-4” (Accession: VKT4_PSETT) (Filippovich *et al.*, 2002). Others have nonsense mutations in the beginning or the end of the transcript (*e.g.* isotig 171 and 172). The proteomic analysis identified textilinin 1, 2 and 3 at the isoform level because of unique

peptides at spots 97, 98 and 106, but no other kind of serine protease inhibitors were identified in this venom.

Since there is an interest in understanding the structure, function and pharmacology of textilinin and other venom serine-protease inhibitors as well as the molecular evolution mechanism of toxin recruitment in the venom gland, it is important to survey all sequences related to this family. A great variety of BPTI transcripts were already described in elapid venom gland transcriptomes and some of them present an additional BPTI or whey-acidic-protein (WAP) domain (Jackson *et al.*, 2013; St Pierre *et al.*, 2008). WAP are peptides from the I17 serine proteases inhibitors family, found in large amounts in many mammals milk, and contain a domain found in many other proteins (Bingle *et al.*, 2002; Hennighausen and Sippel, 1982). Snake waprins were isolated from *Oxyuranus microlepidotus* and *Naja nigricollis* venom, and displayed antimicrobial activity but no haemolytic, no toxicity in mice and no elastase and cathepsin G inhibition (Nair *et al.*, 2007; Torres *et al.*, 2003). WAP transcripts were also sequenced from snake venom glands, such as textwaprins from *P. textilis* (Fry *et al.*, 2008; St Pierre *et al.*, 2008) but the function in the venom is still unknown. Transcripts of fused Kunitz inhibitors and WAP domains (KuWAP) were described in *Sistrurus catenatus edwardsii* (Pahari *et al.*, 2007) and *Suta fasciata* (Jackson *et al.*, 2013) venom gland, but with no further characterization. In humans the Epididymal Protease Inhibitors (EPPIN) containing a BPTI and a WAP domain are found in spermatozoa surface and have an antimicrobial activity and contraceptive function (Wang *et al.*, 2007; Yenugu *et al.*, 2004). In snake venom glands, both inhibitors seem to have a common evolutionary pathway as they share signal peptide and the first exon (Jackson *et al.*, 2013; St Pierre *et al.*, 2008). In this transcriptome, a great variety of BPTI transcripts (isotigs and fragments encoded on singletons) were identified (mono, double and triple domain as well as WAPs and fused BPTI/WAP) and knowledge of their sequences can help to better understand this family of protease inhibitors in snakes and their toxin version in their venom. Surprisingly, a partial sequence of a double BPTI domain serine protease inhibitor similar to those found in leech (Simakov *et al.*, 2013) and tick (Macedo-Ribeiro *et al.*, 2008) saliva was identified in singleton BC91X. In the *Rhipicephalus microplus* saliva, the Boophilin-H2 (Accession: BOOH2_RHIMP) is a double BPTI domain serine protease inhibitor that inhibits the host blood clotting as the animal feeds (Macedo-Ribeiro *et al.*, 2008). Further genetic cloning and venom characterization should elucidate the function of this inhibitor in venom.

4.1.5. C-type lectins.

The most commonly described C-type lectins (CLects) in Elapidae are galactose or mannose binding with ~26 kDa and usually structured into homo or heterodimers (Abreu *et al.*, 2006; Du *et al.*, 2002; Earl *et al.*, 2011; Zha *et al.*, 2001). C-type lectins are common body proteins and the diversity of transcripts observed in the venom gland might be related to house-keeping functions, thus, abundance of transcript may not equate to toxin diversity *per se*. In fact, between the diversity of CLects sequenced, only the isogroup 13 set of transcripts was identified in the proteome analysis in spots 102, 103, 111, 112 and 114. This isogroup is formed by 4 variants of a galactose-binding CLect (isotigs 145 to 148, named “Venom C-type lectin galactose binding variant” numbered from 3 to 6, respectively) differing from each other by some point mutations, not involved in the QPD sugar binding site or metal binding regions. Some other galactose binding CLect described in elapid snake venoms were also identified in this proteome. CLects are being described for the first time at the protein level on *P. textilis* venom.

4.1.6. CRiSP.

Venom Cysteine Rich Secretory Proteins (CRiSPs) have conserved sequence between Colubridae, Viperidae and Elapidae species, with ~23kDa and 16 structural cysteines. Although the 3D structure is resolved, the active sites and function are poorly elucidated (Heyborne and Mackessy, 2009). The first venom CRiSP described was “helothermine” from the venomous lizards *Heloderma*, which modifies the ionic channels in mice causing lethargy, partial paralysis of the hind limbs and hypothermia (Mochca-Morales *et al.*, 1990; Morrisette *et al.*, 1995).

Pseudechetoxin, a CRiSP from *Pseudechis australis* venom inhibits smooth muscle contraction by blocking cyclic nucleotide-gated ion channels (Yamazaki and Morita, 2004). In other species such as *P. textilis*, a pseudechetoxin-like sequence was described but has not been characterized (St Pierre *et al.*, 2005).

The transcriptome revealed a unique truncated sequence (isotig 418) of CRiSP. An adenosine homopolymer might have impaired the exact sequence signal during the pyrosequencing resulting on the insertion of an artificial stop codon at the ORF position 639. However, a more careful analysis of the sequence allowed us to observe the frame shift and, after correction, to identify the full sequence of a pseudechetoxin-like toxin. CRiSP peptides were identified in spots 20, 74, 75 and 109 (Fig. 1).

4.1.7. Snake Venom Metalloproteinases (SVMPs).

Snake venom metalloproteinases are a polygenic family of enzymes, whose sequence, structure and function are well characterized. Most viper venoms contain an abundance of SVMPs that can be responsible for bite site haemorrhage, oedema, myonecrosis, blister formation, dermo-necrosis and inflammatory reactions. Systemic effects of SVMPs in snake venoms include coagulopathy, fibrinolysis, apoptosis induction, and activation of factor X and prothrombin (Markland and Swenson, 2013).

The transcriptome revealed a variety of truncated sequences of a P-III class SVMP. These variations of non-sense mutations can be interpreted as (1) individual genetic variation of the specimen, (2) negative selection of this toxin (loss of trophic adaptation) in the species or population, or even (3) a pre-adaptation or neo-functionalization process. Although the venomomics analysis did not provide reliable evidences of SVMP in the venom, western blotting revealed an immunoreactive band against anti-jararhagin polyclonal antibodies indicating the presence of P-III SVMP. Furthermore, a full P-III SVMP transcript was cloned and sequenced from a venom gland cDNA library. All those truncated sequences identified in the transcriptome match to this cloned P-III SVMP cDNA. The sequence similarity analysis (BLAST) of the cloned SVMP shows a higher similarity to related species (98 to 62% of identity with Elapidae) than to other venomous snakes (62 to 52% of identity with Viperidae).

4.2. Transcripts of interest not identified in the venom.

Some transcripts identified in the transcriptome as toxins or putative toxin transcripts were not identified at the protein level in the venom. This might be explained by the fact that (1) not all transcripts are translated into proteins, and (2) when translated it is not necessarily present in the venom, or (3) individual and regional genetic and epigenetic variations can result into different patterns of expression. Additionally, biased results derived from technical limitations (*e.g.* 2D-PAGE limitation, spot sampling and peptide ionization issues) can lead to underrepresentation of some venom components. Even so, we discuss these toxin transcripts families below.

4.2.1. Natriuretic peptides.

Venom NPs are homologous to physiological ones and act by decreasing the blood volume in the prey vessels, inducing immediate hypotension, impairing locomotion and facilitating predation (Vink *et al.*, 2012). A C-terminal fragment of a new isoform of the “PtNP-a” (St Pierre *et al.*, 2006) was identified in the isotig 227, called “PtNP-b” as well as a fragment of a new NP in isotig 1037.

4.2.2. Cobra venom factor.

The CVF is a structural and functional analogous to the C3b complement system (Fritzinger *et al.*, 1994). Its role in envenomation is not yet clear, nevertheless it has been used for the therapeutic depletion of the immune system in humans (Vogel and Fritzinger, 2010). Partial sequences were identified in singletons A5YBF, A9LX7 e AWSDT.

4.2.3. Hyaluronidase.

The isogroup 46 encodes for a hyaluronidase, an enzyme responsible for degrading the hyaluronan, a glycosaminoglycan component of the extracellular matrix in connective tissues and blood vessels. Thus, its role in envenomation is primarily described as a venom spreading factor (Kemparaju *et al.*, 2009). Isotig 221 encodes a snake venom hyaluronidase with high sequence identity (>80%) to hyaluronidases from *Micrurus fulvius* (Margres *et al.*, 2013) and *Ophiophagus hannah* (Vonk *et al.*, 2013). The isotig 222 encodes an unusual truncated version, already described in *Echis*, *Cerastes* and *Bitis* species (Harrison *et al.*, 2007).

4.2.4. Transferrin.

Transferrins have been identified in the venom proteomes of other Australian elapids including *Pseudechis australis* (Georgieva *et al.*, 2011) and *P. guttatus* (Viala *et al.*, 2014). Antimicrobial activity was detected and suggested as a probable activity in the venom (Georgieva *et al.*, 2011).

4.2.5. Dipeptidylpeptidase-IV.

Dipeptidylpeptidases-IV (DPP4) were previously described in snake venom and cloned from venom glands (Faiz *et al.*, 1996; Gasparello-Clemente and Silveira, 2002; Ogawa *et al.*, 2006; St Pierre *et al.*, 2007a). They may act by interfering in prey homeostasis by inactivating peptides like glucagon or acting upon the immune or neuroendocrine system (Ogawa *et al.*, 2006), or simply processing zymogen toxins. Truncated and fragmented isotigs and singletons were identified.

4.2.6. Veficolin.

Veficolins are venom ficolins, first described in *Cerberus rynchops* (Homoplasidae) venom as ryncolins, which contains an N-terminal collagen domain and C-terminal fibrinogen domain (Ompraba *et al.*, 2010). Ompraba *et al.* (2010) identified this protein in the venom by MS considering “hydroxylation of proline” as a variable post-translation modification. No ficolin was identified in *Pseudonaja* venom using this approach. The singleton AUJZW was

identified as a fragment of transcript encoding for veficolin, similar to ryncolins. Other singletons with truncated sequences were also identified.

4.3. Proteins of interest: venom proteins and toxin candidates.

4.3.1. VIP/Glucagon-like peptides and *Heloderma* lizard toxins.

Peptides derived from four toxins found in Gila-monster lizards (*Heloderma*) were identified in *P. textilis* venom proteome: “exendin-2 long” (Accession: EXE2_HELSU), “exendin-4” (Accession: EXE4_HELSU), “helokinstatin-1” (Accession: HKS_HELHO) and “kallikrein-toxin” (Accession: C6EVG4_HELSC) (Supplementary Material). Exendins form a peptide family similar to secretin hormones (Irwin, 2012). “Exendin-2 long” is an intestinal vasoactive peptide that induces hypotension in prey mediated by the relaxation of the myocardium (Tsueshita *et al.*, 2004) and “exendin-4”, also hypotensive, mimetizes glucagon which led to the development of the antidiabetic Byetta®, an insulin production stimulator (Furman, 2012). “Helokinstatin-1”, derived from a decapeptide, antagonizes the bradykinin vasodilatation activity on B2 receptors of bradykinin (Kwok *et al.*, 2008). Finally, “kallikrein-toxin”, described at the cDNA level (Fry *et al.*, 2010), is a kallikrein serine protease of unknown activity, with a sequence similar to gilatoxin (Utaisincharoen *et al.*, 1993). Gilatoxin, similar to batroxobin (Itoh *et al.*, 1987) and crotalase (Henschen-Edman *et al.*, 1999), has kallikrein activity and can act as a haemorrhagic potentiator of *Heloderma* toxins (Utaisincharoen *et al.*, 1993). The origin of those peptides in the venom is unknown, as no transcripts were identified in the transcriptome. No evident spot clustering was observed in the 2D-PAGE for these toxins and the identification of the peptides was always associated with other toxins (VF5_a, PLA₂, VF5_a+ PLA₂ and CRiSP).

4.3.2. Calreticulin.

Five peptides from spot 31 corresponded to calreticulin, also identified in the transcriptome (isotig 391, with 82% of identity with humans). This calreticulin sequence contains a Concanavalin A-like lectin/glucanases domain (Con A-like) followed by a calreticulin/calnexin P domain. InterPro resource (Mitchell *et al.*, 2014) predicted the whole sequence as non-cytoplasmic and without transmembrane motifs, but its function in the venom is unknown. Toxic activity was previously reported in Con A-like domain containing proteins from bacteria, viruses, lion-fish and snakes: e.g. *Clostridium* neurotoxins responsible for the neuromuscular effects of botulism and tetanus (Swaminathan and Eswaramoorthy, 2000); *Pseudomonas* exotoxin A virulence factor (arrest of protein synthesis in eukaryotic

cells) (Wedekind *et al.*, 2001); *Vibrio cholerae* neuraminidase (Crennell *et al.*, 1994) and the rotaviral outer capsid protein VP4 (cell attachment and membrane penetration) (Dormitzer *et al.*, 2002); the *Dendrochirus zebra* (former *Pterois*) lion-fish toxin (UniProt Accession: A0A068BD83_DENZE); and finally, ohanin and thaicobrin are neurotoxins found in snakes (*Ophiophagus hannah* and *Naja kaouthia* respectively) that induce hypolocomotion and hyperalgesia in mice (Pung *et al.*, 2005). Additionally, Kuwabara *et al.* (1995) show that tick salivary secreted CRT binds to coagulation factors without affecting their coagulant properties and interacts with the endothelium to stimulate release of nitric oxide and inhibit clot formation.

4.3.3. Dipeptidase 2.

The identification of dipeptidase-2 in the proteome was unexpected. Based on 3 peptides in spot 117 (Fig. 1, Supplementary material), the peptides matched with DPP-2 described in the *O. hannah* genome (Uniprot Accession: V8NS94_OPHHA) (Vonk *et al.*, 2011). To our knowledge, there is no description of such a venom component in lizard or snake databases, except for a transcript from *C. adamanteus* venom gland (Accession: J3SEA2_CROAD) (Rokyta *et al.*, 2012). DPP2 are M19 membrane peptidases with a renal dipeptidase active site. Those peptidases are involved in renal metabolism of glutathione and their conjugates (Adachi *et al.*, 1993). *O. hannah* and *C. adamanteus* dipeptidase-2 have a predicted N-terminal signal peptide but also a transmembrane region at the C-terminal region, indicating its original membrane function in the venom gland cells.

5. Conclusion.

We reported here the first full venomomic study of the medically important Australian snake *Pseudonaja textilis*. The combination of proteomics and transcriptomics allowed us to distinguish isoforms, a common limitation in proteomics due to limited tryptic peptide sequence coverage (Birrell *et al.*, 2006). The venom gland *de novo* transcriptome analysis revealed 113 different toxin transcripts, including 17 new sequences, such as the textilotoxin B subunit so far described only at the protein level and most of the toxins UTRs, and splicing variants such as the VF5a-3 (with an additional exon) and the “acidic PLA₂ isoform 4” (with an alternative signal peptide). The new splicing variant VF5a-3 and a new transcript of long neurotoxin LNTx-2 were identified at the protein level. Additionally, a P-III SVMP was cloned and detected in the venom by western blot. Based on those new observations upon

toxin transcripts splicing variants, we suggest that the alternative splicing is an important source of variation within toxin families, therefore, genomic sequences and phenotypic identification of those variants should have more attention in venomomics works.

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Highlights

- *Pseudonaja textilis* venom was analyzed by combining transcriptome and proteome.
- A novel coagulation factor 5a splicing variant was detected by both approaches.
- A new long three finger toxin was detected by both approaches.
- Two identical PLA₂s with different UTRs and signal peptides were sequenced.

Ethical statement.

The animal used in this work was euthanized for tissue collection in accordance with Euthanasia of Animals Used for Scientific Purposes guidelines (2001), Australian and New Zealand Council for the Care of Animals in Research and Teaching, under the monitoring of the SA Pathology/ CHN Animal Ethics Committee, Project Approval 93/12.

Supplementary Material: Summary table of *Pseudonaja textilis* venom MS/MS identification on 2D-PAGE spots. Data generated by PEAKS+MASCOT (InChorus) analysis against reptile and toxin public database, enriched with the in house *P. textilis* venom gland transcriptome. (+15.99): Oxidation of methionine. VF5a: Venom coagulation factor 5a; VF10a: Venom coagulation factor 10a; PLA₂: Phospholipase A₂; CRiSP: Cystein-Rich Secretory Protein; 3FTx: Three-finger toxin; HKS: Helokinestatin; EXE2: Exendin-2; EXE4: Exendin-4; BPTI: bovine protease trypsin inhibitors; CLect: C-lectin; DPP: dipeptidase; m: mass; z: charge; Cov.: Coverage.

Spot	Prot. family	Accession	Score (%)	Top 10 hit description	Hit species	Coverage (%)	Peptide	m/z	z
1	VF5a	FA5V_PSETE	99.2	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	17	K.AQYLDNFSNFIGG.K K.SNVMYTLNGYASDR.T K.WSEGSSYSDGTSOVER.L R.GEVGDSLIIYFK.N R.EYVLMFSVFDESK.N K.WTVLDTDEPTVK.D K.AVEPGQVYTYK.W K.ADVEQHAFVAVFDENK.S R.DALSGLLGPTLR.G R.LSESDLTFKK.I K.ELGLIDDEGNPIQPR.R K.WLISSLVAK.H K.NFATQPVSIHPQSAVYNK.W R.EYHIAAQLEDWDYNPQPEELSR.L K.AQYLDNFSNFIGG.K.Y R.LDDAVPPGQSFK.Y K.SWYLEDNIKK.Y K.DAEGAIYPSDPK.E K.LYHSAVDMTR.D R.LSESDLTFK.K R.GILGPVIK.A K.SWYLEDNIK.K K.LYHSAVDM(+15.99)TR.D R.DVTIVFK.N	758.88 795.8 881.36 670.89 797.4 702.4 627.82 607.01 606.87 584.3 890 508.84 667.72 901.76 548.99 637.31 648.38 631.84 398.2 520.29 398.8 584.34 403.56 461.78	2 2 2 2 2 2 3 2 2 2 2 3 3 2 2 2 2 3 3 2 2 2 2 2
		isotig00199	99.2	VF5a-3	<i>Pseudonaja textilis</i>	28	K.AQYLDNFSNFIGG.K K.SNVMYTLNGYASDR.T K.WSEGSSYSDGTSOVER.L R.GEVGDSLIIYFK.N R.EYVLMFSVFDESK.N K.WTVLDTDEPTVK.D K.AVEPGQVYTYK.W R.DALSGLLGPTLR.G R.LSESDLTFKK.I K.ELGLIDDEGNPIQPR.R K.WLISSLVAK.H K.NFATQPVSIHPQSAVYNK.W R.EYHIAAQLEDWDYNPQPEELSR.L K.AQYLDNFSNFIGG.K.Y R.LDDAVPPGQSFK.Y K.SWYLEDNIKK.Y K.DAEGAIYPSDPK.E K.LYHSAVDMTR.D R.LSESDLTFK.K R.GILGPVIK.A K.SWYLEDNIK.K R.EYELDFKQEKPR.D K.LYHSAVDM(+15.99)TR.D R.DVTIVFK.N	758.88 795.8 881.36 670.89 797.4 702.4 627.82 606.87 584.3 890 508.84 667.72 901.76 548.99 637.31 648.38 631.84 398.2 520.29 398.8 584.34 527.92 403.56 461.78	2 2 2 2 2 2 2 2 2 2 2 3 3 2 2 2 3 3 2 2 3 3 2
		isotig00200	99.2	VF5a isoform 2	<i>Pseudonaja textilis</i>	17	K.AQYLDNFSNFIGG.K K.SNVMYTLNGYASDR.T K.WSEGSSYSDGTSOVER.L R.GEVGDSLIIYFK.N R.EYVLMFSVFDESK.N K.WTVLDTDEPTVK.D K.AVEPGQVYTYK.W R.DALSGLLGPTLR.G R.LSESDLTFKK.I K.ELGLIDDEGNPIQPR.R K.WLISSLVAK.H K.NFATQPVSIHPQSAVYNK.W R.EYHIAAQLEDWDYNPQPEELSR.L K.AQYLDNFSNFIGG.K.Y R.LDDAVPPGQSFK.Y K.SWYLEDNIKK.Y K.DAEGAIYPSDPK.E K.LYHSAVDMTR.D R.LSESDLTFK.K R.GILGPVIK.A K.SWYLEDNIK.K R.EYELDFKQEKPR.D K.LYHSAVDM(+15.99)TR.D R.DVTIVFK.N	758.88 795.8 881.36 670.89 797.4 702.4 627.82 606.87 584.3 890 508.84 667.72 901.76 548.99 637.31 648.38 631.84 398.2 520.29 398.8 584.34 527.92 403.56 461.78	2 2 2 2 2 2 2 2 2 2 2 3 3 2 2 2 3 3 2 2 3 3 2
	FA5V_OXYSU		99.2	Venom prothrombin activator Ocutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	14	K.AQYLDNFSNFIGG.K K.SNVMYTLNGYASDR.T K.WSEGSSYSDGTSOVER.L R.GEVGDSLIIYFK.N R.EYVLMFSVFDESK.N K.WTVLDTDEPTVK.D K.AVEPGQVYTYK.W K.ADVEQHAFVAVFDENK.S R.LSESDLTFKK.I K.WLISSLVAK.H K.NFATQPVSIHPQSAVYNK.W K.AQYLDNFSNFIGG.Y R.LDDAVPPGQSFK.Y K.SWYLEDNIKK.Y K.LYHSAVDMTR.D R.LSESDLTFK.K R.GILGPVIK.A K.SWYLEDNIK.K R.EYELDFKQEKPR.D K.LYHSAVDM(+15.99)TR.D R.DVTIVFK.N	758.88 795.8 881.36 670.89 797.4 702.4 627.82 607.01 584.3 508.84 667.72 548.99 637.31 648.38 398.2 520.29 398.8 584.34 527.92 403.56 461.78	2 2 2 2 2 2 2 3 2 2 3 3 2 2 2 2 2 2 3 3 2
	FA5_PSETE		99.2	Coagulation factor V	<i>Pseudonaja textilis</i>	14	K.AQYLDNFSNFIGG.K K.SNVMYTLNGYASDR.T K.WSEGSSYSDGTSOVER.L R.EYVLMFSVFDESK.N K.WTVLDTDEPTVK.D K.AVEPGQVYTYK.W K.ADVEQHAFVAVFDENK.S R.LSESDLTFKK.I K.WLISSLVAK.H K.NFATQPVSIHPQSAVYNK.W R.EYHIAAQLEDWDYNPQPEELSR.L K.AQYLDNFSNFIGG.Y R.LDDAVPPGQSFK.Y K.SWYLEDNIKK.Y K.DAEGAIYPSDPK.E K.LYHSAVDMTR.D R.LSESDLTFK.K K.SWYLEDNIK.K R.EYELDFKQEKPR.D K.LYHSAVDM(+15.99)TR.D	758.88 795.8 881.36 797.4 702.4 627.82 607.01 584.3 508.84 667.72 901.76 548.99 637.31 648.38 631.84 398.2 520.29 584.34 527.92 403.56	2 2 2 2 2 2 3 2 2 3 3 3 2 2 3 2 2 2 3
	FA5V_OXYMI		99.2	Venom prothrombin activator omicaric-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	13	K.AQYLDNFSNFIGG.K K.SNVMYTLNGYASDR.T K.WSEGSSYSDGTSOVER.L R.EYVLMFSVFDESK.N K.WTVLDTDEPTVK.D K.AVEPGQVYTYK.W K.ADVEQHAFVAVFDENK.S K.WLISSLVAK.H	758.88 795.8 881.36 797.4 702.4 627.82 607.01 508.84	2 2 2 2 2 2 3 2

						K.NFATQPVSIHPQSAVYNK.W	667.72	3
						K.AQYLDNFSNFIGKK.Y	548.99	3
						R.LDDAVPPGQSF.K.Y	637.31	2
						K.SWYLEDNIKK.Y	648.38	2
						K.DAEGAIPSPDK.E	631.84	2
						K.LYHSAVDMTR.D	398.2	3
						R.GILGPVIK.A	398.8	2
						K.SWYLEDNIK.K	584.34	2
						R.EYELDFKQEKPR.D	527.92	3
						K.LYHSAVDM(+15.99)TR.D	403.56	3
						R.DTVTIVFK.N	461.78	2
FA5V_PSETE	99.19	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	21		K.WSEGSSYSDGTS DVER.L	881.35	2
						K.VSTINLVGGASVTADMSVSR.T	982.52	2
						K.SNVMYTLNGYASDR.T	795.87	2
						K.ADVEQHAFVAVFDENK.S	909.97	2
						K.AQYLDNFSNFIGK.K	758.87	2
						K.AVEPGQVYTYK.W	627.82	2
						R.GEVGDSLIIYFK.N	670.88	2
						K.NFATQPVSIHPQSAVYNK.W	1001	2
						R.EYVLMFSVFDESK.N	797.4	2
						K.WTVLDTDEPTVK.D	702.38	2
						K.ELGLIDDEGNPIQPR.R	889.98	2
						K.EEVPVNFVPDPESDALAK.E	978.47	2
						R.DALSGLLGPTLR.G	606.89	2
						K.AQYLDNFSNFIGKK.Y	822.93	2
						R.LSEDLTFKK.I	584.32	2
						K.DAEGAIPSPDK.E	631.82	2
						K.LYHSAVDMTR.D	596.81	2
						K.WLISSLVAK.H	508.86	2
						R.LDDAVPPGQSF.K.Y	637.31	2
						K.SWYLEDNIK.K	584.34	2
						R.GILGPVIK.A	398.8	2
						K.SWYLEDNIKK.Y	648.39	2
						K.NLASRPYSIYVHGVSVSK.D	659.71	3
						R.DTVTIVFK.N	461.77	2
						K.VRDTVIVFK.N	589.39	2
						K.HLQAGMYGLNIK.D	503.37	3
						K.WSEGSSYSDGTS DVER.LDDAVPPGQSF.K.Y	1006.11	3
isotig00199	99.19	VF5a-3	<i>Pseudonaja textilis</i>	31		K.WSEGSSYSDGTS DVER.L	881.35	2
isotig00200	99.19	VF5a-2	<i>Pseudonaja textilis</i>	19		K.SNVMYTLNGYASDR.T	795.87	2
						K.AQYLDNFSNFIGK.K	758.87	2
						K.AVEPGQVYTYK.W	627.82	2
						R.GEVGDSLIIYFK.N	670.88	2
						K.NFATQPVSIHPQSAVYNK.W	1001	2
						R.EYVLMFSVFDESK.N	797.4	2
						K.WTVLDTDEPTVK.D	702.38	2
						K.ELGLIDDEGNPIQPR.R	889.98	2
						K.EEVPVNFVPDPESDALAK.E	978.47	2
						R.DALSGLLGPTLR.G	606.89	2
						K.AQYLDNFSNFIGKK.Y	822.93	2
						R.LSEDLTFKK.I	584.32	2
						K.DAEGAIPSPDK.E	631.82	2
						K.LYHSAVDMTR.D	596.81	2
						K.WLISSLVAK.H	508.86	2
						R.LDDAVPPGQSF.K.Y	637.31	2
						K.SWYLEDNIK.K	584.34	2
						R.GILGPVIK.A	398.8	2
						K.SWYLEDNIKK.Y	648.39	2
						K.NLASRPYSIYVHGVSVSK.D	659.71	3
						R.DTVTIVFK.N	461.77	2
						K.VRDTVIVFK.N	589.39	2
						K.HLQAGMYGLNIK.D	503.37	3
						R.EYELDFKQEKPR.D	791.52	2
						K.WSEGSSYSDGTS DVER.LDDAVPPGQSF.K.Y	1006.11	3
FA5V_OXYSU	99.19	Venom prothrombin activator Oscutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	18		K.WSEGSSYSDGTS DVER.L	881.35	2
						K.SNVMYTLNGYASDR.T	795.87	2
						K.ADVEQHAFVAVFDENK.S	909.97	2
						K.AQYLDNFSNFIGK.K	758.87	2
						K.AVEPGQVYTYK.W	627.82	2
						R.GEVGDSLIIYFK.N	670.88	2
						K.NFATQPVSIHPQSAVYNK.W	1001	2
						R.EYVLMFSVFDESK.N	797.4	2
						K.WTVLDTDEPTVK.D	702.38	2
						K.EEVPVNFVPDPESDALAK.E	978.47	2
						K.AQYLDNFSNFIGKK.Y	822.93	2
						R.LSEDLTFKK.I	584.32	2
						K.LYHSAVDMTR.D	596.81	2
						K.WLISSLVAK.H	508.86	2
						R.LDDAVPPGQSF.K.Y	637.31	2
						K.SWYLEDNIK.K	584.34	2
						R.GILGPVIK.A	398.8	2
						K.SWYLEDNIKK.Y	648.39	2
						K.NLASRPYSIYVHGVSVSK.D	659.71	3
						R.DTVTIVFK.N	461.77	2
						K.VRDTVIVFK.N	589.39	2
						K.HLQAGMYGLNIK.D	503.37	3
						R.EYELDFKQEKPR.D	791.52	2
						K.WSEGSSYSDGTS DVER.LDDAVPPGQSF.K.Y	1006.11	3
FA5V_OXYMI	99.19	Venom prothrombin activator omicarín-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	17		K.WSEGSSYSDGTS DVER.L	881.35	2
						K.SNVMYTLNGYASDR.T	795.87	2
						K.ADVEQHAFVAVFDENK.S	909.97	2
						K.AQYLDNFSNFIGK.K	758.87	2
						K.AVEPGQVYTYK.W	627.82	2
						K.NFATQPVSIHPQSAVYNK.W	1001	2
						R.EYVLMFSVFDESK.N	797.4	2
						K.WTVLDTDEPTVK.D	702.38	2
						K.EEVPVNFVPDPESDALAK.E	978.47	2
						K.AQYLDNFSNFIGKK.Y	822.93	2

							K.DAEGAIYSPDK.E	631.82	2
							K.LYHSAVDMTR.D	596.81	2
							K.WLISSLVAK.H	508.86	2
							R.LDDAVPPGQSF.K.Y	637.31	2
							K.SWYLEDNIK.K	584.34	2
							R.GILGPVIK.A	398.8	2
							K.SWYLEDNIK.Y	648.39	2
							K.NLASRPYSIYVHGVSVSK.D	659.71	3
							R.DTVTIVFK.N	461.77	2
							K.VRDTVIVFK.N	589.39	2
							K.HLQAGMYGYLNIK.D	503.37	3
							R.EYELDFKQEKPR.D	791.52	2
							K.WSEGSSYSDGTSOVERLDDAVPPGQSF.K.Y	1006.11	3
FA5_PSETE	99.19		Coagulation factor V	<i>Pseudonaja textilis</i>	15		K.WSEGSSYSDGTSOVER.L	881.35	2
							K.SNVMYTLNGYASDR.T	795.87	2
							K.ADVEQHAVFAVDENK.S	909.97	2
							K.AQYLDNFSNFIGK.K	758.87	2
							K.AVEPGQVYTYK.W	627.82	2
							K.NFATQPVSIHPQSAVYNK.W	1001	2
							R.EYVLMFSVFDESK.N	797.4	2
							K.WTVLDTDEPTVK.D	702.38	2
							K.AQYLDNFSNFIGK.Y	822.93	2
							R.LSEDLTFKK.I	584.32	2
							K.DAEGAIYSPDK.E	631.82	2
							K.LYHSAVDMTR.D	596.81	2
							K.WLISSLVAK.H	508.86	2
							R.LDDAVPPGQSF.K.Y	637.31	2
							K.SWYLEDNIK.K	584.34	2
							K.SWYLEDNIK.Y	648.39	2
							K.NLASRPYSIYVHGVSVSK.D	659.71	3
							K.HLQAGMYGYLNIK.D	503.37	3
							R.EYELDFKQEKPR.D	791.52	2
							K.WSEGSSYSDGTSOVERLDDAVPPGQSF.K.Y	1006.11	3
3	VF5a	isotig00199	99.19	VF5a-3	<i>Pseudonaja textilis</i>	31	K.WSEGSSYSDGTSOVER.L	881.36	2
		isotig00200	99.19	VF5a-2	<i>Pseudonaja textilis</i>	19	K.AQYLDNFSNFIGK.K	758.88	2
							R.EYVLMFSVFDESK.N	797.4	2
							K.NFATQPVSIHPQSAVYNK.W	1000.97	2
							K.SNVMYTLNGYASDR.T	795.86	2
							K.WTVLDTDEPTVK.D	702.39	2
							R.GEVGDILIIYFK.N	670.91	2
							R.DALSGLLGPTLR.G	606.87	2
							K.AVEPGQVYTYK.W	627.82	2
							K.EEVPVNFVPDPESDALAK.E	978.47	2
							K.AQYLDNFSNFIGK.Y	822.93	2
							R.LSEDLTFKK.I	584.32	2
							K.DAEGAIYSPDK.E	631.83	2
							R.LSEDLTFK.K	520.31	2
							K.GVQNSADVEQHAVFAVDENK.S	768.73	3
							R.LDDAVPPGQSF.K.Y	637.31	2
							K.WLISSLVAK.H	508.87	2
							K.LYHSAVDMTR.D	596.79	2
							K.SWYLEDNIK.K	584.35	2
							K.VRDTVIVFK.N	589.42	2
							K.SWYLEDNIK.Y	648.41	2
							K.NLASRPYSIYVHGVSVSK.D	659.7	3
							K.LYHSAVDM(+15.99)TR.D	604.85	2
							R.GILGPVIK.A	398.78	2
							R.TEVL.R.F	309.23	2
							R.EYELDFKQEKPR.D	527.93	3
FA5V_OXYMI	99.19		Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	16		K.WSEGSSYSDGTSOVER.L	881.36	2
							K.AQYLDNFSNFIGK.K	758.88	2
							R.EYVLMFSVFDESK.N	797.4	2
							K.NFATQPVSIHPQSAVYNK.W	1000.97	2
							K.SNVMYTLNGYASDR.T	795.86	2
							K.WTVLDTDEPTVK.D	702.39	2
							K.AVEPGQVYTYK.W	627.82	2
							K.EEVPVNFVPDPESDALAK.E	978.47	2
							K.AQYLDNFSNFIGK.Y	822.93	2
							R.GEVGDILIIYFK.N	683.94	2
							K.DAEGAIYSPDK.E	631.83	2
							R.LDDAVPPGQSF.K.Y	637.31	2
							K.WLISSLVAK.H	508.87	2
							K.LYHSAVDMTR.D	596.79	2
							K.SWYLEDNIK.K	584.35	2
							K.VRDTVIVFK.N	589.42	2
							K.SWYLEDNIK.Y	648.41	2
							K.NLASRPYSIYVHGVSVSK.D	659.7	3
							K.LYHSAVDM(+15.99)TR.D	604.85	2
							R.GILGPVIK.A	398.78	2
							R.EYELDFKQEKPR.D	527.93	3
FA5_PSETE	99.19		Coagulation factor V	<i>Pseudonaja textilis</i>	14		K.WSEGSSYSDGTSOVER.L	881.36	2
							K.AQYLDNFSNFIGK.K	758.88	2
							R.EYVLMFSVFDESK.N	797.4	2
							K.NFATQPVSIHPQSAVYNK.W	1000.97	2
							K.SNVMYTLNGYASDR.T	795.86	2
							K.WTVLDTDEPTVK.D	702.39	2
							K.AVEPGQVYTYK.W	627.82	2
							K.AQYLDNFSNFIGK.Y	822.93	2
							R.LSEDLTFKK.I	584.32	2
							R.GEVGDILIIYFK.N	683.94	2
							K.DAEGAIYSPDK.E	631.83	2
							R.LSEDLTFK.K	520.31	2
							R.LDDAVPPGQSF.K.Y	637.31	2
							K.WLISSLVAK.H	508.87	2
							K.LYHSAVDMTR.D	596.79	2
							K.SWYLEDNIK.K	584.35	2
							K.SWYLEDNIK.Y	648.41	2
							K.NLASRPYSIYVHGVSVSK.D	659.7	3

ACCEPTED MANUSCRIPT

							K.LYHSAVDM(+15.99)TR.D	604.85	2
							R.EYELDFKQEKPR.D	527.93	3
4	VF5a	FASV_PSETE	99	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	4	R.GEVGDSLIYFK.N	670.89	2
							K.AQYLDNFSNFIGK.K	758.93	2
							K.WTVLDTDEPTVK.D	702.38	2
							K.ADVEQHAVFAVDENK.S	607	3
		isotig00199	98.8	VF5a-3	<i>Pseudonaja textilis</i>	6	R.GEVGDSLIYFK.N	670.89	2
		isotig00200	98.8	VF5a-2	<i>Pseudonaja textilis</i>	4	K.AQYLDNFSNFIGK.K	758.93	2
							K.WTVLDTDEPTVK.D	702.38	2
							K.GVQNSADVEQHAVFAVDENK.S	768.72	3
5	VF5a	FASV_PSETE	99.14	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	6	K.AQYLDNFSNFIGK.K	758.9	2
							K.AVEPGQVYTYK.W	627.8	2
							K.ADVEQHAVFAVDENK.S	606.98	3
							K.WTVLDTDEPTVK.D	702.42	2
							K.WLISSLVAK.H	508.87	2
							K.VSTINLVGGASVTADMSVSR.T	982.38	2
							K.LYHSAVDM(+15.99)TR.D	403.58	3
6	VF5a	isotig00199	99.02	VF5a-3	<i>Pseudonaja textilis</i>	5	K.AQYLDNFSNFIGK.K	758.9	2
		FASV_OXYMI	99.02	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	3	K.WTVLDTDEPTVK.D	702.4	2
		FASV_PSETE	99.02	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	3	K.AVEPGQVYTYK.W	627.85	2
		isotig00200	99.02	VF5a-2	<i>Pseudonaja textilis</i>	3	K.LYHSAVDM(+15.99)TR.D	403.55	3
7	VF5a	FASV_OXYSU	99.15	Venom prothrombin activator Oscutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	5	K.AQYLDNFSNFIGK.K	758.9	2
		FASV_OXYMI	99.15	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	5	K.AVEPGQVYTYK.W	627.82	2
		FASV_PSETE	99.15	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	5	K.WTVLDTDEPTVK.D	702.4	2
							K.ADVEQHAVFAVDENK.S	607	3
							K.SWYLEDNIKK.Y	648.41	2
							K.LYHSAVDMTR.D	596.82	2
							R.GILGPVIK.A	398.78	2
8	VF5a	isotig00199	98.85	VF5a-3	<i>Pseudonaja textilis</i>	4	K.AVEPGQVYTYK.W	627.8	2
		FASV_PSETE	98.85	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	2	K.WTVLDTDEPTVK.D	702.39	2
		isotig00200	98.85	VF5a-2	<i>Pseudonaja textilis</i>	2	K.LYHSAVDMTR.D	596.81	2
		FASV_OXYMI	98.84	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	2			
10	VF5a	FASV_OXYMI	96.72	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	2	K.ADVEQHAVFAVDENK.S	1817.8584	2
		FASV_PSETE	96.72	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	2	K.SWYLEDNIKK.Y	1294.6558	1
		FASV_OXYSU	95.61	Venom prothrombin activator Oscutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	2			
		FA5_PSETE	95.61	Coagulation factor V	<i>Pseudonaja textilis</i>	2			
		V8P243_OPHHA	95.61	Coagulation factor V	<i>Ophiophagus hannah</i>	1			
16	PLA2	isotig00264	84.32	Acidic PLA2-A4	<i>Pseudonaja textilis</i>	16	R.FSGPYWNPNYSYK.C	754.83	2
		isotig00263	84.32	Acidic PLA2-A3	<i>Pseudonaja textilis</i>	16	K.GSGSTPVDLDR.C	601.79	2
		PA2A2_PSETE	84.32	Acidic phospholipase A2 2	<i>Pseudonaja textilis</i>	16			
18	VF10a	FA102_PSETE	97.05	Coagulation factor X isoform 2	<i>Pseudonaja textilis</i>	6	K.QDFGIVSGFGR.I	591.83	2
							R.IETGPLLSVDK.I	586.37	2
							K.YGYTK.L	372.76	2
19	VF10a	FAXC_OXYSU	60.64	Oscutarin-C catalytic subunit	<i>Oxyuranus scutellatus</i>	1	K.YGVYTK.V	365.7	2
20	CRISP	isotig00418	64.83	Cysteine-rich venom protein pseudochetoxin-like	<i>Pseudonaja textilis</i>	3	R.NMLQMK.W	382.69	2
		R4G2J3_9SAUR	64.81	CRISP-Pse-4	<i>Pseudonaja modesta</i>	3			
		R4FIS5_9SAUR	64.81	CRISP-Pse-17	<i>Pseudonaja modesta</i>	3			
		CRVP_OXYMI	64.81	Cysteine-rich venom protein pseudochetoxin-like	<i>Oxyuranus microlepidotus</i>	3			
		CRVP_PSETE	64.81	Cysteine-rich venom protein pseudochetoxin-like	<i>Pseudonaja textilis</i>	3			
		CRVP_OXYSC	64.81	Cysteine-rich venom protein pseudochetoxin-like	<i>Oxyuranus s. scutellatus</i>	3			
		R4G7K2_9SAUR	64.81	CRISP-Pse-11	<i>Pseudonaja modesta</i>	3			
		CRVP_PSEPO	62.46	Cysteine-rich venom protein pseudocin	<i>Pseudechis porphyriacus</i>	3	R.NMLQMK.W	382.69	2
		CRVP_PSEAU	62.46	Cysteine-rich venom protein pseudochetoxin	<i>Pseudechis australis</i>	3			
21	VF5a	isotig00200	99.04	VF5a-2	<i>Pseudonaja textilis</i>	4	R.DALSGLLGPTLR.G	606.94	2
							K.NSEITASSYK.K	550.29	2
							K.TWNQYIALR.I	582.86	2
							K.TWWSSWEPFLAR.L	783.39	2
							K.ENHIDPPIAR.Y	425.63	3
							K.NSEITASSYKK.T	614.33	2
							K.HLGILGPIR.A	544.84	2
22	VF5a	FASV_OXYSU	61.55	Venom prothrombin activator Oscutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	1			
		FA5_PSETE	61.55	Coagulation factor V	<i>Pseudonaja textilis</i>	1			
		FASV_OXYMI	61.55	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	1			
		FASV_PSETE	61.55	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	1			
		isotig00200	61.55	VF5a-2	<i>Pseudonaja textilis</i>	1			
		V8P243_OPHHA	61.54	Coagulation factor V	<i>Ophiophagus hannah</i>	1			
23	VF5a	FASV_OXYMI	99.12	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	5	K.SWAYYSGVNPEK.D	700.85	2
		FASV_PSETE	99.12	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	5	K.NSEITASSYK.K	550.3	2
		FA5_PSETE	99.12	Coagulation factor V	<i>Pseudonaja textilis</i>	5	K.TWNQYIALR.I	582.86	2
							K.ENHIDPPIAR.Y	425.59	3
							K.VFTGNINSDGHVK.H	463.26	3
							R.EDNLGVLP LLPGTFASIK.M	1006.56	2
24	VF5a	isotig00200	99.19	VF5a-2	<i>Pseudonaja textilis</i>	11	R.SYLDLDTFQTPSTGGYEYK.H	1019.45	2
							R.AEVDVIEIQFR.N	717.38	2
							K.SWAYYSGVNPEK.D	700.86	2
							R.GMQALFTVIDK.D	611.89	2
							K.NSEITASSYK.K	550.31	2
							R.SYDDKSPFLK.K	664.8	2
							K.IGTWLLETEVGGENQER.G	625.33	3
							K.HLGILGPIR.A	544.89	2
							K.TWNQYIALR.I	582.86	2
							K.TWWSSWEPFLAR.L	783.39	2
							K.DQWLQIDLQHLTK.I	546.66	3
							R.HSETQM(+15.99)HFEGNSDGGTTVK.E	674.34	3
							R.SYDDKSPFLK.K	486.23	3
							K.ENHIDPPIAR.Y	637.86	2
							K.SPFLK.K	360.72	2
							K.NSEITASSYKK.T	614.31	2
25	VF5a	isotig00199	99.19	VF5a-3	<i>Pseudonaja textilis</i>	13	K.AQYLDNFSNFIGK.K	758.86	2
		FASV_PSETE	99.19	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	8	K.AVEPGQVYTYK.W	627.82	2
		isotig00200	99.19	VF5a-2	<i>Pseudonaja textilis</i>	8	K.WTVLDTDEPTVK.D	702.38	2
							K.SNVMTYTLNGYASDR.T	795.91	2
							R.DALSGLLGPTLR.G	606.89	2
							K.WLISSLVAK.H	508.83	2
							K.SNVM(+15.99)YTLNGYASDR.T	803.84	2
							K.AQYLDNFSNFIGKK.Y	822.96	2
							K.SWYLEDNIK.K	584.3	2
							K.SWYLEDNIKK.Y	648.37	2

							K.LYHSAVDMTR.D	596.74	2
							R.GILGPVIK.A	398.82	2
							K.WSEGSSYSDGTSOVER.L	881.34	2
26	VF5a	isotig00199	99.19	VF5a-3	<i>Pseudonaja textilis</i>	12	K.AQYLDNFSNFIGK.K	758.88	2
		FA5V_OXYMI	99.19	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	8	K.AVEPGQVYTYK.W	627.87	2
		FA5V_PSETE	99.19	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	8	K.SNVMYTLNGYASDR.T	795.88	2
		isotig00200	99.19	VF5a-2	<i>Pseudonaja textilis</i>	8	K.WTVLDTDEPTVK.D	702.38	2
							K.WLISSLVAK.H	508.83	2
							R.GILGPVIK.A	398.81	2
							K.NLASRPYSIVVHGVSVK.D	659.73	3
							K.SWYLEDNIIK.K	584.34	2
							R.DTVTIVFK.N	461.81	2
							K.LYHSAVDMTR.D	398.17	3
27	VF5a	isotig00199	99.19	VF5a-3	<i>Pseudonaja textilis</i>	13	K.AQYLDNFSNFIGK.K	758.86	2
		FA5V_OXYMI	99.19	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	8	K.SNVMYTLNGYASDR.T	795.85	2
		FA5V_PSETE	99.19	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	8	K.AVEPGQVYTYK.W	627.84	2
		isotig00200	99.19	VF5a-2	<i>Pseudonaja textilis</i>	8	K.WTVLDTDEPTVK.D	702.38	2
							K.VRDTVTIVFK.N	589.39	2
							K.NLASRPYSIVVHGVSVK.D	659.71	3
							K.AQYLDNFSNFIGK.K.Y	822.89	2
							R.GILGPVIK.A	398.81	2
							K.SWYLEDNIIK.K	584.3	2
							K.KKEEVPVNFVDPFESDALAK.E	738.08	3
							R.DTVTIVFK.N	461.78	2
28	VF5a	FA5V_OXYSU	99.14	Venom prothrombin activator Ocutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	4	K.SNV(+15.99)YTLNGYASDR.T	803.83	2
		FA5V_OXYMI	99.14	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	4	K.SNVMYTLNGYASDR.T	795.85	2
		FA5V_PSETE	99.14	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	4	K.AVEPGQVYTYK.W	627.83	2
		V8P243_OPHHA	98.81	Coagulation factor V	<i>Ophiophagus hannah</i>	2	K.WTVLDTDEPTVK.D	702.38	2
							R.GILGPVIK.A	398.83	2
							K.SWYLEDNIIK.K	584.34	2
29	VF5a	isotig00199	81.75	VF5a-3	<i>Pseudonaja textilis</i>	3	K.AVEPGQVYTYK.W	627.83	2
		FA5V_OXYSU	81.75	Venom prothrombin activator Ocutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	2	K.WTVLDTDEPTVK.D	702.37	2
		FA5_PSETE	81.75	Coagulation factor V	<i>Pseudonaja textilis</i>	2	R.LGEYHINHHRDEGEQER.R	661.34	3
		FA5V_OXYMI	81.75	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	2			
		FA5V_PSETE	81.75	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	2			
		isotig00200	81.75	VF5a-2	<i>Pseudonaja textilis</i>	2			
31	Calreticulin	V8PGR3_OPHHA	98.97	Calreticulin	<i>Ophiophagus hannah</i>	9	K.KPDDWDER.A	530.78	2
		U3FZP8_MICFL	98.97	Calreticulin	<i>Micrurus fulvius</i>	9	K.TLVVQFTVK.H	517.87	2
		isotig00391	98.97	Calreticulin-like	<i>Pseudonaja textilis</i>	9	K.VHVIFNVK.G	510.34	2
		G1KTK9_ANOCA	98.97	Uncharacterized protein	<i>Anolis carolinensis</i>	9	R.QIDNPYK.G	496.3	2
							K.NVLINK.D	350.78	2
33	VF5a	isotig00199	98.38	VF5a-3	<i>Pseudonaja textilis</i>	3	K.SNVMYTLNGYASDR.T	795.89	2
		FA5V_PSETE	98.22	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	2	K.SNV(+15.99)YTLNGYASDR.T	803.85	2
		isotig00200	98.22	VF5a-2	<i>Pseudonaja textilis</i>	2	K.SWYLEDNIIK.Y	648.38	2
37	3FTx	NXS2_PSETE	61.66	Short neurotoxin 2	<i>Pseudonaja textilis</i>	18	R.YLIPATHGNAIPAR.G	747.38	2
		NXS7_PSETE	61.66	Short neurotoxin 7	<i>Pseudonaja textilis</i>	18	R.YIIPATHGNAITYR.G	795.45	2
		isotig00066	61.66	SNTx7	<i>Pseudonaja textilis</i>	15			
		isotig00074	61.66	SNTx7	<i>Pseudonaja textilis</i>	18			
		NXS3_PSETE	61.65	Short neurotoxin 3	<i>Pseudonaja textilis</i>	18	R.YLVPATHGNAIPAR.G	740.42	2
		NXL1_PSETE	56.79	Long neurotoxin 1	<i>Pseudonaja textilis</i>	7	R.TWNDR.G	374.71	2
		R4FK93_9SAUR	56.79	3FTx-Pse-38	<i>Pseudonaja modesta</i>	7			
		R4FIU6_9SAUR	56.79	3FTx-Pse-23	<i>Pseudonaja modesta</i>	7			
		isotig00174	56.79	LNTx 1	<i>Pseudonaja textilis</i>	7			
		isotig00176	56.79	LNTx 1	<i>Pseudonaja textilis</i>	7			
	VF5a	isotig00199	59.84	VF5a-3	<i>Pseudonaja textilis</i>	1	R.NPDDIAGR.Y	429.17	2
		FA5_PSETE	59.53	Coagulation factor V	<i>Pseudonaja textilis</i>	1			
		FA5V_PSETE	59.53	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	1			
		isotig00200	59.53	VF5a-2	<i>Pseudonaja textilis</i>	1			
		V8P243_OPHHA	59.28	Coagulation factor V	<i>Ophiophagus hannah</i>	0			
38	VF10a	FAXC_PSETE	99.15	Venom prothrombin activator pseutarin-C catalytic subunit	<i>Pseudonaja textilis</i>	12	K.QDFGIVSGFGIFER.G	814.92	2
		isotig00188	99.15	VF10a isoform 1	<i>Pseudonaja textilis</i>	12	K.LPSTESSTGRL	574.3	2
		isotig00189	99.15	VF10a isoform 1	<i>Pseudonaja textilis</i>	12	K.LPSTESSTGRL	517.79	2
							R.AETGPLLSVDK.V	565.3	2
							K.VLKVPYVDR.H	363.61	3
							R.QKLPSTESSTGRL	645.83	2
							K.FIPWIKR.I	320.58	3
							K.VPYVDR.H	374.73	2
		FAXC_OXYSU	98.81	Ocutarin-C catalytic subunit	<i>Oxyuranus scutellatus</i>	6	K.LPSTESSTGRL	574.3	2
							K.LPSTESSTGRL	517.79	2
							K.VLKVPYVDR.H	363.61	3
							R.QKLPSTESSTGRL	645.83	2
							K.YGVYTK.V	365.71	2
							K.VPYVDR.H	374.73	2
		FAXD2_DEMVE	81.24	Venom prothrombin activator vestarin-D2	<i>Demansia vestigiata</i>	2	K.VIRVPYVDR.Y	558.87	2
							R.VPYVDR.Y	374.73	2
		V8PHG1_OPHHA	75.66	Coagulation factor X isoform 1 (Fragment)	<i>Ophiophagus hannah</i>	1	K.YGVYTK.V	365.71	2
							K.FIPWIKR.I	320.58	3
		FAXD1_NOTSC	62.97	Venom prothrombin activator notecarin-D1	<i>Notechis s. scutatus</i>	2	K.YGVYTKVSR.F	536.79	2
		FAXD_HOPST	62.97	Venom prothrombin activator hopsarin-D	<i>Hoplocephalus stephensii</i>	2	K.YGVYTK.V	365.71	2
40	VF5a	isotig00199	99.17	VF5a-3	<i>Pseudonaja textilis</i>	10	K.WSEGSSYSDGTSOVER.L	881.35	2
		isotig00200	99.17	VF5a-2	<i>Pseudonaja textilis</i>	6	R.GEVGDSLIYFK.N	670.86	2
							R.DALSGLLGPTLR.G	606.86	2
							K.WLISSLVAK.H	508.82	2
							R.LDDAVPPGQSFK.Y	637.27	2
							K.IVYREYELDFK.Q	737.96	2
							K.HLQAGM(+15.99)YGYLNIIK.D	508.63	3
							R.EYELDFKQEKPR.D	527.9	3
41	VF5a	isotig00200	99.18	VF5a-2	<i>Pseudonaja textilis</i>	5	K.IGTWLLETEVGENQER.G	937.48	2
							R.AEVDVIEIQFR.N	717.39	2
							K.SWAYYSGVNPKEK.D	700.86	2
							R.GEVGDSLIYFK.N	670.87	2
							K.HLGLGPIIR.A	544.83	2
							R.GMQALFTVIDK.D	611.83	2
42	VF5a	isotig00200	99.17	VF5a-2	<i>Pseudonaja textilis</i>	5	K.IGTWLLETEVGENQER.G	937.49	2
							R.AEVDVIEIQFR.N	717.37	2
							K.SWAYYSGVNPKEK.D	700.85	2
							K.HLGLGPIIR.A	544.85	2
							K.DIHVVNFHQFTTEGR.E	662.69	3

						K.SWYFPK.S	414.27	2	
43	VF5a	isotig00200	99.2	VF5a-2	<i>Pseudonaja textilis</i>	9	K.IGTWLLETEVGNER.Q	937.46	2
							R.GMQALFTVIDK.D	611.86	2
							R.AEVDVIEIQFR.N	717.39	2
							R.GM(+15.99)QALFTVIDK.D	619.87	2
							K.VFTGNINSDGHVK.H	694.38	2
							K.TWNQYIALR.I	582.86	2
							K.NSEITASSYK.K	550.32	2
							K.DIHVVNFHGQTFTEGR.E	662.7	3
							K.ENHIDPPIAR.Y	425.59	3
							K.ITSITQGATSMTTSM(+15.99)YVK.T	1025.03	2
							K.HFFKPPILSR.F	414.64	3
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	689.04	3
44	VF5a	isotig00199	99.15	VF5a-3	<i>Pseudonaja textilis</i>	10	K.WSEGSSYSDGTSVDER.L	881.36	2
	FA5V_PSETE	99.15	Venom prothrombin activator	pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	6	R.DALSGLLGPTLR.G	606.9	2
	isotig00200	99.15		VF5a-2	<i>Pseudonaja textilis</i>	6	R.GEVGDSLIIYFK.N	670.9	2
							K.WLISSLVAK.H	508.84	2
							R.LDDAVPPGQSFK.Y	637.36	2
							K.NFATQPVSIHPQSAVYNK.W	667.72	3
							R.EYELDFK.Q	472.26	2
							R.EYELDFKQEKPR.D	527.94	3
45	VF5a	isotig00199	99.19	VF5a-3	<i>Pseudonaja textilis</i>	11	K.WSEGSSYSDGTSVDER.L	881.33	2
	isotig00200	99.19		VF5a-2	<i>Pseudonaja textilis</i>	7	K.AQYLDNFSNFIGK.K	758.89	2
							R.GEVGDSLIIYFK.N	670.88	2
							R.DALSGLLGPTLR.G	606.89	2
							R.LDDAVPPGQSFK.Y	637.35	2
							K.WLISSLVAK.H	508.83	2
							R.DTVTIVFK.N	461.8	2
							R.GILGPVIK.A	398.83	2
							R.EYELDFK.Q	472.25	2
							K.AQYLDNFSNFIGK.K.Y	822.99	2
							R.EYELDFKQEKPR.D	527.9	3
	FA5V_PSETE	99.14		Coagulation factor V	<i>Pseudonaja textilis</i>	5	K.WSEGSSYSDGTSVDER.L	881.33	2
							K.AQYLDNFSNFIGK.K	758.89	2
							R.LDDAVPPGQSFK.Y	637.35	2
							K.WLISSLVAK.H	508.83	2
							R.EYELDFK.Q	472.25	2
							K.AQYLDNFSNFIGK.K.Y	822.99	2
							R.GILGPVIRAK.V	512.39	2
							R.EYELDFKQEKPR.D	527.9	3
46	VF5a	FA5V_PSETE	99.2	Venom prothrombin activator	pseutarin-C non-catalytic subunit	13	K.IGTWLLETEVGNER.Q	937.49	2
							K.SWAYYSGVNPKE.D	700.86	2
							R.GMQALFTVIDK.D	611.87	2
							K.TWNQYIALR.I	582.86	2
							R.SYDDKSPELFK.K	664.79	2
							R.GM(+15.99)QALFTVIDK.D	619.84	2
							K.VFTGNINSDGHVK.H	694.39	2
							K.NSEITASSYK.K	550.31	2
							K.DIHVVNFHGQTFTEGR.E	662.7	3
							K.EHEHPWQIDLQR.Q	567.66	3
							R.LNLEGGTNAWQPEVNNK.D	942.44	2
							K.NSEITASSYK.K.T	614.36	2
							K.ENHIDPPIAR.Y	637.86	2
							R.HSETQMHEFGNSDGTIVK.E	669.01	3
							K.SWYFPK.S	414.25	2
							R.EDNQLGVLPPLPGTFASIK.M	1006.58	2
							R.SYDDKSPELFKK.D	486.26	3
47	VF5a	isotig00200	99.19	VF5a-2	<i>Pseudonaja textilis</i>	22	R.QVVTGIQTQGTVQLK.H	913.53	2
							K.IGTWLLETEVGNER.Q	937.49	2
							R.SYLDLDTFTPTSGGEYK.H	1019.42	2
							K.ITSITQGATSMTTSMYVK.T	1017.04	2
							R.AEVDVIEIQFR.N	717.43	2
							K.SWAYYSGVNPKE.D	700.82	2
							K.ITSITQGATSM(+15.99)TTSMYVK.T	1025.01	2
							R.GMQALFTVIDK.D	611.87	2
							K.HLGILGPIIR.A	544.87	2
							K.NSEITASSYK.K	550.27	2
							R.LNLEGGTNAWQPEVNNK.D	942.45	2
							K.TWWSWEPFLAR.L	783.4	2
							K.VFTGNINSDGHVK.H	694.34	2
							R.EDYQLGVLPPLPGTFASIK.M	1031.05	2
							K.DIHVVNFHGQTFTEGR.E	662.69	3
							R.GM(+15.99)QALFTVIDK.D	619.78	2
							R.SYDDKSPELFK.K	664.78	2
							K.TWNQYIALR.I	582.81	2
							K.ENHIDPPIAR.Y	425.56	3
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.99	3
							K.NSEITASSYK.K.T	409.86	3
							K.AVEPGQVYTYK.W	627.82	2
							R.NLASRPYSLHAHGLLYEK.S	690.34	3
							R.SGPTDNTEK.C	474.69	2
							K.SPELFK.K	360.72	2
							K.EHEHPWQIDLQR.Q	567.65	3
							K.FYNRPTR.I	550.8	2
							R.HSETQM(+15.99)HFEGNSDGTIVK.E	674.28	3
							K.SWYFPK.S	414.19	2
							K.KEHEHPWQIDLQR.Q	458.02	4
							R.SYDDKSPELFKK.D	486.25	3
	FA5V_OXYMI	99.19	Venom prothrombin activator	omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	18	R.QVVTGIQTQGTVQLK.H	913.53	2
							K.IGTWLLETEVGNER.Q	937.49	2
							R.SYLDLDTFTPTSGGEYK.H	1019.42	2
							K.SWAYYSGVNPKE.D	700.82	2
							R.GMQALFTVIDK.D	611.87	2
							K.HLGILGPIIR.A	544.87	2
							K.NSEITASSYK.K	550.27	2
							R.LNLEGGTNAWQPEVNNK.D	942.45	2
							K.TWWSWEPFLAR.L	783.4	2
							K.VFTGNINSDGHVK.H	694.34	2

							K.DIHVVNFHGQTFTEGR.E	662.69	3
							R.GM(+15.99)QALFTVIDK.D	619.78	2
							R.EDNQLGVLPLPGTFASIK.M	1006.57	2
							K.TWNQYIALR.I	582.81	2
							K.ENHIDPPIAR.Y	425.56	3
							K.NSEITASSYK.T	409.86	3
							K.AVEPGQVYTYK.W	627.82	2
							R.NLASRPSYSLHAHGLLYEK.S	690.34	3
							R.SGPTDNTEK.C	474.69	2
							K.EHEHPWQIDLQR.Q	567.65	3
							R.HSETQM(+15.99)HFEGNSDGTTVK.E	674.28	3
							K.SWYFPK.S	414.19	2
							K.KEHEHPWQIDLQR.Q	458.02	4
FA5V_PSETE	99.19	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	19			K.IGTWLLETEVGENQER.G	937.49	2
							R.SYLDLDTFQTPSTGGYEYK.H	1019.42	2
							K.ITSITQGATSMTTSMYVK.T	1017.04	2
							K.SWAYYSGVNPKEK.D	700.82	2
							K.ITSITQGATSM(+15.99)TTSMYVK.T	1025.01	2
							R.GMQALFTVIDK.D	611.87	2
							K.HLGILGPIIR.A	544.87	2
							K.NSEITASSYK.K	550.27	2
							R.LNLEGGTNAWQPEVNNK.D	942.45	2
							K.VFTGNINSDGHVK.H	694.34	2
							K.DIHVVNFHGQTFTEGR.E	662.69	3
							R.GM(+15.99)QALFTVIDK.D	619.78	2
							R.SYDDKSPELFK.K	664.78	2
							R.EDNQLGVLPLPGTFASIK.M	1006.57	2
							K.TWNQYIALR.I	582.81	2
							K.ENHIDPPIAR.Y	425.56	3
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.99	3
							K.NSEITASSYK.T	409.86	3
							K.AVEPGQVYTYK.W	627.82	2
							K.NLASRPSYSLHAHGLLYEK.S	690.34	3
							R.SGPTDNTEK.C	474.69	2
							K.SPELFK.K	360.72	2
							K.EHEHPWQIDLQR.Q	567.65	3
							K.FYNRPTR.F	550.8	2
							R.HSETQM(+15.99)HFEGNSDGTTVK.E	674.28	3
							K.SWYFPK.S	414.19	2
							K.KEHEHPWQIDLQR.Q	458.02	4
							R.SYDDKSPELFK.K	486.25	3
FA5V_OXYSU	99.19	Venom prothrombin activator Ocutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	15			R.QVITGIQTQGTVQLLK.H	913.53	2
							K.IGTWLLETEVGENQER.G	937.49	2
							R.SYLDLDTFQTPSTGGYEYK.H	1019.42	2
							K.ITSITQGATSMTTSMYVK.T	1017.04	2
							K.SWAYYSGVNPKEK.D	700.82	2
							K.ITSITQGATSM(+15.99)TTSMYVK.T	1025.01	2
							R.GMQALFTVIDK.D	611.87	2
							K.HLGILGPIIR.A	544.87	2
							K.TWWSSWEPLAR.L	783.4	2
							K.VFTGNINSDGHVK.H	694.34	2
							R.GM(+15.99)QALFTVIDK.D	619.78	2
							R.EDNQLGVLPLPGTFASIK.M	1006.57	2
							K.TWNQYIALR.I	582.81	2
							K.ENHIDPPIAR.Y	425.56	3
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.99	3
							K.AVEPGQVYTYK.W	627.82	2
							R.NLASRPSYSLHAHGLLYEK.S	690.34	3
							R.SGPTDNTEK.C	474.69	2
							K.EHEHPWQIDLQR.Q	567.65	3
							K.SWYFPK.S	414.19	2
							K.KEHEHPWQIDLQR.Q	458.02	4
48	VF5a	isotig00200	99.21	VF5a-2	<i>Pseudonaja textilis</i>	20	K.IGTWLLETEVGENQER.G	937.49	2
							R.SYLDLDTFQTPSTGGYEYK.H	1019.44	2
							R.AEVDVIEIQFR.N	717.4	2
							K.ITSITQGATSM(+15.99)TTSMYVK.T	1025.01	2
							K.SWAYYSGVNPKEK.D	700.85	2
							R.GMQALFTVIDK.D	611.84	2
							R.GM(+15.99)QALFTVIDK.D	619.84	2
							K.AVEPGQVYTYK.W	627.83	2
							R.SYDDKSPELFK.K	664.78	2
							K.ITSITQGATSMTTSM(+15.99)YVK.T	1024.99	2
							K.VFTGNINSDGHVK.H	694.37	2
							R.DALSGLLGPTLR.G	606.89	2
							K.WTVLDTDEPTVK.D	702.38	2
							K.DIHVVNFHGQTFTEGR.E	662.69	3
							K.HLGILGPIIR.A	544.84	2
							K.TWNQYIALR.I	582.87	2
							K.NSEITASSYK.K	550.3	2
							K.HFFKPPILSR.F	414.59	3
							R.NPDDIAGR.Y	429.25	2
							K.AQYLDNFSNFIGK.K	758.84	2
							K.ENHIDPPIAR.Y	425.59	3
							K.NFATQPVSIHPQSAVYNK.W	667.71	3
							K.SWYFPK.S	414.22	2
							K.NSEITASSYK.T	614.34	2
							R.QVITGIQTQGTVQLLK.H	609.44	3
							R.GILGPVIK.A	398.79	2
							R.TIDIR.E	309.22	2
49	VF5a	isotig00199	98.77	VF5a-3	<i>Pseudonaja textilis</i>	4	K.AQYLDNFSNFIGK.K	758.9	2
		FA5V_OXYMI	98.77	Venom prothrombin activator omicaric-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	2	K.AVEPGQVYTYK.W	627.86	2
		FA5V_PSETE	98.77	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	2	K.WTVLDTDEPTVK.D	702.41	2
		isotig00200	98.77	VF5a-2	<i>Pseudonaja textilis</i>	2			
50	VF5a	isotig00199	98.07	VF5a-3	<i>Pseudonaja textilis</i>	4	K.AQYLDNFSNFIGK.K	758.92	2
		FA5V_OXYSU	98.07	Venom prothrombin activator Ocutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	2	K.AVEPGQVYTYK.W	627.87	2
		FA5V_OXYMI	98.07	Venom prothrombin activator omicaric-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	2	K.WTVLDTDEPTVK.D	702.39	2
		FA5V_PSETE	98.07	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	2			
		FA5_PSETE	98.07	Coagulation factor V	<i>Pseudonaja textilis</i>	2			

		isotig00200	98.07	VF5a-2	<i>Pseudonaja textilis</i>	2		
51	VF5a	isotig00199	99.01	VF5a-3	<i>Pseudonaja textilis</i>	6	K.AQYLDNFSNFIGK.K	758.92
		FA5V_OXYMI	99.01	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	4	K.SNVM(+15.99)YTLNGYASDR.T	803.89
		FA5V_PSETE	99.01	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	4	K.WTVLDTDEPTVK.D	702.4
		isotig00200	99.01	VF5a-2	<i>Pseudonaja textilis</i>	4	K.AVEPGQVYTK.W	627.89
52	VF5a	FA5V_OXYSU	99.01	Venom prothrombin activator Oscutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	4	R.DTVTIVFK.N	461.78
		isotig00199	98.82	VF5a-3	<i>Pseudonaja textilis</i>	6	K.AQYLDNFSNFIGK.K	758.94
		FA5V_PSETE	98.82	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	4	K.WTVLDTDEPTVK.D	702.38
		isotig00200	98.82	VF5a-2	<i>Pseudonaja textilis</i>	4	K.AVEPGQVYTK.W	627.86
53	VF5a	FA5V_OXYMI	98.82	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	4	K.KKEEVPNVFVDPDESALAK.E	553.83
		FA5V_OXYMI	99.12	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	5	K.AQYLDNFSNFIGK.K	758.92
		FA5V_PSETE	99.12	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	5	K.WTVLDTDEPTVK.D	702.39
							K.AVEPGQVYTK.W	627.85
58	VF5a						K.SNVM(+15.99)YTLNGYASDR.T	803.88
							K.ADVEQHAVFAVDENK.S	607.02
							R.DTVTIVFK.N	461.78
59	VF5a	isotig00199	91.38	VF5a-3	<i>Pseudonaja textilis</i>	5	K.SNVMYTLNGYASDR.T	795.73
		FA5V_PSETE	91.38	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	3	K.SWYLEDNIKK.Y	648.22
		isotig00200	91.38	VF5a-2	<i>Pseudonaja textilis</i>	3	K.KKEEVPNVFVDPDESALAK.E	737.92
		FA5V_OXYMI	91.38	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	3		
72	VF5a	HKS	50.62	Helokinestatin-1	<i>Heloderma h. horridum</i>	100	GPYPQLVPR	562.17
		HKS_HELSS	50.62	Helokinestatin-1	<i>Heloderma s. suspectum</i>	100		
		EXE2	45.45	Exendin-2-long	<i>Heloderma suspectum</i>	11	K.YLASILGSR.T	490.1
		EXE2_HELSC	45.45	Exendin-2-long	<i>Heloderma s. cinctum</i>	11		
59	VF5a	HKS	40.17	Helokinestatin-1	<i>Heloderma h. horridum</i>	100	GPYPQLVPR	562.17
		HKS_HELSS	40.17	Helokinestatin-1	<i>Heloderma s. suspectum</i>	100		
		CRISP	34.62	Cysteine-rich venom protein helothermine	<i>Heloderma h. horridum</i>	5	R.RIVEPTASNMLK.M	679.75
		isotig00199	99.13	VF5a-3	<i>Pseudonaja textilis</i>	14	K.NSEITASSYK.K	550.22
72	VF5a	FA5V_PSETE	99.12	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	5	K.HFFKPILSR.F	414.51
		isotig00200	99.12	VF5a-2	<i>Pseudonaja textilis</i>	5	K.TWNQYIALR.I	582.79
							K.ENHIDPPIAR.Y	425.51
							K.ITSITQGATSMTTSMYVK.T	678.31
74	VF5a						K.VFTGNINSDGHVK.H	463.17
							K.ITSITQGATSMTTSM(+15.99)YVK.T	683.65
		FA5V_OXYMI	99.09	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	4	K.NSEITASSYK.K	550.22
							K.HFFKPILSR.F	414.51
75	VF5a						K.TWNQYIALR.I	582.79
							K.ENHIDPPIAR.Y	425.51
							K.ITSITQGATSMTTSMYVK.T	678.31
							K.DSEITASSYK.K	550.66
76	VF5a						K.VFTGNINSDGHVK.H	463.17
							K.ITSITQGATSMTTSM(+15.99)YVK.T	683.65
		EXE2	61.04	Exendin-2-long	<i>Heloderma s. cinctum</i>	11	K.YLASILGSR.T	490.19
		EXE2_HELSC	61.04	Exendin-2-long	<i>Heloderma suspectum</i>	11		
74	VF5a	EXE2	55.82	Exendin-2-long	<i>Heloderma s. cinctum</i>	11	K.YLASILGSR.T	490.22
		EXE2_HELSC	55.82	Exendin-2-long	<i>Heloderma suspectum</i>	11		
		CRISP	55.16	CRISP isoform 1	<i>Suta nigriceps</i>	3	R.NMLQMK.W	382.62
		R4G2J2_DENDV	54.51	CRISP-Den-3	<i>Denisonia devisi</i>	3		
75	VF5a	isotig00418	54.51	Cysteine-rich venom protein pseudochetoxin-like	<i>Pseudonaja textilis</i>	3		
		R4G2J3_9SAUR	54.49	CRISP-Pse-4	<i>Pseudonaja modesta</i>	3		
		R4FIS5_9SAUR	54.49	CRISP-Pse-17	<i>Pseudonaja modesta</i>	3		
		CRVP1_HYDHA	54.49	Cysteine-rich venom protein 1	<i>Hydrophis hardwickii</i>	3		
76	VF5a	CRVP2_HYDHA	54.49	Cysteine-rich venom protein 2	<i>Hydrophis hardwickii</i>	3		
		CRVP_OXYMI	54.49	Cysteine-rich venom protein pseudochetoxin-like	<i>Oxyuranus microlepidotus</i>	3		
		CRVP_PSETE	54.49	Cysteine-rich venom protein pseudochetoxin-like	<i>Pseudonaja textilis</i>	3		
		CRVP_OXYSC	54.49	Cysteine-rich venom protein pseudochetoxin-like	<i>Oxyuranus s. scutellatus</i>	3		
75	VF5a	isotig00418	79.47	Cysteine-rich venom protein pseudochetoxin-like	<i>Pseudonaja textilis</i>	3	R.NMLQMK.W	382.61
		R4G2J3_9SAUR	79.46	CRISP-Pse-4	<i>Pseudonaja modesta</i>	3		
		R4FIS5_9SAUR	79.46	CRISP-Pse-17	<i>Pseudonaja modesta</i>	3		
		CRVP_OXYMI	79.46	Cysteine-rich venom protein pseudochetoxin-like	<i>Oxyuranus microlepidotus</i>	3		
76	VF5a	CRVP_PSETE	79.46	Cysteine-rich venom protein pseudochetoxin-like	<i>Pseudonaja textilis</i>	3		
		CRVP_OXYSC	79.46	Cysteine-rich venom protein pseudochetoxin-like	<i>Oxyuranus s. scutellatus</i>	3		
		R4G7K2_9SAUR	79.46	CRISP-Pse-11	<i>Pseudonaja modesta</i>	3		
77	VF5a	FA5_PSETE	99.14	Coagulation factor V	<i>Pseudonaja textilis</i>	4	K.HLGILGPIIR.A	544.83
							K.VFTGNINSDGHVK.H	463.18
							K.NSEITASSYK.K	550.21
							R.GM(+15.99)QALFTVIDK.D	619.82
78	VF5a						K.NSEITASSYK.T	614.27
							K.ENHIDPPIAR.Y	425.52
		V8P243_OPHHA	97.84	Coagulation factor V	<i>Ophiophagus hannah</i>	1	K.HLGILGPIIR.A	544.83
							R.GM(+15.99)QALFTVIDK.D	619.82
77	VF5a	isotig00199	99.13	VF5a-3	<i>Pseudonaja textilis</i>	9	K.AVEPGQVYTK.W	627.79
		FA5V_PSETE	99.13	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	6	R.GEVGDSLIYFK.N	670.79
		isotig00200	99.13	VF5a-2	<i>Pseudonaja textilis</i>	6	K.WTVLDTDEPTVK.D	702.32
							R.DALSGLLGPTLR.G	606.84
78	VF5a						K.NFATQPVSIIHQSAVYVK.W	667.65
							K.VRDVTIVFK.N	589.31
							K.LYHSAVDM(+15.99)JTR.D	403.47
78	VF5a	isotig00200	99.17	VF5a-2	<i>Pseudonaja textilis</i>	9	R.AEVDVIEIQR.N	717.34
							R.GM(+15.99)QALFTVIDK.D	619.81
							K.HLGILGPIIR.A	544.83
							K.AVEPGQVYTK.W	627.79
79	VF5a						R.DALSGLLGPTLR.G	606.81
							R.NPDDIAGR.Y	429.12
							K.HFFKPILSR.F	414.51
							K.NSEITASSYK.K	550.2
80	VF5a						K.ENHIDPPIAR.Y	425.51
							K.NSEITASSYK.T	614.27
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.98
							K.DIHVVNFHGQFTTEGR.E	662.61
81	VF5a	isotig00199	99.09	VF5a-3	<i>Pseudonaja textilis</i>	6	K.IEESLESR.K	546.21
							K.AVEPGQVYTK.W	627.79
							R.DALSGLLGPTLR.G	606.81
							R.NPDDIAGR.Y	429.12
82	VF5a						K.DKYIYSEENIK.K	701.29

ACCEPTED MANUSCRIPT

						K.YIYSEENIK.K	579.77	2	
79	VF5a	isotig00200	99.17	VF5a-2	<i>Pseudonaja textilis</i>	11	R.GEVGDSLIYFK.N	670.84	2
						K.AVEPGQVYTYK.W	627.8	2	
						R.AEVDDVIEIQFR.N	717.36	2	
						K.AQYLDNFSNFIGK.K	758.83	2	
						K.WTVLDTDEPTVK.D	702.34	2	
						R.DALSGLLGPTLR.G	606.86	2	
						K.WSEGSSYSDGTS DVER.L	881.33	2	
						R.EYVLMFSV FDESK.N	797.35	2	
						K.TWNQYIALR.I	582.79	2	
						K.IGTWLLETEVGENQER.G	625.3	3	
						K.NSEITASSYK.K	550.22	2	
						K.HLGILGPIIR.A	544.84	2	
						K.SWYLEDNIK.K	584.28	2	
						K.SWYLEDNIKK.Y	648.28	2	
						R.GILGPVIA.K	398.72	2	
	EXE2	EXE2_HELSC	59.69	Exendin-2-long	<i>Heloderma s. cinctum</i>	11	K.YLASILGSR.T	490.23	2
		EXE2_HELSE	59.69	Exendin-2-long	<i>Heloderma suspectum</i>	11			
	HKS	HKS_HELHO	59.84	Helokinestatin-1	<i>Heloderma h. horridum</i>	100	GPPYQPLVPR	562.31	2
		HKS_HELSS	59.84	Helokinestatin-1	<i>Heloderma s. suspectum</i>	100			
80	VF5a	isotig00200	99.16	VF5a-2	<i>Pseudonaja textilis</i>	15	R.GMQALFTVIDK.D	611.82	2
						K.SWAYYSGVNPEK.D	700.81	2	
						R.GM(+15.99)QALFTVIDK.D	619.83	2	
						R.AEVDDVIEIQFR.N	717.35	2	
						K.HLGILGPIIR.A	544.83	2	
						K.ENHIDPPIAR.Y	425.53	3	
						K.IGTWLLETEVGENQER.G	625.32	3	
						K.TWNQYIALR.I	582.8	2	
						K.NSEITASSYK.K	550.23	2	
						R.NPDDIAGR.Y	429.13	2	
						K.NSEITASSYK.T	614.28	2	
						K.VFTGNINSDGHVK.H	463.18	3	
						R.SYLDDTFTPTSTGGEYEK.H	679.94	3	
						K.SPELFK.K	360.63	2	
						R.DALSGLLGPTLR.G	606.87	2	
						K.DIHVVNFHGQTFTEEGR.E	662.65	3	
						K.AVEPGQVYTYK.W	627.82	2	
						R.SYDDKSPELFK.K	443.42	3	
						R.HSETQMHFEGNSDGTTVK.E	668.94	3	
						R.SYDDKSPELFKK.D	486.18	3	
						K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	689	3	
	FA5V_OXYSU	99.16	Venom prothrombin activator Ocutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	10	R.GMQALFTVIDK.D	611.82	2	
						K.SWAYYSGVNPEK.D	700.81	2	
						R.GM(+15.99)QALFTVIDK.D	619.83	2	
						K.HLGILGPIIR.A	544.83	2	
						K.ENHIDPPIAR.Y	425.53	3	
						K.IGTWLLETEVGENQER.G	625.32	3	
						K.TWNQYIALR.I	582.8	2	
						K.VFTGNINSDGHVK.H	463.18	3	
						R.SYLDDTFTPTSTGGEYEK.H	679.94	3	
						K.DSEITASSYK.K	550.71	2	
						K.DSEITASSYK.T	614.75	2	
						K.AVEPGQVYTYK.W	627.82	2	
						K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	689	3	
						K.SWAYYSGVNPEK.D	700.8	2	
						R.GM(+15.99)QALFTVIDK.D	619.82	2	
						R.AEVDDVIEIQFR.N	717.35	2	
						R.NPDDIAGR.Y	429.15	2	
						K.ENHIDPPIAR.Y	425.53	3	
						K.TWNQYIALR.I	582.8	2	
						K.HLGILGPIIR.A	544.82	2	
						K.NSEITASSYK.K	550.23	2	
						K.HFFKPPIILSR.F	414.53	3	
						K.IGTWLLETEVGENQER.G	625.31	3	
						R.HSETQMHFEGNSDGTTVK.E	668.95	3	
						K.NSEITASSYK.T	614.3	2	
						K.FYNRPTFR.I	367.47	3	
						R.SYDDKSPELFKK.D	486.2	3	
						K.VFTGNINSDGHVK.H	463.2	3	
	FA5V_OXYSU	99.16	Venom prothrombin activator Ocutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	6	K.SWAYYSGVNPEK.D	700.8	2	
						R.GM(+15.99)QALFTVIDK.D	619.82	2	
						K.ENHIDPPIAR.Y	425.53	3	
						K.TWNQYIALR.I	582.8	2	
						K.HLGILGPIIR.A	544.82	2	
						K.IGTWLLETEVGENQER.G	625.31	3	
						K.VFTGNINSDGHVK.H	463.2	3	
	EXE2	EXE2_HELSC	60.48	Exendin-2-long	<i>Heloderma s. cinctum</i>	11	K.YLASILGSR.T	490.25	2
		EXE2_HELSE	60.48	Exendin-2-long	<i>Heloderma suspectum</i>	11			
82	VF5a	isotig00200	99.18	VF5a-2	<i>Pseudonaja textilis</i>	13	R.GM(+15.99)QALFTVIDK.D	619.82	2
						R.AEVDDVIEIQFR.N	717.35	2	
						K.SWAYYSGVNPEK.D	700.81	2	
						K.ENHIDPPIAR.Y	425.52	3	
						K.HFFKPPIILSR.F	414.52	3	
						K.HLGILGPIIR.A	544.83	2	
						K.IGTWLLETEVGENQER.G	625.3	3	
						K.AVEPGQVYTYK.W	627.82	2	
						K.TWNQYIALR.I	582.79	2	
						R.SYLDDTFTPTSTGGEYEK.H	679.95	3	
						K.NSEITASSYK.K	550.23	2	
						K.VFTGNINSDGHVK.H	694.26	2	
						R.GMQALFTVIDK.D	611.78	2	
						K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.99	3	
						K.FYNRPTFR.I	367.45	3	
						R.SYDDKSPELFKK.D	486.2	3	
						R.TIDIR.E	309.15	2	

83	VF5a	isotig00200	99.17	VF5a-2	<i>Pseudonaja textilis</i>	15	K.SWAYYSGVNPKEK.D	700.8	2
							R.GM(+15.99)QALFTVIDK.D	619.83	2
							R.AEVDVIEIQFR.N	717.34	2
							R.GMQALFTVIDK.D	611.81	2
							K.IGTWLLETEVGENQER.G	625.29	3
							R.NPDDIAGR.Y	429.14	2
							K.ENHIDPPIAR.Y	425.52	3
							K.DIHVVNFHGQTFTEEGR.E	662.62	3
							K.HLGILGPIIR.A	544.82	2
							K.HFFKPPILSR.F	414.53	3
							K.TWNQYIALR.I	582.81	2
							R.DALSGLLGPTLR.G	606.85	2
							K.AVEPGQVYTYK.W	627.81	2
							K.NSEITASSYK.K	550.23	2
							K.VFTGNINSDGHVK.H	463.18	3
							R.SYLDDTFQTPSTGGEYK.H	679.94	3
							K.NSEITASSYK.T	614.28	2
							R.HSETQMHFEGNSDGTTVK.E	668.94	3
							K.SWYFPK.S	414.16	2
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.98	3
FA5V_OXYMI	99.17	Venom prothrombin activator omicarin-C non-catalytic subunit	<i>Oxyuranus microlepidotus</i>	12	K.SWAYYSGVNPKEK.D	700.8	2		
							R.GM(+15.99)QALFTVIDK.D	619.83	2
							R.GMQALFTVIDK.D	611.81	2
							K.IGTWLLETEVGENQER.G	625.29	3
							K.ENHIDPPIAR.Y	425.52	3
							K.DIHVVNFHGQTFTEEGR.E	662.62	3
							K.HLGILGPIIR.A	544.82	2
							K.HFFKPPILSR.F	414.53	3
							K.TWNQYIALR.I	582.81	2
							K.AVEPGQVYTYK.W	627.81	2
							K.NSEITASSYK.K	550.23	2
							K.VFTGNINSDGHVK.H	463.18	3
							R.SYLDDTFQTPSTGGEYK.H	679.94	3
							K.NSEITASSYK.T	614.28	2
							R.HSETQMHFEGNSDGTTVK.E	668.94	3
							K.SWYFPK.S	414.16	2
84	VF5a	isotig00200	99.15	VF5a-2	<i>Pseudonaja textilis</i>	18	R.GM(+15.99)QALFTVIDK.D	619.82	2
							R.AEVDVIEIQFR.N	717.34	2
							R.GMQALFTVIDK.D	611.82	2
							K.SWAYYSGVNPKEK.D	700.79	2
							R.SYLDDTFQTPSTGGEYK.H	1019.4	2
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	1032.98	2
							K.ITSITQGATSMTTSM(+15.99)YVK.T	683.64	3
							K.IGTWLLETEVGENQER.G	625.31	3
							K.DIHVVNFHGQTFTEEGR.E	662.66	3
							R.SYDDKSPELFK.K	664.8	2
							K.ENHIDPPIAR.Y	425.5	3
							K.ITSITQGATSM(+15.99)TTSMYVK.T	1025	2
							K.HLGILGPIIR.A	544.87	2
							K.TWNQYIALR.I	582.8	2
							K.NSEITASSYK.K	550.21	2
							K.NSEITASSYK.T	614.27	2
							K.EHEHPWQIDLQR.Q	567.6	3
							R.HSETQMHFEGNSDGTTVK.E	668.93	3
							K.SPELFK.K	360.64	2
							K.SWYFPK.S	414.15	2
							R.SGPTDNTEK.C	474.66	2
							R.TIDIR.E	309.14	2
							K.VFTGNINSDGHVK.H	694.24	2
							K.HFFKPPILSR.F	414.48	3
							R.SYDDKSPELFK.K	486.18	3
							K.FYNRPTRF.I	367.44	3
							R.NLASRPSYSLHAHGLLYEK.S	690.34	3
FA5V_PSETE	99.15	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	19	R.GM(+15.99)QALFTVIDK.D	619.82	2		
							R.GMQALFTVIDK.D	611.82	2
							K.SWAYYSGVNPKEK.D	700.79	2
							R.SYLDDTFQTPSTGGEYK.H	1019.4	2
							K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	1032.98	2
							K.ITSITQGATSMTTSM(+15.99)YVK.T	683.64	3
							K.IGTWLLETEVGENQER.G	625.31	3
							K.DIHVVNFHGQTFTEEGR.E	662.66	3
							R.SYDDKSPELFK.K	664.8	2
							K.ENHIDPPIAR.Y	425.5	3
							K.TWWSSWEPSLAR.L	753.33	2
							K.ITSITQGATSM(+15.99)TTSMYVK.T	1025	2
							K.HLGILGPIIR.A	544.87	2
							K.TWNQYIALR.I	582.8	2
							K.NSEITASSYK.K	550.21	2
							K.NSEITASSYK.T	614.27	2
							K.EHEHPWQIDLQR.Q	567.6	3
							R.HSETQMHFEGNSDGTTVK.E	668.93	3
							K.SPELFK.K	360.64	2
							R.EDNQLGVLPPLPGTFASIK.M	671.37	3
							K.SWYFPK.S	414.15	2
							R.SGPTDNTEK.C	474.66	2
							R.TIDIR.E	309.14	2
							K.VFTGNINSDGHVK.H	694.24	2
							K.HFFKPPILSR.F	414.48	3
							R.SYDDKSPELFK.K	486.18	3
							K.FYNRPTRF.I	367.44	3
							K.NLASRPSYSLHAHGLLYEK.S	690.34	3
FA5V_OXYSU	99.15	Venom prothrombin activator Oscutarin-C non-catalytic subunit	<i>Oxyuranus scutellatus</i>	14	R.GM(+15.99)QALFTVIDK.D	619.82	2		
							R.GMQALFTVIDK.D	611.82	2
							K.SWAYYSGVNPKEK.D	700.79	2

						R.SYLDDTFQTPSTGGEYK.H	1019.4	2	
						K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	1032.98	2	
						K.ITSITQGATSMTTSM(+15.99)YVK.T	683.64	3	
						K.IGTWLLETEVGENQER.G	625.31	3	
						K.ENHIDPPIAR.Y	425.5	3	
						K.ITSITQGATSM(+15.99)TTSMYVK.T	1025	2	
						K.HLGILGPIIR.A	544.87	2	
						K.TWNQYIALR.I	582.8	2	
						K.DSEITASSYK.K	550.7	2	
						K.EHEHPWQIDLQR.Q	567.6	3	
						R.EDNQLGVLPPLPGTFASIK.M	671.37	3	
						K.DSEITASSYKK.T	614.78	2	
						K.SWYFPK.S	414.15	2	
						R.SGPTDNTEK.C	474.66	2	
						R.TIDIR.E	309.14	2	
						K.VFTGNINSDGHVK.H	694.24	2	
						R.NLASRPYSLHAHGLLYEK.S	690.34	3	
85	VF5a	isotig00200	99.16	VF5a-2	Pseudonaja textilis	15	K.AQYLDNFSNFIGK.K	758.85	2
						R.GEVGDSLIIFYK.N	670.83	2	
						K.AVEPGQVYTYK.W	627.81	2	
						R.DALSGLLGPTLR.G	606.84	2	
						R.AEVDVIEIQFR.N	717.34	2	
						K.WTVLDTDEPTVK.D	702.35	2	
						K.NSEITASSYK.K	550.23	2	
						K.HLGILGPIIR.A	544.81	2	
						K.HLQAGM(+15.99)YGYLNIK.D	508.55	3	
						K.ENHIDPPIAR.Y	425.53	3	
						R.EYELDFK.Q	472.17	2	
						K.NFATQPVSIHPQSAVYNK.W	667.67	3	
						K.HFFKPILLSR.F	414.54	3	
						K.LYHSAVDM(+15.99)TR.D	403.46	3	
						K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.99	3	
						R.GILGPVIK.A	398.72	2	
						K.DIHVVNFHGQTFTEEGR.E	662.65	3	
						K.TWNQYIALR.I	582.8	2	
						K.ITSITQGATSMTTSMYVK.T	678.32	3	
						K.NSEITASSYKK.T	614.27	2	
	FA5_PSETE	99.16	Coagulation factor V	Pseudonaja textilis	12	K.AQYLDNFSNFIGK.K	758.85	2	
						K.AVEPGQVYTYK.W	627.81	2	
						K.WTVLDTDEPTVK.D	702.35	2	
						K.NSEITASSYK.K	550.23	2	
						K.HLGILGPIIR.A	544.81	2	
						K.HLQAGM(+15.99)YGYLNIK.D	508.55	3	
						K.ENHIDPPIAR.Y	425.53	3	
						R.EYELDFK.Q	472.17	2	
						K.NFATQPVSIHPQSAVYNK.W	667.67	3	
						K.HFFKPILLSR.F	414.54	3	
						K.LYHSAVDM(+15.99)TR.D	403.46	3	
						K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	688.99	3	
						K.DIHVVNFHGQTFTEEGR.E	662.65	3	
						K.TWNQYIALR.I	582.8	2	
						K.ITSITQGATSMTTSMYVK.T	678.32	3	
						K.NSEITASSYKK.T	614.27	2	
86	VF5a	isotig00199	99.17	VF5a-3	Pseudonaja textilis	12	R.GEVGDSLIIFYK.N	670.93	2
						K.AQYLDNFSNFIGK.K	758.93	2	
						K.AVEPGQVYTYK.W	627.87	2	
						K.WTVLDTDEPTVK.D	702.42	2	
						R.DALSGLLGPTLR.G	606.92	2	
						K.WSEGSSYSDGTS DVER.L	881.41	2	
						R.EYVLMFSVDESK.N	797.48	2	
						K.NFATQPVSIHPQSAVYNK.W	667.76	3	
	EXE4	EXE4_HEL SU	61.05	Exendin-4	Heloderma suspectum	14	R.HGEGTFTSDLSK.Q	639.85	2
		EXE4_HEL SC	61.05	Exendin-4	Heloderma s. cinctum	14			
	EXE2	EXE2_HEL SC	60.99	Exendin-2-long	Heloderma s. cinctum	11	K.YLASILGSR.T	490.33	2
		EXE2_HEL SU	60.99	Exendin-2-long	Heloderma suspectum	11			
87	VF5a	isotig00199	99.17	VF5a-3	Pseudonaja textilis	11	K.AQYLDNFSNFIGK.K	758.92	2
		FA5V_PSETE	99.17	Venom prothrombin activator pseudarin-C non-catalytic subunit	Pseudonaja textilis	7	R.GEVGDSLIIFYK.N	670.91	2
		isotig00200	99.17	VF5a-2	Pseudonaja textilis	7	R.DALSGLLGPTLR.G	606.95	2
						K.AVEPGQVYTYK.W	627.89	2	
						K.WTVLDTDEPTVK.D	702.42	2	
						K.WSEGSSYSDGTS DVER.L	881.4	2	
						K.NLASRPYSIYVHGVSVSK.D	659.76	3	
						R.EYELDFK.Q	472.26	2	
89	VF5a	isotig00199	55.02	VF5a-3	Pseudonaja textilis	2	R.SGPTDNTEK.C	474.73	2
		FA5V_OXYSU	54.91	Venom prothrombin activator Oscutarin-C non-catalytic subunit	Oxyuranus scutellatus	1			
		FA5_PSETE	54.91	Coagulation factor V	Pseudonaja textilis	1			
		FA5V_OXYMI	54.91	Venom prothrombin activator omicarin-C non-catalytic subunit	Oxyuranus microlepidotus	1			
		FA5V_PSETE	54.91	Venom prothrombin activator pseudarin-C non-catalytic subunit	Pseudonaja textilis	1			
		isotig00200	54.91	VF5a-2	Pseudonaja textilis	1			
		V8P243_OPHHA	54.85	Coagulation factor V	Ophiophagus hannah	0			
90	VF5a	FA5V_PSETE	99.2	Venom prothrombin activator pseudarin-C non-catalytic subunit	Pseudonaja textilis	8	K.IGTWLLETEVGENQER.G	937.52	2
						R.GM(+15.99)QALFTVIDK.D	619.92	2	
						K.ITSITQGATSM(+15.99)TTSM(+15.99)YVK.T	1033.1	2	
						K.VFTGNINSDGHVK.H	694.37	2	
						K.TWWSWVPSLAR.L	753.42	2	
						K.TWNQYIALR.I	582.9	2	
						K.NSEITASSYKK.T	614.38	2	
						K.HFFKPILLSR.F	414.62	3	
						K.NSEITASSYK.K	550.31	2	
						K.DIHVVNFHGQTFTEEGR.E	662.74	3	
						R.GMQALFTVIDK.D	611.93	2	
PLA2	PA2B2_NAJMO	61.66	Basic phospholipase A2 CM-II	Naja mossambica	9	R.YIDANYINLK.E	670.93	2	
EXE2	EXE2_HELSC	61.65	Exendin-2-long	Heloderma s. cinctum	11	K.YLASILGSR.T	490.34	2	

		EXE2_HEL SU	61.65	Exendin-2-long	Heloderma suspectum	11			
91	VF5a	isotig00200	99.13	VF5a-2	Pseudonaja textilis	3	R.AEVDDVIEIQFR.N K.HLGLGPIIR.A R.SYLD DTFQTPTSGGEYK.H	717.46 544.93 1019.51	2 2 2
	PLA2	PA2B2_NAJMO	61	Basic phospholipase A2 CM-II	Naja mossambica	9	R.YIDANYINLK.E	670.92	2
92	VF5a	isotig00199	99.12	VF5a-3	Pseudonaja textilis	8	R.GEVGDSLIIYFK.N	670.95	2
		isotig00200	99.12	VF5a-2	Pseudonaja textilis	5	R.DALSGLLGPTLR.G K.HLQAGM(+15.99)YGYLNIK.D R.EYELDFK.Q R.LDDAVPPGQSFK.Y R.EYVLM(+15.99)FSVFDESK.N	606.89 762.47 472.25 637.39 805.44	2 2 2 2 2
	EXE2	EXE2_HELSC	61.5	Exendin-2-long	Heloderma s. cinctum	11	K.YLASILGSR.T	490.33	2
		EXE2_HEL SU	61.5	Exendin-2-long	Heloderma suspectum	11			
93	VF5a	isotig00200	99.16	VF5a-2	Pseudonaja textilis	6	R.SYLD DTFQTPTSGGEYK.H K.HLGLGPIIR.A R.AEVDDVIEIQFR.N R.GEVGDSLIIYFK.N R.DALSGLLGPTLR.G R.NLASRPYSLHAHGLLYEK.S	1019.51 544.92 717.46 670.94 606.91 690.43	2 2 2 2 2 3
	EXE2	EXE2_HELSC	61.22	Exendin-2-long	Heloderma s. cinctum	11	K.YLASILGSR.T	490.34	2
		EXE2_HEL SU	61.22	Exendin-2-long	Heloderma suspectum	11			
94	VF5a	FASV_PSETE	99.16	Venom prothrombin activator pseutarin-C non-catalytic subunit	Pseudonaja textilis	5	R.GEVGDSLIIYFK.N	670.91	2
		isotig00200	99.16	VF5a-2	Pseudonaja textilis	5	R.DALSGLLGPTLR.G K.DIHVVNFHGQTFTTEGR.E K.AQYLDNFSNFIGK.K K.HFFKPPILSR.F K.VFTGNINSDGHVK.H	606.93 662.72 758.92 414.61 463.25	2 3 2 3 3
	EXE2	EXE2_HELSC	61.42	Exendin-2-long	Heloderma s. cinctum	11	K.YLASILGSR.T	490.32	2
		EXE2_HEL SU	61.42	Exendin-2-long	Heloderma suspectum	11			
95	VF5a	isotig00199	99.16	VF5a-3	Pseudonaja textilis	12	R.GM(+15.99)QALFTVIDK.D	619.91	2
		FASV_PSETE	99.16	Venom prothrombin activator pseutarin-C non-catalytic subunit	Pseudonaja textilis	4	K.DIHVVNFHGQTFTTEGR.E	662.72	3
		isotig00200	99.16	VF5a-2	Pseudonaja textilis	4	K.NSEITASSYK.T K.ENHIDPPIAR.Y K.HFFKPPILSR.F	614.36 425.6 414.61	2 2 3
96	VF5a	isotig00199	99.11	VF5a-3	Pseudonaja textilis	10	R.GM(+15.99)QALFTVIDK.D	619.89	2
		FASV_PSETE	99.1	Venom prothrombin activator pseutarin-C non-catalytic subunit	Pseudonaja textilis	3	K.ENHIDPPIAR.Y	425.61	3
		isotig00200	99.1	VF5a-2	Pseudonaja textilis	3	K.HFFKPPILSR.F K.ITSITQGATSM(+15.99)TSSM(+15.99)YVK.T	414.61 1033.08	3 2
97	BPTI	IVBI2_PSETT	61.21	Protease inhibitor textilinin-2	Pseudonaja t. textilis	17	R.FPSFYNNPDEQK.C R.VRFPSPFYNNPDEQK.C	767.92 597.37	2 3
		isotig00172	60.76	Protease inhibitor textilinin-1 truncated transcript	Pseudonaja textilis	20	R.FPSFYNNPDEK.S	703.91	2
		IVBI1_PSETT	60.76	Protease inhibitor textilinin-1	Pseudonaja t. textilis	13			
98	BPTI	IVBI1_PSETT	61.55	Protease inhibitor textilinin-1	Pseudonaja t. textilis	17	R.FPSFYNNPDEK.K R.VRFPSPFYNNPDEK.K R.FPSFYNNPDEKK.C	703.88 554.65 767.95	2 2 2
		IVBI2_PSETT	61.54	Protease inhibitor textilinin-2	Pseudonaja t. textilis	17	R.FPSFYNNPDEQK.C R.VRFPSPFYNNPDEQK.C	512.29 597.38	3 3
99	PLA2	PA2B2_NAJMO	61.66	Basic phospholipase A2 CM-II	Naja mossambica	9	R.YIDANYINLK.E	670.91	2
	EXE2	EXE2_HELSC	60.79	Exendin-2-long	Heloderma s. cinctum	11	K.YLASILGSR.T	490.34	2
		EXE2_HEL SU	60.79	Exendin-2-long	Heloderma suspectum	11			
101	PLA2	isotig00264	61.52	Acidic PLA2-A4	Pseudonaja textilis	8	K.GGSGTPVDELDR.C	601.82	2
		isotig00263	61.52	Acidic PLA2-A3	Pseudonaja textilis	8			
		PA226_MICAT	61.52	phospholipase A2	Micrurus altirostris	9			
		R4FIM5_9SAUR	61.52	PLA2-Hop-9	Hoplocephalus bungaroides	8			
		PA2A1_AUSSU	61.52	Acidic phospholipase A2 S1-11	Austrelaps surbus	8			
		PA2B_BUNCE	61.52	Basic phospholipase A2 KPA2	Bungarus caeruleus	8			
		PA2PA_OXYMI	61.52	Basic phospholipase A2 paradoxin-like alpha chain	Oxyuranus microlepidotus	8			
		PA2A2_PSETE	61.52	Acidic phospholipase A2 2	Pseudonaja textilis	8			
		R4G2S8_DENDV	61.52	PLA-2-Den-2	Denisonia devisi	8			
		R4FIQ4_9SAUR	61.52	PLA2-Pse-8	Pseudonaja modesta	8			
102	Clect	isotig00146	84.07	Venom Clect-2	Pseudonaja textilis	11	K.YM(+15.99)WEWTD.R.S	601.84	2
		isotig00147	84.07	Venom Clect-3	Pseudonaja textilis	11	R.SRTDFLLWR.K	597.4	2
		R4G2J2_9SAUR	84.07	LP-Pse-1	Pseudonaja modesta	11	K.YMWEWTD.R.S	593.81	2
		R4G7K1_9SAUR	84.07	LP-Pse-2	Pseudonaja modesta	11	R.TDFLLWR.K	475.79	2
		R4FK55_9SAUR	84.07	LP-Pse-5	Pseudonaja modesta	11			
		isotig00145	84.07	Venom Clect-1	Pseudonaja textilis	11			
		isotig00148	84.06	Venom Clect-4	Pseudonaja textilis	10			
103	Clect	LECG1_BUNFA	84.06	C-type lectin Bfl-1	Bungarus fasciatus	9	K.YIWEWTD.R.S	584.83	2
		LECG_PSEPO	84.06	C-type lectin galacte-binding isoform	Pseudechis porphyriacus	9	R.TDFLPWR.K	467.77	2
		LECG_PSEAU	84.06	C-type lectin galacte-binding isoform	Pseudechis australis	9			
		LECG1_BUNMU	84.06	C-type lectin BML-1	Bungarus multicinctus	9			
		R4FIR9_9SAUR	84.02	LP-Pse-3	Pseudonaja modesta	9	K.YIWEWTD.R.S	584.83	2
		R4G314_9SAUR	84.02	LP-Pse-6	Pseudonaja modesta	9	R.TDFLLWR.K	475.78	2
		D2YVJ3_PSETE	83.02	Venom C-type lectin galacte binding isoform variant 1	Pseudonaja textilis	8	R.TDFLPWR.K K.VFNDPK.N	467.77 360.21	2 2
		isotig00146	82.9	Venom Clect-2	Pseudonaja textilis	8	R.TDFLLWR.K	475.78	2
		isotig00147	82.9	Venom Clect-3	Pseudonaja textilis	8	K.VFNDPK.N	360.21	2
		R4G2J2_9SAUR	82.9	LP-Pse-1	Pseudonaja modesta	8			
		isotig00148	82.85	Venom Clect-4	Pseudonaja textilis	7			
104	3FTx	NXL1_LATLA	56.93	Long neurotoxin LLong (Fragment)	Laticauda laticaudata	6	K.TPYVK.S	304.18	2
		isotig01328	56.88	Long neurotoxin 2	Pseudonaja textilis	6			
		NXL20_DRYCN	56.88	Long neurotoxin 20	Drysdalia coronoides	6			
		NXL31_DRYCN	56.88	Long neurotoxin 31	Drysdalia coronoides	6			
		R4FI75_9SAUR	56.88	3FTx-Aca-53	Acanthophis wellsi	6			
		F8J2E3_DRYCN	56.88	Putative long chain neurotoxin 291	Drysdalia coronoides	6			
		F8J2F1_DRYCN	56.88	Putative long chain neurotoxin 178R	Drysdalia coronoides	6			
		F8J2E8_DRYCN	56.88	Putative long chain neurotoxin 472	Drysdalia coronoides	6			
		R4G319_9SAUR	56.88	3FTx-Pse-78	Pseudonaja modesta	6			
		R4G2P6_9SAUR	56.88	3FTx-Aca-18	Acanthophis wellsi	6			
105	3FTx	NXS2_PSETE	61.55	Short neurotoxin 2	Pseudonaja textilis	18	R.YLIPATHGNAIPAR.G	498.65	3
106	3FTx	NXS2_PSETE	61.61	Short neurotoxin 2	Pseudonaja textilis	18	R.YLIPATHGNAIPAR.G	747.46	2
		NXS7_PSETE	61.61	Short neurotoxin 7	Pseudonaja textilis	18	R.YIIPATHGNAITYR.G	530.66	3
		isotig00066	61.61	SNTx7	Pseudonaja textilis	15			

ACCEPTED MANUSCRIPT								
		isotig00074	61.61	SNTx7	<i>Pseudonaja textilis</i>	18		
		NX53_PSETE	61.61	Short neurotoxin 3	<i>Pseudonaja textilis</i>	18	R.YLVPATHGNAIPAR.G	493.94 3
BPTI		IVB13_PSETT	60.46	Protease inhibitor textilinin-3	<i>Pseudonaja t. textilis</i>	7	R.FYYNPR.Q	430.24 2
		R4FK61_9SAUR	60.46	KP-Pse-5	<i>Pseudonaja modesta</i>	7		
		R4G317_9SAUR	60.46	KP-Pse-7	<i>Pseudonaja modesta</i>	7		
107	PLA2	PA2BB_PSETE	98.42	Basic phospholipase A2 homolog textilotoxin B chain	<i>Pseudonaja textilis</i>	27	R.YNSANYNIDIK.T	657.86 2
							DLVEFGFM(+15.99)IR.C	621.91 2
							R.GSGTPVDDVDR.C	559.29 2
							K.RGSGTPVDDVDR.C	637.38 2
		PA2BA_PSETE	59.78	Basic phospholipase A2 textilotoxin A chain	<i>Pseudonaja textilis</i>	6	R.GTPVDDVDR.C	487.25 2
		isotig00814	59.78	PLA2 textilotoxin A	<i>Pseudonaja textilis</i>	6		
108	PLA2	PA2BB_PSETE	98.35	Basic phospholipase A2 homolog textilotoxin B chain	<i>Pseudonaja textilis</i>	27	R.YNSANYNIDIK.T	657.86 2
							K.RGSGTPVDDVDR.C	637.36 2
							DLVEFGFM(+15.99)IR.C	621.91 2
							DLVEFGFMIR.C	613.89 2
							R.GSGTPVDDVDR.C	559.27 2
		PA2BA_PSETE	60.96	Basic phospholipase A2 textilotoxin A chain	<i>Pseudonaja textilis</i>	6	R.GTPVDDVDR.C	487.27 2
		isotig00814	60.96	PLA2 textilotoxin A	<i>Pseudonaja textilis</i>	6		
109	CRISP	isotig00418	76.03	Cysteine-rich venom protein pseudechetoxin-like	<i>Pseudonaja textilis</i>	3	R.NMLQMK.W	382.71 2
		R4G2J3_9SAUR	76.03	CRISP-Pse-4	<i>Pseudonaja modesta</i>	3		
		R4FIS5_9SAUR	76.03	CRISP-Pse-17	<i>Pseudonaja modesta</i>	3		
		CRVP_OXYMI	76.03	Cysteine-rich venom protein pseudechetoxin-like	<i>Oxyuranus microlepidotus</i>	3		
		CRVP_PSETE	76.03	Cysteine-rich venom protein pseudechetoxin-like	<i>Pseudonaja textilis</i>	3		
		CRVP_OXYSC	76.03	Cysteine-rich venom protein pseudechetoxin-like	<i>Oxyuranus s. scutellatus</i>	3		
		R4G7K2_9SAUR	76.03	CRISP-Pse-11	<i>Pseudonaja modesta</i>	3		
		PA2BA_PSETE	60.69	Basic phospholipase A2 textilotoxin A chain	<i>Pseudonaja textilis</i>	6	R.GTPVDDVDR.C	487.25 2
		isotig00814	60.69	PLA2 textilotoxin A	<i>Pseudonaja textilis</i>	6		
110	PLA2	isotig00264	84.18	Acidic PLA2-A4	<i>Pseudonaja textilis</i>	16	R.FSGPYWNPYSYK.C	754.89 2
		isotig00263	84.18	Acidic PLA2-A3	<i>Pseudonaja textilis</i>	16	K.GGSGTPVDELDR.C	601.87 2
		PA2A2_PSETE	84.18	Acidic phospholipase A2 2	<i>Pseudonaja textilis</i>	16		
111	Clect	isotig00146	83.49	Venom Clect-2	<i>Pseudonaja textilis</i>	9	K.YM(+15.99)WEWTD.R.S	601.8 2
		isotig00147	83.49	Venom Clect-3	<i>Pseudonaja textilis</i>	9	R.TDFLLWR.K	475.81 2
		R4G2J2_9SAUR	83.49	LP-Pse-1	<i>Pseudonaja modesta</i>	9		
		R4G7K1_9SAUR	83.49	LP-Pse-2	<i>Pseudonaja modesta</i>	9		
		R4FK55_9SAUR	83.49	LP-Pse-5	<i>Pseudonaja modesta</i>	9		
		isotig00145	83.49	Venom Clect-1	<i>Pseudonaja textilis</i>	9		
		isotig00148	83.48	Venom Clect-4	<i>Pseudonaja textilis</i>	9		
112	Clect	isotig00146	84.02	Venom Clect-2	<i>Pseudonaja textilis</i>	9	K.YM(+15.99)WEWTD.R.S	601.84 2
		isotig00147	84.02	Venom Clect-3	<i>Pseudonaja textilis</i>	9	K.YMWEWTD.R.S	593.8 2
		R4G2J2_9SAUR	84.02	LP-Pse-1	<i>Pseudonaja modesta</i>	9	R.TDFLLWR.K	475.78 2
		R4G7K1_9SAUR	84.02	LP-Pse-2	<i>Pseudonaja modesta</i>	9		
		R4FK55_9SAUR	84.02	LP-Pse-5	<i>Pseudonaja modesta</i>	9		
		isotig00145	84.02	Venom Clect-1	<i>Pseudonaja textilis</i>	9		
		isotig00148	84.01	Venom Clect-4	<i>Pseudonaja textilis</i>	9		
	VF5a	isotig00199	61.41	VF5a-3	<i>Pseudonaja textilis</i>	1	R.DALSGLLGPTLR.G	606.95 2
		FASV_PSETE	61.41	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	1		
		isotig00200	61.41	VF5a-2	<i>Pseudonaja textilis</i>	1		
113	VF10a	FAXC_PSETE	83.3	Venom prothrombin activator pseutarin-C catalytic subunit	<i>Pseudonaja textilis</i>	4	K.VPYVDR.H	374.71 2
		isotig00188	83.3	VF10a isoform 1	<i>Pseudonaja textilis</i>	4	R.AETGPLLSVDR.V	565.38 2
		isotig00189	83.3	VF10a isoform 1	<i>Pseudonaja textilis</i>	4		
114	Clect	isotig00146	82.17	Venom Clect-2	<i>Pseudonaja textilis</i>	9	K.YM(+15.99)WEWTD.R.S	601.82 2
		isotig00147	82.17	Venom Clect-3	<i>Pseudonaja textilis</i>	9	R.TDFLLWR.K	475.79 2
		R4G2J2_9SAUR	82.17	LP-Pse-1	<i>Pseudonaja modesta</i>	9		
		R4G7K1_9SAUR	82.17	LP-Pse-2	<i>Pseudonaja modesta</i>	9		
		R4FK55_9SAUR	82.17	LP-Pse-5	<i>Pseudonaja modesta</i>	9		
		isotig00145	82.17	Venom Clect-1	<i>Pseudonaja textilis</i>	9		
		isotig00148	82.16	Venom Clect-4	<i>Pseudonaja textilis</i>	9		
115	VF10a	FAXC_PSETE	98.17	Venom prothrombin activator pseutarin-C catalytic subunit	<i>Pseudonaja textilis</i>	7	K.QDFGIVSGFGGIFER.G	814.98 2
		isotig00188	98.17	VF10a isoform 1	<i>Pseudonaja textilis</i>	7	R.AETGPLLSVDR.V	565.37 2
		isotig00189	98.17	VF10a isoform 1	<i>Pseudonaja textilis</i>	7	K.VPYVDR.H	374.73 2
116	VF10a	FAXC_PSETE	83.45	Venom prothrombin activator pseutarin-C catalytic subunit	<i>Pseudonaja textilis</i>	4	R.AETGPLLSVDR.V	565.36 2
		isotig00188	83.45	VF10a isoform 1	<i>Pseudonaja textilis</i>	4	K.VPYVDR.H	374.69 2
		isotig00189	83.45	VF10a isoform 1	<i>Pseudonaja textilis</i>	4		
117	DPP	V8NS94_OPHHA	98.86	Dipeptidase	<i>Ophiophagus hannah</i>	11	R.TEVLLEMAASR.L	610.38 2
							R.TEVLLELM(+15.99)AASR.L	618.41 2
							K.LSNTNTNIEK.L	567.36 2
							R.M(+15.99)FYDLGVR.Y	508.81 2
							R.MFYDLGVR.Y	500.82 2
							K.LSTIDLK.L	459.33 2
							R.LTLEQIDVVK.R	579.43 2
							R.M(+15.99)FYDLGVR.Y	508.81 2
							R.MFYDLGVR.Y	500.82 2
							R.LTLEQIDVVKR.M	438.65 3
120	VF5a	isotig00199	99.16	VF5a-3	<i>Pseudonaja textilis</i>	10	R.GEVGDSLIIYFK.N	670.94 2
		FASV_PSETE	99.16	Venom prothrombin activator pseutarin-C non-catalytic subunit	<i>Pseudonaja textilis</i>	6	R.DALSGLLGPTLR.G	606.93 2
		isotig00200	99.16	VF5a-2	<i>Pseudonaja textilis</i>	6	K.WSEGSYSYSDGTSVDR.L	881.39 2
							K.NFATQPVSIHPQSAVYNK.W	667.77 3
							K.WLISLVAK.H	508.87 2
							R.EYELDFK.Q	472.28 2
							K.HLQAGM(+15.99)GYLNIK.D	508.65 3