

Primary light-energy conversion in tetrameric chlorophyll structure of photosystem II and bacterial reaction centers: II. Femto- and picosecond charge separation in PSII D1/D2/Cyt b559 complex

I. V. Shelaev · F. E. Gostev · V. A. Nadtochenko ·
A. Ya. Shkuropatov · A. A. Zabelin · M. D. Mamedov ·
A. Yu. Semenov · O. M. Sarkisov · V. A. Shuvalov

Received: 4 July 2008 / Accepted: 15 September 2008 / Published online: 15 October 2008
© Springer Science+Business Media B.V. 2008

Abstract In Part I of the article, a review of recent data on electron-transfer reactions in photosystem II (PSII) and bacterial reaction center (RC) has been presented. In Part II, transient absorption difference spectroscopy with 20-fs resolution was applied to study the primary charge separation in PSII RC (D1/D2/Cyt b 559 complex) excited at 700 nm at 278 K. It was shown that the initial electron-transfer reaction occurs within 0.9 ps with the formation of the charge-separated state $P680^+Chl_{D1}^-$, which relaxed within 14 ps as indicated by reversible bleaching of 670-nm band that was tentatively assigned to the Chl_{D1} absorption. The subsequent electron transfer from Chl_{D1}^- within 14 ps was accompanied by a development of the radical anion band of $Pheo_{D1}$ at 445 nm, attributable to the formation of the secondary radical pair $P680^+Pheo_{D1}^-$. The key point of this model is that the most blue Q_y transition of Chl_{D1} in RC is allowing an effective stabilization of separated charges. Although an alternative mechanism of charge separation with Chl_{D1}^* as a primary electron donor and $Pheo_{D1}$ as a primary acceptor can not be ruled out, it is less consistent with the kinetics and spectra of absorbance changes induced in the PSII RC preparation by femtosecond excitation at 700 nm.

Keywords Chlorophyll · Pheophytin · Photosystem II · Primary charge separation · Reaction center

Abbreviations

B_A	Bacteriochlorophyll, primary electron acceptor in BRC
BRC	Bacterial reaction center
D1/D2/ Cytb559	PSII RC
CD	Circular dichroism
Chl	Chlorophyll <i>a</i>
Chl_{D1}	Chl located in D1 protein subunit
ET	Electron transfer
HOMO	Highest occupied molecular orbital
LUMO	Lowest unoccupied molecular orbital
Pheo	Pheophytin <i>a</i>
$Pheo_{D1}$	Pheo located in D1 protein subunit
PSII	Photosystem II
Q_A	Primary plastoquinone electron acceptor
RC	Reaction center

Introduction

In Part I of the article, a review of recent data on the mechanism of primary charge separation in bacterial reaction centers (BRC) and photosystem II RC (PSII RC) has been presented. The role of tetrameric (bacterio)chlorophyll structure in efficient charge separation in bacterial and oxygenic photosynthesis was discussed. In Part II, we describe new data on the kinetics and spectra of absorbance changes in isolated PSII RCs (D1/D2/Cyt b559) at 278 K obtained by the pump–probe method with 20-fs time resolution and with excitation centered at 700 nm.

I. V. Shelaev · F. E. Gostev · V. A. Nadtochenko ·
O. M. Sarkisov
NN Semenov Institute of Chemical Physics, Russian Academy
of Sciences, 117991 Moscow, Russia

A. Ya. Shkuropatov · A. A. Zabelin · V. A. Shuvalov (✉)
Institute of Basic Biological Problems, Russian Academy
of Sciences, Pushchino, Moscow Region 142290, Russia
e-mail: shuvalov@issp.serpukhov.su

M. D. Mamedov · A. Yu. Semenov
AN Belosersky Institute of Physical–Chemical Biology,
Moscow State University, 119992 Moscow, Russia

Materials and methods

Preparation

Isolated PSII RCs (D1/D2/Cytb₅₅₉ complexes) were prepared from spinach by the method described in Van Leeuwen et al. (1991). The RC preparations were suspended in a buffer consisting of 20 mM Bis-Tris (pH 6.5), 0.03% (w/v) *n*-dodecyl- β -D-maltoside, and 200 mM sucrose. The optical density at the Q_y absorption maximum (675.5 nm) was adjusted to about 1.0/mm. The measurements were done at 278 K under anaerobic conditions.

Femtosecond laser photolysis setup

Transient absorption spectra were measured by the femtosecond pump-supercontinuum probe setup. The output of a Ti:sapphire oscillator (800 nm, 80 MHz, 80 fs, «Tsunami», «Spectra-Physics», USA) was amplified by a regenerative amplifier system («Spitfire», «Spectra-Physics», USA) up to 1 mJ/pulse at the repetition rate of 1 KHz. The amplified pulses were splitted into two beams. One of the beam was directed in a noncollinearly phase-matched optical parametric amplifier. Its output centered at 700 nm was compressed by a pair of quartz prisms. The gauss pulse of 20 fs at 700 nm with the bandwidth of ~ 40 nm (full width at half-maximum) was used as a pump. The spectral profile of the pump pulses gives three times more excitation of PSII RC at 680 nm than at 670 nm. The second beam was focused onto a thin quartz cell with H₂O to generate supercontinuum probe pulses. The pump and probe pulses were time-delayed with respect to each other by means of a computer-controlled delay stage. They were then attenuated, recombined, and focused onto the sample cell. The pump and probe light spots have the diameters of 200 and 120 μ m, respectively. The pump pulse energy was attenuated at 100 nJ to get optimal excitation on a linear part of the light curve. The pump pulse operation frequency was 50 Hz, low enough to exclude permanent bleaching of the sample due to photochemical processes in RC. The relative polarizations of the pump and the probe beams were adjusted to 54.7° (magic angle). After the sample, the supercontinuum was dispersed by a polychromator («Acton SP-300») and detected by a CCD camera («Roper Scientific SPEC-10»). Transient spectra of absorbance changes $\Delta A(t, \lambda)$ were recorded over the ranges of 400 to 740 nm. The measured spectra were corrected for group delay dispersion in the supercontinuum using the procedure described previously (Ushakov et al. 2004). Experiments were carried out at 278 K in a 0.5-mm path length flow optical cell. Together with operation frequency, the circulation rate in the flow cell was fast enough to avoid multiple excitation of the same sample volume.

Results

Figure 1 shows the spectra of absorbance changes (ΔA) at 278 K obtained with isolated PSII RCs in the range of 400 to 710 nm at various delays (between 0.1 and 28.5 ps) relative to the 20-fs excitation pulse centered at 700 nm. The main changes are related to the bleaching of Soret and Q_y bands of Chl/Pheo molecules at ~ 430 and 682 nm, respectively, including stimulated emission from those molecules, which gives more intensive bleaching in red than in blue. In agreement with the previous measurements (Shkuropatov et al. 1999) indicating that both Pheo molecules contribute to the most red absorption in the RC around 680 nm, the bleaching of the Q_x band of Pheo at about 545 nm is observed at early delay times (0.1–0.2 ps) and persists up to 28.5 ps, the longest delay time employed in this work. The amplitude of the 545-nm bleaching is almost constant within this time period. This observation suggests that the excited state of the RC includes partially Pheo*_{D1,D2} that is eventually converted to the charge-separated state P680⁺Pheo_{D1}[−] with similar bleaching at 545 nm. It should be noted that the bleaching of Pheo narrow band at 515 nm is increased, but that at 420 nm is relaxed in ps-time domain showing a transformation of (Pheo_{D1}/Pheo_{D2})* to (Pheo_{D1})[−].

Inset of Fig. 1 shows that kinetics of ΔA at 665 nm includes fast (completed within 2.5 ps) bleaching with subsequent relaxation with time constant of 13.3 ps. As it is shown later, these absorbance changes are related to the primary electron acceptor (Chl670) reduction and its reoxidation is due to further electron transfer to Pheo_{D1}.

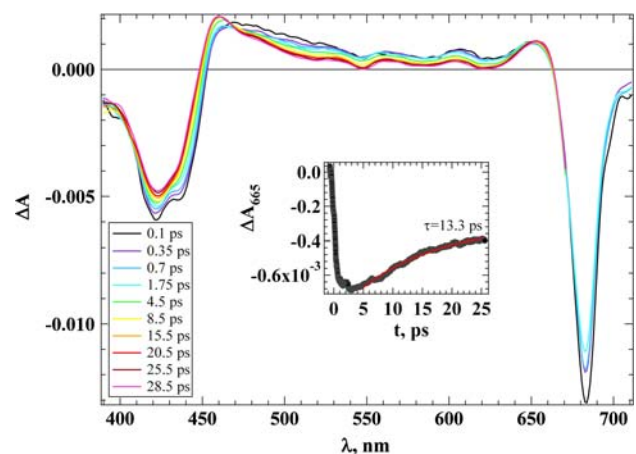


Fig. 1 Difference absorption spectra of D1/D2/Cytb559 complex excited by 20-fs pulses centered at 700 nm with energy of 100 nJ at 278 K. The spectra measured at different delays is indicated in inset A. Inset B shows the kinetics of ΔA at 665 nm revealed directly from the experiment. Fast bleaching at 665 nm is completed to ~ 2.5 ps and followed by the relaxation with time constant of 13.3 ps

Most profound changes of ΔA spectra are observed in the spectral region of 410 to 470 nm (Fig. 1) suggesting the fs- and ps-formation of the radical anion bands of Chl and/or Pheo, which are known to absorb near 450 nm (Fujita et al. 1978). In order to reveal the dynamics of this process, the ΔA spectrum of the RC excited state measured at the earliest delay of about 0.1–0.15 fs was subtracted from the spectra taken at later delays to get the spectra of $\Delta\Delta A$. This was done suggesting that the bleaching of the Soret band is similar for (Chl/Pheo)* and (Chl/Pheo)⁻. Then, the difference in $\Delta\Delta A$ near 450 nm is mostly related to the radical anion bands of (Chl/Pheo)⁻. The result of the subtraction is shown in Fig. 2 for the range of 405 to 585 nm. The appearance of the 445-nm radical anion band very similar to the radical anion band (see Fujita et al. 1978) can be observed already at the delays shorter than 1 ps. It is followed by a development approximately within ≥ 30 ps. The kinetics of this development (Fig. 2, inset B) was approximated by the exponential components with the time constants of 1.4 ± 0.2 ps (amplitude $\sim 26\%$) and 14.7 ± 5 ps ($\sim 74\%$). The indicated amplitudes (in parentheses) were changed up to $\sim 50\%$ for both the components after improving resolution.

Apparently, the development of the radical anion band at 445 nm should be accompanied by a decay of the excited state of RC. The broad, negative $\Delta\Delta A$ band in the region of 470–580 nm centered at ~ 510 nm (Fig. 2), which is developed simultaneously with excitation of the RC and then is decreased within fs and ps time domains (Fig. 2), can be used as an indicator for the formation and decay of the excited state of RC.

Figure 2, inset C, shows that the kinetics of the decay at 510 nm can be described by two exponential components

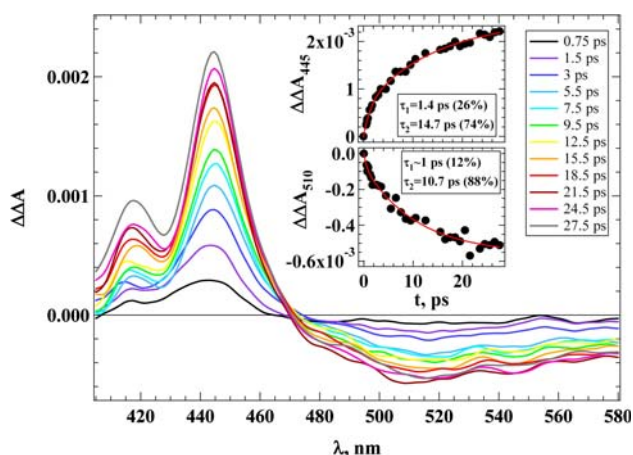


Fig. 2 Data of Fig. 1 were used to calculate the difference–difference absorption spectra in the range of 415 to 580 nm, which are obtained as a result of the subtraction of the spectrum in time domain of 0.1–0.15 ps from the spectra at later delays indicated in inset A. Insets B and C show the kinetics of $\Delta\Delta A$ at 445 and 510 nm, respectively. Two kinetic components of $\Delta\Delta A$ are revealed with time constants of 1–1.4 and 10–15 ps

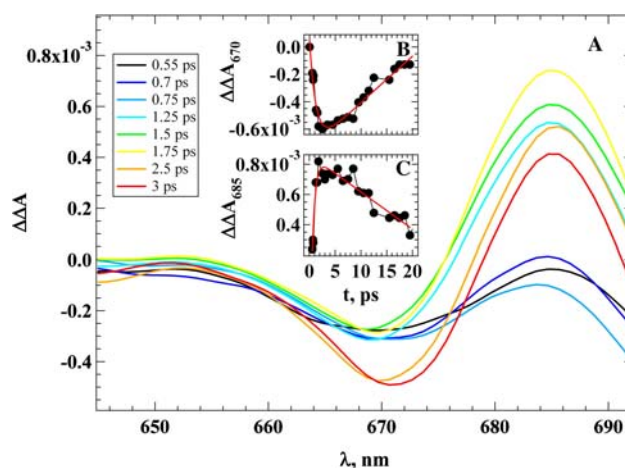


Fig. 3 Data of Fig. 1 were used to calculate the difference–difference absorption spectra in the range of 645 to 692 nm, which are obtained as a result of the subtraction of the spectrum in time domain of 0.1–0.15 ps from the spectra at later delays indicated in inset A. Insets B and C show the kinetics of $\Delta\Delta A$ at 670 and 685 nm, respectively. Two kinetic components of $\Delta\Delta A$ are revealed with time constants of ~ 1 and ~ 14 ps

with time constants of 0.6 ± 0.6 ps ($\sim 12\%$) and 10.7 ± 2 ps ($\sim 88\%$), respectively, which are similar to those observed for the kinetics of $\Delta\Delta A_{445}$ within the experimental error.

The nature of the two kinetic components observed for $\Delta\Delta A_{445}$ and $\Delta\Delta A_{510}$ can be further clarified by the measurements of $\Delta\Delta A$ in the region of the Q_y absorption of the RC pigments. For this purpose, Fig. 3 shows the calculated difference spectra ($\Delta\Delta A$) for different delays that are similar to those depicted in Fig. 2, but are extended to the range of 645 to 740 nm. Here, fs- and ps-dynamics show the main spectral features including bleaching at 670 nm and development centered at 685 nm, close to the spectral position of the main absorption bleaching in RC upon the 700-nm excitation (Fig. 1). The amplitudes of the changes around 670 and 685 nm measured as differences

$$\Delta\Delta A_{670} = \Delta A_{670} - (\Delta A_{645} + \Delta A_{685})/2 \text{ and}$$

$$\Delta\Delta A_{685} = \Delta A_{685} - (\Delta A_{670} + \Delta A_{700})/2$$

were used as an indicator for kinetic bleaching and development. Figure 3 insets show the kinetics of $\Delta\Delta A_{670}$ and $\Delta\Delta A_{685}$. Both kinetics were approximated by the two exponential components with opposite signs and time constants of ~ 1 and ~ 15 ps.

Discussion

Although the DI/DII/Cyt b559 RC complex binds a minimum number of cofactors (six Chls and two Pheos), the dynamics of energy transfer between chromophores and the kinetics and mechanism of primary charge separation in

RC are not yet well understood. The controversy (see Part I of this article) is, to great extent, caused by a significant overlap in the Q_y spectral region together with several difficulties in distinguishing excited states from charge-separated states. Also, rapid equilibration among excited states, between excited states and charge-separated states, and among charge-separated states seems to be observed in the PSII RC that makes it difficult to determine the intrinsic rate constants for charge separation (see, for example, Germano et al. 2004; Groot et al. 2005).

Van Brederode et al. (1997, 1999) were the first to show that direct excitation of the accessory BChl B_A in RCs of purple bacteria can lead to primary charge separation with time constant shorter than 1 ps. Apparently, this reaction should have a low quantum yield in native bacterial RCs because it is in competition with very fast (~ 100 fs) energy transfer from B_A^* to P870 (Vulto et al. 1997), or it might be related to two-photon process. This finding has triggered, however, a discussion on the nature of the primary electron donor in PSII RCs (Van Brederode et al. 1999; Dekker and van Grondelle 2000). Recently, the data have been presented suggesting that the lowest excited state in the PSII RC was not necessarily localized on the “special pair” P680* and could be on Chl_{D1}^* (Prokhorenko and Holzwarth 2000; Diner et al. 2001; Diner and Rappaport 2002; Barter et al. 2003; Germano et al. 2004; Groot et al. 2005; Holzwarth et al. 2006; Pawlowicz et al. 2007; Raszewski and Renger 2008; Raszewski et al. 2008). However, for example, the photon echo data (Prokhorenko and Holzwarth 2000) are mostly based on excitonic calculations including the site energies which are not well defined yet. It is not yet clear at present, whether a single mechanism is realized in PSII RCs or both Chl_{D1}^* and P680* can initiate charge separation. In this respect, it is interesting that a large variation in the initial rates of charge separation has been reported for PSII RC, depending on the experimental conditions such as the wavelengths of excitation and temperature (see Germano et al. 2004 and the references therein).

It is well known that the Q_y absorption of all the eight pigments in the PSII RC appears as a single band centered at 675–676 nm (675.5 nm in our preparation) at room temperature, which is only partially resolved at cryogenic temperatures, displaying overlapping bands around 670 and 680 nm, with a shoulder at about 684 nm. The Pheo Q_x transition is well resolved from that of the Chls, peaking at about 542–543 nm at room and low temperatures. The main purpose of femtosecond pump light pulses at 700 nm employed in this work was to minimize the excitation of the pigments absorbing at 670 nm. As indicated in [Materials and methods](#), these pulses give three times more excitation at 680 nm than at 670 nm in RC.

Figure 1 shows that the absorption difference spectra obtained at early delay times (0.1–0.2 ps) display the Pheo

bleaching at ~ 545 nm, about the same wavelength as the Q_x maximum in the ground state absorption (not shown). The negative band at approximately 580 nm attributable to the bleaching of the Chl Q_x absorption band is also seen in the difference spectra. Between 470 and 620 nm, the bleaching of the Pheo and Chl Q_x transitions is superimposed on a broad development due to excited state absorption of the pigments. In the Q_y region, the early time difference spectra (Fig. 1) show an asymmetric negative band that is red-shifted (to 682 nm) with respect to the main ground state absorption band peaking at 675.5 nm. This 682-nm band represents the ground state bleaching/stimulated emission contributions from the Pheo and Chl molecules excited by the 700-nm pulses. A broad, positive signal of relatively small amplitude between 620 and 660 nm results from the excited state absorption of those molecules.

The low temperature experiments performed with Pheo-modified D1/D2/Cytb559 RC complexes have shown that the central pigments of RC, two Pheos, and four Chls seem to absorb in the 676–685-nm range (Germano et al. 2001). Delocalized excited states were detected in the active cofactor branch, while the inactive branch Pheo and the nearby Chl mainly contributed to localized transitions at 676 and 680 nm, respectively (Germano et al. 2001). At least two distant chlorophylls Chl_{Z1} and Chl_{Z2} , which transfer excitation energy to the 680-nm pigment pool contribute to the PSII RC absorption band around 670 nm. With this assignment, the pump pulse centered at 700 nm could potentially excite several chromophores, with some selection for transitions in the active cofactor branch. It is therefore reasonable to assume that at least P680 and Pheo $_{D1}$ are the cofactors excited at 700 nm. The Q_y transition of Chl_{D1} is also a possible candidate for excitation with the 700-nm flash (Germano et al. 2001); however, the sum of the data obtained in this report seems to be better consistent with its spectral position at 670 nm (see below).

Our interpretation of the results of measurements in PSII RC excited by 20-fs pulses centered at 700 nm and presented in Figs. 1, 2, and 3 is as follows. Amplitude of the bleaching of Pheos band (at 545 nm) simultaneously with the excitation is similar to that induced by the formation of Pheo $^-$ at ≥ 30 ps delay (Fig. 1). It means that the contribution of Pheo $_{D1}^*$ (and possibly Pheo $_{D2}^*$) to the bleaching at 545 nm in excited RC is almost equal to the contribution of Pheo $_{D1}^-$ bleaching at 545 nm connected to radical pair formation. If one suggests that the extinction coefficients are equal for the two bleachings, it would mean that Pheos* are converted to Pheo $_{D1}^-$ when RC are excited at 700 nm. One can find that the ratio of the bleachings at 420/545 nm equal to ~ 13 in the excited state is slightly greater than the 420/545 nm ratio (~ 8) in the photoaccumulation of Pheo $_{D1}^-$ (Klimov et al. 1977, 1980). Furthermore, the

bleaching at 435 nm is observed in the excited state of RC and not in the photoaccumulation of Pheo⁻. It means that in RC* excited by 700-nm pulses, the bleachings include some other pigments, probably P680, having an absorption at 435 nm (Doring et al. 1967, 1969). This is consistent with the 682-nm bleaching that includes the stimulated emission as well.

There are two possible explanations for the results presented in Figs. 2 and 3: (1) the current model of charge separation in PSII RC (see above and Part I of this article) suggesting that the excited state Chl_{D1}* transfers an electron to Pheo_{D1} with the formation of the first radical pair Chl_{D1}⁺Pheo_{D1}⁻; (2) the BRC-like model suggesting that an electron is transferred from P680* to Pheo_{D1} possibly via Chl_{D1}. In both cases, an electron from Pheo_{D1}⁻ is transferred to Q_A within 200 ps (Nuijs et al. 1986). Further, we shall denote these models as Model 1 and Model 2, respectively. Some of the results presented here are consistent with both models, but some are not. Model 1 suggests that the radical anion band of Pheo⁻ at 445 nm (Fujita et al. 1978) should be developed with fastest time constant as well as the bleaching of Chl_{D1} bands due to the formation of Chl_{D1}⁺. Only after the completion of this fast reaction, an electron is transferred from P680 to Chl_{D1}⁺ with a slower time constant. According to Model 2, the fast reaction reflects the formation of P680⁺ Chl_{D1}⁻ while slow one corresponds to the electron transfer from Chl_{D1}⁻ to Pheo_{D1}.

Indeed, according to the described results, there are two kinetic components with average time constants of 0.9 and 14 ps. The subtraction of the spectrum of ΔA, measured at 0.1–0.15 ps, from the spectra at later delays reveals some spectral features (Fig. 2, 3), which show the decay of the excited RC measured as a decay of broad band of RC* centered at 510 nm (Fig. 2, inset C) accompanied by the formation of the anion radical band at 445 nm (Fig. 2, inset B) having two components with the average time constants of 0.9 and 14 ps. These results should be discussed according to the assumptions about two mechanisms of the charge separation presented in Models 1 and 2. According to Model 1, the radical anion band of Pheo_{D1}⁻ at 445 nm cannot be developed in slow kinetics if we do not suggest some special influence of Chl_{D1}⁺ on the spectral band of Pheo_{D1}⁻, which disappears when an electron is transferred from P680 to Chl_{D1}⁺. In agreement with Model 2, the Pheo_{D1}⁻ band at 445 nm should be developed in fast and slow kinetics as it can be seen from Fig. 2, inset B.

According to Model 1, the excited state RC* with pre-dominant excitation of Chl_{D1} should disappear synchronously with the formation of the state Chl_{D1}⁺Pheo_{D1}⁻ in contrast to fast and slow component kinetics of the RC* broad band observed at 510 nm (Fig. 2, inset C). This contradiction can be avoided if one suggests

that the excitation of the red band of RC with bleaching at 682 nm (Fig. 1) do not include the excitation of Chl_{D1}. In this case, a new contradiction appears: the Chl_{D1} cannot be a trap for excited RC (at least by fs pulses centered at 700 nm). Model 2 suggests that P680 is excited by fs broad band centered at 700 nm together with the excitation of Pheo molecule(s) in some RC. Then P680* transfers an electron to Chl_{D1} and then to Pheo_{D1} like in BRC. Limited contribution (~26%, which can be increased up to 50% after better resolution) of fast component to ΔA₄₄₅ can be explained by a smaller extinction coefficient for Chl_{D1}⁻ due to the overlapping of the bleaching and appearing bands or by ~50% mixing of the states P680* and P680⁺Chl_{D1}⁻.

The spectra presented in Fig. 3 can be considered from the point of view of dynamic hole burning. This suggestion is unlikely since the sub-ps relaxation of bleached 670-nm band should be observed but that is not the case. According to Model 2, the bleaching at 670 nm seen in Fig. 3 can be an indication of the involvement of Chl-670 in the charge separation process. Kinetics of the bleaching at 670 nm as well as of the development at 685 nm includes two components with similar time constants mentioned above, but with opposite directions: fast component (0.9 ps) shows the bleaching at 670 nm and development at 685 nm, while slow component (14 ps) shows the relaxations of both processes. The bleaching at 670 nm can be interpreted as a photoreduction of Chl_{D1} due to the primary charge separation between P680* and Chl_{D1}. Since this process is a conversion of RC* to the charge separated state, it should be accompanied by a loss of stimulated emission at 685 nm which can be observed as positive ΔΔA at 685 nm with similar kinetics (Fig. 3, inset C). Further appearance of bleaching at 685 nm (14 ps) could be due to slower electron transfer from Chl_{D1}⁻ to Pheo_{D1}. Furthermore, the photoreduction of Chl_{D1} is accompanied by similar amplitude development at 445 nm (Fig. 2) at earlier delays, as it should be observed according to the reduction of Chl (Fujita et al. 1978). Since the band bleaching at 670 nm was observed always when Pheo⁻ in PSII was photoaccumulated (Klimov et al. 1977, 1980), it can be suggested that this bleaching is a result of the quantum mechanical exchange interaction between Chl_{D1} and Pheo_{D1}⁻. In fact, the bleaching of the 670-nm band is clearly observed even at 77 K (Shuvalov et al. 1989). The fast (~0.9 ps) bleaching and slow (~14 ps) relaxation of ΔA at 665 nm and ΔΔA at 670 nm (Figs. 1, 3) are not consistent with a role of that bleaching as a simple indicator of Pheo reduction since Pheo is constantly reduced in ps time domain (Fig. 2). If Chl_{D1} absorbs at 670 nm, it is clear that Chl_{D1} cannot be a trap for excitation energy in contrast to Model 1. Then the excitation centered at 700 nm allows observing fs kinetics of photoreduction of

Chl_{D1} and ps reoxidation of Chl_{D1}[−] by further ET to Pheo_{D1}.

The Model 2 is consistent with data on CD and LD measurements in PSII RC at low temperature. At 6 K, the CD spectrum of PSII RC includes two positive bands at 680 and 673 nm and a negative band at 667 nm (Germano et al. 2001). The bands at 680 and 667 nm seem to represent the excitonic interaction in the dimer P680 with splitting energy of $\sim 300 \text{ cm}^{-1}$ in good agreement with the calculated coupling energy of $\sim 150 \text{ cm}^{-1}$ (Raszewski et al. 2005, 2008). This conservative CD spectrum is not distorted by the reduction of Pheo_{D1} except a few nm blue shift of P680 bands observed in CD (Tetenkin et al. 1989) and in absorbance changes spectra (Ganago et al. 1982). Therefore, the positive CD band at 673 nm can reflect a rotation strength of the transition of Chl_{D1} which is similar to that of B_A in BRC at 800 nm. In both cases, some magnetic dipole moment of the transitions appears to be observed in non-conservative CD spectra at 673 nm (PSII RC) and 800 nm (BRC). Meaning of that for an electron acceptor molecule should be clarified. Linear dichroism measurements at 6 K show (Germano et al. 2001) that both transitions at 673 and 680 nm have positive polarization. This polarization is increased (upto approximately +0.25) after replacement of Pheo_{D1,D2} with 13'-OH-Pheo showing that transitions belong to Chl_{D1/D2} and P680, respectively. In addition, the replacement of Pheo's leads to the decrease of the negative polarization in the 680-nm region caused by the disappearance of Pheo's transitions at 680 nm with perpendicular orientation (Germano et al. 2001). Interestingly, the oxidation of P870 in BRC is accompanied by a blue shift of the primary electron acceptor B_A band at 800 nm. Similar blue shift of the band is observed at 670 nm, when P680 is oxidized at 77 K (Shuvalov et al. 1989). Another interesting feature is revealed under the photoreduction of Pheo_{D1} at 77 K accompanied by the bleaching of the 668-nm band ($\sim 670 \text{ nm}$ under correction taking into account a development at 675 nm), which is similar to the bleaching of the B_A band at 800 nm under photoreduction of BPheo_A in BRC (Shuvalov and Klimov 1976). Furthermore, the transitions of Chl_{D1,D2} should have a negative polarization according to their orientation in X-ray model (Ferreira et al. 2004) which is consistent with the negative band at 668 nm in LD spectra independent of the presence of Pheo_{D1,D2} in RCII (Germano et al. 2001). Thus, one can conclude that the transition at 673 nm has a positive polarization in LD and a positive rotation strength in CD, and these features are not distorted by the replacement of Pheo_{D1,D2} with 13'-OH-Pheo. These features show evidently that the transition at 673 nm belongs to Chl_{D1/D2}. Taking into account that wavelength at 673 nm is shifted to the red by $\sim 3 \text{ nm}$ by the presence of negative bands at 668 nm in LD spectra (due to Chl_{D1,D2})

and at 667 nm in CD spectra (probably, due to short wavelength exciton transition of P680), the Chl_{D1/D2} transition is located around 670 nm. Participation of the Chl-670 in the electron transfer shows that it is Chl_{D1} (Fig. 3).

Let us consider the energies of the pigment HOMO and LUMO orbitals participating in the electron transfer in PSII RC and BRC. In BRC, most red Q_Y absorption belongs to P870 ($870\text{--}11,494 \text{ cm}^{-1}$), intermediate to B_A ($800\text{--}12,500 \text{ cm}^{-1}$), and most blue absorption belongs to H_A ($760\text{--}13,158 \text{ cm}^{-1}$). Energy difference $\Delta E_{BP} = 1,006 \text{ cm}^{-1}$ and $\Delta E_{HB} = 658 \text{ cm}^{-1}$. If energy of P* is above LUMO of B by $\sim 450 \text{ cm}^{-1}$ (Shuvalov and Yakovlev 1998; Nowak et al. 1998), then the energy of HOMO of P⁺ is above that of B[−] by $1,450 \text{ cm}^{-1}$. When an electron is transferred from P* to B_A, the back reaction from B_A[−] to LUMO of P⁺ has a barrier of 450 cm^{-1} and from original HOMO of B[−] to HOMO of P⁺ has a barrier of $1,450 \text{ cm}^{-1}$. These barriers are enough to block the back reactions at room temperature. If energy of B* is above LUMO of H by $\sim 1,500 \text{ cm}^{-1}$ then the energy of HOMO of B⁺ is above that of original H[−] by $2,158 \text{ cm}^{-1}$. When an electron is transferred from B* to H, the back reaction from H[−] to LUMO of B⁺ has a barrier of $1,500 \text{ cm}^{-1}$ and from HOMO of H[−] to original HOMO of B⁺ has a barrier of $2,158 \text{ cm}^{-1}$. These barriers are again enough to block the back reactions at room temperature.

If in PSII RC most red absorption belongs to Pheo_{D1} and P680 ($680\text{--}14,706 \text{ cm}^{-1}$) and most blue to Chl_{D1} ($670\text{--}14,925 \text{ cm}^{-1}$), the energy difference is $\Delta E_{CP} = \Delta E_{CH} = 220 \text{ cm}^{-1}$. If energy of P* is above LUMO of Chl by $\sim 300 \text{ cm}^{-1}$, then the energy of HOMO of P⁺ is above that of Chl[−] by 520 cm^{-1} . According to Model 2, if an electron is transferred from P* to Chl_{D1} then the back reaction from Chl[−] to LUMO of P⁺ has a barrier of 300 cm^{-1} and from original HOMO of Chl[−] to HOMO of P⁺ has a barrier of 520 cm^{-1} . These barriers are good enough to block the back reactions at room temperature. Further, electron transfer to Pheo_{D1} (-0.64 V , Rutherford et al. 1981)) stabilizes the separated charges for several nanoseconds. In the case when all P680, Chl_{D1}, and Pheo_{D1} have transitions near 680 nm then $|\Delta E|$ for the electron transfer is equal to ΔE for the back transfer between primary electron donor and acceptor (P680* and Chl_{D1}, or Chl_{D1}* and Pheo_{D1}) allowing exchange interaction without the stabilization of separated charges. Considerable part of the energy should disappear as fluorescence or dark dissipation in this case.

Thus, in agreement with the charge separation in BRC, the efficient charge separation in PSII RC is consistent with the most blue transition for Q_Y of Chl_{D1} at 670 nm which is revealed from Fig. 3. In other words, the primary charge separation occurs in tetrameric Chl complex of PSII RC between P680* and Chl_{D1} ($\sim 0.9 \text{ ps}$) and then an electron is transferred to Pheo_{D1} ($\sim 14 \text{ ps}$) like in BRC except some

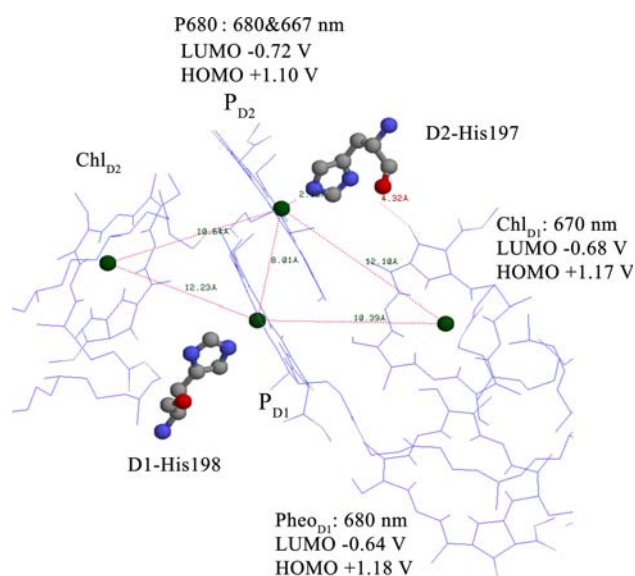
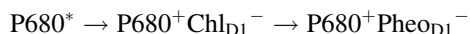


Fig. 4 Arrangement of pigments in PSII RC is found using X-ray analysis by Loll et al. (2005). According to Model 2, P_{D1} and P_{D2} form a dimer, the primary electron donor P680, with excitonic transitions at 680 and 667 nm in PSII RC preparation. Chl_{D1} is a primary electron acceptor with the site energy around 670 nm. Pheo_{D1} is a secondary electron acceptor with site energy around 680 nm and a redox potential of -0.64 V (Rutherford et al. 1981). Using this value and $-\Delta E$ of ~ 0.038 V for each step of electron transfer, the redox potentials for HOMO and LUMO orbitals of the electron donor and acceptors were found and indicated in Fig. 4. According to Model 2, the electron transfer from P680* to Chl_{D1} occurs within 0.9 ps and from Chl_{D1}⁻ to Pheo_{D1} within 14 ps in PSII RC. According to Model 1, Chl_{D1} is a primary electron donor, Pheo_{D1} is a primary electron acceptor, and P680 is a secondary electron donor

difference in time constants (see part I and below). So, the main conclusion from this discussion is consistent with the mechanism of Model 2. The key point of this Model 2 is that the most blue Q_y transition of Chl_{D1} at 670 nm in RC is allowing the effective stabilization of separated charges.

In conclusion, although the results of the femto- pico-second measurements presented in this report cannot rule out Model 1, they seem to be better consistent with the Model 2 for measurements of PSII RC preparation. (It should be noted that according to the recent data of Okubo et al. (2007), the structure of P680 might be perturbed in the PSII RC preparation.) The Model 2 is very similar to that for BRC suggesting that the sequential electron transfer



in PSII RC is realized upon excitation at 700 nm with the time constants for the initial and secondary reactions of 0.9 and 14 ps, respectively. Interestingly, the time constants are different from those observed for purple bacterial RC in which the electron transfer rate from P870* to B_A is slower (~ 3 ps), but from B_A to H_A is faster (~ 1 ps). From recent X-ray analysis, it is known (Fig. 4, Loll et al. 2005) that the center-to-center distances between P_{D1} and P_{D2} is

8.01 Å, between P_{D1} and Chl_{D1} is 10.2 Å, between Chl_{D1} and Pheo_{D1} is 10.6 Å, between P_{D1} and Pheo_{D1} is about 17 Å, and between P_{D2} and Pheo_{D1} is about 20 Å. From this picture, it follows that P_{D1} and P_{D2} are in closest distance, Chl_{D1,D2} are closest pigments to dimer P680, and Chl_{D1} is located between P680 and Pheo_{D1} like a location of B_A between P870 and BPheo_A in BRC structure. Thus, the Model 2 is consistent with the dimer nature of P680 having maximal calculated coupling (158 cm^{-1}) in transition monopole approximation (Raszewski et al. 2005, 2008). Model 2 has an advantage with respect to primary charge separation occurring in excited dimer P680 since it has a parallel arrangement of macrocycles of Chls P_{D1} and P_{D2} (Ferreira et al. 2004; Loll et al. 2005) important for the formation of excimer or exciplex with charge transfer character (Terenin 1967; Beens and Weller 1975; Frese et al. 2003; Hughes et al. 2006). Study of PSII RC and BRC similarities is in progress with respect to the charge separation starting in the dimers P680 and P870 (see also part I).

Acknowledgments We thank Prof. VV Klimov for preparations of PSII RC and for discussion, and MM Leonova for assistance in manuscript preparation. This work was supported by MCB and “Femtosecond optics and new optical materials” grants from the Russian Academy of Sciences, by the State contract 02.512.11.2085, by the President of RF grant SS-4525.2008.4 and by RFFI grants.

References

- Barter LMC, Durrant CJR, Klug DR (2003) A quantitative structure-function relationship for photosystem II reaction center: supermolecular behavior in natural photosynthesis. *Proc Natl Acad Sci USA* 100:946–951. doi:10.1073/pnas.0136891100
- Beens H, Weller A (1975) Excited molecular π -complexes in solution. In: Birks JB (ed) *Organic molecular photophysics*, vol 11. Wiley, London, pp 159–215
- Dekker JP, van Grondelle R (2000) Primary charge separation in photosystem II. *Photosynth Res* 63:195–208. doi:10.1023/A:1006468024245
- Diner BA, Rappaport F (2002) Structure, dynamics, and energetics of the primary photochemistry of photosystem II of oxygenic photosynthesis. *Annu Rev Plant Biol* 53:551–580. doi:10.1146/annurev.arplant.53.100301.135238
- Diner BA, Schlodder E, Nixon PJ, Coleman WJ, Rappaport F, Laverge J, Vermaas WFJ, Chisholm DA (2001) Site-directed mutation at D1-His198 and D2-His197 of photosystem II in *Synechocystis* PCC 6803: sites of the primary charge separation and cation and triplet stabilization. *Biochemistry* 40:9265–9281. doi:10.1021/bi010121r
- Doring G, Stiehl HH, Witt HT (1967) A second chlorophyll reaction in the electron chain of photosynthesis—registration by the repetitive excitation technique. *Z Naturforsch B* 22:639–644
- Doring G, Renger G, Vater J, Witt HT (1969) Properties of the photoactive chlorophyll- a_{711} in photosynthesis. *Z Naturforsch B* 24:1139–1143
- Ferreira KN, Iverson TM, Maghlaoui K, Barber J, Iwata S (2004) Architecture of the photosynthetic oxygen-evolving center. *Science* 303:1831–1838. doi:10.1126/science.1093087

- Frese RN, Germano M, de Weerd FL, van Stokkum IHM, Shkuro-patov AY, Shuvalov VA, van Gorkom HJ, van Grondelle R, Dekker JP (2003) Electric field effects on the chlorophyll, pheophytins, and β -carotenes in the reaction center of photosystem II. *Biochemistry* 42:9205–9213. doi:[10.1021/bi0273516](https://doi.org/10.1021/bi0273516)
- Fujita I, Davis MS, Fajer Y (1978) Anion radicals of pheophytin and chlorophyll *a*: their role in the primary charge separations of plant photosynthesis. *J Am Chem Soc* 100:6280–6282. doi:[10.1021/ja00487a079](https://doi.org/10.1021/ja00487a079)
- Ganago IB, Klimov VV, Ganago AO, Shuvalov VA, Erokhin YE (1982) Linear dichroism and orientation of pheophytin, the intermediary electron acceptor in photosystem II reaction centers. *FEBS Lett* 140:127–130. doi:[10.1016/0014-5793\(82\)80536-X](https://doi.org/10.1016/0014-5793(82)80536-X)
- Germano M, Shkuropatov AY, Permentier H, de Wijn R, Hoff AJ, Shuvalov VA, van Gorkom HJ (2001) Pigment organization and their interactions in reaction centers of photosystem II: optical spectroscopy at 6 K of reaction centers with modified pheophytin composition. *Biochemistry* 40:11472–11482. doi:[10.1021/bi010439j](https://doi.org/10.1021/bi010439j)
- Germano M, Gradinaru CC, Shkuropatov AY, van Stokkum IHM, Shuvalov VA, Dekker JP, van Grondelle R, van Gorkom HJ (2004) Energy and electron transfer in photosystem II reaction centers with modified pheophytin composition. *Biophys J* 86:1664–1672
- Groot ML, Pawlowicz NP, van Wilderen LJGW, Breton J, van Stokkum IHM, van Grondelle R (2005) Initial electron donor and acceptor in isolated photosystem II reaction centers identified with femtosecond mid-IR spectroscopy. *Proc Natl Acad Sci USA* 102:13087–13092. doi:[10.1073/pnas.0503483102](https://doi.org/10.1073/pnas.0503483102)
- Holzwarth AR, Müller MG, Reus M, Nowaczyk M, Sander J, Rögner M (2006) Kinetics and mechanism of electron transfer in intact photosystem II and in the isolated reaction center: pheophytin is the primary electron acceptor. *Proc Natl Acad Sci USA* 103:6895–6900. doi:[10.1073/pnas.0505371103](https://doi.org/10.1073/pnas.0505371103)
- Hughes JL, Smith P, Pace R, Krausz E (2006) Charge separation in photosystem II core complexes induced by 690–730 excitation at 1.7 K. *Biochim Biophys Acta* 1757:841–851. doi:[10.1016/j.bbabi.2006.05.035](https://doi.org/10.1016/j.bbabi.2006.05.035)
- Klimov VV, Klevanik AV, Shuvalov VA, Krasnovsky AA (1977) Reduction of pheophytin in the primary light reaction of photosystem II. *FEBS Lett* 82:183–186. doi:[10.1016/0014-5793\(77\)80580-2](https://doi.org/10.1016/0014-5793(77)80580-2)
- Klimov VV, Allakhverdiev SI, Shutilova NI, Krasnovsky AA (1980) The study of photoreduction of pheophytin and photooxidation of P680 in photosystem II preparations. *Plant Physiol* 27:315–326 (in Russian)
- Loll B, Kern J, Saenger W, Zouni A, Biesiadka J (2005) Towards complete cofactor arrangement in the 3.0 Å resolution structure of photosystem II. *Nature* 438:1040–1044. doi:[10.1038/nature04224](https://doi.org/10.1038/nature04224)
- Nowak FR, Kennis JTM, Franken EM, Shkuropatov AY, Yakovlev AG, Gast P, Hoff AJ, Aartsma TJ, Shuvalov VA (1998) The energy level of P^+B^- in plant pheophytin exchanged bacterial reaction centers probed by the temperature dependence of delayed fluorescence. In: *Proceedings of the XI international congress on photosynthesis*, Budapest, Hungary, Kluwer Academic Publishing, Dordrecht, pp 783–786
- Nuijs AM, van Gorkom HJ, Plijter JJ, Duysens LNM (1986) Primary charge separation and excitation of chlorophyll *a* in photosystem II particles from spinach as studied by picosecond absorbance difference spectroscopy. *Biochim Biophys Acta* 848:167–175. doi:[10.1016/0005-2728\(86\)90038-1](https://doi.org/10.1016/0005-2728(86)90038-1)
- Okubo T, Tomo T, Sugiura M, Noguchi T (2007) Perturbation of the structure of P680 and the charge distribution on its radical cation in isolated reaction center complexes of photosystem II as revealed by fourier transform infrared spectroscopy. *Biochemistry* 46:4390–4397. doi:[10.1021/bi700157n](https://doi.org/10.1021/bi700157n)
- Pawlowicz NP, Groot ML, van Stokkum IH, Breton J, van Grondelle R (2007) Charge separation and energy transfer in the photosystem II core complex studied by femtosecond midinfrared spectroscopy. *Biophys J* 93:2732–2742. doi:[10.1529/biophysj.107.105452](https://doi.org/10.1529/biophysj.107.105452)
- Prokhorenko V, Holzwarth AR (2000) Primary processes and structure of the photosystem II reaction center: a photon echo study. *J Phys Chem B* 104:11563–11578. doi:[10.1021/jp002323n](https://doi.org/10.1021/jp002323n)
- Raszewski G, Renger T (2008) Light harvesting in photosystem II core complexes is limited by the transfer to the trap: can the core complex turn into a photoprotective mode? *J Am Chem Soc* 130:4431–4446. doi:[10.1021/ja7099826](https://doi.org/10.1021/ja7099826)
- Raszewski G, Saenger W, Renger T (2005) Theory of optical spectra of photosystem II reaction centers: location of the triplet state and the identity of the primary electron donor. *Biophys J* 88:986–998. doi:[10.1529/biophysj.104.050294](https://doi.org/10.1529/biophysj.104.050294)
- Raszewski G, Diner BA, Schlodder E, Renger T (2008) Spectroscopic properties of reaction center pigments in photosystem II core complexes: revision of the multimer model. *Biophys J* 95:105–119. doi:[10.1529/biophysj.107.123935](https://doi.org/10.1529/biophysj.107.123935)
- Rutherford AW, Mullet JE, Crofts AR (1981) Measurements of the midpoint potential of the pheophytin acceptor of photosystem II. *FEBS Lett* 123:235–237. doi:[10.1016/0014-5793\(81\)80295-5](https://doi.org/10.1016/0014-5793(81)80295-5)
- Shkuropatov AY, Khatypov RA, Shkuropatova VA, Zvereva MG, Owens TG, Shuvalov VA (1999) Reaction centers of photosystem II with a chemically-modified pigment composition: exchange of pheophytins with 13^1 -deoxo- 13^1 -hydroxy-pheophytin *a*. *FEBS Lett* 450:163–167. doi:[10.1016/S0014-5793\(99\)00486-X](https://doi.org/10.1016/S0014-5793(99)00486-X)
- Shuvalov VA, Klimov VV (1976) The primary photoreactions in the complex cytochrome-*P*-890 · *P*-760 bacteriopheophytin₇₆₀ of *Chromatium minutissimum* at low redox potentials. *Biochim Biophys Acta* 440:587–599. doi:[10.1016/0005-2728\(76\)90044-X](https://doi.org/10.1016/0005-2728(76)90044-X)
- Shuvalov VA, Yakovlev AG (1998) Energy level of P^+B^- relative to P^* , found by measurements of recombination fluorescence in plant-pheophytin-modified reaction centers from *Rhodospira sphaeroides* R-26. *Membr Cell Biol* 12:563–569 (in Russian)
- Shuvalov VA, Heber U, Schreiber U (1989) Low temperature photochemistry and spectral properties of a photosystem II reaction center complex D1/D2/containing the proteins D1 and D2 and two hemes of Cyt b-559. *FEBS Lett* 258:27–31. doi:[10.1016/0014-5793\(89\)81607-2](https://doi.org/10.1016/0014-5793(89)81607-2)
- Terenin AN (1967) Photonics of dye molecules. Nauka, Moscow
- Tetenkin VL, Gulyaev BA, Seibert M, Rubin AB (1989) Spectral properties of stabilized D1/D2/cytochrome b-559 photosystem II reaction center complex Effects of Triton X-100, the redox state of pheophytin, and β -carotene. *FEBS Lett* 250:459–463. doi:[10.1016/0014-5793\(89\)80776-8](https://doi.org/10.1016/0014-5793(89)80776-8)
- Ushakov EN, Nadochenko VA, Gromov SP, Vedernikov AI, Lobova NA, Alfimov MV, Gostev FE, Petrukhin AN, Sarkisov OM (2004) Ultrafast excited state dynamics of the bi- and termolecular stilbene-viologen charge transfer complexes assembled via host–guest interactions. *Chem Phys* 298:251–261. doi:[10.1016/j.chemphys.2003.12.002](https://doi.org/10.1016/j.chemphys.2003.12.002)
- Van Brederode ME, Jones MR, van Mourik F, van Stokkum IHM, van Grondelle R (1997) A new pathway for transmembrane electron transfer in photosynthetic reaction centers of *Rhodospira sphaeroides* not involving the excited special pair. *Biochemistry* 36:6855–6861. doi:[10.1021/bi9703756](https://doi.org/10.1021/bi9703756)
- Van Brederode ME, van Mourik F, van Stokkum IHM, Jones MR, van Grondelle R (1999) Multiple pathways for ultrafast transduction of light energy in the photosynthetic reaction center of *Rhodospira sphaeroides*. *Proc Natl Acad Sci USA* 96:2054–2059. doi:[10.1073/pnas.96.5.2054](https://doi.org/10.1073/pnas.96.5.2054)

- Van Leeuwen PJ, Nieveen MC, Van de Meent EJ, Dekker JP, Van Gorkom HJ (1991) Rapid and simple isolation of pure photosystem II core and reaction center particles from spinach. *Photosynth Res* 28:149–153. doi:[10.1007/BF00054128](https://doi.org/10.1007/BF00054128)
- Vulto SIE, Streltsov AM, AYa Shkuropatov, Shuvalov VA, Aartsma TJ (1997) Subpicosecond excited-state relaxation of the accessory bacteriochlorophylls in native and modified reaction centers of *Rhodobacter sphaeroides* R26. *J Phys Chem* 101:7249–7255