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# Evaluation of selective human MAO inhibitory activities of some novel pyrazoline derivatives

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**Abstract** A series of 1-[2-((5-methyl/chloro)-2-benzoxazolinone-3-yl)acetyl]-3,5-diaryl-4,5-dihydro-1*H*-pyrazole derivatives were prepared by reacting 2-((5-methyl/chloro)-2-benzoxazolinone-3-yl)acetylhydrazine with appropriate chalcones. The chemical structures of all compounds were confirmed by elemental analyses, IR, <sup>1</sup>H NMR and ESI-MS. All the compounds were investigated for their ability to selectively inhibit monoamine oxidase (MAO) by in vitro tests. MAO activities of the compounds were compared with moclobemide and selegiline and all the compounds were found to inhibit human MAO-A selectively. The inhibition profile was found to be competitive and reversible for all compounds by in vitro tests. Among the compounds examined, compounds **5ae**, **5af** and **5ag** were more selective than moclobemide, with respect to the *K<sub>i</sub>* values experimentally found. In addition, the compound **5bg** showed MAO-A inhibitor activity as well as moclobemide. A series of experimentally tested compounds (**5ae–5ch**) were docked computationally to the active site of the MAO-A and MAO-

B isoenzyme. The AUTODOCK 4.01 program was employed to perform automated molecular docking.

**Keywords** 2-Pyrazoline · 2-Benzoxazolinone · Chalcone · Monoamine oxidase inhibitory activity · Molecular docking

## Introduction

Human monoamine oxidases A and B (MAO-A and B) are the most intensively investigated flavin-dependent amine oxidases and play an important role in the control of intracellular concentration of monoaminergic neurotransmitters. The development of human MAO inhibitors led to important breakthroughs in the therapy of several neuropsychiatric disorders. MAO-A inhibitors are prescribed for the treatment of mental depression and anxiety (Yamada and Yasuhara 2004). MAO-B inhibitors are used with L-DOPA and/or dopamine agonists in the symptomatic treatment of Parkinson's disease (Drukarch and van Muijswinkel 2000; Schapira 2007).

Most current monoamine oxidase inhibitors lead to side effects by a lack of affinity and selectivity toward one of the isoforms. So, it remains fundamental to design new more potent, reversible and selective inhibitors of MAO-A and MAO-B.

Different families of heterocycles containing 2 or 4 nitrogen atoms have been used as scaffolds for synthesizing selective monoamine oxidase inhibitors, but the early period of the MAO-inhibitors started with hydrazine derivatives. Pyrazole, pyrazoline, and pyrazolidine derivatives can be considered as a cyclic hydrazine moiety. This scaffold also displayed promising antidepressant and anti-convulsant properties as demonstrated by different and

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established animal models. Diversely substituted pyrazoles, embedded with a variety of functional groups, are important biological agents and a significant amount of research activity has been directed toward this chemical class (Secci et al. 2011). On the basis of this observation, in previous communications, it was reported that N<sub>1</sub>-acetyl, N<sub>1</sub>-thiocarbamoyl, 1,3,5-triphenyl and 1-quinazolinone-3,5-diphenylpyrazolines exhibited high potency along with good selectivity due to their synthetic accessibility permitted a number of chemical changes (Bilgin et al. 1993; Palaska et al. 2001, 2008; Manna et al. 2002; Gökhan et al. 2003; Chimenti et al. 2004, 2005, 2006a, b, 2007, 2008a; Gökhan-Kelekçi et al. 2007, 2009; Özdemir et al. 2007, 2008). These observations motivated us to link different heterocyclic moieties to synthesize a new series of pyrazoline derivatives by combining the benzoxazolinone moiety at the first position in order to evaluate the effect of this substitution on monoamine oxidase inhibitory effects.

## Materials and methods

### Chemistry

All chemicals and solvents used in the present study were purchased from Merck A.G., Aldrich Chemical. Melting points of the compounds were determined with a Thomas Hoover Capillary Melting Point Apparatus and were uncorrected. Infrared (IR) spectra were obtained with a Perkin Elmer SpectrumOne, Nicolet 520 FT-IR spectrometer and the results were expressed in wave number (cm<sup>-1</sup>). <sup>1</sup>H NMR spectrums were recorded on a Bruker 400 MHz UltraShield spectrometer using dimethylsulfoxide (DMSO-*d*<sub>6</sub>) with chemical shifts reported as  $\delta$  (ppm) from TMS. Mass spectrums were undertaken using Waters 2695 Alliance Micromass ZQ LC/MS spectrometer in methanol according to the EI technique. Elemental analyses (C, H, N) were performed on an LECO CHNS 932 analyzer at the laboratory of Ankara University. The purity of the compounds was assessed by TLC on silicagel HF<sub>254+366</sub> (E.Merck, Darmstadt, Germany).

### General procedure for the preparation of 1,3-diaryl-2-propen-1-ones (4e–h) (chalcones)

Chalcone derivatives were synthesized by condensing acetophenone (10 mmol) and appropriate benzaldehydes (10 mmol) in the presence of sodium hydroxide (12.5 mmol) in water and ethanol (5/3 mL) at 0 °C for 1 h. The solid mass separated out was filtered, dried and crystallized from methanol (Dawey and Tivey 1958). 4e: m.p. 58–58.5 °C (Irie and Watanabe 1980; Lipson et al. 2005), 4f: m.p. 60–62 °C (Irie and Watanabe 1980), 4g: m.p.

75–76 °C (Ueno et al. 1983; Dong et al. 2008), 4h: m.p.: 135–137 °C (Sarabhai and Mathur 1963; Kubota et al. 2006).

### General procedure for the preparation of 1-[2-((5-methyl/chloro)-2-benzoxazolinone-3-yl)acetyl]-3,5-diaryl-4,5-dihydro-1H-pyrazoles (5)

2-((5-Methyl/chloro)-2-benzoxazolinone-3-yl)acetylhydrazine (1 mmol) was dissolved in 2 mL of DMF and 20 mL of *n*-propanol. 1,3-Diaryl-2-propen-1-one (1 mmol) and eight drops of hydrochloric acid was added to this solution and was refluxed for approximately 120 h (Gökhan-Kelekçi et al. 2009). The reaction mixture was then cooled and the solid precipitated was recrystallized. If solid was not precipitated, the solution was purified by chromatography on a silica gel column.

### Biochemistry

#### Chemicals

hMAO-A (recombinant, expressed in baculovirus infected BTI insect cells), hMAO-B (recombinant, expressed in baculovirus infected BTI insect cells), *R*-(–)-deprenyl hydrochloride, resorufin, dimethyl sulfoxide, and other chemicals were purchased from Sigma-Aldrich TM (Germany). Moclobemide was donated (Roche Pharmaceuticals, Germany). The Amplex<sup>®</sup>-Red MAO Assay Kit (Molecular Probes, USA) contained benzylamine, *p*-tyramine, Clorgyline (MAO-A inhibitor), Pargyline (MAO-B inhibitor), and horseradish peroxidase.

#### Determination of inhibitory activities of the compounds on human MAO-A and -B

The activity of hMAO-A and hMAO-B (using *p*-tyramine as common substrate for both isoforms) was found to be 185.60 ± 9.50 pmol/mg/min (*n* = 3). The interactions of the synthesized compounds with hMAO isoforms were determined by a fluorimetric method described and modified previously (Anderson et al. 1993; Yáñez et al. 2006; Chimenti et al. 2008b). The production of H<sub>2</sub>O<sub>2</sub> catalyzed by MAO isoforms was detected using 10-acetyl-3,7-dihydroxyphenoxazine (Amplex<sup>®</sup>-Red reagent), a non-fluorescent, highly sensitive, and stable probe that reacts with H<sub>2</sub>O<sub>2</sub> in the presence of horseradish peroxidase to produce the fluorescent product resorufin. The reaction was started by adding (final concentrations) 200  $\mu$ M Amplex Red reagent, 1 U/mL horseradish peroxidase, and *p*-tyramine (concentration range 0.1–1 mM).

Control experiments were carried out simultaneously by replacing the test drugs (novel pyrazoline derivatives and

reference inhibitors) with appropriate dilutions of the vehicles. In addition, the possible capacity of novel compounds to modify the fluorescence generated in the reaction mixture due to non-enzymatic inhibition (e.g., for directly reacting with Amplex Red reagent) was determined by adding these compounds to solutions containing only the Amplex Red reagent in a sodium phosphate buffer.

#### *Kinetic experiments*

Newly synthesized compounds were dissolved in dimethyl sulfoxide, with a maximum concentration of 1 %, and used in the final concentration range of 0.1–1,000 nM. Kinetic data for interaction of the enzyme with the compounds were determined using the Microsoft Excel package program. The slopes of the Lineweaver–Burk plots were plotted versus the inhibitor concentration and the  $K_i$  values were determined from the  $x$  axis intercept as  $-K_i$ . Each  $K_i$  value is the representative of single determination where the correlation coefficient ( $R^2$ ) of the replot of the slopes versus the inhibitor concentrations was at least 0.98. SI ( $K_i(\text{MAO-A})/K_i(\text{MAO-B})$ ) was also calculated. The protein was determined according to the Bradford method (Bradford 1976), in which bovine serum albumin was used as a standard.

#### *Reversibility experiments*

Reversibility of the MAO inhibition with novel derivatives was evaluated by a centrifugation-ultrafiltration method (Chimenti et al. 2010). In brief, adequate amounts of the recombinant hMAO-A or B were incubated together with a single concentration of the newly synthesized compounds or the reference inhibitors in a sodium phosphate buffer (0.05 M, pH 7.4) for 15 min at 37 °C. After this incubation period, an aliquot was stored at 4 °C and used for the measurement of MAO-A and -B activity. The remaining incubated sample was placed in an Ultrafree-0.5 centrifugal tube (Millipore, USA) with a 30 kDa Biomax membrane in the middle of the tube and centrifuged at 9,000× $g$  for 20 min at 4 °C. The enzyme retained in the 30 kDa membrane was resuspended in a sodium phosphate buffer at 4 °C and centrifuged again two successive times. After the third centrifugation, the enzyme retained in the membrane was resuspended in sodium phosphate buffer (300 mL) and an aliquot of this suspension was used for MAO-A and -B activity determination.

Control experiments were performed simultaneously (to define 100 % MAO activity) by replacing the test drugs with appropriate dilutions of the vehicles. The corresponding values of percent (%) MAO isoform inhibition were separately calculated for samples with and without repeated washing.

#### *Molecular docking studies*

The crystal structures of MAO-A and MAO-B were extracted from the protein data bank (PDB) [<http://www.rcsb.org>]. (for MAO-A pdb code: 2Z5X; human monoamine oxidase in complex with harmine, resolution 2.2 Å (Son et al. 2008) and for MAO-B pdb code: 2V5Z; human MAO-B in complex with inhibitor safinamide, resolution 1.6 Å (Binda et al. 2007)). Each structure was cleaned of all water molecules and inhibitors as well as all non-interacting ions before being used in the docking studies. The initial oxidized form of the FAD was used in all docking studies. For MAO-A and MAO-B, one of the two subunits was taken as the target structure. Using a fast Dreiding-like force field, each protein's geometry was first optimized and then submitted to the “Clean Geometry” toolkit of Discovery Studio (Accelrys, Inc.) for a more complete check. Missing hydrogen atoms were added based on the protonation state of the titratable residues at a pH of 7.4. Ionic strength was set to 0.145 and the dielectric constant was set to 10. The ADT (V. 1.5.4) (ADT) (Morris et al. 2009) graphical user interface program was employed to setup the enzymes for molecular docking.

#### *Ligand setups*

The 3D structures of ligand molecules were built, optimized at (PM3) level and saved in pdb format. The ADT package was also employed here to generate the docking input files of ligands. AutoDock 4.2 was used for all dockings; the detailed docking procedure has been given elsewhere (Yelekçi et al. 2007).

## **Results and discussion**

### *Chemistry*

A novel series of 1-[2-((5-methyl/chloro)-2-benzoxazolinone-3-yl)acetyl]-3,5-diaryl-4,5-dihydro-1*H*-pyrazole derivatives were synthesized and investigated for the ability to inhibit the activity of the A and B isoforms of human MAO. The synthesis pathway of the compounds was given in Scheme 1. 5-Methyl-2-benzoxazolinone **1b**, was synthesized as per the methods in the literature using 4-methyl-2-aminophenol and urea (Close et al. 1949). Treatment of (5-methyl/chloro)-2-benzoxazolinone with ethyl chloroacetate in  $\text{K}_2\text{CO}_3$ /acetone gave the N-alkylated product ethyl ((5-methyl/chloro)-2-benzoxazolinone-3-yl)acetate **2a–2c** (Milcent et al. 1996; Potts et al. 1980; Ünlü et al. 1992). The acid hydrazides **3a–3c** were prepared by the reaction of ethyl ((5-methyl/chloro)-2-benzoxazolinone-3-yl)acetate and hydrazine hydrate in ethanol

(Çakır et al. 2001; Gökçe et al. 2001; Önkol et al. 2008; Salgın-Gökşen et al. 2007). On the other hand,  $\alpha,\beta$ -unsaturated carbonyl compounds (chalcones) **4e–4h** were prepared by reacting appropriate aldehydes and acetophenone derivatives under basic condition according to the Claisen–Schmidt condensation (Dawey and Tivey 1958). The reaction of hydrazides **3a–3c** with chalcones **4e–4h** in *n*-propanol under acidic condition gave compounds 1-[2-((5-methyl/chloro)-2-benzoxazolinone-3-yl)acetyl]-3,5-diaryl-4,5-dihydro-1*H*-pyrazoles **5ae–5ch**.

The purity of the synthesized compounds was checked by elemental analyses and the results were within  $\pm 0.4$  % of the theoretical values. The structures of the synthesized compounds were determined on the basis of spectral data analysis; such as IR,  $^1\text{H}$  NMR and ESI–MS (Table 1).

Two C=O stretching bands viewed at  $1,789\text{--}1,753\text{ cm}^{-1}$  and  $1,679\text{--}1,664\text{ cm}^{-1}$  in the IR spectra of compounds **5ae–5ch**. The IR spectra of all the compounds showed C=C and C=N stretching bands at  $1,609\text{--}1,440\text{ cm}^{-1}$ .

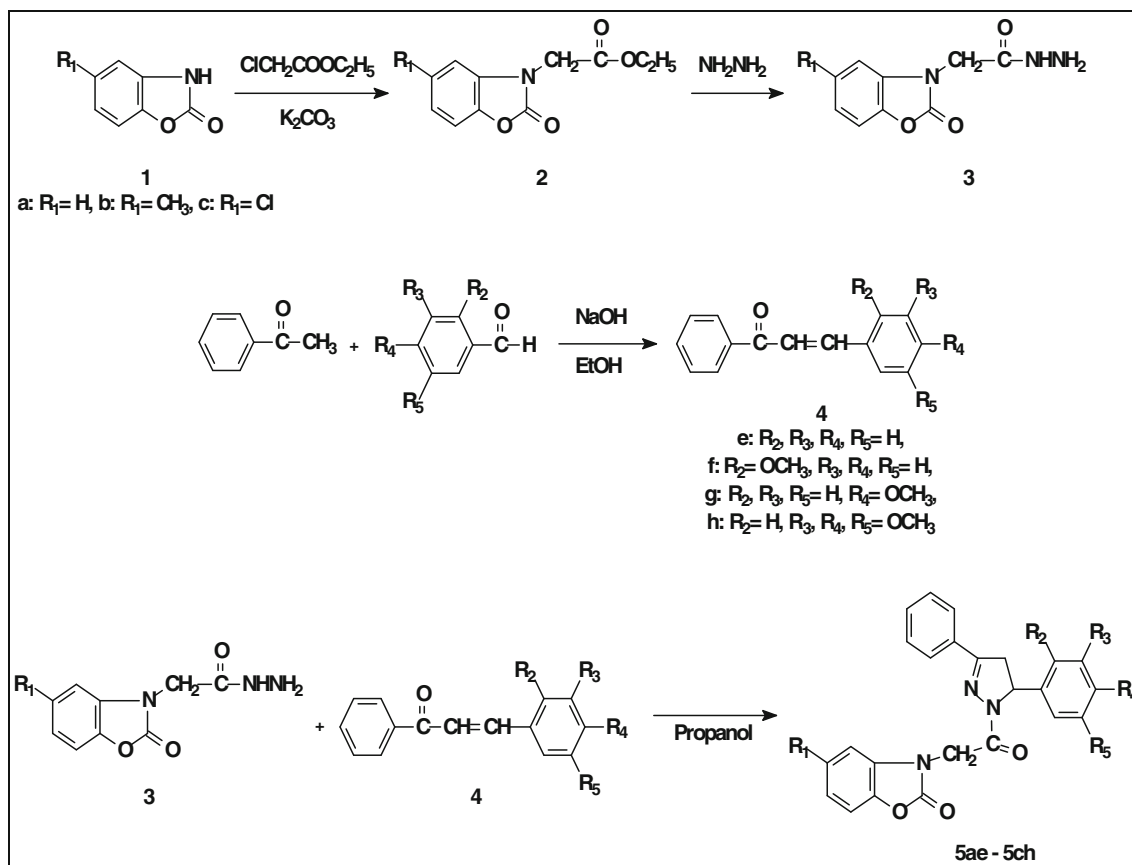
In the  $^1\text{H}$  NMR spectrum of the compounds **5ae–5ch**, it was observed three distinct doublet of doublets of the ABX system at  $\delta$  5.71–3.09 ppm due to pyrazoline ring (Shekarchi et al. 2008). The CH ( $\text{H}_\text{x}$ ) proton appeared between  $\delta$  5.71 and 5.52 ppm due to vicinal coupling with the two

magnetically non-equivalent protons of the methylene group at position 4 of the pyrazoline ring. The signals of  $\text{H}_\text{A}$  and  $\text{H}_\text{B}$  of pyrazoline ring were observed as doublet of doublets in the regions 3.95–3.88 ppm ( $\text{H}_\text{B}$ ) and 3.26–3.09 ppm ( $\text{H}_\text{A}$ ). The  $\text{CH}_2$  protons between the benzoxazolinone and pyrazoline ring resonated as a pair of doublet of doublets between  $\delta$  5.37–5.15 and 5.12–5.03 ppm. The signals for methoxy and methyl appeared at  $\delta$  3.80–3.59 ppm and  $\delta$  2.31–2.27 ppm, respectively (Holla et al. 2000; Chen et al. 2011).

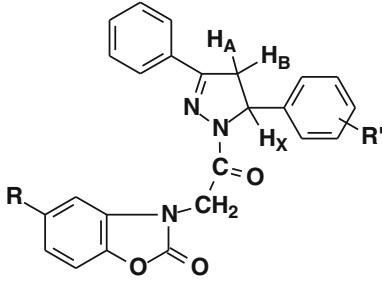
The characteristic peaks were observed in the mass spectra of the compounds. The ions produced under ESI showed a characteristic  $[\text{M} + \text{Na}]^+$  ion peak as the base signal for all compounds. Characteristic  $[\text{M} + \text{Na} + 2]^+$  isotope peaks were observed in the mass spectra of the compounds having chloride ion (compounds **5ce**, **5cf**, **5cg**, **5ch**).

### Biochemistry

MAO-A and MAO-B inhibitory activities of newly synthesized pyrazoline derivatives were determined using hMAO isoforms by a fluorimetric method. All the tested compounds were found to inhibit MAO-A selectively and



**Scheme 1** Synthesis of the compounds

**Table 1** Some characteristic and spectroscopic data of the synthesized compounds (**5ae–5ch**)


Compounds	Melting point (°C)	IR $\nu$ (cm <sup>-1</sup> )	<sup>1</sup> H NMR (DMSO-d <sub>6</sub> ) $\delta$ ppm ( <i>J</i> in Hz)	Mass <i>m/z</i>
<b>5ae</b>	192–194	3,057, 2,934 (C–H), 1,763, 1,673 (C=O), 1,486, 1,440 (C=C, C=N)	3.23 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.4 Hz, J <sub>AX</sub> :4.5 Hz), 3.95 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.6 Hz, J <sub>BX</sub> :11.8 Hz), 5.11 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.8 Hz), 5.25 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.7 Hz), 5.61 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.6 Hz, J <sub>AX</sub> :4.6 Hz), 7.13 (t, 1H, 2-benzox.–H <sub>5</sub> ), 7.18 (t, 1H, 2-benzox.–H <sub>6</sub> ), 7.24–7.28 (m, 4H, 2-benzox.–H <sub>4</sub> ve phenyl-3H), 7.32–7.37 (m, 3H, 2-benzox.–H <sub>7</sub> ve phenyl-2H), 7.51–7.52 (m, 3H, phenyl-3H), 7.87–7.88 (m, 2H, phenyl-2H)	436, 421, 420 (100 %), 398
<b>5af</b>	198–200	2,934, 2,838 (C–H), 1,777, 1,673 (C=O), 1,599, 1,489, 1,440 (C=C, C=N)	3.09 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.0 Hz, J <sub>AX</sub> :4.6 Hz), 3.80 (s, 3H, –OCH <sub>3</sub> ), 3.90 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.0 Hz, J <sub>BX</sub> :11.8 Hz), 5.10 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.7 Hz), 5.28 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.7 Hz), 5.71 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.8 Hz, J <sub>AX</sub> :4.6 Hz), 6.89 (t, 1H, phenyl-H), 7.04 (t, 2H, phenyl-2H), 7.13 (t, 1H, 2-benzox.–H <sub>5</sub> ), 7.19 (t, 1H, 2-benzox.–H <sub>6</sub> ), 7.25 (d, 1H, 2-benzox.–H <sub>4</sub> , J:7.7 Hz), 7.28 (d, 1H, phenyl-H, J:7.6 Hz), 7.36 (d, 1H, 2-benzox.–H <sub>7</sub> , J:7.8 Hz), 7.47–7.51 (m, 3H, phenyl-3H), 7.84–7.86 (m, 2H, phenyl-2H)	466, 451, 450 (100 %), 428
<b>5ag</b>	214–215	2,957, 2,941, 2,828 (C–H), 1,779, 1,669 (C=O), 1,603, 1,516, 1,490, 1,447 (C=C, C=N)	3.22 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.4 Hz, J <sub>AX</sub> :4.8 Hz), 3.72 (s, 3H, –OCH <sub>3</sub> ), 3.90 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.4 Hz, J <sub>BX</sub> :11.6 Hz), 5.08 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:18.0 Hz), 5.21 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.54 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.6 Hz, J <sub>AX</sub> :4.8 Hz), 6.88 (d, 2H, 4-methoxyphenyl-2H, J:8.8 Hz), 7.10–7.19 (m, 4H, 2-benzox.–H <sub>5</sub> , 2-benzox.–H <sub>6</sub> ve 4-methoxyphenyl-2H), 7.25 (d, 1H, 2-benzox.–H <sub>4</sub> , J:7.6 Hz), 7.35 (d, 1H, 2-benzox.–H <sub>7</sub> , J:7.2 Hz), 7.50–7.52 (m, 3H, phenyl-3H), 7.86–7.88 (m, 2H, phenyl-2H)	466, 451, 450 (100 %), 428
<b>5ah</b>	236.5–237.5	2,997, 2,941, 2,825 (C–H), 1,766, 1,679 (C=O), 1,590, 1,457, 1,443 (C=C, C=N)	3.26 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.2 Hz, J <sub>AX</sub> :5.2 Hz), 3.62 (s, 3H, –OCH <sub>3</sub> ), 3.75 (s, 6H, –OCH <sub>3</sub> ), 3.91 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.3 Hz, J <sub>BX</sub> :11.9 Hz), 5.12 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.34 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.7 Hz), 5.55 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.8 Hz, J <sub>AX</sub> :5.1 Hz), 6.52 (s, 2H, 3,4,5-trimethoxyphenyl-2H), 7.14 (t, 1H, 2-benzox.–H <sub>5</sub> ), 7.19 (t, 1H, 2-benzox.–H <sub>6</sub> ), 7.32 (d, 1H, 2-benzox.–H <sub>4</sub> , J:7.6 Hz), 7.37 (d, 1H, 2-benzox.–H <sub>7</sub> , J:7.8 Hz), 7.51–7.52 (m, 3H, phenyl-3H), 7.85–7.87 (m, 2H, phenyl-2H)	526, 511, 510 (100 %), 488, 320, 176

**Table 1** continued

Compounds	Melting point (°C)	IR $\nu$ (cm <sup>-1</sup> )	<sup>1</sup> H NMR (DMSO-d <sub>6</sub> ) $\delta$ ppm ( <i>J</i> in Hz)	Mass <i>m/z</i>
<b>5be</b>	201.5–202.5	3,063, 2,925 (C–H), 1,755, 1,677 (C=O), 1,499, 1,441 (C=C, C=N)	2.30 (s, 3H, –CH <sub>3</sub> ), 3.23 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.2 Hz, J <sub>AX</sub> :4.8 Hz), 3.94 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.0 Hz, J <sub>BX</sub> :11.6 Hz), 5.05 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:18.0 Hz), 5.19 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.60 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.8 Hz, J <sub>AX</sub> :4.8 Hz), 6.92 (d, 1H, 2-benzox.–H <sub>6</sub> , J <sub>67</sub> :8.4 Hz), 7.06 (s, 1H, 2-benzox.–H <sub>4</sub> ), 7.21 (d, 1H, 2-benzox.–H <sub>7</sub> , J <sub>67</sub> :8.0 Hz), 7.23–7.28 (m, 3H, phenyl-3H), 7.32–7.35 (m, 2H, phenyl-2H), 7.49–7.52 (m, 3H, phenyl-3H), 7.86–7.88 (m, 2H, phenyl-2H)	450, 435, 434 (100 %), 412
<b>5bf</b>	206–207	3,472 (O–H), 3,055, 2,913, 2,834 (C–H), 1,753, 1,675 (C=O), 1,597, 1,499, 1,443 (C=C, C=N)	2.31 (s, 3H, –CH <sub>3</sub> ), 3.09 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.0 Hz, J <sub>AX</sub> :4.7 Hz), 3.79 (s, 3H, –OCH <sub>3</sub> ), 3.89 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.0 Hz, J <sub>BX</sub> :11.8 Hz), 5.04 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.27 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.71 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.7 Hz, J <sub>AX</sub> :4.6 Hz), 6.89 (t, 1H, phenyl-H), 6.93 (d, 1H, 2-benzox.–H <sub>6</sub> , J <sub>67</sub> :8.16 Hz), 7.03–7.06 (m, 2H, phenyl-2H), 7.08 (s, 1H, 2-benzox.–H <sub>4</sub> ), 7.22 (d, 1H, 2-benzox.–H <sub>7</sub> , J <sub>67</sub> :8.12 Hz), 7.26 (t, 1H, phenyl-H), 7.47–7.51 (m, 3H, phenyl-3H), 7.84–7.86 (m, 2H, phenyl-2H)	481, 480, 465, 464 (100 %), 443, 442
<b>5bg</b>	172–173	3,074, 2,952, 2,925, 2,830 (C–H), 1,776, 1,674 (C=O), 1,515, 1,495, 1,441 (C=C, C=N)	2.30 (s, 3H, –CH <sub>3</sub> ), 3.22 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.4 Hz, J <sub>AX</sub> :4.8 Hz), 3.72 (s, 3H, –OCH <sub>3</sub> ), 3.90 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.0 Hz, J <sub>BX</sub> :11.6 Hz), 5.03 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:18.0 Hz), 5.15 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.54 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.4 Hz, J <sub>AX</sub> :4.8 Hz), 6.88 (d, 2H, 4-methoxyphenyl-2H, J:8.8 Hz), 6.92 (d, 1H, 2-benzox.–H <sub>6</sub> , J <sub>67</sub> :8.4 Hz), 7.04 (s, 1H, 2-benzox.–H <sub>4</sub> ), 7.16 (d, 2H, 4-methoxyphenyl-2H, J:8.8 Hz), 7.21 (d, 1H, 2-benzox.–H <sub>7</sub> , J <sub>67</sub> :8.4 Hz), 7.50–7.52 (m, 3H, phenyl-3H), 7.86–7.88 (m, 2H, phenyl-2H)	480, 465, 464 (100 %), 442
<b>5bh</b>	237–238	2,929, 2,834 (C–H), 1,766, 1,673 (C=O), 1,609, 1,503, 1,436 (C=C, C=N)	2.27 (s, 3H, –CH <sub>3</sub> ), 3.23 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.6 Hz, J <sub>AX</sub> :5.2 Hz), 3.59 (s, 3H, –OCH <sub>3</sub> ), 3.72 (s, 6H, –OCH <sub>3</sub> ), 3.88 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.2 Hz, J <sub>BX</sub> :11.6 Hz), 5.03 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.27 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:18.0 Hz), 5.52 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.6 Hz, J <sub>AX</sub> :5.2 Hz), 6.49 (s, 2H, 3,4,5- trimethoxyphenyl-2H), 6.91 (d, 1H, 2-benzox.–H <sub>6</sub> , J <sub>67</sub> :8.0 Hz), 7.09 (s, 1H, 2-benzox.–H <sub>4</sub> ), 7.20 (d, 1H, 2-benzox.–H <sub>7</sub> , J <sub>67</sub> :8.4 Hz), 7.47–7.49 (m, 3H, phenyl-3H), 7.82–7.85 (m, 2H, phenyl-2H)	540, 525, 524 (100 %), 502
<b>5ce</b>	146–148	3,055, 2,929 (C–H), 1,755, 1,668 (C=O), 1,487, 1,440 (C=C, C=N)	3.24 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.4 Hz, J <sub>AX</sub> :4.8 Hz), 3.95 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.2 Hz, J <sub>BX</sub> :11.6 Hz), 5.12 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:18.0 Hz), 5.27 (d, 1H, N–CH <sub>2</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.61 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.8 Hz, J <sub>AX</sub> :4.8 Hz), 7.18 (dd, 1H, chlorzox.–H <sub>6</sub> , J <sub>67</sub> :8.6 Hz, J <sub>46</sub> :2.0 Hz), 7.25–7.29 (m, 3H, phenyl-3H), 7.33–7.36 (m, 2H, phenyl-2H), 7.40 (d, 1H, chlorzox.–H <sub>7</sub> , J <sub>67</sub> :8.4 Hz), 7.51–7.53 (m, 4H, chlorzox.–H <sub>4</sub> ve phenyl-3H), 7.86–7.89 (m, 2H, phenyl-2H)	470, 457, 456, 455, 454 (100 %), 434, 432



**Table 1** continued

Compounds	Melting point (°C)	IR $\nu$ (cm <sup>-1</sup> )	<sup>1</sup> H NMR (DMSO-d <sub>6</sub> ) $\delta$ ppm ( <i>J</i> in Hz)	Mass <i>m/z</i>
<b>5cf</b>	176–177	3,059, 2,948, 2,842 (C–H), 1,767, 1,672 (C=O), 1,487, 1,455, 1,440 (C=C, C=N)	3.09 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.1 Hz, J <sub>AX</sub> :4.6 Hz), 3.80 (s, 3H, –OCH <sub>3</sub> ), 3.89 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :17.9 Hz, J <sub>BX</sub> :11.7 Hz), 5.10 (d, 1H, N–CH <sub>1</sub> H <sub>2</sub> –CO, J:17.7 Hz), 5.29 (d, 1H, N–CH <sub>1</sub> H <sub>2</sub> –CO, J:17.7 Hz), 5.71 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.7 Hz, J <sub>AX</sub> :4.6 Hz), 6.89 (t, 1H, phenyl-H), 7.05 (t, 2H, phenyl-2H), 7.18 (dd, 1H, chlorzox.–H <sub>6</sub> , J <sub>67</sub> :8.5 Hz, J <sub>46</sub> :2.1 Hz), 7.26 (t, 1H, phenyl-1H), 7.39 (d, 1H, chlorzox.–H <sub>7</sub> , J <sub>67</sub> :8.5 Hz), 7.49–7.50 (m, 4H, chlorzox.–H <sub>4</sub> ve phenyl-3H), 7.84–7.86 (m, 2H, phenyl-2H)	487, 486, 485, 484 (100 %), 462, 354
<b>5cg<sup>a</sup></b>	–	–	–	–
<b>5ch</b>	262–263	3,063, 2,944, 2,822 (C–H), 1,771, 1,664 (C=O), 1,593, 1,491, 1,440 (C=C, C=N)	3.26 (dd, 1H, H <sub>A</sub> , J <sub>AB</sub> :18.2 Hz, J <sub>AX</sub> :5.2 Hz), 3.61 (s, 3H, –OCH <sub>3</sub> ), 3.75 (s, 6H, –OCH <sub>3</sub> ), 3.91 (dd, 1H, H <sub>B</sub> , J <sub>AB</sub> :18.2 Hz, J <sub>BX</sub> :11.6 Hz), 5.11 (d, 1H, N–CH <sub>1</sub> H <sub>2</sub> –CO, J:17.6 Hz), 5.37 (d, 1H, N–CH <sub>1</sub> H <sub>2</sub> –CO, J:18.0 Hz), 5.55 (dd, 1H, H <sub>X</sub> , J <sub>BX</sub> :11.8 Hz, J <sub>AX</sub> :5.2 Hz), 6.53 (s, 2H, 3,4,5-trimethoxyphenyl-2H), 7.19 (dd, 1H, chlorzox.–H <sub>6</sub> , J <sub>67</sub> :8.4 Hz, J <sub>46</sub> :2.0 Hz), 7.41 (d, 1H, chlorzox.–H <sub>7</sub> , J <sub>67</sub> :8.0 Hz), 7.50–7.52 (m, 3H, chlorzox.–H <sub>4</sub> ve phenyl-2H), 7.55 (d, 1H, phenyl-H, J:2.4 Hz), 7.85–7.87 (m, 2H, phenyl-2H)	562, 560, 547, 546, 545, 544 (100 %), 522, 182

<sup>a</sup> Şahin et al. 2011

competitively (Table 2). These novel compounds were reversible inhibitors of hMAO-A since the enzyme activity was restored after centrifugation-ultrafiltration steps (Table 2).

Except compounds with **h** substitution (trimethoxy) in phenyl ring, all the compounds were found to be a potent MAO-A inhibitors with  $K_i$  values in nM range and with  $SI_{MAO-A}$  in the magnitude of  $10^3$ – $10^4$ . Compounds **5ae**, which is unsubstituted, and **5af**, which has a methoxy substitution on R<sub>2</sub> position were appeared as the most potent MAO-A inhibitors within this series with  $K_i$  values of  $0.003 \pm 10^{-5}$  and  $0.010 \pm 10^{-3}$   $\mu$ M, respectively. Docking results given in Table 2 are in agreement with the biochemical evaluations. The high inhibitory potency and selectivity of **5ae** through hMAO-A were discussed in detail in the next part according to the computational data obtained.

Compound **5bg**, which carries a methyl group at R<sub>1</sub> position of benzoxazolinone ring and a methoxy group at para position of phenyl ring inhibited hMAO-A with  $K_i$  value of  $0.090 \pm 10^{-3}$  (Table 2). Experimental selectivity index for this compound was found as 0.004, which is satisfactory and comparable with  $SI_{MAO-A}$  of known MAO-A inhibitor; moclobemide (0.004).

It was suggested that in case that the benzoxazolinone ring is unsubstituted or substituted with methyl group (**a** or **b** substitution), MAO-A inhibitory activity is better compared to chloride substitution (**c** substitution), except in the

case of compound **5be**. Furthermore, trimethoxy substitution in phenyl ring (**h** substitution) has been found unfavorable in terms of MAO-A inhibitory activity. Among compounds that benzoxazolinone ring is unsubstituted or substituted with chloride group (**a** or **c** substitution), compounds carrying **e** substitution was found the most potent MAO-A inhibitor among the substitutions of e, f and g.

In the present study, we have successfully identified new compounds which are reversible and selective inhibitor of hMAO-A. It was suggested that unsubstituted benzoxazolinone ring favors MAO-A inhibitory activity whereas methoxy substitutions of phenyl ring at meta and para positions reveals a significant decrease in MAO-A inhibition activity. Results of this study will provide a useful information for designing a new series of potent, selective and reversible MAO-A inhibitors in future.

#### Molecular docking studies

To figure out the detailed interactions of the docked poses of the inhibitors, compound **5ae** was selected for visualization. The binding modes for inhibitor **5ea** (Fig. 1) in the MAO-A and MAO-B active site cavities are shown in below images. A careful analysis of the binding mode of the compound **5ea** in the MAO-A cavity revealed that the benzoxazolinone ring of this compound inserted into the hydrophobic pocket lined with the TYR444, TYR407 and FAD cofactor. Two phenyl rings of inhibitor **5ae** make two



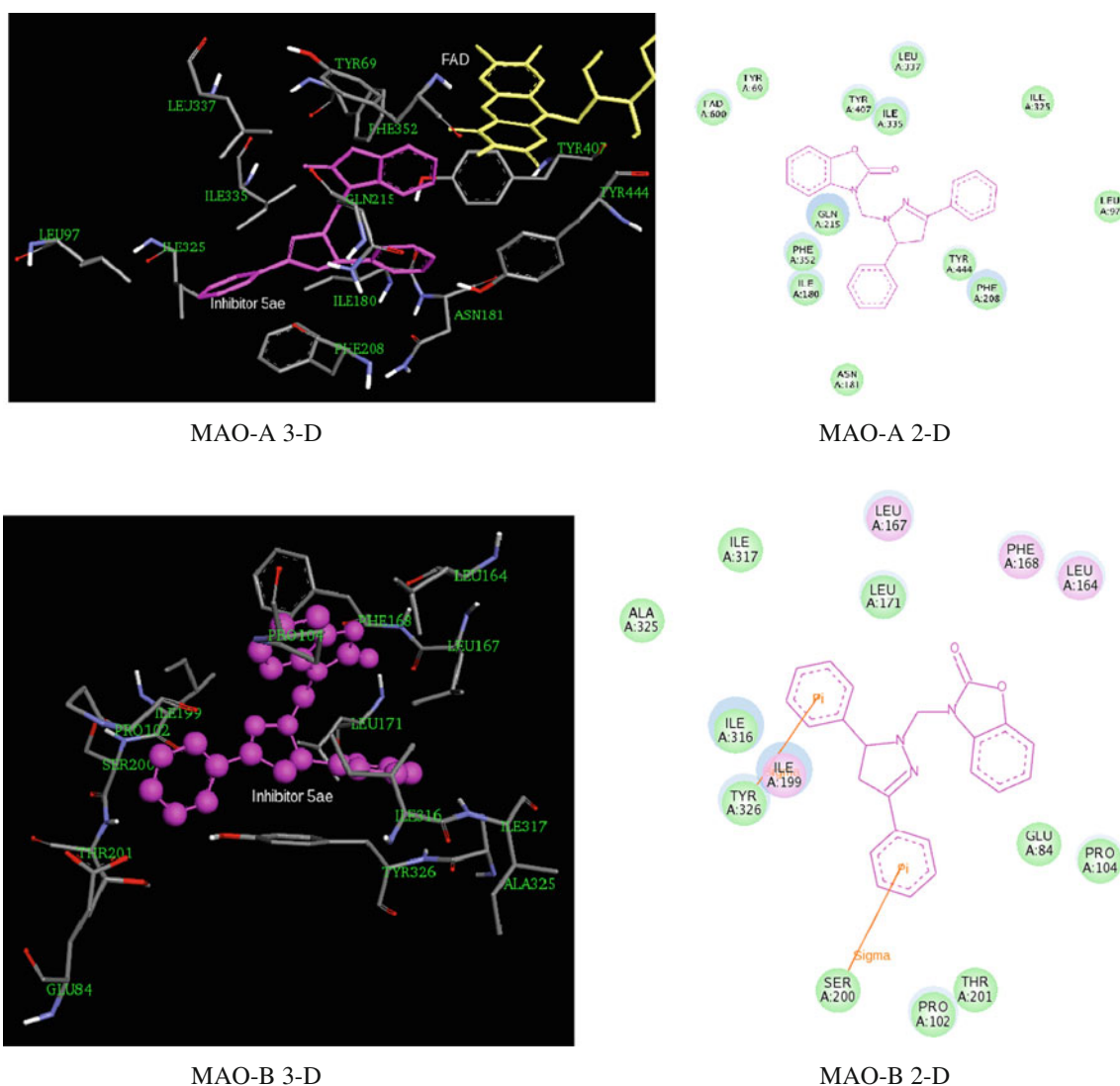
**Table 2** Calculated and experimental  $K_i$  values corresponding to the inhibition of MAO isoforms by the newly synthesized 2-pyrazoline derivatives

Compounds	Calculated $K_i$ value for MAO-A ( $\mu\text{M}$ )	Calculated $K_i$ value for MAO-B ( $\mu\text{M}$ )	Calculated SI*	Experimental $K_i$ value for MAO-A ( $\mu\text{M}$ )**	Experimental $K_i$ value for MAO-B ( $\mu\text{M}$ )**	Experimental SI*	Inhibition type, selectivity, reversibility
<b>5ae (R)</b>	0.001	1.20	0.000833	$0.003 \pm 0.00001$	$1.80 \pm 0.13$	0.002	MAO-A, competitive, reversible
<b>5ae (S)</b>	0.001	1.97	0.000508				
<b>5af (R)</b>	0.009	3.69	0.002	$0.010 \pm 0.001$	$3.80 \pm 0.17$	0.003	MAO-A, competitive, reversible
<b>5af (S)</b>	0.007	5.82	0.001				
<b>5ag (R)</b>	0.00093	11.29	0.0000823	$0.050 \pm 0.002$	$32.00 \pm 1.60$	0.002	MAO-A, competitive, reversible
<b>5ag (S)</b>	0.031	55.15	0.000562				
<b>5ah (R)</b>	24.32	63.85	0.381	$45.260 \pm 1.250$	$590.00 \pm 15.21$	0.076	MAO-A, competitive, reversible
<b>5ah (S)</b>	38.81	566.75	0.068				
<b>5be (R)</b>	0.003	3.74	0.000802	$0.100 \pm 0.009$	$3.00 \pm 0.01$	0.03	MAO-A, competitive, reversible
<b>5be (S)</b>	0.133	3.29	0.040				
<b>5bf (R)</b>	0.070	9.74	0.007	$0.080 \pm 0.002$	$1.00 \pm 0.09$	0.080	MAO-A, competitive, reversible
<b>5bf (S)</b>	0.070	0.70	0.1				
<b>5bg (R)</b>	0.002	12.71	0.000157	$0.090 \pm 0.001$	$23.10 \pm 1.60$	0.004	MAO-A, competitive, reversible
<b>5bg (S)</b>	0.044	19.94	0.002				
<b>5bh (R)</b>	31.61	254.54	0.124	$15.20 \pm 1.05$	$153.00 \pm 8.06$	0.099	MAO-A, competitive, reversible
<b>5bh (S)</b>	461.06	58.55	7.875				
<b>5ce (R)</b>	0.014	2.37	0.006	$0.050 \pm 0.002$	$1.90 \pm 0.009$	0.027	MAO-A, competitive, reversible
<b>5ce (S)</b>	0.063	1.15	0.055				
<b>5cf (R)</b>	0.054	29.39	0.002	$0.095 \pm 0.002$	$7.00 \pm 0.23$	0.014	MAO-A, competitive, reversible
<b>5cf (S)</b>	0.034	0.298	0.114				
<b>5cg (R)</b>	0.009	18.39	0.000489	$0.120 \pm 0.015$	$7.00 \pm 2.11$	0.017	MAO-A, competitive, reversible
<b>5cg (S)</b>	0.548	10.53	0.052				
<b>5ch (R)</b>	8.980	755.08	0.012	$9.26 \pm 0.35$	$805.20 \pm 36.45$	0.011	MAO-A, competitive, reversible
<b>5ch (S)</b>	62.189	292.00	0.213				
<b>Selegiline</b> (MAO-B inhibitor)	22.02	34.07	0.646	$9.06 \pm 0.44$	$0.09 \pm 0.004$	100.67	MAO-B, competitive irreversible
<b>Moclobemide</b> (MAO-A inhibitor)	5.71	250.74	0.023	$0.005 \pm 0.001$	$1.22 \pm 0.08$	0.004	MAO-A, competitive, reversible

\* Selectivity index. It was calculated as  $K_i(\text{MAO-A})/K_i(\text{MAO-B})$ \*\* Each value represents the mean  $\pm$  SEM of three independent experiments

significant  $\sigma$ - $\pi$  interactions with the side chains of PHE352 and PHE208. ASN181, ILE325, LEU97, GLN215, ILE335, LEU337, and TYR69 contribute to the other attractions. The last two pictures of Fig. 1 show the poses of **5ae** in the active side of MAO-B in 3-D and 2-D depictions, respectively. On the contrary, MAO-A compound **5ae** occupies a space in the entrance cavity of MAO-B very far from the

main cavity and hydrophobic packet. The phenyl rings of **5ae** make two  $\sigma$ - $\pi$  interactions with SER200 and TYR326. The selectivity and potency of compound **5ae** on MAO-A compared to MAO-B can be noted in the above poses in MAO-A and MAO-B. The experimental data given in Table 2 are in agreement with these observations. All the computational results may suggest why the MAO-A



**Fig. 1** Docked pose of compound **5ae** (**R**) in MAO-A and MAO-B active site in 3-D and 2-D, respectively. Amino acid side chains are shown as *sticks*, the inhibitor is shown as a *ball and stick* (magenta), and the cofactor FAD is depicted as a *yellow stick*. Residues involved

in hydrogen bonding or polar interactions are represented by *magenta-colored circles*, and residues involved in vdW and hydrophobic interactions are shown by *green circles* in all 2-D figures

inhibitory potency of inhibitor **5ae** ( $K_i = 0.001 \mu\text{M}$ ) is much better and more selective in comparison to MAO-A ( $K_i = 1.20 \mu\text{M}$ ).

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