

High pressure CO₂-controlled reactors: enzymatic chiral resolution in emulsions

 Cite this: *RSC Adv.*, 2014, 4, 24083

Wenting Shang, Xiaogang Zhang,* Xiaoxi Yang and Shujuan Zhang

In this work we have reported the formulation of a CO₂-based micelle stabilized by nontoxic TMN series surfactants. Enantioselection of racemic ibuprofen catalyzed by *Candida antarctica* lipase B (CALB) was used as a model reaction. The effect of reactive parameters, such as temperature, pH, pressure, and water content on reactive environment and conversion has been discussed. For the resolution of racemic ibuprofen in CO₂-based micelles, the enzymatic activity reached a high level at 45 °C, with pressure 250 bar, pH 7.4, and water to surfactant ratio *W*₀ 25. In addition, the relatively long-chain length in TMN-10 could help the esterification and trans-esterification processes, which resulted in an efficient reaction rate in a CO₂-based micelle system. Enzymatic catalysis has been conducted in a CO₂-based system rather than in the conventional media to make the enzyme reaction greener. The better resolution efficiency in high pressure CO₂-based micelles could be achieved within a relatively short period of time compared with other traditional reactive systems.

 Received 11th March 2014
Accepted 20th May 2014

DOI: 10.1039/c4ra02131b

www.rsc.org/advances

1 Introduction

With the growing demand for environmentally friendly processes working at ambient conditions, the use of biocatalysts in organic synthesis has become an interesting alternative to conventional chemical methods.^{1,2} Enzymes are eco-friendly biocatalysts that can catalyze chemical reactions under mild conditions, such as ambient temperature, pressure, and neutral pH.^{3,4} The use of enzymes generally circumvents the need for functional group activation and avoids protection and deprotection steps required in traditional organic syntheses.⁵

From a biotechnological viewpoint, conducting enzymatic reactions in nonaqueous solvents offer new possibilities for producing valuable chemicals. Their versatility would be further expended if the enzymatic reactions could be performed in reverse micelles. Reverse micelles are thermodynamically stable water droplets dispersed in an organic phase by means of a surfactant. One of the most important properties of reverse micelles is their ability to entrap enzymes and other biomolecules in their water droplets. In addition, the micelles are dynamic structures, the substrates and products can be exchanged between water droplets and bulk organic solvent. Reverse micelles have wide applications in a variety of fields including chemical reactions,^{6–8} material synthesis,^{9–11} protein delivery,^{12–14} drug release¹⁵ and so on. A reversed micellar system is especially suitable for lipases, which are activated in the presence of a water–oil interface. These systems can provide a

high interfacial area, with the enzyme anchoring on the aqueous side of the surfactant interface.

The water content which is necessary to favour synthesis reactions in organic media can be achieved by micro-encapsulation of the biocatalyst within reverse micelles. For reverse micelle system, its challenge is recovering the product from reverse micelles at large scale due to the presence of the surfactant and other components of the system, making the separation and purification of the product be more difficult, especially when it is to be used in foods, pharmaceuticals or other products that require either non-toxic or highly pure products.

High pressure carbon dioxide has been identified as a ‘green’ solvent with its potential applications for industrial use as it provides a clean, non-toxic, non-flammable and tunable solvent system which is easily removed to leave reaction products free from undesirable organic residues. Therefore, the combination of biocatalysis and carbon dioxide is extremely attractive. Enzymatic catalysis has been conducted in CO₂-based system rather than in the conventional media to make enzyme reaction greener.

The first report on enzyme catalyzed reactions in high pressure CO₂ was in 1985 by Randolph *et al.*,¹⁶ Hammond *et al.*,¹⁷ and in 1986 by Nakamura *et al.*¹⁸ Recently, a number of groups have explored the use of enzymes in high pressure CO₂.^{19–24} However, it was reported that native enzymes were easily deactivated in CO₂-based medium because CO₂ molecules could react with ε-amino groups of lysine residues placed on the enzyme surface to form inhibitory carbamates.²⁵ One attempt to prevent this phenomenon has been made by using reverse micelles to stabilize the enzyme in a water pool, and meantime maintain the benefit of high mass diffusivity of high pressure CO₂. For the CO₂-based systems, their applications are probably

Department of Chemistry, Renmin University of China, Beijing, 100872, P.R. China.
E-mail: zhang_xg@ruc.edu.cn; Fax: +86-10-62515601; Tel: +86-10-62515601

hindered by the weak solubility for many hydrophiles and proteins of interest.²⁶ Recently, many of amphiphiles have been shown to form micelle in carbon dioxide media. For general case, the reactive rate in high pressure, especially supercritical fluid system is faster than that performed at atmospheric pressure because of the reduced mass-transfer.^{27,28} The extra advantages associated with use of CO₂ as media for enzymatic reactions are not only related to its low environmental impact but also due to its chemical properties such as high diffusivity, tunable solubility with CO₂ density, and easy separation of solvent from products.

There is an increasing trend towards the use of optically pure enantiomers for drugs and agrochemicals because they are more effective and have fewer side effects compared with their racemic mixtures. Ibuprofen, 2-(4-isobutylphenyl)propionic acid, is a racemic carboxylic acid, is a widely used nonsteroidal anti-inflammatory drug that belongs to the family of 2-arylpropionic acid derivatives. Due to its high medical activity and low toxicity, ibuprofen is one of the most popular non-prescription medicines in the world. The pharmaceutical activity of ibuprofen is dependent on the chirality of the compound. Only the *S*-isomer exhibits an anti-inflammatory property and is valuable.²⁹ Enzyme kinetic separation was reported as one of the best ways to obtain pure (*S*)-ibuprofen.³⁰ Utilization of lipase in the resolution of ibuprofen received great interests because of the mild reaction condition, high enantioselectivity, less side reactions and environment pollution of enzyme-catalyzed reactions. However, the biological catalytic processes for chiral separation often taken a very long reaction time, and stability could not be a good guarantee, recycling and reuse were also encountered many challenges.

In this paper, the CO₂-based micelles with the TMN series surfactants, which were related to poly(ethylene glycol)-2,6,8-trimethyl-4-nonyl ethers, were used as reactive medium. Enantioselection of racemic ibuprofen catalyzed by *Candida antarctica* lipase B (CALB) was used as a model reaction. The better resolution efficiency in high pressure CO₂-based micelles could be achieved within a relatively shorter period of time comparing with other reactive systems. The effect of reactive parameters, such as temperature, pH, pressure, and water content on reactive environment and conversion has been discussed.

2 Materials and method

2.1 Materials

Racemic ibuprofen was supplied by Xian Lang Hong Biotechnology Co. Ltd. Propanol was supplied by Sinopharm Chemical Reagent Co., Ltd. The standard reference of (*S*)- and (*R*)-ibuprofen were purchased from Sigma. Ibuprofen propyl ester was synthesized using the method of Patricia de *et al.*³¹ in our lab. Racemic(*R,S*)-ibuprofen and propanol were dropped into a round-bottom flask containing hydrochloric acid as the catalyst and the solution was refluxed for 6 hours. After the reaction, ibuprofen propyl ester was leached by adding right amount of *n*-hexane. From the reaction mixture ibuprofen propyl ester was extracted and purified by repeated extraction with *n*-hexane; ibuprofen could be detected in the product as tested

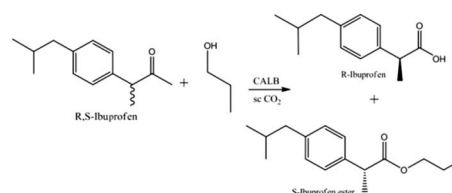
by thin-layer chromatography (TLC). *Candida antarctica* lipase B (CALB) (1000 units per mg solid) was purchased from Sigma (China). CALB was used as received. Non-ionic surfactants Tergitol®TMN-3 (99% 5b-C₁₂E₃, $n = 2.98$, $M_w = 335$), Tergitol®TMN-6 (90% 5b-C₁₂E₈, 10 wt% water, $n = 8.32$, $M_w = 552$), Tergitol®TMN-10 (90% 5b-C₁₂E₁₂, 10 wt% water, $n = 11.55$, $M_w = 694$) were purchased from Sigma-Aldrich and were used as received. The chemicals were of G.R. grade and used without further purification. Carbon dioxide (99.995% purity) was purchased from Beijing Analytical Instruments Inc. Double distilled water was used throughout.

2.2 Enzymatic dynamic kinetic resolution in CO₂-based micelle system

The apparatus in which the chiral resolution of racemic ibuprofen catalyzed by CALB (shown in Scheme 1) in CO₂-based micelle was carried out was shown in Fig. 1. In typical experiments, racemic ibuprofen (20.6 mg), propanol (18 μ l), sodium phosphate buffer (25 μ l, pH = 7.4) mixed with surfactant TMN-10 (35 μ l) and 3 mg CALB solution were loaded in a high-pressure cell with a working volume of 8 ml. The cell was sealed and then heated to 45 °C with stirring. CO₂ was introduced slowly into the cell until to desired pressure. The mixture was stirred at 600 rpm during the reaction, carried out for 36 hours. After reaction, the cell was cooled to 0 °C and the gas stream was then vented slowly to ambient pressure through the traps containing 10 ml *n*-hexane. Thereafter, the cell was opened, and the remaining residue was extracted with another 20 ml of *n*-hexane. The mixture of the two solutions was then filtered through a water-soluble membrane in order to remove the lipase. The resulting solution was conducted by HPLC spectroscopy analysis. All experiments were performed three times to calculate the mean \bar{x} , stand deviation s , and confidence interval of mean Δx of 95%.

2.3 HPLC analysis

The analysis of both enantiomers of ibuprofen was conducted by an Agilent 1100 high performance liquid chromatography (HPLC) instrument equipped with a OD-H chiral column (5 μ m, 4.6 mm \times 250 mm) and a UV detector. The wavelength of the detector and the environment of the column oven were set up at 254 nm and 40 °C respectively during the measurement. The column is capable of separating the (*R*)- and (*S*)-enantiomers of both acid and ester derivatives. The HPLC was performed with *n*-hexane-isopropyl alcohol (98/2, HPLC grade, Fisher Scientific



Scheme 1 Kinetic resolution of ibuprofen catalyzed by CALB in CO₂-based micelle.

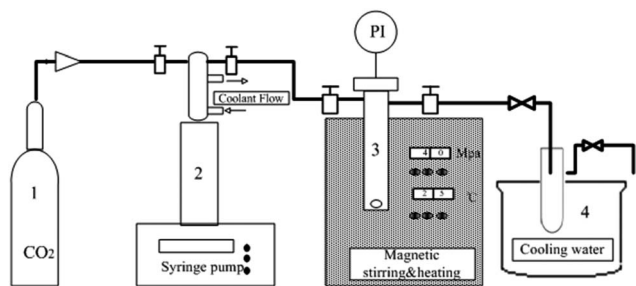


Fig. 1 Schematic diagram of reaction apparatus. (1) CO₂ reservoir tank; (2) high pressure syringe pump; (3) reaction vessel, pressure and temperature control system; (4) cooling water.

Co.) as the mobile phase at a flow rate of 0.5 ml min⁻¹. The concentration of the (*R*)- and (*S*)-ibuprofen ester was determined periodically, and the ee_s of the substrate was calculated by eqn (1).

$$ee_s = \frac{[S] - [R]}{[R] + [S]} \quad (1)$$

3 Results and discussion

The asymmetric synthesis of esters was the most active area of research; the several lipases combined with the unique properties of high pressure CO₂ have successfully permitted the chiral resolution of a large number of racemates.^{32–35}

In this study, we compared the chiral resolution of racemic ibuprofen catalyzed by CALB performed in CO₂-based and hexane-based micelle reactive systems. Fig. 2 showed that in the case of resolution of racemic ibuprofen catalyzed by enzyme, reactive equilibrium could be easily achieved in high pressure CO₂-based than in atmosphere hexane-based micelle. The reason might be attributed to the faster mass transfer effects in high pressure CO₂-based micelle. From the comparison results of enantio-separation for racemic ibuprofen in different reactive systems shown in Table 1, we saw that the better resolution efficiency in high pressure CO₂-based micelles could be achieved within a relatively shorter period of time. The results

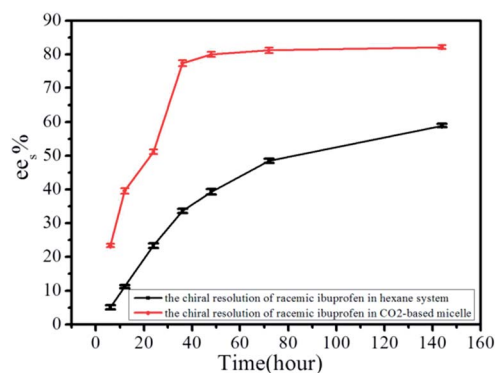


Fig. 2 ee_s values of the chiral resolution of racemic ibuprofen in hexane-based and CO₂-based micellar systems.

Table 1 Comparison of the chiral resolution of racemic ibuprofen in different reactive systems

Entry	Lipase	Solvent	Time [hour]	Conversion [%]	ee _s [%]
1 (ref. 35)	<i>Aspergillus niger</i>	Isooctane	168	>48	>70
2 (ref. 31)	<i>Mucor miehel</i>	SC CO ₂	60	>20	>70
3 (ref. 36)	APE1547	<i>n</i> -Heptane	96	>50	>90
4 ^a	CALB	CO ₂ -based micelle	36	62	83

^a The data in this work.

clearly demonstrated that the CO₂-based micelle stabilized by TMN-10 could be used as a suitable reaction medium for the resolution of racemic ibuprofen. Fig. 2 also showed that the ee_s levelled out in CO₂-based micelle after 36 h. The reaction time was designated as 36 h in following experiments.

The dependence of enzymatic activity on the different EO chain length in the TMN surfactant series were studied also (Fig. 3). For the TMN series surfactants, the ee_s values were increased with an increase in the chain length of EO in TMN. Possibly, the EO groups in TMN surfactant were favourable for stabilization of the enzymes. In addition, the relative long-chain length in TMN-10 could help the esterification and transesterification processes, which resulted in efficient reactive rate in micelle system.

Temperature, pH, pressure and water content were the most important environmental factors affecting enzymatic catalysis in CO₂-based micelle system, particularly their activity, enantioselectivity and stability. The effects of these factors on the chiral resolution of racemic ibuprofen in CO₂-based micelle were systematically investigated.

3.1 The effect of temperature

The reactive temperature was an important factor that affects conversion of the substrate and the enzyme activity. For different reaction systems, the optimum reactive temperature was determined by experiment. In the CO₂-based reverse micelle system, the dependence of temperature on the yield was

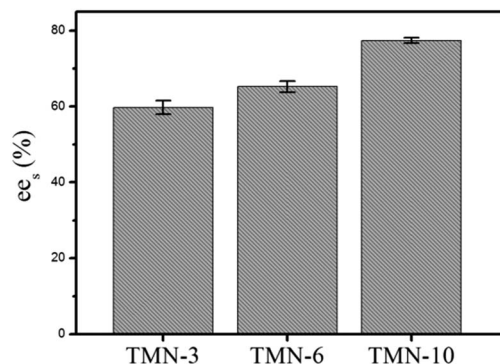


Fig. 3 ee_s values of the chiral resolution of racemic ibuprofen in CO₂-based micelle involved TMN surfactants with different EO chain length.

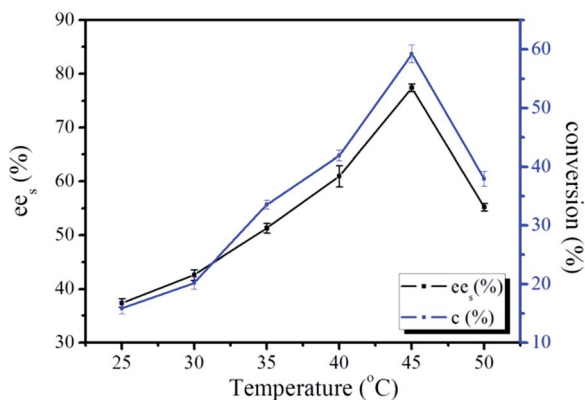


Fig. 4 The conversion and ee_s values of the chiral resolution of racemic ibuprofen in CO_2 -based micelle with TMN-10 at different temperatures.

measured at 250 bar. Fig. 4 showed the conversion and ee_s values of racemic ibuprofen ester under the temperature range of 25–50 °C at 36 hour of the reaction. The results demonstrated that the conversion of the substrate and ee_s increased as the reaction temperature increased until it reached an optimum reaction temperature of 45 °C. The ee_s and conversion of the racemic ibuprofen obtained at this temperature were 85% and 55% respectively. However, the ee_s and conversion decreased rapidly when the temperature was further elevated. A sharp decay in conversion and ee_s at 50 °C was mainly due to the exposure of the enzyme above its optimum temperature for a long time. The accumulated heat could result in partial deactivation and denaturation of enzyme structure. Temperature had a positive effect on kinetics, but it had also a negative effect on enzyme activity. Both effects could be counteracted each other, and resulted in the optimum temperature. This result agreed with other reported findings that the stability of enzyme decreased with the elevated temperatures, although the initial reaction rate was high.³⁷ Therefore, based on the results obtained, the reactive temperature of 45 °C was selected for the resolution of racemic ibuprofen, for which both rapid reactive rate and high enzyme activity were essential for an optimum operation. The actual desired temperature for a reaction was a temperature at which the enzyme exhibited a constant activity over a period of time.

3.2 The effect of pH

In the enzymatic reaction, the pH effect on lipase activity was also carefully investigated. In most enzymatic resolution processes, it has been suggested that the variation in pH of buffer solution might influence the chiral selectivity, since the conformation of an enzyme depended on its ionization state.^{38,39} For general case, the lipase might contain both positively and negatively charged groups, these ionizable groups constituted part of the active sites and often involved in acid–base catalysis. For the CO_2 -based micelle system, the measurement of pH inside the water pool was difficult; the values of pH mentioned here referred to that of the buffer before it was

solubilized in the micelles. In fact, the buffer solution played a great role in pH control for the CO_2 -based micelle.⁴⁰ The results for the pH profile were shown in Fig. 5. It is observed that the pH profile would no longer be bell-shaped fashion; the catalytic activity of CALB was relatively low in a pH 7 buffer solution. The enzymatic activity increased to a maximum value at pH 7.4 and quickly dropped with an increase in the pH value.

3.3 The effect of pressure

The effect of pressure on enantioselection was investigated by the lipase-catalyzed resolution of ibuprofen at pressures ranging from 70 to 300 bar with maintaining the temperature at 45 °C. As shown in Fig. 6, the ee_s values decreased from 66% to 52% when pressure was increased from 70 to 120 bar, and then increased from 52% to the maximum value 78% with the pressure increasing from 120 to 250 bar. A further raise of pressure started to decline the ee_s . The conversion profiles followed similar trends with the ee_s changes with pressure, just a small rise at 80 bar.

Generally, density change with pressure was very sensitive near the critical region for CO_2 -based systems. A large change in density significantly changed the interactions between CO_2 and

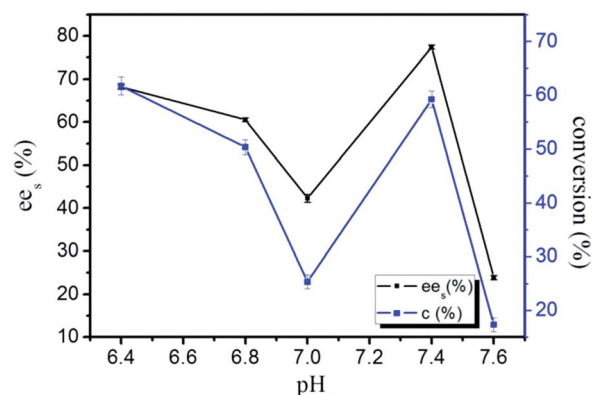


Fig. 5 The conversion and ee_s values of the chiral resolution of racemic ibuprofen in CO_2 -based micelle with TMN-10 at different pH.

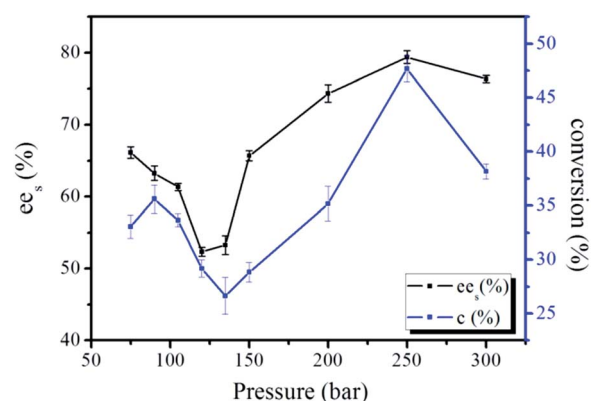


Fig. 6 The conversion and ee_s values of the chiral resolution of racemic ibuprofen in CO_2 -based micelle with TMN-10 at different pressure ranges.

enzyme, causing the formation of carbamates from CO₂ and free amine groups on the surface of the enzyme.²⁵ The interactions might gradually change the conformation of the enzyme in response to pressure, and further resulting in a continuous change in enantioselectivity.^{41–43} The effect of pressure on enzymatic reaction in CO₂-based systems had been the subject of investigation in several studies. However, the influence of pressure on the biocatalysis reactions was still not fully understood due to contradictory results.^{44,45} The rising pressure was accompanied by an increase of the solvating power of high pressure CO₂, which meant the increase of the solubility of the substrates in CO₂. Due to this solvation effect, the partitioning of the substrates between the high pressure CO₂ phase of the reverse micelle and the immediate vicinity of the enzyme was changed. The enzyme environment was depleted with regard to the substrates, which caused a decline in the reaction rate.⁴⁶ When the partitioning of the substrates shifted to the supercritical phase, a further increase of pressure could no longer result in a further depletion in the enzyme environment, *i.e.* the reactive rate could no longer be affected by the rise of pressure. For micelle reactive system, the extent of interfacial area offered more reactive opportunity with the forming of the micelles. The solubility of water in high pressure CO₂ is raised drastically when the pressure is larger than 100 bar.⁴⁷ According to the phase diagrams of water–CO₂ microemulsion formed by TMN series surfactants in the domain of CO₂,⁴⁸ the reactive system tended to form microheterogeneous CO₂-based micelles with the pressure increasing from 150 to 300 bar. Significantly increased interfacial areas favored the substrates approached the interfacial regions in the micelles, accelerating the reaction rate. Therefore, the *ee_s* values and conversion increased when the pressure was larger than 150 bar. The *ee_s* and conversion reached a maximum at 250 bar, where the interfacial areas of the microreactors reached a steady state. With further increase of the pressure at constant volume, the diluting effect caused a decrease in conversion at higher CO₂ pressure. A common rule could not be found to identify how pressure affected the activity of enzyme. From a standpoint of reaction rate, pressure could affect the reaction rate by changing the rate constant directly. An increase in pressure resulted in enhancing fluid density, and improved solvating power of the fluid.

3.4 The effect of water

Water concentration was a key factor which played a significant role in the enzyme-catalyzed reactions in CO₂-based micelles. Control of the water content was important for optimizing the activity in enzymatic systems. The amount of water required to retain catalytic activity was enzyme dependent and varied for different reactive systems. The size of the water pool could be tailored by controlling the *W₀* value (water to surfactant ratio) to make the micelle mimic a biological system or any other restricted environment. In common with many micelles, water-in-CO₂ micelle showed a spherical droplet structure for which the droplet radius was directly proportional to *W₀* value,⁴⁹ and the microenvironment around the enzyme could be tuned directly by simply changing the *W₀* value. The results shown in

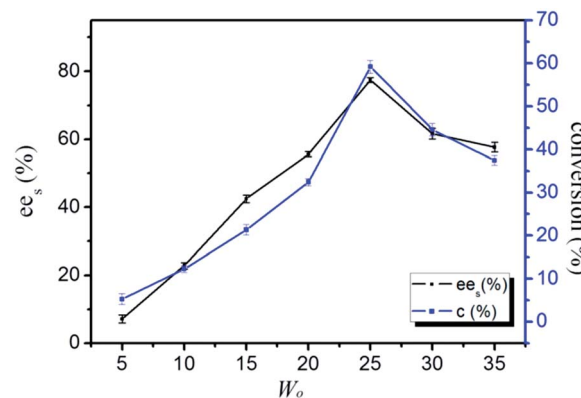


Fig. 7 The conversion and *ee_s* values of the chiral resolution of racemic ibuprofen in CO₂-based micelle with TMN-10 at different water contents.

Fig. 7 demonstrated that increasing the amount of water was expected to increase the enzymatic activity of the lipase in the CO₂-based micelle. The *ee_s* values and conversion gradually decreased as the *W₀* was increased up to 25, after which further increasing the *W₀* could result in the decrease of the *ee_s* and conversion. Generally, a minimum amount of water in the enzyme vicinity was necessary for maintaining catalytic activity, *i.e.* the enzyme needed to be sufficiently hydrated in micelles. If the water content in micelle was too high, the micelle system might be separated two phases; the increased humidity might lead to enzyme deactivation.

4 Conclusions

The chiral resolution of racemic ibuprofen catalyzed by CALB in CO₂-based micelle was explored. The experimental results demonstrated that CO₂-based micelle was used instead of the conventional oil micelle to improve the greenness of the enzyme reactions. For the CO₂-based micelle with surfactant TMN series, the *ee_s* was enhanced with an increase in the chain length of EO in TMN. The effects of reaction parameters such as temperature, pH, pressure, and water content on the reactive behavior were discussed. The combination of enzyme with high pressure CO₂ represented a promising “green” reaction system for bioconversions. In such combination, the amount of less green substances such as surfactants was very small. Studies along these are now in progress.

Acknowledgements

The authors are grateful to the National Nature Science Foundation of China (Grant no. 20876169) for financial support.

Notes and references

- 1 R. Azerad, *Curr. Opin. Biotechnol.*, 2001, **12**, 533–534.
- 2 B. M. Nestl, S. C. Hammer, B. A. Nebel and B. Hauer, *Angew. Chem., Int. Ed.*, 2014, **53**, 3070–3095.

- 3 J. Kim, J. W. Grate and P. Wang, *Trends Biotechnol.*, 2008, **26**, 639–646.
- 4 K. M. Koeller and C.-H. Wong, *Nature*, 2001, **409**, 232–240.
- 5 R. A. Sheldon, *Adv. Synth. Catal.*, 2007, **349**, 1289–1307.
- 6 M. Pileni, *J. Phys. Chem.*, 1993, **97**, 6961–6973.
- 7 D. Goswami, J. K. Basu and S. De, *Crit. Rev. Biotechnol.*, 2013, **33**, 81–96.
- 8 R. N. Mitra, A. Dasgupta, D. Das, S. Roy, S. Debnath and P. K. Das, *Langmuir*, 2005, **21**, 12115–12123.
- 9 J. Zhang and B. Han, *Acc. Chem. Res.*, 2012, **46**, 425–433.
- 10 W. Shang, X. Kang, H. Ning, J. Zhang, X. Zhang, Z. Wu, G. Mo, X. Xing and B. Han, *Langmuir*, 2013, **29**, 13168–13174.
- 11 M. A. Hillmyer, *Science*, 2007, **317**, 604–605.
- 12 C. Müller-Goymann, *Eur. J. Pharm. Biopharm.*, 2004, **58**, 343–356.
- 13 N. Nasongkla, X. Shuai, H. Ai, B. D. Weinberg, J. Pink, D. A. Boothman and J. Gao, *Angew. Chem., Int. Ed.*, 2004, **116**, 6483–6487.
- 14 C. M. Stegmann, D. Seeliger, G. M. Sheldrick, B. L. de Groot and M. C. Wahl, *Angew. Chem., Int. Ed.*, 2009, **121**, 5309–5312.
- 15 J. P. Tan, S. H. Kim, F. Nederberg, E. A. Appel, R. M. Waymouth, Y. Zhang, J. L. Hedrick and Y. Y. Yang, *Small*, 2009, **5**, 1504–1507.
- 16 T. W. Randolph, H. Blanch, J. Prausnitz and C. Wilke, *Biotechnol. Lett.*, 1985, **7**, 325–328.
- 17 D. Hammond, M. Karel, A. Klivanov and V. Krukonsis, *Appl. Biochem. Biotechnol.*, 1985, **11**, 393–400.
- 18 K. Nakamura, Y. M. Chi, Y. Yamada and T. Yano, *Chem. Eng. Commun.*, 1986, **45**, 207–212.
- 19 S. V. Kamat, E. J. Beckman and A. J. Russell, *J. Am. Chem. Soc.*, 1993, **115**, 8845–8846.
- 20 T. Mori and Y. Okahata, *Chem. Commun.*, 1998, 2215–2216.
- 21 K. Chaudhary, S. V. Kamat, E. J. Beckman, D. Nurok, R. M. Kley, P. Hajdu and A. J. Russell, *J. Am. Chem. Soc.*, 1996, **118**, 12891–12901.
- 22 T. Matsuda, T. Harada and K. Nakamura, *Green Chem.*, 2004, **6**, 440–444.
- 23 H. R. Hobbs, B. Kondor, P. Stephenson, R. A. Sheldon, N. R. Thomas and M. Poliakoff, *Green Chem.*, 2006, **8**, 816–821.
- 24 T. Matsuda, T. Harada and K. Nakamura, *Curr. Org. Chem.*, 2005, **9**, 299–315.
- 25 J. Mesiano, E. J. Beckman and A. J. Russell, *Chem. Rev.*, 1999, **99**, 623–634.
- 26 S. Cummings, K. Trickett, R. Enick and J. Eastoe, *Phys. Chem. Chem. Phys.*, 2011, **13**, 1276–1289.
- 27 P. G. Jessop, T. Ikariya and R. Noyori, *Chem. Rev.*, 1999, **99**, 475–494.
- 28 J. Jin, Z. M. Cheng, J. G. Li and S. C. Wu, *Chem. Eng. Sci.*, 2013, **100**, 69–73.
- 29 M. C. Hillier and P. J. Reider, *Drug Discovery Today*, 2002, **7**, 303–314.
- 30 K. Williams, R. Day, R. Knihinicki and A. Duffield, *Biochem. Pharmacol.*, 1986, **35**, 3403–3405.
- 31 P. d. O. Carvalho, F. J. Contesini, R. Bizaco, S. A. Calafatti and G. A. Macedo, *J. Ind. Microbiol. Biotechnol.*, 2006, **33**, 713–718.
- 32 T. Matsuda, K. Watanabe, T. Harada, K. Nakamura, Y. Arita, Y. Misumi, S. Ichikawa and T. Ikariya, *Chem. Commun.*, 2004, 2286–2287.
- 33 E. Celia, E. Cernia, C. Palocci, S. Soro and T. Turchet, *J. Supercrit. Fluids*, 2005, **33**, 193–199.
- 34 G. Paggiola, A. J. Hunt, C. R. McElroy, J. Sherwood and J. H. Clark, *Green Chem.*, 2014, **16**, 2107–2110.
- 35 P. Lozano, *Green Chem.*, 2010, **12**, 555–569.
- 36 D. Zhao, E. Xun, J. Wang, R. Wang, X. Wei, L. Wang and Z. Wang, *Biotechnol. Bioprocess Eng.*, 2011, **16**, 638–644.
- 37 M. Romero, L. Calvo, C. Alba, M. Habulin, M. Primožič and Ž. Knez, *J. Supercrit. Fluids*, 2005, **33**, 77–84.
- 38 V. L. Schramm, *Annu. Rev. Biochem.*, 2011, **80**, 703–732.
- 39 J. M. Palomo, G. Fernández-Lorente, C. Mateo, M. Fuentes, R. Fernández-Lafuente and J. M. Guisan, *Tetrahedron: Asymmetry*, 2002, **13**, 1337–1345.
- 40 P. K. Chan and G. T. Rochelle, *ACS Symp. Ser.*, 1982, **188**, 75–98.
- 41 T. Matsuda, *J. Biosci. Bioeng.*, 2013, **115**, 233–241.
- 42 R. L. Silveira, J. Martinez, M. S. Skaf and L. Martinez, *J. Phys. Chem. B*, 2012, **116**, 5671–5678.
- 43 J. C. Erickson, P. Schyns and C. L. Cooney, *AIChE J.*, 1990, **36**, 299–301.
- 44 C. Kohlmann, S. Leuchs, L. Greiner and W. Leitner, *Green Chem.*, 2011, **13**, 1430–1436.
- 45 S. Kara, W. S. Long, M. Berheide, S. Peper, B. Niemeyer and A. Liese, *J. Biotechnol.*, 2011, **152**, 87–92.
- 46 C. Blattner, M. Zoumpanti, J. Kröner, G. Schmeer, A. Xenakis and W. Kunz, *J. Supercrit. Fluids*, 2006, **36**, 182–193.
- 47 K. Nakamura, *Trends Biotechnol.*, 1990, **8**, 288–292.
- 48 W. Ryoo, S. E. Webber and K. P. Johnston, *Ind. Eng. Chem. Res.*, 2003, **42**, 6348–6358.
- 49 M. Sagisaka, S. Iwama, S. Ono, A. Yoshizawa, A. Mohamed, S. Cummings, C. Yan, C. James, S. E. Rogers and R. K. Heenan, *Langmuir*, 2013, **29**, 7618–7628.