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## Boron nitride nanotubes functionalized with mesoporous silica for intracellular delivery of chemotherapy drugs†

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**Boron nitride nanotube (BNNT)@mesoporous silica hybrids with controllable surface zeta potential were fabricated for intracellular delivery of doxorubicin. The materials showed higher suspension ability, doxorubicin intracellular endocytosis efficiency, and LNcap prostate cancer cell killing ability compared with naked BNNTs.**

Boron nitride nanotubes (BNNTs), as structural analogues of carbon nanotubes (CNTs), have recently attracted a lot of attention in the biomedical field, *e.g.* with respect to bone tissue engineering,<sup>1</sup> drug delivery,<sup>2</sup> boron neutron capture cancer therapy,<sup>3</sup> irreversible lethal electroporation cancer treatment,<sup>4</sup> *etc.* Overall, multi-walled BNNTs show better biocompatibility than CNTs,<sup>1,5</sup> although too long BNNTs result in structural cytotoxicity.<sup>6</sup> The exploration of BNNTs for cancer therapy is a promising direction. BNNTs<sup>3</sup> and CNTs with C<sub>2</sub>B<sub>10</sub> carborane cages attached<sup>7</sup> have shown higher concentration of boron atoms in tumour cells than in blood and other organs, which makes them attractive nanovehicles for the delivery of B to tumour cells for effective B neutron capture for cancer therapy and diagnostics. On the other hand, the exploration of BNNTs as an anti-cancer drug delivery system could provide an integrated vector system by combining chemotherapy with radiation therapy or electroporation cancer therapy. Up to now, little has been done to devise rational strategies for binding anti-cancer drug molecules onto BNNTs, although theoretical calculations have shown that BNNTs are more ideal delivery capsules, compared with other nanotubes, such as carbon, silicon and boron carbide nanotubes.<sup>8</sup>

It is known that the sidewalls of pristine BNNTs are highly hydrophobic and are easily aggregated. The BNNT functionalization for aqueous solubility is necessary. Although some organic molecules, such as polymers<sup>9</sup> and peptides,<sup>10</sup> have been used to

improve the stability of BNNT suspensions, the organic molecules are easily degraded or detached due to the presence of some enzymes *in vivo*. Oxidation and shortening of BNNTs improved the suspension ability and DNA loading to only a certain extent,<sup>11</sup> which is still not satisfactory. The functionalization of BNNTs with silica, a well-proved material with excellent biocompatibility and abundant surface groups, to fabricate core-shell structure,<sup>12</sup> gives the opportunity to improve the suspension ability and hydrophilicity, to control the surface charge and thus to manipulate the intracellular drug delivery.

Herein, we confirm that functionalization of BNNTs with mesoporous silica (i) improved their aqueous suspension ability; (ii) helped to easily control the surface zeta potential; and (iii) enabled binding of drug molecules to functionalized BNNTs, increasing the endocytosis efficiency of doxorubicin (dox) 3–4 times. The designed pathways may not only be effective for isolating BNNTs in aqueous media but may also promote fabrication of functionalized BNNTs and expansion of their future biomedical applications, especially in cancer therapy.

BNNT@mesoporous silica composites with a negative charge (BNNT@mesoporous silica composites, BNNT@MS) and a positive charge (BNNT@mesoporous silica composites grafted with the amino group, BNNT@MS-NH<sub>2</sub>) were fabricated. As illustrated in Scheme 1, the strategy to form mesoporous silica on the outer surface of BNNTs is as follows: the cationic surfactant, cetyltrimethyl ammonium bromide (CTAB), interacted strongly with BNNTs in solution *via* van der Waals forces and was adsorbed onto their outer surfaces.<sup>13</sup> This resulted in the formation of stable BNNT-CTAB surfactant supermolecular structure and gave a high local concentration of CTAB on the surface of BNNTs compared to the bulk solution. After tetraethyl orthosilicate (TEOS) or a mixture of TEOS and 3-aminopropyltrimethoxysilane (APTMS) was added into the above solution, it hydrolyzed and condensed into silicate poly-anions, which cooperatively self-assembled with positively charged groups of cationic surfactant CTAB on the BNNT surfaces. The obtained product was washed several times by centrifugation and redispersion in ethanol to remove the template.

The as-prepared BNNTs synthesized by chemical vapour deposition (CVD)<sup>14</sup> were oxidized in air at 1000 °C for 5 h and

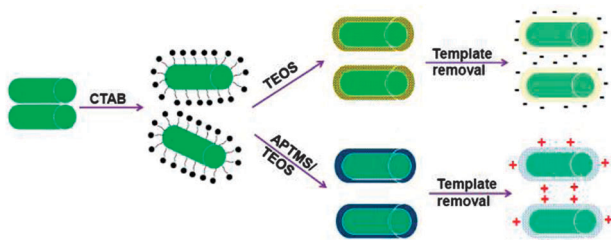
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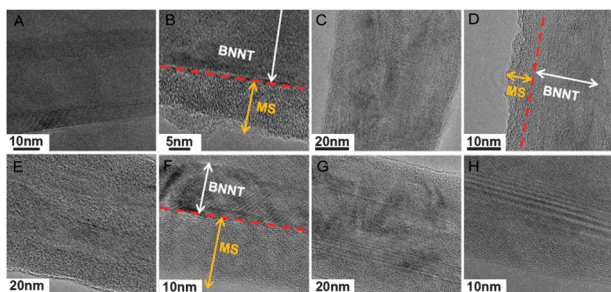
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**Scheme 1** Formation process of BNNT@mesoporous silica.

sonicated in water to obtain the shortened BNNTs, around 1  $\mu\text{m}$  long (Fig. S1, ESI<sup>†</sup>). Synthesis of BNNT@MS was carried out by the hydrolysis of TEOS in the presence of CTAB. By fixing the ratio of CTAB/BN/H<sub>2</sub>O and changing the amount of TEOS, the shell thickness of mesoporous silica was adjusted from 5 nm to 20 nm, as revealed by TEM (Fig. 1). By changing the silica source to the mixture of TEOS and 3-aminopropyltrimethoxysilane (APTMS), the positively charged BNNT@MS-NH<sub>2</sub> hybrids grafted with amino groups were obtained. The obtained products were dispersed in water for more than 24 h and dispersed again by slight shaking when sedimentation had occurred. Whereas the naked BNNTs could be dispersed in water only for 3 h after strong sonication treatment. And once sedimentation had occurred for BNNTs, the dispersion status was difficult to recover by shaking. Thus, it can be concluded that the mesoporous silica shell enhanced the suspension ability and hydrophilic properties of BNNTs in an aqueous solution. In addition, the oxidation treatment of the as-prepared BNNTs in air was important for the subsequent formation of mesoporous silica on the BNNT surface. When H<sub>2</sub>O<sub>2</sub> oxidation treated BNNTs were used as precursors, the mesoporous silica shell could not uniformly form on their surface (Fig. S2, ESI<sup>†</sup>). The N<sub>2</sub> adsorption-desorption isotherms (Fig. S3, ESI<sup>†</sup>) exhibited the type IV isotherm curves with a step between 0.4 and 0.9  $P/P_0$  for BNNT@MS and BNNT@MS-NH<sub>2</sub>, which indicates the presence of mesoporous structure. Both hybrids showed the increased specific surface area of 799 and 465  $\text{m}^2 \text{g}^{-1}$ , respectively, compared with that of pristine BNNTs of about 131  $\text{m}^2 \text{g}^{-1}$ . The pore sizes of both BNNT@MS and BNNT@MS-NH<sub>2</sub> were centred at approximately 1.8 nm.

The surface functionalization of BNNTs after their modification with mesoporous silica was analyzed using the zeta potential measurements and Fourier transform infrared (FTIR) spectroscopy. In contrast to the zeta potential of  $-5 \text{ mV}$  for BNNTs, the values of zeta potential for BNNT@MS and BNNT@MS-NH<sub>2</sub> became negative, around  $-15 \text{ mV}$ , and positive, around  $6 \text{ mV}$ , respectively (Fig. 2A).

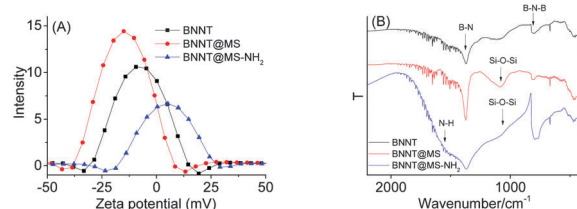


**Fig. 1** TEM images of BNNT (A), BNNT@MS (B), BNNT@MS(-) (C and D), BNNT@MS(+) (E and F) and BNNT@MS-NH<sub>2</sub> (G and H) samples.

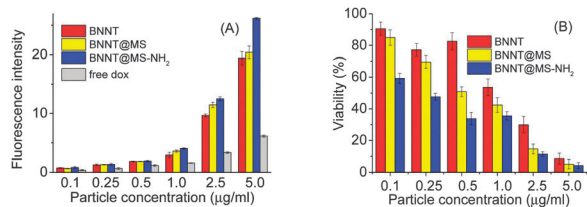
FTIR spectroscopy measurements were carried out to comparatively analyze BNNT, BNNT@MS and BNNT@MS-NH<sub>2</sub> samples, as shown in Fig. 2B. BNNTs showed a typical adsorption spectrum of B-N stretching vibration at  $1372 \text{ cm}^{-1}$  and B-N-B bending vibration at  $800 \text{ cm}^{-1}$ .<sup>15</sup> For BNNT@MS, the adsorption band at  $1080 \text{ cm}^{-1}$  corresponding to Si-O-Si groups appeared.<sup>16</sup> BNNT@MS-NH<sub>2</sub> revealed new shoulder bands at around  $1600 \text{ cm}^{-1}$  associated with the asymmetric and symmetric bending of amino groups ( $-\text{NH}_2$ )<sup>17</sup> and obscure bands at  $1080 \text{ cm}^{-1}$  associated with Si-O-Si vibration. The amount of amino group was determined to be  $(2.5 \pm 1.2) \times 10^{-4} \text{ mol g}^{-1}$ .

The chemotherapy drug dox was then loaded onto BNNTs and BNNTs functionalized with mesoporous silica to check the drug loading ability. The pH-dependent dox loading on BNNTs was observed, as illustrated in Fig. S4A (ESI<sup>†</sup>). Similar pH-dependent loading of dox was found for BNNT@MS and BNNT@MS-NH<sub>2</sub> (data not shown). The loading efficiency of dox on BNNTs increased with an increase in the pH value. The  $\text{pK}_a$  of dox is 8.2. Dox is considered hydrophilic at lower pH because of the presence of the positively charged amino  $-\text{NH}_3^+$  group. The increased deprotonation of  $-\text{NH}_2$  groups on dox at a higher pH = 9.0 accounts for the increased hydrophobicity and decreased solubility of dox,<sup>18</sup> which results in the increase in the interaction between dox and BNNTs. TEM images revealed the presence of amorphous substances on BNNTs, which provides the evidence for the loading of dox on them (Fig. S5, ESI<sup>†</sup>). When doxorubicin was loaded at a pH of about 9.0, BNNTs, BNNT@MS and BNNT@MS-NH<sub>2</sub> showed almost the same doxorubicin loading amounts (Fig. S4B, ESI<sup>†</sup>). BNNTs and functionalized BNNTs allow  $\pi$ -stacking of dox with an ultrahigh loading capacity of  $\sim 60\%$  by weight at a pH value of 9.0. It should be pointed out here that the hydrophobic nature of BNNTs facilitated the loading of dox at such high pH. And the presence of interconnected mesopores in BNNT@MS and BNNT@MS-NH<sub>2</sub> provides the route for the diffusion of dox and loading on the uncovered surface of BNNTs.

To investigate the cellular internalization of dox loaded on different BNNT samples, *i.e.* those functionalized with mesoporous silica, the dox loaded BNNT samples were added to LNCap prostate cancer cells and incubated for 5 h at  $37^\circ\text{C}$  (Fig. 3A and Fig. S6, ESI<sup>†</sup>). For BNNTs, BNNT@MS or BNNT@MS-NH<sub>2</sub>, the loading of dox onto them significantly enhanced the amounts of internalized dox uptaken by LNCap prostate cancer cells, about 3–4 times compared with a free dox. The enhanced internalization of dox through loading onto nanotubes is thought to be due to the different uptake mechanisms, as the dox-loaded particles are internalized by the endocytosis process,<sup>19</sup> compared with a free dox, which enters cells by passive diffusion.<sup>20</sup> As for BNNTs, BNNT@MS



**Fig. 2** (A) Zeta potential and (B) FTIR spectra of BNNT, BNNT@MS and BNNT@MS-NH<sub>2</sub> samples.



**Fig. 3** (A) Fluorescence intensity of intracellular uptake of dox by LNCap prostate cancer cells for BNNT, BNNT@MS and BNNT@MS-NH<sub>2</sub> samples loaded with dox and medium with corresponding amount of free dox after being cultured at 37 °C for 5 h; (B) viability of LNCap prostate cancer cells after culturing with a medium containing different amounts of BNNT, BNNT@MS and BNNT@MS-NH<sub>2</sub> loaded with dox at 37 °C for 24 h.

or BNNT@MS-NH<sub>2</sub> samples, the amounts of the internalized dox were BNNT concentration-dependent. With the increase in nanotube concentration, the internalized dox amounts increased. BNNT@MS and BNNT@MS-NH<sub>2</sub> hybrids showed higher dox intracellular delivery ability than BNNTs. BNNTs functionalized with mesoporous silica exhibited increased suspension ability, which may account for increased dox internalization. Generally, the uptake efficiency depends on the particle size, zeta potential and so on. Particles with a small size and good suspension ability exhibit higher uptake efficiency than those with a large size and poor suspension ability.<sup>21</sup> Particles with a neutral surface possess the lowest uptake efficiency, while particles with a positive charge reveal higher uptake efficiency than those with a negative charge.<sup>22</sup> Functionalization of BNNTs with mesoporous silica improved their suspension ability, which resulted in a higher uptake of dox-loaded particles by cancer cells. In addition, BNNT@MS-NH<sub>2</sub> samples displayed higher dox intracellular delivery ability than BNNT@MS ones, because the zeta potential of cell membranes is normally negative ranging from -40 to -80 mV, and more positively charged particles generally displayed higher uptake by cells.<sup>22</sup> Thus through the adjustment of the surface functional groups on BNNTs, the cellular uptake properties of the loaded drug molecules were effectively manipulated.

To further examine whether the surface functionalization of BNNTs with mesoporous silica can effectively influence the chemotherapy efficacy of dox, the dox loaded BNNTs, and BNNT@MS and BNNT@MS-NH<sub>2</sub> hybrids were cocultured with LNCap prostate cancer cells to check their cancer cell killing ability. As shown in Fig. 3B, BNNTs, BNNT@MS or BNNT@MS-NH<sub>2</sub> samples with dox all showed cancer cell killing ability in a concentration-dependent manner. The cancer cell killing ability follows the sequence: BNNT@MS-NH<sub>2</sub> > BNNT@MS > BNNTs, which is in accordance with the amounts of internalized dox. As mentioned above, the increased suspension ability and hydrophilic properties, and the increased dox internalization by cancer cells were obtained when BNNTs were functionalized with mesoporous silica, which resulted in increased cancer cell killing ability. Similarly, positively charged BNNT@MS-NH<sub>2</sub> exhibited higher cancer cell killing ability than negatively charged BNNT@MS due to the increased dox uptake by cancer cells.

In conclusion, BNNT@mesoporous silica composites with a negative charge and a positive charge were fabricated for

intracellular delivery of chemotherapy drug dox into LNCap prostate cancer cells. The loading of dox onto functionalized BNNTs increased the endocytosis efficiency of dox 3–4 times compared with free dox. BNNTs functionalized with mesoporous silica showed increased suspension ability and hydrophilic properties, increased dox internalization by cancer cells and increased cancer cell killing ability. Positively charged BNNT@MS-NH<sub>2</sub> samples displayed higher dox internalization and cancer cell killing ability than negatively charged BNNT@MS samples. These results provide exciting opportunities for the functionalization of BNNT vehicles in biomedical applications, especially in cancer therapy.

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