

Convergent *de novo* synthesis of vineomycinone B<sub>2</sub> methyl ester†Cite this: *Chem. Commun.*, 2013, **49**, 6806Received 29th May 2013,  
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An efficient *de novo* synthesis of vineomycinone B<sub>2</sub> methyl ester has been achieved. The longest linear route required only 14 steps from achiral commercially available starting materials (4.0% overall yield). The key transformations included the *de novo* asymmetric synthesis of two key fragments, which were joined by a convergent late stage Suzuki's glycosylation for the construction of the aryl β-C-glycoside. A subsequent BBr<sub>3</sub> one-pot debenzoylation, demethylation and air oxidation provided vineomycinone B<sub>2</sub> methyl ester.

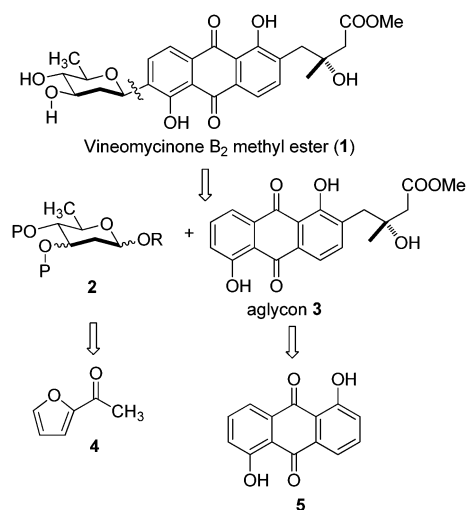
The anthraquinone containing vineomycin members of the angucycline family of antibiotics have been of particular interest to both the synthetic and biological communities due to their unique structures and potent antitumor and antibacterial activities.<sup>1</sup> Vineomycin B<sub>2</sub> is a secondary metabolite of *Streptomyces matensis* subsp. *vineus*, which displays potent antitumor/antibiotic activity with a pharmacologic profile similar to that of the clinically important anthracyclines.<sup>2,3</sup> Acid-catalyzed methanolysis of vineomycin B<sub>2</sub> afforded its aglycon vineomycinone B<sub>2</sub> methyl ester (1) which is the result of *ortho*-C-glycosylation (β-D-olivose) of the methyl ester of fridamycin E (3).<sup>4</sup>

The total synthesis of vineomycinone B<sub>2</sub> methyl ester has been achieved by several groups.<sup>5</sup> While these approaches vary in terms of overall efficiency, the routes tend to focus on the construction of the aglycon portion, rather than β-C-glycoside. These methods varied from the use of cycloadditions with siloxydienes, metalation–stannylation reaction, Bradsher cycloaddition and Cp<sub>2</sub>HfCl<sub>2</sub>/AgClO<sub>4</sub> promoted C-glycosylation to the use of tandem intramolecular benzyne–furan cycloadditions.<sup>5</sup> Despite the success and range of these previous approaches, we felt there was room for a convergent *de novo* asymmetric approach to this class of C-aryl glycoside natural products. In addition to being able to provide access to enantio- and diastereomeric isomers, a convergent *de novo* approach could provide material in a reduced number of steps.

Previously, we described the synthesis and anticancer activity of a sans-C-glycoside trisaccharide portion of vineomycin B<sub>2</sub>.<sup>6</sup> More recently, we finished the asymmetric synthesis of both enantiomers of fridamycin E. These synthetic efforts enable the biological evaluation of these natural product structural motifs. Ultimately we envision these efforts leading to a generalizable late stage C-glycosylation strategy for the synthesis and structure activity relationship studies of this class of C-glycoside natural products.<sup>7</sup>

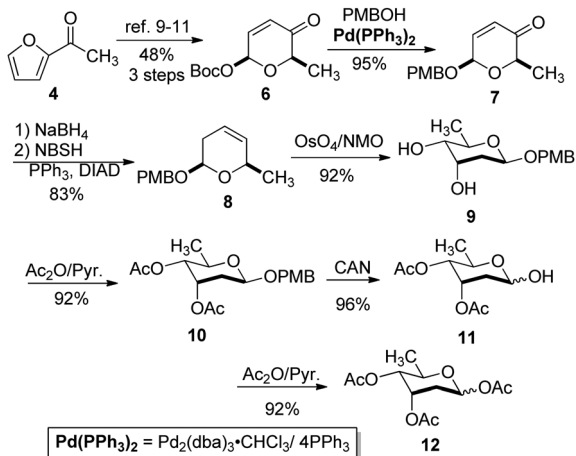
Herein we describe our successful approach to vineomycinone B<sub>2</sub> methyl ester, which is based upon our *de novo* Achmatowicz approach to carbohydrates<sup>6</sup> and our successful syntheses of fridamycin E<sup>8</sup> from achiral starting materials (acyl furan and anthrarufin, respectively). We believe this convergent *de novo* asymmetric approach that utilizes a late stage Suzuki's glycosylation to install the β-C-glycoside provides synthetic material in a more efficient manner than the previous routes.

Our approach to vineomycinone B<sub>2</sub> methyl ester (1) is depicted in Scheme 1. We envisioned that 1 could be accessed from a protected D-olivose or D-digitoxose sugar 2 as the glycosyl donor and the methyl ester of the natural product fridamycin E

Scheme 1 Retrosynthesis of vineomycinone B<sub>2</sub> methyl ester.<sup>a</sup> Guangdong University of Technology, Guangzhou 510006, China<sup>b</sup> Northeastern University, Boston, MA 02115, USA. E-mail: G.O'Doherty@neu.edu

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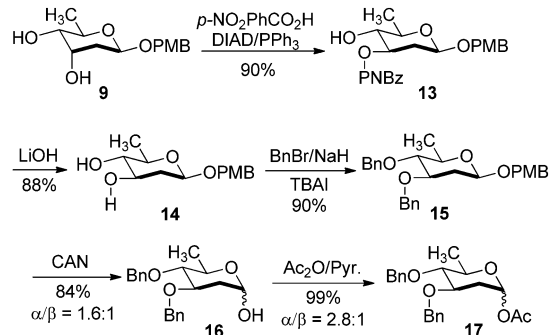


Scheme 2 Approach to glycosyl donor 12.

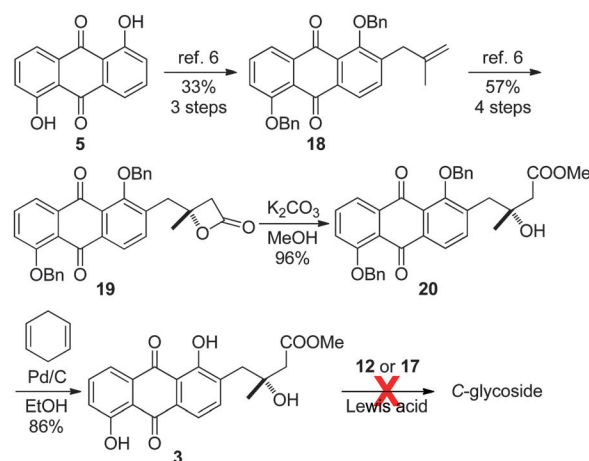
(3) as the aglycon. A Lewis acid promoted  $\beta$ -C-glycosylation could be used to install the glycosidic bond of 1. While the olivose sugar is more direct synthetically, the digitoxose diastereomer has the potential for directing group effects that might be required for selectivity in the C-glycosylation. Conveniently, our *de novo* Achmatowicz strategy was envisioned to produce both diastereomers of the glycosyl donor 2 from commercially available acyl furan 4.<sup>5a</sup> So, the synthetic advantage to the olivose diastereomer boils down to a step before *versus* after the point of convergence. Building upon our fridamycin E work, we imagined an aglycon subunit suitable for coupling (*e.g.*, 3) could be prepared from commercially available anthrarufin 5.<sup>8</sup>

Previously, we have shown that the required Pd glycosyl donor  $\beta$ -D-pyranone 6, as well as its enantiomer and  $\alpha$ -diastereoisomers, could be prepared from acyl furan 4 (Scheme 2). The stereodivergent route employed an enantioselective Noyori reduction,<sup>9,10</sup> an Achmatowicz oxidation, and diastereoselective *tert*-butyl carbonate formation.<sup>11,12</sup> The protected  $\beta$ -D-digitoxose 9 was then completed by a 4-step sequence (Scheme 2).<sup>6</sup> This began with Pd-catalyzed glycosylation of PMBOH and  $\beta$ -D-pyranone 6 to form the  $\beta$ -benzyloxy pyranone 7. A Luche reduction ( $\text{NaBH}_4/\text{CeCl}_3$ ) of 7 followed by the Myers' reductive rearrangement to form 8 and the Upjohn dihydroxylation gave  $\beta$ -D-digitoxose sugar 9 (Scheme 2). The two hydroxyl groups were protected as acetate groups ( $\text{Ac}_2\text{O}/\text{Pyr}$ ) to form 10, which was followed by ceric ammonium nitrate (CAN) oxidative deprotection of the PMB-group to afford lactol 11. Finally the anomeric hydroxyl group was acylated to produce the desired glycosyl donor 12.

We then turned to the preparation of the diastereomeric D-digitoxose glycosyl donor 17 with and anomeric acetate (Scheme 3). This began with a highly regio- and stereospecific Mitsunobu like reaction ( $p\text{-NO}_2\text{PhCO}_2\text{H}/\text{DIAD}/\text{PPh}_3$ ) on *cis*-diol 9 to provide 13 in excellent overall yield. Hydrolysis of the nitrobenzoate 13 with LiOH afforded the *trans*-diol 14 (88%). The two hydroxyl groups were then protected as benzyl ethers (BnBr, NaH/TBAI, 88% yield). A subsequent deprotection of the PMB-group in 15 with CAN smoothly converted it into lactol 16 in 84% yield as an equilibrating mixture of diastereomers ( $\alpha:\beta = 1.6:1$ ). Finally, acylation of 16 gave  $\beta$ -D-olivose acetate 17 (99%,  $\alpha:\beta = 2.8:1$ ) in near quantitative yield.



Scheme 3 Caption approach to glycosyl donor 17.

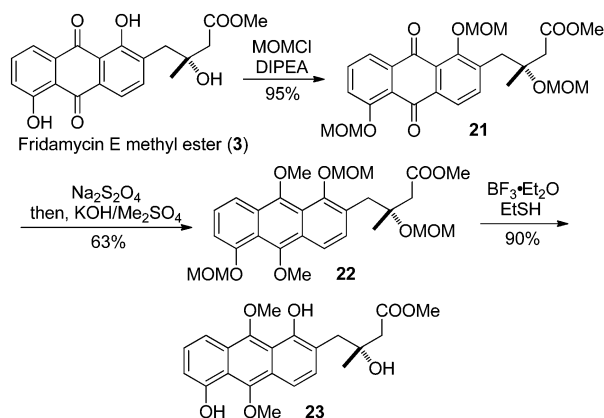


Scheme 4 C-glycosylation attempts of fridamycin E methyl ester 3 with 12 and 17.

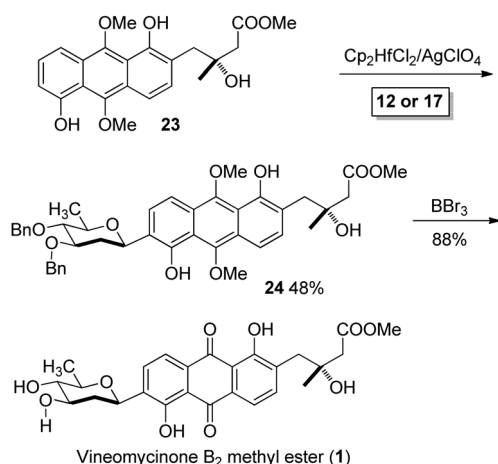
With the glycosyl donor in hand, we then turned to the synthesis of the aglycon 3, the methyl ester of fridamycin E (Scheme 4), this was chosen as it would lead to the latest possible sight of convergence. Previously, we have also developed the asymmetric synthesis of fridamycin E.<sup>8</sup> In this route, the  $\beta$ -lactone 19 was prepared in 7 steps and in 19% overall yield. The synthesis began with the monomethylation of anthrarufin, followed by a redox promoted Claisen rearrangement and benzyl protection to afford *ortho*-methallylated product, was asymmetrically epoxidized in a 3-step protocol (Sharpless dihydroxylation-tosylation-ring-closure) to give epoxide 18. A subsequent Coates carbonylation regio- and stereospecifically provided  $\beta$ -lactone 19, which after methanolysis ( $\text{MeOH}/\text{K}_2\text{CO}_3$ , 96% yield) smoothly afforded  $\beta$ -hydroxyl carboxylic acid methyl ester 20 in excellent yield (96%) with the requisite tertiary alcohol. Finally, a reductive debenzoylation of 20 with 1,4-cyclohexadiene and Pd/C in ethanol gave the methyl ester of fridamycin E 3 in 86% yield.

With both glycosyl donor and aglycon in hand, we explored the possibility of a Lewis acid promoted C-glycosylation of 3 with 12 or 17 (Scheme 4). Unfortunately, our exhaustive survey of various Lewis acids and conditions (*e.g.*,  $\text{BF}_3\cdot\text{Et}_2\text{O}$ , TMSOTf and  $\text{Cp}_2\text{HfCl}_2/\text{AgClO}_4$ )<sup>5e</sup> met with no success. No signs of the desired C-glycoside product were detected.

Thus, we decided to target an earlier synthetic intermediate that would (1) possess a more electron rich aromatic ring and (2) be suitably protected to reveal the aglycon portion of the natural product in a single step.<sup>7e</sup> This led to the choice of the arene 23 with the



Scheme 5 Synthesis of the aglycon 23.

Scheme 6 Synthesis of vineomycinone B<sub>2</sub> methyl ester (1).

reduced quinone ring protected as a bis-methyl ether (Scheme 5). The revised synthesis began with an MOM protection of 3, then *in situ* dithionate reduction of 21 to an anthraquinol intermediate, followed by methylation (KOH/Me<sub>2</sub>SO<sub>4</sub>) to give 22 in 63% yield. Finally deprotection of 22 gave the bis-phenol 23 in 90% yield.

With the second-generation C-glycosylation coupling partner in hand, we turned our focused on the C-glycosylation of 23 with 12 and 17 (Scheme 6). To our delight, treatment of a 1 : 1 mixture of 17 and 23 under the typical Suzuki conditions (Cp<sub>2</sub>HfCl<sub>2</sub>/AgClO<sub>4</sub>) led to formation of the desired β-C-glycoside 24 as a single regio- and diastereo-isomer in 48% yield. While the reaction is envisaged as proceeding through an *O*- to C-glycoside rearrangement,<sup>5</sup> no evidence of an *O*-glycoside product was detected. A clue as to how to best complete the synthesis came from careful observation of the crude reaction mixture. Specifically, under the Lewis acid conditions of the C-glycosylation reaction one can observe the conversion of the acid sensitive starting material 23 into a corresponding anthraquinone. As a result, a one-pot per-deprotection–oxidation was realized with the exposure of 24 to excess BBr<sub>3</sub>. The bis-debenzylation, bis-demethylation and subsequent air oxidation of an anthraquinol intermediate afforded vineomycinone B<sub>2</sub> methyl ester 1 in 88% yield. This synthetic material had physical and spectroscopic data in good agreement with literature values (*e.g.*,  $[\alpha]_{\text{D}}^{20} = +109$  (*c* = 0.27, dioxane),  $[\alpha]_{\text{D}}^{29} = +118$  (*c* = 1.05, dioxane),<sup>5e</sup>  $[\alpha]_{\text{D}}^{24} = +109.8$  (*c* = 0.00091, CDCl<sub>3</sub>)).<sup>5f</sup>

In contrast, the C-glycosylation between 23 and 12 was less successful, as only trace amount of the digitoxose isomer was observed.

In summary, the convergent *de novo* synthesis of vineomycinone B<sub>2</sub> methyl ester was accomplished in only 14 steps with 4.0% overall yield in the longest linear sequence, which is a highly efficient route comparing the shortest previous synthesis (16 steps) of 1.<sup>5</sup> The route shows the efficiency of the convergent *de novo* synthesis of coupling partners for a late stage convergent C-glycosylation followed by a single step deprotection–oxidation to yield the natural product. It is interesting to note that while this is the first synthesis of vineomycinone B<sub>2</sub> methyl ester that uses asymmetric catalysis to install all of the stereocenters, in terms of overall synthetic efficiency it compares quite favorably with the elegant work of Martin and predecessors.<sup>5</sup> Having this improved stereo-divergent access to vineomycinone B<sub>2</sub> methyl ester, its diastereomer and congeners should further enable their biochemical/medicinal chemistry study. Further vineomycins synthetic/biological investigations are ongoing and will be reported in due course.

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