Marine Natural Products

D. J. Faulkner

Scripps Institution of Oceanography, A-012F, University of California, San Diego, La Jolla, CA 92093, USA

Reviewing the literature published between December 1987 and December 1988 (Continuing the coverage of literature in *Natural Product Reports*, 1988, Vol. 5, p. 613)

- 1 Introduction
- 2 Marine Microorganisms and Phytoplankton
- 3 Blue-Green Algae (Cyanobacteria)
- 4 Green Algae
- 5 Brown Algae
- 6 Red Algae
- 7 Sponges
- 8 Coelenterates
- 9 Bryozoans
- 10 Molluscs
- 11 Tunicates
- 12 Echinoderms
- 13 Miscellaneous
- 14 References

1 Introduction

This Report is a review of the literature of marine natural product chemistry for the period 1 December 1987 to the end of 1988. This is the sixth in a series of reviews published in *Natural Product Reports*. The earlier reports¹⁻⁵ cover the period from 1977 to December 1987.

Like the earlier reports in this series, this review does not provide a comprehensive coverage of all research involving chemicals from marine organisms but concentrates on papers that report novel marine natural products with interesting biological and pharmaceutical properties. Biochemical studies involving marine organisms and reports of primary metabolites have been specifically omitted. Research on the biosynthesis of marine natural products has been reviewed in an excellent Report by Garson.⁶ Owing to an expansion of interest in the pharmacological properties of marine natural products, it has become much more difficult to track the literature in this area. While every effort has been made to record the pharmacological activity of new metabolites, studies of mechanisms of action and reports of basic pharmacology in which marine natural products were used as biological probes are reviewed only when they result in a significant change in the biomedical status of a compound. In the area of synthetic organic chemistry, reports of the total synthesis of marine natural products or close analogues are included but papers dealing with methodology directed toward the synthesis of marine natural products have been omitted. No attempt has been made to review the patent literature or conference abstracts and reports.

During the reporting period two volumes of a new series of interpretive reviews have appeared. The subjects covered in the review chapters are 'Natural Product Chemistry and Chemical Defense in Tropical Marine Algae of the Phylum Chlorophyta',7 'Chemical Ecology of the Nudibranchs',8 'Marine Metabolites which Inhibit Development of Echinoderm Embryos',9 'The Search for Antiviral and Anticancer Compounds from Marine Organisms',10 'Chemistry of Aqueous Marine Extracts: Isolation Techniques',11 'Secondary Metabolites from Echinoderms as Taxonomic Markers',12 'The Chemical Ecology of Alcyonarian Corals',13 and 'Chemical Defense in Fishes',14

2 Marine Microorganisms and Phytoplankton

Although there is considerable interest in the potential of marine bacteria and fungi to produce pharmacologically valuable compounds, very little activity has been reported in this field. It has been demonstrated that the three diketopiperazines (1)—(3) that were previously ascribed to the sponge *Tedania ignis*¹⁵ are produced by a species of *Micrococcus* isolated from the sponge. ¹⁶ Dendryphiellin A (4) is an unprecedented metabolite, of possible terpenoid origin, from the marine deuteromycete *Dendryphiella salina*. ¹⁷

Interest in the dinoflagellates arises primarily from their implication in outbreaks of ciguatera poisoning, an illness caused by ingestion of reef fishes. The dinoflagellate most commonly associated with ciguatera outbreaks in the Pacific is Gambierdiscus toxicus, from which the partially characterized maitotoxin was isolated.¹⁸ A new nitrogenous toxin called prorocentrolide (5) was isolated from *Prorocentrum lima*,¹⁹

from which the structurally unrelated toxin okadaic acid (6) had previously been obtained.²⁰ The stereochemistry of prorocentrolide (5) remains to be determined as does that of the antifungal agent goniodomin A (7), which was isolated from Goniodoma pseudogoniaulax.²¹ Amphidinolide C (8) is a 25-membered macrocyclic lactone from a cultured dinoflagellate of the genus Amphidinium that was originally obtained from a flatworm.²² Like previous compounds from this source,^{23, 24} amphidinolide C (8) has potent antineoplastic activity. The sphingosine derivative symbioramide (9), which was isolated from the cultured species of Symbiodinium, is an activator of sarcoplasmic reticulum Ca²⁺-ATPase.²⁵

(12)

3 Blue-Green Algae (Cyanobacteria)

The majority of studies during the past year have featured terrestrial rather than marine cyanophytes. The exceptions are reports of the isolation and structural elucidation of two cytotoxic linear pentapeptides, majusculamide D (10) and deoxymajusculamide D (11),²⁶ and a minor cyclic depsipeptide, 57-normajusculamide C (12),²⁷ from Lyngbya majuscula. A new synthesis of (-)-malyngolide (13), which is a metabolite of Lyngbya majuscula,²⁸ employed an asymmetric reduction using Baker's yeast.²⁹ The cytotoxic styrylchromone hormothamnione (14), that was isolated from Hormothamnion enteromorphoides,³⁰ has been synthesized by two similar routes.^{31,32}

(14)

(25)

4 Green Algae

The chemical defences of tropical green algae continue to be studied in detail, 7.33 but few new metabolites have been described. The only exceptions are two new diterpenoid fish-feeding inhibitors (15) and (16) from *Pseudochlorodesmis furcellata*.34

5 Brown Algae

A review of 'The odoriferous polyene hydrocarbons from marine and terrestrial plants' provides a compact account of research on the pheromones of marine brown algae. To Caudoxirene (17) is a new spermatozoid-releasing and attracting factor from *Perithalia caudata*. The structure of caudoxirene (17) was determined by GC-mass spectral analysis coupled with micro-scale deoxygenation to obtain E-viridiene (18) and was confirmed by synthesis. Five species of brown algae from Victoria, Australia, all use slightly different enantiomeric compositions of (-)-1R,2R hormosirene (19) and (+)-1S,2S hormosirene as gamete attractants. (R)-(+)-4Butyl-2,6-cycloheptadienone (20), which is a minor constituent of the essential oil of a Hawaiian species of *Dictyopteris*, was synthesized in good yield by a route that employed a ring expansion to construct the 7-membered ring.

(26)

GC-MS analysis of the constituents of the Australian algae Zonaria turneriana, Z. crenata, and Z. angustata revealed only known acylphloroglucinol derivatives. ⁴⁰ A second diprenylphenol (21) was found in Perithalia caudata from Australia. ⁴¹ Three new tetraprenyltoluquinols (22)—(24) were isolated from a Sicilian species of Cystoseira. ⁴² A natural hybrid between Cystoseira elegans and C. algeriensis, previously reported as C. algeriensis, ⁴³ ⁴⁴ has been described together with the structural elucidation of two new tetraprenyltoluquinols (25) and (26). ⁴⁵ A model compound (27) incorporating the active portion of the cytotoxin stypoldione (28) ⁴⁶ has been synthesized. ⁴⁷

(43)

A reinvestigation of Cystophora moniliformis resulted in the isolation of two new terpenoids (29) and (30)⁴⁸ in addition to those metabolites reported previously.^{49,50} The methyl acetal (30) is believed to arise by methanolysis of the epoxide (29) during chromatography. Two additional acyclic diterpenes (31) and (32) have been reported from Bifurcaria bifurcata.51 Three new diterpenes of the xenicane class, 4-acetoxydictyolactone (33) and dictyolides A (34) and B (35), together with a norxenicane derivative, nordictyolide (36), are the antitumour constituents of Dictyota dichotoma from Okinawa.52 The conformations of these interesting bicyclic diterpenes were

(42)

determined by interpretation of 2D-NOESY experiments. The structure of crenuladial (37), which is an antimicrobial diterpenoid from Dilophus ligatus, is actually a mixture of bishemiacetals resulting from hydration of a hypothetical 1,4dialdehyde.53 The absolute configurations of dictyodial (38), originally isolated from Dictyota crenulata,54 and the bicyclic isomer sanadaol (39) from Pachydictyon coriaceum⁵⁵ were determined by an enantioselective synthesis of both (+)- and (-)-sanadaol.56 In addition to the known diterpenes (40)—(43),^{57—60} three new hydroazulenoid diterpenes, dictyotriols C (44), D (45) and E (46), were obtained from a Canary

(45) $R^1 = H$, $R^2 = OH$

Islands species of *Dictyota*. ⁶¹ The absolute configurations of the new compounds (44)—(46) and those of dictyol B (41) and dictyotadiol (43) were determined by the CD allylic benzoate method and were found to be opposite to that determined for pachydictyol A (40) by *X*-ray crystallography. ⁵⁷ The novel tricyclic diterpene (47) from *Dilophus okamurai* inhibits feeding by young abalone. ⁶² Two new dolastane diterpenes (48) and (49) have been isolated from *Dictyota cervicornis* from Brazil. ⁶³ Amijitrienol (50), which is a metabolite of *Dictyota linearis*, ⁶⁴ has been synthesized in racemic form by a 10-step sequence. ⁶⁵ Both (+)-isoamijiol (51) and (+)-dolasta-1(15), 7,9-trien-14-ol

(52), which are optical enantiomers of the natural products, $^{66.67}$ have been synthesized from (+)-limonene. 68 The unusual diterpenoid dictymal (53), which is a metabolite of *Dictyota dichotoma*, 69 has been synthesized from two optically active iridoid (monoterpene) synthons. 70 One of the more unusual compounds from a Canary Islands species of *Dictyota* is β -dictalediol monoacetate (54), the structure of which was determined by X-ray analysis. 71

The arsenic-containing carbohydrate (55), which was isolated from the kelp *Ecklonia radiata*, ⁷² has been synthesized from Dribose. ⁷³

6 Red Algae

The eicosanoids, which are a family of biologically active arachidonic acid derivatives, are fairly frequently found in extracts of marine organisms. Ptilodene (56) is an eicosanoid from Ptilota filicina that inhibits both 5-lipoxygenase and Na⁺/K⁺ ATPase. ⁷⁴ 12-(S)-Hydroxyeicosapentaenoic acid (57), which is a potent inhibitor of platelet aggregation, has been isolated in large quantities from Murrayella periclados and has been recognized as the compound previously identified ⁷⁵ as 9-hydroxypentaenoic acid (58) from Laurencia hybrida. ⁷⁶ The unusual unsaturated aldehyde, (E)-2-tridecyl-2-heptadecenal (59), was isolated from a Japanese species of Laurencia and was synthesized by aldol dimerization of pentadecanal. ⁷⁷ Six new

polybrominated 2(5H)-furanones (60)—(65) have been isolated from *Delisea elegans*. Five of the new compounds may be considered as dimers of the fimbrolide (66) that was previously isolated from *Delisea fimbriata*. The structures of dimers (60)—(63) were all determined by X-ray studies and the remaining compounds were identified by interpretation of spectral data. Chilenone B (67) is an unusual trimer of 2-methyl-3(2H)-furanone that was isolated in low yield from Laurencia chilensis. The structures of chilenones A (68)⁸¹ and

B (67), both of which were established by X-ray analysis, are totally unrelated to other Laurencia metabolites.

A new halogenated C₁₅ acetylene, graciosin (69), and the corresponding allene, graciosallene (70), were obtained from a Canary Island specimen of Laurencia obtusa and the structures were determined by X-ray crystallography and chemical methods. ⁸² The known compounds (3E)-laureatin (71) and (3E)-isolaureatin (72)⁸³ have been reported as the major constituents of a specimen of Laurencia nipponica from Shichigahama, Japan. ⁸⁴ (-)-Laurenyne (73) has been synthesized using an acetal-initiated cyclization to form the crucial 8-membered ring. ⁸⁵ This synthesis served to redefine the absolute configuration of (+)-laurenyne, which had earlier been determined by X-ray analysis of material obtained from an Aegean Sea specimen of Laurencia obtusa. ⁸⁶ The enantioselective syntheses of (3Z,6R,7R,9Z,12Z)-6-acetoxy-7-chloropentadeca-3,9,12-trien-1-yne (74) and the corresponding (3E) isomer, which are metabolites of Laurencia pinnatifida, ⁸⁷ have been reported. ⁸⁸

$$CH_2CI$$
 CH_2CI
 R
(75) $R = CI$
(76) $R = H$

(79)
$$R^1 = CI$$
, $R^2 = H$
(80) $R^1 = H$, $R^2 = CI$

(81)
$$X = CI$$

(85) $X = Br$

(86)
$$R^1 = Br$$
, $R^2 = CI$
(87) $R^1 = CI$, $R^2 = Br$

(91)

(95) R = H

Four polyhalogenated linear monoterpenes (75)—(78) and two epimeric bicyclic monoterpenes (79) and (80) were isolated from a Great Barrier Reef sample of *Chondrococcus hornemannii* and their structures were elucidated by interpretation of spectral data. ⁸⁹ Four new polyhalogenated monoterpenes (81)—(84) have been found in two collections of *Plocamium hamatum* from Australia. One collection contained the cyclic monoterpene (81), which is closely related to the known metabolite mertensene (85), ⁹⁰ while the second collection contained two linear monoterpenes (82) and (83) and one cyclic monoterpene (84). ⁹¹ The distribution of aromatic sesquiterpenes in several species of *Laurencia* from the South Australia coast has been reported but no new compounds were encountered. ⁹²

Puertitols A (86) and B (87) are novel halogenated bisabolenes from a Canary Islands sample of Laurencia obtusa. 93 The structures were elucidated by interpretation of spectral data and the absolute configurations determined by use of the CD allylic benzoate method. Majusculone (88) is an unusual norchamigrene derivative from Laurencia majuscula collected in Japan. 94 The structure of 2,10-dibromo-3-chamigren-7-en-9ol (89), previously reported as a metabolite of Laurencia pacifica95 and now found in Laurencia nipponica, has been confirmed and the absolute configuration determined by X-ray analysis.96 A new labile sesquiterpene, dehydrochloroprepacifenol (90), has been obtained from a Mediterranean specimen of L. majuscula.97 The structure of (90) was determined by lowtemperature X-ray diffraction analysis. Pinnatazane (91), the structure of which was determined by X-ray analysis, is a new chamigrene derivative from L. pinnatifida. 98 The known epoxide (92), previously reported from an unidentified species of Laurencia, 99 has also been reported from L. pinnatifida. 100 Five new chamigrene derivatives, laurencenones A-D (93)-(96) and deschloroelatol (97), were obtained from a Jamaican specimen of L. obtusa.¹⁰¹ A specimen of L. scoparia from Jamaica contained several known chamigrenes and the novel epoxide (98).101 The structures of all new compounds were determined by interpretation of spectral data. Two new

ČH₂Br

(103)

.COO

 NH_2

chamigrenes, obtusadiene (99) and isoobtusadiene (100), were isolated from a Puerto Rican specimen of $L.\ obtusa$; the structures were elucidated by analysis of spectral data and the absolute stereochemistry was defined by using the CD benzoate method. 102

(111)

(112)

Venustanol (101) is a brominated labdane diterpene from Laurencia venusta that was identified by spectral methods. 103 Two new brominated diterpenes, sphaeropyrane (102)¹⁰⁴ and sphaeroxetane (103),¹⁰⁵ have been recognized as minor metabolites of *Sphaerococcus coronopifolius* and their structures have been proposed on the basis of spectral data.

The triterpenes from Laurencia species have been the targets of five synthetic efforts. Teurilene (104), which is a metabolite of Laurencia obtusa, ¹⁰⁶ has been synthesized by two very similar routes. ^{107,108} Two syntheses of venustatriol (105) from L. venusta¹⁰⁹ and one synthesis of thyrsiferol (106) from L. thyrsifera¹¹⁰ have been reported. ^{111–113} These routes all involve the formation of cyclic ethers through reactions involving the reaction of alcohols with epoxides.

The red algae often contain unusual amino acids. Recent examples are the quaternary ammonium compounds (107) and (108) from *Pterocladia capillacea*¹¹⁴ and (109) from *Grateloupia proteus*, ¹¹⁵ and the dimethyl sulphonium salt (110) from *Lophocladia lallemandi*. ¹¹⁶ Itomanindoles A (111) and B (112)

are two isomeric methylsulphinyl substituted indoles from Laurencia brongniartii. The structure of itomanindole A (111) was determined by X-ray analysis. Two phenolic metabolites of Polysiphonia lanosa, rhodomelol (113) and methylrhodomelol (114), 117 and delesserine (115), which was obtained from Delesseria sanguinea, 118 have been synthesized from vitamin C. 119 Floridoside (116), 'D'-isofloridoside (117) and 'L'-isofloridoside (118) have been characterized as metabolites of Porphyra perforata. 120

(126) $R^1 = CN$, $R^2 = H$, $R^3 = H$

7 Sponges

Three new polyacetylenic alcohols, melynes A (119), B (120) and C (121), have been isolated from a species of *Xestospongia*

from Vanuatu.121 The structures of the melynes were elucidated from spectral data. Melyne A (119) is active against the intestinal protozoan Giardia. Dysidazirine (122) is a unique azacyclopropene-containing lipid from Dysidea fragilis. 122 The structure of dysidazirine (122), which is cytotoxic and inhibits P. aeruginosa, C. albicans, and S. cerevisiae, was elucidated by chemical and spectroscopic methods. Discodermia calyx has yielded three new cytotoxins - calyculins B (123), C (124), and D (125) - that were identified by comparison of spectral data with those of the known¹²³ metabolite, calyculin A (126).¹²⁴ cytotoxic macrodiolides (dimeric macrolides), bistheonellides A (127) and B (128), have been isolated from a Japanese species of Theonella. 225 Bistheonellide A (127) was shown to be identical to misakinolide-A, for which the incorrect

(128) R = H

(130) ax H-5, eq H-9

(132) eq H-5, ax H-9

(131) ax H-5, eq H-9

(133) eq H-5, ax H-9

monomeric structure (129) had earlier been proposed. 126 It is not immediately clear whether fijianolides A (130) and B (131), isolated from Spongia mycofijiensis from Vanuatu, 127 are identical to isolaulimalide (132) and laulimalide (133), respectively, from an Indonesian species of Hyatella. 128 The spectral data for these cytotoxic metabolites were reported in different solvents making direct comparisons impossible and the proposed structures differ in the conformation assigned at C-5 and C-9, It is hoped that a future paper will resolve this confusion and report the complete stereochemical assignment for these interesting metabolites. A new group of antiviral compounds have been isolated from two unrelated sponges. Mycalamide A (134), which is a metabolite of a New Zealand species of *Mycale*, ¹²⁹ and onnamide A (135), which occurs in an Okinawan species of *Theonella*, ¹³⁰ differ only in the size and nature of the side chain and both compounds resemble pederin (136), which was isolated from a terrestrial beetle Paederus fuscipes. 131 The structures of mycalamide A (134) and onnamide A (135) were elucidated by interpretation of spectral data and the stereochemistry has been assumed to be the same as that of pederin (136).

A species of *Halichondria* from Palau contained five minor macrolides (137)—(141) in addition to the major metabolite halichondramide (142) that was reported previously, ¹³² The structures of dihydrohalichondramide (137), isohalichondramide (138), acid (139), imide (140), and ester (141) were determined by comparison of the spectral data with those of halichondramide (142). ¹³³ Two unusual anthelmintic oxazoles, bengazoles A (143) and B (144), have been isolated from a Fijian sponge of the family Jaspidae. ¹³⁴ The structures proposed for the bengazoles were based on a detailed analysis of ¹H NMR data, assisted by molecular modelling. Mycothiazole (145) is a novel thiazole-containing lipid with anthelmintic properties that was isolated from *Spongia mycofijiensis*. ¹³⁵ The structure of mycothiazole (145) was established by analysis of spectral data.

(143) $R = (CH_2)_{12}CH_3$ (144) $R = (CH_2)_{11}CH(CH_3)_2$

An unusually large number of alkaloids have been reported from sponges during the current review period. The antifungal alkaloid papuamine (146) was isolated from a Papua New Guinea specimen of *Haliclona* and its structure was elucidated by analysis of spectral data. ¹³⁶ The same compound was found to be a minor constituent of a Palauan species of *Haliclona* (cf. H. hornelli) that contained haliclonadiamine (147), which is an unsymmetrical diastereoisomer of papuamine (146), as the major antifungal metabolite. ¹³⁷ The structure of haliclonadiamine (147) was determined by X-ray analysis. The structure of petrosin-A has been revised from (148) to (149) as the result of a 2D-NMR study. ¹³⁸ Four new alkaloids of the manzamine family have been reported from sponges of the genus *Haliclona* and *Xestospongia*. Manzamines B (150) and C (151) were

obtained along with the known alkaloid manzamine A (152)¹³⁹ from an Okinawan species of *Haliclona*.¹⁴⁰ The structures of both new alkaloids were determined by *X*-ray analysis. Manzamines E (153) and F (154), which are the cytotoxic constituents of an Okinawan species of *Xestospongia*, were identified by comparison of their spectral data with those of manzamine A (152).¹⁴¹ Manzamine F (154) was shown to be the same as an alkaloid called keramamine B, to which the incorrect structure (155) had been assigned.¹⁴²

(160)

The cytotoxic alkaloid girolline (156) was isolated from the

axinellid sponge *Pseudaxinyssa cantharella* from New Caledonia. 143 The structure of girolline was elucidated by physicochemical methods. Dysidamide (157) is a new hexachlorinated amino acid derived metabolite from a Red Sea species of *Dysidea*. 144 The structures of two new cyclic peptides, fenestrins A (158) and B (159), from the Fijian sponge *Leucophloeus fenestrata*, were proposed on the basis of spectral analysis. 145 The cyclic depsipeptides jaspamide (160), which is a metabolite of a species of *Jaspis*, 146, 147 and geodiamolide B (161) from a species of *Geodia* have been the subject of

(161)

several synthetic efforts. Both the polypropionate¹⁴⁹ and the peptide¹⁵⁰ portions of jaspamide have been synthesized separately and total syntheses of both jaspamide (160)¹⁵¹ and geodiamolide B (161)¹⁵² have been reported.

Dragmacidin (162) is a cytotoxic bis-indole alkaloid from a deep-water species of *Dragmacidon*. The structure of dragmacidin (162), which showed *in vitro* activity against P388 cells and against human lung, colon, and mammary cancer cell lines, was elucidated by interpretation of spectral data. Four species of *Spongosorites* collected in deep water off the Bahamas have yielded two known alkaloids, topsentins-B1 (163) and -B2 (164), that had previously been found in a Mediterranean shallow-water sponge called *Topsentia genitrix*, ¹⁵⁴ together with a new alkaloid (165). ¹⁵⁵ The synthesis of topsentin-B1 (163) and several analogues were reported, together with their antiviral and antitumour activities. A synthesis of topsentin-A (166), which is the simplest of the alkaloids from *T. genitrix*, has also been reported. ¹⁵⁶ *T. genitrix* also contains the free base 3-methyladenine. ¹⁵⁷ 3-Methylcytidine (167), 3-methyl-2'-deoxycytidine (168), and 3-methyl-2'-deoxyuridine (169) were identified as the metabolites of *Geodia baretti* that caused

(167) $R^1 = \dot{N}H_2$, $R^2 = OH$ (168) $R^1 = \dot{N}H_2$, $R^2 = H$

(169)
$$R^1 = 0$$
, $R^2 = H$

strong contractile activity in the guinea-pig ileum assay. ¹⁵⁸ 1,1-Dimethyl-5,6-dihydroxyindolinium chloride (170) has been isolated from a deep-water species of *Dercitus*, ¹⁵⁹ and 4,5,8-trihydroxyquinoline-2-carboxylic acid (171) was found to be the antimicrobial constituent of the Antarctic sponge *Dendrilla membranosa*. ¹⁶⁰ The indole alkaloid trikentrin A (172), which was isolated from *Trikentrion flabelliforme*, ¹⁶¹ has been synthesized using pyrolysis of an azide to form the 'pyrrole' ring. ¹⁶² Ptilocaulin (173), which is a cytotoxic metabolite of *Ptilocaulis* aff. *P. spiculifer*, ¹⁶³ has been synthesized in racemic form by using a photochemical rearrangement as the key step. ¹⁶⁴

A number of polycyclic heteroaromatic pigments have been isolated from marine sponges. Since some of these pigments are remarkably similar to pigments found in other sessile marine organisms, it has been suggested (but not demonstrated) that they might all be produced by microbial symbionts. Fascaplysin (174) is an unusual antimicrobial pigment from a Fijian species of Fascaplysinopsis. 165 The colour of the pigment petrosamine (175), which was isolated from a Caribbean species of Petrosia, is dependent on the polarity of the solvent and varies from

(190)

green (in THF) to purple (in water). 166 The structures of both fascaplysin (174) and petrosamine (175) were determined by Xray analysis. Dercitin (176) is a violet pigment from a deepwater species of Dercitus that exhibits antitumour, antiviral and immunomodulatory properties in vitro and has in vivo antitumour activity. 167 The structure of this novel acridine alkaloid was proposed on the basis of its spectral properties. Prianosins B (177), C (178), and D (179) are sulphur-containing alkaloids from Prianos melanos that have potent antineoplastic activity.¹⁶⁸ The structures of prianosins B—D (177)—(179) were elucidated by interpretation of spectral data. Discorhabdins A (180), B (181), C (182), and D (183) are cytotoxic pigments from a New Zealand species of Latrunculia. 169,170 Discorhabdin D (183) has also been found in an Okinawan species of Prianos. 170 The structure of discorhabdin A (180) was identical to that previously reported for prianosin A¹⁷¹ and the remaining discorhabdins were identified on the basis of spectral analysis. Amphimedine (184),

(189) $R = NH_2$

which is a cytotoxic constituent of a species of Amphimedon, ¹⁷² has been synthesized in eight steps (21—23% overall yield). ¹⁷³ A synthesis of mimosamycin (185), which was isolated from a species of Reniera, ¹⁷⁴ has been reported. ¹⁷⁵ In addition to the usual dibromotyrosine-derived metabolites found in Aplysina aerophoba, a Canary Islands specimen contained aplysinadiene (186) and the oxazolidinone (187). ¹⁷⁶ The structure of aplysinadiene (186) was elucidated by interpretation of spectral data and confirmed by synthesis, while the structure of the oxazolidinone (187), which appears to be a diastereoisomer of compounds reported previously, ^{177, 178} was determined by X-ray analysis.

(191)

A Red Sea species of *Smenospongia* contains the known prenylated quinone ilimaquinone (188)¹⁷⁹ and a new cytotoxic and antimicrobial aminoquinone called smenospongine (189).¹⁸⁰ A specimen of *Hippospongia* cf. *metachromia* from Okinawa has provided two new prenylated quinones, namely metachromins A (190) and B (191), in addition to the known

(208)

compound isospongiaquinone. 181 The metachromins, which were identified by spectroscopic analysis, exhibited in vitro activity against L1210 leukaemia cells and showed potent coronary vasodilating activity. The known metabolites xestoquinone (192)¹⁸² and halenaquinone (193)¹⁸³ were isolated from a species of *Adocia* from Truk, together with five new minor metabolites (194)—(198). 184 The most interesting of the new metabolites are adociaquinone A (196) and the mildly cytotoxic adociaquinone B (197), which were synthesized by reacting xestoquinone (192) with hypotaurine (a common though rarely reported metabolite of sponges), and 3ketoadociaquinone (198), which was synthesized in like manner from halenaquinone. A total synthesis of halenaquinone (193) and the corresponding hydroquinone from the optically active Wieland-Miescher ketone has provided the absolute configurations of the natural products. 185 Hexaprenylhydroquinone sulphate (199) was identified as an H, K-ATPase

(207)

inhibitor from a Japanese species of *Dysidea*. ¹⁸⁶ Both heptaprenylhydroquinone (200) and octaprenylhydroquinone (201) were isolated from a Tunisian specimen of *Hippospongia communis*. ¹⁸⁷

(209)

The first monoterpenoid to be isolated from a sponge is adriadysiolide (202), which was isolated from a North Adriatic species of *Dysidea*. ¹⁸⁸ The structure proposed for adriadysiolide (202) was confirmed by a stereorational total synthesis. A New Zealand species of *Eurypon* contains four new sesquiterpenes, three of which (203)–(205) possess the carbon skeleton of β -caryophyllene alcohol while the fourth (206) has the axane skeleton. ¹⁸⁹ The structures of (203)–(206) were deduced by interpretation of spectroscopic data. The alcohol (204) and corresponding methyl ether (205) are thought to be artefacts of the isolation procedure. A Japanese species of *Epipolasis* contained the sesquiterpenoids (+)-curcuphenol (207), dehydrocurcuphenol (208), and the related alcohol (209). ¹⁹⁰

21

The curcuphenols (207) and (208) showed strong inhibitory activity against H,K-ATPase. The isolation of thiofurodysinin (210) from an Australian specimen of *Dysidea avara* represents the first report of a sesquiterpene mercaptan from a sponge. ¹⁹¹ 5-Acetoxynakafuran 8 (211), 5-hydroxynakafuran 8 (212) and 5-ketonakafuran 8 (213) have been isolated from *Dysidea etheria* from Bermuda. ¹⁹² A report of a new isonitrile (214), isothiocyanate (215), and formamide (216) trio based on the guai-6-ene skeleton from an unidentified sponge provided some evidence that the formamide might be an artefact of isolation. ¹⁹³ Three new isonitrile-isothiocyanate pairs (217)—(222) have been found as minor metabolites of *Acanthella acuta*. ¹⁹⁴ The structures of (217)—(222) were elucidated by interpretation of

(231)

spectral data. The related sponge A. pulcherrima from Australia has yielded two new isothiocyanates (223) and (224) in addition to the known sesquiterpenes (225)—(227). 195 The syntheses of axisonitrile-1 (228) and axamide-1 (229), which are metabolites of Axinella cannabina, 196—198 and the corresponding 10-epiderivatives have been reported. 199 A simple synthesis of agelasidine A (230) from a species of Agelas employs a hetero-Claisen rearrangement. 201

(232)

Dictyodendrillolide (231) is a prenylated butenolide from a rare species of *Dictyodendrilla* from the Great Barrier Reef.²⁰² Spongia arabica from the Red Sea contains a new spongian derivative called spongialactone A (232).²⁰³ A specimen of *Hyatella intestinalis* contained three novel spongian diterpenes

(233) $R^1 = OAc$, $R^2 = H$

(236) $R^1 = OH$, $R^2 = H$

(237) $R^1 = OAc$, $R^2 = Ac$

(234) R = H

(235) R = OH

(238)

(239)

OR1 OR2

(241) $R^1 = R^2 = Ac$

(242) $R^1 = Ac$, $R^2 = H$

(243) $R^1 = R^2 = H$

(244) $R^1 = R^2 = R^3 = H$

(245) $R^1 = R^3 = H$, $R^2 = OH$

(247) $R^1 = OH$, $R^2 = R^3 = H$

(248) $R^1 = R^2 = H$, $R^3 = OAc$

(249) $R^1 = OH$, $R^2 = H$, $R^3 = OAc$

(246)

(250)

(251)

(253)

(233), (234), and (235), together with the known compounds (236)—(238) that had previously^{204, 205} been isolated from various species of *Spongia*.²⁰⁶ Ten new rearranged spongian diterpenes have been isolated from two Red Sea species of *Dysidea*.²⁰⁷ One species of *Dysidea* contained shahamins A—G (239)—(245), in addition to the known diterpene macfarlandin E (246).²⁰⁸ The second *Dysidea* species contained shahamins F (244), H (247), I (248), and J (249). The structures of the shahamins were proposed on the basis of their spectral data supported by chemical interconversions.²⁰⁷ Two new norditerpenes (250) and (251) have been isolated as minor constituents of *Spongionella gracilis*.²⁰⁹ The nor-diterpene (250) has an unprecedented carbon skeleton. The apocarotenoid xestodiol (252)²¹⁰ and the novel nor-diterpene xestenone (253)²¹¹ are metabolites of *Xestospongia vanilla* that were identified from their spectral properties. Reiswigins A (254) and B (255) are antiviral diterpenes from the deep-water sponge *Epipolasis*

(266)

reiswigi. 212 The structural elucidation by interpretation of spectral data is very sparsely outlined. Two diterpene alkaloids, agelasimine-A (256) and agelasimine-B (257), were obtained from Agelas mauritiana from Enewetak Atoll. 213 The stereochemistry of the diterpene portion of the agelasimines had been determined previously by X-ray analysis of a derivative. 214 The agelasimines (256) and (257) are cytotoxic and may act as calcium channel antagonists and a adrenergic blockers. One additional diterpene isonitrile, isokalihinol F (258), was obtained from a Fijian specimen of Acanthella cavernosa. 215

The absolute configuration of kurospongin (259), which is a novel fish-feeding inhibitor from an Okinawan specimen of Spongia, was determined by application of the Horeau method to the diol formed by reaction of (259) with ethyl magnesium bromide. 216 Carteriospongia flabellifera from the Great Barrier Reef was reported to contain 12,13-didehydrofurospongin-1 (260) and the homosesterterpene (261), both of which were identified from spectral properties. 217 The complete stereochemistry of variabilin (262), which was originally isolated from Ircinia variabilis, 218 has recently been defined. Both variabilin (262) and the corresponding 20E isomer (263) were isolated from a species of Sarcotragus, that also contained the sesterterpenes (264) and (265) and the C₂₁ furanoterpene (266). 219 A New Zealand species of Ircinia produces variabilin (262), the alcohol (265), and a series of four sesterterpenes

(276)

(267)—(270) that are oxygenated at C-5.²²⁰ An Australian species of *Ircinia* contains the antibacterial sesterterpene (271), the structure of which was elucidated by analysis of spectral data and chemical degradation.²²¹ Manoalide 25-monoacetate (272) and the corresponding 'linear' isomer, thorectolide monoacetate (273), were isolated from two different specimens of *Thorectandra excavatus*, while other specimens contained mixtures of the two compounds.²²² Cacospongionolide (274) is an unusual cytotoxic sesterterpene from a specimen of *Cacospongia mollior* from the Tyrrhenian Sea.²²³ The structure of cacospongionolide (274), which belongs to a new skeletal class, was elucidated by interpretation of spectral data. The

(274)

absolute configuration of manoalide (275), which is an antiinflammatory agent from Luffariella variabilis,²²⁴ has been determined by total synthesis of a derivative from 2-deoxy-Dribose.²²⁵ An additional synthesis of manoalide (275), which involves the intermediacy of seco-manoalide (276), has been outlined.²²⁶ The total synthesis of dysideapalaunic acid (277), which is an unreported(!) aldose reductase inhibitor from a Palauan species of Dysidea, has established the absolute configuration.²²⁷ The structure of suvanine, which is an acetyl cholinesterase inhibitor from a species of Coscinoderma found in both Fiji and Palau, has been revised from (278)²²⁸ to (279).²²⁹ A Californian sponge of the family Halichondriidae

(277)

$$R^{4}O$$
 OR^{2}
 OR^{2}
 OR^{3}
(286) $R^{1} = OAc$, $R^{2} = Ac$, $R^{3} = R^{4} = H$
(287) $R^{1} = OAc$, $R^{2} = R^{3} = R^{4} = H$

(288)
$$R^1 = R^3 = R^4 = H$$
, $R^2 = Ac$

(289)
$$R^1 = OAc$$
, $R^2 = R^3 = Ac$, $R^4 = H$

(290)
$$R^1 = OAc$$
, $R^2 = R^4 = Ac$, $R^3 = H$

contained the sulphated sesterterpene hydroquinone (280) and five sulphated sesterterpenes (281)—(285).²³⁰ The structures of halisulphates 1—5 (280)—(284) were determined by interpretation of spectral data and the structure (285) was proposed for halisulphate 6. The halisulphates possess antimicrobial and anti-inflammatory properties.

Pouosides A—E (286)—(290) are triterpene galactosides

from a Trukese species of Asteropus.²³¹ The novel carbon skeleton of the pouosides was identified by interpretation of spectral data. The same sponge also contained sarasinoside A₁ (291).²³² Sarasinosides A₁ (291), B₁ (292), and C₁ (293) are ichthyotoxic and cytotoxic constituents of Asteropus sarasinosum from Palau.²³³ The structure of the sapogenol was determined by X-ray analysis and the structures of the natural

όн

products were elucidated by analysis of spectral data. Two unusual steroids, agnatasterones A (294) and B (295), were isolated from Axinella agnata. Eight new polyhydroxylated sterols based on the 5α -cholest-7-ene- 2α , 3β , 5α , 6β , 9α , 11α , 19-heptol nucleus (296) were found in Dysidea etheria. A related sterol, 11β , 19-epoxycholest-6-en- 3β , 5α , 8α , 9α -tetrol (297) is produced by Dysidea tupha. The ring A contracted sterols, anthosterones A (298) and B (299), were obtained from Anthoracuata graceae. The structure of hipposterol (300), which is a 5,6-secosterol from Hippospongia communis, was confirmed by synthesis. Penasterol (301) is a novel triterpene from an Okinawan species of Penares, that shows potent antileukemic activity. 239

(319) R = OAc

8 Coelenterates

In a departure from previous reviews in this series, the metabolites of coelenterates are grouped by chemical class rather than phylogenetically by Order. The prostanoids and other eicosanoids from coelenterates continue to evoke interest. Leiopathic acid (302) and two known eicosanoids, (303) and (304), were isolated from a black coral, Leiopathes sp., collected at St Paul Island in the South India Ocean. 240 A new epoxyprostanoid (305) which has antiproliferative properties was isolated as a minor metabolite of the Japanese stoloniferan coral Clavularia viridis. 241 Total synthesis of several possible isomers has allowed the structures of (7E)- and (7Z)-punagladin 4 to be revised to (306) and (307) respectively from the corresponding 12S epimers.²⁴² A second synthesis of punaglandin 4 (306) involved the enzymic resolution of the key chlorocyclopentene intermediate.243 The absolute configurations of the lactones (308) and (309) from the

gorgonian Plexaura flava²⁴⁴ were determined by synthesis of their optical enantiomers from (S)-lactic acid. 245 The (S,S) configuration of (-)-bissetone (310), which is a metabolite of Briareum polyanthes, 246 has been established by total synthesis from D-glucose.247

An unnamed soft coral of the genus Primnoeides is the source of a family of five sesquiterpenes (311)—(315) that contain a new carbon skeleton.²⁴⁸ The structure of the alcohol 3-hydroxy-6-methoxyprimnatrienone (312) was determined by X-ray analysis and the remaining primnatriene derivatives were identified by comparison of spectral data. Subergorgic acid (316), which is a cardiotoxic sesquiterpene from the gorgonian Subergorgia suberosa,249 has been synthesized by a stereoselective route.250

Mayolide A (317) is a novel secocembranoid that was isolated from the soft coral Sinularia mayi together with the cembranoid lactones mayolides B (318), C (319), and D (320).251 The

structures of the mayolides were elucidated by interpretation of spectral data and interconversion with known compounds. It has been suggested that some of the six minor cembranoids (321)—(326) isolated from S. mayi might be plausible precursors of the major cembranolides found in the soft coral. The structures of sinulariols C (321) and D (322), sinularial A (323), sinularic acid (324), and sinularones A (325) and B (326) were elucidated by physicochemical methods. ²⁵² The oxidation chemistry of sarcophytol A (327), which is a potent anti-

tumour-promoter from Sarcophyton glaucum, has been reported. 253 Three known cytotoxic cembranolides were isolated from Sinularia mayi: the stereochemistry of cembranolides (328) and (329), previously reported from *Lobophytum* pauciflorum, 254, 255 were determined by X-ray crystallography and spectral methods, respectively, and the absolute configuration of denticulatolide (330), which is a metabolite of L. denticulatum, 256 was determined by X-ray analysis of a p-bromobenzoate derivative. 257 The cembranolide (331), which is a metabolite of the soft coral Lobophytum michelae, 258 has been synthesized.259 The strategies employed in the synthesis of cembranes and cembranolides have been reviewed in detail.²⁸⁰

The minor metabolites of both Atlantic and Pacific specimens of Gersemia rubiformis have been compared.261 The Pacific coast animals yielded two new pseudopterane diterpenoids, isogersemolides A (332) and B (333), and four new cembranoids: isoepilophodiones A (334), B (335), and C (336) and rubifol (337). The Atlantic coast animals contained 3acetoxy- β -cubebene (338). The proposed structures are all based on spectroscopic analysis and chemical interconversion. The cytotoxic activity of a Japanese species of Clavularia is due

(339) R = H

(340) R = Ac

(343) R = H

(344) R = Ac

(346)
$$R^1 = R^2 = H$$

(347) $R^1 = OCOC_3H_7$, $R^2 = H$

OCOC₃H₇

(348)
$$R^1 = H$$
, $R^2 = OH$

(349) $R^1 = H$, $R^2 = Me$

(350) $R^1 = H$, $R^2 = Et$

(351) $R^1 = OH$, $R^2 = Me$

(352) $R^1 = OAc$, $R^2 = Me$

to stolonidiol (339), the acetate (340) and claenone (341), the structures of which were determined on the basis of spectroscopic data and the X-ray analysis of a derivative. ^{262, 263} A related diterpene (342) has been reported from a Chinese species of Clavularia. ²⁶⁴ Sclerophytins A (343) and B (344) are two cytotoxic diterpenes from the soft coral Sclerophytum capitalis from Enewetak that were identified by interpretation of spectral data. ²⁶⁵ A new cladiellane diterpene called alcyonin (345) was isolated from Sinularia flexibilis and identified by using spectroscopic and chemical methods. ²⁶⁶ Three similar diterpenes, litophynins A (346), B (347), and C (348), that inhibit insect growth in the silkworm Bombyx mori, were isolated from a Japanese species of Litophyton. ^{267, 268} The

structures and absolute configuration of the litophynins were established on the basis of spectral and chemical evidence. The Mediterranean stolonifer *Sarcodictyon roseum* contains six novel diterpenoids that are esterified by *N*-methylurocanic acid.^{269,270} The structures of sarcodictyins A (349), B (350), C (351), D (352), E (353), and F (354) were elucidated by spectroscopic methods and the absolute configurations were determined by Horeau's method.

Four new xenicane diterpenes – havannachlorhydrine-11(19) (355), havannadichlorhydrine-7(18), 11(19), (356), havannachlorhydrine-7(18) (357), and 13-desacetylhavannachlorhydrine-11(19) (358) – were isolated from *Xenia membranacea* and their structures were determined by correlation with a derivative

that was subjected to X-ray analysis.²⁷¹ Acalycixeniolides A (359) and B (360) are cytotoxic norditerpenes of the xenicane type that were isolated from Acalycigorgia inermis and identified by spectroscopic studies.²⁷² A related allene, ginamallene (361), was isolated from four Japanese species of Acalycigorgia.²⁷³ Two novel C₂₄ 'acetoacetylated' diterpenes, antheniolides A (362) and B (363), were obtained from Anthelia glauca.²⁷⁴ The structures of the antheniolides were established by using spectroscopic methods and a biosynthetic route from xeniaphyllane²⁷⁵ is proposed.

(380) R = Me

An Indopacific gorgonian of the genus Solenopodium contains six new diterpenes of the briarane class. The structures of solenolides A—F (364)—(369) were assigned on the basis of spectral analyses and chemical modifications. Solenolides A (364), D (367), and E (368) are antiviral and anti-

inflammatory agents while solenolide F (369) shows only antiinflammatory activity. Four new briaranes (370)—(373) have been isolated from *Briareum steckei*.²⁷⁷ The structures of (372) and (373) were determined by X-ray analyses and the major metabolites (370) and (371) were identified by spectroscopic means. Brianthein V (374) is a new cytotoxic and antiviral diterpene from *Briareum asbestinum*.²⁷⁸ The relatively simple briaranes, verecynarmins B (375), C (376), and D (377), that were found in both the sea pen *Veretillum cynomorium* and the nudibranch molluse *Armina maculata*, all undergo very slow conformational interconversion in solution.²⁷⁹ The structures of the verecynarmins were deduced by interpretation of their complex spectral data and by chemical modification.

(382) (epimer at *)

Asperketals A—F (378)—(383) are new diterpenes of the dilophol class from the gorgonian Eunicea asperula, the

(384) $R^1 = Ac$, $R^2 = H$ (385) $R^1 = H$, $R^2 = Ac$

(389) X = H

(390) X = OAc

(391) X = H(1,2-dihydro)

(397) $R^1 = R^2 = R^3 = Ac$

(398) $R^1 = R^2 = Ac$, $R^3 = H$

(399) $R^1 = R^3 = H$, $R^2 = Ac$

(400) $R^1 = R^2 = R^3 = H$

(402) R = Me, X = H

(403) R = Me, X = Br

(404) R = H, X = H

(405) R = H, X = Br

(392) X = Y = H

(393) X = OAc, Y = H

(394) X = H, Y = H (1,2-dihydro)

(395) X = OAc, Y = H(1,2-dihydro)

(396) X = H, Y = OAc (1,2-dihydro)

structures of which were assigned on the basis of chemical and spectroscopic studies.²⁸⁰ A Caribbean gorgonian of the genus Pseudopterogorgia contains three new pseudopterosins (384)—(386) and one new seco-pseudopterosin (387) that were identified by spectral and chemical experiments.²⁸¹ A mechanism linking the two series of compounds through the hydroperoxide (386) is described. (-)-Pseudopterosin A (388), which is an anti-inflammatory diterpene from the gorgonian Pseudopterogorgia elizabethae, 282 has been synthesized from (-)-limonene. 283 Eight new steroidal lactones (389)—(396) of the withanolide class have been isolated from a Trukese species of Minabea²⁸⁴ and five new hippurins (397)—(401) were obtained from an Indian specimen of Isis hippuris. 285

A scleractinian coral of the genus Tubastrea from the Philippines has yielded two new aplysinopsin derivatives, (402) and (403), each of which occurred as a 5:2 mixture of E and Z stereoisomers.²⁸⁶ Two similar compounds (404) and (405) were isolated as 1:1 mixtures of stereoisomers from Leptopsammia pruvoti from the Mediterranean. 286 The structures of the new aplysinopsin derivatives were derived by interpretation of spectral data and confirmed by synthesis. The E/Zisomerization is a photochemical rearrangement. Tubastraine (406) is a novel alkaloid from the stony coral Tubastrea micrantha that was isolated along with heteronemin, which is a well-known sponge metabolite²⁸⁷ and most likely a contaminant.288 Tubastraine looks suspiciously like the bis-pbromobenzoate of the terrestrial alkaloid rohitukine. The

(409) $R^1 = Et$, $R^2 = Br$

hydroid Aglaophenia pluma contains three alkaloids, (407), (408), and (409), that are reminiscent of the β -carbolines found in tunicates.²⁸⁹ The structures of (407)—(409) were deduced from spectral data and confirmed by synthesis.

9 Bryozoans

Chemical studies of bryozoans are clearly limited by collection constraints and few new results have been reported. Four new bromine-containing alkaloids, amanthamides C—F (410)—(413), have been isolated from various collections of Amathia wilsoni from Tasmania. The ratios of the amanthamides, which were identified by spectral analysis, vary according to the collection site. The structure of hinckdentine-A (414), which is a novel alkaloid from the Tasmanian bryozoan Hinksinoflustra denticulata, was determined by X-ray analysis. The ratios of the A (415) and B (416) are the latest β -

lactams to be isolated as minor constituents of *Chartella papyracea*. ²⁹² The structures of the chartellamides were deduced by interpretation of spectral data.

10 Molluscs

The metabolites isolated from sea hares are invariably obtained from an algal dietary source. Aplysiaterpenoids A (81) and B (417) are two new halogenated monoterpenes from Aplysia kurodai.293 Aplysiaterpenoid A (81) has been reported as a major metabolite of *Plocamium hamatum*⁹¹ and (417) is closely related to a known algal metabolite from a species of *Plocamium*.²⁹⁴ The structures of the aplysiaterpenoids were determined by X-ray analyses. The eudesmane sesquiterpenes, lankalapuols A (418) and B (419), that were isolated from a Sri Lankan specimen of Aplysia dactylomela are closely related to certain Laurencia (red alga) metabolites. The structures of the lankalapuols, which are antipodal, were based on spectral studies and the structure and absolute configuration of lankalapuol A (418) was determined by an X-ray analysis performed on the corresponding acetate.²⁹⁵ A Canary Islands specimen of A. dactylomela contained a dolabellane epoxide (420) that is a typical constituent of brown algae. 296 A stereocontrolled synthesis of aplysin (421) and debromoaplysin (422), which are metabolites of Aplysia kurodai, involves the ring expansion of a cyclobutane intermediate.297

HO HO OH

(424)

$$\begin{array}{c}
CH_2OR \\
H \rightarrow OR \\
H \rightarrow OR \\
CH_2OR \\
H \rightarrow OR \\
CH_2OR
\\
CH_2O$$

Yessotoxin (423) is a polyether metabolite from the scallop *Patinopecten yessoensis* that has been implicated in diarrhetic shellfish poisoning. The structure and partial stereochemistry of yessotoxin (423) was deduced from spectral data. *Patinopecten yessoensis* also contains a series of eight polyhydroxylated sterols based on the 3β , 5α , 6β -cholest-7-ene (424) and 3β , 5α , 6β , 9α -cholest-7-ene (425) sterol nuclei. Buccinulin (426) and the known compound kelletinin I (427), which was previously obtained from *Kelletia kelletii*, 300 have been isolated from the Mediterranean whelk *Buccinulum corneum* by two

research groups. 301, 302 The structure of buccinulin (426) was confirmed by synthesis. 302 Two ichthyotoxic diacylglycerols, umbraculumins A (428) and C (429), were isolated from the rare Mediterranean opisthobranch *Umbraculum mediterraneum*. 303 The Spanish dancer nudibranch *Hexabranchus sanguineus* contained dihydrohalichondramide (137) and tetrahydrohalichondramide (430), which were obtained by reduction of halichondramide (142) from the sponge *Halichondria* sp. 133 The distribution of the macrolides within the nudibranch and its egg ribbons and the defensive value of the metabolites have

(443) R = CH2OH

been reported. ³⁰⁴ Pulo'upone (431), which is a metabolite of the cephalaspidean mollusc *Philinopsis speciosa*, ³⁰⁵ has been synthesized by two routes, both of which use an intramolecular Diels-Alder strategy. ^{306, 307} Prosurugatoxin (432), which is a toxin from the Japanese ivory shell *Babylonia japonica*, ³⁰⁸ was synthesized in racemic form. ³⁰⁹

The known sesquiterpene olepupuane (433)³¹⁰ has been isolated from *Dendrodoris limbata* and *D. grandiflora*.³¹¹ The Australian nudibranch *Ceratosoma brevicaudatum* contained the known terpenes dehydrodendrolasin (434),³¹² dehydrolasiosperman (435), and thiofurodysinin acetate (437),³¹³ and three new metabolites: *cis*-dehydrodendrolasin (436),

(methylthio)furodysinin (438), and dithiofurodysinin disulfide (439).³¹⁴ These six sesquiterpenes are all assumed to be of sponge origin. The Mediterranean dorid nudibranch *Doris verrucosa* contains two ichthyotoxic diterpene glycerides, verrucosins-A (440) and -B (441), that were identified from spectral and chemical evidence and an X-ray determination of the structure of (441).³¹⁵ The ascoglossan *Elysia halimedae* feeds preferentially on the green seaweed *Halimeda macroloba* and modifies the major algal diterpenoid, halimadatetraacetate (442), by reduction to the corresponding alcohol (443), which it then uses as a defensive compound.³¹⁶ The metabolites of *Chromodoris funerea* collected from a marine lake in Palau were

$$\begin{array}{c|c} & & \\ & &$$

(446)
$$R^1, R^2 = O$$

(447) $R^1 = H, R^2 = OH$

(450)
$$R^1 = H$$
, $R^2 = OH$

(451)
$$R^1 = OH$$
, $R^2 = H$
(454) $R^1 = H$, $R^2 = OMe$

quite different from those previously reported 317 from specimens taken from a nearby lagoon. Two new sesterterpenes, luffariellins-C (444) and D (445), were identified from their spectral data. 318

The pulmonate mollusc Siphonaria grisea contains two new linear polypropionate metabolites, siphonarienedione (446) and siphonarienolone (447), that were identified by interpret-

ation of spectral data and by chemical interconversion and degradation to (2S,4S,6S)-nonanoic acid.³¹⁹ Aglajne-2 (448) and aglajne-3 (449) are two new propionate-derived compounds found in both *Aglaja depicta* and its prey *Bulla striata*.³²⁰ Their structural and partial stereochemical elucidation was accomplished by using spectroscopic and chemical methods.

11 Tunicates

During the past year there has clearly been a surge of interest in the metabolites of tunicates. This is undoubtedly due to the high incidence of promising pharmacological properties found in the crude extracts of tunicates. The didemnenones (450)—(453) are a series of four cytotoxic cyclopentenone derivatives from didemnid tunicates. ³²¹ Didemnenones A (450) and B (451) were isolated from *Trididemnum cyanophorum* while didemnenones C (452) and D (453) are from *Didemnum voeltzkowi*. The structures of the didemnenones were elucidated by relating their spectral properties to those of a methyl acetal derivative (454), the structure of which was determined by X-ray analysis. Bistramide A is a cytotoxin from *Lissoclinum bistratum* that has been partially characterized by NMR spectroscopy. ³²² Two cytotoxins, iejimalides A (455) and B (456) have been isolated from an Okinawan specimen of *Eudistoma* cf. *rigida*. ³²³ Extensive NMR experiments were used to elucidate the structures of (455) and (456) but their relative

(458)
$$R^1 = H$$
, $R^2 = OH$

(459)
$$R^1 = R^2 = OH$$

stereochemistry remains undefined. Patellazoles A (457), B (458), and C (459) are three cytotoxic macrolides from Lissoclinum patella. The structural elucidations of patellazole B (458)³²⁴ and patellazole C (459)³²⁵ depend on the interpretation of spectral data that did not permit stereochemical assignments. The 2,3-threo-13,14-erythro-stereochemistry of aplidiaspingosine (460), which is an antimicrobial and cytotoxic terpenoid from a species of Aplydium, 326 has been defined by total synthesis. 327 The structures of two thiazoles, (461) and (462), and an imidazole (463) from Aplydium pliciferum were elucidated by interpretation of spectral data and confirmed by synthesis.328 A high-yielding two-phase cyclization reaction has been employed in a synthesis of didemnins A (464), B (465), and C (466). 329, 330 The structural revision, total syntheses and pharmacological properties of didemnins A-E (464)-(468), which are cytotoxic, antiviral, and immunosuppressive agents from Trididemnum solidum, have been reviewed.331 The same review reports the structural elucidation of tunichlorin (469), which is a nickel-containing blue-green coloured porphynoid.

(464) R = H

$$(465) R = -C \qquad N$$

O OH
$$\parallel \ \ \, \parallel$$
(466) R = $-C-CH-CH_3$

22 NPR 7

$$\begin{array}{c} & & & \\ & &$$

Br
$$\stackrel{\text{NMe}_2}{H_2N}$$
 (471) (472)

(481) R = Br (482) R = H

Eudistomin K sulphoxide (470), which is an antiviral agent from the New Zealand ascidian *Ritterella sigillinoides*, ³³² has been synthesized from eudistomin K (471), the structure and absolute configuration of which were determined by *X*-ray analysis. ³³³ *Eudistoma fragum* contained *N,N*-dimethyl-5-bromotryptamine (472), which had previously been found in the sponge *Smenospongia aurea*, ³³⁴ and a new alkaloid, woodinine (473), the structure of which was proposed on the basis of its spectral properties. ³³⁵ The structure of 7-bromo-2,4(1*H*,3*H*)-quinazolinedione (474), which accompanies 6-bromoindole-3-carboxaldehyde in *Pyura sacciformis*, has been confirmed by synthesis. ³³⁶

The structure of shermilamine A (475), which is an orange pigment from a species of Trididemnum from Guam, was determined by X-ray analysis.337 A group of three somewhat similar alkaloids - segoline A (476), isosegoline A (477), and nor-segoline (478) - were obtained from a Red Sea species of Eudistoma. 338 The structure of segoline A (476) was determined by X-ray analysis and the remaining structures were proposed on the basis of comparison of spectral data. A second collection of the same species of Eudistoma from Eilat contained eilatin (479), the structure of which was determined by X-ray analysis.³³⁹ The structure (480) that was assigned by interpretation of long-range C/H coupling data to 2bromoleptoclinidinone, 340 which is an alkaloid from a species of Leptoclinides, is now known to be incorrect. The correct structure of 2-bromoleptoclinidinone (481)341 is that of a brominated derivative of ascididemnin (482), which is a cytotoxic alkaloid from an Okinawan species of Didemnum. 342 The structure of ascididemnin (482) was elucidated by interpretation of spectral data. Cystodytins A (483), B (484), and C (485) are tetracyclic alkaloids from *Cystodytes dellechiajei* that exhibit potent antineoplastic activity. 343 The structures of the cystodytins (483)—(485) were elucidated by interpretation of spectral data.

(486)
$$R^1 = R^3 = H$$
, $R^2 = R^4 = Me$, $R^5 = OH$

(487)
$$R^1 = H$$
, $R^2 = R^3 = R^4 = Me$, $R^5 = OH$

(488)
$$R^1 = Me$$
, $R^2 = R^3 = R^4 = R^5 = H$

$$H_2N$$
 H_2N
 H_2N
 H_2N
 H_3N
 H_4N
 H_5N
 H_5N
 H_5N
 H_7N
 H_7N

(490)
$$R^1 = R^2 = OH$$

(491)
$$R^1 = H$$
, $R^2 = OH$

(492)
$$R^1 = R^2 = H$$

The ascidian *Didemnum characeum* contained a series of four alkaloids (486)—(489) that are closely related to the lamellarins³⁴⁴ previously found in the prosobranch mollusc *Lamellaria* sp. The structure of lamellarin E (486) was determined by *X*-ray analysis and those of lamellarins F (487), G (488), and H (489) were elucidated by interpretation of spectral data.³⁴⁵ The tunichromes are a series of reducing blood pigments from *Ascidia nigra* and *Molgula manhattensis*.³⁴⁶ The structures of the very unstable tunichromes *An*-1 (490), *An*-2 (491), *An*-3 (492), *Mm*-1 (493), and *Mm*-2 (494) were deduced by spectral analyses. Evidence of complex formation between

vanadium and the tunichromes is presented but a stable complex was not isolated.

12 Echinoderms

The sea stars continue to yield new saponins and polyhydroxylated sterols but these compounds are predominantly minor variations of familiar structural types. A review of the secondary metabolites of echinoderms was particularly useful because of its coverage of the Russian literature.¹² In addition to known saponins, the Pacific sea star Asterias amurensis

(495)
$$R^1 = a$$
, $R^2 = XyI$, $R^3 = GaI$, $R^4 = Qui$, $R^5 = H$
(496) $R^1 = b$, $R^2 = XyI$, $R^3 = GaI$, $R^4 = Qui$, $R^5 = H$
(497) $R^1 = c$, $R^2 = XyI$, $R^3 = GaI$, $R^4 = Qui$, $R^5 = H$
(498) $R^1 = d$, $R^2 = XyI$, $R^3 = GIu$, $R^4 = Qui$, $R^5 = H$
(503) $R^1 = e$, $R^2 = Qui$, $R^3 = GIu$, $R^4 = Fuc$, $R^5 = H$
(504) $R^1 = f$, $R^2 = XyI$, $R^3 = GIu$, $R^4 = Fuc$, $R^5 = GaI$
(505) $R^1 = g$, $R^2 = XyI$, $R^3 = GIu$, $R^4 = Fuc$, $R^5 = H$
(506) $R^1 = h$, $R^2 = Qui$, $R^3 = GIu$, $R^4 = Fuc$, $R^5 = H$

Fuc = fucose, Gal = galactose, Qui = quinovose, Xyl = xylose; all sugars are in the pyranose form and linkages are β .

contained four new saponins - asterosides A (495), B (496), C (497), and D (498) - that differ primarily in the structures of their steroidal side chains. 347 The same sea star also contains four new polyhydroxysteroidal glycosides; amurensosides A (499), B (500), C (501), and D (502).347 The Japanese sea star

Asterina pectinifera likewise contains four new saponins pectiniosides A (503), B (504), C (505), and D (506) 348,349 – and two new polyhydroxylated steroids: $(24S)5\alpha$ -cholestane- 3β , 6α , 8, 15α , 24-pentol (507) and (25S)- 5α -cholestane- $3\beta, 4\beta, 6\alpha, 7\alpha, 8, 15\beta, 16\beta, 26$ -octol (508). 350

A number of new polyhydroxysteroidal glycosides have been reported. Indicosides B (509) and C (510) were isolated from Astropecten indicus, 351 thromidioside (511) was found in Thromidia catalia, 352 and glacialosides A (512) and B (513) were obtained from Marthasterias glacialis. 353 All of these compounds are minor metabolites and were accompanied by known saponins. The North Pacific sea star Distolasterias nipon contains asterosaponins D_1 (514), D_2 (515), and D_3 (516), 354

while specimens of *Echinaster sepositus* from Madagascar provided echinasterosides B_1 (517) and B_2 (518), both of which contain an unusual $1 \rightarrow 3$ glycosidic linkage.³⁵⁵

The eicosanoids 8-(R)-HETE (519) and the corresponding 17,18-didehydro derivative have been found in *Patiria miniata*. ³⁵⁶ This finding is of interest since it was known that 8-(R)-HETE (519) can trigger oocyte maturation in starfish. ³⁵⁷ Studies of the glycosphingolipids from *Acanthaster planci* have

 $(520) \ \ R^1 = C_{12}H_{25}, \ \ R^2 = C_{22}H_{46}$

(521) $R^1 = C_{18}H_{37}$, $R^2 = C_{14}H_{29}$

(522) $R^1 = -(CH_2)_8CH \stackrel{Z}{=} CH - C_8H_{17}, R^2 = C_{14}H_{29}$

(523) R = C₂₀H₄₁

(524) R = C₂₁H₄₃

(525) R = C₂₂H₄₅

(526) $R^1 = C_{12}H_{25}$, $R^2 = C_{22}H_{45}$

(527) $R^1 = -(CH_2)_8 \cdot CH \stackrel{Z}{=} CH \cdot C_8 H_{17}, R^2 = C_{14} H_{29}$

(528) $R^1 = H$, $R^2 = H$

(529) $R^1 = OH$, $R^2 = H$

(530) $R^1 = OH$, $R^2 = Me$

View Article Online

yielded six new cerebrosides, acanthacerebrosides A-F (520)—(525), 358, 359and two new ceramide lactosides, acanthalactosides A (526) and B (527).360

13 Miscellaneous

The cytotoxic constituents of the marine worm Cephalodiscus gilchristi (Pterobranchia) have been identified as pyrazine alkaloids derived from two different steroidal subunits. The structure of cephalostatin 1 (528) was determined by X-ray crystallographic analysis361 and the structures of cephalostatins 2 (529), 3 (530), and 4 (531) were elucidated by interpretation of spectral data. 362, 363

The fluorescent compound in the shrimp Euphausia pacifica was found to be the bile pigment (532).364 Compound F (532) was identified by interpretation of spectral data, chemical conversions, and synthesis of a degradation product. The stereochemistry of the 15,16-olefinic bond was not determined.

A new tetrodotoxin derivative, 11-nortetrodotoxin (533), has been isolated as a minor constituent of the liver of the pufferfishes Fugu niphobles, F. pardalis, and F. poecilonotus. 365

14 References

- 1 D. J. Faulkner, Nat. Prod. Rep., 1984, 1, 251.
- 2 D. J. Faulkner, Nat. Prod. Rep., 1984, 1, 551.
- 3 D. J. Faulkner, Nat. Prod. Rep., 1986, 3, 1.
- 4 D. J. Faulkner, Nat. Prod. Rep., 1987, 4, 539.
- 5 D. J. Faulkner, Nat. Prod. Rep., 1988, 5, 613.
- 6 M. J. Garson, Nat. Prod. Rep., 1989, 6, 143. 7 V. J. Paul and W. Fenical, in Bioorganic Marine Chamistry', ed. P. J. Scheuer, Springer-Verlag, Berlin, 1987, Vol. 1, p. 1.
- 8 P. Karuso, in 'Bioorganic Marine Chemistry', ed. P. J. Scheuer, Springer-Verlag, Berlin, 1987, Vol. 1, p. 31. N. Fusetani, in 'Bioorganic Marine Chemistry', ed. P. J. Scheuer,
- Springer-Verlag, Berlin, 1987, Vol. 1, p. 61.
- M. H. G. Munro, R. T. Luibrand, and J. W. Blunt, in 'Bioorganic Marine Chemistry', ed. P. J. Scheuer, Springer-Verlag, Berlin, 1987, Vol. 1, p. 93.
- 11 R. J. Quinn, in 'Bioorganic Marine Chemistry', ed. P. J. Scheuer, Springer-Verlag, Berlin, 1988, Vol. 2, p. 1. V. A. Stonik and G. B. Elyakov, in 'Bioorganic Marine Chem-
- istry', ed. P. J. Scheuer, Springer-Verlag, Berlin, 1988, Vol. 2, p.

- 13 P. W. Sammarco and J. C. Coll, in 'Bioorganic Marine Chemistry', ed. P. J. Scheuer, Springer-Verlag, Berlin, 1988, Vol. 2, p. 87.
- 14 K. Tachibana, in 'Bioorganic Marine Chemistry', ed. P. J. Scheuer, Springer-Verlag, Berlin, 1988, Vol. 2, p. 117.
- 15 F. J. Schmitz, D. J. Vanderah, K. H. Hollenbeak, C. E. L. Enwall, Y. Gopichand, P. K. Sengupta, M. B. Hossain, and D. van der Helm, J. Org. Chem., 1983, 48, 3941.

 16 A. C. Stierle, J. H. Cardellina, II, and F. L. Singleton, Experientia,
- 1988, 44, 1021.
- 17 A. Guerriero, M. D'Ambrosio, V. Cuomo, F. Vanzanella, and F. Pietra, Helv. Chim. Acta, 1988, 71, 57.
- 18 A. Yokoyama, M. Murata, Y. Oshima, T. Iwashita, and T. Yasumoto, J. Biochem., 1988, 104, 184.
- 19 K. Torigoe, M. Murata, T. Yasumoto, and T. Iwashita, J. Am. Chem. Soc., 1988, 110, 7876.
- 20 Y. Murakami, Y. Oshima, and T. Yasumoto, Bull. Jpn. Soc. Sci. Fish., 1982, 48, 69.
- 21 M. Murakami, K. Makabe, K. Yamaguchi, S. Konosu, and M. R. Walchli, *Tetrahedron Lett.*, 1988, **29**, 1149.

 22 J. Kobayashi, M. Ishibashi, M. R. Walchli, H. Nakamura, and T.
- Sasaki, J. Am. Chem. Soc., 1988, 110, 490.
- 23 K. Gustafson and R. J. Andersen, Tetrahedron, 1985, 41, 1101.
- 24 S. J. Coval, G. R. Schulte, G. K. Matsumoto, D. M. Roll, and P. J. Scheuer, Tetrahedron Lett., 1985, 26, 5359.
- 25 J. Kobayahi, M. Ishibashi, H. Nakamura, Y. Hirata, T. Yamasu, T. Sasaki, and Y. Ohizumi, Experientia, 1988, 44, 800.
- 26 R. E. Moore and M. Entzeroth, Phytochemistry, 1988, 27, 3101.
- 27 J. S. Mynderse, A. H. Hunt, and R. E. Moore, J. Nat. Prod., 1988, 51, 1299.
- 28 J. H. Cardellina, II, R. E. Moore, E. V. Arnold, and J. Clardy, J. Org. Chem., 1979, 44, 4039.
- T. Sato, H. Maeno, T. Noro, and T. Fujisawa, Chem. Lett., 1988,
- 30 W. H. Gerwick, A. Lopez, G. D. Van Duyne, J. Clardy, W. Ortiz, and A. Baez, Tetrahedron Lett., 1986, 27, 1979.
- 31 R. Alonso and A. Brossi, Tetrahedron Lett., 1988, 29, 735.
- 32 N. R. Ayyangar, R. A. Khan, and V. H. Deshpande, Tetrahedron Lett., 1988, 29, 2347.
- 33 V. J. Paul and K. L. Van Alstyne, Coral Reefs, 1988, 6, 263.
- 34 V. J. Paul, P. Ciminiello, and W. Fenical, Phytochemistry, 1988, 27, 1011.
- 35 W. Boland, 'Bioflavour '87', ed. P. Schreier, de Gruyter, Berlin, 1988, p. 199.
- 36 D. G. Müller, W. Boland, U. Becker, and T. Wahl, Biol. Chem. Hoppe-Seyler, 1988, **369**, 655. W. Boland, U. Flegel,
- G. Jordt, and D. G. Müller, Naturwissenschaften, 1987, 74, 448.
- 38 R. E. Moore, and G. Yost, J. Chem. Soc., Chem. Commun., 1973,
- 39 M. Asaoka, K. Takenouchi, and H. Takei, Chem. Lett., 1988, 921.
- 40 A. J. Blackman, G. I. Rogers, and J. K. Volkman, J. Nat. Prod., 1988, 51, 158.
- 41 A. J. Blackman, G. I. Rogers, and J. K. Volkman, Phytochemistry,
- 1988, 27, 3686. 42 V. Amico, M. Piattelli, F. Cunsolo, P. Neri, and G. Ruberto, Gazz. Chim. Ital., 1988, 118, 193.
- 43 V. Amico, F. Cunsolo, M. Piattelli, and G. Ruberto, Phytochemistry, 1984, 23, 2017.
- V. Amico, G. Oriente, M. Piattelli, and G. Ruberto, Gazz. Chim. Ital., 1984, 114, 169.
- 45 V. Amico, G. Giaccone, M. Piattelli, and G. Ruberto, Phytochemistry, 1988, 27, 1069.
- 46 W. H. Gerwick and W. Fenical, J. Org. Chem., 1981, 46, 22.
- 47 P. V. Fish, G. Pattenden, and S. T. Hodgson, Tetrahedron Lett., 1988, 29, 3857.
- 48 I. A. Van Altena, Aust. J. Chem., 1988, 41, 49.
- 49 B. N. Ravi, P. T. Murphy, R. O. Lidgard, R. G. Warren, and R. J. Wells, Aust. J. Chem., 1982, 35, 171.
- 50 R. Kazlauskas, P. T. Murphy, and R. J. Wells, Experientia, 1978, **34**, 156.
- 51 L. Semmak, A. Zerzouf, R. Valls, B. Banaigs, G. Jeantry, and C. Francisco, Phytochemistry, 1988, 27, 2347.
- 52 M. O. Ishitsuka, T. Kusumi, and H. Kakisawa, J. Org. Chem., 1988, **53**, 5010.
- 53 C. Tringali, G. Oriente, M. Piattelli, C. Geraci, G. Nicolosi, and E. Breitmaier, Can J. Chem., 1988, 66, 2799.
- 54 J. Finer, J. Clardy, W. Fenical, L. Minale, R. Riccio, J. Battaile, M. Kirkup, and R. E. Moore, J. Org. Chem., 1979, 44, 2044.

- 55 M. Ishitsuka, T. Kusumi, and H. Kakisawa, Tetrahedron Lett., 1982, 23, 3179.
- 56 H. Nagaoka, K. Kobayashi, and Y. Yamada, Tetrahedron Lett., 1988, 29, 5945.
- 57 D. R. Hirschfeld, W. Fenical, G. H. Y. Lin, R. M. Wing, P. Radlick, and J. J. Sims, J. Am. Chem. Soc., 1973, 95, 4049.
- 58 E. Fattorusso, S. Magno, L. Mayol, C. Santacroce, D. Sica, V. Amico, G. Oriente, M. Piattelli, and C. Tringali, J. Chem. Soc., Chem. Commun., 1976, 575.
- 59 B. Danise, L. Minale, R. Riccio, V. Amico, G. Oriente, M. Piattelli, C. Tringali, E. Fattorusso, S. Magno, and L. Mayo, Experientia, 1977, 33, 413.
- 60 D. J. Faulkner, B. N. Ravi, J. Finer, and J. Clardy, Phytochemistry, 1977, 16, 991.
- 61 J. T. Vazquez, M. Chang, K. Nakanishi, E. Manta, C. Pérez, and J. D. Martín, J. Org. Chem., 1988, 53, 4797.
- 62 K. Kurata, K. Shiraishi, T. Takato, K. Taniguchi, and M. Suzuki, *Chem. Lett.*, 1988, 1629.
- 63 A. Kelecom and V. L. Teixeira, Phytochemistry, 1988, 27, 2907.
- 64 M. Ochi, K. Asao, H. Kotsuki, I. Miura, and K. Shibata, Bull, Chem. Soc. Jpn., 1986, 59, 661.
- 65 E. Piers and R. W. Friesen, J. Chem. Soc., Chem. Commun., 1988, 125.
- 66 M. Ochi, M. Watanabe, I. Miura, M. Taniguchi, and T. Tokoroyama, Chem. Lett., 1980, 1229.
- 67 P. Crews, T. E. Klein, E. R. Hogue, and B. L. Myers, J. Org. Chem., 1982, 47, 811.
- 68 G. Mehta and N. Krishnamurthy, Tetrahedron Lett., 1987, 28, 5945.
- 69 M. Segawa, N. Enoki, M. Ikura, K. Hikichi, R. Ishida, H. Shirahama, and T. Matsumoto, *Tetrahedron Lett.*, 1987, 28, 3703.
- 70 N. Kato, S. Tanaka, H. Kataoka, and H. Takeshita, Chem. Lett., 1987, 2295.
- 71 J. Clardy, G. Van Duyne, A. Gallardo, E. Manta, J. D. Martín, C. Pérez, J. L. Ravelo, M. L. Rodríguez, and G. K. Schulte, Tetrahedron Lett., 1987, 28, 6699.
- 72 J. S. Edmonds and K. A. Francesconi, J. Chem. Soc., Perkin Trans. 1, 1983, 2375.
- 73 D. P. McAdam, A. M. A. Perera, and R. V. Stick, Aust. J. Chem., 1987, 40, 1901.
- 74 A. Lopez and W. H. Gerwick, Tetrahedron Lett., 1988, 29, 1505.
- 75 M. D. Higgs, Tetrahedron, 1981, 37, 4255.
- 76 M. Bernart and W. H. Gerwick, Tetrahedron Lett., 1988, 29, 2015.
- 77 M. Suzuki, E. Kurosawa, and K. Kurata, Bull. Chem. Soc. Jpn., 1987, 60, 3793.
- 78 J. D. McCombs, J. W. Blunt, M. V. Chambers, M. H. G. Munro, and W. T. Robinson, *Tetrahedron*, 1988, 44, 1489.
- 79 R. Kazlauskas, P. T. Murphy, R. J. Quinn, and R. J. Wells, Tetrahedron Lett., 1977, 37.
- 80 A. San Martín, J. Rovirosa, C. Xu, H. S. M. Lu, and J. Clardy, Tetrahedron Lett., 1987, 28, 6013.
- 81 A. San Martín, J. Rovirosa, O. Muñoz, M. H. M. Chen, R. D. Guneratne, and J. Clardy, Tetrahedron Lett., 1983, 24, 4063.
- 82 M. Norte, J. L. Fernández, J. Z. Ruano, M. L. Rodríguez, and R. Pérez, *Phytochemistry*, 1988, 27, 3537.
- E. Kurosawa, A. Fukuzawa, and T. Irie, Tetrahedron Lett., 1972, 2121.
- 84 M. Suzuki and E. Kurosawa, Bull. Chem. Soc. Jpn., 1987, 60, 3791.
- 85 C. P. Falshaw, T. J. King, S. Imre, S. Islimyeli, and R. H. Thomson, *Tetrahedron Lett.*, 1980, 21, 4951.
- 86 A. G. González, J. D. Martín, V. S. Martín, M. Norte, R. Pérez, J. Z. Ruano, S. A. Drexler, and J. Clardy, *Tetrahedron*, 1982, 38, 1009.
- 87 L. E. Overman and A. S. Thompson, J. Am. Chem. Soc., 1988, 110, 2248.
- 88 J. M. Palazon and V. S. Martín, Tetrahedron Lett., 1988, 29, 681.
- 89 J. C. Coll and A. D. Wright, Aust. J. Chem., 1987, 40, 1893.
- R. J. Capon, L. M. Engelhardt, E. L. Ghisalberti, P. R. Jefferies,
 V. A. Patrick, and A. H. White, Aust. J. Chem., 1984, 37, 537.
- 91 J. C. Coll, B. W. Skelton, A. H. White, and A. D. Wright, Aust. J. Chem., 1988, 41,1743.
- 92 R. J. Capon, E. L. Ghisalberti, T. A. Mori, and P. R. Jefferies, J. Nat. Prod., 1988, 51, 1302.
- You. 1764. 1764. 1767.
 J. T. Vázquez, M. Chang, K. Nakanishi, J. D. Martín, and R. Pérez, J. Nat. Prod., 1988, 51, 1257.
- 94 M. Suzuki, E. Kurosawa, and K. Kurata, Bull. Chem. Soc. Jpn., 1987, 60, 3795.
- 95 W. Fenical, Phytochemistry, 1976, 15, 511.

96 M. Suzuki, E. Kurosawa, and A. Furusaki, Bull. Chem. Soc. Jpn., 1988, 61, 3371.

NATURAL PRODUCT REPORTS, 1990

- 97 S. Caccamese, A. Compagnini, R. M. Toscano, F. Nicolo, and G. Chapuis, *Tetrahedron*, 1987, 43, 5393.
- 98 Atta-ur-Rahman, V. U. Ahmad, S. Bano, S. A. Abbas, K. A. Alvi, M. S. Ali, H. S. M. Lu, and J. Clardy, *Phytochemistry*, 1988, 27, 3879.
- 99 B. M. Howard and W. Fenical, Tetrahedron Lett., 1975, 1687.
- 100 S. Bano, M. S. Ali, and V. U. Ahmad, Planta Med., 1987, 508.
- 101 D. J. Kennedy, I. A. Selby, and R. H. Thomson, *Phytochemistry*, 1988, 27, 1761.
- 102 W. H. Gerwick, A. Lopez, R. Davila, and R. Albors, J. Nat. Prod., 1987, 50, 1131.
- 103 M. Suzuki, E. Kurosawa, and K. Kurata, Phytochemistry, 1988, 27, 1209.
- 104 F. Cafieri, L. De Napoli, E. Fattorusso, and C. Santacroce, Phytochemistry, 1988, 27, 621.
- 105 S. De Rosa, S. De Stefano, P. Scarpelli, and N. Zavodnik, Phytochemistry, 1988, 27, 1875.
- 106 T. Suzuki, M. Suzuki, A. Furusaki, T. Matsumoto, A. Kato, Y. Imanaka, and E. Kurosawa, Tetrahedron Lett., 1985, 26, 1329.
- 107 M. Hashimoto, M. Yanagiya, and H. Shirahama, Chem. Lett.,
- 108 M. Hashimoto, H. Harigaya, M. Yanagiya, and H. Shirahama, Tetrahedron Lett., 1988, 29, 5947.
- 109 S. Sakemi, T. Higa, C. W. Jefford, and G. Bernardinelli, Tetrahedron Lett., 1986, 27, 4287.
- 110 J. W. Blunt, M. P. Hartshorn, T. J. McLennan, M. H. G. Munro, W. T. Robinson, and S. C. Yorke, *Tetrahedron Lett.*, 1978, 69.
- 111 M. Hashimoto, T. Kan, M. Yanagiya, H. Shirahama, and T. Matsumoto, *Tetrahedron Lett.*, 1987, 28, 5665.
- 112 M. Hashimoto, T. Kan, K. Nozaki, M. Yanagiya, H. Shirahama, and T. Matsumoto, *Tetrahedron Lett.*, 1988, 29, 1143.
- 113 E. J. Corey and D.-C. Ha, Tetrahedron Lett., 1988, 29, 3171.
- 114 S. Sciuto, R. Chillemi, and M. Piattelli, J. Nat. Prod., 1988, 51, 322.
- 115 S. Sciuto, R. Chillemi, R. Morrone, A. Patti, and M. Piattelli, J. Nat. Prod., 1988, 51, 1017.
- 116 J. Tanaka, T. Higa, G. Bernardinelli, and C. W. Jefford, Tetrahedron Lett., 1988, 29, 6091.
- 117 K.-W. Glombitza, I. Sukopp, and H. Weidenfeld, *Planta Med.*, 1985, 437.
- 118 J.-C. Yvin, A.-M. Chevolot-Magueur, L. Chevolot, J.-Y. Lallemand, P. Potier, and J. Guilhem, J. Am. Chem. Soc., 1982, 104, 4497.
- 119 A. J. Poss and R. K. Belter, J. Org. Chem., 1988, 53, 1535.
- 120 J. Meng, K.-G. Rosell, and L. M. Srivastava, Carbohydr. Res. 1987, 161, 171.
- 121 E. Quiñoà and P. Crews, Tetrahedron Lett., 1988, 29, 2037.
- 122 T. F. Molinski and C. M. Ireland, J. Org. Chem., 1988, 53, 2103.
- 123 Y. Kato, N. Fusetani, S. Matsunaga, K. Hashimoto, S. Fujita, and T. Furuya, J. Am. Chem. Soc., 1986, 108, 2780.
- 124 Y. Kato, N. Fusetani, S. Matsunaga, K. Hashimoto, and K. Koseki, J. Org. Chem., 1988, 53, 3930.
- 125 Y. Kato, N. Fusetani, S. Matsunaga, K. Hashimoto, R. Sakai, T. Higa, and Y. Kashman, *Tetrahedron Lett.*, 1987, 28, 6225.
- 126 R. Sakai, T. Higa, and Y. Kashman, *Chem. Lett.*, 1986, 1499.
- 127 D. G. Corley, R. Herb, R. E. Moore, P. J. Scheuer, and V. J. Paul, J. Org. Chem., 1988, 53, 3644.
- 128 E. Quiñoà, Y. Kakou, and P. Crews, J. Org. Chem., 1988, 53, 3642.
- 129 N. B. Perry, J. W. Blunt, M. H. G. Munro, and L. K. Pannell, J. Am. Chem. Soc., 1988, 110, 4850.
- 130 S. Sakemi, T. Ichiba, S. Kohmoto, G. Saucy, and T. Higa, J. Am. Chem. Soc., 1988, 110, 4851.
- 131 C. Cardani, D. Ghiringhelli, R. Mondelli, and A. Quilico, Tetrahedron Lett., 1965, 2537.
- 132 M. R. Kernan and D. J. Faulkner, Tetrahedron Lett., 1987, 28, 2809
- 133 M. R. Kernan, T. F. Molinski, and D. J. Faulkner, J. Org. Chem., 1988, 53, 5014.
- 134 M. Adamczeski, E. Quiñoà, and P. Crews, J. Am. Chem. Soc., 1988, 110, 1598.
- 135 P. Crews, Y. Kakou, and E. Quiñoà, J. Am. Chem. Soc., 1988, 110, 4365.
- 136 B. J. Baker, P. J. Scheuer, and J. N. Shoolery, J. Am. Chem. Soc., 1988, 110, 965.
- 137 E. Fahy, T. F. Molinski, M. K. Harper, B. W. Sullivan, D. J. Faulkner, L. Parkanyi, and J. Clardy, *Tetrahedron Lett.*, 1988, 29, 3427.

- 138 J. C. Braekman, D. Daloze, G. Cimino, and E. Trivellone, Bull. Soc. Chim. Belg., 1988, **97**, 519.
- 139 R. Sakai, T. Higa, C. W. Jefford, and G. Bernardinelli, J. Am. Chem. Soc., 1986, 108, 6404.
- 140 R. Sakai, S. Kohmoto, T. Higa, C. W. Jefford, and G. Bernardinelli, Tetrahedron Lett., 1987, 28, 5493.
- T. Ichiba, R. Sakai, S. Kohmoto, G. Saucy, and T. Higa, Tetrahedron Lett., 1988, 29, 3083.
- 142 H. Nakamura, S. Deng, J. Kobayashi, Y. Ohizumi, Y. Tomotake, T. Matsuzaki, and Y. Hirata, Tetrahedron Lett., 1987, 28, 621.
- 143 A. Ahond, M. B. Zurita, M. Colin, C. Fizames, P. Laboute, F. Lavelle, D. Laurent, C. Poupat, J. Pusset, M. Pusset, O. Thoison,
- and P. Potier, C. R. Acad. Sci. Paris, 1988, 307, 145.
 T. Gebreyesus, T. Yosief, S. Carmely, and Y. Kashman, Tetrahedron Lett., 1988, 29, 3863.
- 145 S. Omar, L. Tenenbaum, L. V. Manes, and P. Crews, Tetrahedron Lett., 1988, 29, 5489.
- 146 T. M. Zabriskie, J. A. Klocke, C. M. Ireland, A. H. Marcus, T. F. Molinski, D. J. Faulkner, C. Xu, and J. Clardy, J. Am. Chem. Soc., 1986, 108, 3123.
- 147 P. Crews, L. V. Manes, and M. Boehler, Tetrahedron Lett., 1986, **27**, 2797.
- 148 W. R. Chan, W. F. Tinto, P. S. Manchand, and L. J. Todaro, J. Org. Chem., 1987, 52, 3091.
- 149 U. Schmidt, W. Siegel, and K. Mundinger, Tetrahedron Lett., 1988, 29, 1269.
- 150 S. Kato, Y. Hamada, and T. Shiori, Tetrahedron Lett., 1988, 29, 6465.
- 151 P. A. Grieco, Y. S. Hon, and A. Pérez-Medrano, J. Am. Chem. Soc., 1988, 110, 1630.
- 152 P. A. Grieco and A. Pérez-Medrano, Tetrahedron Lett., 1988, 29. 4225
- 153 S. Kohmoto, Y. Kashman, O. J. McConnell, K. L. Rinehart, Jr., A. Wright, and F. Koehn, J. Org. Chem., 1988, 53, 3116.
- 154 K. Bartik, J. C. Braekman, D. Daloze, C. Stoller, J. Huysecom, G. Vandevyver, and R. Ottinger, Can. J. Chem., 1987, 65, 2118.
- 155 S. Tsujii, K. L Rinehart, Jr., S. P. Gunasekera, Y. Kashman, S. S. Cross, M. S. Lui, S. A. Pomponi, and M. C. Diaz, J. Org. Chem., 1988, **53**, 5446.
- 156 J. C. Braekman, D. Daloze, and C. Stoller, Bull. Soc. Chim. Belg., 1987, 96, 809.
- 157 C. Stoller, J. C. Braekman, D. Daloze, and G. Vandevyver, J. Nat. Prod., 1988, 51, 383.
- 158 G. Lidgren, L. Bohlin, and C. Christophersen, J. Nat. Prod., 1988, **51**, 1277.
- 159 S. Kohmoto, O. J. McConnell, and A. Wright, Experientia, 1988, 44, 85.
- 160 T. F. Molinski and D. J. Faulkner, Tetrahedron Lett., 1988, 29,
- 161 R. J. Capon, J. K. Macleod, and P. J. Scammells, Tetrahedron 1986, 42, 6545.
- 162 J. K. MacLeod and L. C. Monahan, Tetrahedron Lett., 1988, 29, 391.
- 163 G. C. Harbour, A. A. Tymiak, K. L. Rinehart, Jr, D. W. Shaw, G. H. Hughes, Jr., S. A. Mizsak, J. H. Coats, G. E. Zurenko, L. H. Li, and S. L. Kuentzel, J. Am. Chem. Soc., 1981, 103, 5604.
- 164 T. Uyehara, T. Furuta, Y. Kabawawa, J. Yamada, T. Kato, and Y. Yamamoto, J. Org. Chem., 1988, 53, 3669.
- 165 D. M. Roll, C. M. Ireland, H. S. M. Lu, and J. Clardy, J. Org. Chem., 1988, 53, 3276.
- T. F. Molinski, E. Fahy, D. J. Faulkner, G. D. Van Duyne, and J.
- Clardy, J. Org. Chem., 1988, 53, 1340.

 167 G. P. Gunawardana, S. Kohmoto, S. P. Gunasekera, O. J. McConnell, and F. E. Koehn, J. Am. Chem. Soc., 1988, 110, 4856.
- 168 J. Cheng, Y. Ohizumi, M. R. Wälchli, H. Nakamura, Y. Hirata, T. Sasaki, and J. Kobayashi, J. Org. Chem., 1988, 53, 4621.
- 169 N. B. Perry, J. W. Blunt, and M. H. G. Munro, Tetrahedron, 1988, 44, 1727.
- 170 N. B. Perry, J. W. Blunt, M. H. G. Munro, T. Higa, and R. Sakai, J. Org. Chem., 1988, 53, 4127.
- 171 J. Kobayashi, J. Cheng, M. Ishibashi, H. Nakamura, Y. Ohizumi, Y. Hirata, T. Sasaki, H. Lu, and J. Clardy, Tetrahedron Lett., 1987, 28, 4939.
- 172 F. J. Schmitz, S. K. Agarwal, S. P. Gunasekera, P. G. Schmidt, and J. N. Shoolery, J. Am. Chem. Soc., 1983, 105, 4835.
- 173 A. M. Echavarren and J. K. Stille, J. Am. Chem. Soc., 1988, 110, 4051.
- 174 J. M. Frincke and D. J. Faulkner, J. Am. Chem. Soc., 1982, 104,
- 175 K. A. Parker and D. A. Casteel, J. Org. Chem., 1988, 53, 2847.

- 176 M. Norte, M. L. Rodríguez, J. J. Fernández, L. Eguren, and D. M. Estrada, Tetrahedron, 1988, 44, 4973.
- 177 D. B. Borders, G. O. Morton, and E. R. Wetzel, Tetrahedron Lett., 1974, 2709.
- 178 T. N. Makarieva, V. A. Stonik, P. Alcolado, and G. B. Elyakov, Comp. Biochem. Physiol., 1981, 68, 481.
- 179 R. T. Luibrand, T. R. Erdman, J. J. Vollmer, P. J. Scheuer, J. Finer, and J. Clardy, Tetrahedron, 1979, 35, 609.
- 180 M.-L. Kondracki and M. Guyot, Tetrahedron Lett., 1987, 28, 5815.
- 181 M. Ishibashi, Y. Ohizumi, J. Cheng, H. Nakamura, Y. Hirata, T. Sasaki, and J. Kobayashi, J. Org. Chem., 1988, 53, 2855.
- 182 H. Nakamura, J. Kobayashi, M. Kobayashi, Y. Ohizumi, and Y. Hirata, Chem. Lett., 1985, 713.
- 183 D. M. Roll, P. J. Scheuer, G. K. Matsumoto, and J. Clardy, J. Am. Chem. Soc., 1983, 105, 6177
- 184 F. J. Schmitz and S. J. Bloor, J. Org. Chem., 1988, 53, 3922.
- N. Harada, T. Sugioka, Y. Ando, H. Uda, and T. Kuriki, J. Am. Chem. Soc., 1988, 110, 8483.
- 186 N. Fusetani, M. Sugano, S. Matsunaga, K. Hashimoto, H. Shikama, A. Ohta, and H. Nagano, Experientia, 1987, 43, 1233.
- Y. F. Pouchus, J. F. Verbist, J. F. Baird, and K. Boukef, J. Nat. Prod., 1988, 51, 188.
- 188 I. Mancini, G. Guella, A. Guerriero, A. Boldrin, and F. Pietra, Helv. Chim. Acta, 1987, 70, 2011.
- 189 C. J. Barrow, J. W. Blunt, and M. H. G. Munro, Aust. J. Chem., 1988, **41**, 1755.
- 190 N. Fusetani, M. Sugano, S. Matsunaga, and K. Hashimoto, Experientia, 1987, 43, 1234.
- 191 R. J. Capon and J. K. MacLeod, J. Nat. Prod., 1987, 50, 1136.
- 192 J. H. Cardellina, II, and D. E. Barnekow, J. Org. Chem., 1988, 53,
- 193 H. Tada, T. Tozyo, and M. Shiro, J. Org. Chem., 1988, 53, 3366.
- 194 L. Mayol, V. Piccialli, and D. Sica, Tetrahedron, 1987, 43, 5381.
- 195 R. J. Capon and J. K. MacLeod, Aust. J. Chem., 1988, 41, 979.
- 196 F. Cafieri, E. Fattorusso, S. Magno, C. Santacroce, and D. Sica, Tetrahedron, 1973, 29, 4259.
- 197 M. Adinolfi, L. De Napoli, B. Di Blasio, A. Iengo, C. Pedone, and C. Santacroce, Tetrahedron Lett., 1977, 2815.
- 198 E. Fattorusso, S. Magno, L. Mayol, C. Santacroce, and D. Sica, Tetrahedron, 1975, 31, 269.
- 199 E. Piers, B. W. A. Yeung, and S. J. Rettig, Tetrahedron, 1987, 43,
- 200 H. Nakamura, H. Wu, J. Kobayashi, Y. Ohizumi, Y. Hirata, T. Higashijima, and T. Miyazawa, Tetrahedron Lett., 1983, 24, 4105.
- Y. Ichikawa, Tetrahedron Lett., 1988, 29, 4957.
- 202 R. C. Cambie, P. R. Bergquist, and P. Karuso, J. Nat. Prod., 1988, **51**, 1014.
- 203 S. Hirsch and Y. Kashman, J. Nat. Prod., 1988, 51, 1243.
- 204 R. Kazlauskas, P. T. Murphy, R. J. Well, K. Noack, W. E. Oberhansli, and P. Schonholzer, Aust. J. Chem., 1979, 32, 867.
- 205 N. Capelle, J. C. Braekman, D. Daloze, and B. Tursch, Bull. Soc. Chim. Belg., 1980, 89, 399.
- 206 R. C. Cambie, P. A. Craw, M. J. Stone, and P. R. Bergquist, J. Nat. Prod., 1988, 51, 293.
- S. Carmely, M. Cojocaru, Y. Loya, and Y. Kashman, J. Org. Chem., 1988, 53, 4801.
- 208 T. F. Molinski, D. J. Faulkner, C.-H. He, G. D. Van Duyne, and J. Clardy, J. Org. Chem., 1986, 51, 4564.
- 209 L. Mayol, V. Piccialli, and D. Sica, Gazz. Chim. Ital., 1988, 118,
- 210 P. T. Northcote and R. J. Andersen, J. Nat. Prod., 1987, 50, 1174.
- 211 P. T. Northcote and R. J. Andersen, Tetrahedron Lett., 1988, 29, 4357.
- 212 Y. Kashman, S. Hirsch, F. Koehn, and S. Cross, Tetrahedron Lett., 1987, 28, 5461.
- 213 R. Fathi-Afshar and T. M. Allen, Can. J. Chem., 1988, 66, 45.
- 214 T. Nakatsu, D. J. Faulkner, G. K. Matsumoto, and J. Clardy, Tetrahedron Lett., 1984, 25, 935.
- 215 S. Omar, C. Albert, T. Fanni, and P. Crews, J. Org. Chem., 1988, **53**, 5971.
- 216 J. Tanaka and T. Higa, Tetrahedron, 1988, 44, 2805.
- 217 F. J. Schmitz and J. C. Chang, J. Nat. Prod., 1988, 51, 745.
- 218 D. J. Faulkner, Tetrahedron Lett., 1973, 3821.
- 219 C. J. Barrow, J. W. Blunt, M. H. G. Munro, and N. B. Perry, J. Nat. Prod., 1988, 51, 275.
- 220 C. J. Barrow, J. W. Blunt, M. G. H. Munro, and N. B. Perry, J. Nat. Prod., 1988, 51, 1294.
- 221 R. J. Capon and J. K. MacLeod, Aust. J. Chem., 1987, 40,

- 222 R. C. Cambie, P. A. Craw, P. R. Bergquist, and P. Karuso, J. Nat. Prod., 1988, 51, 331.
- 223 S. De Rosa, S. De Stefano, and N. Zavodnik, J. Org. Chem., 1988, 53, 5020.
- 224 E. D. de Silva and P. J. Scheuer, Tetrahedron Lett., 1980, 21, 1611.
- 225 V. E. Amoo, S. De Bernardo, and M. Wiegele, Tetrahedron Lett., 1988, 29, 2401.
- 226 S. Katsumura, S. Fujiwara, and S. Isoe, Tetrahedron Lett., 1988, 29, 1173.
- 227 H. Hagiwara and H. Uda, J. Chem. Soc., Chem. Commun., 1988,
- 815. 228 L. V. Manes, S. Naylor, P. Crews, and G. J. Bakus, J. Org. Chem., 1985, **50**, 284.
- 229 L. V. Manes, P. Crews, M. R. Kernan, D. J. Faulkner, F. R. Fronczek, and R. D. Gandour, J. Org. Chem., 1988, 53, 570.
- 230 M. R. Kernan and D. J. Faulkner, J. Org. Chem., 1988, 53, 4574.
- 231 M. B. Ksebati, F. J. Schmitz, and S. P. Gunasekera, J. Org. Chem., 1988, 53, 3917.
- 232 F. J. Schmitz, M. B. Ksebati, S. P. Gunasekera, and S. Agarwal, J. Org. Chem., 1988, 53, 5941.
- 233 I. Kitagawa, M. Kobayashi, Y. Okamoto, M. Yoshikawa, and Y. Hamamoto, Chem. Pharm. Bull., 1987, 35, 5036.
- 234 G. Guella and F. Pietra, Helv. Chim. Acta, 1988, 71, 62
- 235 R. R. West and J. H. Cardellina, II, J. Org. Chem., 1988, 53, 2782.
- 236 J. C. Braekman, D. Daloze, B. Moussiaux, G. Vandervyver, and R. Riccio, Bull. Soc. Chim. Belg., 1988, 97, 293.
- 237 M. Tischler, S. W. Ayer, R. J. Andersen, J. F. Mitchell, and J. Clardy, Can. J. Chem., 1988, 66, 1173.
- 238 A. Madaio, V. Piccialli, and D. Sica, Tetrahedron Lett., 1988, 29,
- 239 J. Cheng, J. Kobayashi, H. Nakamura, Y. Ohizumi, Y. Hirata, and T. Sasaki, J. Chem. Soc., Perkin Trans. 1, 1988, 2403.
- 240 A. Guerriero, M. D'Ambrosio, and F. Pietra, Helv. Chim. Acta, 1988, 71, 1094.
- 241 K. Iguchi, S. Kaneta, K. Mori, and Y. Yamada, Chem. Pharm. Bull., 1987, 35, 4375.
- 242 M. Suzuki, Y. Morita, A. Yanagisawa, B. J. Baker, P. J. Scheuer, and R. Noyori, J. Org. Chem., 1988, 53, 286.
- 243 K. Mori and T. Takeuchi, Tetrahedron, 1988, 44, 333.
- 244 B. N. Ravi and R. J. Wells, Aust. J. Chem., 1982, 35, 105.
- 245 R. M. Ortuño, J. Bigorra, and J. Font, Tetrahedron, 1988, 44, 5139.
- 246 J. H. Cardellina, II, R. L. Hendrikson, K. P. Manfredi, S. A. Strobel, and J. Clardy, Tetrahedron Lett., 1987, 28, 727.
- 247 M. Brehm, W. G. Dauben, P. Köhler, and F. W. Lichtenthaler, Angew, Chem., Int. Ed. Engl., 1987, 26, 1271.
- 248 R. C. Cambie, P. A Craw, J. S. Buckleton, G. R. Clark, and C. E. F. Rickard, Aust. J. Chem., 1988, 41, 365.
- 249 A. Groweiss, W. H. Fenical, C.-H. He, J. Clardy, Z. Wu, Z. Yiao, and K. Long, Tetrahedron Lett., 1985, 26, 2379.
- 250 C. Iwata, Y. Takemoto, M. Doi, and T. Imanishi, J. Org. Chem., 1988, 53, 1623.
- 251 M. Kobayashi, Chem. Pharm. Bull., 1988, 36, 488.
- 252 M. Kobayashi and T. Hamaguchi, Chem. Pharm. Bull., 1988, 36,
- 253 M. Kobayashi, K. Kondo, K. Osabe, and H. Mitsuhashi, Chem. Pharm. Bull., 1988, 36, 2331.
- 254 Y. Yamada, S. Suzuki, K. Iguchi, H. Kikuchi, Y. Tsukitani, and H. Horiai, Chem. Pharm. Bull., 1980, 28, 2035.
- 255 B. F. Bowden, J. C. Coll, S. J. Mitchell, and G. J. Stokie, Aust. J. Chem., 1978, 31, 1303.
- Y. Uchio, S. Eguchi, J. Kuramoto, M. Nakayama, and T. Hase, Tetrahedron Lett., 1985, 26, 4487.
- 257 T. Kusumi, I. Ohtani, Y. Inouye, and H. Kakisawa, Tetrahedron Lett., 1988, 29, 4731.
- 258 J. C. Coll, S. J. Mitchell, and G. J. Stokie, Aust. J. Chem., 1977, **30**, 1859.
- 259 J. A. Marshall, S. L. Crooks, and B. S. DeHoff, J. Org. Chem., 1988, **53**, 1616.
- 260 M. A Tius, Chem. Rev., 1988, 88, 719.
- 261 D. E. Williams, R. J. Andersen, J. F. Kingston, and A. G. Fallis,
- Can. J. Chem., 1988, 66, 2928.
 262 K. Mori, K. Iguchi, N. Yamada, Y. Yamada, and Y. Inouye, Tetrahedron Lett., 1987, 28, 5673.
- 263 K. Mori, K. Iguchi, N. Yamada, Y. Yamada, and Y. Inouye, Chem. Pharm. Bull., 1988, 36, 2840.
- L. Jin-Cui, Z. Zhi-Ming, X. Zong-Xiang, N. Chao-Zou, and W. Yu-Lin, Huaxue Xuebao, 1987, 45, 558.
- 265 P. Sharma and M. Alam, J. Chem. Soc., Perkin Trans. 1, 1988, 2537.

- 266 T. Kusumi, H. Uchida, M. O. Ishitsuka, H. Yamamoto, and H. Kakisawa, Chem. Lett., 1988, 1077.
- 267 M. Ochi, K. Futatsugi, H. Kotsuki, M. Ishii, and K. Shibata, Chem. Lett., 1987, 2207.
- 268 M. Ochi, K. Futatsugi, Y. Kume, H. Kotsuki, K. Asao, and K. Shibata, Chem. Lett., 1988, 1661.
- 269 M. D'Ambrosio, A. Guerriero, and F. Pietra, Helv. Chim. Acta., 1987, 70, 2019.
- 270 M. D'Ambrosio, A. Guerriero, and F. Pietra, Helv. Chim. Acta, 1988, 71, 964.
- 271 A. Almourabit, A. Ahond, A. Chiaroni, C. Poupat, C. Riche, P. Potier, P. Laboute, and J.-L. Menou, J. Nat. Prod., 1988, 51, 282.
- 272 N. Fusetani, M. Asano, S. Matsunaga, and K. Hashimoto, Tetrahedron Lett., 1987, 28, 5837.
- 273 S. Hokama, J. Tanaka, T. Higa, N. Fusetani, M. Asano, S. Matsunaga, and K. Hashimoto, Chem. Lett., 1988, 855.
- 274 D. Green, S. Carmely, Y. Benayahu, and Y. Kashman, Tetra-
- hedron Lett., 1988, 29, 1605. 275 A. Groweiss and Y. Kashman, Tetrahedron, 1983, 39, 3385.
- 276 A. Groweiss, S. A. Look, and W. Fenical, J. Org. Chem., 1988, 53,
- 277 B. F. Bowden, J. C. Coll, W. Patalinghug, B. W. Skelton, I. Vasilescu, and A. H. White, Aust. J. Chem., 1987, 40, 2085.
- 278 S. J. Coval, S. Cross, G. Bernardinelli, and C. W. Jefford, J. Nat. Prod., 1988, 51, 981.
- 279 A. Guerriero, M. D'Ambrosio, and F. Pietra, Helv. Chim. Acta, 1988, 71, 472
- 280 J. Shin and W. Fenical, J. Org. Chem., 1988, 53, 3271.
- 281 C. A. Harvis, M. T. Burch, and W. Fenical, Tetrahedron Lett., 1988, 29, 4361.
- 282 S. A. Look, W. Fenical, G. K. Matsumoto, and J. Clardy, J. Org. Chem., 1986, 51, 5140.
- 283 C. A. Broka, S. Chan, and B. Peterson, J. Org. Chem., 1988, 53,
- 284 M. B. Ksebati and F. J. Schmitz, J. Org. Chem., 1988, 53, 3926.
- 285 C. B. Rao, K. V. Ramana, D. V. Rao, E. Fahy, and D. J. Faulkner, J. Nat. Prod., 1988, 51, 954.
- 286 G. Guella, I. Mancini, H. Zibrowius, and F. Pietra, Helv. Chim. Acta, 1988, **71**, 773.
- 287 R. Kazlauskas, P. T. Murphy, R. J. Quinn, and R. J. Wells, Tetrahedron Lett., 1976, 2631.
- 288 M. Alam, R. Sanduja, and G. M. Wellington, Heterocycles, 1988, **27**, 719.
- 289 A. Aiello, E. Fattorusso, S. Magno, and L. Mayol, Tetrahedron, 1987, 43, 5929.
- 290 A. J. Blackman and R. D. Green, Aust. J. Chem., 1987, 40, 1655.
- 291 A. J. Blackman, T. W. Hambley, K. Picker, W. C. Taylor, and N. Thirasasana, Tetrahedron Lett., 1987, 28, 5561.
- 292 U. Anthoni, K. Bock, L. Chevolot, C. Larsen, P. H. Nielsen, and C. Christophersen, J. Org. Chem., 1987, 52, 5638.
- 293 T. Miyamoto, R. Higuchi, N. Marubayashi, and T. Komori, Liebigs Ann. Chem., 1988, 1191.
- 294 D. B. Stierle, R. M. Wing, and J. J. Sims, Tetrahedron, 1979, 35,
- 295 B. Baker, L. Ratnapala, M. P. D. Mahindaratne, E. D. de Silva, L. M. V. Tillekeratne, J. H. Jeong, P. J. Scheuer, and K. Seff, Tetrahedron, 1988, 44, 4695.
- 296 A. G. González, F. Cataldo, and J. Fernández, J. Nat. Prod., 1987, 50, 1158.
- 297 A. Ghosh, S. Biswas, and R. V. Venkateswaran, J. Chem. Soc., Chem. Commun., 1988, 1421.
- 298 M. Murata, M. Kumagai, J. S. Lee, and T. Yasumoto, Tetrahedron Lett., 1987, 28, 5869.
- 299 M. Iorizzi, L. Minale, R. Riccio, J.-S. Lee, and T. Yasumoto, J. Nat. Prod., 1988, 51, 1098.
- 300 A. A. Tymiak and K. L. Rinehart, Jr., J. Am. Chem. Soc., 1983, 105, 7396.
- 301 G. Cimino, S. De Stefano, and G. Strazzullo, J. Nat. Prod., 1987, **50**, 1171.
- 302 P. Ciminiello, E. Fattorusso, and S. Magno, Gazz. Chim. Ital., 1988, 118, 105.
- 303 G. Cimino, A. Crispino, A. Spinella, and G. Sodano, Tetrahedron Lett., 1988, 29, 3613.
- 304 J. R. Pawlik, M. R. Kernan, T. F. Molinski, M. K. Harper, and D. J. Faulkner, J. Exp. Mar. Biol. Ecol., 1988, 119, 99.
- 305 S. J. Coval and P. J. Scheuer, J. Org. Chem., 1985, 50, 3024.
- 306 S. D. Burke, A. D. Piscopio, and J. L. Buchanan, Tetrahedron Lett., 1988, 29, 2757.
- 307 W. Oppolzer, D. Dupuis, G. Poli, T. M. Raynham, and G. Bernardinelli, Tetrahedron Lett., 1988, 29, 5885.

- 308 T. Kosuge, K. Tsuji, K. Harai, T. Fukuyama, H. Nukaya, and H. Ishida, Chem. Pharm. Bull., 1985, 33, 2890.
- 309 S. Inoue, K. Okada, H. Tanino, and H. Kakoi, *Tetrahedron Lett.*, 1988, 29, 1547.
- 310 R. K. Okuda, P. J. Scheuer, J. E. Hochlowski, R. P. Walker, and D. J. Faulkner, J. Org. Chem., 1983, 48, 1866.
- 311 G. Cimino, G. Sodano, and A. Spinella, J. Nat. Prod., 1988, 51, 1010.
- 312 R. Kazlauskas, P. T. Murphy, R. J. Wells, J. J. Daly, and P. Schonholzer, *Tetrahedron Lett.*, 1978, 4951.
- 313 G. Cimino, S. De Stefano, L. Minale, and E. Trivellone, *Tetrahedron*, 1972, 28, 4761.
- 314 M. B. Ksebati and F. J. Schmitz, J. Nat. Prod., 1988, 51, 857.
- 315 G. Cimino, M. Gavagnin, G. Sodano, R. Puliti, C. A. Mattia, and L. Mazzarella, *Tetrahedron*, 1988, 44, 2301.
- 316 V. J. Paul and K. L. Van Alstyne, J. Exp. Mar. Biol. Ecol., 1988, 119, 15.
- 317 B. Carté, M. R. Kernan, E. B. Barrabee, D. J. Faulkner, G. K. Matsumoto, and J. Clardy, J. Org. Chem., 1986, 51, 3528.
- 318 M. R. Kernan, E. B. Barrabee, and D. J. Faulkner, Comp. Biochem. Physiol., 1988, 89B, 275.
- 319 M. Norte, F. Cataldo, and A. G. González, *Tetrahedron Lett.*, 1988, 29, 2879.
- 320 G. Cimino, G. Sodano, and A. Spinella, *J. Org. Chem.*, 1988, **52**, 5326.
- 321 N. Lindquist, W. Fenical, D. F. Sesin, C. M. Ireland, G. D. Van Duyne, C. J. Forsyth, and J. Clardy, *J. Am. Chem. Soc.*, 1988, 110, 1308.
- 322 D. Gouiffès, S. Moreau, N. Helbecque, J. L. Bernier, J. P. Hénichart, Y. Barbin, D. Laurent, and J. F. Verbist, *Tetrahedron*, 1988, 44, 451.
- 323 J. Kobayashi, J. Cheng, T. Ohta, H. Nakamura, S. Nozoe, Y. Hirata, Y. Ohizumi, and T. Sasaki, J. Org. Chem., 1988, 53, 6147.
- 324 D. G. Corley, R. E. Moore, and V. J. Paul, J. Am. Chem. Soc., 1988, 110, 7920.
- 325 T. M. Zabriskie, C. L. Mayne, and C. M. Ireland, J. Am. Chem. Soc., 1988, 110, 7919.
- 326 G. T. Carter and K. L. Rinehart, Jr., J. Am. Chem. Soc., 1978, 100, 7441.
- 327 T. Umemura and K. Mori, Agric. Biol. Chem., 1987, 51, 217.
- 328 L. Arabshi and F. J. Schmitz, Tetrahedron Lett., 1988, 29, 1099.
- 329 U. Schmidt, M. Kroner, and H. Griesser, *Tetrahedron Lett.*, 1988, 29, 3057.
- 330 U. Schmidt, M. Kroner, and H. Griesser, *Tetrahedron Lett.*, 1988, 29, 4407.
- 331 K. L. Rinehart, V. Kishore, K. C. Bible, R. Sakai, D. W. Sullins, and K.-M. Li, J. Nat. Prod., 1988, 51, 1.
- 332 R. J. Lake, M. M. Brennan, J. W. Blunt, M. H. G. Munro, and L. K. Pannell, *Tetrahedron Lett.*, 1988, 29, 2255.
- 333 R. J. Lake, J. D. McCombs, J. W. Blunt, M. H. G. Munro, and W. T. Robinson, *Tetrahedron Lett.*, 1988, 29, 4971.
- 334 P. Djura, D. B. Stierle, B. J. Sullivan, D. J. Faulkner, E. Arnold, and J. Clardy, J. Org. Chem., 1980, 45, 1435.
- 335 C. Debitus, D. Laurent, and M. Pais, J. Nat. Prod., 1988, **51**, 799.
- 336 H. Niwa, Y. Yoshida, and K. Yamada, J. Nat. Prod., 1988, 51, 343.
- 337 N. M. Cooray, P. J. Scheuer, L. Parkanyi, and J. Clardy, J. Org. Chem., 1988, 53, 4619.

- 338 A. Rudi, Y. Benahayu, I. Goldberg, and Y. Kashman, Tetrahedron Lett., 1988, 29, 3861.
- 339 A. Rudi, Y. Benayahu, I. Goldberg, and Y. Kashman, Tetrahedron Lett., 1988, 29, 6655.
- 340 S. J. Bloor and F. J. Schmitz, J. Am. Chem. Soc., 1987, 109, 6134.
- 341 F. S. de Guzman and F. J. Schmitz, *Tetrahedron Lett.*, 1989, 30, 1069.
- 342 J. Kobayashi, J. Cheng, H. Nakamura, Y. Ohizumi, Y. Hirata, T. Sasaki, T. Ohta, and S. Nozoe, *Tetrahedron Lett.*, 1988, 29, 1177.
- 343 J. Kobayashi, J. Cheng, M. R. Wälchli, H. Nakamura, Y. Hirata, T. Sasaki, and Y. Ohizumi, J. Org. Chem., 1988, 53, 1800.
- 344 R. J. Andersen, D. J. Faulkner, C.-H. He, G. D. Van Duyne, and J. Clardy, J. Am. Chem. Soc., 1985, 107, 5492.
- 345 N. Lindquist, W. Fenical, G. D. Van Duyne, and J. Clardy, J. Org. Chem., 1988, 53, 4570.
- 346 E. M. Oltz, R. C. Bruening, M. J. Smith, K. Kustin, and K. Nakanishi, J. Am Chem. Soc., 1988, 110, 6162.
- 347 R. Riccio, M. Iorizzi, L. Minale, Y. Oshima, and T. Yasumoto, J. Chem. Soc., Perkin Trans. 1, 1988, 1337.
- 348 Y. Noguchi, R. Higuchi, N. Marubayashi, and T. Komori, Liebigs Ann. Chem., 1987, 341.
- 349 M.-A. Dubois, Y. Noguchi, R. Higuchi, and T. Komori, *Liebigs Ann. Chem.*, 1988, 495.
- 350 R. Higuchi, Y. Noguchi, T. Komori, and T. Sasaki, Liebigs Ann. Chem., 1988, 1185.
- 351 R. Riccio, L. Minale, S. Bano, N. Bano, and V. U. Ahmad, Gazz. Chim. Ital., 1987, 117, 755.
- 352 R. Riccio, O. Squillace Greco, L. Minale, S. La Barre, and D. Laurent, J. Nat. Prod., 1988, 51, 1003.
- 353 R. Riccio, O. Squillace Greco, and L. Minale, *J. Nat. Prod.*, 1988, 51, 989.
- 354 I. I Kapustina, A. I. Kalinovskii, S. G. Polonik, and V. A. Stonik,
- Chem. Nat. Comp., 1987, 23, 209. 355 E. B. Levina, A. I. Kalinovskii, P. V. Andriyashchenko, and A. A.
- Kich, Chem. Nat. Prod., 1987, 23, 206.
 356 M. V. D'Auria, L. Minale, R. Riccio, and E. Uriarte, Experientia,
- 1988, 44, 719.
 357 L. Meijer, J. Maclouf, and R. W. Bryant, Prostaglandins,
- Leukotrienes Medicine, 1986, 23, 179. 358 Y. Kawano, R. Higuchi, R. Isobe, and T. Komori, Liebigs Ann.
- Chem., 1988, 19.
- 359 S. Sugiyama, M. Honda, and T. Komori, Liebigs Ann. Chem., 1988, 619.
- 360 Y. Kawano, R. Higuchi, R. Isobe, and T. Komori, Liebigs Ann. Chem., 1988, 1181.
- 361 G. R. Pettit, M. Inoue, Y. Kamano, D. L. Herald, C. Arm, C. Dufresne, N. D. Christie, J. M. Schmidt, D. L. Doubek, and T. S. Krupa, J. Am. Chem. Soc., 1988, 110, 2006.
- 362 G. R. Pettit, M. Inoue, Y. Kamano, C. Dufresne, N. D. Christie, M. L. Niven, and D. L. Herald, J. Chem. Soc., Chem. Commun., 1988, 865.
- 363 G. R. Pettit, M. Inoue, Y. Kamano, C. Dufresne, N. D. Christie, M. L. Niven, and D. L. Herald, J. Chem. Soc., Chem. Commun., 1988, 1440
- 364 H. Nakamura, B. Musicki, Y. Kishi, and O. Shimomura, J. Am. Chem. Soc., 1988, 110, 2683.
- 365 A. Endo, S. S. Khora, M. Murata, H. Naoki, and T. Yasumoto, Tetrahedron Lett., 1988, 29, 4127.