See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/275527517

Synthesis and Evaluation of Cationic Norbornanes as Peptidomimetic Antibacterial Agents

ARTICLE in ORGANIC & BIOMOLECULAR CHEMISTRY · APRIL 2015

Impact Factor: 3.56 · DOI: 10.1039/C5OB00621J

READS

21

13 AUTHORS, INCLUDING:



Shane M. Hickey

University of South Australia

7 PUBLICATIONS 16 CITATIONS

SEE PROFILE



Mark Stuart Butler

University of Queensland

94 PUBLICATIONS 2,948 CITATIONS

SEE PROFILE



Jian Li

Monash University (Australia)

132 PUBLICATIONS 3,967 CITATIONS

SEE PROFILE



Johnny X Huang

University of Queensland

32 PUBLICATIONS 194 CITATIONS

SEE PROFILE

Organic & Biomolecular Chemistry



PAPER

View Article Online



Cite this: *Org. Biomol. Chem.*, 2015, **13**, 6225

Synthesis and evaluation of cationic norbornanes as peptidomimetic antibacterial agents†

Shane M. Hickey, ^a Trent D. Ashton, ^a Simren K. Khosa, ^a Ryan N. Robson, ^a Jonathan M. White, ^b Jian Li, ^c Roger L. Nation, ^c Heidi Y. Yu, ^c Alysha G. Elliott, ^d Mark S. Butler, ^d Johnny X. Huang, ^d Matthew A. Cooper ^d and Frederick M. Pfeffer*

A series of structurally amphiphilic biscationic norbornanes have been synthesised as rigidified, low molecular weight peptidomimetics of cationic antimicrobial peptides. A variety of charged hydrophilic functionalities were attached to the norbornane scaffold including aminium, guanidinium, imidazolium and pyridinium moieties. Additionally, a range of hydrophobic groups of differing sizes were incorporated through an acetal linkage. The compounds were evaluated for antibacterial activity against both Gramnegative and Gram-positive bacteria. Activity was observed across the series; the most potent of which exhibited an MIC's $\leq 1~\mu g~mL^{-1}$ against *Streptococcus pneumoniae*, *Enterococcus faecalis* and several strains of *Staphylococcus aureus*, including multi-resistant methicillin resistant (mMRSA), glycopeptide-intermediate (GISA) and vancomycin-intermediate (VISA) *S. aureus*.

Received 30th March 2015, Accepted 27th April 2015 DOI: 10.1039/c5ob00621j

www.rsc.org/obc

Introduction

There is consensus amongst health professionals that a scenario is imminent whereby strains of pathogenic bacteria will be resistant to all current antibiotic treatments. In fact, resistance has been observed for every antibiotic in current clinical use. The development of new antibacterial agents is therefore a priority for continued human health. Naturally occurring cationic antimicrobial peptides are well studied in medicinal chemistry as both antibiotics in their own right and as lead compounds for the development of peptidomimetic antibacterial agents. The mode of action of polymyxin peptide antibiotics (such as colistin and polymyxin B) initially involves electrostatic binding to the lipid A portion of lipopolysaccharide

(LPS) followed by disruption of the Gram-negative cellular membrane ultimately leading to cell lysis.⁶ In order to elicit this response it is accepted that peptides (and peptidomimetics) of this nature must either inherently possess, or adopt, an amphiphilic structure.⁷

Previous work has demonstrated that the norbornene framework, when functionalised with thiourea groups, was capable of binding to phosphoanionic species. As an extension of this study, bisguanidinyl norbornane 1 (Fig. 1) was designed as an amphiphile capable of binding lipid A (a phosphoanionic component of the Gram-negative bacterial cell wall) and exhibited antibacterial activity against *Pseudomonas aeruginosa* and *Acinetobacter baumannii*. Both of these bacterial species present a global medical challenge due to increasing resistance to almost all currently available antibiotics. More recently, a series of compounds bearing two hydrophobic benzyl groups attached to the norbornane framework *via* an ether link (compound 2, Fig. 1), were shown to exhibit modest antibacterial activity against a range of Gramnegative and Gram-positive bacterial strains. Both

Herein, the synthesis of a larger, more diverse set of functionalised norbornane acetals is presented. The antibacterial activity of these compounds was evaluated against a range of Gram-negative and Gram-positive bacteria including members of the ESKAPE pathogens^{1a} using disk diffusion assay (Kirby-Bauer, 50 µg per disk) and micro-broth dilution assay to ascertain the minimum inhibitory concentrations (MICs). From this data a putative structure activity relationship (SAR) has been identified.

^aResearch Centre for Chemistry and Biotechnology, School of Life and Environmental Sciences, Deakin University, Waurn Ponds, Victoria 3216, Australia

^bRight Institute, School of Chemistry, University of Melhourne, Parkville

^bBio21 Institute, School of Chemistry, University of Melbourne, Parkville, Victoria 3010, Australia

^cDrug Delivery, Disposition and Dynamics, Monash Institute of Pharmaceutical Science, Royal Parade, Parkville, Victoria 3052, Australia

^dInstitute for Molecular Bioscience, The University of Queensland, Brisbane, Queensland 4072, Australia. E-mail: fred.pfeffer@deakin.edu.au

[†] Electronic supplementary information (ESI) available: Crystal structure of compound **4a** (CIF), 2D NMR spectra for compounds **4a**, and *exo/endo* isomers from the reaction products of diol 3 and a selected benzaldehye, synthetic procedures for all known compounds and copies of NMR spectra (¹H and ¹³C) for all new compounds. All bacterial strains tested, and all cytotoxicity data. CCDC 1050774. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c5ob00621j

Fig. 1 Previously reported antibacterial agents based on the norbornane scaffold.

Results and discussion

Chemistry

Design. A lipophilic moiety is an essential component of antibacterial agents that act via membrane disruption. Hence, the design of the family of compounds involved variation of the hydrophobic section to include alkyl and mixed alkyl/aryl functionality linked through an acetal to the parent norbornane. To mimic the lysine and arginine residues present in naturally occurring cationic antibacterial peptides, amines and guanidines were used as the cationic groups. Furthermore, alkylated imidazolium and pyridinium, as well as charge neutral anion recognition groups, thioureas and squaramides, were also incorporated.

Synthesis. Initially, a synthetic route that allowed for late stage installation of the lipophilic group was investigated; however, the isolation of key intermediates in reproducible yields was problematic. A synthetic route to access the desired amphiphiles 8a-d (Scheme 1), which allowed for late stage intro-

duction of the hydrophilic (or anion binding) group using 2-(tertbutoxycarbonylamino)ethylamine 6,11 proved to be more robust. Following the method reported by Pandey and co-workers, 12 stirring the appropriate aldehyde with diol 3 in PhMe with catalytic TsOH (0.05 equiv.) at 110 °C gave acetals 4a-d (48-92%).

Unfortunately, in some instances condensation of benzaldehydes resulted in a thermodynamic mixture of inseparable exo/endo acetal isomers. After further functionalisation of the norbornane scaffold, separation of the isomers could sometimes be achieved using column chromatography (see ESI† for further discussion and NMR spectra of individual isomers). It is reasonable to suggest that the stability of the intermediate oxonium species was enhanced by the adjacent aromatic system thus facilitating isomerisation. Given that the final step in the synthesis involved acid mediated Boc deprotection, product degradation was likely to occur. 13 As such this subset of analogues was abandoned.

Aliphatic acetals were not prone to the same isomerisation/ complications as their aryl counterparts. The exo-orientation

HO OME I ROOME OR' III ROOME NHR"

4a-d R' = Me IV R = Boc IV R = A - D R = H-HCI

$$R = a b c d R' = H - C R = B - C R - C R = B - C R - C R = B - C R - C$$

Scheme 1 Reagents and conditions: (i) RCHO, TsOH·H₂O, MgSO₄, PhMe, 3h, 110 °C, 4a-d (48-92%); (ii) NaOH, THF/H₂O, 16 h, 21 °C, 5a-d (78-86%); (iii) 6, EDCI, HOBt, CHCl₃, MW: 30 min, 50 °C, 7a-d (35-58%); (iv) AcCl, MeOH, 24 h, 21 °C, 8a-d (94-99%). ¹H NOESY correlations of H-2, H-4 and H-6 (bottom left) and crystal structure of 4a with the aliphatic chain and hydrogens (except for H-2, H-4 and H-6) have been omitted for clarity (bottom right).

of the alkyl chain was confirmed for norbornane dimethyl ester 4a (Scheme 1) using nuclear Overhauser effect (NOE) spectroscopy. Clear correlations were observed between the acetal H-4 methine and the *endo*-methine protons (H-2 and H-6, see ESI† for full NOE spectra) which can only exist if the aliphatic chain has *exo*-orientation. Single crystal X-ray crystallographic analysis (Scheme 1, see ESI† for details of crystallisation) also clearly depicts the *exo*-isomer.

Methyl ester hydrolysis gave diacids 5a-d in good yields (78–86%, Scheme 1), which were then coupled to amine 6¹¹ using EDCI/HOBt and microwave heating (50 °C for 30 min) to give compounds 7a-d (35–58%). Removal of the Boc-groups was effected using *in situ* generated HCl (AcCl/MeOH) to afford the diamines 8a-d as hydrochloride salts in excellent yields (94–99%). All compounds were isolated as the desired *exo*-acetal with no evidence of isomerisation.

Guanidines **10a–e** were accessed by attaching 2-[2,3-bis(*tert*-butoxycarbonyl)guanidino]ethylamine **11**¹⁴ using the previously described microwave-mediated coupling protocol, followed by removal of the Boc-protecting groups using AcCl in MeOH (Scheme 2). In order to investigate the importance of the counterion in relation to antibacterial activity two analogues (**10a** and **10c**) were also synthesised as their trifluoroacetate salts (**12** and **1** respectively, Scheme 3).

In an earlier communication, computational modelling indicated that an ethyl linker between the norbornane scaffold and the guanidine was sufficient for effective binding of Lipid A. ^{8a,15} However the dependence of activity on spacer length was never experimentally tested, therefore, in this study homologated analogues were prepared. Aminopropylguanidine 17 and aminobutylguanidine 18¹⁴ were attached to the norbornane framework to give diamides 13 (49%) and 14 (52%)

Scheme 2 Reagents and conditions: (i) **10**, EDCI, HOBt, DMF, MW: 30 min, 50 °C, **9a**-e (41–74%); (ii) AcCl, MeOH, 16 h, 21 °C, **10a**-e (74–95%).

(Scheme 4). Subsequent deprotection using AcCl/MeOH gave the desired diguanidines **15** (74%) and **16** (94%) as HCl salts (Scheme 4).

To investigate the influence of net ionic charge, two singularly charged analogues were synthesised. Discrimination between the norbornane *exo*- and *endo*-carboxylic acids was achieved *via* iodolactonisation (95%) of norbornene diacid (compound **S1**, see ESI†). Following Fischer esterification (90%) and Zn-mediated reductive elimination the norbornene **19** was obtained in 93% (Scheme 5). The free *endo*-carboxylic acid was then converted to the *n*-heptylamide **20a** (77%) using microwave-assisted amide coupling conditions. Subsequent dihydroxylation of the alkene gave **21a** in 85% yield. Acetal **22a** was then synthesised in 94% yield, before hydrolysis of the *exo*-methyl ester afforded carboxylic acid **23a**

Scheme 3 Reagents and conditions: (i) TFA, CH₂Cl₂, 1–2 d, 21 °C.

Scheme 4 Reagents and conditions: (i) 17 (or 18), EDCI, HOBt, DMF, MW: 30 min, 50 °C; (ii) AcCl, MeOH, 16 h, 21 °C.

Scheme 5 Reagents and conditions: (i) n-heptylamine or n-hexadecylamine, EDCI, HOBt, CHCl₃, MW: 30 min, 50 °C; (ii) OsO₄, NMO, acetone/H₂O, 3 d, 21 °C; (iii) octanal, TsOH·H₂O, MgSO₄, PhMe, 3 h, 110 °C; (iv) NaOH, THF/H₂O, 16 h, 21 °C; (v) 11, EDCI, HOBt, DMF, MW: 30 min, 50 °C; (vi) AcCI, MeOH, 16 h, 21 °C.

in 64% yield. Incorporation of Boc-protected aminoethylguanidine 11 and subsequent deprotection gave the desired HCl salt 25a in near quantitative yield (99%). Guanidine 25b was synthesised in an analogous fashion, using n-hexadecylamine rather than *n*-heptylamine.

Analogues with other anion recognition groups (both charged and charge-neutral) were also synthesised. Both the imidazolium and pyridinium analogues 26c and 26d were accessed in reasonable yield (74 and 64% respectively) through the α -bromoamide (27, see ESI†) following the procedure reported by Gathergood.¹⁷ Incorporation of squaramides and thioureas onto the norbornane framework was also performed as both have previously been used to bind phosphoanions. 15,18 Stirring diamine 8a and squaramate S2 (synthesised from squaric acid, 19 see ESI† for details) at ambient temperature gave the desired disquaramide 26a (40%, Scheme 6). In a

similar fashion, bis-thiourea 26b was accessed in excellent vield (95%) by treating diamine 8a with phenylisothiocyanate at ambient temperature.

Biological evaluation

The antibacterial activity of these compounds was evaluated against a range of Gram-negative and Gram-positive bacteria, including members of the ESKAPE pathogens; 1a first using the disk diffusion assay (Kirby-Bauer) to identify active compounds then micro-broth dilution assays to ascertain minimum inhibitory concentrations (MICs).²⁰

Zones of inhibition (ZOI) were observed for diamine dihydrochloride 8a (50 μg mL⁻¹) against Pseudomonas aeruginosa (16 mm) and Klebsiella pneumoniae (11 mm) in the disk diffusion assay. Similar activity was observed for diamines 8b (10 and 12 mm respectively) and 8d (7 mm against K. pneumo-

Scheme 6 Reagent and conditions: (i) \$2 or phenylisothiocyanate, Et₃N, MeOH or CH₂Cl₂, 21 °C, 26a (40%) or 26b (95%); (ii) bromoacetyl bromide, Et₃N, CH₂Cl₂, 5 h, -78 °C, **27** (65%); (iii) 1-methylimidazole or pyridine, THF, **26c** (74%) or **26d** (64%).

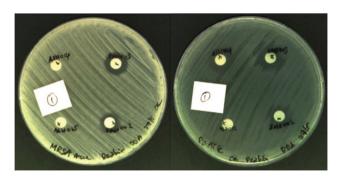


Fig. 2 Antibacterial activity of 10a against MRSA (LHS) and *P. aeruginosa* (RHS).

niae), which had larger hydrophobic portions, including activity against Gram-positive methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant *Enterococcus faecium* (VRE).

Compounds bearing two guanidinium groups displayed similar activity to their diamine counterparts; diguanidines **10a** (Fig. 2), **10b** and **10d** were comparable to diamines **8a**, **8b** and **8d** respectively (Table 1). However, for the imidazolium (**26c**) and pyridinium (**26d**) analogues, only minor inhibition was observed against MRSA (ZOI = 7 and 8 mm, respectively). Furthermore, the effect of the counterion was not significant; trifluoroacetate salt (**12**) possessed similar antibacterial activity to its HCl salt counterpart (**10a**, Table 1).

The effect of the linker between the norbornane framework and the guanidinium moiety was minimal as the three related diguanidine analogues **10a** (ethyl), **15** (propyl) and **16** (butyl),

each had a ZOI \geq 12 mm against *P. aeruginosa*, *K. pneumoniae* and MRSA. When only a single cationic group was present only moderate, or no, activity was observed *e.g.* 25a (ZOI = 8 and 7 mm for MRSA and VRE, respectively, Table 1) and 25b (inactive). This trend continued for the charge neutral disquaramide 26a and bis-thiourea 26b analogues in which only moderate inhibition observed against MRSA (ZOI = 7 and 8 mm, respectively) was noted.

Both the norbornane acetals that showed promising results in the disk diffusion assay and the more lipophilic acetals (**8c**, **10c** and **25b**, Table 1) were subject to micro-broth dilution assay to quantify MICs (Table 2). Again the activity of the aminium and guanidinium analogues was comparable. For example, analogues with a hexadecyl chain (diamine **8c** and diguanidine **10c**) showed almost identical MIC values against all bacterial pathogens (*e.g.* 32 μ g mL⁻¹ against *E. coli* and 0.5 and 1 μ g mL⁻¹ against MRSA respectively, Table 2).

Weak activity in the form of identical MIC values (32 μg mL⁻¹ against MRSA, Table 2) were observed for both compound **16** (which used a butyl linker to separate the guanidinium charge from the norbornane scaffold) and **10a** (which employed an ethyl linker). No activity was observed for the corresponding propyl analogue (**15**). These results are supported by the disk diffusion results discussed earlier (Table 1). The importance of the net overall cationic charge was again emphasised with singularly charged norbornane guanidines **25a** and **25b** showing only modest potency; **25a** displayed MIC values of 32 μg mL⁻¹ against all strains tested apart from MRSA (4 μg mL⁻¹), whilst **25b** was only active against MRSA (32 μg mL⁻¹, Table 2).

Table 1 Antibacterial activity measured with disk diffusion^a

Compound	A. baumannii ATCC 19606	P. aeruginosa ATCC 27853	K. pneumoniae ATCC 13883	S. aureus MRSA ATCC 43300	E. faecium VRE ATCC 700221
Amines					
8a	_	16	11	8	_
8b	_	10	12	12	14
8c	_	_	_	_	_
8d	9	_	7	9	_
Guanidines					
10a	NT^c	15	12	13	12
12	NT	11	11	10	10
10b	_	8	8	9	10
10c	_	_	_	_	_
10d	9	_	8	8	_
10e	10	_	_	_	_
15	_	14	12	16	_
16	_	14	14	14	8
25a	_	_	_	8	7
25b	_	_	_	_	_
Other anion reco	gnition groups				
26a		_	_	7	_
26b	_	_	_	8	_
26c	_	_	_	7	_
26d	_	_	_	8	_
COL^b	20	19	20	_	_

 $[^]a$ Measured after incubation of disk (6 mm diameter, 50 μg per disk) at 37 °C for 20 h. b Tested at 10 μg per disk. c NT = not tested.

Table 2 MIC values ($\mu g \text{ mL}^{-1}$) and cLog P values^a

Compound	A. baumannii ATCC 19606	P. aeruginosa ATCC 27853	K. pneumoniae ATCC 700603	E. coli ATCC 25922	S. aureus MRSA ATCC 43300	cLog P
Amines						
8a	>32	>32	>32	>32	>32	-0.56
8b	>32	16-32	>32	4-8	2	1.46
8c	32	>32	>32	32	0.5	3.48
8d	>32	16-32	16	1-2	1	1.82
Guanidines						
10a	>32	>32	>32	>32	32	-1.93
12	>32	>32	>32	>32	32	-1.93
10b	>32	16	>32	4	2	0.09
10c	16-32	>32	>32	32	1	2.11
1	8-16	>32	>32	32	0.5	2.11
10d	32	16	32	4-8	1	-1.39
10e	>32	>32	>32	>32	>32	0.45
15	>32	>32	>32	>32	>32	-0.85
16	>32	>32	>32	>32	32	2.04
25a	32	32	32	32	4	6.56
25 b	>32	>32	>32	>32	32	-3.07
COL	0.06	0.25	0.03	0.06	>32	

^a Calculated using http://www.molinspiration.com software.

Diamines 8c (hexadecyl chain) and 8d (3-[4-(octyloxy) phenyl propyl chain) both had MIC values of 1 µg mL⁻¹ against MRSA. Indeed, compound 8d was active against a variety of isolates tested with MIC values of 2, 16 and 32 µg mL⁻¹ against E. coli, K. pneumoniae and P. aeruginosa respectively (Table 2). When a dodecanyl tail was present with two guanidine moieties (10b) activity was observed against E. coli and MRSA (MIC values of 2 and 4 µg mL⁻¹ respectively, Table 2). Similarly diguanidine 10d, which contains both alkyl and aryl portions in its hydrophobic tail, showed activity against all bacterial strains tested; highest activity was exhibited against MRSA (1 μg mL⁻¹, Table 2). It is also worth noting that a large calculated Log P (cLog P) range is evident for the active compounds (-3.07-6.56, Table 2), which is in accordance with antibacterial compounds typically showcasing a broader cLog P range when compared to other pharmaceuticals.21

Compounds with MIC values $\leq 2 \mu g \text{ mL}^{-1}$ against Grampositive MRSA (Table 2), were subjected to a second round of micro-broth dilution assay against additional Gram-positive bacterial isolates including: multi-resistant methicillin resistant S. aureus (mMRSA), glycopeptide-intermediate S. aureus (GISA), Vancomycin-intermediate S. aureus (VISA), Streptococcus pneumoniae and Enterococcus faecalis (Table 3). Compounds with a dodecyl chain attached (diamine 8b and diguanidine 10b) exhibited good activity against the Gram-positive bacterial strains shown in Table 3 with MIC values ranging from 2-8 µg mL⁻¹. The larger hexadecyl-substituted analogues (diamine 8c and diguanidines 10c and 1) showed excellent activity with MIC values $\leq 2 \mu \text{g mL}^{-1}$ for all bacterial strains tested in this

Table 3 MIC values (μg mL⁻¹) and cell line cytotoxicity

	Compound							
	8b	8c	8d	10b	10c	1	10d	VAN ^a
S. aureus mMRSA	8	2	1	4	2	2	2	1
S. aureus GISA, NRS 17	2	0.5	1	2	1	0.5	1	4
S. aureus VISA, NRS 1	4	1	1	4	2	1	2	8
S. aureus MRSA	8	1	1	4	2	2	2	2
S. pneumoniae MDR ATCC 700677	2	0.5	1	2	1	0.5	1	2
E. faecalis VanA	8	0.5	2	4	1	1	2	>32
	CC_{50}							
	8b	8c	8d	10b	10c	1	10d	TAM^b
HEK293 ATCC CRL-1573	9	7	12	7	12	6	13	11.1
HepG2 ATCC HB-8065	11	6	12	10	11	9	14	18.7

^a VAN = vancomycin. ^b TAM = Tamoxifen.

assay. Of particular note were diamines **8c** and **8d** which showed MIC comparable to, or better than, vancomycin against all strains tested (Table 3).

Cytotoxicity against human embryonic kidney cells (HEK293) and hepatocellular carcinoma (HepG2) was also determined (Table 3). In all cases the compounds exhibited some cytotoxicity to both HEK293 and HepG2; however, modest selectivity for bacterial cells in preference to human cells for the compounds presented here is apparent (*e.g.* for compound 8d MIC = $1-2 \mu g \, \mathrm{mL}^{-1}$, CC₅₀ = 12).

Conclusions

Herein, we have described a high-yielding and scalable (to multi-gram amounts) method to access highly functionalised norbornane frameworks. A series of cationic amphiphilic antibacterial agents were synthesised which possess a variety of aminium, guanidinium, imidazolium and pyridinium groups.

Detailed SAR indicates that dicationic species were significantly more active than monocationic species, which in turn were more active than charge neutral compounds. The relationship between activity and the cationic group follows the trend: aminium \approx guanidinium > imidazolium \approx pyridinium. The nature of the counterion appears to have, at best, a minor influence on activity.

Compounds containing larger hydrophobic groups typically led to increased antibacterial activity. The hexadecyl analogues (diamine **8c** and diguanidine **10c**) and 3-[4-(octyloxy)phenyl]propyl analogues (diamine **8d** and diguanidine **10d**) were the most potent compounds identified in this study with MIC $\leq 2~\mu g~mL^{-1}$ against all Gram-positive bacterial isolates tested.

The results presented here reinforce the notion that the activity of cationic antimicrobial peptides can be mimicked by relatively small, structurally rigid amphiphiles. Indeed, when compared to other synthetic scaffolds (such as calixarenes) which are used to generate antibacterial amphiphiles, ²² they combine a relatively low molecular weight with potent antibacterial activity.

Experimental

The following compounds were prepared using literature methods and full reaction details can be found in the ESI;† hexadecanal,²³ methyl-3-(4-hydroxyphenyl)propionate,²⁴ 4-(octyloxy)benzaldehyde,²⁵ 1,⁹ 3,⁹ 4a,⁹ 5a,⁹ 5c,⁹ 6,¹¹ 8a,⁹ 9c,⁹ 11,^{14a} 17,^{14a} 18,^{14a} 19,^{16b} S1²⁶ and S2.¹⁹

Methyl-5,6-dihydroxy-endo-3-(heptylcarbamoyl)bicyclo[2.2.1]-heptane-2-exo-carboxylate (21a). To the stirring solution of alkene 20a (1.12 g, 3.82 mmol), NMO·H $_2$ O (570 mg, 4.20 mmol), and a 1:4 ratio of H $_2$ O/acetone (9.3 mL), was added OsO $_4$ (490 μ L, 0.08 mmol) in one addition and the reaction was stirred at ambient temperature for 3 d. The reaction mix was quenched with sat. Na $_2$ S $_2$ O $_5$ (15 mL), and extracted with EtOAc (3 × 30 mL). The combined organic phase was

washed with brine (30 mL), dried (MgSO₄), filtered, and concentrated *in vacuo* to afford a yellow oil (1.06 g, 85%). ¹H NMR (500 MHz, CDCl₃) δ 0.87 (3H, t, J = 6.9 Hz, CH₃), 1.25–1.31 (8H, m, 4 × CH₂), 1.36 (1H, d, J = 10.9 Hz, H7s), 1.47–1.50 (2H, m, NHCH₂CH₂), 1.89 (1H, dd, J = 10.9, 1.2 Hz, H7a), 2.40 (1H, d, J = 3.1 Hz, H4), 2.52 (1H, br s, H1), 2.73 (1H, d, J = 5.7 Hz, H2), 2.92 (1H, dd, J = 5.6, 4.9 Hz, H3), 3.19–3.24 (2H, m, NHCH₂), 3.69 (3H, s, OMe), 3.87 (1H, d, J = 5.6 Hz, H6), 4.02 (1H, d, J = 5.2 Hz, H5), 5.95 (1H, t, J = 5.4 Hz, NH). ¹³C NMR (125 MHz, CDCl₃) δ 14.2, 22.7, 27.0, 29.1, 29.7, 31.8, 32.4, 40.0, 45.1, 47.1, 47.7, 48.1, 52.5, 69.8, 73.4, 171.6, 175.0. HRMS (ESI, m/z) for C₁₇H₂₉NO₅ [M + H]⁺ calc. 328.2119; found 328.2114.

Methyl-5,6-dihydroxy-endo-3-(hexadecylcarbamoyl)bicyclo-[2.2.1]heptane-2-exo-carboxylate (21b). To the stirring solution of alkene 20b (1.34 g, 3.18 mmol), NMO·H₂O (475 mg, 3.50 mmol), CHCl₃ (10 mL), and a 1:4 ratio of H₂O/acetone (8.0 mL), was added OsO₄ (4% in H₂O, 410 μL, 0.06 mmol) and the reaction was stirred at ambient temperature for 16 h, before being heated for a further 24 h at 50 °C. Further OsO₄ (4% in H₂O, 410 μL, 0.06 mmol) was added and the reaction was stirred for another 48 h before being quenched with sat. $Na_2S_2O_5$ (15 mL), and extracted with CHCl₃ (3 × 25 mL). The combined organic phase was washed with brine (25 mL), dried (MgSO₄), filtered, and dried in vacuo to afford a dark green wax (1.28 g, 89%). ¹H NMR (500 MHz, CDCl₃) δ 0.87 (3H, t, J = 6.9 Hz, CH₃), 1.25–1.28 (26H, m, 13 × CH₂), 1.37 (1H, d, J =10.9 Hz, H7s), 1.47-1.50 (2H, m, NHCH₂CH₂), 1.68 (2H, br s, $2 \times OH$), 1.90 (1H, dd, J = 10.9, 1.2 Hz, H7a), 2.29 (1H, d, J = 10.9) 3.2 Hz, H4), 2.52 (1H, br s, H1), 2.74 (1H, d, J = 5.4 Hz, H2), 2.92 (1H, app. t, J = 5.4 Hz, H3), 3.18-3.24 (2H, m, NHC H_2), 3.70 (3H, s, OMe), 3.89 (1H, d, J = 5.8 Hz, H5), 4.04 (1H, d, J =5.7 Hz, H6), 5.85 (1H, t, J = 5.3 Hz, NH). ¹³C NMR (125 MHz, $CDCl_3$) δ 14.3, 22.8, 27.1, 29.4, 29.5 (2 × C), 29.69 (2 × C), 29.74, 29.80 (2 \times C), 29.84 (3 \times C), 32.1, 32.4, 40.0, 45.1, 47.1, 47.7, 48.1, 52.4, 69.8, 73.5, 171.5, 175.0. HRMS (ESI, m/z) for $C_{26}H_{47}NO_5 [M + H]^+$ calc. 454.3520; found 454.3542.

General procedure A: acetal formation

To a stirring suspension of the appropriate diol, $TsOH \cdot H_2O$ (0.05 equiv.), $MgSO_4$ (1.0 equiv.) and PhMe, was treated with the required aldehyde (1.5 equiv.) at 110 °C for 3 h. Solid $MgSO_4$ was removed by filtration and the filtrate was diluted with EtOAc (30 mL), washed with H_2O (2 × 15 mL), brine (15 mL), dried ($MgSO_4$), filtered and concentrated *in vacuo* to give the crude material which was purified by column chromatography (as specified below) to afford the title compound.

Dimethyl 4-undecanyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]**decane-8***endo-9-exo-***dicarboxylate (4b).** Compound **4b** was prepared from diol 3 (947 mg, 3.87 mmol) and dodecanal (1.3 mL, 5.86 mmol) according to general procedure A and was purified by column chromatography (5–10% EtOAc in pet. spirits) to give the title compound (1.33 g, 83%) as a white waxy solid; $R_f = 0.24$ (10% EtOAc in pet. spirits). ¹H NMR (500 MHz, CDCl₃) δ 0.87 (3H, t, J = 6.9 Hz, CH₃), 1.24–1.39 (19H, m, $9 \times \text{CH}_2$, H10s), 1.60–1.65 (2H, m, CHCH₂), 1.78 (1H, dd, J = 6.9 Hz, CH₂), 1.78 (1H, dd, J = 6.9 Hz,

10.9, 1.4 Hz, H10a), 2.40-2.65 (2H, m, H1, H7), 2.72 (1H, d, J = 4.4 Hz, H9), 3.23 (1H, app. t, J = 5.1 Hz, H8), 3.70 (6H, s, $2 \times OMe$), 3.90 (1H, d, J = 5.5 Hz, H6), 4.03 (1H, d, J = 5.6 Hz, H2), 4.65 (1H, t, J = 4.9 Hz, H4). ¹³C NMR (125 MHz, CDCl₃) δ 14.3, 22.8, 24.4, 29.5, 29.6, 29.7 (2 × C), 29.8 (2 × C), 31.8, 32.1, 32.9, 43.4, 43.8, 45.1, 45.4, 52.3, 52.5, 78.9, 81.4, 104.3, 172.9, 174.1. HRMS (ESI, m/z) for $C_{23}H_{38}O_6$ [M + H]⁺ calc. 411.2741; found 411.2745.

4-hexadecyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-Dimethyl endo-9-exo-dicarboxylate (4c). Compound 4c was prepared from diol 3 (239 mg, 0.98 mmol) and hexadecanal S6 (353 mg, 1.47 mmol) according to general procedure A and was purified by column chromatography (5% EtOAc in pet. spirits) to give the title compound (276 mg, 60%) as a white solid; $R_f = 0.31$ (10% EtOAc in pet. spirits). m.p: 71.0-73.7 °C. ¹H NMR (270 MHz, CDCl₃) δ 0.88 (3H, t, J = 6.4 Hz, CH₃), 1.25-1.41 $(27H, m, 13 \times CH_2, H10s), 1.59-1.66 (2H, m, CHCH_2), 1.78 (1H, Max CH_2)$ dd, J = 10.7, 1.4 Hz, H10a), 2.64-2.66 (2H, m, H1, H7), 2.72 (1H, d, J = 4.8 Hz, H9), 3.22 (1H, app. t, J = 5.0 Hz, H8), 3.70(6H, s, 2 × OMe), 3.90 (1H, d, J = 5.3 Hz, H6), 4.03 (1H, d, J =5.5 Hz, H2), 4.65 (1H, t, J = 4.8 Hz, H4). ¹³C NMR (67.5 MHz, $CDCl_3$) δ 14.3, 22.8, 24.4, 29.5, 29.6, 29.7 (2 × C), 29.8 (6 × C), 31.8, 32.1, 32.9, 43.4, 43.8, 45.2, 45.4, 52.3, 52.5, 78.9, 81.4, 104.4, 172.9, 174.1. HRMS (ESI, m/z) for $C_{27}H_{46}O_6$ [M + H]⁺ calc. 467.3367; found 467.3378.

Dimethyl 4-[4'-(octyloxy)phenethyl]-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exo-dicarboxylate (4d). Compound 4d was prepared from diol 3 (275 mg, 1.13 mmol) and aldehyde S10 (437 mg, 1.67 mmol) according to general procedure A and was purified by column chromatography (5-10-20% EtOAc in pet. spirits) to give the title compound (265 mg, 48%) as a yellow oil; $R_f = 0.62$ (20% EtOAc in pet. spirits). ¹H NMR (500 MHz, CDCl₃) δ 0.88 (3H, t, J = 7.1 Hz, CH₃), 1.28–1.38 (9H, m, 4 × CH₂, H10s), 1.41-1.47 (2H, m, CH₂), 1.73-1.78 (2H, m, CH₂), 1.81 (1H, dd, J = 10.8, 1.5 Hz, H10a), 1.91-1.95(2H, m, CH₂), 2.65-2.69 (4H, m, H1, H7, ArCH₂), 2.74 (1H, d, J = 4.5 Hz, H9), 3.24 (1H, app. t, J = 5.0 Hz, H8), 3.70 (3H, s, Me), 3.71 (3H, s, Me), 3.90-3.93 (3H, m, H6, OCH₂), 4.05 (1H, d, J = 5.7 Hz, H2), 4.68 (1H, t, J = 4.8 Hz, H4), 6.79–6.82 (2H, m, ArH), 7.07-7.09 (2H, m, ArH). ¹³C NMR (125 MHz, $CDCl_3$) δ 14.2, 22.8, 26.2, 29.4, 29.5 (3 × C), 31.8, 32.0, 34.6, 43.4, 43.8, 45.2, 45.4, 52.3, 52.5, 68.2, 79.0, 81.5, 103.5, 114.6 (2 × C), 129.3 (2 × C), 133.2, 157.6, 172.9, 174.1. HRMS (ESI, m/z) for $C_{28}H_{40}O_7$ [M + K]⁺ calc. 527.2406; found 527.2410.

Dimethyl 4-ethylbenzene-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exo-dicarboxylate (4e). Compound 4e was prepared from diol 3 (1.01 g, 4.14 mmol) and 3-phenylpropionaldehyde (820 µL, 6.21 mmol) according to general procedure A and was purified by column chromatography (10% EtOAc in pet. spirits) to give the title compound (1.18 g, 79%) as a yellow oil; $R_{\rm f} = 0.22 \ (10\% \ \text{EtOAc} \ \text{in pet. spirits}).$ ¹H NMR (270 MHz, $CDCl_3$) δ 1.38 (1H, dt, J = 10.8, 1.4 Hz, H10s), 1.82 (1H, dd, J = 10.8, 1.5 Hz, H10a), 1.93-2.01 (2H, m, CHCH₂), 2.65-2.77 (5H, m, H1, H7, H9, ArC H_2), 3.25 (1H, app. t, J = 5.1 Hz, H8), 3.71 (3H, s, Me), 3.72 (3H, s, Me), 3.94 (1H, d, J = 5.6 Hz, H6), 4.06

(1H, d, J = 5.6 Hz, H2), 4.70 (1H, t, J = 4.8 Hz, H4), 7.16-7.31(5H, m, ArH). 13 C NMR (67.5 MHz, CDCl₃) δ 30.4, 31.8, 34.4, 43.4, 43.8, 45.2, 45.4, 52.3, 52.5, 79.0, 81.5, 103.4, 126.1, 128.46 (2 × C), 128.53 (2 × C), 141.4, 172.9, 174.1. HRMS (ESI, m/z) for $C_{20}H_{24}O_6$ [M + H]⁺ calc. 361.1646; found 361.1655.

Methyl 4-heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-(heptylcarbamoyl)-9-exo-carboxylate (22a). Compound 22a was prepared from diol 21a (196 mg, 0.60 mmol) and octanal (140 µL, 0.90 mmol) according to general procedure A and was purified by column chromatography (10-20% EtOAc in pet. spirits) to give the title compound (247 mg, 94%) as a clear oil; $R_{\rm f}$ = 0.45 (20% EtOAc in pet. spirits). ¹H NMR (500 MHz, $CDCl_3$) δ 0.85-0.89 (6H, m, 2 × CH_3), 1.27-1.40 (19H, m, $9 \times CH_2$, H10s), 1.47-1.50 (2H, m, NHCH₂CH₂), 1.60-1.65 (2H, m, CHC H_2), 1.80 (1H, d, J = 10.6 Hz, H10a), 2.53 (1H, d, J = 4.5 Hz, H7), 2.66 (1H, br s, H1), 2.77 (1H, d, J = 5.5 Hz, H9), 2.98 (1H, app. t, J = 5.1 Hz, H8), 3.19–3.27 (2H, m, NHC H_2), 3.69 (3H, s, OMe), 4.09 (1H, d, J = 5.7 Hz, H6), 4.14 (1H, 5.7 Hz, H2), 4.65 (1H, t, J = 4.9 Hz, H4), 5.63 (1H, t, J = 5.0 Hz, NH). 13 C NMR (125 MHz, CDCl₃) δ 14.22, 14.23, 22.7, 22.8, 24.4, 27.0, 29.0, 29.3, 29.66, 29.74, 31.8, 31.9, 32.3, 33.0, 40.0, 43.6, 44.1, 45.1, 46.7, 52.5, 78.5, 81.4, 104.1, 170.8, 174.8. HRMS (ESI, m/z) for $C_{25}H_{43}NO_5 [M + H]^+$ calc. 438.3214; found 438.3205.

4-heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-Methyl (hexadecylcarbamovl)-9-exo-carboxylate (22b). Compound 22b was prepared from diol 21b (76 mg, 0.17 mmol) and octanal (40 μL, 0.26 mmol) according to general procedure A and was purified by column chromatography (10-20% EtOAc in pet. spirits) to give the title compound (43 mg, 45%) as a clear oil; $R_{\rm f}$ = 0.47 (20% EtOAc in pet. spirits). ¹H NMR (500 MHz, CDCl₃) δ 0.85-0.89 (6H, m, 2 × CH₃), 1.25-1.32 (35H, m, 17 × CH₂, H10s), 1.35-1.44 (2H, m, CH₂), 1.47-1.50 (2H, m, CH_2), 1.60–1.64 (2H, m, CH_2), 1.80 (1H, dd, J = 10.7, 1.3 Hz, H10a), 2.52 (1H, d, J = 4.4 Hz, H1), 2.65 (1H, br s, H7), 2.76 (1H, dd, J = 5.4, 1.1 Hz, H9), 2.98 (1H, app. t, J = 5.1 Hz, H8),3.18-3.27 (2H, m, NHC H_2), 3.69 (3H, s, OMe), 4.08 (1H, d, J=5.6 Hz, H2), 4.14 (1H, d, J = 5.6 Hz, H6), 4.65 (1H, t, J = 4.9 Hz, H4), 5.66 (1H, t, J = 3.7 Hz, NH). ¹³C NMR (100 MHz, CDCl₃) δ 14.2, 14.3, 22.76, 22.83, 24.4, 27.0, 29.1, 29.2, 29.3, 29.4, 29.5, $29.67, 29.72, 29.75, 29.80, 29.82, 29.84 (3 \times C), 31.9, 32.0, 32.3,$ 33.0, 40.0, 43.6, 44.1, 45.1, 46.7, 52.4, 78.6, 81.4, 104.1, 170.8, 174.8. HRMS (ESI, m/z) calculated for $C_{34}H_{61}NO_5$ [M + H] 564.4623; found 564.4631.

General procedure B: hydrolysis of methyl esters

A biphasic solution of methyl ester in 2 M NaOH/THF (1:4) was stirred at ambient temperature for 16 h. The reaction mixture was extracted with CH_2Cl_2 (2 × 8 mL) and the isolated aqueous phase was acidified to pH = 1 using 2 M HCl and extracted with EtOAc (3 × 15 mL). The combined organic phase was washed with brine (10 mL), dried (MgSO₄), filtered and concentrated in vacuo to afford the title compound.

4-Undecanyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]**decane-8-***endo***-9-***exo***dicarboxylic acid** (**5b**). The title compound was prepared from diester **4b** (137 mg, 0.33 mmol) according to general procedure B and isolated white powder (105 mg, 83%). m.p: 138.1–139.5 °C. ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.8 Hz, CH₃), 1.29–1.42 (19H, m, 9 × CH₂, H10s), 1.59–1.63 (2H, m, CHCH₂), 1.75 (1H, dd, J = 10.6, 1.2 Hz, H10a), 2.56 (1H, dd, J = 5.5, 1.0 Hz, H1), 2.58 (1H, br s, H7), 2.64 (1H, app. t, J = 4.4 Hz, H9), 3.17 (1H, app. t, J = 5.0 Hz, H8), 3.99 (1H, d, J = 5.6 Hz, H6), 4.02 (1H, d, J = 5.6 Hz, H2), 4.67 (1H, t, J = 4.8 Hz, H4). ¹³C NMR (125 MHz, CD₃OD) δ 14.5, 23.8, 25.2, 30.5 (2 × C), 30.7 (3 × C), 30.8, 32.4, 33.1, 33.9, 44.4, 44.9, 46.4, 46.6, 80.2, 82.7, 105.2, 175.5, 176.9. HRMS (ESI, m/z) for C₂₁H₃₄O₆ [M + Na]⁺ calc. 405.2248; found 405.2254.

4-[4'-(Octyloxy)phenethyl]-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exo-dicarboxylic acid (5d). The title compound was prepared from diester 4d (260 mg, 0.532 mmol) according to general procedure B and isolated as a white solid (197 mg, 80%). m.p: 144.9-147.4 °C. ¹H NMR (270 MHz, DMSO-d₆) δ 0.85 (3H, t, J = 5.9 Hz, CH₃), 1.25-1.41 (11H, m, 5 × CH₂, H10s), 1.62-1.72 (3H, m, CH₂, H10a), 1.77-1.85 (2H, m, CH₂), 2.42 (1H, d, J = 5.2 Hz, H1), 2.52–2.60 (4H, m, CH₂, H7, H9), 3.02 (1H, app. t, J = 5.2 Hz, H8), 3.89–3.92 (3H, m, OCH₂, H6), 4.00 (1H, d, J = 5.6 Hz, H2), 4.64 (1H, t, J = 4.7 Hz, H4), 6.80(2H, d, J = 8.5 Hz, ArH), 7.09 (2H, d, J = 8.5 Hz, ArH). ¹³C NMR (67.5 MHz, DMSO- d_6) δ 14.0, 22.1, 25.6, 28.7 (2 × C), 28.8 $(2 \times C)$, 31.2 $(2 \times C)$, 34.2, 42.6, 43.3, 44.4, 45.0, 67.3, 78.3, 80.8, 102.5, 114.3 (2 × C), 129.1 (2 × C), 132.9, 156.9, 173.3, 174.6. HRMS (ESI, m/z) for $C_{26}H_{36}O_7 [M + Na]^+$ calc. 483.2352; found 483.2364.

4-Ethylbenzene-3,5-dioxatricyclo[5.2.1.0^{2,6}]**decane-8-endo-9-exo-carboxylic acid** (5e). The title compound was prepared from diester **4e** (1.13 g, 3.12 mmol) according to general procedure B and isolated as a white solid (805 mg, 78%). m.p: 179.7–181.3 °C. ¹H NMR (270 MHz, DMSO- d_6) δ 1.24 (1H, d, J = 10.8 Hz, H10s), 1.66 (1H, d, J = 9.6 Hz, H10a), 1.83–1.90 (2H, m, CHC H_2), 2.42 (1H, d, J = 5.5 Hz, H9), 2.50–2.52 (1H, m, H1), 2.58 (1H, d, J = 4.7 Hz, H7), 2.64–2.69 (2H, m, ArCH₂), 3.02 (1H, app. t, J = 3.5 Hz, H8), 3.93 (1H, d, J = 5.6 Hz, H6), 4.01 (1H, d, J = 4.8 Hz, H2), 4.67 (1H, t, J = 4.8 Hz, H4), 7.17–7.30 (5H, m, ArH), 12.59 (2H, br s, 2 × OH). ¹³C NMR (67.5 MHz, DMSO- d_6) δ 29.7, 31.2, 33.9, 42.6, 43.3, 44.4, 45.0, 78.3, 80.8, 102.5, 125.9, 128.2 (2 × C), 128.3 (2 × C), 141.2, 173.3, 174.6. HRMS (ESI, m/z) for C₁₈H₂₀O₆ [M + Na]⁺ calc. 355.1152; found 355.1150.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-*endo*(heptylcarbamoyl)-9-*exo*-carboxylic acid (23a). The title compound was prepared from ester 22a (240 mg, 0.55 mmol) according to general procedure B and isolated as a white waxy solid (148 mg, 64%). m.p: 137.4–139.6 °C. ¹H NMR (500 MHz, CD₃OD) δ 0.89–0.91 (6H, m, 2 × CH₃), 1.30–1.41 (19H, m, 9 × CH₂, H10s), 1.50–1.53 (2H, m, NHCH₂CH₂), 1.58–1.62 (2H, m, CHCH₂), 1.75 (1H, dd, J = 10.5, 1.4 Hz, H10a), 2.55 (1H, d, J = 4.1 Hz, H7), 2.59 (1H, br s, H1), 2.68 (1H, d, J = 4.8 Hz, H9), 3.05 (1H, app. t, J = 5.2 Hz, H8), 3.13 (1H, dt, J = 13.3, 7.0 Hz, NHCH₂), 3.25 (1H, dt, J = 13.3, 7.0 Hz, NHCH₂), 3.25 (1H, dt, J = 13.3, 7.0 Hz, NHCH₂), 4.01 (1H, dt, J = 13.4 (1) (1H, dt, J = 13.5) (1H, dt, J = 13.6 (1H, dt, J = 13.7) (1H, dt, J = 13.8 (1H, dt, J = 13.8) (1H, dt, J = 13.9) (1H, dt,

J = 5.6 Hz, H6), 4.03 (1H, d, J = 5.7 Hz, H3), 4.65 (1H, t, J = 4.8 Hz, H4). ¹³C NMR (125 MHz, CD₃OD) δ 14.4, 14.5, 23.66, 23.70, 25.2, 27.9, 30.1, 30.3, 30.4, 30.6, 32.9, 33.0, 33.1, 33.9, 40.5, 44.3, 45.4, 46.3, 47.8, 79.9, 82.6, 105.1, 173.4, 177.2. HRMS (ESI, m/z) for C₂₄H₄₁NO₅ [M + H]⁺ calc. 424.3058; found 424.3044.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo(hexadecylcarbamoyl)-9-exo-carboxylic acid (23b). The title compound was prepared from ester 22b (41 mg, 0.07 mmol) according to general procedure B and isolated as a brown wax (37 mg, 93%). ¹H NMR (270 MHz, CDCl₃) δ 0.85–0.90 (6H, m, $2 \times CH_3$, 1.25-1.52 (39H, m, $18 \times CH_2$, H10s), 1.59-1.67 (2H, m, $CHCH_2$), 1.83 (1H, d, J = 10.1 Hz, H10a), 2.54 (1H, d, J = 10.1 Hz, H10a), 2.55 (1H, d, J = 10.1 Hz, H10a), 2.55 (1H, d, J = 10.1 Hz, H10a), 2.55 (1H, d, J = 10.1 Hz, J = 10.13.3 Hz, H7), 2.71-2.73 (2H, m, H1, H9), 2.95 (1H, dd, J = 5.5, 4.8 Hz, H8), 3.19-3.28 (2H, m, NHC H_2), 4.07 (1H, d, J = 5.7 Hz, H6), 4.14 (1H, d, J = 5.7 Hz, H2), 4.66 (1H, t, J = 4.8 Hz, H4), 5.85 (1H, t, J = 4.1 Hz, NH). ¹³C NMR (67.5 MHz, CDCl₃) δ 14.2, 14.3, 22.75, 22.82, 24.3, 27.0, 29.3, 29.4, 29.5, 29.6, 29.67 $(2 \times C)$, 29.73, 29.80 $(2 \times C)$, 29.83, 29.84 $(3 \times C)$, 31.9, 32.1, 32.5, 32.9, 40.1, 43.7, 43.9, 44.6, 46.9, 78.5, 81.4, 104.3, 171.3, 177.9. HRMS (ESI, m/z) for $C_{33}H_{59}NO_5 [M + H]^+$ calc. 550.4466; found 550.4455.

General procedure C: amide formation

A microwave vial was charged with the appropriate carboxylic acid, EDCI (3.0 equiv.), HOBt (0.1 equiv.) and anhydrous CHCl₃ and was stirred at ambient temperature for 30 min. The appropriate alkylamine (3.0 equiv.) was then added and the reaction was heated at 50 °C for 30 min using microwave irradiation. The resulting homogenous clear solution was diluted with CHCl₃ (20 mL), washed with H₂O (2 × 10 mL), brine (8 mL), dried (MgSO₄), filtered, and concentrated *in vacuo* to afford the crude material that was purified by column chromatography (as specified below) to give the title compound.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylamino)ethylcarbamoyl]-4-heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (7a). Compound 7a was prepared from diacid 5a (333 mg, 1.02 mmol) and amine 6 (490 mg, 3.06 mmol) according to general procedure C and after purification by column chromatography (50-70% EtOAc in pet. spirits-EtOAc) was isolated as a white solid (357 mg, 57%); $R_f = 0.21$ (70% EtOAc in pet. spirits). m.p: 121.7–123.4 °C. ¹H NMR (270 MHz, CDCl₃) δ 0.87 (3H, t, J = 6.9 Hz, CH₃), 1.25–1.44 (28H, m, $5 \times \text{CH}_2$, t-Bu), 1.57-1.65 (3H, m, CHC H_2 , H10s), 1.80 (1H, d, J = 3.6 Hz, H10a), 2.43 (1H, d, J = 5.0 Hz, H1), 2.53–2.57 (2H, m, H7, H9), 2.92 (1H, app. t, J = 5.1 Hz, H8), 3.26-3.41 (8H, m, $4 \times \text{CH}_2$), 3.96 (1H, d, J = 5.8 Hz, H2), 4.13 (1H, d, J = 5.5 Hz, H6), 4.64 $(1H, t, J = 4.8 \text{ Hz}, H4), 5.01-5.08 (2H, m, 2 \times NH), 6.89 (1H, br)$ s, NH), 6.86 (1H, br s, NH). 13 C NMR (67.5 MHz, CDCl₃) δ 14.2, 22.8, 24.4, 28.5, 29.3, 29.6, 29.8, 31.9, 32.6, 33.0, 40.4, 40.6, 41.2, 43.4 (2 × C), 44.5, 44.7, 47.8, 78.8, 79.8, 80.0, 81.6, 104.2, 156.8, 157.1, 172.5, 174.3. HRMS (ESI, m/z) for $C_{31}H_{54}N_4O_8$ $[M + H]^{+}$ calc. 611.4014; found 611.4031.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylamino)ethyl-carbamoyl]-4-undecanyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (7b). Compound 7b was prepared from diacid 5b (512 mg,

1.34 mmol) and amine 6 (670 mg, 4.18 mmol) according to general procedure C and after purification by column chromatography (50% EtOAc in pet. spirits-EtOAc) was isolated as a white solid (542 mg, 61%); $R_f = 0.43$ (EtOAc). ¹H NMR (500 MHz, CDCl₃) δ 0.87 (3H, t, J = 7.1 Hz, CH₃), 1.24-1.39 $(18H, m, 9 \times CH_2), 1.43-1.44 (18H, m, 2 \times t-Bu), 1.49 (1H, d, J =$ 10.5 Hz, H10s), 1.60-1.64 (2H, m, CHC H_2), 1.81 (1H, d, J =9.9 Hz, H10a), 2.40 (1H, d, J = 5.6 Hz, H1), 2.54 (1H, d, J =3.9 Hz, H7), 2.59 (1H, br s, H9), 2.88 (1H, dd, J = 5.8, 4.6 Hz, H8), 3.22-3.44 (8H, m, $4 \times CH_2$), 3.96 (1H, d, J = 5.6 Hz, H2), 4.13 (1H, d, J = 5.6 Hz, H6), 4.64 (1H, t, J = 4.8 Hz, H4), 4.92-5.02 (2H, m, 2 × NH), 6.63 (1H, br s, NH), 6.77 (1H, br s, NH). ¹³C NMR (125 MHz, CDCl₃) δ 14.3, 22.8, 24.4, 28.5, 29.5, 29.67, 29.68, 29.71, 29.76, 29.79, 32.1, 32.6, 33.0, 40.4, 40.5, 40.7, 41.2, 43.3, 44.4, 44.5, 48.0, 78.7, 79.9, 80.1, 81.6, 104.2, 156.8, 157.1, 172.5, 174.3. HRMS (ESI, m/z) for $C_{35}H_{62}N_4O_8$ $[M + H]^{+}$ calc. 667.4640; found 667.4656.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylamino)ethylcarbamoyl]-4-hexapentyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (7c). Compound 7c was prepared from diacid 5c (100 mg, 0.23 mmol) and amine 6 (110 mg, 0.68 mmol) according to general procedure C and after purification by column chromatography (50-70% EtOAc in pet. spirits) was isolated as clear oil (76 mg, 46%); $R_f = 0.07$ (70% EtOAc in pet. spirits). ¹H NMR (500 MHz, CDCl₃) δ 0.87 (3H, t, J = 6.9 Hz, CH₃), 1.24–1.39 $(27H, m, 13 \times CH_2, H10s), 1.43-1.44 (18H, m, 2 \times t-Bu),$ 1.59-1.64 (2H, m, CHC H_2), 1.81 (1H, d, J = 9.8 Hz, H10a), 2.42(1H, d, J = 5.7 Hz, H1), 2.54 (1H, d, J = 3.7 Hz, H7), 2.57 (1H, br)s, H9), 2.91 (1H, app. t, J = 4.9 Hz, H8), 3.22-3.43 (8H, m, $4 \times CH_2$, 3.96 (1H, d, J = 5.5 Hz, H2), 4.13 (1H, d, J = 4.8 Hz, H6), 4.64 (1H, t, J = 4.9 Hz, H4), 5.01-5.08 (2H, m, 2 × NH), 6.69 (1H, br s, NH), 6.84 (1H, br s, NH). ¹³C NMR (125 MHz, $CDCl_3$) δ 14.3, 22.8, 24.4, 28.5, 29.5, 29.67, 29.69, 29.72, 29.80 $(2 \times C)$, 29.81 $(2 \times C)$, 29.84 $(2 \times C)$, 32.1, 32.6, 33.0, 34.0, 40.5, 40.6, 41.1, 43.4, 44.5, 44.6, 47.8, 78.7, 79.8, 80.0, 81.6, 104.2, 156.8, 157.1, 172.5, 174.3. HRMS (ESI, m/z) for $C_{39}H_{70}N_4O_8$ $[M + H]^{+}$ calc. 723.5266; found 723.5263.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylamino)ethylcarbamoyl]-4-[4'-(octyloxy)phenethyl]-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (7d). Compound 7d was prepared from diacid 5d (51 mg, 0.11 mmol) and amine 6 (78 mg, 0.49 mmol) according to general procedure C and after purification by column chromatography (50% EtOAc in CH₂Cl₂-EtOAc) was isolated as a clear oil (29 mg, 35%); $R_f = 0.34$ (EtOAc). ¹H NMR (500 MHz, $CDCl_3$) δ 0.88 (3H, t, J = 6.9 Hz, CH_3), 1.25-1.36 (8H, m, $4 \times CH_2$, 1.42-1.43 (20H, m, CH_2 , t-Bu), 1.51 (1H, d, J =10.1 Hz, H10s), 1.72-1.77 (2H, m, CH_2), 1.81 (1H, d, J =10.0 Hz, H10a), 1.88-1.92 (2H, m, CH_2), 2.51 (1H, d, J = 5.4 Hz, H1), 2.56 (1H, s, H7), 2.59 (1H, d, J = 3.7 Hz, H9), 2.65 (2H, t, J = 8.1 Hz, ArCH₂), 3.05 (1H, app. t, J = 4.7 Hz, H8), 3.25–3.34 (8H, m, $4 \times CH_2$), 3.91 (2H, t, J = 6.6 Hz, OCH_2), 4.00 (1H, d, J =5.5 Hz, H6), 4.13 (1H, d, J = 5.3 Hz, H2), 4.66 (1H, t, J = 4.7 Hz, H4), 5.27 (2H, br s, $2 \times NH$), 6.79 (2H, d, J = 8.6 Hz, ArH), 6.97 (1H, br s, NH), 7.06 (2H, d, J = 8.6 Hz, ArH), 7.20 (1H, br s, NH). ¹³C NMR (125 MHz, CDCl₃) δ 14.2, 22.8, 26.2, 28.5, 29.4 $(2 \times C)$, 29.5, 29.8, 31.9, 32.4, 34.8, 40.4 $(2 \times C)$, 40.8 $(2 \times C)$,

43.7, 44.4, 45.5, 47.1, 68.1, 78.9, 79.7, 79.9, 81.7, 103.2, 114.5 $(2 \times C)$, 129.2 $(2 \times C)$, 133.4, 156.8, 157.0, 157.5, 172.5, 174.4. HRMS (ESI, m/z) for $C_{40}H_{64}N_4O_9$ [M + H]⁺ calc. 745.4746; found 745.4768.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylguanidino)propylcarbamoyl]-4-heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (13). Compound 13 was prepared from diacid 5a (170 mg, 0.52 mmol) and amine 17 (968 mg, 3.06 mmol) according to general procedure C and after purification by column chromatography (20-50% EtOAc in CH2Cl2) was isolated as a white residue (234 mg, 49%); $R_f = 0.41$ (50% EtOAc in CH_2Cl_2). ¹H NMR (500 MHz, CDCl₃) δ 0.86 (3H, t, J = 6.5 Hz, CH₃), 1.23-1.27 (8H, m, 4 × CH₂), 1.36-1.41 (2H, m, CH₂), 1.49-1.51 (36H, m, $4 \times t$ -Bu), 1.57 (1H, d, J = 10.4 Hz, H10s), 1.60–1.72 (6H, m, $3 \times CH_2$), 1.77 (1H, d, J = 9.9 Hz, H10a), 2.54 (1H, br s, H7), 2.56 (1H, d, J = 5.3 Hz, H1), 2.76 (1H, d, J = 4.2 Hz, H9), 3.05-3.19 (3H, m, CH₂, H8), 3.29-3.42 (4H, m, $2 \times \text{CH}_2$), 3.47-3.63 (2H, m, CH₂), 4.03 (1H, d, J = 5.7 Hz, H2), 4.05 (1H, d, J = 5.7 Hz, H6), 4.63 (1H, t, J = 4.8 Hz, H4), 6.85 (1H, t, J =5.1 Hz, NH), 7.62 (1H, t, J = 5.7 Hz, NH), 8.45 (2H, br s, 2 × NH), 11.46 (1H, s, NH), 11.49 (1H, s, NH). ¹³C NMR (125 MHz, CDCl₃) δ 14.2, 22.7, 24.3, 28.2, 28.4, 28.5, 29.3, 29.7, 29.8, 30.0, 31.9, 32.3, 33.0, 35.5, 36.5, 37.4, 38.0, 43.6, 44.1, 45.5, 47.3, 79.0, 79.9, 81.8, 83.5, 83.6, 104.0, 153.29, 153.34, 156.7, 157.2, 163.2 (2 × C), 171.8, 174.0. HRMS (ESI, m/z) for $C_{45}H_{78}N_8O_{12}$ [M + H]⁺ calc. 923.5812; found 923.5822.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylguanidino)butylcarbamoyl]-4-heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (14). Compound 14 was prepared from diacid 5a (251 mg, 0.77 mmol) and amine 18 (1.03 g, 3.12 mmol) according to general procedure C and after purification by column chromatography (20-50% EtOAc in CH2Cl2) was isolated as a white residue (382 mg, 52%); $R_f = 0.48$ (50% EtOAc in CH_2Cl_2). ¹H NMR (500 MHz, CDCl₃) δ 0.86 (3H, t, J = 6.8 Hz, CH₃), 1.25-1.28 (10H, m, 5 × CH₂), 1.37-1.63 (47H, m, 4 × t-Bu, $5 \times CH_2$, H10s), 1.80 (1H, d, J = 9.7 Hz, H10a), 2.43 (1H, d, J =5.6 Hz, H9), 2.56-2.57 (2H, m, H1, H7), 2.93 (1H, dd, J = 5.7, 4.7 Hz, H8), 3.19-3.47 (8H, m, $4 \times CH_2$), 3.98 (1H, d, J = 5.6 Hz, H2), 4.09 (1H, d, J = 5.6 Hz, H6), 4.64 (1H, t, J = 4.8 Hz, H4), 6.25-6.44 (2H, m, 2 × NH), 8.38-8.43 (2H, m, 2 × NH), 11.48 (1H, s, NH), 11.50 (1H, s, NH). ¹³C NMR (125 MHz, CDCl₃) δ 14.2, 22.8, 24.4, 26.5, 26.8, 26.9, 27.0, 28.2, 28.41, 28.42, 29.3, 29.7, 31.9, 32.6, 33.0, 39.3, 39.5, 40.7, 40.8, 43.4, 44.4, 44.6, 47.9, 78.8, 79.9, 80.1, 81.7, 83.5, 83.6, 104.2, 153.4 (2 × C), 156.2 (2 × C), 163.2 (2 × C), 171.9, 173.6. HRMS (ESI, m/z) for $C_{47}H_{82}N_8O_{12}$ [M + H]⁺ calc. 951.6125; found 951.6137.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylguanidino)ethylcarbamoyl]-4-ethylbenzene-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (9e). Compound 9e was prepared from diacid 5e (203 mg, 0.61 mmol) and amine 11 (550 mg, 1.83 mmol) according to general procedure C and after purification by flash column chromatography (50% EtOAc in pet. spirits-EtOAc) was isolated as a white waxy solid (242 mg, 44%); $R_f =$ 0.39 (70% EtOAc in pet. spirits).

¹H NMR (270 MHz, CDCl₃) δ 1.49–1.51 (37H, m, *t*-Bu, H10*s*), 1.82 (1H, d, J = 10.6 Hz, H10*a*), 1.90–1.98 (2H, m, CH₂), 2.46 (1H, d, J = 6.2 Hz, H1), 2.61 (1H, br s, H7), 2.70–2.76 (3H, m, ArCH₂, H9), 2.96 (1H, app. t, J = 5.9 Hz, H8), 3.35–3.62 (8H, m, 4 × CH₂), 3.98 (1H, d, J = 5.6 Hz, H2), 4.07 (1H, d, J = 5.3 Hz, H6), 4.66 (1H, t, J = 4.7 Hz, H4), 6.88 (1H, t, J = 5.3 Hz, NH), 7.16–7.30 (5H, m, ArH), 8.05 (1H, t, J = 3.1 Hz, NH), 8.51 (1H, t, J = 5.9 Hz, NH), 8.65 (1H, t, J = 5.7 Hz, NH), 11.47 (1H, s, NH), 11.48 (1H, s, NH). ¹³C NMR (67.5 MHz, CDCl₃) δ 28.2, 28.4, 30.4, 32.5, 34.5, 40.0, 40.2 (2 × C), 42.3, 43.0, 44.3, 44.4, 47.8, 79.2, 79.7, 79.9, 81.8, 83.4, 83.7, 103.2, 126.1, 128.5 (4 × C), 141.6, 153.2 (2 × C), 157.1, 158.0, 163.0, 163.5, 172.0, 174.8. HRMS (ESI, m/z) for C₄₄H₆₇N₈O₁₂ [M + H]⁺ calc. 901.5030; found 901.5052.

Methyl endo-3-(heptylcarbamoyl)bicyclo[2.2.1]hept-5-ene-2exo-carboxylate (20a). Compound 20a was prepared from acid 19 (533 mg, 2.72 mmol) and n-heptylamine (620 μ L, 4.07 mmol) according to general procedure C and after purification by column chromatography (20% EtOAc in CH2Cl2) was isolated as a clear oil (615 mg, 77%); $R_f = 0.32$ (20% EtOAc in CH_2Cl_2). ¹H NMR (500 MHz, $CDCl_3$) δ 0.87 (3H, t, J = 6.9 Hz, CH_3), 1.26–1.30 (8H, m, 4 × CH_2), 1.44–1.48 (3H, m, NHCH₂CH₂, H7s), 1.55 (1H, d, J = 8.7 Hz, H7a), 2.58 (1H, dd, J = 5.0, 1.8 Hz, H2), 3.10-3.25 (5H, m, NHCH₂, H1, H4, H3),3.72 (3H, s, Me), 5.80 (1H, t, J = 6.0 Hz, NH), 6.17 (1H, dd, J =5.7, 3.0 Hz, H6), 6.23 (1H, dd, J = 5.6, 3.4 Hz, H5). ¹³C NMR $(125 \text{ MHz}, \text{CDCl}_3) \delta 14.2, 22.7, 27.0, 29.1, 29.8, 31.9, 39.7, 45.7,$ 46.9, 47.8, 48.2, 49.8, 52.3, 135.7, 136.6, 172.5, 175.7. HRMS (ESI, m/z) for $C_{17}H_{27}NO_3$ [M + H]⁺ calc. 294.2064; found 294.2056.

Methyl endo-3-(hexadecylcarbamoyl)bicyclo[2.2.1]hept-5ene-2-exo-carboxylate (20b). Compound 20b was prepared from acid 19 (511 mg, 2.60 mmol) and n-hexadecylamine (950 mg, 3.90 mmol) according to general procedure C and after purification by column chromatography (5% EtOAc in CH_2Cl_2) was isolated as a clear oil (793 mg, 73%); $R_f = 0.41$ (20% EtOAc in CH₂Cl₂). m.p: 83.5-84.5 °C. ¹H NMR (500 MHz, $CDCl_3$) δ 0.87 (3H, t, J = 6.8 Hz, CH_3), 1.25–1.29 (26H, m, $13 \times CH_2$, 1.44–1.49 (3H, m, NHCH₂CH₂, H7s), 1.55 (1H, d, J = 8.7 Hz, H7a), 2.58 (1H, dd, J = 5.0, 1.7 Hz, H2), 3.10–3.26 (5H, m, NHCH₂, H1, H3, H4), 3.73 (3H, s, Me), 5.79 (1H, t, J = 5.5 Hz, NH), 6.17 (1H, dd, J = 5.5, 2.8 Hz, H6), 6.24 (1H, dd, J = 5.6, 3.5 Hz, H5). 13 C NMR (125 MHz, CDCl₃) δ 14.3, 22.8, 27.0, 29.4, 29.5, 29.69, 29.72, 29.79 (2 × C), 29.81 (2 × C), 29.83 $(3 \times C)$, 32.1, 39.7, 45.7, 46.9, 47.8, 48.2, 49.8, 52.3, 135.7, 136.6, 172.5, 175.7. HRMS (ESI, m/z) for $C_{26}H_{45}NO_3$ [M + H]⁺ calc. 420.3472; found 420.3483.

9-exo-[2'-(2",3"-Bis-tert-butoxycarbonylguanidino)ethylcarbamoyl]-8-endo-(hexadecylcarbamoyl)-4-heptyl-3,5-dioxatricyclo-[5.2.1.0^{2,6}]decane (24b). Compound 24b was prepared from acid 23b (20 mg, 0.04 mmol) and amine 11 (21 mg, 0.07 mmol) according to general procedure C and after purification by column chromatography (10–20% EtOAc in CH_2Cl_2) was isolated as a pale yellow oil (22 mg, 79%); $R_f = 0.32$ (20% EtOAc in CH_2Cl_2). ¹H NMR (500 MHz, $CDCl_3$) δ 0.85–0.88 (6H,

m, 2 × CH₃), 1.24–1.50 (57H, m, 19 × CH₂, 2 × t-Bu, H10s), 1.59–1.63 (2H, m, CH₂), 1.78 (1H, d, J = 9.6 Hz, H10a), 2.39 (1H, d, J = 5.6 Hz, H9), 2.52 (1H, d, J = 4.1 Hz, H1), 2.56 (1H, br s, H7), 2.94 (1H, dd, J = 5.8, 4.6 Hz, H8), 3.13–3.27 (2H, m, CH₂), 3.40 (2H, dt, J = 5.7, 5.3 Hz, CH₂), 3.51–3.64 (2H, m, CH₂), 3.98 (1H, d, J = 5.7 Hz, H2), 4.19 (1H, d, J = 5.7 Hz, H6), 4.63 (1H, t, J = 4.9 Hz, H4), 6.19 (1H, t, J = 5.6 Hz, NH), 7.06 (1H, t, J = 4.9 Hz, NH), 8.54 (1H, t, J = 5.7 Hz, NH), 11.49 (1H, s, NH). ¹³C NMR (125 MHz, CDCl₃) δ 14.2, 14.3, 22.75, 22.83, 24.4, 27.1, 28.2, 28.4, 29.2, 29.3, 29.4, 29.5, 29.67, 29.68, 29.73, 29.80, 29.83 (3 × C), 31.9 (2 × C), 32.1 (2 × C), 32.3, 33.0, 39.9, 40.0, 40.7, 43.3, 44.5, 44.9, 47.4, 78.7, 79.8, 81.6, 83.6, 104.1, 153.2, 157.3, 163.4, 171.7, 174.1. HRMS (ESI, m/z) for C₄₆H₈₃N₅O₈ [M + Na]⁺ calc. 856.6134; found 856.6134.

General procedure D: amidation of diacids

A microwave vial was charged with the appropriate carboxylic acid, EDCI (1.5 equiv.), HOBt (0.05 equiv.) and dry DMF and was stirred at ambient temperature for 30 min. The appropriate alkylamine (1.5 equiv.) was then added and the reaction was irradiated to 50 °C for 30 min. The resulting homogenous mixture was diluted with EtOAc (15 mL), washed with H₂O (3 × 8 mL), brine (8 mL), dried (MgSO₄), filtered, and concentrated *in vacuo* to afford a solid that was purified by column chromatography (as specified below) which gave the title compound.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylguanidino)ethylcarbamoyl]-4-heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (9a). Compound 9a was prepared from diacid 5a (156 mg, 0.48 mmol) and amine 11 (434 mg, 1.43 mmol) according to general procedure D and after purification by column chromatography (20-70% EtOAc in pet. spirits) was isolated as a white solid (274 mg, 64%); $R_f = 0.38$ (70% EtOAc in pet. spirits). m.p: 102.9-127.5 °C (slow decomposition). ¹H NMR (270 MHz, CDCl₃) δ 0.86 (3H, t, J = 6.7 Hz, CH₃), 1.25 (12H, br s, $6 \times CH_2$), 1.48–1.63 (37H, m, t-Bu, H10s), 1.72–1.83 (1H, m, H10a), 2.44 (1H, d, J = 5.9 Hz, H1), 2.57 (1H, br s, H7), 2.70 (1H, d, J = 3.8 Hz, H9), 2.94 (1H, app. t, J = 4.5 Hz, H8), 3.31-3.40 (4H, m, $2 \times CH_2$), 3.51-3.61 (4H, m, $2 \times CH_2$), 3.95(1H, d, J = 5.5 Hz, H2), 4.03 (1H, d, J = 5.5 Hz, H6), 4.60 (1H, t, H2)J = 4.8 Hz, H4), 6.87 (1H, t, J = 5.4 Hz, NH), 8.04 (1H, t, J = 4.1Hz, NH), 8.51 (1H, t, J = 5.3 Hz, NH), 8.64 (1H, t, J = 5.7 Hz, NH), 11.45 (1H, s, NH), 11.48 (1H, s, NH). ¹³C NMR (67.5 MHz, $CDCl_3$) δ 14.2, 22.8, 24.3, 28.2, 28.4, 29.3, 29.7, 31.9, 32.5, 33.0, 40.0, 40.1, 40.2, 42.3, 43.0, 44.3, 44.4, 47.8, 79.0, 79.6, 79.9, 81.7, 83.4, 83.7, 104.1, 153.2 (2 × C), 157.1, 157.9, 163.0, 163.5, 172.0, 174.3. HRMS (ESI, m/z) for $C_{43}H_{74}N_8O_{12}$ [M + H]⁺ calc. 895.5499; found 895.5511.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylguanidino)-ethylcarbamoyl]-4-undecanyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (9b). Compound 9b was prepared from diacid 5b (95 mg, 0.25 mmol) and amine 11 (230 mg, 0.78 mmol) according to general procedure D and after purification by column chromatography (50–70% EtOAc in pet. spirits) was isolated as a clear oil (116 mg, 49%); $R_{\rm f}$ = 0.50 (70% EtOAc in pet. spirits). ¹H NMR (500 MHz, CDCl₃) δ 0.87 (3H, t, J = 6.2 Hz, CH₃), 1.24–1.37 (22H, m, 11 × CH₂), 1.49–1.50 (37H, m, t-Bu, H10s),

1.57–1.62 (2H, m, CHC H_2), 1.77 (1H, d, J = 10.4 Hz, H10a), 2.45 (1H, d, J = 5.8 Hz, H1), 2.58 (1H, br s, H7), 2.69 (1H, d, J = 4.1 Hz, H9), 2.94 (1H, app. t, J = 5.2 Hz, H8), 3.34–3.44 (4H, m, $2 \times CH_2$, 3.55-3.59 (4H, m, $2 \times CH_2$), 3.95 (1H, d, J = 5.7 Hz, H2), 4.02 (1H, d, J = 5.7 Hz, H6), 4.60 (1H, t, J = 4.9 H4), 6.88(1H, br s, NH), 8.04 (1H, t, J = 4.0 Hz, NH), 8.55 (1H, br s, NH), 8.65 (1H, br s, NH), 11.45 (1H, s, NH), 11.48 (1H, s, NH). 13 C NMR (125 MHz, CDCl₃) δ 14.3, 22.8, 24.4, 28.18, 28.19, $28.4, 29.5 (2 \times C), 29.66 (3 \times C), 29.72, 29.75, 29.8, 32.0, 32.5,$ 33.0, $40.0 (2 \times C)$, 40.3, 42.2, 43.0, 44.2, 44.3, 47.8, 79.0, 79.8, 80.1, 81.6, 83.5, 83.8, 104.1, 153.17, 153.19, 157.0, 157.8, 162.7, 163.2, 172.0, 174.2. HRMS (ESI, m/z) for $C_{47}H_{82}N_8O_{12}$ [M + H] calc. 951.6125; found 951.6131.

8-endo-9-exo-Di[2'-(2",3"-bis-tert-butoxycarbonylguanidino)ethylcarbamoyl]-4-[4'-(octyloxy)phenethyl]-3,5-dioxatricyclo-[5.2.1.0^{2,6}]decane (9d). Compound 9d was prepared from diacid 5d (89 mg, 0.19 mmol) and amine 11 (180 mg, 0.57 mmol) according to general procedure D and after purification by column chromatography (50-70% EtOAc in pet. spirits) was isolated as a clear oil (81 mg, 41%); $R_f = 0.38$ (70%) EtOAc in pet. spirits). ¹H NMR (500 MHz, CDCl₃) δ 0.88 (3H, t, $J = 6.4 \text{ Hz}, \text{CH}_3$, 1.25–1.36 (8H, m, $4 \times \text{CH}_2$), 1.40–1.54 (39H, m, CH_2 , $4 \times t$ -Bu, H10s), 1.73–1.78 (2H, m, CH_2), 1.81 (1H, d, J = 10.2 Hz, H10a), 1.87-1.92 (2H, m, CH₂), 2.46 (1H, d, J =5.5 Hz, H1), 2.60 (1H, s, H7), 2.64-2.68 (2H, m, ArCH₂), 2.74 (1H, d, J = 4.2 Hz, H9), 2.96 (1H, app. t, J = 5.4 Hz, H8), 3.35-3.45 (4H, m, $2 \times CH_2$), 3.55-3.61 (4H, m, $2 \times CH_2$), 3.91(2H, t, J = 6.6 Hz, OCH₂), 3.98 (1H, d, J = 5.6 Hz, H6), 4.06 (1H, d)d, J = 5.7 Hz, H2, 4.64 (1H, t, J = 4.6 Hz, H4), 6.80 (2H, d, J =8.5 Hz, ArH), 6.90 (1H, br s, NH), 7.07 (2H, d, J = 8.5 Hz, ArH), 8.03 (1H, t, *J* = 4.2 Hz, NH), 8.53 (1H, br s, NH), 8.65 (1H, t, *J* = 4.1 Hz, NH), 11.47-11.48 (2H, m, 2 × NH). ¹³C NMR (125 MHz, $CDCl_3$) δ 14.2, 22.8, 26.2, 28.2, 28.4, 29.4, 29.5 (3 × C), 32.0, 32.5, 34.8, 40.1 (2 × C), 40.3, 42.2, 43.0, 44.3, 44.4, 47.8, 68.2, 79.1, 79.9, 80.1, 81.8, 83.5, 83.8, 103.3, 114.6 (2 × C), 129.3 $(2 \times C)$, 133.5, 153.2 $(2 \times C)$, 157.0, 157.5, 157.9, 162.9, 163.3, 172.0, 174.1. HRMS (ESI, m/z) for $C_{52}H_{84}N_8O_{13}$ [M + H]⁺ calc. 1029.6231; found 1029.6253.

9-exo-[2'-(2",3"-Bis-tert-butoxycarbonylguanidino)ethylcarbamoyl]-8-endo-(heptylcarbamoyl)-4-heptyl-3,5-dioxatricyclo-[5.2.1.0^{2,6}]decane (24a). Compound 24a was prepared from acid 23a (43 mg, 0.10 mmol) and amine 11 (50 mg, 0.17 mmol) according to general procedure D and after purification by column chromatography (10-50% EtOAc in CH₂Cl₂) was isolated as a clear oil (21 mg, 30%); $R_f = 0.48$ (50% EtOAc in CH_2Cl_2). ¹H NMR (500 MHz, $CDCl_3$) δ 0.85 (6H, m, 2 × CH_3), 1.25-1.50 (39H, m, $10 \times CH_2$, $2 \times t$ -Bu, H10s), 1.59-1.64 (2H, m, CH_2), 1.78–1.80 (1H, m, H10a), 2.39 (1H, d, J = 5.3 Hz, H9), 2.52 (1H, d, J = 4.1 Hz, H1), 2.56 (1H, br s, H7), 2.94 (1H, dd, J = 5.9, 4.6 Hz, H8), 3.15–3.26 (2H, m, CH₂), 3.40 (2H, dt, J =8.3, 5.5 Hz, CH_2), 3.50–3.62 (2H, m, CH_2), 3.98 (1H, d, J =5.6 Hz, H2), 4.19 (1H, d, J = 5.6 Hz, H6), 4.63 (1H, t, J = 4.9 Hz, H4), 6.17 (1H, t, J = 5.6 Hz, NH), 7.04 (1H, t, J = 4.9 Hz, NH), 8.53 (1H, t, J = 5.8 Hz, NH), 11.48 (1H, s, NH). ¹³C NMR (125 MHz, CDCl₃) δ 14.20, 14.21, 22.7, 22.8, 24.4, 27.0, 28.2, 28.4, 29.0, 29.3, 29.6, 29.7, 31.8, 31.9, 32.3, 34.0, 39.9, 40.0,

40.7, 43.3, 44.5, 44.9, 47.5, 78.8, 79.8, 81.6, 83.6, 104.2, 153.3, 157.3, 163.4, 171.7, 174.1. HRMS (ESI, m/z) for $C_{37}H_{65}N_5O_8$ $[M + H]^{+}$ calc. 708.4906; found 708.4921.

General procedure E: deprotection of Boc groups

To a stirring solution of Boc-protected amine/guanidine and MeOH was added dropwise AcCl (10.0 equiv.), and the reaction was stirred for 24 h at ambient temperature (in instances when ¹H NMR spectroscopy indicated the presence of Boc-groups the crude material was retreated using the aforementioned conditions). The reaction mixture was concentrated under vacuum and co-evaporated with MeOH (2 × 0.5 mL), to afford the title compound.

4-Undecanyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exodicarboxamidoethylamine hydrogen chloride (8b). Compound 8b was synthesised from Boc-protected amine 7b (459 mg, 0.69 mmol) according to general procedure E as a white solid (364 mg, 98%). m.p: 175.3-233.8 °C (slow decomposition). ¹H NMR (500 MHz, DMSO- d_6) δ 0.85 (3H, t, J = 6.6 Hz, CH₃), 1.23-1.30 (19H, m, 9 × CH₂, H10s), 1.49-1.53 (3H, m, CHCH₂, H10a), 2.41 (1H, br s, H1), 2.50 (1H, m, H7), 2.61 (1H, d, J =4.4 Hz, H9), 2.81–2.88 (4H, m, $2 \times CH_2$), 3.10 (1H, app. t, J =4.9 Hz, H8), 3.21–3.36 (4H, m, $2 \times CH_2$), 3.85 (1H, d, J = 5.6 Hz, H6), 3.91 (1H, d, J = 5.6 Hz, H2), 4.59 (1H, t, J = 4.7 Hz, H4), 8.01 (6H, br s, $2 \times NH_3$), 8.24 (1H, t, J = 5.4 Hz, NH), 8.35 (1H, t, J = 5.1 Hz, NH). ¹³C NMR (125 MHz, DMSO- d_6) δ 14.0, 22.2, $23.8, 28.8, 28.99 (3 \times C), 29.04, 29.1, 31.1, 31.3, 32.4, 36.7, 36.8,$ 38.48, 38.52, 42.8, 43.2, 44.5, 46.1, 78.3, 81.1, 103.0, 171.6, 173.5. HRMS (ESI, m/z) for $C_{25}H_{46}N_4O_4$ [M + 2H]²⁺ calc. 234.1823; found 234.1826.

4-Pentadecyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exodicarboxamidoethylamine hydrogen chloride (8c). Compound 8c was synthesised from Boc-protected amine 7c (76 mg, 0.11 mmol) according to general procedure E as a white powder (58 mg, 94%). m.p: 156.3-200.1 °C (slow decomposition). ¹H NMR (500 MHz, DMSO- d_6) δ 0.85 (3H, t, J = 7.1 Hz, CH_3), 1.23-1.31 (27H, m, 13 × CH_2 , H10s), 1.49-1.53 (3H, m, CHCH₂, H10a), 2.41 (1H, br s, H9), 2.49–2.51 (1H, m, H7), 2.60 $(1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 2.81-2.86 (4H, m, 2 \times CH₂), 3.10 (1H, d, J = 4.4 Hz, H1), 3.10$ app. t, J = 4.9 Hz, H8), 3.19–3.32 (4H, m, $2 \times CH_2$), 3.86 (1H, d, J = 5.6 Hz, H6), 3.91 (1H, d, J = 7.8 Hz, H2), 4.59 (1H, t, J = 4.7Hz, H4), 7.96 (6H, br s, $2 \times NH_3$), 8.23 (1H, t, J = 5.4 Hz, NH), 8.33 (1H, t, J = 5.5 Hz, NH). ¹³C NMR (125 MHz, DMSO- d_6) δ 14.0, 22.1, 23.8, 28.7, 28.99 (3 × C), 29.04 (3 × C), 29.07 $(3 \times C)$, 31.1, 31.3, 32.4, 36.7, 36.8, 38.4, 38.5, 42.8, 43.1, 44.5, 46.1, 78.2, 81.0, 102.9, 171.5, 173.5. HRMS (ESI, m/z) for $C_{29}H_{54}N_4O_4 [M + 2H]^{2+}$ calc. 262.2145; found 262.2150.

4-[4'-(Octyloxy)phenethyl]-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exo-dicarboxamidoethylamine hydrogen chloride (8d). Compound 8d was synthesised from Boc-protected amine 7d (28 mg, 0.04 mmol) according to general procedure E as a white solid (23 mg, 99%). m.p: 159.7-197.8 °C (slow decomposition). ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J =3.8 Hz, CH_3), 1.29–1.34 (8H, m, 4 × CH_2) 1.47–1.49 (3H, m, CH₂, H10s), 1.73-1.79 (3H, m, CH₂, H10a), 1.87-1.88 (2H, m, CH₂), 2.53 (1H, br s, H1), 2.64–2.67 (4H, m, ArCH₂, H9, H7),

3.06–3.09 (4H, m, 2 × CH₂), 3.24 (1H, app. t, J = 5.0 Hz, H8), 3.37–3.40 (2H, m, CH₂), 3.50–3.61 (2H, m, CH₂), 3.90–3.93 (2H, m, OCH₂), 4.02–4.03 (1H, m, H6), 4.06 (1H, m, H2), 4.67 (1H, t, J = 4.4 Hz, H4), 6.79–6.81 (2H, m, ArH), 7.05–7.08 (2H, m, ArH). ¹³C NMR (125 MHz, CD₃OD) δ 14.4, 23.7, 27.2, 30.4, 30.5 (2 × C), 32.8, 33.0, 36.0, 38.5 (2 × C), 40.9, 45.0, 45.1, 47.1, 47.3, 69.0, 80.1, 83.0, 104.3, 115.5 (2 × C), 130.2 (2 × C), 134.6, 158.9, 175.1, 177.0. HRMS (ESI, m/z) for C₃₀H₄₈N₄O₅ [M + 2H]²⁺ calc. 273.1855; found 273.1893.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exodicarboxamidoethylguanidine hydrogen chloride (10a). Compound 10a was synthesised from Boc-protected guanidine 9a (156 mg, 0.17 mmol) according to general procedure E as a white solid (94 mg, 95%). ¹H NMR (270 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.5 Hz, CH₃), 1.29 (10H, m, 5 × CH₂), 1.45–1.61 (3H, m, CHCH₂, H10s), 1.74 (1H, d, J = 9.6 Hz, H10a), 2.45 (1H, br s, H9), 2.61–2.63 (2H, m, H1, H7), 3.22 (1H, app. t, J = 4.9 Hz, H8), 3.30–3.39 (8H, m, 4 × CH₂), 4.00 (1H, d, J = 5.6 Hz, H6), 4.05 (1H, d, J = 5.5 Hz, H2), 4.66 (1H, t, J = 4.7 Hz, H4), 7.43–7.46 (1H, m, NH). ¹³C NMR (67.5 MHz, CD₃OD) δ 14.4, 23.7, 25.2, 28.2, 30.3, 30.6, 32.7, 32.9, 33.9, 39.6, 39.7, 41.9, 44.9, 45.1, 46.9, 47.7, 80.0, 82.9, 105.1, 153.6, 158.9, 174.7, 176.6. HRMS (ESI, m/z) for C₂₃H₄₂N₈O₄ [M + 2H]²⁺ calc. 248.1737; found 248.1744.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exodicarboxamidoethylguanidine hydrogen 2,2,2-trifluoroacetate (12). To the stirring solution of Boc-protected guanidine 9a (112 mg, 0.125 mmol) in CH₂Cl₂ (2.5 mL) was added trifluoroacetic acid (700 µL, 9.1 mmol) and the reaction was stirred at ambient temperature for 24 h. The solvent was removed under reduced pressure and the sample was co-evaporated with CHCl₃ (2 × 1 mL) and concentrated in vacuo to give the title compound (54 mg, 60%) as a yellow resin. ¹H NMR (500 MHz, DMSO- d_6) δ 0.85 (3H, t, J = 6.8 Hz, CH₃), 1.23–1.31 $(11H, m, 5 \times CH_2, H10s), 1.49-1.54 (3H, m, CHCH_2, H10a),$ 2.34 (1H, br s, H9), 2.53 (1H, d, J = 4.4 Hz, H7), 3.04-3.28 (10H, m, $4 \times CH_2$, H1, H8), 3.88 (1H, d, J = 5.6 Hz, H6), 3.90 (1H, d, J = 5.6 Hz, H2), 4.59 (1H, t, J = 4.8 H4), 7.21 (6H, br s, NH), 7.56-7.59 (2H, m, NH), 8.13 (1H, t, J = 5.7 Hz, NH), 8.20 (1H, t, J = 5.1 Hz, NH). ¹³C NMR (125 MHz, DMSO- d_6) δ 13.6, 21.7, 23.4, 28.3, 28.6, 30.7, 32.0, 37.7, 37.8, 40.0, 40.1, 40.2, 42.4, 42.9, 44.0, 46.0, 77.8, 78.8, 102.6, 156.6 (2 × C), 171.3, 173.1. HRMS (ESI, m/z) for $C_{23}H_{42}N_8O_4$ [M + 2H]²⁺ calc. 248.1737; found 248.1744.

4-Undecanyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-*endo*-9-*exo*-dicarboxamidoethylguanidine hydrogen chloride (10b). Compound 10b was synthesised from Boc-protected guanidine 9b (93 mg, 0.10 mmol) according to general procedure E as a colourless residue (52 mg, 85%). ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.8 Hz, CH₃), 1.29–1.40 (18H, m, 9 × CH₂), 1.47 (1H, d, J = 10.4 Hz, H10s), 1.57–1.61 (2H, m, CHC H_2), 1.73 (1H, d, J = 10.0 Hz, H10a), 2.46 (1H, br s, H9), 2.62–2.63 (2H, m, H1, H7), 3.22 (1H, app. t, J = 4.9 Hz, H8), 3.30–3.39 (8H, m, 4 × CH₂), 4.00 (1H, d, J = 5.5 Hz, H6), 4.05 (1H, d, J = 5.6 Hz, H2), 4.66 (1H, t, J = 4.7 Hz, H4). ¹³C NMR (125 MHz, CD₃OD) δ 14.5, 23.7, 25.2, 30.5 (2 × C), 30.66 (2 × C), 30.74, 30.8, 32.7,

33.1, 33.9, 39.6, 39.7, 41.9, 42.0, 44.9, 45.1, 46.9, 47.7, 79.9, 82.8, 105.1, 158.8, 158.9, 174.6, 176.5. HRMS (ESI, m/z) for $C_{27}H_{50}N_8O_4\left[M+2H\right]^{2+}$ calc. 276.2050; found 276.2057.

4-Pentadecyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exodicarboxamidoethylguanidine hydrogen chloride (10c). Compound 10c was synthesised from Boc-protected guanidine 9c (58 mg, 0.06 mmol) according to general procedure E as an off-white solid (34 mg, 87%). ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.9 Hz, CH₃), 1.29–1.45 (26H, m, 13 × CH₂), 1.47 (1H, d, J = 10.4 Hz, H10s), 1.58–1.62 (2H, m, CHC H_2), 1.74 (1H, d, J = 9.8 Hz, H10a), 2.45 (1H, br s, H1), 2.61-2.62 (2H, m, H10a)H7, H9), 3.22 (1H, app. t, J = 4.7 Hz, H8), 3.30-3.39 (8H, m, $4 \times CH_2$, 3.99 (1H, d, J = 5.6 Hz, H2), 4.04 (1H, d, J = 5.6 Hz, H6), 4.66 (1H, t, I = 4.8 Hz, H4). ¹³C NMR (125 MHz, CD₃OD) δ 14.4, 23.7, 25.2, 28.2, 30.5, 30.6, 30.70 (2 × C), 30.73, 30.74 $(2 \times C)$, 30.8 $(2 \times C)$, 32.7, 33.1, 33.9, 39.6, 39.7, 41.9, 42.0, 44.9, 45.1, 46.9, 47.7, 80.0, 82.9, 105.1, 158.9 (2 × C), 174.6, 176.6. HRMS (ESI, m/z) for $C_{31}H_{58}N_8O_4$ [M + 2H]²⁺ calc. 304.2363; found 304.2374.

4-[4'-(Octyloxy)phenethyl]-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exo-dicarboxamidoethylguanidine hydrogen chloride (10d). Compound 10d was synthesised from Boc-protected guanidine 9d (78 mg, 0.08 mmol) according to general procedure E as a yellow oil (39 mg, 74%). ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.9 Hz, CH₃), 1.28-1.35 (8H, m, 4 × CH₂) 1.44-1.53 (3H, m, CH₂, H10s), 1.71-1.78 (3H, m, CH₂, H10a), 1.84-1.88 (2H, m, CH₂), 2.49 (1H, br s, H1), 2.62-2.65 (4H, m, ArCH₂, H9, H7), 3.23 (1H, app. t, J = 5.0 Hz, H8), 3.30-3.39 (8H, m, $4 \times CH_2$), 3.92 (2H, t, J = 6.5 Hz, OCH_2), 4.02(1H, d, J = 5.5 Hz, H6), 4.07 (1H, d, J = 5.6 Hz, H2), 4.67 (1H, t, H2)J = 4.4 Hz, H4), 6.79-6.81 (2H, m, ArH), 7.06-7.09 (2H, m, ArH). ¹³C NMR (125 MHz, CD₃OD) δ 14.4, 23.7, 27.2, 30.4, 30.46, 30.49, 30.7, 32.8, 33.0, 36.0, 39.6, 39.7, 41.9, 42.0, 44.9, 45.1, 47.0, 47.6, 69.0, 80.1, 82.9, 104.3, 115.5 (2 × C), 130.2 $(2 \times C)$, 134.6, 158.9 $(3 \times C)$, 174.6, 176.5. HRMS (ESI, m/z) for $C_{32}H_{52}N_8O_5 [M + 2H]^{2+}$ calc. 315.2103; found 315.2109.

4-Ethylbenzene-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9exo-dicarboxamidoethylguanidine hydrogen chloride (10e). Compound 10e was synthesised from Boc-protected guanidine 9e (242 mg, 0.27 mmol) according to general procedure E as a white solid (145 mg, 93%). m.p: 133.1-149.1 °C (slow decomposition). ¹H NMR (500 MHz, CD₃OD) δ 1.50 (1H, d, J = 10.5 Hz, H10s), 1.78 (1H, d, J = 10.5 Hz, H10a), 1.88-1.92 (2H, m, CHCH₂), 2.49 (1H, br s, H9), 2.62-2.66 (2H, m, H1, H7), 2.69-2.72 (2H, m, CH_2), 3.23 (1H, app. t, J = 5.0 Hz, H8), 3.29-3.40 (8H, m, $4 \times CH_2$), 4.02 (1H, d, J = 5.6 Hz, H6), 4.08(1H, d, J = 5.6 Hz, H2), 4.68 (1H, t, J = 4.8 Hz, H4), 7.14-7.18(3H, m, ArH), 7.23-7.27 (2H, m, ArH). ¹³C NMR (125 MHz, CD₃OD) δ 31.3, 32.8, 35.8, 39.6, 39.7, 41.9, 42.0, 44.9, 45.1, 47.0, 47.7, 80.1, 83.0, 101.3, 127.0, 129.3 (2 × C), 129.5 (2 × C), 142.7, 158.9 (2 × C), 174.6, 176.5. HRMS (ESI, m/z) for $C_{24}H_{36}N_8O_4 [M + 2H]^{2+}$ calc. 251.1503; found 251.1510.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-*endo*-9-*exo*-dicarboxamidopropylguanidine hydrogen chloride (15). Compound 15 was synthesised from Boc-protected guanidine 13 (139 mg, 0.15 mmol) according to general procedure E as a

white solid (66 mg, 74%). m.p: 142.4-146.4 °C. ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.8 Hz, CH₃), 1.29–1.42 $(10H, m, 5 \times CH_2)$, 1.48 (1H, d, J = 10.4 Hz, H10s), 1.58-1.62 (2H, m, CHCH₂), 1.71-1.79 (5H, m, 2 × CH₂, H10a), 2.42 (1H, br s, H9), 2.59-2.60 (2H, m, H1, H7), 3.18-3.29 (9H, m, $4 \times CH_2$, H8), 3.99 (1H, d, I = 5.6 Hz, H6), 4.05 (1H, d, I = 5.6Hz, H2), 4.66 (1H, t, J = 4.8 Hz, H4). ¹³C NMR (125 MHz, CD_3OD) δ 14.4, 23.7, 25.2, 29.90, 29.93, 30.3, 30.6, 32.7, 32.9, 33.9, 37.69, 37.71, 39.99, 40.05, 44.9, 45.2, 46.9, 47.9, 80.0, 82.9, 105.1, 158.7 (2 × C), 174.1, 176.1. HRMS (ESI, m/z) for $C_{25}H_{46}N_8O_4 [M + 2H]^{2+}$ calc. 262.1894; found 262.1892.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exodicarboxamidobutylguanidine hydrogen chloride (16). Compound 16 was synthesised from Boc-protected guanidine 14 (78 mg, 0.08 mmol) according to general procedure E as a white solid (48 mg, 94%). m.p: 110.5-124.0 °C. ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.8 Hz, CH₃), 1.29–1.40 (10H, m, $5 \times CH_2$), 1.48 (1H, d, J = 10.3 Hz, H10s), 1.57-1.60 (10H, m, $5 \times CH_2$), 1.71 (1H, d, J = 10.2 Hz, H10a), 2.39 (1H, br s, H9), 2.58-2.59 (2H, m, H1, H7), 3.15-3.27 (9H, m, 4 × CH₂, H8), 3.99 (1H, d, J = 5.6 Hz, H6), 4.05 (1H, d, J = 5.6 Hz, H2), 4.65 (1H, t, J = 4.8 Hz, H4). ¹³C NMR (125 MHz, CD₃OD) δ 14.4, 23.7, 25.2, 27.17, 27.23, 27.7 (2 × C), 30.3, 30.6, 32.7, 32.9, 33.9, 39.9 (2 × C), 42.1 (2 × C), 44.9, 45.3, 46.9, 48.0, 80.0, 82.9, 105.1, 158.6 (2 \times C), 173.9, 175.9. HRMS (ESI, m/z) for $C_{27}H_{50}N_8O_4 [M + 2H]^{2+}$ calc. 276.2050; found 276.2050.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-(heptylcarbamoyl)-9-exo-carboxamidoethylguanidine hydrogen chloride (25a). Compound 25a was synthesised from Boc-protected guanidine 24a (16 mg, 0.02 mmol) according to general procedure E as a colourless oil (10 mg, 99%). ¹H NMR (500 MHz, CD₃OD) δ 0.88-0.91 (6H, m, 2 × CH₃), 1.29-1.40 (18H, m, $9 \times CH_2$, 1.47-1.52 (3H, m, CH₂, H10s), 1.57-1.61 (2H, m, CH_2), 1.72 (1H, d, J = 10.4 Hz, H10a), 2.42 (1H, br s, H9), 2.58-2.60 (2H, m, H1, H7), 3.07-3.13 (2H, m, CH₂), 3.17 (1H, app. t, J = 5.0 Hz, H8), 3.23-3.34 (4H, m, $2 \times \text{CH}_2$), 3.98 (1H, d, J = 5.6 Hz, H6), 4.04 (1H, d, J = 5.7 Hz, H2), 4.65 (1H, t, J =4.8 Hz, H4). 13 C NMR (125 MHz, CD₃OD) δ 14.41, 14.44, 23.6, 23.7, 25.2, 27.9, 30.1, 30.3, 30.4, 30.6, 32.7, 32.91, 33.0, 33.9, 39.7, 40.5, 42.0, 44.8, 45.3, 46.9, 47.8, 80.0, 82.9, 105.1, 158.9, 173.5, 176.8. HRMS (ESI, m/z) for $C_{27}H_{49}N_5O_4$ [M + H]⁺ calc. 508.3857; found 508.3861.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-(hexadecylcarbamoyl)-9-exo-carboxamidoethylguanidine hydrogen chloride (25b). Compound 25b was synthesised from Boc-protected guanidine 24b (21 mg, 0.03 mmol) according to general procedure E as an orange oil (16 mg, 80%). ¹H NMR (500 MHz, CD₃OD) δ 0.90 (6H, t, J = 6.5 Hz, $2 \times \text{CH}_3$), 1.29–1.42 (36H, m, $18 \times CH_2$, 1.47-1.51 (3H, m, CH_2 , H10s), 1.57-1.61 (2H, m, CH_2), 1.72 (1H, d, J = 10.2 Hz, H10a), 2.42 (1H, br s, H9), 2.59-2.61 (2H, m, H1, H7), 3.04-3.10 (2H, m, CH₂), 3.17 (1H, app. t, J = 5.1 Hz, H8), 3.26–3.35 (4H, m, $2 \times \text{CH}_2$), 3.99 (1H, d, J = 5.6 Hz, H6), 4.04 (1H, d, J = 5.6 Hz, H2), 4.65 (1H, t, J =4.8 Hz, H4). 13 C NMR (125 MHz, CD₃OD) δ 14.4, 14.5, 23.69, 23.73, 25.3, 28.0, 30.1, 30.3, 30.35, 30.39, 30.5, 30.6, 30.7 $(2 \times C)$, 30.76, 30.81 $(4 \times C)$, 32.7, 32.9, 33.1 $(2 \times C)$, 33.9

 $(2 \times C)$, 39.7, 40.5, 42.0, 44.8, 45.3, 46.9, 47.8, 80.0, 82.9, 105.1, 158.9, 173.5, 176.8. HRMS (ESI, m/z) for $C_{36}H_{68}N_5O_4$ [M + H]⁺ calc. 634.5266; found 634.5283.

8-endo-9-exo-Di[2'-(2"-bromoacetamide)ethylcarbamoyl]-4heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (27). To the stirring solution of diamine 8a (210 mg, 0.434 mmol), Et₃N (320 μL, 2.26 mmol) and CH₂Cl₂ (4.3 mL) at -78 °C under an inert atmosphere, was added bromoacetyl bromide (110 µL, 1.22 mmol) dropwise. The reaction was stirred for 5 h, before being warmed to -20 °C, quenched with H2O (5 mL) and diluted with CH₂Cl₂ (20 mL). The combined organic phase was washed with H₂O (10 mL), sat. NH₄Cl (10 mL), sat. NaHCO₃ (10 mL) and brine (10 mL), dried (MgSO₄), filtered and concentrated in vacuo to afford a white solid that was purified by column chromatography (EtOAc-10% MeOH in CH2Cl2) to give the title compound (185 mg, 65%) as a white powder; $R_f =$ 0.36 (10% MeOH in CH₂Cl₂). m.p: 65.6-70.4 °C. ¹H NMR (500 MHz, DMSO- d_6) δ 0.85 (3H, t, J = 6.7 Hz, CH₃), 1.23-1.31 $(11H, m, 5 \times CH_2, H10s), 1.50-1.52 (3H, m, CHCH_2, H10a),$ 2.32 (1H, br s, H7), 2.46 (1H, d, J = 4.6 Hz, H1), 2.49-2.50 (1H, m, H9), 3.01-3.19 (9H, m, $4 \times CH_2$, H8), 3.83 (2H, s, BrCH₂), 3.84 (2H, s, BrCH₂), 3.86 (1H, d, J = 5.7 Hz, H2), 3.90 (1H, d, J = 5.7 Hz, H2)5.6 Hz, H6), 4.59 (1H, t, J = 4.7 Hz, H4), 8.00 (1H, t, J = 5.4 Hz, NH), 8.04 (1H, t, J = 5.0 Hz, NH), 8.29–8.30 (2H, m, 2 × NH). 13 C NMR (125 MHz, DMSO- d_6) δ 14.0, 22.1, 23.7, 28.6, 28.9, 29.5 $(2 \times C)$, 31.1, 31.2, 32.3, 38.1 $(2 \times C)$, 38.3 $(2 \times C)$, 42.7, 43.2, 44.5, 46.2, 78.2, 81.1, 102.9, 166.1, 166.2, 171.2, 173.1. HRMS (ESI, m/z) for $C_{25}H_{40}N_4O_6$ [M + H]⁺ calc. 651.1387; found 651.1406.

8-endo-9-exo-Di[2'-([2"-benzylaminocyclobut-1"-ene-3",4"dione]amino)ethylcarbamoyl]-4-heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (26a). Treatment of salt 8a (99 mg, 0.204 mmol) in MeOH (730 μ L) with Et₃N (85 μ L, 0.612 mmol) was followed by addition of squaric ester S2 (90 mg, 0.408 mmol) and the reaction was stirred for 2.5 h at ambient temperature. The white slurry was concentrated under vacuum before being triturated in 2M HCl (3 mL) for 40 min at ambient temperature. The white solid was collected by vacuum filtration before being diluted with MeOH (600 µL), treated with 1,2-ethylenediamine (20 µL) and stirred at ambient temperature for a further 2 h. Again the resulting slurry was triturated in 2M HCl (3 mL) for 20 min at ambient temperature and the off white solid was collected and dried in vacuo to afford the title compound (63 mg, 40%). m.p: 166.4-204.9 °C (slow decomposition). ¹H NMR (500 MHz, DMSO- d_6) δ 0.84 (3H, t, J = 6.7 Hz, CH₃), 1.22–1.35 (11H, m, $5 \times CH_2$, H10s), 1.48-1.49 (3H, CHC H_2 , H10a), 2.29 (1H, br s, H1), 2.46 (1H, d, J = 5.0 Hz, H7), 2.48–2.50 (1H, m, H9), 3.06 (1H, app. t, J = 3.8 Hz, H8), 3.10-3.27 (8H, m, $4 \times NHCH_2$), 3.83 (1H, d, J = 5.6 Hz, H6), 3.87 (1H, d, J =5.6 Hz, H2), 4.54 (1H, t, J = 4.6 Hz, H4), 4.70 (4H, br s, $2 \times ArCH_2$, 7.27–7.38 (10H, m, ArH), 7.49 (2H, br s, $2 \times NH$), 7.89 (2H, br s, $2 \times NH$), 8.11 (1H, t, J = 5.0 Hz, NH), 8.16 (1H, br s, NH). ¹³C NMR (125 MHz, DMSO- d_6) δ 13.9, 22.1, 23.7, 28.6, 28.9, 31.1, 31.2, 32.4, 39.52 (4 × C), 43.0, 43.1, 44.5, 46.2, $46.8 (2 \times C)$, 78.2, 81.0, 102.9, $127.4 (2 \times C)$, $127.5 (4 \times C)$, 128.6 $(4 \times C)$, 139.0 $(2 \times C)$, 167.5 $(2 \times C)$, 168.0 $(2 \times C)$, 171.3, 173.2,

182.5 (4 × C). HRMS (ESI, m/z) for $C_{43}H_{52}N_6O_8$ [M + H]⁺ calc. 781.3919; found 781.3904.

8-endo-9-exo-Di[2'-(phenylthioureido)ethylcarbamoyl]-4heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane (26b). Treatment of salt 8a (52 mg, 0.108 mmol) in CH_2Cl_2 (500 $\mu L) with <math display="inline">Et_3N$ (50 µL, 0.109 mmol) was followed by addition of phenylisothiocyanate (13 µL, 0.109 mmol) and the reaction was stirred for 22 h at ambient temperature. The reaction was diluted with H_2O (10 mL) and extracted with CH_2Cl_2 (2 × 10 mL). The combined organic phase was washed with 2M HCl (10 mL), brine (10 mL), dried (MgSO₄), filtered, and concentrated in vacuo to give a white waxy solid which was purified by column chromatography (50% EtOAc in pet. spirits-EtOAc) to afford the title compound (35 mg, 95%) as a white paste; $R_f = 0.37$ (EtOAc). ¹H NMR (500 MHz, CDCl₃) δ 0.87 (3H, t, J = 6.8 Hz, CH₃), 1.25-1.42 (11H, m, $5 \times \text{CH}_2$, H10s), 1.59-1.63 (2H, m, CHC H_2) 1.75 (1H, d, J = 9.9 Hz, H10a), 2.35 (1H, d, J = 5.8 Hz, H7), 2.52 (1H, br s, H1), 2.59 (1H, d, J = 3.9 Hz, H9), 2.85 (1H, app. t, $J_{\text{app}} = 4.8 \text{ Hz}, \text{H8}, 3.26-3.28 (2H, m, NHC}_2) 3.45-3.51 (2H, m, MHC}_2)$ $NHCH_2$), 3.68-3.87 (4H, m, 2 × $NHCH_2$), 3.89 (1H, d, J = 5.7 Hz, H6), 3.99 (1H, d, J = 5.6 Hz, H2), 4.60 (1H, t, J = 4.9 Hz, H4), 6.68 (2H, br s, $2 \times NH$), 6.81 (1H, t, J = 5.6 Hz, NH), 7.06 (1H, t, J = 5.3 Hz, NH, 7.21-7.31 (6H, m, ArH), 7.41 (4H, t, <math>J = 7.7 Hz,ArH), 8.06 (1H, br s, NH), 8.12 (1H, br s, NH). 13C NMR (125 MHz, CDCl₃) δ 14.2, 22.8, 24.4, 29.3, 29.7, 31.9, 32.6, 33.0, 39.4, 40.4, 43.4, 44.3, 44.4, 44.9, 45.6, 48.0, 78.7, 81.5, 104.2, 125.5 (2 \times C), 125.6 (2 \times C), 127.5 (2 \times C), 130.3 (4 \times C), 136.2 $(2 \times C)$, 172.9 $(2 \times C)$, 174.8 $(2 \times C)$. HRMS (ESI, m/z) for $C_{35}H_{48}N_6O_4S_2[M+H]^+$ calc. 681.3251; found 681.3251.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-endo-9-exo-di-[2'-(ethylcarbamoyl)amidomethyl]di-1-methyl-1H-imidazol-3ium bromide (26c). To the stirring solution of 1-methylimidazole (16 µL, 0.19 mmol) in anhydrous THF (500 µL) at -78 °C under an inert atmosphere, was added alpha bromoamide 27 (63 mg, 0.10 mmol) in THF (500 µL) slowly. The reaction was warmed to -15 °C and stirred for 1 h, before being warmed to ambient temperature and stirred for a further 2 d. The THF was decanted off and the material was washed and decanted with Et₂O (3 × 1 mL). The material was diluted with H₂O (5 mL) and extracted with CH₂Cl₂ (4 mL). The aqueous portion was lyophilised for 48 h to give the title compound (59 mg, 74%) as light yellow residue. ¹H NMR (500 MHz, DMSO- d_6) δ 0.85 (3H, t, J = 6.8 Hz, CH₃), 1.23–1.29 (11H, m, 5 × CH₂, H10s), 1.49-1.51 (3H, m, CHCH₂, H10a), 2.32 (1H, br s, H7), 2.48-2.55 (2H, m, H1, H9), 3.01-3.23 (9H, m, 4 × CH₂, H8), 3.85 (1H, d, J = 5.6 Hz, H2), 3.89 (6H, s, $2 \times NCH_3$), 3.92 (1H, d, J = 5.5 Hz, H6), 4.60 (1H, t, J = 4.7 Hz, H4), 4.97 (4H, s, H4) $2 \times CH_2N$), 7.68-7.71 (4H, m, $4 \times ArH$), 8.08-8.12 (2H, m, 2 × NH), 8.48-8.51 (2H, m, 2 × NH), 9.08 (2H, s, NCHN). ¹³C NMR (125 MHz, DMSO- d_6) δ 14.0, 22.1, 23.8, 28.7, 28.9, 31.1, 31.2, 32.4, 35.9 ($2 \times C$), 38.2, 38.4, 38.6, 38.7, 42.7, 43.4, 44.4, 46.2, 50.5, 50.6, 78.2, 81.1, 102.9, 123.1 (2 × C), 123.8 $(2 \times C)$, 137.7 $(2 \times C)$, 165.1 $(2 \times C)$, 171.2, 173.1. HRMS (ESI, m/z) for C₃₃H₅₀N₈O₆ [M + 2H]²⁺ calc. 328.1999; found 328.1996.

4-Heptyl-3,5-dioxatricyclo[5.2.1.0^{2,6}]decane-8-*endo*-9-*exo*-di[2'-(ethylcarbamoyl)amidomethyl]dipyridin-1-ium bromide

(26d). To the stirring solution of alpha bromoamide 27 (31 mg, 0.05 mmol) in anhydrous THF (480 μL), under inert conditions was added pyridine (100 µL, 1.24 mmol) and the reaction was stirred at 66 °C for 16 h. The reaction was concentrated under vacuum and the crude material was rinsed and decanted with Et₂O (5 × 1 mL). Excess solvent was removed under vacuum before the sample was diluted in H₂O (5 mL) and extracted with CH_2Cl_2 (3 × 4 mL). The aqueous portion was lyophilised for 48 h to give the title compound (25 mg, 64%) as an orange spongy material. ¹H NMR (500 MHz, CD₃OD) δ 0.90 (3H, t, J = 6.9 Hz, CH₃), 1.29–1.45 (11H, m, 5 × CH_2 , H10s), 1.59–1.63 (2H, m, $CHCH_2$) 1.70 (1H, dd, J = 10.3, 1.2 Hz, H10a), 2.44 (1H, br s, H1), 2.58-2.59 (2H, m, H9, H7), 3.18 (1H, app. t, I = 4.8 Hz, H8), 3.33–3.43 (8H, m, $4 \times \text{CH}_2$), 4.01 (1H, d, J = 5.6 Hz, H6), 4.04 (1H, d, J = 5.6 Hz, H2), 4.66(1H, t, J = 4.8 Hz, H4), 5.45 (2H, s, CH₂), 5.46 (2H, s, CH₂),8.14-8.17 (4H, m, ArH), 8.65-8.68 (2H, m, ArH), 8.93-8.94 (4H, m, ArH). 13 C NMR (125 MHz, CD₃OD) δ 14.4, 23.7, 25.3, 30.4, 30.6, 32.8, 32.9, 33.9, 39.9, 40.0, 40.6, 40.7, 45.1 ($2 \times C$), 47.2, $47.5, 63.0 (2 \times C), 78.0, 82.9, 105.1, 129.0 (4 \times C), 147.5 (2 \times C),$ 147.55 (2 × C), 147.60 (2 × C), 166.3 (2 × C), 174.3, 176.2. HRMS (ESI, m/z) for $C_{35}H_{48}N_6O_6$ [M + 2H]²⁺ calc. 325.1890; found 325.1898.

Crystallography

Intensity data were collected with an Oxford Diffraction Super-Nova CCD diffractometer using Cu-K α radiation, the temperature during data collection was maintained at 130.0(1) using an Oxford Cryosystems cooling device. The structure was solved by direct methods and difference Fourier synthesis. Thermal ellipsoid plots were generated using the program ORTEP-3²⁸ integrated within the WINGX²⁹ suite of programs. Disordered solvent, assumed to be ethanol was removed using the Squeeze procedure. ³⁰

Disk diffusion assay

A stock solution of 10 mg mL $^{-1}$ was made for each compound under observation using DMSO as a solvent. Each of these stock solutions was then diluted by a factor of 1:2 to bring the concentration to 5 mg mL $^{-1}$. The diluted solutions were then filter-sterilised using a 0.2 μ m nylon filter, and 10 μ L of the 5 mg mL $^{-1}$ stock was pipetted onto a blank disk (*i.e.* 50 μ g per disk; Oxoid Limited, Hampshire, UK). Suspensions of all bacterial isolates were adjusted to a 0.5 McFarland standard (in 0.9% NaCl) before they were swabbed onto nutrient agar plates. The controls used were a 10 μ g colistin disk (sulphate, Oxoid), 10 μ L of DMSO and a plate swabbed with saline from the dispenser used.

Minimum inhibitory concentration (MIC) determination

Bacteria were obtained either from American Type Culture Collection (ATCC; Manassas, VA, USA), Network on Antimicrobial Resistance in Staphylococcus aureus (NARSA) or from a clinical isolate library at The University of Queensland, Centre for Clinical Research (UQCCR) as listed in Table S1 (see ESI†). Bacteria were cultured in Nutrient broth (NB; Bacto Laboratories,

catalogue no. 234000) or Muller-Hinton broth (MHB; Bacto Laboratories, catalogue no. 211443) at 37 °C overnight with shaking (~180 RPM). A sample of each culture was diluted 50fold in fresh MHB and incubated at 37 °C for 1.5-3 h with shaking (~180 RPM). Compound stock solutions were prepared as 10 mg mL⁻¹ in DMSO and colistin was dissolved in milli-Q water at 5.12 mg mL⁻¹. The compounds, at twice the final desired concentration, were serially diluted 2-fold across the wells of 96-well plates (Non-Binding Surface, Corning, catalogue no. 3641). Mid-log phase bacterial cultures (after 1.5-3 h incubation) were diluted to a final concentration of 5×10^5 colony forming units (CFU) per mL, and 50 µL was added to each well giving a final compound concentration range of 32 $\mu g \text{ mL}^{-1}$ to 0.015 $\mu g \text{ mL}^{-1}$ (DMSO $\leq 1\%$). MICs were determined visually after 20 h of incubation at 37 °C, with the MIC defined as the lowest compound concentration at which no bacterial growth was visible. Determined MIC values are the result of two independent experiments of n = 2, giving a final dataset of n = 4.

Cytotoxicity evaluation

HEK293 (ATCC CRL-1573) and HepG2 (ATCC HB-8065) cells were seeded as 3000 cells per well in a 384-well plate in DMEM medium (GIBCO-Invitrogen #11995-073), in which 10% of FBS was added. Cells were incubated for 24 h at 37 °C, 5% CO2 to allow cells to attach to the plates. A concentration series of compounds was then added into each well. The cells were incubated with the compounds for 24 h at 37 °C, 5% CO₂. After the incubation, 10 µM resazurin (dissolved in PBS) was added to each well. The plates were then incubated for 2 h at 37 °C, 5% CO₂. The fluorescence intensity was read using Polarstar Omega with excitation/emission 560/590. The data was analysed by Prism software. Results are presented as the average percentage of control ± SD for each set of duplicate wells using the following equation:

$$\begin{aligned} \text{Percentage Viability} &= (\text{FITEST} - \text{FI}_{\text{Negative}} / \text{FI}_{\text{UNTREATED}} \\ &- \text{FI}_{\text{Negative}}) \times 100. \end{aligned}$$

Acknowledgements

F.M.P., S.M.H., S.K.K., R.N.R. & T.D.A. thank the ARC (DP140100227) and the Strategic Research Centre for Chemistry and Biotechnology (Deakin University) for financial support and a top-up scholarship for S.M.H. The authors would also like to thank Dr Damien Callahan for his assistance with the collection of HRMS and the Australian Research Council for funding Deakin University's Nuclear Magnetic Resonance Facility through LIEF grant LE110100141. J.L. & R.L.N. are supported by a research grant from the National Institute of Allergy and Infectious Diseases of the National Institutes of Health (R01 AI098771). J.L. is an Australian NHMRC Senior Research Fellow, while M.A.C. is an Australian NHMRC Principal Research Fellow. The content is solely the responsibility of the authors and does not necessarily represent the official

views of the National Institute of Allergy and Infectious Diseases or the National Institutes of Health. The MIC screening was done in collaboration with CO-ADD (Community for Open Antimicrobial Drug Discovery, co-add.org) and we thank David L. Paterson (UQCCR, University of Queensland) for his kind donation of clinical Gram-positive isolates used for testing.

References

- 1 (a) H. W. Boucher, G. H. Talbot, J. S. Bradley, J. E. Edwards, D. Gilbert, L. B. Rice, M. Scheld, B. Spellberg and J. Bartlett, Clin. Infect. Dis., 2009, 48, 1-12; (b) G. H. Talbot, J. Bradley, J. E. Edwards, D. Gilbert, M. Scheld and J. G. Bartlett, Clin. Infect. Dis., 2006, 42, 657-668; (c) W. Yau, R. J. Owen, A. Poudyal, J. M. Bell, J. D. Turnidge, H. H. Yu, R. L. Nation and J. Li, J. Infect., 2009, 58, 138-
- 2 (a) E. Breukink and B. de Kruijff, Nat. Rev. Drug Discovery, 2006, 5, 321-323; (b) G. Taubes, Science, 2008, 321, 356-
- 3 M. A. Cooper and D. Shlaes, Nature, 2011, 472, 32-32.
- 4 (a) J. Harder, R. Gläser and J. M. Schröder, J. Endotoxin Res., 2007, 13, 317; (b) L. L. Ling, T. Schneider, A. J. Peoples, A. L. Spoering, I. Engels, B. P. Conlon, A. Mueller, T. F. Schaberle, D. E. Hughes, S. Epstein, M. Jones, L. Lazarides, V. A. Steadman, D. R. Cohen, C. R. Felix, K. A. Fetterman, W. P. Millett, A. G. Nitti, A. M. Zullo, C. Chen and K. Lewis, *Nature*, 2015, 517, 455-459.
- 5 (a) M. S. Butler, M. A. Blaskovich and M. A. Cooper, J. Antibiot., 2013, 66, 571-591; (b) C. T. Walsh and T. A. Wencewicz, J. Antibiot., 2014, 67, 7-22; (c) R. E. W. Hancock, Nat. Biotechnol., 2014, 32, 66-68; (d) A. Ivankin, L. Livne, A. Mor, G. A. Caputo, W. F. DeGrado, M. Meron, B. Lin and D. Gidalevitz, Angew. Chem., 2010, 122, 8640-8643.
- 6 (a) Y. Shai, Biochim. Biophys. Acta, 1999, 1462, 55-70; (b) E. Guan-Guerra, T. Santos-Mendoza and S. Lugo-Reyes, Clin. Immunol., 2010, 135, 1-11; (c) G. Baumann and P. Mueller, J. Supramol. Struct., 1974, 2, 538-557; (d) D. R. Laver, J. Biophys., 1994, **66**, 355–359.
- 7 (a) L. S. McCoy, K. D. Roberts, R. L. Nation, P. E. Thompson, T. Velkov, J. Li and Y. Tor, ChemBioChem, 2013, 14, 2083-2086; (b) T. Velkov, P. E. Thompson, R. L. Nation and J. Li, J. Med. Chem., 2010, 53, 1898; (c) T. M. Arnold, G. N. Forrest and K. J. Messmer, Am. J. Health-Syst. Pharm., 2007, 64; (d) J. Li, R. L. Nation, R. W. Milne, J. D. Turnidge and K. Coulthard, Int. J. Antimicrob. Agents, 2005, 25, 11-25; (e) D. R. Storm, K. S. Rosenthal and P. E. Swanson, Annu. Rev. Biochem., 1977, 46, 723-763.
- 8 (a) A. J. Lowe, G. A. Dyson and F. M. Pfeffer, Org. Biomol. Chem., 2007, 5, 1343-1346; (b) A. J. Lowe, G. A. Dyson and F. M. Pfeffer, Eur. J. Org. Chem., 2008, 1559-1567;

- (c) A. J. Lowe, B. M. Long and F. M. Pfeffer, *Chem. Commun.*, 2013, **49**, 3376–3388.
- 9 L. C. Henderson, J. Li, R. L. Nation, T. Velkov and F. M. Pfeffer, *Chem. Commun.*, 2010, **46**, 3197–3199.
- 10 S. M. Hickey, T. D. Ashton, J. M. White, J. Li, R. L. Nation, H. Y. Yu, A. G. Elliott, M. S. Butler, J. X. Huang, M. A. Cooper and F. M. Pfeffer, RSC Adv., 2015, 5, 28582– 28596.
- 11 L. D. Van Vliet, T. Ellis, P. J. Foley, L. Liu, F. M. Pfeffer, R. A. Russell, R. N. Warrener, F. Hollfelder and M. J. Waring, J. Med. Chem., 2007, 50, 2326–2340.
- 12 G. Pandey, N. R. Gupta and T. M. Pimpalpalle, *Org. Lett.*, 2009, **11**, 2547–2550.
- 13 A 4-fluorophenyl acetal analogue was converted to the intended diBoc guanidine and the acetal was indeed unstable to the acidic conditions used during the final Bocdeprotection step (as evidenced by ¹H NMR spectroscopy, see ESI†).
- 14 (a) S. M. Hickey, T. D. Ashton, S. K. Khosa and F. M. Pfeffer, *Synlett*, 2012, 1779–1782; (b) S. M. Hickey, T. D. Ashton and F. M. Pfeffer, *Asian J. Org. Chem.*, 2015, 4, 320–326.
- 15 A. J. Lowe and F. M. Pfeffer, *Chem. Commun.*, 2008, 1871–1873.
- 16 (a) N. Hamanaka, T. Seko, T. Miyazaki, M. Naka, K. Furuta and H. Yamamoto, *Tetrahedron Lett.*, 1989, **30**, 2399–2402; (b) S. M. Hickey, S. K. Tripcony, R. Li, R. J. Williams and F. M. Pfeffer, *Supramol. Chem.*, 2015, **27**, 425–435.
- 17 D. Coleman, M. Spulak, M. T. Garcia and N. Gathergood, *Green Chem.*, 2012, **14**, 1350–1356.

- 18 R. Ian Storer, C. Aciro and L. H. Jones, *Chem. Soc. Rev.*, 2011, 40, 2330–2346.
- 19 (a) M. Mohamed, T. P. Gonçalves, R. J. Whitby, H. F. Sneddon and D. C. Harrowven, *Chem. Eur. J.*, 2011, 17, 13698–13705; (b) N. C. Lim, M. D. Morton, H. A. Jenkins and C. Brückner, *J. Org. Chem.*, 2003, 68, 9233–9241.
- L. B. Reller, M. Weinstein, J. H. Jorgensen and M. J. Ferraro, *Clin. Infect. Dis.*, 2009, 49, 1749–1755.
- 21 D. J. Payne, M. N. Gwynn, D. J. Holmes and D. L. Pompliano, *Nat. Rev. Drug Discovery*, 2007, **6**, 29–40.
- 22 X. Chen, R. P. M. Dings, I. Nesmelova, S. Debbert, J. R. Haseman, J. Maxwell, T. R. Hoye and K. H. Mayo, *J. Med. Chem.*, 2006, 49, 7754–7765.
- 23 L. He, H.-S. Byun and R. Bittman, *J. Org. Chem.*, 2000, **65**, 7618–7626.
- 24 V. Rauniyar and D. G. Hall, J. Org. Chem., 2009, 74, 4236–4241.
- 25 G. D. Vilela, R. R. da Rosa, P. H. Schneider, I. H. Bechtold, J. Eccher and A. A. Merlo, *Tetrahedron Lett.*, 2011, 52, 6569–6572.
- 26 D. E. Rajsfus, S. Alter-Zilberfarb and A. A. Frimer, *J. Fluor-ine Chem.*, 2013, 148, 49–58.
- 27 G. M. Sheldrick, Acta Crystallogr., Sect. A, 2007, 64, 112– 122.
- 28 L. J. Farrugia, J. Appl. Crystallogr., 1997, 30, 565-565.
- 29 L. J. Farrugia, J. Appl. Crystallogr., 1999, 32, 837–838.
- 30 P. Van der Sluis and A. Spek, *Acta Crystallogr., Sect. A*, 1990, 46, 194–201.