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Conformational Changes of Polypeptides on Crystallization

We are interested in determining whether polypeptides retain their conformation on crystallizing from various solvent systems.¹⁻³ We give here a brief summary of some systematic infrared measurements on a series of five polypeptides. The spectra of solutions of the polypeptides in pure trifluoroacetic acid, (trifluoroacetic acid—TFA, cis-1,2-dichloroethylene—DCE, poly-L-alanine—PLA, poly-L-leucine—PLL, poly-γ-benzyl-L-glutamate—PBLG, poly-L-valine—PLV, poly-γ-methyl-L-glutamate—PMLG), and in a mixed solvent (trifluoroacetic acid plus dichloroethylene) were obtained, as were the spectra of the solids crystallized from these solvents. The amide I and II bands were used as the principal though not exclusive diagnostic tool.⁴

All solvents were the purest grade available and used as received from the manufacturers; synthetic polypeptides were purchased from Pilot Chemical Company. Infrared difference spectra were determined with a calibrated Perkin-Elmer 521 instrument using a demountable cell with 0.025-mm pathlength for the solution spectra (concentration 10 mg/ml). Solid samples were cast on watch glasses by controlled slow solvent evaporation at $23^{\circ} \pm 0.2^{\circ}\mathrm{C}$; this procedure yielded a wide range of morphologies, 2-4 which were reproducible for each polypeptide under the same casting conditions. Solid samples were usually prepared in KBr pellets or as films sandwiched between KBr discs. No solvent inclusion effects were observed and the spectra were found highly reproducible. The positions and accepted conformational assignments of the observed amide I and II maxima are summarized in Table I for two widely different solvent compositions. (The amide I and II regions of the PLL spectra, shown in Figure 1, will serve as an illustrative example.)

TABLE I
Observed Amide I and II Band Maxima of Polypeptides in Various
Solvents and in Solid States Crystallized from these Solvents

	Solvent	Solution			Solid State		
Poly- peptide	Composition TFA:DCE (v:v)	Amide I (cm ⁻¹)	Amide II (cm ⁻¹)	Con- for- mation	Amide I (cm ⁻¹)	Amide II (cm ⁻¹)	Con- for- mation
PLL	25:75 100:0	1649, 1610 1624	a 1532	α β^{d}	1654 1656	1544 1546	α α
PLA	25:75 100:0	1656, 1619 1624	1539 ^b 1532	α β d	1656 1657	1541 1543	α
PLV	25:75 100:0	1632 1625	a 1531	β β	1633 1628	1543 1544	β
PMLG	35:65 100:0	1640 1636	1539° 1533	$eta \ eta^{ ext{d}}$	1630 1624	1530 1540	β β
PBLG	25:75 100:0	1637 1640	1534 1533	$eta \ eta^{ m d}$	1650 1650	1543 1541	α α

^a Broad band, very poorly resolved, but shifted to higher wavenumber than in pure TFA.

^b Poorly resolved.

^c Shoulder.

^d These polypeptides are assigned a random-coil conformation in TFA by most authors.⁸ It is difficult to reconcile our observed frequencies, which are in the range usually attributed to a β form,^{5–8} to this assignment. Absorption near 1630 cm⁻¹ has been used by others to indicate the presence of β conformation in solution;^{9,10} Bradbury and Rattle, however, refer to "the acid solvated coil frequency of the amide I" peak at ca. 1630 cm⁻¹ for PLA in TFA.¹¹

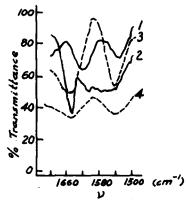


Fig. 1. Amide I and II regions of the ir spectrum of poly-L-leucine; curve 1—PLL in TFA; curve 2—PLL in 25:75 TFA:DCE; curve 3—PLL cast from TFA; curve 4—PLL cast from 25:75 TFA:DCE.

The solid-state conformations agree with those obtained previously by various experimental methods.¹²

One is struck by the fact that while the dominant conformation in solution appears, in six of the ten cases, to be largely retained on crystallization, α helix (or possibly a disordered conformation) is found in the solid state in the remaining four, even though the polypeptide appears to be predominantly in an extended form in solution (PLA, PLL, and PBLG, in pure TFA; PBLG in mixed solvent). Conformations thus may be quite different in the crystalline solid and in the dissolved material. Some similar observations have been made previously, for instance for poly(acetyl) hydroxyproline. These findings would suggest considerable caution in transferring solid-state conformational assignments (for instance based on crystal structures) uncritically to the dissolved state.

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References

- 1. Brumberger, H. & Anderson, L. C. (1972) Biopolymers 11, 679-682.
- Anderson, L. C., Brumberger, H. & Marchessault, R. H. (1967) Nature 216, 52-54.
 - 3. Brumberger, H. (1970) Nature 227, 490-491.
- 4. See Cheng, B. (1969) Ph.D. dissertation, Dept. Chem., Syracuse Univ., Syracuse, N.Y., for additional experimental details and data.
- 5. Miyazawa, T. (1967) in $Poly-\alpha$ -Amino Acids, Fasman, G. D., Ed., Marcel Dekker, New York, ch. 2.
- 6. Bamford, C. H., Elliott, A. & Hanby, W. E. (1956) Synthetic Polypeptides, Academic, New York, ch. 5.
- 7. Susi, H. (1969) in Structure and Stability of Biological Macromolecules, Timasheff, S. N. & Fasman, G. D., Eds., Marcel Dekker, New York, ch. 7.
- 8. Walton, A. G. & Blackwell, J. (1973) Biopolymers, Academic, New York, chs. 5, 9.
 - 9. Wada, A., Tsuboi, M. & Konishi, E. (1961) J. Phys. Chem. 65, 1119-1123.

- 10. Parrish, J. R. & Blout, E. R. (1971) Biopolymers 10, 1491-1512.
- 11. Bradbury, E. M. & Rattle, H. W. E. (1968) Polymer **9**, 201–217.
- 12. See ref. (8), pp. 377-378 and literature cited there.
- 13. Andries, J. C. & Walton, A. G. (1969) Biopolymers 8, 465-474.

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