Letter to the Editor: The formation of arbuscular mycorrhizae by an Ascomycete?

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Received: 4 August 2008/Accepted: 12 August 2008/Published online: 5 September 2008 © Springer Science+Business Media B.V. 2008

A recent publication by Fan et al. (2008) in Biotechnology Letters (30:1489–1494) described the isolation of *Penicillium pinophilum* (anamorphic Ascomycota) from strawberry (Fragaria × ananassa Duch. CV. Zoji) roots and reported the effects of re-inoculation with pure cultures of P. pinophilum on the growth and nutrient uptake of strawberry plants. The authors state that the isolate of P. pinophilum formed arbuscular mycorrhiza, a symbiosis known to be established between the majority of terrestrial plant roots and fungal species of Glomeromycota (Schüßler et al. 2001), yet never with fungi outside of this phylum. Glomeromycota, also known as arbuscular mycorrhizal fungi (AMF) are obligate symbionts, so far known to be fully dependent on carbon delivered from their host plants, which they in turn supply with limiting nutrients from the soil, e.g. phosphate (Smith and Read 1997). It is therefore unlikely that Fan et al. (2008) really isolated an AMF species, as their isolate grew on Melin-Norkans (MMN) media in absence of any host roots. On the other hand, as the authors identified their isolate morphologically and by molecular methods, providing a sequence in Gen-Bank (accession no. EU277738), I have no doubt that their culture at least contained a saprotrophic fungus able to grow on artificial media, and likely this was the dominant, prolifically growing fungus in their culture.

The species *P. pinophilum* has some degrading capabilities (e.g. Rando et al. 1997) and was characterized as a "minor pathogen" causing growth retardation, e.g. in tomato (Gamliel and Katan 1993). There are also studies describing antagonistic effects of *P. pinophilum* against plant pathogenic fungi (e.g. Alagesaboopathi 1994). However, no study so far has reported positive effects on plant growth and nutrient capture as were observed by Fan et al. (2008).

It is noteworthy that several publications reported synergistic effects of AMF and Penicillium spp. (e.g. Babana and Antoun 2006; Cabello et al. 2005; Chandanie et al. 2006; Zaidi and Khan 2007) on plant growth, even specifically involving P. pinophilum (Gryndler et al. 2002b). In addition, strawberry was repeatedly reported to react positively to AMF inoculation (e.g. Gange 2001; Gryndler et al. 2002a). It is therefore likely that the increase in biomass as well as nitrogen and phosphate content of the strawberry plants observed in the study by Fan et al. (2008) was potentially due to undetected contamination of their pots with AMF. This hypothesis is supported by the AMF colonization rates reported in the study. The colonization of plant roots by AMF can be easily identified by the characteristic formation of arbuscules, tree-shaped branching structures that invaginate the plant cell plasmalemma and create a large area of membrane-membrane

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apposition contact (see Vierheilig et al. 2005). Such structures were also clearly visible in the roots of the strawberry plants (personal communication).

The question remains how the contamination could have occurred as Fan et al. (2008) used autoclaved soil as growth substrate and inoculated at least one of their treatments with fresh hyphae from the culture material of P. pinophilum which, due to the obligate biotrophism of Glomeromycota, could not have any AMF propagules. A contamination in the greenhouse is also unlikely as the noninoculated control plants were found to contain no AMF structures. However, due to the stimulatory effects saprobic fungi can have on spore germination and hyphal growth of AMF (Fracchia et al. 2004) it is conceivable that inoculation with P. pinophilum stimulated AMF propagules surviving the autoclaving, and thus the AMF symbiosis was established. In case of the non-inoculated controls this stimulatory effect was not present and therefore the roots were not colonized by AMF.

Given the facts presented here, the finding that *P. pinophilum* is capable to form arbuscular mycorrhizal symbiosis should be critically tested in further experiments with other plants in order to confirm or confute the results. Simple experiments on the resistance of AMF propagules to extreme conditions encountered during autoclaving should also be undertaken.

Acknowledgements I would like to thank Tesfaye Wubet and Dirk Krüger for discussion and comments on this letter.

References

- Alagesaboopathi C (1994) Biological-control of damping-off disease of cotton seedling. Curr Sci 66:865–868
- Babana AH, Antoun H (2006) Effect of Tilemsi phosphate rock-solubilizing microorganisms on phosphorus uptake

- and yield of field-grown wheat ($Triticum\ aestivum\ L.$) in Mali. Plant Soil 287:51–58
- Cabello M, Irrazabal G, Bucsinszky AM et al (2005) Effect of an arbuscular mycorrhizal fungus, *Glomus mosseae*, and a rock-phosphate-solubilizing fungus, *Penicillium thomii*, on *Mentha piperita* growth in a soilless medium. J Basic Microbiol 45:182–189
- Chandanie WA, Kubota M, Hyakumachi M (2006) Interactions between plant growth promoting fungi and arbuscular mycorrhizal fungus *Glomus mosseae* and induction of systemic resistance to anthracnose disease in cucumber. Plant Soil 286:209–217
- Fan YQ, Luan YS, An LJ et al (2008) Arbuscular mycorrhizae formed by *Penicillium pinophilum* improve the growth, nutrient uptake and photosynthesis of strawberry with two inoculum-types. Biotechnol Lett 30:1489–1494
- Fracchia S, Sampedro I, Scervino JM et al (2004) Influence of saprobe fungi and their exudates on arbuscular mycorrhizal symbioses. Symbiosis 36:169–182
- Gamliel A, Katan J (1993) Suppression of major and minor pathogens by fluorescent pseudomonads in solarized and nonsolarized soils. Phytopathology 83:68–75
- Gange AC (2001) Species-specific responses of a root-and shoot-feeding insect to arbuscular mycorrhizal colonization of its host plant. New Phytol 150:611–618
- Gryndler M, Vosatka M, Hrselova H et al (2002a) Effect of dual inoculation with arbuscular mycorrhizal fungi and bacteria on growth and mineral nutrition of strawberry. J Plant Nutr 25:1341–1358
- Gryndler M, Vosatka M, Hrselova H et al (2002b) Interaction between arbuscular mycorrhizal fungi and cellulose in growth substrate. Appl Soil Ecol 19:279–288
- Rando D, Kohring GW, Giffhorn F (1997) Production, purification and characterization of glucose oxidase from a newly isolated strain of *Penicillium pinophilum*. Appl Microbiol Biotechnol 48:34–40
- Schüßler A, Schwarzott D, Walker C (2001) A new fungal phylum, the Glomeromycota: phylogeny and evolution. Mycol Res 105:1413–1421
- Smith SE, Read DJ (1997) Mycorrhizal symbiosis. Academic Press, San Diego
- Vierheilig H, Schweiger P, Brundrett M (2005) An overview of methods for the detection and observation of arbuscular mycorrhizal fungi in roots. Physiol Plantarum 125:393–404
- Zaidi A, Khan MS (2007) Stimulatory effects of dual inoculation with phosphate solubilising microorganisms and arbuscular mycorrhizal fungus on chickpea. Aust J Exp Agric 47:1016–1022

