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Genome Mining in *Streptomyces*. Elucidation of the Role of Baeyer−Villiger Monooxygenases and Non-Heme Iron-Dependent Dehydrogenase/Oxygenases in the Final Steps of the Biosynthesis of Pentalenolactone and Neopentalenolactone[†]

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ABSTRACT: The pentalenolactone biosynthetic gene clusters have been cloned and sequenced from two known producers of the sesquiterpenoid antibiotic pentalenolactone, *Streptomyces exfoliatus* UC5319 and *Streptomyces arenae* TU469. The recombinant enzymes PenE and PntE, from *S. exfoliatus* and *S. arenae*, respectively, catalyze the flavin-dependent Baeyer−Villiger oxidation of 1-deoxy-11-oxopentalenic acid (**7**) to pentalenolactone D (**8**). Recombinant PenD, PntD, and PtlD, the latter from *Streptomyces avermitilis*, each catalyze the Fe^{2+}-α-ketoglutarate-dependent oxidation of pentalenolactone D (**8**) to pentalenolactone E (**15**) and pentalenolactone F (**16**). Incubation of PenD, PntD, or PtlD with the isomeric neopentalenolactone D (**9**) gave PL308 (**12**) and a compound tentatively identified as neopentalenolactone E (**14**). These results are corroborated by analysis of the Δ*penD* and Δ*pntD* mutants of *S. exfoliatus* and *S. arenae*, respectively, both of which accumulate pentalenolactone D but are blocked in production of pentalenolactone as well as the precursors pentalenolactones E and F. Finally, complementation of the previously described *S. avermitilis* Δ*ptlE* Δ*ptlD* deletion mutant with either *pen*E or *pnt*E gave pentalenolactone D (**8**), while complemention of the Δ*ptlE* Δ*ptlD* double mutant with *pntE* plus *pntD* or *penE* plus *pntD* gave pentalenolactone F (**16**).

Terpenoid compounds are ubiquitous in nature, being widely distributed in terrestial and marine plants, fungi, liverworts, and, as is becoming increasingly common, many bacteria. Of the tens of thousands of known monoterpenes, sequiterpenes, and diterpenes, the several hundred parent cyclic hydrocarbon and alcohol products are formed from the universal acyclic C_{10}, C_{15}, and C_{20} precursors geranyl, farnesyl, and geranylgeranyl diphosphate. More than a thousand presumptive terpene cyclase genes from plants and microorganisms have been identified, of which about 10% have been experimentally assigned a confirmed biochemical function by identification of their native substrate and characteristic cyclization product. Building on earlier classical isotopic precursor incorporation studies with intact organisms, probing mechanistic investigations using purified, often recombinant, terpene synthases have established many of the key mechanistic details of the intricate cyclization pathways themselves (*1*−*3*). Such studies have been powerfully supported by an increasing number of detailed crystallographic investigations (*4*, *5*) and recent computational analyses of cyclization mechanisms (*6*, *7*).

The vast majority of terpenoid metabolites are in fact derived by one or more oxidations and other late stage modifications of the parent hydrocarbons or alcohols initially generated by terpene synthases (*3*, *8*−*16*). Nature uses an impressive variety of biochemical strategies for the oxidative cleavage of carbon−carbon bonds, mediated not just by P450s but also by non-heme-Fe^{2+}-α-ketoglutarate-dependent dioxygenases and flavin-dependent monooxygenases. Little is known about the biosynthetic logic governing this common late-stage oxidative metabolism, including the roles of specific oxygenases and the sequential order of such transformations.

Pentalenolactone (**1**) is a sesquiterpenoid antibiotic that has been isolated from more than 30 species of *Streptomyces* (*17*−*20*). The antibiotic action of pentalenolactone is due to the presence of an electrophilic epoxylactone moiety that alkylates the active site cysteine of the target glycolytic enzyme glyceraldehyde-3-phosphate dehydrogenase (GAPDH)[1] (*21*−*24*). Self-resistance in the pentalenolactone producer *Streptomyces arenae* TU469 is due to an inducible, pentalenolactone-insensitive GAPDH encoded by the *gapR* gene (*25*−*28*). The biosynthesis of pentalenolactone has been studied in considerable detail (*29*−*31*) (Scheme 1). Pentalenene synthase, which catalyzes the cyclization of farnesyl diphosphate (**2**, FPP) to the parent sesquiterpene hydrocarbon pentalenene (**3**) (*32*), has been cloned from *Streptomyces exfoliatus* UC5319 and expressed in *Escherichia coli* (*33*), and its

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[1]Abbreviations: BVMO, Baeyer−Villiger monooxygenase; CIAP, calf intestinal alkaline phosphatase; DTT, 1,4-dithiothreitol; FAD, flavin adenine dinucleotide; FPP, farnesyl diphosphate; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; HSV, Herpes simplex virus; IPTG, isopropyl β-D-thiogalactopyranoside; LB, Luria−Bertani medium; NAD^+, β-nicotinamide adenine dinucleotide; NADPH, β-nicotinamide adenine dinucleotide 2′-phosphate reduced tetrasodium salt hydrate; ODS, octadecylsilane; ORF, open reading frame; SFM, soy flour, mannitol; PMSF, phenylmethanesulfonyl fluoride; TAP, transcriptionally active polymerase chain reaction; TB, Terrific broth.

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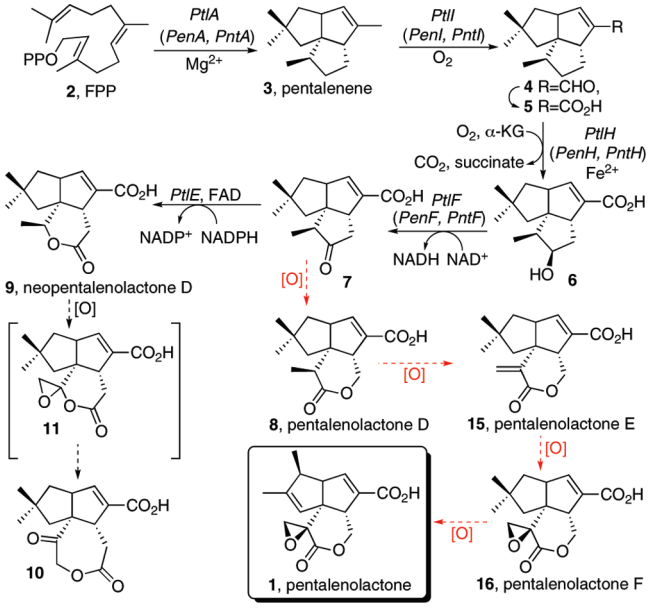
crystal structure has been determined (*34*). The detailed mechanism and stereochemistry of the cyclization itself has been elucidated (*31*, *35*). In addition, a variety of oxidized metabolites that represent plausible intermediates and shunt metabolites in the conversion of pentalenene to pentalenolactone have been isolated from several pentalenolactone-producing strains of *Streptomyces* (*36*−*40*).

We have recently reported that the 13.4-kb *ptl* gene cluster centered at 3.75 Mb of the 9.03-Mb linear genome of *Streptomyces avermitilis* harbors 13 unidirectionally transcribed open reading frames (ORFs) that encode several enzymes involved in the early stages of pentalenolactone biosynthesis (*41*) (Figure 1). We have determined the biochemical function of several of these ORFs, including a pentalenene synthase (PtlA, SAV_2998) (*41*), a P450-dependent monooxygenase (PtlI, SAV_2999) that catalyzes the oxidative conversion of pentalenene (**3**) to pentalenal (**4**) and, most likely, to 1-deoxypentalenic acid (**5**) (*42*), an Fe^{2+}-α-ketoglutarate-dependent dioxygenase (PtlH, SAV_2991) that hydroxylates **5** to 1-deoxy-11β-hydroxypentalenic acid (**6**) (*43*, *44*), and an NAD^+-dependent dehydrogenase (PtlF, SAV_2993) that oxidizes **6** to the corresponding ketone, 1-deoxy-11-oxopentalenic acid (**7**) (*45*) (Scheme 1, Figure 1). At the 5′-end of the cluster is the resistance gene *gap1* (*sav2990*) which encodes a pentalenolactone-insensitive glyceraldehyde-3-phosphate dehydrogenase (*41*). Each of these biochemical reactions has been directly demonstrated using the corresponding recombinant proteins, and the results have been supported using engineered deletion mutants of *S. avermitilis* that harbor defined segments of the *ptl* gene cluster under control of the *ermE* promoter. In addition, *ptlB* (*sav2997*) encodes a presumptive prenyl transferase responsible for the dedicated formation of FPP, while *ptlG* (*sav2992*) is predicted to encode a transmembrane efflux protein proposed to be responsible for export of the final antibiotic.

Scheme 1: Biosynthesis of Neopentalenolactone D and Proposed Biosynthesis of Pentalenolactone



Conversion of 1-deoxy-11-oxopentalenic acid (**7**) to pentalenolactone should require four distinct oxidative tranformations, with the initial step expected to be a Baeyer−Villiger-type oxidation of **7** to the corresponding lactone, pentalenolactone D (**8**) (Scheme 1). In our investigations of the *S. avermitilis ptl* cluster, however, we unexpectedly found that while recombinant PtlE did indeed catalyze a flavin-dependent Baeyer−Villiger reaction of **7**, instead of generating **8** the product of the reaction was the previously unknown isomeric lactone, neopentalenolactone D (**9**) (*46*). In support of this finding, the *S. avermitilis* Δ*ptlE* Δ*ptlD* double deletion mutant accumulated 1-deoxy-11-oxopentalenic acid (**7**) while the Δ*ptlD* deletion mutant accumulated **9** (Figure 2). In fact, it is now clear that wild-type *S. avermitilis* does not produce pentalenolactone itself but instead a group of new metabolites that are neopentalenolactone derivatives. For example, cultures of *S. avermitilis* SUKA5, from which >1.5 Mb of DNA had been deleted from one end of the 9.05-Mb linear genome (*46*, *47*) but in which the *ptl* cluster was still intact, produced the novel metabolite neopentalenoketolactone (**10**), a presumed rearrangement product of the hypothetical labile neopentalenolactone F (**11**). Similarly, *S. avermitilis* SUKA16, a derivative of SUKA5 in which a portion of the *ptl* cluster had been placed under control of the strong constitutive *ermE* promoter, produced two previously unknown metabolites, characterized as the corresponding methyl esters, PL308 (**12**) and

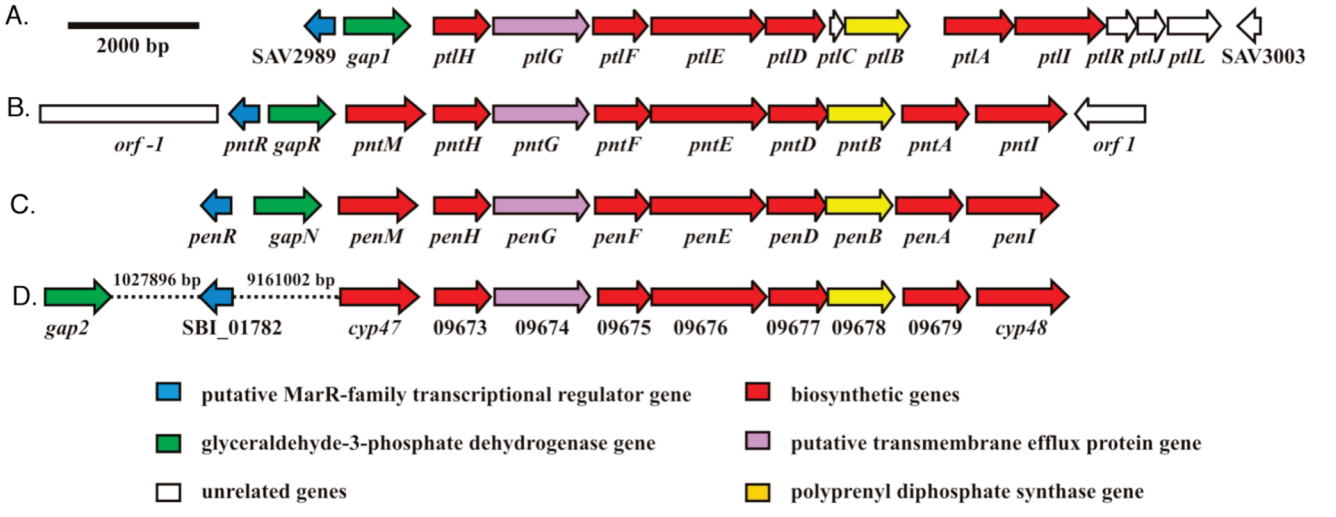


FIGURE 1: Neopentalenolactone and pentalenolactone biosynthetic gene clusters. (A) *S. avermitilis ptl* gene cluster for neopentalenolactone biosynthesis. (B) *S. arenae pnt* gene cluster for pentalenolactone biosynthesis. (C) *S. exfoliatus pen* gene cluster for pentalenolactone biosynthesis. (D) Proposed *S. bingchenggensis* pentalenolactone biosynthetic gene cluster.

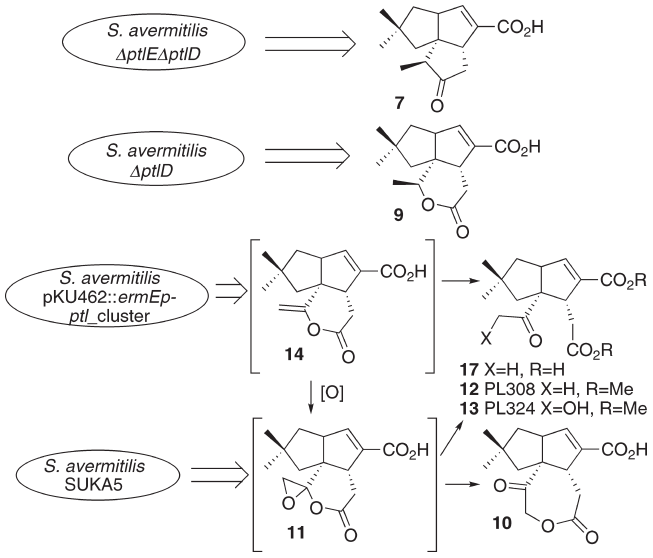


FIGURE 2: Production of neopentalenolactone metabolites and intermediates by *S. avermitilis* mutants.

PL324 (**13**). Both **12** and **13** are thought to result from hydrolysis of the hypothetical products of oxidation of **9**, neopentalenolactones E (**14**) and F (**11**). Significantly, neither **10**, **12**, nor **13** are produced by the *S. avermitilis* Δ*ptlD* mutant, indicating that each of these metabolites must result from oxidation of neopentalenolactone D (**9**), presumably mediated by the *ptlD* gene product.

Although we had discovered a previously unknown branch of the classical pentalenolactone pathway, we were still faced with elucidating the final oxidative steps in the biosynthesis of pentalenolactone itself. We now report the cloning and characterization of the complete pentalenolactone biosynthetic gene clusters from two known producers of pentalenolactone, *S. exfoliatus* UC5319 (*18*) and *S. arenae* TU469 (*19*). We have further established that the orthologous enzymes from the *pen* and *pnt* clusters, PenE and PntE, respectively, catalyze the flavin-dependent Baeyer−Villiger oxidation of 1-deoxy-11-oxopentalenic acid (**7**) to pentalenolactone D (**8**). We also demonstrate that the orthologous proteins PenD and PntD each then mediate the two-stage Fe^{2+}-α-ketoglutarate-dependent oxidation of **8** to pentalenolactones E (**15**) and F (**16**), a reaction that we show can also be catalyzed by the closely related *S. avermitilis* enzyme, PtlD. Similarly, incubation of PtlD with its natural substrate neopentalenolactone D (**9**) gives a mixture of neopentalenolactone E (**14**) and its derived hydrolysis product PL308 (**12**). These biochemical results are strongly corroborated by analysis of the products of blocked mutants of both *S. exfoliatus* and *S. arenae*, as well as by systematic mutant complementation experiments in *S. exfoliatus*, *S. arenae*, and *S. avermitilis*.

EXPERIMENTAL PROCEDURES

Bacterial Strains and Plasmids. *Streptomyces* and *E. coli* strains, plasmids, and cosmids are listed in Table S1 (Supporting Information).

Materials. Reagents and solvents purchased from Sigma-Aldrich or Fisher Scientific were of the highest quality available and were used without further purification. Recombinant 1-deoxy-11-hydroxypentalenic acid dehydrogenase (PtlF) was expressed and purified as previously described (*45*). Restriction enzymes and T4 DNA ligase were purchased from New England Biolabs and used according to the manufacturer's specifications. Isopropyl β-D-thiogalactopyranoside (IPTG) was purchased from Invitrogen. Ni-NTA affinity resin was purchased from Qiagen. Amicon Ultra Centrifugal Filter Units (Amicon Ultra-15, 10000 and 30000 MWCO) were purchased from Millipore. DNA primers were synthesized by Integrated DNA Technologies. Synthetic genes encoding PenE, PntE, PenD, PntD, and PtlD, optimized for expression in *E. coli*, were prepared by DNA2.0 and supplied in the vectors pJexpress401 and pJ201.

Methods. General methods were as previously described (*41*, *46*). Growth media and conditions used for *E. coli* and *Streptomyces* strains and standard methods for handling *E. coli* and *Streptomyces in vivo* and *in vitro* were those described previously (*48*, *49*), unless otherwise noted. All DNA manipulations were performed following standard procedures (*49*). DNA sequencing was carried out at the U. C. Davis Sequencing Facility, Davis, CA. All proteins were handled at 4 °C unless otherwise stated. Protein concentrations were determined according to the method of Bradford, using a Hewlett-Packard 8452A diode array UV/vis spectrophotometer with bovine serum albumin as the standard (*50*). Protein purity was estimated using SDS−PAGE gel electrophoresis and visualized using Coomassie Blue stain. GC-MS analyses were carried out using either GC-MS method 1, Hewlett-Packard Series 2 GC-MSD, at 70 eV electron impact (EI), operating in positive ion mode, using a HP5MS capillary column (30 m × 0.25 mm) with a solvent delay of 3 min and a temperature program of 60 °C for 2 min, followed by a temperature gradient of 60−280 °C for 11 min at 20 °C/min and a hold at 280 °C for 2 min, or GC-MS method 2, Shimadzu GC-17A, 70 eV EI, operating in positive ion mode, using a neutral bond-5 capillary column (5% phenylmethylsilicon; 30 m × 0.25 mm) with a temperature program of 50−280 °C, 20 °C/min. MALDI-TOF measurements were performed on an Applied Biosystems Voyager DE PRO MALDI-TOF benchtop mass spectrometer. LC-ESI-MS analysis of recombinant proteins was carried out on a Thermo LXQ LC-ESI-MS equipped with Surveyor HPLC system and Waters Symmetry C18 column (2.1 mm × 50 mm, 3.5 μm).

Plasmid p56 Harboring the Pentalenene Synthase Gene from S. exfoliatus UC5319. The primer pair MJ-1 (5′-CTGGCCTCCCGTTTCTACCC-3′) and MJ-2 (5′-AGACGC TGCGGCTCTTGGAC-3′) derived from the internal sequence of *S. exfoliatus* UC5319 pentalenene synthase (GenBank Accession No. U05213) was used to amplify genomic DNA from *S. exfoliatus* UC5319. The resultant 601-bp PCR product was used as a probe for Southern hybridization with genomic DNA isolated from *S. exfoliatus* UC5319 that had been digested with different restriction endonucleases, using the Roche DIG High-Prime DNA Labeling and Detection Starter Kit I. A positively hybridizing band was located at 7 kb in the lane derived from *Pst*I-digested chromosomal DNA. Genomic DNA of *S. exfoliatus* UC5319 was digested with *Pst*I, and the ca. 7-kb DNA fragments were recovered and inserted into the *Pst*I site of pBluescriptII SK(+) to generate a library that was screened by PCR with primer pairs MJ-1 and MJ-2 to identify the positive plasmid p56. Sequencing of the ~7-kb insert of p56 confirmed the presence of the complete pentalenene synthase gene, *penA*.

Generation and Probing of S. exfoliatus UC5319 and S. arenae TU469 Cosmid Libraries. Genomic DNA from *S. exfoliatus* UC5319 and from *S. arenae* TU469 was isolated by salting out (*48*) and partially digested with *Bfu*CI to obtain 30−60-kb DNA fragments. The DNA fragments were dephosphorylated with CIAP and cloned into the vector pHZ1357 (*51*)

that had been treated with *Xba*I, then CIAP, and then *Bam*HI. Each of the libraries was packaged using the Stratagene Gigapack III XL packaging extract and transduced into *E. coli* DH10B. LB agar plates with 100 μg/mL ampicillin were used to select transductants. The genomic library of *S. exfoliatus* UC5319 was made up of 2000 colonies, and the genomic library of *S. arenae* TU469 consisted of 2880 colonies. The primer pairs MJ-1 and MJ-2 were used to screen the *S. exfoliatus* UC5319 cosmid library by PCR, giving positive cosmids G21, K5, and O34. The primer pair DQ66F, 5′-CGCCTACACGCAGGACCAGA-3′, and DQ66R, 5′-ACAGGGACGACCCGATGAGC-3′, based on an internal region of the known DNA sequence of *S. arenae gapR* (GenBank Accession No. U44856) was used to screen the *S. arenae* TU469 cosmid library. Six positive cosmids, 1E2, 7D7, 12B2, 21A5, 21F7, and 27H6, were selected by PCR based on the generation of the 461-bp PCR product.

Sequencing and Sequence Assembly. For the *S. exfoliatus* UC5319 cluster, plasmid p56 harboring the pentalenene synthase gene and flanked by a partial *ptl*-like gene cluster of ~6.9 kb was sequenced by primer walking. Cosmids G21, K5, and O34 from the genomic library of *S. exfoliatus* UC5319 were further screened by PCR using the primer pair MJ-3 (5′-TGGCGACGATCCAGGCGGTC-3′) and MJ-4 (5′-CCATCACCACGCCGGATTCG-3′) derived from plasmid p56 and corresponding to the partial *penG* gene. Only cosmid G21 gave an amplicon of the predicted size (573 bp). Cosmid G21 was therefore selected for sequencing by primer walking in order to obtain complete DNA sequence information for the *S. exfoliatus* UC5319 pentalenolactone biosynthetic gene cluster. Each of the six positive cosmids of *S. arenae* TU469 was digested with different restriction endonuclease enzymes to build up restriction enzyme patterns. Several *Bam*HI DNA fragments were then inserted into the *Bam*HI site of vector pSET152 to generate subclones that were sequenced at each end. Four *Bam*HI DNA fragments of about 18 kb were believed to harbor the complete *ptl*-like gene cluster according to the restriction enzyme pattern and primary DNA sequence information. Four subclones, pDQ19 (or pDQ9 from 21F7), pDQ20, pDQ22 (or pDQ7 from 21F7), and pDQ23, containing the four *Bam*HI DNA fragments from cosmid 1E2 and a subclone pDQ31 containing a *Kpn*I DNA fragment about 7 kb from cosmid 7D7 as a supplement were sequenced. The DNA fragments recovered from these plasmids were partially digested with *Bfu*CI to obtain 1−2-kb DNA fragments that were each inserted into the *Bam*HI site of pBluescriptII SK(+) to generate a small library that was submitted for sequencing. Multiple alignments of sequences were constructed by using the BioEdit sequence alignment editor. The gaps were finished by primer walking. The sequence data were analyzed with the online FramePlot program (http://www.nih.go.jp/~jun/cgi-bin/frameplot.pl). DNA and deduced protein sequence homology searches were performed by using the NCBI BLAST server. The *ptl*-like gene cluster of *S. exfoliatus* UC5319 was named the *pen* gene cluster, and the *ptl*-like gene cluster of *S. arenae* TU469 was named the *pnt* gene cluster (Figure 1). Both sequences have been deposited in GenBank with Accession Numbers HQ292066 and HQ292065, respectively.

Expression and Purification of Recombinant PenE and PntE Proteins. The synthetic *penE* and *pntE* genes optimized for expression in *E. coli* carried a 5′-*Nde*I site including the ATG start codon and a *Xho*I site immediately downstream of the stop codon. The plasmid pJexpress401:*penE*_opt_alt2 was digested with *Nde*I and *Xho*I and subcloned into the corresponding sites of pET-28a(+) vector to give the expression plasmid pET28a-*penE* which was then transformed into *E. coli* BL21(DE3). For expression of synthetic *pntE*, a pair of primers DQ75F3 [5′-CGGCAGCCATATGGTTGATCTGGAG-3′ (*Nde*I underlined)] and DQ75R3 [5′-GGCTCCTCGAGGCGCAGCTCCAGGC-3′ (*Xho*I underlined)] was used to amplify the synthetic gene *pntE* in pJexpress401:*pntE*_opt_alt2. The PCR product was digested with *Nde*I and *Xho*I and inserted into the corresponding site of pET-26b to generate expression plasmid pDQ58 (pET26b-*pntE*) that was sequenced to confirm that no mutations had been introduced during PCR amplification.

For overexpression of recombinant PenE with an N-terminal His_6 tag and PntE with a C-terminal His_6 tag, *E. coli* BL21(DE3) harboring pET28a-*penE* or pDQ58 (pET26b-*pntE*) was grown in Terrific broth (TB) media supplemented with 50 μg/mL kanamycin at 37 °C until the OD_{600} reached 0.6−0.8. IPTG was added to a final concentration of 0.4 mM, and the culture was further incubated at 18 °C overnight. The cells were then harvested by centrifugation at 5000*g* for 15 min and resuspended in lysis buffer A (50 mM Tris-HCl, 10% glycerol, 300 mM NaCl, 0.1 mM DTT, 2.7 mM β-mercaptoethanol, 10 mM imidazole, pH 8.0) containing 10 mg/L pepstatin, 10 mg/L phenylmethanesulfonyl fluoride (PMSF), and 0.2 mg/mL benzamidine. After cell disruption by sonication, the cell debris was removed by centrifugation at 20000*g* for 30 min, and the supernatant was loaded into a Ni-NTA column preequilibrated with lysis buffer. After collecting the flow-through and washing with 20 mM imidazole in lysis buffer A at a flow rate of 2 mL/min, the PenE protein was eluted with elution buffer A (50 mM Tris-HCl, 10% glycerol, 300 mM NaCl, 0.1 mM DTT, 2.7 mM β-mercaptoethanol, 200 mM imidazole, pH 8.0) at a flow rate of 1 mL/min. The fractions were analyzed by SDS−PAGE. The pooled PenE protein was concentrated and buffer-exchanged into storage buffer (50 mM NaH_2PO_4, 0.1 mM DTT, pH 7.0) using an Amicon Ultra Centrifugal Filter (Ultracel-30K) while the recombinant PntE was concentrated and buffer-exchanged using a PD-10 gel filtration column into the storage buffer containing 10% glycerol (50 mM NaH_2PO_4, 0.1 mM DTT, pH 7.0, containing 10% glycerol). The yield of purified recombinant PenE was 35 mg/L of culture and of PntE 27 mg/L of culture. UV: Pen E, λ_{max} 375, 450 nm; PntE, λ_{max} 380, 450 nm. MALDI-TOF: His_6-PenE 67480 ± 30 Da (predicted P-Met, 67518); PntE-His_6 tag 66915 ± 30 Da (predicted 67018). ESI-MS: PntE-His_6 tag, m/z 67008.

Expression and Purification of Recombinant PenD, PntD, and PtlD Proteins. The synthetic *penD* and *pntD* genes in the pJexpress401 vector and synthetic *ptlD* in pJ201 were prepared by DNA2.0 with codons optimized for expression in *E. coli*. Each of the plasmids was digested with *Nde*I and *Xho*I, and the *penD*, *pntD*, and *ptlD* products were ligated into doubly digested pET-28a(+) to give the expression vectors pET28a-*penD*, pET28a-*pntD*, and pET28a-*ptlD*, respectively, which were then individually transformed into *E. coli* BL21(DE3). Cultures of each of the recombinant *E. coli* transformants were grown in TB media containing 50 μg/mL kanamycin at 37 °C to OD_{600} of 0.6−0.8. After addition of 0.4 mM IPTG, the culture was further incubated at 18 °C overnight. The harvested cells were resuspended in lysis buffer B (50 mM NaH_2PO_4, 300 mM NaCl, 10 mM imidazole, pH 8.0), and the suspension was then sonicated. After removal of cell debris, the cell-free extract was applied to a Ni-NTA column and washed with 20 mM imidazole in lysis buffer B. The proteins were finally eluted with elution buffer B (50 mM NaH_2PO_4, 300 mM NaCl, 250 mM imidazole, pH 8.0).

The concentrated protein was exchanged into the storage buffer (20 mM Tris-HCl, 20% glycerol, pH 7.5) using an Amicon ultracentrifugal filter (Ultracel-10K). The purified PenD, PntD, and PtlD concentrations were 42, 36, and 48 mg/L of culture, respectively. ESI-MS for His_6 tag-PenD, m/z 35472 (predicted 35532); ESI-MS for His_6 tag-PntD, m/z 35626 (predicted 35624); ESI-MS for His_6 tag-PtlD, m/z 36268 (predicted P-Met, 36269).

Incubations of PenE and PntE with 1-Deoxy-11-oxopentalenic Acid (**7**). 1-Deoxy-11β-hydroxypentalenic acid (**6**) (0.1 mM), prepared as described below, was incubated at room temperature with recombinant PtlF (14.4 μM) and β-NAD^+ (1.6 mM) in 1 mL of Tris buffer (100 mM Tris-HCl, 1.5 mM DTT, 2% DMSO, pH 8.0) to generate 1-deoxy-11-oxopentalenic acid (**7**). After 20 min, purified PenE (24.5 μM) or PntE (5.4 μM) protein was added along with β-NADPH (1 mM) and FAD (50 μM). After an additional 2 h, the reaction mixture was quenched by addition of 10% HCl to adjust the pH to 2.0, followed by extraction with 3× 1 mL of dichloromethane. The combined organic extracts were dried over anhydrous Na_2SO_4 and concentrated on a rotovap. The residue was resuspended in methanol and treated with trimethylsilyldiazomethane (TMS-CHN_2) to the yield pentalenolactone D methyl ester (**8-Me**), which was identified by direct GC-MS comparison with an authentic sample of **8-Me** (GC-MS method 1).

Incubations of PenD, PntD, and PtlD with Pentalenolactone D (***8***)*, Neopentalenolactone D* (***9***)*, or Pentalenolactone E* (***15***). Enzymatic reactions were performed in 10 mM imidazole buffer (5 mL, pH 6.5) containing 1 mM α-ketoglutarate, 1 mM sodium ascorbate, 0.1 mM $Fe(NH_4)_2(SO_4)_2$, 0.5 mM DTT, 1 mg/mL bovine catalase, and 0.1 mM pentalenolactone D (**8**), neopentalenolactone D (**9**), or pentalenolactone E (**15**). The reaction was initiated by addition of purified PntD, PenD, or PtlD protein (8.4 μM), and the mixture was incubated at 30 °C for 1.5 h. After quenching by addition of 10% HCl to adjust the pH to 2.0, the reaction mixture was extracted with dichloromethane. The combined organic extracts were dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The residue was resuspended in methanol and treated with TMS-CHN_2 followed by GC-MS analysis of the derived methyl esters (GC-MS method 1).

Construction of pntD Mutant S. arenae ZD18. The plasmid pJTU1278 is a derivative of pHZ1358 with a cassette carrying multiple cloning sites and a marker for *lacZ* selection (*52*). The *Xba*I and *Spe*I sites were removed by digestion of pJTU1278 with *Xba*I and *Spe*I and religation to generate the vector pDQ44. The method of PCR-targeted *Streptomyces* gene replacement was used to generate mutants of *S. arenae* TU469 (*53*). A 6412-bp *Kpn*I DNA fragment from pDQ34 harboring *pntD* was inserted into the *Kpn*I site of pDQ44 to generate pDQ45. The plasmid pDQ45 was transformed into *E. coli* BW25113/pIJ790. A pair of primers, DQ79F (5′-ATGGAAGTGACACCGATACCCGGCGCCCCTCTCGGCGCCATTCCGGGGATCCGTCGACC-3′) and DQ79R (5′-TCATGCGGCGGCCTCCGCGCGCCAGGGGCCGGCCGGGTCTGTAGGCTGGAGCTGCTTC-3′), was used for PCR amplification, using as template the 1382-bp *Eco*RI + *Hin*dIII DNA fragment from pIJ773 carrying *oriT* and *aac*(*3*)*IV* to give a 1447-bp product that was transformed into *E. coli* BW25113/pIJ790 containing pDQ45 to generate plasmid pDQ46 in which the 822-bp DNA fragment of *pntD* from nt 40 to nt 861 nt had been replaced by *oriT* and *aac*(*3*)*IV* (~1369 bp) (Figure 4). The plasmid pDQ46 recovered from *E. coli* BW25113 was transformed into *E. coli* ET12567/pUZ8002, and the reisolated unmethylated pDQ46 was digested with *Xba*I and religated to generate pDQ47 from which the *oriT* and *aac*(*3*)*IV* DNA fragment had been deleted leaving a 81-bp scar.

Plasmid pDQ46 was conjugated into *S. arenae* TU469 using apramycin to select exconjugants. The single crossover strains were inoculated onto SFM agar plates (20 g of soy flour, 20 g of mannitol, 20 g of agar in 1 L of water (*48*)) without any added antibiotic in order to obtain double crossover strains under relaxed conditions with apramycin resistance and without thiostrepton resistance. The resultant mutants were confirmed by PCR using the primer pair DQ80F (5′-TGCGAAAGGAGGCAACGC-3′) and DQ80R (5′-GGTGATCCAGCCGAAGTGGTAG-3′). The PCR product of the wild-type *S. arenae* strain is 1041 bp, and the PCR product of the *pntD* mutant, *S arenae* ZD17, was 1588 bp in which an internal 822 bp of TU469 *pntD* from nt 40 to nt 861 had been replaced by the 1369-bp fragment harboring *oriT* and *aac*(*3*)*IV* (Supporting Information Figure S5).

Plasmid pDQ47 was conjugated into *S. arenae* ZD17, and thiostrepton was used to select exconjugants. The single crossover strains were inoculated on SFM plates without antibiotic in order to obtain double crossover strains that had lost both apramycin and thiostrepton resistance. PCR amplification with the primer pair DQ80F/R gave a 300-bp product from the double crossover strain *S. arenae* ZD18 in which the 1369-bp *oriT* and *aac*(*3*)*IV* DNA fragment in ZD17 has been deleted leaving a 81-bp scar corresponding to an in-frame deletion mutant of *pntD* (Supporting Information Figure S5).

Construction of penD Mutant S. exfoliatus ZD20. A ca. 7-kb *Bam*HI + *Hin*dIII DNA fragment harboring *penD* from plasmid p56 was inserted into doubly digested pDQ44 to generate pDQ49. pDQ49 was transformed into *E. coli* BW25113/pIJ790. The 1382-bp *Eco*RI + *Hin*dIII DNA fragment from pIJ773 carrying *oriT* and *aac*(*3*)*IV* was used as tempate for PCR amplification with the primer pair MJ-26 (5′-ATGGACGTAACGCCGATACCCGGCGCAGCTCTCGGGGCAATTCCGGGGATCCGTCGACC-3′) and MJ-27 (5′-TCATGCGGCGACCTCCGCGTGCCAGGGGTCGGTGGGGTCTGTAGGCTGGAGCTGCTTC-3′) to give a 1447-bp DNA fragment that was transformed into *E. coli* BW25113/pIJ790 containing pDQ49 to generate plasmid pDQ50 in which an internal 819 bp of *penD* from nt 40 to nt 858 had been replaced by the ca. 1369-bp *oriT* and *aac*(*3*)*IV*. The plasmid pDQ50 isolated from *E. coli* BW25113 was transformed into *E. coli* ET12567/pUZ8002, and the resultant unmethylated pDQ50 was digested with *Xba*I and religated to generate pDQ51 in which the 1369-bp *oriT* and *aac*(*3*)*IV* DNA fragment had been deleted and replaced with a 81-bp scar. Plasmid pDQ51 was conjugated into *S. exfoliatus* UC5319, and thiostrepton was used to select exconjugants (Figure 3). The single crossover strains were inoculated on SFM plates without any antibiotic under relaxed conditions in order to get double crossover strains that had lost thiostrepton resistance. The primer pair DQ81F (5′-GGCAGGCGAGAAGGAATGG-3′) and DQ81R (5′-TGGTGACGTGAGCGGTGGTC-3′) was used to select the *penD* mutant, *S. exfoliatus* ZD20, which gave a PCR product of 503 bp compared to 1241 bp with wild type strain *S. exfoliatus* UC5319. The in-frame deletion mutant *S. exfoliatus* ZD20 has an 81-bp scar that has replaced an internal 819 bp of *penD* from nt 40 to nt 858 (Supporting Information Figure S5).

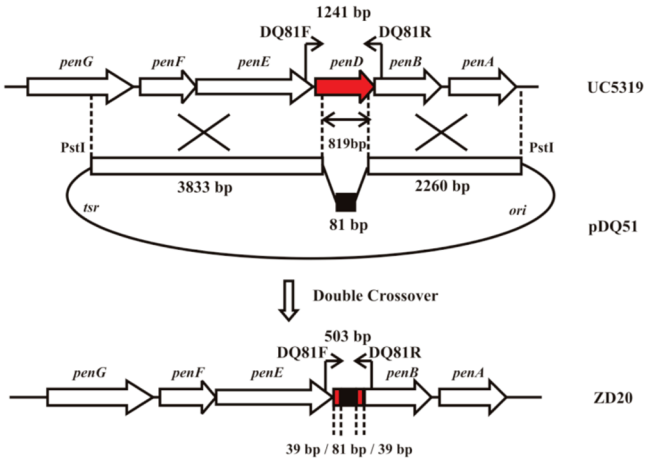


FIGURE 3: Construction of Δ*penD* deletion mutant *S. exfoliatus* ZD20. (See also Supporting Information Figure S5.)

Analysis of Pentalenolactone Production in Wild-Type S. arenae TU469 and Deletion Mutant ZD18(Δ*pntD*). The liquid synthetic medium (*28*) used for fermentation of *S arenae* TU469 and ZD18 contained 4% mannitol, 0.25% asparagines, 0.2% $(NH_4)_2SO_4$, 0.1% NaCl, 0.3% $K_2HPO_4 \cdot 3H_2O$, 0.1% $MgSO_4 \cdot 7H_2O$, 0.04% $CaCl_2 \cdot 2H_2O$, 0.002% $FeSO_4 \cdot 7H_2O$, and 0.001% $ZnSO_4 \cdot 7H_2O$, pH 6.2. The strains of *S. arenae* TU469 and ZD18 were fermented at 30 °C for 6 days. The fermentation broth was adjusted to pH 2.5 with HCl and extracted with chloroform. The organic layers were dried over anhydrous Na_2SO_4, concentrated, methylated with TMS-CHN_2, and analyzed by GC-MS.

Analysis of Pentalenolactone Production in Wild-Type S. exfoliatus UC5319 and Deletion Mutant ZD20(Δ*penD*). The liquid medium used in fermentation of *S. exfoliatus* UC5319 and ZD20 contained 0.2% NaCl, 0.5% $CaCO_3$, 1% corn gluten meal, 0.1125% bactodextrose, 0.2% blackstrap molasses, and 2% corn starch, pH 7.2 (*37*). After incubation at 30 °C for 6 days, the culture was acidified to pH 2.4 with H_2SO_4 and extracted with chloroform. The organic layer was dried over anhydrous Na_2SO_4, concentrated, methylated with TMS-CHN_2, and analyzed by GC-MS.

Complementation of S. avermitilis ΔptlE ΔptlD Double Deletion Mutant with penE, pntE, penD, and pntD. The construction of the *S. avermitilis* SUKA16 pKU462::*ermEp-ptl*-cluster Δ*ptlE* Δ*ptlD* double deletion mutant has been previously described (*46*). The recombinant plasmid pDQ34 (*pnt* cluster) was digested with *Acc*65I, and the largest 6441-bp fragment carrying the *pnt* cluster was introduced into the unique *Acc*65I site of pKU464*aac*(*3*)*IV* to generate pKU464*aac*(*3*)*IV::pnt*-cluster. A second recombinant plasmid p56 (*pen*-cluster) was digested with *Pst*I, and the largest 6906-bp fragment carrying the *pen* cluster was introduced into the unique *Pst*I site of pKU464*aac*(*3*)*IV* to generate pKU464*aac*(*3*)*IV::pen*-cluster. To express *pntE-pntD* or *penE-penD* under the control of an alternative constitutive promoter, *ermEp* was introduced upstream of *pntE* or *penE* using the λ recombination system (*46*). The *ermEp* promoter with *aac*(*3*)*I* was amplified by PCR using the primer pair, forward, 5′-**TCGCGCTCCTGCCGGTACTTCTCCCTCA CCGCTTCCAGGTCCAT**ATGTAGATCCTACCAACCGG-C-3′ (bold characters indicate the N-terminal region of *pntE*, and underlined characters correspond to the start condon of *pntE*), and reverse, 5′- **GTGCTGGCGAGCAGCGAT CCAGGCGGTCAC-GGCGGGGCAGG**GAAGCTAGCGATCTCGGCTTGAAC-3′ (bold characters indicate a part of the region for *pntG*), for DNA from the *pnt* cluster and the primer pair, forward, 5′-**CCTACG-CTGCGCTTGTCGCGTTCCTGCCGGTATTTCTCCCTCAT**-ATGTAGATCCTA CCAACCGGC-3′ (bold characters indicate N-terminal region of *penE*, and underlined characters correspond to the start codon of *penE*), and reverse, 5′-**GAAGTTAGCA GCCCTTGCGCCCTGAGTGCTTGCGGCAGCGTG** AAGC-TAGCGATCTCGGCTTGAAC-3′ (bold characters indicate the region for *lacZ*α of the vector upstream of the cloning site), for the *pen* cluster, respectively. The initial denaturation step (95 °C, 3 min) was followed by 25 cycles of amplification (95 °C, 30 s; 65 °C, 30 s; 72 °C, 30 s) and final incubation at 72 °C (10 min) using Phusion DNA polymerase (New England Biolabs, USA). Each amplicon was cotransformed into L-arabinose-induced *E. coli* BW25113 with linearized (*Sst*I digestion) pKU464*aac*-(*3*)*IV::pnt*-cluster or pKU464*aac*(*3*)*IV::pen*-cluster by electro-poration as described previously (*46*). The desired recombinant plasmids were obtained by the selection for resistance with 25 μg/mL apramycin [*aac*(*3*)*IV*] and 25 μg/mL fortimicin [*aac*(*3*)*I*], and the isolated plasmids were confirmed by restriction digestion to obtain pKU464*aac*(*3*)*IV::ermEp::pnt*-cluster and pKU464*aac*(*3*)*IV::ermEp::pen*-cluster, respectively.

To construct expression cassettes for *pntE*, *pntE-pntD*, *penE*, and *penE-penD*, the downstream region of each plasmid was removed by the λ-recombination system and Cre/*loxP* site-specific recombination. An antibiotic gene *aad*(*3″*) was amplified for each targeted deletion derivative using the designated primer pairs. For *pntE* expression, forward primer 5′-**GACCGTCCCCC-ACCGTCTCTGCGAAAGGAGGCAACGCCATG**CAGTGA-GTTCGAGCGACTCGAGT-3′ (bold characters indicate down-stream of *pntE*, and underlined characters correspond to the start codon of *pntD*) and reverse primer *pnt* 5′-**TCGAAGCGG-CCCGCGCGCTCGGCGAGGTCGACCGTGGGCTG**CTGG-TACCGAGCGA ACGCGTT-3′ (bold characters indicate the region for *pntA*) for the construction of the *ermEp-pntE* cassette. For *pntE-pntD* expression, forward primer 5′-**TGGCGCGCG-GAGGCCGCCGCATGAGCGCCGCTCCCGCCTGC**CAGT-GAGTTCGAGCGACTCGAGT-3′ (bold characters indicate the region between *pntD* and *pntB*, and underlined characters correspond to the stop codon of *pntD*) and the above reverse primer *pen* for the construction of *ermEp-pntE-pntD* cassette. For *penE* expression, forward primer 5′-**GCTGACCACCCCACTCTCAG-CAAAAGGCGGCAACGCCC*ATG***CAGTGAGTTCGAGC-GACTCGAG T-3′ (bold characters indicate the region between *penD* and *penE*, underlined characters correspond to the stop codon of *penE*, and italic characters are the start codon of *penD*) and reverse primer *pen* 5′-**GGTCATGAGTTTCTCAACTTCC-CGTCTCTACAGATCTGTTG**CTGGTACCGAGCGAACG CGTT-3′ (bold characters indicate the downstream of *penA*) for the construction of the *ermEp-penE* cassette. For *penE-penD* expression, 5′-**CCGACCCCTGGCACGCGGAGGTCGCCGC-ATGAGCACCGCTC**CAGTG AGTTCGAGCGACTCGAG-T-3′ (bold characters indicate the region between *penB* and *penD*, and underlined characters correspond to the stop codon of *penD*) and reverse primer *pen* for the construct of *ermEp-penE-penD* cassette. Each pair of primers was used for the amplification of the *loxP-aad*(*3″*)*-loxP* segment using pKU473 as template DNA. The initial denaturation step (95 °C, 3 min) was followed by 25 cycles of amplification (95 °C, 30 s; 65 °C, 30 s; 72 °C, 60 s) and then a final incubation at 72 °C (10 min). Each amplified segment was introduced by electroporation into L-arabinose-induced *E. coli* BW25113 carrying the pKU464*aac*(*3*)*IV::ermEp::pnt*-cluster or

pKU464*aac*(*3*)*IV*::*ermEp*::*pen*-cluster. Transformants were selected by resistance to 25 μg/mL apramycin, 25 μg/mL fortimicin, 50 μg/mL streptomycin, and 100 μg/mL spectinomycin. To remove the resistance gene *aad*(*3″*), pTH19cs1::*cre*, which is a derivative of pSC101 in which *cre* is controlled by the *lacZ* promoter and whose replication is temperature-sensitive (*54*), was introduced into the transformants by the selection of chloramphenicol resistance (30 μg/mL). The chloramphenicol-resistant transformants were spread on LB agar containing 0.1 mM IPTG and 25 μg/mL apramycin, and the plates were incubated overnight at 37 °C to express *cre* and to cure pTH19cs1::*cre*. The plates were replicated onto LB agar containing 25 μg/mL apramycin, 50 μg/mL streptomycin, and 100 μg/mL spectinomycin and LB agar containing 25 μg/mL apramycin and 30 μg/mL chloramphenicol, respectively. After plates were incubated overnight at 30 °C, chloramphenicol-, streptomycin-, and spectinomycin-sensitive clones were selected. The desired recombinant plasmids were confirmed by restriction digestion to obtain pKU464*aac*(*3*)*IV*::*ermEp*::*pntE*, pKU464*aac*(*3*)*IV*::*ermEp*::*pntE*-*pntD*, pKU464*aac*(*3*)*IV*::*ermEp*::*penE*, and pKU464*aac*(*3*)*IV*::*ermEp*::*penE*-*penD*. To introduce each plasmid into the *S. avermitilis* deletion mutants, each recombinant plasmid was first introduced into *E. coli* GM2929 *hsdS*::Tn*10* to obtain unmethylated DNA preparations. Protoplasts of *S. avermitilis* SUKA16 Δ*ptlE* or Δ*ptlE* Δ*ptlD* were transformed by each unmethylated recombinant plasmid preparation using polyethylene glycol as described previously (*55*). Each transformant was selected by overlaying apramycin. Spores of each transformant were prepared from growth on YMS medium (*56*), and each spore suspension in 20% glycerol (v/v) was stored at −30 °C.

Analysis of Products from Complementation of the S. avermitilis Δ*ptlE* Δ*ptlD Double Deletion Mutants*. The cultivation of transformants was performed as previously described (*41*, *46*), and the products were isolated as previously described (*41*, *46*, *57*). For the analysis of the ratio of pentalenolactone D (**8**) and neopentalenolactone D (**9**) of the *S. avermtilis* SUKA16 Δ*ptlE* Δ*ptlD* double mutant carrying *pntE* or a *ptlE*/*pntE* hybrid gene, cultures of each transformant were centrifuged at 3000 rpm for 5 min to obtain the fermentation broth which was adjusted to pH 2.0 with 2 N HCl and extracted with chloroform. The organic layers were dried over anhydrous Na_2SO_4, concentrated, and methylated with TMS-CHN_2. The extracts were analyzed by GC-MS (method 2). For HPLC analysis, the culture extracts were subjected to ODS-HPLC (Pegasil 5 μm, 4.6ϕ × 250 mm) developed with 50% CH_3CN in water at a flow rate of 0.8 mL/min (detection at 210 nm).

Isolation of 1-Deoxy-11β-hydroxypentalenic Acid (***6***)*, Pentalenolactone D* (***8***)*, and Pentalenolactone E* (***15***). The metabolites 1-deoxy-11β-hydroxypentalenic acid (**6**) and pentalenolactone D (**8**) were each isolated as the derived methyl esters, as previously described, from large-scale cultures of the appropriate *S. avermitilis* SUKA16 mutant (*46*). After treatment of the acidic organic extracts of each culture with TMS-CHN_2, the resulting mixtures of methyl esters were purified chromatographically (preparative silica gel and ODS column chromatography) as previously described (*46*) to yield **6-Me** (from the *S. avermitilis* SUKA16 pKU462::*ermEp*-*ptl*-cluster Δ*ptlF* deletion mutant) and **8-Me** (from the *penE*-complemented *S. avermitilis* SUKA16 pKU462::*ermEp*-*ptl*-cluster Δ*ptlE*-Δ*ptlD*/*penE*$^+$ mutant). Pentalenolactone E methyl ester (**15-Me**) was isolated from *S. exfoliatus* UC5319 as previously described (*58*).

To obtain the corresponding free acids, individual solutions of **6-Me** (1.2 mg), **8-Me** (5 mg), **9-Me** (5 mg), and **15-Me** (1.5 mg) in 2 mL of 5% aqueous K_2CO_3 and 4 mL of methanol were heated at reflux for 24 h. After evaporation of the methanol via rotovap, the aqueous solution was acidified to pH 2.0 with 10% HCl and then extracted with chloroform. The combined organic phase was dried over anhydrous Na_2SO_4 and concentrated to give respectively 1.0 mg of 1-deoxy-11β-hydroxypentalenic acid (**6**), 4.0 mg of pentalenolactone D (**8**), 4.2 mg of neopentalenolactone D (**9**), and 1.2 mg of pentalenolactone E (**15**).

Construction of ptlE/*pntE Hybrid Genes*. Chimeric *ptlE* and *pntE* hybrids were prepared by the TAP method (*59*). The hybrid genes containing *ptlE* and *pntE* were constructed by PCR using overlapped primer pairs as follows: a part of the *ptlE* gene was amplified by PCR using template DNA from pKU462::*ptl*-cluster and the primer pairs, ptl-pnt_fwd0 forward, 5′-GGCCGGCCAT***ATG***GTGGATATCGAGGCAGTGA GAGCGAAGTACCGGGAGGAACGCGAC-3′ (bold characters correspond to the N-terminal region of *ptlE*, underlined characters indicate *Nde*I site, and italic characters correspond to the start codon of *ptlE*), and either ptl-pnt_rev300 reverse, 5′-**GTAGCTCTCCACGTCGCACCGGAC**CCCGGGAAAACG-GTTCCAGTACCAGGTCCC-3′ (bold characters indicate *pntE* at 295 to 318 nt, and underlined characters are *ptlE* at 277 to 306 nt), or ptl-pnt_rev600 reverse, 5′-**GGTGCTGTCGCCGCCG-GTGTAGCC**GAAATCCCAGCGGCTGGTGTGG AACG-3′ (bold characters indicate *pntE* at 637 to 660 nt, and underlined characters are *ptlE* at 623 to 648 nt), or ptl-pnt_rev1200 reverse, 5′-**CGCGGTGAGCCGTTCGACGCCCTG**TCC CTGGGTGTC-CACCAGGGTGA-3′ (bold characters indicate *pntE* at 1198 to 1221 nt, and underlined characters are *ptlE* at 1184 to 1206 nt). The corresponding 3′-portions of the *pntE* gene were amplified by PCR with template DNA from pKU464*aac*(*3*)*IV*::*ermEp*::*pntE* using the primer pairs, either ptl-pen_fwd300 forward, 5′-**TGGTACTGGAACCGTTTTCCCGGG**GTCCGGTGCGAC-GTGGAGAGCTACGTCTACATGCC-3′ (bold characters indicate *ptlE* at 283 to 306 nt, and underlined characters are *pntE* at 295 to 329 nt), or ptl-pnt_fwd600 forward, 5′-**TTCCACAC-CAGCCGCTGGGATTTC**GGCTACACCGGCGGCGACAG-CACCGG-3′ (bold characters indicate *ptlE* at 625 to 648 nt, and underlined characters are *pntE* at 637 to 662 nt), or ptl-pnt_fwd1200 forward, 5′-**GTCACCCTGGTGGACACCCAGGGA-**CAGGGCGTCGAACGGCTCACCGC-3′ (bold characters indicate *ptlE* at 1183 to 1206 nt, and underlined characters are *pntE* at 1198 to 1220 nt), and ptl-pnt_rev0 reverse, 5′-GCTCTA-GAGGTGGGGGACGG***TCA***GCGCAGTTCGAGGCCCGC-CAGG-3′ (bold characters indicate C-terminal region of *pntE*, underlined characters are *Xba*I site, and italic characters correspond to the stop codon of *pntE*). The initial denaturation step (95 °C, 3 min) was followed by 5 cycles of amplification (95 °C, 30 s; 50 °C, 30 s; 72 °C, 60 s) followed by 15 cycles using an annealing temperature of 65 °C and then a final incubation at 72 °C (10 min). Each amplicon was treated with T4 DNA polymerase in the presence of dNTPs to remove 3′-nucleotide overhangs at both ends. The second PCR amplification used template DNA from 50-fold dilution of both amplification mixtures (amplicons derived from ptl-pnt_fwd0/ptl-pnt_rev300 primer pair and ptl-pnt_fwd300/ptl-pnt_rev0 primer pair, ptl-pnt_fwd0/ptl-pnt_rev600 primer pair and ptl-pnt_fwd600/ptl-pnt_rev0 primer pair, or ptl-pnt_fwd0/ptl-pnt_rev1200 primer pair and ptl-pnt_fwd1200/ptl-pnt_rev0 primer pair) and Phusion DNA polymerase using 10-fold concentrated primer pair,

ptl-pnt_fwd0 and ptl-pnt_rev0. The initial denaturation step (95 °C, 3 min) was followed by 20 cycles of amplification (95 °C, 30 s; 65 °C, 30 s; 72 °C, 100 s) and then a final incubation at 72 °C (10 min). Each amplified fragment was digested with *Nde*I and *Xba*I, and the resulting fragment was ligated with the large *Nde*I/*Xba*I fragment of pKU460*aac*(*3*)*IV::ermEp*. After confirmation of the sequence of each hybrid gene, the desired recombinant plasmids (pKU460*aac*(*3*)*IV::ermEp::ptlE*/*pntE300*, pKU460*aac*(*3*)*IV::ermEp::ptlE*/*pntE600*, and pKU460*aac*(*3*)*IV::ermEp::ptlE*/*pntE1200*, respectively) were propagated in *E. coli* GM2929 *hsdS*::Tn*10* to prepare unmethylated DNA. The *S. avermitilis* SUKA16 Δ*ptlE* Δ*ptlD* double deletion mutant was transformed by PEG-mediated protoplast transformation using the unmethylated DNA preparation (*55*).

RESULTS

Pentalenolactone Biosynthetic Gene Clusters of S. exfoliatus UC5319 and S. arenae TU469. To isolate the pentalenolactone biosynthetic gene clusters of the known pentalenolactone producers *S. exfoliatus* UC5319 and *S. arenae* TU469, we used probes based on the previously cloned *S. exfoliatus* pentalenene synthase gene and the *S. arenae gapR* resistance gene to screen DNA libraries obtained from the corresponding parent organisms. Thus PCR screening of an *S. arenae* TU469 cosmid library with a primer pair based on the sequence of the *S. arenae gapR* gene led to isolation of six candidate cosmids that together were found to harbor the 13-kb *pnt* cluster encoding 11 ORFs (Figure 1B). Similarly, screening of an *S. exfoliatus* UC5319 plasmid library with a 601-bp PCR probe derived from the internal sequence of the pentalenene synthase gene allowed isolation of plasmid p56 harboring a 7-kb insert that included the complete pentalenene synthase gene, *penA*. PCR screening of an *S. exfoliatus* cosmid library then served to identify a single cosmid G21 harboring the entire ~13-kb *pen* biosynthetic cluster consisting of 11 ORFs that were identical in organization to those in the *pnt* biosynthetic cluster (Figure 1C).

Analysis of the sequence of the *S. exfoliatus pen* biosynthetic gene cluster (Figure 1C) revealed that the furthest upstream gene, designated *penR*, encodes a 153 aa MarR-family transcriptional regulator with strong similarity to the deduced SAV_2989 protein that is found just upstream of the previously characterized *gap1* (*sav2990*) pentalenolactone resistance gene of *S. avermitilis* (Table 1). All but 1 of the remaining 10 unidirectionally transcribed ORFs in the *pen* cluster correspond closely in gene organization and deduced amino acid sequence to the corresponding ORFs of the previously characterized *S. avermitilis ptl* cluster. Thus the *S. exfoliatus gapN* gene has a high level of similarity to *S. avermitilis gap1* that has been shown to encode a pentalenolactone-insensitive GAPDH. The downstream 8 ORFs from *penH* to *penI* have the same gene order as *ptlH* to *ptlI* and exhibit 65−78% identity and 76−87% similarity at the deduced amino acid level to the corresponding *ptl* gene products. Only the *penM* gene, which encodes a predicted 398 aa cytochrome P450 monooxygenase, has no homologue in the *ptl* cluster. In addition to PenA, the previously characterized *S. exfoliatus* pentalenene synthase, PenI is assigned as a cytochrome P450 monooxygenase, PenH as a Fe^{2+}-α-ketoglutarate-dependent hydroxylase, and PenF as an NAD^+-dependent dehydrogenase, based on their close matches to the corresponding PtlI, PtlH, and PtlF proteins, each of established biochemical function (*42*−*45*). Together, PenI, PenH, and PenF are expected to catalyze the oxidative conversion of pentalenene (**3**) to 1-deoxy-11-oxopentalenic acid (**7**), as previously established for the othologous *S. avermitilis* proteins (Scheme 1). The *penB* gene is predicted to encode a polyprenyl diphosphate synthase, based on its similarity to *ptlB* and other prenyltransferase-encoding genes, while *penG* encodes an apparent transmembrane efflux protein, by analogy to the predicted function of *ptlG*. Interestingly, the *S. exfoliatus pen* cluster carries no genes homologous to the *S. avermitilis* genes *ptlC*, *ptlR*, *ptlJ*, and *ptlL* which have as yet no assigned function. Most importantly, the *S. exfoliatus penE* gene is predicted to encode a 584 aa protein that is a homologue of the known Baeyer−Villiger monooxygenase from *S. avermitilis*, PtlE (70% identity and 81% similarity), while *penD* encodes a 298 aa protein that is a presumptive Fe^{2+}-α-ketoglutarate-dependent dioxygenase with 65% identity and 82% similarity to the *S. avermitilis* enzyme PtlD.

The 11 ORFs of the 13-kb *S. arenae pnt* pentalenolactone biosynthetic gene cluster are identical in organization to those of the closely related *pen* cluster (Figure 1). Thus the divergently transcribed *pntR* gene encodes a 153 aa MarR-family transcriptional regulator with 80% identity and 87% similarity to PenR and 73% identity and 85% similarity to *S. avermitilis* SAV_2989 (Table 1). Besides the previously described *gapR* resistance gene encoding a pentalenolactone-insensitive GAPDH, the remaining 9 ORFs from PntM to PntI correspond closely in amino acid sequence to the orthologous ORFs of the *pen* cluster, with levels of amino acid sequence identity from 81% to 93% and similarity from 87% to 96%. Interestingly, the levels of identity and similarity between the orthologous Pnt and Pen proteins are higher by about 15−24% and 8−15%, respectively, than the corresponding relationships between the individual Pnt or Pen proteins and the homologous proteins of the *ptl* cluster. No matches were found in the *pnt* cluster to any of the predicted proteins encoded by the *ptlC*, *ptlR*, *ptlJ*, and *ptlL* genes of *S. avermitilis*.

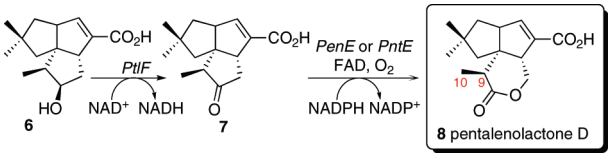
Biochemical Characterization of the PenE and PntE Baeyer−Villiger Monooxygenases. With the full sequences in hand for the individual pentalenolactone biosynthetic gene clusters from the two pentalenolactone producers, *S. exfoliatus* UC5319 and *S. arenae* TU469, we turned our attention to the enzymatic Baeyer−Villiger oxidation of the common intermediate 1-deoxy-11-oxopentalenic acid (**7**), the key branch point that distinguishes the pentalenolactone and neopentalenolactone biosynthetic pathways. A synthetic *penE* gene with codons optimized for *E. coli* was inserted into pET-28a and expressed in *E. coli* BL21(DE3) to give recombinant PenE carrying an N-terminal His_6 tag. Purification by metal ion affinity chromatography using a Ni^{2+}-NTA resin gave recombinant PenE protein, >95% pure by SDS−PAGE (Supporting Information Figure S1). PenE carried a bound FAD cofactor, as deduced from the characteristic UV maxima at 375 and 450 nm. MALDI-TOF MS analysis of His_6 tag-PenE gave the expected M_D value for the parent flavin-free protein lacking the N-terminal Met. In similar manner, a synthetic *pntE* gene was inserted into pET-26b, and the resultant recombinant PntE with an appended C-terminal His_6 tag was purified by immobilized metal ion affinity chromatography. The purified recombinant PntE-His_6 tag protein, >95% pure by SDS−PAGE (Supporting Information Figure S1), carried a bound FAD (λ_{max} 380 and 450 nm) and exhibited the expected M_D values for the cofactor-free protein by both MALDI-TOF and ESI-MS.

Table 1: Comparison of Predicted Proteins of the *ptl*, *pen*, and *pnt* Biosynthetic Gene Clusters

Ptl (aa)[a]	Pen (aa, % identity/ similarity to Ptl)	Pnt (aa, % identity/ similarity to Ptl)	Pen:Pnt (% identity/similarity)	known or predicted function
Sav_2989 (161)	PenR (153, 75/88)	PntR (151, 73/85)	82/89	predicted MarR family transcriptional regulator
Gap1 (334)	GapN (336, 63/76)	GapR (334, 88/92)	60/75	glyceraldehyde-3-phosphate dehydrogenase
	PenM (398)	PntM (398)	81/87	predicted cytochrome P450 monooxygenase
PtlH (285)	PenH (283, 73/83)	PntH (283, 73/84)	89/94	1-deoxypentalenic acid 11β-hydroxylase
PtlG (484)	PenG (484, 65/76)	PntG (484, 65/77)	84/90	predicted transmembrane efflux protein
PtlF (270)	PenF (275, 69/80)	PntF (282, 70/80)	91/94	1-deoxy-11β-hydroxypentalenic acid dehydrogenase
PtlE (594)	PenE (584, 70/81)	PntE (589, 69/82)	93/96	Baeyer−Villiger monooxygenase
PtlD (306)	PenD (298, 65/82)	PntD (299, 64/81)	86/93	predicted dioxygenase
PtlC (58)				hypothetical protein
PtlB (337)	PenB (337, 78/84)	PntB (337, 76/83)	93/94	predicted polyprenyl diphosphate synthase
PtlA (336)	PenA (337, 76/86)	PntA (337, 76/87)	91/95	pentalenene synthase
PtlI (449)	PenI (463, 74/87)	PntI (457, 74/86)	89/95	cytochrome P450 monooxygenase
PtlR (153)				putative AraC-family transcriptional regulator
PtlJ (144)				putative lyase
PtlL (256)				hypothetical protein

[a]For annotations of the *ptl* cluster, see cited papers as well as refs *60* and *61* and the *S. avermitilis* Genome Project Web site (http://avermitilis.ls.kitasato-u.ac.jp/metabolite/).

Scheme 2: PenE- and PntE-Catalyzed Baeyer−Villiger Oxidation of **7** to Pentalenolactone D (**8**)

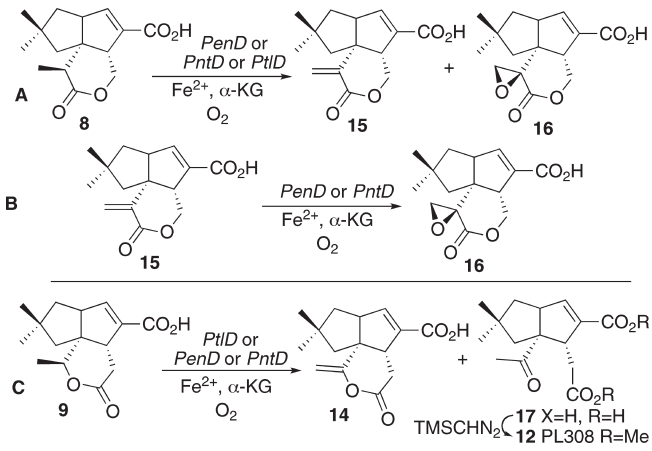


To identify the heretofore cryptic enzymatic activity of PenE and PntE, each recombinant enzyme was incubated with 1-deoxy-11-oxopentalenic acid (**7**) (Scheme 2). The requisite substrate **7** was generated *in situ* from 1-deoxy-11β-hydroxypentalenic acid (**6**) using recombinant PtlF dehydrogenase, due to the tendency of **7** to undergo facile epimerization at C-9 during attempted purification. The reduced flavin cofactor was generated continuously with excess NADPH and catalytic FAD. Treatment of the organic extracts with TMS-CHN_2 followed by capillary GC-MS analysis of the derived methyl esters showed in each case exclusive formation of the Baeyer−Villiger oxidation product, pentalenolactone D methyl ester (**8-Me**) (Scheme 2), which was identical by direct comparison with an authentic sample of **8-Me** in both retention time and mass spectrum (Supporting Information Figure S2). Control incubations with boiled PenE or PntE gave only unoxidized **7-Me** and 9-*epi*-**7-Me**. Neopentalenolactone D methyl ester (**9-Me**) could not be detected in any of the incubation mixtures.

Biochemical Characterization of the Fe^{2+}-α-Ketoglutarate-Dependent Dioxygenases PenD, PntD, and PtlD. Synthetic genes for *penD*, *pntD*, and *ptlD*, each optimized for expression in *E. coli*, were individually cloned into pET-28a, and the derived expression vectors were used to transform *E. coli* BL21(DE3). The resulting recombinant PenD, PntD, and PtlD proteins, each carrying an N-terminal His_6 tag, were each purified to >95% purity by Ni-NTA chromatography (Supporting Information Figure S1). Each of the three proteins exhibited the predicted molecular mass M_D upon ESI-MS analysis.

Individual incubations of recombinant PenD or PntD with pentalenolactone D (**8**) in the presence of Fe^{2+} and α-ketoglutarate, followed by treatment of the organic products with TMS-CHN_2, gave predominantly the double oxidation product

Scheme 3: Oxidation of Pentalenolactone D (**8**) and Neopentalenolactone D (**9**): (A) Conversion of Pentalenolactone D (**8**) to Pentalenolactone E (**15**) and Pentalenolactone F (**16**) Catalyzed by PenD, PntD, or PtlD; (B) Epoxidation of **15** to **16** Catalyzed by PenD or PntD; (C) Oxidation of Neopentalenolactone D (**9**) Catalyzed by PtlD, PenD, or PntD



pentalenolactone F methyl ester (**16-Me**) accompanied by small quantities of pentalenolactone E methyl ester (**15-Me**) (Scheme 3A and Supporting Information Figure S3). Each of the products was identical to authentic reference standards in both retention time and mass spectrum by direct GC-MS comparison. Under the incubation conditions, the substrate was consumed within 90 min. Control incubations using boiled PenD or PntD gave only recovered (**8-Me**). Incubation of pentalenolactone E (**15**) with either PenD or PntD followed by treatment with TMS-CHN_2 gave pentalenolactone F methyl ester (**16-Me**), as established by GC-MS analysis and comparison with authentic **16-Me** (retention time 12.84 min, M^+ *m*/*z* 292) (Scheme 3B, Supporting Information Figure S3).

The natural substrate for the corresponding *S. avermitilis* dioxygenase, PtlD, is expected to be the isomeric lactone neopentalenolactone D (**9**), as suggested by the accumulation of **9** in *S. avermitilis* mutants lacking the *ptlD* gene, as well as the observed formation of **9** by PtlE-catalyzed Baeyer−Villiger oxidation of the common cyclopentanone intermediate **7** (*46*)

(Scheme 1). Very interestingly, we found that recombinant *S. avermitilis* PtlD could also catalyze the two-step oxidation of its unnatural substrate pentalenolactone D (**8**) to pentalenolactone F (**16**), as established by GC-MS analysis of the derived methyl esters (Scheme 3A and Supporting Information Figure S4). Together, these results establish that PtlD, PenD, and PntD not only share a high level of mutual sequence similarity but utilize a common mechanism of action as well, in spite of the isomeric structures of their native substrates.

To examine directly the proposed native biochemical function of PtlD, we incubated recombinant PtlD with its natural substrate **9** in the presence of Fe^{2+} and α-ketoglutarate (Scheme 3C). After 90 min incubation, extraction with dichloromethane and methylation of the crude organic extract, the major product was PL308 (**12**), which was identical in retention time and mass spectrum (retention time 11.92 min, *m*/*z 308*) by direct capillary GC-MS comparison with authentic **12** (*46*) (Supporting Information Figure S4). Although neither the more highly oxidized PL324 (**13**, 10-hydroxy-PL308) nor the closely related ketolactone **10-Me** could be detected by GC-MS analysis of the resulting methylated organic extracts, an unstable coproduct, *m*/*z* 276 (retention time 12.10 min), could be isolated which has tentatively been assigned the structure of neopentalenolactone E methyl ester (**14-Me**). The same mixture of PL308 (**12**) and **14-Me** was also generated upon incubation of neopentalenolactone D (**9**) under identical conditions with either recombinant PenD or PntD. To confirm the structures of the two neopentalenolactone D-derived oxidation products, we carried out a preparative scale incubation of **9** with recombinant PtlD and examined the individual products by ^{1}H NMR after column chromatographic separation of the derived methyl esters. The major component was confirmed to be PL308 (**12**), as established by direct comparison with the spectrum of authentic **12** (*46*). The second component, **14-Me** (*m*/*z* 276), exhibited several key ^{1}H NMR signals strongly supporting the proposed neopentalenolactone E methyl ester structure. Besides the methyl ester singlet at δ 3.74, a characteristic pair of upfield-shifted olefinic singlets at δ 4.88 (H-10*syn*) and 4.73 (H-10*anti*) indicated the presence of the enol lactone (*62*), while the conjugated olefinic proton signal at δ 6.81 (H-7) is typical of all pentalenolactone metabolites (*37*). Importantly, a proton signal at δ 4.39 corresponding to the saturated H-9 carbinyl proton of neopentalenolactone D methyl ester (**9-Me**) was entirely absent from the NMR spectrum of **14-Me**. Due to the intrinsic chemical instability of the isolated submilligram quantities of **14-Me** the remaining signals were insufficiently intense to allow complete assignment of the proposed structure or were obscured by residual solvent or other impurities. In a GC-MS time-course study, after 10 min incubation with PtlD, 23% of the neopentalenolactone D substrate had been consumed, with 15% formation of the seco acid **17** corresponding to PL308 and 8% neopentalenolactone E. After 45 min, the products consisted of an equal mixture of PL308 and neopentalenolactone E, with only 2% of residual **9**. Finally, after 90 min, the fraction of PL308 increased to 73% while the proportion of neopentalenolactone E fell to 26%, consistent with the proposed PtlD-catalyzed oxidation of **9** to the unstable enollactone **14** followed by its nonenzymatic hydrolysis to **17**.

Characterization of the penD and pntD Deletion Mutants. To investigate the *in vivo* role of the *penD* and *pntD* genes, we generated the corresponding in-frame *S. exfoliatus*/Δ*penD* ZD20 and *S. arenae*/Δ*pntD* ZD18 deletion mutants, using PCR-targeted *Streptomyces* gene replacement. To this end, an internal

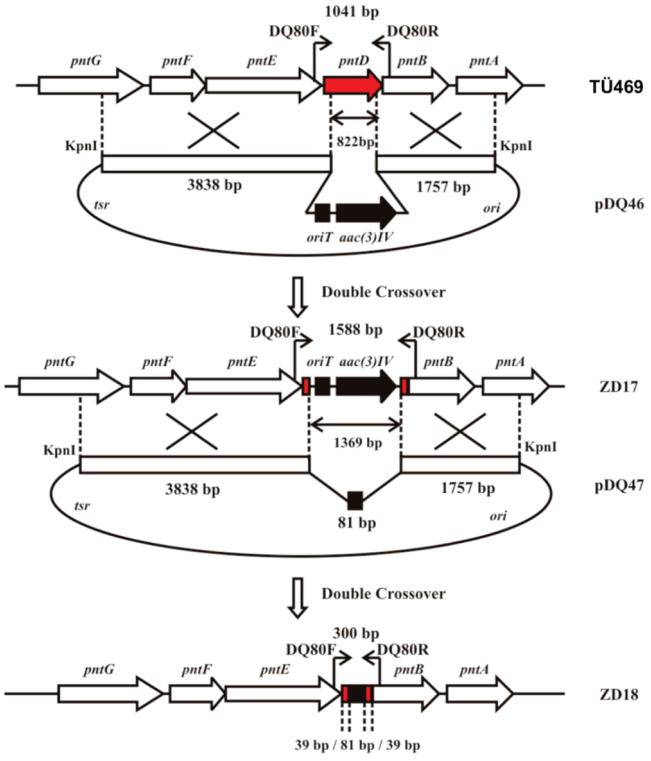


FIGURE 4: Construction of Δ*pntD* deletion mutant *S. arenae* ZD18. See Supporting Information Figure S5.

819-bp fragment of *penD* carried in a 7-kb segment of the *pen* cluster was first replaced by DNA from *oriT* and *aac(3)IV*, most of which was then deleted by digestion with *Xba*I and religation to leave an in-frame 81-bp DNA scar within the orginal *penD* gene (Figure 3). The derived plasmid, pDQ51, was then conjugated into *S. exfoliatus* UC5319, and single crossover exconjugants were selected with thiostrepton. Double crossover mutant strains were then obtained by growth in the absence of antibiotic, and the desired Δ*penD* deletion mutant, *S. exfoliatus* ZD20, was identified by PCR screening using primer pairs based on the remaining upstream and downstream portions of *penD* (Supporting Information Figure S5).

In like manner, a 822-bp fragment of *pntD* harbored within a 6.4-kb segment of the *pnt* cluster was replaced by a 81-bp scar (Figure 4). The desired in-frame Δ*pntD* deletion mutant, *S. arenae* ZD18, was then obtained by two successive rounds of homologous recombination, using PCR to screen the exconjugants and to confirm the targeted deletion (Supporting Information Figure S5).

Wild-type *S. exfoliatus* UC5319 and *S. arenae* TU469 and the corresponding Δ*penD* and Δ*pntD* deletion mutants, *S. exfoliatus* ZD20 and *S. arenae* ZD18, were each grown in liquid production cultures at 30 °C for 6 days. After acidification and chloroform extraction, analysis of the derived mixture of methyl esters by capillary GC-MS confirmed that each of the wild-type parent strains produced pentalenolactone (**1**) as well as varying proportions of pentalenolactones D (**8**), E (**15**), and F (**16**). By contrast, neither *S. exfoliatus* ZD20 (Δ*penD*) nor *S. arenae* ZD18 (Δ*pntD*) produced any detectable pentalenolactone (**1**) or either of the late-stage intermediates **15** and **16**. Instead, both deletion mutants accumulated the Baeyer−Villiger oxidation product pentalenolactone D (**8**), the demonstrated *in vitro* substrate of both PenD and PntD (Figure 5, Supporting Information Figure S6). As expected, neither the two wild-type strains nor the Δ*penD* and

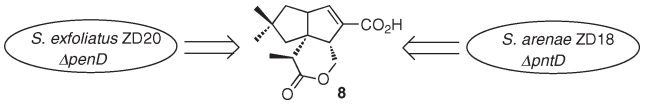


FIGURE 5: Production of pentalenolactone D (**8**) by Δ*penD* deletion mutant *S. exfoliatus* ZD20 and Δ*pntD* deletion mutant *S. arenae* ZD18.

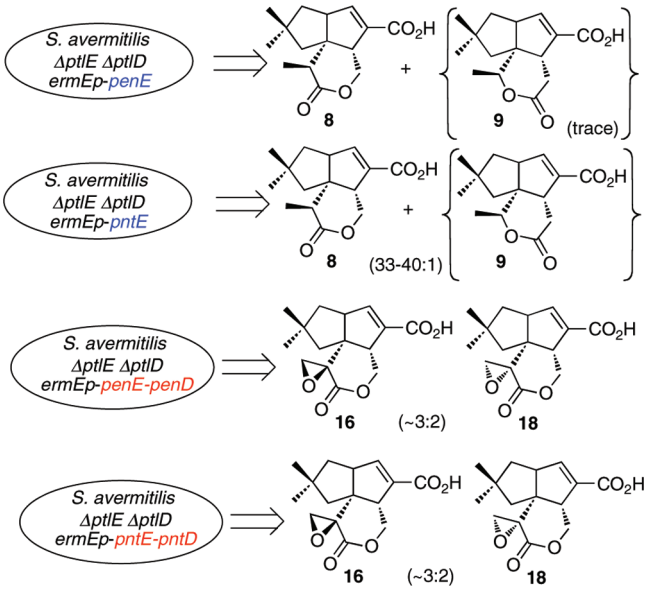


FIGURE 6: Complementation of *S. avermitilis* Δ*ptlE*Δ*ptlD* deletion mutants.

Δ*pntD* deletion mutants produced any detectable neopentalenolactone D (**9**).

Complementation of the S. avermitilis ΔptlE ΔptlD Deletion Mutant with penE, pntE, penE-penD, and pntE-pntD. The previously described *S. avermitilis* SUKA16 Δ*ptlE* Δ*ptlD* double deletion mutant accumulates 1-deoxy-11-oxopentalenic acid (**7**) (Figure 2) but not neopentalenolactone D (**9**) (*46*). In the wild-type strain, **7** serves as the natural substrate for the PtlE-catalyzed Baeyer−Villiger oxidation to **9** (*46*). To test the *in vivo* function of the corresponding Baeyer−Villiger oxygenase genes from the pentalenolactone biosynthetic clusters, *penE* and *pntE* under control of the constitutive *ermEp* promoter were each introduced by protoplast transformation into the *S. avermitilis* Δ*ptlE* Δ*ptlD* double deletion mutant. Complementation with either *penE* or *pntE* resulted in formation of pentalenolactone D (**8**), as established by GC-MS analysis of the methylated organic extracts of the corresponding transformants (Figure 6, Supporting Information Figures S7 and S8). Both GC-MS and LC-MS analysis of a concentrated sample from cultures of the *S. avermitilis* double deletion mutant complemented with *pntE* (*S. avermitilis ermEp-ptl*-cluster Δ*ptlE*Δ*ptlD ermEp-pntE*) also revealed the presence of a very low level of the isomeric Baeyer−Villiger product neopentalenolactone D (**9**) as no more than 2−3% of the mixture (Supporting Information Figure S8). Trace amounts of **9** were also observed in the culture of the *S. avermitilis* double deletion mutant complemented with *penE*.

We also complemented the *S. avermitilis* double deletion mutants with both *penE* and *penD* or *pntE* and *pntD*, respectively, again under control of the *ermE* promoter. GC-MS analysis of the derived methyl esters and comparison with authentic standards established that the resulting transformants produced the expected product pentalenolactone F (**16**) resulting from two-step PenD- or PntD-catalyzed desaturation and epoxidation of **8**, accompanied by the isomeric epoxylactone, 9,10-*epi*-pentalenolactone F (**18**) (Figure 6, Supporting Information Figures S7, S7-2, and S9). The observed ratio of **16** to **18** was ~3:2. The complementation by *penE-penD* was less efficient, and only very small amounts of **16** and **18** were detected as the derived methyl esters. Notably, the intermediate pentalenolactone E (**15**) that was observed in the enzymatic reaction of PtlD, PenD, and PntD with **8** could not be detected in the extracts of the mutants complemented with either *penE-penD or pntE-pntD*.

In Vivo Analysis of Baeyer−Villiger Products Generated by Hybrid PtlE/PntE Proteins. Although the amino acid sequences of PtlE and PntE were extremely similar, each enzyme catalyzed a distinct regiospecific Baeyer−Villiger reaction. To explore the protein structural basis for this difference in function, we constructed a progressive series of three different *ptlE*/*pntE* hybrid genes, in which the crossover points were engineered by the TAP method between the NADPH-binding and FAD-binding motifs (91−102 aa; *ptl-pntE300*), inside the FAD-binding motif (206−216 aa; *ptlE*/*pntE600*), and between the FAD-binding motif and C-terminus region (395−407 aa; *ptlE*/*pntE1200*), respectively (Supporting Information Figure S10) (*59*). Each hybrid gene was expressed under the control of the *ermE* promoter in *S. avermitilis* Δ*ptlE* Δ*ptlD* double mutants. Transformants carrying *ptlE*/*pntE300* or *ptlE*/*pntE600* produced both pentalenolactone D (**8**) and neopentalenolactone D (**9**). The production of **9** in the *S. avermitilis* Δ*ptlE* Δ*ptlD* double mutant carrying *ptlE*/*pntE300* increased slightly in comparison to that of the double mutant carrying *pntE* alone, with a ratio of **8** to **9** of ca. 20:1. The double mutant carrying *ptlE*/*pntE600* produced increased levels of **9**, resulting in a ratio of **8** to **9** of ca. 5:1. By contrast, **8** was not observed at all in the extracts of the double mutant carrying *ptlE*/*pntE1200*, while the overall productivity of this latter transformant was reduced (Supporting Information Figure S11).

DISCUSSION

Pentalenolactone Biosynthetic Gene Clusters. The 11 ORFs of the *pen* and *pnt* pentalenolactone biosynthetic gene clusters of *S. exfoliatus* and *S. arenae* are not only identical in organization but exhibit a very high degree of mutual sequence identity (Figure 1 and Table 1). Both the pentalenolactone and the neopentalenolactone biosynthetic pathways involve the multistep conversion of FPP (**2**) to a common intermediate, 1-deoxy-11-oxopentalenic acid (**7**), mediated in turn by the orthologous PenB/PntB/PtlB, PenA/PntA/PtlA, PenI/PntI/PtlI, PenH/PntH/PtlH, and PenF/PntF/PtlF gene products (Scheme 1). At this point the two pathways branch. PenE and PntE each catalyze the Baeyer−Villiger oxidation of **7** to pentalenolactone D (**8**) (Schemes 1 and 2) while PtlE catalyzes the complementary oxidation of **7** to the isomeric lactone, neopentalenolactone D (**9**) (Scheme 1).

A BLASTP search of the NCBI nonredundant (nr) protein database with the *S. exfoliatus* PenA sequence revealed that the predicted SBI_09679 protein (UniProt ID D7CBC0; GenBank ADI12797.1) from the recently reported genome sequence of *Streptomyces bingchenggensis* BCW-1 (*63*) corresponds to a pentalenene synthase with a high degree of certainty. In addition to the convincing 91% identity and 96% similarity between the predicted SBI_09679 gene product and the amino acid sequence of PenA, SBI_09679 is also found within a cluster

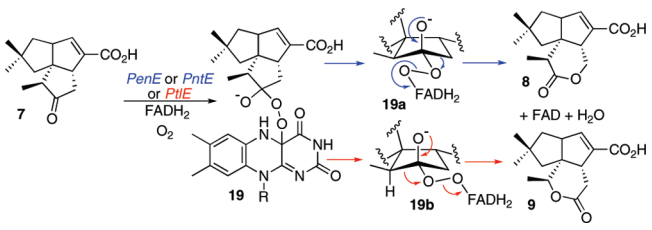
of 9 unidirectionally transcribed ORFs (GenBank ADI2790.1−ADI2798.1) that is identical in organization to both the *pen* and *pnt* biosynthetic gene clusters, with typically >90% pairwise predicted amino acid sequence identity and similarity to the orthologous *pen* or *pnt* proteins (Figure 1D). Interestingly, the individual ORFs of the predicted *S. bingchenggensis* pentalenolactone biosynthetic cluster more closely resemble those of the validated pentalenolactone producers, *S. exfoliatus* and *S. arenae*, than they do the corresponding ORFs of the *S. avermitilis* neopentalenolactone *ptl* cluster. The only significant differences in the organization of the presumptive *S. bingchenggensis* biosynthetic cluster from that of the corresponding *pen*, *pnt*, and *ptl* biosynthetic gene clusters are the absence of an upstream MarR-like *penR*/*pntR*/*sav2989* homologue and the fact that the apparent pentalenolactone resistance gene *SBI_00942* (UniProt D7C675, GenBank ADI04063.1), with 64−68% amino acid sequence identity and 77−80% similarity to Gap1, GapR, and GapN, is separated by some 2.0 Mb from the pentalenolactone biosynthetic cluster in the 11.9-Mb *S. bingchenggensis* linear genome. Although *S. bingchenggensis* is known to be a prolific producer of a wide variety of natural products, including several anthelminthic milbemycins, the polyether ionophore nanchangmycin, and the cyclic bingchamide pentapeptides (*63*), there are no reports of the isolation of pentalenolactone from this organism.

The BLASTP search also turned up nearly a dozen additional bacterial terpene synthases that have been erroneously annotated as "pentalenene synthase" in spite of the fact that they have much lower overall sequence similarity (<35%) to any of the validated pentalenene synthases. For example, the recently reported genome sequence of *Streptomyces clavuligerus* ATCC 27064 alone contains seven such misidentified "pentalenene synthase" genes (*64*, *65*). Additional incorrectly attributed "pentalenene synthases" can be found in the reported genome sequences of *Streptomyces hygroscopicus* ATCC 53653, *S. bingchenggensis* BCW-1, *Streptomyces flavogriseus* ATCC 33331, and *Sorangium cellulosum* "So ce 56" as well as the fungus *Aspergillus flavus* NRRL3357. Besides the obviously low level of overall pairwise similarity to either of the previously reported pentalenene synthase sequences for PenA or PtlA, the two apparent Mg^{2+}-binding domains in all of these predicted proteins also deviate significantly from the highly conserved DDFLD and NDIASLEKE motifs found only in the three bona fide pentalenene synthases, PenA, PtlA, and PntA, as well as SBI_09679. Of equal importance, none of the numerous missannotated "pentalenene synthases" are located within biosynthetic gene clusters containing orthologues of any of the 10 remaining functional ORFs that are found in the *pen*, *pnt*, and *ptl* biosynthetic gene clusters or the presumptive pentalenolactone biosynthetic cluster of *S. bingchenggensis*. These erroneously inferred electronic annotations might originally have been due to the fact that, until relatively recently, the *S. exfoliatus* pentalenene synthase was the only bacterial terpene synthase sequence of experimentally assigned biochemical function. Unfortunately, such annotation errors, once they are in the public databases, are readily propagated and will be difficult to correct in uncurated databases in the absence of agreed standards for annotation of terpene synthases of unknown function (*66*, *67*).

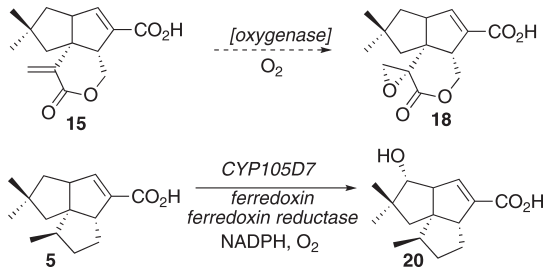
The role of the both PenE and PntE in mediating the Baeyer−Villiger oxidation of 1-deoxy-11-oxopentalenic acid (**7**) to pentalenolactone D (**8**) is conclusively supported by three independent but complementary lines of evidence: (1) the demonstrated flavin-dependent enzymatic conversion of **7** to **8** catalyzed by both recombinant PenE and PntE (Scheme 2); (2) the accumulation of **8** by the in-frame deletion mutants of *S. exfoliatus* and *S. arenae* lacking *penD* or *pntD*, respectively, and the concurrent abolition of formation of pentalenolactone (**1**) and both of the late-stage intermediates **15** and **16** (Figure 6); and (3) the production of the heterologous natural product pentalenolactone D (**8**) by *S. avermitilis* deletion mutants lacking both *ptlE* and *ptlD* that have been complemented by either *penE* or *pntE* (Figure 6). PenE and PntE should therefore be considered as paralogues of *S. avermitilis* PtlE, which catalyzes the analogous Baeyer−Villiger oxidation of the common substrate **7** to a distinct product, the isomeric lactone, neopentalenolactone D (**9**) (*46*). Each of the three flavin-dependent monooxygenases, PenE, PntE, and PtlE, catalyze the committed step at the branch point of the pentalenolactone and neopentalenolactone pathways. Notably, all three Baeyer−Villigerases are highly regiospecific with respect to product formation, generating no more than minor amounts (2−3% or less) of the unnatural isomeric lactone. Detailed sequence comparisons do not reveal any obvious residues that can account for the differences in product specificity. Interestingly, the 70% levels of pairwise identity and 80% similarity between either PenE or PntE and the paralogous oxygenase PtlE are not meaningfully distinguishable from the comparable levels of identity (69−73%) and similarity (80−84%) between the orthologous sets of proteins PenH or PntH compared to PtlH that each catalyze identical biochemical reactions on identical substrates or from the comparable levels of identity between the PenF or PntF proteins and PtlF. Nonetheless, by constructing a series of hybrid *ptlE-pntE* genes and *in vivo* complementation of the *S. avermitilis* Δ*ptlE* Δ*ptlD* double mutant with these *ptlE*/*pntE* hybrids, we have provided important clues regarding the protein structural basis for the regiospecificity of the individual Baeyer−Villiger reactions. The two transformants carrying either *ptlE*/*pntE300* or *ptlE*/*pntE600* each produced both neopentalenolactone (**8**) and its isomer pentalenolactone D (**9**). The ratio of **8** to **9** was only slightly perturbed in the former transformant compared to wild type while the productivity of **9** was dramatically increased in the *ptlE*/*pntE600* transformant. By contrast, the transformant carrying *ptlE*/*pntE1200* produced only **9**. These results indicate that the N-terminal region of these Baeyer−Villigerases may influence the regiospecificity of the Baeyer−Villiger oxidation reaction, especially the region around the FAD-binding motif.

Baeyer−Villiger Monooxygenases. The majority of known or predicted biochemical Baeyer−Villiger monooxygenases (BVMOs) appear to be opportunistic enzymes that catalyze catabolic degradation of exogenous ketone substrates such as cyclohexanone, cyclopentanone, cyclododecanone, or 4-hydroxyacetophenone to the corresponding lactones or esters with very high chemo-, regio-, and stereospecificity (*68*−*70*). The PtlE, PenE, and PntE proteins, which are class B BVMOs characterized by a tightly bound FAD cofactor and a dependence on external NADPH (*71*), are among the first such enzymes that have been demonstrated to play a role in a specific biosynthetic pathway and for which the natural substrate has been defined (*46*, *72*). All three proteins harbor several conserved BVMO fingerprint motifs, including **FXGXXXHXXXW**(D/P) (*68*). The enzymatic formation of **8** or **9** is presumably controlled by differences in the conformation of the peroxide bond in the common Criegee-type ketone−4a-hydroperoxyflavin adduct **19**, leading to migration of either the methylene or methane carbon of **7** (*70*, *73*−*75*) (Scheme 4). Although we detected minor

Scheme 4: Baeyer−Villiger Oxidation of **7** to Pentalenolactone D (**8**) and Neopentalenolactone D (**9**) by Fragmentation of Discrete Conformers of the Common Criegee Adduct **19**



Scheme 5: Formation of Shunt Metabolites 9,10-*epi*-Pentalenolactone F (**18**) and Pentalenic Acid (**20**)



amounts (2−3%) of neopentalenolactone D (**9**) along with the major product **8** in culture extracts of *S. avermitilis* double deletion mutants that had been complemented with *pntE*, we could not detect this minor isomer in the GC-MS analysis of the *in vitro* incubations with recombinant PenE or PntE, possibly due to the substantially smaller quantities of lactone that were analyzed in these experiments.

Desaturation−Epoxidation Catalyzed by the Fe^{2+}*-α-Ketoglutarate-Dependent Dioxygenases PenD, PntD, and PtlD*. The role of PenD and PntD in the conversion of pentalenolactone D (**8**) to the epoxylactone **16** is also supported by three interlocking lines of experimental evidence: (1) direct demonstration that both PenD and PntD catalyze the two-step, Fe^{2+}-α-ketoglutarate-dependent, oxidation of **8** to **16** through the intermediacy of the exomethylenelactone, pentalenolactone E (**15**) (Scheme 3A); (2) the accumulation of **8** and the absence of pentalenolactone (**1**) as well as both **15** and **16** in cultures of the in-frame deletion mutants *S. exfoliatus* ZD20 and *S. arenae* ZD18 that lack *penD* or *pntD*, respectively (Figure 5); and (3) the production of pentalenolactone F (**16**) by *S. avermitilis* deletion mutants lacking both *ptlE* and *ptlD* that have been complemented by *penE* plus *penD* or by *pntE* plus *pntD* (Figure 6).

Accompanying **16** in both of the latter *S. avermitilis* mutant complementation experiments is the stereoisomeric epoxide metabolite, 9,10-*epi*-pentalenolactone F (**18**). Although **18** has previously been isolated from cultures of wild-type *S. exfoliatus* UC5319 as a cometabolite of **16** (*37*, *38*, *40*, *76*), this epoxide epimer is not produced by the enzymatic incubations of pentalenolactone D (**8**) with either PenD, PntD, or PtlD. The *in vivo* formation of the epimeric epoxylactone **18** as a shunt metabolite is therefore most likely due to oxidation of the intermediate α-methylenelactone pentalenolactone E (**15**) by an oxygenase that is distinct from both PenD or PntD and that is encoded by a gene that is external to either the *pen* or *pnt* biosynthetic gene clusters (Scheme 5). In a similar manner, we have recently reported that the hydroxylation of the normal biosynthetic pathway intermediate 1-deoxypentalenic acid (**5**) to the commonly occurring shunt metabolite pentalenic acid (**20**) is catalyzed by CYP105D7, a typical heme-dependent monooxygenase that is encoded by the *sav7469* gene, located in the 9.05-Mb *S. avermitilis* linear chromosome more than 5 Mb from the *ptl* biosynthetic gene cluster (*77*).

The PenD-, PntD-, and PtlD-catalyzed dehydrogenation of the α-methyl substituent of pentalenolactone D (**8**) to generate the intermediate α-methylenelactone **15** is reminiscent of the desaturase activity of the multifunctional Fe^{2+}-α-ketoglutarate-dependent clavaminate synthase (CAS), which catalyzes the oxidative conversion dihydroclavaminate to clavaminate (*78*). Although PenD, PntD, and PtlD all belong to the CAS superfamily, these enzymes have no significant linear sequence similarity to CAS itself. Moreover, although the PenD and PntD desaturase−epoxidases on the one hand and the PenH and PntH hydroxylases on the other are all Fe^{2+}-α-ketoglutarate-dependent dioxygenases that act on structurally related substrates, pentalenolactone D (**8**) and 1-deoxypentalenic acid (**5**), respectively, the two groups of enzymes have no primary sequence similarity and catalyze two completely different biochemical reactions: two-step desaturation/epoxidation versus simple hydroxylation.

While most known biochemical epoxidations are catalyzed by flavin-dependent or cytochrome P450-dependent monooxygenases (*73*, *79*), direct epoxidation catalyzed by a Fe^{2+}-α-ketoglutarate-dependent dioxygenase is exceedingly rare (*80*), the only prior example being the recently discovered epoxidation of the unsaturated tripeptide dapdiamide D catalyzed by the DdaC protein of *Pantoea agglomerans* (GenBank Accession No. ADN39482) (*81*). In fact, the DdaC protein has no sequence similarity to either PenD or PntD. Although henbane hyoscamine-6β-hydroxylase is another Fe^{2+}-α-ketoglutarate-dependent dioxygenase that catalyzes a consecutive two-step epoxidation of the saturated substrate hyoscyamine to the corresponding epoxide scopolamine, the reaction differs significantly from the PenD- and PntD-catalyzed transformation since it involves the intermediacy of 6β-hydroxyhyoscyamine which is oxidized directly to scopolamine without the involvement of 6,7-dehydrohyoscyamine (*82*, *83*). An analogous hydroxylation−epoxidation mechanism for the PenD-, PntD-, or PtlD-catalyzed conversion of pentalenolactone D (**8**) to the epoxylactone **16** is firmly excluded by the demonstrated formation of the unsaturated α-methylenelactone **15**, as well as the absence of any detectable 9-hydroxypentalenolactone D in the incubation mixtures.

The natural substrate for *S. avermitilis* PtlD is neopentalenolactone D (**9**), as established by both *in vivo* accumulation of **9** in cultures of *S. avermitilis* Δ*ptlD* deletion mutants and the results of *in vitro* incubation of **9** with recombinant PtlD. Intriguingly, PtlD has a broad enough substrate specificity to permit oxidative desaturation of the isomeric substrate pentalenolactone D (**8**) to pentalenolactone E (**15**) and thence further oxidation to the epoxylactone **16**. Although we have not carried out detailed kinetic comparisons, PenD, PntD, and PtlD each catalyzed the consecutive two-step dehydrogenation−epoxidation of **8** with approximately equal efficiency.

In contrast to the observed two-step desaturation−epoxidation of pentalenolactone D (**8**), *in vitro* incubation of recombinant *S. avermitilis* PtlD or the surrogate enzymes PenD and PntD with neopentalenolactone D (**9**) resulted only in a single-step two-electron dehydrogenation to give a product tentatively identified as neopentalenolactone E (**14**), which underwent facile hydrolysis to the seco acid **17**, that upon methylation yielded PL308 (**12**). Interestingly, we were unable to detect either the expected

epoxidation product, neopentalenolactone F (**11**), or either of the corresponding hydrolysis or rearrangement products, PL324 (**13**, 10-hydroxy-PL308) or neopentalenoketolactone (**10**), that have previously been isolated from engineered mutants of *S. avermitilis* harboring an induced *ptl* biosynthetic cluster (*46*). Whether this difference in product profile is due to the intrinsic instability of the neopentalenolactone E intermediate under the incubation conditions or the absence of an essential cofactor or protein that is present in *S. avermitilis* is not yet known. It is curious, however, that the PenD-, PntD-, or PtlD-catalyzed epoxidation of the electron-poor conjugated double bond of the α-methylene lactone **15** appears to be proceed more favorably than oxidation of the electron-rich double bond of the isomeric enollactone neopentalenolactone E (**14**).

It is also noteworthy that while the PenE, PntE, and PtlE proteins each catalyze the flavin-dependent oxidation of the identical ketone substrate **7**, the individual Baeyer−Villiger oxidations yield isomeric lactone products **8** or **9**. By comparison, although PenD and PntD on the one hand and PtlD on the other normally act on a specific lactone substrate, **8** or **9**, each of these three dioxygenases is able to catalyze the oxidation of either **8** or **9**, giving in each case the same mixture of corresponding products.

CONCLUSION

For the biosynthesis of pentalenolactone, nature has orchestrated the oxidative conversion of the parent tricyclic sesquiterpene hydrocarbon pentalenene to the final metabolite pentalenolactone by calling on a full range of the oxidative instruments in its biochemical repertoire. A heme-dependent, P450 monooxygenase (PenI/PntI) first catalyzes the three-step oxidative conversion of an allylic methyl group to a conjugated carboxylic acid, after which an Fe^{2+}-α-ketoglutarate-dependent dioxygenase (PenH/PntH) hydroxylates an unactivated saturated methylene group. Following oxidation to the derived ketone by a $NADP^+$-dependent dehydrogenase (PenF/PntF), an NADPH−FAD-dependent oxygenase (PenE/PntE) mediates the Baeyer−Villiger reaction to yield the corresponding α-methyl-substituted δ-lactone. This intermediate then serves as the substrate for a two-step dehydrogenation−epoxidation that is catalyzed by a second Fe^{2+}-α-ketoglutarate-dependent dioxygenase (PenD/PntD). The final movement in this virtuoso biosynthetic performance is an unprecedented oxidative rearrangement to give pentalenolactone (**1**), a reaction that we have recently shown to be catalyzed by the heme-dependent P450 monooxygenases PenM and PntM (*84*).

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SUPPORTING INFORMATION AVAILABLE

Table of strains and plasmids, GC-MS and LC-MS data, and construction of deletion mutants and complemented bacterial mutants. This material is available free of charge via the Internet at http://pubs.acs.org.

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