

# Benfluorex, a Hypotriglyceridemic Drug, Reduces Lipid Peroxidation and Alleviates Adverse Metabolic Complications of Copper Deficiency

MEIRA FIELDS, PHD, AND CHARLES G. LEWIS, PHD

*From the USDA, ARS, Beltsville Human Nutrition Research Center, Metabolism and Nutrient Interactions Laboratory, Beltsville, Maryland, USA*

Date accepted: 18 February 1997

## ABSTRACT

The pathologies associated with copper deficiency in rats fed fructose may be induced, in part, by hypertriglyceridemia and lipid peroxidation. Reducing triacylglycerol levels in plasma may result in lowering lipid peroxidation, which in turn could ameliorate metabolic effects resulting from the combination of fructose feeding and copper deficiency. Benfluorex, a hypolipidemic factor able to reduce hypertriglyceridemia, was administered to weanling male rats fed either copper-deficient ( $0.6 \mu\text{g Cu/g}$ ) or adequate ( $6.0 \mu\text{g Cu/g}$ ) diets containing fructose as the sole dietary carbohydrate. In copper-deficient rats, benfluorex ( $50 \mu\text{g} \cdot \text{kg}^{-1} \cdot \text{d}^{-1}$ ) reduced plasma triacylglycerols from 45 to 31 mg/dL, reduced lipid peroxidation by approximately 50%, and prevented the enlargements of heart and liver size and the atrophy of the pancreas, and ameliorated anemia. It is suggested that lipid peroxidation associated with hypertriglyceridemia may be responsible for the pathologies induced by the combination of fructose consumption and copper deficiency. *Nutrition* 1997;13:895–899. ©Elsevier Science Inc. 1997

Key words: benfluorex, hypertriglyceridemia, lipid peroxidation, copper, iron, pancreas

## INTRODUCTION

Copper deficiency is associated with anemia, hypercholesterolemia, hypertriglyceridemia, abnormal glucose tolerance, pancreatic atrophy, heart hypertrophy, and premature mortality.<sup>1–5</sup> These abnormalities are due to the presence of simple sugars, such as sucrose or fructose in the diet.<sup>1–5</sup> When rats consume a copper-deficient diet that contains complex carbohydrates, such as starch, the pathologies associated with copper deficiency are either ameliorated or prevented, and the animals survive.<sup>1–6</sup> Since copper deficiency by itself is not sufficient to induce these pathologies, we hypothesized that metabolic pathways that are specific for fructose but not for starch are responsible for the dramatic differences between fructose-fed and starch-fed rats.

Fructose feeding has been shown to produce elevations in plasma triacylglycerols.<sup>7–10</sup> Increased superoxide production has been demonstrated in patients with hypertriglyceridemia.<sup>11</sup> Similarly, increased levels of reactive oxygen species have been re-

ported to be induced in copper deficiency due to the reduced activity of the copper-containing enzyme superoxide dismutase (CuSOD) and the selenoenzyme glutathione peroxidase (GSH-Px).<sup>12–14</sup> The combination of fructose with copper deficiency produced reactive oxygen species detected by electron spin resonance (ESR) and increased lipid peroxidation.<sup>12,15,16</sup> The consumption of a copper-deficient, starch-based diet did not have this effect.<sup>12,15,16</sup>

Because reactive oxygen species are suspected to play an important role in the initiation of cellular damage, we investigated whether lowering hypertriglyceridemia will reduce levels of lipid peroxidation, which in turn should alleviate some metabolic and physiologic abnormalities associated with copper deficiency. Benfluorex, (2-[1-methyl-2-[3-(trifluoromethyl)-phenyl]ethyl]amino] ethanol benzoate ester), a pharmaceutical hypolipidemic agent, was able to reduce hepatic triacylglycerol in alcoholic rats and in rats fed fructose and fat.<sup>17,18</sup> It also inhibited de novo synthesis of

Correspondence to: Dr. Meira Fields, USDA, ARS, BHNRC, Metabolism and Nutrient Interactions Laboratory, Room 323, Building 307, BARC-East, Beltsville, MD 20705, USA

TABLE I.

FINAL BODY MASS, RELATIVE ORGAN SIZE, HEMATOCRIT AND HEPATIC COPPER AND IRON CONCENTRATIONS

	Copper-deficient		Copper-adequate	
	-Benfluorex	+Benfluorex	-Benfluorex	+Benfluorex
Body mass (g)	159 ± 3	140 ± 4	194 ± 3	172 ± 2
Heart mass (g)	0.99 ± 0.05	0.75 ± 0.02	0.75 ± 0.02	0.66 ± 0.02
Relative heart size (g/100 g)	0.62 ± 0.03	0.53 ± 0.02	0.39 ± 0.09	0.39 ± 0.01
Liver mass (g)	7.34 ± 0.12	5.97 ± 0.18	7.07 ± 0.18	6.27 ± 0.13
Relative liver size (g/100)	4.6 ± 0.10	4.2 ± 0.12	3.6 ± 0.07	3.6 ± 0.06
Hematocrit (%)	25.8 ± 0.6	32.5 ± 1.4	44.3 ± 0.5	44.3 ± 0.5
Liver Cu (μg/g wet wt)	0.93 ± 0.07	1.15 ± 0.10	5.59 ± 0.21	5.71 ± 0.18
Liver Fe (μg/g wet wt)	174.8 ± 11.3	184.3 ± 19.5	102.4 ± 9.5	101.2 ± 2.9
Hepatic lipid peroxidation (nmol MDA/g)	33.8 ± 3.1	17.7 ± 3.5	21.7 ± 3.2	12.7 ± 2.7
ANOVA ( <i>P</i> values)				
	Copper	Benfluorex	Copper × Benfluorex	
Body mass	0.0001	0.0001	NS	
Heart mass	0.0001	0.0001	0.0156	
Relative heart size	0.0001	0.0348	0.0240	
Liver mass	NS	0.0001	NS	
Relative liver size	0.0001	NS	NS	
Hematocrit	0.0001	0.0004	0.0004	
Liver Cu	0.0001	NS	NS	
Liver Fe	0.0001	NS	NS	
Lipid peroxidation	0.0150	0.0010	NS	

Means ± SEM of 10 observations/group. MDA, malondialdehyde; ANOVA, analysis of variance; NS, not significant.

fatty acids, triacylglycerol synthesis and improved insulin sensitivity.<sup>17-19</sup> We predicted that these suppressive effects of benfluorex on lipid metabolism might alleviate the pathologies associated with copper deficiency in rats fed fructose. The work presented here was designed to test this hypothesis.

#### MATERIALS AND METHODS

Weanling male rats weighing approximately 40–50 g each were randomly assigned to four dietary groups according to copper levels and benfluorex: group 1, copper-deficient; group 2, copper-deficient + benfluorex; group 3, copper-adequate; group 4, copper-adequate + benfluorex. The copper-deficient diets contained 0.6 μg Cu/g diet, and the copper-adequate diets contained 6.0 μg Cu/g diet as determined by atomic absorption spectrophotometry. The description of the diets has been previously reported.<sup>20</sup>

Benfluorex hydrochloride (Sigma Chemical Co., St. Louis, MO, USA) was introduced to the diet on day 2 at a concentration designed to provide 12.5 mg · kg<sup>-1</sup> · d<sup>-1</sup> and gradually increased to 50 mg · kg<sup>-1</sup> · d<sup>-1</sup> over a 3-d period.

The rats were fed their respective diets for 4 wk. Following an overnight fast, they were decapitated. Blood was collected into capillary tubes for hematocrit. Blood was also collected into heparinized test tubes and centrifuged. Plasma was used to measure the concentration of triacylglycerols by the automated procedure of the centrifichem. Plasma insulin was measured by using a radio immunoassay (RIA) using rat antibody (Linco Research Inc., St. Charles, MO, USA). Livers, pancreata, and hearts were removed and weighed, and portions of livers were used for the determination of copper and iron concentrations.<sup>21</sup> Other liver

portions were used to assess lipid peroxidation by measuring malondialdehyde formation using the thiobarbituric acid (TBA) reactive technique as described elsewhere.<sup>22,23</sup>

The pancreas was trimmed of fat and weighed again. It was homogenized in phosphate-buffered saline (PBS) using a polytron homogenizer equipped with stainless steel blades. Aliquots of homogenate were used to measure the concentration of insulin using the same RIA kit that utilizes an antibody made specifically against rat insulin (Linco Research Inc.). Other aliquots were used to measure the activities of pancreatic amylase using Sigma enzymatic procedure no. 577, respectively, and pancreatic copper and iron concentrations by atomic absorption spectrophotometry.<sup>21</sup>

All data were subjected to a 2 × 2 analysis of variance (ANOVA), two levels of copper, and two levels of benfluorex. Main effects and interactions of significance level of *P* < 0.05 were considered statistically significant.

#### RESULTS

Final body mass, relative organ sizes, hematocrit, and hepatic copper and iron concentrations are summarized in Table I. Body mass was reduced by copper deficiency. Rats fed benfluorex exhibited lower body mass than rats that did not consume the drug. Food intake was not affected by benfluorex (data not shown). The largest heart and liver sizes were found in copper-deficient rats that did not receive benfluorex. Heart weight and relative heart sizes were reduced by benfluorex in copper-deficient rats. Liver weight was reduced by benfluorex. When expressed per 100 g body weight, the largest relative liver size was noted in copper-deficient rats that were not

TABLE II.

PANCREAS SIZE, COPPER, IRON AND INSULIN CONCENTRATIONS AND AMYLASE ACTIVITY				
	Copper-deficient		Copper-adequate	
	- Benfluorex	+ Benfluorex	- Benfluorex	+ Benfluorex
Pancreas weight (g)	0.73 ± 0.05	0.96 ± 0.04	1.40 ± 0.03	1.32 ± 0.03
Pancreas size (g/100 g)	0.46 ± 0.03	0.69 ± 0.02	0.72 ± 0.02	0.76 ± 0.02
Copper (μg/g wet wt)	1.78 ± 0.12	1.66 ± 0.11	2.31 ± 0.12	1.29 ± 0.09
Iron (μg/g wet wt)	51.6 ± 3.8	49.4 ± 2.3	17.0 ± 0.9	18.9 ± 0.8
Insulin (ng/pancreas)	6348 ± 1650	1718 ± 458	332 ± 87	351 ± 57
Amylase (U/pancreas)	2988 ± 587	5783 ± 889	7695 ± 431	9912 ± 860
ANOVA ( <i>P</i> values)				
	Copper	Benfluorex	Copper × Benfluorex	
Pancreas weight	0.0001	0.0001	0.0002	
Pancreas size	0.0001	NS	0.0006	
Copper	NS	0.0001	0.0006	
Iron	0.0001	NS	NS	
Insulin	0.0001	0.0046	0.0047	
Amylase	0.0001	0.0005	NS	

Means ± SEM of 10 observations/group. NS, not significant.

treated with benfluorex. Copper-deficient rats exhibited the lowest hematocrits. Benfluorex raised hematocrits in copper-deficient rats. Liver copper was the lowest in rats that consumed the low copper diet. Liver iron was elevated by copper deficiency. Benfluorex had no effect on either liver copper or liver iron concentrations. The highest degree of lipid peroxidation was evident in copper-deficient rats that did not receive benfluorex. Benfluorex reduced hepatic lipid peroxidation in copper-deficient and copper-adequate rats.

Table II summarizes data pertaining to pancreas size and contents. The smallest pancreas weight and size was found in copper-deficient rats. Benfluorex raised the weight and relative size of the pancreas in copper-deficient rats. The concentration of copper in the pancreas was lowered but that of iron was elevated by copper deficiency. The highest insulin concentration of the pancreas was found in copper-

deficient rats not receiving benfluorex. The administration of benfluorex greatly reduced the concentration of pancreatic insulin. The lowest insulin content was recorded in copper-adequate controls. The administration of benfluorex doubled the activity of amylase in the pancreas of copper-deficient rats.

Fasting levels of triacylglycerols and insulin in plasma are summarized in Table III. The highest levels of triacylglycerols were found in plasma of copper-deficient rats. Concentration of triacylglycerols was reduced by benfluorex in copper-deficient and adequate rats. Copper-deficient rats had the lowest level of plasma insulin. The administration of benfluorex raised fasting levels of insulin in all rats.

#### DISCUSSION

The present study was designed to determine whether a reduction of plasma triacylglycerols will lower lipid peroxidation,

TABLE III.

FASTING PLASMA TRIACYLGLYCEROLS AND INSULIN				
	Copper-deficient		Copper-adequate	
	- Benfluorex	+ Benfluorex	- Benfluorex	+ Benfluorex
Triacylglycerols (mg/dL)	45.1 ± 3.5	31.3 ± 3.3	30.7 ± 1.3	22.3 ± 1.2
Insulin (ng/mL)	0.165 ± 0.02	0.313 ± 0.06	0.454 ± 0.09	0.520 ± 0.06
ANOVA ( <i>P</i> values)				
	Copper	Benfluorex	Copper × Benfluorex	
Triacylglycerol	0.0001	0.0001	NS	
Insulin	0.0603	0.0015	NS	

Means ± SEM of 10 observations/group. NS, not significant.

which in turn will prevent some of the pathologies associated with copper deficiency such as liver enlargement, heart hypertrophy, pancreatic atrophy, and anemia. The administration of benfluorex, a hypolipogenic and hypotriacylglycerolmic drug reduced plasma triacylglycerols. It also lowered hepatic lipid peroxidation and prevented abnormalities induced by copper deficiency. It is suggested that suppressing effects of benfluorex on lipid metabolism were responsible for the amelioration of pathologies associated with copper deficiency.

The reduced hepatic lipid peroxidation and levels of plasma triacylglycerols were not due to changes in copper or iron concentrations, but rather to drug mechanism. Benfluorex via its metabolites inhibits *de novo* synthesis of fatty acids and triacylglycerol synthesis, and improves insulin sensitivity.<sup>17-19</sup> Because of these functions, benfluorex has been recommended as a potential tool for alleviating some of the adverse effects of diabetes, obesity, and atherosclerosis.<sup>17-19</sup>

It is well established that fatty liver leads to cirrhosis and fibrosis.<sup>24,25</sup> Similarly, in iron-storage diseases, the liver is the main target organ for injury, and hepatic fibrosis and cirrhosis are the major pathologic hallmarks of the disease.<sup>24,25</sup> Excess tissue iron provides a direct stimulus to collagen synthesis.<sup>26</sup> High stored iron by itself is not sufficient to cause tissue damage. It is well established that iron is highly reactive under a certain redox environment.<sup>27-29</sup> This reactivity can be associated with damage to cellular systems. Fructose consumption could provide this redox environment. Excess amounts of NADH and NADPH are known to occur during the metabolism of fructose.<sup>30-32</sup> In addition, fructose feeding should increase the production of superoxide since it stimulates the activity of xanthine oxidase.<sup>30</sup> In copper deficiency the combination of fructose feeding with hepatic iron retention is responsible for liver damage, as shown by fat vacuoles and foci of individual cell necrosis of hepatocytes.<sup>33</sup> Prevention of lipogenesis should reduce the availability of substrates necessary for lipid peroxidation by iron. Similarly, the reduction of hepatic iron in copper deficiency also lowers reactive oxygen species and lipid peroxidation.<sup>15,16</sup>

It is well established that liver injury, which is caused by hepatic iron overload, is capable of inducing pathologies to other organs such as the heart and the pancreas.<sup>24</sup> Liver iron overload has been associated with pancreatic damage and diabetes.<sup>34-36</sup> Hepatic iron retention occurs in copper deficiency.<sup>37,38</sup> Prevention of hepatic iron retention in copper deficiency either by adminis-

tration of deferoxamine or lowering dietary intake of iron reduced the size of the liver and prevented hypertrophy of the heart and atrophy of the pancreas.<sup>15,16</sup> In the present study, however, levels of hepatic iron were not reduced by benfluorex, and yet liver enlargement and pancreatic atrophy were prevented. It is suggested that other mechanisms not related to iron but associated with prevention of hepatic lipogenesis were responsible for beneficial effects of benfluorex.

In addition to preventing pancreatic atrophy, functional ability of the pancreas to secrete insulin and to synthesize amylase was restored by benfluorex. In copper deficiency the exocrine pancreas atrophies, and the activity of digestive enzymes is reduced.<sup>39-41</sup> In addition, the ability of the endocrine pancreas to secrete insulin is impaired.<sup>42</sup> Copper-deficient rats of the present study accumulated insulin in the pancreas that was not secreted to the general circulation. The ability of the endocrine pancreas to secrete insulin was greatly restored by benfluorex. This improved capacity of the pancreas to release insulin was associated with higher levels of plasma insulin. Amylase activity was also raised by benfluorex. The improvement in insulin secretion may be responsible for amylase activity. It has been reported that amylase activity is regulated by insulin.<sup>43,44</sup>

Both copper-deficient and copper-adequate rats of the present study exhibited a reduction of body mass when treated with benfluorex. It is well established that growth rate affects essential metalhomeostasis. Due to a depressed growth rate, the requirements for copper are reduced and higher concentrations of stored copper but lower iron were reported.<sup>45-47</sup> The elevated levels of copper and the reduced levels of iron were responsible for the amelioration of the pathologies associated with copper deficiency.<sup>45-47</sup> In the present study, however, the reduced body mass of rats treated with benfluorex did not affect levels of copper or iron. Alleviated signs associated with copper deficiency in rats treated with benfluorex should be due to the direct effect of either benfluorex or its metabolites on metabolic pathways.

The results of this study show that rats fed a copper-deficient diet developed hypertriglyceridemia, lipid peroxidation, and numerous other pathologies. The ability of benfluorex to ameliorate or prevent these abnormalities may indicate that the mechanism involves alterations in lipid metabolism. Once circulating triacylglycerols were lowered and reactive oxygen species were reduced, a cluster of other abnormalities associated with copper deficiency were ameliorated.

## REFERENCES

1. Fields M, Ferretti RJ, Smith JC, Reiser S. The effect of copper deficiency on metabolism and mortality in rats fed sucrose or starch diets. *J Nutr* 1983;113:1335
2. Redman RS, Fields M, Reiser S, Smith JC. Dietary fructose exacerbates the cardiac abnormalities of copper deficiency in rats. *Atherosclerosis* 1988;74:203
3. Fell BF, King TP, Davies NT. Pancreatic atrophy in copper-deficient rats: histochemical and ultrastructural evidence of a selective effect on acinar cells. *Histochem J* 1982;14:665
4. Allen KGD, Klevay LM. Cholesterolemia and cardiovascular abnormalities in rats caused by copper deficiency. *Atherosclerosis* 1987;29:81
5. Fields M, Ferretti RJ, Smith JC, Reiser S. Impairment of glucose tolerance in copper deficient rats: dependency on the type of dietary carbohydrate. *J Nutr* 1984;114:393
6. Fields M, Ferretti RJ, Smith JC, Reiser S. The severity of copper deficiency is determined by the type of dietary carbohydrate. *Proc Soc Exp Biol Med* 1984;175:530
7. Thorburn AW, Storlien LH, Jenkins AB, Khouri S, Kraegen EW. Fructose-induced *in vivo* insulin resistance and elevated plasma triacylglycerol levels in rats. *Am J Clin Nutr* 1989;49:1155
8. Zavaroni I, Chen Y-DI, Reaven GM. Studies on the mechanism of fructose induced hypertriglyceridemia in the rat. *Metabolism* 1982; 31:1077
9. Sleder J, Chen Y-DI, Cully MD, Reaven GM. Hyperinsulinemia in fructose-induced hypertriglyceridemia in the rat. *Metabolism* 1980; 29:303
10. Hallfrisch J, Reiser S, Prather ES. Blood lipid distribution of hyperinsulinemic men consuming three levels of fructose. *Am J Clin Nutr* 1983;37:740
11. Hiramatsu K, Arimori S. Increased superoxide production by mononuclear cells of patients with hypertriglyceridemia and diabetes. *Diabetes* 1988;37:832
12. Fields M, Ferretti RJ, Smith JC, Reiser S. Effects of the interaction of type of dietary carbohydrate with copper deficiency on lipid peroxidation in rat tissues. *Biol Trace Elem Res* 1984;6:379
13. Paynter DI, Moir RJ, Underwood EJ. Changes in activity of Cu-Zn superoxide dismutase enzyme in tissue of the rat with changes in dietary copper. *J Nutr* 1979;109:1570
14. Prohaska JR, Sunde RA, Zinn KR. Livers from copper-deficient rats have lower glutathione peroxidase and mRNA levels but normal liver selenium. *J Nutr Biochem* 1992;3:429
15. Fields M, Lewis CG, Lure MD, Antholine WE. The severity of copper

- deficiency can be ameliorated by deferoxamine. *Metabolism* 1991;40:105
16. Fields M, Lewis CG, Lure MD, Burns WA, Antholine WE. Low dietary iron prevents free radical formation and heart pathology of copper-deficient rats fed fructose. *Proc Soc Exp Biol Med* 1993;202:225
  17. Geelen MJH. Mechanisms responsible for the inhibitory effects of benfluorex on hepatic intermediary metabolism. *Biochem Pharmacol* 1983;32:1765
  18. Storlien LH, Oakes ND, Pan DA, Kusunoki M, Jenkins AB. Syndromes of insulin resistance in the rat. Inducement by diet and amelioration with benfluorex. *Diabetes* 1993;42:457
  19. Brindley DN, Akester H, Derrick GP, et al. Effects of chronic administration of benfluorex to rats on the metabolism of corticosterone, glucose, triacylglycerols, glycerol and fatty acids. *Biochem Pharmacol* 1988;37:695
  20. Fields M, Lewis CG, Lure MD, Antholine WE. The influence of gender on developing copper deficiency and on free radical generation of rats fed a fructose diet. *Metabolism* 1992;41:989
  21. Hill AD, Patterson KY, Veillon C, Morris ER. Digestion of biological materials for mineral analysis using a combination of wet and dry ashing. *Anal Chem* 1986;58:340
  22. Fields M, Lewis CG, Lure MD. Allopurinol, an inhibitor of xanthine oxidase, reduces uric acid levels and modifies the signs associated with copper deficiency in rats fed fructose. *Free Rad Biol Med* 1996;20:595
  23. Paynter DI. The role of dietary copper, manganese, selenium and vitamin E in lipid peroxidation in tissues of the rat. *Biol Trace Elem Res* 1980;2:121
  24. Robbins SL. *Pathology*. Philadelphia: W.B. Saunders Company, 1968
  25. Bonkovsky HL. Iron and the liver. *Am J Med Sci* 1991;301:32
  26. Pietrangeli A, Gualdi R, Casalgrandi G, et al. Enhanced hepatic collagen type I mRNA expression into fat storing cells in a rodent model of hemochromatosis. *Hepatology* 1994;19:714
  27. Braughler JM, Duncan LA, Chase RL. The involvement of iron in lipid peroxidation. Importance of ferric to ferrous ratios in initiation. *J Biol Chem* 1986;261:10282
  28. Cantoni O, Furmo M, Cattaberi F. Role of metal ions in oxidant cell injury. *Biol Trace Elem Res* 1989;21:277
  29. Rowley D, Halliwell B. Superoxide-dependent information of hydroxyl radicals from NADH and NADPH in the presence of iron salts. *FEBS Lett* 1982;142:39
  30. Van den Berghe G. Fructose: metabolism and short-term effects on carbohydrate and purine metabolic pathways. *Prog Biochem Pharmacol* 1986;21:1
  31. Van den Berghe G. Metabolic effects of fructose in the liver. *Curr Top Cell Regul* 1978;13:97
  32. Bellomo G, Comstock JP, Wen D. Prolonged fructose feeding and aldose reductase inhibition: effect on the polyol pathway in kidney of normal rats. *Proc Soc Exp Biol Med* 1987;186:348
  33. Fields M, Lewis CG, Lure MD, Burns WA. Dietary ferric vs. ferrous iron in copper-deficient rats fed fructose-based diets. *J Am Coll Nutr* 1995;4:399
  34. Dymock LW, Cassar J, Pyke DA, Oakley WG, Williams R. Observations on the pathogenesis, complications and treatment of diabetes in 115 cases of haemochromatosis. *Am J Med* 1972;52:203
  35. Cutler P. Deferoxamine therapy in high-ferritin diabetes. *Diabetes* 1989;38:1207
  36. Williams R, Smith PM, Spicer EJP, Barry M, Sherlock S. Venesection therapy in idiopathic haemochromatosis. *Q J Med* 1969;38:1
  37. Williams DM, Kennedy FS, Green BF. Hepatic iron accumulation in copper deficient rats. *Br J Nutr* 1983;50:653
  38. Owen JR. Effects of iron on copper metabolism and copper on iron metabolism in rats. *Am J Physiol* 1973;224:514
  39. Fell BF, King TP, Davies NT. Pancreatic atrophy in copper-deficient rats. Histochemical and ultrastructural evidence of a selective effect on acinar cells. *Histochem J* 1982;14:665
  40. Folsch UR, Creunzfeldt MD. Pancreatic duct cells in rats: secretory studies in response to secretin, cholecystokinin-pancreozymin and gastrin in vivo. *Gastroenterology* 1977;73:1053
  41. Lewis CG, Fields M, Craft N, Yang C-Y, Reiser S. Changes in pancreatic enzyme specific activities of rats fed a high-fructose, low-copper diet. *J Am Coll Nutr* 1988;7:27
  42. Recant L, Voyles NR, Timmers K, Zalenski C, Fields M, Bhathena SJ. Copper deficiency in weanling rats increased pancreatic contents of enkephalin-containing peptides and insulin. *Peptides* 1986;7:1061
  43. Soling HD, Under KO. The role of insulin in the regulation of alpha amylase synthesis in the rat pancreas. *Eur J Clin Invest* 1972;2:199
  44. Korc M, Owenbach D, Quinto C, Rutter WJ. Pancreatic islets-acinar cell interaction: amylase messenger RNA levels are determined by insulin. *Science* 1981;213:351
  45. Saari JT, Johnson WT, Reeves PG, Johnson LK. Amelioration of effects of severe dietary copper-deficiency by food restriction in rats. *Am J Clin Nutr* 1993;58:891
  46. Werman MJ, Bhathena SJ. Restricted food intake ameliorates the severity of copper-deficiency in rats fed a copper-deficient, high-fructose diet. *Med Sci Res* 1993;21:309
  47. Fields M, Lewis CG, Lure MD. Hypothesis: the requirements for copper are determined by growth rate in rats fed fructose but not in rats fed starch. *J Appl Nutr* 1993;45:85