

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/5925940>

Product Deuterium Isotope Effect for Orotidine 5'-Monophosphate Decarboxylase: Evidence for the Existence of a Short-Lived Carbanion Intermediate

ARTICLE in JOURNAL OF THE AMERICAN CHEMICAL SOCIETY · NOVEMBER 2007

Impact Factor: 12.11 · DOI: 10.1021/ja076222f · Source: PubMed

CITATIONS

38

READS

19

6 AUTHORS, INCLUDING:



Tina L Amyes

University at Buffalo, The State University of ...

108 PUBLICATIONS 3,380 CITATIONS

SEE PROFILE



Bryant McKay Wood

Harvard Medical School

19 PUBLICATIONS 452 CITATIONS

SEE PROFILE



John P. Richard

University at Buffalo, The State University of ...

219 PUBLICATIONS 6,902 CITATIONS

SEE PROFILE

Published in final edited form as:

J Am Chem Soc. 2007 October 31; 129(43): 12946–12947.

Product Deuterium Isotope Effect for Orotidine 5'-Monophosphate Decarboxylase: Evidence for the Existence of a Short-Lived Carbanion Intermediate

Krisztina Toth[†], Tina L. Amyes[†], Bryant M. Wood[#], Kui Chan[#], John A. Gerlt[#], and John P. Richard[†]

Department of Chemistry, University at Buffalo, Buffalo, New York 14260 and Departments of Biochemistry and Chemistry, University of Illinois, Urbana, Illinois 61801

We report that equal yields of [6-¹H]-uridine 5'-monophosphate (50%) and [6-²H]-uridine 5'-monophosphate (50%) are obtained from the decarboxylation of orotidine 5'-monophosphate (**OMP**) catalyzed by orotidine 5'-monophosphate decarboxylase in a solvent of 50/50 (v/v) H₂O/D₂O. This observation of an unusually small product isotope effect of unity eliminates a proposed mechanism in which proton transfer from Lys-93¹ to C-6 provides electrophilic *push* to the loss of CO₂ from **OMP** in a concerted reaction.^{2,3} It provides evidence that proton transfer from the ammonium cation side-chain of Lys-93 to a vinyl carbanion intermediate is *faster* than the bond rotation that exchanges the positions of the acidic N-L⁺ hydrons of this side-chain.

Orotidine 5'-monophosphate decarboxylase (OMPDC) is a remarkable enzyme because it employs no metal ions or other cofactors but yet effects an enormous 10¹⁷-fold acceleration of the chemically very difficult decarboxylation of **OMP** to give uridine 5'-monophosphate (**UMP**).^{4,5} It has been shown that a large fraction of the enzymatic rate acceleration results directly from utilization of the intrinsic binding energy of the remote nonreacting 5'-phosphodianion group of **OMP** in transition state stabilization.⁶ The decarboxylation reaction is often proposed to proceed in two steps through a vinyl carbanion intermediate (Scheme 1). However, it has also been suggested that this unstable intermediate might be avoided in a concerted reaction in which decarboxylation and proton transfer to C-6 occur in a single step.^{2,3}

Experimental and computational studies on OMPDC have focused largely on the partly rate-determining and highly unfavorable loss of CO₂ from **OMP**.⁷⁻⁹ There are few data pertaining to the proton transfer to C-6 of the pyrimidine ring. Experimental characterization of this proton transfer step is essential for insight into the existence and lifetime of the putative enzyme-bound vinyl carbanion intermediate.

OMPDC catalyzes incorporation of a hydron from solvent into the **UMP** product and it has been reported that the decarboxylation of saturating **OMP** is 30% faster in H₂O than in D₂O.

⁷ While the origin of this solvent isotope effect on *k*_{cat} is unclear, it may represent a secondary

jrichard@chem.buffalo.edu.

[†]University at Buffalo

[#]University of Illinois

solvent kinetic isotope effect (SKIE). By contrast, a product isotope effect (PIE) determined in experiments in which H and D in a mixed solvent of H₂O/D₂O compete for reaction with enzyme-bound **OMP** to form **UMP** labeled at C-6 (Scheme 2) would provide insight into the changes in bonding at the transferred hydron that occur on proceeding to the transition state for the product-determining step.¹⁰ PIEs are more precise and easier to interpret than SKIEs determined as the ratio of rate constants for reactions in H₂O and D₂O because: (1) There are no complications from any secondary SKIE when the H- and D-labeled products are formed in the same mixed H₂O/D₂O solvent. (2) There are no errors due to differences in the conditions for separate reactions in H₂O and D₂O, such as enzyme concentration, temperature and pL.

The product distribution for the decarboxylation of **OMP** catalyzed by OMPDC in 50/50 (v/v) H₂O/D₂O was determined by ¹H NMR spectroscopy at 500 MHz. Figure 1 shows the partial ¹H NMR spectrum of **UMP** obtained from the decarboxylation of **OMP** (2 mM) catalyzed by OMPDC from *S. cerevisiae* (C155S mutant, 24 nM, 1 hr, >90% reaction) in 50/50 (v/v) H₂O/D₂O at pL 7.3 and 25 °C (*I* = 0.10, NaCl).^{11,12} The value of PIE = 1.0 was calculated using eq 1, where *A_H* is the integrated area of the doublet due to the C-6 proton of [6-¹H]-**UMP** (7.990 ppm), and *A_D* is the integrated area of the singlet due to the C-5 proton of [6-²H]-**UMP** (5.865 ppm).¹³ By comparison, PIEs of 7.3 – 8.1 for proton transfer to ring-substituted aryl vinyl ethers from lyonium ion in 50/50 (v/v) H₂O/D₂O have been reported recently.¹⁰

$$\text{PIE} = A_{\text{H}}/A_{\text{D}} \quad (1)$$

We used similar procedures to determine values of PIE = 1.0 for decarboxylation of **OMP** (2 mM) catalyzed by OMPDC from both *E. coli* (40 nM) and *M. thermoautotrophicum* (40 nM) in 50/50 (v/v) H₂O/D₂O at pL 7.3 and 25 °C (*I* = 0.10, NaCl). The essentially identical PIEs determined for OMPDC from different sources is significant, because these enzymes exhibit somewhat different architectures at their active sites.^{2,9a,14,15}

The value of PIE = 1.0 for the OMPDC-catalyzed decarboxylation of **OMP** in 50/50 (v/v) H₂O/D₂O shows that the deuterium enrichment of the hydron used to protonate **OMP** or an intermediate carbanion at the reaction transition state (50%) is the same as that of the 50/50 (v/v) H₂O/D₂O solvent. The product-determining step is thought to be proton transfer from the NL₃⁺ group of the side-chain of Lys-93 to **OMP** or to a reaction intermediate (Scheme 3).¹⁵ Values of $\phi_{\text{NL}_3^+} \approx 1.0$ have been reported for the H/D fractionation between L₂O and R-NL₃⁺, so that the deuterium enrichment of the NL₃⁺ group of Lys-93 should be similar to that of the solvent L₂O.¹⁶ Therefore the PIE of 1.0 is essentially equal to the primary kinetic isotope effect for reaction of the H- and D-labeled NL₃⁺ group of Lys-93 to form [6-¹H]-**UMP** and [6-²H]-**UMP**.

A significant primary product isotope effect is expected for a reaction in which there is *movement* of the proton in the transition state for the product-determining step,^{17a} and there is no precedent for PIEs as small as 1.0 when carbanion protonation is the product-determining step.^{17a,18} The observed PIE of 1.0 requires that all of the zero point energy present in the N-L⁺ bonds of Lys-93 be maintained at the transition state for the step that determines whether the **UMP** product is labeled at C-6 with H or D. This PIE is not consistent with a mechanism in which proton transfer from Lys-93 to C-6 of **OMP** provides electrophilic *push* to the loss of CO₂ in a concerted reaction that avoids formation of an unstable vinyl carbanion intermediate (bottom pathway, Scheme 3).^{2,3,19}

We suggest that the essentially statistical yields of [6-¹H]-**UMP** and [6-²H]-**UMP** from the OMPDC-catalyzed decarboxylation of **OMP** are established at a step that occurs prior to hydron transfer to a vinyl carbanion intermediate. This could be the decarboxylation step, if an N-L⁺ bond of Lys-93 is already correctly positioned to deliver a hydron to a vinyl carbanion (*k_{dc}*, Scheme 3). Alternatively it may be a step that orients an N-L⁺ bond of Lys-93 into a

“reactive position” where hydron transfer to a vinyl carbanion intermediate can occur. In both cases the PIE of 1.0 *requires* that the chemical step of hydron transfer to the carbanion be *faster* than any molecular motion that allows its discrimination between reaction with H and D at the NL_3^+ group of Lys-93.¹⁷ We therefore propose that hydron transfer from the side-chain of Lys-93 to a vinyl carbanion intermediate (k_p) is faster than any movement that exchanges the positions of the N-L^+ hydrons and which would allow the carbanion to *select* for reaction with H or D.¹⁷ In water, the rate constant for such a step is ca. 10^{11} s^{-1} .²⁰

The X-ray crystal structure of yeast OMPDC complexed with 6-hydroxyuridine 5'-monophosphate shows that the $\text{CH}_2\text{-NH}_3^+$ group of Lys-93 is anchored by two hydrogen bonds to the carboxylate groups of Asp-91 and Asp-96 that are proposed to direct the third ammonium hydron of Lys-93 towards the putative vinyl carbanion intermediate.¹⁵ These hydrogen bonds should also restrict rotation about the carbon-nitrogen bond of the terminal $\text{CH}_2\text{-NL}_3^+$ group of Lys-93 ($k_{\text{rot}} \ll 10^{11} \text{ s}^{-1}$). This would favor the observed unselective proton transfer from the remaining free (non-hydrogen-bonded) hydron to a vinyl carbanion intermediate.

Acknowledgement

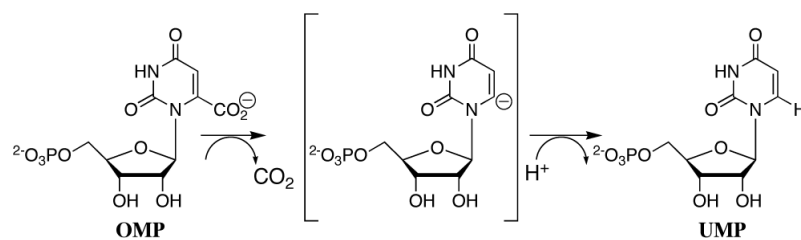
We acknowledge the National Institutes of Health (GM39754 to JPR and GM65155 to JAG) for generous support of this work.

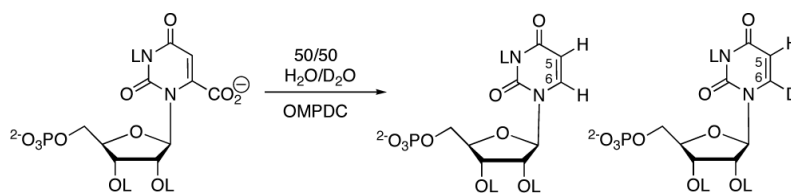
References

1. Residues are numbered according to the sequence for the enzyme from yeast.
2. Begley TP, Appleby TC, Ealick SE. *Curr. Opin. Struct. Biol* 2000;10:711–718. [PubMed: 11114509]
3. Begley TP, Ealick SE. *Curr. Opin. Chem. Biol* 2004;8:508–515. [PubMed: 15450493]
4. Miller BG, Wolfenden R. *Annu. Rev. Biochem* 2002;71:847–885. [PubMed: 12045113]
5. Radzicka A, Wolfenden R. *Science* 1995;267:90–93. [PubMed: 7809611]
6. Amyes TL, Richard JP, Tait JJ. *J. Am. Chem. Soc* 2005;127:15708–15709. [PubMed: 16277505]
7. Ehrlich JI, Hwang C-C, Cook PF, Blanchard JS. *J. Am. Chem. Soc* 1999;121:6966–6967.
8. Rishavy MA, Cleland WW. *Biochemistry* 2000;39:4569–4574. [PubMed: 10769111]
9. a Wu N, Mo Y, Gao J, Pai EF. *Proc. Natl. Acad. Sci. U.S.A* 2000;97:2017–2022. [PubMed: 10681441]
b Gao J, Byun KL, Kluger R. *Topics in Current Chemistry* 2004;238:113–136. c Warshel A, Strajbl M, Villa J, Florian J. *Biochemistry* 2000;39:14728–14738. [PubMed: 11101287]
10. Tsang W-Y, Richard JP. *J. Am. Chem. Soc* 2007;129:ASAP.
11. ¹H NMR spectra were recorded on a Varian Unity Inova-500 spectrometer using a sweep width of 6000 Hz, a 90° pulse angle, an acquisition time of 6 s, a relaxation delay between pulses of 80 s (> 7T₁) and with suppression of the water peak. Baselines were subjected to first-order drift correction before integration of the signals.
12. Pentz L, Thornton ER. *J. Am. Chem. Soc* 1967;89:6931–6938. Reaction mixtures were buffered with 50 mM 3-(N-morpholino)propanesulfonic acid (50% free base). Values of pL were obtained by adding 0.18 to the reading of the pH meter
13. Control experiments showed that $A_{\text{H}}/A_{\text{D}}$ (eq 1) remains constant over a period of ca. 20 hours. The variation in this ratio determined from integration of different NMR spectra is less than 3%.
14. Harris P, Poulsen J-CN, Jensen KF, Larsen S. *J. Mol. Biol* 2002;318:1019–1029. [PubMed: 12054799]
15. Miller BG, Hassell AM, Wolfenden R, Milburn MV, Short SA. *Proc. Natl. Acad. Sci. U.S.A* 2000;97:2011–2016. [PubMed: 10681417]
16. Schowen KB, Schowen RL. *Methods Enzymol* 1982;87:551–606. [PubMed: 6294457]
17. a Thibblin A, Jencks WP. *J. Am. Chem. Soc* 1979;101:4963–4973. b Jencks WP. *Acc. Chem. Res* 1980;13:161–169.
18. Fishbein JC, Jencks WP. *J. Am. Chem. Soc* 1988;110:5075–5086.
19. The primary ¹³C KIE on OMPDC-catalyzed decarboxylation of **OMP** labeled at the carboxylate carbon decreases from 1.043 for reaction in H₂O to 1.034 for the 25% slower reaction in D₂O [Ref.

7], which shows that decarboxylation is *less* rate-determining in D₂O than in H₂O. This result is not easily rationalized by a mechanism in which proton transfer from Lys-93 to C-6 of **OMP** is concerted with the loss of CO₂ because the change from H₂O to D₂O should raise the barrier to a reaction in which proton transfer is concerted with loss of CO₂, as a result of a normal primary KIE. This would cause the loss of CO₂ to become more rate-determining in a multistep enzymatic reaction in D₂O and would result in an increase, rather than the observed decrease, in the ¹³C isotope effect.

20. Richard JP, Tsuji Y. J. Am. Chem. Soc 2000;122:3963–3964.

**Scheme 1.**

**Scheme 2.**

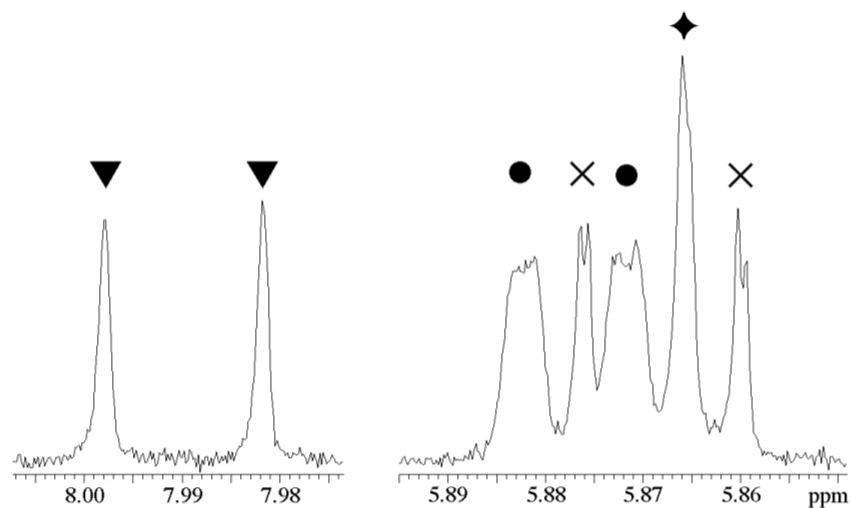
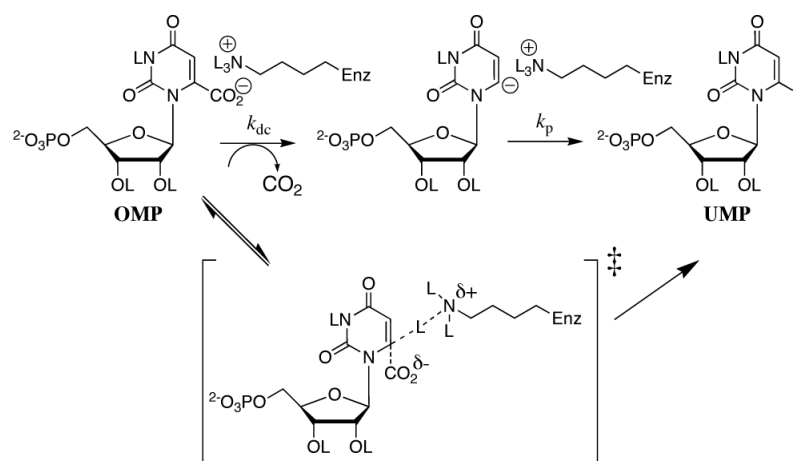


Figure 1. Partial ^1H NMR spectrum (500 MHz) of **UMP** from decarboxylation of **OMP** (2 mM) catalyzed by OMPDC from *S. cerevisiae* (24 nM) in 50/50 (v/v) $\text{H}_2\text{O}/\text{D}_2\text{O}$ at pH 7.3 and 25°C . Key: (▼) Doublet due to the C-6 proton of $[6\text{-}^1\text{H}]\text{-UMP}$; (●) Doublets (not resolved) due to the anomeric protons of $[6\text{-}^1\text{H}]\text{-UMP}$ and $[6\text{-}^2\text{H}]\text{-UMP}$; (×) Doublet due to the C-5 proton of $[6\text{-}^1\text{H}]\text{-UMP}$; (✧) Singlet due to the C-5 proton of $[6\text{-}^2\text{H}]\text{-UMP}$.



Scheme 3.