

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/258921320>

Modulation of Quinone PCET Reaction by Ca^{2+} Ion Captured by Calix[4]quinone in Water

ARTICLE in JOURNAL OF THE AMERICAN CHEMICAL SOCIETY · NOVEMBER 2013

Impact Factor: 12.11 · DOI: 10.1021/ja410406e · Source: PubMed

CITATIONS

2

READS

79

9 AUTHORS, INCLUDING:



Yang-Rae Kim

Seoul National University

45 PUBLICATIONS 800 CITATIONS

SEE PROFILE



Rebecca Soyoung Kim

Seoul National University

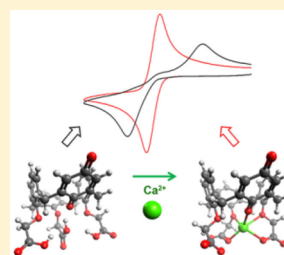
6 PUBLICATIONS 8 CITATIONS

SEE PROFILE

Modulation of Quinone PCET Reaction by Ca^{2+} Ion Captured by Calix[4]quinone in WaterYang-Rae Kim,^{†,‡} R. Soyoung Kim,^{†,‡} Sun Kil Kang,^{†,§} Myung Gil Choi,^{||} Hong Yeong Kim,^{||} Daeheum Cho,[⊥] Jin Yong Lee,^{*,⊥} Suk-Kyu Chang,^{*,||} and Taek Dong Chung^{*,†}[†]Department of Chemistry, Seoul National University, Seoul 151-747, Korea^{||}Department of Chemistry, Chung-Ang University, Seoul 156-756, Korea[⊥]Department of Chemistry, Sungkyunkwan University, Suwon 440-746, Korea

S Supporting Information

ABSTRACT: Calix[4]arene-triacid-monoquinone (CTAQ), a quinone-containing water-soluble ionophore, was utilized to investigate how proton-coupled electron transfer (PCET) reactions of quinones were influenced by redox-inactive metal ions in aqueous environment. This ionophoric quinone derivative captured a Ca^{2+} ion that drastically altered the voltammetric behavior of quinone, showing a characteristic response to pH and unique redox wave separation. Spectroelectrochemistry verified significant stabilization of the semiquinone, and electrocatalytic currents were observed in the presence of Ca^{2+} -free CTAQ. Using digital simulation of cyclic voltammograms to clarify how the thermodynamic properties of quinones were altered, a simple scheme was proposed that successfully accounted for all the observations. The change induced by Ca^{2+} complexation was explained on the basis of the combined effects of the electrostatic influence of the captured metal ion and hydrogen bonding of water molecules with the support of DFT calculation.



■ INTRODUCTION

Electron transfer (ET) processes are often coupled to the transfer of positively charged species, e.g. protons or metal ions, to avoid unstable states. When the positively charged entity is a proton, such an electron transfer is called proton-coupled electron transfer (PCET), which has been extensively studied and reviewed.^{1–3} Analogously, redox-inactive metal ions in the vicinity of redox centers can also influence ET. The term metal ion-coupled electron transfer (MCET) has been coined recently in analogy to PCET,^{4–6} as both protonation and metal ion binding bring about a positive shift in the reduction potential that depends on $\text{p}K_{\text{a}}$ or the binding constant difference.⁷ Redox-inactive metal ions such as Mg, Ca, and Zn ions are important components in enzymes; in addition to providing structural support, they sometimes reside at active sites and take part in catalysis by activating the substrate electrophilically.⁸ MCET has also inspired electrochemical energy storage (EES) research to explore the potential of organic molecules that bind with metal ions upon reduction.^{9,10}

Naturally, it is not surprising that PCET processes would be altered in the presence of metal ions. However, not many studies have aimed to probe the simultaneous influence of protons and metal ions on ET, i.e. how metal ions affect PCET, not just ET, processes. A few examples come from computational works: a DFT study of formamide self-exchange reaction showed that PCET was changed to hydrogen-atom transfer (HAT) by metal ion coordination, which shifted the ET site from the oxygen pair to the nitrogen pair.¹¹ Similarly, the metal ion was shown to influence radical type and ET channels in imide PCET reactions,¹² and nucleoside acidity was found to

be heightened by metal ion complexation.¹³ As for experimental works, flavin *E*-pH diagrams in aqueous environment were found to be transformed by the addition of metal ions,¹⁴ and more recently the influence of metal ions, protonation, and hydrogen bonding on flavin was investigated in aprotic solutions.¹⁵ Though these imply that PCET reactions may be affected by metal ions in interesting ways, presumably because of the complicated interpretation of results, most studies were confined to MCET or PCET alone, and a systematic experimental approach to elucidate metal ion influence on PCET is largely absent as yet.¹⁶

Quinone reduction/oxidation is a classic example of PCET. Because of their important roles in nature and the interesting chemical properties of the redox reactions of various quinones in different solvents and pH range, extensive literature is present.^{17–22} The $\text{p}K_{\text{a}}$ values of quinone, semiquinone, and hydroquinone are vastly different, making the shift in redox potential discriminatively sensitive to proton transfer or even hydrogen bonding.^{23–27} Electrochemistry of quinones is also sensitive to metal ion binding. Metal ions were found to stabilize the semiquinone radical anion ($\text{Q}^{\bullet-}$),²⁸ enhancing intermolecular^{29–32} and intramolecular^{33,34} one-electron transfers to quinone in aprotic solvents. There is a biological example that utilizes the quinone–metal ion interaction; a series of dehydrogenases contain a Ca^{2+} –pyrroloquinoline quinone (PQQ) complex in the active sites.³⁵ The removal of Ca^{2+} ion resulted in loss of activity because electron transfer

Received: October 10, 2013

Published: November 25, 2013



from the substrate to the quinone cofactor required the presence of a Ca^{2+} ion.³⁶ Searching for studies that probed metal ion influence on quinone PCET reactions in water, an electrochemical investigation of coenzyme Q (CoQ) derivatives was found to show that hydroxylated CoQ gives rise to a Ca^{2+} -sensitive/pH-insensitive redox peak, whereas the native CoQ shows a Ca^{2+} -insensitive/pH-sensitive redox peak.³⁷ However, in this example, proton and Ca^{2+} hardly compete with each other because hydroxyl and native CoQ have completely different properties in terms of pK_a and metal ion binding strength. Some studies observed that quinone autooxidation by oxygen was enhanced by metal ions,^{38–40} but they were mostly limited to observation of apparent kinetics and provided no comprehensive picture about how metal ions affect the PCET reactions. Moreover, in existing studies the high dielectric constant of water or even polar organic solvents required addition of an excess amount of metal ions so that complexities arose from metal ion concentration dependency and different binding modes,³¹ and in nearly all cases quinones had to be ortho or hydroxylated to chelate metal ions.

For a straightforward assessment of metal ion influence on PCET in aqueous solutions, it would be desirable to anchor the metal ion near the redox center rather than let it ion-pair with it upon ET, eliminating the metal ion concentration dependence and simplifying the redox reaction. In this study, we utilized a quinone-containing water-soluble ionophore as a model compound, calix[4]arene-triacid-monoquinone (CTAQ, Figure 1). Calixquinones were first synthesized in 1989⁴¹ and were

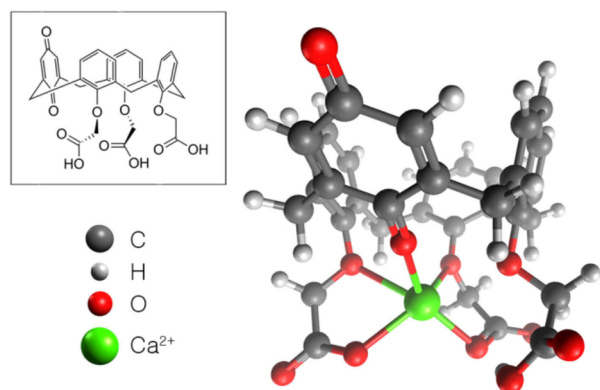


Figure 1. Structures of (inset, 2D) CTAQ and (3D) CaCTAQ. The geometry of CaCTAQ was optimized at MO6/6-31+G**. Two-dimensional (2D) and three-dimensional (3D) images were drawn with ChemDraw and Avogadro, respectively.

expanded in diversity^{42,43} to a water-soluble version through the attachment of carboxylic acid groups to the lower rim.⁴⁴ These pendant carboxylates bind most metal ions effectively, and little metal ion concentration dependency was found at >1 equiv of metal ions. Voltammetric behavior of CTAQ was also significantly altered by the presence of metal ions. Surprisingly, in alkaline-buffered solutions, binding of some metal ions such as Ca^{2+} caused the single two-electron redox wave of the quinone to split into two distinct waves that were attributed to separate one-electron transfer steps.⁴⁵ To the best of our knowledge, this is very unique in that such a clear separation of electron transfer steps to quinone in buffered aqueous solutions has not been reported elsewhere. In spite of these interesting attributes of water-soluble calixquinone and its metal ion complexes, rigorous investigation of its PCET behavior was put

off up to this day. This may be due to the fact that researchers were primarily concerned with analytical applications regarding calixquinones.

Here we give a detailed account of the electrochemistry of CTAQ and Ca^{2+} -complexed CTAQ (CaCTAQ) in aqueous environment. Along with the biological significance of Ca^{2+} ,^{46,47} we chose to focus on Ca^{2+} after preliminary experiments with other metal ions, as will be explained below. Electrochemical methods were primarily employed as they are useful for thermodynamic investigation of PCET.²² The nature of the unusual separation of faradaic reaction steps was verified with electrochemical and spectroscopic evidence. Digital simulation of voltammograms yielded quantitative estimates of the thermodynamic parameters involved, and simple arguments based on those parameters led to the observation of electrocatalytic currents at less than 1 equiv of Ca^{2+} . The change induced by Ca^{2+} pointed to the crucial contribution of the surrounding aqueous environment, in addition to the effect of Ca^{2+} itself.

■ EXPERIMENTAL DETAILS

Reagents. *tert*-Butyl bromoacetate, barium oxide, thallium(III) nitrate trihydrate, trifluoroacetic acid (TFA), 2-(cyclohexylamino)-ethanesulfonic acid (CHES), 3-(cyclohexylamino)-1-propanesulfonic acid (CAPS), 4-(2-hydroxyethyl)-piperazine-1-ethanesulfonic acid (HEPES), and 2-(*N*-morpholino)ethanesulfonic acid (MES) were purchased from Aldrich. Tetraethyl ammonium hydroxide (TEAOH) (20 wt% in aqueous solution) was obtained from Acros Organics. Calix[4]arene was procured from Janssen and used without further purification. Chloride salts of metal ions were obtained from Aldrich. All aqueous solutions were prepared using deionized water purified in a Nano Pure System (Barnsted). Column chromatography was performed using silica gel 60 (230–400 mesh, ASTM, Merck).

Caution: Thallium salts are toxic and must be handled with care.

Synthesis. CTAQ was synthesized according to procedures previously reported by our group (Scheme S1 in SI).⁴⁴ Tri-*tert*-butyl-calix[4]arene **2** was obtained by selective trialkylation of calix[4]arene **1** using *tert*-butyl bromoacetate and BaO in *N,N*-dimethylformamide (DMF) and BaO in dichloromethane. The phenol moiety of **2** was oxidized to quinone using thallium(III) nitrate trihydrate in methanol and chloroform mixture (1:1, v/v). Finally, CTAQ **4** was prepared by hydrolysis of the tri-*tert*-butyl ester monoquinone derivative **3** using TFA in dichloromethane. CTAQ is moderately soluble in basic aqueous solution, acetone, and methanol.

Electrochemistry. Electrochemical experiments were performed using a Windows-driven electrochemical analyzer (BAS100B/W, Bioanalytical Systems) employing positive-feedback routines to compensate for resistance. The surface of the glassy carbon working electrode (3 mm dia., BAS) was polished with 0.3 and 0.05 μm alumina (Buehler, Lake Bluff). An Ag/AgCl (in 3 M NaCl, 0.209 V vs NHE) reference electrode and a platinum wire counter electrode were used for voltammetric experiments. Solutions with pH values of 5.7, 6.2, 6.6, 7.0, 7.4, 7.8, 8.2, 8.6, 9.0, 9.4, 9.8, 10.2, 10.6, 11.0, and 11.4 were used. The voltammetry experiments were carried out in 0.1 M MES buffer (pH 5.7, 6.2, 6.6), HEPES buffer (pH 7.0, 7.4, 7.8, 8.2), CHES buffer (pH 8.6, 9.0, 9.4, 9.8), and CAPS buffer (pH 10.2, 10.6, 11.0, 11.4), respectively. The pH values of the buffer solutions used in the experiment were adjusted by adding hydrochloric acid (HCl) solution or TEAOH solution. Stock solutions of metal ions added to CTAQ solutions were in the form of 50 mM chloride salt. Ultrapure N_2 gas was bubbled through the solutions in all electrochemical experiments. All experiments were carried out at ambient temperature. Digital simulations of cyclic voltammograms were performed using the software package *DigiElch - Professional* (version 7.FD) purchased from Gamry Instruments; details of simulation conditions are given in the Supporting Information (SI).

Spectroelectrochemistry. All absorption UV–visible spectra were obtained with a Cary 60 UV–vis spectrophotometer (Agilent). A SEC-C05 thin layer quartz glass spectroelectrochemical cell (ALS, path length 0.5 mm) with a Teflon cap contained the solution and electrodes. An SEC-C gauze working electrode (ALS) was used along with an Ag/AgCl reference electrode and a Pt wire counter electrode. Spectra were recorded every 6 s during electrolytic reduction. The solution was purged with ultrapure N₂ prior to electrolysis.

Electron Paramagnetic Resonance. The model JES-TE200 (JEOL) operating at X-band (9.4 GHz) and employing 100 kHz modulation was used. Solution at pH 7.4 containing 1.0 mM CTAQ and one-equivalent of Ca²⁺ was bulk electrolyzed under ultrapure N₂ purging for 700 s at −0.6 V (vs Ag/AgCl) utilizing Pt as working and counter electrodes. The solution was transferred to a flat cell, and the EPR spectrum was recorded after 18 min from the end of electrolysis. Electrolysis and EPR measurement were conducted at ambient temperature.

DFT Calculation. All of the geometries of the free CTAQ and metal ion-bound CTAQ (MCTAQ) were optimized at the DFT level of theory (M06 hybrid-meta functional⁴⁸) together with LANL2DZ effective core potential (ECP)⁴⁹ for Sr and Ba atoms and 6-31+G** basis set for the other atoms. M06 functional is well-known to exhibit good performance on various systems such as main group thermochemistry, kinetics, organometallics, and especially noncovalent interaction.⁴⁸ In addition, we employed diffuse function basis set for better description of noncovalent interactions between guest metal ion and carboxylic oxygen atoms and hydrogen bonding between the carboxylic acid groups.⁵⁰ All calculations were carried out using a suite of Gaussian 09 software packages.⁵¹

RESULTS AND DISCUSSION

Electrochemical Behavior of CTAQ and CaCTAQ. The cyclic voltammograms of quinones in buffered aqueous solutions, in general, show a quasi-reversible pair of redox waves with relatively large peak-to-peak separation. Attributed to the coupled transfer of protons, this is what we see for CTAQ (Figure 2a, black curves). The only redox-active moiety of CTAQ is the quinone in the potential window employed. Therefore, CTAQ undergoes two-electron/two-proton redox reaction like a simple *p*-quinone, though the details of its reaction may be affected by the pendant carboxylic acid groups, as will be discussed later.

When one-equivalent of Ca²⁺ was added to CTAQ solution, however, the voltammogram was completely transformed both in neutral and basic solutions (Figure 2a, red curves). Acidic condition was avoided as CTAQ molecules became protonated and precipitated. First, at pH 7.4, peak-to-peak separation was drastically reduced from ~400 mV to ~80 mV, and sharp, symmetric peaks appeared in both cathodic and anodic scans. At pH 10.2, even more surprisingly, the single two-electron wave of CTAQ is split into two peaks as observed in aprotic solvents, strongly implying that sequential one-electron transfers occur.

In order to clarify the electrochemical behavior of CaCTAQ, cyclic voltammetry was carried out at various pHs ranging from slightly acidic to basic and midpoint potentials (E_m) were plotted against pH (Figure 2b, see Figure S1, SI for the actual voltammograms). E_m simply indicates the average of cathodic and anodic peak potentials and roughly corresponds to the standard reduction potentials for the redox couple involved. The resulting diagram showed a remarkably systematic variation of E with pH, and redox reactions of CaCTAQ could be inferred from it (eqs 1–3). In the low-pH region, E_m decreased with increasing pH at a slope close to −59 mV/pH, as it should for two-electron/two-proton transfer. The single peak at low pH splits into two at higher pH, beginning at

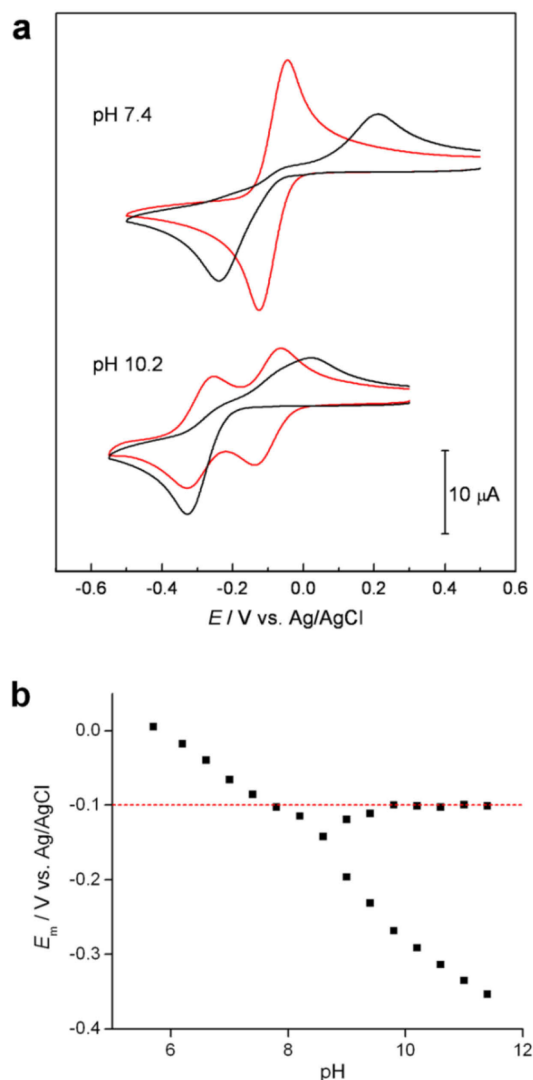
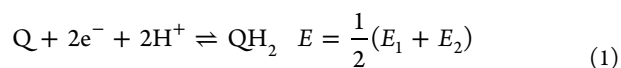


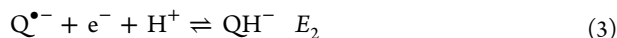
Figure 2. (a) Cyclic voltammograms of (black) CTAQ and (red) CaCTAQ at pH 7.4 and 10.2. Scan rate was 50 mV s^{−1}. (b) The pH dependence of the midpoint potentials (E_m) of CaCTAQ. Midpoint potentials were obtained from cyclic voltammograms of 1.0 mM CTAQ solution in the presence of one equivalent of Ca²⁺ (Figure S1, SI). Around the branching point (8.2 < pH < 9.0), the close distance between peaks hindered accurate determination of E_m from cyclic voltammograms.

around pH 8.6. In the high-pH region, the first peak remained nearly constant, whereas the second peak decreased with a slope of about −59 mV/pH (E -pH diagram with superimposed lines of the designated slopes is shown in Figure S2, SI). Presuming that each peak involves sequential one-electron transfer, the pH-independent first peak should involve only one-electron transfer without proton transfer, whereas the second should involve one-electron/one-proton transfer. Such interpretation of the E -pH diagram is summarized below. E_1 and E_2 each designate the standard potentials for the first and second one-electron reduction at each pH, i.e. $E_1 = E_{Q/Q\bullet-}$ and $E_2 = E_{Q\bullet-/QH_2}$ or $E_{Q\bullet-/QH-}$ or an intermediate value, depending on the pH.

pH < 8.6



pH > 8.6



It should be noted that, as mentioned earlier, such a clear separation of the two redox steps for quinones in aqueous solutions is highly unusual. Due to the remarkable stabilization of the reduced quinone by protonation, the second electron transfer mostly follows the first reduction immediately and produces a single two-electron peak. Even in the absence of protonation, strong hydrogen bonding of water molecules disallows the separation of the two redox steps.²³

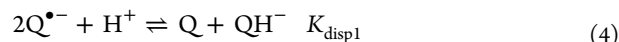
In fact, the appearance of two peaks in itself does not guarantee sequential one-electron transfers. In unbuffered solutions at a pH where proton concentration is comparable to that of the quinone, two peaks were observed and it was once interpreted as separate one-electron transfers.⁵² But it was later revealed that the two peaks each corresponded to two-electron reductions with different fates of the reduced species, namely protonation (QH_2) or stabilization by hydrogen bonding ($Q^{2-} \cdots (H_2O)_n$).²³ It is also possible to conceive an alternative interpretation of Figure 2b, assigning the pH-independent peak at -0.1 V to hydrogen-bond stabilized two-electron transfer and the other peak with a slope of -59 mV/pH to two-electron/two-proton transfer. However, for CaCTAQ, we confirmed the number of electrons transferred with chronoamperometry (see SI), and the results of the following section supports the extraordinary one-electron scheme as well.

Incidentally, the influence of other redox-inactive metal ions was also investigated. Other alkaline earth metal ions showed similar trends with different degree of influence increasing in the order of $Mg^{2+} < Ba^{2+} < Sr^{2+} < Ca^{2+}$ (Figure S3, SI). The cyclic voltammogram of SrCTAQ was nearly identical to that of CaCTAQ, and Ba^{2+} showed intermediate behavior. As the binding constants of Ca^{2+} , Sr^{2+} and Ba^{2+} with CTAQ are all in the 10^6 range,⁵³ this difference may be ascribed to their different Lewis acidities. Mg^{2+} had little effect, presumably owing to tight hydration⁴⁶ that prevents effective host–guest interaction with CTAQ. Alkali metal ions (Li^+ , Na^+ , K^+ , Rb^+ , Cs^+) had little effect,⁴⁴ even at >50 -equivalent, though their presence in large excess affected the voltammograms of MCTAQ ($M = Ca, Sr$ or Ba), indicating that they competitively occupy the binding pocket of CTAQ when present in large amounts. Unfortunately, biologically significant Zn^{2+} was bound too tightly and formed precipitates with CTAQ. Therefore, we focused our attention on CaCTAQ in this study.

Stabilization of Semiquinone Radical Anion by Ca^{2+} .

Separation of the one-electron reductions of quinone implies that the intermediate semiquinone is stable. The stability of the semiquinone radical anion, $Q^{\bullet-}$, can be quantitatively expressed by the equilibrium constant for the disproportionation/comproportionation reaction (eq 4), which is kinetically very facile for quinones.⁵⁴ K_{disp} is related to the rate of hydroquinone autoxidation by O_2 and superoxide production in biological systems and thus has been of much interest.^{55,56} In the literature, K_{disp} values for various quinones in aqueous solutions were estimated from the equilibrium concentration of $Q^{\bullet-}$ determined by EPR.^{57,58} However, for CaCTAQ, the separation of E_1 and E_2 at basic pH allows direct calculation of K_{disp} from electrochemical data.

The disproportionation reaction for CaCTAQ in the basic region above pH 8.6 can be written as below, involving one proton.



Because of the involvement of one proton, K_{disp1} is a function of pH. A modified parameter, $K_{disp1,pH}$, can be defined as follows.

$$K_{disp1,pH} = K_{disp1}[H^+] = \frac{[QH^-][Q]}{[Q^{\bullet-}]^2} \quad (5)$$

$$\log K_{disp1,pH} = -pH + \log K_{disp1} \quad (6)$$

Simple derivation with the Nernst equation gives eq 9 from eqs 5, 7 and 8.

$$E = E_{Q/Q^{\bullet-}} + \frac{RT}{F} \ln \frac{[Q]}{[Q^{\bullet-}]} \quad (7)$$

$$\begin{aligned} E &= E_{Q^{\bullet-}/QH^-} + \frac{RT}{F} \ln \frac{[Q^{\bullet-}][H^+]}{[QH^-]} \\ &= E_{Q^{\bullet-}/QH^-,pH} + \frac{RT}{F} \ln \frac{[Q^{\bullet-}]}{[QH^-]} \end{aligned} \quad (8)$$

$$\log K_{disp1,pH} = 2.303 \frac{RT}{F} (E_{Q^{\bullet-}/QH^-,pH} - E_{Q/Q^{\bullet-}}) \quad (9)$$

Because $E_{Q/Q^{\bullet-}}$ and $E_{Q^{\bullet-}/QH^-,pH}$ equals E_1 and E_2 at each pH in the basic region, $\log K_{disp1,pH}$ could be plotted against pH to give a linear plot (Figure 3) using the data in Figure 2b. The

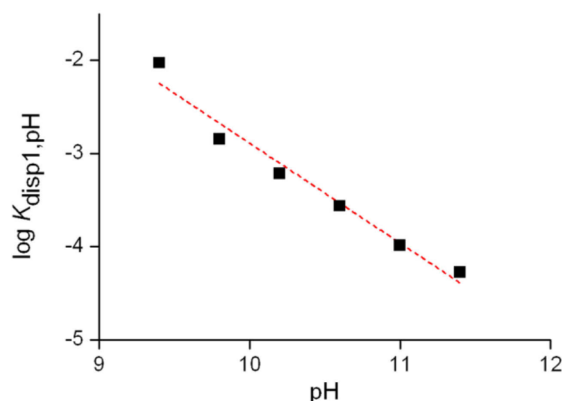


Figure 3. Plot of $\log K_{disp1,pH}$ against pH for CaCTAQ. The slope is -1.07 ($R^2 = 0.96$).

slope of -1.07 was in good agreement with eq 6. Extrapolating to neutral pH, $K_{disp1,pH}$ was estimated to be $10^{-0.1}$, or 0.8, at pH 7.4. Considering the fact that the major reduced species at pH 7.4 is QH_2 , and assuming $pK_{a,QH^-/QH_2}$ to be 8.6, the effective disproportionation constant at pH 7.4 was calculated to be $10^{1.1}$, or 13. (Note: eqs 1 – 3 suggest that $pK_{a,QH^-/QH_2}$ should be no larger than 8.6. Because a large pK_a value favors the right side of the equilibrium in eq 4, K_{disp1} is larger for higher pK_a value. It can be seen that, even when assuming the largest plausible value of pK_a for CaCTAQ, $K_{disp1,pH}$ is still very small.) This is 5 orders of magnitude smaller than the value of $\sim 10^6$ for 2,6-dimethyl-1,4-benzoquinone (DMBQ), which is a simple structural analogue for the quinone moiety of CTAQ.⁵⁷ The

value suggests that the semiquinone radical anion becomes more stable in the presence of Ca^{2+} .

The conclusion that $\text{CaCTAQ}^{\bullet-}$ is very stable even at pH 7.4 called for spectroscopic verification of this species. While cyclic voltammetry at neutral pH was inherently unable to confirm their existence, spectroelectrochemical experiment utilizing optically transparent thin layer electrochemical (OTTLE) cells⁵⁹ was expected to reveal the existence of $\text{Q}^{\bullet-}$ produced via comproportionation. Bulk electrolysis in neutral solution at a fixed potential was performed along with simultaneous observation of the UV-vis spectra.

The UV-vis absorption spectra in Figure 4 demonstrate the concentration changes of various species of CTAQ and

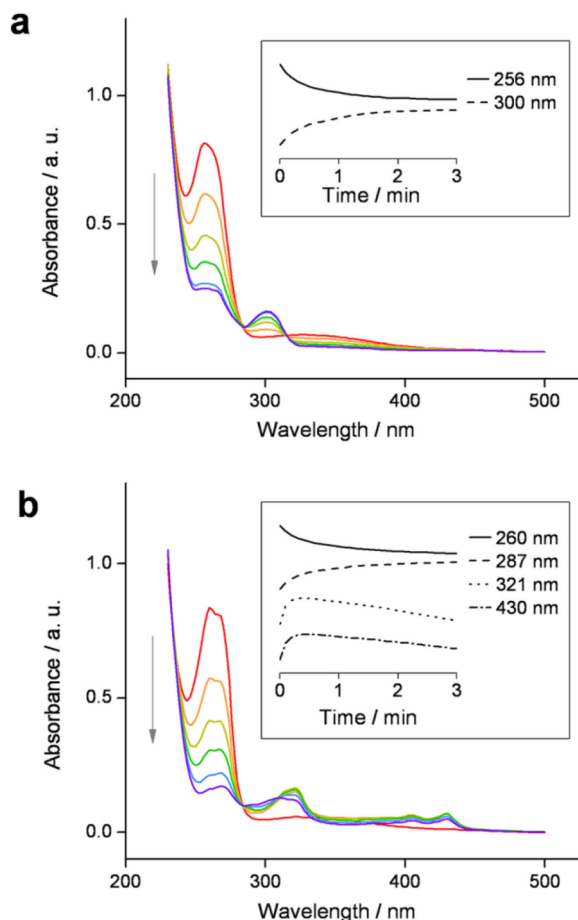


Figure 4. UV-visible absorption spectra obtained during electrochemical reduction of CTAQ in the (a) absence and (b) presence of one-equivalent of Ca^{2+} in pH 7.4 buffered solutions. The lines designate the absorbance spectra at 0, 12, 30, 60, 120, 180 s after the onset of electrolysis, respectively, in the order indicated by the arrow (i.e., from red to purple). (Inset) Time-dependent absorbance profiles at different wavelengths obtained during and after electrolysis. The absorbance values have been normalized for ease of comparison. Concentration of CTAQ was 1.0 mM and applied potential was -0.6 V vs Ag/AgCl.

CaCTAQ during electrolysis. In the absence of Ca^{2+} ion (Figure 4a), the appearance of clear isosbestic points implied that no intermediate species was present. The absorption peak at 256 nm rapidly decayed while the peak at 300 nm kept increasing during electrolysis. These two peaks were assigned to CTAQ and CTAQH_2 , respectively, as their wavelengths were

similar to those of 1,4-benzoquinone (BQ) and BQH_2 in the literature, which are 246 and 289 nm.⁶⁰

In the presence of one-equivalent of Ca^{2+} ion (Figure 4b), the absorption peak at 260 nm decreased in a similar way to the 256 nm peak of CTAQ, suggesting that the peak at 260 nm can be assigned to the unreduced CaCTAQ . However, unlike CTAQ, the isosbestic point above 300 nm was blurred and new peaks emerged at 321, 405, and 430 nm. These new peaks rose and fell during electrolysis, implying that these peaks represented intermediate species with a significantly long lifetime. The only plausible intermediate species in CaCTAQ reduction is the semiquinone radical anion, $\text{CaCTAQ}^{\bullet-}$. Moreover, these wavelengths were close to where those for the semiquinone radical anion of BQ reportedly appeared (~ 315 and ~ 430 nm).^{60,61} Because the electrode reaction produces the fully reduced CaCTAQH_2 , the semiquinone radical anion, $\text{Q}^{\bullet-}$, must have come from comproportionation between CaCTAQ and CaCTAQH_2 . The peak representing CaCTAQH_2 seemed to overlap with that of $\text{CaCTAQ}^{\bullet-}$ at 300 nm, but a small peak at 287 nm could be picked out and assigned to CaCTAQH_2 only, judging from its monotonous increase throughout the electrolysis. Hence, UV-vis spectra recorded during potentiostatic reduction of CTAQ and CaCTAQ imply that semiquinone becomes very stable in the presence of Ca^{2+} .

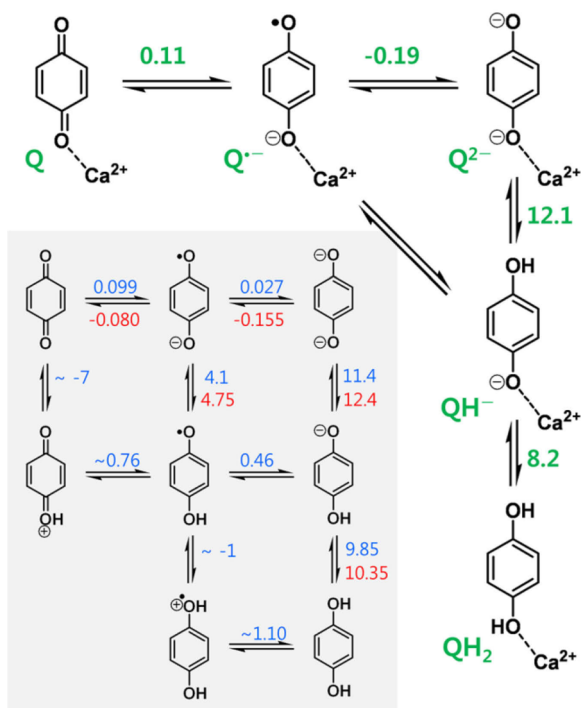
For added proof, the radical character of the transient species in UV-vis spectra was confirmed by EPR. The bulk-electrolyzed CaCTAQ solution showed a very strong and clear EPR signal that slowly decayed with time and was similar to that of $\text{BQ}^{\bullet-}$.^{61,62} (Figure S4, SI). Incidentally, an EPR signal was observed in the case of free CTAQ (not shown) as well but was far weaker in strength and barely resolved.

Digital Simulation of Cyclic Voltammograms of CaCTAQ .

The preceding sections revealed the unusual redox mechanism and semiquinone radical stability of CaCTAQ by investigating the cyclic voltammograms obtained at various pHs. Digital simulation of CaCTAQ voltammograms was performed in order to corroborate the proposed mechanism and obtain additional quantitative information. Scheme 1 summarizes the mechanism employed in the simulation. Considering the low pK_a of semiquinones, e.g. below 5 for DMBQ, the protonated neutral semiquinone radical, QH^\bullet , was expected to participate little in the pH region investigated. Moreover, the complexation of Ca^{2+} is expected to lower the pK_a of QH^\bullet further. Thus, the contribution of QH^\bullet was neglected in the simulation after verifying its incorporation had little effect for reasonable pK_a values. For the second electron transfer, which is a PCET, including a concerted proton-electron transfer (CPET) pathway enabled us to better reproduce the experimental voltammogram, in line with several recent reports on CPET.^{22,63–65} When modeling protonation steps, both specific and general acid catalysis were considered by using two reaction equations for each protonation reaction, one involving H^+ (equivalent to H_3O^+) and the other involving the acid form of the buffer species. The pH was set by adjusting the concentrations of the acid and base forms of the buffer. Diffusion coefficient for the CaCTAQ species was estimated from chronoamperometric currents recorded at an ultramicroelectrode (UME) (see SI). Full tabulation of all the reactions and associated parameters is available in SI (Table S1).

The values of the standard reduction potentials, acid dissociation constants and various unknown rate constants

Scheme 1. Partial Nine-Member Square Scheme for CaCTAQ Utilized in the Digital Simulation and Analogous Schemes for BQ and DMBQ^a



^aNumbers indicate E (V vs NHE) and pK_a values for (Outer, green) CaCTAQ, (inner, blue) BQ and (inner, red) DMBQ, which were obtained from simulation for CaCTAQ and from literature for BQ⁶⁶ and DMBQ^{56,67,68} (Note: some of these values are not directly obtained but calculated from other E and pK_a . Also, slightly different values were previously reported in the literature.^{57,69})

were determined by fitting the simulated voltammograms to the experimental data. As a rule, all protonation reactions were given a diffusion-limited rate⁷⁰ and disproportionation reaction rates were set by referring to the literature.⁵⁴ Applying an identical set of parameters to pH values ranging from 7.0 to 11.4 allowed us to narrow down the uncertainty in the obtained parameters. In fact, it is normally difficult to unambiguously simulate voltammograms of multiple electron transfers coupled to proton transfers because of the large number of unknown parameters involved, even if pH or scan rate is varied to check the validity of the simulation.^{22,71} However, for CaCTAQ, the separation of two-electron redox wave into one-electron waves at high pH enabled relatively straightforward simulation. E for the $Q/Q^{\bullet-}$ couple (E_1) was already evident from Figure 2b, and $pK_{a,QH-/QH_2}$ could be roughly estimated from it as well, as discussed previously. Initial guesses of other E and pK_a values were taken from those of DMBQ. As for estimating the individual heterogeneous rate constants, for which wildly disparate estimations exist in the literature, the wave separation at basic pH again helped. The ET rate constant for $Q/Q^{\bullet-}$, as it was a simple one-electron transfer, could be estimated directly by fitting. For the second ET, the concerted pathway was predominant so that k_{CPET} could also be directly estimated by fitting. The ET rate constant for $Q^{\bullet-}/Q^{2-}$ mattered little for reasonable values of k_{ET} , and was assumed to be the same as the k_{ET} for the first ET. Although plausible values were obtained for rate constants in this way, we do not claim these rate constants

to be accurate values, but rather focus on discussing the implications of the thermodynamic parameters.

As shown in Figure 5, good fit was achieved over the pH range employed. The peak potentials were well reproduced and

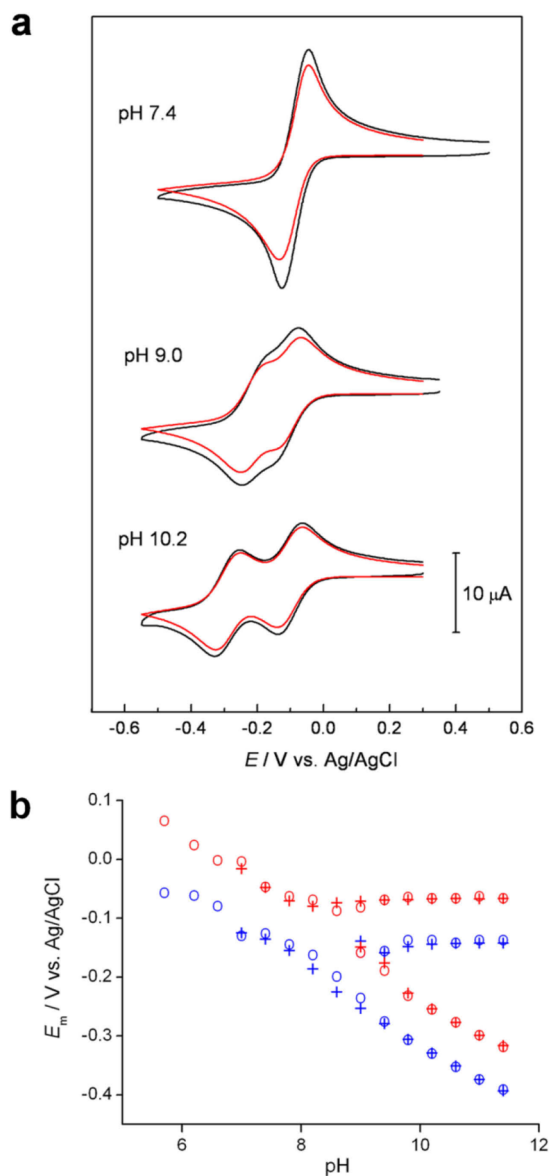


Figure 5. (a) (Red) Simulated and (black) experimental cyclic voltammograms of CaCTAQ. (b) (Red) Cathodic and (blue) anodic peak potentials of (+) simulated and (O) experimental voltammograms.

shapes of the cyclic voltammograms were similar in general. A small difference in the peak currents might be due to limited accuracy in the estimation of the diffusion coefficient. The Pourbaix diagram constructed from simulation results and overlaid over experimental data is given in Figure S2, SI. The good correlation overall again supports the validity of the proposed mechanism. Only the region between pH 5.7 and 6.6, where MES buffer was employed, did not show good agreement between experimental and simulated voltammograms. The protonated semiquinone, QH^{\bullet} , might have begun to appear in considerable amount at this low pH; or metal ion

complexation could have become weaker, as Ca^{2+} is known to be released from complexes in acidic condition.⁴⁷

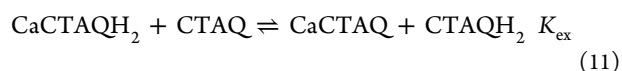
Now with the results of simulation at hand, we can compare Ca^{2+} -complexed quinone with free quinone in terms of thermodynamic parameters. For the sake of comparison with CaCTAQ, it was deemed appropriate to take the E and $\text{p}K_{\text{a}}$ values of DMBQ to represent those of Ca^{2+} -free quinone, on the basis of structural similarity and the fact that complexation of Ca^{2+} precludes the participation of carboxylic acid groups in the PCET of quinone. (Note: Simulation of CTAQ, which possesses three pendant carboxylic acid groups, was a formidable task; experimental E -pH diagram of CTAQ showed a slope of -50 mV/pH throughout the pH range investigated, slightly deviating from the theoretical value of -59 mV/pH, and simulation with the simple scheme used for CaCTAQ did not work, implying the participation of intramolecular acid-base equilibria, namely those of the pendant carboxylic acid groups.) According to Scheme 1, $E_{\text{Q}^{\bullet-}/\text{Q}^{2-}}$ becomes more positive and $E_{\text{Q}^{\bullet-}/\text{Q}^{2-}}$ more negative in the presence of Ca^{2+} . Also, $\text{p}K_{\text{a,QH}/\text{QH}_2}$ is 2 orders of magnitude smaller for CaCTAQ than for DMBQ. Calculating E_2 from solution pH, E and $\text{p}K_{\text{a}}$ values of QH_2 (eq 10), E_1 and E_2 were found to shift ~ 200 mV positively and negatively, respectively, for CaCTAQ with respect to DMBQ at pH 7.4. E_1 and E_2 at pH 10.2 were also shifted positively and negatively. In eq 10, $\text{p}K_{\text{a}1}$ and $\text{p}K_{\text{a}2}$ each stands for $\text{p}K_{\text{a,QH}/\text{QH}_2}$ and $\text{p}K_{\text{a,Q}^{\bullet-}/\text{Q}^{\bullet}}$. Derivation for eq 10 is given in SI.

$$E_2 = E_{\text{Q}^{\bullet-}/\text{Q}^{2-}} + \frac{RT}{F} \ln \frac{[\text{H}^+]^2 + [\text{H}^+]K_{\text{a}1} + K_{\text{a}1}K_{\text{a}2}}{K_{\text{a}1}K_{\text{a}2}} \quad (10)$$

Such positive and negative shift in E_1 and E_2 comprise all the essence of the change induced by Ca^{2+} . The peak separation at basic pH is a natural result, because it is exactly the fact that E_2 is more positive than E_1 , namely potential inversion, that caused the two redox waves to overlap and result in a merged, two-electron peak. Also, as demonstrated by the calculation of K_{dispr} , more negative value of $E_2 - E_1$ directly translates into greater stability of the semiquinone (eq 9). Because Ca^{2+} shifts E_1 positively and E_2 negatively, it decreases the value of $E_2 - E_1$ and stabilizes the semiquinone.

Moreover, the vast difference between the voltammograms of CTAQ and CaCTAQ at neutral pH (Figure 2a) can be easily understood. Since the inherent ET rates of quinones are quite slow (0.005 cm s^{-1} for CaCTAQ), the onset potentials for reduction and oxidation occur near E_1 and E_2 , respectively.⁷² When there is a large potential inversion, i.e. $E_2 - E_1 \gg 0$, E_1 is much more negative than the overall thermodynamic reduction potential, E_{ave} , so a significant overpotential is required to drive reduction. The same applies to oxidation, as E_2 is much more positive than E_{ave} . In the literature, this concept is contained in the apparent rate constant k_{app} for the overall two-electron redox reaction, a parameter that is determined by the individual E , $\text{p}K_{\text{a}}$ values and solution pH, following the theoretical framework of Laviron.^{17,18} For interpreting voltammograms it seems more intuitive to think in terms of E_1 and E_2 . As $E_2 - E_1$ decreases, i.e. there is less potential inversion, the required overpotential decreases, and the overall, apparent kinetics for the electrode reaction is enhanced. This accounts for the transformation of the voltammogram induced by Ca^{2+} at neutral pH.

Electrocatalytic Reduction of CTAQ by Ca^{2+} . The enhanced kinetics of CaCTAQ implies that in principle CaCTAQ can act as an electron mediator for CTAQ. Because of the relative values of E_1 and E_2 , if a potential sweep is carried out in a solution containing both CTAQ and CaCTAQ species, the CaCTAQ species will always be reduced and oxidized before CTAQ or CTAQH₂. Also, CaCTAQ and CTAQ were found to easily undergo homogeneous reactions with each other exchanging electrons and protons (eq 11), just as their comproportionation and disproportionation reactions are very facile, and similarly to other simple quinones.⁷³ Therefore, CTAQ will be reduced and oxidized via CaCTAQ before sufficiently high electrode potential is reached for direct electron transfer at the electrode. In other words, CTAQ will be reduced and oxidized electrocatalytically.



The actual cyclic voltammograms, recorded after adding less than one-equivalent of Ca^{2+} to a solution of CTAQ at pH 7.4, are presented in Figure 6a. The high binding constant of CTAQ implies that CaCTAQ mole fraction should approximately equal the equivalence of Ca^{2+} added. As explained above, catalytic currents were observed on both forward and reverse scans. On the cathodic scan, even adding as little as 0.05 equiv of Ca^{2+} was enough to pull forward the reduction peak so that it nearly overlapped with the reduction peak for CaCTAQ.

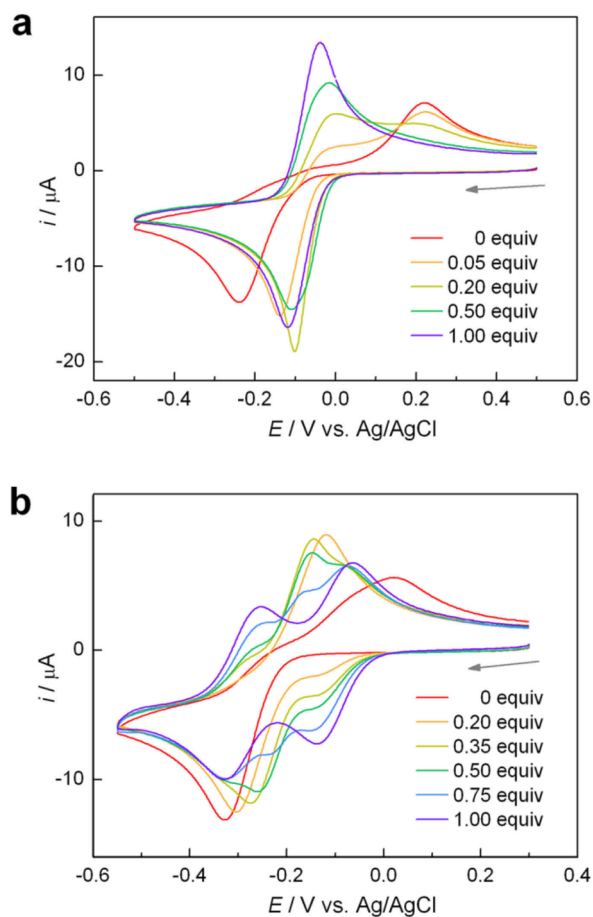


Figure 6. Cyclic voltammograms of 1.0 mM CTAQ in the presence of various amounts of Ca^{2+} in (a) pH 7.4 or (b) pH 10.2 buffered solution. Scan rate was 50 mV s^{-1} .

This indicates that the CaCTAQH₂ generated at the electrode surface rapidly reduces the free CTAQ molecules around it. Incidentally, both the peak current and peak potential implied more enhanced reduction at 0.20 equiv of Ca²⁺ than at 1 equiv. A possible explanation is that the electron relay from CaCTAQH₂ to CTAQ was faster than the diffusion of CaCTAQ to the electrode, so that the reduction current reached a maximum earlier when both CTAQ and CaCTAQ were present than when there were CaCTAQ only. On the anodic scan, two oxidation peaks are found that obviously correspond to Ca²⁺-complexed and Ca²⁺-free reduced quinones, but their relative heights did not reflect their molar ratio. For instance, at 0.20 equiv of Ca²⁺, the peak heights were close to 1:1 rather than 1:4. This indicates that the CaCTAQ oxidized at the electrode subsequently oxidizes CTAQH₂ around it.

Therefore, electrocatalytic activity of CaCTAQ was found on both scan directions, but it was clearly stronger during the cathodic scan. In other words, the equilibrium of eq 11 lies more toward the right. K_{ex} being greater than 1, the direct correlation between standard reduction potentials and equilibrium constants dictates that the overall reduction potential of CaCTAQ should be more negative than CTAQ (eqs 12 – 13).

$$K_{\text{ex}} = \frac{[\text{CaCTAQ}][\text{CTAQH}_2]}{[\text{CaCTAQH}_2][\text{CTAQ}]} \quad (12)$$

$$\ln K_{\text{ex}} = \frac{2F}{RT}(E_{\text{ave,CaCTAQ}} - E_{\text{ave,CTAQ}}) \quad (13)$$

At pH 10.2, where wave separation occurs, the voltammograms with <1 equiv Ca²⁺ were symmetrically shaped and showed different features from those at pH 7.4 (Figure 6b). A close examination on the basis of previous discussions makes the mechanism behind it clear. The initial peak in either cathodic or anodic scan grows roughly in proportion with the mole fraction of CaCTAQ and thus implies little catalytic current. It is because this peak corresponds to the one-electron reduction or oxidation of CaCTAQ or CaCTAQH[•], producing the semiquinone CaCTAQ[•]. Its homogeneous reaction with either CTAQ or CTAQH₂ is unfavorable because of the high instability of the semiquinone CTAQ[•] that will be produced. The CaCTAQ species act as an electron mediator only after their two-electron transfer products are formed. They may come from electrode reaction or disproportionation. As the mole fraction of CaCTAQ is increased, the peak for CTAQ shifts in potential so that less overpotential is required, and decreases in height at the same time. The potential shift is due to the catalytic action of CaCTAQ species, as explained for the case of neutral pH. The decrease in peak height, on the other hand, follows the decreasing mole fraction of the Ca²⁺-free quinone. This can be understood considering that unlike in neutral solution, the peaks for catalytic redox of CTAQ and heterogeneous redox of CaCTAQ are not merged. It is because the potential for the second one-electron reduction of CaCTAQ (E_2) is more negative than that for the two-electron reduction of CTAQ (E_{ave}), and CaCTAQH[•] required for the homogeneous reduction of CTAQ is produced via disproportionation before the electrode potential reaches E_2 . Analogous reasoning can be applied to the oxidation reactions. In addition to deciphering the reactions involved in the voltammograms, it was inferred from their symmetric shape that K_{ex} is close to 1, i.e. that E_{ave} of CTAQ and CaCTAQ are similar at this pH.

Assuming $E_{\text{ave,CTAQ}} = E_{\text{ave,CaCTAQ}}$ at pH 10.2, E_{ave} of CTAQ was found to be ~60 mV more positive than the E_{ave} of DMBQ. This may be due to the stabilization of CTAQH₂ by the pendant carboxylate groups.

However, it is counterintuitive that E_{ave} of CaCTAQ is more negative than that of CTAQ at pH 7.4. The complexation of Ca²⁺, a metal cation, is making it harder to reduce the quinone. It is common sense in organic chemistry that complexation of a Lewis acid like Ca²⁺ makes the substrate electrophilic and more reducible. On the other hand, observing the color change of CTAQ solution over time implied otherwise. Nucleophilic attack by OH[•] is known to occur at unsubstituted quinones^{74,75} and to change the color of the quinone solution from yellow to brownish-red to violet-red.³⁷ Such color change was also observed with CTAQ, and its rate dramatically accelerated in the presence of Ca²⁺; at high pH, the solution turned brownish after less than 10 min from adding Ca²⁺. Hence, it is evident that Ca²⁺ renders the quinone moiety electrophilic and susceptible to nucleophilic attack by OH[•]. This apparent contradiction will be resolved as we clarify the influence of Ca²⁺ in the next section.

Elucidating the Role of Ca²⁺ and Aqueous Environment in Shifting E_1 and E_2 . To summarize the results obtained so far, interesting new properties emerged when Ca²⁺ is bound to CTAQ. All of those observations are in essence results of positive and negative shift in E_1 and E_2 of the quinone. Reported many times in the literature from electrochemical and photoinduced electron transfer experiments, the positive shift of E_1 has been attributed to the Lewis acidity of Ca²⁺ and the stabilizing effect that it has on the semiquinone radical anion by electrostatic attraction. The term MCET designates this metal ion-assisted one-electron transfer to quinone. The negative shift of E_2 is a little bit less straightforward, and has appeared less frequently in the literature than the positive shift of E_1 . We could only find a single case that directly reported the negative shift of E_2 .³⁶ As for studies with less direct relevance, autooxidation of quinones by dissolved oxygen was reported to be generally enhanced by the presence of metal ions, as noted earlier.^{38,39,76} These studies explained their observations by assigning two roles to the metal ion: promoting deprotonation of the fully reduced quinone to destabilize it and stabilizing the semiquinone radical anion.

While these explanations are valid and roughly recapitulate the role of Ca²⁺ in CaCTAQ, our case requires a little more clarification. Since CTAQ is a *p*-quinone, the Ca²⁺ is bound to one of the oxygens only, whereas the Ca²⁺ was dually coordinated by the quinone in previous studies, as they all dealt with *o*- or hydroxy-quinones. Thus, the tendency to promote deprotonation is weaker in CaCTAQ. Indeed, the pK_a of CaCTAQH₂ compared to DMBQH₂ shows that Ca²⁺ is not very effective in hindering protonation; the oxygen on the other side is especially little affected (Scheme 1). Moreover, not only $E_{\text{Q}^{\bullet-}/\text{QH}_2}$ but also $E_{\text{Q}^{\bullet-}/\text{Q}_2^-}$ is more negative for CaCTAQ than DMBQ. As E for CTAQ seems to be even more positive than DMBQ (vide supra), it can be reasonably assumed that E_2 of CTAQ, both with and without protonation, became more negative by the complexation of Ca²⁺.

As for the stabilization of Q[•] by Ca²⁺, if it is wholly attributed to electrostatic effects, QH[•] and Q²⁻ should be equally stabilized by Ca²⁺ as well. However, as the thermodynamic redox potential depends on the relative stability of the reduced and oxidized forms, equal stabilization of the semiquinone and fully reduced quinone anions cannot induce a

negative shift in E_2 . Some literature has referred to the seemingly extraordinary stabilization of the semiquinone radical anion by metal ions with the term spin stabilizer.⁷⁷ But Ca^{2+} is a diamagnetic species, and our effort to identify any physical grounds for additional stabilization of $\text{Q}^{\bullet-}$ over Q^{2-} large enough to account for the negative shift of E_2 was unsuccessful. In fact, in aprotic solvents, metal ions were shown to stabilize Q^{2-} more than $\text{Q}^{\bullet-}$,^{30,33} in good accord with classic electrostatic considerations.⁷⁸ (Note: Ref 30 does not directly state that Q^{2-} is more stable than $\text{Q}^{\bullet-}$ when complexed with metal ion. However, upon metal ion addition, the one-electron transfer seems to become a two-electron transfer, which indicates that the positive shift of E_2 is larger than that of E_1 ; therefore, we can say that Q^{2-} was more stabilized than $\text{Q}^{\bullet-}$.)

Therefore, we rather attributed the extra stabilization of semiquinone or destabilization of fully reduced quinone by Ca^{2+} to the aqueous environment surrounding CaCTAQ and CTAQ. A survey of the literature on quinone electrochemistry indicates that the stabilizing effect of hydrogen bonding is weaker upon the semiquinone than upon the fully reduced quinone. In aprotic solvents, quinones show two clearly separated, one-electron waves, which move toward positive potential upon incremental addition of hydrogen bonding agents such as water.²⁶ The two waves eventually overlap because the second wave corresponding to $\text{Q}^{\bullet-} \rightarrow \text{Q}^{2-}$ moves much more rapidly than the first wave corresponding to $\text{Q} \rightarrow \text{Q}^{\bullet-}$. Actually, the first wave is scarcely shifted, indicating that $\text{Q}^{\bullet-}$ is much less stabilized by hydrogen bonding. Electrochemical study of quinones in unbuffered aqueous solutions also reported stronger hydrogen bonds formed on Q^{2-} than on $\text{Q}^{\bullet-}$.²³ Intramolecular hydrogen bonding is known to stabilize $\text{Q}^{\bullet-}$ considerably for some hydroxyl quinones, but this may as well be due to wider delocalization and not solely due to hydrogen bonding.^{79,80} In addition, documented pK_a values of QH^{\bullet} and QH_2 vastly differ and thus show that protonation is likewise less effective in stabilizing the radical than the fully reduced form. Parenthetically, this trend of extremely dissimilar pK_a values holds for the superoxide and peroxide couple as well (4.8⁸¹ and 11.6⁸² for HO_2^{\bullet} and H_2O_2 , respectively). In short, an aqueous environment is more effective in stabilizing the fully reduced quinone species than the semiquinone radical.

In CaCTAQ, hydrogen bonding as well as protonation to the quinone is partially blocked. As hydrogen bonding and protonation promote the stability of the fully reduced quinone more than that of the semiquinone, the net effect of Ca^{2+} in water will be to stabilize the semiquinone and destabilize the fully reduced quinone. Hence, by Ca^{2+} a negative shift in E_2 and an overall negative shift in E_{ave} result, although the quinone itself becomes electrophilic.

DFT calculations on the three redox states of CTAQ and MCTAQ ($\text{M} = \text{Ca}, \text{Sr}, \text{Ba}, \text{Mg}$) were also carried out to examine the validity of our explanation, and the results agreed with our discussion (see SI). Upon geometry optimization, a proton from one of the carboxylic acid groups of CTAQ coordinated to the quinone oxygen in an analogous manner to guest metal ions. The distance between the quinone oxygen and proton or metal ions in the quinone, semiquinone, and fully reduced quinone forms varied differently for CTAQ and MCTAQ and indicated that the acid groups were more effective in stabilizing the fully reduced quinone, whereas the metal ion was more effective in stabilizing the radical. The relative stability of each redox pair, assessed with the change in

quinone group charge density, also pointed to the same conclusion.

In essence, the fact that Ca^{2+} can stabilize $\text{Q}^{\bullet-}$ more than water but cannot for Q^{2-} , which is well-stabilized by water, gives rise to the interesting properties of CaCTAQ. However, if the Lewis acidity of the metal ion is so large that it can stabilize both $\text{Q}^{\bullet-}$ and Q^{2-} more than water, E_2 will not be shifted negatively. In fact, the initial, transient voltammogram of ZnCTAQ, which slowly precipitated out of the solution, implied such a possibility. We plan to further investigate this issue by synthesizing thiol-CTAQ that can be immobilized on an electrode surface to prevent precipitation. Incidentally, the intermediate Lewis acidity of Ca^{2+} may account for its various roles in nature, as implied by a recent study that suggested Ca^{2+} acts to modulate the redox potential in the oxygen-evolving complex (OEC) moderately.⁸³

CONCLUSIONS

In summary, CTAQ was taken as a model compound to study the influence of redox-inactive Ca^{2+} ion on quinones in buffered aqueous solutions. Its voltammetric response and properties were significantly altered in the presence of Ca^{2+} ; an unusual wave separation was seen at basic pH and observed for the first time for quinones in buffered aqueous solutions, and the semiquinone radical anion was extremely stabilized as shown through the large reduction in K_{disp} and spectroscopic observation. Digital simulation, which was straightforward owing to wave separation, yielded quantitative information to account for these results, namely, the positive and negative shift of E_1 and E_2 . It also foretold a catalytic current in the presence of less than one equivalent of Ca^{2+} arising from CaCTAQ species reducing and oxidizing CTAQ homogeneously. Then the shifts of E_1 and E_2 were explained by taking into account the surrounding aqueous environment, which has much stronger stabilizing effect on the dianion than on the radical, contrary to metal ions. The analysis presented in this work is expected to contribute to a clearer understanding of how metal ions influence PCET reactions in aqueous solutions. It is hoped that this will lead to interesting new applications that depend on tuning redox and acid–base properties, as hinted at by some of our observations.

ASSOCIATED CONTENT

Supporting Information

Synthesis procedure of CTAQ, cyclic voltammograms at various pHs and annotated Pourbaix diagram of CaCTAQ, cyclic voltammograms obtained with other metal ions, EPR spectrum for bulk-electrolyzed CaCTAQ, analysis of chronoamperometry data obtained with a UME, information about digital simulation, derivation of eq 10, and description of DFT calculation results. This material is available free of charge via the Internet at <http://pubs.acs.org>.

AUTHOR INFORMATION

Corresponding Authors

*tdchung@snu.ac.kr

*skchang@cau.ac.kr

*jinylee@skku.edu

Present Address

§Advanced Research Institute, LG Electronics Inc., Korea.

Author Contributions

‡Y.-R.K. and R.S.K. contributed equally.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

This work was supported by the Global Frontier R&D Program on Center for Multiscale Energy System (No. 2012M3A6A7055873) and the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT&Future (No. 2007-0056095, 2011-0030268 and No. 2012R1A2A1A03011289). We dedicate this paper in memorial to the late Dr. Su-Moon Park.

■ REFERENCES

- (1) Weinberg, D. R.; Gagliardi, C. J.; Hull, J. F.; Murphy, C. F.; Kent, C. A.; Westlake, B. C.; Paul, A.; Ess, D. H.; McCafferty, D. G.; Meyer, T. J. *Chem. Rev.* **2012**, *112*, 4016–4093.
- (2) Huynh, M. H. V.; Meyer, T. J. *Chem. Rev.* **2007**, *107*, 5004–5064.
- (3) Cukier, R. I.; Nocera, D. G. *Annu. Rev. Phys. Chem.* **1998**, *49*, 337–369.
- (4) Fukuzumi, S. In *Roles of Metal Ions in Controlling Bioinspired Electron-Transfer Systems. Metal Ion-Coupled Electron Transfer*; Prog. Inorg. Chem.; John Wiley & Sons, 2009; Vol. 56, pp 49–153.
- (5) Fukuzumi, S.; Ohkubo, K. *Coord. Chem. Rev.* **2010**, *254*, 372–385.
- (6) Fukuzumi, S. *Org. Biomol. Chem.* **2003**, *1*, 609–620.
- (7) Ge, Y.; Lilienthal, R. R.; Smith, D. K. *J. Am. Chem. Soc.* **1996**, *118*, 3976–3977.
- (8) Andreini, C.; Bertini, I.; Cavallaro, G.; Holliday, G. L.; Thornton, J. M. *J. Biol. Inorg. Chem.* **2008**, *13*, 1205–1218.
- (9) Lee, M.; Hong, J.; Seo, D.-H.; Nam, D. H.; Nam, K. T.; Kang, K.; Park, C. B. *Angew. Chem.* **2013**, *125*, 8480–8486.
- (10) Hernández-Burgos, K.; Rodríguez-Calero, G. G.; Zhou, W.; Burkhardt, S. E.; Abruña, H. D. *J. Am. Chem. Soc.* **2013**, *135*, 14532.
- (11) Chen, X.; Bu, Y. *J. Am. Chem. Soc.* **2007**, *129*, 9713–9720.
- (12) Chen, X.; Xing, D.; Zhang, L.; Cukier, R. I.; Bu, Y. *J. Comput. Chem.* **2009**, *30*, 2694–2705.
- (13) Aliakbar Tehrani, Z.; Fattahi, A.; Pourjavadi, A. *Sci. Iran.* **2012**, *19*, 535–545.
- (14) Ksenzhek, O. S.; Petrova, S. A. *Bioelectrochem. Bioenerg.* **1983**, *11*, 105–127.
- (15) Fukuzumi, S.; Kojima, T. *J. Biol. Inorg. Chem.* **2008**, *13*, 321–333.
- (16) Costentin, C.; Louault, C.; Robert, M.; Savéant, J.-M. *Proc. Natl. Acad. Sci. U.S.A.* **2009**, *106*, 18143–18148.
- (17) Laviron, E. *J. Electroanal. Chem. Interfac. Electrochem.* **1983**, *146*, 15–36.
- (18) Laviron, E. *J. Electroanal. Chem. Interfac. Electrochem.* **1982**, *134*, 205–212.
- (19) Gómez, M.; González, F. J.; González, I. *J. Electroanal. Chem.* **2005**, *578*, 193–202.
- (20) Guin, P. S.; Das, S.; Mandal, P. C. *Int. J. Electrochem.* **2011**, *2011*, 1–22.
- (21) Chambers, J. Q. In *The Chemistry of Quinonoid Compounds*; Patai, S.; Rappaport, Z., Eds.; John Wiley & Sons: Great Britain, 1988; Vol. II, pp 719–757.
- (22) Costentin, C.; Robert, M.; Savéant, J.-M. *Chem. Rev.* **2010**, *110*, PR1–PR40.
- (23) Quan, M.; Sanchez, D.; Wasylkiw, M. F.; Smith, D. K. *J. Am. Chem. Soc.* **2007**, *129*, 12847–12856.
- (24) Hui, Y.; Chng, E. L. K.; Chng, C. Y. L.; Poh, H. L.; Webster, R. D. *J. Am. Chem. Soc.* **2009**, *131*, 1523–1534.
- (25) Eggins, B. R.; Chambers, J. Q. *J. Electrochem. Soc.* **1970**, *117*, 186–192.
- (26) Gupta, N.; Linschitz, H. *J. Am. Chem. Soc.* **1997**, *119*, 6384–6391.
- (27) Tessensohn, M. E.; Hirao, H.; Webster, R. D. *J. Phys. Chem. C* **2013**, *117*, 1081–1090.
- (28) Eaton, D. R. *Inorg. Chem.* **1964**, *3*, 1268–1271.
- (29) Itoh, S.; Fukuzumi, S. *J. Mol. Catal. B: Enzym.* **2000**, *8*, 85–94.
- (30) Kawashima, T.; Ohkubo, K.; Fukuzumi, S. *Phys. Chem. Chem. Phys.* **2011**, *13*, 3344.
- (31) Yuasa, J.; Suenobu, T.; Fukuzumi, S. *ChemPhysChem* **2006**, *7*, 942–954.
- (32) Itoh, S.; Kawakami, H.; Fukuzumi, S. *J. Am. Chem. Soc.* **1998**, *120*, 7271.
- (33) Wu, H.; Zhang, D.; Su, L.; Ohkubo, K.; Zhang, C.; Yin, S.; Mao, L.; Shuai, Z.; Fukuzumi, S.; Zhu, D. *J. Am. Chem. Soc.* **2007**, *129*, 6839–6846.
- (34) Fukuzumi, S.; Okamoto, K.; Yoshida, Y.; Imahori, H.; Araki, Y.; Ito, O. *J. Am. Chem. Soc.* **2003**, *125*, 1007–1013.
- (35) Kay, C. W. M.; Mennenga, B.; Görisch, H.; Bittl, R. *J. Am. Chem. Soc.* **2005**, *127*, 7974–7975.
- (36) Sato, A.; Takagi, K.; Kano, K.; Kato, N.; Duine, J.; Ikeda, T. *Biochem. J.* **2001**, *357*, 893–898.
- (37) Bogeski, I.; Gulaboski, R.; Kappl, R.; Mirceski, V.; Stefova, M.; Petreska, J.; Hoth, M. *J. Am. Chem. Soc.* **2011**, *133*, 9293–9303.
- (38) Lebedev, A. V.; Ivanova, M. V.; Timoshin, A. A.; Ruuge, E. K. *ChemPhysChem* **2007**, *8*, 1863–1869.
- (39) Lebedev, A. V.; Ivanova, M. V.; Ruuge, E. K. *Arch. Biochem. Biophys.* **2003**, *413*, 191–198.
- (40) Itoh, S.; Kawakami, H.; Fukuzumi, S. *Chem. Commun.* **1997**, 29–30.
- (41) Morita, Y.; Agawa, T.; Kai, Y.; Kanehisa, N.; Kasai, N.; Nomura, E.; Taniguchi, H. *Chem. Lett.* **1989**, *18*, 1349–1352.
- (42) Gomez-Kaifer, M.; Reddy, P. A.; Gutsche, C. D.; Echegoyen, L. *J. Am. Chem. Soc.* **1994**, *116*, 3580–3587.
- (43) Chung, T. D.; Kim, H.-S. *J. Incl. Phenom. Mol. Recognit. Chem.* **1998**, *32*, 179–193.
- (44) Chung, T. D.; Kang, S.-K.; Kim, H.-S.; Kim, J. R.; Oh, W. S.; Chang, S.-K. *Chem. Lett.* **1998**, 1225.
- (45) Kang, S.-K.; Chung, T. D.; Kim, H.-S. *Electrochim. Acta* **2000**, *45*, 2939–2943.
- (46) Eugene, A. P.; Robert, H. K. In *Calcium Binding Proteins*; Wiley Series in Protein and Peptide Science; John Wiley & Sons: Singapore, 2011.
- (47) Yocum, C. F. *Coord. Chem. Rev.* **2008**, *252*, 296–305.
- (48) Zhao, Y.; Truhlar, D. G. *Theor. Chem. Acc.* **2008**, *120*, 215–241.
- (49) Hay, P. J.; Wadt, W. R. *J. Chem. Phys.* **1985**, *82*, 270.
- (50) Papajak, E.; Truhlar, D. G. *J. Chem. Theory Comput.* **2010**, *6*, 597–601.
- (51) Frisch, M. J. *Gaussian 09*, revision B.01; Gaussian Inc.: Wallingford, CT, 2009.
- (52) Shim, Y.-B.; Park, S.-M. *J. Electroanal. Chem.* **1997**, *425*, 201–207.
- (53) Kang, S.-K.; Lee, O.-S.; Chang, S.-K.; Chung, D.-S.; Kim, H.-S.; Chung, T.-D. *Bull. Korean Chem. Soc.* **2011**, *32*, 793–799.
- (54) Roginsky, V. A.; Pisarenko, L. M.; Bors, W.; Michel, C.; Saran, M. *J. Chem. Soc., Faraday Trans.* **1998**, *94*, 1835–1840.
- (55) Osyczka, A.; Moser, C. C.; Dutton, P. L. *Trends Biochem. Sci.* **2005**, *30*, 176–182.
- (56) Song, Y.; Buettner, G. R. *Free Radical Biol. Med.* **2010**, *49*, 919–962.
- (57) Roginsky, V. A.; Pisarenko, L. M.; Bors, W.; Michel, C. *J. Chem. Soc., Perkin Trans. 2* **1999**, 871–876.
- (58) Alegria, A. E.; Lopez, M.; Guevara, N. *J. Chem. Soc., Faraday Trans.* **1996**, *92*, 4965.
- (59) Kaim, W.; Fiedler, J. *Chem. Soc. Rev.* **2009**, *38*, 3373.
- (60) Zhao, X.; Imahori, H.; Zhan, C.-G.; Sakata, Y.; Iwata, S.; Kitagawa, T. *J. Phys. Chem. A* **1997**, *101*, 622–631.
- (61) Tang, Y.; Wu, Y.; Wang, Z. *J. Electrochem. Soc.* **2001**, *148*, E133.
- (62) Park, H.; Won, M.-S.; Cheong, C.; Shim, Y.-B. *Electroanalysis* **2002**, *14*, 1501–1507.
- (63) Anxolabéhère-Mallart, E.; Costentin, C.; Policar, C.; Robert, M.; Savéant, J.-M.; Teillout, A.-L. *Faraday Discuss.* **2010**, *148*, 83–95.
- (64) Medina-Ramos, J.; Oyesanya, O.; Alvarez, J. C. *J. Phys. Chem. C* **2013**, *117*, 902–912.

- (65) Song, N.; Gagliardi, C. J.; Binstead, R. A.; Zhang, M.-T.; Thorp, H.; Meyer, T. J. *J. Am. Chem. Soc.* **2012**, *134*, 18538–18541.
- (66) Laviron, E. *J. Electroanal. Chem.* **1984**, *169*, 29.
- (67) Bishop, C. A.; Tong, L. K. *J. Am. Chem. Soc.* **1965**, *87*, 501–505.
- (68) Patel, K. B.; Willson, R. L. *J. Chem. Soc. Faraday Trans. 1* **1973**, *69*, 814–825.
- (69) Wardman, P. *J. Phys. Chem. Ref. Data* **1989**, *18*, 1637.
- (70) Batchelor-McAuley, C.; Li, Q.; Dapin, S. M.; Compton, R. G. *J. Phys. Chem. B* **2010**, *114*, 4094–4100.
- (71) Batchelor-McAuley, C.; Kozub, B. R.; Menshynkau, D.; Compton, R. G. *J. Phys. Chem. C* **2011**, *115*, 714–718.
- (72) Compton, R. G.; Banks, C. E. *Understanding Voltammetry*; 2nd ed.; World Scientific Publishing: Singapore, 2007.
- (73) Laviron, E. *J. Electroanal. Chem.* **1986**, *208*, 357.
- (74) Pedersen, J. A. *Spectrochim. Acta, Part A* **2002**, *58*, 1257–1270.
- (75) Fukuzumi, S.; Nakanishi, I.; Maruta, J.; Yorisue, T.; Suenobu, T.; Itoh, S.; Arakawa, R.; Kadish, K. M. *J. Am. Chem. Soc.* **1998**, *120*, 6673–6680.
- (76) Zhang, L.; Bandy, B.; Davison, A. J. *Free Radical Biol. Med.* **1996**, *20*, 495–505.
- (77) Stegmann, H. B.; Bergler, H. U.; Scheffler, K. *Angew. Chem., Int. Ed. Engl.* **1981**, *20*, 389–390.
- (78) Evans, D. H.; Lehmann, M. W. *Acta Chem. Scand.* **1999**, *53*, 765–774.
- (79) Aguilar-Martinez, M.; Macías-Ruvalcaba, N. A.; Bautista-Martínez, J. A.; Gómez, M.; González, F. J.; González, I. *Curr. Org. Chem.* **2004**, *8*, 1721–1738.
- (80) Ashnagar, A.; Bruce, J. M.; Dutton, P. L.; Prince, R. C. *Biochim. Biophys. Acta* **1984**, *801*, 351–359.
- (81) Bielski, B. H. J.; Cabelli, D. E.; Arudi, R. L.; Ross, A. B. *J. Phys. Chem. Ref. Data* **1985**, *14*, 1041–1100.
- (82) Dean, J. A. *Lange's Handbook of Chemistry*, 14th ed.; McGraw-Hill: New York, 1992.
- (83) Tsui, E. Y.; Tran, R.; Yano, J.; Agapie, T. *Nat. Chem.* **2013**, *5*, 293–299.