

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/7283771>

Efficient Glycosidation of a Phenyl Thiosialoside Donor with Diphenyl Sulfoxide and Triflic Anhydride in Dichloromethane

ARTICLE *in* ORGANIC LETTERS · APRIL 2006

Impact Factor: 6.36 · DOI: 10.1021/ol060030s · Source: PubMed

CITATIONS

53

READS

18

2 AUTHORS, INCLUDING:



David Crich

Wayne State University

483 PUBLICATIONS 12,028 CITATIONS

SEE PROFILE

Published in final edited form as:

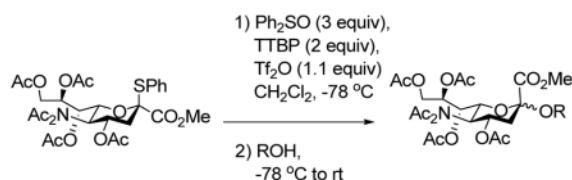
Org Lett. 2006 March 2; 8(5): 959–962. doi:10.1021/ol060030s.

Efficient Glycosidation of a Phenyl Thiosialoside Donor with Diphenyl Sulfoxide and Triflic Anhydride in Dichloromethane

David Crich and Wenju Li

Department of Chemistry, University of Illinois at Chicago, 845 West Taylor Street, Chicago, Illinois 60607-7061, Dcrich@uic.edu

Abstract

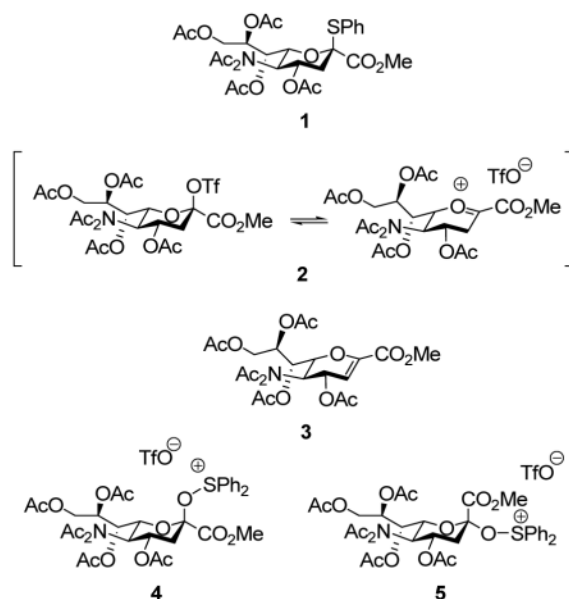


The formation of sialic glycosides using a thiosialic acid derivative, diphenyl sulfoxide and trifluoromethanesulfonic anhydride is reported. With an excess of diphenyl sulfoxide glycal formation can be completely suppressed and excellent yields are obtained for coupling to a wide range of primary, secondary and tertiary acceptors.

The sialic acids, discovered in vertebrates by Blix and Klenk in 1941, are a family of more than 40 2-keto-3-deoxy-nonulosonic acids that are incorporated in oligosaccharides and glycoconjugates which play important biological roles in high animals and human beings.¹ The lead member of the series, *N*-acetylneuraminic acid (Neu5Ac), is often found at the terminal positions of glycoproteins and glycolipids and connected to galactosides or 2-acetamido-galactosides by $\alpha(2\rightarrow3)$ or $\alpha(2\rightarrow6)$ linkages. The biological significance of the sialic acid glycosides combines with the widely acknowledged difficulty inherent in their synthesis to render this class of glycosidic bonds one of the most widely studied in the field of carbohydrate chemistry.^{2,3} Both the challenge and the importance of the problem are readily appreciated from the application of almost every major class of glycosidic bond forming reaction to the sialic acid glycosides in recent years.^{2,3,4} Thioglycosides are no exception to this rule, with most common promoters having been evaluated in this respect,^{2,3,4} with the exceptions of the recent 1-benzenesulfinyl piperidine (BSP)/triflic anhydride system developed in this laboratory,^{5,6} the related diphenyl sulfoxide (Ph₂SO)/triflic anhydride method as adapted to the thioglycosides by van Boom,^{6,7} and the Kahne sulfoxide method.^{6,8} We are interested in developing rapid, low temperature α -selective sialylations from thioglycoside donors, avoiding the use of acetonitrile and its stereodirecting effect, which is common to much work in the area,^{2,3,4,9} and report here on our preliminary observations.

Among the various 2-thiosialyl donors, we chose methyl 2- β -phenylthio-4,7,8,9-tetra-*O*-acetyl *N,N*-diacetylneuraminate **1**, because of its crystalline nature and enhanced selectivity, compared to the *N*-monoacetyl counterpart.^{3n,10} Initially, we investigated our BSP/Tf₂O/TTBP activation method,⁷ but saw no activation even after changing the hydroxy protecting groups from esters to more armed silyl ethers. We explored therefore the more potent promotion system Ph₂O/Tf₂O, which was developed by Gin for dehydrative glycosylation strategies,¹¹

including sialytations.^{11d} Hoping to generate the β -anomeric sialyl triflate as intermediate species, we first applied a stoichiometric quantity of the $\text{Ph}_2\text{SO}/\text{Trf}_2\text{O}$ combination in the presence of the mild, hindered base 2,4,6-tri-*tert*-butylpyrimidine (TTBP).¹² Unfortunately, while activation was complete at -78°C in a matter of minutes, it was followed by immediate and quantitative elimination to the glycal **3**. However, raising the amount of Ph_2SO to 3 equivalents, with otherwise unchanged conditions, we found that the coupling reaction proceeded in excellent yield and good selectivity on subsequent addition of isopropanol (Table 1, entry 1, 97%, 2.3:1 α/β). We postulate that the initial intermediate oxacarbenium ion/anomeric triflate pair **2** undergoes instantaneous decomposition to the glycal **3** in the absence of a suitable nucleophile. However, in the presence of excess sulfoxide this species is trapped to give two covalent sulfonium salts **4** and **5**, which serve as reservoirs for the oxacarbenium ion in the subsequent glycosylation. Low temperature NMR experiments in CD_2Cl_2 support this hypothesis, with two apparently isomeric species being observed in the ratio 1.5:1 when the reaction was conducted with 3 equiv of Ph_2SO .¹³ These intermediates, tentatively assigned to **4** and **5**, were immediately converted to the two methyl glycosides, also in the ratio 1.5:1 on addition of methanol at -78°C .



This protocol,¹⁴ employing three molar equivalents of diphenyl sulfoxide, was then applied to a range of acceptors (Table 1), with anomeric stereochemistry assigned on the basis of the chemical shift of the sialic acid H-3,¹⁵ H-4,¹⁶ and, preferably, the sialic acid $^3J_{\text{C1,H3ax}}$.¹⁷ With the exception of alcohol **18**, for which no coupled product was isolated, the isolated yields of these coupling reactions ranged from good to excellent. Stereoselectivity, on the other hand, varied considerably and was very substrate dependent. Optimal α -selectivities were obtained for the important galactose 6-OH (Table 1, entries 7–9) but, interestingly, considerably lower selectivity was obtained with a glucose 6-OH derivative (Table 1, entry 6). With the 3,4-diols no improvement in selectivity was seen on switching from the 2,6-di-*O*-benzyl ether **15** to the 2,6-di-*O*-propargyl ether, despite the reduced steric bulk of the latter acceptor (Table 1, entries 10 and 11).¹⁸ Perhaps not surprisingly, the selectivity was obtained with the less reactive 4-OH group (Table 1, entry 12). All in all, it is clear that correct stereochemical matching¹⁹ of the acceptor with the donor is important in these couplings, as is becoming increasingly apparent in glycosylation reactions in general.²⁰

With a view to improving the yield, and especially selectivities, we investigated the use of propionitrile as additives, and of several other sulfoxides in place of diphenyl sulfoxide (Table 2).

While a marginally better yield was obtained when the standard solvent was replaced by a 1:1 mixture of acetonitrile and dichloromethane, the $\alpha:\beta$ selectivity was diminished on coupling to isopropanol (Table 2, entries 1 and 2). The use of propionitrile alone as solvent also failed to bring about the desired improvement in selectivity (Table 2, entry 3). Reverting to dichloromethane as solvent, the replacement of diphenyl sulfoxide by the more electron-deficient 4-nitrophenyl phenyl sulfoxide brought about a small increase in selectivity (Table 2, entry 4), while the more electron-rich 4-methoxyphenyl phenyl sulfoxide gave no increase in selectivity and a substantially lower yield (Table 2, entry 5). The use of dibenzothiophene oxide, while giving only a 50% yield of coupled product under the standard activation conditions did result, however, in a significant increase in selectivity to 6:1 in favor of the α -glycoside with model alcohol isopropanol (Table 2, entry 6). The improvement of selectivity on replacing diphenyl sulfoxide by dibenzothiophene oxide must result from a differing population and/or reactivity of the glycosyl sulfonium ions corresponding to **4** and **5**. The relatively low yield obtained with dibenzothiophene oxide as activating agent resulted from poor conversion of the donor **1**, which is presumably due to reduced electrophilicity of its *O*-trifluoromethanesulfonylated adduct. Accordingly, we developed a protocol in which donor **1** was activated with Tf_2O in the presence of both diphenyl sulfoxide and dibenzothiophene oxide and applied to several coupling reactions (Table 3).

Unfortunately, the improvement in selectivity seen with isopropanol did not extend to any of the other examples investigated.

In summary, we have shown that the challenging α -sialylation with 2-thiosialic acid donors can be efficiently performed using the diphenyl sulfoxide/trifluoromethanesulfonic anhydride promotion system. The use of excess diphenyl sulfoxide is shown to be important in these couplings and serves to suppress the formation of the glycal by trapping the first-formed oxacarbenium ion, as suggested by low temperature NMR studies. Sialylations of a series of alcohol nucleophiles showed satisfactory yields and variable anomeric selectivities that are sensitive to the exact nature of the acceptors. With this new procedure, the Neu5Ac $\alpha(2\rightarrow6)$ Gal glycosidic linkages can be installed with excellent yield and selectivity. Sialylation of secondary sugar acceptors and even the simple tertiary alcohol 1-adamantanol proceeded with good yield but modest selectivity.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgements

We thank the NIH (GM 62160) for support of this work.

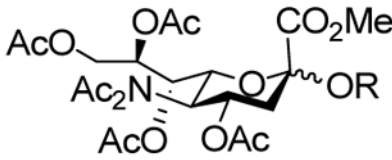

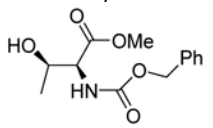
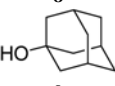
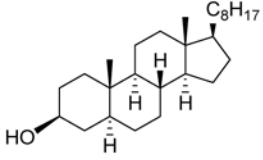
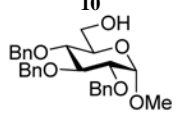
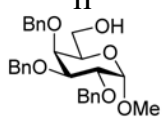
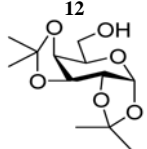
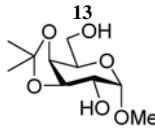
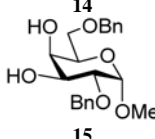
References

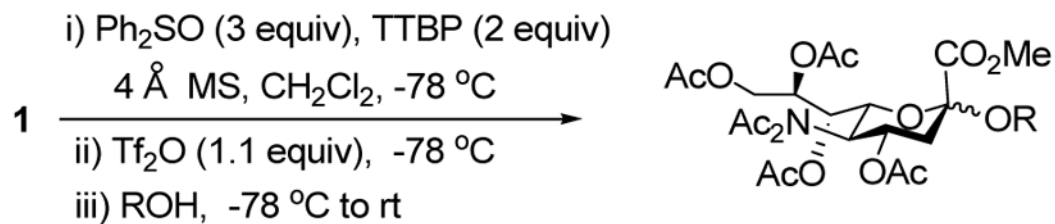
1. Schauer, R., editor. Sialic Acids: Chemistry, Metabolism and Function. 10. Springer-Verlag; Wien, New York: 1982. Rosenberg, A., editor. Biology of Sialic Acids. Plenum Press; New York: 1995.
2. Recent reviews: (a) Boons GJ, Demchenko AV. Chem Rev 2000;100:4539–4565. [PubMed: 11749357] (b) Ress DK, Linhardt RJ. Current Organic Synthesis 2004;1:31–46. (c) Boons GJ, Demchenko AV, Wong C-H. Carbohydrate-based Drug Discovery Wiley-VCH Weinheim 2003;155102 (d) Kiso M, Ishida H, Ito H, Ernst B, Hart G, W. Sinaý PC. Carbohydrates in Chemistry and Biology Wiley-VCH Weinheim 2000;1345365 (e) Halcomb RL, Chappell MD, Wang P, Bertozzi CR. Glycochemistry:

Principles, Synthesis, and Applications Dekker New York 2001:177220 (f) Toshima K, Tatsuta K. Chem Rev 1993;93:1503–1531. (g) Boons GJ. Contemp Org Synth 1996;3:173–200. (h) Roy R. Top Curr Chem 1997;187:241–274. (i) Hasegawa A Khan SHO'Niell R A Modern Methods in Carbohydrate Synthesis Harwood Amsterdam 1996:277300 (j) Hasegawa A Kiso M Hanessian S Preparative Carbohydrate Chemistry Dekker New York 1997:357379

3. Recent examples: (a) Tanaka K, Goi T, Fukase K. Synlett 2005:2958–2962. (b) De Meo C, Parker O. Tetrahedron: Asymmetry 2005;16:303–307. (c) Lin CC, Huang KT, Lin CC. Org Lett 2005;7:4169–4172. [PubMed: 16146379] (d) Tanaka H, Adachi M, Takahashi T. Chem Eur J 2005;11:849–862. (e) Meijer A, Ellervik U. J Org Chem 2004;69:6249–6256. [PubMed: 15357583] (f) Adachi M, Tanaka H, Takahashi T. Synlett 2004:609–614. (g) Cai S, Yu B. Org Lett 2003;5:3827–3830. [PubMed: 14535720] (h) Ishiwata A, Ito Y. Synlett 2003:1339–1343. (i) Ando H, Koike Y, Ishida H, Kiso M. Tetrahedron Lett 2003;44:6883–6886. (j) De Meo C, Demchenko AV, Boons GJ. Aust J Chem 2002;55:131–134. (k) Ye XS, Huang X, Wong CH. Chem Commun 2001:974–975. (l) Yu C-S, Niikura K, Lin CC, Wong CH. Angew Chem Int Ed 2001;113:2984–2987. (m) De Meo C, Demchenko AV, Boons GJ. J Org Chem 2001;66:5490–5497. [PubMed: 11485473] (n) Demchenko AV, Boons GJ. Chem Eur J 1999;5:1278–1283. (o) Martichonok V, Whitesides GM. J Am Chem Soc 1996;118:8187–8191. (p) Martichonok V, Whitesides GM. J Org Chem 1996;61:1702–1706. [PubMed: 11667039]
4. Barresi F, Hindsgaul O. J Carbohydr Chem 1995;14:1043–1087.
5. Crich D, Smith M. J Am Chem Soc 2001;123:9015–9020. [PubMed: 11552809]
6. Crich D, Lim LBL. Org React 2004;64:115–251.
7. Codée JDC, van den Bos LJ, Litjens REJN, Overkleeft HS, van Boeckel CAA, van Boom JH, van der Marel GA. Tetrahedron 2004;60:1057–1064.
8. Kahne D, Walker S, Cheng Y, Engen DV. J Am Chem Soc 1989;111:6881–6882.
9. Boons, G-J. Handbook of Reagents for Organic Synthesis: Reagents for Glycoside, Nucleotide, and Peptide Synthesis. Crich, D., editor. Wiley; Chichester: 2005. p. 9-15.
10. Demchenko AV, Boons GJ. Tetrahedron Lett 1998;39:3065–3068.
11. (a) Garcia BA, Poole JL, Gin DY. J Am Chem Soc 1997;119:7597–7598. (b) Garcia BA, Gin DY. J Am Chem Soc 2000;122:4269–4279. (c) Nguyen HM, Poole JL, Gin DY. Angew Chem Int Ed 2001;40:414–417. (d) Haberman JM, Gin DY. Org Lett 2003;5:2539–2541. [PubMed: 12841775]
12. Crich D, Smith M, Yao Q, Picione J. Synthesis 2001:323–326.
13. The *H-3eq* peaks of the two anomers (at δ 3.1 and 2.9 ppm) are characteristic of the two species in the ^1H NMR spectrum.
14. General experimental procedure for glycosylation reactions: The 2-thiosialic acid donor 1 (0.11 mmol), diphenyl sulfoxide (Ph_2SO , 0.32 mmol), TTBP (0.22 mmol), and activated 4 Å powdered molecular sieves were mixed in anhydrous dichloromethane (2 mL) and cooled down to -78°C under positive argon pressure. The mixture was stirred at -78°C for 30 min before Tf_2O (0.12 mmol) was added. After 10 min, a solution of acceptor (0.22 mmol) in anhydrous dichloromethane (1 mL) was added. The reaction mixture was stirred from 1 to 6 h at -78°C and then warmed up to room temperature, filtered, washed with saturated aqueous NaHCO_3 solution, and brine. The organic layer was dried over Na_2SO_4 , and concentrated under reduced pressure. The glycosides were isolated by silica gel column chromatography or by preparative HPLC.
15. Dabrowski U, Friebohn H, Brossmer R, Supp M. Tetrahedron Lett 1979;20:4637–4640.
16. (a) Paulsen H, Tietz H. Angew Chem Int Ed 1982;21:927–928. (b) van Halbeek H, Dorland L, Vliegthart JFG, Pfeil R, Schauer R. Eur J Biochem 1982;122:305–311. [PubMed: 7060578]
17. (a) Hori H, Nakajima T, Nishida Y, Ohru H, Meguro H. Tetrahedron Lett 1988;29:6317–6320. (b) Prytulla S, Lauterwein J, Klessinger M, Thiem J. Carbohydr Res 1991;215:345–349.
18. Crich D, Jayalath P. Org Lett 2005;7:2277–2280. [PubMed: 15901188]
19. Masamune S, Choy W, Peterson JS, Sita LR. Angew Chem Int Ed Engl 1985;24:1–76.
20. Paulsen, H. Selectivity, A Goal for Synthetic Efficiency. Bartmann, W.; Trost, BM., editors. Verlag Chemie; Weinheim: 1984. p. 169-190. (b) Spijker NM, van Boeckel CAA. Angew Chem Int, Ed Engl 1991;30:180–183. (c) Fraser-Reid B, López JC, Gómez AM, Uriel C. Eur J Org Chem 2004:1387–1395.

Table 1
Coupling the Presence of 300% Diphenyl Sulfoxide

<p> i) Ph₂SO (3 equiv), TTBP (2 equiv) 4 Å MS, CH₂Cl₂, -78 °C ii) Tf₂O (1.1 equiv), -78 °C iii) ROH, -78 °C to rt </p>		
entry	acceptor	product: % yield ^a (α:β) ^b
1		19 : 97% (2.3:1)
2		20 : 94% (1.5:1)
3		21 : 91% (1:1)
4		22 : 89% (3.8:1)
5		23 : 92% (2:1)
6		24 : 97% (1.2:1)
7		25 : 92% (6:1)
8		26 : 94% (10:1)
9		27 : 92% (only α) ^c
10		28 : 76% (1:1.2) ^d

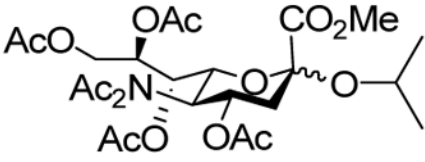
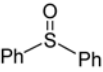
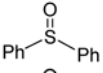
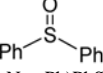
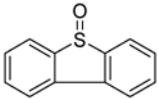


entry	acceptor	product: % yield ^a ($\alpha:\beta$) ^b
11	 16	29 : 81% (1:1.7) ^d
12	 17	30 : 82% (1:7)
13	 18	-

^a isolated yields^b determined by ^1H NMR on the crude reaction mixture^c coupled to the 6-OH^d coupled to the 3-OH

Table 2

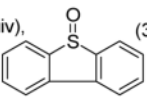
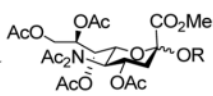

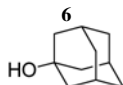
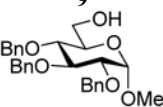
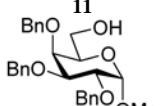
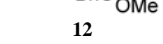
The Effect of Additives and Sulfoxides

i) activator (3 equiv), TTBP (2 equiv), Tf ₂ O (1.1 equiv) 1 $\xrightarrow{\hspace{1.5cm}}$ ii) isopropanol (2 equiv)			
			19
entry	activator	solvent (temp.)	% yield ^a (α:β) ^b
1		CH ₂ Cl ₂ (– 60 °C)	82% (2.2:1)
2		CH ₂ Cl ₂ /CH ₃ CN 1:1 (– 78 °C)	89% (1.8:1)
3		CH ₃ CH ₂ CN (– 78 °C)	77% (2:1)
4	(4-NO ₂ -Ph)PhSO	CH ₂ Cl ₂ (– 78 °C)	75% (2.7:1)
5	(4-MeO-Ph)PhSO	CH ₂ Cl ₂ (– 78 °C)	50% (2:1)
6		CH ₂ Cl ₂ (– 78 °C)	50% (6:1)

^a isolated yields^b determined by ¹H NMR on the crude reaction mixture

Table 3

Use of Dibenzothiophene as Additive

i) Ph ₂ SO (3 equiv), TTBP (2 equiv), Tf ₂ O (1.1 equiv),  (3 equiv) 1 $\xrightarrow{\text{ii) ROH}}$ 		
entry	acceptor	product: % yield ^a (α:β) ^b
1		19 : 95% (6:1)
2	 6	22 : 62% (3.2:1)
3	 9	24 : 65% (1:1)
4	 11  12	25 : 67% (5:1)

^a isolated yields^b determined by ¹H NMR on the crude reaction mixture