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CRITICAL REVIEW

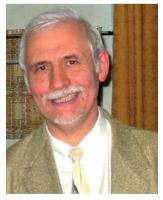
Bulgarian analytical atomic spectroscopists in the new millennium—integrated in the European research area†

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The studies from Bulgarian analytical atomic spectroscopists in the fields of atomic absorption spectrometry (AAS), atomic fluorescence spectrometry (AFS), inductively coupled plasma optical emission spectrometry (ICP-OES), inductively coupled plasma mass spectrometry (ICP-MS) and hyphenated atomic spectrometric methods after 2000 are reviewed with an emphasis on direct methods of analysis (slurry and solid sampling), rational sample pretreatment and trace elements enrichment, alternative pretreatments such as alkaline solubilization by tetraalkylammonium hydroxides, in situ treatment in a graphite atomizer by means of chemical modification, speciation and fractionation analysis. There is a clear focus on the development of sample preconcentration methods coupled with atomic spectrometric techniques, particularly in flow and flow injection systems to alleviate interferences and to improve limits of detection (LODs). The most notable publications concerned with methodological developments in the atomic spectroscopic analysis of environmental samples, biological and advanced materials, food and beverages, as well as speciation and fractionation studies are tabulated. Recent reviews, book chapters, bibliographies and monographs with Bulgarian authorship/co-authorship are presented. The integration of Bulgarian atomic spectroscopists in the European research area is illustrated. Some trends in the publication activities are presented.

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The aim of this review is to present research and publication activities of Bulgarian scientists in the field of analytical atomic spectrometry (AS) in the new millennium. X-ray fluorescence (XRF) techniques are not included in this review since the XRF field is outside of research and publication activities of the present authors. Articles on purely physical aspects of atomic spectroscopy and publications in Bulgarian language fall outside of its scope as a rule, hence ca. 15–20% of the initially collected publications have been excluded from the present discussion. We acknowledge that the review is not comprehensive as a further ca. 40% of relevant publications were considered in the review but not included in the final reference list. Our aim was to be critical and to provide readers with an overview of high quality Bulgarian spectroscopy research. Several pre-2000 references are given as relevant sources of information in the recent bibliographies, 1,2 reviews, 3-6 book chapters, 7-9 and monographs 10,11 on ETAAS, 8,9 hydride generation (HG) AAS, 6,10 chemical modification,^{2,7} hyphenated vapor generation (VG) AAS techniques,⁶ and applications of AAS in occupational and environmental health practice.5,11

Early contributions of Bulgarian analysts to the field of AAS in the years 1964–1994 (456 references) were compiled and indexed by author, analyte, AAS technique, application area and other keywords by Tsalev.¹ A positive trend of the research activities and an increased percentage of publications in international editions were clearly shown in that article as mean values for earlier 5-year increments, viz.: nil(!), 35, 22, 33, 44 and 55% for the periods of 1965–1970, 1971–1975, 1976–1980, 1981–1985, 1986–1990 and 1991–1994, respectively. During the last 12 years about 75% of the scientific works of Bulgarian analytical atomic spectroscopists have been published in renowned international scientific journals, books and proceedings of international conferences; ca. 25% are published in English language in national specialized scientific journals. About one-half of scientific contributions are only of Bulgarian co-authorship.

The main Bulgarian scientific groups working in the field of analytical atomic spectroscopy, covering ca. 94% of examined papers, are situated at three institutions: the Faculty of Chemistry and Pharmacy (formerly Faculty of Chemistry), University of Sofia "St. Kliment Ohridski" (S. Arpadjan, R. Djingova, I. Karadjova, P. Kovacheva, I. Kuleff, V. Lyubomirova, D. L. Tsaley, B. Zlateva); the Institute of General and Inorganic Chemistry of the Bulgarian Academy of Sciences (N. Daskalova, A. Detcheva, G. Gentscheva, I. Havezov, E. Ivanova, S. Velichkov); and the Faculty of Chemistry, University of Plovdiv "Paisii Hilendarski" (G. Andreev, V. Kmetov, K. Simitchiev, V. Stefanova). Other contributions come from the Institute of Mineralogy and Crystallography, BAS (N. Lihareva); Geological Institute, BAS (M. Karadjov, E. N. Pentcheva, N. Velitchkova); Agricultural University, Plovdiv (K. Ivanov); Regional Laboratory – Burgas, Environmental Executive Agency, Ministry of Environment and Waters (L. Chepanova), National Center for Public Health Protection, Sofia (R. B. Georgieva), etc.

Fruitful international cooperation with scientists and laboratories from 16 countries has been established as depicted in Fig. 1. From about 109 publications with co-authors from Europe, Turkey and Japan, productive co-works with international authors from laboratories in Belgium (21), Czech Republic (4), France (7), Germany (13), Italy (9), Spain (9), Switzerland



Fig. 1 Recent research cooperation of Bulgarian analytical spectroscopists.

(4), etc. and about 30 papers with scientists from neighboring Balkan countries in Bursa (3), Ljubljana (2), Skopje (24) and Thessaloniki (1) are known.

The research activities are directed towards optimization studies on the instrumental and chemical parameters of the AS methods, improvement of their metrological and technical characteristics and application studies for trace analysis of various types of samples. Approximate distribution pattern of 211 examined publications between several application fields indicates preference for environmental (\sim 43%), agricultural, food and beverage (~22%), industrial and advanced materials $(\sim16\%)$, geological $(\sim3.5\%)$ and miscellaneous including applications to aqueous and model solutions (\sim 5.5%). If compared with previous bibliography on AAS only,1 this distribution demonstrates about a 2-2.5-fold decline of clinical/biological, geological and industrial application yet ca. 2.5-fold increase of environmental applications. Interestingly, the speciation and fractionation works (~45 papers, partly presented in Tables 2 and 3) are increased five-fold vs. previous periods¹ to ca. 23% of all application papers for the reviewed period. The approximate percentage distribution of the main analytical techniques: AAS $(\sim61\%)$, AFS $(\sim4\%)$, ICP-OES $(\sim21\%)$, ICP-MS $(\sim15\%)$ and VG with in-atomizer trapping of chemically generated species (VG-ETAAS) (\sim 3%) is in agreement with current trends of increased role and accessibility of multielement, powerful ICP techniques. Articles on procedures and applications reveal proportions between various AAS techniques by themselves: flame atomic absorption spectrometry (FAAS) (\sim 28%), electrothermal AAS (ETAAS) (~54%), chemical vapor generation (CVG) AAS (VGAAS) (~13%) and VG-ETAAS (4%). Thus comparison of this splitting of AAS between techniques with available data for the previous three decades 1964–1994 (60, 31, 9 and 0% for FAAS, ETAAS, VGAAS and VG-ETAAS, respectively)1 demonstrates a reasonable contemporary decline of FAAS research in favor of ETAAS and VGAAS. The role of VG techniques with hydride generation (HG) and the cold vapor technique (CV, CVT) for mercury is also increasing, resulting in new methods such as CVAAS, HGAAS, CVAFS, HGAFS, CV-ICP-OES, HG-ICP-OES, VG-ICP-MS, as well as further hyphenations like electrothermal vaporization (ETV) in ETV-ICP-OES for multielement determinations^{12,13} and VG

Table 1 Atomic spectrometric procedures and applications briefly outlined

| Analyte | Matrix | Brief procedure outline | Ref. |
|---------|--|--|------|
| As | Algae; plants (citrus leaves, grass) | $\mbox{HNO}_3\mbox{-H}_2\mbox{O}_2\mbox{-HF}$ dign; USN-ICP-OES; validation with CRMs | 98 |
| As | Aq. solns, org. solvents | CF-HGAAS; org. solvent interf. (MeOH, EtOH, i-PrOH, ethanolamine, acetonitrile, ethylene glycol, glycerol) studied. Atomization interf. by MeOH, EtOH and PrOH compared; three atomizers compared: QTA, Ar–H ₂ MDF and FIF; interf. minimized in FIF mode | 99 |
| As | Essential oils (lavender, rose) | 4–5 fold diln with EtOH or i-PrOH; ZETAAS; THGA; L-cysteine–Pd–citric acid chem. modifier on Zr–Ir treated platforms; LODs 4.4–4.7 ng g ⁻¹ ; also LTPA in oxygen; LODs 2.5–2.7 ng g ⁻¹ | 100 |
| As | Fish | TMAH solubilization; ZETAAS with Pd modifier; $T_{\rm pyr}$ 1400 °C, $T_{\rm at}$ 2000 °C; LOD 0.45 µg g ⁻¹ | 50 |
| As | Marine tissue CRMs | MW-assisted LLE of lyophilized tissue with TMAH or ethanol; ZETAAS; Zr-Ir permanent modifiers; FI-HGAAS; LODs 0.003–0.02 mg kg ⁻¹ | 51 |
| As | Plant, sediment, soil CRMs | SS-ETAAS; US agitation of slurry; LOD 30 µg kg ⁻¹ ; validation with CRMs | 60 |
| As | Urine | 10 mL urine + 5 mL 0.5 M Cys + 2.5 mL 1 M HCl dild to 50 mL; batch HGAAS; LOD 0.5 ng; 15 h ⁻¹ | 101 |
| As | Wine | 1 mL sample + 1 mL 1% m/v Cys–0.1 M HCl; 1–10× diln; CF-HG-FAFS with Ar–H ₂ MDF; LODs 0.3–0.6 μg L ⁻¹ ; alternatively bomb decomposition with HNO ₃ –H ₂ O ₂ and CF-HG-FAFS or ETAAS | 20 |
| As | Wine | Evaporation of ethanol for inorg. As; MW-assisted HNO ₃ –H ₂ O ₂ dign for total As; HGAAS | 65 |
| Au | Gold alloys | Aq. regia dign; FAAS with ASDI and ensemble summation; calibrn by a bracketing method; tutorial treatment of traceability, validation and uncertainty budget calculation | 102 |
| В | Tourmaline | Na ₂ CO ₃ –ZnO (3.5 : 0.6) dign; boron leached with H ₂ O; ICP-OES; LOD 4 μ g g ⁻¹ | 103 |
| Cd | Natural waters | Preconcn by flotation on Pb(II) heptyl DTC and Co(III) heptyl DTC; ZETAAS; LOD 3.0 ng L ⁻¹ | 72 |
| Cd | Seawater, urine | VG-ETAAS; Cd° trapped on Zr–Ir or W–Ir treated platforms; THGA®; T_{coll} 350 °C; T_{at} 1300 °C; m_{o} 2.8 pg; LOD 2 ng L ⁻¹ ; memory effects observed | 14 |
| Со | Aq. solns | SPE in a KR pre-coated with oxine derivatives; FI-ETAAS; 34× enrichment | 96 |
| Co | Natural waters | FI; KR pre-coated with PMBP; ETAAS; LOD 8.1 ng L^{-1} | 73 |
| Co | Natural waters, cod muscle, | FI; KR pre-coated with nitroso-R-salt/TBABr; ETAAS; LOD 5 ng | 74 |
| Cr | mussel tissue Serum, urine | L^{-1} ; validation with CRMs Direct ZETAAS; H_2O_2 modifier; validation with CRMs; LOD 0.08 $\mu g L^{-1}$ | 104 |
| Cu | Seawater | SPE on a new IIP Cu(II)–PAR; FAAS; LOD 0.001 μmol L ⁻¹ | 75 |
| Fe | Seawater | SPE on anatase TiO ₂ nanoparticles; ICP-MS; validation with CRM; LOD 69 ng kg ⁻¹ | 76 |
| Hg | Fish, liver, kidney, plant, soil CRMs | Direct SoS-ZETAAS; specially designed furnace; lack of matrix effects; calibrn with CRMs; LOQ $0.008~\mu g~g^{-1}$ | 61 |
| Hg | Wine | MW-assisted or under reflux dign with HCl-H ₂ O ₂ ; LLE of APDC, chloride or iodide complexes with IBMK; ETAAS using different Pd modifiers; CVAAS; LOD 0.2 µg L ⁻¹ | 67 |
| Hg | Seawater, mineral water | SPE on a new IIP; CF-CVAAS; LOD 0.006 μg L ⁻¹ . Synthesis and characterization of IIP microbeads by cross-linking dispersion copolymerization of methacrylic acid and trimethylolpropane trimethacrylate in acetonitrile detailed | 77 |
| Na | Single crystals of $KTiOPO_4$ or $KGd(WO_4)_2$ | Slurry in 2% HNO ₃ –0.005% Triton-X-100; SS-ETAAS; TiC-coated or W-coated GT, resp.; RSD 3%; good agreement with FAAS after digestion | 63 |
| Ni | Serum, urine | ZETAAS; H_2O_2 modifier; LOD 0.2 μ g L^{-1} ; validation with CRMs | 105 |
| Pb | Ag, Cu, and their alloys | Electrochemical Pb sepn; ICP-OES; AAS; lead isotope ratios detn by ICP-MS; archaeometry; validation with CRMs | 95 |
| Pb | Plant, sea water | HNO ₃ –H ₂ O ₂ dign; ID-ICP-MS; ID-HG-ICP-MS; pH effect critical; ID mode preferred for better accuracy and precision | 45 |
| Pb | Wine | ETAAS; Pd modifier; 1 : 1 sample diln with 0.2 M $\bar{\text{H}}\text{NO}_3$; LOD $0.91.8~\mu\text{g L}^{-1}$ | 68 |
| Pb | Wine | $10\times$ diln; CF-HGAFS with miniature Ar/H ₂ diffusion flame in the presence of 1% m/v K ₃ [Fe(CN) ₆]–0.1 M HCl; LOD 0.3 µg L ⁻¹ ; alternatively direct ETAAS | 69 |

Table 1 (Contd.)

| Analyte | Matrix | Brief procedure outline | Ref. |
|----------------|--|---|------|
| Pd | Blood, road dust | FI enrichment by sorption of Pd(SCN) ₄ ⁻² onto KR pre-coated with K ⁺ 18-crown-6; ETAAS; LOD 16 ng L ⁻¹ | 97 |
| Pd | Tunnel dust | FI-ETAAS, sepn and preconcn using a cyclic polyether (18-crown-6) previously immobilized on a KR; LOD 16 ng L ⁻¹ | |
| REEs | Eu_2O_3 , Lu_2O_3 | ICP-OES; spectral interferences in REE matrices assessed; O-values tabulated | 17 |
| REEs | Soils, sediments | Cation-exchange sepn; ICP-OES; validation with CRMs; LODs | 93 |
| Se | Blood serum, hair, urine | 0.05–0.5 mg kg ⁻¹ 1 + 2 diln of serum; 10 μg Rh modifier; 1 + 2 to 1 + 4 diln of urine, 20 μg Rh modifier; hair solubilized with TEAH; ZETAAS (THGA®); LOD 6 μg L ⁻¹ ; alternatively FI-HG-ETAAS or | |
| Tl | Natural waters | CF-HG-NDAFS procedures for urine digests SPE; Empore [™] chelating disc; ZETAAS; uncoated GT; modifier- free; effect of humic acids | 78 |
| Tl | Wine | HNO ₃ -H ₂ O ₂ dign; LLE from 0.5 M KI soln with IBMK; ZETAAS; tartaric acid or Pd + ascorbic acid chemical modifiers; LOD 0.05 μg L ⁻¹ | 66 |
| As, Hg | Sunflower oil | Sample solubilization with TMAH for As; LLE of As and Hg with NH ₄ OH/EDTA; ZETAAS; LODs 2 ng g ⁻¹ As, 3 ng g ⁻¹ Hg | 49 |
| As, Se | Plants, soils, sediments | SS-ETAAS; mixed Ir–Mg modifier; var. mineralization modes evaluated; validation with CRMs | 62 |
| Cd, Pb | Blood, tissues (hair, liver, muscle), urine, water | Direct injection for liq. samples; TEAH solubilization for biol. tissues; phosphate modifier stabilized on pre-treated platforms with Zr-Ir or W-Ir; m _o 0.7–1.0 pg Cd, 26–31 pg Pb | 38 |
| Cd, Pb | Bovine liver and orchard leaves SRMs | APDC complexes sorbed on C18 column; TS-flame-furnace AAS; LODs 7 ng L ⁻¹ Cd; 170 ng L ⁻¹ Pb; validation with CRMs | 44 |
| Na, Fe | Single crystals of RbTiOPO ₄ | Slurry in 2% HNO ₃ -0.005% Triton-X-100; SS-ETAAS; TiC-coated GT | 64 |
| Pd, Pt | Airborne particulate matter, | MW-assisted SPE; thioureidopropyl functionalized silica gel; | 26 |
| Pd, Pt | serum, urine Street dust, road dust | ICP-MS; LOD 0.2 ng L ⁻¹ ; validation by CRMs IE on Dowex 1 × 10; elution with thiourea at 60 °C; ICP-OES; R 98%; LOD 15 ng g ⁻¹ ; agreement with ICP-MS results | 27 |
| As, Sb, Tl | Aq. solns | ETAAS; GT with refractory metal platforms; W coil on a Ta platform recommended; 2 µg Pd modifier | 41 |
| Au, Pd, Pt | Cu ore and concentrate; soils, | SPE with Cys-modified SiG; ZETAAS; ICP-OES; LODs 0.02–0.005 μg L ⁻¹ | 28 |
| Ca, Fe, Mn | river water, seawater Moss | ASDI-FAAS flame microsampling for plant digests; comparison with ICP-MS | 106 |
| Cd, Pb, Zn | Oak leaves | Dign with HNO ₃ -H ₂ O ₂ ; USN-ICP-OES. Oak leaves as bioindicators; evaluation of air pollution; drawing of heavy metal fallout maps | 90 |
| Ga, Ho, Tm | Optical crystals of potassium titanyl phosphate | Dign with HF- H_2SO_4 - H_2O_2 ; ICP-OES; study of spectral interferences in a K-, P- and Ti-containing matrix | 31 |
| Pd, Pt, Rh | Auto catalyst (recycled), CRM SARM 7, Pt ore | ICP-OES with radial viewing 40.68 MHz plasma; study of spectral interferences in environmental matrices; LODs in ng g ⁻¹ : 1700 Pt, 1440 Pd and 900 Rh; LODs at var. λ tabulated; validation with CRMs | 29 |
| Pd, Pt, Rh | Pharmaceutical products (Tritace, Vivace, Laprilen, Enalapril) | Acceleration of CPE by MW (9× faster) and US irradiation; Triton-X-100; 2-mercaptobenzothiazole; acceleration of complex formation by reducing agents (KI and SnCl ₂) examined; surfactant-rich concentrate phase dild with 1 M HCl prior to | 46 |
| Pd, Pt, Rh | Road dust | CF-ICP-MS; LODs 1–6 ng L ⁻¹ ICP-MS; spectral interferences investigation; uncertainty budget | 107 |
| Pd, Pt, Rh | Road dust, serum, urine | analysis and interference alleviation by preliminary acid leaching On-line FI enrichment in a KR with diethylthiourea; ICP-TOFMS | 25 |
| REEs, Sc, Y | Dy_2O_3 , Ho_2O_3 , Tb_2O_3 , Tm_2O_3 | with USN; LODs 0.36–2.1 ng L ⁻¹ ICP-OES; spectral interf. in REE matrices assessed; best λ and LODs. | 34 |
| REEs, Sc, Y | Gd ₂ O ₃ , Er ₂ O ₃ | LODs tabulated ICP-OES; spectral interf. in REE matrices assessed; best λ and LOPs tabulated | 30 |
| REEs, Sc, Y | Yb_2O_3 | LODs tabulated ICP-OES; spectral interf. in REE matrix assessed; best λ and LODs | 32 |
| Cd, Co, Cu, Ni | Aq. solns | tabulated FAAS; uncertainty budget contributions from the flame, light | 108 |
| Cd, Cr, Ni, Pb | Plant CRMs | source and optics; combined instrumental uncertainty evaluated Dry mineralization followed by acid leaching; fast furnace programme ETAAS; Ir permanent modifier; validation with CRMs | 37 |

Table 1 (Contd.)

| Analyte | Matrix | Brief procedure outline | Ref. |
|--|--|---|------|
| Cd, Cu, Ni, Pb | Seawater | SPE on monodisperse, submicrometer silica spheres modified with 3-aminopropyl-trimethoxysilane; <i>in situ</i> sampling and preconcn; | 79 |
| Cd, Cu, Pb, Zn | Effervescent vitamins with micro-minerals | ETAAS; LODs 0.002–0.02 µg L ⁻¹ MW-assisted CPE with APDC and Triton X-100 at pH 7.5; collected surfactant-rich phase dissolved with 3 M HNO ₃ ; prior to | 42 |
| Cd, Pb, Cu, Zn | Plant oil crops (peanuts, rapeseed, sesame); plant parts (roots, stems, leaves, seeds, fruit shell); soil | ASDI-FAAS microsampling (350 μL) Aq. regia dign of soils. Soil extracts in 5 mM DTPA–0.1 M TEA, pH 7.3. Plant tissues dry ashed at 400 °C; 1 + 1 HNO ₃ treatment; final solns in 20% v/v HCl; ICP-OES; std solns in 2% v/v HNO ₃ . The effect of pollution at different distances from a non-ferrous metal plant detailed | 109 |
| Cd, Hg, Mn, Pb | Algae | Direct SoS-ZETAAS; LODs 0.0013–0.008 μg g ⁻¹ ; validation with CRMs | 110 |
| Ca, K, Mg, Mn, Na | Fluid inclusions in quartz | Opening of fluid inclusions by crushing or decrepitation of the quartz samples; leaching of the released components by HNO ₃ or CDTA; FAAS; ETAAS | 111 |
| Cd, Cu, Hg, Ni, Pb | Surface waters | Preconcn on Cys-modified chitosan membrane; elution with thiourea–HCl; ZETAAS; LOQs 0.001–0.05 μg L ⁻¹ ; LOD 0.012 μg L ⁻¹ for Hg by CF-CVAAS | 80 |
| Cd, Hg, Mn, Pb, Sn | Seafood CRMs (cod muscle, mussel tissue, tuna fish, plankton, salmon, trout) | Direct SoS-ZETAAS (modifier-free); A_p measurements for Hg, A_{int} and 3-field mode for other analytes; LODs (pg): Cd 1.3, Hg 40, Mn 12, Pb 8 and Sn 70; calibrn with CRMs of organic matrices | 112 |
| Fe, Li, Nb, Sr, Ti | Single crystals of the SrO– Li ₂ O–Fe ₂ O ₃ –TiO ₂ system | HF–H ₃ BO ₃ dign; stabilization by 6.5% HNO ₃ –0.05% HBF ₄ ; ICP-OES; 0.04–0.08% H ₃ BO ₃ well tolerated; matrix-matched calibrn; | 58 |
| Ir, Pd, Pt, Rh, Ru | Moss, Pt ore, road dust, sediment | detn of single crystals composition MW-assisted dign with HCl-HNO ₃ ; ICP-MS; validation with CRMs; LODs 0.015–0.03 ng g ⁻¹ | 113 |
| As, Bi, Hg, Se, Sn, Tl | Marine sediments, water | ICP-OES; quantification of spectral interferences in environmental matrices; LODs 0.3–0.9 µg g ⁻¹ ; validation with CRMs | 56 |
| Ba, Cd, Cr, Mn, Pb, Zn | Atmospheric particulate matter | Samples collected by the Bergerhoff method; aq. regia dign; ICP-OES; FAAS; d.c. arc OES | 55 |
| Cd, Co, Cu, Ni, Fe, Pb | Natural waters, sea water | SPE on a polycarboxylic microsphere gel; ETAAS; validation with CRM; LODs $0.005-0.05 \mu g L^{-1}$ | 81 |
| Cd, Cr, Cu, Fe, Mn, Pb | Lavender plant and oil | Oil dild $1 + 1$ or $1 + 2$ m/m with 1,4-dioxane. Wet dign of plant with HNO ₃ and H ₂ O ₂ ; FAAS; ZETAAS; permanent modification of GT with W and Pt; $T_{\rm pyr}$ 900, 1300, 1200, 1200, 1200 and 900 °C for Cd, Cr, Cu, Fe, Mn and Pb, resp. | 114 |
| Ag, Cd, Co, Ni, Pb, U, Y | Natural waters | FI enrichment in a KR pre-coated with PMBP; ICP TOFMS with USN; LODS 0.3–15.2 ng L ⁻¹ | 82 |
| Be, Bi, REEs, Te, Th, U, Y | Soils, sediments | Overnight stay with HF–HNO ₃ ; subsequent stepwise MW dign with HF–H ₃ BO ₃ ; ICP-MS; validation with CRMs | 47 |
| Cd, Co, Cr, Cu, Fe, Ni, Pb | Milk, cheese, chocolate | Wet digestion with HNO ₃ -HClO ₄ -H ₂ O ₂ ; slurry preparation in HNO ₃ -H ₂ O ₂ ; ETAAS analysis of solutions and slurries, compared with DPCSV; validation with CRMs | 115 |
| Be, Bi, Ga, REEs, Te, Tl, Th, U | Leaves (poplar) | MW-assisted dign with HNO ₃ -H ₂ O ₂ ; ICP-MS; poplar leaves as bioindicator studied; distribution patterns; background conc. levels of elements tabulated | 116 |
| Cd, Co, Cr, Cu, Mn, Ni, Pb, Zn | Sewage sludge | US-assisted extraction (20 min sonication at 90 °C in HNO ₃ –HCl (1 : 1); ICP-OES | 87 |
| Ag, Cd, Co, Cu, In, Mo, Ni, Pb, Sb | Natural waters, cod muscle, mussel tissue | FI enrichment of APDC complexes in a KR; ICP-TOFMS with USN; LODs between 0.5 and 26 ng L ⁻¹ ; validation with CRMs | 83 |
| Al, As, Ba, Mg, Pb, Sr, V, Zn, Zr | Archaeological bones | Dry ashing followed by MW-assisted dign with HNO ₃ ; ICP-MS; validation with CRM; reconstruction of paleodiet; archaeometric | 92 |
| As, Cd, Bi, Hg, Pb, Sb, Se, Sn, Tl | Aq. solns | study ETAAS; $100 \mu g$ Ag modifier for atomization from the wall or from a platform; $1.2-1.8 \times$ sens. enhancement by an Ag modifier | 36 |
| Bi, Cd, Cr, Cu, Fe, Mn, Ni, Pb, Sb | Lavender oil | Diln with i-PrOH (1 + 1) or 1,4-dioxane (1 + 2); ETAAS; W-treated or W-Pt treated GTs | 117 |
| As, Cd, Co, Cu, Hg, Fe, Mn, Ni, Pb, Zn | Sediment, sludge, soil | MW-assisted dign with <i>aq. regia</i> ; FAAS; ICP-MS; ICP-MS with temp. controlled MW-assisted dign recommended; validation with CRMs | 48 |
| Ba, Cd, Co, Cr, Cu, Mn, Ni, Pb, Sr, Zn | Archaeological bones | ICP-OES; IE enrichment on Dowex 1×10 resin for Cu(II); 3 nebulizers compared: concentric-flow (Meinhard), cross-flow and V-groove; spectral interf. tabulated; validation with CRM; reconstruction of paleodiet; archaeometric study | 91 |

Table 1 (Contd.)

| Analyte Matrix | | Brief procedure outline | |
|---|---|--|-----|
| Ce, Ir, La, Nd, Pb, Pd, Pt, Rh, Ru, Zr | Street dust, plants (dandelion, annual ryegrass, plantain, moss, mushrooms) | MW-assisted dign with HNO ₃ and HCl for ICP-MS; LLE of Pd(II)—DEDTC with CHCl ₃ ; aq. regia dign for ICP-OES; validation with CRMs; accumulation of PGE by plants along | |
| Al, Ca, Co, Cr, Cu, Fe, K, Na, Mg, Mn, Ni, Sb | Archaeological glasses | highways evaluated Direct ETV-ICP-OES analysis of powdered microsamples; calibration with CRMs; CHF ₃ gaseous modifier | 13 |
| Ag, Bi, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Pd, Pt, Sb | Au | Reductive pptn of Au matrix by oxalic acid; ETAAS; ICP-OES; LOQs 0.02–0.1 $\mu g \ g^{-1}$ or 0.3–2.4 $\mu g \ g^{-1}$, resp. | 94 |
| As, Ba, Ca, Cd, Cr, Cu, Fe, Mn, Pb, Sr, U, V, Zn | Moss | Comparison between 3 methodologies for discrete sample introduction in ICP-MS; Every Sweep Internal Standardisation algorithm (ESIS) recommended; LOQs 0.02–66 mg kg ⁻¹ | 43 |
| As, Bi, Cd, Co, Cr, Cu, Fe, Hg, Mn, Ni, Pb, Sb, Se | Ag (high-purity) | Total or partial reductive sepn of Ag matrix with ascorbic acid; ZETAAS; LODs $0.001 \mu g g^{-1} Cd$, $0.005 \mu g g^{-1} Mn$, $0.01 \mu g g^{-1} Cr$ and Cu, $0.02 \mu g g^{-1} Fe$, Co and Pb, $0.05 \mu g g^{-1} As$, Bi, Ni, Sb and Se, $0.2 \mu g g^{-1} Hg$ | 35 |
| Ce, Dy, Er, Eu, Gd, Ho, La, Lu, Nd, Pr, Sm, Tb, Tm, Y, Yb | Natural waters | FI enrichment in KR pre-coated with PMBP; ICP-TOFMS with USN; LODs 3–670 pg L ⁻¹ | 84 |
| Al, B, Ba, Cd, Co, Cr, Cu, Fe, Mn, Ni, P, Pb, S, Sr, Ti, V, Zn | Plant CRMs (apple leaves, sargasso, tobacco) | Sample pretreatment by carbonization; ETV-ICP-OES; gaseous modifiers of the Ar inner gas (CCl ₄ , CHCl ₃ , CCl ₂ F ₂ , C ₃ H ₈); freon CCl ₂ F ₂ recommended; LODs 5–250 ng g $^{-1}$; calibrn with CRMs | 12 |
| As, Be, Bi, Cd, Co, Cr, Cu, Ga, Ni, Pb, Rb, REEs, Sr, Th, Te, V, Zn | Plants | MW-assisted dign with HNO ₃ –H ₂ O ₂ ; ICP-MS; validation with CRMs | 118 |
| Al, Bi, Cd, Co, Cr, Cu, Fe, Ga, In, Mn, Mo, Ni, Pb, Tl, V, Sb, Sn, Zn | Waters | Preconcn on high-surface area ZrO ₂ ; FI-ICP-OES | 85 |
| Al, Ce, Cr, Er, Fe, Ga, Ge, Ho, Mn, Na, Nb, Nd, Ni, Rb, Tb, Tm, Yb, Zr | Optical crystals of potassium titanyl phosphate | Dign with HF-H ₂ SO ₄ ; ICP-OES; assessment of dopants incorporation into the optical crystal | 57 |
| Al, Bi, Co, Cd, Cu, Cr, Fe, Hg, Ga, In, Mo, Mn, Ni, Pb, Sc, Tl, Ti, Sn, U, V, W, Y, Zn, Zr | Natural waters | On-line preconcn on Muromac A-1 resin; ICP-OES; LODs 0.001–0.009 $\mu g \ L^{-1};$ validation with CRMs | 86 |

with in-atomizer trapping of chemically generated species (VG-ETAAS, CV-ETAAS)¹⁴ (Table 1). Couplings with chromatographic techniques for speciation analysis such as gas chromatography (GC) with cryogenic trapping (CT) and AFS detection for Hg,^{15,16} liquid chromatography (LC) and high performance liquid chromatography (HPLC) for As,¹⁷⁻²⁰ Pd,²¹ Pt²¹ and Se¹⁹ are compiled in Table 2 – see also recent *reviews*.^{6,22-24}

Along with the application of well-established methods, novel developments for the AS analytical techniques are reported (Table 1), where applications to numerous chemical elements are given with analytes arranged in an alphabetical order. Among the most intensively studied analytes are As, Cd, Co, Cr, Fe, Hg, Mn, Ni, Pb, Sb, Se, Tl, Zn, and platinum group metals Pd, Pt, Rh (PGMs) as emerging pollutants.^{25–28} Rare earth elements (REEs) are components of advanced materials and analytical challenge because of their spectral interferences.^{29–34} Application studies of 27 chemical elements representing interest for speciation and fractionation studies and procedures are known, partly presented

in Tables 2 and 3, respectively, with a focus on environmentally, toxicologically and nutritionally important elements As, Cd, Cr, Cu, Fe, Hg, Pb, Sb, Se, Sn and Zn, and PGMs.

Systematic studies are performed on the optimization of the instrumental parameters of AS detection methods. Silver is proposed as a new modifier for the ETAAS determination of volatile elements; ^{35,36} four gaseous chemical modifiers in ETV-ICP-OES are studied; ¹² the advantages of an Ir permanent modifier in fast ETAAS programs are demonstrated. ³⁷ Several contributions on chemical modification with an emphasis on permanent modifiers in ETAAS and VG-ETAAS are published. ^{3,4,7,38,39} Chemically modified platforms ^{38–40} and refractory metal platforms are studied for ETAAS. ⁴¹ A bibliography on earlier chemical modification literature in ETAAS for the period 1973–1989 with 560 references indexed by authors, matrix, modifier and other keywords is published.²

Sample introduction in AS methods has been the subject of thorough studies, e.g., the development of air-segmented discrete

Table 2 Speciation studies and procedures briefly outlined

| Element | Species | Matrix | Brief procedure outline | Ref. |
|---------|--|---|---|------|
| As | As(III) + As(v) + MMA + DMA; total As | Marine tissues (algae, aq. plant, fish, liver, mussel, oyster CRMs) | Pressurized MW extrn with TMAH or aq. 80% v/v MeOH; ZETAAS with THGA [®] ; Pd modifier on a Zr-Ir treated platform; LOD 1.2 mg kg ⁻¹ total As. Hydride active fraction by FI-HG-ETAAS in 60-90 mM HCl-75 mM Cys; LODs 3-20 μg kg ⁻¹ . The behavior of arsenosugars addressed. The std addn approach is inefficient in the case of species-dependent slope of the calibrn | 51 |
| As | As(III), As(v), MMA, DMA | Algae, seawater | graph ('intrinsic element speciation interference') Biouptake and biotransformation of var. As species on a green microalga <i>Chlorella salina</i> in seawater studied; the effect of phosphate discussed; wet dign of algae with HNO ₃ ; | 52 |
| As | As(v), MMA, DMA | Aq. solns | FI-HGAAS and ZETAAS as detailed in ref. 51 On-line pre-redn and complexation of As species with Cys in FI-HGAAS studied. HPLC-HGAAS sepn of complexes demonstrated on a strongly acidic cation exchange column at pH 3.3–3.5; | 18 |
| As | As(III) + As(V) + MMA + DMA | Fish | LOD 0.5 ng or 10 µg L ⁻¹ MW extrn in 0.075% m/v TMAH or 20% v/v MeOH; FI-HG-ETAAS; total As by direct ETAAS; validation with CRMs | 50 |
| As | As(III), As(v), MMA, DMA | Natural waters | Selective CF-HGAAS from var. reaction media; LOD $0.1 \mu g L^{-1}$ | 121 |
| As | As(III) + As(v) + MMA + DMA | Sediment and soil extracts | Extrn with 50 mM EDTA, pH 6–7; CF-HGAAS; LOD 0.2 mg kg ⁻¹ ; RSD 3–7% | 127 |
| As | As(III) + As(V) + MMA + DMA | Urine | 10–25× diln; HG from 50 mM L ⁻¹ HCl–70 mM Cys medium; FI-HG-ETAAS; Zr–Ir permanent modifier; THGA [®] ; T_{coll} , T_{pyr} , T_{at} , T_{cl} 450, 500, 2100 and 2150 °C, resp.; m_{o} 39 pg; LODs 0.4–1 μg L ⁻¹ ; 25 h ⁻¹ | 130 |
| As | As(III) + As(V) + MMA + | Urine | 5× diln; batch HGAAS from 50 mM HCl-70 mM | 101 |
| As | DMA Inorg. As; total As | Wine | Cys medium; LOD 0.5 ng; RSD 2-6%; 15 h ⁻¹ Evaporation of ethanol for inorg. As; MW- | 65 |
| As | As(III); $As(III) + MMA$; $As(III) + As(V)$; MMA | Wine | assisted HNO ₃ –H ₂ O ₂ dign for total As; HGAAS (i) CF-HG-FAFS with Ar–H ₂ MDF; 5–10× diln; selective HG from var. reaction media/pH; (ii) HPLC-HG-FAFS on a strongly acidic anion | 20 |
| Cr | Cr(III), Cr(VI) | Natural waters | exchange column SPE on high-surface area TiO_2 (anatase); ETAAS; LODs 30 ng L ⁻¹ for Cr^{3+} and 24 ng L ⁻¹ for $Cr(vi)$ | 122 |
| Cr | Cr(III), Cr(VI) | Natural waters | Preconcn and speciation analysis on high-surface | 123 |
| Fe | Fe(II), Fe(III) | Wines | area TiO ₂ ; FI-ICP-OES; validation with CRMs LLE with thiocyanate/IBMK or 1,10- phenanthroline/ClO ₄ ⁻ /IBMK; column SPE with TOMACl on silica gel; FAAS | 70 |
| Hg | Hg(II), MeHg | Fish | MW-assisted extrn with TMAH, methanolic KOH or 5 M HCl; CF-CVAAS, FI-CVAAS and CF-CVG-ICP-OES compared with LODs 5–7 ng g ⁻¹ for inorgHg(II) and 12–16 ng g ⁻¹ for MeHg, resp.; validation with CRMs | 53 |
| Hg | Hg(II), MeHg | Natural water | HG-CT-GC-AFS; validation with CRMs | 15 |
| Hg | Hg(II), MeHg | Sediment, water | Derivatization by HG or ethylation; cryofocussing; GC-AFS; LODs 0.22 ng L ⁻¹ for Hg ²⁺ , 0.02 ng L ⁻¹ for MeHg ⁺ | 16 |
| Hg | Hg(II), MeHg | Surface waters (spiked) | New sorbent L-Cys-grafted SiG evaluated; on-site SPE enrichment; CF-CVAAS vs. ICP-MS; LOQs 1.5 and 5 ng L ⁻¹ for dissolved inorgHg(II) and CH ₃ Hg(I) by CF-CVG-AAS and 1 and 2.5 ng L ⁻¹ by ICP-MS with RSD between 7–12% and 7–14%, resp. | 124 |
| Sb | Sb(III), Sb(v) | Natural waters | LSME of DTC complexes into xylene at pH 5–8 for Sb(III) and pH 0–1.2 for total Sb; ZETAAS; LOD 2 ng L ⁻¹ ; validation with CRM | 125 |
| Pt | Pt(II) and Pt(IV) chloride species | Soil (spiked) | SPE on Dowex 1 × 10; Pt(II)–DEDTC elution with IBMK; PtCl ₆ ²⁻ elution with thiourea; ICP-OES; LOD 15 ng g ⁻¹ | 128 |
| As, Se | As(III), As(v), MMA, DMA, AsB; Se(IV), Se(VI) | Groundwater, soil extracts | Anion exchange; IC-ICP-MS; LODs 0.4–0.8 μ g L ⁻¹ As, 4 μ g L ⁻¹ Se | 19 |

Table 2 (Contd.)

| Element | Species | Matrix | Brief procedure outline | Ref. |
|---------|---|--------------------------------|--|------|
| As, Sn | As(III), As(v), MMA, DMA, AsB, AsC, TMAO, Me ₄ As ⁺ ; inorgSn(II/IV), Me ₂ Sn, Me ₃ Sn, Et ₃ Sn, Pr ₃ Sn, Ph ₃ Sn, BuSn, Bu ₂ Sn, Bu ₃ Sn | Aq. solns; urine (for As only) | On-line UV photooxidation with $K_2S_2O_8$ in a KR for 6 orgAs and 10 orgSn species in a FI-HGAAS system evaluated with a view to coupling with a HPLC-UV-HGAAS system. LODs 7 and 4 μ g L ⁻¹ for As in FI-UV-HGAAS and HPLC-UV-HGAAS systems, resp. Var. reaction mixtures evaluated for VG from var. As and Sn species; low | 17 |
| Pd, Pt | Humic acid complexes of Pd(II), Pt(II), Pt(IV) | Soil and street dust (spiked) | recoveries for butylated organotin species Complexation of Pd(II), PtCl ₄ ²⁻ and PtCl ₆ ²⁻ with humic acids studied by SEC with UV-DAD; ICP-MS quantification | 21 |

Table 3 Selected fractionation studies and procedures briefly outlined

| Element | Species | Matrix | Brief procedure outline | Ref. |
|----------------|---|---|---|------|
| As, Cd, Pb | Anion, cation, macromolecular fractions | Medicinal herbs | Fractionation scheme; MW-assisted dign with HNO ₃ ; herbal infusions in hot water; SPE on an anion-exchange resin; pptn of protein-bound element species; ICP-MS | 119 |
| Ce, La, Zr | Fractions of extracted species | Road dust | Fractionation <i>via</i> conventional sequential extraction protocol (BCR); ICP-MS; complexation with humic substances; validation with CRMs | 131 |
| Cu, Fe, Zn | Three fractions; labile complexes of Cu, Fe, Zn | Wines | Fractionation; speciation; separation of wine polyphenols, wine proteins and polysaccharides on XAD-8; separation of cationic and anionic species of analytes on Dowex ion exchange resins; FAAS; ETAAS | 71 |
| Pd, Pt, Rh | Fractions of extracted species | Soil, street dust | A 3-step BCR sequential extrn procedure; solubilization; ICP-MS; complexation with humic substances studied | 129 |
| Cd, Cu, Ni, Pb | Free ions; labile complexes; kinetically inert org. complexes | Surface waters of Black Sea coastal area | Fractionation by HF-PLM and DGT; ETAAS following LLE of DTC complexes into IBMK; the concns of Cd, Cu, Ni and Pb detd by HF-PLM are lower compared to those by DGT, viz. 'free' metal ions by HF-PLM and dynamic (mobile and labile) species by the DGT. Bioavailability to green microalga Chlorella salina studied | 126 |
| Cu, Fe, Mn, Zn | Hot water extractable elements; anion, cation, macromolecular fractions | Medicinal plants | Herbal infusions in hot water; speciation; fractionation by IE, selective pptn and LLE; ICP-MS; FAAS; bioavailability of extracted and separated fractions discussed | 120 |

sample introduction (ASDI) for flame AAS;⁴² the studies on three discrete sample introduction systems in ICP-MS, flow injection, air segmented discrete introduction and injection into an air carrier, eventually recommending the use of the Every Sweep Internal Standardization (ESIS) algorithm for quantitative purposes.⁴³ The use of thermospray (TS) flame furnace AAS for the determination of trace amounts of Cd and Pb has been demonstrated.⁴⁴ A comparison is made between hydride generation and nebulization for sample introduction in the determination of Pb by ICP-MS with external calibration or internal standard (IS).⁴⁵ Semi-batch and FI-VG-ETAAS with different flow systems and in-atomizer trapping were applied for determination of Cd in water and urine.¹⁴

Microwave (MW) treatments are steadily adopted for intensification and 'greening' of sample pretreatment. MW assisted cloud point extraction (CPE) was successfully applied for trace amounts of Pd, Pt and Rh in various pharmaceutical products.⁴⁶

More efficient and environmentally friendly MW digestions of soils and sediments for determination of lanthanoids⁴⁷ and other elements by FAAS and ICP-MS have been developed.^{47,48} Alkaline solubilization of small samples of biological matrices such as fish and other marine tissues, food, hair, nail, liver, *etc.* with strong organic bases tetramethylammonium hydroxide (TMAH) or tetraethylammonium hydroxide (TEAH) has proved convenient and successful in ETAAS and VGAAS procedures, being compatible with VG speciation analysis procedures, as shown in some examples for As,^{49–52} Cd and Pb,³⁸ Hg,⁵³ Se,⁵⁴ *etc.*¹¹ in Tables 1 and 2.

Systematic studies are performed on the quantification of the spectral interferences in ICP-OES, which is of particular importance for the trace analysis in complex matrices, *e.g.*, the determination of trace REEs in "pure" rare earth matrices;^{30,32–34} determination of toxic and essential trace elements in environmental and geological matrices;^{55,56} and determination of

dopants and major components in optical single crystals. 31,57,58 Best wavelength (λ) selection is addressed. The basis of these investigations is the Q-concept using the "true detection limits criterion", developed by Boumans et~al. The quantification of the interferences is achieved in terms of Q-values for line interference $Q_1(\lambda_a)$ and wing background interference $Q_W(\Delta \lambda_a)$. The "best" analytical lines with a view to minimizing spectral interferences were selected and the true LODs by using these lines were calculated. Up to 15 REEs (Ce, Dy, Er, Eu, Gd, Ho, La, Lu, Nd, Pr, Sm, Tb, Tm, Y, Yb) have been treated in these systematic studies by Daskalova and co-workers (Table 1).

A special emphasis is put on the development of slurry sampling (SS) and solid sampling (SoS) ETAAS methods for trace analysis in complex, hardly soluble matrices. These approaches are environmentally friendly and exhibit lower reagent/procedural blanks, lower contamination risk, reduced sample consumption and other positive assets. For example, SS or SoS ETAAS methods are developed for the determination of toxic trace elements such as As, Cd, Hg, Mn, Pb, and Sn in biological tissues, algae, soils and sediments, 60-62 as well as for the determination of trace element content in single crystals of optical materials. 63,64

Since Bulgaria is a major producer of wines, recent research work has been devoted to the analysis of wines by developing robust, sensitive and precise methods for the determination of total concentrations of trace elements like As, Cd, Cr, Cu, Hg, Fe, Ni, Pb, Tl and Se in wines using various AS techniques (ETAAS, HGAAS, CV-AAS, VG-FAFS, FAAS),^{20,65-68} as well as their relevant fractions and chemical species.^{20,65,69-71}

Several studies are devoted to the determination of toxic trace elements in a broad variety of environmental samples such as air, ²⁶ natural waters, ^{14,72–86} soils and sediments, ^{48,62,87} and road dusts ^{25,27,88,89} with a view to evaluating environmental pollution by industrial, agricultural and traffic activities, drawing heavy metal fallout maps, ⁹⁰ etc.

The composition of a large variety of archaeological bones has been addressed in archaeometric studies. The obtained results are used to reconstruct paleodiet.^{91,92} Major and minor constituents of medieval glasses are determined with a view to shed light on medieval recipe norms for glass production.¹³

As a result of the studies on sample pretreatment coupled to AS techniques, numerous separation and enrichment procedures are developed and optimized. Special attention is paid to the flow injection (FI) or CF (continuous flow) on-line coupling of preconcentration procedures with the subsequent AS detection techniques such as ETAAS, ICP-OES and ICP-MS. Solid phase extraction (SPE) procedures using sorbent nano- and microbeads are proposed for the enrichment of metal ions from waters and biological fluids;26,28,76,79-81,85 ion exchangers and chelating resins are evaluated with the same purpose. 27,78,86,93 Preconcentration on new ion imprinted polymers (IIPs) is used for the determination of Cu in waters.⁷⁵ Preconcentration of a large number of analytes is realized by liquid-liquid extraction (LLE) of their chelate complexes, chlorides or iodides in isobutylmethyl ketone (IBMK)^{66,67,87} or by flotation.⁷² Cloud point extraction (CPE) preconcentration procedures are applied to the determination of PGMs in pharmaceutical preparations.⁴⁶ Reductive precipitation of the matrix is applied for the determination of trace elements in high-purity gold⁹⁴ and silver.³⁵

Electrochemical separation of Pb is used prior to its isotope ratio determinations in Ag, Cu and their alloys in archaeometric investigations.⁹⁵

A series of studies is devoted to the optimization of the FI preconcentration of trace elements as chelate complexes in a knotted reactor (KR) and its on-line coupling with ETAAS using various chelating reagents and preconcentration techniques. 73,74,88,96,97 The FI system is further modified with a view to its on-line coupling with ICP-time-of-flight(TOF)-MS. 25,82-84

Analytical procedures using solid phase extraction (SPE), ion exchange (IE), LLE, as well as fractionation schemes were developed for the speciation determination of toxic elements like As, Cr, Hg, Tl, Fe, Pt, Sb, and Se in various food items, 50,51,53 beverages, 118-120 natural waters, 15,19,52,121-126 soils and sediments, 16-21,127-129 body fluids and tissues, 17,101,130 road dust, 131 etc., with a view to assess bioavailability, biotransformations and distribution patterns.

The metrological characterization of the AS methods, providing metrological traceability and uncertainty budget evaluation, is addressed in several papers. 61,102,107,108

Bulgarian analytical atomic spectroscopists have contributed *chapters* in encyclopedia¹³² and scientific *books* on AAS,¹³² AAS in environmental, biological and food analysis,²⁴ palladium in environmental materials,¹³³ analysis of Au alloys by FAAS as a practical example on traceability, measurement uncertainty and validation,¹⁰² as well as *reviews* on ICP-MS using FI on-line preconcentration and time-of-flight mass analyser (ICP-TOF-MS),¹³⁴ "VG or ETAAS—Both!",²² advances, anticipations and reality with permanent modification in ETAAS,³⁹ speciation analysis for Hg in the aquatic environment,²³ SPE in the determination of Au, Pd and Pt,¹³⁵ VG techniques for Cd,¹⁴ and analysis of wine.¹³⁶

The above survey indicates that the research activities of Bulgarian atomic spectroscopists in the new millennium have led to noticeable contributions to the field of analytical atomic spectroscopy like methodological improvements in sample pretreatment, chemical modification, trace element enrichment, quantification of spectral interferences in ICP-OES with an emphasis on REEs, development of speciation and coupled techniques, as well as in numerous application studies. Substantial progress in international visibility of research results and their documentation has been reached in the recent two decades. The role of international cooperation between Bulgarian and European scientists, realized by joint projects, research stays, and informal contacts and collaborations, is gradually increased by involving younger researchers.

An outlook on near future for Bulgarian atomic spectroscopy, although uncertain, is worth attempting. On the basis of the authors' experience and knowledge of available research facilities, current (mainly academic) staff resources, international research links and proximate projects it could be expected that some traditional research and application fields such as trace element enrichment, speciation analysis, characterization of novel/advanced materials and metal complexes, environmental analysis, improving analytical methodology for emerging (micro, nano) pollutants, food and agriculture, water (with high TDS) and plant analysis, and archaeometric investigation would be continued. There are good prospects for analytical developments and applications to food, beverages and food additives,

particularly local food items such as fish, wine, spirits, juices, honey, herbs and teas, nuts, plant extracts, vegetable oils, essential oils, as well as pharmaceutical research and development. Metrological aspects of analysis (thorough validation, uncertainty budgeting, robustness, selectivity, reference materials) will be continuative. The general trends to environmentally friendly treatments and methodologies (UV, sonication, MW, scaling down, etc.) in digestion, enrichment and quantification would be considered. Moderate progress in research on novel materials in sample preparation schemes (IIPs, MIPs, nanosized materials, novel reagents, etc.) is anticipated. Some decline of FAAS, ETAAS and CVG would be expected, while research and novel developments on multielement plasma-based techniques will grow. Publication activities are expected to stabilize at a reasonable level of 75-80% in international journals, with over 50% international co-authorship. Further success owing to the imagination and creativity of the younger generation of students and researchers is an optimistic expectation.

List of abbreviations

| | | GC | Gas chromatography |
|------------------------|--|-------------|---|
| 1 | Integrated absorbance (peak area | GT | Graphite tube |
| $A_{ m int}$ | measurements) | HF-PLM | Hollow fiber permeation liquid membrane |
| 1 | Absorbance (peak height measurements) | HG | Hydride generation |
| $A_{ m p}$ AAS | Atomic absorption spectrometry | HGAAS | Hydride generation atomic absorption |
| addn | Addition | | spectrometry |
| AFS | Atomic fluorescence spectrometry | HMA- | Hexamethylene ammonium |
| APDC | Ammonium pyrrolidinedithiocarbamate | HMDTC | hexamethylenedithiocarbamate |
| | Aqueous | HPLC | High performance liquid chromatography |
| aq. | • | IBMK | Isobutylmethyl ketone |
| aq. regia AS | Aqua regia (HNO ₃ –HCl, 1 : 3 v/v) | IC | Ion chromatography |
| | Analytical atomic spectrometry Arsenobetaine | ICP | Inductively coupled plasma |
| AsB AsC | Arsenobetaine | ICP-MS | Inductively coupled plasma mass spectrometry |
| | | ICP-OES | Inductively coupled plasma optical emission |
| ASDI BCR | Air-segmented discrete sample introduction | | spectrometry |
| BICIN | Community Bureau of Reference | ID | Isotope dilution |
| | N,N-Bis(2-hydroxyethyl)-glycine Characteristic concentration | IIP | Ion imprinted polymer |
| C _o calibrn | | inorg. | Inorganic |
| | Calibration | interf. | Interference |
| CDTA | Cyclohexane-1,2-diamine-N,N,N',N'- | IRMM | Institute for Reference Materials and |
| CE | tetraacetic acid | | Measurements (Geel, Belgium) |
| CF | Continuous flow | IS | Internal standard |
| corrn | Correction | IE | Ion exchange |
| CPE | Cloud point extraction | KR | Knotted reactor |
| CRM | Certified reference material | LC | Liquid chromatography |
| CT | Cryogenic trap (trapping) | LLE | Liquid-liquid extraction |
| CV | Cold vapor | LOD | Limit of detection |
| CVAAS | Cold vapor atomic absorption spectrometry | LOQ | Limit of quantitation |
| CVAFS | Cold vapor atomic fluorescence spectrometry | LSME | Liquid semi-microextraction |
| CVG | Chemical vapor generation | LTPA | Low temperature plasma ashing |
| CVT | Cold vapor technique | $m_{\rm o}$ | Characteristic mass for integrated absorbance |
| Cys | L-Cysteine | | measurements |
| DAD | Diode array detector | $m_{ m p}$ | Characteristic mass for peak height |
| d.c. | Direct current | r | measurements |
| DEDTC | Diethyldithiocarbamate | MDF | Miniature diffusion flame |
| detd | Determined | MMA | Monomethylarsonate |
| detn | Determination | MS | Mass spectrometry |
| DGT | Diffusion gradients in thin-film gels | | 1 |

dign

dild

diln

dissoln

DMA

DTC

DTPA

EDTA

eqn

ETV

extrn

FAAS

FAFS

FΙ

FIF

FIT

EPXMA

ETAAS

DPCSV

Digestion

Diluted

Dilution

Dissolution

voltammetry

Equation

spectrometry

Flow injection

Flame-in-flame

Flame-in-tube

Extraction

Dimethylarsinate

Dithiocarbamate

Differential pulse cathodic stripping

Diethylenetriaminepentaacetic acid

Electron probe X-ray microanalysis

Electrothermal atomic absorption

Flame atomic absorption spectrometry

Flame atomic fluorescence spectrometry

Electrothermal vaporization

Ethylenediaminetetraacetic acid

m/v Mass/volume MW Microwave ND Non-dispersive

OES Optical emission spectroscopy (spectrometry)

org. Organic (organo-)
PAR 4-(2-Pyridylazo)resorcinol
PGM Platinum group metal

PMBP 1-Phenyl-3-methyl-4-benzoyl pyrazolone-5

ppt Precipitate
pptn Precipitation
preconcn Preconcentration
QTA Quartz tube atomizer

R Recovery

REE Rare earth element RM Reference material

RSD Relative standard deviation

sens. Sensitivity sepn Separation

SEC Size exclusion chromatography

SiG Silica gel
SoS Solid sampling
SPE Solid phase extraction
SRM® Standard reference material

SS Slurry sampling std Standard

STPF Stabilized temperature platform furnace

 $T_{\rm at}$ Atomisation temperature $T_{\rm cl}$ Clean temperature

 T_{coll} Collection (trapping) temperature

 $T_{
m pyr}$ Pyrolysis temperature $T_{
m vap}$ Vaporization temperature TBA Tributylammonium TDS Total dissolved solids TEA Triethanolamine

TEAH Tetraethylammonium hydroxide
THGA® Transverse heated graphite atomizer
TMAH Tetramethylammonium hydroxide

TMAO Trimethylarsineoxide TOF Time-of-flight

TOFMS Time-of-flight mass spectrometry
TOMACl Tri-*n*-octylmethylammonium chloride

TS Thermospray US Ultrasonic

USN Ultrasonic nebulizer
UV Ultraviolet (treatment)
var. Various; variable
VG Vapor generation

VGAAS Vapor generation atomic absorption

spectrometry

VGAFS Vapor generation atomic fluorescence

spectrometry

VG-FAFS Vapor generation flame atomic fluorescence

spectrometry

vol. Volume
v/v Volume/volume
XRF X-ray fluorescence

ZETAAS ETAAS with Zeeman-effect background

correction

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