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Economically viable sensitive and selective luminescent sensor for the determination of Au(III) in environmental samples†

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This paper describes the ultrasensitive and selective determination of Au(III) in an aqueous solution using a functionalized mercapto thiadiazole ligand. UV-visible and spectrofluorimetry studies of functionalized mercapto thiadiazole ligands such as 2,5-dimercapto-1,3,4-thiadiazole (DMT), 2-amino-5-mercapto-1,3,4-thiadiazole (AMT) and 2-mercapto-5-methyl-1,3,4-thiadiazole (MMT) were carried out in the presence of Au(III) ions in solution. DMT, AMT and MMT exhibit an absorption maximum at 330, 310 and 299 nm, respectively. The emission intensities of the respective compounds were enhanced at 435, 428 and 442 nm after the addition of 1 nM Au(III); furthermore, the color of the solutions also changed to yellow. The observed color changes and emission intensity enhancement are ascribed to the effective complex formation of Au(III) with DMT, AMT and MMT ligands. When 8 nM Au(III) was added into the aqueous solutions of DMT, AMT and MMT, the emission intensity was enhanced to 102, 8 and 5-fold, respectively. The binding constants for DMT, AMT and MMT-Au(iii) complexes were found to be 1.52, 1.05 and 1.04×10^5 mol⁻¹ L, respectively. The higher emission intensity and binding constant value obtained for DMT reveals that the DMT-Au(III) complex is highly fluorescent compared to the other two complexes. Thus, DMT was chosen as a fluorophore for the determination of Au(III). Interestingly, even after the addition of 1 pM Au(III) into DMT solution, the emission intensity was enhanced at 435 nm. Based on the enhancement of emission intensity, we have determined the concentration of Au(iii) and the detection limit was found to be 1 pg L^{-1} (S/N = 3). Furthermore, 60 000-fold higher concentrations of common interferences and 500-fold higher concentration of Cu(II), Pb(II), Ag(II), Ag(III) and Ag(III) did not interfere for the determination of 8 nM Au(III). The proposed method was successfully applied to determine Au(III) in different water samples and the obtained results were validated with ICP-AES. The present method of determination has several advantages, including low cost and environmental friendly nature.

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Introduction

Gold is one of the most significant noble metals, and it is widely disseminated in nature and its concentration in soil and rocks has been found to be about 1 and 4 ng g⁻¹, respectively, and 50 and 200 pg mL⁻¹ have been found in sea water and river water, respectively.¹⁻³ Gold has many practical uses due to its unique physical and chemical properties, and it is very useful in pollution control, mobile phones, laptops, space travel and dentistry.^{4,5} For example, gold was used as a colouring agent in cranberry glass to produce an intense red color.⁶ Since gold can

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† Electronic supplementary information (ESI) available: Effect of ligands concentrations on the fluorescence intensity of 8 nM Au($\rm III$); effect of pH; Job's plots; possible coordination mechanism scheme; emission spectra of 0.5 mM DMT in the presence 10 pM to 1 nM Au($\rm III$). See DOI: 10.1039/c4ra05217j

reflect the infrared, visible light and radio waves, it is used as a coating material of many satellites and planes. It has also an anti-inflammatory property; thus, it is used in pharmaceuticals for the treatment of arthritis and tuberculosis.^{7,8}

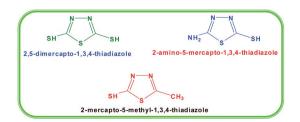
Gold-based drugs are valuable for the treatment of a wide variety of diseases, including rheumatic arthritis, asthma, tuberculosis, malaria, cancer, HIV and brain lesions. 9-11 In recent years, gold nanoparticles have been used as sensors, drug and gene deliveries and molecular imaging probes. 12-14 Moreover, some gold complexes have been utilized as effective drugs in treating rheumatoid arthritis, psoriatic arthritis and bronchial asthma. 9-11 On the other hand, excess of gold may be toxic to human and animal organisms. It inhibits the effect upon the activity of many enzymes and DNA separation because of its strong binding ability. 15,16 Au(III) is one of the most potent sensitizers with a high incidence of allergic reactions like contact dermatitis, rhinitis, conjunctivitis, asthma and urticarial, and it causes cell toxicity in living organisms. 17 Soluble gold salts such as gold chloride is known to cause damage to the

liver, kidneys, and the peripheral nervous system of animals. ¹⁷ The guideline values of Au(III) for various aquatic organisms are reported to be 6–75 $\mu g \ L^{-1}$. Although gold and its complexes have been extensively used for different applications, they are also toxic to living beings; therefore, it is essential to determine a simple and cost effective method for its concentration determination.

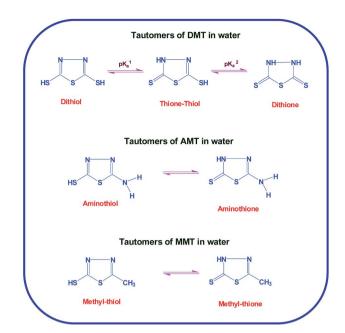
Several methods have been used to determine Au(III), which includes atomic absorption spectrometry,18 inductively coupled plasma atomic emission spectrometry,19 cloud point extraction, 20 solid phase extraction, 21a,b colorimetry 22 and voltammetry.23 However, these methods have several demerits such as long analysis time, requiring sample pretreatment, low sensitivity and low selectivity. 18-21 On the other hand, spectrofluorimetric method of determination has several merits, including high sensitivity and selectivity and less time consumption.24-28 To date, only few papers were reported for the determination of Au(III) in environmental samples by a fluorimetry method.24-28 In these examples, Au(III) was determined by using different organic fluorophores, including propargylamide,24 a spirolactam derivative,25 rhodamine hydrazide cyanatobenzene,26 N-propargyl-rhodamine lactam27 and fluorescent red GK.28

However, many of the above organic fluorophores are insoluble in water and therefore are unsuitable for real sample analysis; ^{29a,b} moreover, the organic solvents are harmful to environment. Furthermore, these conventional organic fluorescent molecules usually suffer from notorious limitations such as low signal intensities and photobleaching and most of them tend to have narrow excitation spectra and exhibit a broad emission band with red tailing. ^{29a,b} In addition, these fluorophores failed to determine Au(III) with high sensitivity and selectivity.

DMT, AMT and MMT are interesting heterocyclic compounds containing more than four coordinating sites (Scheme 1). These compounds are bioactive and are used as a metal chelating agents, lubricant additives like corrosion inhibitors and anti-wear agents, cross-linkers for polymers and as components of cathode material battery systems. ^{30–32} DMT is used as a bactericide ^{33,34} while AMT is extensively used in the field of electroanalysis to sense many biological and toxic chemicals with high selectivity and sensitivity. ³² MMT is used as a lubricant additive in corrosion inhibitors and anti-wear agents, cross-linkers for polymers and as components of cathode material battery systems. ³⁵ DMT, AMT and MMT are



Scheme 1 Structures of DMT, AMT and MMT.



Scheme 2 Tautomers of DMT, AMT and MMT in water.

used as a capping ligand for the synthesis of metal nanoparticles.³⁶ They exhibit tautomeric equilibria in protonated and deprotonated forms (Scheme 2).³⁷ These can facilitate formation of metal complexes.

In the solid state, DMT exists as a thiolate tautomer, while in solution it exists as dithiol, thiolate and dithione forms (Scheme 2A).³⁸ It has acid dissociation constants of $pK_a^1 = -1.36$ and $pK_a^2 = 7.5$, making its neutral form a very strong acid and its anionic form a weak acid.^{39,40} The pK_a values for AMT and MMT are 6.94 and 5.8, respectively, and they act as chelating ligands.⁴⁰ These compounds can coordinate through either both nitrogen atoms or both the thiocarbonyl sulfur atoms or one nitrogen and one sulfur atom on either the same side or different sides. The amine group of AMT can also coordinate with metal ions; moreover, the π -electron in the aromatic heterocyclic ring can also coordinate with metal ions.⁴¹

In this paper, we are reporting the absorption and emission spectral studies of DMT, AMT and MMT in the presence of Au(III) in aqueous solution. UV-visible spectra of DMT, AMT and MMT exhibit absorption peaks at 330, 310 and 299 nm, respectively. Interestingly, while adding even 1 nM concentration of Au(III) into a DMT, AMT or MMT solution, the emission intensity was increased to 102, 8 and 5-fold, respectively. Among the three compounds, DMT showed extreme sensitivity and selectivity by determining 8 nM Au(III) in the presence of 60 000fold higher concentration of common interferences, excluding Cu(II), Pb(II), Ag(I), Ag(II), Ag(III) and Hg(II). Therefore, it was successfully used for the determination of Au(III) in different water samples. The present method was also validated with ICP-AES. Importantly the advantages of present method include low cost, being environmentally friendly and highly selective and sensitive.

2. Experimental

2.1. Chemicals

2,5-Dimercapto-1,3,4-thiadiazole, 2-amino-5-mercapto-1,3,4-thiadiazole, 2-mercapto-3-methyl-1,3,4-thiadiazole and hydrogen tetrachloroaurate trihydrate (HAuCl₄·3H₂O) were purchased from Sigma-Aldrich and were used as received. The phosphate buffer solutions (PBS) with different pH values were prepared by using Na₂HPO₄ and NaH₂PO₄. All other chemicals used in this investigation were of analytical grade and used directly without further purification. Millipore Milli-Q (18 M Ω cm) water was used in all experiments. For the real sample analysis, we have collected the tap water from Gandhigram, river water from Vaigai River, and lake and pond water samples from Nilakottai village, Tamilnadu, India.

2.2. Reagents

Standard stock solutions of Au(III) (1 mM) and ligands (DMT, AMT, MMT) (5 mM) were prepared with millipore water. The working solution of Au(III) was obtained by appropriate dilution of the stock solution with millipore water. The solutions of Fe²⁺, Co²⁺, Cd²⁺, Ni²⁺, Mn²⁺, Cu²⁺, Mg²⁺, Zn²⁺, Ca²⁺, Na⁺, K⁺, Cr³⁺, Pb²⁺, Hg²⁺, Ag⁺, Ag²⁺ and Ag³⁺ (1 mM) were prepared from their respective salts.

2.3. Instrumentation

Absorption spectra were measured by using JASCO V-550 UV-visible spectrophotometer. Fluorescence spectral measurements were performed on a JASCO FP-6500 spectrofluorimeter equipped with a xenon discharge lamp using a 1 cm quartz cell at room temperature. Inductively coupled plasma atomic emission spectra were measured by using thermo electron IRIS intrepid II XSP DUO model ICP-AES.

2.4. Procedure for absorption and fluorescence spectral measurements

A known amount of Au(III) and ligands from stock solution were added to 10 mL flask, and then diluted to the mark with millipore water. 3.0 mL of each solution was transferred to the quartz cell and the absorbance was measured against a corresponding reagent as reference. The fluorescence measurements were carried out with excitations at 330, 310 and 299 nm.

3. Results and discussion

3.1. Absorption and emission spectra of DMT, AMT and MMT

UV-visible spectrum of DMT exhibits the absorption peak at 330 nm with a shoulder band at 257 nm and these bands are ascribed to the dithiol and thiolate forms of DMT, respectively (Fig. 1A; curve a). ^{38,39} The UV-visible spectra of AMT and MMT exhibit the absorption band at 310 and 299 nm, respectively (Fig. 1B and C; curve a). The emission spectra of DMT, AMT and MMT showed the emission wavelength maximum ($\lambda_{\rm em}$) at 435, 428 and 442 nm with an excitation wavelength ($\lambda_{\rm ex}$) of 330, 310 and 299 nm, respectively (Fig. 1; curve a).

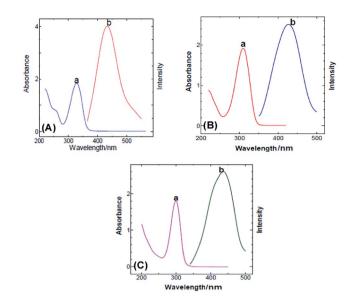


Fig. 1 (a) UV-visible and (b) emission spectra of (A) DMT, (B) AMT and (C) MMT. ((A) $\lambda_{\rm ex}/\lambda_{\rm em}$: 330/435 nm; (B) $\lambda_{\rm ex}/\lambda_{\rm em}$: 310/428 nm; (C) $\lambda_{\rm ex}/\lambda_{\rm em}$: 299/442 nm).

3.2. Absorption spectral studies of DMT, AMT and MMT in the presence of Au(III) ions

The absorption spectra of 0.5 mM DMT in the presence of different concentrations of Au($_{\rm III}$) are shown in Fig. 2A. DMT exhibits the absorption band at 330 nm with a shoulder band at 257 nm. After the addition of 1 $_{\rm IM}$ Au($_{\rm III}$), the absorbance at 330 nm was decreased (curve b), and the color of the solution was

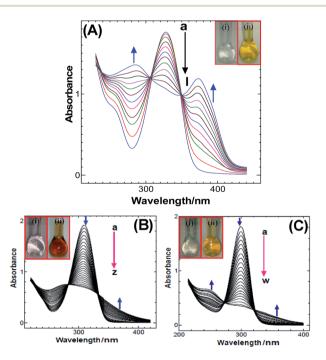


Fig. 2 UV-visible spectra of (A) DMT, (B) AMT and (C) MMT in the presence of different concentrations of Au(III): (a) 0, (b–z) each increment 1×10^{-6} M Au(III). Insets: (i) images of 0.5×10^{-3} M (A) DMT, (B) AMT and (C) MMT (i) absence and (ii) in the presence of 1×10^{-6} M Au(III).

changed from colorless to yellow (inset of Fig. 2A). When 3 µM

Au(III) was added, the absorption at 330 nm decreased and a new broad band was observed at 380 nm. Furthermore, on increasing the concentration of Au(III), the absorbance at 330 nm was decreased and the broad band became a predominant peak. After the addition of 9 µM Au(III), the shoulder band at 257 nm had vanished and a new band was observed at 290 nm. The color of the solution also becomes an intense yellow. When 11 μM Au(III) was added, the peak at 330 nm had completely vanished and the two broad bands were predominant at 280 and 380 nm. We have calculated the molar extinction coefficient of peaks at 280 and 380 nm after the addition of 11 µM Au(III), and they were found to be 1.3×10^5 and 1.1×10^5 M⁻¹ cm⁻¹, respectively. Interestingly, two well-defined isosbestic points were observed at 300 and 350 nm. The observed spectral and color changes suggested that DMT forms a complex with Au(III). Furthermore, the obtained isosbestic point reveals that the neat conversion of free DMT into complexed DMT-Au(III) species.

Fig. 2B shows the absorption spectra of AMT in the presence of different concentrations of Au(III). When 1 µM Au(III) was added, the absorbance at 310 nm decreased and the color of the solution changed from colorless to yellow (curve b; inset of Fig. 2B). Furthermore, on increasing the concentration of Au(III), the absorbance was dramatically decreased and a broad band was observed at 350 nm. Similar to DMT, in this case also two well-defined isosbestic points were observed at 285 and 330 nm. After the addition of 26 µM Au(III), the absorbance at 310 nm was completely vanished and the absorbance of broad band at 350 nm was increased. The observed spectral and color changes are ascribed to the complex formation between AMT and Au(III).

The absorption spectrum of MMT exhibits an absorption maximum at 299 nm (Fig. 2C; curve a). After adding 1 μM Au(III), the absorption at 299 nm was decreased (Fig. 2C; curve b). The color of the solution was changed from colorless to yellow (Inset of Fig. 2C). Furthermore, on increasing the concentrations of Au(III), the absorption at 299 nm was decreased and new broad bands were observed at 265 and 340 nm. Similar to DMT and AMT, two well-defined isosbestic points were also observed for MMT at 270 and 320 nm. The observed spectral and color changes confirm the complex formation between DMT and Au(III). Generally, gold ions have the strong thiophilic tendency with sulfur containing ligands that form strong complexes. 42,43 DMT, AMT and MMT have one sulphur and two nitrogen atoms in their structure. Gold ions coordinate with sulfur and nitrogen donors of DMT, AMT and MMT and form strong complexes.44

3.3. Emission spectral studies of DMT, AMT and MMT with Au(III) ions

The emission spectra of DMT, AMT and MMT in the presence of different concentrations of Au(III) are shown in Fig. 3. Fig. 3A shows the emission spectrum of DMT, which exhibits an emission maximum at 435 nm with excitation at 330 nm. While adding of 1 nM Au(III) into DMT, the emission intensity was increased 102-fold at 435 nm. Furthermore, on increasing the concentration of Au(III) from 2 to 8 nM, the emission intensity at 435 nm was dramatically increased.

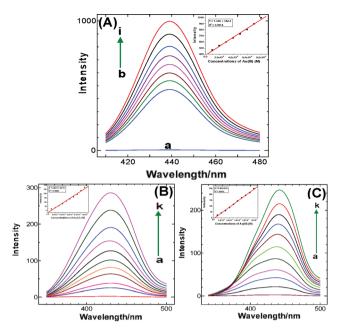


Fig. 3 Emission spectra of 0.5×10^{-3} M (A) DMT, (B) AMT and (C) MMT in the presence of different concentrations of Au(III): (a) 0, (b-z) each increment is 1×10^{-9} M of Au(III) ((A) $\lambda_{\rm ex}/\lambda_{\rm em}$: 330/435 nm; (B) $\lambda_{\rm ex}/\lambda_{\rm em}$: 310/428 nm; (C) $\lambda_{\rm ex}/\lambda_{\rm em}$: 299/442 nm). Insets: linearity plot of (A) DMT, (B) AMT and (C) MMT in the presence of different nanomolar concentrations of Au(III).

Fig. 3B (curve a) shows the emission spectrum of AMT. It exhibits an emission maximum at 428 nm with excitation at 310 nm. Furthermore, on increasing the concentration of Au(III), the emission intensity at 428 nm was increased. A good linearity was observed from 1 nM to 0.1 μ M ($R^2 = 0.9901$). MMT exhibits an emission maximum at 442 nm with an excitation wavelength of 299 nm (Fig. 3C; curve a). After the addition of 1 nM Au(III), the emission intensity at 442 nm was enhanced (Fig. 3C; curve b). Furthermore, on increasing the concentration of Au(III), the emission intensity at 442 nm was dramatically enhanced. Good linearity was observed from 1 nM to 0.1 μ M ($R^2 = 0.9972$). Generally, the terminal sulfur atoms of DMT, AMT and MMT and amine group of AMT establish strong σ-bonds with Au(III) that are stabilized by $p\pi$ -d π back-bonding and result in the formation of strong complexes.45 The observed emission intensity enhancement of DMT, AMT and MMT at 435, 428 and 442 nm, respectively, after the addition of Au(III) are ascribed to the photo-induced charge transfer and chelation enhanced fluorescence.46,47

3.4. Effect of ligand concentration, pH and stoichiometric composition

The effect of DMT, AMT and MMT concentration on the fluorescence intensity was examined in the range from 0.05 to 0.6 mM in the presence of 8 nM Au(III) (Fig. S1; ESI†). The fluorescence intensity was rapidly enhanced as the concentration of DMT, AMT and MMT was increased. The emission spectra of 0.5 mM DMT, AMT and MMT in 8 nM Au(III) showed the maximum fluorescence intensity, and it remains constant beyond this

concentration (Fig. S1; ESI†). Therefore, we have fixed the concentration of 0.5 mM DMT, AMT and MMT for all the experiments.

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In order to establish an optimum pH of DMT, AMT and MMT-Au(III) complexes, we have taken phosphate buffer solutions in the pH range of 1.0–10.0 and monitored the fluorescence intensity. The fluorescence intensity of the compounds in the presence of 8 nM Au(III) was enhanced from pH 1–4 and showed a maximum at pH 5 (Fig. S2; ESI†). It started to decrease beyond pH 5. It has been reported that the formation of complexes are favourable in mildly acidic conditions. ⁴⁵ Hence, we obtained maximum emission intensity at pH 5.0 (Fig. S2; ESI†). Thus, we have maintained pH 5.0 for all the experimental solutions. Job's method was applied to ascertain the stoichiometric composition of the complex formed between Au(III) and DMT, AMT and MMT. The stoichiometric composition of DMT–Au(III) was found to be 1:1. AMT–Au(III) and MMT–Au(III) stoichiometric compositions were found to be 1:2 (Fig. S3; ESI†).

3.5. Binding constant and possible binding mechanism of DMT, AMT and MMT-Au(III) complexes

The binding constant (K_A) value can be estimated by using the following equation:⁴⁸

$$\log[F_0 - F/F_0] = \log K_A + n \log[Q]$$

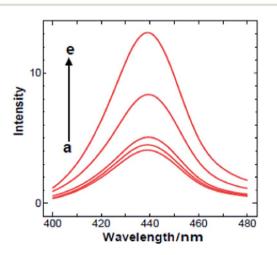
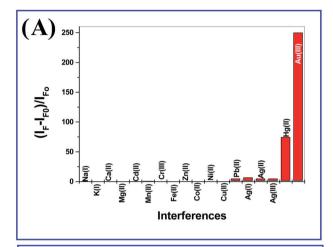


Fig. 4 Emission spectra of 0.5 mM DMT in the presence of different concentrations of Au(III): (a) 0, (b) 1, (c) 2, (d) 5 and (e) 10 pM Au(III) ($\lambda_{ex}/\lambda_{em}$: 330/435 nm).

where F_0 is the fluorescence intensity of ligands, F is the fluorescence intensity of ligand in the presence of Au(III), K_A is the binding constant, n is the number of binding sites and Q is the concentration of Au(III). K_A and n can be measured from the intercept and slope obtained through plotting $\log[F_0 - F/F_0]$ against $\log[Q]$. The binding constant (K_A) value of DMT-Au(III), AMT-Au(III) and MMT-Au(III) were found to be 1.5225 \times 10⁵ mol⁻¹ L, 1.0473 mol⁻¹ L and 1.04488 mol⁻¹ L for DMT-Au(III), AMT-Au(III) and MMT-Au(IIII) complexes, respectively. The binding sites (n) of all the compounds were found to be \sim 1. The



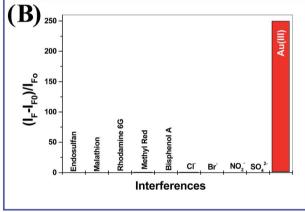


Fig. 5 Selectivity of the DMT towards Au(III). The emission response of Au(III) (8 nM) in 0.5 mM DMT solution in the presence of (A) 0.48 mM of Na⁺, K⁺, Ca²⁺, Mg²⁺, Fe²⁺, Cd²⁺, Cr³⁺, Mn²⁺, Zn²⁺, Co²⁺, Ni²⁺, Cu²⁺ and 40 μ M Pb²⁺, Ag⁺, Ag²⁺, Ag³⁺ and 8 nM Hg²⁺. (B) 0.48 mM endosulfan, malathion, rhodamine 6G, methyl red, bisphenol A, Cl⁻, Br⁻, NO₂⁻ and SO₄²⁻.

Table 1 Comparison of Au(III) detection limit obtained in the present study with the reported fluorophores by the fluorimetry method

| S. no. | Ligand | Medium | Detection limit | Ref. |
|--------|---------------------------------------|--------------------|----------------------------|-----------|
| 1 | Propargylamide | DMSO/acetonitrile | $63~\mu\mathrm{g~L}^{-1}$ | 24 |
| 2 | Spirolactam rhodamine derivative | Acetonitrile/PBS | $0.4~\mathrm{mg~L^{-1}}$ | 25 |
| 3 | Rhodamine hydrazide isocyanatobenzene | DMF/PBS | 56 $\mu g L^{-1}$ | 26 |
| 4 | N-Propargyl-rhodamine lactam | DMSO/HEPES | 78 $\mu g L^{-1}$ | 27 |
| 5 | Fluorescent red GK | Acetonitrile/HEPES | $0.33~{ m mg}~{ m L}^{-1}$ | 28 |
| 6 | 2,5-Dimercapto-1,3,4-thiadiazole | Water | 1.0 pg L^{-1} | This work |

Table 2 Real sample analysis

| Samples | Au(III) added (ng L ⁻¹) | Au(III) found (ng L ⁻¹) | RSD | Recovery (%) |
|-------------|--|--|------|--------------|
| Tap water | 0 | _ | | _ |
| rup water | 10 | 9.90 ± 0.11 | 0.47 | 99.1 |
| | 20 | 20.00 ± 0.04 | 0.23 | 100.2 |
| River water | 0 | _ | | _ |
| | 10 | 10.01 ± 0.01 | 0.66 | 100.2 |
| | 20 | 19.81 ± 0.11 | 1.02 | 99.6 |
| Lake water | 0 | _ | _ | _ |
| | 10 | 9.90 ± 0.09 | 0.58 | 99.9 |
| | 20 | 19.59 ± 0.19 | 0.42 | 98.9 |
| Pond water | 0 | _ | _ | _ |
| | 10 | 10.02 ± 0.02 | 0.35 | 100.4 |
| | 20 | $\textbf{19.98} \pm \textbf{0.01}$ | 0.47 | 99.9 |
| | | | | |

obtained higher binding constant value of DMT–Au(III) suggests that there is a strong binding force between DMT and Au(III).

Based on the pH effect, binding sites and binding constant values, we have proposed the possible coordination sites of DMT, AMT and MMT with Au(III) (Scheme S1; ESI†). DMT in water exists in the thione-thiol and dithione forms. At pH 1, it mainly exists in the dithiol form. While increasing the pH of the solution, the amount of thione-thiol form increases. Based on the obtained high emission intensity of DMT at pH 5, we have concluded that the thione-thiol form is higher at pH 5. It has been well established that the formation of DMT-metal complex is favorable in mildly acidic conditions.39,40 Therefore, the effective complex formation is favourable at pH 5. Beyond pH 5, the fluorescence intensity decreases due to the decrease in concentration of the thione-thiol form.³⁸ These results suggests that high pH is not favorable for the effective formation of DMT complexes. Similar behavior was also observed for AMT and MMT ligands. 40-45 From the observed higher emission intensity at pH 5, we have concluded that the amino-thione and methylthione forms of AMT and MMT, respectively, at pH 5 more favorably form AMT- and MMT-Au(III) complexes. 40-45 DMT, AMT and MMT can coordinate with Au(III) through the sulfur and nitrogen atoms. The π -electron in the aromatic heterocyclic ring can also coordinate with Au(III) and forms the polymeric structure⁴¹ (Scheme S1; ESI†).

3.6. Determination of Au(III) using DMT as fluorophore

Interestingly, the emission intensity was enhanced to 102-fold at 435 nm while adding 1 nM Au(III) into DMT in contrast to AMT and MMT. Thus, we have chosen DMT for the sensitive determination of Au(III). Obviously, the emission intensity of DMT at 435 nm was enhanced even after the addition of 1 pM

Au(III) (Fig. 4; curve b). We have systematically increased the concentration of Au(III) from 1 pM to 8 nM; therefore, the emission intensity at 435 nm dramatically enhanced (Fig. 3A, 4 and S4 (ESI†)). Note that good linearity was observed form 1 pM to 8 nM ($R^2 = 0.9932$). Based on the enhancement of emission intensity, the concentration of Au(III) was determined and the detection limit was found to be 1 pg L⁻¹ (S/N = 3). The detection limit of Au(III), fluorophore, medium obtained in the present method were compared with the other fluorophores and are given in Table 1. As seen from Table 1, the present method of detecting Au(III) is more sensitive than the previously reported fluorophores.

3.7. Effect of interferences

The effect of various interferences for the determination of Au(III) concentration was investigated using the sample containing 8 nM Au(III). As shown in Fig. 5, DMT can selectively bind with Au(III) even in the presence of 60 000-fold higher concentrations of Na⁺, K⁺, Ca²⁺, Mg²⁺, Fe²⁺, Cd²⁺, Cr³⁺, Mn²⁺, Zn²⁺, Co²⁺ and Ni²⁺. In our previous reports, we have reported DMT can selectively bind with Pb(II) and Hg(II) even in the presence of 50 000-fold higher concentration of common interferences. 49a,b In this study, we have found that the presence of 60 000-fold concentrations of Cu(II), Pb(II), Ag(I), Ag(II), Ag(III) and Hg(II) interfere for the determination of 8 nM Au(III). However, with the addition of 1 mM ETDA as a masking agent, no interference was observed in the presence of 500-fold higher concentrations of Cu(II), Pb(II) and Ag(I), Ag(II) and Ag(III). It should be noted that 60 000-fold higher concentration of Hg(II) did interfere for the determination of 8 nM Au(III) even in the presence of EDTA.

Furthermore, we have examined the interference of other toxic chemicals such as pesticides (endosulfan and malathion), organic dyes (rhodamine 6G and methyl red), plastic contaminant (bisphenol A) and anions (Cl $^-$, Br $^-$, NO $_2$ $^-$ and SO $_4$ 2 $^-$) for the determination of 8 nM Au(III). We have found that 60 000-fold higher concentration of above mentioned toxic contaminants do not interfere for the determination of 8 nM Au(III) (Fig. 5B). Hence, excluding Cu(II), Pb(II), Ag(I), Ag(II), Ag(III) and Hg(II), 60 000-fold higher concentration of all other metal ions do not interfere with the concentration determination of 8 nM Au(III).

3.8. Real sample analysis and validation with ICP-AES

The present method was utilized to determine Au(III) in tap water, river water, lake water and pond water samples (Table 2). The recovery of 98.9% and 100.4% was observed and good

Table 3 Validation of the present method determination of Au(III) concentration with ICP-AES

| | $Au(III)$ found (mg L^{-1}) | | | | |
|-------------------------------------|-------------------------------------|-----------------|------|--------|--------|
| Au(III) added (mg L ⁻¹) | Fluorimetry method (this work) | ICP-AES method | RSD | T-test | F-test |
| 0.05 | 0.05 ± 0.002 | 0.05 ± 0.005 | 0.52 | 0.4390 | 1.0 |
| 0.1 | $\textbf{0.101} \pm \textbf{0.001}$ | 0.101 ± 0.002 | 0.38 | 0.3152 | 1.0 |

agreement was obtained between spiked and measured Au(III). Furthermore, we have found that the tap water, river water and lake water do not contain any amount of Au(III). The present method was validated with ICP-AES method (Table 3). On the basis of T-test and F-test, the results obtained from the present method are in good agreement with the results obtained from ICP-AES method.

4. Conclusions

In this paper, we have systematically studied the UV-visible and emission spectral behavior of DMT, AMT and MMT in the presence of Au(III) in aqueous solution. The emission intensity was enhanced while adding Au(III) into these ligands due to the effective complex formation between these ligands and Au(III). Interestingly, the emission intensity was enhanced to 102-fold after adding 1 nM Au(III) to DMT. On the other hand, AMT and MMT showed only 8- and 5-fold of emission intensity enhancement. Surprisingly, while adding 1 pM concentration of Au(III) into DMT solution, the emission intensity was enhanced at 435 nm. Thus, we have successfully used DMT as a fluorophore for the determination of Au(III). Moreover, we have calculated the molar extinction coefficient and binding constant of AMT, DMT and MMT-Au(III) complexes, and good linearity was observed from 1 pM to 8 nM ($R^2 = 0.9932$) and the detection limit was found to be 1 pg L⁻¹. Furthermore, 60 000fold higher concentration of common interferences, excluding Hg(II), does not interfere for the determination of 8 nM Au(III) using DMT as fluorophore. We have successfully utilized the present method for the determination of Au(III) in tap, river, lake and pond water samples, and the results were validated with the ICP-AES method. The present method is economically viable and environmentally friendly.

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