

Am Chem Soc. Author manuscript; available in PMC 2013 July 25.

Published in final edited form as:

J Am Chem Soc. 2012 July 25; 134(29): 11952–11955. doi:10.1021/ja305321e.

De Novo Asymmetric Synthesis of all D-, all L- and D-/L-Oligosaccharides Using Atom-less Protecting Groups

Ravula Satheesh Babu[†], Qian Chen[†], Sang-Woo Kang[†], Maoquan Zhou[†], and George A. O'Doherty^{*}

Department of Chemistry and Chemical Biology, Northeastern University, Boston, MA 02115

Abstract

Oligosaccharide synthesis is hindered by the need for multiple steps as well as numerous selective protections and deprotections. Herein we report a highly efficient *de novo* route to various oligosaccharide motifs, of use for biological and medicinal structure activity studies. The key to the overall efficiency is the judicious use of asymmetric catalysis and synthetic design. These green principles include the bidirectional use of highly stereoselective catalysis (Pd(0)-glycosylation/post-glycosylation). In addition, the chemoselective use of C-C and C-O π -bonds functionality, as atom-less protecting groups as well as an anomeric directing group (via a Pd- π -allyl), highlights the atom economical aspects of the route to a divergent set of natural and unnatural oligosaccharides (*i.e.*, various D-/L-diastereomers of oligosaccharide as well as deoxysugars which lack *C*-2 anomeric directing groups). For example, in only 12 steps, the construction of a highly branched hepta-saccharide with 35 stereocenters was accomplished from an achiral acylfuran.

Of the biopolymers, the carbohydrates present the greatest synthetic challenges. In contrast to peptides and nucleic acids, there is a dearth of synthetic methods for the incorporation of unnatural monomers into oligosaccharide structures. While there are many methods for the de novo asymmetric synthesis of monosaccharides, the translation of these methods to oligosaccharides synthesis is severely limited.

Due to the stereochemical complexity, the hexoses have long stood as targets for synthesis. In particular, they serve as test substrates for asymmetric catalysis. As these *de novo* asymmetric routes evolved from flexible, albeit laborious, approaches to all eight hexoses to quite efficient approaches to specific hexoses, the utility of these catalytic asymmetric approaches have been limited to the synthesis of unnatural and rare monosaccharides.

As the recognition of the importance of oligosaccharides to biological processes has grown,⁵ the commensurate need for synthetic oligosaccharides intermediates has been largely met with traditional carbohydrate approaches, which draw from naturally occurring carbohydrate starting materials.⁶ While these approaches have been widely successful for supplying natural oligosaccharide structural motifs for biological studies, they have limited potential for medicinal chemistry studies.^{1,5,7} The obvious solution of merging of these two

Notes

Corresponding Author: G.O' Doherty@neu.edu.

Author Contributions

These authors contributed equally to this work and are listed alphabetically.

The authors declare no competing financial interest.

approaches, *de novo* monosaccharide synthesis followed by oligosaccharide assembly, is mired by the need for additional protection and deprotection steps that makes these processes uneconomical, not to mention un-atom economical.⁸ Thus, more convenient and efficient methods are still needed.⁹

Oligosaccharide motifs, like the nona-saccharide Man-9, are involved in a variety of important biological functions (*e.g.*, protein and cellular recognition and signaling). While multistep synthesis and isolation and semi-synthesis have provided access to these types of oligosaccharides for biological studies, these studies have been limited to the natural structural motifs. ¹⁰ For instance, having access to a hepta-saccharide motif with unnatural branching and its enantiomer (cf., **23**, Scheme 4) would enable structure activity relationship (SAR) studies of the role these complex oligosaccharide arrays (*e.g.*, Man-9) play in many biological processes. For instance, having access to both D- and L-glycoconjugates could allow for the differentiation of specific protein interaction from pharmacokinetic properties. ¹¹

To address this need, we have developed a complementary *de novo* approach to carbohydrates that is compatible with diversification and oligosaccharide assembly (Scheme 1).¹² Our approach allows for the facile incorporation of simple pyranyl-sugars (*i.e.*, under functionalized sugars) into various oligosaccharide motifs without the need for any protection/deprotection steps with complete stereocontrol.¹³ The appeal of this approach is that: the double bond in pyranone glycosyl donor (2a/2b) serves as an anomeric directing group in the Pd(0)-catalyzed glycosylation; the enone functionality of the pyranone products (3a/3b) serves as atom-less protecting groups for the *C*-2 to *C*-4 triol portion of the hexose (4a/4b); ¹⁴ and the route has equal efficiency to access to various D- and L-isomer. ^{11,12,15}

Previously we have developed a diastereoselective Pd(0)-catalyzed glycosylation reaction and demonstrated its use in the *de novo* synthesis of all D- or all L-1,4- and 1,6- α -mannotrisaccharides. 11,12 Herein, we present our discovery of an expeditious route to various highly branched/all D-, all L-and mixed D-/L-oligosaccharides using asymmetric and diasteroselective catalysis for stereocontrol. These studies resulted in a synthesis of a library of enantiopure α -rhamno/manno-trisaccharide (Schemes 2 and 3), as well as a 12-step synthesis of a highly branched hepta-saccharide with up to 35 stereocenters from achiral starting materials. This greener approach to oligosaccharides not only reduces the number of chemical transformations it also limits the number of protecting groups used (one TBS-group/hexose for hepta-saccharide 23).

We first explored the library synthesis of 1,4-α-rhamno-trisaccharides (i.e., D-D-D, D-D-L, D-L-D, D-L-L trisaccharides, 12 to 15). The route commenced with the allylic alcohol D-5, which was prepared from commercially available achiral acylfuran 1a (Scheme 1). 16 Using a Pd-catalyzed glycosylation with pyranone D-2a and reduction, monosaccharide D-5 was diastereoselectively elongated into the disaccharide 6 (Scheme 2). By changing the pyranone to its enantiomer (\alpha-Boc pyranone L-2a) the diasteromeric D-L disaccharide 7 was prepared. Repeating this divergent diastereoselective elongation of disaccharides 6 and 7 (glycosylation/reduction) provided the four possible D-/L-diastereomeric trisaccharides 8, 9, 10 and 11. These four diastereomeric routes occurred with almost the same synthetic efficiency and with virtually complete stereocontrol. A similarly diastereoselective osmium catalyzed global dihydroxylation of allylic alcohol 8, 9, 10 and 11 afforded the four desired diastereomeric 1,4-linked α-rhamno-pyranoses D-D-D-12a, D-D-L-13a, D-L-D-14a, D-L-L-15a. A final element of synthetic divergence can be employed with a global reduction of the double bonds in trisaccharide 8, 9, 10 and 11 with excess diimide 17 to give the α -1,4linked 2,3-dideoxyoligosaccharides 12b, 13b, 14b and 15b in nearly quantitative yields. An important feature of this route is that the ketone reductions and alkene dihydroxylations

occur with complete stereocontrol with respect to the local pyran stereochemistry and not the stereochemistry of the adjacent pyran rings (a fundamental requirement for atom-less protecting groups).

To further explore the stereochemical robustness of this approach, we next explored the bidirectional glycosylation of the *C*-4 and *C*-6 alcohols. The desired diol **D-16** was prepared from Boc pyranone **D-2b** in 3 steps with 69% overall yield (Scheme 3). Subjecting alcohol **D-16** to our typical Pd-catalyzed glycosylation reaction conditions provided the trisaccharide precursor trispyran **17** in 75% yield. Diastereoselective 1,2-reduction of pyranone **17** with NaBH₄ and OsO₄/NMO oxidation of the resulting allylic alcohols afforded the branched mixed D-/L-trisaccharide **18** with α-*rhamno*/*manno*-stereochemistry as a single diastereomer in 68% yield. Alternatively, by switching the dihydroxylation conditions to diimide reduction, the branched mixed D-/L-trisaccharide **19** with two 2,3-dideoxy-α-*rhamno* and one 2,3-dideoxy-*manno*-pyranose rings was prepared as a single diastereomer.

Encouraged by the results, we were emboldened to try the application of this approach to the target hepta-oligomannoside 23 (Scheme 4). Gratifyingly, the preparation of hyper-branched hepta-saccharide occurred with the same efficiency and high degree of stereocontrol (cf., Schemes 2 and 3). Palladium catalyzed glycosylation of pyranone L-2b with the enantiomeric diol L-16, followed NaBH₄ reduction provided the allylic alcohol 20 in 65% yield (Scheme 4). Treatment of 20 with TBAF successfully removed the two TBS-groups and the resulting tetraol 21 then served as a glycosyl acceptor for per-glycosylation. Simply repeating the two-step protocol (glycosylation/reduction) on 21 gave the hepta-saccharide/ tetra-allylic alcohol 22 in 49% yield. To our delight, per-dihydroxylation of 22 using the Upjohn conditions (OsO₄/NMO) gave exclusively the α-manno-hepta-saccharide 23 in 86% yield, which could be purified by simple silica gel chromatography (ether/MeOH). Similarly, a global reduction with excess diimide resulted in the 2,3-dideoxy-D-erythrohexo-hepta-saccharide 24, which was easily isolated in 74% yield. While a similarly short route to ent-23 (e.g., the all D-hepta-saccharide) can be envisioned using a traditional glycosylation strategy (Scheme S1), the route would require the use of significantly more protecting groups and offer no practical approach to the D-/L-stereoisomers and deoxycongeners. It is important to note the acid sensitive of the glycosidic bonds in deoxy heptasaccharides 23 and 24, which might not survive the strongly acidic conditions of a traditional glycosylation.

In summary, a highly efficient, stereocontrolled and divergent route to various natural and unnatural oligosaccharide motifs has been developed via asymmetric catalysis. This *de novo* route featured the bidirectional use of a Pd(0)-catalyzed glycosylation/post-glycosylation transformations. The four possible D-/L-diastereomeric α-1,4-linked *rhamno*-trisaccharides and dideoxy-congeners were prepared in 10 steps starting from achiral acylfuran **1a**. Similarly, a highly branched hepta-saccharide was stereoselectively constructed from an achiral acylfuran **1b** in 12 steps. This approach is quite mild, amenable to the acid sensitive deoxysugars capable of being diversified into any of the D- and/or L-sugar diastereomers with complete stereocontrol. Finally, this new method enables the rapid assembly of unnatural oligosaccharide motifs (e.g., enantiomer, deoxysugars, D-/L-diastereomers) and in turn the possibility for numerous mode of action studies in glycobiology. Effort along these lines are ongoing and will be reported in due course.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We are grateful to NIH (GM090259) and NSF (CHE-0749451) for financial support of this research.

References

 For reviews of other approaches to hexoses, see: Gijsen HJM, Qiao L, Fitz W, Wong CH. Chem Rev. 1996; 96:443. [PubMed: 11848760] Hudlicky T, Entwistle DA, Pitzer KK, Thorpe AJ. Chem Rev. 1996; 96:1195. [PubMed: 11848785] Yu X, O'Doherty GA. ACS Symposium Series. Chemical Glycobiology. 2008; 990:3–28.

- (a) Ko SY, Lee AWM, Masamune S, Reed LA III, Sharpless KB, Walker FJ. Science. 1983;
 220:949. [PubMed: 17816019] (b) Ko SY, Lee AWM, Masamune S, Reed LA III, Sharpless KB, Walker FJ. Tetrahedron. 1990; 46:245.
- 3. (a) Northrup AB, MacMillan DWC. Science. 2004; 305:1752. [PubMed: 15308765] (b) Northrup AB, MacMillan DWC. J Am Chem Soc. 2002; 124:6798. [PubMed: 12059180] (c) Pizzarello S, Weber AL. Science. 2004; 303:1151. [PubMed: 14976304] (d) Casas J, Engqvist M, Ibrahem I, Kaynak B, Córdova A. Angew Chem Int Ed. 2005; 44:1343.(e) Ahmed MM, Berry BP, Hunter TJ, Tomcik DJ, O'Doherty GA. Org Lett. 2005; 7:745. [PubMed: 15704940] (f) Harris JM, Keranen MD, O'Doherty GA. J Org Chem. 1999; 64:2982. [PubMed: 11674384] (g) Shan M, Xing Y, O'Doherty GA. J Org Chem. 2009; 74:5961. [PubMed: 20560564] (h) Ahmed MM, O'Doherty GA. Carbohydr Res. 2006; 341:1505. [PubMed: 16616898]
- (a) Mangion IK, MacMillan DWC. J Am Chem Soc. 2005; 127:3696. [PubMed: 15771494] (b) Ahmed MM, O'Doherty GA. Tetrahedron Lett. 2005; 46:4151.(c) Shan M, O'Doherty GA. Org Lett. 2006; 8:5149. [PubMed: 17048865] (d) Gao D, O'Doherty GA. Org Lett. 2005; 7:1069. [PubMed: 15760141] (e) Gao D, O'Doherty GA. J Org Chem. 2005; 70:9932. [PubMed: 16292824] (f) Balachari D, O'Doherty GA. Org Lett. 2000; 2:4033. [PubMed: 11112636] (g) Guo H, O'Doherty GA. Org Lett. 2006; 8:1609. [PubMed: 16597122] (h) Abrams JN, Babu RS, Guo H, Le D, Le J, Osbourn JM, O'Doherty GA. J Org Chem. 2008; 73:1935. [PubMed: 18237188]
- 5. Bertozzi CR, Kiessling LL. Science. 2001; 291:2357. [PubMed: 11269316]
- 6. For oligosaccharide synthesis, see: Danishefsky SJ, McClure KF, Randolph JT, Ruggeri RB. Science. 1993; 260:1307. [PubMed: 8493573] Plante OJ, Palmacci ER, Seeberger PH. Science. 2001; 291:1523. [PubMed: 11222853] Sears P, Wong CH. Science. 2001; 291:2344. [PubMed: 11269314] Yamada H, Harada T, Miyazaki H, Takahashi T. Tetrahedron Lett. 1994; 35:3979.
- 7. (a) Wang LX. Curr Opin Drug Discov Devel. 2006; 9:194.(b) Ni J, Song H, Wang Y, Stamatos N, Wang LX. Bioconjugate Chem. 2006; 17:493.(c) Zeng Y, Wang J, Li B, Hauser S, Li H, Wang LX. Chem Eur J. 2006; 12:3355. [PubMed: 16470771] (d) Wang J, Le N, Heredia A, Song H, Redfield R, Wang LX. Org Biomol Chem. 2005; 3:1781. [PubMed: 15858664]
- 8. Trost BM. Science. 1991; 254:1471. [PubMed: 1962206]
- Wang CC, Lee JC, Luo SY, Kulkarni SS, Huang YW, Lee CC, Chang KL, Hung SC. Nature. 2007; 446:896. [PubMed: 17443183]
- 10. (a) Singh S, Ni J, Wang LX. Bioorg Med Chem Lett. 2003; 13:327. [PubMed: 12565922] (b) Calarese DA, Lee HK, Huang CY, Best MD, Astronomo RD, Stanfield RL, Katinger H, Burton DR, Wong CH, Wilson IA. Proc Natl Acad Sci USA. 2005; 102:13372. [PubMed: 16174734] (c) Dudkin VY, Orlova M, Geng X, Mandal M, Olson WC, Danishefsky SJ. J Am Chem Soc. 2004; 126:9560. [PubMed: 15291558]
- 11. (a) Wang HYL, Wu B, Zhang Q, Rojanasakul Y, O'Doherty GA. ACS Med Chem Lett. 2011; 2:259. [PubMed: 21572583] (b) Wang HYL, Rojanasakul Y, O'Doherty GA. ACS Med Chem Lett. 2011; 2:264. [PubMed: 21660118] (c) Iyer A, Zhou M, Azad N, Elbaz H, Wang L, Rogalsky DK, Rojanasakul Y, O'Doherty GA, Langenhan JM. ACS Med Chem Lett. 2010; 1:326. [PubMed: 21103068] (d) Wang HYL, Xin W, Zhou M, Stueckle TA, Rojanasakul Y, O'Doherty GA. ACS Med Chem Lett. 2011; 2:73. [PubMed: 21643465]
- 12. (a) Babu RS, O'Doherty GA. J Am Chem Soc. 2003; 125:12406. [PubMed: 14531673] (b) Babu RS, Zhou M, O'Doherty GA. J Am Chem Soc. 2004; 126:3428. [PubMed: 15025462] (c) Babu RS, O'Doherty GA. J Carb Chem. 2005; 24:169.(d) Guo H, O'Doherty GA. Angew Chem Int Ed. 2007; 46:5206.(e) Guo H, O'Doherty GA. J Org Chem. 2008; 73:5211. [PubMed: 18563936]

13. For examples of protecting group free synthesis, see: Crabtree RH. Science. 2007; 318:756. [PubMed: 17975055] Chen MS, White MC. Science. 2007; 318:783. [PubMed: 17975062]

- 14. Trost BM. Science. 1983; 219:245. [PubMed: 17798254]
- 15. Haukaas MH, O'Doherty GA. Org Lett. 2001; 3:401. [PubMed: 11428024]
- 16. Li M, Scott JG, O'Doherty GA. Tetrahedron Lett. 2004; 45:1005.Guo H, O'Doherty GA. Org Lett. 2005; 7:3921. [PubMed: 16119932] Guo H, O'Doherty GA. J Org Chem. 2008; 73:5211.
 [PubMed: 18563936] Or from vinylfurans: Bushey ML, Haukaas MH, O'Doherty GA. J Org Chem. 1999; 64:2984. [PubMed: 11674385] Harris JM, Keranen MD, Nguyen H, Young VG, O'Doherty GA. Carbohydr Res. 2000; 328:17. [PubMed: 11005573]
- 17. (a) Haukaas MH, O'Doherty GA. Org Lett. 2002; 4:1771. [PubMed: 12000295] (b) Houk KN, Li Y, McAllister MA, O'Doherty G, Paquette LA, Siebrand W, Smedarchina ZK. J Am Chem Soc. 1994; 116:10895.

$$O \longrightarrow R^1 \xrightarrow{a, b, c} O \longrightarrow C \longrightarrow R^2 O \longrightarrow R^1 \longrightarrow R^2 O \longrightarrow R^1$$

1a: R¹ = CH₃

L-2a: $R^1 = CH_3$

L-3a: $R^1 = CH_3$

L-4a: $R^1 = CH_3$

1b: $R^1 = CH_2OTBS$

L-2b: $R^1 = CH_2OTBS$

L-3b: $R^1 = CH_2OTBS$

L-4b: $R^1 = CH_2OTBS$

a. Noyori(S,S); b. NBS/ H_2O ; c. Boc_2O , DMAP; d. R^2OH , $Pd(0)/PPh_3$; e. $NaBH_4$; f. OsO_4 , NMO

Scheme 1.

De Novo Achmatowicz Approach to the Hexoses

Scheme 2. Synthesis of a Library of 1,4-Linked α -Rhamno-trisaccharides Containing both D- and L-Pyranoses

Scheme 3.
Synthesis of Branched α-*Rhamno/Manno*-trisaccharide Containing both D- and L-Pyranoses

Scheme 4. A 12-Step Synthesis of Highly Branched All-L-α-*Manno*-hepta-pyranoside