

Mechanisms for the Deamination Reaction of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$ and $2\text{H}_2\text{O}/\text{OH}^-$: A Computational Study

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Mechanisms for the deamination reaction of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ and $2\text{H}_2\text{O}/\text{OH}^-$ to produce uracil were investigated using ab initio calculations. Optimized geometries of reactants, transition states, intermediates, and products were determined at MP2 and B3LYP using the 6-31G(d) basis set and at B3LYP/6-31+G(d) levels of theory. Single point energies were also determined at MP2/G3MP2Large and G3MP2 levels of theory. Thermodynamic properties (ΔE , ΔH , and ΔG), activation energies, enthalpies, and free energies of activation were calculated for each reaction pathway investigated. Intrinsic reaction coordinate (IRC) analysis was performed to characterize the transition states on the potential energy surface. Seven pathways for the deamination reaction were found. All pathways produce an initial tetrahedral intermediate followed by several conformational changes. The final intermediate for all pathways dissociates to product via a 1–3 proton shift. The activation energy for the rate-determining step, the formation of the tetrahedral intermediate for pathway D, the only pathway that can lead to uracil, is $115.3 \text{ kJ mol}^{-1}$ at the G3MP2 level of theory, in excellent agreement with the experimental value ($117 \pm 4 \text{ kJ mol}^{-1}$).

1. INTRODUCTION

A detailed knowledge of the structure of cytosine (Cyt) and its nucleoside as well as the tautomerism of nucleic acid bases is an important prerequisite in understanding the molecular basis underlying their biological and medicinal functions. The genetic code can be affected by tautomers of cytosine which can be recognized as other nucleic acid bases (i.e., uracil or thymine); in particular, rare tautomers are thought to induce alterations in the normal base pairing leading to spontaneous mutations and errors in the genetic code, in the DNA or RNA helices.¹ Cytosine occurs naturally in all nucleic acids, both DNA and RNA. It is chemically bound to the sugar moiety and interacts with other nucleic acid–bases via hydrogen bonds, most frequently with guanine.² For a more detailed review, see our previous work³ and the references cited therein.

Mutations or changes to the nucleotide sequences of DNA can arise in a number of ways. Mistakes can be made during DNA replication that result in the incorporation of an incorrect base. Once altered, these changes may then be propagated by further DNA replication. Finally, large scale changes can sometimes occur in the form of DNA insertions and/or deletions. Spontaneous mutations can arise as a result of chemical changes to individual bases in DNA. One such chemical change is the conversion of cytosine to uracil (Ura) which is classified as a deamination reaction (the loss of an amino group from a tetrahedral carbon). The hydrolytic deamination reaction of cytosine to yield uracil is shown in Figure 1. Uracil is found in RNA and can base pair with adenine. It has firmly been established that deamination of the DNA base cytosine is an extremely rare event under

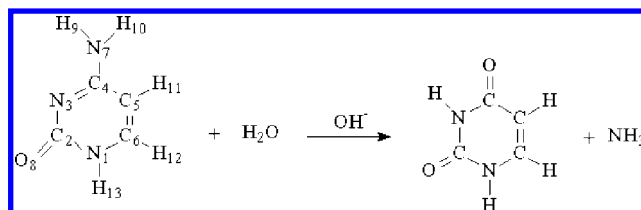


Figure 1. Deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ and the atom labelling in cytosine.

normal physiological conditions (40–100 deamination in human genome per day, pH 7.4) although the rate of deamination can be significantly increased in the presence of various reagents such as NO, HNO_2 and bisulfite.

If uracil is found in DNA, it poses a very serious problem. The cell, however, has a specific enzyme to remove it from DNA, called DNA uracil-*N*-glycosylase. The uracil formed by cytosine deamination is potentially mutagenic, changing the coding information during DNA replication and RNA transcription, resulting in altered base pairs in the genome.⁴

The deamination of cytosine, in particular, and a number of its derivatives have been the subject of many experimental studies.^{5–15} Federico et al.¹⁵ were able to determine the rate constant of cytosine deamination for single- and double-stranded DNA at physiological relevant conditions (37 °C and pH 7.4) by a sensitive genetic assay in which a mutant bacteriophage with a CCC codon is used with *E. coli* host cells defective in repair of uracil. Deamination to yield UCC or CUC lead to the generation of blue plaques, whereas, the CCC results in colorless plaques. Their measured rate constants for single- and double-stranded DNA are 1×10^{-10} and $7 \times 10^{-13} \text{ s}^{-1}$, respectively, with an activation energy of $117 \pm 4 \text{ kJ mol}^{-1}$. This value agrees well with the previous value of 121 kJ mol^{-1} found by Lindahl and

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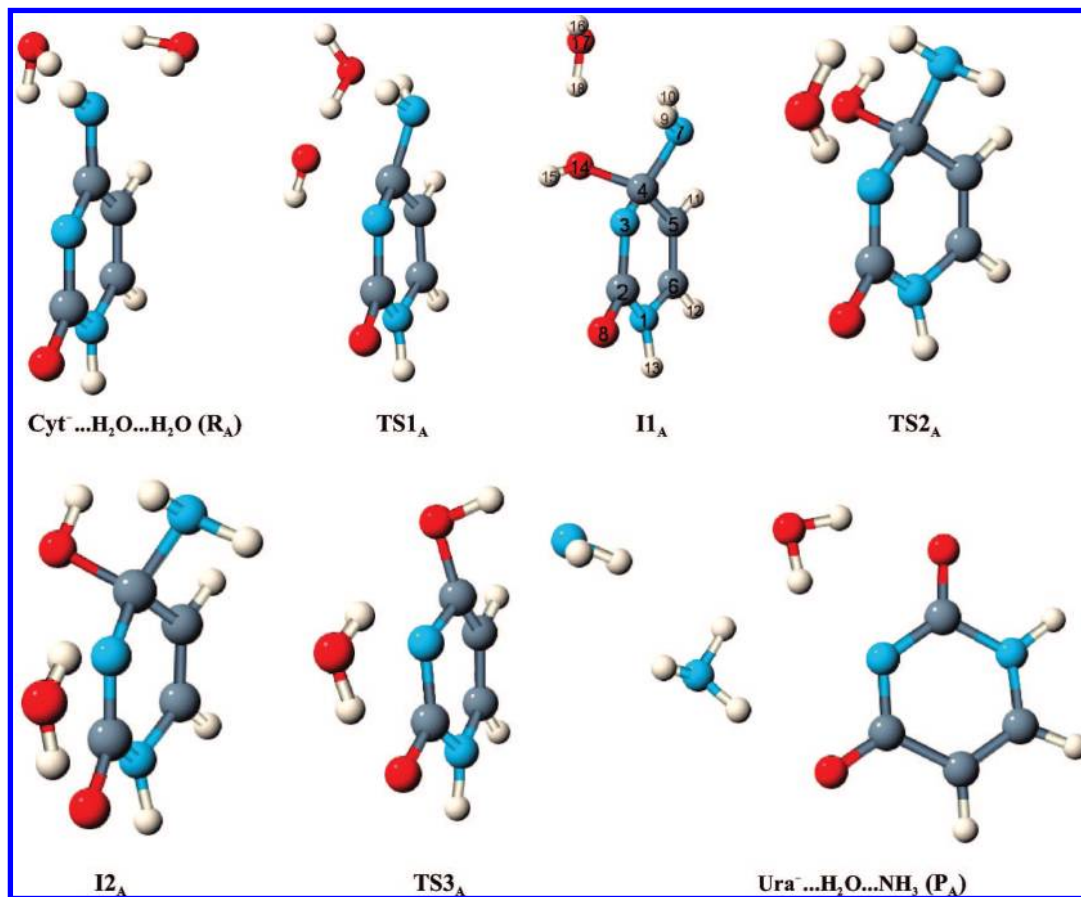


Figure 2. Optimized structures for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (pathway A).

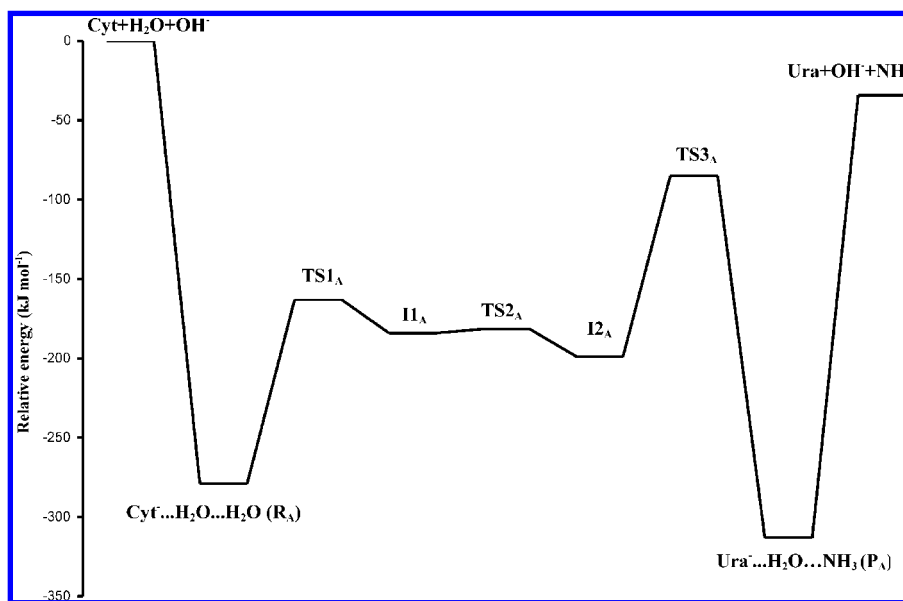


Figure 3. Reaction pathway A for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$. Relative energies are at the G3MP2 level of theory.

Nyberg¹⁶ obtained over a 25 °C temperature range. As a part of their investigation of the deamination of 1-methylcytosine and 1,5-dimethylcytosine with Pt^{II} complex, both experimentally and using DFT calculations, Sponer et al.¹⁷ also studied the deamination of cytosine with OH^- using the PCM model to account for solvation effects. However, their reported activation energy barrier ($213.4 \text{ kJ mol}^{-1}$ at B3LYP/6-31G(d)) is not very close to the experimentally accepted value^{15,16} ($117 \pm 4 \text{ kJ mol}^{-1}$), and the mechanism

reported in their paper differs in a number of ways from the one reported in our previous work.³ Rayat et al.¹⁸ studied the nitrosative deamination of cytosine in DNA experimentally. They provided a mechanistic hypothesis for nitrosative cytosine deamination which involves a pyrimidine ring-opened intermediate and proposed a number of possible reaction channels. More recently, Matsubara et al.¹⁹ studied the catalysis for cytidine deaminase. They also studied the uncatalysed hydrolytic deamination of cytosine with H_2O .

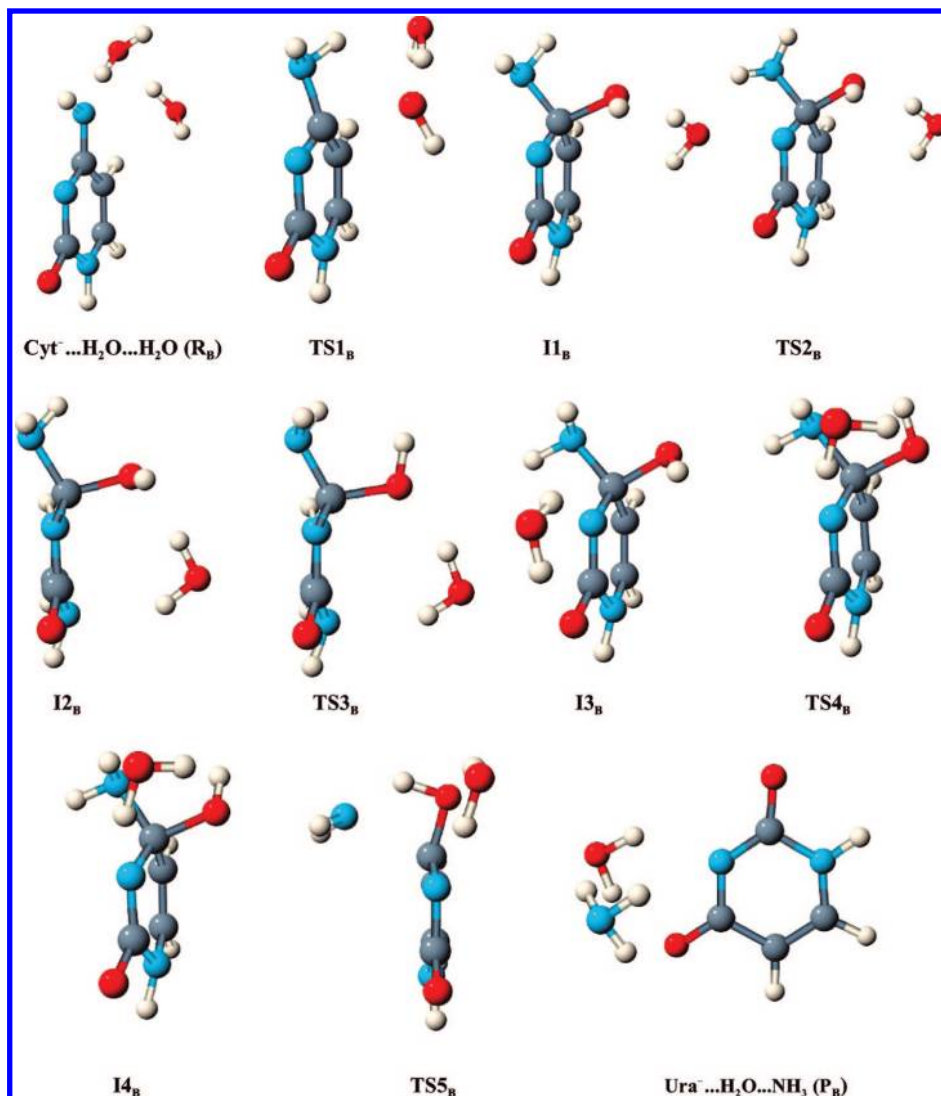


Figure 4. Optimized structures for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (pathway B).

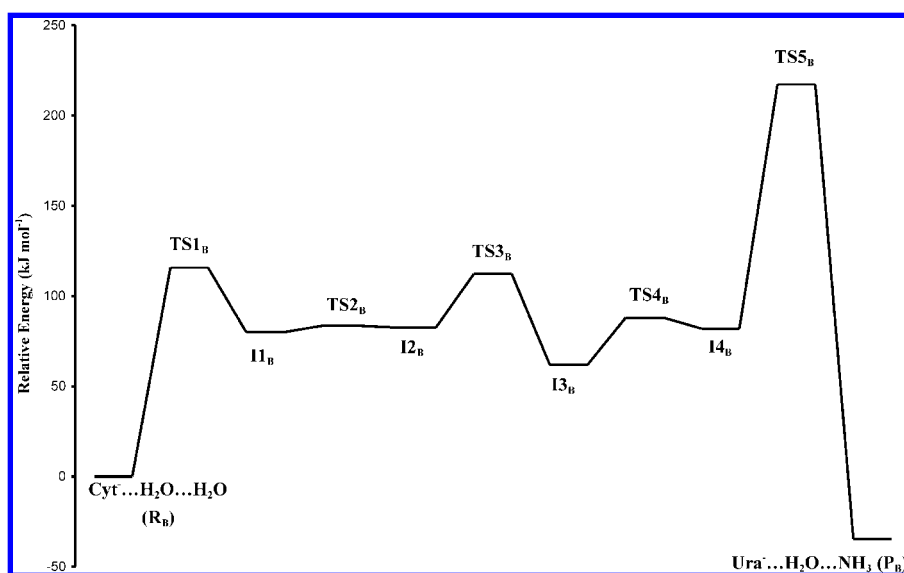


Figure 5. Reaction pathway B for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$. Relative energies are at the G3MP2 level of theory.

They found that the catalytic action of cytidine deaminase is effectively enhanced by the participation of the extra water molecule. Their reported activation energy barrier with one

water molecule is $237.4 \text{ kJ mol}^{-1}$ at B3LYP/6-31G(d,p)). We have shown in previous work on the deamination of cytosine with H_2O , that this reaction is not the most likely

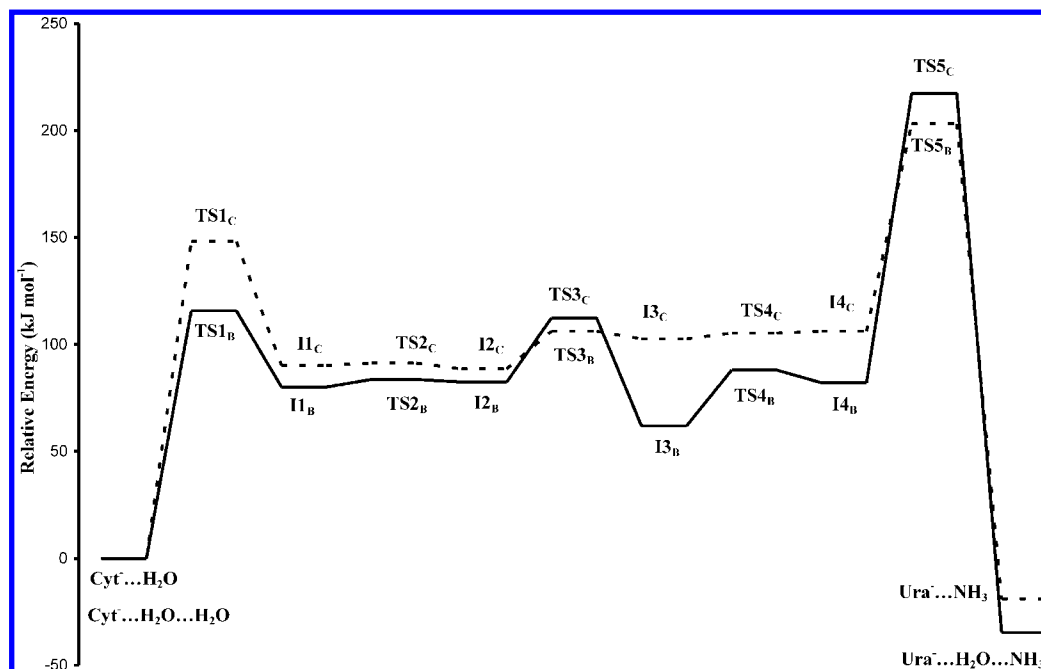


Figure 6. Comparison of reaction pathways for the deamination of cytosine with OH^- (dotted line)³ and reaction pathway B for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ at the G3MP2 level of theory.

Table 1. Activation Energies, Enthalpies of Activation, and Free Energies of Activation for the Deamination of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (in kJ mol^{-1}) at 298.15 K (Pathway A)

	MP2/ 6-31G(d)	B3LYP/ 6-31G(d)	B3LYP/ 6-31+G(d)	G3MP2
$\Delta E_{\text{act}}^{\ddagger}$, TS1 _A	137.1	130.7	118.4	115.3 (116.5) ^a
ΔH^{\ddagger} , TS1 _A	129.8	124.6	114.7	112.1
ΔG^{\ddagger} , TS1 _A	139.2	133.4	122.9	122.0
$\Delta E_{\text{act}}^{\ddagger}$, TS2 _A	8.2	7.5	4.6	2.5 (4.4)
ΔH^{\ddagger} , TS2 _A	5.6	4.4	2.4	0.9
ΔG^{\ddagger} , TS2 _A	5.1	4.6	7.7	2.5
$\Delta E_{\text{act}}^{\ddagger}$, TS3 _A	141.7	107.6	103.7	114.0 (125.8)
ΔH^{\ddagger} , TS3 _A	122.6	92.9	90.9	115.1
ΔG^{\ddagger} , TS3 _A	118.1	89.4	87.8	113.8

^a The values in bracket are for MP2/G3MP2Large.

mechanism to account for the deamination reaction of cytosine and hence not a model for similar systems, such as cytosine derivatives (Cytidine).³

Our previous results^{3,20} were useful starting points for the proposed mechanisms in this paper. The deamination of cytosine with H_2O was found³ to have a high activation energy ($221.3 \text{ kJ mol}^{-1}$ for pathway A and $260.3 \text{ kJ mol}^{-1}$ for pathway B). The activation energy for the deamination reaction with OH^- was very high, with an overall activation energy of 203 kJ mol^{-1} at the G3MP2 level of theory, compared to the experimental value ($117 \pm 4 \text{ kJ mol}^{-1}$). This reaction takes place at physiological condition, pH 7.4, which is slightly basic. Federico et al.¹⁵ suggested that the probability of deamination would increase by any process that would facilitate OH^- attack on the C₄ residue of cytosine. Since most proton transfers are mediated by water, one must consider the role of water molecules in the proton transfer. OH^- forms a strong hydrogen bond with H_2O and a stable complex. Computational studies have shown that interaction with water changes the relative energies of the cytosine tautomers, the canonical tautomer (the amino-oxo tautomer as shown in Figure 1) being better hydrated than

Table 2. Activation Energies, Enthalpies of Activation, and Free Energies of Activation for the Deamination of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (in kJ mol^{-1}) at 298.15 K (Pathway B)

	MP2/ 6-31G(d)	B3LYP/ 6-31G(d)	B3LYP/ 6-31+G(d)	G3MP2
$\Delta E_{\text{act}}^{\ddagger}$, TS1 _B	132.6	127.3	116.8	115.7 (120.2) ^a
ΔH^{\ddagger} , TS1 _B	129.3	122.7	114.0	113.3
ΔG^{\ddagger} , TS1 _B	144.4	135.8	123.3	122.4
$\Delta E_{\text{act}}^{\ddagger}$, TS2 _B	8.5	2.3	2.7	3.5 (4.2)
ΔH^{\ddagger} , TS2 _B	5.4	0.2	0.7	2.0
ΔG^{\ddagger} , TS2 _B	6.5	3.4	3.9	5.0
$\Delta E_{\text{act}}^{\ddagger}$, TS3 _B	45.4	41.4	35.1	29.9 (34.1)
ΔH^{\ddagger} , TS3 _B	41.2	36.8	30.4	29.0
ΔG^{\ddagger} , TS3 _B	42.1	37.5	30.8	29.9
$\Delta E_{\text{act}}^{\ddagger}$, TS4 _B	26.5	23.5	15.0	26.0 (26.3)
ΔH^{\ddagger} , TS4 _B	23.9	20.7	11.8	24.2
ΔG^{\ddagger} , TS4 _B	29.7	26.3	15.1	28.3
$\Delta E_{\text{act}}^{\ddagger}$, TS5 _B	144.7	112.2	100.7	115.2 (128.3)
ΔH^{\ddagger} , TS5 _B	124.6	95.9	88.1	113.5
ΔG^{\ddagger} , TS5 _B	115.3	88.5	84.7	70.5

^a The values in bracket are for MP2/G3MP2Large.

the other tautomers.²¹ The mechanism for the deamination of cytosine in DNA is unknown and the experimental activation energy was determined for cytosine deamination in single-stranded DNA. For these reasons, we have performed a detailed study of possible mechanisms for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ and $2\text{H}_2\text{O}/\text{OH}^-$ in which the water molecules act as a solvent (and do not directly participate in the proton transfer process), as well as when they mediate the hydrogen transfer step.

2. COMPUTATIONAL METHOD

All the computations were performed with the Gaussian03 suite of programs.²² The geometries of all reactants, transition states, intermediates, and products were fully optimized at MP2 and B3LYP levels of theory using the 6-31G(d) basis set and at B3LYP/6-31+G(d). Single point energies were determined at G3MP2 and the MP2/G3MP2Large levels of theory. We

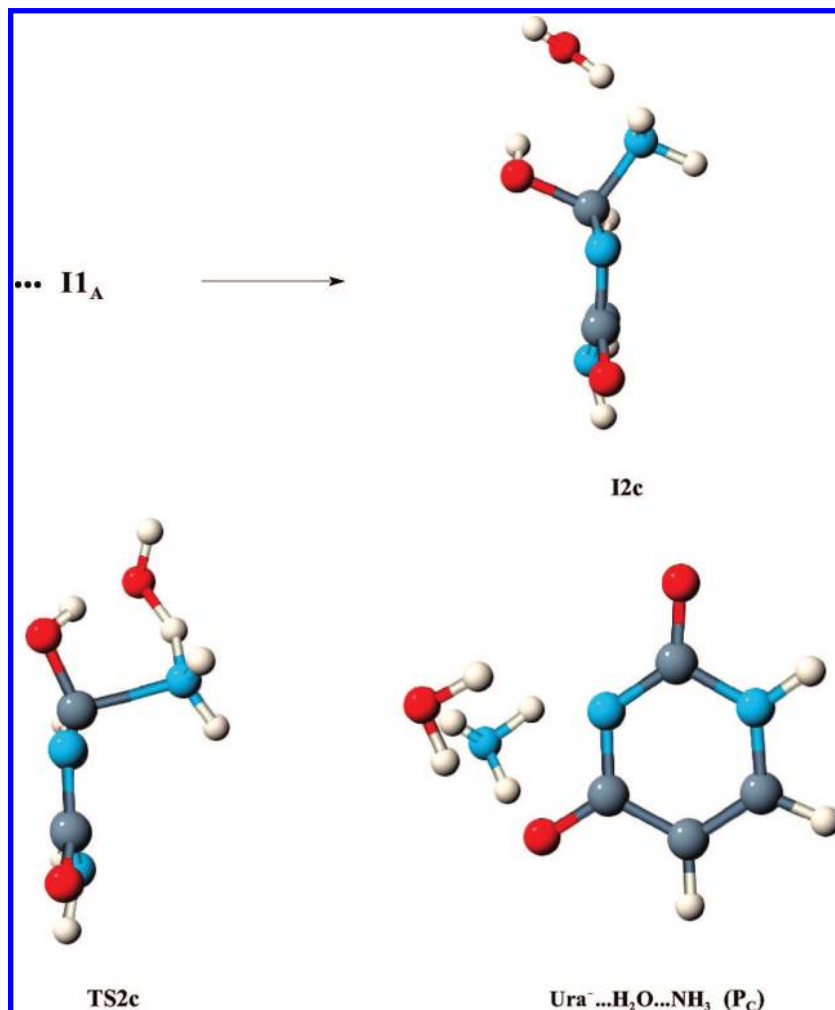


Figure 7. Optimized structures for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (pathway C).

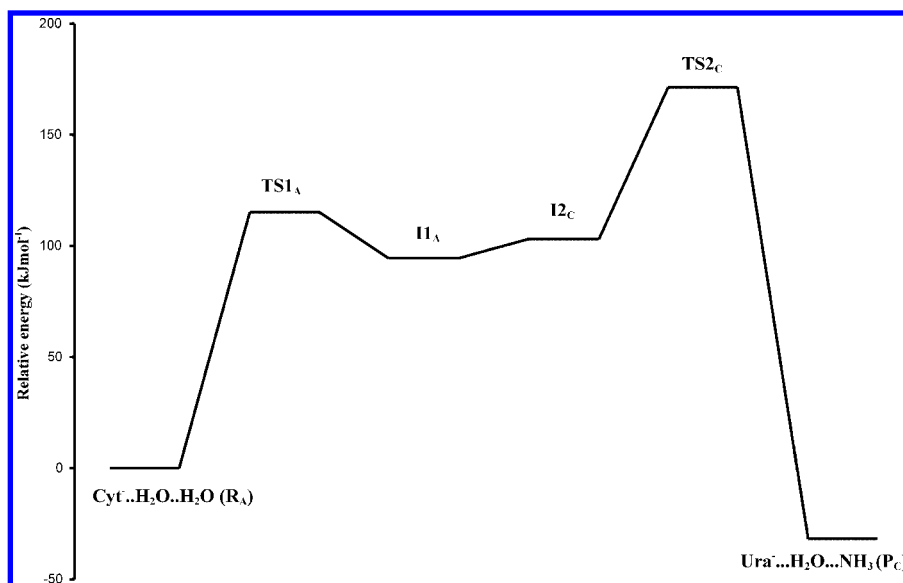


Figure 8. Reaction pathway C for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$. Relative energies are at the G3MP2 level of theory.

chose the G3MP2 level of theory which is known to give reliable energetics as confirmed from our previous results.²³ The complete reaction pathway for each mechanism discussed in this paper has been verified using intrinsic reaction coordinate (IRC) analysis of all transition states. The structures at the last IRC points in both directions were further optimized, in order

to positively identify the reactant and product to which each transition state is connected. A frequency analysis was performed for each stationary point in order to ensure that all minima had no imaginary frequencies on the potential-energy surface and that transition states have a single imaginary frequency.

Table 3. Activation Energies, Enthalpies of Activation, and Free Energies of Activation for the Deamination of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (in kJ mol^{-1}) at 298.15 K (Pathway C)

	MP2/ 6-31G(d)	B3LYP/ 6-31G(d)	B3LYP/ 6-31+G(d)	G3MP2
$\Delta E_{\text{act}}^{\ddagger}$, TS1 _A	137.1	130.7	118.4	115.3 (116.5) ^a
ΔH^{\ddagger} , TS1 _A	129.8	124.6	114.7	112.1
ΔG^{\ddagger} , TS1 _A	139.2	133.4	122.9	122.0
$\Delta E_{\text{act}}^{\ddagger}$, TS2 _C	73.0	52.2	50.2	68.3 (65.3)
ΔH^{\ddagger} , TS2 _C	59.7	36.6	41.4	65.2
ΔG^{\ddagger} , TS2 _C	66.3	42.1	51.7	68.3

^a The values in bracket are for MP2/G3MP2Large.

3. RESULTS AND DISCUSSION

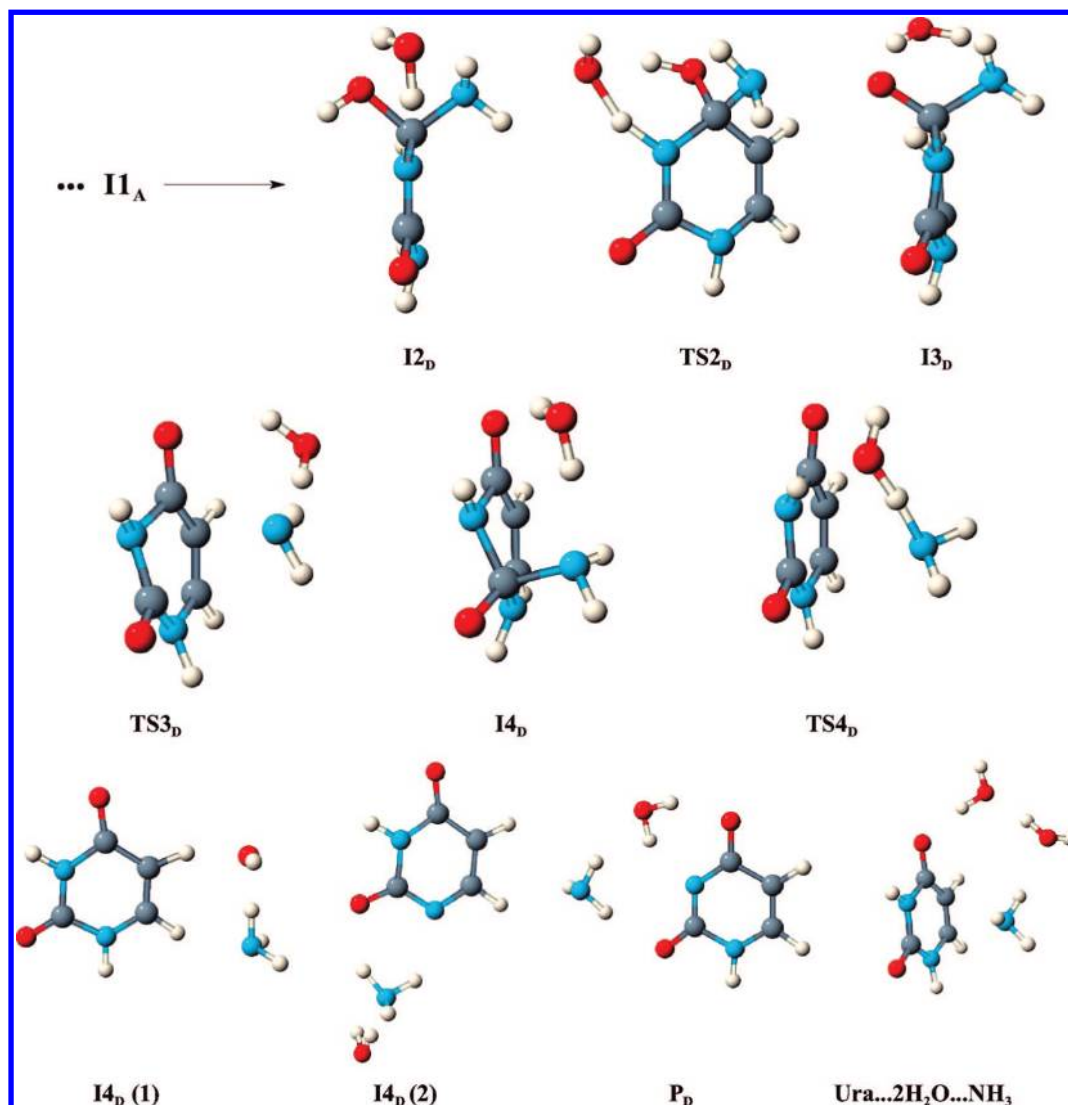
Cytosine has significant hydrogen bonding abilities; in particular, it possesses both hydrogen bond donor and acceptor groups. For this reason, we have examined a number of possible hydrogen-bonded complexes between H_2O , OH^- , and cytosine. We found that adding H_2O and OH^- to the $\text{N}_7\text{--C}_4\text{--C}_5\text{--C}_6$ side produced higher activation energies than adding them to the $\text{N}_7\text{--C}_4\text{--N}_3\text{--C}_2$ side of cytosine (see Figure 1). In addition, since it is the $\text{N}_7\text{--C}_4\text{--N}_3\text{--C}_2$ side of cytosine that hydrogen bonds with other nucleic acid-bases (normally guanine), it is more sterically hindered.²

Adding one water molecule stabilized the transition states and destabilized the reactant complex $\text{Cyt}^- \cdots \text{H}_2\text{O}$. Deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ and $2\text{H}_2\text{O}/\text{OH}^-$ can follow several possible pathways designated as pathways A \rightarrow D and F \rightarrow H.

3.1. Deamination of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$: Pathways A and B. In our previous work, we found two energetically equivalent pathways for the deamination of cytosine with OH^- which involve OH^- attack on both faces of cytosine.³ However, due to lack of symmetry, the extra water molecule now results in two different pathways (A and B) for attack on the two faces of cytosine.

Pathway A. The geometries for the reactants, intermediates, transition states, and products involved in pathway A (OH^- attack on the left face of cytosine with the carbonyl group facing the observer) are shown in Figure 2, and their relative energies are given in Figure 3.

For comparison, Figure 3 shows the relative stability of ($\text{Cyt} + \text{H}_2\text{O} + \text{OH}^-$) and the ($\text{Cyt}^- \cdots \text{H}_2\text{O} \cdots \text{H}_2\text{O}$) complex which is highly stabilized partly due to the delocalized negative charge on the cytosine anion compared to OH^- . Deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ ($\text{Cyt} + \text{OH}^- + \text{H}_2\text{O} \rightarrow \text{Ura} + \text{NH}_3 + \text{OH}^-$) closely follows the mechanism for the deamination of cytosine with H_2O and as well as the deamination with OH^- ,

**Figure 9.** Optimized structures for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (pathway D).

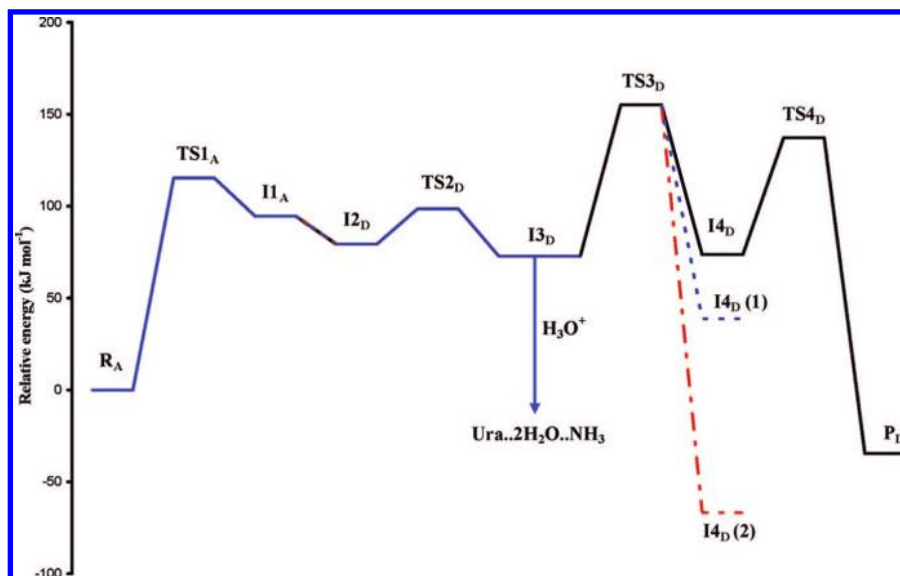


Figure 10. Reaction pathway D for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$. Relative energies are at the G3MP2 level of theory. The $\text{I4}_\text{D}(1)$ optimized structure is at B3LYP/6-31G(d), and $\text{I4}_\text{D}(2)$ is at B3LYP/6-31+G(d) and HF/6-31G(d) (see the Supporting Information).

Table 4. Activation Energies, Enthalpies of Activation, and Free Energies of Activation for the Deamination of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (in kJ mol^{-1}) at 298.15 K (Pathway D)

	MP2/ 6-31G(d)	B3LYP/ 6-31G(d)	B3LYP/ 6-31+G(d)	G3MP2
$\Delta E_\text{act}^\ddagger$, TS1 _A	137.1	130.7	118.4	115.3 (116.5)^a
ΔH^\ddagger , TS1 _A	129.8	124.6	114.7	112.1
ΔG^\ddagger , TS1 _A	139.2	133.4	122.9	122.0
$\Delta E_\text{act}^\ddagger$, TS2 _D	37.9	27.0	21.6	19.2 (25.3)
ΔH^\ddagger , TS2 _D	23.8	12.3	8.0	16.6
ΔG^\ddagger , TS2 _D	28.6	16.5	16.1	22.8
$\Delta E_\text{act}^\ddagger$, TS3 _D	120.9	95.2	69.4	82.4 (93.6)
ΔH^\ddagger , TS3 _D	108.3	76.7	58.0	83.9
ΔG^\ddagger , TS3 _D	107.3	68.4	59.4	81.5
$\Delta E_\text{act}^\ddagger$, TS4 _D	87.7	55.0	73.4 ^b	63.6 (78.7)
ΔH^\ddagger , TS4 _D	73.4	47.0	51.6	64.4
ΔG^\ddagger , TS4 _D	78.8	59.4	58.0	62.3

^a The values in bracket are for MP2/G3MP2Large. Bold numbers are indicating the rate-determining step for this pathway. ^b TS4_D did not converge to a first-order saddle point at B3LYP/6-31+G(d) but converged to the product; the value reported is for B3LYP/6-31+G(d)//HF/6-31G(d).

particularly, in relation to the two rate-determining steps. The first rate-determining step involves formation of a tetrahedral intermediate (I1_A), followed by a conformational change and a second rate-determining step which is a 1–3 proton shift from the hydroxy group to the exocyclic nitrogen atom.

Pathway A has two rate-determining steps. Deprotonation of cytosine occurs easily without forming a $\text{Cyt} \cdots \text{H}_2\text{O} \cdots \text{OH}^-$ complex. The H_{10} immediately transfers to OH^- forming a water molecule and a more stable ($\text{Cyt}^- \cdots 2\text{H}_2\text{O}$) complex which is the reactant for pathways A, B, C, and D. In the first rate-determining step, the addition of a water molecule stabilizes the hydroxide ion being formed in the transition state (TS1_A), dropping the activation energy. Nucleophilic attack by the water molecule on the C_4 carbon atom occurs with simultaneous proton transfer from H_2O to the exocyclic imine nitrogen of the cytosine anion to form a tetrahedral intermediate (I1_A). This is followed by a conformational change to give a second intermediate I2_A in which the water molecule has migrated to be in the same

plane as the cytosine ring, with an activation energy of only 2.5 kJ mol^{-1} at G3MP2. Conformers I1_A and I2_A only differ with respect to the position of the water molecule, the torsion of H_{15} ($\text{H}_{15}\text{O}_{14}\text{C}_4\text{N}_3$), and the torsion of the hydrogen atoms (H_9 and H_{10}) which are connected to N_7 . See the atom labeling for I1_A in Figure 2. The conformational change is followed by an intramolecular 1–3 proton shift from the hydroxyl group to the amino group to yield a $\text{Ura}^- \cdots \text{H}_2\text{O} \cdots \text{NH}_3$ complex. This mechanism is similar to the deamination of cytosine with OH^- .³ However, addition of a single water molecule stabilizes the transition states of both rate-determining steps lowering the overall activation energy by 8.9 kJ mol^{-1} .

Pathway B. The optimized structures involved in pathway B are shown in Figure 4, while the relative energies are given in Figure 5. Pathway B also has two rate-determining steps. As in pathway A, one of the H_2O attacks the C_4 carbon atom with simultaneous proton transfer to the imino group (sp^2 exocyclic nitrogen) of cytosine producing a tetrahedral intermediate, I1_B . This is followed by several low barrier conformational changes connecting intermediates I1_B and I4_B . In our previous work on the deamination of cytosine with H_2O and OH^- , we obtained two intermediate conformers with H_2O and three with OH^- . These conformers are different due to the existence of the two functional groups ($-\text{NH}_2$ and $-\text{OH}$) and H_2O in this system resulting in extra degrees of freedom. These structures are very similar, differing mainly in the torsion and the angles of these functional groups. The activation energies for the conformational changes are very small as expected and do not have a significant effect on the mechanism of this reaction. The final step is a 1–3 proton shift from the hydroxy group to the exocyclic nitrogen of the tautomer (I4_B) which results in the uracil anion deamination product; see Figure 4.

Figure 6 shows a comparison of the reaction pathways for the deamination of cytosine with OH^- and with $\text{H}_2\text{O}/\text{OH}^-$ for pathway B. It can be seen from Figure 6 that the addition of a water molecule results in a slightly higher overall barrier for deamination.

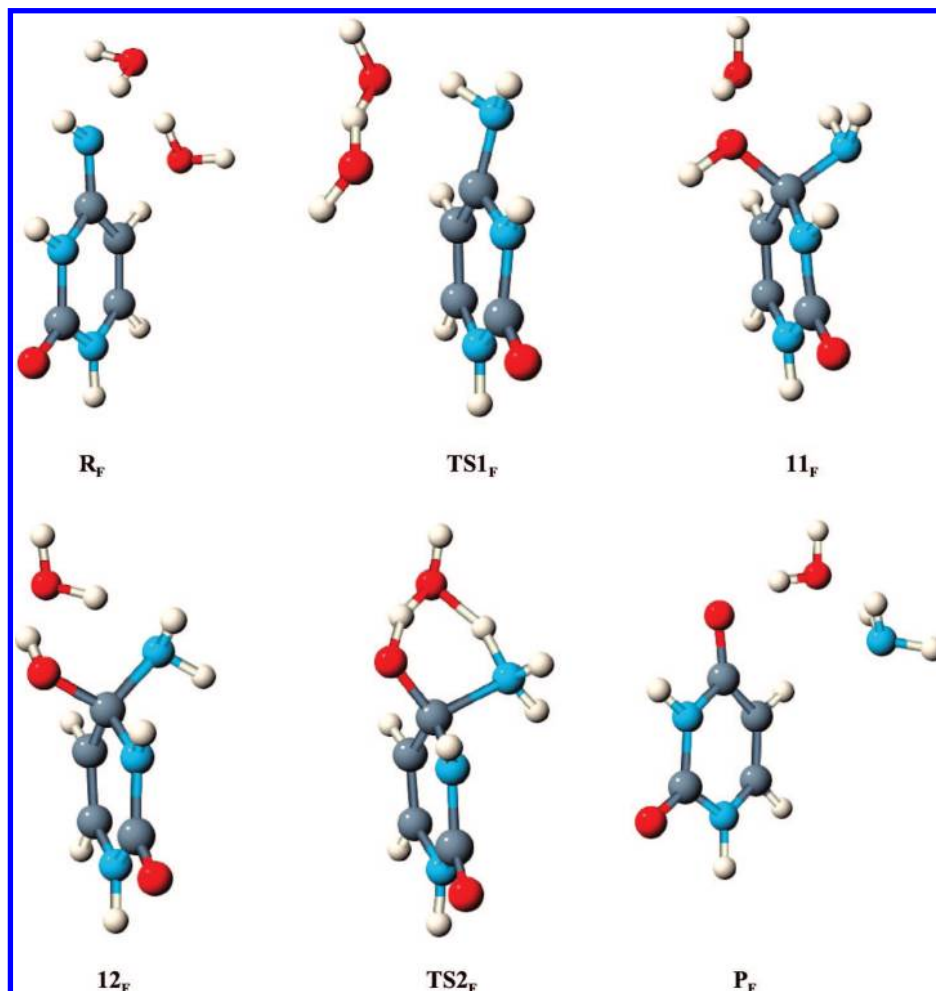


Figure 11. Optimized structures for the deamination of the amino-oxo tautomer of cytosine with $2\text{H}_2\text{O}$ (pathway F).

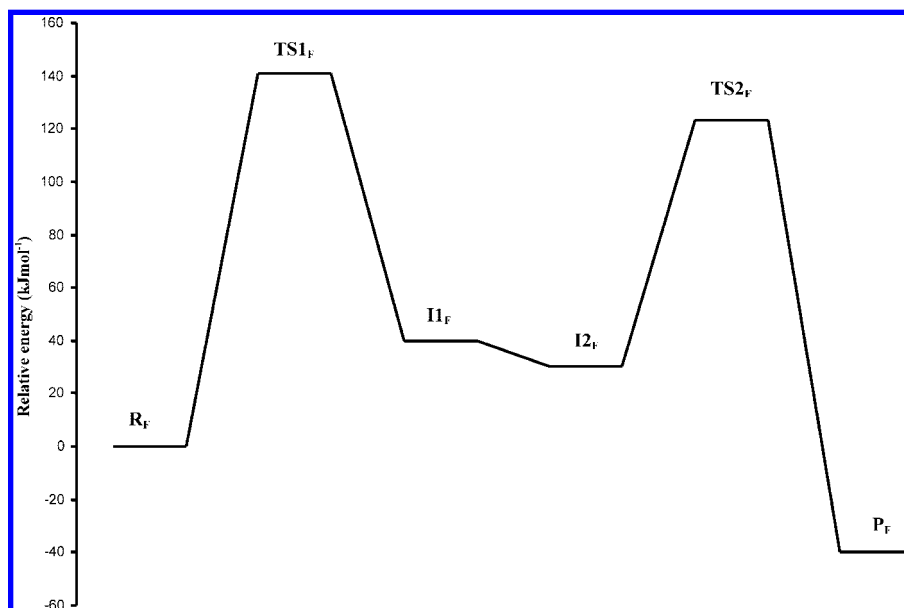


Figure 12. Reaction pathway F for the deamination of the amino-oxo tautomer of cytosine with $2\text{H}_2\text{O}$. Relative energies are at the G3MP2 level of theory.

The activation energies, enthalpies of activation, and free energies of activation for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ at the MP2 and B3LYP levels of theory using the 6-31G(d) basis set, B3LYP/6-31+G(d), and G3MP2 levels of theory for both pathways A and B are listed in Tables 1 and 2, respectively. Activation energies of $117 \pm$

4 and 121 kJ mol^{-1} were reported experimentally for this reaction.^{15,16} In our preliminary computational study,³ the activation energies for the two rate-determining steps are 148.0 and 97.0 kJ mol^{-1} resulting in an overall activation energy of $203.1 \text{ kJ mol}^{-1}$; see Figure 6. In this study, the activation energies for the two rate-determining steps are 115.3 and 114.0

Table 5. Activation Energies, Enthalpies of Activation, and Free Energies of Activation for the Deamination of the Amino-oxo Tautomer of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$ (in kJ mol^{-1}) at 298.15 K (Pathway F)

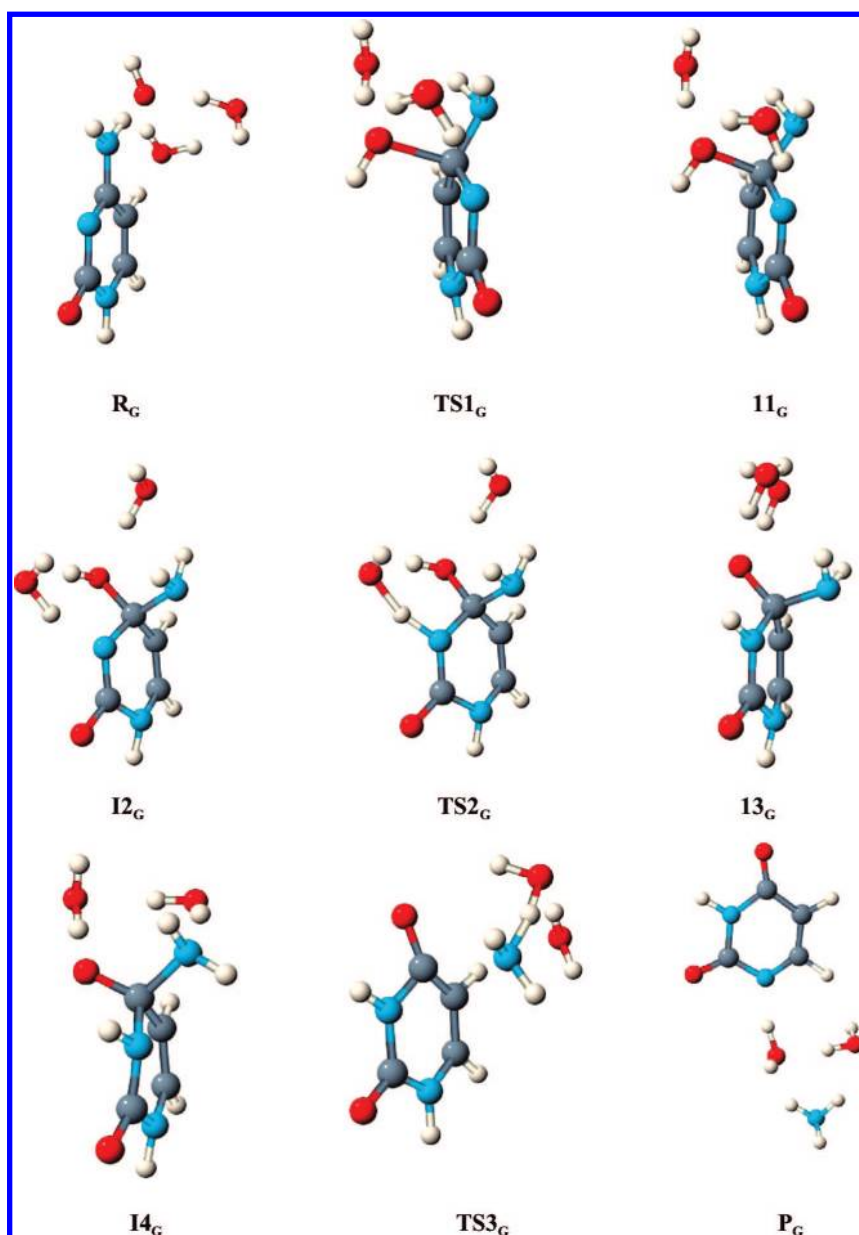
	MP2/ 6-31G(d)	B3LYP/ 6-31G(d)	B3LYP/ 6-31+G(d)	G3MP2
ΔE_{act} , TS1 _F	155.9	140.4	129.6	140.9 (141.8) ^a
ΔH^\ddagger , TS1 _F	140.9	125.0	116.3	135.1
ΔG^\ddagger , TS1 _F	159.4	140.1	129.5	153.8
ΔE_{act} , TS2 _F	93.8	80.6	81.6	92.6 (91.1)
ΔH^\ddagger , TS2 _F	74.4	61.4	63.4	88.2
ΔG^\ddagger , TS2 _F	80.8	68.0	70.7	99.2

^a The values in bracket are for MP2/G3MP2Large.

kJ mol^{-1} for pathway A and 115.7 and 115.2 kJ mol^{-1} for pathway B at the G3MP2 level of theory. Therefore, the overall activation energies for pathways A and B are 194.2 and 197.1 kJ mol^{-1} at G3MP2 and 192.7 and 193.7 kJ mol^{-1} at the B3LYP/6-31+G(d) level of theory, respectively. The MP2/G3MP2Large results for the barriers, are in excellent agreement

with the G3MP2 values, differing by no more than 13 kJ mol^{-1} as shown in Tables 1 and 2. Furthermore, Tables 1 and 2 show that the computationally less expensive B3LYP/6-31+G(d) results are in good agreement with the G3MP2 differing by no more than 18 kJ mol^{-1} in this study.

3.2. Deamination of Cytosine with $\text{H}_2\text{O}/\text{OH}^-$: Pathways C and D. The overall activation energies for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ for both pathways A and B are still high compared to the experimental value. In addition, the extra water molecule for pathways A and B did not participate in the 1–3 proton shift. It was previously reported that the formation of uracil (second step) from the tetrahedral intermediate is the rate-determining step.^{6,24} For this step, we examined the role of the water molecule as a mediator on the 1–3 proton shift from the $-\text{OH}$ group to the $-\text{NH}_2$ group which is one of the rate-determining steps of the catalytic process. It is well-known that water can act not only as a solvent but as a catalyst where it can donate or accept a proton to promote long-range proton transfer.^{20,23} From our previous study on the decomposition reaction of

**Figure 13.** Optimized structures for the deamination of cytosine with $2\text{H}_2\text{O}/\text{OH}^-$ (pathway G).

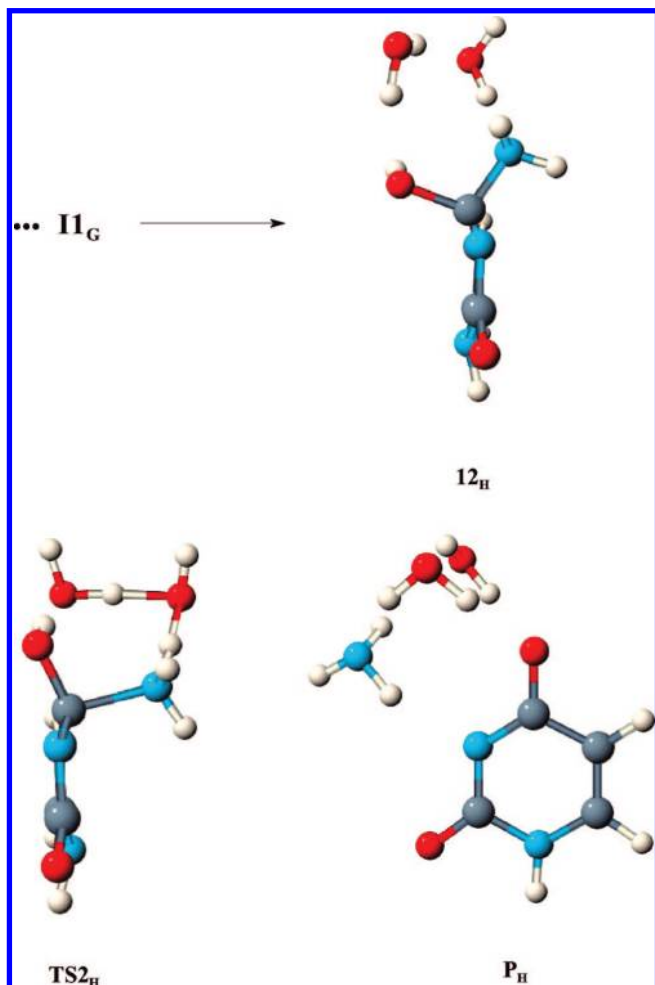


Figure 14. Optimized structures for the deamination of cytosine with $2\text{H}_2\text{O}/\text{OH}^-$ (pathway H).

formamidine,²³ we found that adding one water molecule catalyzed the 1–3 proton shift by forming a cyclic hydrogen-bonded transition state reducing the barrier by 88.4 kJ mol^{-1} at G3MP2. Addition of a second water molecule, further reduced the barrier by 17.2 kJ mol^{-1} at G3MP2.

Pathway C. The optimized structures and the relative energies involved in pathway C are shown in Figures 7 and 8, respectively. The activation energies, enthalpies of activation, and free energies of activation for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ at MP2 and B3LYP levels of theory using the 6-31G(d) basis set, B3LYP/6-31+G(d), and G3MP2 levels of theory for pathway C are listed in Table 3. The first transition state for pathway C is the same as pathway A.

Intermediates I1_A and I2_C are connected with several conformational changes, which do not have a significant effect on the mechanism of this reaction and hence are not included in the figures. The barrier for the water-mediated 1–3 proton shift is reduced by 45.7 kJ mol^{-1} at G3MP2 and 53.5 kJ mol^{-1} at B3LYP/6-31+G(d) levels of theory. The activation energy for this step is 68.3 kJ mol^{-1} at G3MP2 and 50.2 kJ mol^{-1} at B3LYP/6-31+G(d). However, the overall activation energy is still somewhat high, $171.1 \text{ kJ mol}^{-1}$ ($160.6 \text{ kJ mol}^{-1}$ at B3LYP/6-31+G(d)); see Table 3.

Pathway D. The optimized structures involved in pathway D are shown in Figure 9, and their relative energies given in Figure 10. The activation energies, enthalpies of activation,

and free energies of activation for the deamination of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ at MP2 and B3LYP levels of theory using the 6-31G(d) basis set, B3LYP/6-31+G(d), and G3MP2 levels of theory for pathway D are listed in Table 4.

Of all the pathways investigated, pathway D (Figure 9) is the only pathway that can lead to neutral uracil for this system. The first step for pathway D is the same as pathway A. In this case, intermediate I1_A is converted to I2_D by conformational changes. Intermediate I2_D undergoes a water-mediated 1–3 proton shift, from the $-\text{OH}$ to the N_3 atom, to form intermediate I3_D through TS2_D . The activation energy for TS2_D is 19.2 kJ mol^{-1} at G3MP2; see Table 4. At the MP2/6-31G(d) level of theory, intermediate I3_D is converted to intermediate I4_D through transition state TS3_D , where the $-\text{NH}_2$ group is migrating from C_4 to C_2 (the carbonyl carbon), with the negative charge localized at O_8 . In this case the IRC analysis resulted in two other possible intermediates, $\text{I4}_\text{D}(1)$ at B3LYP/6-31G(d) and $\text{I4}_\text{D}(2)$ at HF/6-31G(d) (see the Supporting Information) and B3LYP/6-31+G(d) levels of theory. From intermediate I4_D , a proton transfers from the oxygen of H_2O to the $-\text{NH}_2$ group with simultaneous transfer of a proton from the N_3 atom of uracil, through TS4_D , to form a $\text{Ura}^- \cdots \text{H}_2\text{O} \cdots \text{NH}_3$ complex (P_D) with the negative charge at N_3 . For pathway D (R to P_D), the overall activation energy is 155.2 and $135.0 \text{ kJ mol}^{-1}$ at G3MP2 and B3LYP/6-31+G(d) levels of theory, respectively.

However, in reality (in DNA or in solution), intermediate I3_D , as shown in Figure 9, would most likely be protonated at the $-\text{NH}_2$ group to form ammonia resulting in a complex of $\text{Ura} \cdots 2\text{H}_2\text{O} \cdots \text{NH}_3$. In this case, the first step is the rate-determining step with an activation energy of 115.3 and $118.4 \text{ kJ mol}^{-1}$ at G3MP2 and B3LYP/6-31+G(d) levels of theory, respectively, which accounts for the observed experimental value.

3.3. Deamination of a Tautomer of Cytosine with $2\text{H}_2\text{O}$: Pathway F. We also considered the possibility of the $\text{Cyt}^- \cdots \text{H}_2\text{O} \cdots \text{H}_2\text{O}$ complex being protonated. Computational studies also have predicted that cytosine protonation in aqueous solution or in the gas phase should occur at the N_3 position^{25–29} (see Figure 1). Protonation occurred at the N_3 site, known to be more favorable site for protonation of cytosine,²⁹ resulting in a complex of the amino-oxo tautomer of cytosine with two water molecules. This resulted in a new pathway designated as pathway F. The geometries for the reactant, intermediates, transition state, and product are shown in Figure 11, while the relative energies are shown in Figure 12. Activation energies, enthalpies of activation, and free energies of activation for pathway F at MP2 and B3LYP levels of theory using the 6-31G(d) basis set, B3LYP/6-31+G(d), and G3MP2 levels of theory are listed in Table 5.

Pathway F is a mechanism with a single rate-determining step and several conformational changes connecting intermediates I1_F and I2_F . The first and last steps are similar to the previous pathways. Interestingly, the barriers for TS1_F and TS2_F increase for pathway F compared to TS1 and TS2_C for pathway C from 115.3 to $140.9 \text{ kJ mol}^{-1}$ and from 68.3 to 92.6 kJ mol^{-1} at G3MP2 level of theory, respectively. The first step is the rate-determining step with a barrier of 140.9 and $129.6 \text{ kJ mol}^{-1}$ at G3MP2 and B3LYP/6-31+G(d) levels of theory, respectively, which is high compared to the experimental value.

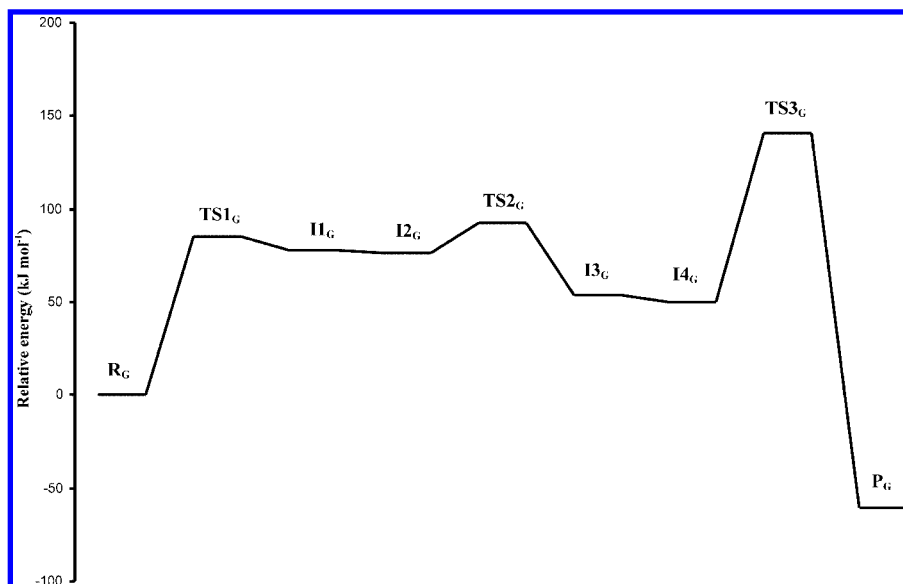


Figure 15. Reaction pathway G for the deamination of cytosine with $2\text{H}_2\text{O}/\text{OH}^-$. Relative energies are at the G3MP2 level of theory.

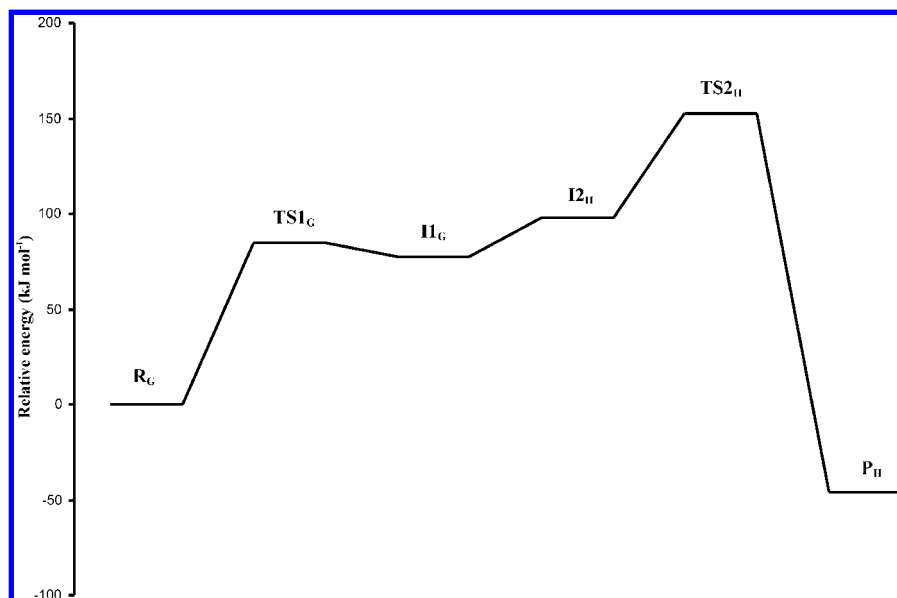


Figure 16. Reaction pathway H for the deamination of cytosine with $2\text{H}_2\text{O}/\text{OH}^-$. Relative energies are at the G3MP2 level of theory.

Table 6. Activation Energies, Enthalpies of Activation, and Free Energies of Activation for the Deamination of Cytosine with $2\text{H}_2\text{O}/\text{OH}^-$ (in kJ mol^{-1}) at 298.15 K (Pathway G)

	MP2/ 6-31G(d)	B3LYP/ 6-31G(d)	B3LYP/ 6-31+G(d)	G3MP2
$\Delta E_{\text{act}}, \text{TS1}_G$	76.1	96.6	110.9	84.8 (86.5) ^a
$\Delta H^\ddagger, \text{TS1}_G$	79.3	99.8	112.3	82.0
$\Delta G^\ddagger, \text{TS1}_G$	91.1	112.6	123.9	93.5
$\Delta E_{\text{act}}, \text{TS2}_G$	22.8	16.4	16.7	16.2 (18.9)
$\Delta H^\ddagger, \text{TS2}_G$	9.9	4.2	4.7	12.4
$\Delta G^\ddagger, \text{TS2}_G$	17.4	10.6	12.5	23.4
$\Delta E_{\text{act}}, \text{TS3}_G$	106.1	83.0	75.5	90.3 (94.5)
$\Delta H^\ddagger, \text{TS3}_G$	86.7	63.8	61.4	90.2
$\Delta G^\ddagger, \text{TS3}_G$	83.6	58.1	54.8	89.5

^a The values in bracket are for MP2/G3MP2Large.

Table 7. Activation Energies, Enthalpies of Activation, and Free Energies of Activation for the Deamination of Cytosine with $2\text{H}_2\text{O}/\text{OH}^-$ (in kJ mol^{-1}) at 298.15 K (Pathway H)

	MP2/ 6-31G(d)	B3LYP/ 6-31G(d)	B3LYP/ 6-31+G(d)	G3MP2
$\Delta E_{\text{act}}, \text{TS1}_G$	76.1	96.6	110.9	84.8 (86.5) ^a
$\Delta H^\ddagger, \text{TS1}_G$	79.3	99.8	112.3	82.0
$\Delta G^\ddagger, \text{TS1}_G$	91.1	112.6	123.9	93.5
$\Delta E_{\text{act}}, \text{TS2}_H$	57.1	35.2	57.6	55.3 (50.6)
$\Delta H^\ddagger, \text{TS2}_H$	41.3	21.4	46.9	51.5
$\Delta G^\ddagger, \text{TS2}_H$	53.8	30.5	55.9	63.6

^a The values in bracket are for MP2/G3MP2Large.

3.4. Deamination of Cytosine with $2\text{H}_2\text{O}/\text{OH}^-$: Pathways G and H. Since the presence of a single water molecule had a significant effect on the various reaction pathways, we have investigated the effect of a second water molecule on the potential energy surface for pathways C and D. The

geometries of the reactants, intermediates, transition states, and products for pathways G and H are shown in Figures 13 and 14, respectively. The relative energies for pathways G and H are shown in Figures 15 and 16, respectively. The activation energies, enthalpies of activation, and free energies of activation for pathways G and H are listed in Tables 6 and 7, respectively.

The key steps for pathways G and H are similar to the previous pathways C and D. Addition of the second water molecule further reduces the barriers for both rate-determining steps. For pathway G, the product (P_G) is a complex of the uracil anion, two water molecules, and ammonia ($\text{Ura}^- \cdots 2\text{H}_2\text{O} \cdots \text{NH}_3$) at the MP2/6-31G(d), B3LYP/6-31G(d), and B3LYP/6-31+G(d) levels of theory with a negative charge at the N_1 atom, which is not possible in DNA and RNA (cytosine is chemically bound to the sugar moiety at N_1 in DNA).

For pathways G and H, we note that adding the second water molecule reduces the activation energy of the first-step (which is the same for both pathways) by 30.5 kJ mol^{-1} at G3MP2 and by a lesser amount for the other steps. This is due to the fact that the second water molecule can both stabilize the transition state and act as a catalyst for this system. The activation energies for the two rate-determining steps are 84.8 and 90.3 kJ mol^{-1} for pathway G and 84.8 and 55.3 kJ mol^{-1} for pathway H at G3MP2 level of theory. However, the overall activation energies for pathways G and H are 140.5 and $152.9 \text{ kJ mol}^{-1}$ at G3MP2 (134.6 and $155.6 \text{ kJ mol}^{-1}$ at B3LYP/6-31+G(d)), respectively, see Tables 6 and 7, which are still high compared to the experimental value. For pathway G intermediate I_{3G} , similarly to intermediate I_{3D} , would most likely be protonated at the $-\text{NH}_2$ group. The first step would then be the rate-determining step with an activation energy of only 93 kJ mol^{-1} at G3MP2. This lower barrier may account for the difference between the deamination of free cytosine versus cytosine in single-stranded DNA.

4. CONCLUSIONS

The mechanism for the deamination reaction of cytosine with $\text{H}_2\text{O}/\text{OH}^-$ and $2\text{H}_2\text{O}/\text{OH}^-$ to produce uracil was investigated using ab initio calculations. Seven pathways for the deamination reaction were found. This paper shows the first detailed study of possible mechanisms for the deamination reaction of cytosine with OH^- including the effect of the presence of one and two water molecules. In all the mechanisms, a series of conformational changes connect the transition states of the two rate-determining steps. Pathway D is the first plausible mechanism reported for the deamination of cytosine, where the calculated activation energy ($115.3 \text{ kJ mol}^{-1}$ at G3MP2 level of theory) agrees very well with the experimentally determined activation energy ($117 \pm 4 \text{ kJ mol}^{-1}$).

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Supporting Information Available: Full geometries and energies of all structures for all pathways investigated at all levels of theory used. This information is available free of charge via the Internet at <http://pubs.acs.org>.

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