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Dioxygen Reactivity of Mononuclear Heme and Copper Components Yielding A High-Spin Heme-Peroxo-Cu Complex

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> > Received March 6, 2001

In this report, we describe the formation of a pentacoordinate high-spin heme-peroxo-Cu complex, [(F₈TPP)Fe^{III}-(O₂²⁻)- $Cu^{II}(TMPA)]^+$ (2), formed upon addition of O_2 to a 1:1 mixture of the reduced heme complex (F₈TPP)Fe^{II} (1a) and copper complex [(TMPA)Cu^I(CH₃CN)]⁺ (**1b**) {Scheme 1}. Remarkably, mixing of dioxygen with Fe and Cu mononuclear components²⁻⁵ leads to the heterobinuclear complex 2, in preference to homobinuclear μ -peroxo or μ -oxo heme-only^{3,6} or copper-only products. 4,5,7 The present system provides significant new electronic, structural, and mechanistic advances into the dioxygen chemistry of heme-Cu systems. Insights obtained may have relevance to heme-copper oxidase O₂-binding and reduction,⁸⁻¹¹ other chemistries or biochemistries involving dioxygen-binding to metals, 8,12,13 O₂-activation (e.g., cytochrome P-450 monooxygenase, 14 copper complex O₂-reactivity, 5,15,16 and Cu monooxygenases^{15,17,18}), and O-O reductive cleavage.

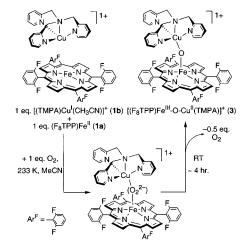
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- (1) Abbreviations used: $F_8TPP = tetrakis(2,6-difluorophenyl)$ porphyrinate; TMPA = tris(2-pyridylmethyl)amine; P = porphyrinate; MALDI-TOF-MS Matrix Assisted Laser Desorption Ionization Time Of Flight Mass
- Spectrometry. (2) The dioxygen chemistry of $(F_8TPP)Fe^{II}$ and $[(TMPA)Cu^I(MeCN)]^+$ have

- been previously studied. See refs 3-5.
 (3) Ghiladi, R. A.; Kretzer, R. M.; Guzei, I.; Rheingold, A. L.; Neuhold, Y.-M.; Hatwell, K. R.; Zuberbühler, A. D.; Karlin, K. D. Submitted for publication.
- (4) Tyeklár, Z.; Jacobson, R. R.; Wei, N.; Murthy, N. N.; Zubieta, J.; Karlin, K. D. J. Am. Chem. Soc. 1993, 115, 2677-2689.

- (5) Karlin, K. D.; Kaderli, S.; Zuberbühler, A. D. Acc. Chem. Res. 1997, 30, 139-147
- (6) Karlin, K. D.; Nanthakumar, A.; Fox, S.; Murthy, N. N.; Ravi, N.; Huynh, B. H.; Orosz, R. D.; Day, E. P. *J. Am. Chem. Soc.* **1994**, *116*, 4753–
- 4763. (7) $(F_8TPP)Fe^{III} (O_2^{2^-}) Fe^{III}(F_8TPP)$ ($\delta_{pyrrole} = 17.5 \text{ ppm}$, $\lambda_{max} = 414$ (Soret), 536 nm; CH_2CI_2 , 193 K), $^3(F_8TPP)Fe^{III} O Fe^{III}(F_8TPP)$ ($\delta_{pyrrole} = 13.1 \text{ ppm}$, $\lambda_{max} = 400$ (Soret), 561 nm; CH_2CI_2 , 193 K), 6 and $[(TMPA)Cu^{II} (O_2^{2^-}) Cu^{II}(TMPA)]^{2^+}$ ($\lambda_{max} = 525 \text{ nm})^4$ products are not observed by UV-visible or NMR spectroscopies.
 - (8) Momenteau, M.; Reed, C. A. Chem. Rev. **1994**, 94, 659–698. (9) Gennis, R. B. Science **1998**, 280, 1712–1713.

 - (10) Michel, H. Proc. Natl. Acad. Sci. U.S.A. 1998, 95, 12819-12824.
- (11) Babcock, G. T. Proc. Natl. Acad. Sci. U.S.A. 1999, 96, 12971–12973.
 (12) Ho, R. Y. N.; Liebman, J. F.; Valentine, J. S. In Active Oxygen in Biochemistry; Valentine, J. S., Foote, C. S., Greenberg, A., Liebman, J. F., Eds.; Blackie Academic and Professional, Chapman & Hall: Glasgow, 1995;
- (13) Valentine, J. S., Foote, C. S., Greenberg, A., Liebman, J. F., Eds. *Active Oxygen in Biochemistry*; Chapman & Hall: New York, 1995. (14) Sono, M.; Roach, M. P.; Coulter, E. D.; Dawson, J. H. *Chem. Rev.*
- 1996, 96, 2841-2887.

Scheme 1



 $[(F_8TPP)Fe^{III}-(O_2^{2-})-Cu^{II}(TMPA)]^+$ (2)

Upon addition of dioxygen at -40 °C in MeCN solvent to an equimolar solution of (F₈TPP)Fe^{II} (1a)^{3,19} and [(TMPA)Cu^I(CH₃-CN)](ClO₄) (**1b**)⁴ {reduced spectrum: $\lambda_{\text{max}} = 414$ (sh), 421 (Soret), 526 nm}, UV-visible spectroscopy (Figure 1) reveals the formation of a new species $\{\lambda_{max} = 412 \text{ (Soret)}, 558 \text{ nm}\}$

We formulate this O_2 -adduct as the peroxo complex [(F_8 TPP)- $Fe^{III} - (O_2^{2-}) - Cu^{II}(TMPA)]^+$ (2) based upon the following: (1) The resonance Raman spectrum of 2 (Figure 2A) presents a peroxo O-O stretching vibration at 808 cm⁻¹ that downshifts by 46 cm⁻¹ with ¹⁸O-labeled dioxygen (Figure 2B). In the scrambled isotope experiment, the ¹⁶O-¹⁸O stretch corresponds to a single component at 785 cm⁻¹ (Figure 2C), and indicates that the peroxide species is bound in a symmetric fashion. (2) MALDI-TOF-MS of 2 (formed in MeCN solvent) gives a parent peak at m/z 1239 {(M - ClO₄⁻ + MeCN)⁺} when $^{16}O_2$ is employed.²⁰ The expected increase in mass of 4 is observed when 2 forms from $^{18}O_2$, m/z 1243. (3) Dioxygen-uptake measurements {spectrophotometric titration: MeCN, -40 °C} revealed an oxygenation stoichiometry of $1a:1b:O_2 = 1:1:1$.

Further characterization of [(F₈TPP)Fe^{III}-(O₂²⁻)-Cu^{II}(TMPA)]⁺ (2) comes from NMR spectroscopy.²¹ In MeCN at −40 °C, the $(F_8TPP)Fe^{II}/[(TMPA)Cu^I(CH_3CN)]^+$ system (1a:1b = 1:1) has a single pyrrole resonance at δ 10 ppm (Figure 3A), consistent with a low spin (S = 0) system {Evans NMR method, $\mu_B = 0$ }. Oxygenation of the 1a/1b mixture leads to a downfield shifting of the pyrrole resonances²² for **2** (Figure 3B: $\delta_{\text{pyrrole}} = 68 \text{ ppm}$, s, br), with upfield shifted pyridyl peak resonances also observed at -11 and -20 ppm. An overall S = 2 spin state for **2** is assigned (Evans method, $\mu_{\rm B} = 5.1$, -40 °C), arising from the antiferromagnetic coupling of the S = 5/2 high spin ferric heme to the S

⁽¹⁵⁾ Karlin, K. D.; Zuberbühler, A. D. In Bioinorganic Catalysis: Second Edition, Revised and Expanded; Reedijk, J., Bouwman, E., Ed.; Marcel Dekker: New York, 1999; pp 469–534.

(16) Blackman, A. G.; Tolman, W. B. Struct. Bond. 2000, 97, 179–211.

(17) Solomon, E. I.; Sundaram, U. M.; Machonkin, T. E. Chem. Rev. 1996,

^{96, 2563-2605.}

⁽¹⁸⁾ Klinman, J. P. Chem. Rev. 1996, 96, 2541-2561.

⁽¹⁹⁾ Kopf, M.-A.; Neuhold, Y.-M.; Zuberbühler, A. D.; Karlin, K. D. Inorg. Chèm. 1999, 38, 3093-3102.

⁽²⁰⁾ Coordination of the MeCN to the iron is inconsistent with the pentacoordinate high-spin configuration deduced from the resonance Raman signature of the F₈TPP ligand in 2. Coordination to the copper center is under investigation.

⁽²¹⁾ All pyrrole resonances have been confirmed by ²H NMR on complexes made with pyrrole-deuterated F₈TPPH₂ ligand.

⁽²²⁾ Consistent with a high-spin porphyrinate-iron(III) center.

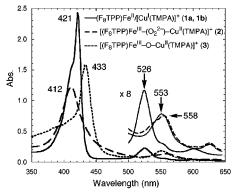


Figure 1. UV-visible spectra of the $(F_8TPP)Fe^{II}$ (1a)/[(TMPA)Cu^I(CH₃-CN)](ClO₄) (1b) oxygenation reaction in MeCN at -40 °C.

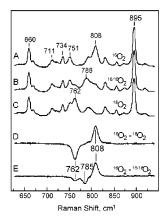


Figure 2. Resonance Raman spectra of $[(F_8TPP)Fe^{III}-(O_2^{2-})-Cu^{II}-(TMPA)]^+$ (2), formed by oxygenation of $(F_8TPP)Fe^{II}/[(TMPA)Cu^I(CH_3-CN)]^+$ (1a:1b = 1:1) in MeCN at -40 °C using $^{16}O_2$ (A), a scrambled mixed-isotope gas containing 25% $^{16}O_2$, 50% $^{16}O^{-18}O$, and 25% $^{18}O_2$ (B), and a pure $^{18}O_2$ gas (C). All spectra were obtained at room temperature with a 413 nm excitation in MeCN solvent. The difference spectra A minus C, and A minus B are also shown as traces D and E, respectively.

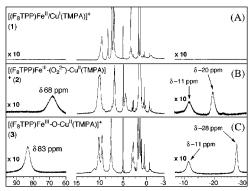


Figure 3. ¹H NMR spectra (400 MHz, CD₃CN, -40 °C) of heme—copper complexes. See text for further explanation.

= $^{1}/_{2}$ copper(II) through the bridging peroxide ligand. 23,24 We have previously observed and reported this characteristic pattern of downfield shifted pyrrole resonances and upfield shifted peaks in similar (P)Fe^{III}-X-Cu^{II} (X = O_{2}^{2-} , O^{2-}) systems (including 3, Scheme 1) having S=2 spin states. 19,23,24 In fact, thermal decomposition of 2 yields μ -oxo complex [(F₈TPP)Fe^{III}-O-Cu^{II}-(TMPA)]⁺ (3) {Scheme 1; μ _B = 5.1, Evans method, -40 °C, λ _{max} = 433 (Soret), 553 nm}, with previously assigned down-

field shifted pyrrole resonances (δ 83 ppm), and upfield shifted peaks at - 11 (pyridyl 5-H) and - 28 (pyridyl 3-H) ppm (Figure 3C).^{6,24}

The electronic structure of complex **2** was further probed by Mössbauer spectroscopy (4.2 K, zero field), which shows a sharp quadrupole doublet (Figure S1) with parameters $\{\Delta E_Q = 1.14 \text{ mm/s}, \delta = 0.57 \text{ mm/s}\}$ typical for high-spin ferric compounds. The isomer shift is significantly larger than those (0.33–0.45 mm/s) observed for high-spin ferric-heme compounds, ²⁵ and is consistent with binding of an electron-rich peroxide ligand. ²⁶ The magnetic field dependence (data not shown) is also consistent with **2** being an integer spin system. ²⁷ Porphyrin skeletal modes in the high-frequency region of the resonance Raman spectra confirm these conclusions, and reveal a pentacoordinate high-spin configuration of the heme iron in the peroxo intermediate **2** (data not shown).

Stopped-flow UV-visible spectroscopy (500–700 nm monitoring, acetone, -94 to -75 °C) revealed the presence of a hemesuperoxo (F₈TPP)Fe^{III}-(O₂⁻) intermediate^{3,19} { $\lambda_{\rm max}=537$ nm}, formed within mixing time (~1 ms) prior to formation of the heme-peroxo-Cu complex 2 { $\lambda_{\rm max}=556$ nm}, with little or no Cu-only O₂-adducts observed. The overall kinetics are complicated by 2–3 side reactions with minor absorbance changes, but the main 537 \rightarrow 556 nm heme-superoxo to heme-peroxo-Cu transformation can be reasonably described by a first-order rate constant with $\Delta H^{\ddagger}=45\pm1$ kJ mol $^{-1}$ and $\Delta S^{\ddagger}=-19\pm6$ J mol $^{-1}$ K $^{-1}$ (k=0.07 s $^{-1}$, -90 °C, k=0.32 s $^{-1}$, -80 °C).

As mentioned, μ -peroxo complex $[(F_8TPP)Fe^{III}-(O_2{}^2)-Cu^{II}(TMPA)]^+$ (2) transforms thermally to the μ -oxo complex $[(F_8TPP)Fe^{III}-O-Cu^{II}(TMPA)]^+$ (3). We find that this occurs in a slow reaction $\{t_{1/2}=1016\pm20\text{ s; MeCN, }22\text{ °C, }0.28\text{ mM}\}$, with concomitant release of 0.40-0.45 equiv of $O_2.^{28}$ Given the $1a+1b+O_2\to2$ stoichiometry (vide supra), and that the subsequent decomposition of 2 yielding 3 releases \sim 0.5 equiv O_2 , the fate of all oxygen atoms in the formation and decomposition of 2 is known. The mechanism of O-O reductive cleavage in the transformation $2\to3+\frac{1}{2}O_2$ will be the object of future study.²⁹

In conclusion, complex $[(F_8\text{TPP})Fe^{\text{III}}-(O_2^{2^-})-Cu^{\text{II}}(\text{TMPA})]^+$ (2) contains a symmetrically bound peroxide (i.e., most likely μ -1,2 or μ - η^2 : η^2) in a high-spin heme—Cu antiferromagnetically coupled S=2 system. The resonance Raman mixed-isotope experiment, the Mössbauer spectroscopic data, the stopped-flow kinetics, and the observation of O_2 evolution in the crudely biomimetic reductive O—O cleavage reaction {i.e., thermal decomposition of 2} are all significant new advances.

Acknowledgment. We are grateful to the National Institutes of Health (K.D.K., GM28962; R.J.C., GM54882; P.M.-L., GM34468 to Professor T. M. Loehr; B.H.H., GM58778) and Swiss National Science Foundation (A.D.Z.) for support of this research.

Supporting Information Available: Mössbauer spectrum (Figure S1) for complex **2** (4.2 K, MeCN) (PDF). This material is available free of charge via the Internet at http://pubs.acs.org.

JA010602Y

⁽²³⁾ Ghiladi, R. A.; Ju, T. D.; Lee, D.-H.; Moënne-Loccoz, P.; Kaderli, S.; Neuhold, Y.-M.; Zuberbühler, A. D.; Woods, A. S.; Cotter, R. J.; Karlin, K. D. *J. Am. Chem. Soc.* **1999**, *121*, 9885–9886.

⁽²⁴⁾ Nanthakumar, A.; Fox, S.; Murthy, N. N.; Karlin, K. D. *J. Am. Chem. Soc.* **1997**, *119*, 3898–3906.

⁽²⁵⁾ Debrunner, P. G. In *Iron Porphyrins*; Lever, A. B. P., Gray, H. B., Eds.; VCH Publisher: New York, 1990; pp 139–234.

⁽²⁶⁾ Burstyn, J. N.; Roe, J. A.; Miksztal, A. R.; Shaevitz, B. A.; Lang, G.; Valentine, J. S. *J. Am. Chem. Soc.* **1988**, *110*, 1382–1388.

⁽²⁷⁾ A detailed full study is in progress.

⁽²⁸⁾ Dioxygen evolution was confirmed by a quantitative alkaline pyrogallol test for O₂ release.

⁽²⁹⁾ Non-heme (DuBois, J. L.; Mizoguchi, T. J.; Lippard, S. J. *Coord. Chem. Rev.* **2000**, 200, 443–485) and heme (Chin, D.-H., LaMar, G. N., Balch, A. L. *J. Am. Chem. Soc.* **1980**, 102, 4344–4350) peroxo-diiron systems are known to undergo similar peroxo-to-oxo transformations which release O₂.