

Spectroscopic Rationale for Efficient Stimulated-Emission Depletion Microscopy Fluorophores

Jun-ichi Hotta,^{*,†} Eduard Fron,[†] Peter Dedecker,[†] Kris P. F. Janssen,[‡] Chen Li,[§] Klaus Müllen,[§] Benjamin Harke,^{||} Johanna Bückers,^{||} Stefan W. Hell,^{||} and Johan Hofkens[†]

Department of Chemistry, Katholieke Universiteit Leuven, Celestijnenlaan 200F, B-3001 Heverlee, Belgium, BIOSYST - MeBioS, Katholieke Universiteit Leuven, Willem de Croylaan 42, B-3001 Heverlee, Belgium, Max-Planck-Institut für Polymerforschung, Ackermannweg 10, 55128 Mainz, Germany, and Department of NanoBiophotonics, Max Planck Institute for Biophysical Chemistry, Am Fassberg 11, 37077 Göttingen, Germany

Received January 5, 2010; E-mail: junichi.hotta@chem.kuleuven.be

For decades, the power of far-field fluorescence microscopy has been hampered by the diffraction resolution limit,^{1,2} $d = \lambda/(2NA) > 200$ nm,^{3,4} where λ denotes the light wavelength and NA the numerical aperture of the lens. Identical fluorophores closer than d cannot be discerned because their signals cannot be separated by the detector. However, in the recent past, it has been demonstrated that the diffraction limit can be fundamentally overcome⁵ by modulating or switching the ability of the dye to emit fluorescence, causing adjacent objects to be registered sequentially. To this end, stimulated-emission depletion (STED) microscopy^{5,6} and its derivatives modulate the fluorescence capability of dye ensembles using defined spatial patterns of light, whereas the techniques known as (f)PALM,^{7,8} (d)STORM,^{9,10} and related concepts¹¹ involve switching individual fluorophores stochastically in space followed by mathematical localization of their coordinates. Hence, all of these concepts harness a molecular mechanism that keeps the dye nonresponsive when it is exposed to excitation light.

In STED microscopy (Figure 1),¹² the dyes are kept non-responsive (dark) by subjecting them to red-shifted light, called the STED beam, which induces stimulated emission from their fluorescent state, S_1 , to their ground state, S_0 . The maximum occupation probability of the S_1 state decreases nearly exponentially with the intensity I of the STED beam. Applying the condition $I \gg I_s$, where I_s is a dye constant, virtually confines the dye to the dark S_0 state, even if it is subject to excitation light. Usually formed as a donut, the STED beam is superimposed upon the regularly focused excitation spot to keep dark all of the dye molecules that are potentially present except those at the center of the donut. Only dyes that happen to be located within half the distance from the center given by $\rho \approx \lambda(1 + I_m/I_s)^{-0.5}/(2NA) \ll d$, in which I_m is the intensity at the donut crest, are allowed to be bright (ρ is the resolution in STED). Scanning the superimposed beams across the object sequentially registers fluorophores that are further than ρ apart. The resolution ρ can in principle become arbitrarily high for $I \gg I_s$. Clearly, the fluorophores that had been kept dark by STED must instantly become bright when they fall into the donut minimum, but this is the case only if they have not been transferred to a long-lived dark state. This aspect is critical for small ρ , because larger values of I increase the probability that a dye in the S_1 state passes to such a long-lived dark state, or simply bleached. In contrast, stimulated emission is instant, reversible, and so fundamental that almost any fluorophore can be transiently turned off by a STED beam.¹³ Thus, if long-term

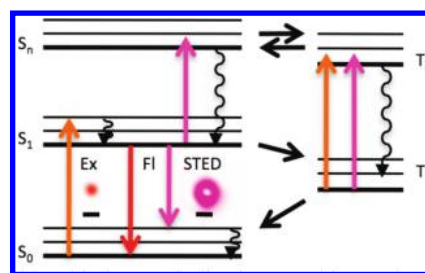


Figure 1. Jablonski diagram indicating transitions relevant for STED. Scale bars under the excitation and focal spots represent 500 nm.

darkening is minimized, STED should provide nanoscale resolution on the ensemble level with nearly any fluorophore.

Therefore, avoiding any process that competes with stimulated emission is prudent. Tuning the STED beam wavelength to the red edge of the fluorescence spectrum largely avoids excitation from the S_0 state, but transitions originating from excited states may still occur, such as crossing to the triplet states, T_n , as well as absorption to S_n and T_n states with $n > 1$ (Figure 1). In fact, many bleaching pathways involve triplet states,^{14,15} which are effectively populated through the S_n states. The higher S_n states can also lead to bleaching directly.¹⁶ Both processes compete with favorable ultrafast non-radiative $S_n \rightarrow S_1$ (or $T_n \rightarrow T_1$) relaxation followed by spontaneous or stimulated emission.¹³

With these conditions in mind, we explored the use of rylene dyes for STED microscopy. In view of their large extinction coefficient, brightness, and photostability (especially when applied in materials sciences^{4,17}), these dyes should, at face value, be ideal for STED. Specifically, we investigated tetraphenoxo-substituted perylenediimide (pPDI) and tetraphenoxo-substituted terylenediimide (pTDI)¹⁸ dyes, which have revealed remarkable photostability and low intersystem crossing in single-molecule experiments.¹⁷ Single pPDI and pTDI molecules can be registered for over 1 h without photobleaching and with few excursions to dark states.¹⁷

STED microscopy has been carried out with excitation light pulses followed by pulses for STED or with continuous wave (CW) beams for excitation and STED. The pulsed-mode setup has been reported elsewhere,^{6,19} while the CW system used here is detailed in the Supporting Information. First attempts with pPDI in the CW mode using a STED wavelength of 700 nm exhibited pronounced bleaching, as only <40% of the molecules seen in the confocal image were also found in the STED image (Figure 2a,b). Using 76 MHz pulse pairs with a STED wavelength of 740 nm slightly reduced the bleaching; a resolution gain by a factor of ~ 3 was observed (~ 65 nm; see Figure S2 in the Supporting Information) when a STED power of 33 mW was applied in the focal plane.

[†] Department of Chemistry, Katholieke Universiteit Leuven.

[‡] BIOSYST - MeBioS, Katholieke Universiteit Leuven.

[§] Max-Planck-Institut für Polymerforschung.

^{||} Max-Planck-Institute for Biophysical Chemistry.

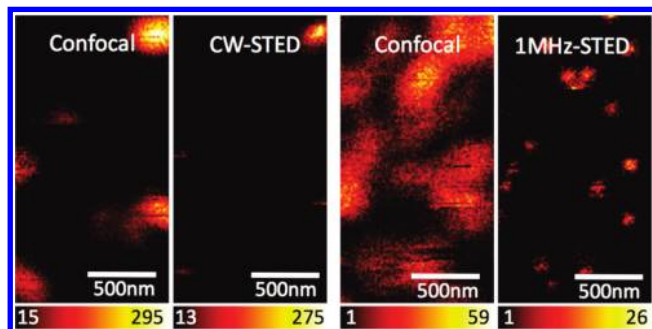


Figure 2. CW-STED image of pPDI exhibiting bleaching. In the CW mode, the excitation (543 nm) and STED (700 nm) beam powers were 2.0 μ W and 16 mW, respectively. Bleaching was slightly reduced upon application of pulsed-mode STED microscopy with a 1 μ s interpulse time interval using a 2.0 μ W (543 nm) excitation beam and a 670 μ W (710 nm) STED beam. The pixel dwell times for each pixel were 2 and 1 ms for CW and 1 MHz pulsed STED, respectively.

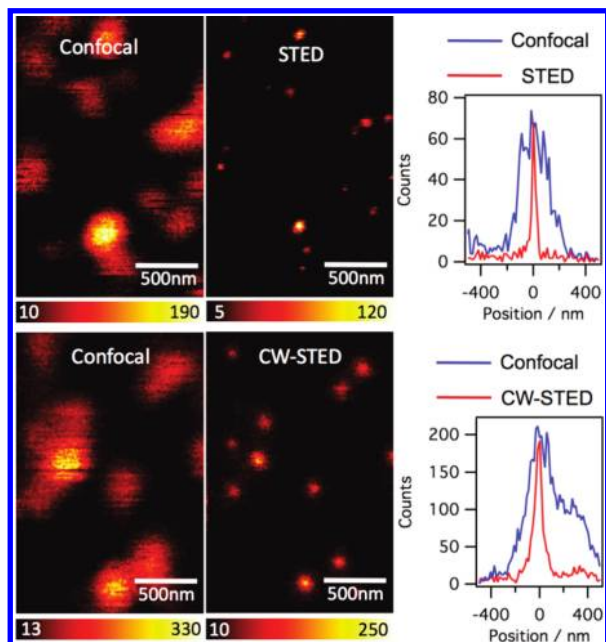


Figure 3. Confocal vs STED images of single pTDI molecules for (top) 76 MHz pulsed STED and (bottom) CW-STED. The bin times for each pixel were 0.5 and 2 ms for the pulsed and CW-STED modes, respectively. In the pulsed mode, the average excitation power was 4.0 μ W (635 nm), whereas in the CW mode, 2.5 μ W (633 nm) was employed. The average power of the pulsed STED beam (775 nm) was 32 mW, whereas that of the CW-STED beam (780 nm) was 58 mW. The fwhm of pulsed STED and CW-STED images of single molecules (point spread functions) were 34 and 78 nm, respectively.

For this power setting, $\sim 50\%$ of the molecules could be observed after acquisition of the STED image. Using a lower repetition rate of 1 MHz reduced the bleaching to some extent, since the associated time between the pulse pairs (1 μ s) allowed relaxation of dark states before the arrival of the succeeding pulse pair^{15,20} and gave slightly better resolution (55 nm; see Figure S3). In contrast, as shown in Figure 3, pTDI exhibited excellent performance at 76 MHz, with $\sim 95\%$ of the molecules in the confocal image also emerging in the STED counterpart. The resolution obtained with the employed STED beam power was 35 nm (without further data treatment). pTDI showed equally good performance in the CW mode too, with 90% recovery and 75 nm resolution. Longer interpulse breaks were not required.²⁰

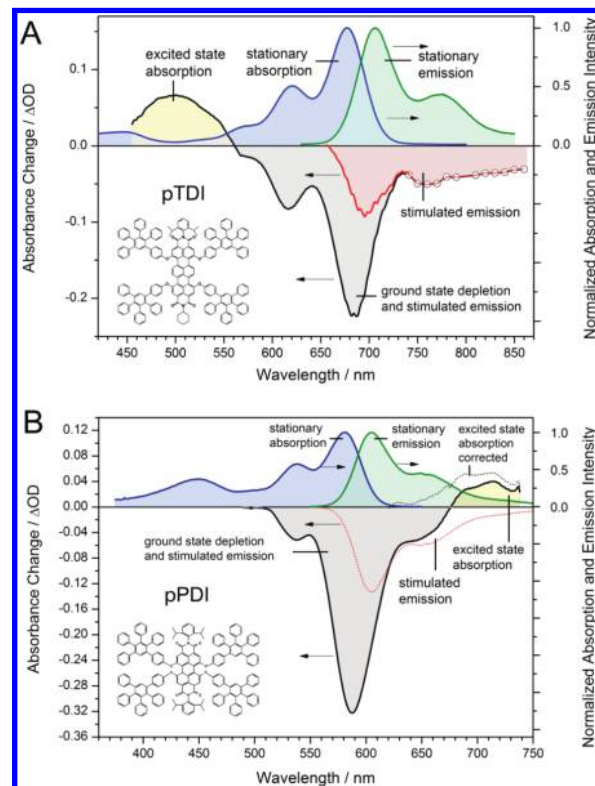


Figure 4. Transient absorption spectra of pTDI and pPDI. The transient absorption band ($S_1 \rightarrow S_n$) is blue-shifted for pTDI and well-separated from the stimulated-emission band and STED de-excitation beam wavelength (780 nm) used. The excited-state absorption band of pPDI overlaps with the stimulated-emission spectrum; at the STED beam wavelength (700 nm), strong $S_1 \rightarrow S_n$ absorption occurs.

Since in the reported single-molecule (excitation only) experiments the photostability of pPDI was comparable to that of pTDI, we found the differences observed with STED rather puzzling. We therefore carried out femtosecond transient absorption measurements to elucidate the photophysics and photochemistry of the excited states of the two fluorophores.²¹

Figure 4 shows the transient absorption spectra recorded 5 ps after excitation for pPDI and pTDI in toluene. In these plots, augmented optical density indicates absorption of the S_1 state populated after excitation, while negative values are due to a superposition of ground-state depletion (the disappearance of the S_0 absorption due to the formation of an excited state) and stimulated emission from the S_1 state.

Both pPDI and pTDI show strong negative contributions originating from ground-state depletion and stimulated emission. More interesting, however, are their $S_1 \rightarrow S_n$ absorption bands. In the case of pTDI, this band is blue-shifted relative to the $S_0 \rightarrow S_1$ absorption. In contrast, pPDI exhibits $S_1 \rightarrow S_n$ absorption at the red edge of the emission spectrum.

To avoid anti-Stokes excitation by the STED beam, its wavelength is typically chosen to lie in the red tail of the emission spectrum. For pTDI, this is a safe choice that leads to the excellent STED performance demonstrated above. Altogether, these results indicate that a good STED dye shows negligible $S_1 \rightarrow S_n$ absorption at the STED wavelength, thereby avoiding population of higher excited states.

In the case of pPDI, the situation is different. While stimulated emission undoubtedly occurs, the fluorophore can effectively absorb a photon and move on to a higher-energy S_n state. The fluorophore can rapidly (within <5 ps) convert back to the S_1 state, from which

it fluoresces, undergoes stimulated emission, or re-enters the same cycle, each with a probability corresponding to the transition probability of the respective process. Unfortunately, non-negligible excited-state absorption fosters repeated excitation to the S_n state and hence photobleaching, either by facilitating crossing to the T_n state or by starting a bleaching reaction straight from the S_n state.²¹ Clearly, the average number of cycles into higher excited states per fluorophore increases with smaller ρ .

The deconvolution of the transient absorption spectrum (500–740 nm) into the corresponding ground-state absorption, stimulated-emission, and excited-state absorption spectra reveals that pPDI has no “good wavelength band” to achieve de-excitation without substantial excitation to higher states. Nevertheless, when the (photoinduced oxidation) bleaching pathways from the higher states or the triplet state are blocked by working under a nitrogen atmosphere, improved STED images of pPDI can be recorded, as shown in Figure S4. Like the 1 MHz repetition rate experiment, this finding indicates the participation of long-lived (triplet) states in bleaching while also underscoring the relevance of avoiding pumping to higher states in STED microscopy.

In conclusion, our work provides a rationale for the suitability of dyes for STED microscopy. We have demonstrated that the excited-state properties are essential for transiently keeping the fluorophore dark by stimulated emission. Dyes for which the $S_1 \rightarrow S_n$ absorption spectrum is shifted with respect to its stimulated-emission counterpart satisfy the prerequisite for excellent performance in STED microscopy. For a given fluorophore, the location of the excited-state absorption band is difficult to predict, but for many dyes, the excited-state properties have been published. For example, a dye with little $S_1 \rightarrow S_n$ absorption is the red-emitting dye Atto647N, which to date has been the most popular dye for STED at ~ 750 nm. Hence, our investigations suggest suitable fluorophores and guide the optimization of the experimental settings. Moreover, they pinpoint a chief criterion, namely, spectral decoupling of the stimulated-emission and $S_1 \rightarrow S_n$ bands, for designing and selecting dyes for a non-diffraction-limited far-field fluorescence microscopy that is based just on molecular state transitions.

Acknowledgment. We acknowledge D. Schwarzer, E. Rittweger, and A. Walter (all of Max Planck Institute for Biophysical Chemistry) for valuable discussions. Financial support from the “Fonds voor Wetenschappelijk Onderzoek FWO” (Grant G.0366.06), the K. U. Leuven Research Fund (GOA 2006/2, Center of Excellence CECAT, CREA2007), the Flemish Government (long-term structural funding—Methusalem), and the Federal Science Policy of Belgium (IAP-VI/27) is gratefully acknowledged. P. D. thanks the FWO for a postdoctoral fellowship. This work, as part of the European Science Foundation EUROCORES Program SONS, was supported by funds from the FWO and the E.C. Sixth Framework Program (ERAS-CT-2003-980409).

Supporting Information Available: Structures and complete chemical names of pPDI and pTDI, sample preparation for STED microscopy, details of the experimental setup for CW STED microscopy, STED images of pPDI with the 76 MHz system, CW STED images of pPDI under nitrogen, pulse-energy dependence of STED resolution, and a reference for triplet absorption. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- (1) (a) Abbe, E. *Arch. Mikrosk. Anat.* **1873**, 9, 413. (b) Lord Rayleigh, *Philos. Mag., Ser. 5* **1896**, 42, 167.
- (2) Webb, R. H. *Rep. Prog. Phys.* **1996**, 59, 427.
- (3) (a) Weiss, S. *Nat. Struct. Biol.* **2000**, 7, 724. (b) Moerner, W. E. *J. Phys. Chem. B* **2002**, 106, 910. (c) Tinnefeld, P.; Sauer, M. *Angew. Chem., Int. Ed.* **2005**, 44, 2642.
- (4) (a) Roelfaers, M. B. J.; Sels, B. F.; Uji-i, H.; De Schryver, F. C.; Jacobs, P. A.; De Vos, D. E.; Hofkens, J. *Nature* **2006**, 439, 572. (b) Dedecker, P.; Muls, B.; Deres, A.; Uji-i, H.; Hotta, J.; Sliwa, M.; Soumillion, J.-P.; Müllen, K.; Enderlein, J.; Hofkens, J. *Adv. Mater.* **2009**, 21, 1079.
- (5) Hell, S. W.; Wichmann, J. *Opt. Lett.* **1994**, 19, 780.
- (6) Willig, K. I.; Harke, B.; Medda, R.; Hell, S. W. *Nat. Methods* **2007**, 4, 915.
- (7) Betzig, E.; Patterson, G. H.; Sougrat, R.; Lindwasser, O. W.; Olenych, S.; Bonifacio, J. S.; Davidson, M. W.; Lippincott-Schwartz, J.; Hess, H. F. *Science* **2006**, 313, 1642.
- (8) Hess, S. T.; Girirajan, T. P. K.; Mason, M. D. *Biophys. J.* **2006**, 91, 4258.
- (9) Rust, M. J.; Bates, M.; Zhuang, X. *Nat. Methods* **2006**, 3, 793.
- (10) Heilemann, M.; van de Linde, S.; Schüttelpelz, M.; Kasper, R.; Seefeldt, B.; Mukherjee, A.; Tinnefeld, P.; Sauer, M. *Angew. Chem., Int. Ed.* **2008**, 47, 6172.
- (11) (a) Heilemann, M.; Dedecker, P.; Hofkens, J.; Sauer, M. *Laser Photonics Rev.* **2009**, 3, 180. (b) Dedecker, P.; Hotta, J.; Flors, C.; Sliwa, M.; Uji-i, H.; Roelfaers, M. B. J.; Ando, R.; Mizuno, H.; Miyawaki, A.; Hofkens, J. *J. Am. Chem. Soc.* **2007**, 129, 16132. (c) Flors, C.; Hotta, J.; Uji-i, H.; Dedecker, P.; Ando, R.; Mizuno, H.; Miyawaki, A.; Hofkens, J. *J. Am. Chem. Soc.* **2007**, 129, 13970. (d) Fujita, K.; Kobayashi, M.; Kawano, S.; Yamanaka, M.; Kawata, S. *Phys. Rev. Lett.* **2007**, 99, 228105.
- (12) (a) Keller, J.; Schönl, A.; Hell, S. W. *Opt. Express* **2007**, 15, 3361. (b) Dedecker, P.; Muls, B.; Hofkens, J.; Enderlein, J.; Hotta, J. *Opt. Express* **2007**, 15, 3372.
- (13) Rittweger, E.; Rankin, B. R.; Westphal, V.; Hell, S. W. *Chem. Phys. Lett.* **2007**, 442, 483.
- (14) Song, L.; Varmaa, C. A.; Verhoevena, J. W.; Tanke, H. J. *Biophys. J.* **1996**, 70, 2959.
- (15) Donnert, G.; Keller, J.; Medda, R.; Andrei, M. A.; Rizzoli, S. O.; Luehrmann, R.; Jahn, R.; Eggeling, C.; Hell, S. W. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, 103, 11440.
- (16) Tinnefeld, P.; Hofkens, J.; Herten, D. P.; Masuo, S.; Vosch, T.; Cotlet, M.; Habuchi, S.; Müllen, K.; De Schryver, F. C.; Sauer, M. *ChemPhysChem* **2004**, 5, 1786.
- (17) (a) Uji-i, H.; Melnikov, S. M.; Deres, A.; Bergamini, G.; De Schryver, F.; Herrmann, A.; Müllen, K.; Enderlein, J.; Hofkens, J. *Polymer* **2006**, 47, 2511. (b) Braeken, E.; De Cremer, G.; Marsal, P.; Pepe, G.; Müllen, K.; Vallee, R. A. L. *J. Am. Chem. Soc.* **2009**, 131, 12201. (c) Kirstein, J.; Platschek, B.; Jung, C.; Brown, R.; Bein, T.; Brauchle, C. *Nat. Mater.* **2007**, 6, 303.
- (18) Herrmann, A.; Müllen, K. *Chem. Lett.* **2006**, 35, 978.
- (19) Wildanger, D.; Rittweger, E.; Kastrop, L.; Hell, S. W. *Opt. Express* **2008**, 16, 9614.
- (20) Donnert, G.; Eggeling, C.; Hell, S. W. *Photochem. Photobiol. Sci.* **2009**, 8, 481.
- (21) Fron, E.; Pilot, R.; Schweitzer, G.; Qu, J.; Herrmann, A.; Müllen, K.; Hofkens, J.; Van der Auwerter, M.; De Schryver, F. C. *Photochem. Photobiol. Sci.* **2008**, 7, 597.

JA100079W