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Exploring the Use of Conformationally Locked Aminoglycosides as a New Strategy to Overcome Bacterial Resistance

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Abstract: The emergence of bacterial resistance to the major classes of antibiotics has become a serious problem over recent years. For aminoglycosides, the major biochemical mechanism for bacterial resistance is the enzymatic modification of the drug. Interestingly, in several cases, the oligosaccharide conformation recognized by the ribosomal RNA and the enzymes responsible for the antibiotic inactivation is remarkably different. This observation suggests a possible structure-based chemical strategy to overcome bacterial resistance; in principle, it should be possible to design a conformationally locked oligosaccharide that still retains antibiotic activity but that is not susceptible to enzymatic inactivation. To explore the scope and limitations of this strategy, we have synthesized several aminoglycoside derivatives locked in the ribosome-bound "bioactive" conformation. The effect of the structural preorganization on RNA binding, together with its influence on the aminoglycoside inactivation by several enzymes involved in bacterial resistance, has been studied. Our results indicate that the conformational constraint has a modest effect on their interaction with ribosomal RNA. In contrast, it may display a large impact on their enzymatic inactivation. Thus, the work presented herein provides a key example of how the conformational differences exhibited by these ligands within the binding pockets of the ribosome and of those enzymes involved in bacterial resistance can, in favorable cases, be exploited for designing new antibiotic derivatives with improved activity in resistant strains.

Introduction

Aminoglycoside antibiotics are highly potent, wide-spectrum bactericidals.^{1–4} These drugs bind to the decoding region aminoacyl-tRNA site (A-site) inducing codon misreading and inhibiting translocation, eventually resulting in cell death.^{1–4} From a chemical point of view, aminoglycosides are polycationic oligosaccharides carrying several hydroxyl and amino functional groups. Most include one or more amino sugars attached to a 2-deoxystreptamine ring (ring II in Scheme 1). The structures of several aminoglycosides, such as gentamycin, tobramycin, and paromomycin, bound to the target ribosomal A-site RNA have been determined recently by both X-ray and NMR methods.^{5–7} These structures have provided an extremely

detailed knowledge about the interactions that stabilize the drug/RNA complexes.

Several years after the introduction of aminoglycosides in human antibacterial chemotherapy, resistant organisms began to appear. Indeed, the emergence of bacterial resistance to the major classes of antibiotics has become a serious problem over recent years.^{8,9} Acquired resistance to aminoglycoside antibiotics can occur via three different mechanisms: mutation of the ribosomal target, reduced permeability for the antibiotics, and enzymatic modification of the drugs, thus leading to inactivation. The most prevalent source of clinically relevant resistance is conferred by the third mechanism, the enzymatic inactivation of the drugs.^{8,9} Enzymes involved in bacterial defense against aminoglycoside antibiotics can be broadly classified as *N*-acetyltransferases (AACs), *O*-adenyl transferases (ANTs), and

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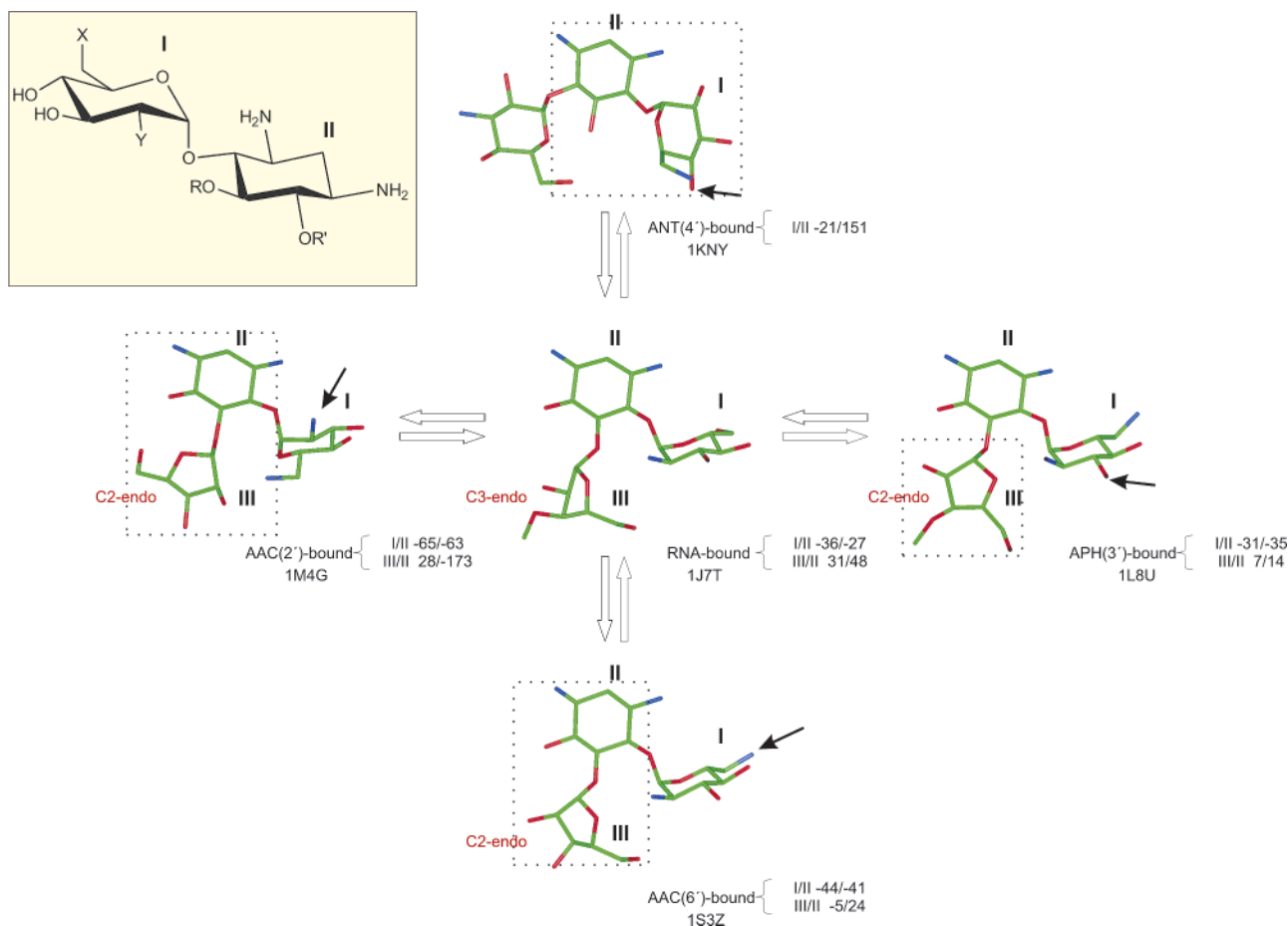
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Scheme 1. Schematic Representation of the Conformational Differences (Highlighted with a Dotted Box) Exhibited by Aminoglycosides in the Binding Pockets of RNA and Some of the Enzymes Involved in Bacterial Resistance^a



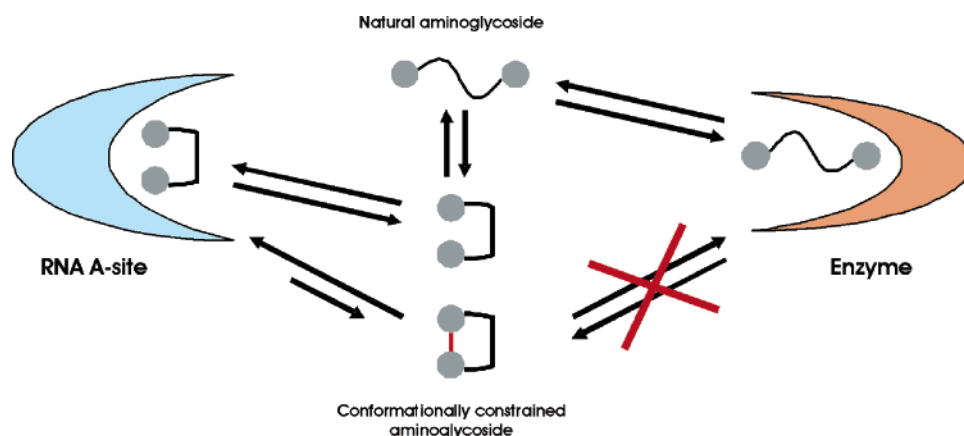
^a The glucose, 2-deoxy-streptamine, and ribose units are numbered as I, II, and III, respectively. The values of the glycosidic torsion angles Φ (H1–C1–Ox–Cx) and Ψ (Hx–Cx–O1–C1) for each glycosidic linkage in the bound state, together with the pdb codes of the complexes, are indicated. The position of the aminoglycoside modified by the enzyme is marked by an arrow. A schematic representation of the Glc α (1–4)-2-deoxy-streptamine fragment present in neomycin (X = NH₂, Y = NH₂, R = 2,6-diamino-2,6-dideoxy-L-Ido β (1–3)Rib, R' = H), paromomycin (X = OH, Y = NH₂, R = 2,6-diamino-2,6-dideoxy-L-Ido β (1–3)Rib, R' = H), ribostamycin (X = NH₂, Y = NH₂, R = Rib, R' = H), and kanamycin (X = NH₂, Y = OH, R = H, R' = 3-amino-3-deoxy- α -Glc) is shown in the top left-hand corner.

O-phosphotransferases (APHs). It is evident that a detailed knowledge of the structures of these enzymes and their interactions with the drugs should provide a framework that will facilitate the rational design either of novel aminoglycosides, not susceptible to modification, or of inhibitors of these enzymes, which could be administered in tandem with existing aminoglycosides. Indeed, the search for new derivatives not susceptible to modification by bacterial defense proteins constitutes an active field of research.^{10–13}

The 3D structures of several aminoglycoside–enzyme complexes have been described recently by X-ray methods.^{9,14} Interestingly, in some cases, the 3D structure of the antibiotic

in the enzyme binding site significantly differs from that observed in the corresponding aminoglycoside/A-site RNA complex (see Scheme 1). For example, *Staphylococcus aureus* ANT(4') selects a high energy conformation of the ligand, kanamycin.¹⁵ In particular, the Glc α (1–4)-2-deoxy streptamine fragment (numbered as units I and II in Scheme 1), present in all aminoglycosides with different patterns of amination/hydroxylation at the glucose unit (I), adopts an anti- Ψ geometry (Φ/Ψ , –21/151). In contrast, for tobramycin and paromomycin, the analogous glycosidic linkage is defined by a syn- Ψ conformation ($\Phi/\Psi \approx -40/-30$) in the RNA-bound state. The “syn- Ψ ” geometry also constitutes the most populated minimum for the free antibiotics, according to NMR data.¹⁶ The drug conformation selected by ANT(4') represents a high-energy minimum that must be stabilized by specific aminoglycoside–protein interactions. In a similar way, the X-ray structure of *Mycobacterium tuberculosis* AAC(2') in complex with ribostamycin shows that the Rib β (1–5)-2-deoxy streptamine fragment (num-

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Chart 1. Schematic Representation of the Proposed Structure-Based Chemical Strategy to Overcome Bacterial Resistance

bered as units III and II in Scheme 1) also adopts a high energy “anti- Ψ ” conformation ($\Phi/\Psi = 28/-173$).¹⁷ This geometry is significantly different to the *bioactive* “syn- ψ ” orientation adopted by the same glycosidic bond in the paromomycin/A-site complex.^{5,6} An additional key example of conformational differences of aminoglycosides between the RNA- and protein-bound states is provided by the recently reported structure of the *Salmonella enteritidis* AAC(6′) in complex with ribostamycin.¹⁸ In this case, the Rib β (1–5)-2-deoxy streptamine fragment (units III and II in Scheme 1) of the oligosaccharide presents a rather unusual nonexo-anomeric conformation (characterized by a Φ angle of -5°). In addition, the furanose ring (numbered as III in Scheme 1) is bound in a C2-endo conformation, in contrast with the C3-endo geometry selected by the ribosomal RNA. A similar furanose puckering is also recognized by the *Enterococcus faecalis* APH(3′) enzyme according to the X-ray structure of its complex with neomycin-B.¹⁹

The different 3D-shapes adopted by aminoglycosides in the RNA- and enzyme-bound states suggest a possible structure-based chemical strategy to obtain antibiotics with a better activity against resistant bacteria. Assuming that, in these cases, some degree of conformational distortion of the substrates is required for enzymatic activity, it should be possible to design a conformationally locked oligosaccharide that still retains antibiotic activity but that is not susceptible to enzymatic inactivation (Chart 1).²⁰

To test this hypothesis, we have designed and synthesized several neomycin-B analogues locked in the RNA-bound (*bioactive*) structure of the antibiotic.²¹ Thus, we have studied the effect that structural preorganization has on the strength of the aminoglycoside–RNA interaction. In a second step, we have determined the susceptibility of the conformationally locked derivatives to enzymatic inactivation, employing the enzymes *Staphylococcus aureus* ANT(4′), *Mycobacterium tuberculosis* AAC(2′), and *Enterococcus faecalis* APH(3′) as model systems.

The design of our conformationally constrained aminoglycosides is based on the structural information available for the RNA–paromomycin complex.^{5,6} The structure of neomycin-B (1) along with the numbering employed for the different sugar units along this manuscript (I, II, III, and IV for the glucose, 2-deoxy-streptamine, ribose, and idose units, respectively) is shown in Scheme 2. It is known that positions O5_{III} and N2_I are hydrogen bonded within the ribosome bound state. Thus, in our analogues, this interaction has been replaced by either a direct covalent bond, as in 2, a two-methylene bridge, as in 3, or by a salt bridge, as in 4 (see Scheme 2).

It should be pointed out that OH_{5_{III}} is involved in RNA recognition^{5,6} and might also have an influence on the enzymatic inactivation of the antibiotics. Since cyclization of neomycin-B to give 2 or 3 required the removal of this OH group, the deoxy-derivatives 5 and 6 were also synthesized as controls, to isolate the effect of rigidification.

In the first part of the manuscript we present a detailed description of the synthesis of the proposed neomycin-B derivatives. The next section deals with the structural NMR analysis of the obtained aminoglycosides. This study, essential in order to establish any structure–activity relationship, shows that, as expected, the obtained antibiotics exhibit a significantly reduced flexibility with respect to natural neomycin-B and, in addition, closely resemble its *bioactive* ribosome-bound conformation. In the next two sections the effect of the conformational restriction on the RNA binding properties and the biological activity of these analogues is determined. Finally the susceptibility of the obtained derivatives to enzymatic modification has been tested employing three different enzymes involved in bacterial resistance. This final analysis suggests that conformational restriction might constitute a useful approach to prevent aminoglycoside inactivation by bacterial defense proteins.

Results and Discussion

Synthesis of the Proposed Neomycin-B Derivatives.

A. Synthesis of Neomycin-B Conformationally Constrained Analogue 2. Compound 2 was prepared following the simple four-step procedure shown in Scheme 3. Thus, the hexa-*N*-(benzyloxycarbonyl) derivative (7), readily prepared from neomycin-B (1),²² was treated with 2,4,6-triisopropylbenzene-sulfonyl chloride in pyridine²³ to yield the primary sulfonate

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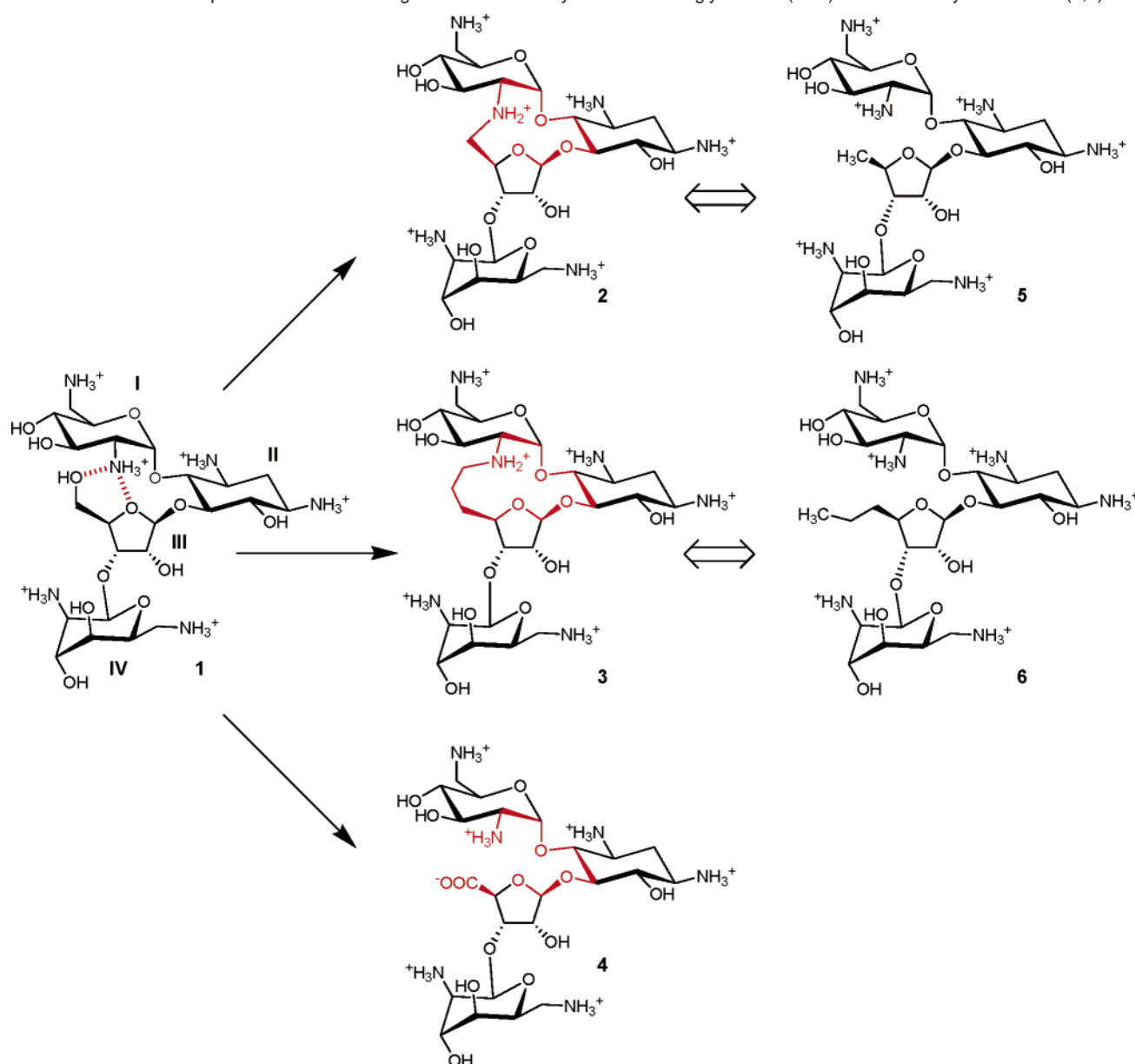
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Scheme 2. Schematic Representation of the Target Conformationally Locked Aminoglycosides (**2–4**) and the Deoxy Derivatives (**5,6**)^a

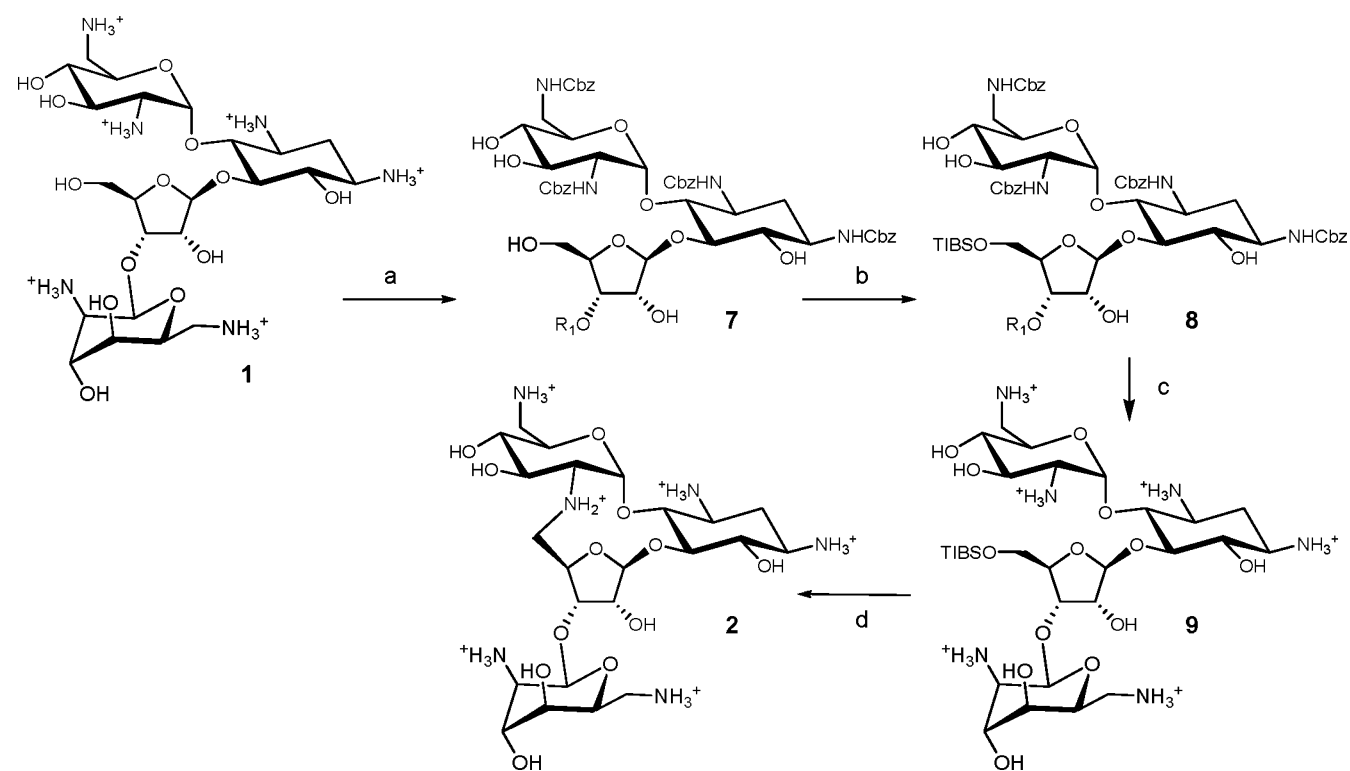
^a The numbering employed for the different sugar units is indicated for neomycin-B (compound **1**).

(**8**), regioselectively. Deprotection of the amino groups by palladium-catalyzed hydrogenolysis in MeOH/trifluoroacetic acid gave crude sulfonate (**9**) that smoothly cyclized in a highly regioselective way by mild heating in water at pH 7.0 to give **2** in 56% yield.

B. Synthesis of Neomycin-B Conformationally Constrained Analogue 3. The synthesis of compound **3** is shown in Scheme 4. The key step involves a two-carbon atom elongation of ring III side chain through a Wittig-type reaction on aldehyde **12**. The following four-step strategy was used to obtain this aldehyde in good yield. First, regioselective protection of the primary hydroxyl group of **7**, by treatment with triisopropylsilyl trifluoromethanesulfonate and 2,6-lutidine, followed by acetylation of the secondary hydroxyl groups gave compound **10**, which was desilylated by treatment with HF·pyridine to give **11**. Finally, oxidation of **11** using the Dess–Martin periodinane gave the required aldehyde **12** in 57% overall yield from **7**. Treatment of **12** with (methoxycarbonylmethylen)-

triphenylphosphorane in acetonitrile gave the corresponding α,β -unsaturated ester **13**, which was reduced with NaBH₄ in the presence of catalytic LiCl to give alcohol **14** in 89% yield. Initially, we tried the selective activation of the primary alcohol in **14**, by treatment with 2,4,6-triisopropylbenzenesulfonyl chloride in pyridine. However, the low regioselectivity obtained in the reaction forced us to use a three-step strategy similar to that described above. Thus, regioselective protection of the primary hydroxyl group in **14** as a triisopropylsilyl ether, yielded derivative **15**. Acetylation of the secondary hydroxyl groups followed by desilylation gave derivative **16**, which was transformed into the corresponding primary sulfonate by treatment with *p*-toluenesulfonyl chloride in pyridine. Finally, clean deacetylation of this derivative was best performed by mild KCN-catalyzed methanolysis,²⁴ to give **17**. The carbamate groups were removed by palladium-catalyzed hydrogenolysis

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Scheme 3^a

^a (a) CbzCl, Na₂CO₃, MeOH/H₂O 3:1, 0 °C, 3 h, 92%; (b) 2,4,6-triisopropylbenzenesulfonyl chloride (TIBSCl), Py, rt, 72 h, 44%; (c) H₂ (1 atm), Pd/C, MeOH, trifluoroacetic acid, rt, 8 h, 90%; (d) H₂O, pH 7.0, 60 °C, 5 d, 56%. In **7** and **8**, R₁ = 2,6-diamino-2,6-di-*N*-(benzyloxycarbonyl)-2,6-dideoxy-β-L-idose.

in MeOH/TFA. The resulting crude sulfonate was cyclized at 40 °C and pH 7.0 in H₂O to give **3** in 62% yield.

The cyclization proceeded with an astonishing high regioselectivity considering the number of amino groups present in this neomycin-B derivative (see Figure S1). In fact, compound **3** is the only macrocyclic derivative isolated from the reaction mixture. Close inspection of the corresponding molecular model shows that the two amino groups present in unit IV could also participate in the nucleophilic attack to the side chain of III. However, preliminary molecular mechanics calculations suggest that the cyclization involving NH₂ at position 2 of unit IV would proceed through a high energy transition state. The preferred cyclization through the NH₂ at position 2 of ring I instead of that at position 6 of unit IV can be explained considering the different pK_a values of these groups (7.4 vs 8.5, respectively). In fact, under the employed experimental conditions (pH 7.0), the later amino group is fundamentally in its protonated state.

Compound **6** was readily obtained from **17** by palladium-catalyzed hydrogenolysis of the Cbz protecting groups and subsequent reduction of the primary sulfonate under the same conditions using a longer reaction time.

Finally derivatives **4** and **5** were easily obtained from **11** as described in the Supporting Information.

NMR Analysis of the Conformationally Locked Amino-glycosides 2–4. Selective 1D-NOE experiments were carried out at 313 K and pH 5.0 for analogues **2–4**. In addition, ³*J* values were measured for all the sugar units (Table 1). To get an experimentally derived ensemble, MD-tar (molecular dynamics with time-averaged restraints) simulations²⁵ were carried out

by including all the experimental NOE-derived distances and *J* data as time-average restraints with the AMBER 5.0²⁶ program, as described.¹⁶

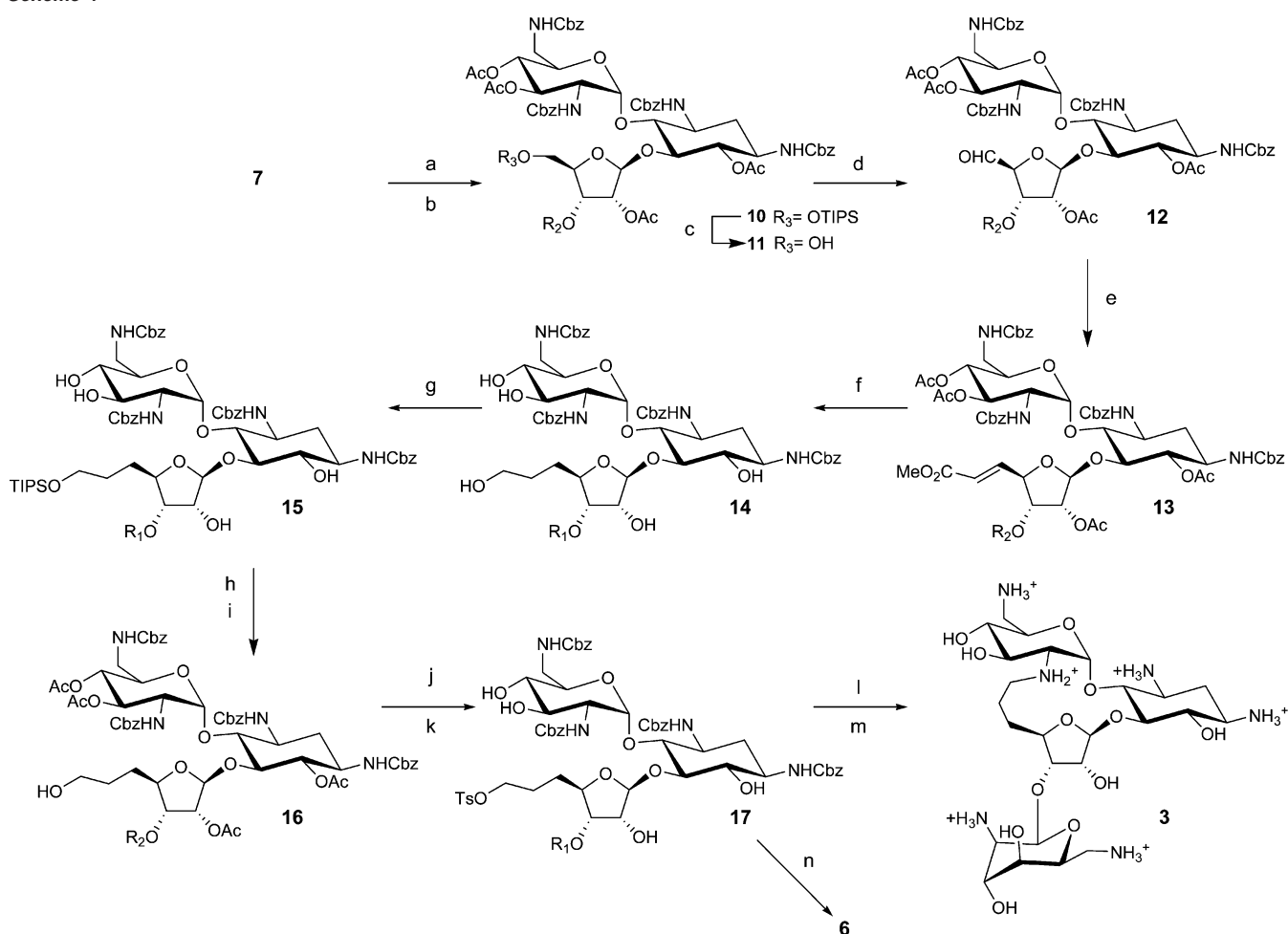
The obtained trajectories were able to quantitatively reproduce all the NMR experimental data. In all cases, the analyses of the *J* couplings for units I and II unambiguously show that the ⁴C₁ conformer is the very major one in water (>98%). In contrast, for ring IV the ¹C₄ conformer with three axial and two equatorial substituents is the only one experimentally detected. A similar behavior has been reported for the natural antibiotic,¹⁶ and therefore the cyclization of neomycin-B to give **2–3** does not produce significant distortions in any of the three pyranose rings.

According to the experimental data, derivative **2** is extremely rigid. Both *J* and NOE values (see Table 1 and Figure 1) are consistent with a pure “gt” (ω_{O–C4–C5–O5} ≈ 60°) orientation for the side chain of unit III and a unique puckering for its furanose ring. In fact, only minor fluctuations around the I/II and III/II glycosidic linkages are compatible with this macrocyclic structure, according to the MD-tar simulations.

A summary of the experimental data employed in the NMR analysis of **3** is shown in Figure 2. In this case, a large number of lower bound constraints had to be included in the MD-tar simulations (together with the NOE derived distances and *J* values) to properly define its structure in solution. As expected, the experimental data indicates the presence of a larger degree of flexibility. Thus, NOEs and *J* values are consistent with the

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Scheme 4^a

^a (a) Triisopropylsilyl trifluoromethanesulfonate (TIPSOTf), 2,6-lutidine, THF, rt, 1 h, 80%; (b) Ac₂O, Py, rt, 5 h, ~100%; (c) HF·Py, THF, rt, 5 h, 90%; (d) Dess–Martin periodinane, CH₂Cl₂, rt, 2 h, 79%; (e) Ph₃P=CHCO₂Me, CH₃CN, rt, 3 h, 82%; (f) NaBH₄, cat. LiCl, EtOH/THF, rt, 12 h, 89%; (g) TIPSOTf, 2,6-lutidine, THF, 0 °C, 1 h, 60%; (h) Ac₂O, Py, rt, 5 h, ~100%; (i) HF·Py, THF, rt, 5 h, 79%; (j) *p*-toluenesulfonyl chloride (TsCl), Py, cat. DMAP, rt, 3 h, 79%; (k) cat. KCN, MeOH/THF 2:1, rt, 10 h, 58%; (l) H₂ (1 atm), Pd/C, MeOH, trifluoroacetic acid, rt, 8 h, ~88%; (m) H₂O, pH 7.0, 40 °C, 10 d, 62%; (n) H₂ (1 atm), Pd/C, MeOH, trifluoroacetic acid, rt, 48 h, ~60%; In **14**, **15**, and **17** R₁ = 2,6-diamino-2,6-di-*N*-(benzyloxycarbonyl)-2,6-dideoxy-β-*L*-idose. In **10–13** and **16** R₂ = 3,4-di-*O*-acetyl-2,6-diamino-2,6-di-*N*-(benzyloxycarbonyl)-2,6-dideoxy-β-*L*-idose.

presence of several orientations for the propylidene bridge that links units I and III. For the side chain of ring III, a major (>80%) “gt” ($\omega_{O-C4-C5-C6} \approx 60^\circ$) and a minor (<20%) “gg” ($\omega_{O-C4-C5-C6} \approx -60^\circ$) orientation are detected. Moreover, the furanose ring presents two different puckerings characterized by phase angles of 25° (“North”) and 160° (“South”) (for phase angles the Altoona and Sundaralingam²⁷ convention is employed) and relative populations of 40:60, respectively. According to the MD-trajectories, the macrocyclic structure present in **3** is compatible with relatively large fluctuations around the I/II and III/II glycosidic linkages (see Figure 3).

Finally, our combined NMR/MD approach indicates that derivative **4** also exhibits a significant degree of flexibility (Figure 4). As previously observed for **3**, ring III presents two different puckerings characterized by phase angles around 5° and 170° and relative populations of 30:70, respectively. The polar interaction between NH₂I and the carboxylate present in III (see Figure 4) seems to be compatible with the presence of relatively wide fluctuations around the I/II and III/II glycosidic linkages.

The obtained ensembles of conformers for **2–4** that quantitatively reproduce the experimental data are shown in Figure 5. These ensembles were compared to that obtained for the natural antibiotic **1** in the free (experimentally derived from the NMR data)¹⁶ and RNA-bound (derived from unrestrained MD simulations of the neomycin-B/A-site complex) states.¹⁶ As expected, analogues **2–4** are more rigid than **1** in the free state. The RMSD for heavy atom superimposition for the ensemble corresponding to **1** in the free state is 2.1 Å changing to 0.4 Å for the bound ligand. The corresponding values for **3** and **4** are 1.6 Å. The conformational restriction imposed by covalently bonding C5_{III} to N2_I in **2** is especially severe with an RMSD value of 0.9 Å.

Interestingly, the locked aminoglycosides **2–4** closely resemble the bioactive conformation of the natural antibiotic. The most representative minimum energy conformations present in solution for **2** and **3**, superimposed on the X-ray structure of paromomycin in complex with A-site RNA, are shown in Figure 6. For **2**, deviations in Φ/Ψ values with respect to the paromomycin bound structure are less than 11° and 21° for the I/II and III/II linkages, respectively. The corresponding devia-

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Table 1. Experimental and Theoretical Distances (above) and Coupling Constant Values (below) Obtained for Compounds **2**, **3**, and **4** at pH 5.0^a

$d(\text{\AA})$	compound 2		compound 3		compound 4	
	expt	MD-tar (5 ns)	expt	MD-tar (80 ns)	expt	MD-tar (5 ns)
H1 _I –H4 _{II}	2.4	2.4	2.4	2.3	2.5	2.5
H1 _I –H5 _{II}			>3.3	3.8	3.3	3.2
H1 _I –H1 _{III}			>3.3	4.5	3.9	4.0
H1 _I –H2 _{III}			>3.3	3.7	3.4	3.2
H1 _I –H3 _{III}			>3.3	3.3		
H1 _I –H5 _{III} R	2.4	2.3	>3.3	5.1		
H1 _I –H5 _{III} S			>3.3	3.3		
H1 _I –H6 _{III} R/S			>3.3/>3.3	3.3/4.6		
H1 _I –H7 _{III} R/S			>3.3/>3.3	3.3/3.6		
H2 _I –H5 _{III} R	<2.4	2.4				
H2 _I –H7 _{III} R			2.6	2.6		
H3 _I –H7 _{III} R/S			>3.3/>3.3	3.5/3.3		
H1 _{III} –H5 _{II}	2.6	2.6	2.2	2.3	2.5	2.5
H1 _{III} –H4 _{II}	>3.5	4.5	>3.3	4.4		
H1 _{III} –H6 _{II}	3.5	3.6	>3.3	3.7	3.6	3.3
H1 _{III} –H4 _{III}	3.6	3.7	3.1	3.0	3.5	3.3
H2 _{III} –H7 _{III} R/S			>3.3/>3.3	3.3/4.9		
H2 _{III} –H6 _{II}			>3.3	4.2	3.2	3.3
H2 _{III} –H6 _{III} R/S			>3.3/>3.3	4.9/3.4		
H2 _{III} –H5 _{III} R	<2.4	2.2				
H3 _{III} –H7 _{III} R/S			>3.3/>3.3	3.3/4.6		
H3 _{III} –H6 _{III} R/S			>3.3/>3.3	4.3/3.2		
H4 _{III} –H6 _{III} R/S			>3.3/>3.3	3.2/3.8		
H1 _{IV} –H2 _{III}	>4	4				
H1 _{IV} –H3 _{III}	2.5	2.6	2.5	2.3	2.4	2.5
H1 _{IV} –H4 _{III}	3.5	3.5	>3.3	4.0	3.8	3.9
$J(\text{Hz})$	expt	MD-tar (5 ns)	expt	MD-tar (80 ns)	expt	MD-tar (5 ns)
H1 _{III} –H2 _{III}	3.9	4.5	3.5	4.1	3.9	4.0
H2 _{III} –H3 _{III}	4.9	4.2			4.0	4.1
H3 _{III} –H4 _{III}	<1.0	1.1	3.5	4.5	4.4	4.3
H4 _{III} –H5 _{III} R	11.6	11.0	<3.5	4.1		
H4 _{III} –H5 _{III} S	4.0	4.7	9.5	8.8		
H7 _{III} R–H6 _{III} R			3.0	3.4		
H7 _{III} R–H6 _{III} S			8.5	7.9		
H7 _{III} S–H6 _{III} R			7.0	7.2		

^a Molecular dynamic simulations with time-averaged restraints (MD-tar) for compounds **2** and **4** were carried out using explicit solvent, counterions, periodic boundary conditions (PBC), and Ewald sums for the treatment of electrostatic interactions. For compound **3**, in vacuo 80 ns MD-tar trajectories were collected (see the Materials and Methods). The stereochemistry of the protons involved in the distance constraints is indicated when appropriate.

tions for **3** are larger (15° and 45°). However, the global shape of the molecule is very similar to that observed for paromomycin in the RNA-bound state, as shown by the low RMSD values for heavy atom superimposition (see Figure 6).

Previous studies have shown that neomycin-B in solution is characterized by a remarkable flexibility, with different conformations around the three glycosidic linkages and two different conformational populations for the unit III furanose ring (with phase angles ~20° and ~160° and populations of 60–70% and 30–40%, respectively) in fast exchange.¹⁶ On the other hand, MD simulations of neomycin-B complexed to ribosomal A-site RNA suggest that its flexibility is severely reduced upon binding and only one of the furanose conformations is selected by the RNA receptor. Interestingly, in analogue **2**, the unit III furanose is locked in a very similar conformation to that present in the A-site/paromomycin complex, with an almost identical orientation of the key polar groups, O2 and O3 (Figure 6).

In conclusion, according to our structural analysis, derivatives **2–4** satisfy the basic requirements for an optimal conforma-

tionally constrained derivative; they are all more rigid than the natural antibiotic neomycin-B and closely resemble its bioactive, RNA-bound conformation.

Fluorescence-Based Analysis of the Interaction between Aminoglycosides 1–6 and the Ribosomal Decoding Site. As a next step, the binding properties of compounds **1–4** to ribosomal RNA were analyzed. It is known that, for natural paromomycin, OH5_{III} is involved in RNA recognition.^{5,6} Cyclization of neomycin-B to give **2** and **3** requires the removal of this OH group. To isolate the effect of rigidification on the aminoglycoside–RNA interaction, the binding properties of the deoxyderivatives **5** and **6** were also tested. The fluorescence-based approach, developed independently by Pilch²⁸ and Hermann,²⁹ was employed. This methodology makes use of a modified 27-mer RNA fragment (A-site(2AP)) including the A-site sequence and the fluorescent base analogue 2-amino-purine (2AP) replacing A1492 (Figure 7a). It is well established that major groove binding of aminoglycosides to the A-site induces a significant conformational change in the RNA so that the adenine residues at positions 1492 and 1493 are displaced from the helical stack toward the minor groove.^{5–7} As the fluorescence of 2AP is modulated by hydrophobic stacking interactions, the employed RNA fragment (shown in Figure 7a) provides an adequate means for monitoring and quantifying the destacking of A1492 that accompanies complex formation.

It has been shown that paromomycin binding to the A-site(2AP) induces a marked increase in the fluorescence intensity at 369 nm. These changes were monitored as a function of the [drug]/[RNA] ratio in titration experiments, to derive K_d values. Some typical binding curves are shown in Figure 7b. The affinity constants measured for compounds **1–6** are shown in Figure 8. According to our titration experiments, natural neomycin-B binds to the A-site with a K_d value of 0.07 μM in 100 mM NaCl, 10 mM phosphate and pH 7.6. This result is in agreement with previously reported values obtained by other methods under comparable experimental conditions.³⁰

The maximum fluorescence increases observed for the neomycin-B derivatives (**2**, **3**, **5**, and **6**) are similar to that detected for the parent compound (**1**), suggesting the presence of similar binding modes. The corresponding K_d values are shown in Figure 8a. It can be observed that deoxy derivatives **5** and **6** present a 10-fold reduction in affinity with respect to the natural compound **1**. This result is not unexpected as OH5_{III} is involved in a hydrogen bond with G1491 in the paromomycin/A-site complex. Therefore, the ΔG contribution of this hydrogen bond can be estimated as ~1.3 kcal/mol. The observed affinities for the macrocyclic derivatives **2** and **3** are in the same range of those observed for the corresponding open deoxy-analogues **5** and **6**, respectively. According to these data, under the employed experimental conditions, the ΔG contribution resulting from the conformational restriction is almost negligible.^{31,32}

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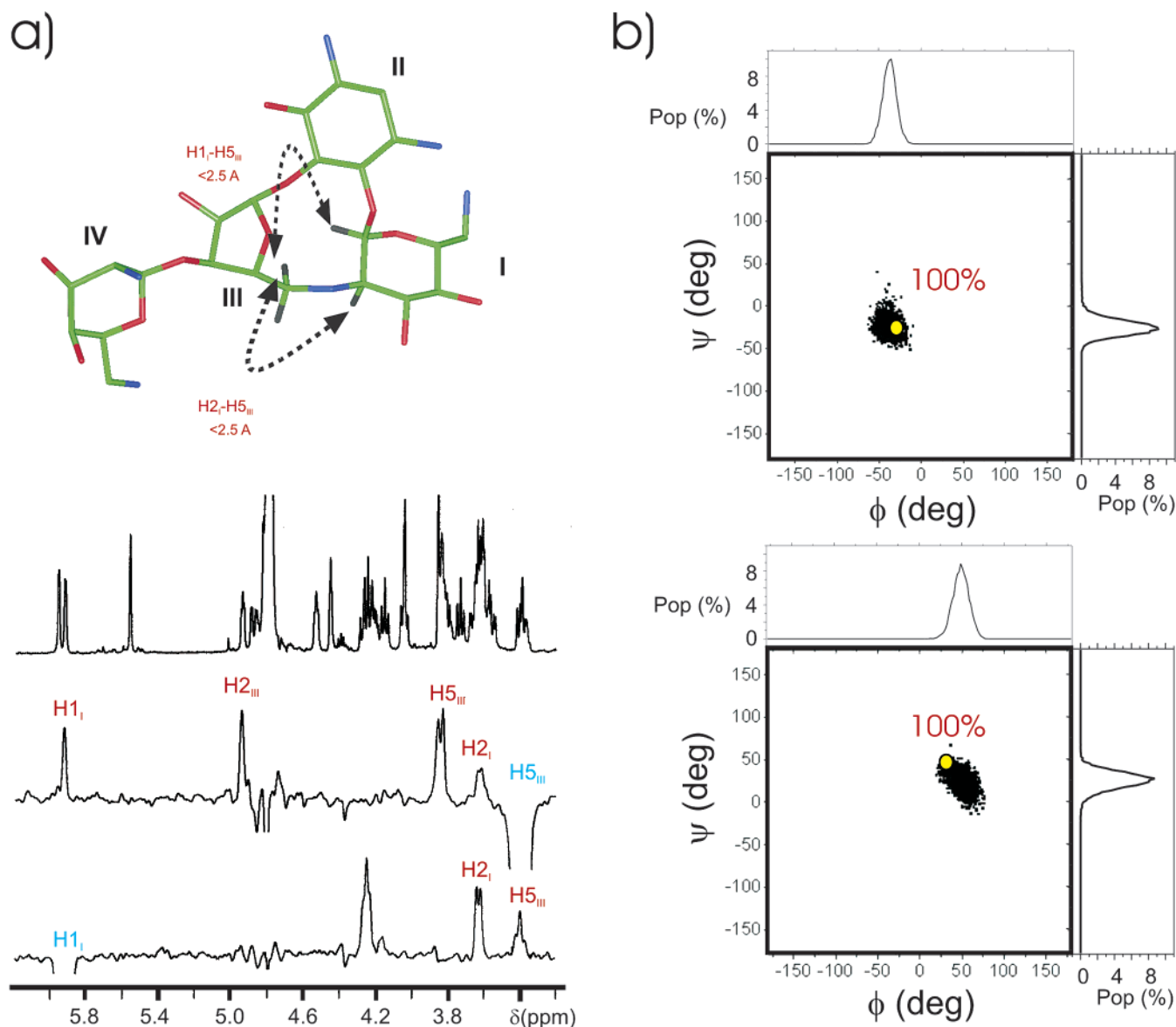


Figure 1. (a) Selective 1D NOE experiments with the 1D-DPFGSE NOE pulse sequence, corresponding to the inversion of H5_{III} and H1_I in compound **2**. Key NOEs probing the spatial proximity of units I and III are highlighted in the upper part of the figure. The numbering employed for the different sugar units is also indicated. (b) MD-tar Φ/Ψ distributions obtained for the I/II (upper panel) and III/II (lower panel) glycosidic linkages. The values corresponding to the paromomycin–RNA bound form are indicated with a yellow dot.

Conformational freedom has been shown to play a key role in the recognition of ligands by receptors.³³ Several attempts have been made to exploit the decrease of conformational entropy of a particular ligand by either locking or biasing conformations toward the bioactive one. In some cases, the

constrained derivatives presented significantly larger affinities for the receptor than the natural ligand.³⁴ In contrast, reported results for carbohydrate–protein interactions range from modest Δ*G* benefits (<0.5 kcal/mol)³⁵ to small penalties^{35,36} (<0.3 kcal/mol). In these cases, the small effect of the conformational constraint on the global Δ*G* has been attributed to an entropy–enthalpy compensation phenomenon.³⁶ Our results indicate that despite the different chemical nature of both the ligand and the receptor, aminoglycoside–RNA interactions exhibit a similar behavior to those previously observed in the recognition of neutral carbohydrates by proteins. Especially striking is the lack

- (31) While this manuscript was in preparation, Tor and co-workers reported the interaction of both the locked derivative **2** and the corresponding paromomycin analogue with the same labeled A-site RNA fragment. Although the buffer conditions employed for the fluorescence measurements differ from ours, the reported decrease in affinity for the locked derivatives is 22–14-fold with respect to the parent aminoglycosides, in agreement with our results; Blount, K. F.; Zhao, F.; Hermann, T.; Tor, Y. *J. Am. Chem. Soc.* **2005**, *127* (27), 9818–9829.
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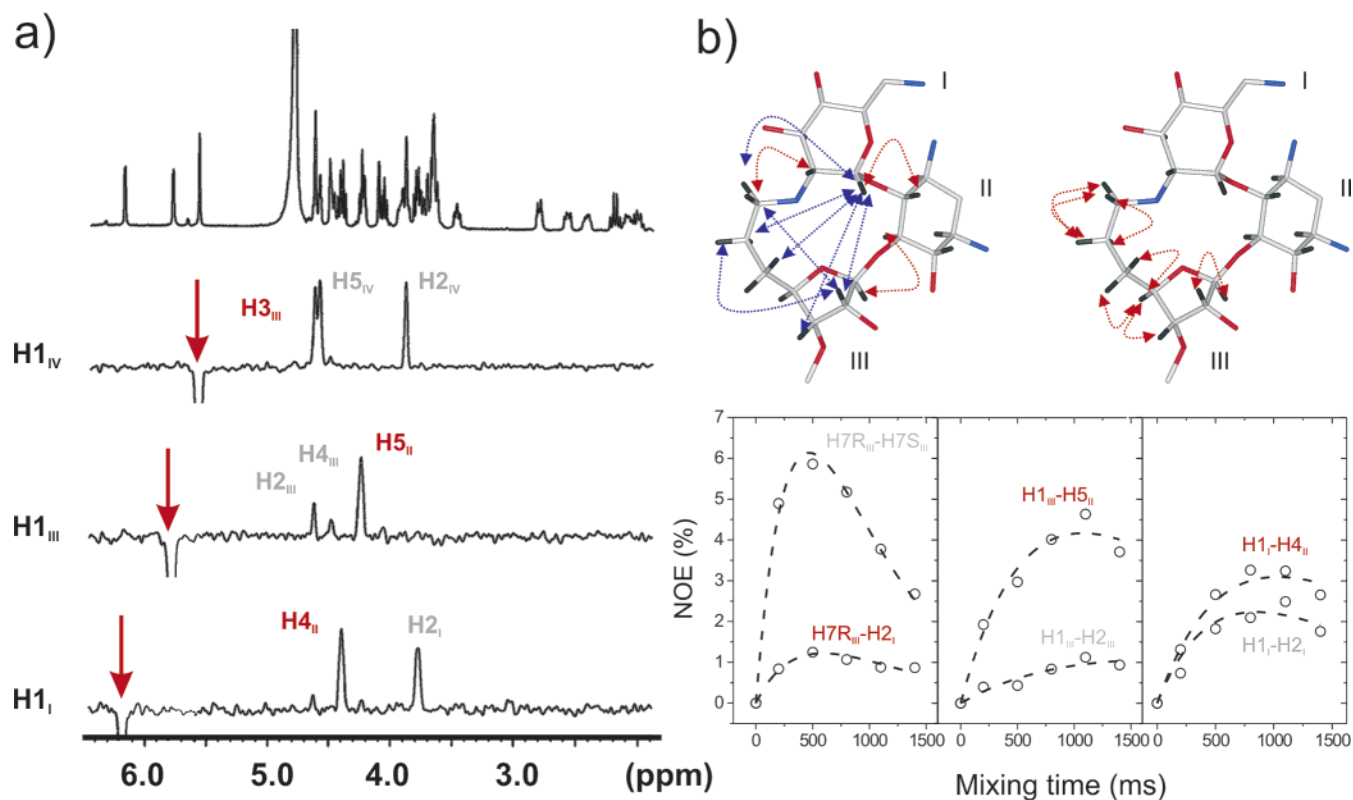


Figure 2. (a) Selective 1D NOE experiments with the 1D-DPGSE NOE pulse sequence, corresponding to the inversion of the anomeric protons in compound **3**. (b) Upper panel: Schematic representation of some of the experimental information employed in the structural analysis of **3**. The upper left panel shows the observed NOEs as red arrows and those not observed as blue arrows. The upper right panel shows the structurally relevant experimental J couplings measured for **3**. The ring IV is omitted for simplicity. Lower panel: some representative NOE buildup curves. NOEs corresponding to known fixed distances, employed as references, are labeled in gray. NOEs corresponding to unknown distances are labeled in red.

of effect that preorganization of the unit III furanose has on the binding strength. Close inspection of molecular models shows that the global shape of the antibiotic is extremely dependent on the precise puckering of this ring. In fact, the specific drug–RNA contacts observed in the paromomycin–RNA complex,^{5,6} involving units III and IV and presumably required for tight binding, are only feasible when the furanose adopts a puckering in the “South” region of the pseudorotational path, which is the minor one in the free state. Nevertheless, the original NMR studies by Puglisi and co-workers⁷ showed that, for paromomycin, these rings are dynamic in the ribosome-bound state, and therefore the contacts observed in the crystal structure of the complex might be transient. In addition, the Westhof and Hanessian groups have recently described, for a paromomycin derivative, a new binding mode to the A-site, for which the ribose ring (unit III) is recognized by the RNA receptor in a C2-endo conformation.³⁷

Finally, for compound **4**, the maximum increase in fluorescence observed upon complex formation is less than half the value observed for neomycin-B, suggesting a smaller structural change in the receptor upon complex formation. It is known that, in the paromomycin–A-site complex, OH5_{III} is located in a strongly electronegative region of the RNA binding pocket. Probably, the substitution of this OH group by a negatively charged carboxylate precludes a deep penetration of the ligand in the major groove. However, despite this modification, the observed affinity for **4** is in the micromolar range.

Effect of the Conformational Constraint on the Antibiotic Activity. The biological activity of the locked neomycin-B derivatives **2–4** was tested against *E. coli* (see Table 2). For derivative **2**, a more extensive testing was performed employing both Gram+ and Gram– bacteria. Despite the observed decrease in activity with respect to the natural compound **1** (3–20-fold depending on the bacteria and the compound), derivatives **2** and **3** are still significantly active (with MIC values in the 2.5–20 $\mu\text{g/mL}$ range). In contrast, derivative **4** showed a poor antibiotic activity. This suggests that **4** is adversely affected by other factors such as a different binding mode (as suggested by fluorescence experiments above), different kinetics for crossing the bacterial membrane, digestion or modification by bacterial enzymes, etc. With respect to **2** and **3**, the obtained results show a good correlation with the binding affinities previously measured for ribosomal RNA and indicate that these locked derivatives might potentially be employed as antibiotics. Therefore, the inactivation of these analogues by aminoglycoside modifying enzymes was tested.

Effect of the Conformational Constraint on Aminoglycoside Inactivation by Bacterial Defense Proteins. To analyze the effect that conformational restriction has on the antibiotic enzymatic inactivation, three different enzymes were chosen as model systems: *Staphylococcus aureus* ANT(4'), *Mycobacterium tuberculosis* AAC(2'), and *Enterococcus faecalis* APH(3'). These proteins are representative of the three main families of enzymes that modify aminoglycosides: adenylyltransferases, acyltransferases, and phosphotransferases. In addition, there is high-resolution X-ray structural information available

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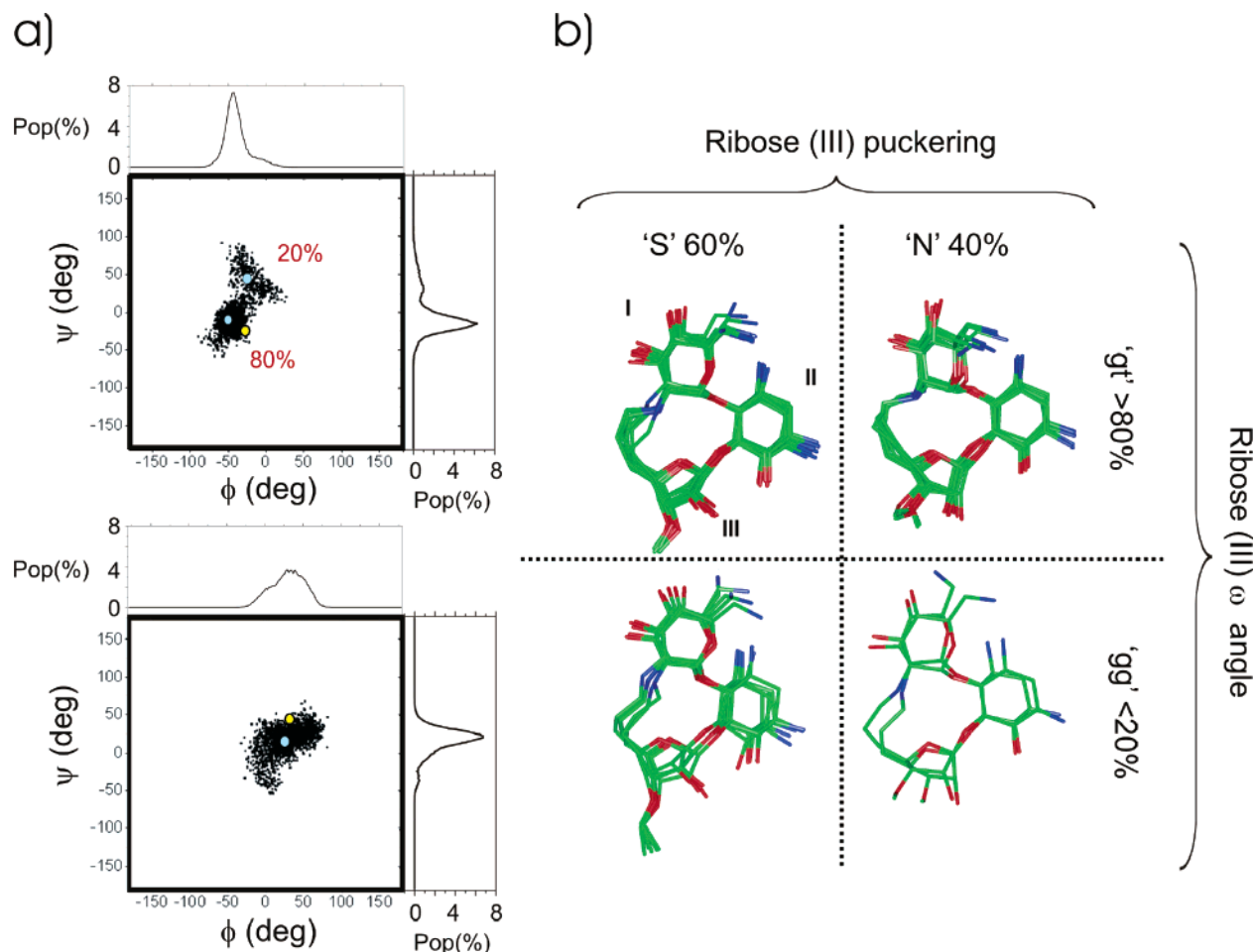


Figure 3. (a) MD-tar Φ/Ψ distributions obtained for the I/II (upper panel) and III/II (lower panel) glycosidic linkages in compound **3**. Minimum energy geometries are indicated with a cyan dot. The Φ/Ψ values observed for the ligand in the paromomycin-RNA complex are indicated with a yellow dot. (b) Representation of the major conformational populations detected in solution for **3**. Unit IV is omitted for simplicity.

for the three enzymes in complex with several antibiotics. Analysis of these data permits the deduction of either minor (as in APH(3')) or major (as in ANT(4') or AAC(2')) conformational differences, for the aminoglycoside, in the protein-bound state with respect to the RNA-bound state (see Scheme 1).^{15,17,19}

Staphylococcus aureus ANT(4') catalyzes the transfer of one adenylyl group from ATP to position O-4 (see Scheme 1) of ring I. This transfer leads to a sharp decrease in the drug affinity for its target RNA. The 3D structure of ANT(4') complexed to kanamycin has been determined by using X-ray.¹⁵ In addition, the structures of related analogues, such as gentamycin, tobramycin, and paromomycin bound to the target ribosomal RNA have also been determined recently.^{5,6} Interestingly, the oligosaccharide conformation recognized by the RNA and by the enzyme is remarkably different, as previously mentioned (see Scheme 1). If it is assumed that some degree of conformational distortion at the I/II fragment is in fact required for enzymatic activity, the conformational restriction introduced in derivatives **2** and **3** might provide a certain degree of protection against the enzymatic inactivation of the antibiotic.

To test this hypothesis, enzymatic reactions were monitored in NMR tubes. ATP (100 μ M–3 mM) and the aminoglycosides (100 μ M–1.5 mM) were dissolved in phosphate buffer (10 mM phosphate, 2.5–5.0 mM MgCl_2 , pH 7.0). After addition of the enzyme (0.2–1.0 μ M), the evolution of the reaction mixture at

310 K was monitored by 1D NMR, and the final products were characterized by 2D NMR and MALDI-TOF MS. As shown in Figure 9, at 100 μ M neomycin-B, 100 μ M ATP, 2.5 mM MgCl_2 , and 0.2 μ M ANT(4'), the adenylation reaction is completed within 22 min. NMR analysis permitted us to confirm that ANT(4') selectively modifies OH_{4I}. In fact, if only 1 equiv of ATP is employed, this is the only product detected. Interestingly, if 2 ATP equiv are used, nonselective secondary adenylation are detected but at much longer reaction times. For example, at 3 mM ATP, 1.5 mM neomycin-B, 5 mM MgCl_2 (pH 7.0, 310 K), and 1 μ M ANT(4'), the primary adenylation is completed in 2 min. In contrast, the enzyme requires more than 12 h to consume the second ATP equivalent, once the primary adenylation is completed. The presence of diadenylated neomycin-B derivatives were confirmed by MALDI-TOF.

For the rigid analogue **2**, at 100 μ M aminoglycoside, 100 μ M ATP, 2.5 mM MgCl_2 , and 0.2 μ M ANT(4'), no reaction was detected even 24 h after enzyme addition (see Figure 9). At larger ANT(4') concentrations (up to 1 μ M ANT(4')), slow, nonselective adenylation processes (similar to those described above for **1**) were observed. In contrast, the related deoxy derivative **5** reacted smoothly to give the monoadenylated compound within a few minutes, which demonstrates that the behavior exhibited by **2** has its origin in the extra rigidity of the antibiotic and that it is not related to the absence of OH_{5III}. Therefore, the simple modification leading to conformationally

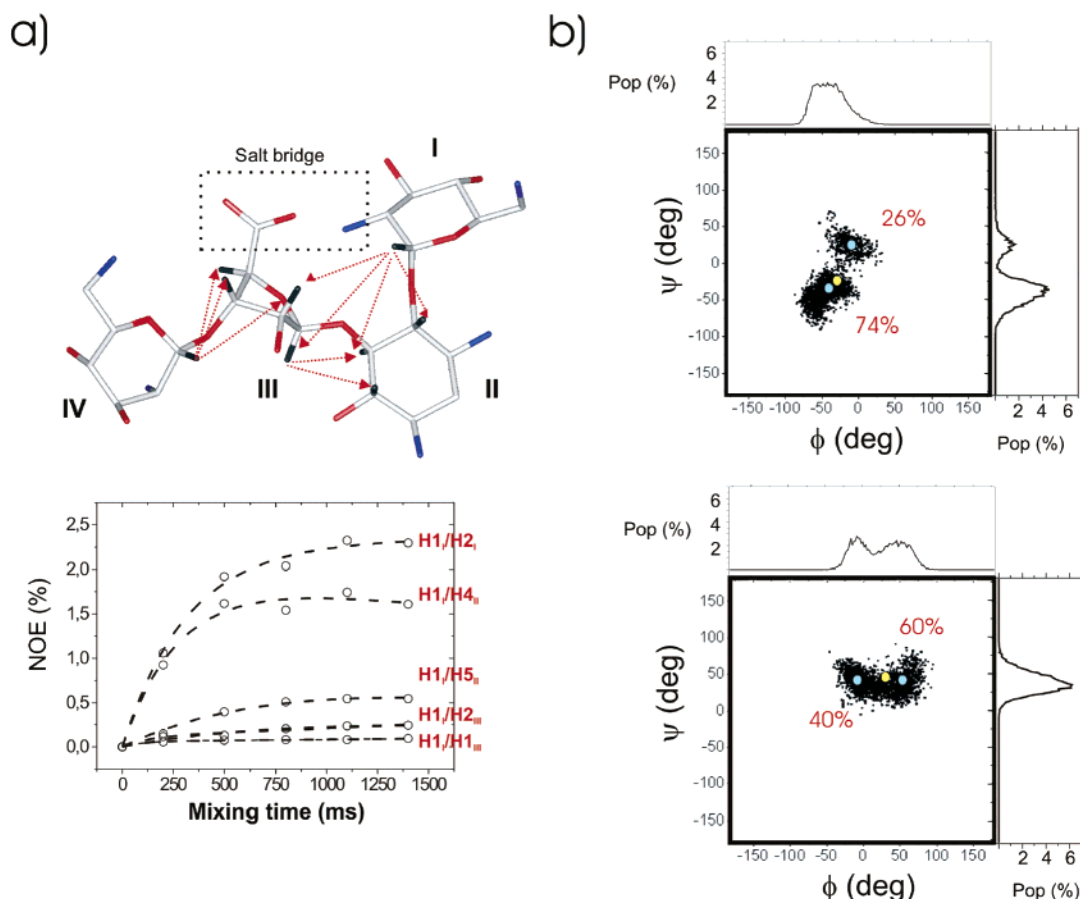


Figure 4. (a) Upper panel: Schematic representation of some of the observed NOEs employed in the structural analysis of **4**. The salt bridge interaction between units III and I is highlighted with a dotted box. Lower panel: Representative NOE build-up curves. (b) MD-tar Φ/Ψ distributions obtained for the I/II (upper) and III/II (lower) glycosidic linkages. Minimum energy geometries are indicated with a cyan dot. The Φ/Ψ values corresponding to the paromomycin–RNA bound form are indicated with a yellow dot.

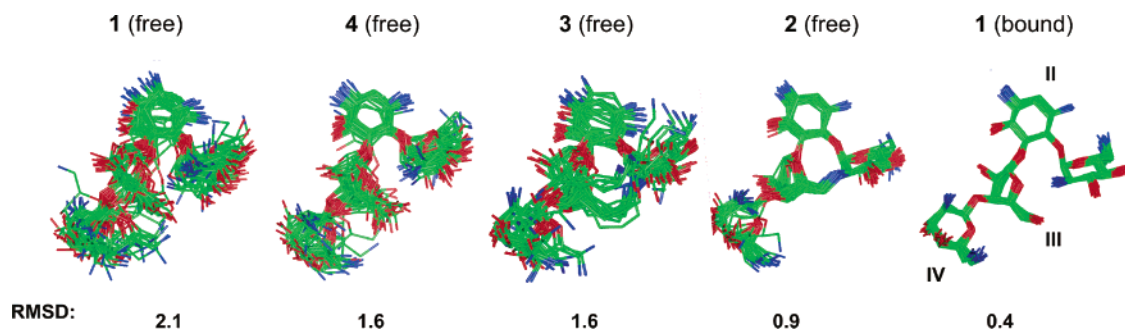


Figure 5. MD-tar ensembles of conformers for **2–4**. The experimental ensemble corresponding to **1** in the free state (left) and that obtained from MD simulations of the neomycin–RNA complex¹⁶ (right) are shown for comparison. RMSDs (Å) for heavy atom superimposition are given below. The numbering employed for the different sugar units is shown on the right.

restricted **2** provides an effective protection against aminoglycoside inactivation by *S. aureus* ANT(4').

Surprisingly, the locked analogue **3** exhibited a similar behavior to that observed for the natural antibiotic, neomycin-B. Indeed, the specific activities of ANT(4') for aminoglycosides **1** and **3** are almost identical (see Figure 9). According to these data, it seems that ANT(4') is able to recognize and inactivate aminoglycoside conformations different from the anti- Ψ geometry observed in the enzyme–kanamycin complex¹⁵ by X-ray methods. In fact, this conformation is not accessible for derivative **3**. In any case, these results prove that some degree of conformational flexibility in the ligand is critical for

enzymatic activity. For ANT(4') a strong conformational restriction (as present in **2**) prevents the antibiotic enzymatic adenylation, although the limited flexibility present in **3** restores the enzymatic activity.

In vivo activities of neomycin-B (**1**) and mimic **2** were tested employing the bacteria *E. coli* DH5 α (pBBR1MCS-2), which expresses the resistance enzyme ANT(4'). As expected, the MIC value for the natural antibiotic is significantly increased (from 3 to 60 $\mu\text{g/mL}$). In contrast, the cyclic derivative **2** maintains the same activity observed for the nonresistant bacteria (20 $\mu\text{g/mL}$). In conclusion, this simple modification leading to the conformationally restricted **2** provides an effective protection

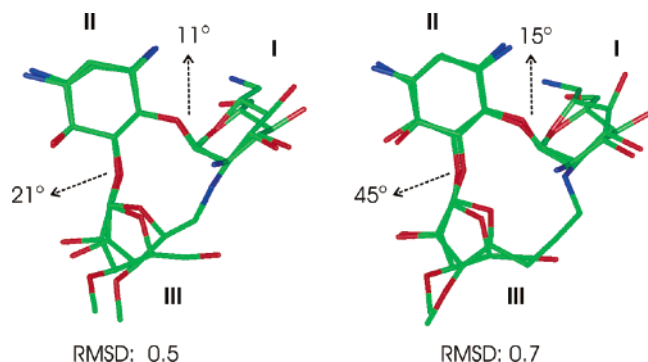


Figure 6. Structures representative of the main conformational families present in solution for neomycin-B derivatives **2** (left) and **3** (right) superimposed on the X-ray structure of paromomycin in the complex with ribosomal RNA, according to X-ray data. The maximum Φ/Ψ deviations (for each glycosidic linkage) and RMSD (Å) values for heavy atom superimpositions are shown. Unit IV is omitted for simplicity.

against aminoglycoside inactivation by *S. aureus* ANT(4'), both in vivo and in vitro, while maintaining a significant antibiotic activity.

Mycobacterium tuberculosis AAC(2') catalyzes the transfer of one acetyl group from Acetyl CoA to position N-2 (see Scheme 1) of the ring I. The 3D structure of AAC(2') complexed to ribostamycin has been recently determined by X-ray.¹⁷ As previously mentioned, the oligosaccharide conformations bound by the RNA and by the enzyme are remarkably different (see Scheme 1). Interestingly, the recognition of a high energy conformation of the antibiotic seems essential for the enzymatic activity. Thus, by forcing an anti- Ψ geometry around the III/II linkage, the protein increases the accessibility of the reacting NH₂I group which, otherwise, would be hydrogen-bonded to the nonvicinal unit III in the low energy syn- Ψ minimum. According to the conformational analysis described above, for cyclic derivatives **2** and **3**, this conformational change is no longer possible. Therefore, the geometrical restriction introduced in these derivatives is expected to fully protect the antibiotics from the enzymatic acetylation. To test this hypothesis, enzymatic reactions were performed in NMR tubes employing similar conditions as those previously described for ANT(4') (see Materials and Methods). As shown in Figure 10, at 100 μ M neomycin-B, 100 μ M AcCoA, 2.5 mM MgCl₂, and 0.2 μ M AAC(2'), the acetylation reaction is completed within 1 h. A similar behavior was observed for the deoxy derivatives **5** and **6**. In contrast, no reaction was observed for the locked aminoglycosides **2** and **3**, even after 24 h and higher enzyme

Table 2. Experimental MIC Values (μ g/mL) Measured for Neomycin-B and Its Conformationally Constrained Analogues **2**–**4**

	MICs (μ g/mL)			
	1	2	3	4
<i>E. coli</i> (DH5 α)	3	20	9	> 100
<i>E. coli</i> (BL21)	0.50	10	2.5	
<i>S. epidermis</i>	2	10		
<i>Bacillus cereus</i>	1	5		
<i>Alcaligenes faecalis</i>	1	20		

concentrations (0.2–20 μ M). Therefore, the conformational constraint introduced in **2** and **3** provides complete protection against aminoglycoside modification by AAC(2').

The in vivo activities of neomycin-B (**1**) and mimic **3** were tested employing the bacteria *E. coli* BL21(DE3)(pET23a-AAC(2')), which expresses the resistance enzyme AAC(2'). The MIC values measured for **1** and **3** were 2 and 2.5 μ g/mL, respectively. As expected, **3** is not affected by the expression of AAC(2'), and this locked aminoglycoside exhibits a good antibiotic activity. However, under the employed experimental conditions, the AAC(2') enzymatic activity seems to have a low influence on the antibiotic properties of the natural compound **1**. Although merely speculative, acetylation at position 2 of ring I might have a small influence on the stability of the drug/A-site complex. In fact, according to the structural information available,^{5–7} the amino group located at this position of the antibiotic is not involved in any key contact with the RNA target, and therefore acetylated **1** might retain a significant activity.

Finally, the effect of neomycin-B flexibility on its enzymatic inactivation by *Enterococcus faecalis* APH(3') was tested. This enzyme catalyzes the transfer of a phosphate group from ATP to position 4 (see Scheme 1) of ring I. In this case, the X-ray structure of the enzyme in complex with neomycin-B shows that the antibiotic structure recognized by the enzyme and the target RNA are remarkably similar (see Scheme 1). In fact, they mainly differ in the conformation adopted by the furanose ring of III (C3-endo and close to C2-endo for the RNA- and enzyme-bound conformations, respectively).

As shown in Figure 11, at 100 μ M neomycin-B, 100 μ M ATP, 2.5 mM MgCl₂, and 0.2 μ M APH(3'), a 50% antibiotic transformation is achieved in 17 min, and the reaction is almost complete in 40 min. Under identical conditions, the enzyme needs 35 min to modify 50% of analogue **3**. For the more rigid derivative **2**, the reaction is even slower (around 100 min for a 50% transformation). In conclusion, although the extra rigidity

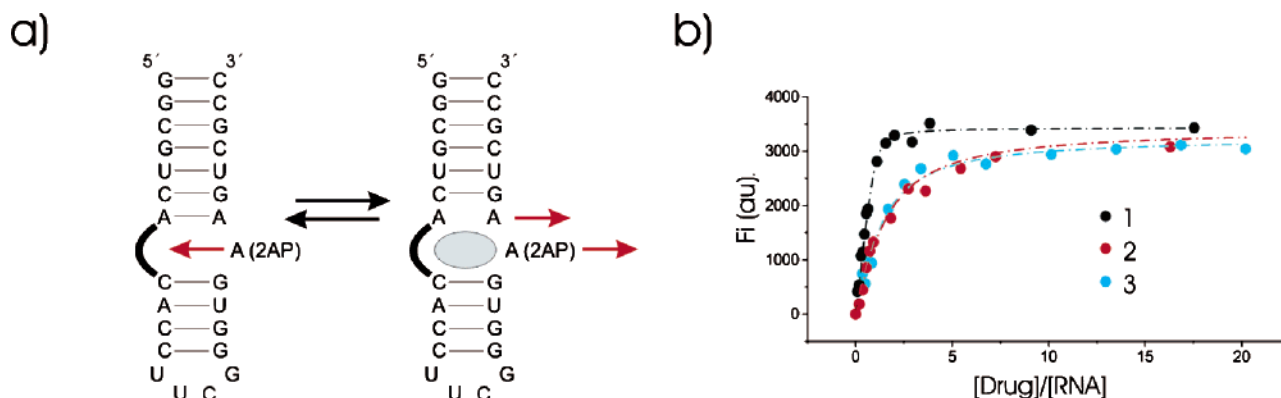


Figure 7. (a) Schematic representation of the modified RNA fragment employed in our binding studies and the structural change induced by aminoglycoside binding. (b) Some representative binding curves obtained for derivatives **1** (black), **2** (red) and **3** (cyan) under identical conditions.

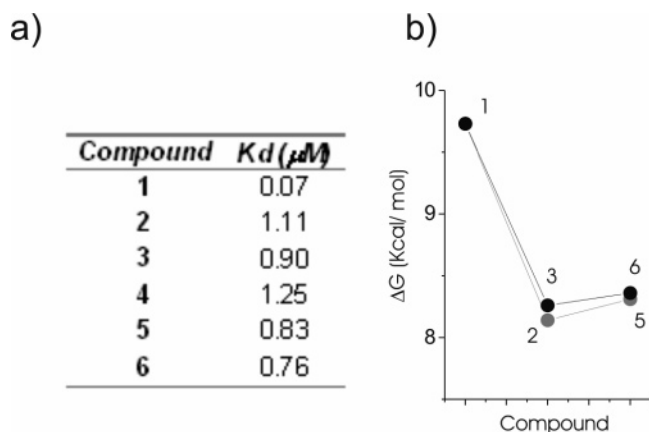


Figure 8. (a) K_d values obtained for aminoglycosides 1–6. (b) Schematic representation of the ΔG values measured for the binding of aminoglycosides 1, 2, 3, 5 and 6 to the A-site.

of the locked derivatives provides a certain protection against modification by APH(3'), in this case, it is insufficient to prevent the enzymatic inactivation of the drugs. This result is not totally unexpected, given the close similarity of the aminoglycoside structures in the binding pockets of both receptors (the A-site RNA and the enzyme).^{6,7,19}

The in vivo activities of neomycin-B (1) and mimic 2 were tested employing the bacteria *E. coli* DH5 α (pACYC177), which expresses the resistance enzyme APH(3'). The MIC values for the natural antibiotic and cyclic derivatives 2 and 3 were in all cases $>150 \mu\text{g/mL}$. Therefore, in this case, although the extra rigidity present in compound 2 provides a certain in vitro protection against the antibiotic enzymatic inactivation, this is clearly insufficient to warrant a good biological activity.

Conclusions

We have shown that the conformational differences exhibited by aminoglycosides within the binding pockets of the ribosome and of those enzymes involved in bacterial resistance can, in certain favorable cases, be exploited for designing new antibiotic derivatives not susceptible to enzymatic inactivation. Thus, the

neomycin-B locked derivatives 2 and 3 have been specifically designed to mimic the bioactive RNA-bound structure of the drug. In fact, both antibiotics maintain a reasonable affinity for the target A-site RNA and exhibit significant biological activities. For compound 2, the conformational constraint provides full protection against the aminoglycoside modification by *Staphylococcus aureus* ANT(4') and *Mycobacterium tuberculosis* AAC(2') and partially protects against modification by *Enterococcus faecalis* APH(3'). In fact, 2 presents an improved biological activity with respect to neomycin-B in bacteria expressing *Staphylococcus aureus* ANT(4'). In addition, derivative 3 is not susceptible to modification by *Mycobacterium tuberculosis* AAC(2'). However, for derivatives 2 and 3, the price paid in terms of RNA binding affinity is not negligible (around 1.6 kcal/mol in ΔG). Clearly, more structure activity studies are needed in the future to address this problem. Interestingly, our RNA binding studies show that for these compounds the observed decrease in affinity has its origin, mainly, in the loss of the OH at position 5 of ring III. In contrast, the conformational restriction does not seem to be critical. According to X-ray studies OH5_{III} is involved in a hydrogen bond with G1491 in the neomycin-B/ribosome complex. Our results indicate that this particular interaction plays a significant role in the stabilization of the complex. Thus by participating in a direct ligand–RNA interaction the OH5_{III} might help to maintain an appropriate orientation of units III and IV within the RNA binding pocket.³² Therefore, efforts should be made in future to optimize the ligand–RNA contacts established by rings III and IV while maintaining the conformational restriction.

In our opinion, this example represents a test case for the validity of a structure-based approach for the design and preparation of ligands that specifically interact with a given receptor and that, in this particular case, might potentially be used as antibiotics. Finally, we would like to stress that the utility of conformational restriction to prevent the enzymatic inactivation of a particular drug while preserving its biological activity might not be restricted to aminoglycosides and could find further applications within medicinal chemistry.

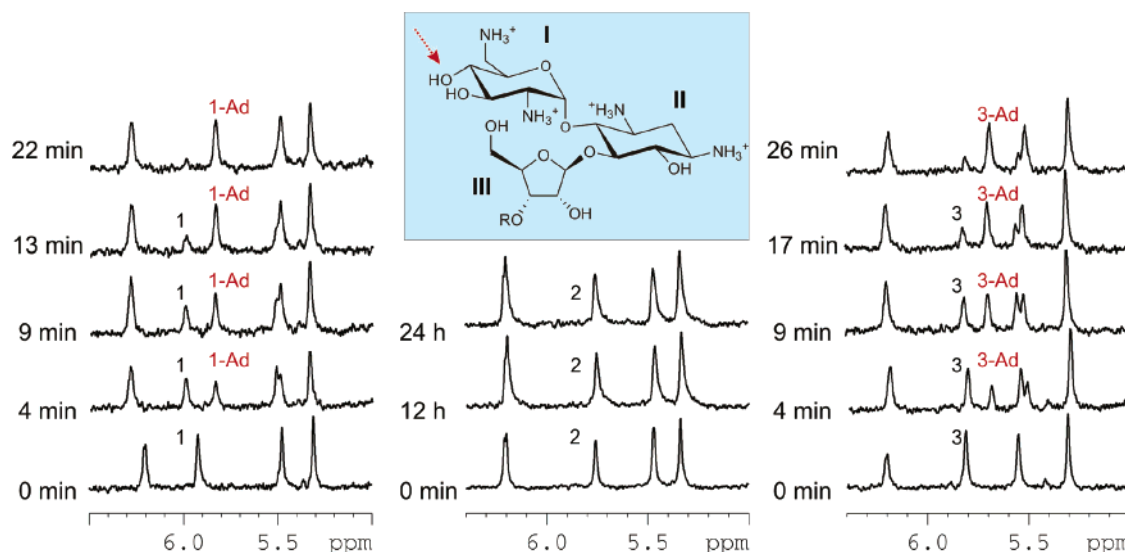


Figure 9. Time evolution (at pH 7.0 and 310 K) of neomycin-B (1, left), derivative 2 (middle), and derivative 3 (right) at 100 μM concentration in the presence of ATP (100 μM), phosphate (10 mM), and MgCl_2 (2.5 mM) after addition of 0.2 μM ANT(4') followed by 1-D NMR. For derivative 2, no reaction was detected. 1-Ad and 3-Ad stand for the adenylated derivatives 1 and 3, respectively. The aminoglycoside position modified by the enzyme is shown schematically in the inset.

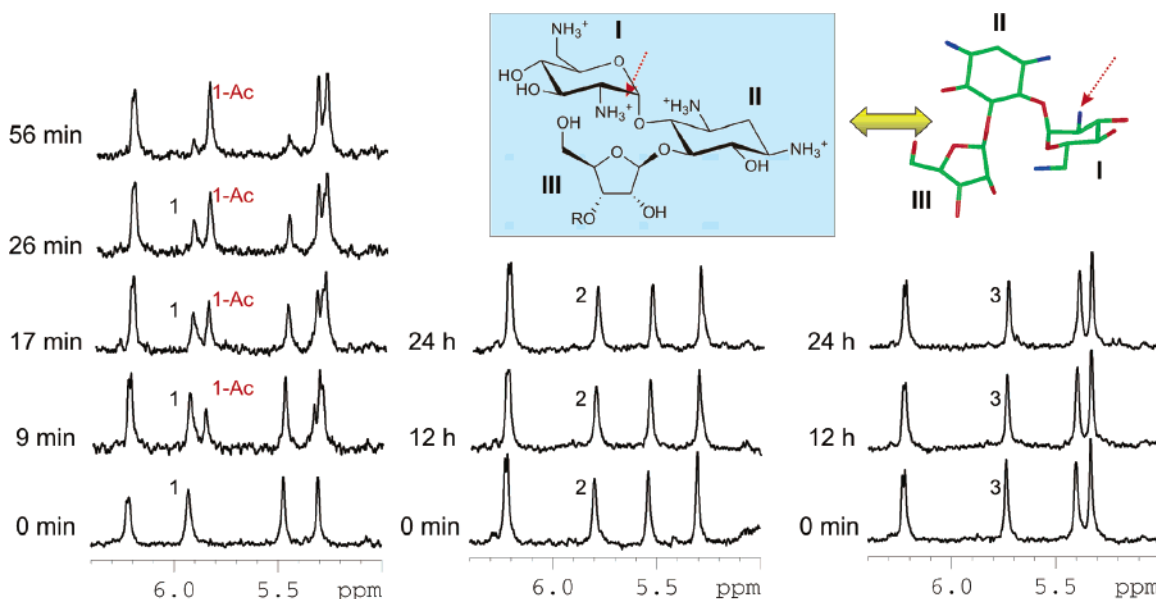


Figure 10. Time evolution (at pH 7.0 and 310 K) of neomycin-B (left), derivative **2** (middle), and derivative **3** (right) at 100 μ M concentration in the presence of AcCoA (100 μ M) and phosphate (10 mM) after addition of 0.2 μ M AAC(2'), followed by 1-D NMR. For derivatives **2** and **3**, no reaction was detected even after 24 h, independently of the concentration of the enzyme employed. **1-Ac** stands for acetylated neomycin-B. The aminoglycoside position modified by the enzyme is shown schematically in the inset. The X-ray structure of ribostamycin in complex with AAC(2') is also represented.

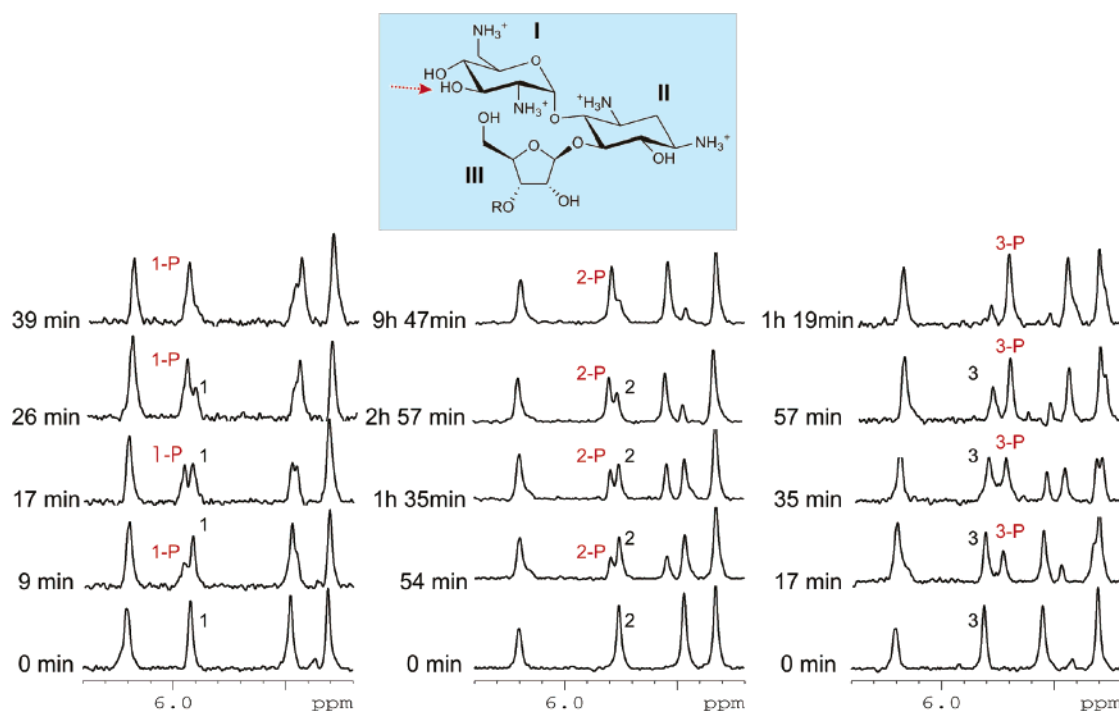


Figure 11. Time evolution (at pH 7.0 and 310 K) of neomycin-B (left), derivative **2** (middle), and derivative **3** (right) at 100 μ M concentration in the presence of ATP (100 μ M), phosphate (10 mM), and MgCl_2 (2.5 mM) after addition of 0.2 μ M APH(3') followed by NMR. **1-P**, **2-P**, and **3-P** stand for the modified antibiotics **1**, **2** and **3**, respectively. The aminoglycoside position modified by the enzyme is shown schematically in the inset.

Materials and Methods

Synthesis of the Aminoglycoside Derivatives. Compound 8. Compound **7** (500 mg, 0.34 mmol) was dissolved in pyridine (10 mL), and 2,4,6-triisopropylbenzene sulfonyl chloride (3.2 g, 10.6 mmol) was added under argon at rt. After stirring for 72 h, the mixture was diluted with EtOAc (300 mL) and H_2O (100 mL) and then neutralized with 1 N HCl. The aqueous phase was extracted with EtOAc (2×100 mL), and the combined organic layers were washed with brine (50 mL), dried over Na_2SO_4 , and evaporated at reduced pressure; the crude was purified by column chromatography on silica gel ($\text{CH}_2\text{Cl}_2/\text{MeOH}$, 20:1) to give **8** (250 mg, 44%). $R_f = 0.5$ ($\text{CH}_2\text{Cl}_2/\text{MeOH}$ 10:1). ^1H NMR (500.13

MHz, CD_3OD): δ 7.4–7.2 (m, 30H, CH_2Ph), 5.43 (broad s, anomeric proton), 5.24 (broad s, anomeric proton), 5.08 (broad s, anomeric proton), 5.2–5.0 (m, 12H, CH_2Ph), 2.91 (m, 3H, $\text{CH}(\text{CH}_3)_2$), 1.21 (18H, $\text{CH}(\text{CH}_3)_2$). ^{13}C NMR (75.47 MHz, CD_3OD): 130–129 (CH_2Ph), 111.0 (anomeric carbon), 99.8 (anomeric carbon), 99.0 (anomeric carbon), 69–68, (CH_2Ph), 35.5 ($\text{CH}(\text{CH}_3)_2$), 35.4 ($\text{C}_{2\text{H}}$), 25.4 ($\text{CH}(\text{CH}_3)_2$), 81.8, 81.1, 71.8, 71.2, 69.3, 57.7, 54.2, 52.8, 52.6, 43.3, 42.8. MALDI-TOF m/z 1708.3 [$\text{M} + \text{Na}^+$].

Compound 9. Palladium on charcoal (1:5 catalyst/substrate by weight) was added to a solution of **8** (50 mg, 0.030 mmol) and trifluoroacetic acid (20 μL , 0.26 mmol) in MeOH (3 mL). After the

suspension was stirred under H_2 (1 atm), at rt for 8 h, the catalyst was removed by filtration, and the solvent was evaporated to give **9** (24 mg, 90%), which was used in the next step without further purification. $R_f = 0.21$ ($NH_4OH/CHCl_3/n-BuOH/EtOH$ 3:1:2:2.5). 1H NMR (500.13 MHz, D_2O): δ 5.55 (broad s, 1H, H_{1I}), 5.25 (broad s, 1H, H_{1III}), 5.01 (broad s, 1H, H_{1IV}), 1.90 (m, 1H, H_{2II} equatorial), 1.15 (m, 1H, H_{2II} axial). MALDI-TOF m/z 903.7 [$M + Na^+$].

Compound 2. A diluted solution of **9** (50 mg, 0.056 mmol) in H_2O (100 mL, pH 7.0) was stirred at 60 °C for 5 days. The solvent was removed at reduced pressure, and the crude was purified by ion exchange column chromatography (0.5 cm \times 7 cm) with Amberlite CG-50 (0–7.5% NH_4OH) to give, after lyophilization, compound **2** (18 mg, 56%). $R_f = 0.16$ ($NH_4OH/CHCl_3/n-BuOH/EtOH$ 3:1:2:2.5). 1H NMR (500.13 MHz, D_2O): δ 5.75 (d, $J = 3.9$ Hz, 1H, H_{1III}), 5.20 (d, $J = 3.5$ Hz, 1H, H_{1I}), 5.03 (broad s, 1H, H_{1IV}), 2.72 (1H, H_{5III}), 2.55 (1H, H_{2I}), 1.95 (m, 1H, H_{2II} equatorial), 1.16 (m, 1H, H_{2II} axial). ^{13}C NMR (75.47 MHz, D_2O): δ 100.3 (C_{1IV}), 109.5 (C_{1III}), 98.2 (C_{1I}), 60.7 (C_{2I}), 48.4 (C_{5III}). MALDI-TOF m/z 619.1 [$M + Na^+$].

Compound 10. To a solution of compound **7** (1.0 g, 0.7 mmol) in dry THF (7 mL) at 0 °C under argon was added 2,6-lutidine (0.24 mL, 2.1 mmol) and TIPSOTf (0.28 mL, 1.05 mmol). The mixture was allowed to warm to rt and stirred for 1 h. The solvent was evaporated, and the crude was purified by flash chromatography ($CH_2Cl_2/MeOH$, 15:1) to give the silyl ether derivative (0.88 g, 80%). $R_f = 0.4$ ($CH_2Cl_2/MeOH$ 10:1). To a solution of this derivative (1.0 g, 0.63 mmol) in pyridine (15 mL) was added at rt Ac_2O (7 mL). After the mixture was stirred for 5 h at rt, the solvent was removed at reduced pressure, and the crude was purified by column chromatography on silica gel ($EtOAc/hexane$, 1:1) to give **10** (1.02 g, 100%). $R_f = 0.6$ ($EtOAc/hexane$ 2:1). 1H NMR (500.13 MHz, CD_3OD): δ 7.4–7.2 (m, 30H, CH_2Ph), 5.2–5.0 (12H, CH_2Ph), 2.1–1.8 (18H, CH_3CO_2), 1.97 (m, 1H, H_{2II} equatorial), 1.36 (m, 1H, H_{2II} axial), 1.32 (m, 3H, $SiCH(CH_3)_2$), 1.10 (18H, $SiCH(CH_3)_2$). ^{13}C NMR (75.47 MHz, CD_3OD): 130–129 (CH_2Ph), 69–68, (CH_2Ph), 35.4 (C_{2II}), 21–20 (CH_3CO_2), 18.5 ($SiCH(CH_3)_2$). MALDI-TOF m/z 1850.2 [$M + Na^+$].

Compound 11. To a solution of the silyl ether **10** (0.5 g, 0.28 mmol) in dry THF (5 mL), in a polyethylene vessel at 0 °C, was added HF/pyridine (0.3 mL). After stirring at rt for 5 h, the mixture was diluted with Et_2O and neutralized with aqueous saturated $NaHCO_3$. The combined organic layer was dried, evaporated, and purified by flash chromatography ($EtOAc/hexane$, 1:1) to give alcohol **11** (0.42 g, 90%). $R_f = 0.3$ ($AcOEt/hexane$ 2:1). 1H NMR (500.13 MHz, CD_3OD): δ 7.4–7.2 (m, 30H, CH_2Ph), 5.81 (broad s, anomeric proton), 5.72 (broad s, anomeric proton), 5.18 (broad s, anomeric proton), 5.2–4.9 (m, 12H, CH_2Ph), 2.2–1.9 (18H, CH_3CO_2), 1.95 (m, 1H, H_{2II} equatorial), 1.64 (m, 1H, H_{2II} axial). ^{13}C NMR (75.47 MHz, CD_3OD): 130–129 (CH_2Ph), 110.5 (anomeric carbon), 98.2 (anomeric carbon), 68–67, (CH_2Ph), 34.4 (C_{2II}), 21–20 (CH_3CO_2), 84.1, 83.4, 76.1, 74.1, 72.4, 69.8, 62.4, 54.9, 51.1, 42.1. MALDI-TOF m/z 1694.6 [$M + Na^+$].

Compound 12. Dess–Martin periodinane (95 mg, 0.26 mmol) was added to a solution of alcohol **11** (250 mg, 0.15 mmol) in CH_2Cl_2 (5 mL). After the mixture was stirred for 2 h at rt, Et_2O was added (20 mL), and the mixture was poured into saturated aqueous $NaHCO_3$ solution (25 mL) and $Na_2S_2O_5$ (100 mg). The organic layer was washed with saturated aqueous $NaHCO_3$ (25 mL) and brine (25 mL) and dried, and the solvent evaporated at reduced pressure to give, after purification by flash chromatography ($EtOAc/hexane$, 2:1), aldehyde **12** (199 mg, 79%). $R_f = 0.3$ ($AcOEt/hexane$ 2:1). 1H NMR (500.13 MHz, $CDCl_3$): δ 9.4 (broad s, 1H, CHO), 7.4–7.2 (m, 30H, CH_2Ph), 5.2–4.9 (12H, CH_2Ph), 2.1–1.9 (18H, CH_3CO_2), 1.92 (m, 1H, H_{2II} equatorial), 1.41 (m, 1H, H_{2II} axial). ^{13}C NMR (75.47 MHz, $CDCl_3$): 130–129 (CH_2Ph), 68–67, (CH_2Ph), 34.6 (C_{2II}), 21–20 (CH_3CO_2). MALDI-TOF m/z 1691.58 [$M + Na^+$].

Compound 13. To a solution of **12** (250 mg, 0.15 mmol) in dry acetonitrile (6 mL) was added (methoxycarbonylmethylenetriphenylphosphorane (66 mg, 0.19 mmol). After stirring at rt for 3 h, the solvent

was removed at reduced pressure, and the crude was purified by column chromatography on silica gel ($EtOAc/hexane$, 1:1) to give compound **13** (215 mg, 82%). $R_f = 0.3$ ($AcOEt/hexane$ 2:1). 1H NMR (500.13 MHz, $CDCl_3$): δ 7.4–7.2 (m, 30H, CH_2Ph), 7.16 (broad m, 1H), 6.33 (broad s, 1H), 5.2–4.9 (12H, CH_2Ph), 3.90 (s, 3H, CO_2CH_3), 2.1–1.8 (18H, CH_3CO_2), 1.98 (m, 1H, H_{2II} equatorial), 1.40 (m, 1H, H_{2II} axial). ^{13}C NMR (75.47 MHz, $CDCl_3$): 130–129 (CH_2Ph), 68–67 (CH_2Ph), 34.6 (C_{2II}), 21–20 (CH_3CO_2). MALDI-TOF m/z 1748.7 [$M + Na^+$].

Compound 14. To a solution of compound **13** (200 mg, 0.12 mmol) in THF/ $EtOH$ (1:2, 10 mL) at rt was added $NaBH_4$ (54 mg, 1.46 mmol) and LiCl (6 mg, 0.15 mmol). After the mixture was stirred at rt for 12 h, $EtOAc$ (25 mL) was added, and the mixture was washed with aqueous saturated $NaHCO_3$ and brine. The organic phase was dried over Na_2SO_4 , the solvent was removed at reduced pressure, and the crude was purified by flash chromatography ($CH_2Cl_2/MeOH$, 20:1) to give **14** (155 mg, 89%). $R_f = 0.4$ ($CH_2Cl_2/MeOH$ 10:1). 1H NMR (500.13 MHz, CD_3OD): δ 7.4–7.2 (m, 30H, CH_2Ph), 5.16 (broad s, 1H, H_{1I}), 5.04 (broad s, 1H, H_{1III}), 5.2–5.0 (12H, CH_2Ph), 4.78 (broad s, 1H, H_{1IV}), 1.99 (m, 1H, H_{2II} equatorial), 1.62–1.60 (m, 4H, $-CH_2-CH_2-CH_2-OH$), 1.39 (m, 1H, H_{2II} axial). ^{13}C NMR (75.47 MHz, CD_3OD): 130–129 (CH_2Ph), 111.3 (C_{1III}), 101.2 (C_{1I}), 100.9 (C_{1IV}), 69–68, (CH_2Ph), 35.1 (C_{2II}), 30.6 ($-CH_2-CH_2-CH_2-OH$), 29.5 ($-CH_2-CH_2-CH_2-OH$), 86.8, 82.8, 76.4, 74.9, 73.6, 73.4, 72.9, 71.6, 69.4, 63.0, 57.7, 54.2, 52.6, 43.1, 42.5. MALDI-TOF m/z 1469.7 [$M + Na^+$].

Compound 15. Treatment of compound **14** (150 mg, 0.10 mmol) at 0 °C following the same procedure described for the preparation of compound **10** gave the silyl ether **15** (97 mg, 60%). $R_f = 0.4$ ($CH_2Cl_2/MeOH$ 10:1). 1H NMR (500.13 MHz, CD_3OD): δ 7.4–7.2 (m, 30H, CH_2Ph), 5.26 (broad s, 1H, H_{1I}), 5.07 (broad s, 1H, H_{1III}), 5.2–5.0 (12H, CH_2Ph), 4.85 (broad s, 1H, H_{1IV}), 1.96 (m, 1H, H_{2II} equatorial), 1.66–1.63 (m, 4H, $-CH_2-CH_2-CH_2-OTIPS$), 1.38 (m, 1H, H_{2II} axial), 1.04 (18H, $CH(CH_3)_2$). ^{13}C NMR (75.47 MHz, CD_3OD): 130–129 (CH_2Ph), 111.0 (C_{1III}), 100.7 (C_{1IV}), 100.5 (C_{1I}), 69–68, (CH_2Ph), 35.1 (C_{2II}), 31.1 ($-CH_2-CH_2-CH_2-OTIPS$), 26.7 ($-CH_2-CH_2-CH_2-OTIPS$), 18.31 ($CH(CH_3)_2$), 86.7, 82.7, 82.2, 75.8, 74.7, 73.5, 73.1, 72.0, 69.3, 64.8, 57.9, 54.3, 53.3, 52.5, 43.4, 42.5. MALDI-TOF m/z 1626.2 [$M + Na^+$].

Compound 16. Acetylation of **15** (97 mg, 0.060 mmol) employing the same methodology described for **10** gave the fully acetylated derivative in quantitative yield. $R_f = 0.3$ ($CH_2Cl_2/MeOH$ 20:1). From this compound (111 mg, 0.060 mmol), derivative **16** (81 mg, 0.048 mmol) was obtained following a procedure identical to that described for **11**; yield: 79%. $R_f = 0.25$ ($CH_2Cl_2/MeOH$ 20:1). 1H NMR (500.13 MHz, CD_3OD): δ 7.4–7.2 (m, 30H, CH_2Ph), 5.1–4.9 (12H, CH_2Ph), 2.1–1.9 (18H, CH_3CO_2), 1.94 (m, 1H, H_{2II} equatorial), 1.63 (m, 1H, H_{2II} axial). ^{13}C NMR (75.47 MHz, CD_3OD): 130–129 (CH_2Ph), 68–67, (CH_2Ph), 34.5 (C_{2II}), 21–20 (CH_3CO_2). MALDI-TOF m/z 1721.8 [$M + Na^+$].

Compound 17. To a solution of compound **16** (81 mg, 0.048 mmol) in pyridine (2 mL) was added DMAP (1 mg, 0.008 mmol) and *p*-toluenesulfonyl chloride (14 mg, 0.073 mmol). After stirring the mixture, at room temperature under argon for 3 h, the solvent was evaporated and the crude was purified by flash chromatography ($EtOAc/hexane$, 1:1) to give the primary sulfonate (70 mg, 79%). $R_f = 0.4$ ($CH_2Cl_2/MeOH$ 20:1). To a solution of this derivative (70 mg, 0.038 mmol) in $MeOH/THF$ (2:1, 300 μ L) was added KCN (2 mg, 0.031 mmol). After stirring at room temperature for 10 h, the solvent was removed and the crude was purified by column chromatography on silica gel ($CH_2Cl_2/MeOH$, 20:1) to give the deacetylated derivative **17** (36 mg, 58%). $R_f = 0.44$ ($CH_2Cl_2/MeOH$ 10:1). 1H NMR (500.13 MHz, CD_3OD): δ 7.75 (d, $J = 8$ Hz, 2H, O-Ts), 7.36 (d, $J = 8$ Hz, 2H, O-Ts), 7.4–7.2 (m, 30H, CH_2Ph), 5.24 (broad s, 1H, H_{1I}), 5.01 (broad s, 1H, H_{1III}), 5.2–5.0 (12H, CH_2Ph), 4.81 (broad s, 1H, H_{1IV}), 1.97 (m, 1H, H_{2II} equatorial), 1.65–1.60 (m, 4H, $-CH_2-CH_2-CH_2-OTs$), 1.36 (m, 1H, H_{2II} axial). ^{13}C NMR (75.47 MHz, CD_3OD): 130–129 (CH_2Ph), 111.6 (C_{1III}), 100.6 (C_{1IV}), 100.5 (C_{1I}), 69–68, (CH_2Ph),

35.4 (C_{2II}), 30.8 (−CH₂−CH₂−CH₂−OTs), 26.5(−CH₂−CH₂−CH₂−OTs), 86.8, 82.5, 81.9, 76.3, 73.7, 73.3, 72.2, 69.5, 68.4, 57.8, 54.6, 52.8, 52.5, 43.5, 42.6. MALDI-TOF *m/z* 1623.3 [M + Na⁺].

Compound 3. Treatment of **17** (12 mg, 0.0075 mmol) under the same conditions described for the preparation of compound **9** gave the fully deprotected tosylate (5 mg, 88%) which was used in the next step without further purification. *R_f* = 0.16 (NH₄OH/CHCl₃/*n*-BuOH/EtOH, 3:1:2:2.5). A diluted solution of this compound (5 mg, 0.0066 mmol) in H₂O (100 mL, pH 7.0) was stirred at 40 °C for 10 days. The solvent was removed at reduced pressure, and the crude was purified by ion exchange column chromatography (0.5 cm × 7 cm) with Amberlite CG-50 (0–7.5% NH₄OH) to give, after lyophilization, compound **3** (3 mg, 62%). *R_f* = 0.16 (NH₄OH/CHCl₃/*n*-BuOH/EtOH 3:1:2:2.5). ¹H NMR (500.13 MHz, D₂O): 5.9 (d, *J* = 3.9 Hz, 1H, H_{1I}), 5.5 (d, *J* = 3.5 Hz, 1H, H_{1III}), 5.2 (d, *J* = 1.5 Hz, 1H, H_{1IV}), 4.3 (1H, H_{2III}), 4.2 (1H, H_{4III}), 3.6 (1H, H_{2IV}), 3.6 (1H, H_{7III}), 3.5 (1H, H_{2I}), 3.1 (1H, H_{7III}), 2.5 (m, 1H, H_{2II} equatorial), 2.3 (1H, H_{5III}), 2.1 (1H, H_{6III}), 1.8 (m, 1H, H_{2II} axial), 1.7 (1H, H_{5III}), 1.7 (1H, H_{6III}). ¹³C NMR (75.47 MHz, D₂O): 108.7 (C_{1III}), 96.4 (C_{1I}), 96.1 (C_{1IV}), 73.2 (C_{2III}), 59.3 (C_{2I}), 50.8 (C_{2IV}), 49.8 (C_{7III}), 31.3 (C_{5III}), 28.2 (C_{2II}), 21.8 (C_{6III}). MALDI-TOF *m/z* 648.1 [M + Na⁺].

Compound 6. Palladium on charcoal (1:5 catalyst/substrate by weight) was added to a solution of **17** (25 mg, 0.016 mmol) and trifluoroacetic acid (20 μL, 0.26 mmol) in MeOH (3 mL). After stirring the suspension under H₂ (1 atm) at room temperature for 48 h, the catalyst was removed by filtration, and the solvent was evaporated at reduced pressure. Derivative **6** (6.3 mg, 0.010 mmol) was purified by ion exchange column chromatography (0.5 cm × 7 cm) with Amberlite CG-50 (0–7.5% NH₄OH); yield: ~60%. *R_f* = 0.24 (NH₄OH/CHCl₃/*n*-BuOH/EtOH, 3:1:2:2.5). ¹H NMR (500.13 MHz, D₂O): δ 5.9 (broad s, 1H, H_{1I}), 5.3 (d, *J* = 3.5 Hz, 1H, H_{1III}), 5.2 (broad s, 1H, H_{1IV}), 4.3 (1H, H_{2III}), 3.5 (1H, H_{2IV}), 3.4 (1H, H_{6I}), 3.4 (1H, H_{6IV}), 3.4 (1H, H_{6IV}), 3.3 (1H, H_{2I}), 3.2 (1H, H_{6I}), 2.3 (m, 1H, H_{2II} equatorial), 1.7 (m, 1H, H_{2II} axial). ¹³C NMR (75.47 MHz, D₂O): 109.7 (C_{1III}), 95.7 (C_{1IV}), 95.1(C_{1I}), 79.4 (C_{2III}), 53.5 (C_{2I}), 50.9 (C_{2IV}), 40.5 (C_{6IV}), 40.2 (C_{6I}), 29.7 (C_{2II}). MALDI-TOF *m/z* 649.8 [M + Na⁺].

NMR and Molecular Modeling of 2–4. RESP atomic charges for neomycin-B analogues **2–4** were derived by applying the RESP module of the AMBER 5.0 package²⁶ to the HF/6-31G(d) ESP charges calculated with Gaussian-94.³⁸ All MD simulations were carried out using the SANDER module within AMBER and the Cornell et al. force field.³⁹ Parameters for the acetalic functions were taken from GLYCAM.⁴⁰

NMR Structure of Derivatives 2–4. NMR experiments were recorded on a Varian Unity 500 and Bruker Avance 500 at 313 K. Selective 1D NOE experiments were recorded employing the 1D-DPGSE NOE pulse sequence. NOE intensities were normalized with respect to the diagonal peak at zero mixing time. Selective T₁ measurements were performed on the anomeric and several other protons to obtain the above-mentioned values. Experimental NOEs were fitted to a double exponential function, $f(t) = p_0(e^{-p_1 t})(1 - e^{-p_2 t})$ with *p*₀, *p*₁, and *p*₂ being adjustable parameters.⁴¹ The initial slope was determined from the first derivative at time *t* = 0, $f'(0) = p_0 p_2$. From the initial slopes, interproton distances were obtained by employing the isolated spin pair approximation.

To obtain a NMR-derived ensemble, MD-tar²⁵ simulations were performed for compounds **2–4** employing a protocol identical to that described by Asensio et al.¹⁶ NOE-derived distances were included as time averaged distance constraints, and scalar coupling constants, as

time averaged *J* coupling constraints. A $\langle r^{-6} \rangle^{-1/6}$ average was used for the distances, and a linear average was used for the coupling constants. For derivatives **2** and **4**, final trajectories were carried out using explicit solvent, counterions, periodic boundary conditions (PBC) and Ewald sums for the treatment of electrostatic interactions. An exponential decay constant of 500 ps and simulation lengths of 5 ns were employed.

For compound **3**, the experimental data could only be quantitatively reproduced after (in vacuo ϵ = 80) 80 ns MD-tar simulations. Preliminary tests showed that, for this flexible molecule, the use of exponential decay constants shorter than 5 ns produced unstable trajectories and led in some cases to severe distortions of the sugar rings. This problem could be solved by increasing the exponential decay constant value to 8–10 ns. It has been estimated that simulation lengths of ca. 1 order of magnitude larger than the exponential decay constant should be used to generate reliable estimates of average properties. Therefore the final simulation length used was 80 ns. We would like to point out that for MD-tar runs the sensitivity of the results to the simulation conditions (solvent, counterions, cutoffs, dielectric constant, etc.) used is extremely dependent on the amount of experimental information included as time-averaged constraints. For derivative **3** we employed 23 distance constraints and 7 *J* values. Under these conditions, the conformational regions sampled by the trajectory (within the esterically allowable regions) are mainly determined by the constraints, and the fine details of the simulation are not critical. In fact similar results were obtained using ϵ = 40 and ϵ = 20.

Finally, to further clarify the conformational properties of **3**, we performed additional calculations. Thus, a geometry representative of each of the five most populated conformational families present for derivative **3** in solution, according to the in vacuo ϵ = 80 MD-tar calculations, was subjected to 5 ns unconstrained MD simulations in the presence of explicit water, counterions, periodic boundary conditions, and Ewald sums for the treatment of electrostatic interactions. The obtained trajectories show that, as deduced from the in vacuo MD-tar simulations, the macrocyclic structure of compound **3** is compatible with wide fluctuations around the glycosidic linkages, different puckerings for ring III and different orientations for the three methylene bridge. In fact, the conformational regions populated by **3** according to these simulations are similar those deduced from the in vacuo 80 ns MD-tar trajectories (Figure S7).

Binding Experiments. The labeled 27mer A-site model was obtained in its polyacrylamide gel electrophoresis- (PAGE-) purified sodium salt form from Dharmacon Research, Inc. (Lafayette, CO). Buffer conditions for the binding experiments were 100 mM NaCl, 10 mM sodium phosphate (pH 7.6). Fluorescence experiments were conducted on a Fluorolog-3 spectrophotometer (Jobin Yvon-Spex Instruments S.A., Inc., New Jersey) equipped with a thermostat. A quartz cell with a 1 cm path length in both the excitation and emission directions was used in all the experiments that were performed at 25 °C. The RNA concentration was, in all cases, 1 μM in strand. For renaturing the RNA, it was placed in a water bath at 20 °C, heated to 85 °C for 1 min and then slowly cooled back to 20 °C over a 2 h period. With the excitation wavelength set to 310 nm, the change in the fluorescence emission at 369 nm upon complex formation was followed in titration experiments. Aliquots of the antibiotic derivatives were added sequentially, and 2 min of equilibration time were allowed before each fluorescence measurement. Fluorescence intensities were corrected for volume changes according to the following equation: $F_{i,corr} = F_{i,obs} * V_i/V_o$ where $F_{i,corr}$ is the corrected intensity for point *i* of the titration, $F_{i,obs}$ is the measured intensity at point *i*, *V_i* is the volume after the *i*th addition and *V_o* is the initial volume (typically 2 mL). The K_d values for simple binding were determined by plotting $F_{i,corr}$ against the ligand to RNA ratio. In all cases, the data were found to fit well assuming a 1:1 complex.

Cloning, Overexpression, and Purification of the Aminoglyside Modifying Enzymes. The plasmid codifying for the AAC(2')-Ic from *Mycobacterium tuberculosis* was kindly provided by J. S. Blanchard

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(Albert Einstein College of Medicine, New York). This plasmid was transformed in the *E. coli* strain BL21(DE3). The expression and purification of *Mycobacterium tuberculosis* AAC(2') was performed as described by J. S. Blanchard and co.⁴²

Enterococcus faecalis APH(3') and *Staphylococcus aureus* ANT(4') were subcloned from the plasmids pACYC177 and pBBR1MCS-2, respectively. PCR amplifications were performed in a 100 μ L reaction mixture containing DNA (3 μ L) of the plasmids as a template, water (70.5 μ L), buffer (100 mM Tris-HCL, 500 mM KCl, pH 8.0, 10 μ L), MgCl₂ (2 mM, 8 μ L), dNTPs (200 μ M), primers (1 μ M), and Taq DNA polymerase (2.5 U). The primers employed for *Enterococcus faecalis* APH(3') were Nt (NdeI) (5'tatacatatgagccatattcaagg3') and Ct (EcoRI)- (5'tatagaattcttagaaaaactcatc3'), and those used for *Staphylococcus aureus* ANT(4') were Nt(Hind III)(5'gccgcaagattctgaatggaccaataata3') and Ct(Xho I) (5'gcctcgagtcaaatggtatcggtt3'). The reaction was subjected to 30 cycles of amplification. The cycle conditions were set as follows: denaturation at 94 °C for 1 min, annealing at 50 °C/1.0 min for *Enterococcus faecalis* APH(3'), and 55 °C/1.0 min for *Staphylococcus aureus* ANT(4') and elongation at 72 °C for 1 min. The genes obtained from the PCR were purified with the Wizard PCR Preps DNA Purification System (Promega) and ligated in pET-28b(+). The pET28b-APH3' and pET28b-ANT4' expression vectors constructed in this way were then transformed into *E. coli* BL21(DE3) competent cells.

Selected clones for *Enterococcus faecalis* APH(3') and *Staphylococcus aureus* ANT(4') were grown on 100 mL of LB medium containing 26 μ g/mL kanamycin at 37 °C. When the cell growth reached an optical density at 600 nm (OD₆₀₀) of 0.5, the temperature was switched to 30 °C and the culture was induced with 0.5 mM IPTG. After 24 h of induction, the expression level was analyzed by SDS-PAGE using gels with 13% polyacrylamide in the separation zone. Proteins were purified employing a Ni²⁺-IDA-agarose column. The enzymes after this step were found to be more than 90% pure as assessed by SDS-polyacrylamide gel electrophoresis.

Enzymatic Activity. Enzymatic reactions were performed in NMR tubes. The nucleoside triphosphate (ATP 100 μ M–3.0 mM) and the aminoglycoside (100 μ M–1.5 mM) were dissolved in 10 mM phosphate buffer, pH 7.0, 2.5–5.0 mM MgCl₂. After addition of the enzyme (0.2–

20 μ M), the evolution of the reaction mixture at 37 °C was monitored by 1D ¹H NMR, and the final products were characterized by 2D NMR and MALDI-TOF MS.

Biological Activity. Minimal inhibitory concentrations (MIC) were measured according to a published method.⁴³ The different bacteria were grown in 1 mL of Mueller–Hinton broth to an optical density OD₆₀₀ of 0.5 units. The desired concentrations of antibiotic were added from stock solutions. The samples were incubated at 37 °C for 24 h. After this time the control culture (with no antibiotic) had an OD₆₀₀ of 1.2–1.6. Then the OD₆₀₀ of each sample was read, and the MIC was taken as the lowest antibiotic concentration inhibiting bacterial growth by greater than 90%.

To determine the effect that expression of AAC(2') has on the MIC values the previously described protocol was slightly modified. The different bacteria were grown in 1 mL of Mueller–Hinton broth to an optical density OD₆₀₀ of 0.3 units. The temperature was switched to 30 °C, and the culture was induced with 0.5 mM IPTG. When the OD₆₀₀ reached 0.5 units, the desired concentrations of antibiotic were added from stock solutions. The samples were incubated at 30 °C for 24 h, and the MIC was taken as the lowest antibiotic concentration inhibiting bacterial growth by greater than 90%.

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Supporting Information Available: Synthesis of derivatives 4 and 5. Supplementary Figures S1 to S7 showing NMR spectra of aminoglycosides 2–6 and solvated MD simulations for derivative 3. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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