See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/224006534

Correction to Context-Dependent Effects of Asparagine Glycosylation on Pin WW Folding Kinetics and Thermodynamics.

ARTICLE in JOURNAL OF THE AMERICAN CHEMICAL SOCIETY · MARCH 2012

Impact Factor: 12.11 · DOI: 10.1021/ja300899d · Source: PubMed

READS

30

8 AUTHORS, INCLUDING:



Joshua L Price

Brigham Young University - Provo Main Cam...

29 PUBLICATIONS 714 CITATIONS

SEE PROFILE



Jeffery W Kelly

The Scripps Research Institute

340 PUBLICATIONS 24,031 CITATIONS

SEE PROFILE



Am Chem Soc. Author manuscript; available in PMC 2012 September 11.

Published in final edited form as:

JAm Chem Soc. 2012 March 7; 134(9): 4450–4451. doi:10.1021/ja300899d.

Correction to Context-Dependent Effects of Asparagine Glycosylation on Pin WW Folding Kinetics and Thermodynamics

Joshua L. Price, Dalit Shental-Bechor, Apratim Dhar, Maurice J. Turner, Evan T. Powers, Martin Gruebele, Yaakov Levy, and Jeffery W. Kelly

In extracting folding and unfolding rate information from our apparent rate constant data, and in fitting the extracted folding rates as a function of temperature to Kramer's model, we inadvertently used the pre-jump equilibrium temperature instead of the post-jump temperature: a difference of 12 °C. The absolute folding and unfolding rate values for each WW variant in Tables 3 and 4 are different than reported in the original paper by < 2-fold for all but 2 cases that are slightly > 2-fold, but most of the folding and unfolding rate ratios (comparing the folding or unfolding rates of the glycosylated vs. non-glycosylated WW variants) are similar to the values presented in the original paper. The changes to the data do not change the conclusion of our paper that specific, evolved protein-glycan contacts must also play a role in mediating the beneficial energetic effects on protein folding that glycosylation can confer.

The corrected data appear in Tables 3 and 4 below. The corrected versions of Figures S44 to S55 (the data that Tables 3 and 4 summarize) are provided in the Supporting Information. The following sentences in the Results section text referring to our kinetic data (pp. 15364):

The modest stabilizing effect of the Asn to Asn-GlcNAc substitution at positions 20 and 30 appears to be a result, in part, of an increased folding rate. Glycosylation at position 20 (compare **20** with **20g**) increases the folding rate []1.1-fold (Table 3). Glycosylation at position 30 (compare **30** with **30g**) has a similar effect. These small folding rate increases agree with the predictions of the computational model and could be consistent with a small amount of denatured-state destabilization as a consequence of glycosylation. However, the decreased unfolding rate of **20g** relative to **20** (also predicted by the model) could indicate that glycosylation at position 20 actually stabilizes the folding transition state and native state simultaneously.

are changed to:

The modest stabilizing effect of the Asn to Asn-GlcNAc substitution at position 20 appears to be primarily due to an increased folding rate (**20g** folds 1.5 times faster than **20**). This folding rate increase agrees with the predictions of the computational model, and could be consistent with a small amount of denatured-state destabilization as a consequence of glycosylation. However, the unfolding rates of **20g** and **20** are indistinguishable, which disagrees with the predicted decrease in unfolding rate upon glycosylation. The stabilizing effect of glycosylation at position 30 appears to come primarily from a decrease in unfolding rate (**30g** unfolds 0.7 times as fast as non-glycosylated **Pin WW**), as predicted by the model. This decrease compensates for an unexpected decrease in folding rate: **30g** folds

Price et al. Page 2

0.8 times as fast as **Pin WW** whereas the model predicted a large increase in folding rate. Results for **33** and **33g** are similarly inconsistent with the predictions of the model.

In addition, the following sentence in the Discussion section text about our kinetic data (pp. 15366):

For example, the observed increased stability and increased folding rate of **20g** relative to **20**, and of **30g** relative to **30**, could be the result of simultaneous transition state and native state stabilization (rather than destabilization of the denatured ensemble), reflecting the presence of favorable glycan-protein contacts at these positions.

is changed to:

For example, the observed increased stability of **20g** and **30g** relative to **20** and **30**, respectively, could be the result of favorable native-state GlcNAc-protein contacts at these positions.

Table 3

Experimentally Measured and Computationally Predicted Folding Rates for Pin WW Variants Having Either Asn or Asn-GlcNAc at the Indicated Positions^a

Price et al.

				(k _f [Asn-GlcN.	(k _f [Asn-GlcNAc])/(k _f [Asn])
structural context	protein	residue at indicated position	measured folding rate b (10 3 s^{-1})	measured	$predicted^c$
β-strand 1	14	Asn	p ⁻	p ⁻	2
	14g	Asn-GlcNAc	p^{-}		
loop 1	17	Asn	5.8 ± 0.2	0.80 ± 0.04	3.6
	17g	Asn-GlcNAc	4.7 ± 0.1		
	18	Asn	8.0 ± 0.2	0.71 ± 0.03	3.7
	18g	Asn-GlcNAc	5.6 ± 0.2		
	19	Asn	6.6 ± 0.2	0.77 ± 0.06	2.8
	19g	Asn-GlcNAc	5.1 ± 0.4		
	20	Asn	4.5 ± 0.2	1.48 ± 0.09	7.7
	20g	Asn-GlcNAc	6.6 ± 0.3		
β-strand 2	23	Asn	p ⁻	p ⁻	1.1
	23g	Asn-GlcNAc	<i>p</i> ⁻		
	PinWW	Asn	11.6 ± 0.7	p ⁻	6.0
	26g	Asn-GlcNAc	<i>p</i> ⁻		
loop 2	Pin WW	Asn	11.6 ± 0.7	0.81 ± 0.05	3.4
	30g	Asn-GlcNAc	9.4 ± 0.2		
β-strand 3	33	Asn	6.3 ± 0.7	0.70 ± 0.16	8.0
	33g	Asn-GlcNAc	4.4 ± 0.8		

 $^{^{3}}$ Variants for which experimental observations agree with computational predictions are italicized.

Page 3

beasured folding rates at 55 °C (328.15 K) were calculated based on relaxation data from laser temperature jump experiments on 100 µM solutions of each Pin WW domain variant in 20 mM sodium phosphate buffer (pH 7; see supporting information for details). Uncertainties represent the standard error in folding rates.

Credicted folding rate ratios at the indicated positions were calculated based on the all-atom native-topology model at the calculated melting temperature of the corresponding non-glycosylated Asncontaining Pin WW variant (in contrast with the measured folding rate ratios, which are all at 55 °C).

The folding kinetics of peptides 14, 14g, 23, 23g, and 26g could not be analyzed via laser temperature-jump experiments due to their low thermal stability.

Price et al. Page 4

Table 4

Experimentally Measured and Computationally Predicted Unfolding Rates for Pin WW Variants Having Either Asn or Asn-GlcNAc at the Indicated Positions^a

Price et al.

				(ku[Asn-GlcN	$(k_u[Asn\text{-}GlcNAc])/(k_u[Asn])$
structural context	protein	residue at indicated position	measured unfolding rate b $(10^3\mathrm{s}^{-1})$	measured	$predicted^c$
β-strand 1	14	Asn	ρ^{-}	p ⁻	0.50
	14g	Asn-GlcNAc	p ⁻		
loop 1	17	Asn	6.1 ± 0.4	0.89 ± 0.08	0.58
	17g	Asn-GlcNAc	5.4 ± 0.3		
	18	Asn	6.9 ± 0.3	0.99 ± 0.09	95.0
	18g	Asn-GlcNAc	6.9 ± 0.5		
	19	Asn	5.4 ± 0.3	1.17 ± 0.12	0.33
	19g	Asn-GlcNAc	6.4 ± 0.6		
	20	Asn	5.9 ± 0.4	1.15 ± 0.11	0.45
	20g	Asn-GlcNAc	6.8 ± 0.4		
β-strand 2	23	Asn	p ⁻	p ⁻	22.0
	23g	Asn-GlcNAc	ρ^{-}		
	PinWW	Asn	8.0 ± 0.7	p -	4.3
	26g	Asn-GlcNAc	ρ^{-}		
loop 2	Pin WW	Asn	8.0 ± 0.7	0.68 ± 0.08	02'0
	30g	Asn-GlcNAc	5.4 ± 0.4		
β-strand 3	33	Asn	42.5 ± 7.3	1.13 ± 0.39	2.4
	33g	Asn-GlcNAc	48.0 ± 14.5		

 $^{^{3}}$ Variants for which experimental observations agree with computational predictions are italicized.

Page 5

beasured unfolding rates at 55 °C (328.15 K) were calculated based on relaxation data from laser temperature jump experiments on 100 μ M solutions of each Pin WW domain variant in 20 μ M sodium phosphate buffer (pH 7; see supporting information for details). Uncertainties represent the standard error in folding rates.

^CPredicted unfolding rate ratios at the indicated positions were calculated based on the all-atom native-topology model at the calculated melting temperature of the corresponding non-glycosylated Asncontaining Pin WW variant (in contrast with the measured unfolding rate ratios, which are all at 55 °C).

d. The unfolding kinetics of peptides 14, 14g, 23, 23g, and 26g could not be analyzed via laser temperature-jump experiments due to their low thermal stability.

Price et al. Page 6