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Chemical Modification of Chitosan. 10.1 Synthesis of Dendronized Chitosan-Sialic Acid Hybrid Using Convergent Grafting of Preassembled Dendrons Built on Gallic Acid and Tri(ethylene glycol) Backbone

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ABSTRACT: Sialic acid dendrons bearing a focal aldehyde end group were synthesized by a reiterative amide bond strategy. Polyamine-ending trivalent (G=1) and nonavalent (G=2) dendrons having gallic acid as branching unit and tri(ethylene glycol) as spacer arm were prepared and initially attached to a sialic acid p-phenylisothiocyanate derivative. The focal aldehyde sialodendrons were then convergently grafted onto the polysaccharide chitosan backbone by reductive amination in 76–80% yields. The degrees of substitution (DS) of the sialodendrimer in the hybrids were 0.13 (G=1) and 0.06 (G=2), which indicates that 87% and 40% of the sialodendrons were attached to the primary amino groups of chitosan. The water solubility of these novel hybrids was further improved by N-succinylation of the remaining amine functionality.

### Introduction

Chitin and the deacetylated form, chitosan, are attractive linear amino polysaccharides found in the shell of crustaceans. These abundant biopolymers, composed mainly of poly( $\beta$ -1,4)-2-acetamido/2-amino-2-deoxy-D-glucopyranose repeating units, constitute valuable biomass resources useful in the preparation of functional biomaterials. Since chitosan itself is nontoxic and biodegradable and shows widespread biological activities, it is an appealing bioactive polymer for further development. Unfortunately, its poor solubility in both organic solvents and aqueous solutions has hampered its widespread development. Recently, this problem has been partly overcome by using counteranions of organic acids.  $^{1b,e}$ 

Alternatively, glycodendrimers<sup>4</sup> are monodispersed and chemically well-defined semi-macromolecules bearing exposed carbohydrate moieties that are finding useful applications as viral and pathogenic cell adhesion inhibitors.<sup>5</sup> Increasing scientific efforts have gone into the design and synthesis of dendrimers.<sup>6</sup> Dendrimers are attractive molecules owing to their multifunctional properties and are useful as scaffold for pharmacophores, neoglycoconjugates, probes, catalysts, and so on.6 Dendronized polymers, on the other hand, are also attractive because of their rodlike conformation and nanostructures.7 Although several investigations have been published toward the synthesis of dendronized polymers,7 there is only scarce reports on dendronized polysaccharide, especially related to chitosan backbone. 1c We report herein the preparation of chitosan-sialodendrimer hybrids using gallic acid as focal point and tri-(ethylene glycol) as spacer arm. The present report complements our previous finding1c using poly(amidoamine) (PAMAM) dendrons as scaffold that lacked the capacity for high incorporation of preformed dendrons on chitosan.

## **Experimental Section**

**Materials.** Chitosan (Flonac C, NHAc = 0.2, DP = 140, FW of unit = 169) was purchased from Kyowa Tecnos Co., Japan. Dialysis membranes (MW 1200 and MW 12 000 cutoff) were purchased from Sigma Co. Aminoacetaldehyde diethyl acetal (**2**) and other reagents were from Aldrich Co. and used without further purification.

**General Methods.** The  $^1H$  and  $^{13}C$  NMR spectra were recorded on a Bruker 500 MHz AMX NMR spectrometer. Proton chemical shifts ( $\delta$ ) are given relative to internal CHCl<sub>3</sub> for CDCl<sub>3</sub>, DMSO for DMSO- $d_6$ , and 3-(trimethylsilyl)propionic- $2,2,3,3-d_4$  acid sodium salt (water-soluble "TMS", TMSP) for D<sub>2</sub>O or 0.5 M DCl in D<sub>2</sub>O solution. Carbon chemical shifts are also given relative to CDCl<sub>3</sub>, DMSO, or TMSP. The degree of polymerization (DP) of original chitosan was determined by GPC using pullulan as standard.

Preparation of Dendron Moieties. 3,4,5-Tris(8-azido-3,6dioxaoctyloxy)benzamidoacetaldehyde Diethyl Acetal (3). 3,4,5-Tris(8-azido-3,6-dioxaoctyloxy)benzoic acid (1) was prepared according to previous literature.  $^8$  To a solution of 1 (0.5 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added diisopropylethylamine (DIPEA: 1.5 mmol), aminoacetaldehyde diethyl acetal (2) (0.75 mmol), and 1-hydroxybenzotriazole hydrate (HOBt: 0.75 mmol). After stirring at room temperature for 5 min under N<sub>2</sub>, 1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydrochloride (EDC: 0.75 mmol) was added. After stirring at room temperature for 5 h, the mixture was evaporated, and the resulting residue was purified by silica gel column chromatography using hexane/ethyl acetate (1/1) as eluent to afford 3 (0.39 mmol) in 77% yield. Compound **3:**  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  (ppm) 1.19 (t, J = 7.1 Hz, 6 H,  $CH_3$  of OEt), 3.34 (t, 6 H,  $CH_2N_3$ ), 3.50-3.85 (m, 30 H, OCH<sub>2</sub> and CONHCH<sub>2</sub>), 4.17 (t, J = 4.8Hz, 6 H,  $CH_2OAr$ ), 4.57 (t, J = 5.3 Hz, 1 H, CH of acetal), 6.35 (t, J = 5.8 Hz, 1 H, CONH), 7.00 (s, 2 H, Ar). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  (ppm) 14.4 (CH<sub>3</sub> of OEt), 42.5 (CONH-CH<sub>2</sub>), 50.6 (CH<sub>2</sub>N<sub>3</sub>), 63.0 (CH<sub>2</sub> of OEt), 69.1-72.4 (CH<sub>2</sub>O), 100.8 (CH of acetal), 107.1, 129.6, 141.5, and 152.5 (Ar), 166.9 (NHCO). FAB-MS (pos): calcd for  $C_{31}H_{52}N_{10}O_{12}$  756, found 757.4 (M + 1, 4.0% base peak).

3,4,5-Tris(8-amino-3,6- $dioxaoctyloxy)benzamidoacetal-dehyde Diethyl Acetal (4). To a solution of <math>\bf 3$  (0.21 mmol) in EtOH (10 mL) was added 50 mg of 10% palladium on carbon.

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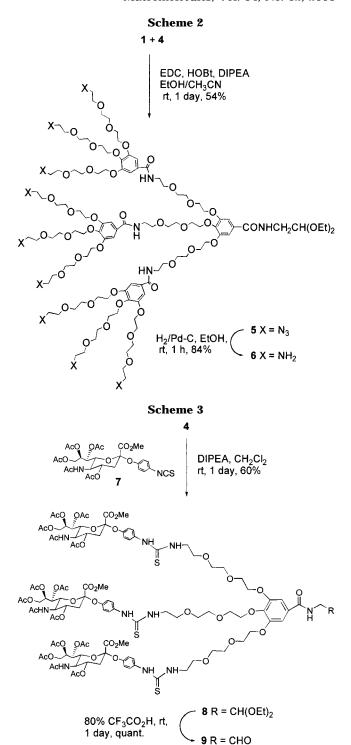
# Scheme 1 N<sub>3</sub> O O O CO<sub>2</sub>H O 1 N<sub>3</sub> H<sub>2</sub>NCH<sub>2</sub>CH(OEt)<sub>2</sub> EDC, HOBt, DIPEA EtOH/CH<sub>3</sub>CN, rt, 5 h 77% O CONHCH<sub>2</sub>CH(OEt)<sub>2</sub> 3 X = N<sub>3</sub> H<sub>2</sub>/Pd-C, EtOH, rt, 4 h, 90% 4 X = NH<sub>2</sub>

Hydrogen was constantly bubbled through the solution at room temperature. After 4 h, the catalyst was removed by filtration, and the filtrate was evaporated under vacuum to dryness. The crude product 4 (0.19 mmol, 90% yield) was used in the next step without further purification. Compound 4:  $^1\mathrm{H}$  NMR (CDCl\_3)  $\delta$  1.14 (m, 6 H, C $H_3$  of OEt), 2.69 (br, 6 H, N $H_2$ ), 2.80 (t, J=4.6 Hz, 6 H, C $H_2$ NH $_2$ ), 3.45–3.80 (m, 30 H, OC $H_2$  and CONHC $H_2$ ), 4.16 (t, J=5.1 Hz, 6 H, C $H_2$ OAr), 4.59 (t, J=5.3 Hz, 1 H, CH of acetal), 7.06 (s, 2 H, Ar).  $^{13}\mathrm{C}$  NMR (CDCl\_3)  $\delta$  14.7 (CH $_3$  of OEt), 41.3 (C $H_2$ NH $_2$ ), 42.6 (CONH– $CH_2$ ), 62.7 (C $H_2$  of OEt), 69.0–72.6 (C $H_2$ O), 100.7 (CH of acetal), 107.4, 129.9, 140.9, and 152.3 (Ar), 166.3 (NHCO). FAB-MS (pos): calcd for C $_{31}\mathrm{H}_{58}\mathrm{N}_4\mathrm{O}_{12}$  678, found 679.2 (M + 1, 1.0% base peak).

3,4,5-Tris{8-[3,4,5-tri-(8-azido-3,6-dioxaoctyloxy)benzamido]-3,6-dioxaoctyloxy\benzamidoacetaldehyde Diethyl Acetal (5). To a solution of amine 4 (0.36 mmol) in EtOH/CH<sub>3</sub>CN (1/1: 20 mL) was added DIPEA (1.8 mmol), 1 (1.3 mmol: 1.2 equiv/ NH<sub>2</sub>), and HOBt (1.8 mmol). After stirring at room temperature for 5 min under N2, EDC (1.8 mmol) was added. After stirring at room temperature for 1 day, the mixture was evaporated, and the crude product was purified by silica gel column chromatography using hexane/ethyl acetate (1/1) to afford 5 (0.194 mmol) in 54% yield. Data for 5: 1H NMR (CDCl<sub>3</sub>)  $\delta$  1.19 (t, J = 7.1 Hz, 6 H, CH<sub>3</sub> of OEt), 3.34 (t, 18 H,  $CH_2N_3$ ), 3.50-3.85 (m,  $OCH_2$  and  $CONHCH_2$ ), 4.17 and 4.30 (m, 24 H, C $H_2$ OAr), 7.27 (m, 8 H, Ar). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  14.3 (CH<sub>3</sub> of OEt), 50.6 (CH<sub>2</sub>N<sub>3</sub>), 61.6 (CH<sub>2</sub> of OEt), 68.3-72.7 (CH<sub>2</sub>), 106.0, 109.0, 110.8, 125.3, 143.7, and 152.3 (Ar), 165.7 and 166.0 (NHCO).

Hydrogenation of **5** was performed under the same conditions described above for **4** and gave **6** in 84% yield. Data for **6**:  $^{1}$ H NMR (CDCl<sub>3</sub>) δ 1.30 (m, 6 H, C $H_3$  of OEt), 2.50 (br, 18 H, N $H_2$ ), 2.80 (m, 18 H, C $H_2$ NH<sub>2</sub>), 3.40–3.85 (m, OC $H_2$  and CONHC $H_2$ ), 4.15 and 4.30 (m, 24 H, C $H_2$ OAr), 7.20–7.25 (m, 8 H, Ar).  $^{13}$ C NMR (CDCl<sub>3</sub>) δ 14.3 (CH<sub>3</sub> of OEt), 41.5 (CH<sub>2</sub>-NH<sub>2</sub>), 41.6 (CONH–CH<sub>2</sub>), 61.4 (CH<sub>2</sub> of OEt), 67.7–73.3 (CH<sub>2</sub>), 104.02, 108.7, 111.4, 125.4, 152.0, and 152.5 (Ar), 165.8, 166.0, and 166.6 (NH CO).

**Preparation of Sialodendrimer.** p-Isothiocyanatophenyl  $\alpha$ -sialoside (7) was prepared from acetochloroneuraminic acid and p-nitrophenol according to published data. <sup>9</sup> To a solution of amine **4** (0.3 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) was added DIPEA (1.2 mmol) and **7** (1.0 mmol: 1.1 equiv/NH<sub>2</sub>). After stirring at room temperature for 1 day, the mixture was evaporated to dryness. The residue was dissolved in MeOH (5 mL) and the



mixture dialyzed against MeOH/H<sub>2</sub>O (2/1) with dialysis membrane (MW 1200 cutoff) for 2 days. The resulting solution was evaporated to dryness to give trivalent  $\alpha\text{-sialodendrimer}$  acetal **8** (G=1) in 60% yield.

Data for **8:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.30 (m, 6 H,  $CH_3$  of OEt), 1.8–2.1 (m, 45 H, OAc and NHAc of Neu5Ac), 2.75 (br, 3 H, H-3ax of Neu5Ac), 3.40–3.80 (m, OC $H_2$ , CONHC $H_2$ , and CO<sub>2</sub>-Me of Neu5Ac), 4.10 (m, 6 H,  $CH_2$ OAr), 4.20–5.30 (H-4, 7, 8, 9 of Neu5Ac), 7.00–7.50 (m, 14 H, Ar), 7.80 (br, NHC(S)NHAr), 9.80 (brs, NHC(S)NHAr). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  15.3 (CH<sub>3</sub> of OEt), 20.7 and 20.9 (OAc), 23.0 (NHAc), 39.0 (C-3 of Neu5Ac), 41.6 (CONH-CH<sub>2</sub>), 44.1 (CH<sub>2</sub>-NHCS), 47.8 (C-5 of Neu5Ac), 53.1 (CO<sub>2</sub>Me), 61.0 (CH<sub>2</sub> of OEt), 62.2 (C-9 of Neu5Ac), 66.8–73.5 (CH<sub>2</sub> and C-4, 6, 8 of Neu5Ac), 99.4 (CH of acetal), 106.0, 123.9, 137.6, and 151.8 (Ar), 167.1–171.9 (NHCO), 181.5 (C=S).

# Scheme 4 6 7, DIPEA, CH<sub>2</sub>Cl<sub>2</sub> rt, 1 day, 60% CO<sub>2</sub>Me AcO CO<sub>2</sub>Me AcHN CO<sub>2</sub>Me AcHN-ÇO<sub>2</sub>Me QAc AcHN<sup>-</sup> ÓAc AcHN<sup>2</sup> NHAc 10 R = CH(OEt)<sub>2</sub> 80% TFA rt, 1 day quant. 11 R = CHO MeO<sub>2</sub> NHAc OAc AcO<sub>1</sub> AcO NHAC

Acetal 8 (0.13 mmol) was dissolved in aqueous 80% CF<sub>3</sub>-CO<sub>2</sub>H (TFA, 3 mL). After stirring at room temperature for 1 day, the solution was evaporated to dryness to afford aldehyde **9** in quantitative yield. Data for **9:** <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.8– 2.0 (m. 45 H. OAc and NHAc of Neu5Ac), 2.60 (br. 3 H. H-3ax of Neu5Ac), 3.40-4.30 (m, OC $H_2$  and Neu5Ac), 7.00-7.60 (m, 14 H, Ar), 7.80 (br, NHC(S)NHAr), 9.80 (brs, NHC(S)NHAr).

The nonavalent  $\alpha$ -sialodendrimer acetal **10** (G = 2) was obtained 60% yield, and aldehyde **11** (G = 2) was also obtained quantitatively in a similar manner as above. Data for 10: 1H NMR (DMSO- $d_6$ )  $\delta$  1.30 (m, 6 H, C $H_3$  of OEt), 1.7–2.05 (m, 45 H, OAc and NHAc of Neu5Ac), 2.57 (br, 3 H, H-3ax of Neu5Ac), 3.20-4.00 (m, OC $H_2$ , CONHC $H_2$ , and CO<sub>2</sub>Me of Neu5Ac), 4.10 (m, 6 H, CH<sub>2</sub>OAr), 4.20-5.20 (H-4, 7, 8, 9 of Neu5Ac), 7.00-7.60 (m, 14 H, Ar), 7.90 (br, NHC(S)NHAr), 9.80 (brs, NHC-(S)NHAr). <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  14.3 (CH<sub>3</sub> of OEt), 20.2–20.8 (OAc), 22.3, and 22.6 (NHAc), 43.7 (CH2-NHCS), 50.0 (C-5 of Neu5Ac), 53.0 (CO<sub>2</sub>Me), 61.0 (CH<sub>2</sub> of OEt), 62.2 (C-9 of Neu5Ac), 66.8-73.5 (CH<sub>2</sub> and C-4, 6, 8 of Neu5Ac), 98.0 (CH of acetal), 108.0, 124.0, 137.5, and 151.8 (Ar), 168.0-171.7 (NHCO), 180.2 (C=S). Data for **11:** <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta$  1.8– 2.0 (m, 45 H, OAc and NHAc of Neu5Ac), 2.60 (br, 3 H, H-3ax of Neu5Ac), 3.40-4.60 (m, OCH<sub>2</sub> and Neu5Ac), 7.00-7.60 (m, 14 H, Ar), 7.80 (br, NHC(S)NHAr), 9.80 (brs, NHC(S)NHAr).

Preparation of Sialodendrimer-Chitosan Hybrids. A typical procedure is as follows: To a solution of chitosan (200

mg: 1.18 mmol of hexosamine residue) in water (10 mL) and MeOH (30 mL) containing AcOH (30 mg) was added aldehyde 9 (0.18 mmol, 0.15 equiv/hexosamine residue) in MeOH (10 mL). After stirring at room temperature for 1 h, NaCNBH<sub>3</sub> (2.4 mmol) was added, and the solution was continuously stirred at room temperature for 1 day. The reaction mixture was evaporated to dryness and immediately treated with 0.5 M NaOH at room temperature for 2 h. Dialysis against distilled water and lyophilization gave trivalent chitosansialodendrimer hybrid ( $\hat{G} = 1$ ) **12** (370 mg, 76%). Data for **12** (DS = 0.13): <sup>1</sup>H NMR (0.5 M DCl/D<sub>2</sub>O)  $\delta$  1.95-2.12 (m, 2.16 H, NHAc and H-3ax of Neu5Ac), 2.88 (br, 0.39 H, H-3eq of Neu5Ac), 3.26 (brs, 0.67 H, H-2 of unsubstituted GlcN), 3.4-5.0 (m,  $CH_2$  of dendrimer, CH and  $CH_2$  of chitosan and Neu5Ac backbone), 7.30-7.60 (m, 1.82 H, Ar). <sup>13</sup>C NMR (0.5 M DCl/ D<sub>2</sub>O) δ 25.0 (NHAc), 43.6 (C-3 of Neu5Ac), 58.7 (C-5 of Neu5Ac and C-2 of chitosan), 62.9 (C-6 of chitosan), 65.5 (C-9 of Neu5Ac), 71.0-74.8 (CH<sub>2</sub> of dendrimer, C-4, 6, 7, 8 of Neu5Ac and C-3 of chitosan), 77.6 (C-5 of chitosan), 79.4 (C-4 of chitosan), 100.3 (C-1 of chitosan), 104.1 (C-2 of Neu5Ac), 127.7, 140.3, 140.7, and 154.7 (Ar), 174.1-178.1 (C=O).

Nonavalent chitosan-sialodendrimer hybrid (G = 2) **14** was also obtained in a similar manner as above (80%). Data for 14 (DS = 0.06): <sup>1</sup>H NMR (0.5 M DCl/D<sub>2</sub>O)  $\delta$  1.94 (br, 0.54 H, H-3ax of Neu5Ac), 2.00-2.12 (m, 2.22 H, NHAc), 2.88 (br, 0.54 H, H-3eq of Neu5Ac), 3.26 (brs, 0.74 H, H-2 of unsubstituted GlcN), 3.4-5.0 (m,  $CH_2$  of dendrimer, CH and  $CH_2$  of chitosan and Neu5Ac backbone), 7.00-7.60 (m, 2.64 H, Ar).  $^{13}$ C NMR (0.5 M DCl/D<sub>2</sub>O)  $\delta$  25.0 (NHAc of chitosan and Neu5Ac), 42.5 (C-3 of Neu5Ac), 54.6 (C-5 of Neu5Ac), 58.8 (C-2 of chitosan), 63.0 (C-6 of chitosan), 65.6 (C-9 of Neu5Ac), 70.8-74.7 (CH<sub>2</sub> of de ndrimer, C-4, 6, 7, 8 of Neu5Ac and C-3 of chitosan), 77.8 (C-5 of chitosan), 79.5 (C-4 of chitosan), 100.4 (C-1 of chitosan), 104.3 (C-2 of Neu5Ac), 127.9, 140.3, and 154.7 (Ar), 174.2–178.0 (C=O). The DS of sialodendrimer was estimated by  $^{1}$ H NMR from the peak ratio at  $\delta$  3.26 (H-2 of unsubstituted GlcN) and 7.3–7.6 (Ar). The DS estimated from the ratio at  $\delta$  2.88 (H-3eq of Neu5Ac) and 3.26 (H-2) also confirmed the result.

N-Succinylation of primary amino groups in hybrids (12, 14) was performed according to the previous literature.  $^{(a,d,16)}$  Hybrid (14: 140 mg) was dispersed in H<sub>2</sub>O (20 mL) and MeOH (60 mL) containing AcOH (100 mg). Succinic anhydride (3.0 mmol) was added in excess and stirred at 70 °C. After 1 day, the mixture was concentrated to ca. 10 mL, 1.0 M NaOH (10 mL) was then added, and the mixture (in 0.5 M NaOH) was stirred at room temperature. After 2 h, the mixture was dialyzed (H2O) for 2 days and lyophilized to afford watersoluble N-succinylated hybrid (15: 140 mg) in 90% yield. Data for **15** (DS of dendrimer = 0.06, DS of succinyl group = 0.60): <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  1.89 (brt, J = 11.7 Hz, 0.54 H, H-3ax of Neu5Ac), 2.10 (s, 1.62 H, NHAc of Neu5Ac), 2.13 (s, 0.60 H, NHAc), 2.54 (br. 1.20 H, NHC(O)– $CH_2$  of N-succinvl group), 2.62 (br, 1.20 H, CH<sub>2</sub>-CO<sub>2</sub>Na of N-succinyl), 2.93 (H-3eq of Neu5Ac), 3.4-4.6 (m,  $CH_2$  of dendrimer, CH and  $CH_2$  of chitosan and Neu5Ac backbone), 7.00-7.60 (m, 2.64 H, Ar).  $^{13}$ C NMR (D<sub>2</sub>O)  $\delta$  25.0 (NHAc of Neu5Ac), 25.1 (NHAc of chitosan), 35.5 and 35.8 (CH<sub>2</sub> of succinyl), 43.8 (C-3 of Neu5Ac), 54.6 (C-5 of Neu5Ac), 58.1-59.1 (C-2 of chitosan), 63.1-63.4 (C-6 of chitosan), 65.5 (C-9 of Neu5Ac), 71.2-75.1 (CH2 of de ndrimer, C-4, 6, 7, 8 of Neu5Ac and C-3 of chitosan), 77.7 (C-5 of chitosan), 80.6 (C-4 of chitosan), 100.4 (C-1 of chitosan), 104.3 (C-2 of Neu5Ac), 126.2, 127.6, 139.4, and 154.1 (Ar), 176.2-179.2 (NHCO), 183.9 (CO2Na). The DS of succinyl group was estimated by  ${}^{1}H$  NMR from the peak ratio at  $\delta$  1.89 (H-3ax of Neu5Ac: 0.54 H) and at  $\delta 2.54-2.62$  (C $H_2$  of N-succinyl group). Although a part of water-insoluble materials was found in N-succinylated hybrid 13 (90%), the corresponding signals of N-succinyl group were observed in  $^1H$  NMR (D<sub>2</sub>O) at  $\delta$  2.54 (br, NHC( $\mathring{O}$ )- $\mathring{C}H_2$ ) and 2.62 (br,  $\mathring{C}H_2$ - $\mathring{C}O_2$ Na).

## **Results and Discussion**

3,4,5-Trihydroxybenzoic acid (gallic acid)-based sialodendrons were synthesized using commercially available tri(ethylene glycol) as a spacer arm. The latter was chosen to ensure advantageous water solubility of the resulting hybrid and to counteract the hydrophobic effect of the aromatic gallic acid used as focal backbone. As shown in Scheme 1, the terminal carboxyl group of known precursor 18 was modified into focal acetalprotected 3 by simple coupling with commercially available aminoacetaldehyde diethyl acetal (2) in good yield (77%) using carbodiimide (EDC) and hydroxybenzotriazole, followed by hydrogenation of the azide groups to afford trivalent amine 4 in 90% yield. To build up the next generation of dendron backbone (Scheme 2), the coupling of 4 with 3.6 equiv (1.2 equiv/NH<sub>2</sub>) of acid 1 and hydrogenation sequence were carried out as described above for 3 to give a nonavalent amine 6 in moderate yields (5: 54%; 6: 84%). Trivalent sialodendrimer acetal (8) was prepared with amine 4 and peracetylated p-isocyanatophenyl  $\alpha$ -sialoside (7) (3.3) equiv, 1.1 equiv/ NH<sub>2</sub>) at room temperature for 1 day (Scheme 3). Although the purification of **8** (FW = 2451) by silica gel column was difficult owing to its high molecular weight, crude 8 was successfully separated from the remaining sialoside 7 (FW = 627) by dialysis (MW 1200 cutoff)<sup>9</sup> with a mixed solvent of MeOH/H<sub>2</sub>O (2/1) and gave pure 8 in 60% yield. The hydrolysis of

acetal **8** was carried out with aqueous 80%  $CF_3CO_2H$  at room temperature for 1 day to provide aldehyde **9**. <sup>1c</sup> Noteworthy is the fact that the O-sialoside linkage has resisted the hydrolysis step owing to the presence of the ester protecting groups which confer higher stability to the molecule. As shown in Scheme 4, the protected nonavalent sialodendron acetal **10** (FW = 7956) was also successfully prepared and purified in 60% yield and provided aldehyde **11** under the same trifluoroacetolysis conditions (80%). The extent of coupling could be readily established using a negative ninhydrin test and the exact integration of NMR signals.

13 R = H, or  $C(O)(CH_2)_2CO_2Na$ 

0.5 M NaOH rt. 2 h

Scheme 5 shows the efficient reductive amination of trivalent sialodendrimer aldehyde 9 and chitosan with NaCNBH<sub>3</sub> according to the previous literature. <sup>1a,c,d</sup> The aldehyde 9 (0.15 equiv/hexosamine residue) was successfully attached to the chitosan backbone and gave trivalent chitosan-sialodendrimer hybrid (G = 1) 12 in 76% yield. The degree of substitution (DS) of hybrid 12 was shown to be 0.13 by NMR, which indicates that 87% of aldehyde 9 reacted to the primary amino group of chitosan. The nonavalent chitosan-sialodendrimer hybrid (G = 2, Scheme 6) **14** was also successfully prepared with aldehyde 11 (0.15 equiv) in 80% yield. The DS of hybrid (G = 2) was 0.06, and only 40% of aldehyde 11 could be attached to chitosan. The lower reactivity in *G* = 2 would be due to the steric hindrance of the higher molecular weight aldehyde **11** (MW = 7866), compared with aldehyde 9 (MW = 2361). Both hybrids 12 and 14, obtained after protecting group hydrolysis (0.5 M NaOH,

rt, 2 h), were only slightly soluble in neutral water and thus would not be useful in biological evaluation. To further improve their water solubility, the remaining unmodified amino groups of the chitosan backbone were succinylated with excess succinic anhydride (i: AcOH, H<sub>2</sub>O, MeOH, 70 °C, 1 day; ii: 0.5 M NaOH, rt, 2 h, then dialyzed and lyophilized) to give freely water-soluble N-succinylated hybrids 13 and 15 in 90% yields.

0.5 M NaOH, rt, 2 h

In conclusion, water-soluble dendronized chitosansialic acid hybrids were successfully prepared using gallic acid as focal point and tri(ethylene glycol) as spacer arm. Further biological evaluation of these promising hybrids is being investigated toward the inhibition of viral pathogens including the flu virus.5

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## **References and Notes**

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