

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/269717442>

Evolutionary Importance of the Intramolecular Pathways of Hydrolysis of Phosphate Ester Mixed Anhydrides with Amino Acids and Peptides

ARTICLE *in* SCIENTIFIC REPORTS · DECEMBER 2014

Impact Factor: 5.58 · DOI: 10.1038/srep07440 · Source: PubMed

CITATIONS

2

READS

17

4 AUTHORS, INCLUDING:



Damien Beaufils

Université de Montpellier

3 PUBLICATIONS 6 CITATIONS

SEE PROFILE



Rossi jean christophe

Université de Montpellier

18 PUBLICATIONS 186 CITATIONS

SEE PROFILE



Robert Pascal

CNRS - University of Montpellier

80 PUBLICATIONS 879 CITATIONS

SEE PROFILE



OPEN

SUBJECT AREAS:

CHEMICAL BIOLOGY

REACTION MECHANISMS

ORIGIN OF LIFE

MOLECULAR EVOLUTION

Evolutionary Importance of the Intramolecular Pathways of Hydrolysis of Phosphate Ester Mixed Anhydrides with Amino Acids and Peptides

Ziwei Liu, Damien Beaufils, Jean-Christophe Rossi & Robert Pascal

Institut des Biomolécules Max Mousseron, UMR5247 CNRS – University of Montpellier.

Received

17 October 2014

Accepted

21 November 2014

Published

11 December 2014

Correspondence and
requests for materials
should be addressed to

R.P. (rpascal@univ-
montp2.fr)

Aminoacyl adenylates (aa-AMPs) constitute essential intermediates of protein biosynthesis. Their polymerization in aqueous solution has often been claimed as a potential route to abiotic peptides in spite of a highly efficient CO₂-promoted pathway of hydrolysis. Here we investigate the efficiency and relevance of this frequently overlooked pathway from model amino acid phosphate mixed anhydrides including aa-AMPs. Its predominance was demonstrated at CO₂ concentrations matching that of physiological fluids or that of the present-day ocean, making a direct polymerization pathway unlikely. By contrast, the occurrence of the CO₂-promoted pathway was observed to increase the efficiency of peptide bond formation owing to the high reactivity of the *N*-carboxyanhydride (NCA) intermediate. Even considering CO₂ concentrations in early Earth liquid environments equivalent to present levels, mixed anhydrides would have polymerized predominantly through NCAs. The issue of a potential involvement of NCAs as biochemical metabolites could even be raised. The formation of peptide-phosphate mixed anhydrides from 5(4*H*)-oxazolones (transiently formed through prebiotically relevant peptide activation pathways) was also observed as well as the occurrence of the reverse cyclization process in the reactions of these mixed anhydrides. These processes constitute the core of a reaction network that could potentially have evolved towards the emergence of translation.

The biosynthesis of peptides involves aminoacyl adenylates (aa-AMPs), formed through the reaction of ATP with α -amino acids (aas) (Fig. 1), that are subsequently used to aminoacylate tRNA. Their standard free energy of hydrolysis value $\Delta G^{\circ'} = ca. -70 \text{ kJ mol}^{-1}$, determined for Tyr-AMP¹, ranks them among the energy-richest biochemicals. Aa-AMPs possess a phosphate group transfer potential much higher than ATP¹ and might then constitute adenylating agents as well as aminoacylating agents^{2,3}. The otherwise unfavourable¹ reaction of ATP with α -amino acids ($K = 3.5 \times 10^{-7}$) is driven towards completion by selective stabilization of aa-AMPs in the active sites of aminoacyl tRNA synthetases (aaRSs). They usually remain sequestered by the enzyme and are not released in solution before reacting with tRNA. The importance of this process can be appreciated by considering that the set of aaRS enzymes, responsible for the association of amino acids with their cognate tRNAs, actually holds the key of the genetic code. The evolutionary path through which adenylates were introduced in the process remains unidentified. In addition of being thermodynamically unfavourable, the spontaneous reaction is indeed very slow in the absence of enzyme^{4,5}, so that the emergence of the biochemical amino acid activation pathway remains unexplained before a set of catalysts (very probably ribozymes) could lead to an embryo of the genetic code for prebiotically available amino acids⁶. In spite of this obstacle, the evolution of this pathway from an abiotic process of random peptide formation *via* the polymerization of α -amino acid mixed anhydrides with phosphate (aa-PMAs) or phosphate esters (aa-PEMAs) and adenylates (aa-AMPs) has prompted much work^{7–10}. However, the abiotic formation of adenylates or their analogues from phosphate anhydrides did not receive any experimental support. As a matter of fact, the claim¹¹ that ATP is capable of driving the polymerization of α -amino acids on clays through aa-AMP intermediates turned out to be non-reproducible¹². Though the genetic code might have evolved late in the hypothesis of an “RNA world” without needing ATP activation as shown by the successful selection of ribozymes capable of aminoacylating RNAs using either amino acid esters¹³ or activated RNAs¹⁴, an early co-evolution involving the chemistries of nucleotides and amino acids is consistent with the comparatively higher abundance of the latter as the products of abiotic processes. Therefore, selecting the co-

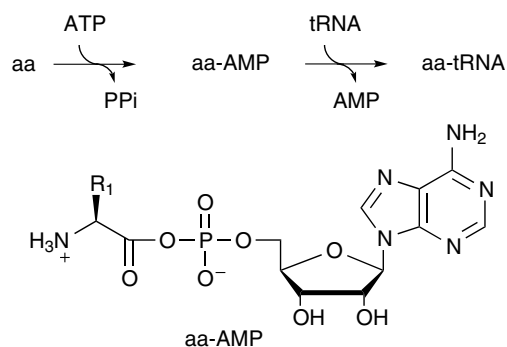


Figure 1 | The aaRS-catalyzed reaction of α -amino acids with ATP.

evolutionary option, the elucidation of the potential evolutionary process through which aa-AMPs could have been introduced requires the identification of simple pathways capable of leading to these intermediates. A likely possibility is the reaction of α -amino acid *N*-carboxyanhydrides (NCAs) with inorganic phosphate¹⁵ and its esters including adenylates that takes place spontaneously at moderate pH^{16,17} (Fig. 2a). This possibility is supported by the role of NCAs deduced from the literature² and the disclosure of realistic abiotic pathways for their formation during the last decade^{18,19}. Since the activation of the C-terminus in peptides has recently been identified as a plausible prebiotic pathway and involves the formation of 5(4*H*)-oxazolone intermediates²⁰, it is reasonable that similar mixed anhydrides with phosphates involving acylated amino acids (acyl-aa-PEMAs) or peptides (peptidyl-PEMAs) could be formed by reaction of the energy-rich cyclic intermediate (Fig. 2b). The occurrence of abiotic pathways leading to aa-PEMA or peptidyl-PEMA must have preceded their involvement in chemical evolution. However, the low stability of these mixed anhydrides and the availability of highly reactive cyclic intermediates prone to polymerize more easily renders their role in early abiotic processes of peptide formation highly questionable.

The kinetic stability of aa-AMPs and of other aa-PEMAs has been studied in aqueous solution leading to contradictory results in the literature^{21–24}. Of particular interest with regard to an evolutionary context is the description of a highly efficient CO₂-catalyzed path of hydrolysis^{21–23}. No definitive mechanism has been proposed but the intermediacy of NCAs is highly probable^{2,25,26} since other activated

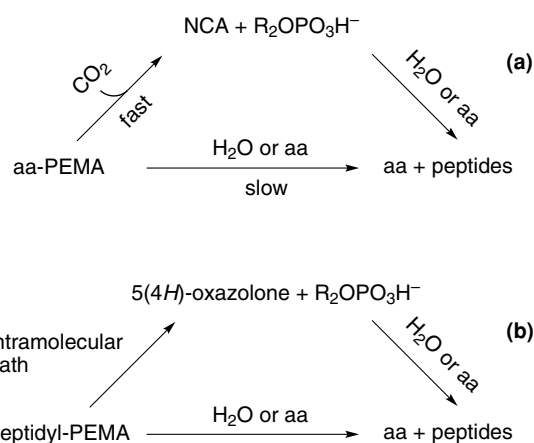


Figure 3 | Intramolecular pathways competing with direct nucleophilic reactions for the conversion of mixed phosphate anhydrides. (a) The efficient hydrolytic pathway and conversion into peptides of amino acid phosphate ester mixed anhydrides promoted by carbon dioxide through NCAs. (b) Cyclization into 5(4*H*)-oxazolone competing with direct nucleophilic reaction of acyl-aa-PEMA and peptidyl-PEMA.

amino acids (nitrophenyl esters, thioesters) proved to undergo conversion into NCAs in hydrogen carbonate buffers²⁵. This analysis casts doubts on the possibility that aa-AMPs constitute efficient monomers for the abiotic formation of peptides in aqueous solutions^{2,3,26} since most early Earth aqueous environments are likely to have contained CO₂ or HCO₃[−]. The present investigations were aimed at providing data on the efficiency of the CO₂-promoted pathway (Fig. 3a) in aqueous solution at neutral pH and in the presence CO₂ concentrations compatible with early Earth environments and at clearly identifying the NCA as an intermediate. They address both the issues of the stability of aa-AMPs and of other aa-PEMAs and that of the path of peptide formation. They demonstrate the prevalence of the CO₂-promoted pathway in the hydrolysis of adenylates. More importantly, using model amino amide reactants, they additionally demonstrate that peptide bond formation takes place predominantly from the cyclic intermediates rather than directly from the mixed anhydrides ruling out any possibility of considering the latter as direct peptide precursors at early stages of chemical or bio-

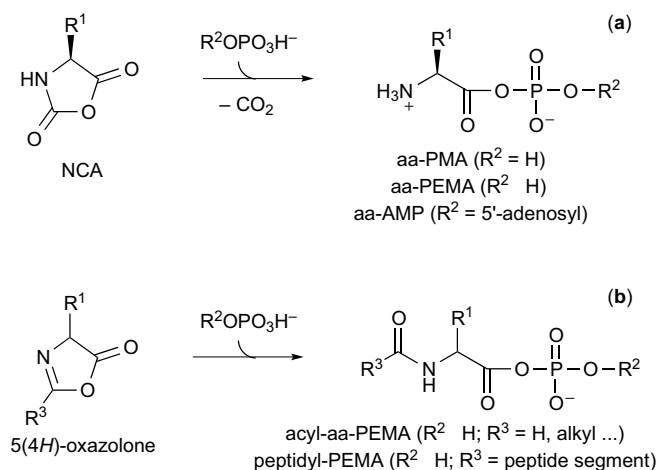


Figure 2 | Potential pathways for the abiotic formation of mixed anhydrides of α -amino acids and peptides with phosphate (PMA) and phosphate esters (PEMA) including adenylates (AMP). (a) Reaction of NCAs with phosphate esters; (b) The hypothesized similar reaction of 5(4*H*)-oxazolones.

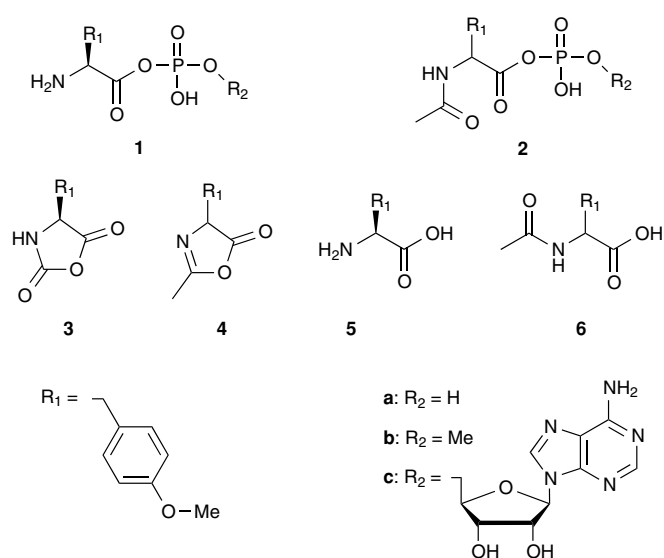


Figure 4 | Structure of the reactants and products related to *O*-methyl-tyrosine (H-Tyr(Me)-OH) 5 studied in this work.



chemical evolution. Lastly, considering NCAs as likely precursors of aa-AMPs and aa-PEMAs, the hypothesis of an abiotic formation of non-coded peptides through these mixed anhydrides becomes unnecessary. The evolution of translation must then have proceeded through a pathway independent from abiotic polymerization. This work also addresses the more general goal of understanding the stability of phosphate mixed anhydrides of amino acids and peptides in aqueous media at moderate pH. As a matter of fact, though *N*-acylation is an obvious way to prevent CO₂ participation, another intramolecular path of breakdown through 5(4*H*)-oxazolones is possible in the case of acyl-aa-PEMAs (Fig. 3b). Therefore, the issues of the importance of the NCA and 5(4*H*)-oxazolone pathways in the reactions of the corresponding mixed anhydrides (Fig. 3) are raised as well as that of the potential role of these cyclic intermediates as potential prebiotic precursors of these mixed anhydrides (Fig. 2). The consequences of these chemical pathways as factors determining early biological evolution of amino acid activation processes and their constraints on the contemporary biochemistry of adenylates will also be discussed.

Results

Experiments were carried out from model systems derived from *O*-methylated tyrosine **5** (Fig. 4) likely to be representative of the reactivity of usual amino acid derivatives. The UV-absorption of the tyrosine side chain ($\lambda_{\text{max}} = 273$ nm) was selected to monitor reactions by HPLC at a reasonably low (0.05–1 mM) concentration range in which activated intermediates have a lifetime sufficient for their behaviour to be determined. Furthermore, phenol methylation was introduced to simplify analyses by avoiding any side-reaction of this group. Reactions were carried out in non-nucleophilic MES or MOPS buffers at pH values of 6.5 or 7.5, respectively, whereas 50 mM phosphate or methyl phosphate buffers were used for studying the transient formation of mixed anhydrides. Analyses were performed to monitor the reaction progress of samples stored in the HPLC systems located in a room maintained at the temperature of 20°C. Fast reactions were monitored by withdrawing 1 mL samples from the reaction medium and the reaction was blocked by addition of a formic acid solution to bring the pH to a value below 4 (Supplementary information).

NCAs as intermediates of aa-PEMA reactions promoted by CO₂. The hydrolysis of methyl phosphate mixed anhydride **1b** was studied in buffered solutions in the presence of varying contents of CO₂/HCO₃[−]. The reaction rates were observed to strongly depend on the presence of CO₂ as shown by a *c.a.* 4 fold increase in rate using pH 6.5 MES buffers previously equilibrated with air as compared with a solution flushed with N₂ for 60 min (Fig. 5, panel A). The rates could be reduced by further *c.a.* 35% by extensive degasification through cycles of freezing at −95°C/gas removal under vacuum/melting in a closed vessel. Under the conditions of the experiment displayed in the panel A of Fig. 5, the starting material **1b** (HPLC retention time, *r.t.* 4.6 min, method A) disappeared slowly and several species containing the methoxyphenyl moiety (λ_{max} 273 nm) were observed, namely the free amino acid **5** (*r.t.* 8.4 min) representing the main product of hydrolysis but also several peaks corresponding to the dipeptide H-Tyr(Me)-Tyr(Me)-OH (*r.t.* 22.7 min) and the diketopiperazine *cyclo*-Tyr(Me)-Tyr(Me) (*r.t.* 23.6 min), very probably resulting of the cyclization of the mixed anhydride H-Tyr(Me)-Tyr(Me)-OPO₃Me[−], which has not been properly identified. The presence of these two products was confirmed by HPLC-MS analysis ($[M + H] = 373.2$ at *r.t.* 1.52 min and 355.2 at *r.t.* 1.88 min, method C). By contrast, the addition of 2 or 10 mM NaHCO₃ to the buffer led to the fast disappearance ($\leq 1\%$ after 3 min) of the mixed anhydride **1b** as monitored by HPLC analysis (Fig. 5, panel B). An intermediate (*r.t.* 23.1 min, method A) formed in proportion yields as high as

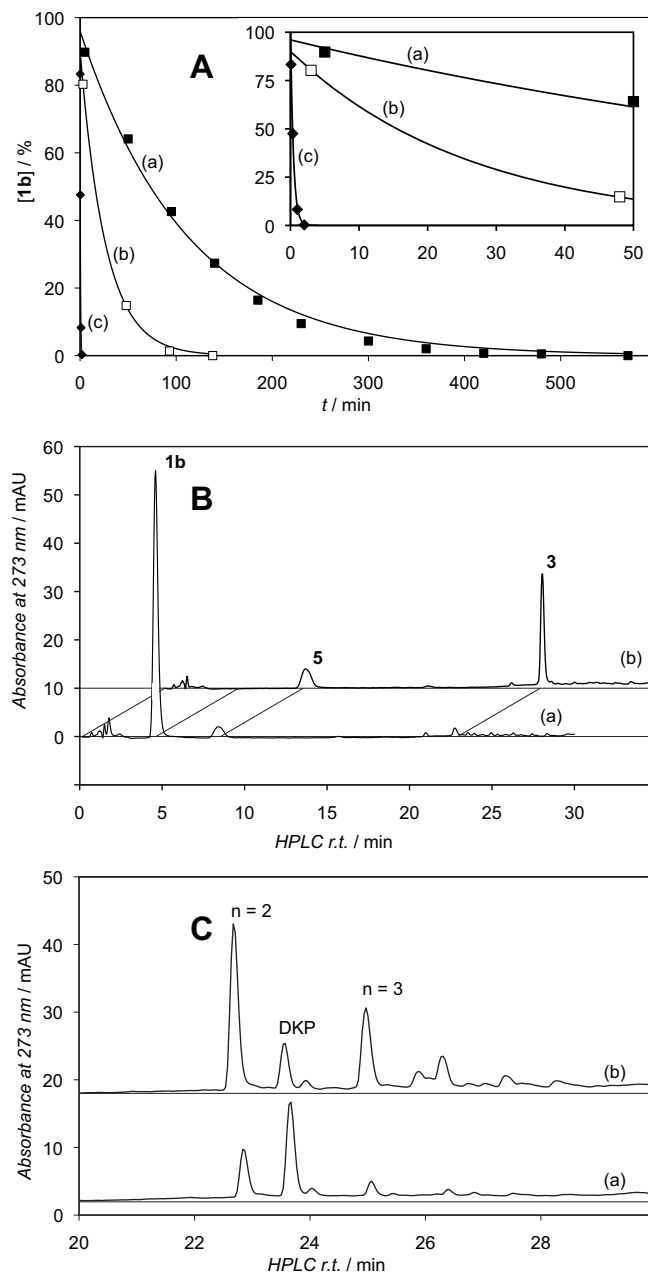


Figure 5 | Hydrolysis of aa-PEMA **1b.** Panel A – Monitoring by the evolution of its HPLC peak area (%), method A): (a) in a 100 mM pH 6.5 MES buffer flushed with N₂ for 60 min (half-life *c.a.* 80 min, filled squares); (b) in a similar buffer equilibrated with air (half-life *c.a.* 18 min, open squares); (c) in a similar buffer to which was added 10 mM NaHCO₃ (half-life *c.a.* 0.3 min, filled diamonds, 1 mL samples were withdrawn and acidified with 20 μ L of 2 M formic acid before analysis); the expanded time scale in the inset shows that reaction (c) is completed within 2 min. Panel B – HPLC traces (method A) of samples withdrawn at 3 min from experiments carried out (a) in a 100 mM pH 6.5 MES buffer flushed with N₂ for 60 min indicating the presence of unreacted mixed anhydride **1b** and a minor conversion to α -amino acid **5** and (b) in a 100 mM pH 6.5 MES buffer to which was added 2 mM NaHCO₃ demonstrating a complete conversion of the starting material into NCA **3**. Panel C – HPLC traces corresponding to the range of retention times of the peptide products (method A, H-(Tyr(Me))_n-OH with *n* = 2 to 5, *r.t.* 22.7, 24.9 min for *n* = 2, 3, respectively and diketopiperazine, *cyclo*-Tyr(Me)-Tyr(Me), DKP, *r.t.* 23.6 min) of the reactions after completion (a) in a 100 mM pH 6.5 MES buffer flushed with N₂ for 60 min and (b) in a 100 mM pH 6.5 MES buffer to which was added 10 mM NaHCO₃. HPLC peaks were identified by both injections of authentic samples and HPLC-ESI-MS analyses (method C) from similar experiments.



60% and was identified as the NCA **3** by a retention time identical to the authentic product and by HPLC-ESI-MS (negative mode $[M-H]^- = 220.07$, r.t. 1.96 min, method C). This intermediate was rapidly converted into the product **5** accompanied by the dipeptide H-Tyr(Me)-Tyr(Me)-OH. The presence of the dipeptide was confirmed by HPLC-ESI-MS (positive mode $[M+H]^+ = 373.2$) as well as that of higher oligomers H-[Tyr(Me)]_n-OH (with $n = 3$ to 5, $[M+H]^+ = 550.2, 727.3, 904.4$ for peaks at r.t. 1.76 min, 1.93 min, 2.06 min, respectively, method C). By contrast reduced amounts of diketopiperazine *cyclo*-Tyr(Me)-Tyr(Me) formed confirming that the starting material lifetime was not sufficient for it to behave as a polymerization initiator leading to a dipeptide mixed anhydride prone to cyclization²⁷. Under these conditions involving the presence of HCO_3^- , the polymerization into peptides thus proceeds through the NCA rather than directly from the starting material. An NCA intermediate was also observed to form rapidly at pH 7.5 in 100 mM MOPS buffers in the presence of added HCO_3^- (Supplementary Information, Fig. S1). This behaviour indicates that the formation of long peptides from adenylates reported in the literature^{9,10} results probably from the polymerization of NCAs rather than from that of adenylates. The conversion of aminoacyl adenylates into NCA in the presence of CO_2/HCO_3^- was investigated starting from the Tyr(Me) derivative **1c** (Supplementary Information, Fig. S2). The conversion of **1c** into NCA was observed to proceed with rates similar to that observed for mixed anhydride **1b**. The release of AMP (r.t. 1.5 min, method A) accompanying the formation of NCA **3** could be detected by HPLC allowing the reaction to be monitored at 50 μ M concentrations of reactant **1c** (r.t. 6.8 min, method A). The lifetime of the adenylate decreased with increasing concentrations of CO_2/HCO_3^- ($t_{1/2} \sim 80$ min, ~ 25 min, and < 2 min at pH 6.5 in N_2 -flushed buffer, air equilibrated buffer and in the presence of 500 μ M HCO_3^- , respectively). At pH 7.5 the lifetime of adenylate **1c** was reduced to less than 1 min in the presence of 500 μ M HCO_3^- , which means that this mixed anhydride is likely to be converted into NCA within a few seconds at concentrations of CO_2/HCO_3^- above 2 mM and at pH value close to neutrality, which are representative of the present day ocean or physiological fluids. It is worth noting that this lifetime is not sufficient for peptides to be significantly formed by a direct reaction with adenylate so that any observation of peptide products under these conditions results for the most part from the intermediacy of NCAs.

At pH 4, the hydrolysis of mixed anhydride **1b** was much slower ($t_{1/2} =$ ca. 550 min) and CO_2 catalysis was not observed (Supplementary Information, Fig. S3). This result is consistent with the results obtained by Kluger from alanyl ethyl phosphate²⁴. The protonation of the amino group of **1b** increases the electrophilic character of its acyl group and then the rates of nucleophilic attack, but it also prevents any possibility of reaction with CO_2 according the pathway of Fig. 3a. The hydrolysis of the acetylated mixed anhydride **2b** was indeed observed to be slower ($t_{1/2} \sim 950$ min at pH 6.5) and was not affected by addition of 10 mM $NaHCO_3$ (Fig. 6) in a way consistent with this explanation and with previously reported analyses²². However, it is important to emphasize that the CO_2 -catalyzed pathway does not only constitute a process leading to the deactivation and the hydrolysis of mixed anhydrides since peptide formation can be improved significantly by this means. As a matter of fact, with regard to peptide formation, the prevalence of the NCA pathway was demonstrated by studying the model reaction of 1 mM mixed anhydride **1b** with 5 mM glycylamide either in a nitrogen-flushed sample or in the presence of 2 mM $NaHCO_3$ (Fig. 7). Importantly, less than 2 min were sufficient for the starting material to be exhausted in the presence of carbonate, whereas CO_2 removal increased the reaction times to much higher values ($t_{1/2} \sim 50$ min) and reduced the final yield in dipeptide (Fig. 7). This reaction remained faster than that observed for the acetylated mixed anhyd-

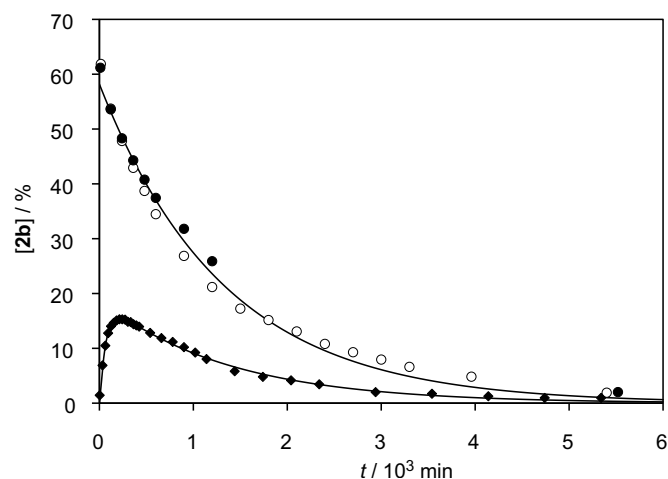


Figure 6 | The hydrolysis of acyl-aa-PEMA **2b compared with the behaviour of the intermediate formed from the independent reaction of 5(4H)-oxazolone **4** with methyl phosphate.** Evolution of **2b** monitored by the evolution of HPLC peak areas (%; method B) in several experiments. (a) Reaction of acyl-aa-PEMA **2b** in a 100 mM pH 6.5 MES buffer ($t_{1/2} =$ ca. 950 min, open circles) and (b) in a similar buffer to which 10 mM $NaHCO_3$ was added (filled circles). (c) Formation ($t_{1/2} =$ ca. 50 min) and hydrolysis ($t_{1/2} =$ ca. 950 min) of mixed anhydride **2b** from 1 mM 5(4H)-oxazolone **4** in a 50 mM methyl phosphate buffer at pH 6.5 (filled diamonds).

ride **2b** ($t_{1/2} \sim 260$ min) unable to undergo the conversion into NCA, but that will be demonstrated below to partly undergo cyclization into 5(4H)-oxazolones. These experiments carried out using glycylamide for mimicking a growing peptide chain show that the polymerization of adenylates and other aa-PEMA is improved in the presence of CO_2 by the occurrence of the NCA pathway owing to both the higher reactivity of the latter intermediate and its ability to suppress diketopiperazine formation.

The interconversion of 5(4H)-oxazolones and acyl-aa-PEMA and peptidyl-PEMA. The reaction of Ac-Tyr(Me)-OH-derived oxazolone **4** in methyl phosphate-buffered aqueous solution (pH 6.5) at 20°C was monitored by HPLC and compared with the hydrolysis of mixed anhydride **2b** in MES buffers (Fig. 6). Comparable rates were observed and the intermediate of the 5(4H)-oxazolone **4** reaction was identified *in situ* by HPLC-ESI-HRMS (negative mode, calcd for $C_{13}H_{17}NO_7P^-$, 330.0743; found 330.0747) as the mixed anhydride **2b**. A similar behaviour was observed from a reaction of inorganic phosphate (Supplementary Information, Fig. S5). The hydrolysis of mixed anhydride **2b** was monitored by HPLC at 20°C in buffered solutions (Fig. 6). The reaction was also carried out in D_2O to detect any hydrogen/deuterium exchange resulting from the transient formation of 5(4H)-oxazolone^{20,28} and compared to the product of a similar reaction of pure oxazolone **4** (Table 1). The values obtained demonstrate the occurrence of an intramolecular pathway already suspected from the higher rate of conversion of acylated aa-AMPs compared to simple acyl-adenylates²⁹. At pH values below 5, the hydrolysis of anhydride **2b** (Supplementary Information, Fig. S4) has been observed to become faster in a way similar to the observation made by Lacey's group for Ac-Phe-AMP²². The identification of an intramolecular pathway made in the present work strongly suggests that the acid catalysis of acyl-aa-PEMA hydrolysis is the consequence of a facilitated cyclization from a good neutral phosphate leaving group. However, the absence of H/D exchange from the reaction of neither acyl-aa-PEMA **2b** nor 5(4H)-oxazolone **4** at this pH (Table 1) prevented any

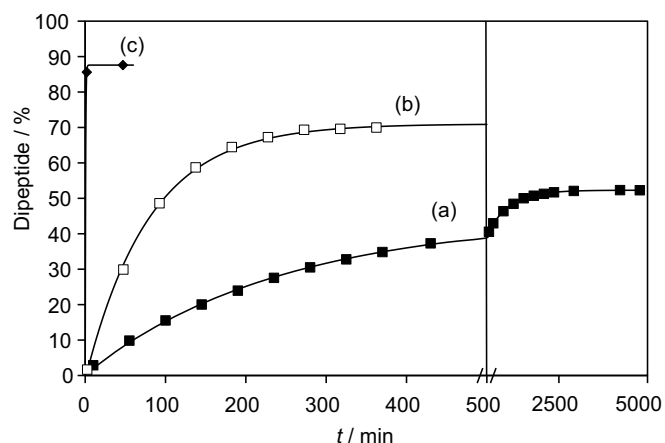


Figure 7 | Formation of the dipeptides Ac-Tyr(Me)-Gly-NH₂ or H-Tyr(Me)-Gly-NH₂ by reaction of 5 mM H-Gly-NH₂ with mixed anhydrides **2b or **1b**, respectively, in 100 mM pH 6.5 MES buffers.** Ratio of the peak area of formed peptides (%): (a) Reaction of 1 mM acyl-aa-PEMA **2b** (half-life c.a. 220 min, method A, r.t. 11.43 min, filled squares); (b) Reaction of aa-PEMA **1b** in the N₂-flushed MES buffer (half-life c.a. 55 min, method A, r.t. 5.85 min, open squares); (c) Reaction of aa-PEMA **1b** in the MES buffer to which 2 mM NaHCO₃ was added (half-life ≤1 min, method A, r.t. 5.85 min, filled diamonds). The peptide products were identified by ESI-MS.

determination of the actual pathway of hydrolysis of mixed anhydride **2b**.

Similarly, we analyzed the degree of D/H exchange during the reaction of **2b** with L-Ala-NH₂ in D₂O at pH 6.5 (Table 1). The observation of a partial deuteration of the two diastereoisomers of the dipeptide product demonstrates that even when a better nucleophile is present, the α -proton is exchanged to a significant extent before the subsequent reaction of the 5(4H)-oxazolone takes place. The fast reaction of acyl-aa-AMP²⁹ and other acyl-aa-PEMA results therefore, at least for a noticeable part, from a transient conversion into 5(4H)-oxazolones. Interestingly, the different degrees of deuteration of the two diastereoisomers indicate that the intramolecular path of Fig. 3b has a higher stereoselectivity as compared to the direct path (the reactants **2b** and **4** were prepared under a racemic form²⁸).

Discussion

As regards aa-PEMA reactions, it is noteworthy that CO₂ catalysis proceeds through a pathway involving induced intramolecularity³⁰. This kind of process shares one of the most important components of enzymatic activity, which corresponds to the utilization of binding energy to non-reacting portions of the substrate to bring about catalysis³¹. It was also proposed to constitute the easiest path for enzyme evolution under the name of *uniform binding*³² and is moreover

necessary for enzymes to exceed a physical limit³³. Induced intramolecularity has also been used to drive highly stereoselective catalysis in organic synthesis^{34,35}. The efficiency of this kind of catalysis relies on the rates of intramolecular reactions³⁶. Carbon dioxide present at total concentrations of ca. 30–40 μ M in pH 6.5 solutions equilibrated with air (as deduced from the Henry's coefficient of CO₂³⁷ and the pK_a of carbonic acid) brings about a rate increase sufficient to render the catalytic pathway largely predominating, which is remarkable by considering a simple three-atom molecule compared to the efficiency of enzymes³⁸. The ease of formation of 5-membered cycles from α -amino acid mixed anhydrides is also demonstrated by the conversion of acyl-aa-PEMA into 5(4H)-oxazolones.

These experiments demonstrating that the NCA path is prevailing at pH values close to neutrality in solutions equilibrated with air at present atmospheric levels of CO₂ (ca. 0.04%) suggest that the pathway must be overwhelming in natural environments with higher contents. The experiments at 2 mM HCO₃[−] are representative of present day ocean total concentration of dissolved carbonate³⁹ showing that the lifetime of aa-PEMA is expressed in tens of seconds in these media at pH 7.5. In biological media, with total carbonate concentrations approaching or exceeding 10 mM, the lifetime of mixed anhydrides would be even shorter. The early atmosphere had a CO₂ content that remains poorly constrained⁴⁰ but values similar to the present atmospheric levels⁴¹, or representing up to hundred times this value^{40,42}, are often considered. Under these conditions, aa-PEMAs would be rapidly converted into NCA before any direct conversion into peptides could take place, which discards the earlier proposed contribution of aa-AMPs in the formation of prebiotic peptides^{7–11}. Moreover, a less efficient polymerization ability of aa-PEMA and the diketopiperazine side-reaction make them improbable peptide precursors. The possibility that a very low content of CO₂ in the atmosphere could have transiently permitted mixed anhydrides to be stabilized²³ is made unlikely because it would have also required a very efficient removal of the most part of CO₂ in the whole ocean (≥ 2 mM in HCO₃[−]). On the contrary, the development of the activation pathway leading to translation must have occurred in an environment in which the role of NCA was unavoidable rather than in a local environment in which the mixed anhydrides were preserved from the presence of CO₂ and HCO₃[−] by any kind of geochemical processes. NCA can be considered not only as intermediates of the degradation pathway of adenylates but also as precursors of any kind of aa-PEMA mixed anhydrides including adenylates as well as precursors of peptides through a pathway suppressing diketopiperazine side-reaction. From this point of view, the catalysis by carbon dioxide may lead to a fast exchange among different energy-rich species capable of linking activated amino acids to phosphorylating species. This distribution of energy in a reaction network, that may have anticipated the role of ATP as an energy currency, ensured a global far from equilibrium situation that was essential even at early stages of chemical evolution⁴³. Considered from the point of view of a co-evolutionary development of peptide

Table 1 | Results of H/D exchange experiments carried out from either acetylated mixed anhydride **2b or oxazolone **4** under various conditions**

Nucleophile	Buffer	Ratio of [M + 1] MS peak in the product corrected from natural abundance ^a	
		Mixed anhydride 2b	Oxazolone 4
H ₂ O	100 mM formate ^b	<1%	<1%
H ₂ O	100 mM MES ^c	55%	≥93%
H-Ala-NH ₂	100 mM MES ^c	27% (L, L-isomer) 16% (D, L-isomer)	99% (L, L-isomer) 98% (D, L-isomer)

^aDetermination by ESI-MS: after the reaction in D₂O the residue was freeze-dried and then submitted three times to cycles of addition of H₂O and freeze-drying to convert any easily exchangeable hydrons and finally analyzed by MS;

^bBuffer ratio equivalent to that of a pH 4 buffer in H₂O;

^cbuffer ratio equivalent to that of a pH 6.5 buffer in H₂O.



and nucleotide chemistries⁴⁴ the CO₂-catalyzed pathway may then constitute a key-element in the systemic integration of the two sub-systems⁴⁵.

The fast conversion of adenylates, and more generally mixed anhydrides aa-PEMAs, into NCAs at low concentrations of CO₂ in water questions the way through which the biochemical amino acid activation evolved. As a matter of fact, aa-AMPs, possibly produced from ATP through ribozyme activity⁴⁶, would rapidly be converted into NCAs impeding the evolution of translation. Conversely, the catalytic activity of aaRSs might have evolved by acting on the thermodynamically favourable reverse reaction of aa-AMPs (formed spontaneously from NCAs) as a primitive pathway to produce ATP^{2,3}. One could argue that the NCA pathway of Fig. 3a is still active in living cells but this speculation is not supported by any experimental data. However, the mechanism of pretransfer editing of misactivated aaRSs (through which adenylates are hydrolyzed) remains uncertain⁴⁷. Any possible release of adenylates from the active site to solution⁴⁸ during this step would lead to the formation of the corresponding NCA within seconds. Whatever NCA is actually or not a biochemical metabolite, the present results indicate that living organisms probably had to limit the importance of the release of adenylates into solution after translation evolved since a conversion into NCA would certainly lead to random aminoacylation of pending amino groups likely to be harmful to protein functional integrity. From this point of view, the *N*-formylation of methionine needed to initiate ribosomal peptide synthesis in bacteria might be considered as a remnant of a period in which NCA could be released in the cytoplasm. Therefore, we conclude that the potential formation of NCAs at least influenced the development of the translation apparatus and that of the aaRS family of enzymes in order to avoid random aminoacylation and that the NCA pathway must be taken into account in evolutionary studies.

Our analyses confirm the observations made by Lacey that CO₂ is a very efficient catalyst for the conversion of adenylates. However, taking into account the probable role of NCAs and the diversity of processes made available through their intermediacy leads us to the very different conclusion that the process could be favourable to the development and evolution of life rather than solely detrimental to the role of adenylates as intermediates of peptide formation. It is also worth noting that acyl-aa-PEMA that were considered by Lacey as blocked equivalents of aa-AMPs^{22,23} does actually not constitute models of the reactivity of their parent compounds since they also undergo a spontaneous cyclization into 5(4*H*)-oxazolone. The transient formation of 5(4*H*)-oxazolone intermediates may be responsible for their efficiency in peptide formation²⁰. The mixed anhydrides formed from free amino acids as well as peptide segments turn out to constitute unlikely precursors of peptides since their reactions are actually preceded by a very efficient cyclization into uncharged intermediates that thus constitute better electrophilic agents. This observation can be related to the evolutionary advantage of phosphate derivatives⁴⁹ that is partly related to their negative charge reducing spontaneous hydrolytic degradation with respect to their enzyme-promoted reactions. From this perspective, their involvement required specific and efficient catalysts. However, the fact that NCA and 5(4*H*)-oxazolone also constitute precursors of mixed anhydrides through spontaneous processes provides a potential path through which these intermediates may have led for example to aminoacyl esters of RNA at predisposed locations^{16,23,50}.

Methods

Reagents and solvents were purchased from Bachem, Sigma-Aldrich, or Euriso-Top and used without further purification. Starting materials and products samples were prepared according to standard procedures and characterized by ¹H, ¹³C and ³¹P NMR spectrometry and HRMS (Supplementary Information). NMR analyses were performed on a Bruker Avance 300 apparatus. HPLC analyses were performed on a Waters Alliance 2690 system with a photodiode array detector 996 using a Thermo Scientific BDS Hypersil C18 5 μm 2.1 × 50 mm column; mobile phase: A: H₂O + 0.1% TFA, B: CH₃CN + 0.1% TFA; flow rate: 0.2 mL/min and two different gradients;

method A: 0 min (5% B), to 15 min (15% B), 25 min (60% B) and 26 min (100% B); method B: 0 min (5% B), to 10 min (20% B), 11 min (100% B). HPLC-ESI-MS analyses were carried out on a Waters Synapt G2-S system connected to a Waters Acquity UPLC H-Class apparatus equipped with a Acquity UPLC BEH C18, 1.7 μm 2.1 × 50 mm column; method C: A: H₂O + 0.01% formic acid, B: acetonitrile + 0.01% formic acid; flow rate: 0.5 mL/min; linear gradient 0% to 100% B over 3 min.

- Wells, T. N. C., Ho, C. K. & Fersht, A. R. Free Energy of Hydrolysis of Tyrosyl Adenylate and Its Binding to Wild-Type and Engineered Mutant Tyrosyl-tRNA Synthetases. *Biochemistry* **25**, 6603–6608 (1986).
- Pascal, R., Boiteau, L. & Commeyras, A. From the prebiotic synthesis of α-amino acids towards a primitive translation apparatus for the synthesis of peptides. *Top. Curr. Chem.* **259**, 69–122 (2005).
- Pascal, R. & Boiteau, L. Energetic constraints on prebiotic pathways: application to the emergence of translation. In Gargaud, M., Lopez-Garcia, P. & Martin, H. (eds.) *Origin and Evolution of Life: an astrobiology perspective* 247–258 (Cambridge University Press, Cambridge, 2011).
- Pham, Y. et al. Tryptophanyl-tRNA synthetase Urzyme: a model to recapitulate molecular evolution and investigate intramolecular complementation. *J. Biol. Chem.* **285**, 38590–38601 (2010).
- Li, L., Francklyn, C. & Carter, C. W. Aminoacylating urzymes challenge the RNA world hypothesis. *J. Biol. Chem.* **288**, 26856–26863 (2013).
- Wong, J. T.-F. A Co-Evolution Theory of the Genetic Code. *Proc. Natl. Acad. Sci. U.S.A.* **72**, 1909–1912 (1975).
- Katchalsky, A. & Paecht, M. Phosphate Anhydrides of Amino Acids. *J. Am. Chem. Soc.* **76**, 6042–6044 (1954).
- Lewinsohn, R., Paecht-Horowitz, M. & Katchalsky, A. Polycondensation of amino acid phosphoanhydrides. III. Polymerization of alanine adenylate. *Biochim. Biophys. Acta* **140**, 24–36 (1967).
- Paecht-Horowitz, M., Berger, J. & Katchalsky, A. Prebiotic synthesis of polypeptides by heterogeneous polycondensation of amino acid adenylates. *Nature* **228**, 636–639 (1970).
- Armstrong, D. W., Seguin, R., McNeal, C. J., Macfarlane, R. D. & Fendler, J. H. Spontaneous Polypeptide Formation from Amino Acyl Adenylates in Surfactant Aggregates. *J. Am. Chem. Soc.* **100**, 4605–4606 (1978).
- Paecht-Horowitz, M. & Katchalsky, A. Synthesis of aminoacyl-adenylates under prebiotic conditions. *J. Mol. Evol.* **2**, 91–98 (1973).
- Warden, J. T., McCullough, J. J., Lemmon, R. M. & Calvin, M. A re-examination of the zeolite-promoted, clay-mediated peptide synthesis. *J. Mol. Evol.* **4**, 189–194 (1974).
- Lee, N., Bessho, Y., Wei, K., Szostak, J. W. & Suga, H. Ribozyme-catalyzed tRNA aminoacylation. *Nat. Struct. Biol.* **7**, 28–33 (2000).
- Xu, J., Appel, B., Balke, D., Wichert, C. & Müller, S. RNA Aminoacylation Mediated by Sequential Action of Two Ribozymes and a Nonactivated Amino Acid. *ChemBioChem* **15**, 1200–1209 (2014).
- Biron, J.-P. & Pascal, R. Amino acid *N*-carboxyanhydrides: activated peptide monomers behaving as phosphate-activating agents in aqueous solution. *J. Am. Chem. Soc.* **126**, 9198–9199 (2004).
- Biron, J.-P., Parkes, A. L., Pascal, R. & Sutherland, J. D. Expeditious, potentially primordial, aminoacylation of nucleotides. *Angew. Chem. Int. Ed.* **44**, 6731–6734 (2005).
- Leman, L. J., Orgel, L. E. & Ghadiri, M. R. Amino Acid Dependent Formation of Phosphate Anhydrides in Water Mediated by Carbonyl Sulfide. *J. Am. Chem. Soc.* **128**, 20–21 (2006).
- Leman, L., Orgel, L. & Ghadiri, M. R. Carbonyl Sulfide-Mediated Prebiotic Formation of Peptides. *Science* **306**, 283–286 (2004).
- Danger, G., Boiteau, L., Cottet, H. & Pascal, R. The peptide formation mediated by cyanate revisited. *N*-carboxyanhydrides as accessible intermediates in the decomposition of *N*-carbamoylamino acids. *J. Am. Chem. Soc.* **128**, 7412–7413 (2006).
- Danger, G. et al. 5(4*H*)-oxazolones as intermediates in the carbodiimide- and cyanamide-promoted peptide activations in aqueous solution. *Angew. Chem. Int. Ed.* **52**, 611–614 (2013).
- Mullins, Jr, D. W., Senaratne, N. & Lacey, Jr, J. C. Aminoacyl-nucleotide reactions: studies related to the origin of the genetic code and protein synthesis. *Orig. Life* **14**, 597–604 (1984).
- Lacey, Jr, J. C., Senaratne, N. & Mullins, Jr, D. W. Hydrolytic properties of phenylalanyl- and *N*-acetylphenylalanyl adenylate anhydrides. *Orig. Life* **15**, 45–54 (1984).
- Wickramasinghe, N. S. M. D., Staves, M. P. & Lacey, Jr, J. C. Stereoselective, nonenzymatic, intramolecular transfer of amino acids. *Biochemistry* **30**, 2768–2772 (1991).
- Kluger, R., Loo, R. W. & Mazza, V. Biomimetically activated amino acids. Catalysis in the hydrolysis of alanyl ethyl phosphate. *J. Am. Chem. Soc.* **119**, 12089–12094 (1997).
- Brack, A. Origins Life, 1987, **17**, 367–379. Brack, A. Selective emergence and survival of early polypeptides in water. *Orig. Life* **17**, 367–379 (1987).
- Danger, G., Plasson, R. & Pascal, R. Pathways for the formation and evolution of peptides in prebiotic environments. *Chem. Soc. Rev.* **41**, 5416–5429 (2012).



27. Brack, A., Ehler, K. W. & Orgel, L. E. *N*, *N'*-carbonyldiimidazole-induced diketopiperazine formation in aqueous solution in the presence of adenosine-5'-monophosphate. *J. Mol. Evol.* **8**, 307–310 (1976).
28. Beaufils, D., Danger, G., Boiteau, L., Rossi, J.-C. & Pascal, R. Diastereoselectivity in prebiotically relevant 5(4H)-oxazolone-mediated peptide couplings. *Chem. Commun.* **50**, 3100–3102 (2014).
29. Schall, O., Suzuki, I., Murray, C., Gordon, J. & Gokel, G. Characterization of Acyl Adenyl Anhydrides: Differences in the Hydrolytic Rates of Fatty Acyl-AMP and Aminoacyl-AMP Derivatives. *J. Org. Chem.* **63**, 8661–8667 (1998).
30. Pascal, R. Catalysis through Induced Intramolecularity: What Can Be Learned by Mimicking Enzymes with Carbonyl Compounds that Covalently Bind Substrates? *Eur. J. Org. Chem.* 1813–1824 (2003).
31. Jencks, W. P. Binding energy, specificity, and enzymic catalysis: the CIRCE effect. *Adv. Enzymol. Relat. Areas Mol. Biol.* **43**, 219–410 (1975).
32. Albery, W. J. & Knowles, J. R. Evolution of enzyme function and the development of catalytic efficiency. *Biochemistry* **15**, 5631–5640 (1976).
33. Pascal, R. Do enzymes bind their substrates in the ground state because of a physico-chemical requirement? *Bioorg. Chem.* **31**, 485–493 (2003).
34. Tan, K. L. Catalysis: Temporary intramolecularity. *Nat. Chem.* **4**, 253–254 (2012).
35. Beauchemin, A. M. Site-selective reactions: Exploiting intramolecularity. *Nat. Chem.* **5**, 731–732 (2013).
36. Page, M. I. & Jencks, W. P. Entropic contribution to rate accelerations in enzymic and intramolecular reactions and the chelate effect. *Proc. Natl. Acad. Sci. U.S.A.* **68**, 1678–1683 (1971).
37. Lide, D. R. *Handbook of Chemistry and Physics*, 88th edition, 8–84 (CRC Press, Boca Raton, FL, 2008).
38. Wolfenden, R. & Snider, M. J. The Depth of Chemical Time and the Power of Enzymes as Catalysts. *Acc. Chem. Res.* **34**, 938–945 (2001).
39. Millero, F. J., Feistel, R., Wright, D. G. & McDougall, T. J. The composition of Standard Seawater and the definition of the Reference-Composition Salinity Scale 17. *Deep-Sea Res. I* **55**, 50–72 (2008).
40. Reinhard, C. T. & Planavsky, N. J. Mineralogical constraints on Precambrian pCO₂. *Nature* **474**, E1–E2 (2011).
41. Rosing, M. T., Bird, D. K., Sleep, N. H. & Bjerrum, C. J. No climate paradox under the faint early Sun. *Nature* **464**, 744–747 (2010).
42. Dauphas, N. & Kasting, J. F. Low pCO₂ in the pore water, not in the Archean air. *Nature* **474**, E2–E3 (2011).
43. Pascal, R., Pross, A. & Sutherland, J. D. Towards an evolutionary theory of the origin of life based on kinetics and thermodynamics. *Open Biol.* **3**, 130156 (2013).
44. Borsenberger, V. *et al.* Exploratory Studies to Investigate a Linked Prebiotic Origin of RNA and Coded Peptides. *Chem. Biodivers.* **1**, 203–246 (2004).
45. Ruiz-Mirazo, K., Briones, C. & de la Escosura, A. Prebiotic systems chemistry: new perspectives for the origins of life. *Chem. Rev.* **114**, 285–366 (2014).
46. Kumar, R. K. & Yarus, M. RNA-Catalyzed Amino Acid Activation. *Biochemistry* **40**, 6998–7004 (2001).
47. Nordin, B. E. & Schimmel, P. Transiently Misacylated tRNA Is a Primer for Editing of Misactivated Adenylates by Class I Aminoacyl-tRNA Synthetases. *Biochemistry* **42**, 12989–12997 (2003).
48. Manandhar, M. & Cronan, J. E. Proofreading of noncognate acyl adenylates by an acyl-coenzyme A ligase. *Chem. Biol.* **20**, 1441–1446 (2013).
49. Westheimer, F. H. Why nature chose phosphate. *Science* **235**, 1173–1178 (1987).
50. Her, S. & Kluger, R. Biomimetic protecting-group-free 2',3'-selective aminoacylation of nucleosides and nucleotides. *Org. Biomol. Chem.* **9**, 676–678 (2011).

Acknowledgments

This work was supported by a grant from the Simons Foundation (Grant Number 293065 to Z.L.). The authors thank the COST action CM1304 Emergence and Evolution of Complex Chemical Systems.

Author contributions

Z.L., J.-C.R. and R.P. designed research; Z.L. performed research; D.B. contributed new reagents and analytic tools; Z.L., J.-C.R. and R.P. analyzed data and wrote the paper.

Additional information

Supplementary information accompanies this paper at <http://www.nature.com/scientificreports>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Liu, Z., Beaufils, D., Rossi, J.-C. & Pascal, R. Evolutionary Importance of the Intramolecular Pathways of Hydrolysis of Phosphate Ester Mixed Anhydrides with Amino Acids and Peptides. *Sci. Rep.* **4**, 7440; DOI:10.1038/srep07440 (2014).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder in order to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>