

Direct N.m.r. Observation of Cell-free Conversion of (L- α -Amino- δ -adipyl)-L-cysteinyl-D-valine[†] into Isopenicillin N

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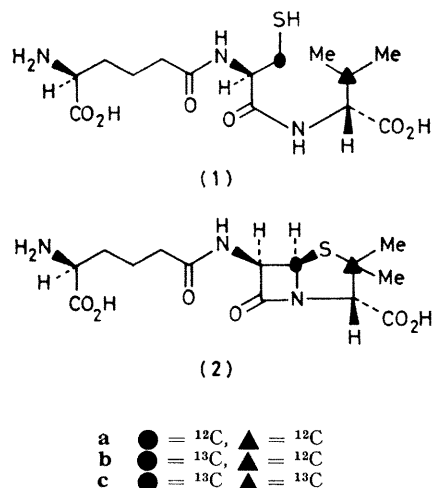
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Summary Carbon-13 n.m.r. spectroscopy has been used to observe the efficient conversion of (L- α -amino- δ -adipyl)-

L-cysteinyl-D-valine into isopenicillin N in cell-free extracts of *Cephalosporium acremonium*.

[†] α -Amino- δ -adipyl = 5-amino-5-carboxypentanoyl.

RECENT investigations with *Cephalosporium acremonium* by O'Sullivan *et al*¹ and Konomi *et al*² into the cell-free conversion of (L- α -amino- δ -adipyl)-L-cysteinyl-D-valine (LLD-ACV) (**1a**) into the corresponding penicillins have shown that the first-formed penicillin is isopenicillin N (**2a**). Furthermore, the results of O'Sullivan *et al*¹ were consistent with the hypothesis that the carbon skeleton of the LLD-ACV tripeptide was incorporated intact into the isopenicillin N molecule. We now report the direct observation of this conversion by ¹³C n.m.r. spectroscopy, which entirely confirms the previous work



When labelled tripeptide, (L- α -amino- δ -adipyl)-L-[3-¹³C]-cysteinyl-D-valine³ (**1b**) was incubated with cell-free extracts⁴ of *C. acremonium* at 5 °C in the probe of an n.m.r. spectrometer (Bruker WH 300, 25 mm n.m.r. tube), the intensity of the ¹³C n.m.r. signal[†] at 26.7 p.p.m. due to the labelled atom of the tripeptide decreased with time, whilst a new signal appeared at 67.7 p.p.m. and showed a corresponding increase in intensity with time. The latter signal appeared as a doublet in the off-resonance proton-decoupled spectrum and was ascribed to C-5 of the isopenicillin N (**2b**). A plot of the change in peak height of this signal *versus* time at 5 °C, and subsequently 9 °C, is shown in Figure 1B. Bioassay⁴ of samples of the incubation mixture showed a similar increase in β -lactam antibiotic activity with time (Figure 1A).

The yield of the conversion was high after completion of the incubation the height of the ¹³C n.m.r. signal due to labelled substrate (**1b**) was less than 20% of that due to the product (**2b**). It was not possible under these conditions

to calculate molar proportions from the signal intensities but bioassay of the final mixture indicated that the conversion was $80 \pm 8\%$.

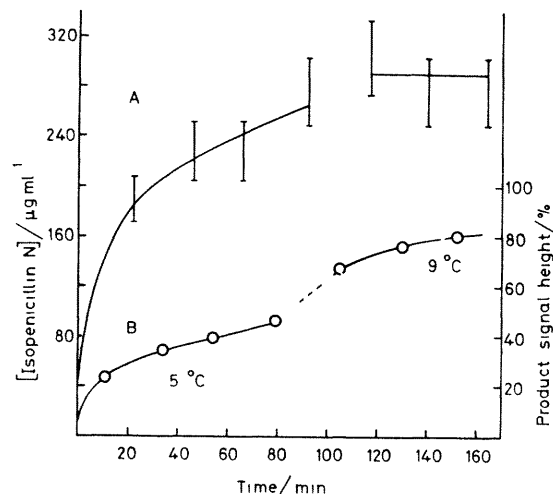


FIGURE 1 Incubation of a mixture of the LLD-ACV isotopomer (**1b**) with cell free extract of *C. acremonium* at a probe temperature of 5 °C and subsequently 9 °C. Curve A Level of antibacterial activity of samples removed from the incubation mixture as determined by a hole plate assay⁴ against *Staphylococcus aureus*. Error bars represent a reading of the assay plate to ± 0.5 mm. Curve B Increase in the height of the ¹³C n.m.r. signal at 67.7 p.p.m. due to C-5 of isopenicillin N (**2b**). The signal height is expressed as a percentage of the combined signal heights of reactant and product. The change in bioactivity with time (Curve A) shows that the change in n.m.r. signal height is proportional to concentration change of isopenicillin N.

Entirely consistent results were obtained in an analogous experiment using the doubly labelled tripeptide, (L- α -amino- δ -adipyl)-L-[3-¹³C]-cysteinyl-D-[3-¹³C]-valine³ (**1c**). During incubation the ¹³C n.m.r. signals at 26.7 and 31.8 p.p.m. of the labelled atoms of the substrate (**1c**) showed a uniform decrease in intensity whilst two new signals which appeared at 67.7 and 65.6 p.p.m. showed a corresponding increase in intensity with time (see, for example, Figure 2). The signals at 67.7 and 65.6 p.p.m. were assigned[§] to C-5 and C-2, respectively, of the isopenicillin N isotopomer (**2c**). Off-resonance proton-decoupled ¹³C n.m.r. spectra of the final incubation mixture, after removal of the protein,[¶] confirmed these assignments: the signal at 67.7 p.p.m. occurred as a doublet, whilst that at 65.6 p.p.m. was a singlet. Geminal ¹³C-¹³C coupling between C-2 and C-5 of (**2c**) was not observed under conditions where the width at half-height of the signals was 2.5 Hz.

[†] Chemical shifts are given in p.p.m. downfield from Me₄Si and were measured relative to the high field signal of the buffer, sodium 3-(N-morpholino)propanesulphonate, which had been calibrated relative to dioxan (δ 67.4 p.p.m.).

[§] The chemical shifts of C-2 and C-5 of penicillin N are given as δ 65.1 and 67.4 p.p.m., respectively, in ref. 5.

[¶] An aqueous extract of the separated protein did not show bioactivity and gave negligible signal intensity in the ¹³C n.m.r. spectrum recorded under conditions similar to those of all other spectra.

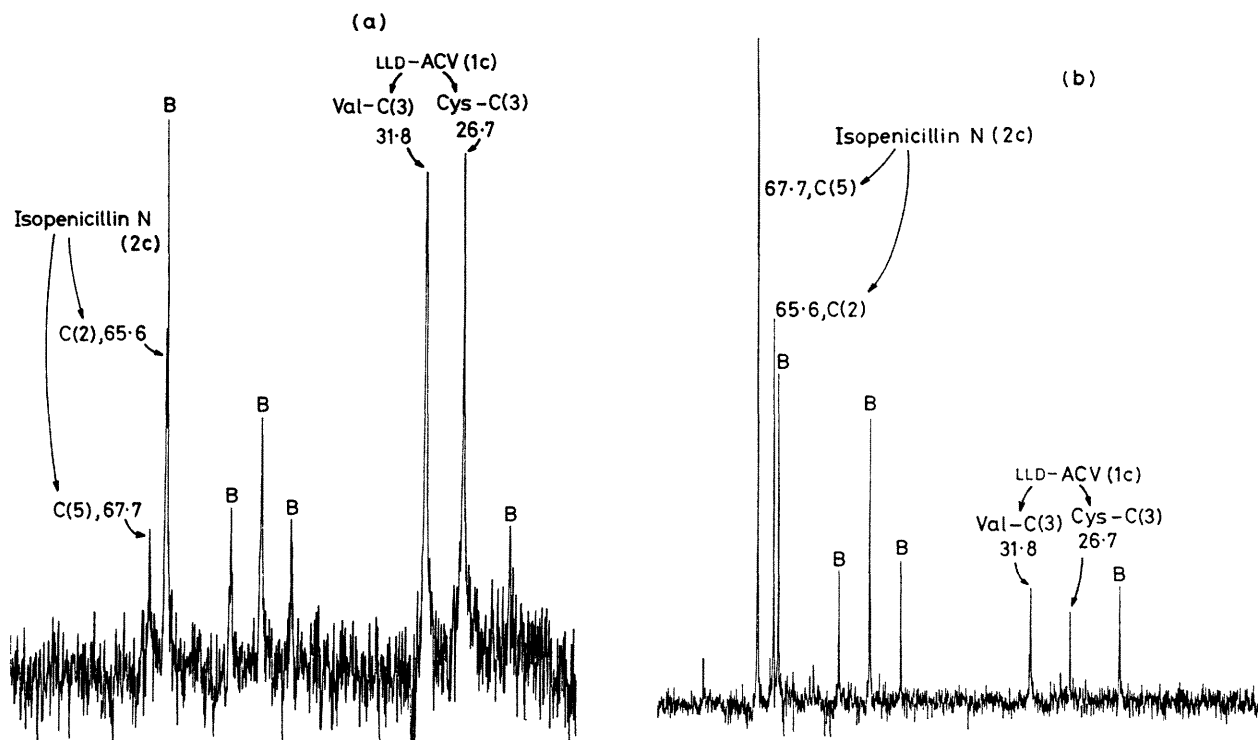


FIGURE 2. 75.47 MHz Broad-band proton-decoupled ^{13}C n.m.r. spectra (25 mm tube) of a mixture of cell-free extract and (L- α -amino- δ -adipyl)-L-[3- ^{13}C]cysteinyl-D-[3- ^{13}C]valine (**1c**) (initial concentration 1mM): (a) after incubation at 5 °C for 25 min; (b) after incubation for a further 65 min, precipitation of the protein with acetone, lyophilisation of the supernatant, and resuspension in D_2O . The incubation mixture also contained dithiothreitol (2mM), L-ascorbic acid (1mM), and FeSO_4 (0.05mM) in sodium 3-(N-morpholino)-propanesulphonate buffer at pH 7.2 (10mM) (B). Transients (1000) were accumulated in a data length of 16,384 with a sweep-width of 18.5 kHz.

It is significant that no other ^{13}C n.m.r. signals were observed in any of these experiments. If enzyme-free intermediates are involved in the cell-free conversion of LLD-ACV tripeptide to isopenicillin N, their concentrations do not appear to reach levels in the present system sufficient to permit detection by this method.

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