

Simple Ligand Effects Switch a Hydrogenase Mimic between H₂ and O₂ ActivationKyoungmok Kim,^[a] Takahiro Matsumoto,^[a, b] Andrew Robertson,^[c]
Hidetaka Nakai,^[a, b] and Seiji Ogo^{*,[a, b, d]}

Abstract: Herein, we report a [NiRu] biomimetic system for O₂-tolerant [NiFe]hydrogenases and demonstrate that electron donation to the [NiRu] center can switch the system between the activation of H₂ and O₂ through simple ligand effects by using hexamethylbenzene and pentamethylcyclopentadienyl ligands, respectively. Furthermore, we present the synthesis and direct obser-

vations of a [NiRu]–peroxo species, which was formed by the oxygenation of a Ni-SIa model [NiRu] complex, that we propose as a biomimetic analogue of O₂-bound species (OBS) of

Keywords: bioinorganic chemistry • biomimetic compounds • hydrogen • ligand effects • oxygen

O₂-tolerant [NiFe]hydrogenases. The [NiRu]–peroxo complex was fully characterized by X-ray analysis, X-ray photoelectron spectroscopy (XPS), mass spectrometry, and ¹H NMR spectroscopy. The OBS analogue was capable of oxidizing *p*-hydroquinone and sodium borohydride to turn back into the Ni-SIa model complex.

Introduction

Hydrogenases (H₂ases) are a class of enzymes that are currently receiving much interest owing to their ability to extract electrons, and hence energy, from H₂.^[1] Their active centers are based on an [Fe], [FeFe], or [NiFe] core^[2–4] and they are usually deactivated by the presence of trace amounts of O₂.^[1,5–12]

However, a subset of [NiFe]H₂ases exhibit O₂ tolerance,^[7–12] and the two currently favored proposals to explain this O₂ tolerance are physical active site blocking^[7–9] and the reduction of bound O₂ by electron-transport from {Fe-S} clusters.^[9–12] Evidence for the physical protection of the active site comes from computational modeling and X-ray structures that suggest the presence of small, hydrophobic gas channels in *Ralstonia eutropha* H16;^[7] these gas channels

would be expected to prevent the diffusion of O₂ to the active site. Support for the electron-transport mechanism has been provided in the form of structural, experimental, and kinetic studies on [NiFe]H₂ases of *Hydrogenovibrio marinus* H110^[10] and *Ralstonia eutropha* H16.^[11]

It has been proposed that electron transport followed by protonation allows the enzyme to reduce O₂ and eliminate it as water (Figure 1). However, there has been no direct experimental evidence that Ni-SIa (the EPR-silent active state) is able to activate O₂ in this manner and no O₂-bound species (OBS) have been observed so far.

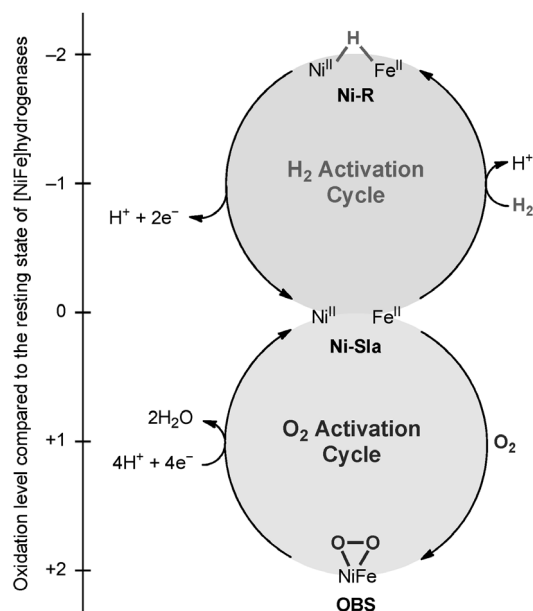
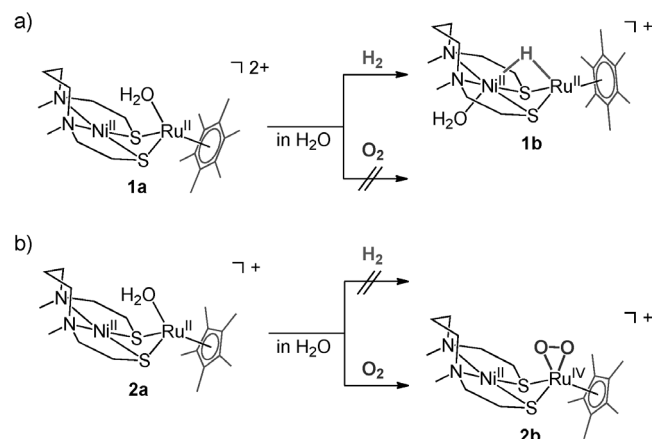


Figure 1. A proposed electron-transport cycle of O₂-tolerant [NiFe]-H₂ases.^[11] Ni-SIa: EPR-silent active state. Ni-R: EPR-silent reduced state. OBS: O₂-bound species.

- [a] K. Kim, Dr. T. Matsumoto, Dr. H. Nakai, Prof. S. Ogo
Department of Chemistry and Biochemistry
Graduate School of Engineering, Kyushu University
744 Moto-oka, Nishi-ku, Fukuoka 819-0395 (Japan)
Fax: (+81)92-802-2823
E-mail: ogotcm@mail.cstm.kyushu-u.ac.jp
- [b] Dr. T. Matsumoto, Dr. H. Nakai, Prof. S. Ogo
International Institute for Carbon-Neutral Energy Research (WPI-I2CNER)
Kyushu University
744 Moto-oka, Nishi-ku, Fukuoka 819-0395 (Japan)
- [c] Dr. A. Robertson
International Education Center, Kyushu University
744 Moto-oka, Nishi-ku, Fukuoka 819-0395 (Japan)
- [d] Prof. S. Ogo
Core Research for Evolutional Science and Technology (CREST)
Japan Science and Technology Agency (JST)
Kawaguchi Center Building, 4-1-8 Honcho
Kawaguchi-shi, Saitama 332-0012 (Japan)

As part of our efforts to apply insights from H_2 ases to find solutions for sustainable power, we have previously reported the synthesis of a biomimetic analogue of $[NiFe]-H_2$ ases: $[Ni^{II}LRu^{II}(H_2O)(\eta^6-C_6Me_6)](NO_3)_2$ (**1a**) (NO_3)₂; $L = N,N'$ -dimethyl- N,N' -bis(2-mercaptoethyl)-1,3-propanediamine). This organometallic catalyst was capable of reproducing all of the defining chemical aspects of these enzymes: heterolytic activation of H_2 , electron extraction from H_2 , and isotope-exchange with D_2O to simultaneously generate HD and D_2 .^[13] This unique ability to thoroughly mimic the properties of $[NiFe]H_2$ ases gave us confidence in using it as an organometallic substitute for the real enzymes. In addition, because this system was wholly artificial and relatively simple, we were able to make a systematic evaluation of its structural and electronic characteristics.^[13]

In common with O_2 -tolerant $[NiFe]H_2$ ases, compound **1a** could activate H_2 even in the presence of O_2 (Scheme 1 a). Whilst we could not “catch it in the act” of interacting with O_2 , we considered that a relatively small change to its electronic properties might enhance any interactions to the point of observability.



Scheme 1. Reactivity of a) compound **1a** and b) compound **2a** towards H_2 and O_2 in water.

With this goal in mind, we synthesized a variation of compound **1a** that replaced the hexamethylbenzene ligand with a pentamethylcyclopentadienyl ligand: $[Ni^{II}LRu^{II}(H_2O)(\eta^5-C_5Me_5)](NO_3)_2$, **2a**) (NO_3)₂. The pentamethylcyclopentadienyl anion is a stronger σ -donor than hexamethylbenzene and thus this substitution allowed us to better establish the importance of aromatic electronic donors to the reactivity of our system, whilst providing an insight into O_2 activation. Electron donation was especially relevant to the study of these H_2 ases because it has been suggested that a proximal $\{Fe-S\}$ cluster can provide electrons to activate bound O_2 at the active site and thereby facilitate its removal.

Herein, we report that σ -donation to the Ru center of compound **2a** switched the activity of the system between H_2 and O_2 activation. We also provide crystallographic evidence for a $[NiRu]$ -peroxo complex, which may act as a biomimetic analogue of the OBS of H_2 ases, and thereby shed light on the mechanism of the O_2 tolerance.

Results and Discussion

Two versions of our system were synthesized: compounds **1a** and **2a**. The synthesis of compound **1a** has been reported previously,^[13] whilst complex **2a**) (NO_3)₂ was synthesized by the treatment of a Ni^{II} complex, $[Ni^{II}L]$,^[14] with Ru^{II} complex $[Ru^{II}(\eta^5-C_5Me_5)(CH_3CN)_3](NO_3)_2$ ^[15] in water and was isolated as a black powder. The structure of compound **2a** was characterized by electrospray ionization mass spectrometry (ESI-MS), 1H NMR and IR spectroscopy, and elemental analysis.

We examined the reactivity of compounds **1a** and **2a** toward 0.1 MPa of H_2 or O_2 in water at 25 °C (Scheme 1). H_2 or O_2 gas was bubbled through aqueous solutions of compounds **1a** or **2a**, and the reactions were monitored by ESI-MS and UV/Vis spectroscopy. As previously reported, compound **1a** activated H_2 to form a hydride complex, $[Ni^{II}(H_2O)L(\mu-H)Ru^{II}(\eta^6-C_6Me_6)](NO_3)_2$ (**1b**) (NO_3)₂,^[13] but showed no reactivity toward O_2 . By contrast, compound **2a** did not react with H_2 but acquired O_2 to afford a $Ni^{II}Ru^{IV}$ -peroxo complex, $[Ni^{II}LRu^{IV}(\eta^2-O_2)(\eta^5-C_5Me_5)](NO_3)_2$ (**2b**) (NO_3)₂. Compound **2b** was characterized by X-ray analysis (Figure 2), X-ray photoelectron spectroscopy (XPS, Figure 3), ESI-MS (Figure 4), 1H NMR and IR spectroscopy, and elemental analysis.

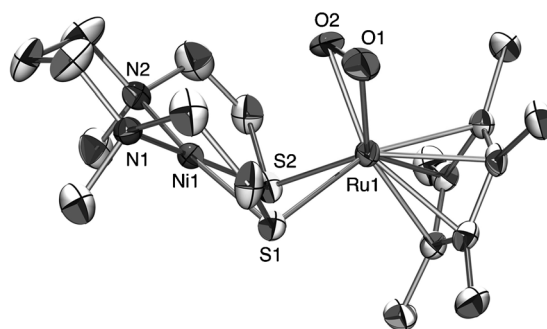


Figure 2. An ORTEP of complex **2b**) (CF_3SO_3), ellipsoids set at 50% probability. The counteranion (CF_3SO_3) and hydrogen atoms are omitted for clarity. Selected distances [Å] and angles [°]: $Ni1 \cdots Ru1$ 3.0781(7), $O1-O2$ 1.404(5), $Ru1-O1$ 2.000(3), $Ru1-O2$ 2.027(3), $Ru1-S1$ 2.406(1), $Ru1-S2$ 2.393(1), $Ni1-S1$ 2.179(1), $Ni1-S2$ 2.171(1), $Ni1-N1$ 1.968(4), $Ni1-N2$ 1.966(4); $Ni1-S1-Ru1$ 84.17(4), $Ni1-S2-Ru1$ 84.67(4).

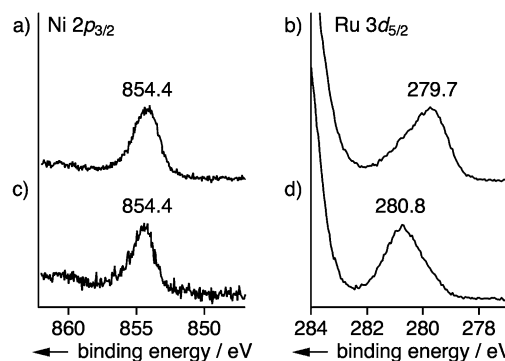


Figure 3. XPS spectra of a) $Ni\ 2p$ and b) $Ru\ 3d$ regions for complex **2a**) (NO_3)₂. XPS spectra of c) $Ni\ 2p$ and d) $Ru\ 3d$ regions for complex **2b**) (NO_3)₂.

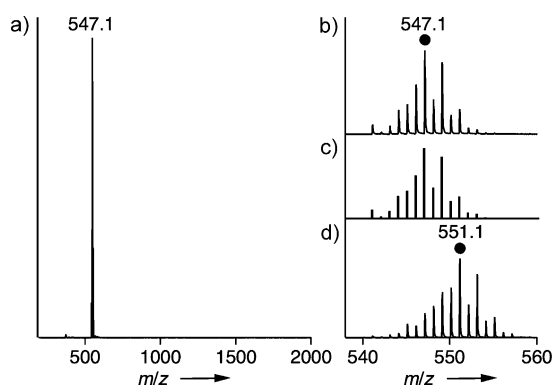


Figure 4. a) Positive-ion ESI mass spectrum of complex **[2b](NO₃)** in water. b) The signal at m/z 547.1 corresponds to complex **[2b]⁺**. c) Calculated isotopic distribution for complex **[2b]⁺**. d) Positive-ion ESI mass spectrum of complex **[¹⁸O-labeled 2b](NO₃)** in water.

This drastic change in reactivity towards H₂ and O₂ arose from the electronic properties of the aromatic ligands. The negatively charged pentamethylcyclopentadienyl ligand has a stronger electron-donating ability than the neutral hexamethylbenzene ligand, which presumably facilitated the Ru^{II} center in reaching the high-valent Ru^{IV} state.

To confirm the presence of the O₂ adduct, complex **[2b](CF₃SO₃)** was isolated and an X-ray structure was successfully obtained (Figure 2). Dark-brown crystals of complex **[2b](CF₃SO₃)** that were suitable for X-ray analysis were obtained by diffusion of diethyl ether into a solution of complex **[2b](CF₃SO₃)** in acetonitrile, which was prepared from the replacement of the counteranion of NO₃[−] in complex **[2b](NO₃)** by CF₃SO₃[−]. The framework of compound **2b** is based around a NiS₂Ru “butterfly” core, in a similar manner to that of compound **1a**, but with minor parametric differences. The Ni1···Ru1 separation was 3.0781(7) Å, which was slightly shorter than that in compound **1a** (3.1611(6) Å). The Ni-S-Ru bite angles were 84.17(4) and 84.67(4)°, slightly smaller than those in compound **1a** (86.81(4) and 87.20(5)°, respectively). The peroxo ligand in compound **2b** was coordinated to the Ru center in a side-on fashion, with an O–O distance of 1.404(5) Å. This distance corresponded to that reported previously for the two-electron-reduction of O₂ into peroxide (O₂^{2−})^[16] and was within the range found for the previously-reported [Ru(η²-O₂)(η⁵-C₅Me₅)] complexes (1.316(6)–1.416(5) Å).^[17–19] This distance was slightly longer than that found in another dinuclear [NiRu] peroxo complex, [Ni(S₂N₂)Ru(η²-O₂)(η⁵-C₅Me₅)]⁺ (1.371(8) Å, S₂N₂ = *N,N'*-bis(2-mercaptoethyl)-*N,N'*-dimethyl-1,3-diaminoethane),^[17] and comparable to that reported in a dinuclear [RhRu] peroxo complex, [Rh(Tp*)(μ-SPh)₂Ru(η²-O₂)(η⁵-C₅Me₅)] (1.397(3) Å, Tp* = hydrotris(3,5-dimethylpyrazol-1-yl)).^[18]

Given the similarities between compound **1a** and [NiFe]-H₂ases, in view of the active site structure and function,^[13] we propose that compound **2b** is a good candidate as a mimic of the OBS of [NiFe]H₂ases. Although this O₂ adduct is not unique in the field of such catalysts, it is the

first time that its precise relationship pertaining to H₂ activation has been studied, together with an investigation of the relevant chemical properties.

The peroxo ligand in compound **2b** was formed by two-electron reduction of O₂ by compound **2a**. The two electrons came from the Ru^{II} center in compound **2a**, which was investigated by XPS. Figure 3 shows that the binding energies of Ni 2p_{3/2} and Ru 3d_{5/2} in compound **2a** are 854.4 and 279.7 eV, respectively. The formation of compound **2b** by the oxygenation of compound **2a** resulted in an increase in the binding energy of Ru 3d_{5/2} (280.8 eV) and no change in the binding energy of Ni 2p_{3/2} (854.4 eV). These results indicated that Ni^{II}Ru^{II} complex **2a** underwent two-electron oxidation by O₂ to form Ni^{II}Ru^{IV}-peroxo complex **2b**.^[20]

Figure 4 shows a positive-ion ESI mass spectrum of complex **[2b](NO₃)** in water under a N₂ atmosphere. The prominent signal at m/z 547.1 ($I=100\%$ in the range m/z 200–2000) displayed a characteristic isotopic distribution that matched well with the calculated isotopic distribution for complex **[2b]⁺**. To establish the origin of the peroxo ligand of compound **2b**, ¹⁸O-labeled **2b** was synthesized by the reaction of compound **2a** with ¹⁸O₂ in water. ESI-MS results showed that the signal at m/z 547.1 was shifted to m/z 551.1. This result indicated that the two ¹⁸O atoms from gaseous ¹⁸O₂ had been incorporated into the ¹⁸O-labeled **2b**.

Peroxo complex **2b** was formed within 1 second by exposure of compound **2a** to O₂ in water at 25 °C. This rapid conversion of compound **2a** into compound **2b** resulted in the growth of an absorption band at 390 nm; kinetic studies of the oxygenation of compound **2a** were subsequently performed at −20 °C in CH₃CN. The reaction of compound **2a** with excess O₂ in O₂-saturated CH₃CN at −20 °C obeyed pseudo-first-order kinetics with respect to compound **2a** ($v=k_{\text{obs}}[\mathbf{2a}]$) over 5 half-lives, as monitored by UV/Vis spectroscopy (Figure 5). The rate constant (k_{obs}) was determined as $k_{\text{obs}}=1.2\times10^{-3}\text{ s}^{-1}$.

Following the isolation of the OBS model compound (**2b**), we turned to study its ability to oxidize an external

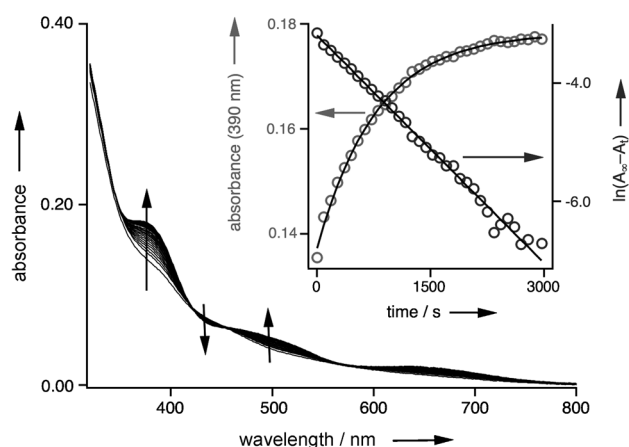


Figure 5. Changes in the UV/Vis spectra during the reaction of complex **[2a](NO₃)** (7.1 μM) with O₂ in O₂-saturated CH₃CN at −20 °C. Inset: time profile of the absorbance at 390 nm and first-order kinetic plots for the change in absorbance at 390 nm of complex **[2a](NO₃)**.

substrate. We found that compound **2b** was capable of oxidizing *para*-hydroquinone in water (pH 2.0, under a N₂ atmosphere) into the corresponding *para*-benzoquinone in quantitative yield. Analysis of the kinetic data for the oxidation of *para*-hydroquinone was performed by monitoring the decrease in the absorption band of compound **2b** at 390 nm, which revealed that the reaction of compound **2b** with 10 equivalents of *para*-hydroquinone in water (pH 2.0, 15°C) followed pseudo-first-order kinetics with respect to compound **2b** ($v = k_{\text{obs}}[\mathbf{2b}]$) over 5 half-lives (Figure 6). The rate constant (k_{obs}) was obtained as a slope of the linear first-order plot ($k_{\text{obs}} = 2.4 \times 10^{-3} \text{ s}^{-1}$).

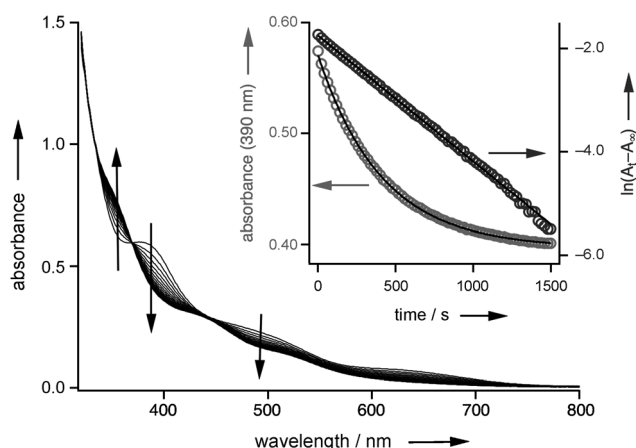
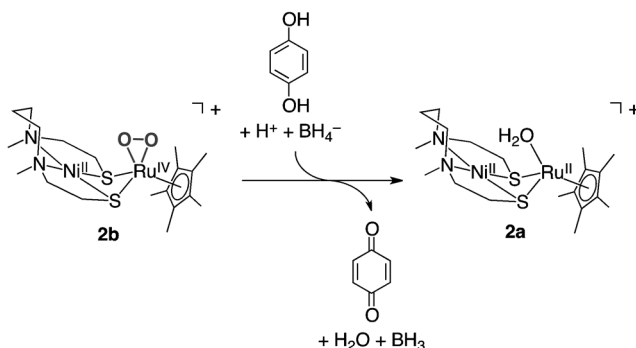


Figure 6. Changes in the UV/Vis spectra for the reaction of complex **[2b](NO₃)** (21 μM) with 10 equivalents of *p*-hydroquinone in water (pH 2.0, 15°C). Inset: time profile of the absorbance at 390 nm and first-order kinetic plots for the change in absorbance at 390 nm of complex **[2b](NO₃)**.

Compound **2b** was able to acquire electrons and protons from *para*-hydroquinone and NaBH₄ in water to regenerate into the initial Ni-SIa model compound (**2a**; Scheme 2).^[21] This recovery process was monitored by ESI-MS. The signal at m/z 547.1, which was derived from compound **2b**, completely disappeared following the addition of 10 equivalents



Scheme 2. Regeneration of the Ni-SIa model complex (**2a**) from the OBS model complex (**2b**) by acquiring electrons and protons from *p*-hydroquinone and NaBH₄ in water.

of *para*-hydroquinone and NaBH₄ in water, with a concomitant appearance of a signal at m/z 515.2, which was derived from compound **2a** (pH 2.0, 15°C, under a N₂ atmosphere, 30 min, Figure 7). Exposure of the regenerated compound **2a** to O₂ led again to the formation of compound **2b**.

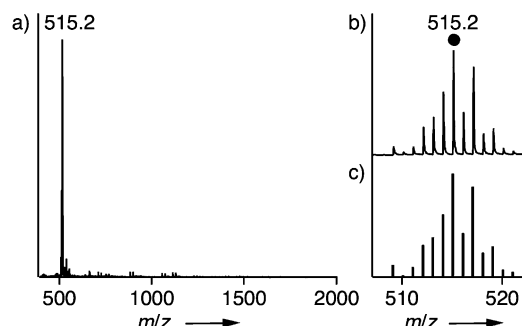


Figure 7. a) Positive-ion ESI mass spectrum obtained from the reaction of complex **[2b]** (NO₃) with 10 equivalents of *p*-hydroquinone and NaBH₄ in water (pH 2.0, 15°C, under a N₂ atmosphere, 30 min). b) Signal at m/z 515.2 fragment **[2a-H₂O]⁺**. c) Calculated isotopic distribution for fragment **[2a-H₂O]⁺**.

These results allowed us to construct a cycle of O₂ activation for [NiRu] biomimetic models of O₂-tolerant [NiFe]-H₂ases (Figure 8). Exposure of the Ni-SIa model compound (**2a**) to O₂ in water to form the OBS model compound (**2b**) corresponded thereby to the deactivation step proposed for natural [NiFe]H₂ases. The deactivated species (**2b**) then ob-

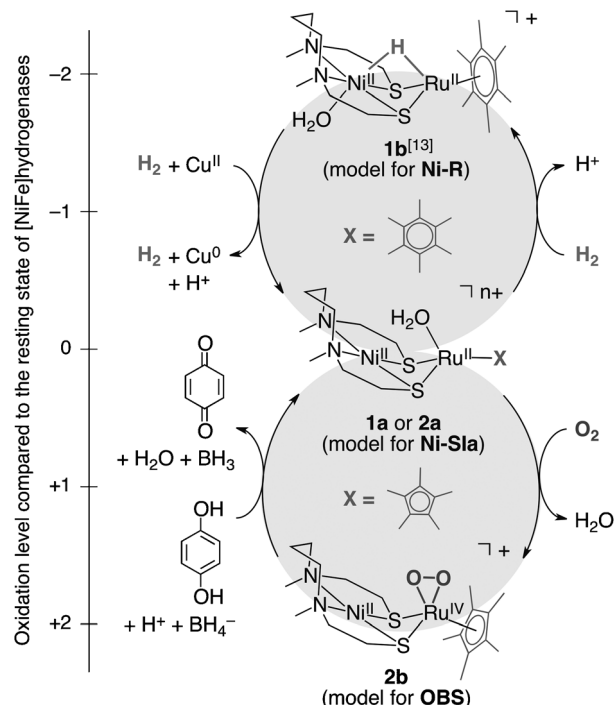


Figure 8. The electron-transport O₂-tolerant mechanism with [NiRu] biomimetic models for [NiFe]H₂ases. **1a**: X = C₆Me₆, $n = 2$; **2a**: X = C₅Me₅, $n = 1$.

tained electrons and protons to return to Ni-SIIa species (**2a**); this step corresponded to the recovery step.

Whilst we required two closely related, but different, ligands for our system to follow both cycles, natural O₂-tolerant enzymes can accomplish a similar change in electron donation by simply switching the conveyance of electrons from a proximal {Fe-S} cluster. Hence, although we had to construct our H₂- and O₂-activating systems from different starting materials, the system as a whole was a good mimic for the ability of native enzymes to switch between different activating modes of its active site.

Conclusions

The σ -donating ability of two closely related ligands in [Ni-Fe]H₂ase mimics determined whether the [NiRu] catalyst was H₂-activating, to form a reactive hydride, or O₂-activating, to form an active [NiRu]-peroxo complex. This behavior was considered to mimic the ability of certain [NiFe]-H₂ases to switch between H₂ and O₂ activation by enabling/disabling electron transport from {Fe-S} clusters. The O₂ activation cycle proceeded through a peroxo adduct, and we propose that a similar adduct may be found in O₂-tolerant [NiFe]H₂ases. These findings are expected to be of use in further investigating the properties of natural [NiFe]H₂ases and in designing future catalysts for the controlled activation of H₂ and O₂.

Experimental Section

Materials and Methods

All experiments were carried out under a N₂ atmosphere by using standard Schlenk techniques and a glove box. CH₃CN was distilled over CaH₂ under a N₂ atmosphere prior to use. Mono-Ni^{II} complex [Ni^{II}L] (L = *N,N'*-dimethyl-*N,N'*-bis(2-mercaptoethyl)-1,3-propanediamine),^[14] mono-Ru^{II} complex [Ru^{II}(η^5 -C₅Me₅)(CH₃CN)₃](NO₃)₃,^[15] Ni^{II}Ru^{II}-aqua complex [Ni^{II}LRu^{II}(H₂O)(η^6 -C₆Me₆)](NO₃)₂ (**[1a]**(NO₃)₂),^[13] and Ni^{II}Ru^{II}-hydride complex [Ni^{II}(H₂O)L(μ -H)Ru^{II}(η^6 -C₆Me₆)](NO₃) (**[1b]**(NO₃))^[13] were prepared according to literature procedures. D₂O was purchased from Cambridge Isotope Laboratories, Inc., ¹⁸O₂ was purchased from SHOKO CO., LTD., and water was purchased from Wako Pure Chemical Industries, Ltd.; these chemicals were used without further purification.

¹H NMR spectra were recorded on a JEOL JNM-AL300 spectrometer at 25 °C. ¹H NMR experiments (D₂O) were performed in a NMR tube (diameter = 1.5 mm), with [D₄]3-(trimethylsilyl)propionic-2,2,3,3 acid sodium salt (TSP, 100 mM) as the reference compound (the methyl protons or the carbon resonance were set at 0.00 ppm). ESI-MS data were obtained by using a JEOL JMS-T100LC AccuTOF mass spectrometer. IR spectra of solid compounds (KBr disks) were recorded on a Thermo Nicolet NEXUS 8700 FTIR instrument from 650 to 4000 cm⁻¹, by using 2 cm⁻¹ standard resolution at 25 °C. UV/Vis spectra were recorded on an Otsuka Electronics photodiode array spectrometer MCPD-2000 with an Otsuka Electronics optical fiber attachment (light path length 0.80 cm). Elemental analysis was obtained on a PerkinElmer 2400II series CHNS/O analyzer. The oxidation states of the metal centers were determined by X-ray photoelectron spectroscopy (XPS), by using a Nissan Arc PHI 5800 system with an Al/Mg dual-anode X-ray source. Binding energies were collected with a C 1s binding energy of the carbon atoms of the ligand in the specimens as 284.5 eV.^[22] The pH (pD) value of the solution

was adjusted to 2.0 by using 10 mM HNO₃/water (DNO₃/D₂O) and was determined with a pH meter (TOA, HM-20J) that was equipped with a pH combination electrode (TOA, GST-5725C). pD values were corrected by adding 0.4 to the observed values (i.e., pD = pH meter reading + 0.4).^[23]

[Ni^{II}LRu^{II}(H₂O)(η^5 -C₅Me₅)](NO₃) (**[2a]**(NO₃))

A solution of a Ni^{II} complex ([Ni^{II}L], 280 mg, 1.0 mmol) in water (25 mL) was added to a solution of the Ru^{II} complex [Ru^{II}(η^5 -C₅Me₅)(CH₃CN)₃](NO₃) (420 mg, 1.0 mmol) in water (25 mL) under a N₂ atmosphere to afford a black solution. After stirring for 2 h, the solvent was removed under reduced pressure to yield complex **[2a]**(NO₃) as a black powder. The powder was washed with a small amount of water (2.0 mL) and dried in vacuo (yield: 85%, based on [Ru^{II}(η^5 -C₅Me₅)(CH₃CN)₃](NO₃)). ¹H NMR (300 MHz, D₂O, 25 °C): δ = 1.68–1.89, 1.94–2.10, and 2.88–2.95 (m, 14H; CH₂), 2.16 (s, 15H; C₅(CH₃)₅), 2.69 ppm (s, 6H; NCH₃); ESI-MS (in water): *m/z* (%) 515.2 **[2a-H₂O]**⁺, 200–2000 (100); FTIR (KBr disk): $\tilde{\nu}$ = 3430 (O–H), 2895 (aliphatic C–H), 1625 (aromatic C=C), 1460 (aromatic C=C), 1431 (aromatic C=C), 1384 cm⁻¹ (NO₃); elemental analysis calcd (%) for C₁₆H₃₇N₃O₄S₂NiRu: C 38.33, H 6.26, N 7.06; found: C 38.49, H 6.31, N 7.26.

[Ni^{II}LRu^{IV}(η^2 -O₂)(η^5 -C₅Me₅)](NO₃) (**[2b]**(NO₃))

Complex **[2a]**(NO₃) (66 mg, 0.11 mmol) was dissolved in water (30 mL) and stirred under an O₂ atmosphere for 2 h to afford a dark-brown solution. The solvent was evaporated to yield complex **[2b]**(NO₃) as a dark-brown powder (yield: 96%, based on **[2a]**(NO₃)). ¹H NMR (300 MHz, in D₂O, 25 °C): δ = 2.03–2.45, 2.73–2.83, and 3.40–3.50 (m, 14H; CH₂), 1.62 (s, 15H; C₅(CH₃)₅), 2.72 ppm (s, 6H; NCH₃); ESI-MS (in water): *m/z* (%) 547.1 **[2b]**⁺, 200–2000 (100); FTIR (KBr disk): $\tilde{\nu}$ = 3420 (O–H), 2917 (aliphatic C–H), 1629 (aromatic C=C), 1459 (aromatic C=C), 1430 (aromatic C=C), 1384 cm⁻¹ (NO₃); elemental analysis calcd (%) for C₁₆H₃₆N₃O₅S₂NiRu (**[2b]**(NO₃)·0.5H₂O): C 36.90, H 5.87, N 6.79; found: C 36.98, H 5.80, N 7.09.

[Ni^{II}LRu^{IV}(η^2 -¹⁸O₂)(η^5 -C₅Me₅)](NO₃) (**[18O-labeled 2b]**(NO₃))

Isotope-labeled complex [¹⁸O-labeled **2b]**(NO₃) was obtained in the same manner as described for the preparation of complex **[2b]**(NO₃) except that ¹⁸O₂ was used instead. ESI-MS (in water): *m/z* (%) 551.1 [¹⁸O-labeled **2b]**⁺, 200–2000 (100).

Typical Procedure for the Regeneration of Complex **[2a]**(NO₃) from Complex **[2b]**(NO₃)

To a solution of complex **[2b]**(NO₃) (30 mg, 50 μ mol) in water (3.0 mL) at pH 2.0 was added *p*-hydroquinone (55 mg, 0.50 mmol) and the resulting solution was stirred for 1 h under a N₂ atmosphere to afford a red-brown solution. The addition of NaBH₄ (19 mg, 0.50 mmol) to this solution afforded a black solution of complex **[2a]**(NO₃) with gas evolution. This reaction was monitored by ESI-MS and by ¹H NMR and UV/Vis spectroscopy. The yield of complex **[2a]**(NO₃) was 20%, based on complex **[2b]**(NO₃), which was determined by ¹H NMR spectroscopy, using 1,4-dioxane as an internal standard in D₂O.

Quantitative Analysis of *p*-Benzoquinone from the Reaction of Complex **[2b]**(NO₃) with *p*-Hydroquinone by ¹H NMR Spectroscopy

To a solution of complex **[2b]**(NO₃) (4.5 mg, 7.4 μ mol) in D₂O (1.0 mL) at pD 2.0 was added *p*-hydroquinone (8.1 mg, 74 μ mol) under a N₂ atmosphere. The resulting solution was stirred for 30 min. The yield of *p*-benzoquinone as an oxidized product was 95%, based on complex **[2b]**(NO₃), which was determined by ¹H NMR spectroscopy, using 1,4-dioxane as an internal standard in D₂O (*p*-benzoquinone: δ = 6.83 ppm (s, 4H; C₆H₄)).

Kinetic Measurements of the Reaction of Complex **[2a]**(NO₃) with O₂

The reaction of complex **[2a]**(NO₃) with O₂ in O₂-saturated CH₃CN at –20 °C under an O₂ atmosphere was followed by UV/Vis spectroscopy (390 nm). O₂ gas was bubbled through CH₃CN (6.9 mL) in a Schlenk flask (10 mL) at –20 °C for 30 min to give an O₂-saturated CH₃CN solu-

tion. To the resulting solution was added a solution of complex **[2a]**(NO₃) in CH₃CN (0.50 mM, 0.10 mL). The final concentration of complex **[2a]**(NO₃) was 7.1 μM. The rate constant (k_{obs}) was determined by a least-squares curve fit ($k_{\text{obs}} = 1.2 \times 10^{-3} \text{ s}^{-1}$).

Kinetic Measurements of the Reaction of Complex **[2b]**(NO₃) with *p*-Hydroquinone

The reaction of complex **[2b]**(NO₃) with *p*-hydroquinone in water (pH 2.0) at 15 °C under a N₂ atmosphere was followed by UV/Vis spectroscopy (390 nm). A solution of *p*-hydroquinone (15 mM, 0.10 mL) in water was added at 15 °C to water (pH 2.0, 6.8 mL) in a Schlenk flask (25 mL). To the resulting solution was added a solution of complex **[2b]**(NO₃) in water (1.5 mM, 0.10 mL). The final concentrations of complex **[2b]**(NO₃) and *p*-hydroquinone were 21 μM and 210 μM, respectively. The rate constant (k_{obs}) was determined by a least-squares curve fit ($k_{\text{obs}} = 2.4 \times 10^{-3} \text{ s}^{-1}$).

X-ray Crystallographic Analysis

Dark-brown crystals of complex **[2b]**(CF₃SO₃) that were suitable for X-ray analysis were obtained by diffusion of Et₂O into a solution of complex **[2b]**(CF₃SO₃) in CH₃CN, which was prepared from the replacement of the counteranion of NO₃[−] in complex **[2b]**(NO₃) by CF₃SO₃[−] by addition of NaCF₃SO₃ in CH₃CN. Measurements were made on a Rigaku/MS Saturn CCD diffractometer with graphite monochromated MoK_α radiation ($\lambda = 0.71070$). All calculations were performed by using the teXsan crystallographic software package of the Molecular Structure Corporation. CCDC 785544 (**[2b]**(CF₃SO₃)) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre at www.ccdc.cam.ac.uk/data_request/cif. Crystal data: C₂₀H₃₅F₃N₂NiO₅RuS₃; $M_r = 696.45$; orthorhombic; $a = 13.0153(21) \text{ \AA}$, $b = 14.4222(23) \text{ \AA}$, $c = 27.9367(47) \text{ \AA}$; $V = 5244.0(15) \text{ \AA}^3$; space group *Pbca*; $Z = 8$; $T = 123.2 \text{ K}$; 38506 reflns measured; 5971 unique reflns ($R_{\text{int}} = 0.109$); $R_1 = 0.048$; $wR = 0.123$; GOF on F^2 : 0.998.

Acknowledgements

This work was supported by the World Premier International Research Center Initiative (WPI), the Global COE Program “Science for Future Molecular Systems”, the G30 Program, and Grants-in-Aid: 18065017 (Chemistry of Concerto Catalysis), 19205009, and 23655053 from the Ministry of Education, Culture, Sports, Science and Technology (MEXT), Japan, and by the Basic Research Programs CREST Type “Development of the Foundation for Nano-Interface Technology” from the JST, Japan.

- [1] a) C. Tard, C. J. Pickett, *Chem. Rev.* **2009**, *109*, 2245–2274; b) F. A. Armstrong, N. A. Belsey, J. A. Cracknell, G. Goldet, A. Parkin, E. Reisner, K. A. Vincent, A. F. Wait, *Chem. Soc. Rev.* **2009**, *38*, 36–51; c) M. Rakowski DuBois, D. L. DuBois, *Chem. Soc. Rev.* **2009**, *38*, 62–72; d) F. Gloaguen, T. B. Rauchfuss, *Chem. Soc. Rev.* **2009**, *38*, 100–108; e) S. Vogt, E. J. Lyon, S. Shima, R. K. Thauer, *J. Biol. Inorg. Chem.* **2008**, *13*, 97–106; f) G. J. Kubas, *Chem. Rev.* **2007**, *107*, 4152–4205; g) A. L. de Lacey, V. M. Fernández, M. Rousset, R. Cammack, *Chem. Rev.* **2007**, *107*, 4304–4330; h) W. Lubitz, E. Reijerse, M. van Gastel, *Chem. Rev.* **2007**, *107*, 4331–4365; i) P. E. M. Siegbahn, J. W. Tye, M. B. Hall, *Chem. Rev.* **2007**, *107*, 4414–4435; j) V. Artero, M. Fontecave, *Coord. Chem. Rev.* **2005**, *249*, 1518–1535; k) I. P. Georgakaki, L. M. Thomson, E. J. Lyon, M. B. Hall, M. Y. Darensbourg, *Coord. Chem. Rev.* **2003**, *238–239*, 255–266; l) R. H. Morris in *Concepts and Models in Bioinorganic Chemistry* (Eds.: H.-B. Kraatz, N. Metzler-Nolte), Wiley-VCH: Weinheim, Germany, **2006**, Chap. 15, pp. 331–362.
- [2] S. Shima, O. Pilak, S. Vogt, M. Schick, M. S. Stagni, W. Meyer-Klaucke, E. Warkentin, R. K. Thauer, U. Ermler, *Science* **2008**, *321*, 572–575.
- [3] a) J. W. Peters, W. N. Lanzilotta, B. J. Lemon, L. C. Seefeldt, *Science* **1998**, *282*, 1853–1858; b) Y. Nicolet, C. Piras, P. Legrand, E. C. Hatchikian, J. C. Fontecilla-Camps, *Structure* **1999**, *7*, 13–23.
- [4] a) A. Volbeda, M.-H. Charon, C. Piras, E. C. Hatchikian, M. Frey, J. C. Fontecilla-Camps, *Nature* **1995**, *373*, 580–587; b) A. Volbeda, E. Garcin, C. Piras, A. L. de Lacey, V. M. Fernandez, E. C. Hatchikian, M. Frey, J. C. Fontecilla-Camps, *J. Am. Chem. Soc.* **1996**, *118*, 12989–12996; c) Y. Higuchi, T. Yagi, N. Yasuoka, *Structure* **1997**, *5*, 1671–1680; d) Y. Higuchi, H. Ogata, K. Miki, N. Yasuoka, T. Yagi, *Structure* **1999**, *7*, 549–556.
- [5] H. Ogata, S. Hirota, A. Nakahara, H. Komori, N. Shibata, T. Kato, K. Kano, Y. Higuchi, *Structure* **2005**, *13*, 1635–1642.
- [6] a) C. Léger, S. Dementin, P. Bertrand, M. Rousset, B. Guigliarelli, *J. Am. Chem. Soc.* **2004**, *126*, 12162–12172; b) S. E. Lamle, S. P. J. Albracht, F. A. Armstrong, *J. Am. Chem. Soc.* **2004**, *126*, 14899–14909.
- [7] a) T. Buhrke, O. Lenz, N. Krauss, B. Friedrich, *J. Biol. Chem.* **2005**, *280*, 23791–23796; b) M. Ludwig, J. A. Cracknell, K. A. Vincent, F. A. Armstrong, O. Lenz, *J. Biol. Chem.* **2009**, *284*, 465–477.
- [8] O. Duché, S. Elsen, L. Cournac, A. Colbeau, *FEBS J.* **2005**, *272*, 3899–3908.
- [9] a) A. Parkin, G. Goldet, C. Cavazza, J. C. Fontecilla-Camps, F. A. Armstrong, *J. Am. Chem. Soc.* **2008**, *130*, 13410–13416; b) M. C. Marques, R. Coelho, A. L. de Lacey, I. A. C. Pereira, P. M. Matias, *J. Mol. Biol.* **2010**, *396*, 893–907; c) C. S. A. Baltazar, M. C. Marques, C. M. Soares, A. M. de Lacey, I. A. C. Pereira, P. M. Matias, *Eur. J. Inorg. Chem.* **2011**, 948–962.
- [10] Y. Shomura, K.-S. Yoon, H. Nishihara, Y. Higuchi, *Nature* **2011**, *479*, 253–256.
- [11] a) T. Burgdorf, O. Lenz, T. Buhrke, E. van der Linden, A. K. Jones, S. P. J. Albracht, B. Friedrich, *J. Mol. Microbiol. Biotechnol.* **2005**, *10*, 181–196; b) J. A. Cracknell, A. F. Wait, O. Lenz, B. Friedrich, F. A. Armstrong, *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 20681–20686; c) T. Goris, A. F. Wait, M. Saggi, J. Fritsch, N. Heidary, M. Stein, I. Zebger, F. Lendzian, F. A. Armstrong, B. Friedrich, O. Lenz, *Nat. Chem. Biol.* **2011**, *7*, 310–318; d) J. Fritsch, P. Scheerer, S. Frielingsdorf, S. Kroschinsky, B. Friedrich, O. Lenz, C. M. T. Spahn, *Nature* **2011**, *479*, 249–252.
- [12] a) A. Volbeda, L. Martin, C. Cavazza, M. Matho, B. W. Faber, W. Roseboom, S. P. J. Albracht, E. Garcin, M. Rousset, J. C. Fontecilla-Camps, *J. Biol. Inorg. Chem.* **2005**, *10*, 239–249; b) S. E. Lamle, S. P. J. Albracht, F. A. Armstrong, *J. Am. Chem. Soc.* **2005**, *127*, 6595–6604; c) M. Pandelia, V. Fourmond, P. Tron-Infossi, E. Lojou, P. Bertrand, C. Léger, M. Giudici-Orticoni, W. Lubitz, *J. Am. Chem. Soc.* **2010**, *132*, 6991–7004; d) M. J. Lukey, A. Parkin, M. M. Roessler, B. J. Murphy, J. Harmer, T. Palmer, F. Sargent, F. A. Armstrong, *J. Biol. Chem.* **2010**, *285*, 3928–3938; e) M. J. Lukey, M. M. Roessler, A. Parkin, R. M. Evans, R. A. Davies, O. Lenz, B. Friedrich, F. Sargent, F. A. Armstrong, *J. Am. Chem. Soc.* **2011**, *133*, 16881–16892.
- [13] a) S. Ogo, R. Kabe, K. Uehara, B. Kure, T. Nishimura, S. C. Menon, R. Harada, S. Fukuzumi, Y. Higuchi, T. Ohhara, T. Tamada, R. Kuroki, *Science* **2007**, *316*, 585–587; b) S. Ogo, *Chem. Commun.* **2009**, 3317–3325.
- [14] G. J. Colpas, M. Kumar, R. O. Day, M. J. Maroney, *Inorg. Chem.* **1990**, *29*, 4779–4788.
- [15] P. J. Fagan, M. D. Ward, J. C. Calabrese, *J. Am. Chem. Soc.* **1989**, *111*, 1698–1719.
- [16] a) R. D. Jones, D. A. Summerville, F. Basolo, *Chem. Rev.* **1979**, *79*, 139–179; b) E. C. Niederhoffer, J. H. Timmons, A. E. Martell, *Chem. Rev.* **1984**, *84*, 137–203; c) C. J. Cramer, W. B. Tolman, K. H. Theopold, A. L. Rheingold, *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 3635–3640.
- [17] M. A. Reynolds, T. B. Rauchfuss, S. R. Wilson, *Organometallics* **2003**, *22*, 1619–1625.
- [18] S. Nagao, H. Seino, M. Hida, Y. Mizobe, *Dalton Trans.* **2005**, 3166–3172.
- [19] a) K. Kirchner, K. Mauthner, K. Mereiter, R. Schmid, *J. Chem. Soc. Chem. Commun.* **1993**, 892–894; b) I. de los Ríos, M. J. Tenorio, J. Padilla, M. C. Puerta, P. Valera, *J. Chem. Soc. Dalton Trans.* **1996**, 377–381; c) M. Sato, M. Asai, *J. Organomet. Chem.* **1996**, *508*, 121–

- 127; d) M. I. Bruce, B. C. Hall, N. N. Zaitseva, B. W. Skelton, A. H. White, *J. Chem. Soc. Dalton Trans.* **1998**, 1793–1803; e) G. Jia, W. S. Ng, H. S. Chu, W.-T. Wong, N.-T. Yu, I. D. Williams, *Organometallics* **1999**, *18*, 3597–3602; f) M. E. Navarro Clemente, P. J. Saavedra, M. C. Vázquez, M. A. Paz-Sandoval, *Organometallics* **2002**, *21*, 592–605; g) M. D. Palacios, M. C. Puerta, P. Valerga, A. Lledós, E. Veilly, *Inorg. Chem.* **2007**, *46*, 6958–6967; h) L. Cuesta, E. Tomat, V. M. Lynch, J. L. Sessler, *Chem. Commun.* **2008**, 3744–3746; i) W. M. Khairul, M. A. Fox, N. N. Zaitseva, M. Gaudio, D. S. Yufit, B. W. Skelton, A. H. White, J. A. K. Howard, M. I. Bruce, P. J. Low, *Dalton Trans.* **2009**, 610–620.
- [20] a) M. Tada, S. Muratsugu, M. Kinoshita, T. Sasaki, Y. Iwasawa, *J. Am. Chem. Soc.* **2010**, *132*, 713–724; b) J. H. Burness, J. G. Dillard, L. T. Taylor, *J. Am. Chem. Soc.* **1975**, *97*, 6080–6088.
- [21] We added excess amounts of NaBH₄ (10 equivalents) to an aqueous solution of compound **2b** and *p*-hydroquinone to proceed the reaction at pH 2.0.
- [22] J. F. Moulder, W. F. Stickle, P. E. Sobol, K. D. Bomben, *Handbook of X-ray Photoelectron Spectroscopy*, Physical Electronics, Inc., Minnesota, **1995**.
- [23] a) P. K. Glasoe, F. A. Long, *J. Phys. Chem.* **1960**, *64*, 188–190; b) K. Mikkelsen, S. O. Nielsen, *J. Phys. Chem.* **1960**, *64*, 632–637.

Received: December 16, 2011
Published online: March 1, 2012