

# *Dopamine D<sub>1</sub> Receptor Ligands: Where Are We Now and Where Are We Going*

*Jing Zhang,<sup>1</sup> Bing Xiong,<sup>2</sup> Xuechu Zhen,<sup>3</sup> Ao Zhang<sup>1</sup>*

<sup>1</sup>Synthetic Organic & Medicinal Chemistry Laboratory (SOMCL), Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

<sup>2</sup>Discovery Chemistry Laboratory, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

<sup>3</sup>Neuropharmacological Laboratory, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

Published online 18 July 2008 in Wiley InterScience (www.interscience.wiley.com).

DOI 10.1002/med.20130



**Abstract:** The dopamine (DA) D<sub>1</sub> receptor is the most highly expressed DA receptor subtype among the DA receptor family. Although the first DA D<sub>1</sub> receptor selective ligand SCH-23390 (**1**) was introduced more than two decades ago, clinically useful D<sub>1</sub> receptor selective ligands are rare. A renewed interest was ignited in the early 1990s by Nichols and Mailman who developed dihydrexidine (**27a**), the first high affinity full efficacy agonist for the D<sub>1</sub> receptor. Since then, a number of D<sub>1</sub> receptor agonists with full intrinsic activity, including A-86929 (**31a**), dinapsoline (**32a**), dinoxylone (**34a**), and doxanthrine (**35a**) were identified. These compounds all contain a conformationally rigid structure. However, the fate of such ligands for clinical use as treatments of Parkinson's disease and other related CNS disorders is not optimistic since the clinical trial with dihydrexidine (**27a**) was not successful. Further investigations on other compounds which are currently in the discovery stage will be crucial for determining the future of the D<sub>1</sub> receptor agonists. © 2008 Wiley Periodicals, Inc. *Med Res Rev*, 29, No. 2, 272–294, 2009

**Key words:** dopamine; dopamine receptor; benzazepine; dihydrexidine; Parkinson's disease

## **1. DOPAMINE AND DOPAMINE RECEPTORS**

Dopamine (DA) is one of the most important neurotransmitters in the central nervous system (CNS). It is involved in almost every aspect of brain functions, including control of movement, cognition,

---

*Contract grant sponsor:* Chinese National Science Foundation; *Contract grant number:* 30672517; *Contract grant sponsor:* Shanghai Commission of Science and Technology; *Contract grant number:* 07pj14104; *Contract grant sponsor:* Ministry of Science and Technology; *Contract grant number:* 2007AA02z163.

*Correspondence to:* Xuechu Zhen, and Ao Zhang, Synthetic Organic & Medicinal Chemistry Laboratory (SOMCL), Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China.

E-mail: xczen@mail.shcnc.ac.cn; aozhang@mail.shcnc.ac.cn

emotion, and regulation of the endocrine system. Dysfunctions of CNS dopaminergic neurotransmission account for a number of neurodegenerative and psychiatric disorders, including Parkinson's disease (PD), bipolar disorder, and schizophrenia.<sup>1,2</sup> In addition, all drugs of abuse influence directly or indirectly DA neurotransmission. Clinical and experimental studies clearly indicated that the positive reinforcement of the drugs of abuse is mediated primarily through the activation of the mesocorticolimbic DA system.<sup>3–5</sup> In the peripheral systems, DA also plays an important role in the functional regulation of cardiovascular and endocrinal systems.<sup>6</sup>

Dopamine (DA) exerts its actions mainly through DA receptors. These receptors exist both in mammalian CNS and peripheral tissues. They belong to a superfamily of large proteins containing seven relatively hydrophobic  $\alpha$ -helical transmembrane spanning segments linked to more hydrophilic segments with an extracellular amino terminus. This is the typical conformation for G-protein-coupled receptors (GPCRs) that interact with many membrane or cytoplasmic effector molecules and regulate corresponding brain functions. The five cloned DA receptors (D<sub>1</sub>–D<sub>5</sub>) are classified on the basis of their cDNA/gene sequence, as well as their pharmacological and biochemical characteristics. The D<sub>1</sub> and D<sub>5</sub> subtypes preferentially bind to phenylbenzazepines and activate G<sub>s</sub> proteins (G proteins that stimulate adenylate cyclase). Members of the D<sub>2</sub>-like DA receptor family (D<sub>2</sub>, D<sub>3</sub>, and D<sub>4</sub>), recognize the butyrophenones and benzamides, and are coupled to G<sub>i</sub> proteins (G proteins that inhibit adenylate cyclase formation). Activation of these D<sub>2</sub>-like receptors results in the inhibition of adenylate cyclase.<sup>7,8</sup>

Regulation of adenylate cyclase results in the change of cAMP that had been considered to be the major responsible secondary messenger system for DA receptor functions. However, recent information indicates that other intracellular signaling pathways (e.g., phospholipase C, protein kinase C and so on) are also involved in transducing DA signals.<sup>9–11</sup> In addition, DA receptors have also been shown to regulate mitogen-activated protein kinase (MAPK) pathways, a central pathway in the regulation of cell proliferation, differentiation, and survival. It was shown that stimulation of the D<sub>2</sub>, D<sub>3</sub>, and D<sub>4</sub> DA receptors activates extracellular signal-regulated kinase (ERK),<sup>9,10</sup> whereas D<sub>1</sub> DA receptor stimulation activates the p38 MAPK.<sup>12</sup> Interestingly, there are some reports indicating that differential regulation of DA receptors on these signaling pathways may underlie the functional differences of D<sub>1</sub> and D<sub>2</sub> DA receptors.

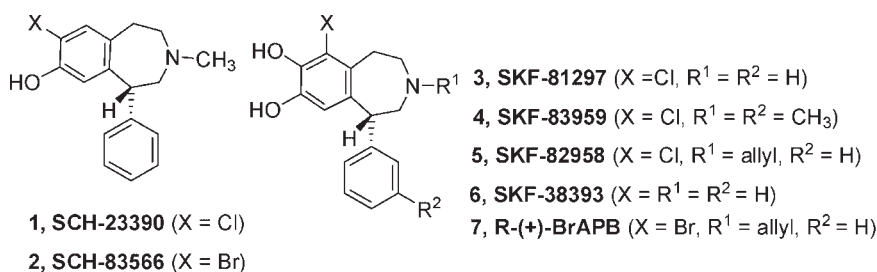
## 2. DA D<sub>1</sub> RECEPTOR AND ITS THERAPEUTIC POTENTIAL

The D<sub>1</sub> receptor is the most highly expressed DA receptor subtype with high levels in the DA rich areas of the mammalian forebrain, such as the caudate nucleus, putamen, substantia nigra, nucleus accumbens, hypothalamus, thalamus, frontal cortex, and olfactory bulb. It plays a crucial role in a variety of cognitive functions and is implicated in substance abuse disorders. The importance of the D<sub>1</sub> receptor in the treatment of PD was only appreciated until the 1990s when dihydrexidine (DHX), the first high affinity D<sub>1</sub> agonist with full intrinsic activity, was developed.<sup>13,14</sup> The pathology of PD is the progressive loss of nigrostriatal dopaminergic neurons. Degeneration of these neurons causes depletion of striatal DA. It is known that dopaminergic projections from the substantia nigra to the striatum are vital to motor control. Depletion of DA leads to motor symptoms clinically characterized by tremor, bradykinesia, rigidity and a late appearing loss of postural reflexes.<sup>15</sup> Thus, DA replacement is the most common therapeutic approach for relieving the PD symptoms. L-3,4-dihydroxy-phenylalanine (L-DOPA, levodopa) remains the most potent therapeutic drug for PD, and often produces dramatic effects in *de novo* patients.<sup>16</sup> Unfortunately, long-term use of levodopa is commonly associated with motor complications such as dyskinesia. Selective DA receptor agonists gained popularity because of their long-lasting and somewhat less severe adverse effects.<sup>17</sup> Another important advantage for using DA receptor agonists in PD therapy is the potential neuroprotective actions. The protective effects of DA agonists on DA neurons, however, seemingly do not involve DA

receptors.<sup>18</sup> It has been believed for quite a long time, that the therapeutic anti-parkinsonian effects of levodopa is primarily attributed to its stimulation of DA D<sub>2</sub> receptor whereas dyskinesias might be mediated by the over-stimulation of the D<sub>1</sub> receptor. As a result, D<sub>2</sub> receptor agonists were first introduced for the PD treatment.<sup>19</sup> The ergoline bromocriptine is the first DA receptor agonist that has been approved for anti-parkinsonian therapy since 1974. It was used initially as adjunct therapy to levodopa in patients experiencing motor fluctuations but was later recommended as mono-therapy for early PD patients.<sup>20</sup>

Little attention was paid to the anti-parkinson potential of D<sub>1</sub> receptor agonists at the beginning of their development. Part of the reasons was due to the lack of ideal D<sub>1</sub> receptor agonists which limited the effort in exploring the role of the D<sub>1</sub> receptor in PD treatment. Selective D<sub>1</sub> agonists of the first generation, such as SKF-38393 (compound **6**, Fig. 1), possess high affinity and selectivity, but have relative low intrinsic activities.<sup>21,22</sup> However, recent information indicates that D<sub>1</sub> receptor may play even greater role in motor control. DHX (**27a**), discovered in the late 1980s, was the first full D<sub>1</sub> DA receptor agonist.<sup>23</sup> DHX has high affinity for the D<sub>1</sub> receptor, with a relative low selectivity (~10-fold) for D<sub>1</sub> versus D<sub>2</sub> receptors.<sup>24</sup> The initial clinical trial with DHX failed due to severe adverse effects.<sup>25</sup> In fact, many clinically approved dopaminergic agents, such as  $\alpha$ -dihydroergocryptine, lisuride, pergolide, pramipexole, and ropinirole, all have multiple actions at many of DA receptor subtypes (including D<sub>1</sub>, D<sub>2</sub>, or D<sub>3</sub> receptors) indicating that a more complex mechanism may be involved. It is therefore reasonable to hypothesize that each of these DA receptors contributes to the therapeutic effects in a unique manner and stimulation of one particular DA receptor may not be sufficient.<sup>26</sup> Although selectivity of DA agonists for DA receptors have been demonstrated to have some benefits for PD patients, the overall advantage of this type of drugs remains uncertain. In terms of the high selective D<sub>1</sub> receptor agonists, it is far from clear to classify their role in PD, simply because there is no such drug available in clinical trial for the time being. A combination therapy with L-DOPA according the stage of PD and therapeutic responses may be a better approach.

Schizophrenia is a devastating mental illness affecting approximately 1% of the population worldwide. It is generally believed that abnormal neurotransmission, especially in DA and glutamate (NMDA) system, plays a critical role in the pathophysiology of schizophrenia. The hyperactivity of DA in subcortical structure is thought to associate with the positive symptoms, whereas D<sub>1</sub> receptor hypoactivity in the frontal cortical area has recently been suggested to attribute to the negative symptom and impaired cognitive function.<sup>27</sup> Indeed, D<sub>1</sub> receptor in frontal cortex is involved in the working memory expression.<sup>28</sup> Low doses of selective D<sub>1</sub> receptor agonists (e.g., DHX, A77636, and SKF-81297) have been shown to enhance the cognitive function in primates.<sup>27,29</sup> Recently, preliminary human studies indicated that DHX is safe and well tolerated with no serious adverse events at a single dose of 20 mg.<sup>30,31</sup> It is now suggested that combined activation of D<sub>1</sub> receptor along with  $\alpha_7$  nicotonic receptor, may be the most promising therapeutic mechanisms for improving cognitive deficits and negative symptoms in schizophrenia.<sup>30,31</sup> Thus, D<sub>1</sub> receptor selective agonists may represent an exciting direction for the treatment of schizophrenia in the future.<sup>32</sup>



**Figure 1.** Representative D<sub>1</sub> benzazepines.

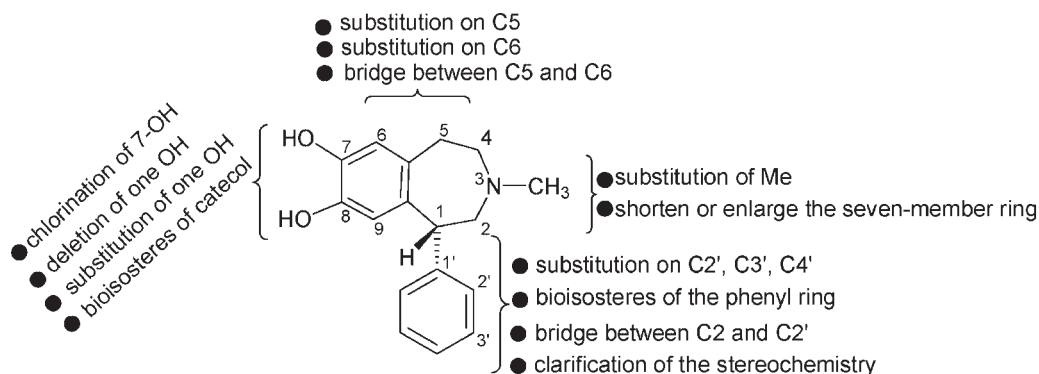
### 3. HISTORICAL REVIEW ON DA D<sub>1</sub> RECEPTOR SELECTIVE LIGANDS

The therapeutic potentials of DA D<sub>1</sub> receptor ligands were re-kindled in 1990s. Studies since then have provided a wealth of information on the structure–activity relationship (SAR) at the D<sub>1</sub> receptor. However, lack of information on the exact three-dimensional orientation of amino acid residues at the binding sites of the DA receptors, together with the limited understanding on interactions between the D<sub>1</sub> receptor and selective ligands still hampers the rational design and development of potent and selective D<sub>1</sub> receptor agents. Current knowledge on D<sub>1</sub> agonist and antagonist pharmacophores, and the intrinsic SAR, remains largely empirical. Most recently, computer-aided analysis on the conformations of the current available full agonists and quantitative structure–activity relationship (QSAR) characterizing D<sub>1</sub> antagonists has greatly stimulated the advances in this field.<sup>33–37</sup>

A number of ligands with high affinity for the D<sub>1</sub> receptor had been developed in 1980s. Most of those research work was limited on a single chemical class: phenyltetrahydrobenzazepines. Therefore, many useful D<sub>1</sub> receptor selective agents possessing partial agonist or antagonist properties with high D<sub>1</sub> affinity, for example, SCH-23390 (**1**), SKF-83959 (**4**) were derived from this group (Fig. 1). Ironically, the high D<sub>1</sub> receptor binding profile of these ligands was poorly relevant to their intrinsic activity. As a result, few D<sub>1</sub> full agonists were available for further investigation.<sup>22,36</sup>

In 1990s, led by Nichols et al.,<sup>22,33–35</sup> the search of DA D<sub>1</sub> full agonists was switched to the development of structurally rigid analogs of  $\beta$ -phenyldopamines. Thus, several compounds with novel structures distinguished from traditional phenylbenzazepines were found possessing exceptional anti-parkinsonian effects in the MPTP-treated monkey model.<sup>33,34,38</sup> Such breakthrough has largely spurred the interests of developing D<sub>1</sub> agonists as potential treatment of PD. Thus, a tentative SAR was established for typical high affinity D<sub>1</sub> full agonists, based on Nichols and Mailman's conceptual model of the agonist state on the D<sub>1</sub> receptor (Fig. 2). They generally contain the following principles:

- (i) A phenylethylamine moiety. It would be optimal for D<sub>1</sub> receptor intrinsic activity if this moiety is in a *trans* extended conformation. An aspartate residue of the D<sub>1</sub> receptor may bind with its protonated amino group and the substituent on the nitrogen atom may greatly influence its D<sub>1</sub> selectivity over the D<sub>2</sub> receptor.<sup>7,39</sup>
- (ii) A catechol moiety, or a catechol ring with halogen replacement of the two hydroxyl groups is necessary. It is believed that such catecholic function would serve as a hydrogen-bond donor to the serine residues in the active site of the D<sub>1</sub> receptor, although D<sub>1</sub> receptor agents without a catechol ring do exist.<sup>40–44</sup>
- (iii) A hydrophobic substituent (e.g., phenyl) attached to the  $\beta$ -carbon of the ethylamine side chain. Such a component is postulated to interact with the hydrophobic binding domain of the



**Figure 2.** Structural modification on benzazepines.

D<sub>1</sub> receptor active site, and is crucial to the agent's affinity and potency as well as selectivity over other DA subtypes. However, whether the coplanar orientation of the  $\beta$ -phenyl moiety with the catechol ring is the optimal conformation remains controversial.<sup>45</sup>

Part of the advances described above has been discussed in several recent articles.<sup>18,46</sup> This review, therefore, will focus on progress of two major categories of D<sub>1</sub> receptor ligands: conformationally constrained benzazepine derivatives and dihydrexidine tetracyclic analogues.

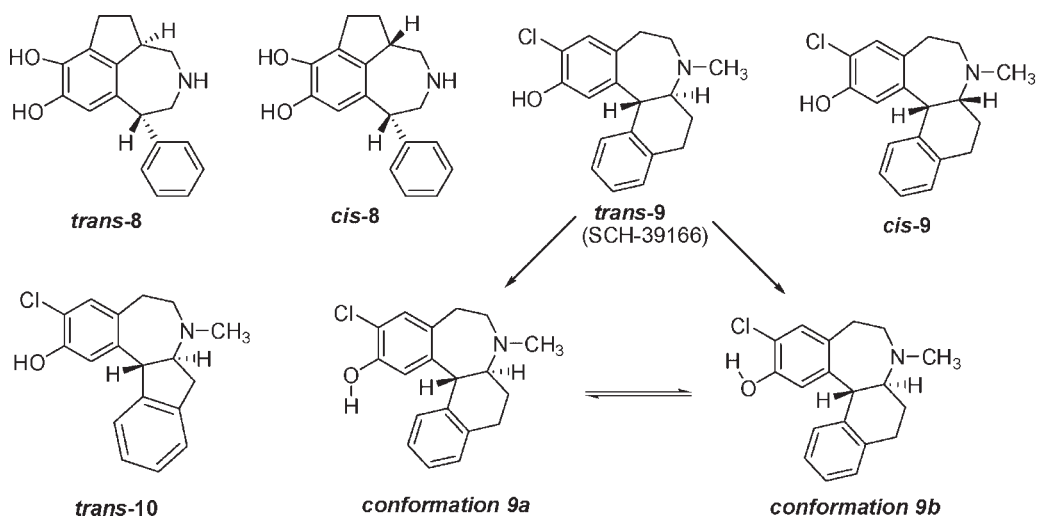
#### 4. RECENT DEVELOPMENT OF DA D<sub>1</sub> RECEPTOR SELECTIVE LIGANDS

##### A. Conformationally Constrained Arylbenzazepine Analogues

The family of phenylbenzazepines represents a unique class of compounds possessing high binding affinity for the D<sub>1</sub> receptors and high selectivity over the D<sub>2</sub> receptors. Considerable efforts have been devoted to this category of compounds in the past several decades, mainly because of its therapeutic application as anti-psychotics and anti-stimulant medications. Although full D<sub>1</sub> agonists with high affinity are rare, a large number of partial agonists and antagonists have been discovered (see Fig. 1), including the first high-affinity and selective D<sub>1</sub>/D<sub>5</sub> antagonist SCH-23390 (**1**) and its conformationally restricted analogue SCH-39166 (**9**, Fig. 3).

The SAR of benzazepine analogues has been studied extensively using fruitful structural modifications (Fig. 2). In general, a chiral center exists on the C1-position where the receptor affinity resides primarily in the R enantiomer.<sup>47</sup> These compounds possess a considerable degree of conformational mobility, and the optimum position of the phenyl ring for interaction with the proposed accessory binding site of the D<sub>1</sub> receptor (e.g., SCH-23390, **1**) is likely to be equatorially.<sup>48,49</sup> There was a hypothesis that the preferred binding of benzazepine analogues to the D<sub>1</sub> receptor might be the consequence of a  $\pi$ - $\pi$  non-bonded interaction between the C1-aromatic ring and a complementary residue of Phe, Tyr, or Trp on the receptor surface.<sup>50</sup> To probe the validity of such a concept, conformationally constrained benzazepines **8**–**10** were developed (Fig. 3).

Insertion of an ethylene bridge between C5 and C6 into the molecule of benzazepine **6** (SKF-38393) resulted in compound **8**, with either a *cis*-, or *trans*-conformation.<sup>49</sup> Compound *cis*-**8** has a high affinity at the D<sub>1</sub> receptor with a K<sub>bind</sub> of 24 nM, whereas the *trans*-**8** is inactive. However, similar to other benzazepine derivatives, this compound also shows partial agonist activity with only



**Figure 3.** Conformationally restrained benzazepine analogues.

62% of DA response. The high affinity of *cis*-isomer may be rationalized by the fact that the side-chain nitrogen of *cis*-isomer is nearly in the plane of the catechol nucleus which is required for dopaminergic pharmacophore and D<sub>1</sub> receptor binding, and *trans*-isomer **8** is unable to reach the D<sub>1</sub> receptor binding site (Table I).

Insertion of an ethylene bridge between C2 and C2' into the molecule of benzazepine **1** (SCH-23390) yielded compounds **9** where the additional ring C was fused in *cis* or *trans* form.<sup>48</sup> Binding studies on both the *cis* and *trans* isomers disclosed that the *trans*-**9** has a K<sub>i</sub> of 3.3 nM and is 143-fold more potent than *cis*-**9**. Resolution of the racemic *trans*-**9** indicated that the *R*-(-)-*trans*-**9** has a significantly greater D<sub>1</sub> affinity and selectivity (K<sub>i</sub>, 1.9 nM). This phenomenon is consistent with the fact that the D<sub>1</sub> receptor affinity in the 1-phenyl-1H-3-benzazepine family is associated specifically with the *R*-enantiomers. Similarly, the C2 and C2' methylene-bridged benzazepine **10** also favors its affinity and selectivity for the D<sub>1</sub> receptor in its *trans* form *R*-enantiomer.<sup>51</sup> Further investigation on *trans*-**9** (SCH-39166) disclosed that in comparison to the prototype SCH-23390, it is a selective DA D<sub>1</sub> receptor antagonist, with reduced affinity for serotonin receptors and with a longer duration of actions in a conditioned avoidance paradigm in primates.<sup>52</sup>

Conformational analysis was also conducted to clarify the exact mode of binding of these conformationally constrained compounds to the D<sub>1</sub> receptors, without reaching a solid conclusion.<sup>53</sup> One additional question raised from these studies is the exact orientation of the C8 hydroxyl group in these compounds. For example, compound **9** has two possible orientations when approaching the receptors, conformation **9a** and conformation **9b** (Fig. 3). The C8 hydroxyl group was recognized a prerequisite for binding to the DA receptors. In 2005, Wu and his colleagues indirectly addressed this question by carefully designing a series of heterocycle-fused compounds **11–16** (Fig. 4).<sup>54</sup> They concluded that a suitably arranged heterocycle with an NH group would be able to mimic the prototypic OH group to probe its binding direction, and more importantly, to serve as metabolically stable bioisosteres.

The triazole compound **11** can roughly represent a rigid analog of conformation A, and the triazole NH is sufficiently acidic to form a hydrogen bond as its prototypic OH. However, the affinity of this compound at the D<sub>1</sub> receptor is disappointing with a K<sub>i</sub> of 583 nM. With proposed hydrogen bonding in opposite orientation, compound **12** represents the conformation B. This methyl substituted pyrrole analog displayed a K<sub>i</sub> of 24.7 nM for binding to the D<sub>1</sub> receptor, which is much higher than that of compound **11**, where the proposed hydrogen bonding is oppositely oriented. Based on these findings, it was concluded that conformation B (similar to conformation A and B for compound **9** in Fig. 3) might be the active binding conformer, while the conformation A is not. According to this hypothesis, benzimidazole **13** and benzotriazole **14** were prepared with proper hydrogen bonding orientation, but the binding affinity of these two analogues at the D<sub>1</sub> receptor turned out to be poor (K<sub>i</sub>, 248 and 146 nM, respectively). The unexpected poor affinity of compounds **13** and **14** may be ascribed to the possible tautomerization of the imidazole and triazole functions in these two compounds, thus the N–H can switch from N1-position to N3 position. This analysis prompted the design and synthesis of benzimidazolone **15**, where two NHs exist in different chemical and magnetic environments. As expected, compounds **15a,b** display a significant enhancement in binding affinity to the D<sub>1</sub> receptor with K<sub>i</sub>s of 7 and 16.5 nM, respectively. Further, compounds **16a,b**, with a stronger acidic NH and without the additional unnecessary NH, give remarkably high D<sub>1</sub> receptor binding affinity. The K<sub>i</sub>s of **16a,b** are 2.1 and 6.5 nM, respectively, threefold higher than that of compound **15a,b**.

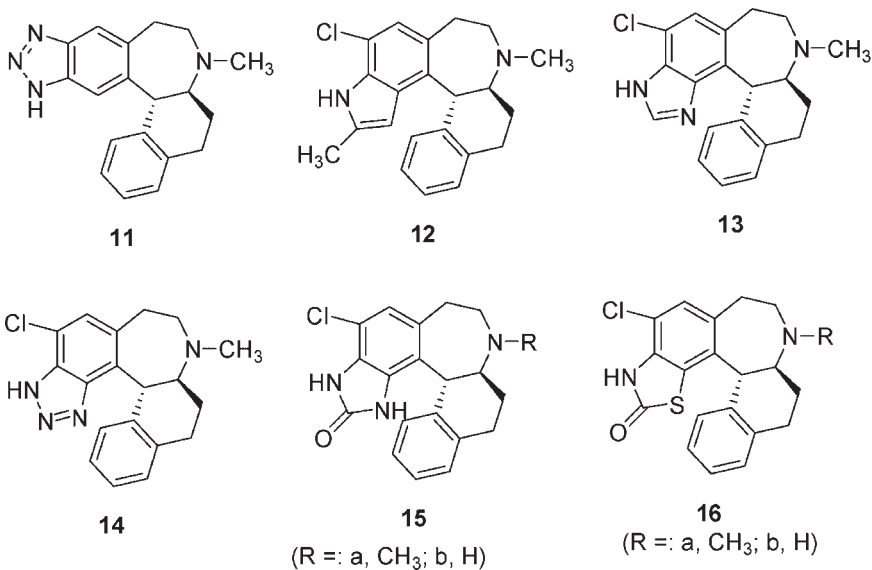
To further explore the potential of the conformationally rigid 2,2'-bridged benzazepines, a large series of substituted analogues **17** and **18** were synthesized (Fig. 5).<sup>55</sup> Variant substitution patterns (3'-, 4'-, 5'-), substituent nature and size were investigated. High binding affinity for the D<sub>1</sub> receptor was observed in compounds **17a–k**, and **18a–l**. All these compounds have K<sub>i</sub>s of less than 5 nM, with the highest affinity of 0.45 nM for compound **18g** which contains a 4'-methanesulfonylamino



**Table 1.** Binding Affinity at the Dopamine D<sub>1</sub>, D<sub>2</sub> and Serotonin 5-HT<sub>2A</sub> Receptors of Benzazepine Analogues (K<sub>i</sub>/IC<sub>50</sub>, nM)<sup>a</sup>

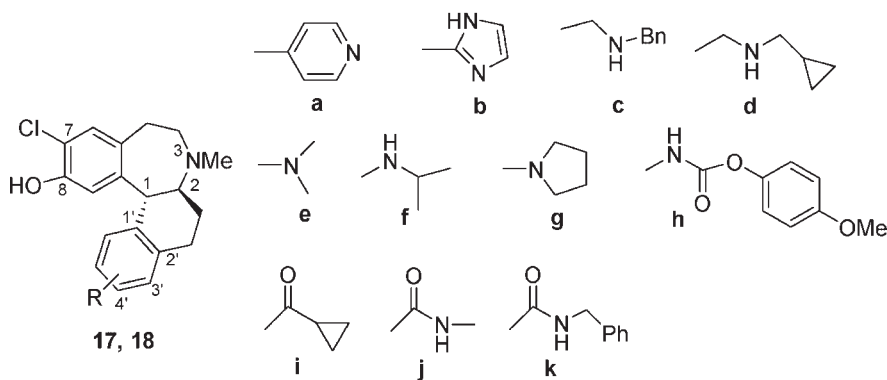
Compound	D <sub>1</sub>	D <sub>2</sub>	D <sub>5</sub>	5-HT <sub>2A</sub>	Reference
<b>1</b> (SCH23390)	0.12	83.8	12	596	36
<b>3</b> (SKF81297)	1.90	1272	-	398	36
<b>4</b> (SKF83959)	1.18	920	7.56	266	36
<b>5</b> (SKF82958)	4.56	264	-	1612	36
<b>6</b> (SKF38393)	26.6	>10000	-	>10000	36
<b>7</b> (BrAPB)	2.29	209	-	-	36
<i>trans</i> - <b>8</b>	1220	>10000	-	-	49
<i>cis</i> - <b>8</b>	24	>10000	-	-	49
<i>cis</i> - <b>9</b>	473	9073	-	-	48
<i>trans</i> - <b>9</b>	3.3	4115	-	-	48
<i>R</i> -(-)- <i>trans</i> - <b>9</b>	1.9	514	-	-	48
<i>S</i> -(+)- <i>trans</i> - <b>9</b>	531	3046	-	-	48
<b>10</b>	7	-	-	-	51
<b>11</b>	583	3000	-	-	54
<b>12</b>	24.7	232	-	-	54
<b>13</b>	248	984	-	-	54
<b>14</b>	146	1530	-	-	54
<b>15a</b>	7	1023	-	-	54
<b>15b</b>	16.5	3270	-	-	54
<b>16a</b>	2.1	257	-	-	54
<b>16b</b>	6.5	661	-	-	54
<b>18g</b>	0.45	>10000	-	-	55
<b>19<sup>b</sup></b>	12	-	-	-	56

<sup>a</sup>Data were taken directly from the corresponding literature.<sup>b</sup>IC<sub>50</sub> value.

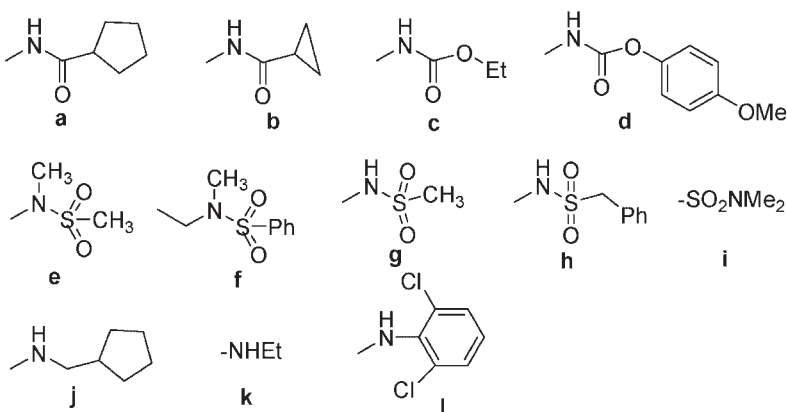


**Figure 4.** 2,2'-Bridged conformationally restrained benzazepine analogues.

### 3' substituted series 17:



### 4' substituted series 18:



**Figure 5.** Substituted 2,2'-bridged benzazepine analogues.



substituent. The selectivity of this compound for the D<sub>1</sub> receptor over the D<sub>2</sub> receptor is more than 6,000-fold.

Another series of compounds is 6,7-heterocycle-fused benzazepines with a general formula **I** (Fig. 6).<sup>56</sup> In these compounds, a furan-, pentane-, hexane-, thiophene-, or 1,3-dioxopentane-, was fused to the C6 and C7 of the benzazepine ring system (**19–25**). In several cases, the C1-phenyl group of benzazepine core was generally substituted with a benzofuran-7-yl, or a 2,3-dihydrobenzofuran-7-yl moiety that has been previously reported to be a good bioisostere of phenyl functionality. These compounds generally retain high affinity at the DA D<sub>1</sub> receptor. Among this series, compound **19** has the highest binding affinity with an IC<sub>50</sub> of 12 nM at this receptor.

### B. Dihydropyridine and Its Polycyclic Analogues

Compound **26** represents a series of 4-phenyl substituted tetrahydroisoquinolines,<sup>34,37,57,58</sup> which has moderate, but selective affinity for the DA D<sub>1</sub> receptors. Rigidification of this template by insertion of an ethylene-bridge yielded a series of novel tetracyclic compounds **27a–c** (Fig. 7).<sup>59,60</sup> These compounds can be viewed as a B-ring modification of the conformationally constrained benzazepine analogue **9** (SCH-39166). The *trans*-configured 10,11-dihydroxy-5,6,6a,7,8,12b-hexahydrobenzo[*a*]phenanthridine **27a** (dihydropyridine, DHX) is a highly potent and selective D<sub>1</sub> receptor agonist in rat brain. It competes for [<sup>3</sup>H]SCH-23390 binding sites in rat striatal homogenate with an IC<sub>50</sub> of 12 nM. EC<sub>50</sub> of DHX in activating DA-sensitive rat striatal adenylate cyclase is 70 nM. The maximal stimulation is equal to or slightly greater than that produced by DA itself. More importantly, compound **27a** is a full DA D<sub>1</sub> agonist, different from the previously available agents which show only partial agonism. The *N*-methyl-(**27b**), *N*-propyl-(**27c**) analogues give a much lower potency in binding to the D<sub>1</sub> receptor, with IC<sub>50</sub>s of 91 and 651 nM, respectively, and their intrinsic activity is substantially reduced. Compound **28**, reported much earlier by Wei and Teitel<sup>61</sup> with a 9,10-dihydroxy substituted catechol moiety is inactive at the D<sub>1</sub> receptor. The *cis*-isomer **29** is also found neither stimulating cAMP synthesis nor inhibiting the cAMP synthesis induced by DA itself.

The discovery of DHX (**27a**) was a breakthrough in the development of DA D<sub>1</sub> receptor full agonists. It was the first high-potency, full efficacy, bioavailable agonist selective for the D<sub>1</sub> receptor, and represents a new conformationally rigid structural scaffold of D<sub>1</sub> receptor ligands. The racemate **27a** has impressive anti-parkinsonian actions in the MPTP primate model and was studied in several clinic trials.<sup>24</sup> Resolution of racemic **27a** resulted in two isomers (+)-**27a** (6aR,12bS) and (–)-**27a**

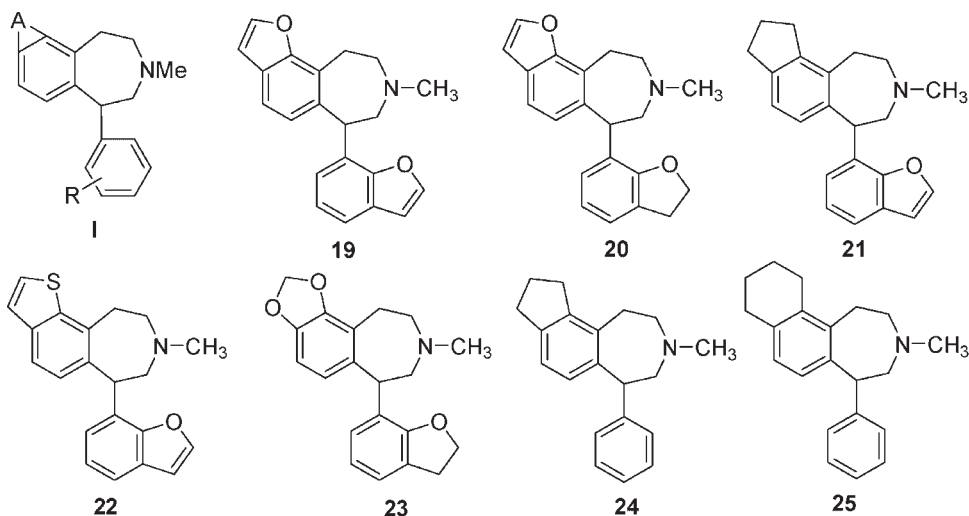
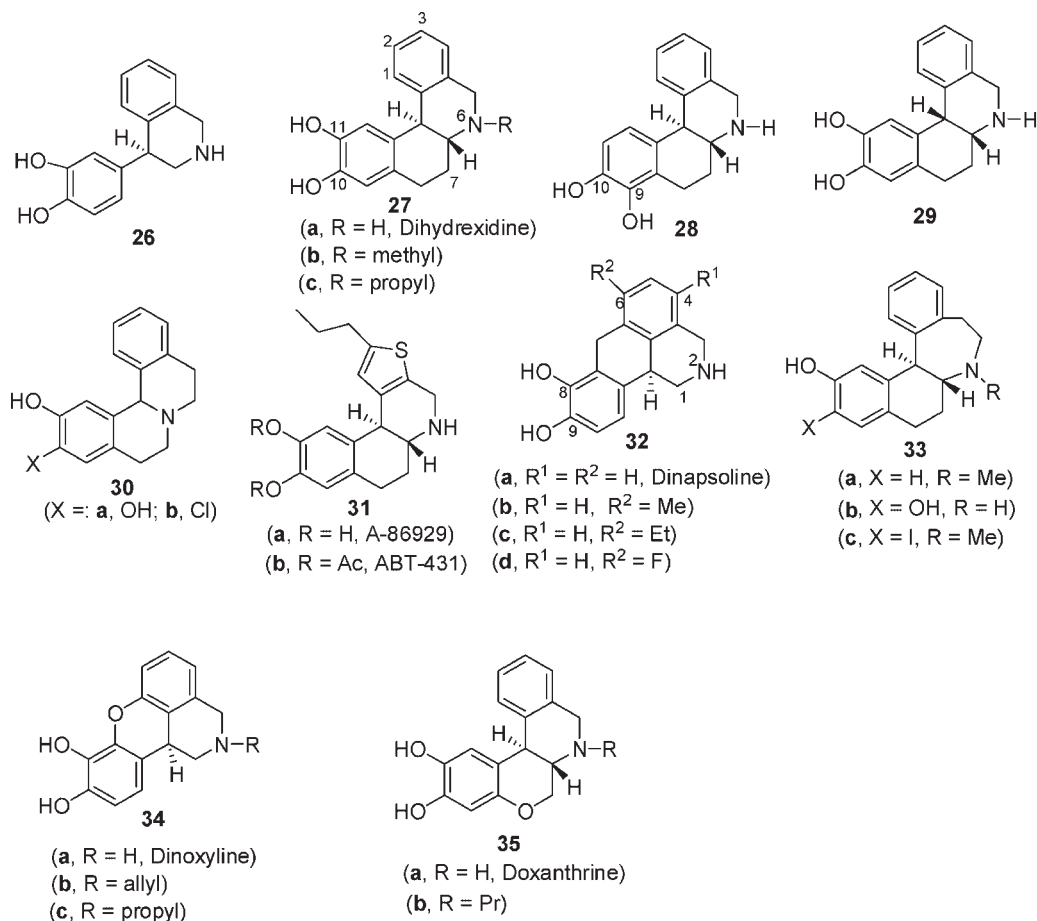


Figure 6. Tricyclic benzazepine analogues.



**Figure 7.** Dihydroxide and its tetracyclic analogues.

(6aS,12bR).<sup>62</sup> (+)-**27a** is 2-, and 25-fold more potent than the racemate and (–)-**27a**, respectively. Functionally, (+)-**27a** is a full agonist, with an EC<sub>50</sub> of 51 nM in activating striatal DA-sensitive adenylate cyclase versus 2.15 μM for the (–)-isomer (–)-**27a**. Thus, it is likely that the DA receptor activity of DHX resides principally in the (6aR,12bS)-(+)-enantiomer conformation (Table II).

An approach to explore the N-atom location was conducted yielding compounds **30a,b**.<sup>63</sup> Such structurally rigid tetrahydroisoquinolines are poor D<sub>1</sub> receptor ligands, with K<sub>i</sub> values at 2700 nM and 830 nM, respectively for compounds **30a** and **30b** in competition for [<sup>3</sup>H]SCH-23390 binding sites in rat striatal homogenate. Thus, the nitrogen position has a crucial pharmacophoric importance in modulating D<sub>1</sub> receptor binding affinity and functionality.

Thiophenes **31a,b** can be viewed as bioisosteres of the A-ring phenyl group of DHX (**27a**).<sup>64</sup> The NH compound **31a** (A-86929) possess high affinity for the cloned human D<sub>1</sub> receptor with a K<sub>i</sub> of 49 nM. Similar to **27a**, this compound also has full intrinsic activity relative to DA in stimulating adenylate cyclase (EC<sub>50</sub>, 9 nM). Resolving this racemic compound showed that (+)-**31a** is approximately 60-fold weaker in both binding affinity and functional activity. In addition, compound **31a** produces robust rotation in the unilaterally lesioned rat after both acute and repeated administration. However, this compound is very unstable, and readily undergoes air-oxidation. As a result, it is generally prepared and used as its diacetate precursor **31b** (ABT-431). This prodrug offers greater solid-state stability and can be easily converted to its parent compound **31a** both *in vitro* and *in vivo*.<sup>64</sup>

**Table II.** Dopamine D<sub>1</sub>, D<sub>2</sub> Receptor Binding affinity of Conformationally Constrained Tetracyclic Ligands<sup>a</sup>

Compound	K <sub>i</sub> /K <sub>0.5</sub> / IC <sub>50</sub> (nM)	D <sub>1</sub>	D <sub>2</sub>	Reference
<b>27a</b> (dihydroxidine)	IC <sub>50</sub>	12	120	59
(+)- <b>27a</b>	K <sub>0.5</sub> (IC <sub>50</sub> )	4.59 (5.6)	43	62, 65
(-)- <b>27a</b>	IC <sub>50</sub>	149	1250	62
<b>27b</b>	IC <sub>50</sub>	91	136	59
<b>27c</b>	IC <sub>50</sub>	651	53	59
<b>28</b>	-	NA <sup>b</sup>	NA <sup>b</sup>	59
<b>29</b>	IC <sub>50</sub>	> 5000	>5000	59
<b>30a</b>	K <sub>i</sub>	830	>5000	63
<b>30b</b>	K <sub>i</sub>	2700	>5000	63
<b>31a (A-86929)</b>	K <sub>i</sub>	49	710	64
<b>32a</b> (dinapsoline)	K <sub>0.5</sub> (IC <sub>50</sub> )	5.93(67)	31.3	65
(+)- <b>32a</b>	IC <sub>50</sub>	33	38	66
(-)- <b>32a</b>	IC <sub>50</sub>	5300	1500	66
<b>32b</b>	K <sub>0.5</sub>	11	57	67
<b>32c</b>	K <sub>0.5</sub>	14	-	68
(-)- <b>32d</b>	K <sub>0.5</sub>	>3000	-	68
(+)- <b>32d</b>	K <sub>0.5</sub>	71	7	68
<b>33a</b>	K <sub>0.5</sub>	3950	>10000	69
<b>33b</b>	K <sub>0.5</sub>	2850	2220	69
<b>33c</b>	K <sub>0.5</sub>	1380	2960	69
<b>34a</b> (dinoxyline)	K <sub>0.5</sub>	8.3	6.2	70
<b>34b</b>	K <sub>0.5</sub>	260	54	70
<b>34c</b>	K <sub>0.5</sub>	250	17	70
<b>35a</b> (doxanthrine)	K <sub>i</sub>	22	3700	71
(+)- <b>35a</b>	K <sub>i</sub>	8	2500	71
(-)- <b>35a</b>	K <sub>i</sub>	270	6800	71
<b>35b</b>	K <sub>i</sub>	330	340	71

<sup>a</sup>Data were taken directly from the corresponding reference, the dash lines indicate no data available.<sup>b</sup>No activity.

Following the original idea to rigidify the tetrahydroisoquinolines (e.g., **26**), and encouraged by successful development of the full D<sub>1</sub> agonist **27a**, Nichols and colleagues introduced a bridge to the structure **26** and its analogues.<sup>65</sup> Thus, a series of 8,9-dihydroxy-2,3,7,11*b*-tetrahydro-1*H*-naph[1,2,3-*de*]isoquinolines **32a–d** were developed (Fig. 7). This approach involves taking the backbone of DHX (**27a**), the first high-affinity full D<sub>1</sub> agonist, and tethering the two phenyl rings

through a methylene-bridge followed by removal of the C(7)-C(8) ethoxy-bridge. Preliminary molecular modeling studies demonstrated that these modifications conserved the essential elements of the pharmacophore required for D<sub>1</sub> receptor activity.<sup>35</sup> Compound **32a** (dinapsoline) has almost identical affinity ( $K_i = 5.9$  nM) to **27a** at the rat striatal D<sub>1</sub> receptors and displays a shallow competition curve ( $n_H = 0.66$ ) that suggests agonist properties. In both the rat striatum and C-6-mD<sub>1</sub> cells, dinapsoline **32a** is a full agonist with an EC<sub>50</sub> of ca. 30 nM in stimulating cAMP synthesis via the D<sub>1</sub> receptor. Resolution of **32a** into a pair of enantiomers showed that the (*R*)-(+)-**32a** is the active enantiomer.<sup>66</sup> In unilateral 6-hydroxydopamine (6-OHDA)-lesioned rats (+)-dinapsoline **32a** induces robust rotational behavior comparable to that of an external benchmark, **31a**. The C6-methyl (**32b**) and C6-ethyl (**32c**) analogues have almost identical binding affinity to the D<sub>1</sub> receptor with  $K_i$ s of 11 and 14 nM, respectively.<sup>67,68</sup> Other C6 or C4 substitutions led to a decay in D<sub>1</sub> receptor affinity. It is of note that the active D-ring analog, 6-fluorodinapsoline **32d**, resides its biological activity in the (+)-enantiomer which is consistent with the activity of (+)-dinapsoline **32a**.

*trans*-6,6a,7,8,9,13b-Hexahydro-5*H*-benzo[*d*]naphth[2,1-*b*]azepines **33a–c** were designed as conformationally restricted homologues of the potent benzophenanthridine D<sub>1</sub> agonist DHX **27a**.<sup>69</sup> Based on previously knowledge, the dihydroxy secondary amine **33b** was predicted to be a D<sub>1</sub> receptor agonist, whereas the *N*-methyl compounds **33a** and **33c** were predicted to be D<sub>1</sub> receptor antagonists. Surprisingly, none of the three compounds shows appreciable affinity for the D<sub>1</sub> receptor ( $K_{0.5}$ : 3.9, 2.8, and 1.38  $\mu$ M, respectively). A comparison of the low-energy conformations of these molecules shows that the pendant phenyl ring of **33b** is twisted for approximately 28° relative to that of the corresponding ring of **27a**. Further, the additional methylene unit used to expand the C ring of **33b** projects toward the  $\alpha$ -face of the molecule, suggesting that steric protrusion in this region of the molecule is not well tolerated.<sup>69</sup>

Different from the approach in designing dihydrexidine (**27a**) and dinapsoline (**32a**), 8,9-dihydroxy-1,2,3,11b-tetrahydrochromeno[4,3,2-*de*] isoquinolines **34a–c** were developed by tethering the two phenyl rings of  $\beta$ -phenyldopamine D<sub>1</sub> pharmacophore using an ether linkage.<sup>70</sup> The resulting compound **34a** (dinoxylene) is found to be a potent full D<sub>1</sub> agonist ( $K_{0.5} = 8.3$  nM; EC<sub>50</sub> = 87 nM) for striatal D<sub>1</sub> receptors, but also has high affinity for brain D<sub>2</sub>-like and cloned D<sub>2</sub> and D<sub>3</sub> receptors. The *N*-allyl (**34b**) and *N*-*n*-propyl (**34c**) derivatives have much reduced affinity for the D<sub>1</sub> receptor with  $K_{0.5}$  of 260 and 250 nM, respectively. However, increased D<sub>2</sub>-like receptor binding affinity is observed. Therefore, this represents the first example of ligands with high affinity for all DA receptors, yet with functional characteristics similar to DA itself.

Tethering the two phenyl rings of  $\beta$ -phenyldopamine D<sub>1</sub> pharmacophore by a –CH<sub>2</sub>O– linkage produced *trans*-2,3-dihydroxy-6a,7,8,12b-tetrahydro-6*H*-chromeno [3,4-*c*]isoquinolines **35a,b**.<sup>71</sup> These compounds can be viewed as heterocyclic bioisosteres of the potent DA D<sub>1</sub>-selective full agonist DHX (**27a**). Compound **35a** (doxanthrine) possesses high affinity ( $K_i = 20$ –30 nM) for the porcine D<sub>1</sub>-like receptors in native striatal tissue and full intrinsic activity at cloned human DA D<sub>1</sub> receptors with much lower affinity at the D<sub>2</sub>-like receptors ( $K_i = 3000$  nM). Again (+)-**35a** gives much higher potency ( $K_i = 8$  nM) in binding to the D<sub>1</sub> receptor than (–)-**35a** ( $K_i = 270$  nM). The binding and functional properties of this compound highlights the effectiveness of constructing DA D<sub>1</sub> agonist ligands using the  $\beta$ -phenyldopamine pharmacophore template. The *N*-propyl analogue **35b** has a substantially weaker affinity at the D<sub>1</sub> receptor ( $K_i = 330$  nM).

### C. Computational Modeling of DA D<sub>1</sub> Receptor

Computer-aided drug design plays an important role in the development of new drugs that target GPCRs. As in the efforts for targeting D<sub>1</sub> DA receptors, many computational modeling methods have been utilized with the aim to investigate the SARs mostly lying in the agonists. However, due to the difficulty in crystallizing GPCRs, and the limitation in obtaining the three-dimensional structures, early investigations mostly used the ligand-based approaches, such as pharmacophore detection and

Quantitative-SAR (QSAR).<sup>35,72–78</sup> In 1996, following the conformational analyses of dihydrexidine (DHX) and other active D<sub>1</sub> agonists, Mottola et al.<sup>79</sup> applied the active analog approach with the help of a SYBYL software package, and built a pharmacophoric model of the DA D<sub>1</sub> receptor. In this model, two distances constraints and one angle constraint are included, namely the distance between the positively charged nitrogen and oxygen of *m*-hydroxyl group of the catechol ring in a range of 7.1–8.0 Å, the distance between the nitrogen and oxygen of *p*-hydroxyl group in a range of 7.1–7.4 Å, and the angle between the planes defined by the catechol ring and the pendent aromatic ring in a range of 50–80°. In comparison to the inactive compounds of several DHX analogs, it is clearly indicated that additional bulk group attached to the nitrogen atom in DHX is intolerable and sterically clashes with binding site residues of the D<sub>1</sub> receptor. Similarly, Wilcox et al.<sup>80,81</sup> also conducted an analysis of a series of D<sub>1</sub> receptor agonists, and summarized a pharmacophore model and a CoMFA based QSAR model. In this model, the distance constraints from the positively charged nitrogen to *m*-hydroxyl and to *p*-hydroxyl groups are slightly shorter than that in Mottola's model, at 5.9–7.8 Å and 5.1–7.5 Å, respectively. In addition to these distance constraints, Wilcox detected a pharmacophoric pattern that modeled how high the nitrogen above the plane defined by the catechol ring. According to their model, the ideal N-plane height is within 0.5–2.0 Å. They also found that the distance between the nitrogen atom and the oxygen atom of the *m*-hydroxyl-, and *p*-hydroxyl is at 7.0 Å, and the N-plane height of 1.2 Å would give agonists with greatest D<sub>1</sub>/D<sub>2</sub> selectivity.

Apart from ligand-based studies on D<sub>1</sub> agonist design, numerous attempts have been done toward modeling the interactions between D<sub>1</sub> receptor ligands and the binding sites of the receptor.<sup>82–90</sup> Kalani et al.<sup>88</sup> performed a structure prediction for D<sub>2</sub> DA receptors, which also shed lights on D<sub>1</sub> agonist interaction mechanism on D<sub>1</sub> receptors. Based on the model they built, they successfully incorporated the structure information with large volume of data from biochemical and pharmacological studies. From the interaction patterns between the D<sub>2</sub> receptor and its endogenous ligand DA, it is apparent that the two hydroxyl groups in the catechol ring form several hydrogen bonds with serine residues (Ser5.42, Ser5.43, Ser5.47, numbered according to the Ballesteros–Weinstein designation) in transmembrane (TM) helix 5, whereas the positively charged amino group forms a tight salt bridge with the carboxyl group of Asp3.32. The studies also revealed that certain hydrophobic residues located at TM3, TM4 TM5, and TM6 form a hydrophobic pocket to accommodate the aromatic ring in several agonists. Since the breakthrough in determining the high-resolution crystal structure of bovine rhodopsin,<sup>89</sup> building the homology model of the D<sub>1</sub> receptor is feasible by utilizing this crystal structure as the 3D template. Recently Xhaard et al.<sup>91</sup> investigated the binding modes of catecholamines in the frame of homology models of adrenoceptors and DA receptors constructed from the crystal structure of bovine rhodopsin (PDB access code 1U19). This study clearly demonstrated the important amino acid residues in the receptor binding site that interact with DA receptor ligands, which are similar to the findings by Kalani et al.<sup>88</sup> They also highlighted that several residues may be responsible for the subtype selectivity among DA receptors, such as the SerX12.52 in the D<sub>1</sub>-like receptors and HisX12.52 in the D<sub>2</sub>-like receptors. In the D<sub>1</sub>-like receptors, a more bulky side chain of Ile3.33 (Val3.33 in D<sub>2</sub>-like receptor) may explain why a slightly smaller hydrophobic group is favorable for selective D<sub>1</sub>-like ligands. Recently, a higher homology GPCR,  $\beta$ -adrenergic receptor was determined<sup>90</sup> at 2.4 Å resolution by X-ray crystallography. A comparison with the bovine rhodopsin structures revealed that these two proteins have large deviation on several TM helices, especially in TM3 and TM5, which in turn cause the different orientations of certain residues at the binding sites. These differences raised the question that how accurate the homology models would be. But as demonstrated by many homology structure-based virtual screening studies,<sup>88–94</sup> these models are relatively useful in lead identification and lead optimization if the model was built within the proper constraints provided by experiments. Nevertheless, additional high-resolution structure of GPCR is needed to clarify the agonist-binding conformational states in the activation process.

In summary, the development of computational model of DA receptors has provided extensive useful information and greatly assisted the explanation and prediction of ligand activity and selectivity at the DA receptors. However, a major obstacle in this field is the lack of exact 3D structures of DA receptors at an atomic level that extremely hinders the effective design and development of DA receptor-selective ligands. Current design of DA receptor (D<sub>1</sub>, D<sub>2</sub>) selective ligands still remains largely empirical and somewhat unpredictable. However, the rapid progress in structural genomics<sup>95–97</sup> of membrane proteins and methodology development in structure prediction<sup>98,99</sup> will gradually shift the way to structure-based drug design for developing high selective D<sub>1</sub> agonist as well as for other GPCR modulators. The efforts of attaining the GPCR 3D structure models will eventually provide an interaction map for finding novel scaffolds of D<sub>1</sub> agonists and optimizing the potency and selectivity of lead compounds. Another exciting strategy in GPCR drug development is the emergence of chemogenomics method that aims to construct the drug-target networks. This will sequentially provide additional clues for lead identification and off-target prediction.<sup>100–105</sup>

## 5. DISCOVERY OF NEW D<sub>1</sub> RECEPTOR (PI-D<sub>1</sub>) AND ITS SPECIFIC LIGANDS

In addition to the well-characterized D<sub>1</sub>- and D<sub>2</sub>-like DA receptors, recent evidence suggests that there may be a new type of D<sub>1</sub>-like DA receptors. Unlike the classical D<sub>1A</sub> or D<sub>5</sub> receptors that couple to Gs proteins and stimulate cAMP formation, this novel D<sub>1</sub> receptor selectively couples to G<sub>q</sub> protein and stimulates the hydrolysis of phosphoinositide (PI) to diacylglycerol (DAG) and inositol 1,4,5-trisphosphate (IP<sub>3</sub>) via the activation of PLCβ.<sup>106–115</sup> DAG activates protein kinase C (PKC) whereas IP<sub>3</sub> induces release of calcium from intracellular stores. The putative PI-linked D<sub>1</sub> receptor (PI-D<sub>1</sub>) is widely distributed in the brain.<sup>113</sup> The functional role of PI-D<sub>1</sub> receptor was completely unknown until the recent identification of a selective agonist (SKF-83959, **4**) for this receptor.

SKF-83959 (compound **4**), as described above, belongs to the benzazepine family, and has been characterized early as an agonist for the classic D<sub>1</sub> receptor but producing a novel behavioral profile in rodents.<sup>116,117</sup> It was recently identified that this drug is a selective agonist for the putative PI-linked DA D<sub>1</sub> receptor (PI-D<sub>1</sub>).<sup>113,118</sup> Previous studies have shown that this compound has excellent anti-parkinsonian effects in the MTTP-treated monkey model or 6-OHDA-lesioned rat model of PD.<sup>119,120</sup> The anti-parkinsonian action of this compound exhibits unique features, including: (1) it is effective in L-DOPA-insensitive animals; (2) side effects, such as dyskinesias that are sometimes severe during L-DOPA treatment,<sup>121</sup> disappear with time during SKF-83959 treatment, and (3) the anti-parkinsonian effects are stable over time of treatment, unlike the case with the L-DOPA treatment. Biochemical and pharmacological studies clearly indicate that the action of SKF-83959 is independent of cAMP/PKA pathway.<sup>117–119,122</sup> It was further demonstrated that the anti-parkinsonian action of SKF-83959 appears to be mediated via the PI-D<sub>1</sub> receptor in the 6-OHDA lesioned PD rat model,<sup>115</sup> indicating that stimulation of striatal PLC/IP<sub>3</sub> pathway by the PI-D<sub>1</sub> receptor may be the underlying mechanism for the drug's anti-parkinsonian action. Moreover, it was found that chronic SKF-83959 produced less severe dyskinesia, and attenuated the development of L-DOPA-induced dyskinesia.<sup>123</sup> In addition, the anti-dyskinesia effect of the PI-D<sub>1</sub> receptor agonist appears to be associated with the neuroprotective effect of SKF-83959.<sup>124</sup> In addition to the role of PI-D<sub>1</sub> receptors in the anti-parkinsonian effects, it was also found that SKF-83959 alters the sensoring-motor gating response of rats, and stimulation of this receptor induces activation of CaMKII in brain tissues.<sup>125–128</sup> In view of the importance of CaMKII in neuroplasticity associated with learning and memory, this observation raised a potential interest in exploring the role of PI-D<sub>1</sub> receptor in brain cognitive function. Taken together, all this indicates that the putative PI-D<sub>1</sub> receptor could be a potential target for PD or other neuropsychotic diseases. However, the existence of a novel PI-linked DA receptor is hampered by the failure in the effort to identify the specific gene for this



receptor, and by other controversial reports.<sup>129–132</sup> The discrepancy could be attributed to the differences in cellular contents or partners that requires for DA-mediated Gq/11 activation. Interestingly, Susan George et al.<sup>133</sup> recently provided evidence in support of their proposal that D<sub>1</sub> and D<sub>2</sub> receptor interaction (hetero-oligomeric association of GPCRs) may underlie the DA receptor-coupled Gq/11 pathway. This is an attractive proposal since it could explain the failure of heterologously expressed D<sub>1</sub> receptor cells to activate Gq/11 pathway.

## 6. CLINICAL DEVELOPMENT ON DA D<sub>1</sub> RECEPTOR LIGANDS

As stated above, development of DA D<sub>1</sub> receptor ligands was started much earlier than for other DA receptors. The classical benzazepine analogues, for example, **1** (SCH-23390) and **6** (SKF-38393), were widely used as pharmacological tools for probing the D<sub>1</sub> receptor functions. Several compounds, with high binding affinity and selectivity for the D<sub>1</sub> receptor had even entered clinical trials as anti-parkinsonian or anti-psychotic drugs, but were discontinued later or no development information reported for several years.<sup>134–137</sup> These ligands include conformationally flexible benzazepines, for example, SKF-83566 (**2**), SKF-81297 (**3**), SKF-82958 (**5**), SKF-38393 (**6**), SKF-77434, SKF-87516, SKF-80273, SKF-83565, A-86929 (**31a**), LY-270411, NNC-112, and NNC-22-0010. In addition to their low intrinsic partial agonist activity, the failure of these compounds was largely due to poor oral bioavailability and rapid tolerance.

The only compound that reached the market is fenoldopam (**36**). Benzazepine **36** is a potent D<sub>1</sub> receptor agonist which acts peripherally and selectively produces systemic vasodilation.<sup>137</sup> It was developed by GlaxoSmithKline and was launched as Corlopam in the US in 1998 for short-term management of severe hypertension with or without deteriorating end-organ function. It was licensed to Abbott in the US in 2001 (Figure 8).

The conformationally rigid benzazepine analogues and the polycyclic derivatives have spurred new interests in exploration of the therapeutic potentials of the D<sub>1</sub> receptor agents.<sup>138–143</sup> Several compounds have already been under development. These include: (1) benzazepine analogues: CEE-03-310 (**37**) by Addex for the treatment of drug abuse in 2002, TSR-1938 (**38**) by CeNeS for the treatment of substance abuse and sleep disorders in 2003, Ecopipam (**9**, SCH-39166) by Schering-Plough for the management of obesity in 2000, and (2) polycyclic derivatives: DHX (**27a**) by Purdue University for drug dependence, PD and schizophrenia, **DAR-201** (dinapsoline, **32a**) by DarPharm Inc as treatment for PD, schizophrenia (+)-**32a** by Bristol-Myers Squibb under development for treatment of PD, and **Doxanthrine** (**35a**) by Purdue University as anti-psychotics and as anti-parkinsonian agent. Among these agents, DHX (**27a**) was in Phase I trials with moderate-to-severe PD in 1995 and was pushed to phase II trials. However, negative results, especially poor pharmacokinetic profiles and adverse effects, have been observed in the development process, which make this compound less likely to be used clinically.

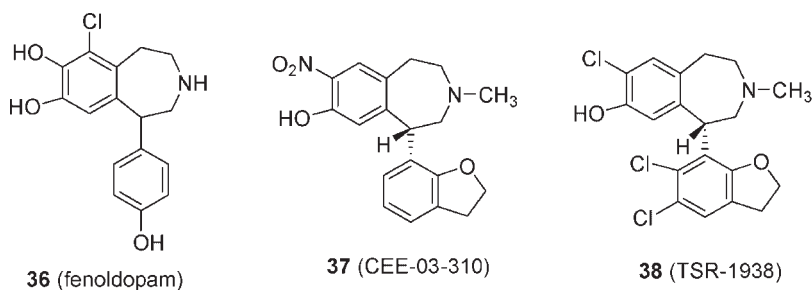


Figure 8. Benzazepine analogues launched or under development.



## 7. CONCLUSION AND PROSPECTIVE

Dopamine D<sub>1</sub> receptor is the most abundant DA receptor subtype among the DA receptor family. Although the first DA D<sub>1</sub> receptor selective ligands SCH-23390 (**1**) was introduced more than two decades ago, clinically useful D<sub>1</sub> receptor selective ligands remain rare. The conformationally flexible 1-phenylbenzazepine analogues have served as the major D<sub>1</sub> receptor agonists and antagonists for many years only resulting in several research tools. The therapeutic utility of D<sub>1</sub> receptor agents was largely suspended for years in 1980s, primarily due to the early hypothesis that D<sub>1</sub> receptor agonists are more or less likely to be associated with dyskinesia than levodopa, and due to the lack of a D<sub>1</sub> full efficacy agonists.<sup>143–148</sup> A renewed interest has not been initiated until the development of polycyclic derivatives, for example, DHX (**27a**) by David E Nichols and Roth B Mailman in the early 1990s. Since then, the development of DA D<sub>1</sub> receptor ligands has fallen in two major categories, dihydroxidine analogues and conformationally rigid benzazepine derivatives with exception of a few others (e.g., isochromans). The former category has opened a new avenue for the development of high affinity full efficacy D<sub>1</sub> receptor agonists, including **31a** (A-86929), **32a** (dinapsoline), **34** (dinoxylene), **35** (doxanthrine). The latter approach has also provided several new benzazepines with D<sub>1</sub> receptor high binding and selectivity, including SCH-39166 (compound **9**).

These advances have highlighted several crucial principles for drug designers to develop full D<sub>1</sub> receptor agonists, and several such agonists have indeed shown full anti-parkinsonian responses that is not supportive of the previous D<sub>1</sub>-induced-dyskinesia hypothesis (e.g., ABT0431).<sup>145,148</sup> Despite of these advances, the chance for any given compound as a treatment of PD or any other CNS disorder is not optimistic because of the many obstacles in pharmacokinetics and toxicity, as evidenced by the example of DHX (**27a**). Further investigations on compounds **9**, **32a**, **34**, and **35**, which are currently in the discovery stage, will be crucial for determining the future of the D<sub>1</sub> receptor agonists or antagonists.

## ACKNOWLEDGMENTS

Financial support for this project was provided by Chinese National Science Foundation (30672517 to AZ), Shanghai Commission of Science and Technology (07pj14104 to AZ), and Ministry of Science and Technology (2007AA02z163 to XZ). Support from Shanghai Institute of Materia Medica is also appreciated.

## REFERENCES

1. Missale C, Nash SR, Robinson SW, Jaber M, Caron MG. Dopamine receptors: From structure to function. *Physiol Rev* 1998;78:189–225.
2. Sidhu A, Niznik HB. Coupling of dopamine receptor subtypes to multiple and diverse G proteins. *Int J Dev Neurosci* 2000;18:669–677.
3. Koob GF. Dopamine, addiction and reward. *Semin Neurosci* 1992;4:139–148.
4. Kuhar MJ, Ritz MC, Boja JW. The dopamine hypothesis of the reinforcing properties of cocaine. *Trends Neurosci* 1991;14:299–302.
5. Witkin JM. Pharmacotherapy of cocaine abuse: Preclinical development. *Neurosci Biobehav Rev* 1994;18:121–142.
6. Stote RM, Dubb JW, Familiar RG, Erb BB, Alexander F. A new oral renal vasodilator, fenoldopam. *Clin Pharmacol Ther* 1983;34:309–315.
7. Civelli O, Bunzow JR, Grandy DK, Zhou QY, Van Tol HH. Molecular biology of the dopamine receptors. *Eur J Pharmacol* 1991;207:277–286.
8. Gingrich JA, Caron MG. Recent advances in the molecular biology of dopamine receptors. *Annu Rev Neurosci* 1993;16:299–321.

9. Luo Y, Kokkonen GC, Wang X, Neve KA, Roth GS. D<sub>2</sub> dopamine receptors stimulate mitogenesis through pertussis toxin-sensitive G proteins and Ras-involved ERK and SAP/JNK pathways in rat C6-D<sub>2</sub>L glioma cells. *J Neurochem* 1998;71:980–990.
10. Zhen X, Zhang J, Johnson GP, Friedman E. D<sub>4</sub> dopamine receptor differentially regulates Akt/nuclear factor- $\kappa$ B and extracellular signal-regulated kinase pathways in D4MN9D cells. *Mol Pharm* 2001;60:857–864.
11. Cai G, Zhen X, Uryu K, Friedman E. Activation of extracellular signal-regulated protein kinases is associated with a sensitized locomotor response to D<sub>2</sub> dopamine receptor stimulation in unilateral 6-hydroxydopamine-lesioned rats. *J Neurosci* 2000;20:1849–1857.
12. Zhen X, Uryu K, Wang HY, Friedman E. D<sub>1</sub> dopamine receptor agonists mediate activation of p38 mitogen-activated protein kinase and c-Jun amino-terminal kinase by a protein kinase A-dependent mechanism in SK-N-MC human neuroblastoma cells. *Mol Pharm* 1998;54:453–458.
13. Schneider JS, Sun ZQ, Roeltgen DP. Effects of dihydrexidine, a full dopamine D-1 receptor agonist, on delayed response performance in chronic low dose MPTP-treated monkeys. *Brain Res* 1994;663:140–144.
14. Kohli JD, Horn PT, Glock D, Brewster WK, Nichols DE. Dihydrexidine: A new potent peripheral dopamine D<sub>1</sub> receptor agonist. *Eur J Pharmacol* 1993;235:31–35.
15. Agid Y. Parkinson's disease: Pathophysiology. *Lancet* 1991;337:1321–1324.
16. Olanow CW, Obeso JA, Stocchi F. Continuous dopamine-receptor treatment of Parkinson's disease: Scientific rationale and clinical implications. *Lancet Neurol* 2006;5:677–687.
17. Jenner P. Dopamine agonists, receptor selectivity and dyskinesia induction in Parkinson's disease. *Curr Opin Neurol* 2003;16: (Suppl 1); S3–S7.
18. Lewis MM, Huang X, Nichols DE, Mailman RB. D<sub>1</sub> and, functionally selective dopamine agonists as neuroprotective agents in Parkinson's disease. *CNS Neurol Disord Drug Targets* 2006;5:345–353.
19. Radad K, Gille G, Rausch WD. Short review on dopamine agonists: Insight into clinical and research studies relevant to Parkinson's disease. *Pharmacol Rep* 2005;57:701–712.
20. Foley P, Gerlach M, Double KL, Riederer P. Dopamine receptor agonists in the therapy of Parkinson's disease. *J Neural Transm* 2004;111:1375–1446.
21. Setler PE, Sarau HM, Zirkle CL, Saunders HL. The central effects of a novel dopamine agonist. *Eur J Pharmacol* 1978;50:419–430.
22. Watts VJ, Lawler CP, Gonzales AJ, Zhou QY, Civelli O, Nichols DE, Mailman RB. Spare receptors and intrinsic activity: Studies with D<sub>1</sub> dopamine receptor agonists. *Synapse* 1995;21:177–187.
23. Lovenberg TW, Brewster WK, Mottola DM, Lee RC, Riggs RM, Nichols DE, Lewis MH, Mailman RB. Dihydrexidine, a novel selective high potency full dopamine D-1 receptor agonist. *Eur J Pharmacol* 1989;166:111–113.
24. Mottola DM, Brewster WK, Cook LL, Nichols DE, Mailman RB. Dihydrexidine, a novel full efficacy D<sub>1</sub> dopamine receptor agonist. *J. Pharmacol Exp Ther* 1992;262:383–393.
25. Blanchet PJ, Fang J, Gillespie M, Sabounjian L, Locke KW, Gammans R, Mouradian MM, Chase TN. Effects of the full dopamine D<sub>1</sub> receptor agonist dihydrexidine in Parkinson's disease. *Clin Neuropharmacol* 1998;21:339–343.
26. Gerlach M, Double K, Arzberger T, Leblhuber F, Tatschner T, Riederer P. Dopamine receptor agonists in current clinical use: Comparative dopamine receptor binding profiles defined in the human striatum. *J Neural Transm* 2003;110:1119–1127.
27. Goldman-Rakic PS, Castner SA, Svensson TH, Siever LJ, Williams GV. Targeting the dopamine D<sub>1</sub> receptor in schizophrenia: Insights for cognitive function. *Psychopharmacology (Berl)* 2004;174:3–16.
28. Arnsten AF, Cai JX, Murphy BL, Goldman-Rakic PS. Dopamine D<sub>1</sub> receptor mechanisms in the cognitive performance of young adult and aged monkeys. *Psychopharmacology (Berl)* 1994;116:143–151.
29. Miyamoto S, Duncan GE, Marx CE, Lieberman JA. Treatments for schizophrenia: A critical review of pharmacology and mechanisms of action of antipsychotic drugs. *Mol Psychiatry* 2005;10:79–104.
30. George MS, Molnar CE, Grenesko EL, Anderson B, Mu Q, Johnson K, Nahas Z, Knable M, Fernandes P, Juncos J, Huang X, Nichols DE, Mailman RB. A single 20 mg dose of dihydrexidine (DAR-0100), a full dopamine D<sub>1</sub> agonist, is safe and tolerated in patients with schizophrenia. *Schizophr Res* 2007;93:42–50.
31. Mu Q, Johnson K, Morgan PS, Grenesko EL, Molnar CE, Anderson B, Nahas Z, Kozel FA, Kose S, Knable M, Fernandes P, Nichols DE, Mailman RB, George MS. A single 20 mg dose of the full D<sub>1</sub> dopamine agonist dihydrexidine (DAR-0100) increases prefrontal perfusion in schizophrenia. *Schizophr Res* 2007;94:332–341.
32. Stone JM, Pilowsky LS. Novel targets for drugs in schizophrenia. *CNS & Neurological Disorders—Drug Targets* 2007;6:265–272.

33. Taylor JR, Lawrence MS, Redmond DE, Jr., Elsworth JD, Roth RH, Nichols DE, Mailman RB. Dihydropyridine, a full dopamine D<sub>1</sub> agonist, reduces MPTP-induced parkinsonism in monkeys. *Eur J Pharmacol* 1991;199:389–391.
34. Charifson PS, Wyrick SD, Hoffman AJ, Simmons RM, Bowen JP, McDougald DL, Mailman RB. Synthesis and pharmacological characterization of 1-phenyl-, 4-phenyl-, and 1-benzyl-1,2,3,4-tetrahydroisoquinolines as dopamine receptor ligands. *J Med Chem* 1988;31:1941–1946.
35. Hoffman B, Cho SJ, Zheng W, Wyrick S, Nichols DE, Mailman RB, Tropsha A. Quantitative structure-activity relationship modeling of dopamine D<sub>1</sub> antagonists using comparative molecular field analysis, genetic algorithms-partial least-squares, and K nearest neighbor methods. *J Med Chem* 1999;42:3217–3226.
36. Neumeyer JL, Kula NS, Bergman J, Baldessarini RJ. Receptor affinities of dopamine D<sub>1</sub> receptor-selective novel phenylbenzazepines. *Eur J Pharmacol* 2003;474:137–140.
37. Charifson PS, Bowen JP, Wyrick SD, Hoffman AJ, Cory M, McPhail AT, Mailman RB. Conformational analysis and molecular modeling of 1-phenyl-, 4-phenyl-, and 1-benzyl-1,2,3,4-tetrahydroisoquinolines as D<sub>1</sub> dopamine receptor ligands. *J Med Chem* 1989;32:2050–2058.
38. Keababian JW, Britton DR, DeNinno MP, Perner R, Smith L, Jenner P, Schoenleber R, Williams M. A-77636: A potent and selective dopamine D<sub>1</sub> receptor agonist with antiparkinsonian activity in marmosets. *Eur J Pharmacol* 1992;229:203–209.
39. Oloff S, Mailman RB, Tropsha A. Application of validated QSAR models of D<sub>1</sub> dopaminergic antagonists for database mining. *J Med Chem* 2005;48:7322–7332.
40. Strange PG. The binding of agonists and antagonists to dopamine receptors. *Biochem Soc Trans* 1996;24:188–192.
41. Claudi F, Cingolani GM, Di Stefano A, Giorgioni G, Amenta F, Barili P, Ferrari F, Giuliani D. Synthesis, resolution, and preliminary evaluation of trans-2-Amino-6(5)-hydroxy-1-phenyl-2,3-dihydro-1H-indenes and related derivatives as dopamine receptors ligands. *J Med Chem* 1996;39:4238–4246.
42. Seiler MP, Hagenbach A, Wüthrich HJ, Markstein R. Trans-hexahydroindolo[4,3-*ab*] phenanthridines (“benzergolines”), the first structural class of potent and selective dopamine D<sub>1</sub> receptor agonists lacking a catechol group. *J Med Chem* 1991;34:303–307.
43. Imperato A, Di Chiara G. CY, 208–243, a novel dopamine D-1 receptor agonist, fails to modify dopamine release in freely moving rats. *Eur J Pharmacol* 1989;160:155–158.
44. Witt T, Hock FJ, Lehmann J. 7-methyl-6,7,8,9,14,15-hexahydro-5H-benz[d]indolo[2,3-*g*]azecine: A new heterocyclic system and a new lead compound for dopamine receptor antagonists. *J Med Chem* 2000;43:2079–2081.
45. Snyder SE, Aviles-Garay FA, Chakraborti R, Nichols DE, Watts VJ, Mailman RB. Synthesis and evaluation of 6,7-dihydroxy-2,3,4,8,9,13b-hexahydro-1H-benzo[6,7] cyclohepta[1,2,3-*ef*][3] benzazepine, 6,7-Dihydroxy-1,2,3,4,8,12b-hexahydroanthr [10,4a,4-*cd*]azepine, and 10-(aminomethyl)-9,10-dihydro-1,2-dihydroxyanthracene as conformationally restricted analogs of β-Phenyldopamine. *J Med Chem* 1995;38:2395–2409.
46. Zhang A, Neumeyer JL, Baldessarini RJ. Recent progress in development of dopamine receptor subtype-selective agents: Potential therapeutics for neurological and psychiatric disorders. *Chem Rev* 2007;107:274–303.
47. Barnett A. Review on dopamine receptors. *Drugs Future* 1986;11:49–56.
48. Berger JG, Chang WK, Clader JW, Hou D, Chipkin RE, Mcphail AT. Synthesis and receptor affinities of some conformationally restricted analogues of the dopamine D<sub>1</sub> selective ligand (5R)-8-Chloro-2,3,4,5-tetrahydro-3-methyl-5-phenyl-1H-3-benzazepin-7-ol. *J Med Chem* 1989;32:1913–1921.
49. Weinstock J, Oh HJ, DeBrosse CW, Eggleston DS, Wise M, Flaim KE, Gessner GW, Sawyer JL, Kaiser C. Synthesis, conformation, and dopaminergic activity of 5,6-ethano-bridged derivatives of selective dopaminergic 3-benzazepines. *J Med Chem* 1987;30:1303–1308.
50. Dandridge PA, Kaiser C, Brenner M, Gaitanopoulos D, Davis LD, Webb RL, Foley JJ, Sarau HM. Synthesis, resolution, absolute stereochemistry, and enantioselectivity of 3',4'-dihydroxynomifensine. *J Med Chem* 1984;27:28–35.
51. Berger JG, Chang WK, Gold EH, Clader JW. New fused benzazepine compounds—having analgesic, anticholinergic, anti-aggressive, tranquillising and renal vasodilator properties. EP230270-A(1987) Schering Corp.
52. Barnett A, McQuade RD, Tedford C. Highlights of D<sub>1</sub> dopamine receptor antagonist research. *Neurochem Int* 1992;20: (Suppl 1); S119–S122.
53. Pettersson I, Liljefors T, Bøgesø K. Conformational analysis and structure-activity relationships of selective dopamine D<sub>1</sub> receptor agonists and antagonists of benzazepine series. *J Med Chem* 1990;33:2197–2204.

54. Wu WL, Burnett DA, Spring R, Greenlee WJ, Smith M, Favreau L, Fawzi A, Zhang H, Lachowicz JE. Dopamine D<sub>1</sub>/D<sub>5</sub> receptor antagonists with improved pharmacokinetics: Design, synthesis, and biological evaluation of phenol bioisosteric analogues of benzazepine D<sub>1</sub>/D<sub>5</sub> antagonists. *J Med Chem* 2005;48:680–693.
55. Burnett DA, Greenlee WJ, Mckirtrick B, Su J, Zhu Z, Sasikumar TK, Mazzola R, Qiang L, Ye Y. New indane-fused 2,3,4,5-tetrahydro-1H-benzo(d)azepine derivatives useful for treating metabolic disorder, eating disorder, diabetes, obsessive-compulsive disorder and autism. US 2005075325 (2005) Schering Corp.
56. Hohlweg R, Nielsen E, Nielsen EB. Novel tricyclic 2,3,4,5-tetrahydro-1H-3-benzazepines—To treat dysfunctions of the dopaminergic receptor systems e.g. psychosis, pain, Parkinson's disease, depression. WO9420472(1994)Novo-Nordisk As.
57. Riggs RM, Nichols DE, Foreman MM, Truex LL, Glock D, Kohli JD. Specific dopamine D-1 and DA<sub>1</sub> properties of 4-(mono- and -dihydroxyphenyl)-1,2,3,4-tetrahydroisoquinoline and its tetrahydrothieno[2,3-c]pyridine analogue. *J Med Chem* 1987;30:1454–1458.
58. Riggs RM, Nichols DE, Foreman MM, Truex LL. Evaluation of isomeric 4-(chlorohydroxyphenyl)-1,2,3,4-tetrahydroisoquinolines as dopamine D-1 antagonists. *J Med Chem* 1987;30:1887–1891.
59. Brewster WK, Nichols DE, Riggs RM, Mottola DM, Lovenberg TW, Lewis MH, Mailman RB. Trans-10,11-dihydroxy-5,6,6a,7,8,12b-hexahydrobenzo[a] phenanthridine: A highly potent selective dopamine D<sub>1</sub> full agonist. *J Med Chem* 1990;33:1756–1764.
60. Wilcox RE, Tseng T, Brusniak MY, Ginsburg B, Pearlman RS, Teeter M, DuRand C, Starr S, Neve KA. CoMFA-based prediction of agonist affinities at recombinant D<sub>1</sub> vs D<sub>2</sub> dopamine receptors. *J Med Chem* 1998;41:4385–4399.
61. Wei CC, Teitel S. Synthesis of a benzo[a]phenanthridine isomeric with apomorphine. *Heterocycles* 1977;8:97–102.
62. Knoerzer TA, Nichols DE, Brewster WK, Watts VJ, Mottola D, Mailman RB. Dopaminergic Benzo[a]phenanthridines: Resolution and pharmacological evaluation of the enantiomers of dihydrexidine, the full efficacy D<sub>1</sub> dopamine receptor agonist. *J Med Chem* 1994;37:2453–2460.
63. Minor DL, Wyrick SD, Charifson PS, Watts VJ, Nichols DE, Mailman RB. Synthesis and molecular modeling of 1-phenyl-1,2,3,4-tetrahydroisoquinolines and related 5,6,8,9-tetrahydro-13bH-dibenzo[a,h]-quinolizines as D<sub>1</sub> dopamine antagonists. *J Med Chem* 1994;37:4317–4328.
64. Michaelides MR, Hong Y, DiDomenico S, Jr., Asin KE, Britton DR, Lin CW, Williams M, Shiosaki K. (5aR,11bS)-4,5,5a,6,7,11b-hexahydro-2-propyl-3-thia-5-azacyclopent-1-ena[c]-phenanthrene-9,10-diol (A-86929): A potent and selective dopamine D<sub>1</sub> agonist that maintains behavioral efficacy following repeated administration and characterization of its diacetyl prodrug (ABT-431). *J Med Chem* 1995;38:3445–3447.
65. Ghosh D, Snyder SE, Watts VJ, Mailman RB, Nichols DE. 9-Dihydroxy-2,3,7,11b-tetrahydro-1H-naph[1,2,3-de]isoquinoline: A potent full dopamine D<sub>1</sub> agonist containing a rigid-beta-phenyldopamine pharmacophore. *J Med Chem* 1996;39:549–555.
66. Sit SY, Xie K, Jacutin-Porte S, Taber MT, Gulwadi AG, Korpinen CD, Burris KD, Molski TF, Ryan E, Xu C, Wong H, Zhu J, Krishnananthan S, Gao Q, Verdoorn T, Johnson G. (+)-Dinapsoline: An efficient synthesis and pharmacological profile of a novel dopamine agonist. *J Med Chem* 2002;45:3660–3668.
67. Qandil AM, Lewis MM, Jassen A, Leonard SK, Mailman RB, Nichols DE. Synthesis and pharmacological evaluation of substituted naphth[1,2,3-de]isoquinolines (dinapsoline analogues) as D<sub>1</sub> and D<sub>2</sub> dopamine receptor ligands. *Bioorg Med Chem* 2003;11:1451–1464.
68. Sit SY, Xie K, Jacutin-Porte S, Boy KM, Seanz J, Taber MT, Gulwadi AG, Korpinen CD, Burris KD, Molski TF, Ryan E, Xu C, Verdoorn T, Johnson G, Nichols DE, Mailman RB. Synthesis and SAR exploration of dinapsoline analogues. *Bioorg Med Chem* 2004;12:715–734.
69. Negash K, Nichols DE, Watts VJ, Mailman RB. Further definition of the D<sub>1</sub> dopamine receptor pharmacophore: Synthesis of trans-6,6a,7,8,9,13b-hexahydro-5H-benzod[2,1-b]azepines as rigid analogues of  $\beta$ -phenyldopamine. *J Med Chem* 1997;40:2140–2147.
70. Grubbs RA, Lewis MM, Owens-Vance C, Gay EA, Jassen AK, Mailman RB, Nichols DE. 8,9-dihydroxy-1,2,3,11b-tetrahydrochromeno[4,3,2,-de]isoquinoline (dinoxyline), a high affinity and potent agonist at all dopamine receptor isoforms. *Bioorg Med Chem* 2004;12:1403–1412.
71. Cueva JP, Giorgioni G, Grubbs RA, Chemel BR, Watts VJ, Nichols DE. trans-2,3-dihydroxy-6a,7,8,12b-tetrahydro-6H-chromeno[3,4-c]isoquinoline: Synthesis, resolution, and preliminary pharmacological characterization of a new dopamine D<sub>1</sub> receptor full agonist. *J Med Chem* 2006;49:6848–6857.
72. Ortore G, Tuccinardi T, Bertini S, Martinelli A. A theoretical study to investigate D<sub>2</sub>DAR/D<sub>4</sub>DAR selectivity: Receptor modeling and molecular docking of dopaminergic ligands. *J Med Chem* 2006;49:1397–1407.

73. Kim HJ, Cho YS, Koh HY, Kong JY, No KT, Pae AN. Classification of dopamine antagonists using functional feature hypothesis and topological descriptors. *Bioorg Med Chem* 2006;14:1454–1461.
74. Samanta S, Debnath B, Gayen S, Ghosh B, Basu A, Srikanth K, Jha T. QSAR modeling on dopamine D<sub>2</sub> receptor binding affinity of 6-methoxy benzamides. *Farmaco* 2005;60:818–825.
75. Klabunde T, Evers A. GPCR antitarget modeling: Pharmacophore models for biogenic amine binding GPCRs to avoid GPCR-mediated side effects. *Chembiochem* 2005;6:876–889.
76. Sukalović V, Zlatović M, Andrić D, Roglić G, Kostić-Rajacčić S, Soskić V. Modeling of the D<sub>2</sub> dopamine receptor arylpiperazine binding site for 1-[2-[5-(1H-benzimidazole-2-thione)]ethyl]-4-aryl piperazines. *Arch Pharm* 2004;337:502–512.
77. Shacham S, Marantz Y, Bar-Haim S, Kalid O, Warshaviak D, Avisar N, Inbal B, Heifetz A, Fichman M, Topf M, Naor Z, Noiman S, Becker OM. PREDICT modeling and in-silico screening for G-protein coupled receptors. *Proteins* 2004;57:51–86.
78. Varady J, Wu X, Fang X, Min J, Hu Z, Levant B, Wang S. Molecular modeling of the three-dimensional structure of dopamine 3 (D<sub>3</sub>) subtype receptor: Discovery of novel and potent D<sub>3</sub> ligands through a hybrid pharmacophore- and structure-based database searching approach. *J Med Chem* 2003;46:4377–4392.
79. Mottola DM, Laiter S, Watts VJ, Tropsha A, Wyrick SD, Nichols DE, Mailman RB. Conformational analysis of D<sub>1</sub> dopamine receptor agonists: Pharmacophore assessment and receptor mapping. *J Med Chem* 1996;39:285–296.
80. Wilcox RE, Tseng T, Brusniak MY, Ginsburg B, Pearlman RS, Teeter M, DuRand C, Starr S, Neve KA. CoMFA-based prediction of agonist affinities at recombinant D<sub>1</sub> vs D<sub>2</sub> dopamine receptors. *J Med Chem* 1998;41:4385–4399.
81. Wilcox RE, Huang WH, Brusniak MY, Wilcox DM, Pearlman RS, Teeter MM, DuRand CJ, Wiens BL, Neve KA. CoMFA-based prediction of agonist affinities at recombinant wild type versus serine to alanine point mutated D<sub>2</sub> dopamine receptors. *J Med Chem* 2000;43:3005–3019.
82. Homan EJ, Wikström HV, Grol CJ. Molecular modeling of the dopamine D<sub>2</sub> and serotonin 5-HT<sub>1A</sub> receptor binding modes of the enantiomers of 5-OMe-BPAT. *Bioorg Med Chem* 1999;7:1805–1820.
83. Hobrath JV, Wang S. Computational elucidation of the structural basis of ligand binding to the dopamine 3 receptor through docking and homology modeling. *J Med Chem* 2006;49:4470–4476.
84. Shi L, Javitch JA. The second extracellular loop of the dopamine D<sub>2</sub> receptor lines the binding-site crevice. *Proc Natl Acad Sci USA* 2004;101:440–445.
85. Kortagere S, Welsh WJ. Development and application of hybrid structure based method for efficient screening of ligands binding to G-protein coupled receptors. *J Comput Aided Mol Des* 2006;20:789–802.
86. Al-Fulaij MA, Ren Y, Beinborn M, Kopin AS. Identification of amino acid determinants of dopamine 2 receptor synthetic agonist function. *J Pharmacol Exp Ther* 2007;321:298–307.
87. Ishiguro M. Ligand-binding modes in cationic biogenic amine receptors. *Chembiochem* 2004;5:1210–1219.
88. Kalani MY, Vaidehi N, Hall SE, Trabanino RJ, Freddolino PL, Kalani MA, Floriano WB, Kam VW, Goddard WA. The predicted 3D structure of the human D<sub>2</sub> dopamine receptor and the binding site and binding affinities for agonists and antagonists. *Proc Natl Acad Sci USA* 2004;101:3815–3820.
89. Gether U. Uncovering molecular mechanisms involved in activation of G protein-coupled receptors. *Endocr Rev* 2000;21:90–113.
90. Cherezov V, Rosenbaum DM, Hanson MA, Rasmussen SG, Thian FS, Kobilka TS, Choi HJ, Kuhn P, Weis WI, Kobilka BK, Stevens RC. High-resolution crystal structure of an engineered human beta2-adrenergic G protein-coupled receptor. *Science* 2007;318:1258–1265.
91. Xhaard H, Rantanen VV, Nyrönen T, Johnson MS. Molecular evolution of adrenoceptors and dopamine receptors: Implications for the binding of catecholamines. *J Med Chem* 2006;49:1706–1719.
92. Evers A, Hessler G, Matter H, Klabunde T. Virtual screening of biogenic amine-binding G-protein coupled receptors: Comparative evaluation of protein- and ligand-based virtual screening protocols. *J Med Chem* 2005;48:5448–5465.
93. Evers A, Klabunde T. Structure-based drug discovery using GPCR homology modeling: Successful virtual screening for antagonists of the alpha1A adrenergic receptor. *J Med Chem* 2005;48:1088–1097.
94. Bissantz C, Bernard P, Hibert M, Rognan D. Protein-based virtual screening of chemical databases. II. Are homology models of G-protein coupled receptors suitable targets? *Proteins* 2003;50:5–25.
95. Lundstrom K. Structural genomics and drug discovery. *J Cell Mol Med* 2007;11:224–238.
96. Lundstrom K, Wagner R, Reinhart C, Desmyter A, Cherouati N, Magnin T, Zeder-Lutz G, Courtot M, Prual C, Andre N, Hassaine G, Michel H, Cambillau C, Pattus F. Structural genomics on membrane proteins: Comparison of more than 100 GPCRs in 3 expression systems. *J Struct Funct Genomics* 2006;7:77–91.



97. Rosenbaum DM, Cherezov V, Hanson MA, Rasmussen SG, Thian FS, Kobilka TS, Choi HJ, Yao XJ, Weis WI, Stevens RC, Kobilka BK. GPCR engineering yields high-resolution structural insights into beta2-adrenergic receptor function. *Science* 2007;318:1266–1273.
98. Ginalski K. Comparative modeling for protein structure prediction. *Curr Opin Struct Biol* 2006;16:172–177.
99. Yarov-Yarovoy V, Schonbrun J, Baker D. Multipass membrane protein structure prediction using Rosetta. *Proteins* 2006;62:1010–1025.
100. Bredel M, Jacoby E. Chemogenomics: An emerging strategy for rapid target and drug discovery. *Nat Rev Genet* 2004;5:262–275.
101. Bender A, Young DW, Jenkins JL, Serrano M, Mikhailov D, Clemons PA, Davies JW. Chemogenomic data analysis: Prediction of small-molecule targets and the advent of biological fingerprint. *Comb Chem High Throughput Screen* 2007;10:719–731.
102. Rognan D. Chemogenomic approaches to rational drug design. *Br J Pharmacol* 2007;152:38–52.
103. Harris CJ, Stevens AP. Chemogenomics: Structuring the drug discovery process to gene families. *Drug Discov Today* 2006;11:880–888.
104. Martin RE, Green LG, Guba W, Kratochwil N, Christ A. Discovery of the first nonpeptidic, small-molecule, highly selective somatostatin receptor subtype 5 antagonists: A chemogenomics approach. *J Med Chem* 2007;50:6291–6294.
105. Klabunde T, Jager R. Chemogenomics approaches to G-protein coupled receptor lead finding. *Ernst Schering Res Found Workshop* 2006; 31–46.
106. Felder CC, Jose PA, Axelrod J. The dopamine-1 agonist, SKF82526, stimulates phospholipase-C activity independent of adenylyl cyclase. *J Pharmacol Exp Ther* 1989;248:171–175.
107. Undie AS, Friedman E. Stimulation of a dopamine D<sub>1</sub> receptor enhances inositol phosphates formation in rat brain. *J Pharmacol Exp Ther* 1990;253:987–992.
108. Fraill DE, Manelli AM, Witte DG, Lin CW, Steffey ME, Mackenzie RG. Cloning and characterization of a truncated dopamine D<sub>1</sub> receptor from goldfish retina: Stimulation of cyclic AMP production and calcium mobilization. *Mol Pharmacol* 1993;44:1113–1118.
109. Undie AS, Weinstock J, Sarau HM, Friedman E. Evidence for a distinct D<sub>1</sub>-like dopamine receptor that couples to activation of phosphoinositide metabolism in brain. *J Neurochem* 1994;62:2045–2048.
110. Wang HY, Undie AS, Friedman E. Evidence for the coupling of Gq protein to D<sub>1</sub>-like dopamine sites in rat striatum: Possible role in dopamine-mediated inositol phosphate formation. *Mol Pharmacol* 1995;48:988–994.
111. Pacheco MA, Jope RS. Comparison of [<sup>3</sup>H] phosphatidylinositol and [<sup>3</sup>H] phosphatidylinositol 4, 5-bisphosphate hydrolysis in postmortem human brain membranes and characterization of stimulation by dopamine D<sub>1</sub> receptors. *J Neurochem* 1997;69:639–644.
112. Jin LQ, Cai GP, Wang HY, Smith C, Friedman E. Characterization of the phosphoinositide-linked dopamine receptor in a mouse hippocampal-neuroblastoma cell line. *J Neurochem* 1998;71:1935–1943.
113. Jin L, Goswami S, Cai G, Zhen X, Friedman E. SKF83959 selectively regulates phosphatidylinositol-linked D<sub>1</sub> dopamine receptors in rat brain. *J Neurochem* 2003;85:378–386.
114. Ming Y, Zhang H, Chen J, Zhen X. SKF83959 stimulates Ca<sup>2+</sup> signaling in primary cultured hippocampal neurons. *J Neurochem* 2005;98:1316–1323.
115. Zhen X, Gaswami S, Abdali SA. Regulation of cdk5 and CaMKII by PI-linked dopamine receptor in rat brain. *Mol Pharmacol* 2004;66:1500–1507.
116. Deveney AM, Waddington JL. Pharmacological characterization of behavioral responses to SKF 83959 in relation to “D<sub>1</sub>-like” dopamine receptors not linked to adenylyl cyclase. *Br J Pharmacol* 1995;116:2120–2126.
117. Andringa G, Drukarch B, Luyen JE, Cools AR, Stoof JC. The alleged dopamine D<sub>1</sub> receptor agonist SKF 83959 is a dopamine D<sub>1</sub> receptor antagonist in primate cells and interacts with other receptors. *Eur J Pharmacol* 1999;364:33–41.
118. Panchalingam S, Undie AS. SKF83959 exhibits biochemical agonism by stimulation [<sup>35</sup>S] GTPγS binding and phosphoinositide hydrolysis in rat and monkey brain. *Neuropharmacol* 2001;40:826–837.
119. Arnt J, Hyttel J, Sánchez C. Partial and full dopamine D<sub>1</sub> receptor agonists in mice and rats: Relation between behavioral effects and stimulation of adenylyl cyclase activity in vitro. *Eur J Pharmacol* 1992;213:259–267.
120. Andringa G, Stoof JC, Cools AR. Sub-chronic administration of the dopamine D<sub>1</sub> antagonist SKF 83959 in bilaterally MPTP-treated rhesus monkeys: Stable therapeutic effects and wearing-off dyskinesia. *Psychopharmacol* 1999;146:328–334.
121. Togasaki DM, Tan L, Protell P, DiMonte DA, Quik M, Langston JW. Levodopa induces dyskinesias in normal squirrel monkeys. *Ann Neurol* 2001;50:254–257.

122. Adachi K, Ikeda H, Hasegawa M, Nakamura S, Waddington JL, Koshikawa N. SKF 83959 and non-cyclase-coupled dopamine D<sub>1</sub>-like receptors in jaw movements via dopamine D<sub>1</sub>-like/D<sub>2</sub>-like receptor synergism. *Eur J Pharmacol* 1999;367:143–149.
123. Zhang H, Chen JG, Zhen X. SKF83959 attenuated L-DOPA-induced dyskinesia in 6-OHDA-lesioned rat model of Parkinson's disease. *Neuropharmacol* 2007;53:125–133.
124. Yu Y, Wang JR, Sun PH, Guo Y, Zhang Z, Jin GZ, Zhen X. Neuroprotective effects of atypical D<sub>1</sub> receptor agonist SKF83959 are mediated via D<sub>1</sub> receptor dependent inhibition of glycogen synthase kinase-3 $\beta$  and a receptor independent anti-oxidative action. *J Neurochem* 2007; (in press).
125. Zhang ZJ, Jiang XL, Zhang SE, Zhang Y, Hough CJ, Li H, Zhen X. The paradoxical effects of SKF83959, a novel dopamine D<sub>1</sub>-like receptor agonist, in the rat acoustic startle reflex paradigm. *Neurosci Lett* 2005; 382:134–138.
126. Zhen X, Goswami S, Friedman E. Activation of phosphatidylinositol (PI)-linked D<sub>1</sub> dopamine receptor is associated with the antiparkinsonian action of SKF83959. *Biochem Pharm Behav* 2005;80:597–601.
127. Friedman E, Jin LQ, Cai GP, Hollon TR, Drago J, Sibley DR, Wang HY. D<sub>1</sub>-like dopaminergic activation of phosphoinositide hydrolysis is independent of D<sub>1A</sub> dopamine receptors: Evidence from D<sub>1A</sub> knockout mice. *Mol Pharmacol* 1997;51:6–11.
128. Gnanalingham KK, Hunter AJ, Jenner P, Marsden CD. Stimulation of adenylate cyclase activity by benzazepine D-1 dopamine agonists with varying efficacies in the 6-hydroxydopamine lesioned rat—Relationship to circling behaviour. *Biochem Pharmacol* 1995;49:1185–1193.
129. Mailman RB, Schulz DW, Kilts CD, Lewis MH, Rollema H, Wyrick S. Multiple forms of the D<sub>1</sub> dopamine receptor: Its linkage to adenylate cyclase and psychopharmacological effects. *Psychopharmacol Bull* 1986;22:593–598.
130. Mailman RB, Schulz DW, Kilts CD, Lewis MH, Rollema H, Wyrick S. The multiplicity of the D<sub>1</sub> dopamine receptor. *Adv Exp Med Biol* 1986;204:53–72.
131. Leonard SK, Anderson CM, Lachowicz JE, Schulz DW, Kilts CD, Mailman RB. Amygdaloid D<sub>1</sub> receptors are not linked to stimulation of adenylate cyclase. *Synapse* 2003;50:320–333.
132. Braun A, Fabbri G, Mouradian MM, Serrati C, Barone P, Chase TN. Selective D-1 dopamine receptor agonist treatment of Parkinson's disease. *J Neural Transm* 1987;68:41–50.
133. Rashid AJ, So CH, Kong MM, Furtak T, El-Ghundi M, Cheng R, O'Dowd BF, George SR. D<sub>1</sub>-D<sub>2</sub> dopamine receptor heterooligomers with unique pharmacology are coupled to rapid activation of Gq/11 in the striatum. *Proc Nat Acad Sci USA* 2007;104:654–659.
134. Rashid AJ, O'Dowd BF, Verma V, George SR. Neuronal Gq/11-coupled dopamine receptors: An uncharted role for dopamine. *Trends Pharmacol Sci* 2007;28:551–555.
135. Emre M, Rinne UK, Rascol A, Lees A, Agid Y, Lataste X. Effects of a selective partial D<sub>1</sub> agonist, CY 208–243, in de novo patients with Parkinson disease. *Mov Disord* 1992;7:239–243.
136. Mailman R, Huang X, Nichols DE. Parkinson's disease and D<sub>1</sub> dopamine receptors. *Curr Opin Invest Drugs* 2001;2:1582–1591.
137. Brogden RN, Markham A. Fenoldopam: A review of its pharmacodynamic and pharmacokinetic properties and intravenous clinical potential in the management of hypertensive urgencies and emergencies. *Drugs* 1997;54:634–635.
138. Fornai F, di Poggio AB, Pellegrini A, Ruggieri S, Paparelli A. Noradrenaline in Parkinson's disease: From disease progression to current therapeutics. *Curr Med Chem* 2007;14:2330–2334.
139. Contin M, Riva R, Albani F, Baruzzi A. Pharmacokinetic optimization of dopamine receptor agonist therapy for Parkinson's disease. *CNS Drugs* 2000;14:439–455.
140. Kebabian JW, Tarazi FI, Kula NS, Baldessarini RJ. Compounds selective for dopamine receptor subtypes. *Drug Discov Today* 1997;2:333–340.
141. Zhang A, Kan Y, Li F. Recent advances towards the discovery of dopamine receptor ligands. *Expert Opin Ther Pat* 2006;16:587–630.
142. Rezak M. Current Pharmacotherapeutic treatment options in Parkinson's disease. *Dis Mon* 2007;53:214–222.
143. Salmi P, Isacson R, Kull B. Dihydropyridine—The first full dopamine D<sub>1</sub> receptor agonist. *CNS Drug Rev* 2004;10:230–242.
144. Grondin R, Bedard PJ, Britton DR, Shiosaki K. Potential therapeutic use of the selective dopamine D<sub>1</sub> receptor agonist, A-86929: An acute study in parkinsonian levodopa-primed monkeys. *Neurology* 1997;49:421–426.
145. Rascol O, Nutt JG, Blin O, Goetz CG, Trugman JM, Soubrouillard C, Carter JH, Currie LJ, Fabre N, Thalamas C, Giardina WJ, Wright S. Induction by dopamine D<sub>1</sub> receptor agonist ABT 431 of dyskinesia similar to levodopa in patients with Parkinson disease. *Arch Neurol* 2001;58:249–254.



146. Bourne JA. SCH 23390: The first selective dopamine D<sub>1</sub>-like receptor antagonist. *CNS Drug Rev* 2001;7:399–414.
147. Peacock L, Gerlach J. Aberrant behavioral effects of a dopamine D<sub>1</sub> receptor antagonist and agonist in monkeys: Evidence of uncharted dopamine D<sub>1</sub> receptor actions. *Biol Psychiatry* 2001;50:501–509.
148. Rascol O, Blin O, Thalamas C, Descombes S, Soubrouillard C, Azulay P, Fabre N, Viallet F, Lafnizegger K, Wright S, Carter JH, Nutt JG. ABT-431, a D<sub>1</sub> receptor agonist prodrug, has efficacy in Parkinson's disease. *Ann Neurol* 1999;45:736–741.

---

**Jing Zhang** received his B.S. degree in Chemistry Engineering from Nanjing University of Science and Technology, China in 2006. Since then, she is a graduate student under the supervision of Professor Ao Zhang at the Shanghai Institute of Materia Medica, Chinese Academy of Sciences. Her research topics focus on the design and synthesis of novel benzazepines with activities at dopamine receptors (D<sub>1</sub>, D<sub>2</sub>) and/or serotonin receptors (5-HT<sub>1A</sub>, 5-HT<sub>2A</sub>).

**Bing Xiong** received his Ph.D. in medicinal chemistry from Shanghai Institute of Materia Medica in 2003. He then moved to McGill University in Canada for post-doctoral training in structure-based drug design both in *in-silico* and in *web-lab* parts. He returned to Shanghai Institute of Materia Medica in 2006 as an associate professor. Dr Xiong's research interests include: Informatics-aided drug design, drug-target interaction, off-target prediction and lead compound identification.

**Xuechu Zhen**, Professor of Neuropharmacology at Shanghai Institute of Materia Medica (SIMM), Chinese Academy of Sciences, was formerly associate professor of the Department of Physiology and Pharmacology at the City University of New York. He received the doctoral degree in 1996 at University of Geneva, and then moved to Medical College of Pennsylvania, and Pennsylvania State University Medical School for postdoctoral training. He was appointed as a faculty member in 1999 at Drexel University Medical School prior to moving to City University of New York as an associate professor. Since 2006, he has had the position of Principal Investigator and Professor in SIMM and has headed the Laboratory of Neurotransmitter and related CNS diseases. Dr. Zhen's research interests include: (1) neurotransmitter receptor functional regulation and signaling transduction; (2) dopamine system and related disorders such as Parkinson's disease, schizophrenia; (3) preclinical study of tetrahydropprotoberberine (THPBs) and derivative compounds in schizophrenia and drug abuse.

**Ao Zhang** was born in Sichuan of China in 1969. He received his B.S. degree in Chemistry in 1992 from Sichuan Normal College, now as Xihua Normal University. He was awarded a M. S. degree in Organic Chemistry in 1995 from Nankai University. After that he worked in Shanghai Institute of Organic Chemistry for two years and then was recommended to the predoctoral program of the same Institute in 1997 under the supervision of Professor Biao Jiang and obtained his Ph.D. in Organic Chemistry in 2000. From 2001 to 2002, he worked as a Postdoctoral Fellow in Professor Alan P Kozikowski's Drug Discovery Group at Georgetown University Medical Center. From 2002, he joined Professor John L. Neumeyer's Medicinal Chemistry Group as a Research Investigator at McLean Hospital, Harvard Medical School, where he was promoted to Instructor at Harvard Medical School in 2003 and was appointed to Assistant Director of the Medicinal Chemistry Program at McLean Hospital in 2004. From 2006, he was awarded a Hundred Talent Project from the Chinese Academy of Sciences, and worked as a full Professor and Principal Investigator at Shanghai Institute of Materia Medica where he is the Director of Synthetic Organic & Medicinal Chemistry Laboratory (SOMCL). Dr Zhang's research interests include: (1) Design and synthesis of novel small molecules targeted to G-protein-coupled receptors, especially dopamine and opioid receptors; (2) total synthesis of naturally bioactive products and structural modification; (3) development of new agents as structural and functional probes for the diagnosis and treatment of neurological and psychiatric disorders.