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SPECIAL REPORT

The Role of the Envelope Glycoprotein in the Depletion of T Helper Cells in Human Immunodeficiency Virus Infection*

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Infection with the human immunodeficiency virus (HIV) causes gradual depletion of CD4+ T helper lymphocytes and destruction of the lymphoid tissue, which ultimately leads to a fatal defect of the cellular immune system. Paramount to the understanding of the pathogenesis of HIV infection is to elucidate the mechanism which underlies the loss of T helper cells. Various ideas have been proposed

in order to explain this issue. Several hypotheses have focused on the role of the envelope glycoprotein in this process. This review summarizes the data obtained and concepts proposed regarding the involvement of the HIV glycoprotein in the pathology of CD4+ T cell depletion. (Pathology Oncology Research Vol 3, No 1, 62-67, 1997)

Key words: HIV, AIDS, envelope, T helper lymphocyte, pathogenesis, apoptosis

Introduction

Recent studies which demonstrate rapid turnover i. e. death and replacement of CD4+ T cells during human immunodeficiency virus (HIV) infection, support the concept that the gradual deterioration of the cellular immune functions in HIV infected individuals is the result of a physical loss of CD4+ T helper cells.¹⁻³ In addition; the observation that the proliferative response of mononuclear cells to mitogens and recall antigen is reduced in infected individuals suggests that functional defects of the T helper cell population develop during disease progression.^{4,5} The decline of the CD4+ T lymphocyte number in the peripheral blood and the CD4+ T lymphocyte malfunction is accompanied by continued destruction of the lymphoid tissue.^{6,7} Multiple hypotheses have been proposed in order to

explain the mechanisms underlying the alterations in the CD4+ T helper lymphocyte population during the course of HIV-1 infection. Most of these concepts were established from experimental in vitro investigations or are based on empirical data obtained by studies using primary mononuclear cells from infected individuals. However, it has not been clarified which of the ideas are relevant to the in vivo situation. Several of the hypotheses focus on the envelope glycoprotein and suggest that this molecule plays a particular role in the depletion of CD4+ T lymphocytes during the course of infection with HIV. This review summarizes and evaluates the data that were generated regarding the function of the glycoprotein in this process.

The HIV-1 envelope glycoprotein

The HIV-1 glycoprotein gp160 is composed of the two glycoprotein molecules gp41 and gp120. Upon budding of the virus from the infected cell, it is incorporated into the viral lipid membrane envelope. The gp41 molecule is anchored in the lipid bilayer and contains a cytoplasmic region, a transmembrane portion and an extracellular part which is non-covalently linked to the gp120 molecule. The gp120 glycoprotein is located completely outside the membrane and easily shed from the viral surface.^{8,9} This molecule contains several regions whose genetic sequence is relatively well preserved in different viral isolates, des-

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Abbreviations: HIV: Human Immunodeficiency Virus

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ignated C1-C4, and the regions V1-V5 which display substantial genetic variability. The cellular receptor for gp160 is the T cell receptor molecule CD4. Contact of gp120 with the CD4 molecule is mediated by a discontinuous CD4-binding domain located in the C2, V4, C4 and C5 regions.^{10,11} In addition, for entry of the virus into the cell, additional sequences possibly located in the V3 region have to interact with the coreceptor.^{12,13} These coreceptors are members of the chemokine receptor family such as CCR-5 and CXCR-4 (fusin/LESTR) and possibly CCR-2b and -3.¹⁴⁻²⁰ Interaction of gp120 with CD4 and the coreceptor may result in conformational changes of the glycoprotein which exposes the fusogenic peptide of gp41 for interaction with the plasma membrane of the host cell.²¹

Cell-cell fusion and the formation of syncytia

After virus infection and upon replication of HIV in the cell, gp160 is inserted in the cellular membrane before viral particles bud from the surface. Interaction of cells expressing the HIV envelope glycoprotein on the surface with cells expressing the CD4 molecule and members of the chemokine receptor family causes fusion of the cells.¹⁶ The fusion process requires the expression of the complete gp160 molecule in a fashion that allows intracellular cleavage to gp41 and gp120 and mutations in the cleavage site abolish the formation of syncytia.²² Cell-cell fusion can be extensive and result in the formation of large syncytia, a phenomenon which is readily observed in HIV-infected CD4+ T lymphoblast cultures.²³⁻²⁵ Because syncytia are not viable for an extended time period in vitro and the formation of syncytia coincides with the death of the infected cell culture, it was hypothesized that cell-cell fusion may underly T cell depletion in the infected individual.²³⁻²⁵ However, an important argument raised against this hypothesis is the experience that syncytia are rarely detectable in vivo, and lymphoid tissue does not exhibit massive syncytium formation.^{6,26-28}

Although this argument lead to the introduction of multiple additional hypotheses, several lines of evidence support the view that cell-cell fusion plays a role in vivo. For instance, cell-cell fusion products which contain dendritic cells possibly fused with T helper lymphocytes have been found in adenoidal lymphoid tissue.²⁹⁻³¹ Similarly, syncytia are readily detectable in the central nervous system where they are composed primarily of cells of the monocytic lineage including microglial cells and macrophages.^{32,33} The view that syncytia may occur in lymphoid tissue in vivo is further supported by an in vitro model system in which blocks of human tonsils were kept in long-term histoculture and small syncytia could be generated by implantation of glycoprotein-expressing cells.³⁴ In addition, the experience that syncytia are not easily detected in lymphoid tissue may in part be due to the fact that the lymphoid organs are densely packed with lymphocytes, a fact which makes it difficult to identify small syncytia by histologic analysis.³⁵

An additional point which has to be considered is that formation of large syncytia in cell culture is not observed with all viral isolates. HIV-1 strains have therefore been termed as either syncytium-inducing (SI) or non-syncytium-inducing (NSI) according to the ability to form syncytia and to cause cytopathology after infection of MT-2 cells.^{36,37} In addition, virus isolates of the SI phenotype are typically T cell-tropic and NSI virus strains are macrophage-tropic. The phenotypic difference is mirrored by genotypic differences between these groups in the V1-V2 and V3 region^{38,39} and the ability to form syncytia corresponds to the presence of a particular HIV coreceptor.^{17,18,40} It was previously suggested that a shift in the dominance from NSI to SI isolates in the peripheral blood of infected individuals correlates with the progression of disease; a concept which favors the assumption that SI viruses are more virulent than NSI isolates and implies that syncytium formation plays a role in T helper cell depletion.^{41,42} However, this may not always be the case, since the viral phenotype does not necessarily correlate with progression to AIDS.^{43,44} Moreover, it was demonstrated that isolates which have been classified as NSI strains in MT-2 cells may still cause fusion of a few cells without progression to large syncytia⁴⁵ or even result in overt syncytium formation and cytopathology in primary T cells⁴⁶ and other cell culture systems.⁴⁷

The notion that cytopathology associated with cell-cell fusion may constitute a relevant mechanism of T cell depletion in vivo is further supported by the observation that cell-cell fusion products may be more fragile than previously appreciated. For instance, contact of primary CD4+ T cells with envelope-expressing B lymphoblasts results in rapid lysis of a significant fraction of the cells in a few hours and the formation of relatively few and small syncytia. In contrast, contact of glycoprotein-expressing cells with transformed T cell lines like Jurkat and CEM induces the generation of large syncytia and no detectable cell lysis for more than 8 hours of incubation.⁴⁸ Additional data from our laboratory obtained by quantitative flow cytometric analysis demonstrate that coincubation of envelope glycoprotein-expressing cells with unstimulated primary PBMC causes selective disappearance of the majority of normal CD4+ T cells in a matter of hours by both syncytium formation and rapid cell death.⁴⁹

Accumulation of glycoprotein-CD4 complexes

Several viral proteins including the envelope glycoprotein have been implicated in mediating direct cytopathicity (reviewed in: 50). For instance, expression of gp160 (but not gp120) in CD4+ T lymphoblasts induces the death

of single cells by apoptosis.⁵¹⁻⁵³ Since the expression of gp160 causes retention of the CD4 molecule in the endoplasmic reticulum,⁵⁴ it was hypothesized that the cytopathicity observed is due to the accumulation of gp160-CD4 complexes at nuclear pores which may affect the transport of biomolecules to and from the nucleus.⁵⁵

Binding of gp120 to the CD4 molecule: induction of anergy and mediation of antibody-dependent cellular cytotoxicity

There is some evidence that gp120 shed from the virion surface and from infected cells is present in sera of HIV-infected individuals⁵⁶ and may bind to the CD4 molecule on T lymphocytes.⁵⁷ It was proposed that this interaction may interfere with normal antigen-specific activation of T cells simply by masking the CD4 antigen for interaction with MHC class II molecules.⁵⁸ This interaction may contribute to the functional defects of the cellular immune response which precedes the decline of CD4+ T-cells in HIV-infected individuals.⁵⁹ Moreover, it was demonstrated in vitro that glycoprotein-specific antibodies can crosslink CD4 molecules when they are coated with soluble gp120. This process reduces interleukin-2 production,^{60,61} inhibits proliferation of the T helper lymphocytes⁶² and renders the cells anergic,⁶³ possibly by initiating phosphorylation and activation of the tyrosin protein kinase p56^{lck}.⁶⁴ Upon subsequent activation of the cells through the T cell receptor these cells will be hyporesponsive⁶⁴ and undergo programmed cell death.⁶⁵⁻⁶⁷ Finally, by adhesion of the HIV glycoprotein to the CD4 molecule, uninfected cells will be recognized and lysed by natural and lymphokine-activated killer cells which present anti-gp120 antibodies on their surface bound to Fc receptors. This antibody-dependent cellular cytotoxicity has been shown by several groups to be effective in vitro.⁶⁸⁻⁷¹

Autoimmunity

In addition to the mechanisms described above several other characteristics have been attributed to the HIV envelope glycoprotein. These include the induction of autoimmunity based on structural similarities detected between the HIV-1 envelope glycoprotein and immunologically important molecules such as MHC class II antigens, the Fas/Apo-1 protein and functional domains of immunoglobulins^{72,73} and superantigen-like activation of particular V T cell subsets in vitro.⁷⁴ However, these observations have not been uniformly confirmed and are still a matter of dispute.^{75,76}

Apoptosis

Cells may die either by necrosis or by apoptosis. Necrotic cell death may be regarded as non-physiologic because it is usually the result of physical or chemical alterations in the

environment causing damage to the cellular metabolism or structure. This type of cell death causes in vivo secondary damage to neighbouring cells by enzymes and toxic products released by the dying cell and induces an inflammatory response in the affected tissue. In contrast, apoptotic cell death is the outcome of a process intrinsic to a particular cell during which the cell actively starts and executes its own death program upon signaling from outside or infection of the cell. In the course of apoptosis, the cell disintegrates into membrane-enveloped subcellular particles, so called apoptotic bodies, which contain cytoplasm, morphologically intact organelles and parts of the nucleus. Apoptotic bodies are subsequently taken up and digested by neighbouring cells including macrophages and epithelial cells. The apoptotic cell death does not cause an inflammatory reaction (reviewed in: 77,78). Similar to the induction of necrosis, multiple stimuli and events can give rise to apoptotic cell death. Therefore, the presence of apoptotic cells in a particular tissue does not provide any indication as to which mechanism underlies the cell death observed.

Several studies have demonstrated an increased incidence of apoptotic cell death in HIV infection.^{66,79} However, in HIV-infected individuals an increased incidence of apoptosis after in vitro stimulation has not only been observed with CD4+ T cells but also with CD8+ T cells when compared with cells from uninfected controls.^{80,81} A possible explanation for this observation is that in HIV infection both CD4+ and CD8+ T cells are activated. Stimulation of lymphocytes in the wake of a viral infection may cause increased levels of apoptotic lymphocyte death.⁸² Alternatively, apoptosis of CD4+ and CD8+ T lymphocytes are due to different processes.

Signs of apoptotic cell death were detected in various in vitro studies, most of which have been linked to the envelope glycoprotein. As outlined above, antigenic activation of helper cells, which were previously rendered anergic by crosslinking of CD4 molecules, results in lymphocyte apoptosis.⁶⁵ In addition, accumulation of gp160-CD4 complexes in infected cells may cause single cell apoptosis.⁵³ Alternatively, it was demonstrated that contact of HIV glycoprotein-expressing cells with CD4+ T cells causes the CD4+ T cells to rapidly die by apoptosis in cell culture.^{83,84} In this system, apoptotic cell death occurs upon contact of gp160 and the CD4 molecule⁸⁵⁻⁸⁹ and depends on the presentation of a cleavable gp160 molecule on the cellular surface.^{86,90} In addition to the CD4-binding region, the V3 loop seems to be critical because point mutations in this area and monoclonal Ab directed at this region which inhibit cell-cell fusion but not binding to the CD4 molecule abolish induction of apoptosis⁹¹ and rapid cell lysis.⁴⁸ Apoptotic cell death upon interaction of infected with uninfected cells can be detected in single cells and in cells fused in syncytia,^{86,88} both in primary CD4+ T lymphocytes^{85,89} and in T lymphoblast cell lines.^{83,84,88,89,92}

Conclusions

Although the hypotheses presented are not mutually exclusive, they cannot be unified to a single concept. However, several of the hypotheses are linked by a common prerequisite for induction of apoptosis or may describe related processes. For instance, gp120 shed by infected cells constitutes the basis of antibody-dependent cellular cytotoxicity, CD4 masking, crosslinking and induction of anergy with subsequent apoptosis. The exertion of such mechanisms requires some kind of immune response, either cellular or humoral. Alternatively, syncytium formation, rapid lysis and induction of apoptosis were described as a result of the interaction of gp160 on the cell surface of infected cells with uninfected CD4+ lymphocytes. Crucial to the latter concept is that HIV replicates *in vivo*. In these circumstances, the cytopathicity observed should correlate with the proportion of HIV-infected and virus-replicating cells. The fact that primary CD4+ T cells die in a matter of days after infection with HIV in cell culture indicates that virus infection *in vitro* is highly cytotoxic in the absence of any immune response.

Common to the two groups of hypotheses is that they deliver an explanation for the death of uninfected CD4+ T cells. This is in contrast to the idea that accumulation of gp 160-CD4 complexes causes cytotoxicity, a process which would account only for the death of HIV-infected cells. This concept has to compete with other mechanisms involved in killing of HIV-infected cells; like lysis by cytotoxic T lymphocytes.^{93,94}

However, several lines of evidence suggest that not only infected but also uninfected cells are destroyed during HIV infection. For instance, histopathologic studies point to the fact that loss of CD4+ T cells is not restricted to infected cells but uninfected CD4+ T cells are similarly affected *in vivo*.⁸¹

In addition, the number of HIV-infected cells is markedly lower than the number of CD4+ T cells dying and being replaced day by day.⁹⁵ Finally, recent measurements in HIV-infected individuals undergoing antiviral combination therapy and mathematical modelling suggest that the number of virions produced each day exceeds the number of CD4+ T cells lost by a factor of approximately 10.^{1,2,3,96} Since up to 99.99% of the virus particles produced are infection-incompetent⁹⁷ the ratio of virions produced and cells destroyed each day may be too low to account for lysis of only infected cells in the course of HIV disease.

In conclusion, although the cause of CD4+ T cell depletion in HIV-1 infection might be multifactorial and is still elusive, several lines of evidence indicate that the envelope glycoprotein may contribute to this process. However, the ideas raised regarding the role of the gly-

coprotein are diverse and can only partially be harmonized into common concepts for the understanding of the role of this molecule in the depletion of T helper cells.

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References

1. Ho DD, Neumann AU, Perelson AS, et al: Rapid turnover of plasma virions and CD4 lymphocytes in HIV-1 infection. *Nature* 373:123-126, 1995.
2. Wei X, Ghosh SK, Taylor ME, et al.: Viral dynamics in human immunodeficiency virus type 1 infection. *Nature* 373:117-122, 1995.
3. Perelson AS, Neumann AU, Markowitz M, et al: HIV-1 dynamics in vivo: virion clearance rate, infected cell life-span, and viral generation time. *Science* 271:1582-1586, 1996.
4. Clerici M, Stocks NI, Zajac RA, et al: Detection of three distinct patterns of T helper cell dysfunction in asymptomatic human immunodeficiency virus-positive patients. *J Clin Invest* 93:768-775, 1989.
5. Shearer GM and Clerici M: Early T helper cell defects in HIV infection. *Aas* 5:245-253, 1991.
6. Racz P: Spectrum of morphologic changes of lymph nodes from patients with Aas or AIDS related complexes. *Progr Allergy* 37:81-181, 1985.
7. Fauci AS: Multifactorial nature of human immunodeficiency virus disease: Implications for therapy. *Science* 262:1011-1018, 1993.
8. Gelderblom HR, Reupke H and Pauli G: Loss of envelope antigens of HTLV-III/LAV, a factor in AIDS pathogenesis. *Lancet* 2:1016-1017, 1985.
9. McKeating JA, McKnight A and Moore JP: Differential loss of envelope glycoprotein from virions of human immunodeficiency virus type 1 isolates: effect on infectivity and neutralization. *J Virol* 65:852-860, 1991.
10. Olshevsky K, Helseth E, Furman C, et al: Identification of individual human immunodeficiency virus type 1 gp 120 amino acids important for CD4 binding. *J Virol* 64:5701-5707, 1990.
11. Pollard S, Rosa M, Rosa J and Wiley D: Truncated variants of gp 120 bind CD4 with high affinity and suggest a minimum CD4 binding region. *EMBO J* 11:585-591, 1992.
12. Trkola A, Dragic T, Arthos J, et al.: CD4-dependent, antibody-sensitive interactions between HIV-1 and its coreceptor CCR-5. *Nature* 384:184-187, 1996.
13. Wu L, Gerardi NP, Wyatt R, et al.: CD4-induced interaction of primary HIV-1 gp 120 glycoproteins with the chemokine receptor CCR-5. *Nature* 384:179-183, 1996.
14. Alkhatib G, Combadiere C, Broder CC, et al: CCR5: A RANTES, MIP-1, MIP-1 receptor as a fusion cofactor for macrophage-tropic HIV-1. *Science* 272:1955-1958, 1996.
15. Berson F, Long D, Doranz BJ, et al: A seven transmembrane domain receptor involved in fusion and entry of T-cell tropic human immunodeficiency virus type 1 strains. *J Virol* 70:1996.
16. Choe H, Farzan M, Sun Y, et al.: The j3-chemokine receptors CCR3 and CCR5 facilitate infection by primary HIV-1 isolates. *Cell* 85:1135-1148, 1996.
17. Cocchi F, DeVico AL, Garzino-Demo A, et al: The V3 domain of the HIV-1 gp120 envelope glycoprotein is critical for chemokine-mediated blockade of infection. *Nature Med* 2:1244-1247, 1996.
18. Doranz BJ, Rucker J, Yi Y, et al.: A dual tropic primary HIV-1 isolate that uses fusion and the -chemokine receptors CKR-5, CKR-3, and CKR-2b as fusion cofactors. *Cell* 85:1149-1158, 1996.

19. Dragic T, Litwin V, Allaway GP, et al.: HIV-1 entry into CD4+ T cells is mediated by the chemokine receptor CC-CKR-5. *Nature* 381:667-673, 1996.
20. Feng Y, Broder CC, Kennedy PE and Berger EA: HIV-1 entry cofactor: Functional cDNA cloning of a seven-transmembrane G protein-coupled receptor. *Science* 272:872-877, 1996.
21. Sattentau QJ, Moore JP, Vignaux F, et al: Conformational changes induced in the envelope glycoproteins of the human and simian immunodeficiency viruses by soluble receptor binding. *J Virol* 67:7383-93, 1993.
22. McCune JM, Rabin LB, Feinberg MB, et al: Endoproteolytic cleavage of gp 160 is required for the activation of the human immunodeficiency virus. *Cell* 53:55-67, 1988.
23. Lifson JD, Feinberg MB, Reyes GR, et al.: Induction of CD4-dependent cell fusion by the HTLV-III/LAV envelope glycoprotein. *Nature* 323:725-728, 1986.
24. Lifson JD, Reyes GIZ, McGrath MC, et al: AIDS retrovirus induced cytopathology: giant cell formation and involvement of CD4 antigen. *Science* 232:1123-1127, 1986.
25. Sodroski J, Goh WC, Rosen C, et al: Role of HTLV-III/LAV envelope in syncytium formation and cytopathicity. *Nature* 322:470-474, 1986.
26. Harper M, Marselle LM, Gallo RC and Wong-Staal F: Detection of lymphocytes expressing human T-lymphotropic virus type III in lymph nodes and peripheral blood from infected individuals by in situ hybridization. *Proc Natl Acad Sci* 83:772-776, 1986.
27. Tenner-Racz K, Racz P, Bofill M, et al.: HTLVII/LAV antigens in lymph nodes of homosexual men with persistent generalized lymphadenopathy and AaS. *Am J Pathol* 123:9-15, 1986.
28. Baroni CD, Pezzella F, Pezzella M, et al: Expression of HIV in lymph node cells of LAS patients. *Am J Pathol* 133:498-506, 1988.
29. Rinfret A, Latendresse H, Lefebvre R, et al: Human immunodeficiency virus-infected multinucleated histiocytes in oropharyngeal lymphoid tissue from two asymptomatic patients. *Am J Pathol* 138:421-426, 1991.
30. Kaaya E, Li SL, Feichtinger H, et al.: Accessory cells and macrophages in the histopathology of SIVsm-infected cynomolgus monkeys. *Res Virol* 144:81-92, 1993.
31. Frankel SS, Wenig BM, Burke AP, et al.: Replication of HIV-1 in dendritic cell-derived syncytia at the mucosal surface of the adenoid. *Science* 272:115-117, 1996.
32. Navia BA, Cho E-S, Petit CK and Price RW: The AIDS dementia complex. II. Neuropathology. *Ann Neurol* 19: 525-535, 1986.
33. Gulevich SJ and Wiley CA: HIV infection and the brain. *AIDS* 5:S49-S54, 1991.
34. Margolis LB, Glushakova S, Baibakov B and Zimmerberg J: Syncytium formation in cultrated human lymphoid tissue: fusion of implanted HIV glycoprotein 120/41-expressing cells with native CD4+ cells. *AIDS Res Hum Retroviruses* 11:697-704, 1995.
35. Soll DR and Kennedy RC: The role of T cell motility and cytoskeletal reorganization in HIV-induced syncytium formation. *AIDS Res Hum Retroviruses* 10:325-327, 1994.
36. Cheng-Mayer C, Seto D, Tateno M and Levy JA: Biologic features of HIV that correlate with virulence in the host. *Science* 240:80-82, 1988.
37. Tersmette M, Goede REY de, Bert JM, et al.: Differential syncytium inducing capacity of human immunodeficiency virus isolates: frequent detection of syncytium-inducing isolates in patients with acquired immunodeficiency syndrome (AaS) and AIDS-related complex. *J Virol* 62:2026-2032, 1988.
38. Fenyő EM, Morfeldt-Manson L, Chiodi F, et al.: Distinct replicative and cytopathic characteristics of human immunodeficiency virus isolates. *J Virol* 62:4414-4419, 1988.
39. Fouchier RAM, Groenink M, Koostra NA, et al: Phenotype-associated sequence variation in the third variable domain of the human immunodeficiency virus type 1 gp 120 molecule. *J Virol* 66:3183-3187, 1992.
40. Zhang L, Huang Y, He T, Cao Y and Ho DD: HIV-1 subtype and second-receptor use. *Nature* 383:768, 1996.
41. Schuitemaker H, Koot M, Koostra NA, et al.: Biological phenotype of human immunodeficiency virus type 1 clones at different stages of infection: progression of disease is associated with a shift from monocytopathic to T-cell-tropic virus population. *J Virol* 66:1354-1360, 1992.
42. Nielsen C, Pedersen C, Lundgren JD and Gerstoft J: Biological properties of HIV isolates in primary HIV infection: consequences for the subsequent course of infection. *AIDS* 7:1035-1040, 1993.
43. Karlsson A, Parsmyr K, Sandstrom E, et al: MT-2 cell tropism as prognostic marker for disease progression in human immunodeficiency virus type 1 infection. *J Clin Microbiol* 32:364-370, 1994.
44. Richman DD and Bozzette SA: The impact of the syncytium-inducing phenotype of human immunodeficiency virus on disease progression. *J Infect Dis* 169:1994.
45. Weiss CD, Barnett SW, Cacalano N, et al: Studies of HIV-1 envelope glycoprotein-mediated fusion using a simple fluorescence assay. *AIDS* 10:241-246, 1996.
46. Todd BJ, Kedar P and Pope JH: Syncytium-induction in primary CD4+ T-cell lines from normal donors by human immunodeficiency virus type 1 isolates with non-syncytium-inducing genotype and phenotype in MT-2 cells. *J Virol* 69:7099-7105, 1995.
47. Pope M, Betjes MGH, Romani N, et al.: Conjugates of dendritic cells and memory T lymphocytes from skin facilitate productive infection with HIV-1. *Cell* 78: 389-398, 1994.
48. Heinkelein M, Sopfer S and Jassoy C: Contact of human immunodeficiency virus type 1-infected and uninfected CD4+ T lymphocytes is highly cytolytic for both cells. *J Virol* 69:6925-6931, 1995.
49. Jassoy C, Muller M, Kutsch O, et al: Quantitative analysis of the loss of uninfected CD4 T cells upon contact with HIV-infected cells. *Convergence of AIDS and Cancer Research*, Budapest, August 25-28, 1996; (Abstr. p.
50. Levy JA: HIV and the pathogenesis of AIDS. ASM Press, Washington, 1994.
51. Koga Y, Sasaki M, Yoshida H, et al: Cytopathic effect determined by the amount of CD4 molecules in human cell lines expressing gp 160 of human immunodeficiency virus. *J Immunol* 144:94-102, 1990.
52. Koga Y, Nakamura K, Sasaki M, et al: The difference in gp 160 and gp 120 of HIV type 1 in induction of CD4 downregulation precedes single-cell killing. *Virology* 201:137-141, 1994.
53. Lu YY, Koga Y, Tanaka M, Sasaki G, et al: Apoptosis induced in CD4+ cells expressing gp 160 of human immunodeficiency virus type 1. *J Virol* 68:390-399, 1994.
54. Crise B, Bounocore L and Rose JK: CD4 is retained in the endoplasmic reticulum by human immunodeficiency virus type 1 precursor. *J Virol* 64:5585-5593, 1990.
55. Koga Y, Sasaki M, Yoshida H, et al: Disturbance of nuclear transport of proteins in CD4+ cells expressing gp 160 of human immunodeficiency virus. *J Virol* 65:5609-5612, 1991.
56. Oh SK, Cruikshank WW, Raina J, et al: Identification of HIV-1 envelope glycoproteins in the serum of AIDS and ARC patients. *J Acquir Immune Def Syndr* 5:251-256, 1992.
57. Daniel V, Susal C, Prodeus AP, et al: CD4+ lymphocyte depletion in HIV-infected patients is associated with gp 120-immunoglobulin

- lin-complement attachment to CD4+ cells. *Vox Sang* 64:31-36, 1993.
58. *Rosenstein Y, Burakoff SJ and Herrmann SH*: HIV gp 120 can block CD4-class II MHC-mediated adhesion. *J Immunol* 144:526-531, 1994.
 59. *Shearer GM, Bernstein DC, Tung KSK, et al*: A model for the selective loss of major histocompatibility complex self restricted T-cell immune responses during the development of acquired immune deficiency syndrome (AIDS). *J Immunol* 137:2514-2521, 1986.
 60. *Diamond DC, Sleckman BP, Gregory T, et al*: Inhibition of CD4+ T cell function by the HIV envelope protein gp120. *J Immunol* 141:3715-3721, 1988.
 61. *Oyaizu N, Chirmule N, Kalyanaraman VS, et al*: Human immunodeficiency virus type 1 envelope glycoprotein gp 120 produces immune defects in CD4+ T lymphocytes by inhibiting interleukin 2 mRNA. *Proc Natl Acad Sci* 87:2379-2382, 1990.
 62. *Mann DL, Lasane F, Popovic M, et al*: HTLV III large envelope protein (gp 120) suppresses PHA-induced lymphocyte pathogenesis. *J Immunol* 138:2640-2648, 1987.
 63. *Mittler RS and Hoffmann MK*: Synergism between HIV gp 120 and gp 120-specific antibody in blocking human T cell activation. *Science* 245:1380-1382, 1989.
 64. *Goldman F, Jensen WA, Johnson GL, et al*: gp 120 ligation of CD4 induces p56lck activation and TCR desensitization independent of TCR tyrosine phosphorylation. *J Immunol* 153:2905-2917, 1994.
 65. *Banda NK, Bernier J, Kurahara DK, et al*: Crosslinking CD4 by human immunodeficiency virus gp 120 primes T cells for activation-induced apoptosis. *J Exp Med* 176:1099-1106, 1992.
 66. *Groux H, Torpier G, Monte D, et al*: Activation-induced death by apoptosis in CD4+ T cells from human immunodeficiency virus-infected asymptomatic individuals. *J Exp Med* 175:331-340, 1992.
 67. *Finkel TH and Banda NK*: Indirect mechanisms of HIV pathogenesis: how does HIV kill T cells? *Curr Opin Immunol* 6:605-615, 1994.
 68. *Blumberg RS, Paradis T, Hartshorn KL, et al*: Antibody-dependent cell-mediated cytotoxicity against cells infected with the human immunodeficiency virus. *J Infect Dis* 156:878-884, 1987.
 69. *Ljunggren K, Bottiger B, Biberfeld G, et al*: Antibody-dependent cellular cytotoxicity-inducing antibodies against human immunodeficiency virus. *J Immunol* 139:2263-2267, 1987.
 70. *Lyerly HK, Matthews TJ, Langlois AJ, et al*: Human T-cell lymphotropic virus IIIb glycoprotein (gp 120) bound to CD4 determinants on normal lymphocytes and expressed by infected cells serves as target for immune attack. *Proc Natl Acad Sci* 84:4601-4605, 1987.
 71. *Rook AH, Lane HC, Folks T, et al*: Sera from HTLV-III/LAV antibody-positive individuals mediate antibody-dependent cellular cytotoxicity against HTLVIII/LAV-infected cells. *J Immunol* 138:1064-1067, 1987.
 72. *Clerici M, Shearer G, Hounsell EF, et al*: Alloactivated cytotoxic T cells recognize the carboxy-terminal domain of human immunodeficiency virus-1 gp 120 envelope glycoprotein. *Eur J Immunol* 23:2022-2025, 1993.
 73. *Silvestris F, Williams RC and Dammaco F*: Autoreactivity in HIV-1 infection: the role of molecular mimicry. *Clin Immunol Immunopathol* 75:197-205, 1995.
 74. *Akolkar PN, Chirmule N, Gulwani-Akolkar B, et al*: V beta-specific activation of T cells by the HIV glycoprotein gp 160. *Scand J Immunol* 41:487-98, 1995.
 75. *Boldt-Houle DM, Rinaldo CR and Ehrlich GD*: Random depletion of T cells that bear specific T cell receptor V beta sequences in AIDS patients. *J Leukoc Biol* 54: 486-491, 1993.
 76. *Boyer V, Smith LR, Erre F, et al*: T cell receptor V beta repertoire in HIV infected individuals: lack of evidence for selective V beta depletion. *Clin Exp Immunol* 92:437-441, 1993.
 77. *Wyllie AH, Kerr JFR and Currie AR*: Cell death: the significance of apoptosis. *Int Rev Cytol* 68:251-306, 1980.
 78. *Kerr JFR, Winterford CM and Harmon BV*: Morphologic criteria for identifying apoptosis. In: *Morphologic criteria for identifying apoptosis*. (Eds: J. Celis). Academic Press, 1994, pp. 319-329.
 79. *Meyaard L, Otto SA, Jonker RR, et al*: Programmed death of T cells in HIV-1 infection. *Science* 257:217-219, 1992.
 80. *Gougeon M L, Garcia S, Heeney J, et al*: Programmed cell death in AIDS-related HIV and SIV infections. *AIDS Res Human Retrov* 9:553-563, 1993.
 81. *Finkel TH, Tudor-Williams G, Banda NK, et al*: Apoptosis occurs predominantly on bystander cells and not in productively infected cells of HIV- and SIV-infected lymph nodes. *Nature Med* 1:129-134, 1995.
 82. *Meyaard L and Miedema F*: Programmed death of T cells in HIV infection: Result of immune activation? *Curr Top Microbiol Immunol* 200:213-221, 1995.
 83. *Laurent-Crawford AG, Krust B, Muller S, et al*: The cytopathic effect of HIV is associated with apoptosis. *Virology* 185:829-839, 1991.
 84. *Teraï C, Kornbluth RS, Pauza CD, et al*: Apoptosis as a mechanism of cell death in cultured T lymphoblasts acutely infected with HIV-1. *J Clin Invest* 87:1710-1715, 1991.
 85. *Heinkelein M, Jassoy C*: Unpublished data.
 86. *Laurent-Crawford AG, Krust B, Riviere Y, et al*: Membrane expression of HIV envelope glycoproteins triggers apoptosis in CD4 cells: *AIDS Res Hum Retrov* 9:761-773, 1993.
 87. *Corbeil J and Richman DD*: Productive infection and subsequent interaction of CD4-gp 120 at the cellular membrane is required for HIV-induced apoptosis of CD4+ T cells. *J Gen Virol* 76:681690, 1995.
 88. *Maldarelli F, Sato H, Berthold E, et al*: Rapid induction of apoptosis by cell-to-cell transmission of human immunodeficiency virus type 1. *J Virol* 69:6457-6465, 1995.
 89. *Nardelli B, Gonzales CJ, Schechter M and Valentine FT*: CD4+ blood lymphocytes are rapidly killed in vitro by contact with autologous human immunodeficiency virus-infected cells. *Proc Natl Acad Sci* 92:7312-7316, 1995.
 90. *Kruger U, Pfeiffer T and Bosch V*: Generation of lymphocyte cell lines coexpressing CD4 and wild-type or mutant HIV type 1 glycoproteins: Implications for HIV type 1 env-induced cell lysis. *AIDS Res Hum Retrov* 12:783-792, 1996.
 91. *Laurent-Crawford AG, Coccia E, Krust B and Hovanessian AG*: Membrane-expressed HIV envelope glycoprotein heterodimer is a powerful inducer of cell death in uninfected CD4+ target cells. *Res Virol* 146:5-17, 1995.
 92. *Martin SJ, Matear PM and Vyakarnam A*: HIV-1 infection of human CD4+ T cells in vitro. *J Immunol* 152:330-342, 1994.
 93. *Zinkernagel RM and Hengartner H*: T-cell mediated immunopathology versus direct cytolysis by virus: implications for HIV and AIDS. *Immunol Today* 15:262-268, 1994.
 94. *Jassoy C and Walker BD*: HIV-specific cytotoxic T lymphocytes and the control of HIV-1 replication. *Springer Sem Immunopathol* 18:in press, 1996.
 95. *Haase AT, Henry K, Zupancie M, et al*: Quantitative image analysis of HIV-1 infection in lymphoid tissue. *Science* 274:985-989, 1996.
 96. *Levy JA, Ramachandran B, Barker E, et al*: Plasma viral load, CD4+ cell counts, and HIV-1 production by cells. *Science* 271:670-671, 1996.
 97. *Piatk M, Saag MS, Yang LC, et al*: High levels of HIV-1 in plasma during all stages of infection determined by competitive PCR. *Science* 259:1749-1754, 1993.