

## REVIEW ARTICLE

# Chemistry and Pharmacology of The Citrus Bioflavonoid Hesperidin

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**Hesperidin, a bioflavonoid, is an abundant and inexpensive by-product of Citrus cultivation. A deficiency of this substance in the diet has been linked with abnormal capillary leakiness as well as pain in the extremities causing aches, weakness and night leg cramps. No signs of toxicity have been observed with the normal intake of hesperidin or related compounds.**

**Both hesperidin and its aglycone hesperetin have been reported to possess a wide range of pharmacological properties. This paper reviews various aspects of hesperidin and its related compounds, including their occurrence, physical and chemical properties, analysis, pharmacokinetics, safety and toxicity and the marketed products available. A special emphasis has been laid on the pharmacological properties and medicinal uses of these compounds. Copyright © 2001 John Wiley & Sons, Ltd.**

*Keywords:* hesperidin; hesperetin; bioflavonoid; pharmacology; analysis.

## INTRODUCTION

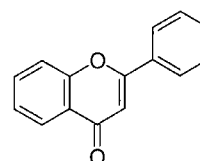
Flavonoids comprise a large group of naturally occurring, low molecular weight, polyphenolic compounds widely distributed in the plant kingdom as secondary metabolites. They represent one of the most important and interesting classes of biologically active compounds and occur both in the free state and as glycosides. They are based on the parent compound, flavone (2-phenyl chromone or 2-phenyl benzopyrone), characterized by a C<sub>6</sub>-C<sub>3</sub>-C<sub>6</sub> carbon skeleton where the C<sub>6</sub> components are aromatic rings (Fig. 1). Flavonoids occur in practically all parts of plants including fruit, vegetables, nuts, seeds, leaves, flowers and bark (Middleton, 1984).

Dr Albert Szent-Györgi, a famed Hungarian researcher, found that Citrus peel flavonoids were effective in preventing capillary bleeding and was the first to report the biological activity of flavonoids on capillary fragility associated with scurvy (Armentano *et al.*, 1936; Szent-Györgi, 1938). The broad spectrum of biological activities within the group and the multiplicity of actions displayed by certain individual members make the flavonoids one of the most intriguing classes of biologically active compounds, and thus these are often termed 'bioflavonoids'.

Hesperidin is an abundant and inexpensive by-product of Citrus cultivation and is the major flavonoid in sweet orange and lemon. In young immature oranges it can account for up to 14% of the fresh weight of the fruit (Barthe *et al.*, 1988). It is usually found in association with vitamin C. Some symptoms originally thought to be

due to vitamin C deficiency such as bruising due to capillary fragility were found in early studies to be relieved by crude vitamin C extract but not by purified vitamin C. The bioflavonoids, formerly called 'vitamin P', were found to be the essential components in correcting this bruising tendency and improving the permeability and integrity of the capillary lining. These bioflavonoids include hesperidin, citrin, rutin, flavones, flavonols, catechin and quercetin.

Of historical importance is the observation that 'citrin', a mixture of two flavonoids, eriodictyol and hesperidin, was considered to possess a vitamin-like activity, as early as in 1949 (Scarborough and Bacharach, 1949). This material was later termed 'vitamin P' to indicate that it could decrease capillary permeability and fragility, prolong the life of marginally scorbutic guinea-pigs and reduce the signs of hypovitaminosis C. Although the term vitamin P was subsequently abandoned, there was a strong indication that the material had potent antioxidant-dependent vitamin C sparing activity (Clemetson, 1989). Hesperidin deficiency has since been linked with abnormal capillary leakiness as well as pain in the extremities causing aches, weakness and night leg cramps. Supplemental hesperidin also helps in reducing oedema or excess swelling in the legs due to fluid accumulation. As with other bioflavonoids, hesperidin works best when administered concomitantly with



**Figure 1.** The flavone nucleus (2-phenylbenzopyrone)

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vitamin C. No signs of toxicity have been observed with normal intake of hesperidin.

Hesperidin was first discovered in 1827, by Lebreton, but not in a pure state and has been under continuous investigation since then (Fluckiger and Hanbury, 1986). Though it has been found to possess a wide range of pharmacological properties, no comprehensive report is available on this compound. The following provides a comprehensive review of this particular bioflavonoid.

## OCCURRENCE

Hesperidin is isolated in large amounts from the discarded rinds of the ordinary orange *Citrus aurantium* L. (Kanes *et al.*, 1993; Emim *et al.*, 1994), *C. sinensis* (Horowitz and Gentili, 1963), *C. unshiu* (Kawaguchi *et al.*, 1997) and other species of the genus *Citrus* (family Rutaceae). It has been reported to occur in many plants other than *Citrus*, such as in genera Fabaceae (Bhalla and Dakwake, 1978), Betulaceae (Pawlowska, 1980), Lamia-ceae (Kokkalou and Kapetanidis, 1988) and Papilionaceae. Its presence in the bark of *Zanthoxylum avicennae* and *Z. cuspidatum* (family Rutaceae) has been reported (Arthur *et al.*, 1956). These plants are indigenous to Hong Kong. The presence of hesperetin-7- $\beta$ -neohesperidoside (neohesperidin) was recently reported in *C. humilis* and *C. cornigera*, two *Cynara* species growing in Greece (Chinou and Harvala, 1997). Hesperidin has also been isolated from the roots of *Acanthopanax setchuenensis* (family Araliaceae), collected in Sichuan Province, China (Zhao *et al.*, 1999).

Hesperidin occurs in greatest concentration in green fruit and its concentration in the fruit increases during storage (Higby, 1941). Its distribution in the epicarp, mesocarp, endocarp and juice of *Citrus* fruits has been reported (Kawaguchi *et al.*, 1997). The distribution and concentration of hesperidin within the different tissues of mature fruit of *C. sinensis* has been measured using a radioimmunoassay method and it was found to present in high levels in the albedo, membranes and the pith whereas the concentration was much lower in the juice vesicles and seeds. In seeds, the hesperidin content increases after germination suggesting that there is a net production of this compound in the developing seedling, which is partly stimulated by light (Barthe *et al.*, 1988).

Hesperidin occurs in crystalline, feather-like aggregates or sphaerocrystalline masses in the cells (Evans, 1996). Di Mauro *et al.* (1999) have recently described a novel procedure for obtaining hesperidin from waste orange peel of the *Citrus* industry based on the adsorption of dilute extracts of hesperidin on a styrene-divinylbenzene resin.

## PROPERTIES

Pure hesperidin occurs as long hair-like needles, tan or pale yellow in colour. Its melting point ranges from 258° to 262°C (softens at 250°C). It has a molecular formula  $C_{18}H_{34}O_{15}$  and a molecular weight of 610.57 daltons. It is easily soluble in dilute alkali and in pyridine giving a clear yellow solution, slightly soluble in methanol, hot glacial acetic acid and almost insoluble in acetone,

benzene and chloroform. The solubility in water is 1 in 50 (Budavari, 1996). It has a property of forming complex crystals with other similar glucosides, which greatly affects its solubility and other physical properties, making it difficult to obtain in a pure state (Higby, 1941). It can, however, be purified by washing with hot water and extraction with 95% methyl alcohol, followed by crystallization (King and Robertson, 1931). It is tasteless and odourless (Kometani *et al.*, 1996).

## CHEMISTRY

Hesperidin (Fig. 2) is a flavanone glycoside, comprising of an aglycone, hesperetin (Fig. 3) or methyl eriodictyol (Evans, 1996) and an attached disaccharide, rutinose. Hesperidin is, therefore, a  $\beta$ -7-rutinoside of hesperetin (Preston *et al.*, 1953). The disaccharide unit ( $C_{12}H_{22}O_{10}$ ) is composed of one molecule of rhamnose and one of glucose and may assume one of the two isomeric forms, rutinose or neohesperidose. Rutinose (Fig. 4) is chemically 6-O-(6-deoxy- $\alpha$ -L-mannopyranosyl)-D-glucose or O- $\alpha$ -L-rhamnosyl-(1  $\rightarrow$  6)glucose. Neohesperidose is chemically O- $\alpha$ -L-rhamnosyl-(1  $\rightarrow$  2)glucose, differing only in the configuration of the two sugar units (Harborne, 1994). The position of the sugars in hesperidin was determined by partial hydrolysis with dilute acid leading to the formation of L-rhamnose and hesperetin-7- $\beta$ -D-glucoside which could be cleaved by the enzyme  $\beta$ -D-glucosidase (Fox *et al.*, 1953). Hence, in hesperidin, glucose is attached to hesperetin and rhamnose is attached to the glucose. Hesperetin ( $C_{16}H_{14}O_6$ ) chemically is 3', 5, 7-trihydroxy-4'-methoxy flavanone. Hesperidin is thus 3', 5, 7-trihydroxy-4'-methoxyflavanone-7-(6- $\alpha$ -L-rhamnopyranosyl)- $\beta$ -D-glucopyranoside or -7-rutinoside (Calomme *et al.*, 1996).

Hesperidin, upon alkaline hydrolysis yields phloroglucinol and hesperetic acid (Higby, 1941). Upon acid

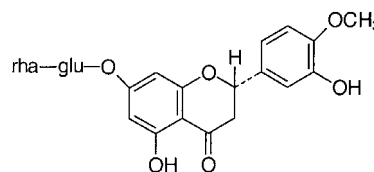


Figure 2. Hesperidin (hesperetin-7-rhamnoglucoside)

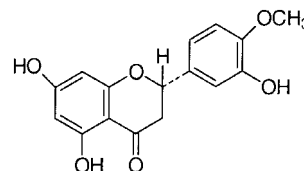


Figure 3. Hesperetin (3, 5, 7-trihydroxy-4-methoxyflavanone)

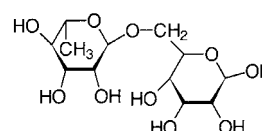


Figure 4. Rutinose [O- $\alpha$ -L-rhamnosyl-(1  $\rightarrow$  6)glucose]

hydrolysis, hesperidin produces one mole each of the aglycone hesperetin, D-glucose and L-rhamnose (Asahina *et al.*, 1930). Hydrolysis with either dilute sulphuric acid or sulphuric acid in ethylene glycol yields an optically active mixture of (±)- and (–)-hesperetin, which can be separated by fractional recrystallization. Both have been characterized by conversion into derivatives (Arthur *et al.*, 1956). Under carefully controlled conditions of acid hydrolysis, hesperidin yields the optically active laevo-rotatory aglycone, (–)-hesperetin, and this on ozonization, gives L- malic acid, indicating that (–)-hesperetin has the 2S-configuration (Arakawa and Nakazaki, 1960).

There is a known relation between the structure of the disaccharide and the presence or absence of bitterness in the compound (Horowitz and Gentili, 1963). Rutinosides are tasteless, whereas neohesperidosides are intensely bitter. Hesperidin itself is a flavonoid rutinoside and is thereby non-bitter (Kometani *et al.*, 1996). The bitter neohesperidosides mainly accumulate in grapefruit whereas the non-bitter rutinosides predominate in orange and lemon (Horowitz, 1961).

## IDENTIFICATION AND ANALYSIS

Both hesperidin and hesperetin show a characteristic flavanone absorption spectrum with UV maxima at 286 and 289 nm, respectively and an inflection of low intensity at 330 nm (Jurd, 1962).

The  $^1\text{H}$  NMR spectra of TMS ether of both hesperidin and hesperetin in  $\text{CCl}_4$  have been published (Mabry *et al.*, 1970). The  $^1\text{H}$  NMR spectra of hesperidin in  $\text{DMSO-d}_6$  solution has also been studied and the values for chemical shifts and coupling constants presented (Nieto and Gutierrez, 1986). Aged samples of hesperidin in  $\text{DMSO}$  show NMR lines from most protons split into two sets of signals, the second being assigned to an isomer of hesperidin with a change of the natural configuration at the  $\text{C}_2$  atom. A  $^{13}\text{C}$  NMR spectrum of hesperidin has been published (Agarwal, 1989). Markham and Ternai (1976) have described the  $^{13}\text{C}$  NMR of hesperidin and compared it with those of other major flavonoid groups including glycosides, relating to the one basic solvent system,  $\text{DMSO-d}_6$ .

A method for the *fluorimetric determination* of hesperidin has been developed, based on the reaction between rutin and dimethylformamide to form a fluorescent chelate. This chelate shows two excitation maxima at 295 and 370 nm and one emission maximum at 460 nm, indicating a 1:1 molar ratio of ligand to metal ion (Kaito *et al.*, 1979). Manual and flow injection methods are also used along with the spectrofluorimetry (Perez-Ruiz *et al.*, 1999).

Other methods have been developed for the *spectrophotometric determination* of hesperidin by forming its complexes with certain metals. Copper (II)–hesperidin complex (Kuntic *et al.*, 1999), uranyl (II)–hesperidin complex (Kuntic *et al.*, 1998), zirconium (IV)–hesperidin complex (Radovic *et al.*, 1996) and aluminium (III)–hesperidin complexes (Malesev *et al.*, 1997) have been formed in water–methanol systems and the determination of the compound is found to be precise (Radovic *et al.*, 1996).

*Radioimmunoassay (RIA) methods*, which combine sensitivity, specificity, high sample capacity, speed and

relative simplicity, are finding wide application for the quantification of plant secondary compounds. A RIA method has been developed for hesperidin utilizing antibodies raised against a hesperidin-4-O-carboxymethyl-oxime hapten and a tritiated radiotracer prepared by direct reduction of hesperidin with  $\text{NaB}[^3\text{H}]_4$ . The method possesses a detection limit and measuring range at nanogram levels, a low coefficient of variation and a good correlation with HPLC (Barthe *et al.*, 1988).

*Capillary electrophoresis* is another method that has been used to simultaneously analyse hesperidin and neohesperidin, along with a range of other compounds. Separation could be easily achieved and the compounds could be subsequently quantified. The method allows rapid monitoring with great specificity (Cancalon, 1999).

A large number of scientists have worked on the development of *chromatographic methods* for the estimation of hesperidin and its analogues. Höerhammer and Wagner (1962), separated and purified Citrus flavonoids by preparative TLC (silica gel) using butanol, acetic acid and water (4:1:5) as the solvent system. The separation of hesperidin, hesperetin, eriodictin, naringin and naringenin was successfully accomplished. Various scientists have worked on the separation and quantification of hesperidin and/or hesperetin from a mixture of a number of flavonoids, using HPLC. Most of these utilize a reversed phase C-18 column and an aqueous mobile phase consisting of water, acetonitrile, methanol and tetrahydrofuran coupled with minor amounts of acids (Fisher, 1978; Castele *et al.*, 1982; Rouseff, 1988; Rouseff *et al.*, 1992; Mouly *et al.*, 1993; Wang *et al.*, 1994; Mouly *et al.*, 1998; Saija *et al.*, 1998; Ross *et al.*, 2000). The detection is accomplished using a UV detector set at 280 nm. Most of these methods are precise and accurate and can be used for quality control of industrial concentrates and juices. Sheu and Lu (1995) developed a HPLC method for the simultaneous determination of the constituents of a Chinese herbal formula, Hsiao-cheng-chi-tang, containing hesperidin as one of the ingredients. This method utilizes a Cosmosil-5 C-18 column with acetate buffer as the mobile phase and the publication considers the effects of pH, buffer concentration and column selectivity.

A reversed phase HPLC system has been used to separate 141 flavonoids including hesperidin, hesperetin and neohesperidin (Castele *et al.*, 1982). The column used was Lichrosorb RP-18, the mobile phase a combination of an isocratic and gradient elutions (5% aqueous formic acid and methanol) and a UV detector set at 280 nm. They also discussed the correlations between the structure and retention time values of different flavonoids. Thirty four selected flavonoids including hesperidin and hesperetin were analysed by HPLC, using a  $\mu\text{Bondapak}$  C-18 column and UV detector at 254 nm (Daigle and Conkerton, 1982). This method employs two pumps, one eluting water–acetic acid (495:5) and the other eluting methanol. The retention times were measured to calculate two chromatographic parameters—the capacity factors and the relative retentions. They emphasized that ‘adsorption’ and not the size of the molecule is operative in its separation. Another simple and precise method has been established for the simultaneous determination of five selected marker components, including hesperidin, in an oriental pharmaceutical decoction—‘Hei-San’, by means of HPLC using tetra-N-amylammonium bromide (TAA) as an ion-

pair reagent. An ODS column has been used with multi-step gradient elution with 10 mM TAA ( $\text{H}_3\text{PO}_4$ , pH 4.0)-acetonitrile. The acetonitrile increased linearly from 24% to 90%. This method has also been compared with three other methods, i.e. a water-acetonitrile method, a phosphate buffer method and an ion-suppression method, and was found to give comparable and satisfactory results (Yamauchi *et al.*, 1996). In an attempt to screen the flavonoids from *Larix* species, needles of different larch species were extracted and analysed on HPLC using Dupont 830 chromatograph with a Zorbax ODS column using a gradient of 45% methanol in water to 100% methanol, both with 0.1% acetic acid at 50°C, with double detection at 254 and 360 nm. Hesperidin and hesperetin were eluted at a retention time of 6.5 and 9.9 min, respectively (Niemann and Koerselman-Kooy, 1977).

Hesperidin and neohesperidin have been analysed by liquid chromatography/mass spectrometry (LC-MS) with a turbo-ion-spray (TBS) interface. Characterization of isomers differing in the glycosylation was found to be possible on the basis of different mass spectra (Careri *et al.*, 1999). Another LC-MS method using a capillary scale particle beam interface has been developed for the sensitive detection of flavonoid compounds in complex matrices (Capiello *et al.*, 1999).

Other methods used in the analysis of hesperidin include *spectrodensitometry* (El-Bayoumi, 1999), *gas-liquid chromatography* of the trimethylsilyl ether (Drawert *et al.*, 1980) and *micellar electrokinetic capillary chromatography*, MECK (Ferrerres *et al.*, 1994). *Voltammetry* has also been utilized (Obendorf and Reichart, 1995; Zoulis and Efstathiou, 1996; Volikakis and Efstathiou, 2000).

## COMBINATION PRODUCTS

Hesperidin is commonly used in traditional medicines as a combination product. The most widely used combination product of hesperidin is Daflon- 500 mg<sup>®</sup> (Servier; Switzerland), which contains hesperidin (50 mg) and diosmin (450 mg). Diosmin (Fig. 5) is another bioflavonoid with a molecular weight of 608.6 daltons and is chemically diosmetin-7-rhamnoglucoside. The aglycone diosmetin can be described by the systematic name 3', 5,7-trihydroxy-4'-methoxyflavone or 5,7-dihydroxy-2-(3-hydroxy-4-methoxyphenyl)-4H-1-benzopyran-4-one. It differs from hesperetin only in having a double bond between C2 and C3, thereby being a flavone, whereas hesperetin is a flavanone.

Treatment with Daflon-500 mg<sup>®</sup> has demonstrated a wide variety of pharmacological activities including beneficial effects in human subjects with chronic venous insufficiency. Two hundred and fifteen human subjects

were administered one tablet of the drug, twice daily for one year. Subjects recorded an overall assessment of efficacy and relief from venous symptoms such as cramps and evening oedema. Clinical laboratory values remained within normal range throughout the study year (Guillot *et al.*, 1989).

## SAFETY AND TOXICITY

In general, Citrus bioflavonoids including hesperidin appear to be extremely safe and without side effects even during pregnancy (Pizzorno Jr and Murray, 1999). Based on an experiment in male and female mice (Sieve, 1952), it was established that phosphorylated hesperidin (PH) was nontoxic both to the organism and to tissues, easily assimilated, nonaccumulative and caused no allergic reactions. Based on these results Sieve performed a study on humans and reported that phosphorylated hesperidin could be given to humans clinically as an anti fertility agent, along with other substitution factors such as vitamins, endocrines, amphetamine derivatives and decholic acid derivatives. Besides this, trauma, infectious diseases or systemic diseases did not inhibit its antifertility effect.

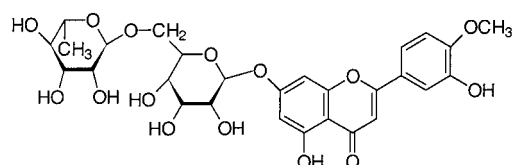
In a study in rats, when Daflon-500 mg<sup>®</sup> was administered by gastric intubation for 26 weeks, no deaths, changes in weight or abnormalities of standard functional tests were observed. A mixture of pure compounds in a ratio of 1:9 was used to obtain the reported effect (Damon *et al.*, 1987). Methyl hesperidin, when administered orally to mice at a level as high as 5% in the diet, exerted no mutagenic or carcinogenic effects. No obvious toxic effects in mice of either sex were observed (Kawabe *et al.*, 1993). Moreover, the ingestion of hesperidin did not affect the daily food intake, body weight gain or food efficiency (Kawaguchi *et al.*, 1997).

In a study in humans hesperidin administration was found to result in minor side effects in only 10% of the subjects compared with 13.9% of those treated with placebo (Meyer, 1994). However, there have been some reports of interactions between the aglycone hesperetin (Mitsunaga *et al.*, 2000), hesperidin (Melzig *et al.*, 1997) and conventional drugs.

## DRUG-FOOD INTERACTIONS

Hesperidin, as such has not been widely reported to interact with drugs or food substances (Kawaguchi *et al.*, 1997). In an early study conducted on human volunteers, PH was found to have no interaction when given concomitantly with vitamins, endocrines, amphetamine derivatives and decholic acid derivatives (Sieve, 1952).

In a recent study conducted to see whether the grapefruit bioflavonoids alter the permeation of vincristine (an agent used in cancer therapy), across the blood-brain barrier, it was observed that hesperetin increased the [3H] vincristine uptake in the 10–50  $\mu\text{M}$  range, but the glycoside did not have any effect (Mitsunaga *et al.*, 2000). This effect was thought to be brought about by the stimulation of P-glycoprotein-mediated drug efflux through the cells. This indicated that the patients taking drugs that are P-glycoprotein substrates, may need to



**Figure 5.** Diosmin (diosmetin-7-rhamnoglucoside)

restrict their intake of bioflavonoid containing foods and beverages. Hesperidin along with other flavonoids was also found to interact with daunomycin (Melzig *et al.*, 1997).

## PHARMACOKINETICS

The pharmacokinetics of hesperidin has not been discussed in great detail in the literature but a few references are available.

### Absorption

To evaluate the oral absorption of hesperidin from Citrus products, healthy white males (human volunteers), 25 years of age, were given 500 mg of the drug in water and equivalent amounts of grapefruit and orange juice. It was absorbed from the gastrointestinal tract after oral administration in any form, but cumulative urinary recovery indicated low bioavailability (<25%). The aglycone, hesperetin, was detected in both urine and plasma. Absorbed Citrus flavanones were thought to undergo glucuronidation before urinary excretion (Ameer *et al.*, 1996). Intestinal permeability to hesperidin glycosides was investigated by using a cultured monolayer of Caco-2 cells as a model for the small intestinal epithelium. Whereas hesperidin did not permeate across the Caco-2 monolayer probably owing to its low solubility, its glycosides did permeate, in a time- and dose-dependent manner. This permeation was thought to occur via a paracellular pathway (Kim *et al.*, 1999). The absorption of orally administered hesperidin in rabbits has been found to depend on the diet. It was absorbed when given with a synthetic ration but not with a commercial pelleted ration (Williams, 1964).

### Metabolism

The metabolic fate of six flavonoids including hesperidin and hesperetin was studied following oral ingestion in rats. The major metabolic product in the urine was *m*-hydroxyphenylpropionic acid along with lesser amounts of *m*-coumaric acid and the aglycones. The aglycones were free as well as conjugated with glucuronic acid. This indicated that absorption had occurred from the intestinal tract followed by dehydroxylation, demethoxylation or demethylation followed by dehydroxylation to yield *m*-hydroxyphenylpropionic acid. This study also indicated that hesperetin was more readily absorbed than hesperidin in rats, rabbits, as well as in humans (Fig. 6) (Booth *et al.*, 1958). When human volunteers ingested hesperidin, a marked difference was observed in the metabolism. 3-Hydroxy-4-methoxyphenylhydracrylic acid was obtained as the major urinary metabolite, indicating that the pyran ring of hesperetin splits to yield this hydracrylic acid. A small amount of the glucuronide of hesperetin was also detected (Booth *et al.*, 1958; Williams, 1964). Hesperidin was found to be transformed to its aglycone, hesperetin, in the intestine by the bacteria producing  $\alpha$ -rhamnosidase and  $\beta$ -glucosidase or endo- $\beta$ -glucosidase. It was also found that the antiplatelet activity and cytotoxicity of the metabolite formed

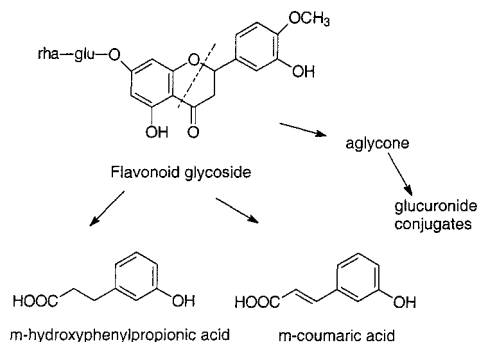


Figure 6. Metabolic fate of a flavonoid glycoside in the rat

in the human intestine was more pronounced than that of the parent compound (Kim *et al.*, 1998).

### Excretion

Recently a study was carried out to study the plasma kinetics and urinary excretion of hesperetin and naringenin from grapefruit and orange juice. Healthy volunteers ingested the juices; blood and urine samples were collected and analysed by HPLC with electrochemical detection. Hesperetin was found to be bioavailable from the study juices with marked inter-individual variations. The urine levels were variable enough to conclude that urinary excretion levels could not be used as biomarkers for dietary intake (Erlund *et al.*, 2001). Animal studies showed virtually complete elimination of Daflon-500 mg<sup>®</sup>, 96 h after administration, without any untoward accumulation in any particular organ (Meyer, 1994).

A resorption and excretion study was performed on <sup>14</sup>C-hesperidin methylchalcone (a hydrosoluble, semi-synthetic derivative of natural hesperidin) in rats and it was observed that, at a dose of 10 mg/kg body weight, it was absorbed 1–2 h after oral administration. The blood kinetic patterns suggested an entero-hepatic cycle, demonstrated by the intravenous administration of the compound at the same dose. The blood profiles demonstrated good bioavailability of the drug. Urinary excretion was lower than faecal excretion after oral ingestion but both were comparable after administration by intravenous route. Moreover, excretion mainly occurred in the first 24 h following administration, by either route (Chanal *et al.*, 1981).

## PHARMACOLOGICAL EFFECTS

Hesperidin is effectively used as a supplemental agent in treatment programmes and protocols of broadly complementary settings.

### Effects on the vascular system

Hesperidin supplementation has been used in patients suffering from blood vessel disorders including fragility and permeability complaints resulting in easy bruising and varicosities. This effect of hesperidin has been of

considerable interest in the last decade and a large number of publications have addressed this therapeutic application.

Increased permeability of blood capillaries is a feature of several disease states and is manifested in such symptoms as oedema, bleeding and hypertension. Diseases which are usually associated with increased capillary permeability include diabetes, chronic venous insufficiency, haemorrhoids, scurvy, various ulcers and bruising. Early research on flavonoids from plant extracts established that they reduced the permeability and fragility of capillary walls (Bisset *et al.*, 1991). Hesperidin, along with other flavonoid compounds has been widely reported to inhibit an increase of capillary permeability. As early as in 1939, Morii found that daily administration of 30 mg of hesperidin decreased the capillary permeability and increased capillary resistance in various clinical cases suffering from pleurisy, tuberculosis, Grave's disease and beriberi (Morii, 1939). Since then, hesperidin has increasingly been used as a therapeutic agent for increasing capillary resistance.

Scarborough (1940) used hesperidin in the treatment of purpura, resulting from the use of arsenicals and in the treatment of syphilis. He reported that hesperidin, as well as hesperetin, were active for increasing capillary resistance. Higby (1941) reported the successful use of hesperidin in the clinical treatment of haemorrhagic purpura and disorders arising from abnormal capillary fragility. This role of hesperidin in increasing capillary resistance has been attributed to its inhibitory effect on the action of hyaluronidase enzyme, which in turn has been known to accentuate capillary permeability and fragility (Beiler and Martin, 1947).

Struckmann and Nicolaides (1994) used Daflon-500 mg<sup>®</sup> to treat chronic venous insufficiency. This effect has been ascribed to an inhibition of inflammatory processes in the ischaemia-induced hyperpermeability that characterizes venous stasis (Jean and Bodinier, 1994). The micronized purified flavonoid fraction containing diosmin and hesperidin (9:1) was found to prevent ischaemia/reperfusion induced leukocyte adhesion in skeletal muscles localized in the ischaemic region (Korthuis and Gute, 1999). An open pilot study, on 24 human patients with third stage chronic venous insufficiency, indicated a beneficial haemorheological effect on oral administration, resolving the stasis with an increase in the blood cell velocity. Furthermore a concomitant increase in relative packed cell volume and red blood cell velocity after therapy suggests an improvement in the flexibility of red blood cells (Allegra *et al.*, 1995). Daflon is also found to possess venotonic and vasculo-protective pharmacological properties and reinforces venous tone by prolonging the activity of parietal norepinephrine (Amiel and Barbe, 1998).

An antihypercholesterolaemic activity of hesperidin in CCl<sub>4</sub>-induced hypercholesterolaemic rats has been reported (Son *et al.*, 1991). Intragastric hesperidin has been found to significantly lower cholesterol, low density lipoproteins (LDL), total lipids and triglyceride levels in normo-lipidaemic rats and in diet- or Triton-induced dyslipidaemic rats but increased high density lipoprotein (HDL) levels (Monforte *et al.*, 1995). Tangerine peel extract and a mixture of hesperidin and naringin were also found to significantly lower levels of plasma and hepatic cholesterol, and hepatic triglycerides compared with those of the control. They also reduced the excretion

of faecal neutral sterols compared with the control (Bok *et al.*, 1999). These effects are accompanied by a reduction in the plasma and hepatic activities of 3-hydroxy-3-methyl-glutaryl-CoA (HMGCoA) reductase and acyl CoA: cholesterol transferase (Bok *et al.*, 1999). The hypocholesterolaemic effect of hesperetin in rats may also be mediated via these enzymes (Lee *et al.*, 1999a).

Hesperidin has also been reported to exert antihyperlipidaemic effects and the methanolic extracts of the stems of *Prunus davidiana* Fr. (Rosaceae), containing hesperetin-5-glucoside and other flavonoids, was found to be useful for the treatment of hyperlipidaemia. Intraperitoneal administration of hesperetin-5-glucoside to rats fed on a high fat diet was found to significantly reduce the total cholesterol level compared with the control group but had no effect on the serum triglyceride level (Choi *et al.*, 1991). In another study, the antilipidaemic activity of hesperidin in normolipidaemic and induced hypercholesterolaemic rats has been demonstrated (Monforte *et al.*, 1995). Hesperidin exerted significant activity, which may be due to increased hepatic cholesterol catabolism. In yet another study, the flavonoids from *C. unshiu* were screened for lipase inhibitory effect and hesperidin was found to inhibit lipase activity from porcine pancreas and that from *Pseudomonas*. Moreover, hesperidin (10%, p.o), decreased plasma triglyceride concentration and increased the amount of faecal lipids excreted in rats (Kawaguchi *et al.*, 1997).

The calcium channel blocker activity of hesperidin has been patented (Morita *et al.*, 1992). It is found to inhibit nitrendipine binding to skeletal muscle membrane protein in rabbits. The capillary antihemorrhagic activity of hesperidin in mice has also been reported (Jurisson, 1973). Kubo *et al.* (1992a,b) have reported and filed patents relating to the myocardial depressant activity of hesperidin in human adults.

Galati *et al.* (1996) have demonstrated significant antihypertensive and diuretic effects of hesperidin in rats following oral administration of the drug at a dose of 200 mg/kg body weight and ascribed this hypotensive effect to increased diuresis. However, other mechanisms are probably involved. It is well established that various flavanones, including hesperidin, influence various enzymes such as protein kinase, lipooxygenase and cyclooxygenase. This effect on enzymes is responsible for the flavonoids' activity on some haematic parameters, lowering of erythrocytic adhesion, platelet aggregation and blood viscosity. The antihypertensive activity of hesperidin might therefore be due to its activity on enzymatic systems that influence blood rheology. In addition, various flavonoids are potent inhibitors of cyclic-AMP phosphodiesterase, and probably this activity is the basis for the observed diuretic effect (Emim *et al.*, 1994; Galati *et al.*, 1994).

### Antiinflammatory effects

Hesperidin has been reported to possess significant anti-inflammatory and analgesic effects (Galati *et al.*, 1994). Emim *et al.* (1994) presented pharmacological data favouring the use of hesperidin as an inexpensive anti-inflammatory agent or as a lead compound, especially for patients with hypersensitivity to the ordinarily used non-steroidal antiinflammatory agents. Hesperidin, though ineffective after oral administration, was found to be

active on subcutaneous injection without remarkable changes in rat behaviour or apparent tissue damage at the site of injection even after repeated administration. At high doses, it also reduced dextran induced paw oedema in rats. This effect could be attributed to the inhibition of the release of histamine from basophils by hesperidin or its metabolic products.

Hesperidin in combination with diosmin, shows a marked protective effect against inflammatory disorders, both *in vivo* and *in vitro*, possibly through a mechanism involving an inhibition of eicosanoid synthesis and/or antioxidant free radical scavenger activity (Jean and Bodinier, 1994). Lonchamp *et al.* (1989) studied the scavenging properties of Daflon 500 mg<sup>®</sup> on active oxygen radicals *in vitro* and *in vivo*, implying its pharmacological action on capillary hyperpermeability as well as antiinflammatory and antioedematous actions.

S-5682 (Daflon) was demonstrated to improve multiple histological aspects of the acute inflammatory reaction (diapedesis of polymorphs, lymphocytes, histocytes and macrophages) and features of the chronic inflammatory reactions (newly formed micro vascularization of the granuloma tissues, perivascular oedema, presence of collagen fibres). Its effects in the chronic treatment of inflammatory granuloma in the rat were studied. It possessed potent antioedematous activity and reduced the synthesis of PGE<sub>2</sub> and PGF<sub>2α</sub> in inflammatory granuloma (Damon *et al.*, 1987).

The antiinflammatory effect of hesperidin in the acute stage of trinitrobenzene sulphonic acid (TNBS) model of rat colitis was evaluated. Pretreatment with hesperidin reduced colonic damage compared with TNBS control rats. In addition, hesperidin after oral administration reduced the areas of colonic necrosis and hyperaemia, scored according to the severity and extension of involved tissue compared with the control group. A significant reduction in the colonic weight/length ratio was also observed. This intestinal antiinflammatory effect was also shown biochemically since hesperidin was able to reduce colonic myeloperoxidase activity, an enzyme that is considered as a biochemical marker for neutrophilic infiltration in damaged tissue (Crespo *et al.*, 1999). Another possible mechanism that contributes to this effect is its antioxidant property (Jean and Bodinier, 1994). Hesperidin has been reported to ameliorate colonic oxidative stress that occurs in an experimental model of inflammatory bowel disease (Loguercio *et al.*, 1996). When given intragastrically to mice at a dose of 25 mg/kg, it was found to reduce adjuvant-induced arthritis (Kim *et al.*, 1990).

Pretreatment with Daflon-500<sup>®</sup> prior to the induction of tourniquet ischaemia significantly lowered the number of adherent leukocytes thereby controlling oedema in clinical situations (Friesenecker *et al.*, 1995). This protective effect is associated with the decreased platelet and complement system activation, leading to a lowered release of histamine and decreased leukocyte-dependent endothelial damage.

### Action on enzymes

Hyaluronidase is an enzyme which depolymerizes the mucopolysaccharide hyaluronic acid. It has been known to play a part in the typical changes in the morphology of the connective tissue, particularly of the interfibrillator

cement substance, which is believed to contain hyaluronic acid. This enzyme plays a role in regulating the permeability of capillary walls and supporting tissues. Hyaluronidase causes a breakdown of hyaluronic acid, thereby increasing tissue permeability. Also, various bacteria produce hyaluronidase, thereby increasing the permeability of the tissue and favouring the invasion of tissue by these and other microorganisms (Hahn, 1952). Various scientists and medical researchers have been working on the hyaluronidase inhibitory effects of different flavonoids. Beiler and Martin, as early as in 1948, prepared various derivatives of hesperidin as hyaluronidase inhibitors. The sulphonated and phosphorylated hesperidins proved to be extremely potent inhibitors, and acetylated hesperidin caused a potentiation in the inhibitory action of ascorbic acid on hyaluronidase. They demonstrated that, although pure hesperidin and hesperetin were active as hyaluronidase inhibitors, this action could be greatly potentiated by the formation of the above mentioned derivatives of the pure compound (Beiler and Martin, 1948). Based on an investigation on the isolated connective membranes from mice, it has been reported that the permeability increasing action of hyaluronidase could be abolished by contact with phosphorylated hesperidin for 15 min or more. It not only abolishes the hyaluronidase action but also restores the original degree of permeability of the membranes. It even reduces the permeability of normal (untreated) membranes (Steincke, 1956).

Besides hyaluronidase, hesperidin has also been reported to inhibit human acrosin, a sperm enzyme, *in vitro* (Jackson, 1959). It also shows an inhibitory effect on aldol reductase (lens) *in vitro* (Varma and Kinoshita, 1976), alkaline phosphatase in rat serum (Son *et al.*, 1991) and a weak activity against alpha-glucosidase *in vitro* (Iio *et al.*, 1984). Inhibition of alkaline phosphatase was, however, found to be absent in a study on rat liver (Son *et al.*, 1991) and another *in vitro* study where the enzyme was derived from calf intestinal mucosa (Iio *et al.*, 1980). Hesperidin, however, did not inhibit xanthine oxidase *in vitro* (Iio *et al.*, 1985). It was also found to be inactive as an inhibitor of the enzyme reverse transcriptase, when tested on virus-raucher murine leukaemia *in vitro* (Ono *et al.*, 1990).

In a study carried out on some flavonoids, on nonenzymatic lipid oxidation and enzymatic oxidation of arachidonic acid *in vitro*, hesperidin was found to stimulate the enzyme cyclooxygenase (Iio *et al.*, 1984). However, inhibition of 1,5-lipoxygenase by Citrus peel flavonoids has been reported (Malterud and Rydland, 2000).

When a range of flavonoids were screened for inhibitory effects against partially purified aromatase prepared from human placenta, hesperetin was found to possess a significant inhibitory effect on the same (Jeong *et al.*, 1999). A series of flavonoids were tested for their effects on low K<sub>m</sub> phosphodiesterase with cyclic AMP as the substrate and for their lipolytic activity, employing rat adipocytes. Hesperetin showed inhibition of epinephrine induced lipolysis, in a dose dependent manner but did not inhibit phosphodiesterase significantly (Kuppusamy and Das, 1992).

Hesperidin and diosmin, alone and in combination (1:9), were found to inhibit prostaglandins, when used against sponge-induced granuloma in rats, significantly lowering the prostaglandin levels of the treated animals

compared with those of untreated controls, through 16 days after intragastric sponge implantation (Damon *et al.*, 1987).

### Antimicrobial activity

Hesperidin and hesperetin, among other flavonoids, have shown anti-infective and antireplicative activities *in vitro* against several plant and animal microbes.

**Antibacterial.** In a study involving the investigation of anti-*Helicobacter pylori* (HP) activity, *in vitro*, of some flavonoids and their metabolites, hesperetin and other flavonoids were found to inhibit the growth of HP (Bae *et al.*, 1999). In patients with chronic gastritis, HP promotes the alteration from gastritis to gastric cancer (Correa, 1988). However, Islam and Ahsan (1997), recently showed hesperidin to be inactive *in vitro* on agar plates, against *Bacillus subtilis*, *Staphylococcus aureus*, *Streptococcus hemolyticus*, *Escherichia coli*, *Klebsiella* species, *Pseudomonas aeruginosa*, *Salmonella typhi*, *Shigella dysenteriae*, *Shigella flexneri* and *Vibrio cholera*.

**Antifungal.** The antifungal activity of hesperidin has been reported at a dose ranging from 1 to 10 µg, against *Botrytis cinerea*, *Trichoderma glaucum* and *Aspergillus fumigatus* (Krolicki and Lamer-Zarawska, 1984). It was, however, found to be inactive against *Aspergillus fumigatus*, *Aspergillus niger* and *Trichoderma* species (Islam and Ahsan, 1997).

**Antiviral.** Wacker and Eilmes, in their two studies, reported hesperidin to be active as an antiviral agent, *in vitro*, at different concentrations on cell culture, against virus-vesicular stomatitis. Hyaluronidase was found to reverse its antiviral activity suggesting that this activity of hesperidin was mediated by its antihyaluronidase effect. They also reported hesperidin to be active against influenza virus (Wacker and Eilmes, 1975; Wacker and Eilmes, 1978). In a recent study, hesperidin was found to possess a weak activity against herpes simplex virus (Lee *et al.*, 1999b). It was found that certain flavonoids, including hesperidin, possessed antiviral activity against herpes type-I, para influenza-3, poliovirus type-I and respiratory syncytial virus (RSV) in tissue cell monolayers (Middleton, 1984). Mucsi and Pragai (1985) demonstrated the inhibitory effect of hesperidin among other flavonoids on human herpes simplex virus type-I (HSV-I) and suid (alpha) herpes virus type-I (*Pseudorabies* virus). A direct relationship between the antiviral activity of the flavonoid and its ability to stimulate cyclic AMP synthesis in the cells was exhibited. Quercetin and hesperetin are also known to possess antireplicative activity (inhibition of virus replication after established infection) if introduced at the time of cell activation. Hesperetin was, however, found to be inactive against HIV-virus (Hu *et al.*, 1994), pseudorabies virus, herpes simplex virus (Mucsi and Pragai, 1985) and rhinovirus (Tsuchiya *et al.*, 1985). In a recent study, the inhibitory effects of some flavonoids was tested on infectivity of rotavirus, which predominantly causes sporadic diarrhoea in infants and young children (Bae *et al.*, 2000) and hesperidin was found to have a potent inhibitory effect. Hesperetin, however, did not possess this activity, indicating that the rutinoside moiety is essential in

protecting against the invasion of rotavirus into the cells. The fruit of *C. aurantium*, having hesperidin and neohesperidin as main constituents, was also found to possess a potent inhibitory effect on rotavirus infectivity (Kim *et al.*, 2000).

When tested for an effect on the infectivity and replication of HSV-1, polio-virus type-1 and para-influenza virus type-3, hesperetin was found to have no effect on the infectivity but reduced intracellular replication of each of the viruses (Kaul *et al.*, 1985). Hesperidin was also found to have a significant effect against *Staphylococcus aureus* on infected mice (Panasiak *et al.*, 1989).

**Anti yeast.** Hesperidin was found to have no inhibitory effect on *Candida albicans* and *Saccharomyces cerevisiae*, when tested *in vitro* on agar plates (Islam and Ahsan, 1997).

### Anti fertility activity

Hesperidin and its derivatives have been under investigation as anti fertility factors for a very long time. As early as 1948, phosphorylated hesperidin (PH) was reported to act as an effective anti fertility agent mediating this effect by inhibiting the sperm enzyme hyaluronidase (Beiler and Martin, 1948; Martin and Beiler, 1952). In a study on rats, PH was found to have an antifertility effect when given orally or intraperitoneally (Martin and Beiler, 1952). In another attempt to assess the antifertility effect, PH (20 mg/kg) was administered to both male and female mice, intraperitoneally for 8 days. On mating, the percent pregnancies were significantly reduced. The oestrus cycle of the females remained unchanged. Microscopic examination of the seminal fluid from males showed a normal sperm count and sperm motility. No permanent sterility in either sex was observed. Based on these results, a study was further carried out on 300 married human couples. PH tablets (100 mg) were given orally to both the partners, over varying periods, up to 30 months. Contraception occurred in all the cases except for two, where the couples were non cooperative regarding the study parameters. Merely omitting the drug for a period of 48 h could restore fertility. It thereby offers promise as an oral contraceptive without any toxic effects or permanent inhibition of sterility. Trauma, infectious diseases or systemic diseases did not inhibit the antifertility effect (Sieve, 1952).

Later it was reported that the fertilizing capacity of rabbit sperm was inhibited to a certain extent when the sperm was suspended in a 1% solution of PH and inseminated into female rabbits (Chang and Pincus, 1953). PH also exhibited significant contraceptive efficacy on vaginal application in rabbits, even at non-spermicidal concentrations (Joyce and Zaneveld, 1985). In yet another study, PH was proven to inhibit bovine fertilization by mouse spermatozoa by specifically inhibiting the sperm enzyme hyaluronidase. Hyaluronidase is reported to be required for sperm penetration through the follicle cell layer of the egg. PH, by inhibiting sperm hyaluronidase can thereby successfully be used as an antifertility agent (Joyce *et al.*, 1986).



## Anticarcinogenic activity

During the past decade, a considerable amount of research has been carried out on the anticancer activities of hesperidin and its aglycone hesperetin. Encouraging results of carcinogenesis inhibition were observed by using a hesperidin/diosmin combination on male mouse urinary bladder. The combination inhibited *N*-butyl-*N*-(4-hydroxybutyl)nitrosamine-induced carcinogenesis when treated with the combination for 8 weeks during the tumour initiation phase (Yang *et al.*, 1997). An inhibition of carcinogenesis in rats by hesperidin, at a concentration of 500 ppm/kg body weight has also been reported. The compound was also found to inhibit 4-nitroquinoline-1-oxide-induced oral carcinogenesis and to decrease the number of lesions, polyamine levels in tongue tissue and cell proliferation activities (Tanaka *et al.*, 1994). The same group later reported a similar activity of hesperidin alone, and in combination with diosmin, where it not only inhibited 4-nitroquinoline-1-oxide initiated tumorigenesis but also showed chemoprevention of azoxymethane-induced rat colon carcinogenesis. These results are thought to be due to increased suppression of cell proliferation (Tanaka *et al.*, 1997a,c).

Hesperidin, when administered subcutaneously to CD-1 mice, did not inhibit 7,12-dimethylbenz( $\alpha$ )anthracene-induced tumour initiation but inhibited 12-O-tetradecanoyl-13-phorbol acetate (TPA)-induced tumour promotion, thus indicating its potential use as a chemopreventive agent (Berkarda *et al.*, 1998). Later they established the protective effect of hesperidin against TPA-stimulated infiltration of neutrophils. It also afforded significant protection against TPA induced hyperplasia in the dorsal skin of CD-1 mouse, through multiple applications prior to TPA (Koyunku *et al.*, 1999).

The mutagenic activity of hesperetin was assessed along with several other flavonoids and their derivatives using *Salmonella typhimurium* mutants that reveal base pair substitutions and frameshift mutagens. Neither hesperetin nor neohesperetin were found to be mutagenic (Bjeldanes and Chang, 1977). Much later when the antimutagenicity of various Citrus flavonoids including naringin, hesperidin, nobletin and tangeritin was evaluated, hesperidin was found to possess a weak antimutagenic effect on *Salmonella typhimurium* against benz( $\alpha$ )pyrene-induced mutations (Choi *et al.*, 1994; Calomme *et al.*, 1996). An antimutagenic effect has also been reported against *N*-methyl-*N*-amyl nitrosamine-induced tumorigenesis, in rats (Tanaka *et al.*, 1997b). When a number of plant flavonoids were tested for mutagenicity in the *Salmonella*/microsomal activation system, both hesperidin and hesperetin were found to require metabolic activation for any mutagenicity (Hardigree and Epler, 1978). Hesperetin has shown an antimutagenic effect against aflatoxin B1 with more than a 70% inhibition rate in *S. typhimurium*, in the presence of a mammalian metabolic activation system (Choi *et al.*, 1994).

When the effects of predominant dietary flavonoids were tested for inhibiting neoplastic transformation induced by 3-methylcolanthrene in C3H 10T1/2 murine fibroblasts, hesperidin and hesperetin were found to be the most potent agents inhibiting this transformation completely (Franke *et al.*, 1998). Structure-activity comparisons were also made and favourable structures discussed.

## Platelet and cell aggregation inhibition

Hesperidin was found to increase the survival time of rats by 16–71 days when they were placed on a thrombogenic diet but produced variable results when the rats were on an atherogenic diet (Robbins, 1967). In studies carried out on human adult platelets (Zaragoza *et al.*, 1985; 1986), hesperidin was reported to inhibit epinephrine- and ADP-induced platelet aggregation at a concentration of 0.08 mg/ml. In yet another study carried out on horses, hesperidin was found to effectively reduce aggregation of erythrocytes. This decrease in the blood cell aggregation may explain the beneficial effects of hesperidin on abnormal capillary permeability and fragility, the reduction of disease symptoms and their protection against various traumas and stresses (Robbins, 1971). Hesperidin and diosmin, in combination, were found to decrease leukocyte adhesion to the endothelium in post capillary venules, after ischaemia/reperfusion, as assayed through skin fold window chamber in guinea-pigs following intragastric administration of the compounds (Friesenecker *et al.*, 1994). This antiischaemic effect of hesperidin was again reported in the hamster, at a dose of 20 mg/kg body weight after intragastric administration (Bouskela and Donyo, 1995). In addition a micronized purified flavonoid fraction containing hesperidin and diosmin was found to significantly decrease ADP-induced platelet aggregation and increase platelet disaggregation, in rats. Fibrinogen binding to ADP-induced platelets was also reduced significantly (McGregor *et al.*, 1999).

## Ultraviolet protecting activity

Recently much research has been focused on the potential use of flavonoids as free radical scavengers to prevent oxidative skin damage (Schoemaker *et al.*, 1995; Mortimer, 1997). The oxidative stress caused by ultraviolet irradiation might be an initiator in the pathogenesis of skin cancer and photoaging (Dalle Carbonare and Pathak, 1992; Darr and Fridovich, 1994). Bonina *et al.* (1996, 1998) investigated the ability of topically applied hesperetin, alone and in a crude extract of *C. sinensis* along with other flavonoids, to reduce UV-B-induced skin erythema. Employing phosphatidylcholine (PC) vesicles as model membranes they studied the effect of hesperetin on UV irradiation induced peroxidation. *In vitro* human skin permeation of these compounds was also measured. Hesperetin was effectively found to protect PC liposomes from UV-irradiation induced peroxidation probably by scavenging oxygen free radicals generated by UV irradiation. It was also able to permeate through the stratum corneum. It was later concluded from *in vitro* and *in vivo* data, obtained after applying a gel formulation uniformly on the skin sites (human volunteers) and exposing them to UV-B irradiation and monitoring the extent of erythema by means of reflectance spectrophotometry, that hesperetin might be successfully employed as a topical photo-protective agent (Saija *et al.*, 1998).

In another study, transglycosylation of hesperidin by cyclodextrin glucanotransferase was conducted and its mono- and di-glucosides were prepared, both having the same absorption spectra as that of hesperidin. When these were exposed to ultraviolet light, they seemed to stabilize

the colour of the pigments by absorbing the UV rays. In addition they did not have strong spectra in visible light. The authors propose its use as a colour stabilizer in food products (Kometani *et al.*, 1994).

### Miscellaneous effects

**Analgesic and antipyretic activity.** Hesperidin has exhibited analgesic activity in mice on subcutaneous administration. This effect has been explained as being exerted through a peripheral and not a central mechanism (Galati *et al.*, 1994). It also showed analgesic activity in mice following intraperitoneal administration. Also, hesperidin decreased the fever induced by yeast in rats. This effect may be related to the inhibition of yeast-induced prostaglandin biosynthesis. Hesperidin is known to inhibit both histamine and prostaglandin release, thereby acting as a defensive gastric factor and preventing acid secretions and lesions of the gastric mucosa (cited in Emim *et al.*, 1994).

**Antioxidant effect.** Hesperidin has been reported to possess antioxidant properties. For example, it has been found to reduce superoxide in electron transfer plus concerted proton transfer reaction *in vitro* (Jovanovic *et al.*, 1994). This activity was also exhibited in liver homogenates for hydroperoxide-induced chemiluminescence (Fraga *et al.*, 1987). It has been suggested that the hesperidin/diosmin combination could function as an antioxidant, which may explain its beneficial therapeutic effect in chronic venous insufficiency where oxidative stress is involved in the pathological mechanism (Bouskela *et al.*, 1997). A number of researchers have examined the antioxidant activity and radical scavenging properties of hesperidin using a variety of assay systems (Kroyer, 1986; Brasseur *et al.*, 1986; Ratty and Das, 1988; Yuting *et al.*, 1990; Wang and Zheng, 1992; Miyake *et al.*, 1997; Deng *et al.*, 1997; Miller and Rice-Evans, 1997; Suarez *et al.*, 1998; Malterud and Rydland, 2000). Results from different assays varied considerably, but in most, hesperidin was found to be inactive or only moderately active in comparison with other flavonoids and nonflavonoidal antioxidants. Hesperidin was also found not to inhibit the liberation of reactive oxygen species from stimulated neutrophils (Limasset *et al.*, 1993). Thus hesperidin appears not to be a particularly active antioxidant in comparison with most other flavonoids.

**Immuno-modulatory activity.** It has been reported that hesperidin possesses an immuno-suppressant activity (Kim and Cho, 1991). It suppresses the bacterial alpha-amylase antibody production in mice on intragastric administration at a dose of 50 mg/kg. In another study, intragastric administration of 50 mg/kg of hesperidin to male mice, significantly increased the development of immunological memory in cellular immune response (Kim and Cho, 1991).

Recently, the colony stimulating factor (CSF) inducing activity of various bioflavones was tested, in order to evaluate their immuno-modulating activities. Samples of different bioflavonoids, suspended in saline were injected intraperitoneally into mice at a dose of 1 mg/kg, 6 h before bleeding. Hesperidin exhibited the strongest CSF

inducing activity and the response was dose dependent (Kawaguchi *et al.*, 1999).

**Antiallergic effects.** Hesperidin's antiallergic and anti-anaphylactic activities have been reported and patented (Kubo and Matsuda, 1992). It was shown to possess antianaphylactic activity where it delayed hypersensitivity against sheep red blood cells, prevented passive cutaneous anaphylaxis, induced mast cell degranulation and also prevented histamine-induced anaphylaxis after intragastric administration to mice at different doses (Kim and Chung, 1990). Later, it was found to inhibit vascular permeability in the homologous passive cutaneous anaphylaxis test conducted on rats after intragastric administration, at a dose of 0.1 mmol/kg (Kubo and Matsuda, 1992). An inhibition of 48 h homologous passive cutaneous anaphylaxis in rats has also been reported with the use of hesperidin (Matsuda *et al.*, 1991).

**Activity on haemorrhoids and IUCD-induced bleeding.** Oral tablets of a hesperidin/diosmin combination were clinically tested in situations involving intrauterine contraceptive device (IUCD)-induced bleeding. Thirty one women were given the combination over a period of three consecutive cycles. Eighty three percent of the patients showed improvement in the signs and symptoms associated with the bleeding. The acceptability was excellent with minimal side effects (Daftary *et al.*, 1995). Daflon-500 mg<sup>®</sup> has also been found to reduce inflammation by inhibition of prostaglandin and thromboxane B2 release from macrophages and also to reduce the free radicals. The investigators postulated that inhibition of the inflammatory response in the haemorrhoid surgical wound site reduced defaecation stress and bacterial fibrinolysis, decreasing the risk of secondary bleeding (Ho *et al.*, 1995). The effectiveness and tolerability of a similar hesperidin/diosmin combination was evaluated for the treatment of haemorrhoids. Overall improvement of signs and symptoms was quicker and greater with good acceptability and minimal side effects (Godeberge, 1995). Recently, a similar combination was provided in a micronized form, for a median of 8 weeks before delivery and 4 weeks after delivery, to 50 pregnant women with acute haemorrhoids and was found to be very successful with 66% of the patients reporting relief of symptoms by day 4 of treatment and fewer suffering relapses during the antenatal period. The treatment was well accepted and did not affect pregnancy, fetal development, birth weight, infant growth or feeding (Buckshee *et al.*, 1997).

**Hormonal disorders.** Hesperidin was reported (Smith, 1964) to help in regulating oestrogen levels and decreasing related pain, inflammation and swelling. In a clinical study, 94 women suffering from hot flashes and other menopausal symptoms were given a formula containing 900 mg of hesperidin, 300 mg of hesperidin methylchalcone and 1200 mg of vitamin C daily. At the end of 1 month, symptoms of hot flashes were relieved in 53% of the patients and reduced in 34%. Improvements were also noted in nocturnal leg cramps, nose bleeds and easy bruising. The only side effect noted was a slightly offensive body odour with a tendency for the perspiration to discolour the clothing.

**Antiulcer activity.** Hesperidin demonstrated no ulcero-

genic effect in male rats at a dose of 100 mg/kg by subcutaneous administration (Emim *et al.*, 1994). However, when given to rats orally at a dose of 100 mg/kg, the compound exhibited a positive antiulcer effect on cold stress-induced ulcers but had no effect on ethanol-induced ulcers. No increase in gastric mucus was observed. It was found to stimulate hexosamine secretion in cold stress-induced ulcers but inhibited the same in ethanol-induced ulcers (Suarez *et al.*, 1996).

**Effect on wound healing.** Recently a micronized flavonoid fraction, comprising of 90% diosmin and 10% hesperidin, was tested for its effect on clean wounds and those infected with *S. aureus* on oral and topical administration. The study showed that while there was no significant effect on clean wounds, the combination had a beneficial effect on the infected wounds, when administered both orally and topically (Hasanoglu *et al.*, 2001).

## MARKETED THERAPEUTIC SYSTEMS

Hesperidin, though not yet very widely used therapeutically, has been listed by the Martindale Extra Pharmacopoeia to be an ingredient of various formulations internationally used for vascular disorders (Angiopan—Gentili, Italy; Circovenil—Wyeth, Spain; Daflon—therapia, Germany; Varico Sanol Forte—Sanol, Germany), capillary fragility (Cepevit-K—Darcy, France), haemorrhoids (Daflon 500—Servier, Switzerland; Hamamelis

complex—Blackmores, Australia), rheumatic and joint disorders (Guaiaicum complex—Blackmores, Australia; Ostochort—Adenylchemic, Germany), vitamin C deficiency and dietary supplement (HY-C—Solgar, USA; Min-Detox-C—Eagle, Australia), skin trauma (Proveno—Madaus, Germany; Ondascora—Servier, France), obstetric disorders (Rubus complex—Blackmores, Australia), gingival inflammation (Peridin-C—Hamilton, Australia), fluid retention and gastrointestinal disorders (Hepanephrol—Rosa Phytopharma, France) (Reynolds, 1996).

## CONCLUSION

From the above review, it can be concluded that hesperidin, its aglycone hesperetin, and their various derivatives possess significant potential as therapeutic agents for a wide range of diseases and disorders. This promise needs to be effectively studied at the clinical level, so as to firmly establish the usefulness of these compounds in the treatment or prevention of disease in humans.

## Acknowledgements

The authors wish to thank TOPCAD, Chicago, USA, for financing this project and for helping to further research in this area.

## REFERENCES

- Agarwal PK. 1989. In *Carbon-13 NMR of flavonoids*. Elsevier: Amsterdam.
- Allegra C, Bartolo M, Jr, Carioti B, Cassiani D. 1995. An original microhaemorheological approach to the pharmacological effects of Daflon-500 mg in severe chronic venous insufficiency. *Int J Microcirc Clin Exp* **15** Suppl 1: 50–54.
- Ameer B, Weintraub RA, Johnson JV, Yost RA, Rouseff RL. 1996. Flavanone absorption after naringin, hesperidin and citrus administration. *Clin Pharmacol Ther* **60**: 34–40.
- Amiel M, Barbe R. 1998. Study of the pharmacodynamic activity of Daflon-500 mg. *Ann Cardiol Angiol (Paris)* **47**: 185–188.
- Arakawa H, Nakazaki M. 1960. Absolute configuration of (–) hesperetin and (–) liquiritigenin. *Chem Ind (Rev)* **73**.
- Armentano L, Bentsath A, Beres T, Rusznyak S, Szent-Györgi A. 1936. Über den Einfluss von substanzen der flavon gruppe auf die permeabilität der kapillaren. *Vitamin P. Dtsch Med Wochenschr* **62**: 1325–1328.
- Arthur HR, Wui WH, Ma CN. 1956. An examination of the Rutaceae of Hong Kong. Part-I. Flavonoid glycosides from *Zanthoxylum* species and the occurrence of optically active hesperidin. *J Chem Soc (I)*: 632–635.
- Asahina Y, Nakagome G, Inubuse M. 1930. Flavanone glucosides(V): Reduction of flavanone and flavonol derivatives. *J Pharm Soc-Japan* **50**: 217–223.
- Bae EA, Han MJ, Kim DH. 1999. *In vitro* anti-Helicobacter activity of some flavonoids and their metabolites. *Planta Med* **65**: 442–443.
- Bae EA, Han MJ, Lee M, Kim DH. 2000. *In vitro* inhibitory effect of some flavonoids on rotavirus infectivity. *Biol Pharm Bull* **23**: 1122–1124.
- Barthe GA, Jourdan PS, McIntosh Ca, Mansell RL. 1988. Radioimmunoassay for the quantitative determination of hesperidin and analysis of its distribution in *Citrus sinensis*. *Phytochemistry* **27**: 249–254.
- Beiler JM, Martin GJ. 1947. Inhibitory action of vitamin-P compounds on hyaluronidase. *J Biol Chem* **171**: 507–511.
- Beiler JM, Martin GJ. 1948. Inhibition of hyaluronidase action by derivatives of hesperidin. *J Biol Chem* **174**: 31–35.
- Berkarda B, Koyuncu H, Soybir G, Baykut F. 1998. Inhibitory effect of hesperidin on tumour initiation and promotion in mouse skin. *Res Exp Med (Berl)* **198**: 93–99.
- Bhalla NP, Dakwake RN. 1978. Chemotaxonomy of *Indigofera* Linn. *J Indian Bot Soc* **57**: 180–185.
- Bisset NG, Houghton PJ, Hylands PJ. 1991. In *The medicinal plant industry. Medicinal Plant Industry*. CRC Press: Boca Raton, FL.
- Bjeldanes LF, Chang GW. 1977. Mutagenic activity of quercetin and related compounds. *Science* **197**: 577–578.
- Brasseur T, Angenot L, Pincemail J, Deby C. 1986. Propriétés antiradicalaires antilipoperoxydantes et antioxydantes de flavonoides. *Bull Liaison Groupe Polyphenols* **13**: 557–559.
- Bok SH, Lee SH, Park YB *et al.* 1999. Plasma and hepatic cholesterol hepatic activities of 3-hydroxy-3-methyl-glutaryl-CoA reductase and acyl CoA; Cholesterol transferase are lower in rats fed Citrus peel extract or a mixture of Citrus bioflavonoids. *J Nutr* **129**: 1182–1185.
- Bonina F, Lanza M, Montenegro L *et al.* 1996. Flavonoids as potential protective agents against photo-oxidative skin damage. *Int J Pharm* **145**: 87–94.
- Bonina F, Saija A, Tomaino A, Lo Cascio R, Dederen JC. 1998. *In vitro* antioxidant activity and *in vivo* photoprotective effect of a red orange extract. *Int J Cosm sci* **20**: 331–342.
- Booth AN, Jones FT, DeEds F. 1958. Metabolic fate of hesperidin, eriodictyol, homoeriodictyol and diosmin. *J Biol Chem* **230**: 661–668.
- Bouskela E, Cyrino FZ, Lerond L. 1997. Effects of oral administration of different doses of purified micronized flavonoid fraction on microvascular reactivity after ischemia/reperfusion in the hamster cheek pouch. *Br J Pharmacol* **122**: 1611–1616.

- Bouskela E, Donyo KA. 1995. Effects of oral administration of purified micronised flavonoid fraction on increased microvascular permeability induced by various agents and on ischemia/reperfusion in diabetic hamsters. *Int J Microcirc Clin Exp* **15**: 293–300.
- Buckshee K, Takkar D, Aggarwal N. 1997. Micronised flavonoid therapy in internal hemorrhoids in pregnancy. *Int J Gynaecol Obstet* **57**: 145–151.
- Budavari S. 1996. In *The Merck Index*. Merck and Co., Inc.: NJ.
- Calomme M, Pieters L, Vlietinck A, Berghe DV. 1996. Inhibition of bacterial mutagenesis by Citrus flavonoids. *Planta Med* **62**: 222–226.
- Canclon PF. 1999. Analytical monitoring of citrus juices by using capillary electrophoresis. *J AOAC Int* **82**: 195–206.
- Capiello A, Famiglini G, Mangani F, Careri M, Lombardi P, Mucchino C. 1999. Liquid chromatographic/mass spectrometric determination of phenolic compounds using a capillary scale particle beam interface. *J Chromatogr A* **855**: 515–527.
- Careri M, Elviri L, Mangia A. 1999. Validation of a liquid chromatography ion spray mass spectrometry method for the analysis of flavanones, flavones and flavonols. *Rapid Commun Mass Spectrom* **13**: 2399–2405.
- Castele KV, Geiger H, Van Sumere CF. 1982. Separation of flavonoids by reversed phase high performance liquid chromatography. *J Chromatogr* **240**: 81–94.
- Chanal JL, Cousse H, Sicart MT, Bonnaud B, Marnigan R. 1981. Absorption and elimination of (<sup>14</sup>C) hesperidin methylchalcone in the rat. *Eur J Drug Metab Pharmacokin* **6**: 171–177.
- Chang MC, Pincus G. 1953. Does phosphorylated hesperidin affect fertility? *Science* **117**: 274–276.
- Chinou I, Harvala C. 1997. Polyphenolic constituents from the leaves of *Cynara* species growing in Greece. *Planta Med* **63**: 469–470.
- Choi JS, Park KY, Moon SH, Rhee SH, Young HS. 1994. Antimutagenic effects of plant flavonoids in the Salmonella assay system. *Arch Pharm Res* **17**: 71–75.
- Choi JS, Yokozawa T, Oura H. 1991. Antihyperlipidemic effect of flavonoids from *Prunus davidiana*. *J Nat Prod* **54**: 218–224.
- Clemetson CAB. 1989. *Vitamin C*. CRC Press: Boca Raton, Florida, 101–128.
- Correa P. 1988. A human model of gastric carcinogenesis. *Cancer Res* **48**: 3554–3560.
- Crespo ME, Galvez J, Cruz T, Ocete MA, Zarzuelo A. 1999. Anti-inflammatory activity of diosmin and hesperidin in rat colitis induced by TNBS. *Planta Med* **65**: 651–653.
- Daftary SN, Irani JS, Tsouderos Y. 1995. Therapeutic activity of micronized flavonoid fraction (Daflon-500 mg) in IUCD-induced bleeding. *Drugs Today* **31**: 41–45.
- Daigle DJ, Konkerton EJ. 1982. High-performance liquid chromatography of 34 selected flavonoids. *J Chromatogr* **240**: 202–205.
- Dalle Carbonare M, Pathak MA. 1992. Skin photosensitising agents and the role of reactive oxygen species in photoaging. *J Photochem Photobiol B* **14**: 105–124.
- Damon M, Flandre O, Michel F, Labrid C, Crastes de Paulet A. 1987. Effect of chronic treatment with a purified flavonoid fraction on inflammatory granuloma in the rat. Study of prostaglandin E<sub>2</sub> and F<sub>2</sub> alpha and thromboxane B<sub>2</sub> release and histological changes. *Arzneimittelforsch* **37**: 1149–1153.
- Darr D, Fridovich I. 1994. Free radicals in cutaneous biology. *J Invest Dermatol* **102**: 671–675.
- Deng W, Fang WL, Wu J. 1997. Flavonoids function as antioxidants: by scavenging reactive oxygen species or by chelating iron? *Radiat Phys Chem* **50**: 271–276.
- Di Mauro A, Fallico B, Passerini A, Rapisarda P, Maccarone E. 1999. Recovery of hesperidin from orange peel by concentration of extracts on styrene-divinylbenzene resin. *J Agric Food Chem* **47**: 4391–4397.
- Drawet F, Leupold G, Pivernetz H. 1980. Quantitative gas chromatographic determination of Rutin, Hesperidin und Naringenin in Orangensaft. *Chem Mikrobiol Technol Lebensm* **6**: 189–191.
- El-Bayoumi A. 1999. Modified H-point standard addition method and logarithmic function for the spectrometric and spectrophotometric determination of hesperidin and diosmin in mixtures. *Anal Lett* **32**: 383–400.
- Emim JADS, Oliveira AB, Lapa AJ. 1994. Pharmacological evaluation of the anti-inflammatory activity of a Citrus bioflavonoid, hesperidin and the isoflavonoids dauricin and claussequinone, in rats and mice. *J Pharm Pharmacol* **46**: 118–122.
- Erlund I, Meririnne E, Alfthan G, Aro A. 2001. Plasma kinetics and urinary excretion of the flavanones naringenin and hesperetin in humans after ingestion of orange juice and grapefruit juice. *J Nutr* **131**: 235–241.
- Evans WC. 1996. In *Trease and Evans' Pharmacognosy*. W. B. Saunders: The Alden Press, Oxford, UK.
- Ferreres F, Blazquez MA, Gil MA, Tomas-Barberan FA. 1994. Separation of honey flavonoids by micellar electrokinetic capillary chromatography. *J Chromatogr A* **669**: 268–274.
- Fisher JF. 1978. A HPLC method for the quantification of hesperidin in orange juice. *J Agric Food Chem* **26**: 1459–1460.
- Fluckiger FA, Hanbury D. 1986. In *Pharmacographia—A History of the Principle Drugs of Vegetable Origin*. International Book Distributors: Delhi, India; 104–105.
- Fox DW, Savage WL, Wender SH. 1953. Hydrolysis of flavonoid rhamnoglucosides to flavonoid glucosides. *J Am Chem Soc* **75**: 2504–2405.
- Fraga CG, Martino VS, Ferraro GE, Goussio JD, Boveris A. 1987. Flavonoids as anti-oxidants evaluated by *in vitro* and *in situ* liver chemiluminescence. *Biochem Pharmacol* **36**: 717–720.
- Franke AA, Cooney RV, Custer LJ, Mordan LJ, Tanaka Y. 1998. Inhibition of neoplastic transformation and bioavailability of dietary flavonoid agents. *Adv Exp Med Biol* **439**: 237–248.
- Friesenecker B, Tsai AG, Intaglietta M. 1995. Cellular basis of inflammation, edema and the activity of Daflon 500 mg. *Int J Microcirc Clin Exp* **15**: 17–21.
- Friesenecker B, Tsai AG, Allegra C, Intaglietta M. 1994. Oral administration of purified micronised flavonoid fraction suppresses leukocyte adhesion in ischemia-reperfusion injury: *In-vivo* observations in the hamster skin fold. *Int J Microcirc Clin Exp* **14**: 50–55.
- Galati EM, Monforte MT, Kirjavainen S, Forestieri AM, Tripodo MM. 1994. Biological effects of hesperidin, a Citrus flavonoid. Part 1. Anti-inflammatory and analgesic activity. *Farmaco* **49**: 709–712.
- Galati EM, Trovato A, Kirjavainen S, Forestieri AM, Monforte MT. 1996. Biological effects of hesperidin, a Citrus flavonoid. Part 3. Antihypertensive and diuretic activity in the rat. *Farmaco* **51**: 219–221.
- Godeberge P. 1995. Daflon-500 mg: International assessment of therapeutic interest for haemorrhoids. *Drugs Today* **31**: 57–62.
- Guillot B, Guihou JJ, De Champvallins M. 1989. A long term treatment with a venotropic drug: Results on efficacy and safety of Daflon-500 mg in chronic venous insufficiency. *Int Angiol* **8**: 67–71.
- Hahn L. 1952. Inhibitors of hyaluronidase. *Nature* **170**: 282–283.
- Harborne JB. 1994. In *The Flavonoids—Advances in Research Since 1986*. Chapman and Hall: London; 340.
- Hardigree AA, Epler JL. 1978. Comparative mutagenesis of plant flavonoids in microbial systems. *Mutat Res* **58**: 231–239.
- Hasanoglu A, Ara C, Ozen S, Kali K, Ertas E. 2001. Efficacy of micronised flavonoid fraction in healing of clean and infected wounds. *Int J Angiol* **10**: 41–44.
- Higby RH. 1941. The chemical nature of hesperidin and its experimental medicinal use as a source of vitamin-P: A review. *J Am Pharm Assoc Sci Ed* **30**: 629.
- Ho YH, Foo CL, Seow-Choen F. 1995. Prospective randomised controlled trial of a micronised flavonoid fraction to reduce bleeding after haemorrhoidectomy. *Br J Surg* **82**: 1034–1035.
- Höerhammer L, Wagner H. 1962. Citrus flavonoids. *Deut Apotheker Ztg* **102**: 759–765.
- Horowitz RM. 1961. *The Orange*. University of California Press: Berkeley, CA.
- Horowitz RM, Gentili B. 1963. Flavonoids of Citrus- VI. *Tetrahedron* **19**: 773–782.
- Hu CQ, Chen K, Shi Q, Kilkuskie RE, Cheng YC, Lee KH. 1994. Anti-AIDS agents. *J Nat Prod* **57**: 42–51.
- Iio M, Moriyama A, Matsumoto Y, Takaki N, Fukumoto M.

1985. Inhibition of xanthine oxidase by flavonoids. *Agr Biol Chem* **49**: 2173–2176.
- lio M Ushijima K, Fujita M, Matsumura M, Miyatake S. 1980. Effects of flavonoids on alkaline phosphatase. *Nippon Noge Kagaku Kaishi* **54**: 171–175.
- lio M Yoshioka A, Imayoshi Y, Koriyama C, Moriyama A. 1984. Effect of flavonoids on alpha-glucosidase and beta-fructosidase from yeast. *Agr Biol Chem* **48**: 1559–1563.
- Islam SKN, Ahsan M. 1997. Biological activities of the secondary metabolites isolated from *Zieria smithi* and *Zanthoxylum elephantiasis* on micro-organisms and brine shrimps. *Phytother Res* **11**: 64–66.
- Jackson H. 1959. Antifertility substances. *Pharmacol Rev* **11**: 135–172.
- Jean T, Bodinier MC. 1994. Mediators involved in inflammation: Effects of Daflon-500mg on their release. *Angiology* **45**: 554–559.
- Jeong HJ, Shin YG, Kim IH, Pezzuto JM. 1999. Inhibition of aromatase activity by flavonoids. *Arch Pharm Res* **22**: 309–312.
- Jovanovic SV, Steenken S, Tosic M, Marjanovic B, Simic MG. 1994. Flavonoids as anti-oxidants. *J Am Chem Soc* **116**: 4846–4851.
- Joyce CL, Mack SR, Anderson RA, Zaneveld LJD. 1986. Effect of hyaluronidase, beta-glucuronidase and beta-N-acetyl glucosaminidase inhibitors on sperm penetration of the mouse oocytes. *Biol Reprod* **35**: 336–346.
- Joyce CL, Zaneveld LJD. 1985. Vaginal contraceptive activity of hyaluronidase and cyclooxygenase inhibitors in the rabbit. *Fertil Steril* **44**: 426–428.
- Jurd L. 1962. Spectral properties of flavonoid compounds. In *The Chemistry of Flavonoid Compounds*, Geissman TA (ed). Macmillan: New York; 107–155.
- Jurisson S. 1973. Flavonoid substances of *Capsella Bursa Pastoris*. *Farmatsiya (Moscow)* **22**: 34–38.
- Kaito T, Yoshida T, Sagara K. 1979. Fluorimetric determination of hesperidin. *J Pharm Sci Technol Japan* **39**: 210–215.
- Kanes K, Tisserat B, Berhow M, Vandercook C. 1993. Phenolic composition of various tissues of Rutaceae species. *Phytochemistry* **324**: 967–974.
- Kaul TN, Middleton EJ, Ogra PL. 1985. Antiviral effects of flavonoids on human viruses. *J Med Virol* **15**: 71–79.
- Kawabe M, Tamano S, Shibata MA, Hirose M, Fukushima S, Ito N. 1993. Subchronic toxicity study of methyl hesperidin in mice. *Tox Lett* **69**: 37–44.
- Kawaguchi K, Kikuchi S, Takayanagi K, Yoshikawa T, Kumazawa Y. 1999. Colony stimulating factor inducing activity of hesperidin. *Planta Med* **65**: 365–366.
- Kawaguchi K, Mizuno T, Aida K, Uchino K. 1997. Hesperidin as an inhibitor of lipases from porcine pancreas and *Pseudomonas*. *Biosci Biotech Biochem* **61**: 102–104.
- Kim CJ, Cho SK. 1991. Pharmacological activities of flavonoids (II)—Structure activity relationships of flavonoids in immunosuppression. *Arch Pharmacol Res* **14**: 147–159.
- Kim CJ, Chung JM. 1990. Pharmacological activities of flavonoids (I)—Relationships of chemical structure of flavonoids and their inhibitory activity on hypersensitivities. *Yakhak Hoe Chi* **34**: 348–364.
- Kim CJ, Su HK, Joo JH, Cho SK. 1990. Pharmacological activities of flavonoids. (II) Relationships of anti-inflammatory and anti-granulomatous actions. *Yakhak Hoe Chi* **34**: 407–414.
- Kim DH, Jung EA, Sohng IS, Han JA, Kim TH, Han MJ. 1998. Intestinal bacterial metabolism of flavonoids and its relation to some biological activities. *Arch Pharm Res* **21**: 17–23.
- Kim DH, Song MJ, Bae EA, Han MJ. 2000. Inhibitory effect of herbal medicines on rotavirus infectivity. *Biol Pharm Bull* **23**: 356–358.
- Kim M, Kometani T, Okada S, Shimizu M. 1999. Permeation of hesperidin glycosides across Caco-2 cell monolayers via the paracellular pathway. *Biosci Biotechnol Biochem* **63**: 2183–2188.
- King FE, Robertson A. 1931. Natural glucosides, Part 3. *J Chem soc (III)*: 1704–1709.
- Kokkalou E, Kapetanidis I. 1988. Flavonoids of the arial parts of *Acinos suaveolens*. *Pharm Acta Helv* **636**: 170–173.
- Kometani T, Nishimura T, Nakae T, Takii H, Okada S. 1996. Synthesis of neohesperidin glycosides and naringin glycosides by cyclodextrin gluconotransferase from an alkalophilic *Bacillus* Species. *Biosci Biotech Biochem* **60**: 645–649.
- Kometani T, Terada Y, Nishimura T, Takii H, Okada S. 1994. Transglycosylation of hesperidin by cyclodextrin gluconotransferase from an alkalophilic *Bacillus* species in alkaline pH and properties of hesperidin glycosides. *Biosci Biotech Biochem* **58**: 1190–1194.
- Korthuis RJ, Gute DC. 1999. Adhesion molecule expression in post ischemic microvascular dysfunction: Activity of a micronised purified flavonoid fraction. *J Vasc Res* **36**: 15–23.
- Koyunko H, Berkada B, Baykut F *et al.* 1999. Preventive effect of hesperidin against inflammation in CD-1 mouse skin caused by tumour promoter. *Anticancer Res* **19**: 3237–3241.
- Krollicki Z, Lamer-Zarawska E. 1984. Investigation of antifungal effect of flavonoids. Part 1. *Herb Pol* **30**: 53–57.
- Kroyer G. 1986. Über die antioxidative Aktivität von Zitrusfruchtschalen. *Z Ernährungswiss* **25**: 63–69.
- Kubo M, Matsuda H. 1992a. Anti-allergic agents containing hesperidin. *Patent Japan Kokai Tokkyo Koho* 64295, 428.
- Kubo M, Yanagisawa T, Sasaki H, Kaizu R. 1992b. Lipooxygenase inhibitors as pharmaceuticals from *Schizonepeta tenuifolia*. *Patent Japan Kokai Tokkyo Koho* 04 208, 221.
- Kuntic V, Blagojevic S, Malesev D, Radovic Z. 1999. Spectrophotometric investigation of the Cu(II)-hesperidin complex. *Pharmazie* **54**: 548.
- Kuntic V, Kosanic M, Malesev D, Radovic Z, Mico U. 1998. Spectrophotometric investigation of the uranyl(II)-hesperidin complex in 70% methanol. *J Serb Chem Soc* **63**: 565–572.
- Kuppusamy UR, Das NP. 1992. Effects of flavonoids on cyclic AMP phosphodiesterase and lipid mobilisation in rat adipocytes. *Biochem Pharmacol* **44**: 1307–1315.
- Lee SH, Jeong TS, Park YB, Kwon YK, Choi MS, Bok SH. 1999a. Hypocholesterolemic effect of hesperetin mediated by inhibition of 3-hydroxy-3-methylglutaryl coenzyme A reductase and acyl coenzyme A: cholesterol acyltransferase in rats fed high-cholesterol diet. *Nutr Res* **19**: 1245–1258.
- Lee JH, Kim YS, Lee CK, Lee HK, Han SS. 1999b. Antiviral activity of some flavonoids on Herpes-simplex virus. *Korean Pharmacog* **30**: 34–39.
- Limasset B, Le Doucen C, Dore JC, Ojasoo T, Damon M, Crastes de Paulet A. 1993. Effects of flavonoids on the release of reactive oxygen species by stimulated human neutrophils. *Biochem Pharmacol* **46**: 1257–1271.
- Loguercio C, D'Argenio G, Delle cave M *et al.* 1996. Direct evidence of oxidative damage in acute and chronic phases of experimental colitis in rats. *Dig Dis Sci* **41**: 1204–1211.
- Lonchamp M, Guardiola B, Sicot N, Bertrand N, Perdrix L, Duhault J. 1989. Protective effect of a purified flavonoid fraction against reactive oxygen radicals. *Arzneimittelforsch* **39**: 882–885.
- Mabry TJ, Markham KR, Thomas MB. 1970. In *The NMR Spectra of Flavonoids*. Springer-Verlag: New York.
- Malesev D, Radovic Z, Kuntic K, Kosanic M. 1997. Spectrophotometric determination of hesperidin by Al (III)—hesperidin complex in water—methanol solution. *Anal Lett* **30**: 917–926.
- Malterud KE, Rydland KM. 2000. Inhibitors of 15-lipoxygenase from orange peel. *J Agric Food Chem* **48**: 5576–5580.
- Markham KR, Ternai B. 1976. <sup>13</sup>C NMR of flavonoids-II. *Tetrahedron* **32**: 2607–2612.
- Martin GJ, Beiler JM. 1952. Effect of phosphorylated hesperidin, a hyaluronidase inhibitor on fertility in the rat. *Science* **115**: 402.
- Matsuda H, Yano M, Kubo M, Iinuma M, Oyama M, Mizuno M. 1991. Pharmacological study of Citrus fruits (II) Anti-allergic effect of fruit of *Citrus unshiu*. *Yakugaku Zasshi* **111**: 193–198.
- McGregor L, Bellangeon M, Chignier E, Lerond L, Rousselle C, McGregor JL. 1999. Effect of a purified micronised fraction on *in vivo* platelet functions in the rat. *Thromb Res* **94**: 235–240.
- Melzig MF, Loose R, Schonherr G. 1997. Effects of flavonoids on daunomycin-induced toxicity in cultivated endothelial cells. *Pharmazie* **52**: 793–796.

- Meyer OC. 1994. Safety and security of Daflon-500 mg in venous insufficiency and in haemorrhoids disease. *Angiology* **45**: 579–584.
- Middleton EJ. 1984. The flavonoids. *Trends Pharmacol Sci* **8**: 335–338.
- Miller NJ, Rice-Evans CA. 1997. The relative contributions of ascorbic acid and phenolic antioxidants to the total antioxidant activity of orange and apple fruit juices and blackcurrant drink. *Food Chem* **60**: 331–337.
- Mitsunaga Y, Takanaga H, Matsuo H *et al.* 2000. Effect of bioflavonoids on vincristine transport across blood-brain barrier. *Eur J Pharmacol* **395**: 193–201.
- Miyake Y, Yamamoto K, Morimitsu Y, Osawa T. 1997. Isolation of glucosylflavone from lemon peel and anti-oxidative activity of flavonoid compounds in lemon fruit. *J Agric Food Chem* **45**: 4619–4623.
- Monforte MT, Trovato A, Kirjavainen S, Forestieri AM, Lo Curto RB. 1995. Biological effects of hesperidin, a Citrus flavonoid. Part 2. Hypolipidemic activity on experimental hypercholesterolemia in the rat. *Farmacologia* **50**: 595–599.
- Morii S. 1939. Research for vitamin-P. *J Biochem-Tokyo* **29**: 487–501.
- Morita O, Sasaki H, Sato S. 1992. Calcium antagonists containing phenols. *Patent Japan Kokai Tokkyo Koho*, 04 243, 822.
- Mortimer PS. 1997. Therapy approaches for lymphedema. *Angiology* **48**: 87–91.
- Mouly P, Gaydou EM, Auffray A. 1998. Simultaneous separation of flavanone glycosides and polymethoxylated flavones in citrus juices using liquid chromatography. *J Chromatogr A* **800**: 171–179.
- Mouly P, Gaydou EM, Estienne J. 1993. Column liquid chromatographic determination of flavanone glycosides in Citrus. Application to grapefruit and sour orange juice adulterations. *J Chromatogr* **634**: 129–134.
- Mucsi I, Pragai BM. 1985. Inhibition of virus multiplication and alteration of cyclic AMP level in cell cultures by flavonoids. *Experientia* **41**: 930–931.
- Niemann GJ, Koerselman-Kooy JW. 1977. Phenolics from Larix needles. *Planta Med* **31**: 297–301.
- Nieto JL, Gutierrez AM. 1986. <sup>1</sup>H NMR spectra at 360 MHz of diosmin and hesperidin in DMSO solution. *Spectro Lett* **19**: 427–434.
- Obendorf D, Reichart E. 1995. Determination of hesperidin by cathode stripping voltammetry in orange juice and helopyrin, a phytopharmaceutical preparation. *Electroanalysis* **7**: 1075–1081.
- Ono K, Nakane H, Fukushima M, Chermann JC, Barre-Sinoussi F. 1990. Differential inhibitory effects of various flavonoids on the activities of reverse transcriptase and cellular DNA and RNA polymerases. *Eur J Biochem* **190**: 469–476.
- Panasiak W, Wleklik M, Oraczewska A, Luczak M. 1989. Influence of flavonoids on combined experimental infections with EMC virus and *Staphylococcus aureus* in mice. *Acta Microbiol Pol* **38**: 185–188.
- Pawlowska L. 1980. Flavonoids of *B. pendula* Roth and *B. obscura* Kot leaves. *Acta Soc Bot Pol* **493**: 281–296.
- Perez-Ruiz T, Martinez-Lozano C, Tomas V, Fenoll J. 1999. Spectrofluorimetric determination of hesperidin by manual and flow injection methods. *Fresenius J Anal Chem* **364**: 279–283.
- Pizzorno Jr JE, Murray MT. 1999. In *Textbook of natural medicine*. Churchill Livingstone: Edinburgh; 79.
- Preston RK, Avakian S, Beiler JM, Moss JN, Martin GJ. 1953. *In-vivo* and *in-vitro* inhibition of hyaluronidase by organic phosphates. *Exp Med Surg* **11**: 1–8.
- Radovic Z, Malesev D, Jelick-Stankov M. 1996. Spectrophotometric determination of hesperidin by Zr(IV)-hesperidin complexation. *Pharmazie* **51**: 604.
- Ratty AK, Das NP. 1988. Effects of flavonoids on non-enzymatic lipid peroxidation: structure activity relationship. *Biochem Med Metab Biol* **39**: 69–79.
- Reynolds JEF. 1996. In *Martindale, The Extra Pharmacopoea*. The Royal Pharmaceutical society: London.
- Robbins RC. 1967. Effect of flavonoids on survival time of rats fed thrombogenic or atherogenic regimens. *J Atheroscler Res* **7**: 3–10.
- Robbins RC. 1971. Effects of phenyl-benzo-gamma-pyrone derivatives (flavonoids) on blood cell aggregation: Basis for a concept of mode of action. *Clin Chem* **17**: 433–437.
- Ross SA, Ziska DS, Zhao K, Elshohly MA. 2000. Variance of common flavonoids by brand of grapefruit juice. *Fitoterapia* **71**: 154–161.
- Rouseff RL. 1988. Liquid chromatographic determination of naringin and neohesperidin as a detector of grapefruit juice and orange juice. *J Assoc Anal Chem* **71**: 798–802.
- Rouseff RL, Dettweiler GR, Swaine RM, Naim M, Zehavi U. 1992. Solid-phase extraction and HPLC determination of 4-vinyl guaicol and its precursor, ferulic acid, in orange juice. *J Chromatogr Sci* **30**: 383–387.
- Saija A, Tomaino A, Trombetta D, Giacchi M, De Pasquale A, Bonina F. 1998. Influence of different penetration enhancers on *in vitro* skin permeation and *in vivo* photoprotective effects of flavonoids. *Int J Pharm* **175**: 85–94.
- Scarborough H. 1940. Deficiency of vitamin-C and vitamin-P in man. *Lancet* **6117**: 644–647.
- Scarborough H, Bacharach L. 1949. Vitamins. In *"Vitamins and Hormones"*, Harris RS, Thimann KV (eds). Academic Press: New York; 1–55.
- Schoemaker JH, Schoemaker MT, Zijlstra H, Vander Horst FA. 1995. Treatment of erythropoietic protoporphyria with hydroxyethylrutinosides. *Dermatology* **191**: 36–38.
- Sheu SJ, Lu CF. 1995. Determination of eight constituents of Hsia-cheng-chi-tang by high performance liquid chromatography. *J Chromatogr A* **704**: 518–523.
- Sieve BJ. 1952. A new antifertility factor. *Science* **116**: 373–385.
- Smith CJ. 1964. Non hormonal control of vasomotor flushing in menopausal patients. *Chic Med* **67**: 193–195.
- Son HS, Kim HS, Ju JS. 1991. Effects of rutin and hesperidin on total cholesterol concentration, transaminase and alkaline phosphatase activity in carbon tetrachloride treated rats. *Hanguk Nonghwa Hakhoe Chi* **34**: 318–326.
- Steincke K. 1956. The hyaluronidase inhibiting and permeability reducing properties of hesperidin in isolated connective tissue membranes. *Acta Pharmacol Toxicol* **12**: 126–136.
- Struckmann JR, Nicolaides AN. 1994. Flavonoids: A review of the pharmacology and therapeutic efficacy of Daflon-500mg in patients with chronic venous insufficiency and related disorders. *Angiologon* **45**: 419–428.
- Suarez J, Herrera MD, Marhuenda E. 1996. Hesperidin and neohesperidin dihydrochalcone on different experimental models of induced gastric ulcer. *Phytother Res* **10**: 616–618.
- Suarez J, Herrera MD, Marhuenda E. 1998. *In vitro* scavenger and antioxidant properties of hesperidin and neohesperidin dihydrochalcone. *Phytomedicine* **5**: 469–473.
- Szent-Gyorgi A. 1938. Preparation of citrin. *Physiol Chem* **225**: 126–131.
- Tanaka T, Makita H, Kawabata K, Mori H. 1997a. Chemoprevention of azoxymethane-induced rat colon carcinogenesis by the naturally occurring flavonoids, diosmin and hesperidin. *Carcinogenesis* **18**: 957–965.
- Tanaka T, Makita H, Kawabata K, Mori H, Kakumoto M. 1997b. Modulation of N-methyl-N-aminonitrosamine induced tumorigenesis by dietary feeding of diosmin and hesperidin, alone and in combination. *Carcinogenesis* **18**: 761–769.
- Tanaka T, Makita H, Ohnishi M, Hirose Y. 1994. Chemoprevention of 4-nitroquinoline-1-oxide-induced oral carcinogenesis by dietary curcumin and hesperidin: Comparison with protective effects of beta carotene. *Cancer Res* **54**: 4653–4659.
- Tanaka T, Makita H, Ohnishi M *et al.* 1997c. Chemoprevention of 4-nitroquinoline-1-oxide-induced oral carcinogenesis in rats by flavonoids diosmin and hesperidin, each alone and in combination. *Cancer Res* **57**: 246–252.
- Tsuchiya Y, Shimizu M, Hiyama Y, Itoh K. 1985. Antiviral activity of natural occurring flavonoids *in vitro*. *Chem Pharm Bull* **33**: 3881–3886.
- Varma SD, Kinoshita JH. 1976. Inhibition of lens aldose reductase by flavonoids—Their possible role in the prevention of diabetic cataracts. *Biochem Pharmacol* **25**: 2505.
- Volikakis GJ, Efstathiou CE. 2000. Determination of rutin and other flavonoids by flow-injection/adsorptive stripping

- voltammetry using nujol-graphite and diphenylether-graphite paste electrodes. *Talanta* **51**: 775–785.
- Wacker VA, Eilmes HG. 1975. Virus inhibition by hesperidin. *Naturwissenschaften* **62**: 301.
- Wacker VA, Eilmes HG. 1978. Antiviral activity of plant components. *Arzneimittelforsch* **28**: 347–350.
- Wang PF, Zheng RL. 1992. Inhibition of the autooxidation of linoleic acid by flavonoids in micelles. *Chem Phys Lipids* **63**: 37–40.
- Wang JZ, Chen DY, Wang J. 1994. Determination of hesperidin in *Pericarpium Citri-Reticulatae* by HPLC. *Chung Kuo Chung Yao Tsa Chih* **19**: 424–425.
- Williams RT. 1964. Metabolism of phenolics in animals. In *"Biochemistry of Phenolic Compounds"*. Harborne JB (ed.). Academic Press: New York; 240–241.
- Yamauchi Y, Ueda J, Ohsawa K. 1996. A simultaneous determination of various main components in oriental pharmaceutical decoction "Heii-san" by ion-pair high-performance liquid chromatography. *Yakugaku Zasshi* **116**: 776–782.
- Yang M, Tanaka T, Hirose Y, Deguchi T, Mori H, Kawada Y. 1997. Chemopreventive effects of diosmin and hesperidin on *N*-butyl-*N*-(4-hydroxybutyl) nitrosamine induced urinary bladder carcinogenesis in male ICR mice. *Int J Can* **73**: 719–724.
- Yuting C, Rongliang Z, Zhongjian J, Yong J. 1990. Flavonoids as superoxide scavengers and antioxidants. *Free Rad Biol Med* **9**: 19–21.
- Zaragoza F, Fdez-Corbeira P, Iglesias I, Benedi J. 1986. New natural inhibitors of platelet aggregation *in vivo*. Part I. Citroflavonoids and hesperidin. *An Real Acad Farm* **52**: 497–504.
- Zaragoza F, Iglesias I, Benedi J, Folez Corbeira P. 1985. Effect of the citroflavonoids on platelet aggregation. *Fitoterapia* **56**: 343–347.
- Zhao WH, Qin GW, Xu RS *et al.*, 1999. Constituents from the roots of *Acanthopanax setchuenensis*. *Fitoterapia* **70**: 529–531.
- Zoulis NE, Efstathiou CE. 1996. Preconcentration at a carbon-paste electrode and determination by adsorptive-stripping voltammetry of rutin and other flavonoids. *Anal Chim Acta* **320**: 255–261.