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Susceptibility of never-dried and freeze-dried bacterial cellulose towards esterification with organic acid

Koon-Yang Lee · Alexander Bismarck

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Abstract The susceptibility of (1) never-dried and (2) freeze-dried bacterial cellulose (BC) towards organic acid esterification is reported in this work. When never-dried BC (BC which was solvent exchanged from water through methanol into pyridine) was modified with hexanoic acid, it was found that the degree of substitution (DS) was significantly lower than that of hexanoic acid modified freeze-dried BC. The crystallinity of freeze-dried BC hexanoate was found to be significantly lower compared to neat BC and never-dried BC hexanoate. This result, along with the high DS indicates that significant bulk modification occurred during the esterification of freeze-dried BC. Such results were not observed for never-dried BC hexanoate. All these evidence point towards to fact that freeze-dried BC was more susceptible to organic acid esterification compared to never-dried BC. A few hypotheses were explored to explain the observed behaviour and further investigated to elucidate our observation; the effect of residual water in cellulose, the accessibility of hydroxyl groups and the crystal structure of never-dried and freeze-dried BC on the susceptibility of cellulose fibrils to esterification, respectively. However, the investigation of these

hypotheses raised more questions and we are still left with the main question; why do BC nanofibres behave differently when modifying freeze-dried BC or never-dried BC?

Keywords Esterification · Organic acids · Bacterial cellulose · Freeze-dried state · Never-dried state

Introduction

Cellulose is used in numerous industries; the pulp and paper industry (Sczostak 2009), the pharmaceutical and cosmetic industry (Klemm et al. 2005) and more recently as reinforcement for polymers (Eichhorn et al. 2010; Habibi et al. 2010; Gardner et al. 2008; Blaker et al. 2011). Due to the rapid advancement and interest in nanotechnology, significant effort has been poured into the utilisation and isolation of nano-sized cellulose to produce nanocellulose for various applications. Nanocellulose can be obtained via two approaches; top-down or bottom-up. The top-down approach involves the disintegration of plant cellulose, such as wood fibres, using high shear forces (Turbak et al. 1983; Herrick et al. 1983). The bottom-up approach, on the other hand, utilises the biosynthesis of cellulose by bacteria, such as from the *Acetobacter* species (Brown 1886).

Although nanocellulose can be produced using the two approaches, the quality of bacterial cellulose (BC)

K.-Y. Lee · A. Bismarck (✉)
Polymer and Composites Engineering (PaCE) Group,
Department of Chemical Engineering, Imperial College
London, South Kensington Campus,
SW7 2AZ London, UK
e-mail: a.bismarck@imperial.ac.uk

was found to be superior over wood-derived nanocellulose (Klemm et al. 2009). This can be attributed to the high purity of BC. Additionally, no further processing is required to obtain nano-sized cellulose. In addition to this, BC possesses a high degree of crystallinity of up to 90% (Czaja et al. 2004). Hsieh et al. (2008) determined the mechanical properties of a single BC nanofibre using Raman spectroscopy. The authors estimated that a single BC nanofibre possesses a Young's modulus of 114 GPa. Cellulose crystals possess a theoretical Young's modulus of between 130 and 160 GPa depending on the crystal form (Eichhorn and Davies 2006; Reiling and Brickmann 1995). These properties are highly favourable for the utilisation of nanocellulose as filler for polymers. However, the hydrophilic nature of cellulose often resulted in poor interfacial adhesion between the cellulose and hydrophobic polymer matrices (Pommet et al. 2008; Siro and Plackett 2010), such as polypropylene and polylactic acid. Surface modification of nanocellulose is often performed to improve the compatibility and, therefore, the stress transfer between the cellulose and the matrix to produce nanocomposites with improvements in both modulus and strength (Lee et al. 2009; de Menezes et al. 2009; Ly et al. 2010; Blaker et al. 2011).

Nanocellulose modification is typically conducted using acetic anhydride (Tingaut et al. 2010). BC, in particular, had been modified by introduction of acetate functional groups, i.e. by reaction with acetic acid, in toluene using perchloric acid as catalyst to produce BC with hydrophobic characteristics (Kim et al. 2002; Ifuku et al. 2007). However, these modifications are limited to the synthesis of short chain cellulose esters of alkanic acids. In order to modify nanocellulose with long-chain aliphatic acids, a two-step reaction has to be used to first produce long chain aliphatic anhydride moieties using trifluoroacetic anhydride, followed by cellulose modification (Heinze et al. 2006). The use of pyridine along with long chain aliphatic carboxylic acids and acid chlorides allowed us to activate the long chain carboxylic acids and produce nanocellulose modified with long chain aliphatic moieties, such as cellulose hexanoate or dodecanoate (Lee et al. 2009).

The chemical modification of BC presents an interesting scenario. Since BC is produced by the biosynthesis of bacteria in an aqueous culture medium, BC stays hydrated and so is never-dried. We have previously reported the successful surface-only

modification of never-dried BC with long chain organic acids (Lee et al. 2009, 2011). The procedure we followed involved the solvent exchange of never-dried BC from water through methanol into pyridine. Attempts were made to reduce the laborious solvent exchange step by freeze-drying BC from water and disperse freeze-dried BC, which does not hornify, directly into pyridine to carry out the same esterification with organic acids. To our surprise we observed that freeze-dried BC is much more susceptible towards esterification with organic acid compared to never-dried BC. Possible hypotheses of the observed susceptibility of freeze-dried BC towards organic esterification are presented and have been further investigated in this study.

Experimental procedure

Materials and methods

Methanol (GPR, purity $\geq 99\%$), ethanol (GPR, purity $\geq 99.7\%$), benzene (analaR NORAMPUR, purity $\geq 99.9\%$) and pyridine (analaR NORAMPUR, purity $\geq 99.7\%$) were purchased from VWR. Hexanoic acid (Aldrich, purity $\geq 99\%$), *p*-toulenesulfonyl chloride (Aldrich, purity $\geq 99\%$), dimethyl carbonate (Aldrich reagent plus, purity $\geq 99\%$), deuterium oxide (Aldrich, purity ≥ 99.99 atom% D), cellulose acetate (Aldrich, 39.7 wt% acetyl content) and cellulose triacetate (Aldrich, 43–49 wt% acetyl content) were purchased from Sigma-Aldrich. Sodium hydroxide (purum grade, pellets) was purchased from Acros Organics. All the materials were used as received without further purification. BC was extracted and purified from commercially available *Nata-de-coco* (CHAOKOH gel in syrup, Ampol Food Processing Ltd, Nakorn Pathom, Thailand) following previously described work (Blaker et al. 2009).

Hexanoic acid esterification of BC

In order to modify never-dried BC, which was always kept in water, 2 g (on dry weight basis) of purified BC was solvent exchanged from water through methanol ($3 \times 600 \text{ cm}^3$) into pyridine ($2 \times 600 \text{ cm}^3$) to ensure the complete removal of water and methanol. The mixture was homogenised using a homogeniser (Polytron PT 10-35 GT, Kinematica, CH) at 20,000 rpm for at least 1 min during each solvent exchange step to

completely disperse the BC in the solvent. BC was retained through centrifugation at 14,000g for 15 min. The excess solvent was decanted prior to re-dispersion in the subsequent solvent. This homogenisation/centrifugation step was repeated three times. After the second solvent exchange step into pyridine, another solvent exchange step was performed to adjust the final concentration of BC in pyridine to 0.5% (g mL⁻¹). This BC-pyridine mixture was then poured into a 1 L 3-neck round bottom flask and stirred using a magnetic stirrer. 92 g of *p*-toluenesulfonyl chloride was added into this BC-pyridine mixture and hexanoic acid was added at an equimolar concentration relative to *p*-toluenesulfonyl chloride into the same reaction vessel. The reaction was conducted under nitrogen flow to create an inert atmosphere for 2 h at 50 °C. After 2 h, the reaction was quenched with 1.5 L of ethanol. The product was washed three times with 800 mL of ethanol using the previously described homogenisation-centrifugation steps.

In a separate experiment, never-dried BC was homogenised in water at a concentration of 0.4% (g mL⁻¹), flash frozen in a Petri dish and subsequently freeze-dried (Edwards Modulyo freeze dryer, West Sussex, UK). This BC is termed freeze-dried BC throughout this study. 2 g of freeze-dried BC was dispersed into 400 mL (0.5% (g mL⁻¹)) of pyridine directly in a 1 L 3-neck round bottom flask and the reaction was conducted as previously described. After 2 h, the hexanoic acid modified freeze-dried BC was purified following the procedure described above. In order to characterise the hexanoic acid modified wet BC and freeze-dried BC, they were solvent exchanged from ethanol through water into dimethyl carbonate at a concentration of 0.4% (g mL⁻¹), flash frozen in a Petri dish and subsequently freeze-dried from dimethyl carbonate to obtain dry modified samples that were further characterised. The never-dried BC hexanoate and freeze-dried BC hexanoate are termed C₆-NDBC and C₆-FDBC, respectively.

Characterisation of the modified (never-dried and freeze-dried) BC

Attenuated total reflection infrared spectroscopy (ATR-IR)

ATR-IR spectra were recorded using a Spectrum One FTIR-spectrometer (Perkin Elmer, Massachusetts, USA). The spectra were collected at a resolution of

2 cm⁻¹, in the range of 600 and 4,000 cm⁻¹. A total of 16 scans was used to collect each spectrum.

Degree of substitution of modified BC

The degree of substitution (DS) of C₆-NDBC and C₆-FDBC was determined using the procedure developed by Sassi and Chanzy (1995); the DS was calculated based on the ATR-IR spectra using the absorption band of the pyranose ring as an internal standard at a wavenumber of 1,050 cm⁻¹. A calibration master curve was established, relating the ratio of the absorbance intensities of the carbonyl bonds (1,750 cm⁻¹) and pyranose ring's to the known DS for cellulose acetates and cellulose triacetates. By comparing the ratio of the intensities between 1,750 and 1,050 cm⁻¹ of C₆-NDBC and C₆-FDBC to the calibration master curve, the DS of the modified BC was obtained.

Crystallography of BC hexanoate

The X-ray diffraction (XRD) pattern of modified BC was obtained using a PANalytical X'Pert PRO X-ray diffractometer (PANalytical Ltd, Cambridge, UK). The diffractograms were taken from 2θ = 10° to 45° at a step size of 0.02° using a Ni filtered Cu Kα₁ (1.541 Å) as the X-ray source. Segal's method (1959) was used to calculate the crystallinity of (modified) BC:

$$\chi_c = \frac{I_{002} - I_{am}}{I_{002}} \times 100\% \quad (1)$$

where χ_c is the crystallinity of the cellulose, I_{002} and I_{am} are the intensity of the 002 (2θ = 22.5°) and amorphous (2θ = 18°) reflections, respectively. This is an empirical equation for estimating the degree of crystallinity of pure cellulose materials.

Scanning electron microscopy (SEM)

SEM was performed using a high-resolution field emission gun scanning electron microscope (LEO Gemini 1525 FEG-SEM, Oberkochen, Germany). It was used to characterise the morphology of BC nanofibres after esterification with hexanoic acid. The accelerating voltage used was 5 kV. Prior to SEM, the samples were mounted to SEM stubs using carbon tabs and coated with Cr (K550 sputter coater, Emitech Ltd, Ashford, Kent, UK) for 1 min at 75 mA.

Characterisation of neat never-dried and freeze-dried BC

In order to mimic the behaviour of never-dried BC in pyridine prior to the hexanoic acid esterification reaction, the never-dried BC was solvent exchanged from water through methanol into benzene using the previously described concentration and homogenisation-centrifugation steps. Once the never-dried BC was dispersed in benzene, it was subsequently freeze-dried to obtain a dry sample hopefully representative of never-dried BC in pyridine. Benzene was chosen as it is structurally related to pyridine and because of its ability to be freeze-dried.

Determining the residual water content of never-dried and freeze-dried BC

Esterification of cellulose with carboxylic acids is a reversible reaction and the presence of water will affect the conversion of cellulose to cellulose esters. The residual water content of freeze-dried BC was determined using dynamic vapour sorption (DVS-Advantage, Surface Measurement Systems Ltd, Alpertown, UK). 30 mg of freeze-dried BC was loaded into the chamber of DVS held at 0% relative humidity (RH) for 5 h. During this period, the mass change was measured in situ inside the chamber. The residual water content was obtained from the difference between the initial and final mass of the sample.

The residual water content in never-dried BC after successive solvent exchange steps was estimated from the solvent exchange efficiency. To determine the solvent exchange efficiency, 2 g (on a dry weight basis) of never-dried BC was dispersed in 600 cm³ of methanol following the previously described homogenisation step and centrifuged at 14,000g for 15 min after which the supernatant was removed. The water content in this supernatant was determined using Karl-Fischer titration (DL32 Coulometric Titrator, Mettler Toledo, Leicester, UK). The homogenisation-centrifugation-titration step was repeated 5 times (the number of solvent exchange steps in this work). The efficiency of water removal for solvent exchange was calculated using:

$$R_w = \frac{W_{i+1} - W_i}{W_i} \times 100\% \quad (2)$$

where R_w , W_{i+1} and W_i are the water removal efficiency, the water content after the solvent exchange step $i + 1$ and step i , respectively.

Determining the hydroxyl group availability of never-dried and freeze-dried BC

In order to study the difference between the hydroxyl group availability for the esterification reaction of the different starting BC, a method based on hydrogen/deuterium (H/D) exchange was utilised following our previous work (Lee et al. 2011). This measurement was carried out using dynamic vapour sorption (DVS-Advantage, Surface Measurement Systems Ltd, Alpertown, UK). A sample mass of approximately 30 mg of BC was placed in the sample pan and the sample chamber was pre-conditioned at 0% RH of deuterium oxide (D₂O) for 5 h at room temperature to remove adsorbed water molecules. The RH of D₂O was increased to 90% for 2 h to allow for the adsorption of D₂O and for H/D exchange with accessible hydroxyl groups to occur. The RH was then reduced to 0% for 2 h to allow D₂O molecules to desorb. This cycle was repeated 10 times such that the H/D can occur on all accessible hydroxyl groups on BC. A short adsorption-desorption cycle was utilised to avoid bulk sorption of D₂O into BC, as only the accessible hydroxyl groups were of interest. After 10 adsorption-desorption cycles, the sample was post-conditioned at 0% RH for 5 h to remove any residual adsorbed D₂O molecules. As deuterium is one neutron heavier than hydrogen, the mass increase after post-conditioning of BC was measured in situ by an ultra-sensitive microbalance and the amount of accessible hydroxyl groups was back calculated from this mass increase.

Specific surface area measurement of never-dried and freeze-dried BC

Nitrogen adsorption/desorption isotherms were performed to determine the specific surface area of never-dried and freeze-dried BC using a surface area and porosity analyser (TriStar 3000, Micrometrics Ltd, Dunstable, UK). The specific surface area was calculated using the Brunauer-Emmett-Teller (BET) equation. Prior to the measurement, the samples were degassed at 80 °C overnight to remove adsorbed water molecules.

Allomorphs of never-dried and freeze-dried BC

The different allomorphs of crystalline cellulose were evaluated using an empirical mathematical function developed by Wada et al. (2001). This mathematical

function is supposed to discriminate cellulose I_α , which is triclinic, from cellulose I_β , which is monoclinic, based on the XRD pattern of the cellulose of interest. The discrimination factor Z is related to the d-spacings of the cellulose via:

$$Z = 1693 \times d_1 - 902 \times d_2 - 549 \quad (3)$$

where $Z > 0$ represents I_α -rich cellulose and $Z < 0$ represents I_β -rich cellulose. d_1 (nm) and d_2 (nm) represent the d-spacing of the reflection planes at Bragg's angle of 14° and 16° , respectively.

Results and discussion

Degree of substitution of (hexanoic acid modified) BC

ATR-IR was used to study the chemical characteristics of neat and hexanoic acid modified BC. The spectra are shown in Fig. 1. The spectra were normalised to the intensity of the absorption band corresponding to the C–O–C link in cellulose molecules, which is around $1,160\text{ cm}^{-1}$ (Ilharco et al. 1997). The appearance of an absorption band around $1,750\text{ cm}^{-1}$ for hexanoic acid modified BC can be attributed to carbonyl bonds (C = O), which formed during the esterification of BC with hexanoic acid. Additional peaks can also be seen around $2,900\text{ cm}^{-1}$, which correspond to the absorption of methyl ($-\text{CH}_3$) or methylene ($-\text{CH}_2-$) groups. It can be seen from these spectra that the intensity of $1,750\text{ cm}^{-1}$ is higher when freeze-dried BC was used as starting

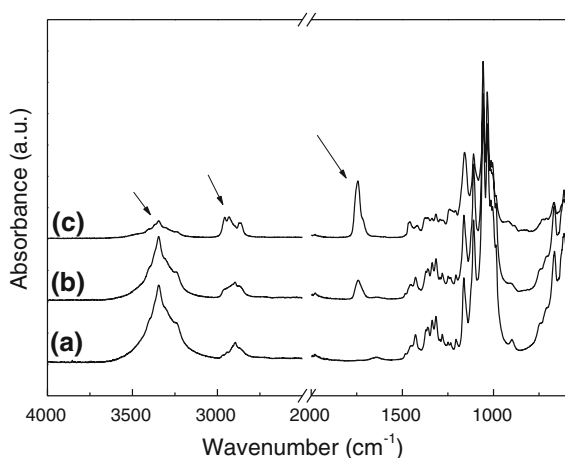


Fig. 1 ATR-IR spectra of neat and hexanoic acid modified BC. **a** Neat BC, **b** C₆-NDBC and **c** C₆-FDBC

Table 1 Degree of substitutions and crystallinity (χ_c) of hexanoic acid modified BC

Sample	Degree of substitution	χ_c (%)
C ₆ -NDBC	0.36 ± 0.01	91 ± 3
C ₆ -FDBC	1.87 ± 0.02	53 ± 4

material. In addition to this, the esterification of freeze-dried BC with hexanoic acid resulted in a significant decrease in the intensity of the hydroxyl groups ($-\text{OH}$) around $3,300\text{ cm}^{-1}$. When never-dried BC was used as the starting material for the esterification reaction, the intensity of the carbonyl absorption band was much lower than that of C₆-FDBC. This is due to the higher degree of substitution when freeze-dried BC was used as the starting material compared to never-dried BC (see Table 1).

Morphology of BC hexanoate

The morphology of BC before and after hexanoic acid esterification is shown in Fig. 2. It can be seen that neat BC possesses a fibrous structure, approximately 50 nm in diameter and several micrometres in length. When BC was modified with hexanoic acid using never-dried BC as the starting material (C₆-NDBC), the fibrous structure of BC is retained (see Fig. 2). This result corroborates the low DS, high crystallinity (Table 1) and low absorbance intensity at $1,750\text{ cm}^{-1}$ in the ATR-IR spectra of C₆-NDBC (Fig. 1). However, when BC hexanoate was synthesised from BC as starting material (C₆-FDBC), the original fibrous structure of BC was lost (Fig. 2 (bottom)). Instead, the modified cellulose resembles a semicrystalline cellulose ester polymer. Similar observations were also reported by Barud et al. (2008), who produced cellulose acetate by the acetylation of BC. This observation is consistent with the high DS and intensity of carbonyl bonds in the ATR-IR spectra, which points towards significant bulk modification during the esterification reaction.

Hypotheses for the bulk modification of freeze-dried BC and surface-only modification of never-dried BC

Could the water adsorbed on starting BC material affect the equilibrium of the esterification reaction?

The esterification reaction of cellulose with a carboxylic acid is reversible with cellulose esters and water

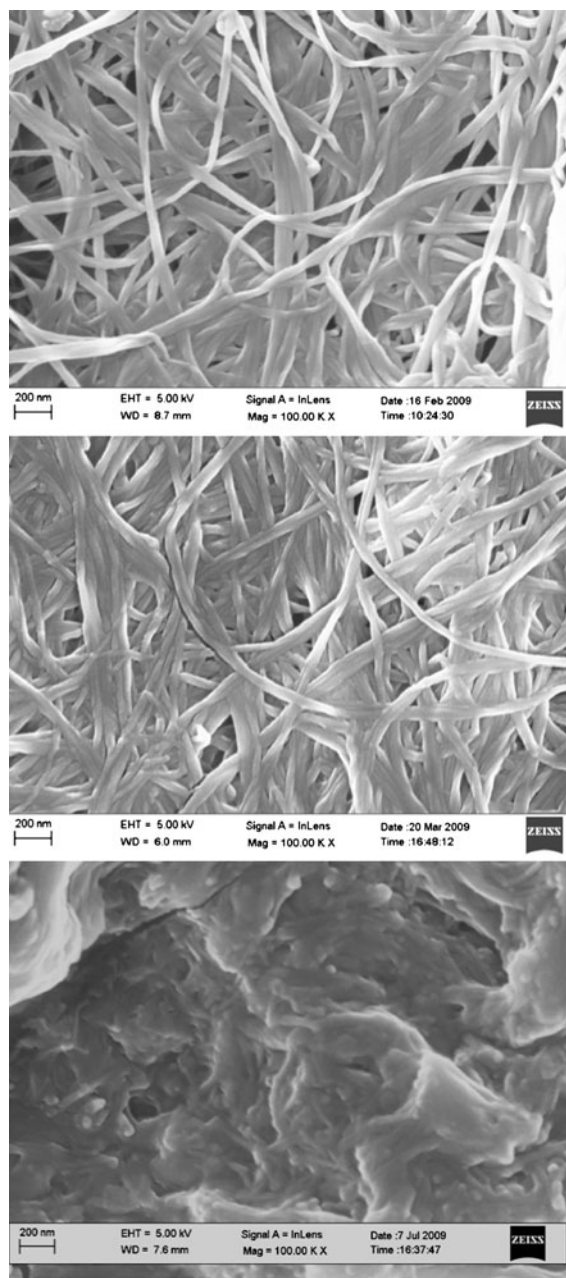


Fig. 2 Scanning electron micrographs showing the morphology of neat and modified BC. *Top*: Neat BC, *middle*: C₆-NDBC and *bottom*: C₆-FDBC

being the product of the reaction. According to Le Chatelier's principle, the removal of water from the reaction will shift the equilibrium of the reaction to favour the production of more cellulose esters. It has also been reported that the starting form of cellulose (never-dried, partially dried or fully dried) affects the

dissolution of cellulose in ionic liquid as the water molecules in the never-dried cellulose reduce the solvent quality (Spinu et al. 2011). The estimated residual water content for both types of BC is tabulated in Table 2. It can be seen that the efficiency of water removal was much greater through solvent exchange method compared to the direct freeze-drying of BC. This can be attributed to the hydrophilic nature of BC, which absorbs water from the atmosphere. The water removal efficiency of the solvent exchange method was found to be 58 ± 14 (wt/wt)%. Through solvent exchange, wet BC is thoroughly mixed with a solvent, which is miscible with water, and the liquid phase is displaced by centrifugation and then decanted. By repeating this process, the water in wet BC is constantly removed and reduced to a minimum. However, this result contradicts with the DS of the modified BC, which showed freeze-dried BC (with higher residual water content) underwent severe bulk modification but never-dried BC (with lower residual water content) underwent surface only modification. The reason for this observation is not clear but it implies that the presence of water is not solely responsible for the observed different behaviour of freeze-dried and never-dried BC.

Could the accessibility of hydroxyl groups be different for freeze-dried and never-dried BC?

The accessible hydroxyl groups (–OH) of both freeze-dried and never-dried BC is tabulated in Table 2. Never-dried BC and freeze-dried BC were found to possess an accessible –OH group availability of 1.12 and 1.24, respectively out of a maximum of 3. The difference in accessible –OH groups availability is about $10 \pm 1\%$. This difference can be attributed to the different dispersing medium (either water or benzene) when preparing BC. Water is known to swell cellulose, exposing more cellulose –OH groups (Venkateswaran and Riemen 1965) but non-polar molecules, like benzene, have been shown to reduce the accessibility of the hydroxyl groups of cellulose (Colombo and Immerguy 1970). As a result, BC freeze-dried from water possesses more accessible hydroxyl groups. Nonetheless, 10% more accessible hydroxyl groups of freeze-dried BC compared to never-dried BC is not significant and does not explain the observed severe modification of freeze-dried BC.

Table 2 A table summarizing the estimated residual water content, accessible –OH groups, specific surface area (S), crystallinity (χ_c), d-spacings of the reflection at 14° (d_1), 16°(d₂) and the discriminant value between I_α and I_β (Z), respectively of never-dried and freeze-dried BC

Types of BC	Residual water (mg/g cellulose)	Accessible OH group ^a	S (m ² g ⁻¹)	χ_c (%)	d_1 (nm)	d_2 (nm)	Z value
Never-dried	13 ± 4	1.12 ± 0.04	92.2 ± 0.1	90 ± 4	0.611	0.614	−0.78 (I_β -rich)
Freeze-dried	23 ± 6	1.24 ± 0.01	44.6 ± 0.1	89 ± 3	0.528	0.531	13.87 (I_α -rich)

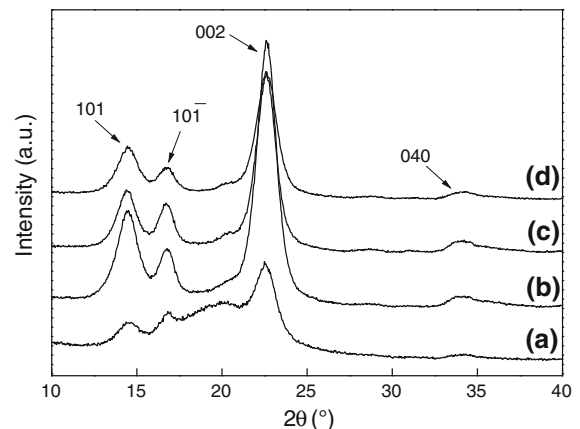
^a Maximum value of 3

Could the exposed surface area of never-dried and freeze-dried BC affect the severity of the esterification reaction?

There is significant evidence in the literature that freeze-drying cellulose from solvents other than water results in a significant increase of the surface area of nanocellulose (Jin et al. 2004; Kuga et al. 2002; Ishida et al. 2004). Similar results were also found in this work. When BC was solvent exchanged into benzene and freeze-dried (never-dried BC), the surface area is twice as large as BC freeze-dried from water (see Table 2). This implies that the exposed surface area of never-dried BC in a solvent is higher than that of freeze-dried BC. The larger surface area in an organic solvent is a result of the low surface tension of the solvent. Merchant (1957) observed that the surface area of cellulose increases with decreasing surface tension of the organic solvent in which the cellulose was dispersed in. Although no exact reason was given at the time of this work, it can be postulated that better wetting can be expected between high surface energy BC and low surface tension liquids. This results in better dispersion of cellulose fibrils in the solvent and therefore the observed high surface area in low surface tension liquids. Nonetheless, our surface area results contradict the observation of severe bulk modification of freeze-dried BC, which possesses a lower surface area.

Could the crystallography of BC change as a result of drying from different solvents?

The XRD pattern of BC hexanoate is shown in Fig. 3a, b and their crystallinity calculated from the Segal's equation in Table 1. Crystallographically, BC possess a cellulose I structure (Vanderhart and Atalla 1984). The characteristic peaks shown in Fig. 3 correspond to

**Fig. 3** XRD spectra of hexanoic acid modified BC. **a** C₆-NDBC, **b** C₆-FDBC, **c** never-dried BC and **d** freeze-dried BC

the diffraction planes of 101, 101̄, 002 and 040, respectively (Freire et al. 2006). The crystallinity of C₆-NDBC is in good agreement with the crystallinity of neat BC of approximately 90% (Czaja et al. 2004). This implies that the modification of never-dried BC resulted in the *surface-only* modification of BC (Lee et al. 2011). When freeze-dried BC was modified with hexanoic acid using identical reaction conditions, the XRD pattern of C₆-FDBC showed a significant reduction the intensities of all the diffraction planes. The crystallinity of C₆-FDBC dropped to only 53%, which is an indication that the cellulose loses its crystal structure and becomes more amorphous. The substitution of hydroxyl groups during the esterification reaction resulted in the reduction of the hydrogen bonding density between cellulose molecules, which in turn partially destroy the crystalline structure of bacterial cellulose (Yin et al. 2007). These results are in good agreement with the ATR-IR spectra and DS of C₆-FDBC shown previously, which points towards

significant bulk modification of cellulose when freeze-dried BC was used as the starting material.

Cellulose is a semi-crystalline composite of two different crystalline structures, namely I_α and I_β , respectively (Horie et al. 1987a). Cellulose I_α possesses a triclinic structure whereas I_β has a monoclinic crystalline structure (Sugiyama et al. 1991). It was found that cellulose I_β is the more thermodynamically favourable structure and cellulose I_α can readily be converted to I_β via routes such as hydrothermal treatment (Horie et al. 1987b). In addition to this, at equivalent hydroxyl group accessibility, cellulose I_α was found to be more reactive towards acetylation compared to cellulose I_β (Sassi et al. 2000). Wada et al. (2001) derived an empirical equation (see Eq. 3) providing the ratio of I_α to I_β based on two equatorial d-spacings of cellulose and these results are tabulated in Table 2, alongside with the crystallinity and d-spacings of never-dried and freeze-dried BC. It can be seen from this table that the crystallinity of the two types of BC are the same ($\sim 90\%$), indicating that the significant bulk modification of freeze-dried BC could not be the direct result of differences in the crystallinity of BC.

The XRD patterns of never-dried and freeze-dried BC are shown Fig. 3c, d. The two diffraction peaks, d_1 and d_2 , are in fact composites of I_α and I_β reflections (Wada et al. 2001). d_1 corresponds to I_α 100 and I_β $1\bar{1}0$ reflections, respectively whereas d_2 corresponds to I_α 010 and I_β 110 reflections, respectively. From the calculated d-spacings for the two types of cellulose, the Z value (which discriminates between I_α -rich and I_β -rich) can be computed. Based on the calculated Z value, never-dried BC is rich in cellulose I_β whereas freeze-dried BC is cellulose I_α dominant. Combining this result with the –OH group accessibility of freeze-dried BC, which is 10% higher than that of never-dried BC, it implies that freeze-dried BC is more susceptible to esterification compared to never-dried BC. This is consistent with our observations. However, this raises the next question; why should BC, which is known to be cellulose I_α , have a cellulose I_β structure when dried from an organic solvent? There was significant evidence that points towards the fact that the surface tension of dispersing medium affects the exposed surface area of cellulose (Merchant 1957; Ishida et al. 2004; Kuga et al. 2002). It was also speculated by Merchant (1957) that the crystal structure of cellulose

would be different when dried from different solvents due to the difference in polarity of the solvents.

Conclusions

We observed that BC freeze-dried from water is more susceptible towards organic acid esterification than never-dried BC. It was found that freeze-dried BC underwent significant bulk modification with degree of substitution of 1.87 compared to never-dried BC of only 0.36. The crystallinity of the freeze-dried BC decreased to 53% from the original value of 90% after the modification whilst the crystallinity of never-dried BC did not change significantly after modification. A few hypotheses to explain the observed behaviour were explored; the effect of residual water, accessibility of –OH groups and crystal structure of the cellulose, respectively. These results were found to be in disagreement with our experimental observations. Never-dried BC, which was solvent exchanged from water through methanol into pyridine, was found to contain less residual water compared to freeze-dried BC. This implies that significant bulk modification should occur in never-dried BC as the lack of water should shift the equilibrium of the esterification reaction to the right, producing more cellulose esters. However, it was freeze-dried BC, which had a higher water content, that underwent significant bulk modification. Freeze-dried BC was found to possess 10% more accessible –OH groups compared to never-dried BC. However, 10% more –OH groups is very unlikely to result in an 80% increase in the degree of substitution of modified cellulose. XRD pattern show that freeze-dried BC is cellulose I_α -rich and never-dried BC is cellulose I_β -rich. It has been shown that cellulose I_α is more susceptible to acetylation compared to cellulose I_β . We postulate that it is this combination of more exposed –OH groups and the cellulose I_α structure of freeze-dried BC which resulted in significantly higher bulk modification of the cellulose compared to never-dried BC under identical reaction conditions. However, the main question that we need to raise is: Why does never-dried BC, which was obtained by freeze-drying from an organic solvent is found to possess a cellulose I_β structure while it is well accepted that BC possesses a cellulose I_α structure?

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