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Synthesis, sensor activity and logic behaviour of a novel bichromophoric system based on rhodamine 6G and 1,8-naphthalimide



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ABSTRACT

A novel fluorescence sensing system based on rhodamine 6G and 1,8-naphthalimide fluorophores was synthesized and investigated. The system was designed as a wavelength-shifting bichromophoric molecule where the 1,8-naphthalimide moiety is an energy “donor” capable of absorbing light and efficiently transferring the energy to a focal Rhodamine 6G “acceptor”. Furthermore, the 1,8-naphthalimide unit was configured on the “fluorophore-spacer-receptor” format. Thus, the distinguishing features of fluorescence resonance energy transfer systems were successfully combined with the properties of photoinduced electron transfer and classical ring-opening sensor systems. The synthesized compound shows excellent signalling properties towards protons as well as Cu^{2+} and Hg^{2+} over the representative metal ions. Due to the remarkable fluorescence changes in the presence of protons, Cu^{2+} and Hg^{2+} ions the novel system is able to act as a two output combinatorial logic circuit with three chemical inputs.

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1. Introduction

Detection of trace amounts of species such as protons and metal ions is of great interest in the fields of chemical, biological, medicinal and environmental sciences [1–6]. In this regard, fluorescence spectroscopy affords an easy way of sensing and imaging cations because of its simplicity and high sensitivity [7–9].

Determination of pH is one of the most important analytical methods in the chemical laboratories and in the industry. pH is a key parameter in clinical analysis, food production, biotechnological processes, waste water treatment procedures, environmental and life sciences [10–16]. Intracellular pH environments play a central modulating role in biosystems, since minor variations in pH

may lead to markedly changed cellular behaviour in proliferation, apoptosis, enzymatic activity and ion transport [17,18]. Accordingly, some illnesses like cancer, stroke and Alzheimer's disease are found to be accompanied with pH changes [19,20]. Hence the development of simple and convenient approaches for detecting pH is of great importance in both cellular analysis and diagnosis.

Developing sensors for selective and sensitive detection of mercury species have received an immense interest due to their extreme toxicity towards humans [21,22]. Mercury is the third most frequently found and second most common toxic heavy metal in the list of the Agency for Toxic Substances and Disease Registry (ATSDR) of the U.S. Department of Health and Human Services [23]. The extreme toxicity of mercury and its derivatives results from its high affinity for thiol groups in proteins and enzymes, leading to the dysfunction of cells and consequently causing health problems [24,25]. Unfortunately, mercury contamination can occur through a variety of natural and anthropogenic sources including oceanic and volcanic emissions, gold mining, and combustion of fossil fuels. Thus, the health concerns over exposure to mercury have

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motivated the exploration of selective and efficient methods for the monitoring of mercury in biological and environmental samples.

Rhodamine fluorophore has consistently demonstrated its potential to construct fluorescent sensors for determination of protons and mercury, ascribing to their spirolactam ring-opening equilibrium [26–30]. The single emission-based probes usually tend to be affected by a variety of factors such as instrumental efficiency and environmental conditions, as well as the concentration of the probe molecule. These interferences can be eliminated by employing ratiometric fluorescent probes, which allow the measurement of changes of the intensity ratio at two emission bands to provide built-in correction for the above-mentioned environmental effects [9,25,27,31,32]. Several strategies, including internal charge transfer (ICT) [33,34], excited-state intramolecular proton transfer (ESIPT) [35] and fluorescence resonance energy transfer (FRET) [36,37], have been adopted to design ratiometric probes. Among them, FRET strategy could provide moderate resolution of the two emission bands and has been widely applied in designing ratiometric probes for bioanalytical applications. In this work, a novel ratiometric fluorescent probe for pH and mercury detection was developed combining the advantages of the FRET based bichromophoric systems and Rhodamine receptor abilities.

Since the first suggestion of Boolean operator at molecular level based on fluorescent chemosensors, the field has rapidly extended from simple switches to more complex molecular systems that are capable of performing a variety of classical logic functions [38–40] and examples of half-adder [41,42], full-adder [43], keypad lock [44], half-subtractor [45], full-subtractor [43], encoder-decoder [46] and a digital comparator [47–49] have been described. Also a smart oligonucleotide-based automaton that plays simple games such as Tic-Tac-Toe have been reported [50]. Such logic functions are of elevated interest for applications such as object coding, intelligent materials, drug delivery and activation, diagnostics or actuation [51,52]. Nevertheless, the physical integration of molecular logic gates is especially important for rational design and implementation towards advanced molecular scale computing. Herein, we propose a cascade of two molecular fluorescent chemosensors that are integrated in logic circuit comprising a FRET bichromophoric system. The examined bichromophoric system was constructed by coupling of a Rhodamine 6G acceptor dye and a photoinduced electron transfer (PET) based 1,8-naphthalimide donor (Scheme 1).

2. Experimental

2.1. Materials

The intermediate compounds **2** and **3** were prepared according to the reported procedure [53]. Commercially available ethylenediamine and methylacrylate (Aldrich) were used without purification. All solvents (Aldrich, Fisher Chemical) were pure or of

spectroscopy grade. Commercial aqueous buffer solution HEPES (Fisher Chemical) was used as received. $\text{Zn}(\text{NO}_3)_2$, $\text{Cu}(\text{NO}_3)_2$, $\text{Ni}(\text{NO}_3)_2$, $\text{Co}(\text{NO}_3)_2$, $\text{Pb}(\text{NO}_3)_2$, $\text{Fe}(\text{NO}_3)_3$, $\text{Hg}(\text{NO}_3)_2$, and AgNO_3 salts were the sources for metal cations (all Aldrich salts at p.a. grade).

2.2. Methods

FT-IR spectra were recorded on a Varian Scimitar 1000 spectrometer. The ^1H NMR spectra (chemical shifts are given in ppm) were recorded on a Bruker DRX-250 spectrometer operating at 250.13 MHz. TLC was performed on silica gel, Fluka F60 254, 20×20 , 0.2 mm. The melting points were determined by means of a Kofler melting point microscope. The UV–VIS absorption spectra were recorded on a spectrophotometer Hewlett Packard 8452A. The fluorescence spectra were taken on a Scinco FS-2 spectrofluorimeter. The fluorescence quantum yields (Φ_F) were measured relatively to Rhodamine 6G ($\Phi_F = 0.95$ in ethanol [54]) or Coumarin 6 ($\Phi_F = 0.78$ in ethanol [55]) as standards. All the experiments were performed at room temperature (25.0 °C). A 1×1 cm quartz cuvette was used for all spectroscopic analysis. To adjust the pH, very small volumes of hydrochloric acid and sodium hydroxide were used. The effect of the metal cations upon the fluorescence intensity was examined by adding portions of the metal cations stock solution to a known volume of the fluorophore solution (10 mL). The addition was limited to 100 μL so that dilution remains insignificant.

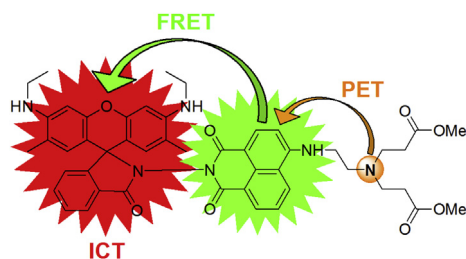
2.3. Synthesis of dyad (**4**)

To a solution of **3** (1.96 g, 3 mmol) in DMF (30 mL), ethylenediamine (6.2 mL, 30 equiv. per reactive nitro group) was added at room temperature. After 48 h, the resulting solution was poured into water. The precipitate was filtered off and washed with water. The final product **4** was obtained as yellow solid after purification by column chromatography using silica gel as a stationary phase and methanol/ammonium hydroxide = 5:0.4 as eluent. In order to remove the silica gel particles, the solid was dissolved in chloroform, resulting solution was filtrated off and the solvent was evaporated in vacuum.

Yield: 1.24 g (62%), m.p. > 250 °C, $R_f = 0.48$ (methanol/ammonium hydroxide = 5:0.4) IR (KBr) cm^{-1} : 3313 and 3224 (ν_{NH} and ν_{NH_2}); 2927 and 2885 (ν_{CH}); 1706 ($\nu^{\text{as}}\text{N}-\text{C}=\text{O}$); 1674 ($\nu^{\text{s}}\text{N}-\text{C}=\text{O}$). ^1H NMR (CDCl_3 - d , 250.13 MHz) ppm: 8.18 (d, 1H, $J = 8.4$ Hz, Naphthalimide H-7); 8.11 (m, 1H, 9-Ph H-3); 7.73 (d, 1H, $J = 8.6$ Hz, Naphthalimide H-2); 7.65 (m, 2H, 9-Ph H-4, 9-Ph H-5); 7.56 (d, 1H, $J = 7.2$ Hz, Naphthalimide H-5); 7.29 (m, 1H, 9-Ph H-6); 7.00 (dd, 1H, $J = 8.4$ Hz, $J = 7.2$ Hz, Naphthalimide H-6); 6.51 (m, 3H, Rhodamine H-4, Rhodamine H-5, Naphthalimide 4-NH); 6.21 (d, 1H, $J = 8.6$ Hz, Naphthalimide H-3); 6.05 (s, 1H, Rhodamine H-1); 6.00 (s, 1H, Rhodamine H-8); 3.41 (br.s, 2H, $2 \times \text{NH}$); 3.22 (m, 2H, $\text{NHCH}_2\text{CH}_2\text{NH}_2$); 3.03 (m, 6H, $\text{NHCH}_2\text{CH}_2\text{NH}_2$, Rhodamine $2 \times \text{CH}_2\text{CH}_3$); 1.98 (s, 3H, Rhodamine CH_3); 1.97 (s, 3H, Rhodamine CH_3); 1.20 (m, 8H, NH_2 , Rhodamine $2 \times \text{CH}_2\text{CH}_3$). Elemental analysis: Calculated for $\text{C}_{40}\text{H}_{38}\text{N}_6\text{O}_4$ (MW 666.8) C 72.05, H 5.74, N 12.60%; Found C 72.31, H 5.85, N 12.44%.

2.4. Synthesis of dyad (**5**)

A suspension of Dyad **4** (1.0 g, 1.5 mmol) in methanol (20 mL) was added dropwise over period of 20–30 min to a solution of methylacrylate (1.4 mL, 10 equiv. per reactive amine group) in 2 mL of cooled to 0 °C methanol. The reaction was allowed to warm slowly to room temperature and then stirred for 3 days. The final product was obtained as yellow solid after purification by column chromatography using silica gel as a stationary phase and toluene/ethyl acetate/ethanol = 3:3:1 as eluent. In order to remove the silica



Scheme 1. Rhodamine 6G/1,8-naphthalimide wavelength-shifting bichromophoric system.

gel particles, the solid was dissolved in chloroform, resulting solution was filtrated off and the solvent was evaporated in vacuum.

Yield: 0.91 g (73%), m.p 194–195 °C, R_f = 0.74 (toluene/ethyl acetate/ethanol = 3:3:1). IR (KBr) cm^{-1} : 3366 (νNH); 2921 and 2878 (νCH); 1722 ($\nu\text{C}=\text{OOME}$); 1676 ($\nu\text{N}-\text{C}=\text{O}$). ^1H NMR (CDCl_3 -d, 250.13 MHz) ppm: 8.26 (d, 1H, J = 8.3 Hz, Naphthalimide H-7); 8.11 (m, 3H, 9-Ph H-3, 9-Ph H-4 и Naphthalimide H-2); 7.64 (m, 2H, 9-Ph H-5, Naphthalimide H-5); 7.40 (t, 1H, J = 7.9 Hz, Naphthalimide H-6); 7.30 (m, 1H, 9-Ph H-6); 6.61 (m, 2H, Rhodamine H-4, Rhodamine H-5); 6.47 (d, 1H, J = 8.5 Hz, Naphthalimide H-3); 6.21 (br.s, 1H, Naphthalimide NH); 6.12 (s, 1H, Rhodamine H-1); 6.10 (s, 1H, Rhodamine H-8); 3.59 (s, 6H, $2 \times \text{CH}_2\text{OOCH}_3$); 3.45 (br.s, 2H, $2 \times \text{NH}$); 3.22 (m, 2H, NHCH_2CH_2); 3.03 (m, 4H, Rhodamine $2 \times \text{CH}_2\text{CH}_3$); 2.86 (t, 6H, J = 6.3 Hz, $3 \times \text{NCH}_2$); 2.51 (t, 4H, J = 6.4 Hz, $2 \times \text{CH}_2\text{COOCH}_3$); 2.03 (s, 6H, $2 \times \text{Rhodamine CH}_3$); 1.20 (m, 6H, Rhodamine $2 \times \text{CH}_2\text{CH}_3$). Elemental analysis: Calculated for $\text{C}_{48}\text{H}_{50}\text{N}_6\text{O}_8$ (MW 839.0) C 68.72, H 6.01, N 10.02%; Found C 69.08, H 6.17, N 9.77%.

3. Results and discussion

3.1. Design and synthesis

It is well known that the spectral overlap between the emission of the donor fluorophore and the absorbance of the acceptor is the main factor that determines the efficiency of FRET systems [56]. When constructing the novel FRET system **5**, the 4-amino-1,8-naphthalimide fluorophore was chosen as an energy donor due to its strong emission in the visible region (450–600 nm) which covers a part of rhodamine absorption, thus providing favourable conditions for the FRET process [57,58]. Also, the 1,8-naphthalimide donor is a PET based chemosensing system with a diester terminated amine receptor [59]. Thus we expect that the PET process in 1,8-naphthalimide will effectively quench the fluorescence of the yellow-green emitting donor and affect the FRET efficiency to the rhodamine unit, which represents the “off-state” of the system. Coupling the amine receptor with the analyte (protons or different metal ions) would increase its oxidation potential, and as such, thermodynamically disallow the electron transfer to the donating 1,8-naphthalimide. Consequently the energy transfer to the acceptor would be “switched on” (Scheme 2).

The synthesis of the desired wavelength-shifting bichromophore **5** was accomplished in four steps as outlined in Scheme 3.

First, intermediate **3** was obtained as we previously reported by condensation of Rhodamine 6G hydrazide **2** with 4-nitro-1,8-naphthalic anhydride in glacial acetic acid at 110 °C for 5 h. Rhodamine 6G hydrazide **2** was obtained by reaction of Rhodamine 6G **1** with hydrazine monohydrate under reflux in absolute ethyl alcohol for 5 h [53].

Thus prepared intermediate **3** was nucleophilically substituted with the commercially available ethylenediamine in DMF solution at room temperature for 48 h. Finally, the reaction of intermediate **4** with methylacrylate in methanol gave the novel Dyad **5**.

The synthesized compounds were characterized by their melting points and TLC retention values (R_f) and identified by conventional techniques - elemental analysis data, UV–Vis, fluorescence, FT-IR and ^1H NMR spectroscopy.

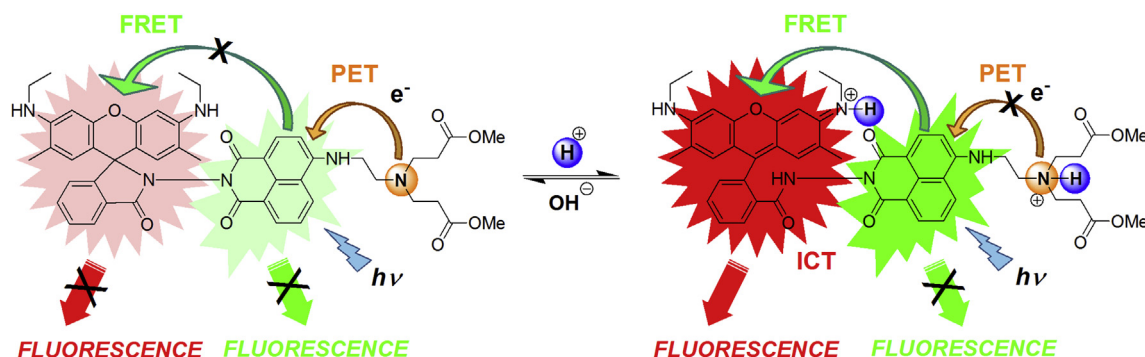
For instance, in the ^1H NMR (CDCl_3 -d, 250.13 MHz) spectrum of Dyad **4** a resonance at 6.21 ppm (Fig. 1a, I) was observed which is characteristic for the proton in position C-3 of the yellow-green emitting 1,8-naphthalimide, substituted in position C-4 with an electron-donating amino group. This resonance is different from the corresponding resonance for the 4-nitro-1,8-naphthalimide moiety in intermediate **3** (8.35 ppm) [53]. This is solid evidence that the 4-nitro-1,8-naphthalimide unit in **3** was completely converted into the yellow-green emitting donor fluorophore (Dyad **4**). Furthermore, in the ^1H NMR spectrum of final Dyad **5** a singlet at 3.59 ppm for 6 protons (Fig. 1b, II) and two triplets at 2.86 ppm for 6 protons (Fig. 1b, III) and at 2.51 ppm for 4 protons (Fig. 1b, IV) appeared which is typical for the desired diester terminated receptor unit [60] suggesting that the starting primary amine in Dyad **4** was successfully branched to give the diester moiety in Dyad **5**.

3.2. Influence of pH on the photophysical properties of dyad **5**

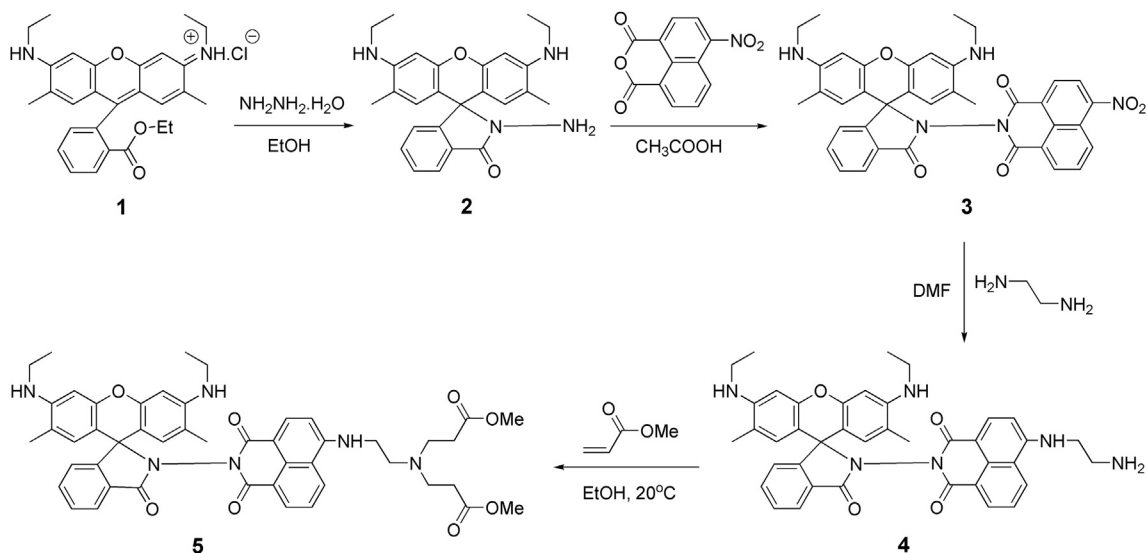
Photophysical properties of Dyad **5** were determined in water/acetonitrile (4:1, v/v) solution. In alkaline and neutral media ($\text{pH} > 6$) the rhodamine moiety adopts a closed, non-fluorescent spirolactam form. This results in absorption of the system at about 420–440 nm, which is attributed to an internal charge transfer process in the 1,8-naphthalimide chromophore. At ca. pH 5 the spirolactam ring of rhodamine is opened and a new absorption (rhodamine) band with well pronounced maximum at 530 nm appears (Fig. 2) which gradually increases upon increasing the acidity of solution up to pH 2. Also, a blue shift of the 1,8-naphthalimide absorption band ($\Delta\lambda_A = 13$ nm) in acid media was observed (Fig. 2). This effect probably is due to protonation of the amine receptor of compound **5** that give rise to some weak charge repulsion on the aromatic amine of the donor fluorophore, which is well correlated with the data for the 1,8-naphthalimide derivatives [61–63].

From the absorption changes at 530 nm, the titration curve of novel compound was obtained. The analysis of the titration plot according to Eq. (1) [64] gives the pK_a value of 4.7, which is similar to the other rhodamine derivatives [65].

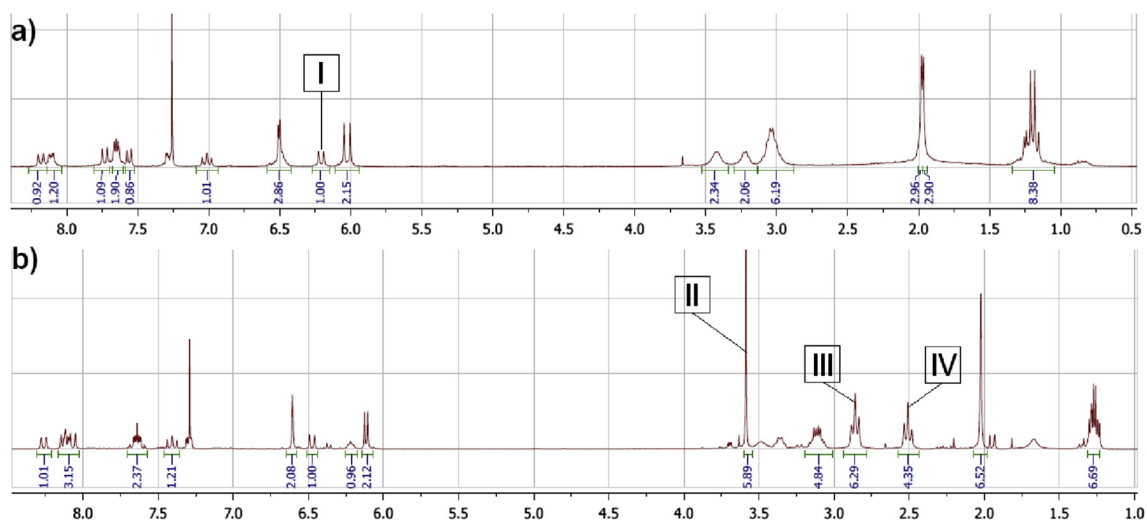
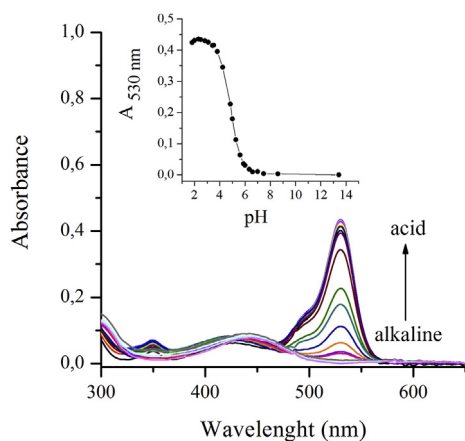
$$\log[(A_{\text{max}} - A)/A - A_{\text{min}}] = \text{pH} - \text{pK}_a \quad (1)$$



Scheme 2. Photophysical behaviour of bichromophoric system **5** as a function of pH after excitation within a spectral region of maximal absorption of the donating fluorophore.



Scheme 3. Synthesis of wavelength-shifting bichromophoric system 5.

Fig. 1. ^1H NMR (CDCl_3 -d, 250.13 MHz) spectra of Dyad 4 (a) and Dyad 5 (b).Fig. 2. Effect of pH on the UV/Vis absorption spectra of Dyad 5. Inset: Titration plot of Dyad 5 at $\lambda_A = 530$ nm in a pH range of ca. 13–2.

In alkaline and neutral media, after excitation at 420 nm (within the spectral region of maximal absorption of the donor fluorophore) the novel fluorescence probe shows only weak emission in the range of 480–700 nm with maximum at 539 nm. Under these conditions, the acceptor dye is in a spirolactam form and energy transfer from the donor fluorophore to the rhodamine is not possible. In this case the system shows a yellow-green emission that is typical for the 4-amino-1,8-naphthalimide derivatives. The quantum yield of fluorescence in water/acetonitrile (4:1, v/v) solution was calculated using Coumarin 6 ($\Phi_F = 0.78$ in ethanol [55]) as a standard according to Eq. (2) [66], where A_{ref} , S_{ref} , n_{ref} and A_{sample} , S_{sample} , n_{sample} represent the absorbance at the excited wavelength, the integrated emission band area and the solvent refractive index of the standard and the sample, respectively.

$$\Phi_F = \Phi_{\text{ref}} \left(\frac{S_{\text{sample}}}{S_{\text{ref}}} \right) \left(\frac{A_{\text{ref}}}{A_{\text{sample}}} \right) \left(\frac{n_{\text{sample}}^2}{n_{\text{ref}}^2} \right) \quad (2)$$

However, the calculated quantum yield of Dyad 5 at pH 7 was 0.02 which is extremely low compared to the quantum yield of the

traditional 4-amino-1,8-naphthalimides, not containing an amine receptor [59]. This phenomenon might be caused by the possible photoinduced electron transfer from the alkylamino receptor to the 4-amino-1,8-naphthalimide fluorophore. Thus the fluorescence of the 4-amino-1,8-naphthalimide fluorophore is quenched (Scheme 2). It is also worth noting that the calculated quantum yield of Dyad **5** water/acetonitrile (4:1, v/v) was even lower than the quantum yield of 1,8-naphthalimide fluorophores with a receptor fragment. The reason for such behaviour of the novel compound could be twofold: (i) It is well known that the water is an effective fluorescence quencher, which additionally decreased the quantum yield in the aqueous medium; (ii) Novel Dyad **5** is highly hydrophobic and probably quenches its emission in the aqueous solution due to the aggregation process.

The used diester terminated naphthalimide acceptor shows pK_a value of about 4.8 [59]. Upon acidification from pH 6 to 2, the tertiary amine receptor is protonated which prevents the PET quenching process in the donor 1,8-naphthalimide. Nevertheless, the 1,8-naphthalimide emission does not increase because the spirolactam ring becomes open under these conditions, which allows the energy transfer to the rhodamine acceptor moiety. This results in remarkable fluorescence intensity enhancement (FE = 358) in the rhodamine emission region at 555 nm (Fig. 3). The quantum yield of fluorescence of rhodamine moiety in Dyad **5** (λ_{ex} = 510 nm) was calculated to be Φ_F = 0.27 at pH 2 in water/acetonitrile (4:1, v/v) using Rhodamine 6G (Φ_F = 0.95 in ethanol [54]) as a standard.

The analysis of the fluorescence changes at 555 nm (Fig. 3) according to Eq. (3) [64] gives the pK_a value of 4.6 which is very similar to the pK_a value calculated according absorption changes and is attributed to the spirolactam ring-opening reaction.

$$\log[(I_{F \max} - I_F)/I_F - I_{F \min}] = pH - pK_a \quad (3)$$

3.3. Influence of metal cations on the fluorescence intensity of dyad **5**

The signalling fluorescent properties of Dyad **5** in the presence of transition metal cations (Co^{2+} , Cu^{2+} , Fe^{3+} , Ni^{2+} , Pb^{2+} , Zn^{2+} , Hg^{2+} and Ag^+) were investigated in acetonitrile. For a constant pH, the solutions were buffered with addition of 1 mmol HEPES solution (pH = 7.2). The HEPES buffer was chosen because it ensures a pH value about pH = 7 where the PET based 1,8-naphthalimide donor and rhodamine acceptor units in Dyad **5** are in their “off-state” and

cation recognition in the both moieties is feasible. The changes in the fluorescence spectrum of bichromophoric system observed after addition of 10 equivalents of examined metal ions are presented in Fig. 4. As can be seen, only Cu^{2+} and Hg^{2+} remarkably affected the fluorescence spectrum of Dyad **5**. The Cu^{2+} ions increased the fluorescence intensity of naphthalimide donor, while the presence of Hg^{2+} ions resulted in a strong fluorescence enhancement of the rhodamine acceptor. Although a fluorescence enhancement of rhodamine unit in Dyad **5** was occurred when Pb^{2+} was added, this effect is negligible compared with the influence of Hg^{2+} and do not affect naphthalimide fluorescence output. This suggests that the ability of Dyad **5** to bind Pb^{2+} ions is a very low.

Upon addition of Cu^{2+} a fluorescence enhancement of the 1,8-naphthalimide donor emission in dyad **5** was observed (Fig. 5A). Obviously, among the examined metal cations only Cu^{2+} is able to bind the PET amine receptor of the 1,8-naphthalimide unit. Upon recognition of Cu^{2+} , which binds the 1,8-naphthalimide receptor engaging its lone-pair electrons, the PET process is no longer possible and the 1,8-naphthalimide fluorescence is increased. The enhancement of the fluorescence emission (FE) has been used as a qualitative parameter. The $FE = I/I_0$ is determined as the ratio between the maximum fluorescence intensity (I - after metal ion addition) and the minimum fluorescence intensity (I_0 - free of metal cations solution). The calculated FE value of 2.2 is consistent with the data for a naphthalimide diester of similar nature that was developed before [59]. The stoichiometry of the complex between Cu^{2+} cation and Dyad **5** was determined using the method of continuous variations (Job's method). Job's plot analysis of the titration revealed a maximum at about 0.5 mol fraction, indicating 1:1 binding stoichiometry (Fig. 5B).

The linear range of the method was found to be at least 2–10 μM Cu^{2+} with a correlation coefficient of $R^2 = 0.99301$. The limit of detection (LOD) was calculated to be 5×10^{-7} mol L^{-1} according to formula $LOD = 3\sigma/b$ where σ is the standard deviation and b is the slope of the calibration plot (Fig. 5A, Inset) [67]. The fluorescence intensity of Dyad **5** without Cu^{2+} was measured by 10 times and the standard deviation of measurements was determined.

In contrast with the effect of Cu^{2+} , the addition of Hg^{2+} decreased the 1,8-naphthalimide emission and a new emission band centered at 555 nm (emission of rhodamine acceptor moiety) appeared (Fig. 6A). This fact is solid evidence that the FRET process in Dyad **5** is “switched on” by Hg^{2+} ions because the excitation of 1,8-naphthalimide at 420 nm results in emission of the rhodamine

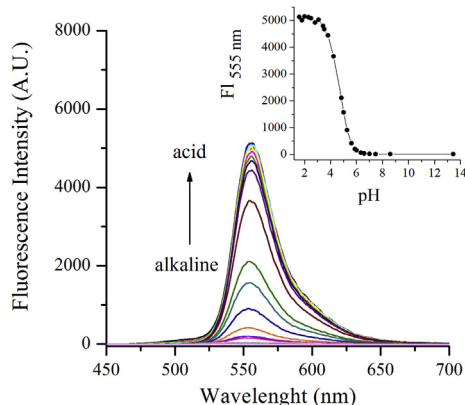


Fig. 3. Effect of pH on the fluorescence spectra of Dyad **5**, excited at 420 nm. Inset: Titration plot of Dyad **5** at λ_F = 530 nm in a pH range of ca. 13–2.

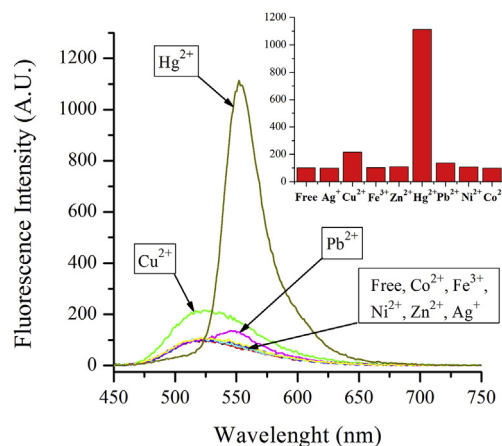


Fig. 4. Effect of the metal cations at concentration $C = 10^{-4}$ mol L^{-1} on the fluorescence of Dyad **5** ($C = 10^{-5}$ mol L^{-1}) in acetonitrile solution buffered with 1 mmol HEPES.

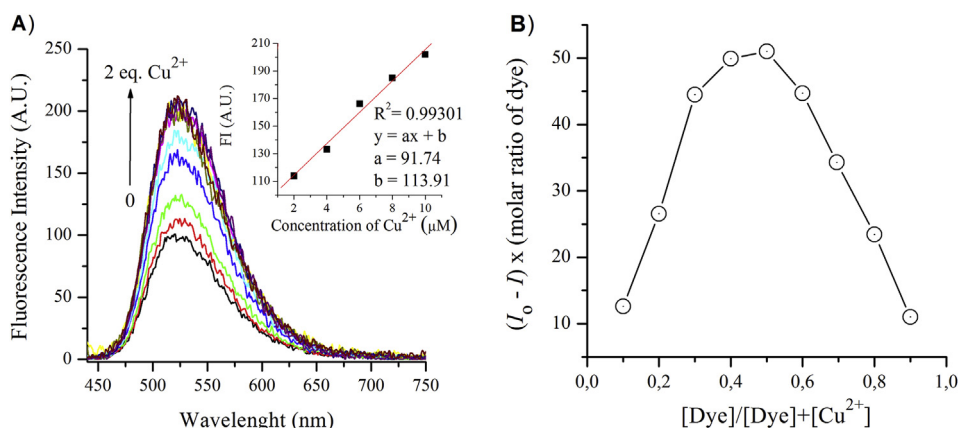


Fig. 5. (A) Effect of Cu^{2+} on the fluorescence of Dyad **5** ($C = 10^{-5} \text{ mol L}^{-1}$) and (B) Job's plot in acetonitrile solution buffered with 1 mmol HEPES. Inset: Calibration plot of Dyad **5**/ Cu^{2+} .

unit with a maximum at 555 nm. Obviously the Hg^{2+} binds the spirolactam receptor moiety and the rhodamine acceptor of Dyad **5** is converted into the fluorescent ring-opened form. The unique selectivity for Hg^{2+} is probably due to the cooperation of several combined influences, such as the suitable coordination geometry, the proper radius and charge density around oxygen atom of the rhodamine carbonyl group, which takes part in the complexation with the Hg^{2+} ion [68,69]. Also it was found that the rhodamine emission peak at 555 nm attained saturation upon addition of 50 eq. of Hg^{2+} . Furthermore the Job's plot analysis (Fig. 6B) revealed 1:1 binding stoichiometry (maximum at about 0.5 mol fraction) suggesting that the Hg^{2+} binding process occurs only in spirolactam receptor fragment in the novel Dyad **5**. The linear range of the method was found to be at least 2–10 μM Hg^{2+} with a correlation coefficient $R^2 = 0.99959$. From the calibration plot (Fig. 6A, Inset) the LOD was calculated to be $9 \times 10^{-8} \text{ mol L}^{-1}$.

3.4. Logic behaviour of dyad 5

Due to the remarkable fluorescence changes of Dyad **5** (1 eq., $C = 10^{-5} \text{ mol L}^{-1}$) in the presence of 2 eq. of protons (HCl, $C = 2 \times 10^{-5} \text{ mol L}^{-1}$), 1 eq. of Cu^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) and 1 eq. of Hg^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) it would be able to act as a two output combinatorial logic circuit with three chemical inputs. By monitoring the donor emission in Dyad **5** (Output 1, 1,8-naphthalimide

fluorescence at 515 nm) a high (coded in binary as 1) fluorescence output was observed only in the presence of Cu^{2+} which cuts off the PET process in the 1,8-naphthalimide fluorophore (Scheme 4). In the presence of protons or Hg^{2+} the rhodamine spirolactam cycle is opened which allows the energy transfer from the 1,8-naphthalimide donor to the rhodamine acceptor, thus quenching the 1,8-naphthalimide emission and preventing the action of Cu^{2+} .

So, the fluorescence of 1,8-naphthalimide moiety in the Dyad **5** at 515 nm (Output 1) is high (Fig. 7A, Table 1) only in the presence of Cu^{2+} (Input 3) and in the absence of Hg^{2+} (Input 1) and protons (Input 2) which is correlated very well with a Three-input Disabled INHIBIT gate [70].

By monitoring the rhodamine emission (Output 2, fluorescence at 550 nm), actually, Dyad **5** works as a two input logic gate because Cu^{2+} (Input 3) does not affect by any way the spirolactam opening reaction which is responsible for the high output value (strong emission). In the presence of 1 eq. Hg^{2+} (Input 1) and/or 2 μL 10% HCl (Input 2) a strong emission at 550 nm was observed owing to the spirolactam opened form (Fig. 8, Table 1). This behaviour correlates very well with OR logic gate [70]. The output is high if either or both of the input values are high. If both inputs are low, then the output is low.

On the basis of the present study and results obtained the logic behaviour of Dyad **5** could be summarized with the electronic representation in Scheme 5.

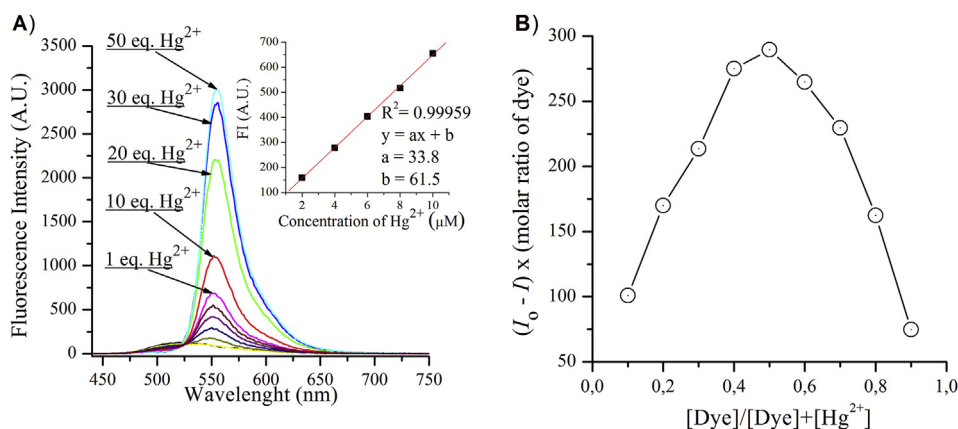
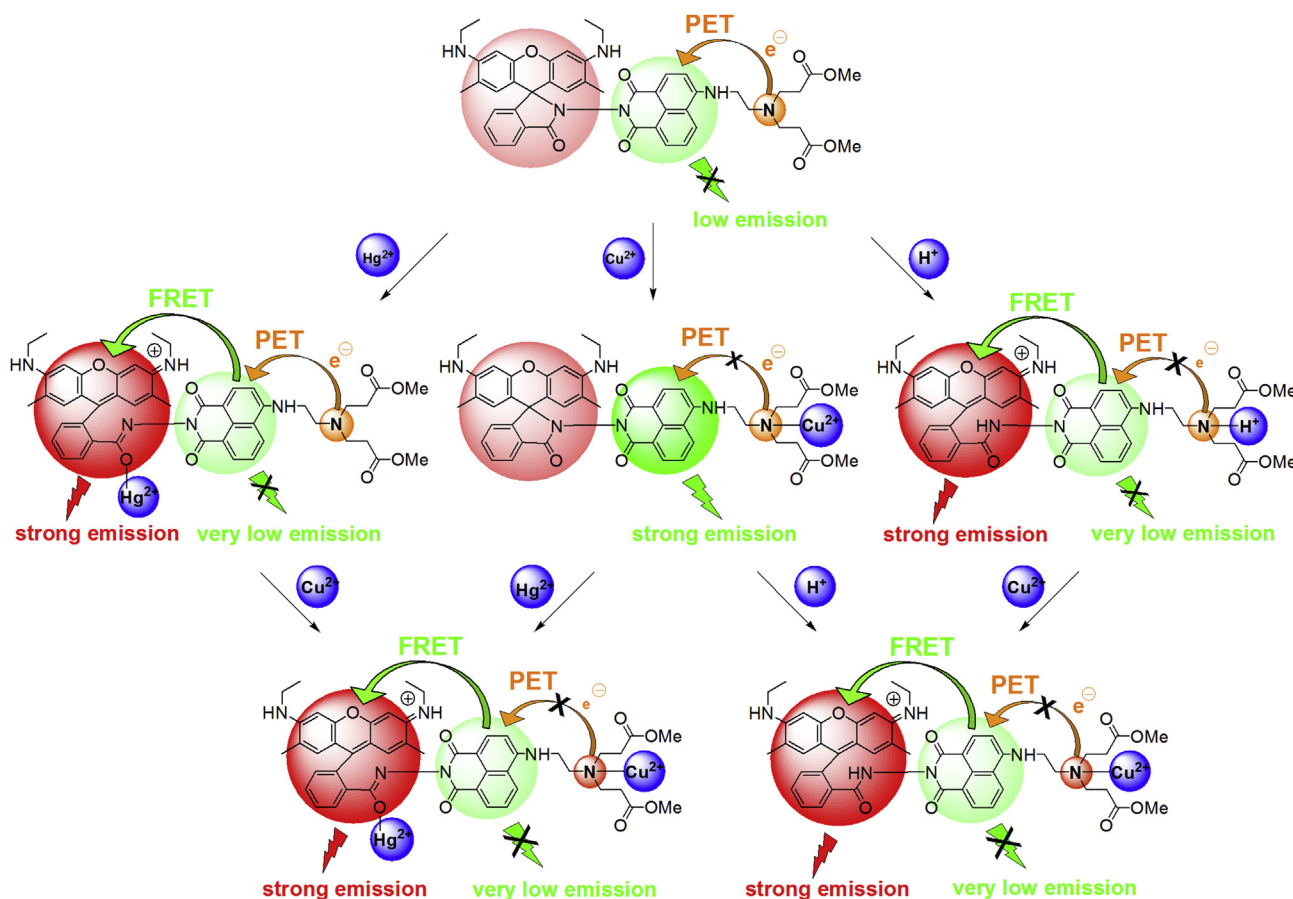


Fig. 6. (A) Effect of Hg^{2+} on the fluorescence of Dyad **5** ($C = 10^{-5} \text{ mol L}^{-1}$) and (B) Job's plot in acetonitrile solution buffered with 1 mmol HEPES. Inset: Calibration plot of Dyad **5**/ Hg^{2+} .



Scheme 4. Fluorescence changes of Dyad **5** (1 eq., $C = 10^{-5} \text{ mol L}^{-1}$) in presence of 2 eq. of protons ($C = 2 \times 10^{-5} \text{ mol L}^{-1}$), 1 eq. of Cu^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) and 1 eq. of Hg^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$).

4. Conclusions

A novel fluorescent wavelength-shifting bichromophoric system with sensing properties based on FRET, PET and ICT, containing yellow-green emitting 1,8-naphthalimide donor and orange-red emitting rhodamine acceptor, was synthesized and its photo-physical behaviour in presence of protons and representative metal ions was studied. The donating 1,8-naphthalimide unit was also designed according to the “fluorophore-spacer-receptor” model and as such it would be able to act as a fluorescence PET based

chemosensor. In alkaline and neutral media, after excitation within the spectral region of maximal absorption of the donor fluorophore the novel fluorescence probe shows only week yellow-green emission, typical for the 1,8-naphthalimides. Acidification from pH 5 to 2 prevents the PET quenching process in the donor unit and opens the rhodamine spirolactam ring, which allows the energy transfer to the rhodamine acceptor moiety with remarkable fluorescence intensity enhancement ($\text{FE} = 358$) in the rhodamine emission region. In the presence of representative metal ions only Cu^{2+} and Hg^{2+} remarkably affect the fluorescence spectrum of the system. It is shown that only Cu^{2+} is able to bind the amine receptor of the 1,8-naphthalimide unit (1:1 binding stoichiometry), thus preventing the PET process in the donor unit. In contrast, the Hg^{2+}

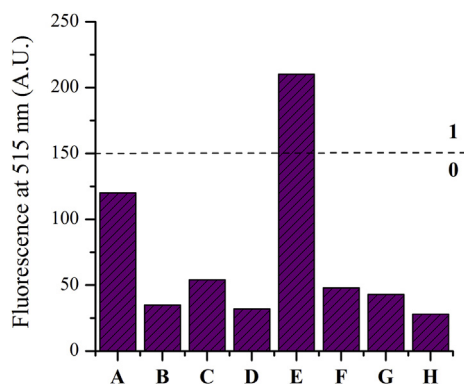


Fig. 7. The changes in fluorescence intensity of Dyad **5** ($C = 10^{-5} \text{ mol L}^{-1}$) at 515 nm with 2 eq. of protons ($C = 2 \times 10^{-5} \text{ mol L}^{-1}$), 1 eq. of Cu^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) and 1 eq. of Hg^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) as chemical inputs.

Table 1

The truth table for the operation of 1 eq. of Dyad **5** ($C = 10^{-5} \text{ mol L}^{-1}$) and 2 eq. of protons ($C = 2 \times 10^{-5} \text{ mol L}^{-1}$), 1 eq. of Cu^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) and 1 eq. of Hg^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) as chemical inputs.

	Input Hg^{2+}	Input H^+	Input Cu^{2+}	Output FI_{515}	Output FI_{550}
A	0	0	0	0	0
B	0	1	0	0	1
C	1	0	0	0	1
D	1	1	0	0	1
E	0	0	1	1	0
F	0	1	1	0	1
G	1	0	1	0	1
H	1	1	1	0	1

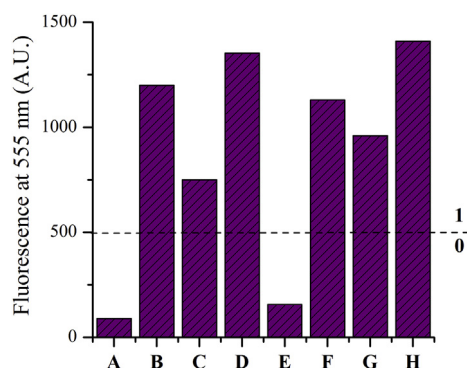
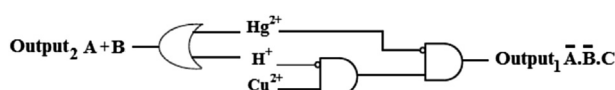


Fig. 8. The changes in fluorescence intensity of Dyad 5 ($C = 10^{-5} \text{ mol L}^{-1}$) at 555 nm with 2 eq. of protons ($C = 2 \times 10^{-5} \text{ mol L}^{-1}$), 1 eq. Cu^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) and 1 eq. Hg^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) as chemical inputs.



Scheme 5. Combinatorial logic circuits on the basis of 1 eq. of Dyad 5 ($C = 10^{-5} \text{ mol L}^{-1}$) and 2 eq. of protons ($C = 2 \times 10^{-5} \text{ mol L}^{-1}$), 1 eq. of Cu^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) and 1 eq. of Hg^{2+} ($C = 10^{-5} \text{ mol L}^{-1}$) as chemical inputs.

binding process occurs only in the receptor unit (1:1 binding stoichiometry), which opens the rhodamine spirolactam ring and allows energy transfer to the rhodamine moiety. Due to the remarkable fluorescence changes in the presence of protons, Cu^{2+} and Hg^{2+} ions the novel system is able to execute OR and INHIBIT logic gates and to act as a two output combinational logic circuit with three chemical inputs.

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