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STABILITY CONSTANT OF THE 1:1 COMPLEX OF SODIUM WITH GUANOSINE 5'-MONOPHOSPHATE

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ABSTRACT The stability of the 1:1 complex of sodium ion with the dianion of guanosine 5'-monophosphate has been determined by means of a potentiometric titration employing a specific ion electrode. The stability constant for the reaction $Na^+ + 5'$ - $GMP^{2-} \rightleftharpoons Na(5'$ - $GMP)^-$ was found to be 2.85 \pm 0.36 M^{-1} at 5°C and an ionic strength of 1.1 \pm 0.1 M. Although 5'-GMP forms ordered self-structures at high concentration in the presence of sodium ions, in dilute solution and at low sodium ion concentrations the Na^+ binding is weak and typical of that for other nucleotides.

INTRODUCTION

In weakly acidic solution (pH \sim 5) guanosine 5'-monophosphate (5'-GMP) spontaneously forms an anisotropic gel (1), but under neutral or slightly basic conditions where the nucleotide is a dianion, soluble aggregates having an ordered structure are formed (2, 3). It has been postulated that self-aggregation occurs through the stacking of planar tetramer units which are formed via interbase hydrogen bonds between donor positions N(1)H and N(2)H and acceptor positions O(6) and N(7) (3-5). The presence of a specific cation is required, and the extent and nature of the self-association appears to be dependent on the size of the cation. Among the alkali metal ions, Na⁺, K⁺, and Rb⁺ promote self-association, whereas Li⁺ and Cs⁺ exhibit little or no tendency to facilitate aggregate formation (6-8). The ordered solution structures of 5'-GMP formed in the presence of Na⁺ are believed to be predominantly octamers. The octamers and species composed of more than two tetrameric units give rise to nonequivalent H(8) environments in the ¹H NMR spectrum (3, 6) and to IR frequency shifts for the C—O and C—N stretching vibrations of the nucleotide.

Although various models regarding the kind and number of Na⁺ binding sites in aggregated 5'-GMP have been proposed (6, 7), the binding of Na⁺ to the monomeric 5'-GMP has not been investigated. In an attempt to understand the role of sodium in the solution chemistry of 5'-GMP, a study of Na⁺ binding to the nucleotide was undertaken using a specific ion electrode. The investigation was carried out at a nucleotide concentration where no ordered structure is present in solution as judged by NMR spectroscopy. It was thought that either the stoichiometry

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of the Na⁺ binding and/or the magnitude of the binding constant might provide insight into the role of the cation in the aggregation of 5'-GMP.

MATERIALS AND METHODS

The 5'-GMP used in the titrations was in the form of the tetramethylammonium (TMA) salt because it has been shown that self-association does not occur in the presence of this cation (9). (TMA)₂(5'-GMP) was prepared by titrating guanylic acid $[H_2(5'-GMP) \cdot H_2O; Sigma Chemical Co., St. Louis, MO; 1 g dissolved in 100 ml of deionized distilled water] with 0.1 M TMA hydroxide (Matheson, Coleman and Bell, Gibbstown, NJ) to a pH of 7.9–8.0. The solid was obtained by freeze drying. Solutions for titration were prepared by dissolving solid (TMA)₂(5'-GMP) or dried NaCl in 0.80 M TMA chloride (Aldrich Chemical Co., Inc., Milwaukee, WI). The 5'-GMP²⁻ concentration was determined spectrophotometrically at 252 nm (<math>\epsilon$ = 13,700).

The ionic medium method of Sillen (10) was used. A portion of 0.1 M (TMA)₂(5'-GMP) was pipetted into the cell and equilibrated with stirring at 5°C. The electrode and thermocouple were placed in the solution. One buret was filled with 0.2 M (TMA)₂(5'-GMP) and another buret was filled with standard NaCl solution (1.00 or 3.00 M). The electrode potential was read and a measured portion of NaCl and approximately equal amount of (TMA)2(5'-GMP) were added. The solution was allowed to equilibrate and the electrode stabilize for 10-15 min before taking a reading. The 5'-GMP²⁻ concentration was held constant at 0.089 M and the TMA+ concentration was 1.0 M throughout the titration. The titrations were done at pH 8.0 \pm 0.1 and an ionic strength of 1.1 ± 0.1 M. The electrode was calibrated immediately preceding a titration by adding portions of standard NaCl solutions in 1.0 M TMA chloride to 1.0 M TMA chloride at 5°C. A plot of electrode potential (mV) vs. log [Na+] was linear for sodium ion concentrations above 10⁻² M. The slope and intercept of the calibration plot were determined using a linear least-squares treatment.

The free [Na $^+$] was measured with an Orion model 9611 (Orion Research Inc., Cambridge, MA) sodium ion combination electrode connected to an Analogic millivolt meter (model AN2570 d XIP; Analogic Corp., Wakefield, MA) via a high impedance buffer. The titrations were carried out inside a Faraday cage. The cell was constructed of glass and consisted of a small jacketed beaker. The temperature was maintained at 5 \pm 1°C by circulating H₂O-ethanol through the cell jacket using a Forma Scientific Model 2095 bath. The

temperature of the solution was determined by a glass-encased copperconstantan thermocouple connected to a calibrated Omega Engineering meter (Omega Engineering, Inc., Stamford, CT). The solution was stirred by means of an air-driven magnetic stirrer. The standard solutions were added to the reaction solution from 5-ml burets having scale divisions of 0.01 ml.

RESULTS AND DISCUSSION

The binding of Na⁺ to disordered 5'-GMP was found to obey the expressions:

$$Na^{+}(aq) + 5' - GMP^{2-}(aq) \rightleftharpoons Na(5' - GMP)^{-}(aq)$$
 (1)

$$K = [Na(5'-GMP)^{-}]/\{[Na^{+}][5'-GMP^{2-}]\}.$$
 (2)

Based on the combined data from two independent titrations, the value of K at 5°C was found to be 2.85 \pm 0.36 M^{-1} . The experimental data and theoretical curve are shown in Fig. 1. Several other models were tried in which the ratio of Na⁺ to 5'-GMP²⁻ in the complex was assumed to be 2:1, 5:4, and 1:n, where n = 2-4. None of these models fit the experimental data.

The 1:1 Na(5'-GMP) complex most likely involves the interaction of Na+ with a phosphate oxygen, because the magnitude of K is typical of the values found for other nucleotide systems. Smith and Alberty (11) found that the apparent stability constant for Na(5'-AMP) was 2.2 ± 0.2 M⁻¹ at 25°C and an ionic strength of 0.2 M with TMA bromide as the supporting electrolyte. They also determined K for the binding of one Na⁺ to the orthophosphate anion and obtained a value of 3.9 \pm 0.4 M⁻¹ under the same conditions. A Donnan equilibrium study by Strauss and co-workers (12) gave values of K for the binding of Na⁺ to DNA in the range 0.7 to 1.6 M⁻¹ at 25°C. Although a direct comparison between these stability constants and that for Na(5'-GMP) cannot be made because the determinations were done at differing ionic strengths and temperatures, it can be seen that they are all of similar magnitude.

Since Na⁺ binding to the disordered nucleotide is quite weak and typical of ion pairing to phosphate oxygen, it

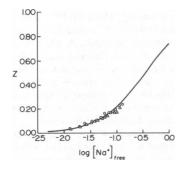


FIGURE 1 Titration curve for the reaction of 5'-GMP²⁻ with Na⁺. O, \triangle , experimental data; —, theoretical curve for 1:1 complex; $Z = ([Na^+]_{total} - [Na^+]_{free})/[5'-GMP^{2-}]_{total}$.

appears that the initial role of Na⁺ is that of partial neutralization of the negative charge of 5'-GMP. It is reasonable to think that this would be a necessary prerequisite for the formation of large 5'-GMP aggregates. However, charge neutralization is probably not the only or the major mechanism by which Na⁺ interacts in the formation of large self-aggregates. The critical aggregation reaction is metal ion-size dependent and may involve the binding of Na⁺ to donor groups from several 5'-GMP units in the ordered species.

Using calorimetry (13) and ultraviolet hypochromicity (1), Chantot and Guschlbauer found evidence for the presence of unstacked tetramers in dilute aqueous solutions of 3'-GMP. Although an unstacked tetramer has not been observed for 5'-GMP, it is a possible structure in dilute solution. Since each 5'-GMP molecule in such a tetramer would be expected to be in rapid exchange with monomer, averaging of the NMR resonances would result. From our measurements, species such as $Na_n(5'-GMP)_n^{n-}$, where n > 1, are indistinguishable from $Na(5'-GMP)^{-}$. Therefore we cannot confirm the existence or absence of an unstacked tetrameric species in the solutions.

Finally, we note that in the titration of 0.089 M 5'-GMP, deviations from 1:1 complex formation were observed at Na⁺ concentrations above 0.15 M. Although we have not determined the nature of the species being formed, analysis indicates an increase in the number of Na⁺ bound per 5'-GMP over that for the 1:1 complex.

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