ELSEVIER

Contents lists available at SciVerse ScienceDirect

Steroids





Mechanistic insights on the reactivity of furospirostanes with the 16β,22:22,25-diepoxy-23-acetoxymethyl-24-methyl side chain



Mariana Macías-Alonso, Marcos Flores-Álamo, Martín A. Iglesias-Arteaga*

Facultad de Ouímica, Universidad Nacional Autónoma de México, Ciudad Universitaria, 04510 México, DF, Mexico

ARTICLE INFO

Article history: Received 21 March 2013 Received in revised form 16 April 2013 Accepted 9 May 2013 Available online 23 May 2013

Keywords: Furospirostanes Bromination Allylic substitution Elimination Bromodeacetoxylation

ABSTRACT

F-ring opening in spirostanes with the 16β ,22:22,25-diepoxy-23-acetoxymethyl-24-methyl side chain produces a Δ^{22} -intermediate with an allylic acetoxy group. For this reason the reactivity profile of these compounds deviates from that observed in other naturally occurring or synthetic spirostanes and furospirostanes

© 2013 Elsevier Ltd. All rights reserved.

1. Introduction

The spiroketal assemblies (see Fig. 1) are present in widespread compounds that can be isolated from many marine and terrestrial organisms: plants, fungi and insects among others. The vast number and the growing pharmacological importance of compounds containing spiroketal assemblies have triggered increasing interest in both their synthesis and studies of their chemical reactivity [1–5]. In addition to the spirostanic sapogenins which bear the 1,6-dioxaspiro[4,5] decane moiety, furospirostanes, an emerging family of steroids bearing the 16β ,22:22,25-diepoxy moiety in the chain, that are considered 1,6-dioxaspiro[4,4] nonane derivatives are attracting increasing attention.

The increasing number of naturally occurring bioactive steroids that bear the furospirostane side chain includes compounds that have shown interesting antitumor activity such as ritterazines (1), cephalostatins (2) [6-10], hippuristanols (3) [11-15], as well as the antihypertensive glycosides of nuatigenin (4) [16] among others (see Fig. 2).

Unlike spirostanic sapogenins, the side-chain reactivity of which has received considerable attention in the past 70 years selected Ref. [17–37], the studies of the reactivity of the furospirostane side chain are rather scarce [38–42]. In particular, Fuchs and coworkers hypothesized that the cytotoxic activity of cephalostatins may be related to the possibility of the generation of oxacarbenium ions around C-22 [43–44]. This fact, added to the increasing

number of cytotoxic steroids bearing the furospirostane side chain that have been identified, prompts the exploration of the reactivity of this moiety.

As a part of our ongoing program directed to the synthesis of potentially bioactive spiroketals derived from rearranged steroid sapogenins, we envisaged the bromination at C-23 of our previously described furospirostane sapogenin **23s-5** [45] as a possible route to the introduction of additional functionality in the F ring (see Scheme 1).

In the course of the bromination experiments employing pyridine hydrobromide perbromide (PyrBr $_2$ ·HBr) we found that, in addition to the desired bromination at C-23, an unexpected substitution of the C-23′ acetoxy moiety also took place. This led us to explore the particular reactivity of this structural fragment. Herein we describe our findings on the reactivity of the 16β ,22:22,25-diepoxy-23-acetoxymethyl-24-methyl side chain.

2. Experimental

Reactions were monitored by TLC on ALUGRAM® SIL G/UV254 plates from MACHEREY–NAGEL. Chromatographic plates were sprayed with a 1% solution of vanillin in 50% HClO₄ and heated until color developed. Purifications and separations were performed in pressurized chromatographic columns packed with MACHEREY–NAGEL silica gel 60 (230–400 mesh ASTM). Melting points were measured on a Melt-Temp II apparatus. Mass spectra were registered in a Thermo-Electron spectrometer model DFS (Double Focus Sector). NMR spectra were recorded in CDCl₃ solution in a Varian INOVA 400 spectrometer using the solvent signal

^{*} Corresponding author. Tel./fax: +52 55 56223803.

E-mail address: martin.iglesias@unam.mx (M.A. Iglesias-Arteaga).

Fig. 1. Common dioxaspiro ketal moieties.

7.26 ppm for ¹H and 77.00 ppm for ¹³C as references. NMR signals assignments were made with the aid of DEPT and a combination of 2D homonuclear (¹H-¹H) and heteronuclear (¹H-¹³C) correlation techniques, which included ¹H-¹H COSY, ¹H-¹H Nuclear Overhauser Effect Spectroscopy (NOESY), and Heteronuclear Single Quantum Correlation (HSQC) [46]. All 2D NMR spectra were recorded using the standard pulse sequences and parameters recommended by the manufacturer.

3. Bromination of 23S-5 with PyH⁺Br₃⁻

 $PyH^+Br_3^-$ (1.28 g, 3.6 mmol) was added to a solution of **235-5** (518 mg, 1.0 mmol) [45] in acetic acid (25 mL). The mixture was stirred for 1 h at 50 °C and poured into ice/water. The produced solid was filtered off washed with water and dissolved in CH_2Cl_2 . The organic solution was dried (anh. $Na_2SO_4)$ and evaporated to

produce a mixture of **23R-6**, **23S-6**, **23R-7** and **23S-7** that were separated in a pressurized chromatographic column packed with silica gel (18.5 g) using hexane/ethyl acetate 15:1 as eluent. Order of elution **23R-7**, **23S-7**, **23S-6**, **23R-6**.

(22S,23R,24S)-16β,22:22,25-diepoxy-23-bromo-23-hydroxymethyl-24-methyl-26,27-dinor-5 β -cholestan-3 α -ol (23R-6). Yield 391.7 mg (0.658 mmol, 65.8%). Mp. 262-264 °C (from ethyl acetate/hexane). ¹H NMR (400 MHz, CDCl₃, δ ppm): 4.71 (m, 1H, H-3), 4.44 (ddd, J = 8.6, 7.6, 6.1 Hz, 1H, H-16), 4.35 (d, J = 11.8 Hz, 1H, H-23'a), 4.23 (d, J = 11.8 Hz, 1H, H-23'b), 4.06(dd, J = 8.1 Hz, 1H, H-25 Pro-S), 3.55 (dd, J = 8.2, 7.1 Hz, 1H, H-25)Pro-R), 2.44 (dq, J = 6.7, 6.7 Hz, 1H, H-20), 2.29 (m, 1H, H-24), 2.11 (s, 3H, CH₃COO-23'), 2.01 (s, 3H, CH₃COO-3), 1.18 (d, I = 6.9 Hz, 3H, H-24'), 1.02 (d, I = 6.9 Hz, 3H, H-21), 0.94 (s, 3H, H.19), 0.88 (s, 3H, H-18). ¹³C NMR (100 MHz, *CDCl*₃, δ ppm): C-1 35.0, C-2 26.6, C-3 74.2, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.5, C-8 35.4, C-9 40.5, C-10 34.7, C-11 20.5, C-12 40.1, C-13 41.6, C-14 56.2, C-15 31.7, C-16 81.0, C-17 63.3, C-18 16.4, C-19 23.3, C-20 35.5, C-21 16.3, C-22 117.4, C-23 73.4, C-23' 69.2, C-24 38.1, C-24' 19.0, C-25 70.7, CH₃COO-23' 170.5, CH₃COO-23 20.8, CH₃-COO-3 170.6, CH₃COO-3 21.4. MS (EI) 515 [M⁺-Br], 454, 426, 394, 366, 344, 329, 284, 269, 255, 215, 161, 147, 137 (100%). HRMS (EI) observed 515.3379 [M^+ -Br]; required for $C_{31}H_{47}O_6$ 515.3372.

(22S,23S,24S)- $16\beta,22:22,25$ -diepoxy-23-bromo-23-hydroxy-methyl-24-methyl-26,27-dinor- 5β -cholestan- 3α -ol diacetate **(23S-6)**. Yield 53.4 mg, (0.09 mmol, 9.0%). Mp. 266-267 °C (*from*

Fig. 2. Some naturally occurring furospirostanes.

a) Nucleophilic substitition; b) elimination

Scheme 1. Functionalization of the furospirostane side chain.

ethyl acetate/hexane). ¹H NMR (400 MHz, *CDCl*₃, δ ppm): 4.71 (m, 1H, H-3), 4.47 (m, 1H, H-16), 4.47 (s, 2H, H-23'), 4.31 (dd, J = 7.9 Hz, 1H, H-25 Pro-S), 3.57 (dd, J = 8.2, 3.9 Hz, 1H, H-25 Pro-R), 2.89 (m, 1H, H-24), 2.52 (m, 1H, H-20), 2.13 (s, 3H, CH₃COO-23'), 2.02 (s, 3H, CH₃COO-3), 1.26 (d, J = 6.9 Hz, 3H, H-21), 1.16 (d, J = 7.3 Hz, 3H, H-24'), 0.93 (s, 3H, H-19), 0.81 (s, 3H, H-18). ¹³C NMR (100 MHz, $CDCl_3$, δ ppm): C-1 35.0, C-2 26.5, C-3 74.2, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.5, C-8 35.4, C-9 40.5, C-10 34.7, C-11 20.4, C-12 39.7, C-13 41.3, C-14 56.3, C-15 31.9, C-16 81.6, C-17 63.9, C-18 16.4, C-19 23.3, C-20 39.4, C-21 18.2, C-22 119.1, C-23 73.8, C-23' 65.9, C-24 46.5, C-24' 16.7, C-25 72.6, CH₃-COO-3 170.6, CH₃COO-23' 170.6, CH₃COO-3 21.5, CH₃COO-23' 21.0. MS (EI) [M⁺-Br], 454, 426, 329, 315, 284, 269, 255, 215, 159, 147, 137 (100%). HRMS (EI) observed 515.3377 [M⁺-Br]; required for C₃₁H₄₇O₆ 515.3372.

Crystals of 23R-6 and 23S-6 mounted on glass fiber were studied with Oxford Diffraction Gemini "A" diffractometer with a CCD area detector ($\lambda_{cuk\alpha}$ = 1.54184 Å, monochromator: graphite) source equipped with a sealed tube X-ray source. Unit cell constants were determined with a set of 15/3 narrow frame/runs (1° in ω) scans. Data sets consisted of 1154 and 684 frames of intensity data collected with a frame width of 1° in ω for **23R-6** and **23S-6** respectively with a crystal-to-detector distance of 55.00 mm. The double pass method of scanning was used to exclude any noise. The collected frames were integrated by using an orientation matrix determined from the narrow frame scans. CrysAlisPro and CrysAlis RED software packages [47] were used for data collection and data integration. Structure solution and refinement were carried out with the programs SHELXS97 and SHELXL97 [48]. For molecular graphics, ORTEP-3 for Windows was employed, and the software used to prepare material for publication was WinGX [49].

Full-matrix least-squares refinement was carried out by minimizing $(Fo^2 - Fc^2)^2$. All non-hydrogen atoms were refined anisotropically. H atoms attached to C atoms were placed in geometrically idealized positions and refined as riding on their parent atoms, with C-H = 0.96–0.99 Å with $U_{\rm iso}({\rm H})$ = 1.2 $U_{\rm eq}({\rm C})$ for methine and methylene groups, and $U_{\rm iso}({\rm H})$ = 1.5 $U_{\rm eq}({\rm C})$ for methyl

group. Crystal data and experimental details of the structure determination are listed in Table 1 [50].

(22S,23R,24S)-16β,22:22,25-diepoxy-23-bromo-23-bromomethyl-24-methyl-26,27-dinor-5 β -cholestan-3 α -ol acetate (23R-**7)**. Yield 24.3 mg, (0.039 mmol, 3.9%). Yellow oil. ¹H NMR (400 MHz, $CDCl_3$, δ ppm): 4.71 (m, 1H, H-3), 4.48 (ddd, J = 7.8, 7.8, 5.1 Hz, 1H, H-16), 4.36 (dd, J = 8.1, 7.1 Hz, 1H, H-25 Pro-S), 4.03 (d, J = 11.6 Hz, 1H, 23'a), 3.70 (d, J = 11.6 Hz, 1H, 23'b), 3.66(dd, J = 8.2, 2.4 Hz, 1H, H-25 Pro-R), 2.80 (qdd, J = 7.2, 7.2, 2.3 Hz, 1H, H-24), 2.42 (m, 1H, H-20), 2.01 (s, 3H, CH₃COO-), 1.28 (d, J = 7.3 Hz, 3H, H-24'), 1.27 (d, J = 6.8 Hz, 3H, H-21), 0.94 (s, 3H, H-19), 0.77 (s, 3H, H-18). ¹³C NMR (100 MHz, *CDCl*₃, δ ppm): C-1 35.0, C-2 26.6, C-3 74.2, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.5, C-8 35.5, C-9 40.5, C-10 34.7, C-11 20.4, C-12 39.6, C-13 41.5, C-14 56.3, C-15 32.0, C-16 81.6, C-17 63.8, C-18 16.3, C-19 23.3, C-20 39.4. C-21 18.7. C-22 119.3. C-23 77.3. C-23' 38.1. C-24 46.6. C-24' 17.0, C-25 73.2, CH₃COO-3 170.6, CH₃COO-3 21.4, MS (EI) 535 $[^{79}Br_2M^+ - ^{79}Br]$, 456, 426, 344, 329, 284, 269, 255, 227, 215, 175, 149, 137 (100%). HRMS (EI) observed 535.2415 [79Br₂M⁺-79Br] required for $C_{29}H_{44}^{79}BrO_4$ 535.2423.

(22S,23S,24S)-16β,22:22,25-diepoxy-23-bromo-23-bromomethyl-24-methyl-26,27-dinor-5 β -cholestan-3 α -ol acetate (23S-**7)** yield 66.0 mg, (0.107 mmol, 10.7%). Yellow oil. ¹H NMR (400 MHz, $CDCl_3$, δ ppm): 4.72 (m, 1H, H-3), 4.44 (m, 1H, H-16), 4.08 (dd, J = 8.2, 8.2 Hz, 1H, H-25 Pro-S), 3.88 (d, J = 10.8 Hz, 1H, H-23'a), 3.75 (d, J = 10.8 Hz, 1H, H-23'b), 3.54 (dd, J = 8.2, 6.8 Hz, 1H, H-25 Pro-R), 2.53 (m, 1H, H-24), 2.47 (m, 1H, H-20), 2.02 (s, 3H, CH₃COO), 1.18 (d, J = 6.8 Hz, 3H, H-21), 1.06 (d, J = 6.9 Hz, 3H, H-24'), 0.95 (s, 1H, H-19), 0.92 (s, 3H, H-18). ¹³C NMR (100 MHz, CDCl₃, δ ppm): C-1 35.0, C-2 26.6, C-3 74.3, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.5, C-8 35.5, C-9 40.5, C-10 34.7, C-11 20.5, C-12 40.1, C-13 41.7, C-14 56.2, C-15 31.7, C-16 81.1, C-17 63.5, C-18 16.42, C-19 23.3, C- 20 35.4, C-21 16.8, C-22 117.5, C-23 75.6, C-23' 43.9, C-24 39.5, C-24' 19.7, C-25 70.4, CH₃COO-3 170.6, CH₃-COO-3 21.5. MS (EI) 535 [⁷⁹Br₂M⁺-⁷⁹Br], 456, 426, 344, 329, 315, 269, 255, 159, 149, 137 (100%). HRMS (EI) observed 535.2413 $[^{79}Br_2M^+ - ^{79}Br]$ required for $C_{29}H_{44}^{79}BrO_4$ 535.2423.

Table 1
Crystal data and structure refinement for 23R-6 and 23S-6.

	23R-6	23S-6
Empirical formula	$C_{31}H_{47}BrO_6$	$C_{31}H_{47}BrO_6$
Formula weight	595.60	595.60
Temperature (K)	298(2) K	298(2) K
Wavelength (Å)	1.54184	1.54184 Å
Crystal system	Monoclinic	Monoclinic
Space group	P21	P21
Unit cell dimensions (Å)	a = 11.0378(4)	a = 13.2911(4)
	b = 7.8265(3)	b = 7.7902(2)
	c = 17.8645(6)	c = 14.5238(3)
	$\beta = 104.658(4)$	$\beta = 98.487(2)$
Volume (Å ³)	1493.04(9)	1487.33(7)
Z	2	2
Crystal size (mm ³)	$0.2171 \times 0.1503 \times 0.0478$	$0.4685 \times 0.257 \times 0.0418$
Density (calculated) (Mg/m ³)	1.325	1.330
Absorption coefficient (mm ⁻¹)	2.202	2.211
F(000)	632	632
Theta range for data collection	4.14°-68.11°	4.21°-68.07°
Index ranges	$-11 \le h \le 13, -8 \le k \le 9, -20 \le 1 \le 21$	$-10 \le h \le 15, -7 \le k \le 9, -17 \le l \le 17$
Reflections collected	10014	5346
Independent reflections	4696 [R(int) = 0.0451]	3737 [R(int) = 0.0220]
Completeness to theta = 66.50°	100.0%	94.6%
Refinement method	Full-matrix least-squares on F ²	Full-matrix least-squares on F ²
Data/restraints/parameters	4696/1/349	3737/1/349
Goodness-of-fit on F ²	0.964	1.103
Final R indices $[I > 2 \text{sigma}(I)]$	R1 = 0.0405, w $R2 = 0.0939$	R1 = 0.0369, w $R2 = 0.0870$
R indices (all data)	R1 = 0.0551, $wR2 = 0.1004$	R1 = 0.0455, w $R2 = 0.0919$
Largest diff. peak and hole	0.299 and -0.255 e.Å ⁻³	$0.485 \text{ and } -0.453 \text{ e.Å}^{-3}$

Scheme 2. Bromination of the furospirostane side chain.

4. Reaction of 23S-5 with HBr in acetic acid

The diacetate **23S-5** (258.4 mg, 0.5 mmol) was added to a 33 wt.% solution of HBr in acetic acid (5 mL, 27.2 mmol of HBr). The mixture was stirred for 45 min., poured into ice/water and extracted with ethyl acetate (40 mL). The organic layer was washed with 5% aqueous Na₂CO₃ (4 \times 25 mL) and water (2 \times 25 mL), dried (anh. Na₂SO₄) and evaporated to produce a mixture of **23R-8** and **23S-8** that were separated in pressurized chromatographic column packed with silica gel (12 g) using hexane/ethyl acetate 40:1 as eluent. Order of elution **23S-8**, **23R-8**.

(22S, 23R, 24R)-16β,22:22,25-diepoxy-23′-bromo-24-methyl-26,27-dinor-5β-cholestan-3α-ol acetate **(23R-8)** yield 41.5 mg, (0.077 mmol, 15.4%). Yellow oil. ¹H NMR (400 MHz, $CDCl_3$, δ ppm): 4.72 (m, 1H, H-3), 4.41 (ddd, J = 8.5, 7.6, 6.0 Hz, 1H, H-16), 3.97 (dd, J = 8.0 Hz, 1H, H-25 Pro-S), 3.57 (dd, J = 10.1, 6.4 Hz, 1H, H-23′a), 3.43 (dd, J = 9.2, 8.3 Hz, 1H, H-25 Pro-R), 3.34 (dd, J = 10.1, 8.3 Hz, 1H, H-23′b), 2.36 (m, J = 13.4, 6.8 Hz, 1H, H-20), 2.27 (m, 1H, H-23), 2.02 (s, 3H, CH₃COO-3), 1.13 (d, J = 6.7 Hz, 3H, H-24′), 1.00 (d, J = 6.9 Hz, 3H, H-21), 0.95 (s, 3H, H-19), 0.82 (s, 3H, H-18). ¹³C NMR (75 MHz, $CDCl_3$, δ ppm): C-1 35.0, C-2 26.6, C-3 74.3, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.5, C-8 35.4,

C-9 40.5, C-10 34.7, C-11 20.6, C-12 40.1, C-13 41.2, C-14 56.2, C-15 31.8, C-16 81.0, C-17 62.9, C-18 16.6, C-19 23.3, C-20 36.1, C-21 16.2, C-22 119.5, C-23 55.7, C-23′ 33.5, C-24 39.9, C-24′ 17.2, C-25 71.4, CH₃COO-3 170.6, CH₃COO-3 21.5. MS (FAB) MH⁺ 539, MH⁺ 537, 459, 457, 255, 147, 133, 91, 83, 73, 57 (100%). HRMS (FAB) observed 537.2500 [MH⁺] required for $C_{29}H_{46}^{79}BrO_{4}$ 537.2579.

(22S, 23S, 24R)-16β,22:22,25-diepoxy-23'-bromo-24-methyl-26,27-dinor-5β-cholestan-3α-ol acetate **(23S-8)** yield 40.6 mg, (0.076 mmol, 15.2%). Yellow oil. 1 H NMR (400 MHz, $CDCl_3$ δ ppm): 4.71 (m, 1H, H-3), 4.44 (m, 1H, H-16), 3.95 (dd, J = 8.3, 6.4 Hz, 1H, H-25 Pro-S), 3.63 (dd, J = 8.3, 2.7 Hz, 1H, H-25 Pro-R), 3.44 (m, 2H, H-23'), 2.47 (m, 1H, H-24), 2.47 (m, 1H, H-23), 2.14 (m, 1H, H-20), 2.02 (s, 3H, CH₃COO-3), 1.13 (d, J = 6.8 Hz, 3H, H-24'), 0.97 (d, J = 6.9 Hz, 3H, H-21), 0.94 (s, 3H, H-19), 0.75 (s, 3H, H-18). 13 C NMR (75 MHz, $CDCl_3$ δ ppm): C-1 35.0, C-2 26.6, C-3 74.3, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.6, C-8 35.3, C-9 40.5, C-10 34.7, C-11 20.6, C-12 40.1, C-13 41.0, C-14 56.1, C-15 31.8, C-16 81.7, C-17 61.1, C-18 16.6, C-19 23.3, C-20 38.2, C-21 14.4, C-22 118.3, C-23 48.9, C-23' 29.2, C-24 34.0, C-24' 14.3, C-25 73.9, CH₃COO-3 170.6, CH₃COO-3 21.5. MS (FAB) MH⁺ 539, MH⁺ 537, 457, 455, 397, 315, 255, 147, 133, 107, 95, 91, 69, 57, 55 (100%).

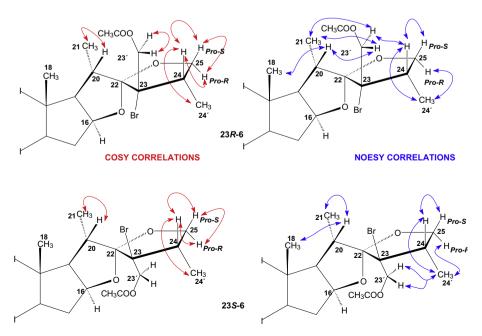


Fig. 3. Selected COSY and NOESY correlation observed in compounds 23R-6 (upper) and 23S-6 (lower).

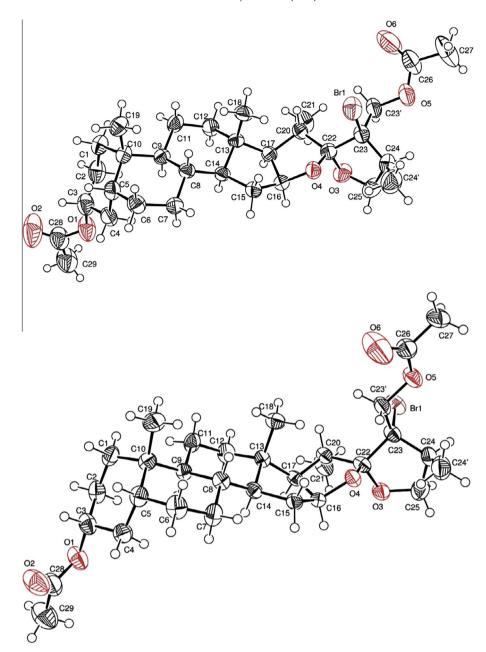


Fig. 4. Crystal structures of 23R-6 (upper) and 23S-6 (lower) with the thermal ellipsoids drawn at 50% of probability.

HRMS (FAB) observed 537.2495 [MH $^{+}$] required for $C_{29}H_{46}^{79}BrO_{4}$ 537.2579.

5. Reaction of 23S-5 with acetic acid and $BF_3 \cdot Et_2O$

BF $_3$ ·Et $_2$ O (1.5 mL, 12.2 mmol) was added to a solution of **23S-5** (518 mg, 1.0 mmol) in acetic acid and the mixture was stirred for 5 h before slow addition of 10% aqueous NaHCO $_3$ solution (20 mL) and extraction with ethyl acetate (25 mL). The organic layer was washed with 10% aqueous NaHCO $_3$ solution (8 × 20 mL) water (3 × 20 mL), dried (anh. Na $_2$ SO $_4$) and evaporated to afford a mixture of the starting material, **23S-5**, the epimeric **23R-5**, and olefin **9** that were separated in pressurized chromatographic column packed with silica gel (15 g) using hexane/ethyl acetate 97:3 as eluent. Order of elution **9**, **23R-5**, **23S-5**.

(22S, 23R, 24R)-16 β ,22:22,25-diepoxy-23-hydroxymethyl-24-methyl-26,27-dinor-5 β -choles-tan-3 α -ol diacetate **(23R-5)**. Yield

86.1 mg, (0.167 mmol, 16.7%). Yellow oil. ¹H NMR (400 MHz, CDCl₃, δ ppm): 4.72 (m, 1H, H-3), 4.45 (m, 1H, H-16), 4.18 (d, J = 6.5 Hz, 2H, H-23'), 3.95 (dd, J = 8.2, 6.7 Hz, 1H, H-25 *Pro-S*), 3.56 (dd, J = 8.3, 3.6 Hz, 1H, H-25 Pro-R), 2.38 (m, 1H, H-23), 2.38 (m, 1H, H-24), 2.23 (dq, J = 6.9. 6.9 Hz, 1H, H-20), 2.04 (s, 3H, CH₃COO-23'), 2.02 (s, 3H, CH₃COO-3), 1.05 (d, J = 6.7 Hz, 3H, H-24'), 0.97 (d, J = 6.9 Hz, 3H, H-21), 0.94 (s, 3H, H-19), 0.76 (s, 3H, H-18). ¹³C NMR (400 MHz, $CDCl_3$, δ ppm): C-1 35.0, C-2 26.6, C-3 74.3, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.6, C-8 35.4, C-9 40.5, C-10 34.7, C-11 20.6, C-12 40.2, C-13 40.9, C-14 56.2, C-15 31.8, C-16 81.6, C-17 61.2, C-18 16.6, C-19 23.3, C-20 38.2, C-21 14.4, C-22 118.4, C-23 44.1, C-23' 61.5, C-24 33.6, C-24' 14.7, C-25 73.9, CH₃COO-3 170.6, CH₃COO-3 21.5, CH₃COO-23' 171.0, CH₃COO-23' 21.0. MS (FAB) 457 [M⁺-CH₃COO], 455, 397, 327, 281, 255, 207, 193, 147, 136, 91, 73(100%). HRMS (FAB) observed 457.3319 [M⁺-CH₃COO] required for C₂₉H₄₅O₄ 457.3318.

(22S, 24R)- 16β ,22:22,25-diepoxy-23-methyliden-24-methyl-26,27-dinor- 5β -choles-tan- 3β -ol diacetate **(9)** yield 30.2 mg,

Scheme 3. Reaction mechanism for the bromination at C-23.

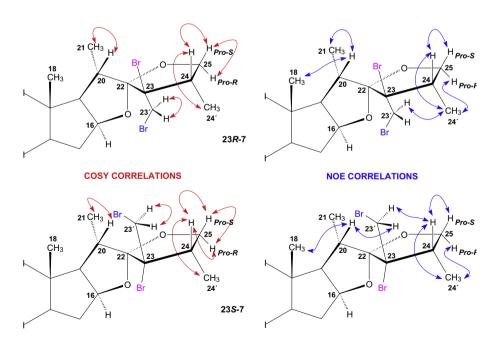


Fig. 5. Selected COSY and NOE correlations in the side chains of compounds 23R-7 (upper) and 23S-7 (lower).

 J = 6.9 Hz, 3H, H-21), 0.87 (s, 3H, H-18). ¹³C NMR (400 MHz, $CDCl_3$, δ ppm): C-1 35.0, C-2 26.6, C-3 74.3, C-4 32.2, C-5 41.8, C-6 26.9, C-7 26.6, C-8 35.4, C-9 40.5, C-10 34.7, C-11 20.6, C-12 40.2, C-13 41.1, C-14 56.4, C-15 31.9, C-16 82.4, C-17 61.6, C-18 16.4, C-19 23.3, C-20 41.1, C-21 14.6, C-22 118.3, C-23 153.9, C-23′ 107.1, C-24 38.3, C-24′ 14.7, C-25 72.1, CH₃COO-3 170.6, CH₃COO-3

Scheme 4. Possible reaction mechanism for bromodeacetoxylation of 23S-5.

Scheme 5. Reaction of 23S-5 with HBr in acetic acid.

21.4. MS (EI) 457 [MH $^{+}$], 456, 426, 396, 366, 329, 315, 284, 269, 255, 215, 161, 149, 137 (100%). HRMS (EI) observed 456.3250 [M $^{+}$] required for $C_{29}H_{44}O_4$ 456.3240.

6. Results and discussion

Treatment of the diacetate **23S-5** with PyrBr₂·HBr in acetic acid afforded the epimeric 23-brominated furospirostanes **23R-6** and

23S-6 together with the unexpected dibrominated compounds **23R-7** and **23S-7** (see Scheme 2).

Although the MS of compound **23R-6** does not show a molecular ion that shows the presence of a bromine, the occurrence of a fragment in m/z 515 may be interpreted as the product of the loss of Br from the molecular ion (M^+ –Br or more likely MH^+ –HBr). The signals of C-16, C-22 and C-26 in 13 C spectrum of compound **23R-6** evidence the integrity of the 16,22:22,25 diepoxy moiety, while the

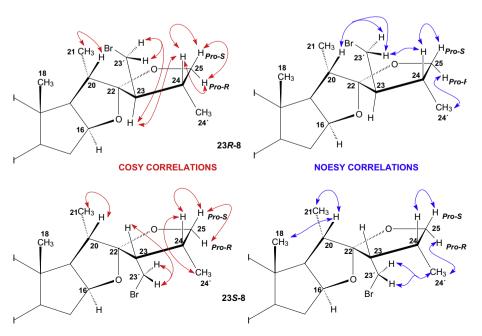


Fig. 6. Selected COSY and NOESY correlations observed in compounds 23R-8 and 23S-8.

Scheme 6. Possible reaction mechanism of the substitution of the C-23'-acetoxy moiety.

Scheme 7. BF3·Et²O-catalyzed reaction of 23S-5 in acetic acid.

downfield shift of the signal of C-23 suggests the substitution of H-23 by a more electronegative atom that, according the reaction conditions, must be no other than a bromine atom. In addition, the substitution of H-23 is evidenced by the disappearance of its NMR signal and the conversion of the H-23' signals from an AB-X system into a doublet, product of the H-23'a ↔ H-23'b geminal coupling, (I = 11.8 Hz). The observed COSY and NOE correlations also corroborate the proposed structure (see Fig. 3). In particular, the NOE correlations of each H-23' with H-20 and H-24 in the respective β -side of rings E and F prove their spatial proximity only possible if the acetoxymethyl moiety that bears the H-23' diaterotopic pair is placed in the β -side that results on the R configuration at C-23. Similar conclusions can be drawn from the MS and NMR spectra of compound 23S-6; the observed NOE correlation between the H-24' methyl group in the α -side of F-ring and the H-23' diasterotopic pair, indicates the α -orientation of the exocyclic acetoxymethyl moiety and consequently the S-configuration at C-23 (see Fig. 3).

X-ray studies on monocrystals of both diasteromers **23R-6** and **23S-6** corroborated the proposed structures and configuration at C-23 (see Fig. 4).

The occurrence of the diastereomeric compounds **23R-6** and **23S-6** can be justified by the acid catalyzed F-ring opening in compound **23S-5** that leads to the intermediate **I**. Bromination at C-23 leads to the mixture of the diastereomeric intermediates **23R-III** and **23S-III** that on F ring closure produce the observed mixture of **23R-6** and **23S-6** (see Scheme 3).

As described for **23R-6** and **23S-6**, in the case of the dibrominated compounds **23R-7** and **23S-7**, the absence of the signal corresponding to H-23 and the doublet multiplicity of the diasterotopic protons at C-23′ suggest the presence of the bromine

atom at C-23. The occurrence of monobrominated fragments in *m*/*z* 535 and 537 may be interpreted as the product of the loss of Br from the molecular ions (M⁺-Br or more likely MH⁺-HBr) of the dibrominated compounds. Additionally the change on the ¹H and ¹³C signals associated to the exocyclic methylene C-23′ (*compared to those of the starting material* **23R-5** *and the monobrominated compounds* **23R-6** *and* **23S-6**) and well as the disappearance of the ¹H and ¹³C signals associated to the acetoxy moiety formerly attached to C-23, evidence the presence of a different atom at C-23 that, according to the reaction conditions, must be no other than a bromine.

NOESY and COSY correlations observed in compounds **23R-7** and **23S-7** corroborated the integrity of the side chain as well as the configuration at C-23. In particular, for compound **23R-7** the observed between H-23' \leftrightarrow H-24' NOESY correlation indicates the α -orientation of the C-23' exocyclic methylene and, as a consequence, the *R*-configuration at C-23. On the other hand, the observed H-23' \leftrightarrow H-20 and H-23' \leftrightarrow H-24 NOESY correlations indicate the β -orientation of the C-23' exocyclic methylene in compound **23S-7** and consequently its *S*-configuration at C-23 (see Fig. 5).

The unexpected occurrence of the dibrominated compounds **23S-7** and **23R-7** can be rationalized as follows. Elimination of the allylic acetate in the intermediate **II** may lead to the intermediate **IV** that undergoes F ring closure to produce the alkene **9** that on bromination leads to the observed dibrominated compounds. In addition, attack of the bromide anion to an allylic carbocation at C-23 may lead to the intermediate **V** that undergoes bromination at C-23 and F-ring closure as described above (see Scheme 4).

To gain a more detailed view on the proposed mechanisms, the possibility of the acid substitution of the acetoxy moiety at

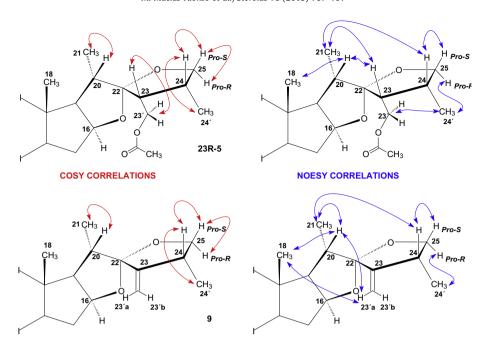


Fig. 7. Selected COSY and NOESY correlations in compounds 23S-6 and 9.

Scheme 8. Possible mechanism for C-23 epimerization and elimination reactions.

C-23'was explored. Treatment of **23S-5** with HBr in acetic acid produced complex mixture from which the C-23' brominated compounds **23S-8** and **23R-8** were isolated in low yields (see Scheme 5).

The Mass spectra of both 23'-brominated compounds are characterized by the presence of molecular ions (MH $^+$, m/z 539 and 537) that indicate the incorporation of a bromine atom. Disappear-

ance of the ¹H and ¹³C signals corresponding to the acetoxy moiety formerly attached to C-23′ indicates the presence of a bromine atom at this position. As in the preceding compounds, the integrity of the spiroketal moiety and the configuration at C-23 in compounds **23R-8** and **23S-8** can be verified by the presence of the ¹H and ¹³C signals as well as the observation of the COSY and NOESY correlations (see Fig. 6).

The occurrence of the C-23′ brominated compounds **23R-8** and **23S-8** evidences the substitution of the acetoxy group attached to C-23′. Acid catalyzed dissociation of **II** leads to the allylic cation **IV** that on reaction with the bromine anion produces **V**. F-ring closure in **V** leads to the observed substitution products **23R-8** and **23S-8**. The observed substitution process strongly suggests the existence of the intermediates **II**, **IV** and **V** in this and the above described reactions (see Scheme 6).

Finally, when **23S-5** was treated with $BF_3 \cdot Et_2O$ in acetic acid to explore the acid-catalyzed inversion at C-23, a mixture of the starting material **23S-5**, its C-23 epimer **23R-5** and the alkene **9** was obtained (see Scheme 7).

Again the integrity of the side chains of compounds **23R-5** and **9**, can be verified by observation of the ^1H and ^{13}C signals corresponding to this fragment as well as the present COSY and NOESY correlations. The configuration at C-23 in compound **23R-5** can be verified by observation of the H-23' \leftrightarrow H-24' both in the α -side of the F-ring (see Fig. 7). In particular the alkene **9** can be recognized by the absence of the signals corresponding to the acetoxy moiety formerly present at C-23' as well as the new ^1H and ^{13}C signals corresponding to the new exocyclic double bond.

Protonation of the intermediate **II** from the *Re* or *Si* sides of C-23, followed by F-ring closure explain the production of the **23R-5/23S-5** mixture. On the other hand, $BF_3 \cdot Et_2O$ -catalyzed concerted elimination/F ring closure may justify the occurrence of the alkene **9**. Alternatively the olefin **9** can be produced through F ring closure in the allylic carbocation **IV** (see Scheme 8).

7. Conclusions

We have found that the presence of the C-23′ acetoxy moiety confers a special reactivity to the furospirostane side chain. The F-ring opening produces a $\Delta^{22}\text{-intermediate}$ in which the C-23′-acetoxy is placed in position allowing its substitution or elimination. This reactivity feature opens new and useful possibilities for the transformation of the 16 β ,22:22,25-diepoxy-23-acetoxy-methyl-24-methyl side chain.

Acknowledgments

The authors thank to the *Dirección General de Asuntos del Personal Académico* (DGAPA-UNAM) for financial support via project IN221911 and CONACyT for scholarship granted to MM-A. Thanks are due to Rosa I. del Villar Morales, Georgina Duarte Lisci (USAI-UNAM) for recording NMR and MS spectra. We want to express our gratitude to Dr. Carlos Cobas from Mestrelab® for assistance with the MestreNova NMR processing program and to Dr. John Boulton for correcting the manuscript.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.steroids.2013. 05.014.

References

- Jacobs MF, Kitching W. Spiroacetals of marine origin. Curr Org Chem 1998;2:395–436.
- [2] Mori K. Synthesis of insect pheromones. In: ApSimon J, editor. The total synthesis of natural products. New York: John Wiley & Sons Inc; 1981.
- [3] Raju R, Saikia AK. Asymmetric synthesis of naturally occuring spiroketals. Molecules 2008;13:1942–2038.
- [4] Mead KT, Brewer BN. Strategies in spiroketal synthesis revisited: recent applications and advances. Curr Org Chem 2003;7:227–56.
- [5] Perron F, Albizati KF. Chemistry of spiroketals. Chem Rev 1989;89:1617-61.

- [6] Moser BR. Review of cytotoxic cephalostatins and ritterazines: isolation and synthesis. J Nat Prod 2008;71:487–91.
- [7] Iglesias MA, Morzycki JW. Cephalostatins and ritterazines. In: Knölker HJ, editor. The alkaloids: chemistry and biology, vol. 72. Amsterdan: Elsevier; 2013. p. 153–279.
- [8] Betancor C, Freire R, Pérez-Martin I, Prangé T, Suárez E. A convenient synthesis of C-22 and C-25 stereoisomers of cephalostatin north 1 side chain from spirostan sapogenins. Org Lett 2002;4:1295-7.
- [9] Lee JS, Fuchs PL. New oxidative tools for the functionalization of the cephalostatin north 1 hemisphere. Org Lett 2003;5:2247–50.
- [10] LaCour TG, Guo C, Bhandaru S, Boyd MR, Fuchs PL. Interphylal product splicing: the first total syntheses of cephalostatin 1, the north hemisphere of ritterazine G, and the highly active hybrid analogue, ritterostatin G_N1_N¹. J Am Chem Soc 1998;120:692–707.
- [11] Rao ChB, Ramana KV, Rao DV, Fahy E, Faulkner DJ. Metabolites of the gorgonian Isis hippuris from India. J Nat Prod 1988;51:954–8.
- [12] González N, Barral MA, Rodriguéz J, Jiménez C. New cytotoxic steroids from the gorgonian Isis hippuris Structure-activity studies. Tetrahedron 2001;57:3487–97.
- [13] Sheu J-H, Chao C-H, Wang G-H, Hung K-C, Duh C-Y, Chiang MY, Wu Y-C, Wud C-C. The first A-nor-hippuristanol and two novel 4,5-secosuberosanoids from the gorgonian *Isis hippuris*. Tetrahedron Lett 2004;45:6413–6.
- [14] Chao C-H, Huang L-F, Yang Y-L, Su J-H, Wang G-H, Chiang MY, Wu Y-C, Dai C-F, Sheu J-H. Polyoxygenated Steroids from the gorgonian *Isis hippuris*. J Nat Prod 2005;68:880–5.
- [15] Ravindar K, Reddy MS, Lindqvist L, Pelletier J, Deslongchamps P. Synthesis of the antiproliferative agent hippuristanol and its analogues via suarez cyclizations and Hg(II)-catalyzed spiroketalizations. J Org Chem 2011;76:1269–84.
- [16] Ibarrola DA, Hellión-Ibarrola MC, Montalbetti Y, Heinichen O, Campuzano MA, Kennedy ML, Alvarenga N, Ferro EA, Dölz-Vargas JH, Momose Y. Antihypertensive effect of nuatigenin-3-O-β-chacotriose from Solanum sisymbriifolium Lam. (Solanaceae) (ñuatî pytâ) in experimentally hypertensive (ARH+DOCA) rats under chronic administration. Phytomedicine 2011;18:634–40.
- [17] Fieser L, Fieser M. Steroids. New York: Reinhold Publishing Corporation; 1959 [and references there in].
- [18] Callow RK, James VHT, Kennard O, Page JE, Paton PN, Riva di Sanseverino L. The structure of the steroidal bromosapogenins. J Chem Soc C 1966:288–97.
- [19] Iglesias-Arteaga MA, Sandoval-Ramírez J, Mata-Esma MY, Viñas-Bravo O, Bernes S. Abnormal Beckmann rearrangement in 23-hydroxyiminodiosgenin acetate. Tetrahedron Lett 2004;45:4921–6.
- [20] Iglesias-Arteaga MA, Velázquez-Huerta GA, Méndez-Stivalet JM, Galano A, Álvarez-Idaboy JR. The Baeyer-Villiger reaction of 23-oxosapogenins. Arkivoc 2005;VI:109-26.
- [21] Iglesias-Arteaga MA, Alvarado-Nuño AA. $BF_3 \cdot Et_2O$ induced Beckmann rearrangement of 23 hydroxyiminosapogenins. A shortcut to bisnorcholaniclactones. Tetrahedron Lett 2006;47:5351-3.
- [22] Iglesias-Arteaga MA, Jastrzębska I, Morzycki JW. Reactions of sapogenins with m-chloroperoxybenzoic acid catalyzed by Lewis acids. Polish J Chem 2006;80:667–71.
- [23] Iglesias-Arteaga MA, Velázquez-Huerta GA. Favorskii rearrangement of 23oxo-3-epi-smilagenin acetate induced by iodosobenzene. Tetrahedron Lett 2005:46:6897-9.
- [24] Iglesias-Arteaga MA, Arcos-Ramos RO. One-step axial acetoxylation at C-23. A new method for the functionalization of the side chain of steroid sapogenins. Tetrahedron Lett 2006;47:8029–31.
- [25] Iglesias-Arteaga MA, Arcos-Ramos RO, Méndez-Stivalet JM. The unexpected course of the reaction of steroid sapogenins with diacetoxyiodobenzene and BF₃:Et₂O in formic acid. Tetrahedron Lett 2007;48:7485–8.
- [26] Hernández R, Marrero-Tellado JJ, Prout K, Suárez E. Lewis acid-mediated isomerization of (25R)-3α-acetoxy-5α-spirostan-23-one, a C-22 spiroacetal: an approach to the synthesis of C-23 spiroacetal steroidal sapogenins. J Chem Soc Chem Commun 1992:275–7.
- [27] Betancor C, Dorta RL, Freire R, Martín A, Prangé T, Suárez E. Stereospecific synthesis of 1,6-dioxadecalins and 2,2 linked ditetrahydrofurans by rearrangement of steroidal spiroacetals. J Org Chem 1998;63:6355-62.
- [28] LaCour TG, Tong Z, Fuchs PL. Consequences of acid catalysis in concurrent ring opening and halogenation of spiroketals. Org Lett 1999;1:1815–8.
- [29] Sandoval-Ramírez J, Meza-Reyes S, del Río RE, Hernández-Linares G, Suárez-Rojas A, Rincón S, et al. Regioselective cleavage of rings E and F in sarsasapogenin. Steroids 2003;68:199–204.
- [30] Cyrański MK, Frelek J, Jastrzębska I, Morzycki JW. Rearrangement of 23oxospirostanes to the 22-oxo-23-spiroketal isomers promoted by Lewis acids. X-ray crystal structure of (23R,25S)-3-acetoxy-16,23:23,26-diepoxy-5cholestan-22-one. Steroids 2004;69:395–400.
- [31] Morzycki JW, Jastrzębska I. Novel transformation of 23-bromosapogenins. Synthesis of (22S, 23R)-22-hydroxy-23,26-epoxyfurostanes. Tetrahedron Lett 2001;42:5989–91.
- [32] Anulewicz-Ostrowska R, Jastrzębska I, Morzycki JW, Wojcik J. An assisted solvolysis of 23 spirostanyl bromides and tosylates. A new rearrangement of spirostanes to the bisfuran systems. J Org Chem 2002;67:6916–24.
- [33] Jastrzębska I, Morzycki JW, Trochimowicz U. Lead tetraacetate-iodine oxidation of 23-spirostanols. Tetrahedron Lett 2004;45:1929–32.
- [34] Jastrzębska I, Morzycki JW. Unusual Baeyer-Villiger oxidation of 23oxosarsasapogenin acetate. Polish J Chem 2005;79:1245–8.

- [35] López Y, Ruíz-Pérez KM, Yépez R, Santillán R, Flores-Álamo M, Iglesias-Arteaga MA. Mechanistic insights and new products of the reaction of steroid sapogenins with NaNO₂ and BF₃·Et₂O in acetic acid. Steroids 2008;73:657–68.
- [36] Ruíz-Pérez KM, Romero-Ávila M, Flores-Pérez B, Flores-Álamo M, Moreno-Esparza R, Iglesias-Arteaga MA. Revisiting 23-iodospirostanes. New facts and full characterization. Steroids 2009;74:996–1002.
- [37] Macías-Alonso M, Morzycki JW, Iglesias-Arteaga MA. Studies on the BF₃-Et₂O catalyzed Baeyer-Villiger reaction of spiroketalic steroidal sapogenins. Steroids 2011;76:317–23.
- [38] Lee JS, Cao H, Fuchs PL. Ruthenium-catalyzed mild C-H oxyfunctionalization of cyclic steroidal ethers1. J Org Chem 2007;72:5820-3.
- [39] Phillips ST, Shair MD. Syntheses of the eastern halves of ritterazines B, F, G, and H, leading to reassignment of the 5,5-spiroketal stereochemistry of ritterazines B and F. J Am Chem Soc 2007;129:6589–98.
- [40] Lee S, Fuchs PL. An efficient C–H oxidation protocol for α -hydroxylation of cyclic steroidal ethers. Org Lett 2004;6:1437–40.
- [41] Betancor C, Freire R, Pérez-Martín I, Prangé T, Suárez E. A convenient synthesis of C-22 and C-25 stereoisomers of cephalostatin north 1 side chain from spirostan sapogenins. Org Lett 2002;4:1295-7.
- [42] Macias-Alonso M, Flores Alamo M, Iglesias Arteaga MA. On the reactivity of 23-methoxycarbonyl furospirostanes. Steroids 2011;76:1021–31.

- [43] Guo C, LaCour TG, Fuchs PL. On the relationship of OSW-1 to the cephalostatins. Bioorg Med Chem Lett 1999;9:419–24.
- [44] LaCour TG, Guo C, Ma S, Jeong JU, Boyd MR, Matsunaga S, Fusetani N, Fuchs PL. On topography and functionality in the B-D rings of cephalostatin cytotoxins. Bioorg Med Chem Lett 1999;9:2587–92.
- [45] Macías-Alonso M, Esturau-Escofet M, Flores-Álamo, Moreno-Esparza R, Iglesias-Arteaga MA. NMR and X-ray characterization of steroids with furospirostane side chains. Arkivoc 2011;xi:165–82.
- [46] For NMR spectra see supplementary information file.
- [47] Oxford diffraction CrysAlis CCD and CrysAlis RED. Abingdon, England: Oxford Diffraction Ltd; 2009.
- [48] Sheldrick GM. A program for crystal structure solution. Acta Crystallogr A 2008;A64:112–22.
- [49] Farrugia LJ. WinGX and ORTEP for windows: an update. J Appl Crystallogr 2012;45:849–54.
- [50] Crystallographic data have been deposited at the Cambridge Crystallographic Data Center as supplementary material numbers CCDC 928324 for 23S-6 and CCDC 928325 for 23R-6. Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK. E-mail: <deposit@ccdc.cam.ac.uk>.