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Conformational study of cationic, zwitterionic, anionic species of aspartic acid, water-added forms and their protonation. A DFT method

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Abstract

The potential energy surfaces of various species of aspartic acid have been computed at the DFT/B3LYP/6-31G(d) level of theory. Conformations of 34, 11, 9 and 3 were found from 324, 162, 81 and 81 possible conformers for species H_3 asp $^+$, H_2 asp, H_3 asp $^-$ and asp $^{2-}$, respectively. The most stable conformation for species H_3 asp $^+$, H_2 asp, H_3 asp $^-$ and asp $^{2-}$ are γ_D , α_L , ϵ_L and β_L conformers, respectively. Three protonation steps due to the tetrahydrated forms of species asp $^{2-}$, H_3 asp $^-$ and H_2 asp of aspartic acid computed at the H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp of aspartic acid computed at the H_3 asp $^+$ and H_3 asp $^+$ are γ_D aspartic acid computed at the H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ are γ_D aspartic acid computed at the H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ are γ_D aspartic acid computed at the H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ are γ_D aspartic acid computed at the H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ are γ_D aspartic acid computed at the H_3 asp $^+$ and H_3 asp $^+$ and H_3 asp $^+$ are γ_D and

Keywords: Aspartic acid; Cationic; Zwitterionic; Anionic species; Conformation; DFT; Protonation

1. Introduction

The conformational equilibria of cationic, zwitterionic and anionic species of aspartic acid were determined within the wide pH range of acidic to basic aqueous solutions [1,2]. Dianionic species of aspartic acid (asp²) found in the aqueous solution at high pH range was introduced and its reaction rates were reported [2,3]. At the wide pH range of aqueous aspartic acid solution, the structures [H₃⁺N-CH(CH-COOH)(COOH)]⁺ for H₃asp⁺, [H₃⁺N-CH(CH $-COOH)(COO^{-})$ for H₂asp, [H₃⁺N-CH(CH-COO⁻)(COO⁻)] for Hasp and [H₂N-CH(CH-COO⁻)(COO⁻)]²⁻ for asp²⁻ have been found and their corresponding acidity constants were reported [1-5]. The zwitterionic form of zero net-charge species of aspartic acid was found that its local energy minima on the potential energy surface (PES) [6] of bare structure has not been similar to its hydrated structure. Many studies have been performed on aspartic acid and aspartate [7], N-acetyl-Laspartic acid-N'-methylamide [8–11] and N-formyl-L-aspartic acid amide and N-formyl-L-aspartatamide [12] using ab initio MO and density functional methods. Thermodynamic quantities of conformational equilibria in the side chains of aspartic

acid in acidic, neutral and basic aqueous solutions were investigated using proton NMR method [13].

In solid state and aqueous solution, the neutral aspartic acid (H_2 asp) is actually present as a zwitterionic α -aspartic acid as [H_3^+N -CH(CH-COOH)(COO $^-$)] [3,6,14]. Nevertheless, the neutral aspartic acid as [H_3^+N -CH(CH-COO $^-$)(COOH)] was also found as a β -carboxylate zwitterion. The stepwise deprotonation process of fully protonated species of aspartic acid (H_3 asp $^+$) can be presented as a pH dependence diagram as shown in Fig. 1.

Definition of atomic numbering for H_3 asp $^+$ as representation of aspartic acid and definition of dihedral angles ω (H1–O1–C1–C2), ψ (O1–C1–C2–N1), φ (H4–N1–C2–C1), χ_1 (N1–C2–C3–C4), χ_2 (C2–C3–C4–O3) and χ_3 (C3–C4–O3–H2) are shown in Fig. 2. The dihedral angles χ_3 and ω were obtained from full optimizations and χ_3 is used to indicate the *endo* and *exo* conformational types of carboxylic group; the *endo* and *exo* forms are defined as χ_3 =0 and 180°, respectively. E and E isomerism of the dihedral angle E was applied for conformational nomenclature. Due to the *endolexo* and E/Z definitions, all aspartic acid structures of various forms can be categorized into species (a) H_3 asp $^+$ -*endo-E*, (b) H_3 asp $^+$ -*endo-Z*, (c) H_3 asp $^+$ -*exo-E*, (d) H_3 asp $^+$ -*exo-Z*, (e) H_2 asp-*endo*, (f) H_2 asp-*exo*, (g) Hasp $^-$ and (h) asp 2 as shown in Fig. 3.

Due to the conformational convention for amino acid residues, [15,16] the number of possible conformations for each aspartic acid structures is 81 which is a combination

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Fig. 1. pH dependence of four typical species of aspartic acid in aqueous solution and protonation reversible mechanism.

number of nine $(\gamma_D,\,\delta_D,\,\alpha_L,\,\epsilon_D,\,\beta_L,\,\epsilon_L,\,\alpha_D,\,\delta_L,\,\gamma_L)$ back-bone (BB) and nine $(g^+g^-,\,a\,g^-,g^-g^-,g^+a,\,a\,a,\,g^-a,\,g^+g^+,\,a\,g^+,\,g^-g^+)$ side-chain (SC) conformations. The nine BB conformations and nine SC conformations are located on Ramachandran map [17,18,19,20] and IUPAC [21] convention as shown in Fig. 4. To employ the Ramachandran and IUPAC conventions for the BB and SC conformations, respectively, the rotational conformation of aspartic acid notified as 'BB[SC]' has been therefore used to call the conformation of aspartic acid.

In present work, the conformations of species $H_3 asp^+$, $H_2 asp$, $Hasp^-$ and asp^{2-} of aspartic acid in gas phase have been located using the potential energy surface (PES) method. The water-added forms of the most stable conformation of each species of aspartic acid have been also studied. Protonation of tetrahydrated forms of the most stable conformation of species $H_3 asp^+$, $H_2 asp$, $Hasp^-$ and asp^{2-} of aspartic acid has been modeled and its reaction energies have been obtained.

2. Computational details

The PESs of all species (H_3 asp⁺-endo-E, H_3 asp⁺-endo-Z, H_3 asp⁺-exo-E, H_3 asp⁺-exo-Z, H_2 asp-endo, H_2 asp-exo, Hasp and asp²) of aspartic acid were computed with 30° increments along four dihedral angles (ψ , ϕ , χ_1 and χ_2) using the hybrid density functional theory (DFT) at B3LYP/6-31G(d) [22,23,24] level of theory.

As the stabilities of bare structure of all species were examined, the species H_2 asp and $Hasp^-$ are not stable without including three water molecules because their amino proton can transfer to their carboxylate oxygen. Therefore, their PESs were computed as trihydrated species by adding three water molecules to stabilize their molecular structures. The B3LYP/6-31G(d)-optimized structures of trihydrated and tetrahydrated forms of the most stable conformation of each species were fully reoptimized at the B3LYP/6-31+G(d, p) level of theory with the zero-point vibrational energy corrections. The stabilization energies of protonation of tetrahydrated form of aspartic acid were derived from their total energies computed at B3LYP/6-31+G(d, p) level of

theory. Therefore, the water-added structures are categorized into three models namely (1) bare, except for the species H₂asp and Hasp⁻, (2) trihydrated and (3) tetrahydrated forms of various species of aspartic acid. All computations were performed using the GAUSSIAN 03 program package [25]. The MOLDEN 3.7 program [26] was utilized to observe molecular energies and geometries convergence via the Gaussian output files. The molecular graphics of all species were generated with the MOLEKEL program [27].

All geometric minima for each species of aspartic acid on the conformational PES are represented by Ramachandran and IUPAC maps as shown in Fig. 4. To combine the conformational convention and each species of aspartic acid, its full name has been therefore specified as 'species type-endolexo-Z/E' of 'BB [SC]' such as conformer of H_3 asp⁺-endo-Z of γ_D [g⁺g⁻]. Energies for each structural conformers of H_3 asp⁺ and H_2 asp species are therefore the functions of E (Ψ , φ , χ_1 , χ_2 , ω) and E (ψ , φ , χ_1 , χ_2 , χ_3), respectively. For the anionic species of Hasp⁻ and asp²⁻, their energies are a function of E (ψ , φ , χ_1 , χ_2).

3. Results and discussion

As the stabilities of bare structure of all species were examined, the proper zwitterionic structures of the species H_2 asp and H_3 p are not stable because their amino proton can

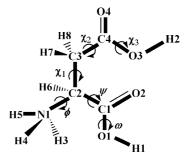


Fig. 2. Definition of atomic numbering for H_3 asp $^+$ as representative of aspartic acid and definition of dihedral angles ω (H1–O1–C1–C2), ψ (O1–C1–C2–N1), ψ (H4–N1–C2–C1), χ_1 (N1–C2–C3–C4), χ_2 (C2–C3–C4–O3) and χ_3 (C3–C4–O3–H2).

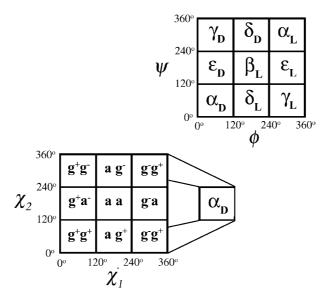


Fig. 3. Aspartic acid species (a) H₃asp⁺-endo-E, (b) H₃asp⁺-endo-Z, (c) H₃asp⁺-exo-E, (d) H₃asp⁺-exo-Z, (e) H₂asp-endo, (f) H₂asp-exo, (g) Hasp⁻ and (h) asp²⁻ species.

easily transfer to their carboxylate oxygen. Due to the stability test of hydrated structures of the species H₂asp and Hasp⁻, it was found that these two species are stabilized by at least three water molecules. The PESs for the species H₂asp and Hasp⁻ were therefore computed as their trihydrated forms but for the species H₃asp⁺ and asp²⁻, their bare structures were employed.

3.1. PESs of various forms of aspartic acid and their geometrical conformations

Conformational structures of H₃asp⁺-endo-E, H₃asp⁺endo-Z, H₃asp⁺-exo-E, H₃asp⁺-exo-Z, H₂asp-endo, H₂aspexo, Hasp and asp are species optimized at B3LYP/6-31G(d) level of theory are shown in Tables S1, S2, S3, S4, S5, S6, S7 and S8 as contained in the Supplementary material. Classification of the types of internal hydrogen bonding for aspartic acid are shown in Fig. 5. Two types of backbone/backbone (BB/BB) and five types of side-chain/backbone (SC/BB) hydrogen bonding were defined. The types and distances of existing hydrogen bonding for various H₃asp⁺ forms of aspartic acid is shown in Table 1 and for various H₂asp, Hasp⁻ and asp²⁻ forms are shown in Table 2. Bond distances of the internal hydrogen bonding of types 1a, 1b, 2a, 2b, 2c, 2d and 2e found in all conformations are within 1.768-1.949, 1.724–2.158, 1.875–1.910, 1.690–1.916 Å, at 2.010 Å and 1.706-1.760 Å, respectively. The numbers of stable conformers for H₃asp⁺-endo-E, H₃asp⁺-endo-Z, H₃asp⁺-exo-E, H₃asp⁺-exo-Z, H₂asp-endo, H₂asp-exo, Hasp⁻ and asp²⁻ are 8, 13, 5, 8, 4, 7, 9 and 3 conformers and their conformational structures are shown in Figs. S1, S2, S3, S4, S5, S6, S7 and S8 as contained in Supplementary material, respectively. The landscape representations of the side-chain conformational PESs, $E=E(\chi_1, \chi_2)$ associated with the backbone conformations of H₃asp⁺-endo-E, H₃asp⁺-endo-Z, H₃asp⁺-exo-E, H₃asp⁺-exo-Z, H₂asp-endo, H₂asp-exo, Hasp⁻ and asp²⁻ are shown in Figs. S9, S10, S11, S12, S13, S14, S15 and S16

(Supplementary material), respectively. The bottoms of the Figs. S9, S10, S11, S12, S13, S14, S15 and S16 show integrated scatter-plot diagrams of the existing conformations which correspond to the PESs located at the top of the figures.

Based on the geometrical parameters of the conformations of species $H_3 asp^+$, $H_2 asp$ or $Hasp^-$, their conformations are identical when their dihedral angles χ_1 and χ_2 are the same value and if their dihedral angles φ differ by 120° or ψ differ by 180°. In case of the species asp^{2-} , their conformations are identical when their dihedral angles χ_2 or ψ differ by 180°.

As the bare structures of the species H_3 asp⁺ and asp²⁻ and the trihydrated forms of the species H_2 asp and Hasp⁻ were employed in the PESs computations using the B3LYP/6-31G(d)

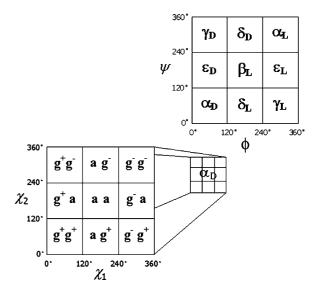


Fig. 4. A schematic representation of relation between a backbone (BB) conformation Ramachandran map (top) and a side-chain (SC) conformation map designated by IUPAC convention (bottom). The Ramachandran and IUPAC-SC conformation maps, defined as 'BB[SC]' conformational notation of aspartic acid.

Fig. 5. Classification of the types of internal hydrogen bonding for various species of aspartic acid.

theoretical level. The most stable conformers of $H_3 asp^+, H_2 asp(3H_2O), Hasp^-(3H_2O)$ and asp^{2-} of aspartic acid in gas phase are γ_D [g $^-$ g $^+$] which is identical to δ_D [g $^-$ g $^+$] (see Table S2 and Fig. S2), α_L [a g $^-$] (see Table S6 and Fig. S6), ϵ_L [g $^+$ a] which is identical to β_L [g $^+$ a] (see Table S7 and Fig. S7) and β_L [g $^+$ a] which is identical to β_L [g $^+$ g $^-$],? δ_L [g $^+$ a] and δ_L [g $^+$ g $^-$] (see Table S8 and Fig. S8), respectively.

3.2. Water-added structures and their protonation

The B3LYP/6-31+G(d, p)-optimized structures of various trihydrated and tetrahydrated species of aspartic acid and their selected geometrical data are tabulated in Table 3. The B3LYP/6-31G(d)-optimized structures of the bare molecule of species $\rm H_3 asp^+$ and $\rm asp^{2-}$, trihydrated ($\rm H_3 asp^+(3H_2O)$, $\rm H_2 asp(3H_2O)$, $\rm Hasp^-(3H_2O)$ and $\rm asp^{2-}(3H_2O)$) and tetrahydrated ($\rm H_3$. $\rm asp^+(4H_2O)$, $\rm H_2 asp(4H_2O)$, $\rm Hasp^-(4H_2O)$ and $\rm asp^{2-}(4H_2O)$) species were fully reoptimized at B3LYP/6-31+G(d, p) level. Conformations of the B3LYP/6-31G(d)-optimized geometries of all aspartic acid species are shown Table 4. The most stable conformation for each aspartic acid species existing as

trihydrated and tetrahydrated forms and their protonation and deprotonation schemes are shown in Fig. 6. The stabilization energies of protonation of tetrahydrated form of aspartic acid derived from their total energies computed at B3LYP/6-31+ G(d, p) level of theory with zero-point vibrational energy (ZPVE)are listed in Table 5. Three protonation steps are exothermic reactions and their stabilization energies of the stepwise protonation are -244.68, -308.57 and -379.97 kcal mol $^{-1}$, respectively.

Based on the B3LYP/6-31+G(d, p) computations of different structural models (the bare, trihydrated and tetrahydrated structures), the conformation for the species $H_3 asp^+ \gamma_D [g-g+]$), $H_2 asp \alpha_L [a g-]$), $Hasp^- (\epsilon_L [g+a])$ and $asp^{2-} (\beta_L [g+a])$ are not changed as shown in Table 4. As compared to other species, the geometrical change of the species $Hasp^-$ has obviously occurred as shown in Fig. 7. Due to the change of trihydrated to tetrahydrated system, Fig. 7(c) illustrates that the side-chain carboxylic group of the species $Hasp^-$ is the main cause of its geometrical change; the change of its dihedral angles χ_1 and χ_2 are ~ 4 and 9° , respectively.

Table 1 The present hydrogen bonding, its type and distance for conformations of the aspartic acid H_3 asp⁺ species

Conformer	Hydrogen bond				
	BB/BB	BB/BB	SC/BB	SC/BB	
	types	distances (Å)	types	distances (Å)	
H ₃ asp ⁺ -endo-E	form				
$\gamma_{\rm D} [a \ a]^{a1}$	1a	1.768	2d	1.780	
$\gamma_D [g^- a]^{b1}$	1a	1.620	2b	1.910	
$\gamma_{\rm D} [g^- g^-]$	1a	1.901	2c	1.820	
$\gamma_{\rm D} [g^- g^+]^{\rm c1}$	1a	1.949	2a	1.750	
α_D [a g ⁺]	1a	1.734	2c	1.694	
$\alpha_{\rm D} \left[g^{+} a \right]^{\rm d1}$	_	_	2b	1.879	
$\alpha_D [g^+g^-]^{e1}$	_	_	2a	1.750	
$\beta_L [g^- g^+]$	_	_	2a	1.759	
H ₃ asp ⁺ -endo-Z	form				
$\gamma_D [a g^-]^{a2}$	_	_	_	_	
$\gamma_{\rm D} [{\rm g}^- {\rm a}]^{\rm b2}$	_	_	2b	1.875	
$\gamma_{\rm D} [{\rm g}^- {\rm g}^+]^{\rm c2}$	_	_	2a	1.662	
$\delta_D [a \ a]^{uz}$	_	_	_	_	
$\varepsilon_{\rm D} [g^{\rm T}a]^{\rm e2}$	_	_	2b	1.888	
$\varepsilon_{\rm D} [g^+g^-]^{\rm f2}$	_	_	2a	1.760	
$\varepsilon_{\rm D} [{\rm a g}^{-}]^{\rm g2}$	1b	2.153	_	_	
$\varepsilon_{\mathrm{D}} [\mathrm{g}^- \mathrm{g}^+]^{\mathrm{h}2}$	_	_	2a	1.750	
$\varepsilon_{\rm D} [{\rm g}^- {\rm a}]$	1b	1.882	2b	1.882	
ε_{L} [a a]	1b	2.158	_	_	
$\alpha_D [a g^+]^{i2}$	_	_	_	_	
$\alpha_{\rm D} [g^+a]^{j2}$	1b	1.883	2a	1.868	
$\alpha_D [g^+g^-]^{k2}$	_	_	2a	1.750	
$H_3 asp^+$ -exo-E f	orm				
$\gamma_{\rm D} [{\rm g}^- {\rm g}^+]^{{\rm a}3}$	_	_	2a	1.688	
$\varepsilon_{\mathrm{D}} \left[\mathrm{g}^{-} \; \mathrm{g}^{+} \right]$	_	_	2a, 2c	1.864, 1.867	
$\alpha_D [g^- a]^{b3}$	_	_	2d	2.010	
$\alpha_{\rm D} [{\rm g}^{+}{\rm g}^{-}]^{{\rm c}3}$	_	_	2a	2.133	
$\delta_{\rm L} [a g^+]^{\rm d3}$	1a	1.724	2c	1.690	
H ₃ asp ⁺ -exo-Z f	orm				
$\gamma_D [a g^-]^{a4}$	_	_	_	_	
$\gamma_{\rm D} [g^- g^+]^{b4}$	_	_	2a	2.236	
$\delta_D [g^+g^-]$	_	_	2a	1.722	
$\varepsilon_{\rm D} [{\rm a g}^{-}]^{\rm c4}$	1b	1.724	_	_	
$\varepsilon_{\rm D} \left[{\rm g}^- {\rm g} + \right]^{\rm d4}$	1b	2.156	2a	2.156	
$\beta_{L} [g^{+}g^{-}]^{e4}$	_	_	2a	1.722	
$\alpha_D [g^- a]^{f4}$	_	_	2c	1.916	
$\alpha_{\rm D} [g^+g^-]^{g4}$	_	_	2a	1.712	

 $^{^{}a1}$ Identical to δ_D [a a] and α_L [a a]. b1 Identical to δ_D [g $^-$ a] and α_L [g $^-$ a]. c1 Identical to δ_D [g $^-$ g $^+$] and α_L [g $^-$ g $^+$]. d1 Identical to δ_L [g $^+$ a] and γ_L [g $^+$ a]. e1 Identical to γ_L [g $^+$ g $^-$]. a2 Identical to δ_D [a g $^-$] and α_L [a g $^-$]. b2 Identical to δ_D [g $^-$ g $^+$]. d2 Identical to α_L [a a]. e2 Identical to δ_D [g $^-$ g $^+$]. d2 Identical to α_L [a a]. e2 Identical to δ_L [g $^+$ g $^-$] and ϵ_L [g $^+$ g $^-$]. d2 Identical to ϵ_L [a g $^-$]. b2 Identical to δ_L [g $^-$ g $^+$] and ϵ_L [g $^+$ g $^-$]. d2 Identical to δ_L [a g $^+$]. b2 Identical to δ_L [g $^+$ g $^-$] and γ_L [g $^+$ g $^-$]. b3 Identical to δ_L [g $^+$ g $^-$] and γ_L [g $^+$ g $^-$]. b3 Identical to δ_L [g $^-$ g $^+$] and γ_L [g $^-$ g $^-$]. b3 Identical to δ_L [g $^-$ g a] and γ_L [g $^-$ g $^-$]. b4 Identical to δ_D [g $^-$ g $^+$] and α_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and α_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_L [g $^-$ g $^+$]. b4 Identical to δ_D [g $^-$ g $^+$] and ϵ_D [g $^-$ g $^+$].

4. Conclusions

The potential energy surfaces for the bare structures of species $H_3 asp^{-+}$ and asp^{2-} and for the trihydrated

Table 2
The present hydrogen bonding, its type and distance for conformations of species H₂asp, Hasp⁻ and asp²⁻ of aspartic acid

Conformer	nformer Hydrogen bond				
	BB/BB	BB/BB	SC/BB	SC/BB	
	types	distances	types	distances	
		(Å)		(Å)	
H ₂ asp-endo fori	m				
$\alpha_{\rm D} [g^+g^-]$	1a	1.570	_	-	
$\varepsilon_{\mathrm{D}} \left[\mathrm{g}^{+} \mathrm{g}^{-} \right]$	1a	1.839		_	
$\delta_{\mathrm{D}} \left[\mathrm{g}^{+} \mathrm{g}^{-} \right]$	1a	1.635	_	-	
$\varepsilon_{\rm L} [g^- g^+]$	_	_	_	-	
H ₂ asp-exo form					
$\varepsilon_{\rm D} [g^+ a]$	_	_	_	_	
$\varepsilon_{\rm D} [{\rm g}^+ {\rm g}^-]$	1a	1.855	_	-	
$\varepsilon_{\rm D} [{\rm a~g}^-]$	_	_	2e	1.760	
$\varepsilon_{\mathrm{D}} \left[\mathrm{g}^{-} \; \mathrm{g}^{+} \right]$	1a	1.733	_	_	
$\delta_{\rm D} \left[{\rm g}^+ {\rm g}^- \right]$	1a	1.655	_	_	
$\gamma_L [g^+g^-]$	1a	1.602	_	_	
$\alpha_L [a g^-]$	_	_	2e	1.706	
Hasp ⁻ form					
$\alpha_D [g^+ a]^{a1}$	_	_	2a	1.407	
$\delta_D [a g^-]$	_	_	_	_	
$\varepsilon_D [g^+g^+]^{b1}$	_	_	2a	1.737	
$\varepsilon_{\rm D} \left[{\rm g}^+ {\rm a} \right]$	_	_	2a	1.532	
$\varepsilon_{\mathrm{D}} \left[\mathrm{g}^{-} \; \mathrm{g}^{+} \right]$	1a	1.850	2a	1.438	
$\gamma_L [g^+g^+]^{c1}$	_	_	2a	1.658	
$\gamma_L [g^- g^+]$	1a	1.801	2a	1.446	
$\varepsilon_L \left[g^+a\right]^{d1}$	_	_	2a	1.552	
$\varepsilon_{\rm L} [g^+g^-]$	_	_	2a	1.632	
Asp ²⁻ form					
$\gamma_D [g^+ a]^{a2}$		_	2a	2.25	
$\alpha_{\rm L} [{\rm g}^- {\rm g}^+]^{\rm b2}$		_	2a	1.947	
$\beta_L [g^+a]^{c2}$	_	_	2a	2.027	

 a1 Identical to γ_L [g ^+a]. b1 Identical to β_L [g $^+g^+$]. c1 Identical to δ_L [g $^+g^+$]. d1 Identical to β_L [g ^+a]. a2 Identical to γ_D [g $^+g^-$], ϵ_D [g ^+a], ϵ_D [g $^+g^-$]. b2 Identical to α_L [g ^-a], ϵ_L [g $^-g^+$], ϵ_L [g ^-a]. c2 Identical to β_L [g $^+g^-$], δ_L [g ^+a], δ_L [g $^+g^-$].

structures of species H₂asp and Hasp of aspartic acid have been performed at the DFT/B3LYP/6-31G(d) level of theory. Conformations of 34, 11, 9 and 3 were found from 324, 162, 81 and 81 possible conformers for species H₃asp⁺, H₂asp, Hasp⁻ and asp²⁻, respectively. Based on the B3LYP/6-31+G(d,p) computations of different structural models (the bare, trihydrated and tetrahydrated structures), the conformation for the species H_3asp^+ γ_D [g-g+]), H_2 asp α_L [a g-]), $Hasp^-$ (ϵ_L [g+a]) and asp^{2-} (β_L [g+a]) are not changed. In gas phase, the species H₂asp and Hasp must be stabilized by at least three water molecules but the bare structures of the species H₃asp⁺ and asp²⁻ are stable. Three protonation steps due to the tetrahydrated species of asp²⁻, Hasp⁻ and H₂asp of aspartic acid computed at B3LYP/6-31+G(d, p) level of theory with zero-point vibrational energy corrections are exothermic reactions and their stabilization energies of the stepwise protonation are -244.68, -308.57 and -379.97 kcal mol⁻¹, respectively.

Table 3 B3LYP/6-31+G(d, p)-optimized conformations of various species of trihydrated and tetrahydrated forms of aspartic acid, their selected geometrical data and total energies

Species	Conformations	Optimized parameters						
		ф	ψ	χ1	χ ₂	χ ₃	ω	$E_{\text{total}}(\text{hartree})^{\text{a}}$
Trihydrated 1	form							
H ₃ asp ⁺	$\gamma_{\rm D} \left[{\rm g}^- {\rm g}^+ \right]$	25.36	-18.98	-50.18	27.13	0.68	0.76	-741.887223
H ₂ asp	$\alpha_L [a g^-]$	-72.92	-18.69	145.70	-106.50	166.30	_	-741.502018
Hasp ⁻	$\varepsilon_{\rm L} [g^+ a]$	-36.74	-149.30	43.77	143.40	_	_	-741.001475
asp ² -	$\beta_L [g^+a]$	-169.70	-163.70	58.26	167.41	_	_	-740.389035
Tetrahydrate	d form							
H ₃ asp ⁺	$\gamma_{\rm D} \left[{\rm g}^- {\rm g}^+ \right]$	30.30	-34.67	-54.37	26.65	0.23	0.62	-818.311997
H ₂ asp	$\alpha_L [a g -]$	-79.39	-19.14	148.89	-108.89	166.20	_	-817.922080
Hasp ⁻	$\varepsilon_{\rm L} [g^+ a]$	-39.11	-150.30	47.78	134.33	_		-817.430348
Asp ² -	$\beta_L [g^+a]$	-165.20	-153.60	58.87	157.00	_	_	-816.824823

^a With ZPVE correction.

Table 4 B3LYP/6-31+G(d, p)-optimized conformations of various species of aspartic acid in gas phase

Species	Conformation	Conformation		
	Bare	Trihydrated	Tetrahydrated	
H ₃ asp ⁺	$\gamma_{\mathrm{D}} \left[\mathrm{g}^{-} \; \mathrm{g}^{+} \right]^{\mathrm{a}}$	$\gamma_{\mathrm{D}} \left[\mathrm{g}^{-} \; \mathrm{g}^{+}\right]^{\mathrm{a}}$	$\gamma_D [g^- g^+]^a$	
H ₂ asp	_b	$\alpha_{\rm L} [{\rm a g}^-]$	$\alpha_{\rm L}$ [a g ⁻]	
Hasp ⁻ Asp ²⁻	_b	$\varepsilon_{\rm L} [{\rm g}^+ {\rm a}]$	$\varepsilon_{\rm L} \left[{\rm g}^+ {\rm a} \right]^{\rm c}$	
Asp ²⁻	$\beta_{\rm L} [g^+a]^{\rm d}$	$\beta_L [g^+a]$	$\beta_L [g^+a]$	

^a Identical to $\delta_D [g^-g^+]$.

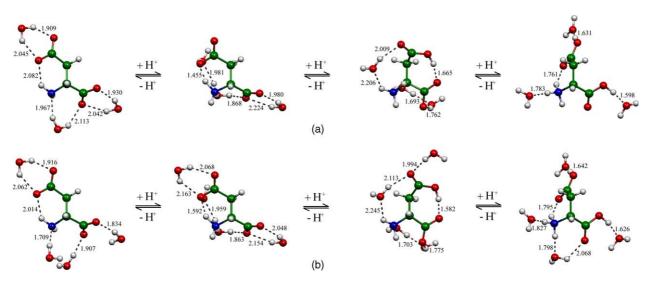


Fig. 6. The B3LYP/6-31+G(d,p) optimized structures of the most stable conformers of four species of aspartic acid as (a) trihydrated and (b) tetrahydrated forms and their protonation schemes. The hydrogen bond distances are in Å.

b Unstable species.

 $[\]begin{array}{c} \text{Climator spectrum} \\ \text{Climator of } \beta_L \ [g^+a]. \\ \text{d} \ \ \text{Identical to} \ \beta_L \ [g^+g^-], \ \delta_L \ [g^+a] \ \text{and} \ \delta_L \ [g^+g^-]. \end{array}$

Table 5 Stabilization energies of protonation reaction of tetrahydrated forms of species asp^{2-} , $Hasp^{-}$ and H_2asp of aspartic acid, computed at B3LYP/6-31+G(d, p) level of theory

Protonation/reaction steps	$\Delta E^{ m a,b}$
ΔE_1 :asp ² - (4H ₂ O) + H ⁺ \rightarrow Hasp - (4H ₂ O)	-244.68
ΔE_2 : Hasp $(4H_2O) + H^+ \rightarrow H_2$ asp $(4H_2O)$	-308.57
ΔE_3 : H ₂ asp(4H ₂ O) + H ⁺ \to H ₃ asp ⁺ (4H ₂ O)	-379.97

- a In kcal mol-1.
- ^b With ZPVE corrections.

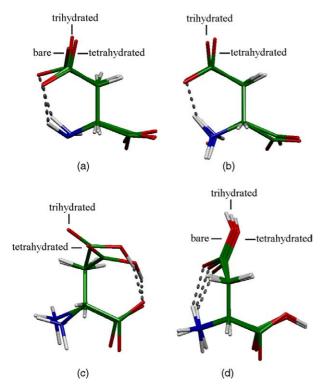


Fig. 7. Superimpose pictures of the geometrical changes of the most stable conformers of species (a) $\rm asp^{2-}$, (b) $\rm Hasp^{-}$, (c) $\rm H_2asp$ and (d) $\rm H_3asp^{+}$. For clarity, water molecules are not displayed.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.theochem.2005.10.036

References

- [1] J.T. Edsall, M.H. Blanchard, J. Am. Chem. Soc. 55 (1933) 2337.
- [2] B. Noszál, P. Sándor, Phys. Chem. 90 (1986) 6345.

- [3] B. Noszál, P. Sándor, Anal. Chem. 61 (1989) 2631.
- [4] P.K. Smith, E.R.B. Smith, J. Biol. Chem. 146 (1942) 187.
- [5] CRC Handbook of Chemistry and Physics, 66 ed., CRC Press, Boca Raton, Florida, 1985.
- [6] P.I. Nagy, B. Noszál, J. Phys. Chem. A 104 (2000) 6834.
- [7] J.C.P. Koo, G.A. Chass, A. Perczel, Ö. Farkas, L.L. Torday, A. Varro, J.G. Papp, I.G. Csizmadia, J. Phys. Chem. A 106 (2002) 6999.
- [8] J.C.P. Koo, J.S.W. Lam, G.A. Chass, L.L. Torday, A. Varro, J.G. Papp, J. Mol. Struct. (THEOCHEM) 620 (2003) 231.
- [9] J.C.P. Koo, J.S.W. Lam, G.A. Chass, D.H. Setiadi, J.M.S. Law, J.G. Papp, B. Penke, I.G. Csizmadia, J. Mol. Struct. (THEOCHEM) 666 (2003) 279.
- [10] J.C.P. Koo, G.A. Chass, A. Perczel, Ö. Farkas, L.L. Torday, A. Varro, J.G. Papp, I.G. Csizmadia, Europhys. J. D 20 (2002) 499.
- [11] J.C.P. Koo, J.S.W. Lam, S.J. Salpietro, G.A. Chass, R.D. Enriz, L.L. Torday, A. Varro, J.G. Papp, J. Mol. Struct. (THEOCHEM) 619 (2002) 143.
- [12] S.J. Salpietro, A. Perczel, Ö. Farkas, R.D. Enriz, I.G. Csizmadia, J. Mol. Struct. (THEOCHEM) 497 (2000) 39.
- [13] T. Kimura, N. Matubayasi, H. Sato, F. Hirata, M. Nakahara, J. Phys. Chem. B 106 (2002) 12336.
- [14] R. Flaig, T. Koritsanszky, D. Zobel, P. Lugar, J. Am. Chem. Soc. 120 (1998) 2227.
- [15] A. Láng, A.K. Füzéry, T. Beke, P. Hudáky, A. Perczel, J. Mol. Struct. (THEOCHEM) 675 (2004) 163.
- [16] D. Siodłak, M.A. Broda, B. Rzeszotarska, J. Mol. Struct. (THEOCHEM) 668 (2004) 75.
- [17] M.F. Masman, M.G. Amaya, A.M. Rodriguez, F.D. Suvire, G.A. Chasse, Ö. Farkas, A. Perczel, R.D. Enriz, J. Mol. Struct. (THEOCHEM) 543 (2001) 203.
- [18] M. Tarditi, M.W. Klipfel, A.M. Rodriguez, F.D. Suvire, G.A. Chasse, Ö. Farkas, A. Perczel, R.D. Enriz, J. Mol. Struct. (THEOCHEM) 545 (2001) 29.
- [19] M.N. Barroso, E.S. Cerutti, A.M. Rodriguez, E.A. Jauregui, Ö. Farkas, A. Perczel, R.D. Enriz, J. Mol. Struct. (THEOCHEM) 548 (2001) 21.
- [20] P. Chakrabarti, D. Pal, Prog. Biophys. Mol. Biol. 76 (2001) 1.
- [21] IUPAC-IUB Commission on Biochemical Nomenclature, Biochemistry, 9 (1970) 3471.
- [22] P.C. Hariharan, J.A. Pople, Theor. Chim. Acta. 28 (1973) 213.
- [23] A.D. Becke, Phys. Rev. A 38 (1988) 3098.
- [24] C. Lee, W. Yang, R.G. Parr, Phys. Rev. B 37 (1988) 385.
- [25] M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria, M.A. Robb, J.R. Cheeseman, J.A. Montgomery, Jr., T. Vreven, K.N. Kudin, J.C. Burant, J.M. Millam, S.S. Iyengar, J. Tomasi, V. Barone, B. Mennucci, M. Cossi, G. Scalmani, N. Rega, G.A. Petersson, H. Nakatsuji, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, M. Klene, X. Li, J.E. Knox, H.P. Hratchian, J.B. Cross, C. Adamo, J. Jaramillo, R. Gomperts, R.E. Stratmann, O. Yazyev, A.J. Austin, R. Cammi, C. Pomelli, J.W. Ochterski, P.Y. Ayala, K. Morokuma, G.A. Voth, P. Salvador, J.J. Dannenberg, V.G. Zakrzewski, S. Dapprich, A.D. Daniels, M.C. Strain, O. Farkas, D.K. Malick, A.D. Rabuck, K. Raghavachari, J.B. Foresman, J.V. Ortiz, Q. Cui, A.G. Baboul, S. Clifford, J. Cioslowski, B.B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R.L. Martin, D.J. Fox, T. Keith, M.A. Al-Laham, C.Y. Peng, A. Nanayakkara, M. Challacombe, P.M.W. Gill, B. Johnson, W. Chen, M.W. Wong, C. Gonzalez, J.A. Pople, GAUSSIAN 03, Revision B.03, Gaussian, Inc., Pittsburgh PA, (2003).
- [26] G. Schaftenaar, MOLDEN 3.7, CAOS/CAMM Center, Nijmegen Toernooiveld, Nijmegen, Netherlands, 1991.
- [27] P. Flükiger, H.P. Lüthi, S. Portmann, J. Weber, MOLEKEL 4.3, Swiss Center for Scientific Computing, Manno (Switzerland), 2000.