

# BFMP: A Method for Discretizing and Visualizing Pyranose Conformations

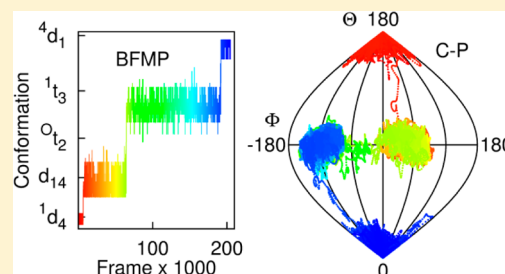
Spandana Makeneni,<sup>†</sup> B. Lachele Foley,<sup>\*,†</sup> and Robert J. Woods<sup>\*,†,‡</sup>

<sup>†</sup>Complex Carbohydrate Research Center, University of Georgia, 315 Riverbend Road, Athens, Georgia 30602, United States

<sup>‡</sup>School of Chemistry, National University of Ireland, Galway, University Road, Galway, Ireland

**S** Supporting Information

**ABSTRACT:** We report a new classification method for pyranose ring conformations called Best-fit, Four-Membered Plane (BFMP), which describes pyranose ring conformations based on reference planes defined by four atoms. The method is able to characterize all asymmetrical and symmetrical shapes of a pyran ring, is readily automated, easy to interpret, and maps trivially to IUPAC definitions. It also provides a qualitative measurement of the distortion of the ring. Example applications include the analysis of data from crystal structures and molecular dynamics simulations.



## INTRODUCTION

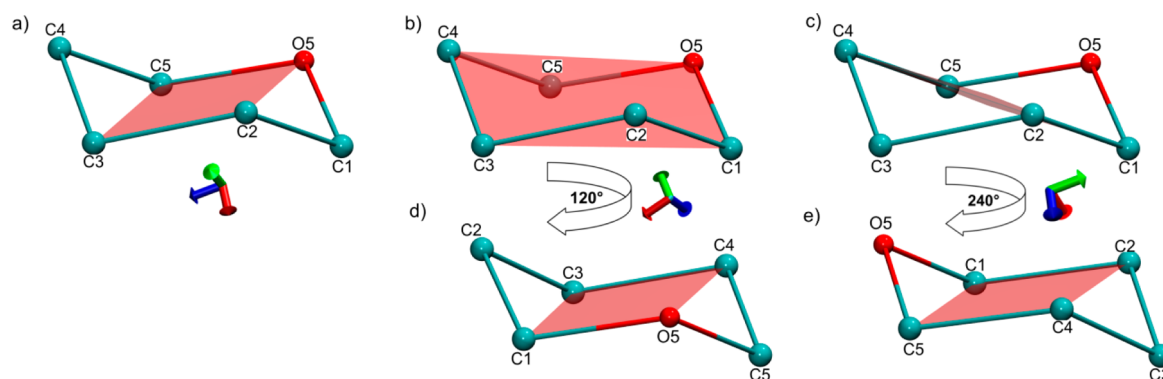
The vast majority of all mammalian glycans exist as branched polymers, in which the monosaccharides are present in their six-membered ring (pyran) form. Pyran ring conformations depend on the nature of the substituents around the ring and may be highly dynamic in solution.<sup>1</sup> In addition, the monosaccharide rings may alter conformation upon binding to enzymes such as glycosyl hydrolases.<sup>2</sup> In some cases, biological function depends on the ability of a protein to bind to a glycan displaying a particular conformation, selected from an ensemble of solution conformations, of the pyran ring. For example, the anticoagulant activity of Antithrombin III depends on the specific interaction of the protein with a bioactive conformation of the polysaccharide heparin.<sup>3,4</sup> Pyran ring conformational propensity has been linked to the chemical reactivity of monosaccharides<sup>5,6</sup> as well as the physiochemical properties, such as elasticity, of the resultant polymers.<sup>7</sup>

The nomenclature adopted by the International Union of Pure and Applied Chemistry (IUPAC) for describing pyran ring conformation<sup>8</sup> divides six-membered ring shapes into 38 distinct conformations: 2 chairs, 6 boats, 6 skew-boats, 12 half-chairs, and 12 envelopes.<sup>9</sup> These descriptors correspond to pyran rings in idealized, symmetrical conformations and do not provide any quantification of the extent to which any given conformation deviates from ideality. However, experimental data from NMR spectroscopy<sup>10</sup> as well as from crystallography<sup>11</sup> show that pyran rings adopt nonidealized, asymmetrical conformations. It is important to precisely quantify the geometry of these structures to understand the process of ring puckering, and methods exist for doing so, but there exist no simple methods for qualitative classification of all ring shapes. Two popular methods are available for the quantification of pyran ring shapes: Whitfield classification<sup>12</sup> and Cremer-Pople parameters.<sup>13</sup> The Whitfield method employs a linear combination of idealized IUPAC shapes to

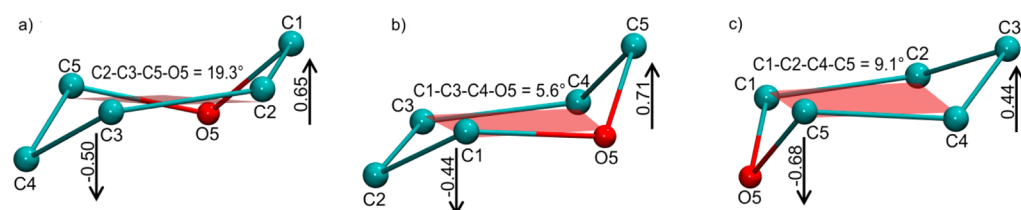
describe ring conformations. For example, a chair form might be characterized as being 89% chair (<sup>1</sup>C<sub>4</sub>) + 8.5% boat (<sup>1</sup><sup>4</sup>B) – 1.9% skew (<sup>0</sup>S<sub>2</sub>).<sup>12</sup> While quantitative, this approach precludes intuitive understanding: it is difficult to construct a mental image from such a linear combination. Cremer-Pople parameters<sup>13</sup> employ a set of abstracted spherical-polar coordinates, *Q*,  $\phi$ , and  $\theta$ , where the polar angle,  $\theta$ , provides a description of the symmetry of puckering around the ring, the phase angle,  $\phi$ , describes the position around the ring where the puckering occurs, and the amplitude, *Q*, describes the magnitude of the puckering. For example, an idealized symmetrical chair might have an amplitude (*Q*) of 0.61, phase angle  $\phi = 0.5^\circ$ , and polar angle  $\theta = 176^\circ$ ; an asymmetrical chair might have *Q* = 0.51,  $\phi = 131^\circ$ , and  $\theta = 157^\circ$ . In practice, *Q* is often ignored, and the values of  $\phi$  and  $\theta$  are plotted on a sphere of constant *Q* (the “*Q*-sphere”). These parameters provide a quantitative description of every possible ring shape, and while mapping the parameters to idealized conformations is straightforward,<sup>14</sup> describing nonstandard ring shapes again requires a linear combination of canonical conformations. More recently Hill and Reilly<sup>15</sup> proposed a quantification method based on a triangular reference plane and a set of three angles. This method is useful to quantify ring puckering and is more intuitive in comparison to the other two methods. However, while basic visualization of the conformation is straightforward, translation to an IUPAC descriptor, where one exists, is not.

Here, we propose a new naming convention, Best-fit Four-Member Plane (BFMP), which can describe all the canonical and asymmetrical conformations adopted by six-membered rings using descriptors comprised of a single letter and one or two numerals. The letters used in the descriptors are derived

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**Figure 1.** a) A pyranose ring in an idealized, symmetrical  ${}^4C_1$  conformation. Atoms C4 and C1 are above and below the reference plane formed by atoms C2, C3, C5, and O5. Two additional planes can also define this conformation: b) with reference atoms C1, C3, C4, and O5 and c) with reference atoms C1, C2, C4, and C5. d) and e) show the planes illustrated in b) and c) from a different view. The reference planes for all other IUPAC shapes are given in Figure S2 and Table S2 in the Supporting Information.



**Figure 2.** a) Distorted  ${}^1C_4$  conformation in PDB ID 1AXM, in which reference atoms C2, C3, C5, and O5 (dihedral =  $19.3^\circ$ ) fit poorly to a plane. b) The best-fit plane in this case is formed by reference atoms C1, C3, C4, and O5 (dihedral =  $5.6^\circ$ ). c) The average dihedral ( $9.1^\circ$ ) for the atoms defining the third  ${}^1C_4$  reference plane falls below the cutoff used in this work, but the fit is still poor when compared to the reference plane in b). Distances from the remaining atoms to the planes are shown in all three cases. The BFMP method may be used to classify this generally as a  ${}^5d_2$  conformation, or with increasing quantitation, as a  ${}^5d_2(5.6^\circ)$ , or  ${}^{5(0.71)}d_2(0.44)(5.6^\circ)$  conformation.

from the number of consecutive atoms in a reference plane, where the reference planes are consistent with those used by IUPAC. For example, a pyranose in a  ${}^4C_1$  conformation has at most two consecutive atoms in the IUPAC reference plane (C2 and C3 or C5 and O, Figure 1) and would be described by BFMP as a  ${}^4d_1$  conformation, where d, for “di”, indicates the two consecutive atoms. Additionally, this method provides quantification of degree of deviation from ideality in two ways. One, the average torsion angle associated with the reference plane represents the coplanarity of the four atoms defining the reference plane. That is, it provides quantification of the degree of distortion of the atoms from their reference plane. Two additional numbers report the distances of the other one or two atoms above or below the reference plane. Any, or none, of these quantifications might be included along with the descriptor. Thus, an idealized (IUPAC) chair conformation would be represented in the BFMP convention as  ${}^4d_1$ , whereas a typical, slightly distorted chair might be represented as  ${}^4d_1$ ,  ${}^4d_1(6^\circ)$ , or as  ${}^{4(0.70)}d_1(0.42)(6^\circ)$ , depending on the information required. This method offers several advantages, including the ability to more precisely describe nonideal conformations without introducing a linear combination of states (Table S1 and Figure S1) as well as retaining a straightforward way to map the new nomenclature back to established IUPAC conformations. In addition, the approach is readily amenable to the automatic detection and characterization of conformational states from experimental or theoretical data. The method and its automation are described below, with applications to an analysis of crystallographic data, as well as data from molecular dynamics (MD) simulations.

## METHODS

**Theory.** Each of the 38 canonical IUPAC shapes contains one, two, or three sets of four coplanar atoms.<sup>16</sup> While 15 unique four-atom sequences exist in a six-membered ring, in only 3 or fewer are the atoms coplanar in any given canonical conformation. For example, a  ${}^4C_1$  conformation contains three such planes. Its standard, reference plane contains atoms C2, C3, C5, and O5, with atoms C4 and C1 above and below the plane, respectively. However, 2 other four-membered planes are also present in, and could be used to define, this canonical, idealized, symmetrical chair conformation (Figure 1).

In the BFMP method, a best-fit plane<sup>13</sup> is calculated for each of the 15 possible four-atom sequences. The plane in which the 4 atoms are closest to being coplanar is chosen as the *reference plane*. Planarity is defined by the average dihedral angles associated with the 4 atoms around the perimeter of each quadrilateral: a value of zero indicates that the 4 atoms are coplanar. The particular conformational assignment also depends on the positions of the remaining 2 atoms above or below the reference plane (Figure 2).

The coplanarity of the four atoms defining the reference plane indicates the maximum extent to which the overall conformation could be symmetrical. That is, a reference plane with a torsion angle close to  $0^\circ$  does not necessarily imply that the remaining two atoms are in symmetrical positions or that the overall conformation approximates one of the 38 idealized IUPAC structures. Therefore, the distances of the remaining two atoms to the reference plane are used to provide additional quantification of the structure’s ideality. If they occupy differing distances from the reference plane, or if they are unusually close to, or far from, it, then the overall shape cannot be symmetrical.

Table 1. BFMP Nomenclature, Including Corresponding IUPAC Nomenclature Where Relevant

Reference Plane Indicator	# Consecutive atoms	Examples
m (mono) <sup>a,b</sup>	n/a	
d (di)	2	 
t (tri)	3	 
q (quatro)	4	 
p (penta)	5	
h (hexa)	6	

<sup>a</sup>Descriptor “m” describes highly distorted structures, for which no set of four atoms is well represented by a plane. <sup>b</sup>Lower case letters are used to avoid confusion with the existing IUPAC nomenclature.

While this combination of dihedral angle and distances cannot be used to perfectly reconstruct the shape, and while it also does not provide strict quantification of ideality, it does provide an intuitive estimate of the minimum deviation from ideality.

While the BFMP descriptors can be used to describe all the ring conformations, an equivalent, dihedral-based method cannot be used to assign descriptors for conformations which employ five (envelope) or six-membered planes. It is not possible to calculate a dihedral for five atoms, and the varying distances between the atoms precludes a simple comparison, for example, of root mean distances of the atoms to reference planes defined by four, five, and six atoms. In such cases, it is useful to employ cutoffs. The procedures used in this work for assigning p-type (envelope) and h-type (planar) conformations are detailed in Section S1. The cutoffs can also be employed to classify highly distorted structures in which none of the atoms in the 15 four-atom sequences are nearly coplanar. For example, in this work, an average dihedral angle greater than 10° is considered too poor an approximation to planarity (Figure S3 and Section S1). Thus, in the case that the best reference plane is described by an average dihedral greater than 10°, the conformation *m* would be assigned. These cutoffs can be altered to suit the requirements of the particular study in which they are employed.

This method not only can readily identify all of the 38 IUPAC conformations but also can be employed to assign asymmetrical shapes to one of an additional 48 conformations. The automation of this method is detailed in the Supporting Information (Section S1 and Figures S3, S4, and S5), and the program is available at <http://glycam.org/publication-materials/bfmp/>. This and future versions of the program and documentation should also be accessible from <http://glycam.org/downloads>.

**BFMP Nomenclature.** The letters used in the nomenclature are derived from the number of consecutive atoms in the reference plane. For example, as stated in the Introduction, a pyranose in a <sup>4</sup>C<sub>1</sub> conformation has 2 consecutive atoms in the reference plane (C2 and C3 or C5 and O, Figure 1) and hence would be described as a <sup>4</sup>d<sub>1</sub> conformation, where “d”, for “di”, indicates the two consecutive atoms. This designation also corresponds to a boat conformation, which has 2 consecutive atoms in its IUPAC reference plane, and hence a <sup>1,4</sup>B boat

conformation would be described as <sup>1,4</sup>d. A skew boat conformation has 3 consecutive atoms, and a half-chair conformation has 4 consecutive atoms; thus, a <sup>1</sup>S<sub>3</sub> can be referred to as <sup>1</sup>t<sub>3</sub> (“t” for tri) and a <sup>2</sup>H<sub>1</sub> as <sup>2</sup>q<sub>1</sub> (“q” for quatro) using the BFMP descriptors. The letters used in the BFMP naming scheme as well as example standard conformations are given in Table 1.

An advantage of the BFMP nomenclature is that it can describe many more conformations than encompassed by the IUPAC descriptions. For example, the iduronic acid (IdoA) residue in the CTX A3–heparin hexasaccharide complex (PDB ID 1XT3)<sup>17</sup> can be described as <sup>1,0</sup>q (4.2°) using a BFMP descriptor. The closest IUPAC conformation is <sup>1</sup>H<sub>O</sub> and shares a reference plane (C2, C3, C4, and C5) with the <sup>1,0</sup>q descriptor. However, the <sup>1</sup>H<sub>O</sub> description incorrectly defines the positions of the out-of-plane atoms (C1 and O5) (Figure 3).

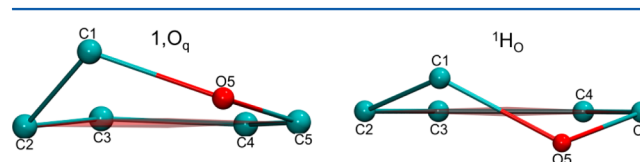
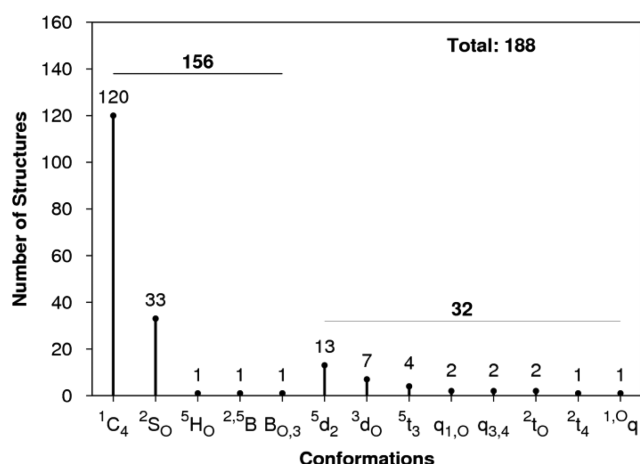


Figure 3. Ring conformation of IdoA in PDB ID 1XT3 (left) and nearest IUPAC structure (right).

**Example Applications. Conformational Analysis of Iduronate Residues in the PDB.** IdoA, predominately in its iduronate form, is a major component of glycosaminoglycans (GAGs), such as heparin and heparan sulfate. Unlike most monosaccharides, which exist primarily in a single ring conformation, IdoA adopts multiple ring conformations.<sup>5,18,19</sup>

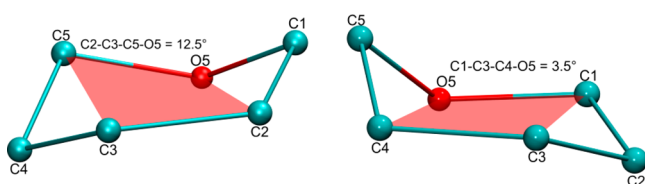
A survey of the PDB<sup>20</sup> identified 188 IdoA residues, of which 83% could be assigned, using the cutoffs described in the SI, to standard IUPAC conformations (<sup>1</sup>C<sub>4</sub>, <sup>2</sup>S<sub>O</sub>, <sup>3</sup>S<sub>1</sub>). Thus, 17% of the conformations of IdoA appeared to be nonstandard (Figure 4).

Of the 32 nonstandard conformations, using BFMP, 24 could be classified as <sup>5</sup>d<sub>2</sub>, <sup>3</sup>d<sub>0</sub> (chair), or <sup>5</sup>t<sub>3</sub> (skew) conformers, which are distorted forms of the <sup>1</sup>C<sub>4</sub> and <sup>2</sup>S<sub>O</sub> forms, respectively. For example, the IdoA ring in the crystal structure NK1–heparin complex<sup>21</sup> has Cremer-Pople parameters of  $\varphi = 131.4$ ,  $\theta =$



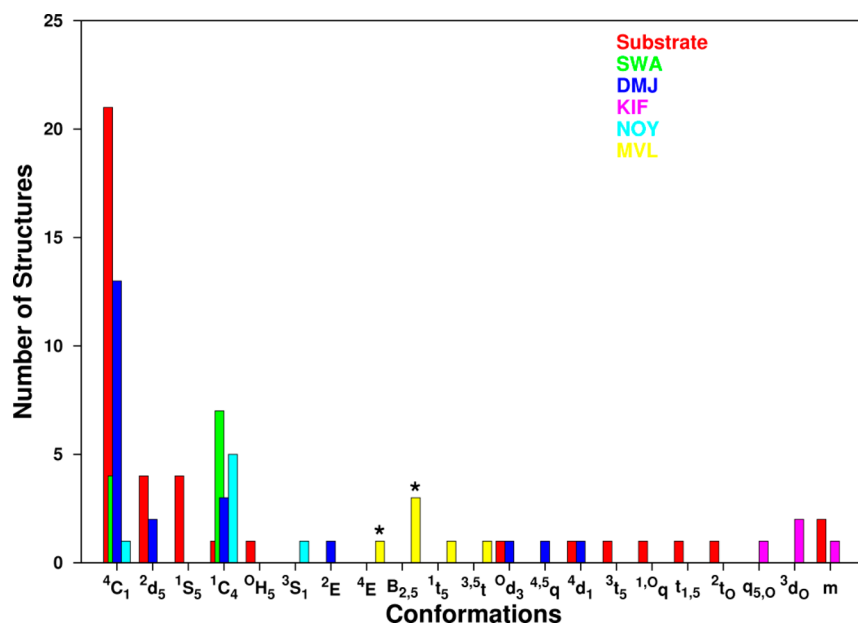
**Figure 4.** BFMP classification of 188 IdoA residues present in crystal complexes, of which 32 are in nonstandard conformations.

157.6, which when mapped onto a constant-Q sphere places the ring between a  ${}^1C_4$  chair and a  ${}^5H_0$  half-chair. Only by visually examining an image of the shape is it possible to deduce that it is a chairlike conformation. In contrast, the BFMP classification directly identifies the structure as a  ${}^5d_2$  chair (Figure 5).



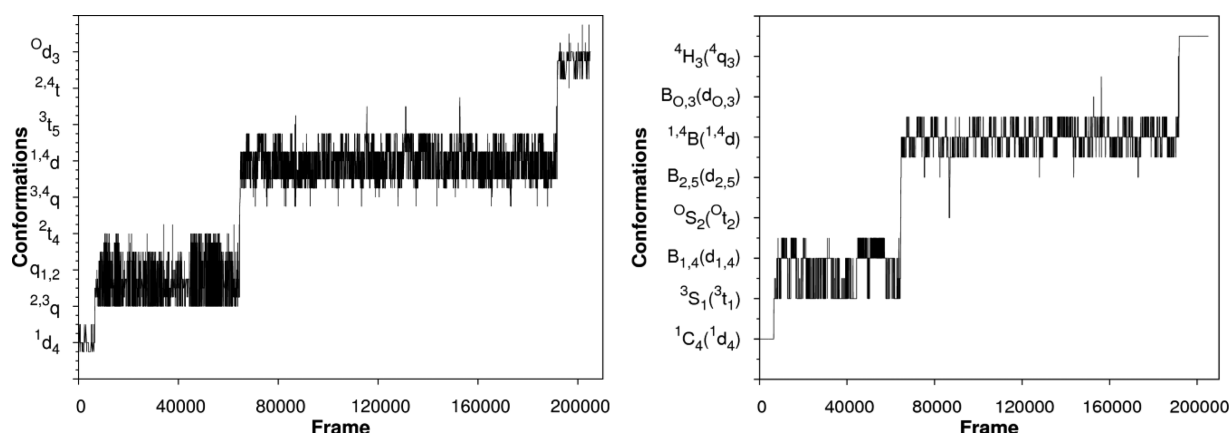
**Figure 5.** Ring conformation of IdoA in PDB ID 1GMO. The residue is in a conformation partway between  ${}^1C_4$  and  ${}^5H_0$  (left). The BFMP method provides a less ambiguous classification as the  ${}^5d_2$  conformer (right) since atoms C1, C3, C4, and O5 best approximate a plane.

**Application in Inhibitor Design.** Aberrant cell-surface glycosylation is an indicator of malignant transformations and is often used to diagnose and monitor the stage of tumor progression.<sup>22–24</sup> Alterations in glycosylation are caused by changes in enzyme activity; for example altered activity of glycosyltransferases and glycosidases in cancer cells leads to modified glycoproteins.<sup>25</sup> Human breast and colon cancers often overexpress *N*-acetylglucosaminyltransferase V (GlcNAcT-V), which leads to an increase in branched *N*-glycans,<sup>26</sup> and inhibition of earlier stage  $\alpha$ -mannosidase activity has shown clinical potential in treating malignancies.<sup>27,28</sup> Inhibitors may be designed to mimic the ground-state or transition-state conformations of the substrate; in the crystal structure of a Golgi  $\alpha$ -mannosidase II in complex with the inhibitor mannoimidazole,<sup>29</sup> the six-membered ring of the inhibitor adopts a conformation similar to that of the presumed ( $B_{2.5}$ ) transition state<sup>30</sup> (see Figure 6). Here we employed the BFMP method to classify the conformations of the terminal mannose ring in exomannosidases present in the PDB (22 complexes) and compared them to the conformations of known inhibitors swainsonine, deoxymannojirimycin, noeumycine, kifunensine, and mannoimidazole extracted from enzyme cocomplexes (49 total complexes) (Table S3). The results showed that 51% of the substrates and 36% of the inhibitors adopted a  ${}^4C_1$  conformation. Of the remainder, 35% of the substrates and 22% of the inhibitors were present in asymmetrical conformations. Kifunensine in particular appeared to exist only in asymmetrical conformations. Very often, these asymmetrical shapes cannot be classified qualitatively and are described as distorted chair or skew-boat conformations. The BFMP method can be used not only to qualitatively describe these conformations but also to quantify the orientation of the atoms relative to the best-fit plane of the ring. This information may be helpful in identifying inhibitors that optimally mimic the orientation of atoms of the substrate in the enzyme substrate complex.<sup>30</sup>

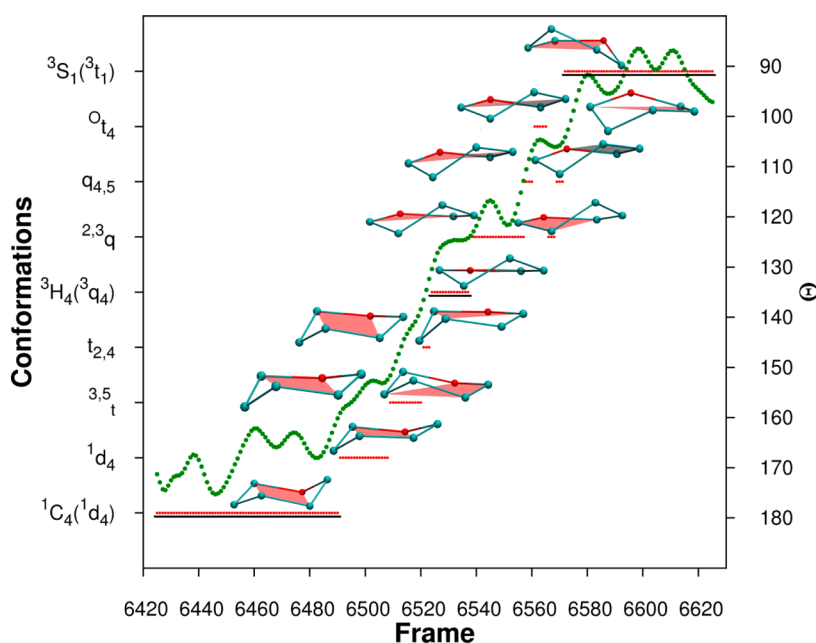


**Figure 6.** Comparison of ring conformations of mannopyranose substrates (red) extracted from cocrystal structures with mannosidases ( $n = 41$ ), and the inhibitors ( $n = 50$ ) swainsonine (SWA, green), deoxymannojirimycin (DMJ, blue), kifunensine (KIF, pink), noeumycine (NOY, cyan), and mannoimidazole (MVL, yellow). Presumed transition states for mannosidases indicated by (\*).





**Figure 7.** Ring conformations, (left) identified by the BFMP method and (right) identified here as being well-described by IUPAC, over the course of a 10 ns MD simulation of Ido plotted as a function of simulation frame. For visual clarity, only subsets of the BFMP and IUPAC descriptors are noted on the y-axes. The complete list of BFMP conformations is provided in Table S4.



**Figure 8.** Frames, corresponding to 200 fs, extracted from the MD simulation shown in Figure 7. The BFMP descriptors are shown by red lines. The lines are underlined for shapes that can be identified as standard IUPAC conformations  ${}^1C_4$ ,  ${}^3H_4$ , and  ${}^3S_1$ . Cremer-Pople  $\theta$  values are plotted as green dots. Schematic representations of the BFMP-defined conformations, in order of occurrence, during the transition from  ${}^1C_4$  to  ${}^3H_4$  to  ${}^3S_1$  conformations are also shown. The structures shown to the left of the green dots are oriented with respect to the IUPAC reference planes, and the structures shown on the right are oriented to reference planes used by BFMP.

**Using BFMP To Define Conformational Interconversions of Pyranose Rings.** The pseudorotation pathways associated with the interconversion of ring forms can provide insight into enzyme mechanisms<sup>31–34</sup> and chemical reactions.<sup>35</sup> Molecular dynamics (MD) simulations can be particularly useful to study the dynamic properties of flexible molecules such as carbohydrates. Here we demonstrate the strengths of the BFMP method monitoring the conformational changes that occur during a 10 ns MD simulation (details in Section S2 of the SI) of  $\alpha$ -L-idopyranose (Ido). The pyranose ring in Ido is known to interchange between at least three conformations in solution:  ${}^4C_1$ ,  ${}^1C_4$ , and  ${}^2S_0$ ,<sup>36,37</sup> and stable nonstandard conformations between the  ${}^2S_0$  and  ${}^1S_3$  regions have been identified.<sup>10</sup> Although 10 ns is far too short a time scale to observe conformational convergence,<sup>1</sup> it is long enough to capture many of the conformational states of the Ido ring.

Analysis of the MD data showed that the ring converted from a  ${}^1C_4$  to a  ${}^4C_1$  conformation, populating over 50 intermediate conformations characterizable by BFMP. Only 15 of these conformers could be described using IUPAC nomenclature (Figure 7). The IUPAC conformers account for only 39% of the total number of conformations.

The BFMP descriptors provide a uniquely detailed insight into the interconversion pathway. In contrast, the use of IUPAC descriptors fails to capture the continuity of the changes in ring shape associated with this process (Figure 8) and implies a rapid flip from the  ${}^1C_4$  to  ${}^3H_4$  to  ${}^3S_1$  states, that is not supported by the MD data.

While CP parameters can be calculated for each of these states, it is difficult to deconvolute these into discrete states. The  $\theta$  values for these states range from  $90^\circ$  to  $180^\circ$  (Figure 8). However, it is difficult to map each of these values to one of the

standard IUPAC conformations. The intermediate values between  $90^{\circ}$ – $105^{\circ}$ ,  $120^{\circ}$ – $135^{\circ}$ , and  $165^{\circ}$ – $180^{\circ}$  can be classified as skew-boat ( $^3S_1$ ) half-chair ( $^3H_4$ ), or chair ( $^1C_4$ ), respectively (Figures S6 and S7). The remaining values cannot be mapped back to a standard IUPAC conformation and hence the conformations for these states cannot be identified without visualization of the shape. Even when there is an obvious IUPAC state as determined by BFMP, it will sometimes map to a different state using the typical IUPAC–CP mappings (Figures S7 and S8). This mapping issue arises because the two classification methods are not equivalent: the Cremer-Pople method quantifies rings based on an average six-membered reference plane, whereas the IUPAC descriptors are based on idealized four-membered planes.

An analysis of the MD data in terms of BFMP descriptors provides a straightforward, objective classification of more conformational states associated with the conformational interconversion of the Ido ring than possible within the IUPAC constraints.

## CONCLUSIONS

IUPAC nomenclature and Cremer-Pople parameters are the most commonly used methods to describe and quantify the conformations adopted by pyran rings. We have shown that the IUPAC conformations cannot describe the nonstandard conformations adopted by these rings. Because of this, while Cremer-Pople parameters can be calculated for any given ring shape, mapping them to one of the IUPAC conformations can be cumbersome if not impossible. The BFMP conformations are described in terms of reference planes, making it trivial to translate to IUPAC nomenclature when relevant, but giving sufficient diversity to classify nonstandard conformations. In addition, the BFMP descriptors provide an objective method for identifying and comparing six-membered ring shapes, facilitating discussions of mechanistic details and transition state structures.

## ASSOCIATED CONTENT

### Supporting Information

Execution protocol for this classification method. Additional tables and figures that aid in understanding the method are also listed here. This material is available free of charge via the Internet at <http://pubs.acs.org>.

## AUTHOR INFORMATION

### Corresponding Authors

\*E-mail: [lfoley@uga.edu](mailto:lfoley@uga.edu) (B.L.F.).

\*E-mail: [rwoods@ccrc.uga.edu](mailto:rwoods@ccrc.uga.edu) (R.J.W.).

### Author Contributions

S.M.: Designed and performed the applications; identified application targets; analyzed the data; wrote the source code for and codesigned the automation of the method; authored the paper and prepared figures. B.L.F.: Conceived, designed, and directed the project; authored the paper. R.J.W.: Specified application targets and methods for data presentation; authored the paper.

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### Notes

The authors declare no competing financial interest.

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## ABBREVIATIONS

IUPAC, International Union of Pure and Applied Chemistry; MD, Molecular Dynamics; CP, Cremer-Pople; PDB, Protein Data Bank

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