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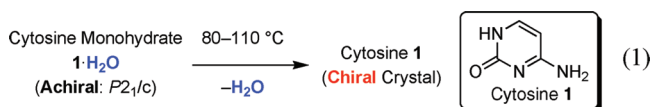
Generation of Absolute Controlled Crystal Chirality by the Removal of Crystal Water from Achiral Crystal of Nucleobase Cytosine

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Crystallization of achiral compounds in enantiomorphs has been considered as one of the candidates for the origin of homochirality.^{1,2} However, such chiral crystallization³ does not show any preference for the formation of one or the other enantiomorphs; that is, the produced crystalline chirality is spontaneously generated to follow the bimodal random distribution.⁴ If it is possible to control the handedness of crystalline chirality formed from an achiral compound without using a chiral auxiliary, this should become another entry for the origin of chirality.⁵



Here we report on the generation of crystal chirality by the thermal dehydration of the crystal water of cytosine⁶ monohydrate $1 \cdot \text{H}_2\text{O}$, which belongs to achiral $P2_1/c$ (eq 1). The thermal direction controls the crystal chirality of the resulting anhydrous crystal **1**. That is, the dehydration of crystal water from the enantiotopic faces of an achiral crystal led to enantiomorphic crystals. Although there were reports on the chiral phenomena based on the enantiotopic faces of the achiral crystal,⁷ to the best of our knowledge, there are no examples of crystal formation in which the enantiomorphs were efficiently controlled by the dehydration of crystal water.

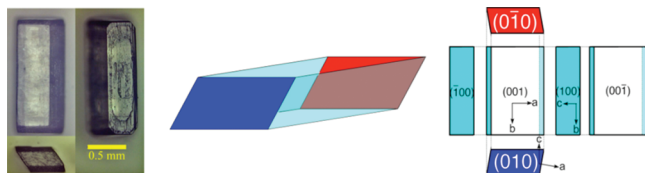


Figure 1. Single crystal of cytosine monohydrate $1 \cdot \text{H}_2\text{O}$.

Although the cytosine crystal obtained from methanol belongs to the *chiral* space group $P2_12_12_1$,^{8,9} crystallization in water affords the achiral monohydrate crystal belonging to the *achiral* $P2_1/c$.¹⁰ Single crystals of $1 \cdot \text{H}_2\text{O}$ with well-defined crystal faces can be obtained from water by slow evaporation at room temperature. The crystal morphology and Miller indices are shown in Figure 1. Each single crystal has parallelogram (010) and (0 $\bar{1}$ 0) faces, which are vertical to the *b*-axis. These nonsuperimposable parallelogram faces are defined as blue colored *b*¹- and red colored *b*²-faces (Figure 2). Then, the crystal was heated from these opposite faces by placing the respective faces on a hot plate (80–110 °C). After the whole crystal became cloudy, i.e., the crystal water had completely disappeared,¹¹ these solid white crystals were characterized by solid-state circular dichroism (CD) analysis¹² as KBr pellets. In the case of the resulting white solid heated from the *b*¹-face, the negative Cotton effect was detected at approximately 310 nm ([CD(–)310_{KBr}]-

1). On the other hand, when the thermal direction was from the *b*²- to the *b*¹-face, the positive Cotton effect was observed at approximately 310 nm ([CD(+)-310_{KBr}]-**1**).

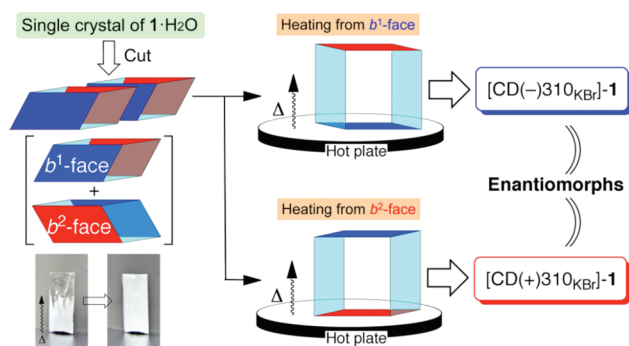


Figure 2. Generation of crystal chirality by the removal of crystal water of cytosine monohydrate $1 \cdot \text{H}_2\text{O}$.

To exclude any effect other than that of the shape of the crystal faces, a single crystal was cut into two pieces vertical to the *b*-axis, and the newly formed faces were placed on the hot plate to perform the dehydration step. In these experiments, the induction sense was the same as observed above without any exception, i.e., dehydration from the *b*₁- and *b*₂-face gave [CD(–)310_{KBr}]- and [CD(+)-310_{KBr}]-**1**, respectively (Figure 2).

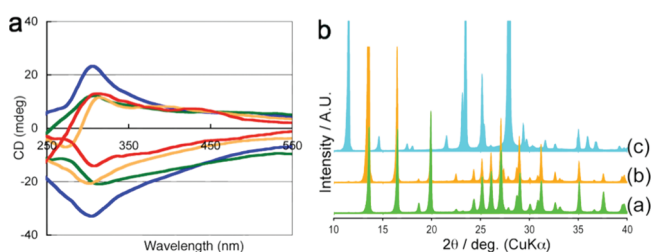


Figure 3. Solid-state properties of cytosine crystals. **a**: Solid-state CD of anhydrous **1**. Lines of the same color indicate the CD signals of the anhydrous crystals that arise from the identical single crystals of $1 \cdot \text{H}_2\text{O}$. **b**: X-ray powder diffraction patterns. (a) Cytosine **1** ($P2_12_12_1$; simulation from single crystal data). (b) Dehydrated cytosine **1** (observed pattern). (c) Cytosine monohydrate $1 \cdot \text{H}_2\text{O}$ ($P2_1/c$; simulation from single crystal data).

The relationship between the face images on the hot plate and the sign of the CD at 310 nm of anhydrous cytosine **1** is highly reproducible, as shown in Figure 3a. Although, the resulting white crystal is not a single crystal, the X-ray powder diffraction patterns of each crystal are shown in Figure 3b. The diffraction pattern of the dehydrated anhydrous cytosine **1** is essentially the same as the simulated pattern of the $P2_12_12_1$ crystal. The pattern of $1 \cdot \text{H}_2\text{O}$ is distinctly different from that of anhydrous **1**; thus, the cytosine

monohydrate $1 \cdot \text{H}_2\text{O}$ is considered to be transferred to the crystal, similar to the case of the $P2_12_12_1$ by the thermal dehydration of crystal water.

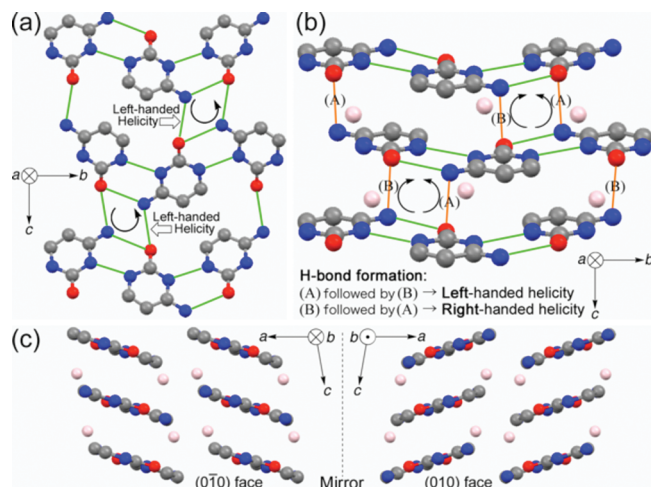
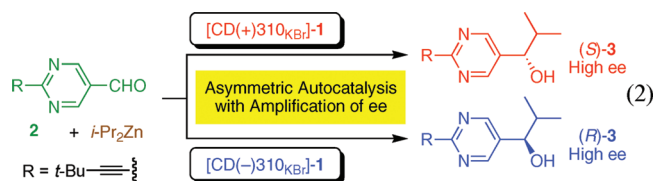


Figure 4. Formation of helical H-bond network by the thermal dehydration of crystal water.

We assumed that the crystal chirality would emerge after reconstitution of the hydrogen bond (H-bond) network in the crystal. In the single crystal structure of anhydrous cytosine **1** ($P2_12_12_1$), shown in Figure 4a, the H-bond between the carbonyl and primary amino group of neighboring molecules (indicated as the green lines) forms a helical arrangement (left-handed crystal was indicated).¹³ A similar (but not helical) H-bond structure can be seen in the crystal of $1 \cdot \text{H}_2\text{O}$ (recrystallized from water), viewed along the a -axis (Figure 4b). The green lines represent the H-bonds originally present in the crystal, which correspond to those of the $P2_12_12_1$ crystal, and the orange lines indicate the newly formed H-bonds (A) and (B). The carbonyl and primary amino groups connected by H-bonds (A) and (B) are originally bonded by water mediated H-bonds. Thus, the removal of crystal water should lead to the creation of (A) and (B), which constitute the helical arrangement in the dehydrated crystal. It can also be seen that the order of formation between (A) and (B) determines the handedness of the resulting helical H-bond network. That is, the initial formation of H-bond (A) followed by H-bond (B) should induce the left-handed helicity, and *vice versa*. Because the direction-selective thermal transfer would control the order of the new H-bond formation, the enantiomorphous dehydrated crystal of cytosine **1** can therefore be obtained by heating from the enantiotopic (0 $\bar{1}$ 0) and (010) faces of the achiral crystal of $1 \cdot \text{H}_2\text{O}$ (Figure 4c).¹⁴



In addition, the dehydrated **1** acted as the origin of chirality in asymmetric autocatalysis^{15,16} to afford enantioenriched products with the absolute configurations correlated to the sign of CD of resulting **1** as shown in eq 2.

In summary, we have demonstrated that the enantioselective formation of the chiral crystal of achiral cytosine can be obtained by the removal of crystal water. Thermal dehydration of the achiral crystal of cytosine monohydrate induces the chirality of the remaining anhydrous cytosine. The heat transfer from the enantiotopic (010) and (0 $\bar{1}$ 0) faces of the achiral single crystal of $1 \cdot \text{H}_2\text{O}$ affords the enantiomorphous anhydrous crystal of cytosine **1** with the chirality corresponding to that of the crystal face.

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Supporting Information Available: TG/DTA of cytosine monohydrate, asymmetric autocatalysis using dehydrated cytosine, comparison between the solid-state CD of anhydrous **1** using KBr and nujol mulls, and the determination of the absolute structure of $P2_12_12_1$ crystal of cytosine. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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