Journal of Chemical Theory and Computation

Free Energy and Kinetics of Conformational Transitions from Voronoi Tessellated Milestoning with Restraining Potentials

Luca Maragliano, † Eric Vanden-Eijnden, ** † and Benoît Roux ** ¶. †

Department of Biochemistry and Molecular Biology, University of Chicago, 5801 South Ellis Avenue, Chicago, Illinois 60637, Courant Institute of Mathematical Sciences, New York University, 251 Mercer Street, New York, New York 10012, USA, and Biosciences Division, Argonne National Laboratory, 9700 South Cass Avenue, Argonne, Illinois 60439, USA

Received May 30, 2009

Abstract: Milestoning is a method aimed at reconstructing the statistical properties of the long-time dynamics of a system by exploiting the crossing statistics of a set of hypersurfaces, called the "milestones", placed along the reaction coordinate [Faradjian and Elber, J. Chem. Phys. 2004, 120, 10880]. Recently, Vanden-Eijnden and Venturoli [J. Chem. Phys. 2009, 130, 194101] showed that when a complete Voronoi tessellation of the configurational space is available, milestoning can be reformulated to utilize the statistics from a series of independent simulations, each confined within a single cell via strict reflections at the boundaries. As a byproduct, this "Voronoi tessellated milestoning" method also permits to compute the free energy of the tessellation. Here, the method is extended to support the usage of differentiable restraining potentials to confine the trajectories within each cell.

1. Introduction

Conformational changes in large biomolecules are complex and slow processes taking place on very long time scales, which often extend well beyond the reach of brute force molecular dynamics (MD) simulations. An emerging class of techniques attacks this problem by first trying to determine an optimal reaction pathway (or pathways) for the transition in a space of very high dimensionality, without making any a priori assumptions about the mechanism.¹⁻⁵ Once such an optimal pathway has been identified, a strategy then consists of inferring the statistical properties of the long-time dynamics by exploiting the information harvested from relatively short independent trajectories.⁶⁻⁹ Of particular interest is the milestoning method introduced by Faradjian and Elber,8 in which the dynamical properties of a reactive event are reconstructed out of a series of short trajectories between a set of hypersurfaces (the "milestones"), disposed sequentially along the reaction pathway between the reactant and the product. 10-14 Recently, Vanden-Eijnden and Venturoli extended milestoning by identifying the edges of the cells of a complete Voronoi tessellation of configurational space as the milestones. 9 It was shown that the "Voronoi tessellated milestoning" could be formulated to require only a series of independent simulations, each confined within a single Voronoi cell, leading to a simplification and an increase in robustness of the original algorithm. The confinement was realized by a simple collision rule via strict hard-wall reflections (velocity inversion) at the boundaries.⁴

Despite its formal simplicity, the hard-wall boundary condition involves modifications of the dynamical propagation algorithm at the heart of MD source codes. This makes its implementation in widely used biomolecular simulation MD packages (e.g., AMBER,15 CHARMM,16 GROMACS,17 and NAMD¹⁸) somewhat cumbersome, and may even affect performance. With the aim of extending the range of applicability of the method, in this work, we propose and test an alternative strategy based on the introduction of continuous and differentiable restraining potentials to confine the system within a given cell. We illustrate how the formalism presented in ref 9 can still be used, with minor modifications, to compute the equilibrium probability and the rates of transitions for the original dynamics. The possibility to use potentials will facilitate the application of the Voronoi tessellated milestoning because the introduction of user-defined external forces is straightfoward. We note that the confinement with potentials can also affect performance because portions of trajectories that are transiently out of the cells are discarded in the analysis, but this effect can be minimized by proper tuning of the parameters in the potentials.

In the remaining part of this letter, we first describe the details of the restraining potentials and the key quantities to be computed for free energy and milestoning calculations, and then

^{*} Corresponding authors. E-mail: eve2@cims.nyu.edu (E.V.-E.); roux@uchicago.edu (B.R.).

[†] University of Chicago.

^{*} New York University.

[¶] Argonne National Laboratory.

we illustrate the method by applying it to two simple examples, a numerical toy model and a solvated alanine dipeptide.

2. Theoretical Developments

2.1. Soft Wall Restraining Potentials. Let us consider a molecular system with coordinates x and potential energy U(x). In most biomolecular applications, the dimensionality of the system is very high, but most of the variables can be unimportant for the description of a reactive mechanism. For this reason, it is customary to introduce a smaller number of collective variables to characterize the process. Let us indicate the set of collective variables, functions of x, by $\{z(x)\}$. Suppose we are given a set of K points in z-space, $(z_1, z_2, ..., z_K)$, that we call centroids. These support a complete and unique Voronoi tessellation of the original Cartesian space of the system, where each cell α is defined such that $||z(x) - z_{\alpha}|| < ||z(x) - z_{\gamma}||$ for all $\gamma \neq \alpha$; $||\cdot||$ indicates the norm in some metric, which shall be assumed Euclidean for the sake of simplicity.

It was shown previously, via a reformulation of the milestoning algorithm,⁸ that the statistical properties of the longtime dynamics of the system can be reconstructed from independent simulations, each confined inside one of the K cells. The key feature required for this result is that the confined dynamics must be equivalent to that from a long unbiased trajectory passing through the same set of cells.^{4,9} More specifically, the confinement must leave unperturbed the dynamical properties of the systems when it is in the interior of the cell as well as the probability flux in and out of the cells. In ref 9, the confinement was realized with a strict reflection rule,⁴ i.e., by reversing the velocity of the atoms when a trajectory attempted to cross the hyperplane between two neighboring cells α and γ . An alternative strategy, explored here, is to introduce planar half-pseudoharmonic restraining potentials to confine the system within each cell:

$$u_{\alpha}(\mathbf{x}) = \sum_{\gamma=1}^{K} \frac{k_{w}}{2} (\hat{\mathbf{n}}_{\alpha\gamma} \cdot [z(\mathbf{x}) - \bar{z}_{\alpha\gamma}])^{2} H(\hat{\mathbf{n}}_{\alpha\gamma} \cdot [z(\mathbf{x}) - \bar{z}_{\alpha\gamma}])$$

where the unit vector $\hat{\mathbf{n}}_{\alpha\gamma} = (z_\gamma - z_\alpha)/\|z_\gamma - z_\alpha\|$ is normal to the hyperplane separating the cells α and γ , $\bar{z}_{\alpha\gamma} = (z_\gamma + z_\alpha)/2$ is the midpoint between the centroids α and γ , and H is the Heaviside step function. The real parameter k_w determines the strength of the restraining potential. Hereafter, we will refer to the present approach as the *soft walls* (SW) restraints, to emphasize the distinction with the strict *hard walls* (HW) reflections used in refs 9 and 4.

An illustration of $u_{\alpha}(x)$ is given in Figure 1. The restraining potential $u_{\alpha}(x)$ is 0 in the interior of the cell α , and it acts as a penalty function toward the cell boundary when the trajectory crosses any of the hyperplanes separating α from other cells.

It can be understood from the following argument that the fundamental requirements of the confined dynamics can be met when the restraints of eq 1 are used. First, since $u_{\alpha}(x) = 0$ inside the cell, using eq 1 guarantees that the equilibrium distribution inside the cell is the same as that of an unbiased simulation, apart for small errors at the boundaries related to time discretization. Moreover, this implies that we also have the correct fluxes in and out of the cell (otherwise, their effect would

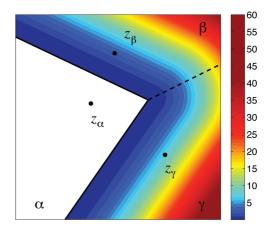


Figure 1. An illustration of the potential $u_{\alpha}(\mathbf{x})$ used to constraint a trajectory in cell α , eq 1. Portions of three cells α , β , and γ with centroids $(\mathbf{z}_{\alpha}, \mathbf{z}_{\beta}, \mathbf{z}_{\gamma})$ are shown. The potential is 0 in cell α (white region), and its units are arbitrary. Cell edges are represented as black lines. Note that the edge between cells β and γ (dashed black line) has no effect on the restraint, and it is shown only for the purpose of clarity.

propagate in and spoil the distribution inside), and hence, the pieces of trajectory of the restricted simulation are indistinguishable from the pieces of an unbiased trajectory passing through the same cell.

It is possible to compute the equilibrium probability and the rates of transitions for the original dynamics from the simulations confined within the Voronoi cells by the HW reflections. In the following sections, we show that, with minor modifications, the formalism can still be used with the SW restraints.

2.2. Free Energy of the Tessellation. The conservation of probability flux through the boundaries of the cell gives a way to compute, π_{α} , the equilibrium probability of the system to be in cell α , and the associated free energy, $G_{\alpha} = -\beta^{-1}\log \pi_{\alpha}$ (where $\beta = 1/k_{\rm B}T$, with $k_{\rm B}$ the Boltzmann constant and T the temperature), as summarized hereafter. The rate of escape from cell α to cell γ , conditional on the system being in the cell α , is defined as $v_{\alpha\gamma} = N_{\alpha\gamma}/T_{\alpha}$, where $N_{\alpha\gamma}$ is the number of collisions with the boundary separating the cells α and γ , and T_{α} is the total simulation time spent inside cell α . The normalized equilibrium probability π_{α} solves the following equations involving the rates of escape $v_{\alpha\gamma}$:

$$\sum_{\substack{\gamma=1\\\gamma\neq\alpha}}^{K} \pi_{\gamma} \nu_{\gamma\alpha} = \sum_{\substack{\gamma=1\\\gamma\neq\alpha}}^{K} \pi_{\alpha} \nu_{\alpha\gamma}, \qquad \sum_{\alpha=1}^{K} \pi_{\alpha} = 1 \qquad (2)$$

This equation expresses that, at statistical steady state, the total probability flux in cell α must be equal to the flux out of α by conservation of the total probability. In ref 9, T_{α} was simply the total simulation time with the system confined in cell α by means of HW. With SW, eq 2 is still valid, as long as one counts only the portion of trajectory spent inside the cell α as the time T_{α} .

2.3. Milestoning. Identifying the edges of the Voronoi tessellation as milestones, the dynamics of the system is reduced to that of a discrete state continuous-time Markov chain in the state space of the milestones indices. By indexing the milestones as i and j, this amounts to defining a rate matrix q_{ij} , whose elements are given by

$$q_{ij} = N_{ij}/R_i \tag{3}$$

The factors N_{ij} and R_i in this equation can be expressed⁹ in terms of the average properties extracted from simulations confined to the cell α , weighted by the equilibrium probability π_{α} of finding the system in cell α :

$$N_{ij} = \sum_{\alpha=1}^{N} \pi_{\alpha} (N_{ij}^{\alpha}/T_{\alpha}), \qquad R_{i} = \sum_{\alpha=1}^{N} \pi_{\alpha} (R_{i}^{\alpha}/T_{\alpha}) \quad (4)$$

Here T_{α} is the duration of the simulation confined in cell α , N_i^{α} is the number of transitions from edges i to j observed during this simulation, and R_i^{α} is the total time that edge i was the last edge to be hit during this simulation (i.e., $\sum_i R_i^{\alpha} = T_{\alpha}$). The only requirement to use eqs 4 with SW is to prune the trajectory to its portion that is strictly inside the cell α before computing N_{ii}^{α} , R_i^{α} , and T_{α} (i.e., the parts spent outside the cell in the SW region must be discarded).

The rate matrix q_{ij} specifies completely the dynamics of the Markov process, and hence, it can be used to compute many important quantities as, for example, the mean first passage times (MFPTs) from any milestone to any other. 9,14 For instance, if τ_i^N with i = 1, ..., N - 1 denote the MFPTs from milestone i to milestone N ($\tau_N^N = 0$ by definition), then these MFPTs can be computed by solving the linear system of equations:

$$\sum_{i=1}^{N-1} q_{ij} \tau_j^N = -1, \qquad i = 1, ..., N-1$$
 (5)

3. Results and Discussion

In the following, we illustrate the implementation of milestoning with SW restraints on two simple systems, the Mueller potential¹⁹ and the solvated alanine dipeptide. Prior to this, an important point concerning the position of the centroids supporting the Voronoi tessellation deserves special attention. While egs 3-5 can be applied to any set of hypersurfaces, it has been shown that the formalism gives exact MFPTs, if the hypersurfaces used as milestones are chosen as the isocommittor surfaces of the reaction. 14 From a practical viewpoint, the isocommittor surfaces can be calculated approximately by the string method and its variants. $^{3-5,20-22}$ In the string method, the transition path is represented as by an ordered sequence of K discrete "images", $\{z^{(1)}, z^{(2)}, ..., z^{(K)}\}\$, in the space of collective variables z(x). This suggests⁹ to first use the string method to determine the transition path by optimizing the position of the K images and then use these images as centroids to support the Voronoi tessellation. Naturally, the edges between the cells are hyperplanes perpendicular to the optimized path and, hence, approximations of the isocommittor surfaces.

3.1. The Mueller Potential. We first consider a simple 2-dimensional system evolving on the Mueller potential energy surface according to Langevin dynamics (i.e., we take $\{z(x,y)\}$ \equiv (x,y), the coordinates of the system). The same model was studied in ref, so for comparison, we took the same centroids as generators of the Voronoi tessellation. These are 18 equidistant images (the red circles in Figure 2), computed with the finite-temperature string method.⁴ All simulations were performed by integrating Langevin equations of motion with the second-order algorithm of ref 23 by taking $\beta^{-1} = 20$, friction

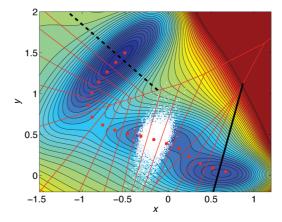


Figure 2. Mueller potential with a set of milestones, corresponding to the edges of Voronoi cells, generated by images along the converged string from the finite-temperature string method. The successive positions of the system at every time step along a piece of simulation with SW are also shown as white dots. MFPTs, discussed in the text, are calculated from the dashed to the continuous black line.

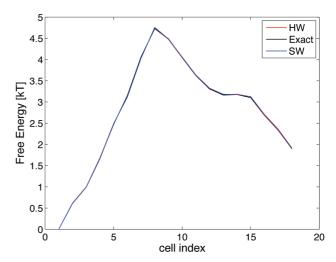


Figure 3. Free energy $G_{\alpha} = -\beta^{-1}\log \pi_{\alpha}$ of the Voronoi tessellation from HW and SW simulations on the Mueller potential compared with the exact one obtained by numerical integration. Numbering of the cells goes from top-left to bottom-right in Figure 2.

400, and a time step $\Delta t = 10^{-4}$. Hard and soft wall trajectories of 108 steps were generated in each cell.

Figure 2 shows the Mueller potential together with the Voronoi tessellation associated to the images from the string. The edges of the cells were taken as milestones. The successive positions of the system at every time step along a piece of simulation with SW are also shown as white dots. Note that, as already pointed out before, the portion of trajectory inside the cell samples the equilibrium distribution associated to the underlying potential in the cell.

Figure 3 shows the free energy $G_{\alpha} = -\beta^{-1} \log \pi_{\alpha}$, where π_{α} is the solution of eq 2, corresponding to the tessellation shown in Figure 2. Results are presented for SW and HW simulations and numerical integration (Exact). The numbering of the cells goes from 1 to 18 from top-left to bottom-right in Figure 2. Table 1 summarizes results for MFPTs calculations considering the transition from the black dashed line to the black continuous line in Figure 2. The table also shows the mean and the extremal

Table 1. Kinetics for the Mueller Potential for the Transition from the Black Dashed Milestone to the Black Continuous Milestone in Figure 2^a

	MFPT	$\langle R_i \rangle$	<i>R_i</i> max	<i>R_i</i> min	$\langle N_{ij} angle$	<i>N_{ij}</i> max	<i>N_{ij}</i> min
soft walls	547 ± 24	0.0065	0.4644	1.55×10^{-8}	1.35×10^{-6}	3.32×10^{-5}	6.05×10^{-11}
hard walls	553 ± 14	0.0065	0.4645	9.99×10^{-9}	1.37×10^{-6}	3.27×10^{-5}	4.10×10^{-11}
free simulation	562 ± 16	0.0065	0.4645	1.89×10^{-8}	1.37×10^{-6}	3.19×10^{-5}	4.02×10^{-11}

^a The agreement of all quantities shows that the dynamical properties of the trajectory inside the cells are left unperturbed by the two different confinement methods, i.e., it is as if the confinement was absent. Errors on MFPTs were obtained from five different runs.

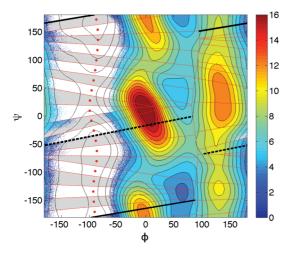


Figure 4. Backbone dihedral free energy surface of solvated alanine dipeptide (units are kcal/mol) with a set of milestones (red lines) corresponding to the edges of Voronoi cells generated by images along the converged minimum free energy path (red circles) and the helical to extended transition. The successive values of dihedral angles at every time step along pieces of simulations with HW are shown as white and gray dots. MFPTs, discussed in the text, are calculated from the dashed to the continuous black line.

values of the elements needed for the calculation of the q_{ij} matrix defined in eq 3, computed from SW and HW simulations and a long free trajectory (2 × 10⁹ steps). The excellent agreement shows that the confinements do not introduce biases in the calculations of q_{ij} .

3.2. Solvated Alanine Dipeptide. In order to test the implementation of SW and HW in a realistic biomolecular context, we examine the transition from helical to extended conformation of the alanine dipeptide solvated in explicit water. We describe the transition here by using two collective variables, the ϕ and ψ backbone dihedral angles, neglecting the role of the solvent degrees of freedom.²⁴ We used the CHARMM¹⁶ code for all calculations with CHARMM22 all-atom force field²⁵ and the TIP3P model²⁶ for water molecules.

The free energy landscape in ϕ and ψ variables was computed with the single-sweep method, ²⁷ and it is shown for reference in Figure 4 (energies are in kcal/mol). A minimum free energy path in (ϕ,ψ) space was computed using the string method. ³ 20 images along this path are represented as circles in Figure 4, and they were used as centroids for the Voronoi tessellation. Note that in our calculation, we focus on the reaction channel with $\phi < 0$, where the main metastable states are. In principle, we could also investigate the kinetics of transitions between the states with $\phi > 0$, but these are less often visited, and hence, their existence does not affect much the rates we compute. The edges of the Voronoi cells are represented as red lines in Figure

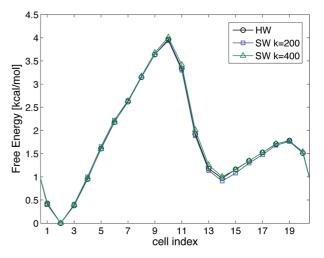


Figure 5. Free energy of the Voronoi tessellation from HW and SW sampling for the solvated alanine dipeptide. Numbering of the cells goes from 1 to 20, counting from top to bottom in Figure 4.

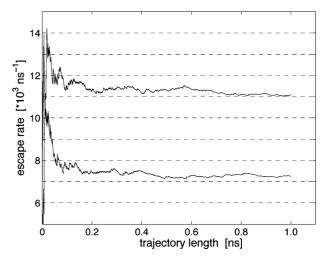


Figure 6. Rates of escape from cell #2 (i.e., associated to the second string image counting from top to bottom in Figure 4), ν_{21} , top line, and ν_{23} , bottom line, as a function of the trajectory length for the alanine dipeptide SW simulation with $k_{\rm w} = 200~{\rm kcal/mol/rads^2}$.

4 (taking into account periodicity). The dashed and continuous black lines are the start and the ending milestones for MFPTs calculation. The HW confinement condition was implemented into the Nosé–Hoover (NH) dynamics subroutine of CHARMM.²⁸ The successive values of dihedral angles at every time step along pieces of simulations with ahrd walls are shown as white and gray dots. NH dynamics was also used for SW simulations. In this case, the forces on atoms coming from the potential (1) were implemented and added to the standard CHARMM forces.

Table 2. Kinetics for the Solvated Alanine Dipeptide for the Transition from the Black Dashed Milestone to the Black Continuous Milestone in Figure 4

	MFPT (ps)	$\langle R_i \rangle$	R_i max	<i>R_i</i> min	$\langle N_{ij} angle$	<i>N_{ij}</i> max	N _{ij} min
soft walls hard walls	28.33 28.13	0.0053 0.0053	0.3242 0.3306	$6.65 \times 10^{-7} \\ 5.00 \times 10^{-7}$	$\begin{array}{c} 9.24 \times 10^{-5} \\ 8.86 \times 10^{-5} \end{array}$	$6.87 \times 10^{-4} \\ 7.02 \times 10^{-4}$	$2.99 \times 10^{-9} \\ 1.97 \times 10^{-8}$

Figure 5 shows the free energy corresponding to the tessellation shown in Figure 4. Results are for HW and SW simulations (with two different values of the penalty constant $k_{\rm w}$). In all cases, the trajectories in each cell were 1 ns long. Numbering of the cells goes from 1 to 20 and top to bottom in Figure 4. In order to assess convergence of the calculation, we monitored, during the simulations, the values of the escape rates from the cells, $\nu_{\alpha\gamma}$. Figure 6 shows the rates from cell two versus the length of the entire confined trajectory for SW simulations with $k_{\rm w} = 200$ kcal/mol/rads². Escape rates from other cells converge on the same time scale.

MFPTs were computed considering the transition from the black dashed to the black continuous line in Figure 4. Results are summarized in Table 2, together with the mean and the extremal values of the elements needed for the calculation of the q_{ij} matrix defined in eq 3. The excellent agreement shows again that the confinements do not introduce biases in the calculations of q_{ii} .

The MFPT for the transition from dashed to continuous milestone in Figure 4 estimated from milestoning with HW and SW simulations is about 28 ps. For comparison, the MFPT between the same two milestones was computed by direct counting of consecutive hittings, along a free, unbiased simulation of 80 ns, obtaining 29.97 ps. These values agree with results from other CHARMM calculations of solvated alanine dipeptide in a similar simulation setup, where the MFPT for the same transition was estimated to be of about 30 ps.^{29,30}

4. Concluding Remarks

Milestoning with Voronoi tessellation⁹ is a method to reconstruct the dynamical properties of complex reactive systems by matching together informations obtained from multiple trajectories, each confined in a different cell of a Voronoi tessellation of configurational space. In this letter, we have illustrated how the formalism can still be applied, with minor modifications, when the confinement in the cells is realized via continuous and differentiable potentials. With respect to the original formulation, where the confinement is realized with strict reflections (velocity inversion at the boundary), the possibility to use potentials will facilitate the application of the method because user-defined external forces are easy to introduce in most MD codes and do not require modification of the dynamical propagators. However, the confinement with potentials can also affect performance because portions of trajectories that are transiently out of the cells are discarded in the analysis. Note also that we did not optimize for efficiency here, which would require adjusting the penalty constant $k_{\rm w}$ in eq 1.

As a final remark, let us point out that the method proposed in this letter, like the one in ref 9, can be trivially distributed on multiple processing nodes, since it is based on independent simulations with no required communication among them.

Acknowledgment. We thank Ron Elber, Maddalena Venturoli, and Albert Pan for many useful discussions. The work of E.V.-E. was partially supported by the National Science Foundation grants DMS-0718172 and DMS-0708140 and the Office of Naval Research grant N00014-04-1-6046. The work of B.R. was supported by National Science Foundation through Grant MCB-0920261.

References

- (1) Elber, R. Curr. Opin. Struct. Biol. 2005, 15, 151.
- (2) Jonsson, H.; Mills, G.; Jacobsen, K. W. Classical and Quantum Dynamics in Condensed Phase Simulations, World Scientific: Singapore, 1998; p 385.
- (3) Maragliano, L.; Fischer, A.; Vanden-Eijnden, E.; Ciccotti, G. J. Chem. Phys. 2006, 125, 024106.
- (4) Vanden-Eijnden, E.; Venturoli, M. J. Chem. Phys. 2009, 130, 194103.
- (5) Pan, A. C.; Sezer, D.; Roux, B. J. Phys. Chem. B 2008, 112, 3432.
- (6) Pan, A. C.; Roux, B. J. Chem. Phys. 2008, 129, 064107.
- (7) van Erp, T. S.; Bolhuis, P. G. J. Comput. Phys. 2005, 205, 157.
- (8) Faradjian, A. K.; Elber, R. J. Chem. Phys. 2004, 120, 10880.
- (9) Vanden-Eijnden, E.; Venturoli, M. J. Chem. Phys. 2009, 130, 194101.
- (10) Shalloway, D.; Faradjian, A. K. J. Chem. Phys. 2006, 124, 054112.
- (11) West, A. M. A.; Elber, R.; Shalloway, D. J. Chem. Phys. 2007, 126, 145104.
- (12) Elber, R. Biophys. J. 2007, 92, L85.
- (13) Kuczera, K.; Jas, G. S.; Elber, R. J. Phys. Chem. A 2009, 113, 7461.
- (14) Vanden-Eijnden, E.; Venturoli, M.; Ciccotti, G.; Elber, R. J. Chem. Phys. 2008, 129, 174102.
- (15) Case, D. A.; Cheatham, T. E.; Darden, T.; Gohlke, H.; Luo, R.; Merz, K. M.; Onufriev, A.; Simmerling, C.; Wang, B.; Woods, R. J. J. Comput. Chem. 2005, 26, 1668.
- (16) Brooks, B. R.; et al. J. Comput. Chem. 2009, 30, 1545.
- (17) Van Der Spoel, D.; Lindahl, E.; Hess, B.; Groenhof, G.; Mark, A. E.; Berendsen, H. J. J. Comput. Chem. 2005, 26, 1701.
- (18) Phillips, J. C.; Braun, R.; Wang, W.; Gumbart, J.; Tajkhorshid, E.; Villa, E.; Chipot, C.; Skeel, R. D.; Kalé, L.; Schulten, K. J. Comput. Chem. 2005, 26, 1781.
- (19) Mueller, K. Angew. Chem., Int. Ed. Engl. 1980, 19, 1.
- (20) Vanden-Eijnden, E. Computer Simulations in Condensed Matter Systems: From Materials to Chemical Biology, Springer-Verlag, Heidelberg, Volume 1, 2007.
- (21) E, W.; Ren, W.; Vanden-Eijnden, E. J. Chem. Phys. 2007, 126, 164103.
- (22) E, W.; Ren, W.; Vanden-Eijnden, E. J. Phys. Chem. B 2005, 109, 6688.

- (23) Vanden-Eijnden, E.; Ciccotti, G. Chem. Phys. Lett. 2006, 429, 310.
- (24) Ma, A.; Dinner, A. R. J. Phys. Chem. B 2005, 109, 6769.
- (25) Mackerell, A. D.; et al. J. Phys. Chem. B 1998, 102, 3586.
- (26) Jorgensen, W. L.; Chandrasekhar, J.; Madura, J. D.; Impey, R. W.; Klein, M. L. J. Chem. Phys. 1983, 79, 926.
- (27) Maragliano, L.; Vanden-Eijnden, E. J. Chem. Phys. 2008, 128, 184110.
- (28) Lamoureux, G.; Roux, B. J. Chem. Phys. 2003, 119, 3025.
- (29) Strodel, B.; Wales, D. J. Chem. Phys. Lett. 2008, 466, 105.
- (30) Zhang, Y.; Pastor, R. W. Mol. Simul. 1994, 13, 25.
 CT900279Z