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Technical Notes

UV Resonance Raman Spectroscopic Detection of Nitrate and Nitrite in Wastewater Treatment Processes

Anatoli Ianoul, † Thomas Coleman, † and Sanford A. Asher*, †

Department of Chemistry, University of Pittsburgh, Pittsburgh, Pennsylvania 15260, and dTEC Systems L.L.C., 3012 16th Avenue West, Seattle, Washington 98119

The 204- and 229-nm excited UV resonance Raman spectra of wastewater solutions containing sodium nitrite and nitrate were measured in the concentration range 7 μM to 3.5 mM (0.1–50 ppm nitrogen). The other chemical species present in wastewater do not interfere with Raman measurements of NO $_2^-/NO_3^-$ bands. We observe detection limits of $<14~\mu M$ (<200 ppb) for both NO $_2^-$ and NO $_3^-$. UV resonance Raman spectroscopy appears to be an excellent tool for on-line monitoring of NO $_2^-/NO_3^-$ in wastewater for the real-time control of water treatment plants.

The reliability and performance of biological nutrient removal (BNR) wastewater treatment plants (WWTPs) can be enhanced by the ability to monitor nitrate and nitrite in BNR activated sludge reactors without the need for reagent additions or complex calibration procedures. Real-time on-line monitoring of these parameters provides input to the process control systems to optimize performance of treatment systems designed for low nitrogen and phosphorus effluent concentrations. According to the Water Environment Research Foundation (WERF), poor sensor performance and high maintenance costs are the major stumbling blocks to improved automation.

In the work here, we demonstrate the utility of UV resonance Raman³⁻⁸ spectroscopy to monitor nitrate and nitrite in wastewater treatment process reactors in real time. Excitation within the strong UV absorption bands of nitrate and nitrite results in the

selective enhancement of the vibrations of these species. We observe little interference from other species in the wastewater. Further, UV excitation avoids fluorescence interference⁵ from other species present in the wastewater, thus yielding high signal-to-noise spectra for determining the nitrate and nitrite concentrations

MATERIALS AND EXPERIMENTAL EQUIPMENT

Chemical Reagents. Nitrate and nitrite solutions were prepared from 0.07 and 0.1 M reagent grade stock solutions in reagent grade type II water, respectively. Both standard stock solutions were traceable to NIST primary standard sources.

Wastewater Solutions. Samples representative of the soluble components of an activated sludge process reactor were obtained from the Grand Coulee, WA municipal wastewater treatment plant (design capacity 0.30 MGD), which uses a cyclic aerobic/anoxic process operating mode. A sample of mixed liquor was drawn from the aeration basin just at the end of an anoxic period when all of the nitrate and nitrite had been depleted. The sample was immediately filtered through a 0.45- μ m membrane filter to obtain a solution containing only the soluble components of the mixed liquor. Sodium nitrite and nitrate were added to this wastewater sample.

Additional wastewater sample solutions were obtained in a similar manner from the Grand Coulee, WA, WWTP, and from the Bingen, WA, WWTP (design capacity 0.80 MGD) which utilizes a cyclic aerobic/anoxic process operating mode very similar to Grand Coulee. Both of these samples were fully depleted of nitrate and nitrite at the time they were collected and filtered.

The third sample was obtained from the Zillah, WA, WWTP (design capacity 0.31 MGD).

When the sample was collected, the Zillah plant was experiencing a mild process upset with a more turbid effluent than normal. The upset conditions began two weeks previously. Because of the timing of the sample collection, it was not possible to collect a sample that was fully depleted of nitrate and nitrite. In lieu of reaching the depletion end point, the filtered sample was analyzed for nitrate plus nitrite (often referred to as NO_X) using the Hach

^{*} Corresponding author: (phone) 412-624-8570; (fax) 412-624-0588; (e-mail) asher@pitt.edu.

[†] University of Pittsburgh.

[‡] dTEC Systems L.L.C.

⁽¹⁾ Lant, P.; Steffens, M. Water Sci. Technol. 1998, 37, 49-54.

⁽²⁾ Water Environment Research Foundation. RFP No. 99-WWF-4, October 1998.

⁽³⁾ Asher S. A.; Munro C. H.; Chi Z. H. Laser Focus World 1997, 33, 99–109.

⁽⁴⁾ Asher, S. A.; Bormett, R. In Raman scattering in materials science; Weber, W. H., Merlin R., Eds.; Springer-Verlag: Berlin, 2000; pp 35–54.

⁽⁵⁾ Asher, S. A.; Johnson, C. R. Science 1984, 225, 311-313.

⁽⁶⁾ Asher, S. A. In Handbook of vibrational spectroscopy, Chalmers, J. M., Griffiths, P. R., Eds.; Wiley & Sons: New York, 2001; pp 1–15.

⁽⁷⁾ Asher, S. A. Anal. Chem. 1993, 65, A59-A66, A201-A210.

⁽⁸⁾ Dudik, J. M.; Johnson C. R.; Asher S. A. J. Chem. Phys. 1985, 82, 1732–1740.

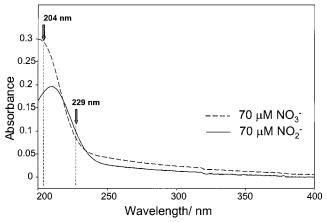


Figure 1. UV absorption spectra of sodium nitrite and nitrate in pure water. Path length, 0.5 cm.

cadmium reduction method. This method usually provides an acceptable approximation for nitrate since nitrite concentrations are typically much lower than nitrate concentrations. The result of this analysis was 0.3 mM NO_X .

UV Resonance Raman Spectroscopy. The UV resonance Raman spectral measurements used either 204-nm excitation obtained by anti-Stokes Raman shifting the third harmonic of a Coherent Inc. Infinity Nd:YAG laser 10 in $\rm H_2$ or 229-nm excitation from a Coherent Inc. CW intracavity frequency-doubled argon ion laser. 11 The Raman scattered light was collected in a $\sim \! 135^{\circ}$ backscattering geometry and dispersed by a Spex double or single monochromator. An intensified CCD detector (Princeton Instrument Co.) was used for detection. Winspec software was used to accumulate the spectra, and Grams software was used for the spectra treatment and analysis.

Typically, an accumulation time of 10 min was used to collect the spectra. The samples were measured in a free-surface flowing stream. The 1379-cm⁻¹ Teflon band was used as an external Raman intensity standard. Raman spectrum of Teflon was recorded before or immediately after acquisition of the sample spectrum. The 1640-cm⁻¹ water band was used as the internal Raman intensity standard.

Absorption spectra were obtained with a Perkin-Elmer Lambda 9 spectrophotometer in a 0.5-cm cuvette.

RESULTS AND DISCUSSION

Absorption Spectra. Figure 1 shows the absorption spectra of nitrate and nitrite. The absorption maximum of NO_3^- is slightly blue-shifted (maximum near 200 nm) compared to that of NO_2^- (maximum near 210 nm). In addition, the maximum NO_3^- molar absorptivity was $8.4 \times 10^3 \, L/(M \cdot cm)$, which is $\sim \! 50\%$ higher than that of NO_2^- ($5.6 \times 10^3 \, L/(M \cdot cm)$).

UV Resonance Raman Spectra. Figure 2 shows the UV resonance Raman spectra of nitrate and nitrite in pure water.

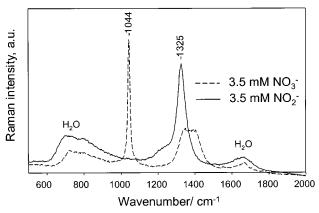


Figure 2. UV resonance Raman spectra of 3.5 mM solutions of $NaNO_2$ and $NaNO_3$. Excitation wavelength, 204 nm; power, 10 mW; accumulation time, 10 min. The spectra were measured in a free-surface stream.

Symmetric N–O stretching vibrations give rise to the strong bands at 1044 cm $^{-1}$ for NO $_3$ $^-$ and at 1325 cm $^{-1}$ for NO $_2$ $^-$. Some medium-intensity bands in the spectrum of NO $_3$ $^-$ are also observed between 1300 and 1500 cm $^{-1}$.

Obviously, nitrate and nitrite are spectrally distinct and can be easily distinguished. There is a partial overlap of the 1325-cm $^{-1}$ NO $_2^-$ band with the weaker NO $_3^-$ band at $\sim\!1400$ cm $^{-1}.$ However, we can easily numerically remove the contributions from nitrate in that spectral interval to obtain the pure nitrite Raman band.

Nitrate and Nitrite Detection Limits. To determine the UV resonance Raman spectral detection limits, we examined the dependence of the UV resonance Raman spectra as a function of nitrate/nitrite concentration in pure water (Figure 3). We observe a linear intensity dependence of the absolute Raman intensities over the analyte concentration range of 7 μ M to 3.5 mM. This indicates that, for 229-nm excitation, self-absorption has little impact on the measured intensities since excitation is on the red edge of the absorption band.

To determine the detection limit for measurements of the nitrate/nitrite concentrations using UV resonance Raman spectroscopy, we carefully analyzed spectra at the lowest concentrations measured: 7, 14, and 35 μM (Figure 3, insets). Raman bands from nitrate and nitrite were clearly observed for analyte concentrations as low as 14 μM . At 7 μM , the Raman signal is almost undetectable because of the noise. Therefore, the detection limits for nitrate and nitrite under these conditions is <14 μM (<200 ppb).

Filtered Activated Sludge Wastewater Samples. Figure 4 shows the $\mathrm{NO_2}^-$ and $\mathrm{NO_3}^-$ concentration dependence of the UV resonance Raman spectra in filtered activated sludge wastewater samples. The spectra of the filtered activated sludge wastewater are essentially identical to those of pure water. As in pure water, the Raman intensity decreases as the concentration of analyte decreases. We observe identical Raman detection limits of <14 $\mu\mathrm{M}$ (<200 ppb).

Choice of Excitation Wavelength. The 204-nm excited UV Raman spectra of NaNO₂ and NaNO₃ are essentially identical to those excited at 229 nm (data not shown). The absolute intensities of the 1325- and 1044-cm $^{-1}$ bands increase with concentration in the range 7 μM to 0.35 mM. However, the absolute intensities do

⁽⁹⁾ Wastewater and Biosolids Analysis Manual, Hach Co.: Loveland, CO, 1999; pp 211–225.

⁽¹⁰⁾ Lednev, I. K.; Karnoup, A. S.; Sparrow, M. C.; Asher, S. A. J. Am. Chem. Soc. 1999, 121, 8074–8086.

⁽¹¹⁾ Asher, S. A.; Bormett, R. W.; Chen, X. G.; Lemmon, D. H.; Cho, N.; Peterson, P.; Arrigoni, M.; Spinelli, L.; Cannon, J. Appl. Spectrosc. 1993, 47, 628–633.

⁽¹²⁾ Larkin, P. J.; Gustafson, W. G.; Asher, S. A. J. Chem. Phys. 1991, 94, 5324– 5330.

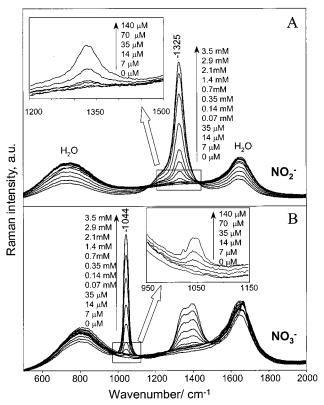


Figure 3. UV resonance Raman spectra of $NaNO_2$ (A) and $NaNO_3$ (B) in pure water as a function of concentration. Excitation wavelength, 229 nm; power, 10 mW; accumulation time, 10 min. The spectra were measured in a free-surface stream.

not linearly increase at higher concentrations due to self-absorption (Figure 5). The relative intensity ratios of the analyte to the 1640-cm $^{-1}$ internal standard water band, however, do increase linearly with concentration (Figure 5).

The data indicate that the NO_2^- relative to the 1640-cm⁻¹ water band¹³ Raman intensity does not significantly change for 204-nm (I(1325)/I(1640)=0.6) excitation compared to 229-nm excitation (I(1325)/I(1640)=0.7). In contrast, we observe an ~2.5-fold increase in the 204-nm NO_3^- relative Raman intensity (I(1044)/I(1640)=1.44) compared to 229-nm (I(1044)/I(1640)=0.72) intensity. Thus, 204-nm excitation is preferable for nitrate detection

Detection of Nitrate and Nitrite in Filtered Activated Sludge Wastewater Samples. Filtered mixed liquor samples from the extended aeration activated sludge treatment plants were measured using 229-nm excitation. A mixed liquor filtrate from the Bingen, WA, activated sludge treatment plant showed Raman spectra similar to those from Grand Coulee (Figure 4), which showed no detectable nitrate or nitrite bands, as well as no bands other than those of water.

The sample from the Zillah, WA, WWTP, which was experiencing a mild process upset, was somewhat turbid and showed the Figure 6A absorption spectrum. The absorption below 240 nm is much higher for the Zillah sample than for the other filtered wastewater samples and the pure water 0.07 mM sodium nitrate and nitrite solutions. This results from the fact that the Zillah sample contains 0.3 mM NO_X .

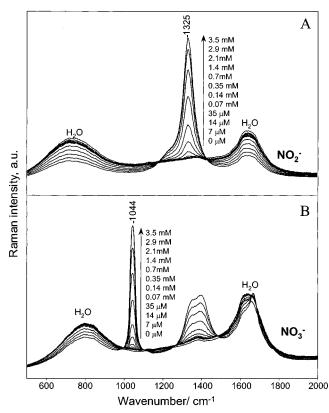


Figure 4. UV resonance Raman spectra of NaNO₂ (A) and NaNO₃ (B) in filtered activated sludge wastewater as a function of concentration. Excitation wavelength, 229 nm; power, 10 mW; accumulation time, 10 min. The spectra were measured in a free-surface stream.

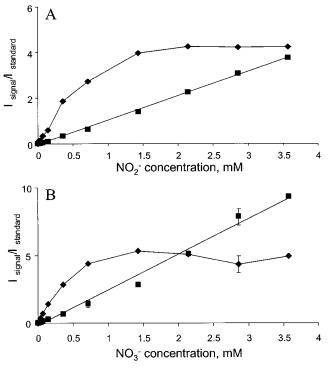


Figure 5. Relative intensities of the 204-nm excited NaNO $_2$ 1325-cm $^{-1}$ (A) and NaNO $_3$ 1044-cm $^{-1}$ (B) Raman bands as a function of concentration. Diamonds, 1379-cm $^{-1}$ Teflon band is used as external standard; squares, 1640-cm $^{-1}$ water band is used as internal standard.

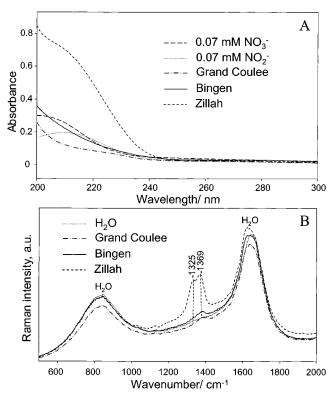


Figure 6. (A) UV absorption spectra of nitrite and nitrate in pure water and of filtered activated sludge wastewater samples. Path length, 0.5 cm. (B) UV resonance Raman spectra of pure water and three filtered activated sludge wastewater samples. Zillah WWTP sample was measured as containing 0.3 mM NO $_X$. Excitation wavelength, 229 nm; power, 10 mW; accumulation time, 10 min. The spectra were measured in a free-surface stream.

Figure 6B shows the Raman spectra at 229-nm excitation for the three filtered activated sludge wastewater samples and pure water. The Raman spectra of the Grand Coulee and Bingen samples are very close to that of pure water. The only additional spectral feature is a weak band at 1369 cm $^{-1}$. Thus, it is easy to analyze for species such as $\rm NO_3^-$ and $\rm NO_2^-$ in wastewater.

The Zillah plant sample, however, shows a $1325\text{-cm}^{-1}\ NO_2^-$ band in addition to another band at $1369\ \text{cm}^{-1}$. There is no

(14) Wen, N.; Brooker, M. H. J. Phys. Chem. 1995, 99, 359-368.

evidence for the NO_3^- at $1044~cm^{-1}$. Thus, all of the NO_X measured by the cadmium reduction method was in the form of nitrite. In retrospect, this result can be explained by the upset conditions at the treatment plant in which the *Nitrobacter* bacteria had been lost from the system. This result directly demonstrates the analytical power of UV Raman spectroscopy, which was able to simultaneously measure nitrate and nitrite down to very low detection levels. These data could be used to provide both process control variable input and diagnostic capabilities for wastewater plant upset conditions.

We are as yet unsure of the origin of the ${\sim}1369\text{-cm}^{-1}$ band, which is weak but evident in every wastewater sample. From its frequency and band shape, we tentatively assign it to the symmetric C–O stretch vibration of HCO_3^- from dissolved carbon dioxide. Whatever the ultimate assignment, this band is far enough from the $1325\text{-cm}^{-1}\ NO_2^-$ band that it does not interfere with NO_2^- determination.

SUMMARY AND CONCLUSIONS

We have successfully demonstrated the potential of UV resonance Raman spectroscopy for on-line monitoring and process control to improve the reliability and performance of biological nutrient removal wastewater treatment plants. Typical wastewater samples do not show interfering spectral bands or significant interfering absorption features. Routine 10-min spectral measurement gives rise to $<\!14~\mu\mathrm{M}$ (200 ppb) UV resonance Raman detection limits for nitrate and nitrite in water. Excitation wavelengths between 230 and 204 nm are suitable for nitrate and nitrite determination.

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