

Chlorinated Ethenes from Groundwater in Tree Trunks

DON A. VROBLESKY*

U.S. Geological Survey, 720 Gracern Road, Suite 129,
Stephenson Center, Columbia, South Carolina 29210-7651

CHRISTOPHER T. NIETCH AND
JAMES T. MORRIS

Department of Biological Sciences, University of South
Carolina, Columbia, South Carolina 29208

The purpose of this investigation was to determine whether tree-core analysis could be used to delineate shallow groundwater contamination by chlorinated ethenes. Analysis of tree cores from bald cypress [*Taxodium distichum* (L.) Rich], tupelo (*Nyssa aquatica* L.), sweet gum (*Liquidambar styraciflua* L.), oak (*Quercus* spp.), sycamore (*Platanus occidentalis* L.), and loblolly pine (*Pinus taeda* L.) growing over shallow groundwater contaminated with *cis*-1,2-dichloroethene (cDCE) and trichloroethene (TCE) showed that those compounds also were present in the trees. The cores were collected and analyzed by headspace gas chromatography. Bald cypress, tupelo, and loblolly pine contained the highest concentrations of TCE, with lesser amounts in nearby oak and sweet gum. The concentrations of cDCE and TCE in various trees appeared to reflect the configuration of the chlorinated-solvent groundwater contamination plume. Bald cypress cores collected along 18.6-m vertical transects of the same trunks showed that TCE concentrations decline by 30–70% with trunk height. The ability of the tested trees to take up cDCE and TCE make tree coring a potentially cost-effective and simple approach to optimizing well placement at this site.

Introduction

The use of vegetation in studies of groundwater contamination has received increasing attention for a variety of reasons. Plants can remove contaminants from the subsurface by direct uptake and degradation (1–6), transpiration of volatile contaminants to the atmosphere (6–8), binding contaminants to plant tissue (4–6), and enhancing microbial growth and bioremediation in the rhizosphere (9–13). Direct uptake of contaminants is controlled by a variety of factors, but in general, moderately hydrophobic organic compounds (octanol–water coefficient, $\log K_{ow} = 0.5–3$), such as trichloroethene (TCE) and *cis*-1,2-dichloroethene (cDCE), readily enter vegetation transpiration streams (10, 14).

This investigation shows that TCE and cDCE are present in tree trunks growing above contaminated shallow groundwater. To our knowledge, this is the first investigation demonstrating that headspace analysis of tree cores can be an inexpensive and rapid method to delineate shallow TCE and cDCE groundwater contamination. In addition, this investigation presents data showing variations in concentra-

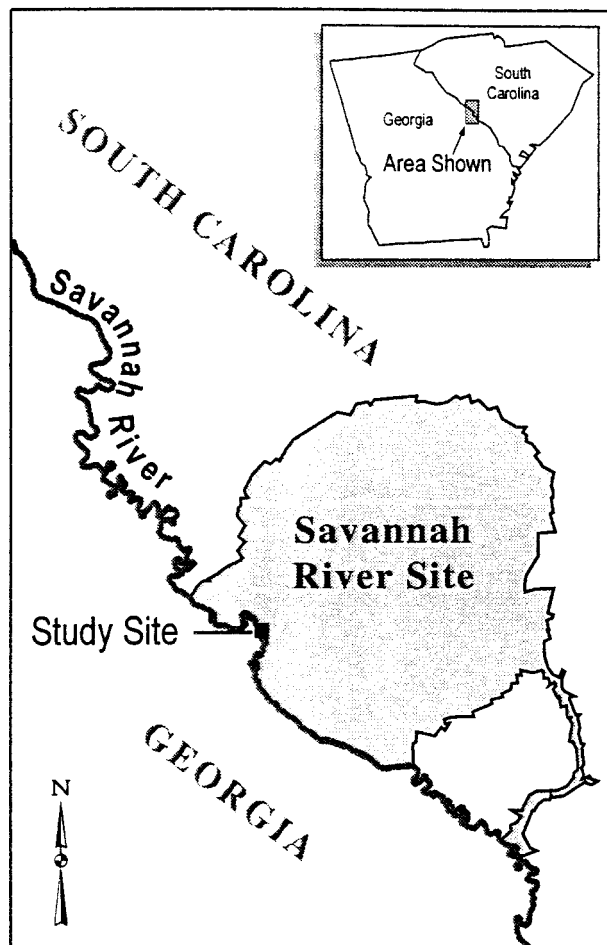


FIGURE 1. Location of study area.

tions of those compounds vertically upward along a tree trunk. Cores were collected from 97 mature trees growing above and in the vicinity of contaminated groundwater. Six different species of trees were examined. Cores from one tree were collected at various heights along the trunk.

Study Area. The study area is a forested flood plain of the Savannah River near the TNX Area, Savannah River site, SC (Figure 1). The flood plain consists of an upper and lower terrace, separated by an embankment ranging from about 1 to 2 m high. Northeast of the upper flood plain, land surface rises sharply to an upland terrace (the TNX Area) containing a former seepage basin. For a few days to a few months per year, the Savannah River inundates the lower flood plain (Figure 2). During most of the year, however, standing water is limited to low-lying areas of the upper and lower flood plains, indicated by marsh symbols in Figure 2.

Chlorinated solvents are present in the groundwater beneath parts of the upper flood plain. The dominant contaminants are TCE and cDCE. The probable source of the chlorinated solvents is leakage from a former seepage basin in the TNX Area (Figure 2). Groundwater flows through unconsolidated sands toward a drainage ditch that empties into the Savannah River. Depth to the water table beneath the flood plain ranges from the land surface to about 1.5 m. The groundwater flow rate in the flood plain is approximately 0.46 m/day (15).

Vegetation in the swamps of the flood plain primarily consists of bald cypress [*Taxodium distichum* (L.) Rich],

* Corresponding author e-mail: vroblesk@usgs.gov; tel: (803)-750-6115; fax: (803)750-6181.

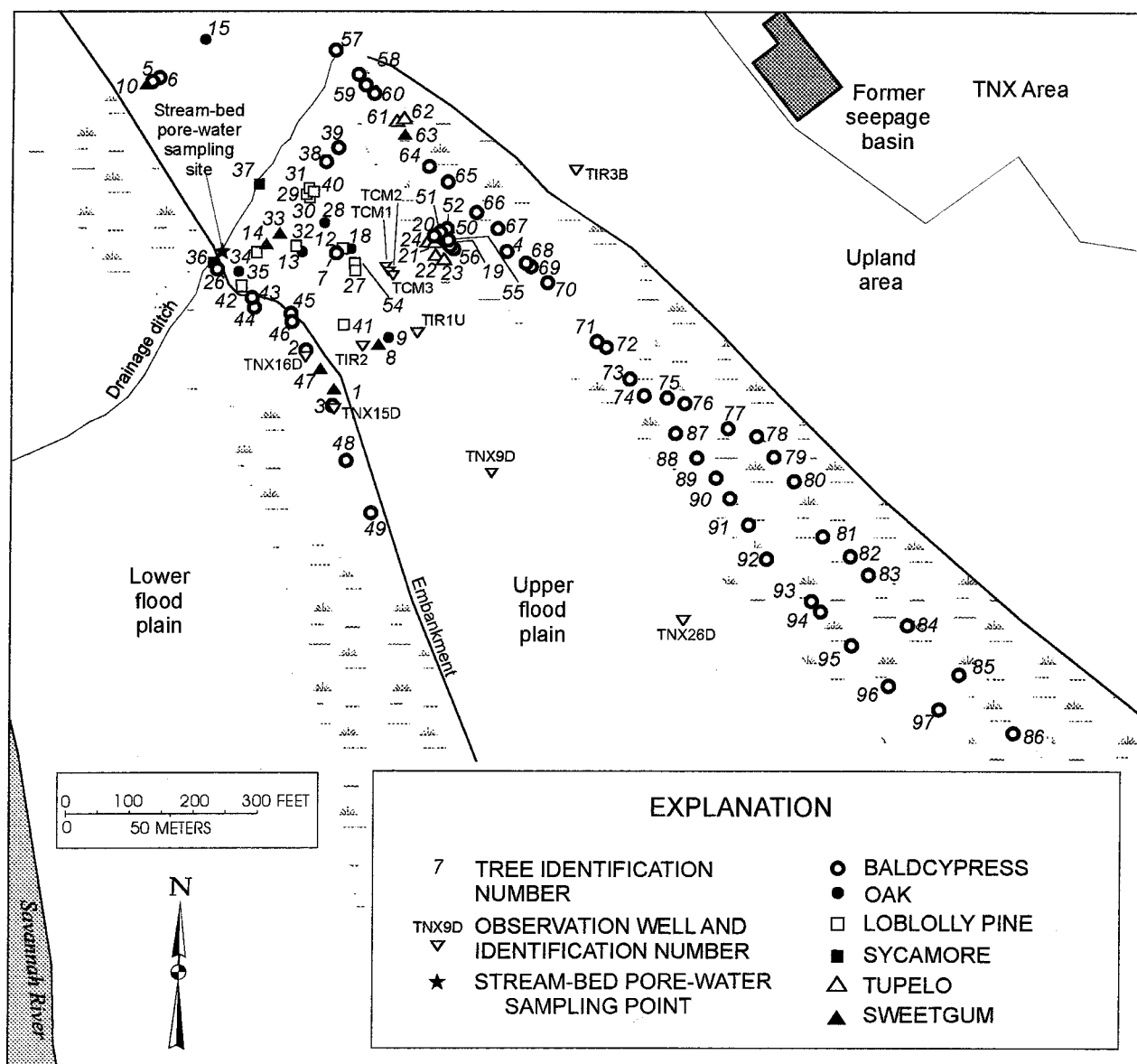


FIGURE 2. Location and identification numbers of test trees and observation wells on the Savannah River flood plain, TNX Area, Savannah River site, SC.

tupelo (*Nyssa aquatica* L.), and sweet gum (*Liquidambar styraciflua* L.). In drier parts of the flood plain, oak (*Quercus* spp.), loblolly pine (*Pinus taeda* L.), and sweet gum are dominant with a relatively small population of bald cypress and sycamore (*Platanus occidentalis* L.).

Methods

Cores were collected with an increment borer from 97 trees on the Savannah River flood plain in South Carolina during January, February, July, August, September, and October 1997 and January and February 1998 (Table 1). The trees included 64 bald cypress, 5 tupelos, 12 loblolly pines, 3 sycamores, 6 oaks, and 7 sweet gums (Figure 2). Part of the area contained groundwater contaminated with TCE and cDCE. Trees 11, 16, 17, and 53 were control oak, loblolly pine, sweet gum, and sycamore trees, respectively, collected from offsite areas not shown in Figure 2.

The cores used to delineate contamination in Figures 3 and 4 were collected from the northeastern side of the respective tree. All cores used to examine areal distribution of contamination were collected from a height of approximately 1.5 m above ground, except for tree 7a (which

was collected from 9 m on August 11, 1997, and from 3 m on October 1, 1997) and trees 20z and 21z (which were collected from about 1.8 m).

At selected trees, two cores were collected approximately 25 mm horizontally from each other to compare replication. The average concentration difference between replicate cores was 15.2% for TCE in six replicate pairs and 2.5% for cDCE in three replicate pairs. A bald cypress (tree 7) also was cored at various heights along its trunk. A bucket truck was used to access the tree, which grew adjacent to an unpaved road.

Upon collection, the cores (approximately 68 mm in length) immediately were removed from the coring tools and placed in 20-mL glass vials. At selected sites, ambient air samples also were collected by waving an empty 20-mL glass vial in the air for several seconds. Teflon-coated septum caps then were crimped onto the vials. The vials were heated at 40 °C for 12 h to vaporize volatile compounds in the cores. The vials then were cooled to room temperature, and a 100 μ L sample of the headspace was collected into a gas syringe. Gas samples were analyzed by photoionization detection on a Photovac 10S Plus gas chromatograph. Concentrations are reported here as nanomoles of gas per liter of core water

TABLE 1. Concentrations of *cis*-1,2-Dichloroethene (cDCE) and Trichloroethene Vapor (TCE) (Expressed as nmol/L of Gas per Volume of Water) in Tree Cores from the TNX Flood Plain, Savannah River Site, SC^a

tree	date	cDCE (nmol/L)	TCE (nmol/L)	species	tree	date	cDCE (nmol/L)	TCE (nmol/L)	species
1	1/22/97	nd	<50 (<50)	swe	45	1/30/98	54	310	cyp
1	10/1/97	60	<20	swe	46	1/30/98	15	536	cyp
2	1/22/97	nd	935 (735)	cyp	47	1/30/98	<10	<50	cyp
2	1/22/97	nd	180	cyp	48	1/30/98	<10	<50	cyp
2	10/1/97	55	220 (180)	cyp	49	1/30/98	<10	<50	cyp
2	1/30/98	<10	376	cyp	50	1/30/98	12	415	cyp
3	1/22/97	nd	245 (380)	cyp	51	1/30/98	18	221	cyp
3	2/13/97	nd	110	cyp	52	1/30/98	25	430	cyp
3	10/1/97	210	100	cyp	53	1/30/98	<10	<50	cyp
3	1/16/98	<10	298	cyp	54	1/30/98	<10	1324	cyp
3	1/30/98	<10	<50	cyp	55	1/14/98	99	966	cyp
4	1/22/97	nd	410 (445)	cyp	56	1/14/98	124	242	cyp
5	2/13/97	nd	<50	cyp	57	1/16/98	<10	<50	cyp
5	8/11/97	<10	<50	cyp	58	1/14/98	<10	<50	cyp
6	2/13/97	nd	<50	cyp	59	1/14/98	<10	<50	cyp
7a	8/11/97	61 (63)	35 040 (35 150)	cyp	60	1/14/98	<10	<50	cyp
7a	10/1/97	175 (176)	4370 (4350)	cyp	60	1/30/98	<10	<50	cyp
7	7/23/97	225 (215)	8590 (9360)	cyp	61	1/14/98	<10	<50	tup
7	9/9/97	nq	3180	cyp	62	1/14/98	<10	<50	tup
7	1/30/98	24 (17) (35)	2093 (2094) (2942)	cyp	63	1/14/98	<10	28	cyp
8	8/11/97	<10	230	swe	64	1/14/98	<10	96	cyp
9	8/11/97	<10	<50	oak	65	1/14/98	<10	272	cyp
9	9/9/97	<10	<50	oak	66	1/14/98	29	167	cyp
9	1/30/98	<10	<50	oak	66	1/26/98	23 (25)	244 (242)	cyp
10	9/9/97	<10	<50	swe	66	2/17/98	55 (53)	216 (212)	cyp
11	9/9/97	<10	<50	oak	67	1/14/98	71	351	cyp
12	9/9/97	110	730	lob	68	1/14/98	<10	46	cyp
12	1/30/98	<10	1742	lob	69	1/14/98	<10 (<10)	310 (265)	cyp
13	9/9/97	40	<50	oak	70	1/14/98	<10 (<10)	49 (45)	cyp
13	1/30/98	<10	<50	oak	71	1/14/98	<10	249	cyp
14	9/9/97	<10	60	swe	71	1/16/98	<10	128	cyp
14	1/30/98	<10	<50	swe	72	1/14/98	<10	219	cyp
15	9/9/97	<10	<50	oak	72	1/16/98	<10	213	cyp
16	9/9/97	<10	<50	lob	73	1/14/98	<10	71	cyp
17	9/9/97	<10	<50	swe	73	1/16/98	<10	63	cyp
18	9/9/97	70	<50	oak	74	1/14/98	<10	78	cyp
18	1/16/98	90	<50	oak	74	1/16/98	<10	31	cyp
18	1/30/98	<10	<50	oak	74	2/17/98	<10	25	cyp
19	1/14/98	47	780	cyp	75	1/16/98	<10	<50	cyp
20z	1/14/98	18	119	cyp	76	1/16/98	<10	44	cyp
20z	1/30/98	23	174	cyp	77	1/16/98	<10	87	cyp
20	1/30/98	16 (<10)	283 (248)	cyp	78	1/16/98	<10	409	cyp
21z	1/14/98	66	1021	tup	78	1/26/98	<10 (<10)	418 (454)	cyp
21z	1/30/98	42	1149	tup	78	2/17/98	<10 (<10)	196 (245)	cyp
21	1/30/98	56 (49)	1469 (1583)	tup	79	1/16/98	<10	200	cyp
23	1/14/98	74	426	tup	79	1/26/98	<10	120	cyp
24	1/14/98	23	1033	tup	80	1/16/98	<10	118	cyp
26	1/26/98	19	362	cyp	80	1/26/98	<10	123	cyp
26	1/30/98	26	457	cyp	81	1/16/98	<10	<50	cyp
27	1/30/98	<10	1241	lob	81	1/26/98	<10	<50	cyp
28	1/30/98	<10	<50	oak	82	1/16/98	<10	<50	cyp
29	1/30/98	<10	308	lob	82	1/26/98	<10	33	cyp
30	1/30/98	<10	318	lob	83	1/26/98	<10	<50	cyp
31	1/30/98	<10	751	lob	84	1/26/98	<10	<50	cyp
32	1/30/98	<10	764	lob	85	1/26/98	<10	<50	cyp
33	1/30/98	<10	74	swe	86	1/26/98	<10	<50	cyp
34	1/30/98	<10	2263	lob	87	2/17/98	<10	46	cyp
35	1/30/98	<10	69	oak	88	2/17/98	<10	43	cyp
36	1/30/98	<10	<50	syc	89	2/17/98	<10	160	cyp
37	1/30/98	<10	130	syc	90	2/17/98	<10	148	cyp
38	1/30/98	<10	106	cyp	91	2/17/98	<10	270	cyp
39	1/30/98	<10	116	cyp	92	2/17/98	<10	48	cyp
40	1/30/98	<10	118	lob	93	2/17/98	<10	<50	cyp
41	1/30/98	<10	1131	lob	94	2/17/98	<10 (<10)	<50 (<50)	cyp
42	1/30/98	<10	479	lob	95	2/17/98	<10	<50	cyp
43	1/30/98	32	296	cyp	96	2/17/98	<10	<50	cyp
43	2/17/98	33	153	cyp	97	2/17/98	<10	<50	cyp
44	1/30/98	11	824	cyp	98	2/17/98	<10	<50	cyp
44	2/17/98	<10 (<10)	270 (229)	cyp	99	2/17/98	<10	<50	cyp

^a nd indicates analysis not done; nq indicates compound not quantified due to peak interferences; () indicates concentration in replicate core sample collected 25 mm laterally from original core sample; cyp indicates bald cypress; lob indicates loblolly pine; tup indicates tupelo; swe indicates sweet gum; syc indicates sycamore.

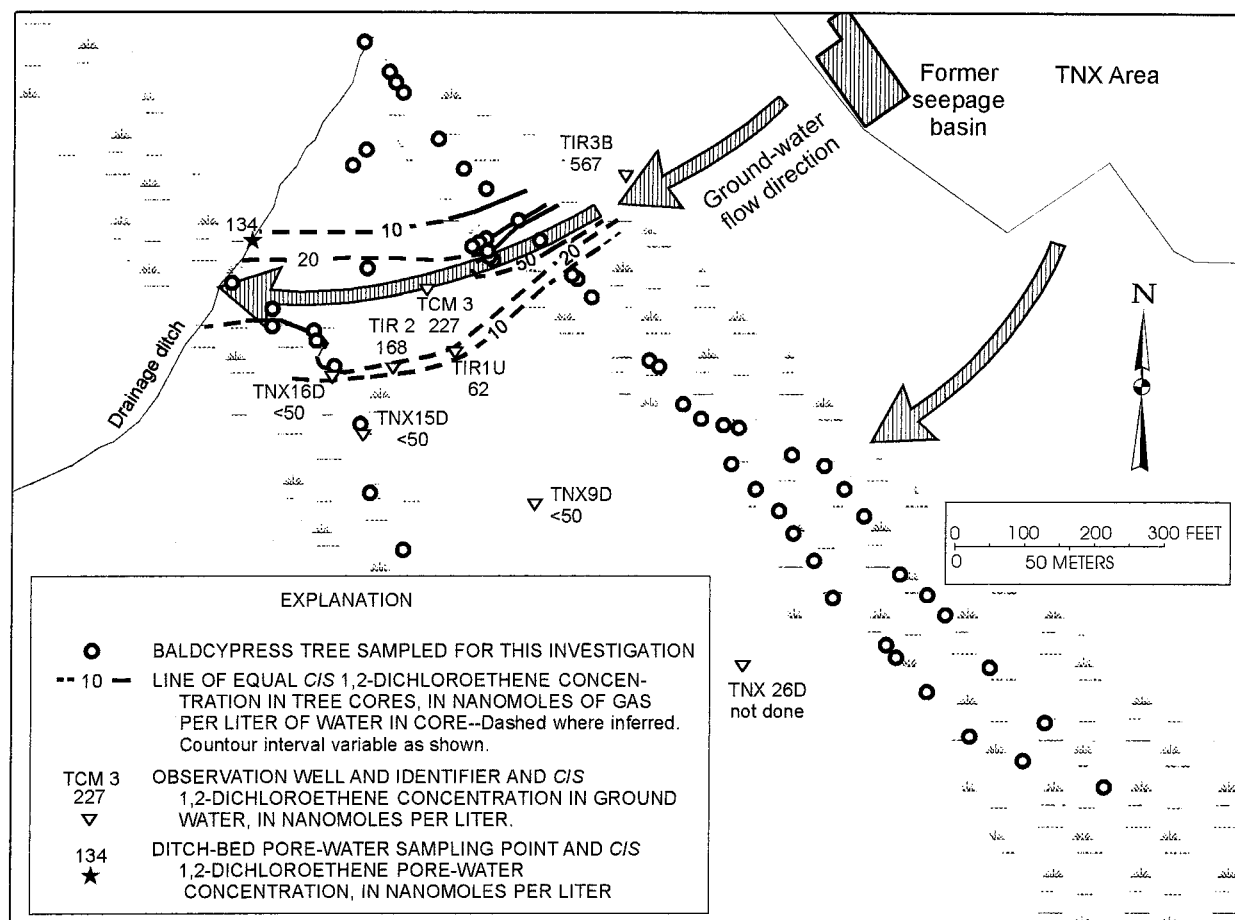


FIGURE 3. *cis*-1,2-Dichloroethene concentrations in bald cypress trunks in January and February 1998 and in groundwater during August 1997 and groundwater flow directions (15), TNX flood plain, Savannah River Site, SC.

(nmol/L). Statistical tests were done on concentrations based on core-water volume, and the core dry weight verified that the sample normalization procedure did not significantly affect the results.

Groundwater was purged and sampled from wells using a positive-displacement submersible pump. Casing water was purged until specific conductance stabilized. Groundwater was sampled from beneath the drainage ditch by pushing a 6.35-cm i.d. stainless steel sampling point into the streambed so that the screened interval (7.8 mm) was approximately 0.6 m below the stream base. Interstitial water was pumped from the piezometer using a peristaltic pump. Water was pumped at a slow rate, and a minimum amount of water (20 mL) was removed prior to collecting a water sample for volatile organic carbons (VOCs) to avoid dilution effects from mixing with streamwater. Water samples were collected in 40-mL glass bottles, capped with Teflon-lined bottle caps, and analyzed using U.S. Environmental Protection Agency Method 8240 (16).

Statistics were performed using the SAS software package (SAS Institute Inc., Cary, NC). Analysis of variance tests (PROC GLM) were used to evaluate differences in TCE and *cDCE* concentrations among tree species and between sites (control vs contaminated). Linear regression analysis was used to test for changes in TCE and *cDCE* with height above ground.

Results and Discussion

Groundwater in the vicinity of several test trees contained TCE and lesser amounts of *cDCE* (Table 1). These compounds were not detected in air samples adjacent to the trees, indicating that the compounds were derived from below land surface. In May 1997, water samples from wells TIR 3B and

TCM 3 contained 3577 and 1187 nmol/L of TCE, respectively (William Pidcoe, Westinghouse Savannah River Company, written communication, July 1997). Well TCM 3 also contained 246 nmol/L of *cDCE*. Numerical simulations (15) suggest that the path of contaminated groundwater flow from the former seepage lagoon is toward a reach of the drainage ditch near the embankment separating the upper and lower flood plains (Figure 3). Groundwater beneath the ditch, collected from a drive-point well screen (Figure 2) in the probable contaminant-discharge zone, contained 134 nmol/L of *cDCE* and 822 nmol/L of TCE in August 1997, providing supporting evidence for the hypothesized path of contaminant transport.

cis-1,2-Dichloroethene was present in cores from several bald cypress trees growing in the area of *cDCE*-contaminated groundwater. The area where *cDCE* was found in bald cypress trees was along a path coincident with the groundwater flow path from the former seepage basin (Figure 3) and coincided with areas where *cDCE* was found in groundwater. The data strongly suggest that the *cDCE* in tree trunks is derived from the contaminated groundwater originating from the former seepage basin.

TCE also was found in bald cypress trunks growing in the area of TCE-contaminated groundwater downgradient from the former seepage basin (Figure 4). The distribution of bald cypress containing TCE was more widespread than the distribution of bald cypress containing *cDCE*. Moreover, TCE also was found in tree 78 and the neighboring trees farther south than the flow path from the former seepage basin (Figure 4). These data suggest a second plume of TCE in the aquifer. Although no data was collected from well TNX 26D during 1997–1998, 41–84 nmol/L of TCE was found in

(18). Additional possible explanations include TCE degradation or sorption within the trunk, the potential for spiral transport of fluids up the trunk (19), or temporal changes in groundwater contaminant concentrations.

Although all of the cores collected for this analysis were from the northeastern part of each tree, selected trees were cored at various locations around the trunk to examine directional variability. Core data from different sides of individual trees showed concentration differences ranging from 44 to 92% for TCE and from 6 to 90% for cDCE. The relatively good replication in cores collected 25 mm apart (15.5% for TCE and 2.5% for cDCE) indicates that the coring approach did not contribute significant inconsistencies to the data. The source of the directional variation is not known but may be related to a variety of factors, such as injuries (20), disease and insect damage (21), gas embolisms (22), and variations in TCE concentration taken up by root systems on differing sides of the tree.

All species of trees examined from areas of groundwater contamination showed evidence of TCE or cDCE in the trunks. Some species appeared to exhibit similar uptake potential. Examination of a cluster of bald cypress and tupelo (trees 19–24, 50–52, and 55) showed no significant differences in concentrations between the species. In January 1998, the TCE concentration in bald cypress 7 was approximately 2000–3000 nmol/L, and high concentrations of TCE also were found in nearby loblolly pines 12, 27, and 54 (1742, 1241, and 1324 nmol/L, respectively). Bald cypress 43 also contained similar TCE concentrations (296 nmol/L) as adjacent loblolly pine 42 (479 nmol/L).

Oaks, however, appeared to contain less TCE than adjacent bald cypress or loblolly pines. In September 1997, oak 11 contained <50 nmol/L of TCE while a nearby loblolly pine (tree 12) contained 730 nmol/L of TCE and a nearby bald cypress tree (tree 7) contained 3180 nmol/L of TCE (Table 1). Similarly, in January 1997, a different oak (tree 35) contained only 69 nmol/L of TCE, whereas loblolly pines on either side of tree 35 contained 2263 (tree 34) and 479 (tree 42) nmol/L. Likewise, oak 13 contained less than 50 nmol/L of TCE while an adjacent loblolly (tree 32) contained 764 nmol/L of TCE. The consistency of the data implies that these findings are function of tree-species differences rather than an artifact of variability due to sampling a particular side of each tree.

Sweet gum also appeared to contain less TCE than loblolly pines. Sweet gums 14 and 33 contained <50 and 74 nmol/L of TCE, respectively, in January 1998, while loblolly pines on various sides of the sweet gums contained 765 (tree 32), 2263 (tree 34), 308 (tree 29), 318 (tree 30), and 751 nmol/L (tree 31) (Table 1).

Previous investigations also have noted concentration differences among species. Selected chlorinated compounds have been found to be degraded faster in the rhizosphere soil of monocot species than dicot species (23). Loblolly pines have been found to take up more TCE than grasses and legumes (11). In this investigation, the concentration differences also may partly be a function of the water-conduction differences between species. Conifers (such as bald cypress and loblolly pine) conduct water through more than the outermost ring, whereas in ring-porous trees (such as oak) nearly all of the water is conducted through the outermost growth ring (24, 25). Thus, the higher concentrations detected in conifers relative to the oaks may be because the cores, being of approximately equal length, incorporated more of the transpiration stream in conifers than in the ring-porous trees.

The extensive areas of swamp and periodic flooding impart difficulties in installing and maintaining effective groundwater sampling wells at this site. The large number of trees available, however, and the ability of those trees to take up

cDCE and TCE, even in areas of standing water, make tree coring a potentially cost-effective and simple approach to optimizing well siting in this type of environment.

Acknowledgments

The use of trade names does not imply endorsement by the U.S. Geological Survey.

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