

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/15636032>

Furanfurin and Thiophenfurin: Two Novel Tiazofurin Analogs. Synthesis, Structure, Antitumor Activity, and Interactions with Inosine Monophosphate Dehydrogenase

ARTICLE *in* JOURNAL OF MEDICINAL CHEMISTRY · OCTOBER 1995

Impact Factor: 5.45 · DOI: 10.1021/jm00019a013 · Source: PubMed

CITATIONS

72

READS

17

10 AUTHORS, INCLUDING:



Loredana Cappellacci

University of Camerino

96 PUBLICATIONS 1,193 CITATIONS

SEE PROFILE



Giuseppe Nocentini

Università degli Studi di Perugia

107 PUBLICATIONS 2,552 CITATIONS

SEE PROFILE

Furanfuran and Thiophenfuran: Two Novel Tiazofurin Analogues. Synthesis, Structure, Antitumor Activity, and Interactions with Inosine Monophosphate Dehydrogenase

Palmarisa Franchetti,* Loredana Cappellacci, Mario Grifantini, Anna Barzi,[†] Giuseppe Nocentini,[†] Hongyaoan Yang,[‡] Ayrn O'Connor,[‡] Hiremagalur N. Jayaram,[‡] Christopher Carrell,[§] and Barry M. Goldstein^{||}

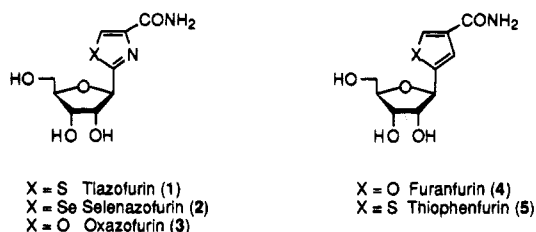
Dipartimento di Scienze Chimiche, Università di Camerino, 62032 Camerino, Italy, Dipartimento di Medicina Clinica, Patologia e Farmacologia, Università di Perugia, 06100 Perugia, Italy, Laboratory for Experimental Oncology, Indiana University School of Medicine, Indianapolis, Indiana 46202-5200, Fox Chase Cancer Center, Fox Chase, Pennsylvania 19111, and Department of Biophysics, University of Rochester Medical Center, Rochester, New York, 14642

Received April 13, 1995*

The syntheses of furan and thiophene analogues of tiazofurin (furanfuran and thiophenfuran, respectively) are described. Direct stannic chloride-catalyzed C-glycosylation of ethyl 3-furan-carboxylate (**6**) or ethyl 3-thiophencarboxylate (**18**) with 1,2,3,5-tetra-*O*-acetyl-D-ribofuranose gave 2- and 5-glycosylated regioisomers, as a mixture of α - and β -anomers, and the β -2,5-diglycosylated derivatives. Deprotection of ethyl 5-(2,3,5-tri-*O*-acetyl- β -D-ribofuranosyl)furan-3-carboxylate (**9** β) and ethyl 5-(2,3,5-tri-*O*-acetyl- β -D-ribofuranosyl)thiophene-3-carboxylate (**20** β) with sodium ethoxide afforded ethyl 5- β -D-ribofuranosylfuran-3-carboxylate (**12** β) and ethyl 5- β -D-ribofuranosylthiophene-3-carboxylate (**23** β) which were converted into 5- β -D-ribofuranosylfuran-3-carboxamide (furanfuran, **4**) and 5- β -D-ribofuranosylthiophene-3-carboxamide (thiophenfuran, **5**) by reaction with ammonium hydroxide. The anomeric configuration and the site of glycosylation were established by ¹H-NMR and proton-proton nuclear Overhauser effect difference spectroscopy. The structure of compound **23** β was confirmed by X-ray crystallography. Thiophenfuran was found to be cytotoxic *in vitro* toward murine lymphocytic leukemia P388 and L1210, human myelogenous leukemia K562, human promyelocytic leukemia HL-60, human colon adenocarcinoma LoVo, and B16 melanoma at concentrations similar to that of tiazofurin. In the same test furanfuran proved to be inactive. Thiophenfuran was found active *in vivo* in BD₂F₁ mice inoculated with L1210 cells with a % T/C of 168 at 25 mg/kg. K562 cells incubation with thiophenfuran resulted in inhibition of inosine monophosphate (IMP) dehydrogenase (63%) and an increase in IMP pools (6-fold) with a concurrent decrease in GTP levels (42%). Incubation of adenosine-labeled K562 cells with tiazofurin, thiophenfuran, and furanfuran resulted in a 2-fold higher NAD analogue formulation by thiophenfuran than by tiazofurin. Furanfuran was converted to the NAD analogue with only 10% efficiency. The results obtained support the hypothesis that the presence of S in the heterocycle in position 2 with respect to the glycosidic bond is essential for the cytotoxicity and IMP dehydrogenase activity of tiazofurin, while the N atom is not.

Tiazofurin (2- β -D-ribofuranosylthiazole-4-carboxamide, NSC-286193, **1**) and selenazofurin (2- β -D-ribofuranosylselenazole-4-carboxamide, NSC-340847, **2**) are two widely studied C-nucleosides endowed with several biological effects. These include effective antitumor activity both *in vitro* and *in vivo*^{1a-f} and the ability to induce differentiation in neoplastic cells,² to inhibit the G protein-mediated cellular signaling mechanism,³ and to downregulate oncogene activity.⁴

The biological effects of these nucleosides, which are structurally related to ribavirin,⁵ appear to be due to inhibition of inosine monophosphate dehydrogenase (IMPDH), which induces the shutdown of guanine nucleotide synthesis.^{1d,f,6,7} In sensitive cells, tiazofurin and selenazofurin are converted into analogues of the co-factor nicotinamide adenine dinucleotide (NAD). These NAD analogues, called TAD (thiazole-4-carboxamide



adenine dinucleotide) and SAD (selenazole-4-carboxamide adenine dinucleotide), are excellent inhibitors of IMPDH as well as of other dehydrogenases.^{1f,7}

Crystallographic studies of tiazofurin and selenazofurin showed close contacts between the thiazole S or selenazole Se heteroatoms and the furanose oxygen.⁸ This close contact has been attributed to an electrostatic attractive interaction between the positively charged sulfur or selenium and the lone pair of electrons on the furanose oxygen, as supported by molecular orbital calculations.⁸ This interaction would be expected to constrain rotation about the C-glycosidic bond in the active anabolites TAD and SAD, influencing the binding of these dinucleotide inhibitors to the target enzyme. This effect was recently observed in crystal structures of enzyme-bound analogues of TAD and SAD.⁹

* Address for correspondence: Dipartimento di Scienze Chimiche, Università di Camerino, Via S. Agostino, 1, 62032 Camerino (MC), Italy. Tel: (737)40336. Fax: (737)37345.

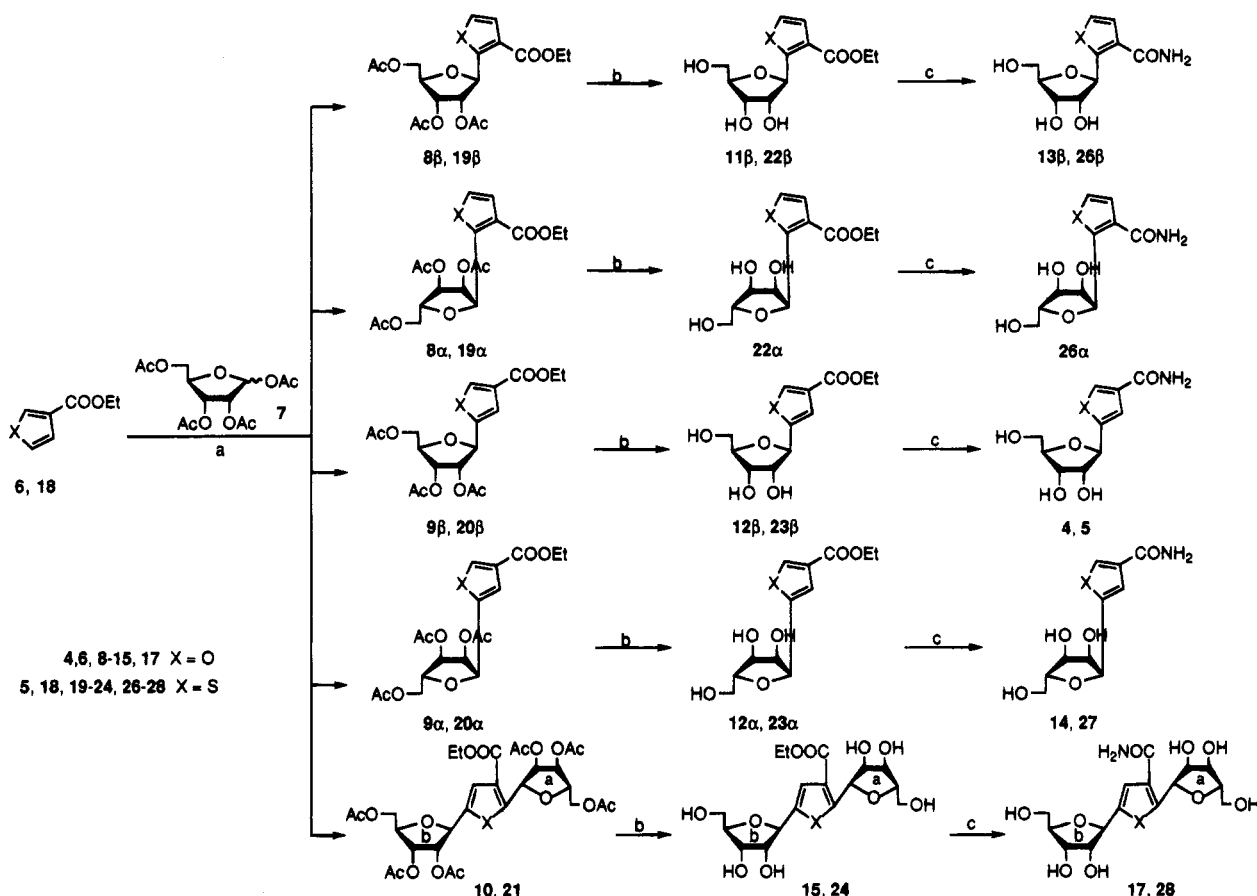
[†] Università di Perugia.

[‡] Indiana University School of Medicine.

[§] Fox Chase Cancer Center.

^{||} University of Rochester Medical Center.

© Abstract published in *Advance ACS Abstracts*, August 1, 1995.

Scheme 1^a

Recently we have synthesized oxazofurin (3), an analogue of tiazofurin and selenazofurin in which the S and Se heteroatoms are substituted by oxygen. This compound was found to be inactive as an antitumor and antiviral agent.¹⁰ The inactivity of oxazofurin might be due to the O—O-1' repulsion which does not allow oxazole and furanose moieties in the putative anabolite oxazole-4-carboxamide adenine dinucleotide (OAD) to assume the right conformation to bind the enzyme. This hypothesis was supported by crystallographic data and quantum-mechanical-based computations.¹¹

As part of our studies dealing with the structure-activity relationships of these types of C-nucleosides, we now report on the synthesis, structure, antitumor activity, and IMPDH inhibitory activity of oxazofurin and tiazofurin analogues in which the base is replaced by other five-membered ring heterocycles such as furan [furanfurin (4)] and thiophene [thiophenfurin (5)].

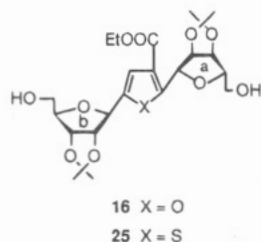
Chemistry

The synthesis of furanfurin (4) was carried out as outlined in Scheme 1 by direct C-glycosylation of ethyl furan-3-carboxylate (6) under Friedel-Crafts conditions as described for heterocycle analogues.¹² The reaction of 6 with tetra-O-acetyl- β -D-ribofuranose (7) in 1,2-dichloroethane in the presence of SnCl_4 gave 2- and 5-glycosylated regioisomers as mixture of α - and β -anomers (8 α , 8 β , 9 α , 9 β , 72%) and the 2,5-di- β -D-ribofuranosyl derivative 10 (12%). The isomers 8 and 9, as anomers α and β , and compound 10 were separated by column chromatography. Compounds 8 β , 9 α , and 9 β were treated with a catalytic amount of sodium ethoxide

in ethanol to give the corresponding deblocked ethyl esters 11 β , 12 α , and 12 β . The glycosylation position was determined by ^1H -NMR and proton-proton nuclear Overhauser effect (NOE) difference spectroscopy. The ^1H -NMR spectrum of 11 β showed that the signal of the H-2 proton of furan had disappeared, indicating that the glycosylation position was at C-2. On the other hand, in the ^1H -NMR spectra of compounds 12 α and 12 β , the signal of the H-5 proton was lacking, supporting C-5 glycosylation. The structures of compounds 12 α and 12 β were further supported by NOE experiments. In fact, when the H-1' signal of these compounds was irradiated, a NOE effect was observed at H-4 confirming that the ribosyl moiety resides at C-5. The anomeric configuration was also assigned on the basis of NOE experiments. In fact, selective irradiation of the anomeric signal of 11 β and 12 β increases the intensity of the H-4' signal (1.8% and 1.7%, respectively), while the intensity enhancement of the H-3' signal was zero; this indicates that H-1' and H-4' are located on the same face of the ribosyl ring.¹³ The anomeric configuration was further supported by studies of ^{13}C -NMR data. Thus, the finding that C-1' in 12 β resonates upfield of C-1' in 12 α is in accord with previous observation of anomeric chemical shifts in other C-ribofuranose compounds as well as N-nucleosides.¹⁴

Treatment of 12 β with ammonium hydroxide (30%) gave furanfurin (4). In a similar way, ethyl esters 11 β and 12 α were converted into the amides 13 β and 14. As far as compound 10 is concerned, the glycosylation positions were established to be at C-2 and C-5 on the basis of the ^1H -NMR spectrum which showed that the

H-2 and H-5 signals were lacking. The anomeric configuration was established to be β on the basis of the observation that in the ^{13}C -NMR spectrum the chemical shift patterns of the bis-isopropylidene derivative of **15** (**16**), measured in CDCl_3 , were very close to the ranges reported by Ohrui *et al.* and Cousineau and Secrist.¹⁵ In fact, the spectrum exhibits methyl signals of the isopropylidene groups at 25.8 and 27.8 ppm, with a $\Delta\delta$ of 2.0 ppm, and the signals of the quaternary isopropylidene carbons at 114.8 and 115.1 ppm, clearly indicative of the β -configuration. Ester **15** was reacted with ammonium hydroxide to give amide **17**.



Thiophenfuran (**5**) was prepared in a similar way as furanfuran starting from ethyl thiophene-3-carboxylate (**18**) (Scheme 1). The reaction with **7** gave a mixture of 2- and 5-glycosylated regioisomers as a mixture of α - and β -anomers (**19** α , **19** β , **20** α , **20** β , 62%) and the 2,5-bis-glycosylated derivative **21** (6%). In this case, the mixture of anomers **19** α , **19** β , and **20** β was separated from **20** α and **21** by column chromatography. Treatment of the mixture of **19** α , **19** β , and **20** β with sodium ethoxide in ethanol gave the deblocked ethyl esters **22** α , **22** β , and **23** β , which were separated by chromatography. In a similar way, starting from **20** α and **21**, the esters **23** α and **24** were obtained. The glycosylation position and the anomeric configurations, assigned by ^1H - and ^{13}C -NMR measurements, were further supported by NOE experiments as described for compounds **11** and **12**. The anomeric configuration of **21** was assigned on the basis of the chemical shift patterns of the bis-isopropylidene derivative of **24** (**25**). Treatment of **23** β with 30% ammonium hydroxide gave thiophenfuran (**5**). In a similar way, starting from **22** α , **22** β , **23** α , and **24** were obtained the amides **26** α , **26** β , **27**, and **28**.

Crystallographic and Computational Studies

Owing to the impossibility of obtaining thiophenfuran and furanfuran in solid form, we have studied the crystal structure of the intermediate **23** β (ethyl 5- β -D-ribofuranosylthiophene-3-carboxylate) by X-ray analysis. The molecular structure of the thiophenfuran carboxylate intermediate shares several features observed in the structures of related thiazole and selenazole nucleosides (Figure 1). The thiophene ring is planar, with the ethyl carboxylate substituent at C-4 coplanar to the heterocycle. The C-glycosidic torsion angle is 46.5° , somewhat higher than that observed in the thiazole nucleosides. However, the thiophene sulfur remains *cis* to the furanose oxygen, with a marginally close nonbonded S–O contact of 3.04 Å. *Ab initio* computations (Figure 2) suggest that this conformation is stabilized by an electrostatic interaction between a positively charged thiophene sulfur and a negatively charged furanose oxygen; this effect is similar to that identified for tiazofurin, selenazofurin, and their analogues.⁸

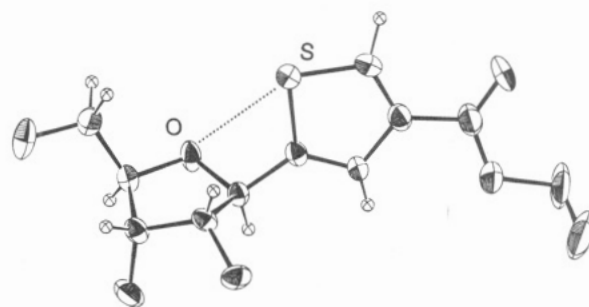


Figure 1. Molecular structure of thiophenfuran-4-carboxylate (**23** β). Non-hydrogen atoms are represented by thermal ellipsoids at the 30% probability level. The dotted line indicates the 3.04 Å nonbonded S–O contact.

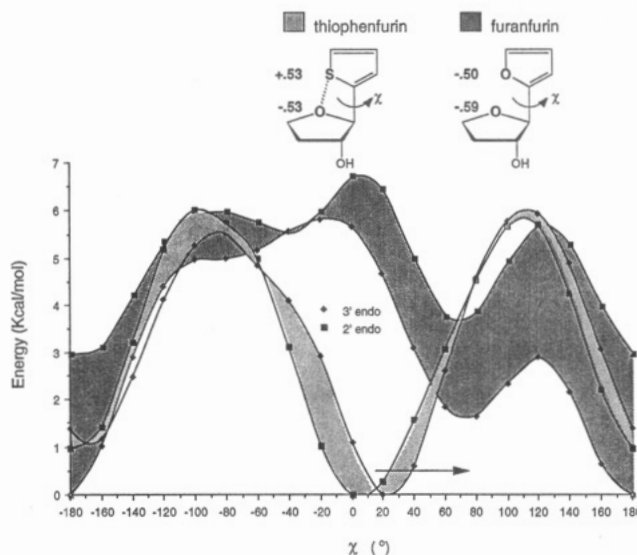


Figure 2. Energy versus C-glycosidic torsion angle χ in the thiophenfuran and furanfuran model fragments. The fragments are illustrated at top in the $\chi = 0^\circ$ conformation. Numbers by the thiophene sulfur and the furan oxygen and both furanose oxygens are natural bond orbital point charges. Rotation about χ is indicated by the curved arrows. Points on each curve were obtained at the RHF/3-21G(*)/3-21G(*) level. Shaded areas for each model represent the span in energy between 2'-endo and 3'-endo conformations. The horizontal arrow indicates the shift to a higher value of χ predicted for furanfuran relative to thiophenfuran.

Calculation of energy versus C-glycosidic torsion angle for a thiophenfuran model fragment is shown in Figure 2. The thiophenfuran fragment shows a global minimum in the range $\chi \sim 0^\circ$ – 25° , depending upon the sugar conformation. A similar value is obtained for tiazofurin.⁸ These results suggest that the minimum energy value of χ for thiophenfuran is somewhat lower than that observed in the carboxylate crystal structure. Conformers at lower values of χ are stabilized by an attractive sulfur–oxygen interaction. Donation of mobile sulfur valence electrons to the conjugated system of the thiophene ring results in a positive charge on the heteroatom (Figure 2). This effect is similar to that seen in the thiazole ring of tiazofurin⁸ and accounts for the electrostatic attraction between the sulfur and the negatively charged furanose oxygen.

The energy profile for the thiophenfuran fragment also shows a local minimum at $\chi = \pm 180^\circ$ (Figure 2). However, this conformer is separated from that at the global minimum by a 5–6 kcal/mol barrier to rotation. The local minimum at 180° is due primarily to interactions between a positively charged proton on C-3 of the

Table 1. Cytotoxicity of Tiazofurin, Furanfuran, and Thiophenfuran

analogue	cytotoxicity (IC ₅₀ , ^a μ M)						
	P388 ^b	L1210 ^b	K562 ^b	HL-60 ^b	LLC ^b	LoVo ^b	B16 ^b
tiazofurin	2.2	7.4	4.5	8.7	1.5	>128	16.2
furanfuran	^c	^c	^c	^c	^c	^c	^c
thiophenfuran	3.2	7.6	8.8	13.9	11.2	102	25.8

^a The concentration of agent required to reduce cell viability by 50%. ^b P388 and L1210 murine lymphocytic leukemias, K562 human myelogenous leukemia, HL-60 human promyelocytic leukemia, Lewis lung carcinoma (LLC), LoVo human colon adenocarcinoma, and B16 melanoma. ^c Inactive until a maximum tested concentration of 200 μ M.

thiophene ring and the furanose oxygen. This minimum is not observed in tiazofurin, whose negative thiazole nitrogen destabilizes the 180° conformer.⁸

Calculation of energy versus C-glycosidic torsion angle for a furanfuran model fragment is also shown in Figure 2. The furanfuran model shows a local minimum in the range $\chi \sim 70$ –80°, depending upon the sugar conformation. This is $\sim 60^\circ$ higher than the global minimum computed for the thiophenfuran fragment (Figure 2) and is similar to values of χ observed in structures of the related inactive analogue oxazofurin.¹¹ Further, the local minimum for furanfuran at $\chi = 70$ –80° is itself 1–2 kcal/mol higher than a global minimum at $\chi = 180^\circ$.

Antitumor Evaluation and Discussion

Furanfuran (4) and thiophenfuran (5) have been evaluated for their ability to inhibit the growth of murine lymphocytic leukemia P388 and L1210, human myelogenous leukemia K562, human promyelocytic leukemia HL-60, Lewis lung carcinoma (LLC), human colon adenocarcinoma LoVo, and B16 melanoma. Tumor growth inhibition was evaluated by the MTT assay after 72 h of cell–compound contact.¹⁶ The IC₅₀ (μ M) values for these C-nucleosides and tiazofurin are summarized in Table 1.

Furanfuran was inactive until a maximum tested concentration of 200 μ M. On the contrary, thiophenfuran demonstrated cytotoxicity against leukemic cells comparable to that of tiazofurin. Thiophenfuran exhibited a similar order of cytotoxicity against solid tumor cells as that observed for tiazofurin.

The cytotoxic activity of thiophenfuran was also tested against normal human bone marrow cells from five different healthy donors. The comparison of the results with those obtained with HL-60 cells (see ratio of the means, Table 2) indicates that thiophenfuran exhibits a good selectivity toward tumor cells. This is especially remarkable since in this system most of the cytotoxic compounds tested are equally or more toxic toward normal cells than tumor cells.

We also evaluated *in vivo* antitumor activity of thiophenfuran in BD₂F₁ mice inoculated intraperitoneally with 1×10^5 L1210 cells. Results reported in Table 3 indicate that thiophenfuran has antitumor activity comparable to that of tiazofurin at a dose of 25 mg/kg. However, the high-dose treatment, which gives a better antitumor activity using tiazofurin, cannot be adopted with thiophenfuran since this agent seems to be toxic. The relatively high toxicity of thiophenfuran was not seen in *in vitro* experiments (see Table 2). This discrepancy could be due to nonhematological toxicity.

As far as the structure–activity relationship is concerned, it was found that the glycosylation position and

the anomeric configuration are important for the cytotoxicity. In fact, the 2-glycosylated isomers **26 β** and **26 α** and the α -anomer **27** were inactive (data not shown), as well as the bis-glycosylated isomer **28**.

The inactivity of furanfuran confirms the hypothesis that the presence of the sulfur in the five-membered ring heterocycle in position 2 with respect to the glycosidic bond is fundamental for the antitumor activity of this type of C-nucleosides, whereas the nitrogen atom contributes little or nothing to the activity of tiazofurin.

In furanfuran, the thiophene sulfur is replaced by the more electronegative furan oxygen, resulting in a negative charge in this region of the heterocycle. Thus, conformers in which the furan and furanose oxygens are *cis* (i.e., low values of χ) are destabilized by electrostatic repulsion between the two negatively charged atoms (Figure 2). A similar effect is seen in oxazofurin.¹¹ Like oxazofurin, furanfuran is inactive.

The activity of tiazofurin requires its enzymatic conversion to the active dinucleotide analogue TAD.^{1,6,7} The activity of thiophenfuran likely requires a similar mechanism. Conversely, the inactivity of furanfuran may be due to its inability to be converted to the dinucleotide or failure of the dinucleotide to bind to the target. Either effect could result from the electronic and conformational differences between furanfuran and thiophenfuran discussed above.

In order to test these hypotheses, we evaluated the formation of NAD analogues by tiazofurin, thiophenfuran, and furanfuran in human myelogenous leukemia K562 cells. As showed in Table 4, incubation of adenosine-labeled cells with C-nucleosides resulted in a 2.5-fold higher NAD analogue formation by thiophenfuran than by tiazofurin. As expected, furanfuran was converted with difficulty to the NAD analogue with only 10% efficiency. These results are in accord with the inhibition of IMPDH activity. For this study, K562 cells in culture were exposed to thiophenfuran (10, 50, or 100 μ M) or saline for 2 h at 37 °C, and then IMPDH activity was assayed (Table 5). The results indicate that IMPDH activity in K562 cells was inhibited by the action of thiophenfuran with a potency similar to that of tiazofurin.

Since inhibition of IMPDH results in perturbation of nucleotide pools, we examined the influence of this compound on the ribonucleotide concentration. The results provided in Table 6 show a dose-dependent increase in IMP levels (6-fold) with a concurrent decrease in guanylate concentration including GMP, GDP, and GTP. However, perturbation of adenylate pools was marginal.

In conclusion, these studies support the hypothesis that the presence of S in the heterocycle in position 2 with respect to the glycosidic bond is essential for cytotoxicity and IMPDH inhibition, while the N atom is not.

Experimental Section

Melting points were determined on a Buchi apparatus and are uncorrected. Elemental analyses were determined on a EA 1108 CHNS-O (Fisons Instruments) analyzer. Ultraviolet spectra were recorded on an HP 8452 A diode array spectrophotometer driven by an Olivetti M 24. Thin layer chromatography (TLC) was run on silica gel 60 F₂₅₄ plates; silica gel 60 (70–230 and 230–400 mesh) (Merck) for column chromatography was used. ¹H- and ¹³C-NMR spectra were deter-

Table 2. Relative Toxicity of Tiazofurin (1) and Thiophenfurin (5) *in Vitro*

compd	bond marrow samples		HL-60		ratio of the means ^a		
	range IC ₅₀ ^b (μM)	range IC ₉₀ ^b (μM)	range IC ₅₀ ^b (μM)	range IC ₉₀ ^b (μM)	IC ₅₀ (BM)/ IC ₅₀ (HL)	IC ₉₀ (BM)/ IC ₉₀ (HL)	IC ₅₀ (BM)/ IC ₉₀ (HL)
1	10.4–176.4	>128	2.5–13.7	7.6–67.7	7.3	>6	1.9
5	21.6–226.7	>128	2.4–4.3	8.3–17.2	21.9	>10	5.9

^a Ratio of antilog (mean log IC₅₀ or IC₉₀) on human bone marrow (BM) and antilog(mean log IC₅₀ or IC₉₀) on human cell line HL-60 (HL). ^b The lower value is equal to antilog(mean log IC₅₀ – 2SD), and the higher value is equal to antilog(mean log IC₅₀ + 2SD).

Table 3. Antitumor Activity of Thiophenfurin and Tiazofurin *in Vivo*

compd	dose ^a (mg/kg)	avg Δwt ^b (%)	T/C ^c (%)
thiophenfurin	400	<i>d</i>	<100
	100	<i>d</i>	<100
	25	–9.8	168
	6.2	+8.5	155
tiazofurin	400	–13.1	260
	25	–4.3	177

^a Female mice were injected intraperitoneally with L1210 cells as detailed in the methods. Treatment was started 24 h later, once a day for 9 consecutive days. ^b Averaged weight change of mice from onset to termination of drug treatment. ^c % T/C represents the ratio of the survival time of treated to control mice × 100. ^d Early death due to toxicity.

Table 4. Formation of NAD Analogues by Tiazofurin, Thiophenfurin, and Furanfuran in Human Myelogenous Leukemia K562 Cells in Culture^a

compd	concentration (μM)	NAD analogue (pmol/10 ⁶ cells)
tiazofurin	50	186.5 ± 19.3
	100	309.9 ± 28.9
thiophenfurin	50	504.9 ± 56.8
	100	885.5 ± 77.2
furanfuran	50	49.4 ± 2.1
	100	124.4 ± 17.2

^a Cells in culture (1 × 10⁷) were labeled with [³H]adenosine for 60 min at 37 °C and then with saline, tiazofurin, thiophenfurin, or furanfuran for 2 h. Cells were centrifuged, extracted with trichloroacetic acid, and neutralized with 0.5 M tri-*n*-octylamine, and an aliquot was analyzed on HPLC.

Table 5. Effects of Thiophenfurin on IMPDH Activity of K562 Cells in Culture

compd	concentration (μM)	IMPDH activity (nmol of XMP formed/ h/mg of protein)	% of control
control		4.97 ± 0.13	100
thiophenfurin	10	2.80 ± 0.18	56 ^a
	50	1.85 ± 0.15	37 ^a
	100	1.07 ± 0.29	22 ^a
tiazofurin	50	1.82 ± 0.08	37 ^a

^a Significantly different from control values (*p* < 0.05).

mined at 300 and 75 MHz, respectively, with a Varian VXR-300 spectrometer. The chemical shift values are expressed in δ values (ppm) relative to tetramethylsilane as an internal standard. All exchangeable protons were confirmed by addition of D₂O. Stationary NOE experiments were run on degassed solutions at 25 °C. A presaturation delay of 1 s was used, during which the decoupler low power was set at 20 dB attenuation. Tiazofurin was obtained through the Drug Synthesis & Chemistry Branch, Developmental Therapeutics Program, Division of Cancer Treatment, National Cancer Institute.

Ethyl 2-(2,3,5-Tri-*O*-acetyl-β-D-ribofuranosyl)furan-3-carboxylate (8β), Ethyl 2-(2,3,5-Tri-*O*-acetyl-α-D-ribofuranosyl)furan-3-carboxylate (8α), Ethyl 5-(2,3,5-Tri-*O*-acetyl-β-D-ribofuranosyl)furan-3-carboxylate (9β), Ethyl 5-(2,3,5-Tri-*O*-acetyl-α-D-ribofuranosyl)furan-3-carboxylate (9α), and Ethyl 2,5-Bis(2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl)furan-3-carboxylate (10). To a stirred solution of ethyl furan-3-carboxylate (6) (14 g, 99.9 mmol) in dry 1,2-

Table 6. Effect of Tiazofurin and Thiophenfurin on Ribonucleotide Levels of K562 Cells in Culture

nucleotide	control (nmol/g)	tiazofurin (% of control)		thiophenfurin (% of control)		
		50 μM	100 μM	10 μM	50 μM	100 μM
AMP	275 ± 14	100	103	156 ^a	140 ^a	131 ^a
ADP	1399 ± 17	103	99	122 ^a	114 ^a	105
ATP	4356 ± 13	101	91	92	88 ^a	81 ^a
NAD	791 ± 43	98	84	81	66 ^a	63 ^a
IMP	131 ± 5	544 ^a	578 ^a	314 ^a	594 ^a	689 ^a
GMP	83 ± 2	62 ^a	55 ^a	98	90	80 ^a
GDP	289 ± 7	82	63 ^a	97	79 ^a	77 ^a
GTP	1496 ± 70	64 ^a	54 ^a	71 ^a	58 ^a	58 ^a
UMP	81 ± 16	113	96	110	115	107
UDP	146 ± 8	120	116	134 ^a	134 ^a	131 ^a
UTP	1017 ± 15	115	90	91	99	87
CDP	358 ± 58	104	85	84	82	99
CTP	312 ± 9	92	32 ^a	32 ^a	32 ^a	32 ^a

^a Significantly different from control values (*p* < 0.05). K562 cells in culture (1 × 10⁷) were incubated with saline (control), tiazofurin, or thiophenfurin for 2 h at 37 °C. Cells were extracted with trichloroacetic acid and neutralized with tri-*n*-octylamine, and an aliquot was analyzed on HPLC.

dichloroethane (150 mL) were added 1,2,3,4-tetra-*O*-acetyl-β-D-ribofuranose (7) (32.4 g, 102 mmol) and SnCl₄ (6.4 mL), and the mixture was reacted at room temperature for 18 h. The black solution was diluted with H₂O, and the mixture was neutralized with NaHCO₃ and extracted with CHCl₃. The combined CHCl₃ extracts were dried over anhydrous Na₂SO₄ and evaporated to dryness. The obtained residue was chromatographed on silica gel eluting with CH₂Cl₂–EtOAc (92:8). Compound 8β was separated as the first fraction as a colorless oil (1.9 g, 4%). TLC (CH₂Cl₂–EtOAc, 92:8): *R*_f = 0.28. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 2.05–2.10 (3s, 9H, OCOCH₃), 4.10–4.40 (m, 5H, H-5', H-4', OCH₂CH₃), 5.33 (t, *J* = 5.2 Hz, 1H, H-3'), 5.45 (t, *J* = 5.7 Hz, 1H, H-2'), 5.60 (d, *J* = 5.9 Hz, 1H, H-1'), 6.81 (d, *J* = 2.0 Hz, 1H, H-4), 7.88 (d, *J* = 2.0 Hz, 1H, H-5). Anal. (C₁₈H₂₂O₁₀) C, H.

It was impossible to obtain a pure sample of compound 8α, which was formed in very low yield (<0.5%). ¹H-NMR (DMSO-*d*₆) (deduced from the mixture 8α, 8β): δ 1.29 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 2.05, 2.06 (2s, 9H, OCOCH₃), 4.28 (m, 5H, H-5', H-4', OCH₂CH₃), 5.15–5.28 (m, 2H, H-3', H-2'), 5.66 (d, *J* = 4.9 Hz, 1H, H-1'), 6.80 (d, *J* = 2.0 Hz, 1H, H-4), 7.85 (d, *J* = 1.9 Hz, 1H, H-5).

Evaporation of the following fraction gave 9β as a colorless oil (27.36 g, 60%). TLC (CH₂Cl₂–EtOAc, 95:5): *R*_f = 0.2. ¹H-NMR (DMSO-*d*₆): δ 1.28 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 2.06, 2.08, 2.11 (3s, 9H, OCOCH₃), 4.10–4.40 (m, 5H, H-5', H-4', OCH₂CH₃), 5.02 (d, 4J = 6.4 Hz, 1H, H-1'), 5.28 (pseudo-t, *J* = 5.5 Hz, 1H, H-3'), 5.41 (pseudo-t, *J* = 6.2 Hz, 1H, H-2'), 6.90 (s, 1H, H-4), 8.46 (s, 1H, H-2). Anal. (C₁₈H₂₂O₁₀), C, H.

Compound 9α was obtained from the third eluate as a white solid (3.18 g, 8%). Mp: 95–97 °C. TLC (CH₂Cl₂–EtOAc, 95:5): *R*_f = 0.15. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, 3H, *J* = 7.1 Hz, 3H, OCH₂CH₃), 1.98–2.08 (3s, 9H, OCOCH₃), 4.10–4.38 (m, 5H, H-5', H-4', OCH₂CH₃), 5.32 (dd, *J* = 4.9, 12.3 Hz, 1H, H-1'), 5.42 (d, *J* = 4.2 Hz, 1H, H-3'), 5.55 (pseudo-t, *J* = 4.6 Hz, 1H, H-2'), 6.70 (s, 1H, H-4), 8.38 (s, 1H, H-5). Anal. (C₁₈H₂₂O₁₀) C, H.

Further elution of the same column with CH₂Cl₂–EtOAc (85:15) as eluent yielded 10 as a colorless oil (7.87 g, 12%). TLC (CH₂Cl₂–EtOAc, 85:15): *R*_f = 0.3. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, *J* = 7.0 Hz, 3H, OCH₂CH₃), 2.0–2.12 (3s, 18H,

OCOCH₃), 4.14–4.45 (m, 8H, H-5'a, H-5'b, H-4'a, H-4'b, OCH₂-CH₃), 5.05 (dd, $J = 2.4$, 6.1 Hz, 1H, H-1'b), 5.20–5.45 (m, 4H, H-3'a, H-3'b, H-2'a, H-2'b), 5.97 (d, $J = 4.6$ Hz, 1H, H-1'a), 6.87 (s, 1H, H-4). Anal. (C₂₉H₃₆O₁₇) C, H.

Ethyl 2-β-D-Ribofuranosylfuran-3-carboxylate (11β). Compound **8β** (1.5 g, 3.76 mmol) was treated with sodium ethoxide (33.1 mL, 15.12 mmol) for 1 h at room temperature. To the reaction mixture was added 0.85 g of Dowex 50w × 8 resin (H⁺) (washed with ethanol), and the suspension was stirred for 1 h. The resin was filtered off, washed with ethanol, and discarded. The filtrate was evaporated to dryness, and the residue was chromatographed on silica gel with CHCl₃-MeOH (92:8) as eluent to give **11β** as a colorless oil (0.61 g, 60%). TLC (CHCl₃-MeOH, 92:8): $R_f = 0.36$. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, $J = 7.1$ Hz, 3H, OCH₂CH₃), 3.48 (dd, $J = 5.7$, 11.6 Hz, 2H, H-5'), 3.70 (dd, $J = 4.5$, 9.8 Hz, 1H, H-4'), 3.95 (dd, $J = 5.0$, 9.9 Hz, 1H, H-3'), 4.16 (dd, $J = 6.3$, 10.2 Hz, 1H, H-2'), 4.25 (q, $J = 7.1$ Hz, 2H, OCH₂CH₃), 4.75 (t, $J = 5.7$ Hz, 1H, OH), 5.02 (d, $J = 5.1$ Hz, 1H, OH), 5.10 (d, 1H, $J = 6.3$ Hz, 1H, OH), 5.30 (d, 1H, $J = 6.2$ Hz, H-1'), 6.73 (d, 1H, $J = 2.0$ Hz, H-4), 7.55 (d, $J = 2.0$ Hz, 1H, H-5). Anal. (C₁₂H₁₆O₇) C, H.

Ethyl 5-β-D-Ribofuranosylfuran-3-carboxylate (12β). The title compound was prepared from **9β** (26 g, 65.26 mmol) by the above method and purified by column chromatography using CHCl₃-MeOH (90:10) as eluent (colorless oil, 12.4 g, 70%). TLC (CHCl₃-MeOH, 90:10): $R_f = 0.38$. ¹H-NMR (DMSO-*d*₆): δ 1.28 (t, $J = 7.0$ Hz, 3H, OCH₂CH₃), 3.45 (dd, $J = 5.1$, 10.1 Hz, 2H, H-5'), 3.79 (q, $J = 4.4$ Hz, 1H, H-4'), 3.91 (q, $J = 5.0$ Hz, 1H, H-3'), 4.08 (q, $J = 5.3$ Hz, 1H, H-2'), 4.25 (q, $J = 7.1$ Hz, 2H, OCH₂CH₃), 4.62 (d, $J = 6.6$ Hz, 1H, H-1'), 4.80 (t, $J = 5.6$ Hz, 1H, OH), 4.98 (d, $J = 5.1$ Hz, 1H, OH), 5.14 (d, $J = 6.4$ Hz, 1H, OH), 6.78 (s, 1H, H-4), 8.35 (s, 1H, H-2). Anal. (C₁₂H₁₆O₇) C, H.

Ethyl 5-α-D-Ribofuranosylfuran-3-carboxylate (12α). Compound **12α** was prepared from **9α** (3 g, 7.5 mmol) by the above method and chromatographed on silica gel (CHCl₃-MeOH, 90:10) as a white foam (1.3 g, 65%). TLC (CHCl₃-MeOH, 90:10): $R_f = 0.36$. ¹H-NMR (DMSO-*d*₆): δ 1.28 (t, $J = 7.1$ Hz, 3H, OCH₂CH₃), 3.45 (pseudo-t, $J = 5.4$ Hz, 1H, H-5'), 3.58 (dd, $J = 2.5$, 5.4 Hz, 1H, H-5'), 3.79 (m, 1H, H-4'), 4.06 (m, 2H, H-2', H-3'), 4.23 (q, $J = 7.1$ Hz, 2H, OCH₂CH₃), 4.70 (t, $J = 5.6$ Hz, 1H, OH), 4.95 (m, $J = 2.8$ Hz, 3H, H-1', 2 OH, changes to a d with deuterium oxide), 6.65 (s, 1H, H-4), 8.3 (s, 1H, H-2). Anal. (C₁₂H₁₆O₇) C, H.

Ethyl 2,5-Di-β-D-ribofuranosylfuran-3-carboxylate (15). Compound **15** was prepared from **10** (7.5 g, 11.42 mmol) by the above method. A foam (2.5 g, 55%) was obtained after purification by flash chromatography on silica gel (CHCl₃-MeOH, 90:10). TLC (CHCl₃-MeOH, 80:20): $R_f = 0.24$. ¹H-NMR (DMSO-*d*₆): δ 1.29 (t, $J = 7.1$ Hz, 3H, OCH₂CH₃), 3.50 (m, 4H, H-5'a, H-5'b), 3.81 (m, 2H, H-4'a, H-4'b), 3.92 (m, 2H, H-3'a, H-3'b), 4.10 (m, 2H, H-2'a, H-2'b), 4.23 (q, $J = 7.1$ Hz, 2H, OCH₂CH₃), 4.61 (d, $J = 6.2$ Hz, 1H, H-1'b), 4.79, 5.0, and 5.15 (3m, 6H, OH), 5.30 (d, $J = 5.1$ Hz, 1H, H-1'a), 6.71 (s, 1H, H-4). Anal. (C₁₇H₂₄O₁₁) C, H.

Ethyl 2,5-Bis(2,3-isopropylidene-β-D-ribofuranosyl)furan-3-carboxylate (16). To stirring solution of **15** (1 g, 2.47 mmol) in acetone (25 mL) and dimethoxypropane (12 mL) cooled at 0 °C was added 0.08 mL of perchloric acid (70%) dropwise. The mixture was reacted for 1.5 h and neutralized at pH 7 by 1 N aqueous sodium hydroxide. After concentration *in vacuo*, the residue was purified by flash chromatography on silica gel eluting with 35% acetone in *n*-hexane yielding **16** as an oil (0.59 g, 50%). ¹H-NMR (CDCl₃): δ 1.30 (t, $J = 7.1$ Hz, 3H, CH₂CH₃), 1.32 (s, 6H, CCH₃), 1.53, 1.57 (2s, 6H, CCH₃), 3.12 (br s, 1H, OH), 3.52–3.88 (m, 4H, H-5'a, H-5'b), 4.12–4.33 (m, 4H, CH₂CH₃, H-4'a, and H-4'b), 4.69–4.84 (m, 5H, H-1'b, H-2'a, H-2'b, H-3'a, H-3'b), 5.52 (d, $J = 3.7$ Hz, 1H, H-1'a), 6.68 (s, 1H, H-4). ¹³C-NMR (CDCl₃): δ 14.7 (CH₂CH₃), 25.8, 27.8 (CH₃CCH₃), 61.4 (CH₂CH₃), 62.9, 63.1, 79.7, 80.1, 81.9, 84.2, 85.0, 85.7, 86.1 (carbons of the ribose skeleton), 110.0 (C-4), 114.8, 115.1 (CH₃CCH₃), 117.1 (C-2), 151.97 (C-5), 157.5 (C-3), 163.1 (C=O). Anal. (C₂₃H₃₂O₇) C, H.

2-β-D-Ribofuranosylfuran-3-carboxamide (13β). Treatment of compound **11β** (0.4 g, 1.5 mmol) with 30% ammonium

hydroxide (16 mL) for 8 h at room temperature and evaporation to dryness afforded a product which was purified by application to the top of a silica gel column followed by elution with CHCl₃-MeOH (70:30). Evaporation of the homogeneous fractions gave 0.19 g (55%) of **13β** as white foam. TLC (CHCl₃-MeOH 70:30): $R_f = 0.39$. ¹H-NMR (DMSO-*d*₆): δ 3.5 (m, 2H, H-5'), 3.75 (q, $J = 4.6$ Hz, 1H, H-4'), 3.92 (q, $J = 5.1$ Hz, 1H, H-3'), 4.15 (q, $J = 6.2$ Hz, 1H, H-2'), 4.72 (t, 1H, $J = 5.7$ Hz, OH), 4.94 (d, $J = 5.1$ Hz, 1H, OH), 5.03 (d, $J = 6.3$ Hz, 1H, OH), 5.38 (d, $J = 6.4$ Hz, 1H, H-1'), 6.85 (d, $J = 2.0$ Hz, 1H, H-4), 7.3, 7.58 (2br s, 2H, NH₂), 7.65 (d, $J = 2.0$ Hz, 1H, H-5). ¹³C-NMR (DMSO-*d*₆): δ 62.5 (C-5'), 71.8, 73.8, 75.4 (C-1', C-2', C-3'), 84.9 (C-4'), 110.3 (C-4), 122.8 (C-5), 142.3 (C-2), 155.1 (C-3), 164.4 (C=O). Anal. (C₁₀H₁₃NO₆) C, H, N.

5-β-D-Ribofuranosylfuran-3-carboxamide (4). Compound **4** was prepared as described for **13β** using **12β** (12 g, 44 mmol) (reaction time 4 h). The residue was chromatographed on silica gel with CHCl₃-MeOH (75:35) to give **4** as a white foam (6.4 g, 60%). TLC (CHCl₃-MeOH, 75:35): $R_f = 0.36$. UV (MeOH) λ_{max} 208 (ϵ 11 050), 242 nm (sh, ϵ 2050). ¹H-NMR (DMSO-*d*₆): δ 3.45 (m, 2H, H-5'), 3.75 (q, $J = 4.7$ Hz, 1H, H-4'), 3.90 (q, $J = 4.5$ Hz, 1H, H-3'), 4.05 (q, $J = 6.1$ Hz, 1H, H-2'), 4.58 (d, $J = 6.8$ Hz, 1H, H-1'), 4.78 (t, $J = 5.6$ Hz, 1H, OH), 4.98 (d, $J = 4.8$ Hz, 1H, OH), 5.1 (d, $J = 6.4$ Hz, 1H, OH), 6.79 (s, 1H, H-4), 7.2, 7.65 (2br s, 2H, NH₂), 8.12 (s, 1H, H-2). ¹³C-NMR (DMSO-*d*₆): δ 62.3 (C-5'), 71.5, 74.1, 77.1 (C-1', C-2', C-3'), 85.3 (C-4'), 107.9 (C-4), 123.6 (C-2), 145.4 (C-5), 154.5 (C-3), 163.7 (C=O). Anal. (C₁₀H₁₃NO₆) C, H, N.

5-α-D-Ribofuranosylfuran-3-carboxamide (14). The title compound was prepared from **12α** (1.2 g, 4.4 mmol). Chromatographic purification on a silica gel column eluting with CHCl₃-MeOH (70:30) afforded **14** as a white foam (0.55 g, 52%). TLC (CHCl₃-MeOH, 70:30): $R_f = 0.24$. ¹H-NMR (DMSO-*d*₆): δ 3.42 (m, 1H, H-5'), 3.56, 3.64 (2dd, $J = 2.4$, 5.4 Hz, 1H, H-5'), 3.80 (m, 1H, H-4'), 4.06 (m, 2H, H-2', H-3'), 4.73 (t, $J = 5.7$ Hz, 1H, OH), 4.92 (3d, $J = 3.5$ Hz, 3H, H-1', 2 OH, changes to a d with deuterium oxide), 6.75 (s, 1H, H-4), 7.15, 7.64 (2br s, 2H, NH₂), 8.07 (s, 1H, H-2). ¹³C-NMR (DMSO-*d*₆): δ 61.6 (C-5'), 72.1, 72.3, 76.3 (C-1', C-2', C-3'), 82.3 (C-4'), 107.7 (C-4), 123.4 (C-2), 144.5 (C-5), 153.6 (C-3), 163.6 (C=O). Anal. (C₁₀H₁₃NO₆) C, H, N.

2,5-Di-β-D-ribofuranosylfuran-3-carboxamide (17). Compound **17** was prepared from **15** (2 g, 4.9 mmol) as described for **13β**. The reaction residue was purified by flash chromatography on silica gel using CHCl₃-MeOH (65:35) to give **17** as a foam (0.69 g, 56%). TLC (CHCl₃-MeOH, 65:35): $R_f = 0.18$. ¹H-NMR (DMSO-*d*₆): δ 3.46–4.15 (m, 10H, H-5'a, H-5'b, H-4'a, H-4'b, H-3'a, H-3'b, H-2'a, H-2'b), 4.52 (d, $J = 6.0$, 1H, H-1'b), 4.72 (q, $J = 5.6$ Hz, 2H, 2 OH), 4.87 (d, $J = 6.7$ Hz, 1H, OH), 4.97 (d, $J = 5.5$ Hz, 1H, OH), 5.08 (d, $J = 6.3$ Hz, 1H, OH), 5.16 (d, $J = 4.7$ Hz, 1H, OH), 5.52 (d, $J = 3.4$ Hz, 1H, H-1'a), 6.79 (s, 1H, H-4), 7.21, 7.7 (2br s, 2H, NH₂). Anal. (C₁₅H₂₁NO₁₀) C, H, N.

Ethyl 2-(2,3,5-Tri-O-acetyl-β-D-ribofuranosyl)thiophene-3-carboxylate (19β), Ethyl 2-(2,3,5-Tri-O-acetyl-α-D-ribofuranosyl)thiophene-3-carboxylate (19α), Ethyl 5-(2,3,5-Tri-O-acetyl-β-D-ribofuranosyl)thiophene-3-carboxylate (20β), Ethyl 5-(2,3,5-Tri-O-acetyl-α-D-ribofuranosyl)thiophene-3-carboxylate (20α), and Ethyl 2,5-Bis(2,3,5-tri-O-acetyl)-β-D-ribofuranosylthiophene-3-carboxylate (21). The title compounds were obtained from ethyl thiophene-3-carboxylate (**18**)¹⁷ (15 g, 96 mmol) utilizing the same procedure used for preparation of **8**–**10**. The residue of CHCl₃ extracts was chromatographed on silica gel eluting with CH₂Cl₂-EtOAc (95:5). Three main fractions were separated. From the fast eluate, a mixture of **19β**, **19α**, and **20β** was obtained as a colorless oil (24.7 g, 62%). TLC (CH₂Cl₂-EtOAc, 95:5): $R_f = 0.43$.

Evaporation of the following fraction gave **20α** as an oil (3.06 g, 7.2%). TLC (CH₂Cl₂-EtOAc, 95:5): $R_f = 0.38$. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, $J = 7.0$ Hz, 3H, OCH₂CH₃), 2.00–2.10 (3s, 9H, OCOCH₃), 4.18–4.32 (m, 5H, H-5', H-4', OCH₂CH₃), 5.40 (dd, $J = 4.6$, 7.2 Hz, 1H, H-3'), 5.55 (t, $J = 3.9$ Hz, 1H, H-2'), 5.68 (d, $J = 3.1$ Hz, 1H, H-1'), 7.40 (d, $J = 1.3$ Hz, 1H, H-4), 8.34 (d, $J = 1.3$ Hz, 1H, H-2). Anal. (C₁₈H₂₂O₉S) C, H.

Evaporation of the last fraction gave ethyl 2,5-bis(2,3,5-tri-O-acetyl- β -D-ribofuranosyl)thiophene-3-carboxylate (**21**) as a colorless oil (3.9 g, 6%). TLC (CH₂Cl₂-EtOAc, 88:12): R_f = 0.1. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, J = 7.0 Hz, 3H, OCH₂CH₃), 1.98–2.18 (m, 18H, OCOCH₃), 4.15–4.40 (m, 8H, H-3'a, H-3'b, H-5'a, H-5'b, OCH₂CH₃), 5.20 (m, 3H, H-2'a, H-4'a, H-4'b), 5.40 (m, 1H, H-2'b), 5.83 (pseudo-t, J = 3.4 Hz, 1H, H-1'b), 6.07 (d, J = 3.3 Hz, 1H, H-1'a), 7.38 (s, 1H, H-4). Anal. (C₂₉H₃₆O₁₆S) C, H.

Ethyl 2- β -D-Ribofuranosylthiophene-3-carboxylate (22 β), Ethyl 2- α -D-Ribofuranosylthiophene-3-carboxylate (22 α), and Ethyl 5- β -D-Ribofuranosylthiophene-3-carboxylate (23 β). The mixture of **19 β** , **19 α** , and **20 β** (20 g) was treated with sodium ethoxide as described for the preparation of **11 β** . Purification of the reaction residue by chromatography on silica gel (CHCl₃-MeOH, 93:7) gave three main fractions which were separated. From the fast eluted fraction, **22 β** (2.22 g, 16%) was obtained as a colorless oil. TLC (CHCl₃-MeOH, 93:7): R_f = 0.47. ¹H-NMR (DMSO-*d*₆): δ 1.32 (t, J = 7.1 Hz, 3H, OCH₂CH₃), 3.6 (m, 2H, H-5'), 3.82 (m, 3H, H-2', H-3', H-4'), 4.25 (q, J = 7.1 Hz, 2H, OCH₂CH₃), 4.81 (t, J = 5.5 Hz, 1H, OH), 4.97 (d, J = 5.0 Hz, 1H, OH), 5.05 (d, J = 5.1 Hz, OH), 5.6 (d, J = 3.6 Hz, 1H, H-1'), 7.34 (d, J = 5.3 Hz, 1H, H-4), 7.5 (d, J = 5.3 Hz, 1H, H-5). Anal. (C₁₂H₁₆O₆S) C, H.

Evaporation of the following fraction gave **22 α** (1.25 g, 9%) as a colorless oil. TLC (CHCl₃-MeOH, 93:7): R_f = 0.42. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, J = 7.1 Hz, 3H, OCH₂CH₃), 3.47, 3.65 (2m, 2H, H-5'), 3.88 (m, 1H, H-4'), 4.10 (m, 2H, H-2', H-3'), 4.25 (q, J = 7.1 Hz, 2H, OCH₂CH₃), 4.75 (t, J = 5.8 Hz, 1H, OH), 4.81 (d, J = 4.7 Hz, 1H, OH), 4.85 (d, J = 7.3 Hz, OH), 5.71 (d, J = 3.1 Hz, 1H, H-1'), 7.33 (d, J = 5.3 Hz, 1H, H-4), 7.45 (d, J = 5.1 Hz, 1H, H-5). Anal. (C₁₂H₁₆O₆S) C, H.

Evaporation of the last fraction gave **23 β** (8.62 g, 62%) as white needles (crystallized from CHCl₃). Mp: 117–119 °C. TLC (CHCl₃-MeOH, 90:10) R_f = 0.45. ¹H-NMR (DMSO-*d*₆): δ 1.32 (t, J = 7.1 Hz, 3H, OCH₂CH₃), 3.60 (m, 2H, H-5'), 3.85 (m, 3H, H-2', H-3', H-4'), 4.27 (q, J = 7.1 Hz, 2H, OCH₂CH₃), 4.83 (t, J = 5.5 Hz, 1H, OH-5'), 5.02 (d, J = 4.0 Hz, 1H, OH), 5.08 (d, J = 5.0 Hz, 1H, OH), 5.60 (d, J = 3.4 Hz, 1H, H-1'), 7.35 (d, J = 5.3 Hz, 1H, H-4), 7.5 (d, J = 5.3 Hz, 1H, H-5). Anal. (C₁₂H₁₆O₆S) C, H.

Ethyl 5- α -D-Ribofuranosylthiophene-3-carboxylate (23 α). Compound **23 α** was obtained from **20 α** (3 g, 7.23 mmol) as described for **22 β** , as a colorless oil (1.4 g, 67%), after chromatography on silica gel with CHCl₃-MeOH (94:6). TLC (CHCl₃-MeOH, 96:4) R_f = 0.20. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, J = 7.1 Hz, 3H, OCH₂CH₃), 3.45 (m, 1H, H-5'), 3.55–3.65 (2dd, J = 4.7, 11.9 Hz, 1H, H-5'), 3.83 (m, 1H, H-4'), 3.96 (dd, J = 4.0, 7.5 Hz, 1H, H-3'), 4.11 (m, 1H, H-2'), 4.25 (q, J = 7.1 Hz, 2H, OCH₂CH₃), 4.73 (t, J = 5.7 Hz, 1H, OH), 4.95 (d, J = 6.3 Hz, 1H, OH), 5.12 (d, J = 4.4 Hz, 1H, OH), 5.20 (d, J = 2.9 Hz, 1H, H-1'), 7.33 (d, J = 1.4 Hz, 1H, H-4), 8.23 (d, J = 1.4 Hz, 1H, H-2). Anal. (C₁₂H₁₆O₆S) C, H.

Ethyl 2,5-Di- β -D-ribofuranosylthiophene-3-carboxylate (24). Compound **24** was prepared from **21** (3.8 g, 5.65 mmol) by the above method. The reaction residue was purified by flash chromatography, using CHCl₃-MeOH (80:20), to give **24** as a foam (1.28 g, 55%). TLC (CHCl₃-MeOH, 80:20): R_f = 0.33. ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, J = 7.1 Hz, 3H, OCH₂CH₃), 3.48 (t, J = 5.1 Hz, 2H, H-5'b), 3.58 (m, 2H, H-5'a), 3.80 (m, 4H, H-4'a, H-4'b, H-3'a, H-3'b), 3.92 (m, 2H, H-2'a, H-2'b), 4.23 (q, J = 7.1 Hz, 2H, OCH₂CH₃), 4.72 (d, J = 7.2 Hz, 1H, H-1'b), 4.80–5.21 (m, 6H, OH), 5.60 (d, J = 3.8 Hz, 1H, H-1'a), 7.28 (s, 1H, H-4). Anal. (C₁₇H₂₄O₁₀S) C, H.

Ethyl 2,5-Bis(2,3-isopropylidene- β -D-ribofuranosyl)-thiophene-3-carboxylate (25). The title compound was prepared from **24** (1 g, 2.38 mmol) as described for **16**. The reaction mixture was purified by flash chromatography eluting with 35% acetone in *n*-hexane to give **25** as an oil (0.59 g, 50%). ¹H-NMR (CDCl₃): δ 1.30 (t, J = 7.1 Hz, 3H, CH₂CH₃), 1.31 (s, 6H, CCH₃), 1.43, 1.46 (2s, 6H, CCH₃), 3.15 (br s, 1H, OH), 3.54–3.94 (m, 4H, H-5'a, H-5'b), 4.06–4.33 (m, 4H, CH₂CH₃, H-4'a, H-4'b), 4.60–4.75 (m, 3H, H-2'b, H-3'a, H-3'b), 4.90–5.01 (m, 2H, H-1'b, H-2'a), 5.81 (d, J = 3.7 Hz, 1H, H-1'a), 7.18 (s, 1H, H-4). ¹³C-NMR (CDCl₃): δ 14.7 (CH₂CH₃), 26.0, 28.1 (CH₃CCH₃), 61.4 (CH₂CH₃), 63.0, 63.2, 81.4, 82.1, 82.2,

82.3, 84.7, 86.1, 86.2, 87.0 (carbons of the ribose skeleton), 114.6, 115.5 (CH₃CCH₃), 124.7 (C-4), 128.4 (C-2), 141.7 (C-5), 148.0 (C-3), 163.4 (C=O). Anal. (C₂₃H₃₂O₆S) C, H.

2- β -D-Ribofuranosylthiophene-3-carboxamide (26 β). Compound **26 β** was obtained from **22 β** (2 g, 6.93 mmol) by the above method (reaction time 8 h). The reaction residue was chromatographed on silica gel with CHCl₃-MeOH (80:20) to give **26 β** as a white foam (1.1 g, 62%). TLC (CHCl₃-MeOH, 80:20): R_f = 0.21. ¹H-NMR (DMSO-*d*₆): δ 3.4 (m, 2H, H-5'), 3.6 (m, 1H, H-4'), 3.85 (m, 1H, H-3'), 4.12 (m, 1H, H-2'), 4.9 (br s, 2H, OH), 5.17 (dd, J = 5.1, 9.1 Hz, 1H, H-1'), 5.20 (d, J = 5.1 Hz, 1H, OH), 7.38 (d, J = 9.16 Hz, 1H, NH), 7.38 (s, 2H, H-4, H-5), 8.31 (d, J = 9.16 Hz, 1H, NH). ¹³C-NMR (DMSO-*d*₆): δ 61.7 (C-5'), 72.6, 73.1, 78.4 (C-1', C-2', C-3'), 82.3 (C-4'), 125.3 (C-5), 129.6 (C-4), 136.1 (C-2), 142.6 (C-3), 164.4 (C=O). Anal. (C₁₀H₁₃NO₅S) C, H, N.

2- α -D-Ribofuranosylthiophene-3-carboxamide (26 α). Compound **26 α** was prepared from **22 α** (1 g, 3.46 mmol) (reaction time 9 h) and purified by chromatography on silica gel (CHCl₃-MeOH, 75:25) to give a white foam (0.52 g, 58%). TLC (CHCl₃-MeOH, 75:25): R_f = 0.41. ¹H-NMR (DMSO-*d*₆): δ 3.45 (m, H, H-5'), 3.62 (m, 1H, H-5'), 3.82 (m, 1H, H-4'), 4.08–4.24 (m, 2H, H-2', H-3'), 4.72 (pseudo-t, 2H, OH), 4.85 (d, J = 6.0 Hz, 1H, OH), 5.77 (d, J = 2.7 Hz, 1H, H-1'), 7.2 (br s, 1H, NH), 7.4 (s, 2H, H-4, H-5), 7.65 (br s, 1H, NH). ¹³C-NMR (DMSO-*d*₆): δ 61.5 (C-5'), 72.5, 72.6, 78.9 (C-1', C-2', C-3'), 82.3 (C-4'), 124.2 (C-5), 126.8 (C-4), 130.9 (C-2), 149.7 (C-3), 165.2 (C=O). Anal. (C₁₀H₁₃NO₅S) C, H, N.

5- β -D-Ribofuranosylthiophene-3-carboxamide (5). The title compound was obtained from **23 β** (5 g, 17.3 mmol) (reaction time 9 h) by the above method. After chromatographic purification of the reaction mixture (CHCl₃-MeOH, 80:20), **5** was obtained as a white foam (3.1 g, 70%). TLC (CHCl₃-MeOH, 80:20): R_f = 0.31. UV (MeOH) λ_{max} 210 (ϵ 35 000), 240 nm (ϵ 10 500). ¹H-NMR (DMSO-*d*₆): δ 3.50 (t, J = 5.1 Hz, 2H, H-5'), 3.81 (m, 2H, H-4', H-3'), 3.93 (m, 1H, H-2'), 4.76 (d, J = 8.0 Hz, 1H, H-1'), 4.80 (t, J = 5.5 Hz, 1H, OH), 5.01 (d, J = 4.7 Hz, 1H, OH), 5.18 (d, J = 6.8 Hz, 1H, OH), 7.20 (br s, 1H, NH), 7.42 (s, 1H, H-4), 7.75 (br s, 1H, NH), 8.02 (s, 1H, H-2). ¹³C-NMR (DMSO-*d*₆): δ 62.3 (C-5'), 71.6 (C-3'), 77.7 (C-2'), 79.3 (C-1'), 85.6 (C-4'), 124.5 (C-4), 128.3 (C-2), 137.4 (C-5), 145.3 (C-3), 165.2 (C=O). Anal. (C₁₀H₁₃NO₅S) C, H, N.

5- α -D-Ribofuranosylthiophene-3-carboxamide (27). Compound **27** was prepared from **23 α** (1 g, 3.46 mmol) (reaction time 9 h) by the above method. The reaction residue was chromatographed on a silica gel column with CHCl₃-MeOH (80:20) to give **27** (0.49 g, 55%) as a white solid. Mp: 155–158 °C. TLC (CHCl₃-MeOH, 80:20): R_f = 0.26. ¹H-NMR (DMSO-*d*₆): δ 3.46 (m, 1H, H-5'), 3.58–3.65 (2m, 1H, H-5'), 3.81 (m, 1H, H-4'), 3.96 (dd, J = 4.3, 7.6 Hz, 1H, H-3'), 4.12 (m, 1H, H-2'), 4.73 (t, J = 5.7 Hz, 1H, OH), 4.92 (d, J = 7.4 Hz, 1H, OH), 5.08 (d, J = 4.7 Hz, 1H, OH), 5.18 (d, J = 2.8 Hz, 1H, H-1'), 7.15 (br s, 1H, NH), 7.35 (s, 1H, H-4), 7.72 (br s, 1H, NH), 8.04 (d, J = 1.3 Hz, 1H, H-2). ¹³C-NMR (DMSO-*d*₆): δ 61.7 (C-5'), 72.6 (C-3'), 73.1 (C-2'), 78.3 (C-1'), 82.3 (C-4'), 125.3 (C-4), 129.6 (C-2), 136.1 (C-5), 142.6 (C-3), 164.4 (C=O). Anal. (C₁₀H₁₃NO₅S) C, H, N.

2,5-Di- β -D-ribofuranosylthiophene-3-carboxamide (28). Compound **28** was obtained from **24** (1 g, 2.37 mmol) by the above method and purified by flash chromatography (CHCl₃-MeOH, 80:20) as a foam (0.45 g, 50%). TLC (CHCl₃-MeOH, 60:40): R_f = 0.16. ¹H-NMR (DMSO-*d*₆) (some signals are doubled for the presence of two conformers): δ 3.46, 3.63 (2m, 4H, H-5'a, H-5'b), 3.83 (m, 4H, H-4'a, H-4'b, H-3'a, H-3'b), 4.12, 4.20 (2m, 2H, H-2'a, H-2'b), 4.65–5.0 (m, 6H, OH), 4.70, 5.12 (2d, 1H, H-1'b), 5.72, 5.80 (2d, 1H, H-1'a), 7.10, 7.17 (2br s, 1H, NH), 7.25, 7.30 (2s, 1H, H-4), 7.57, 7.62 (2br s, 1H, NH). Anal. (C₁₅H₂₁NO₉S) C, H, N.

Crystallographic Study. Small colorless crystals of the carboxylate precursor of thiophenfuran [ethyl 5- β -D-ribofuranosylthiophene-3-carboxylate, C₁₂H₁₆O₆S (**23 β**)] were obtained from chloroform. Crystals are orthorhombic, space group *P*2₁2₁2₁, with cell dimensions *a* = 4.893(2) Å, *b* = 12.764(11) Å, and *c* = 4. Diffraction data were collected on an Enraf-Nonius FAST detector using graphite monochromatized Mo

K α radiation from a rotating anode source. A total of 14 132 reflections were measured to $\theta = 25.3^\circ$ and averaged to yield 1897 unique reflections. The structure was solved by routine application of direct methods and refined to $R = 6.6\%$ ($R_w = 6.5\%$) for the 1312 reflections with values of $F_o > 2\sigma(F_o)$. All non-hydrogen atoms were refined with anisotropic temperature factors using full-matrix least-squares techniques. Thiophene and ribose methylene protons were added in idealized positions and then refined with isotropic temperature factors. High thermal parameters in the ethyl carbons indicated some disorder in this portion of the carboxylate substituent.

Computational Study. Energy as a function of C-glycosidic torsion angle χ was computed for thiophenfurin and furanfurin model fragments using methods similar to those described previously.^{8,11} *Ab initio* calculations were performed using the Gaussian 92 system of programs and internal basis sets.¹⁸

Model fragments for thiophenfurin and furanfurin contained thiophene and furan heterocycles, respectively, with the carboxamide substituent omitted. The heterocycle was connected via a C-glycosidic bond to a 3'-deoxyfuranose ring (Figure 2). By analogy to tiazofurin, χ was defined by atoms O-1'-C-1'-C-2-S-1 and O-1'-C-1'-C-2-O-1 for thiophenfurin and furanfurin, respectively. A value of $\chi = 0^\circ$ refers to the conformation in which the furanose oxygen is *cis* planar to either the furan oxygen or the thiophene sulfur. A positive value of χ indicates a counterclockwise rotation of the C-2-O-1/S bond relative to the C-1'-O-1' bond when viewed down the C-glycosidic bond from C-2 to C-1'.

Energy profiles for the thiophenfurin and furanfurin model compounds were obtained with the furanose ring fixed in both 3'-endo and 2'-endo puckers. The global energy minimum for each compound was normalized to 0 kcal/mol. Energies for each model were obtained for conformers between $\chi_0 = -180^\circ$ and 180° . In each case, the value of χ was incremented in 20° steps and fixed. All remaining geometry variables describing the fragment were then optimized, with the exception of the furanose torsion angles. The starting geometry at each value of χ was the optimized geometry obtained at the previous value. All geometry optimizations used the analytical gradient method.¹⁹

Optimized geometries and associated SCF energies were obtained using the 3-21G basis set for the furanfurin model and the 3-21G* basis set for the sulfur-containing thiophenfurin model. Thus, each point in Figure 2 represents a calculation at either the RHF/3-21G//3-21G or RHF/3-21G**/3-21G* levels.¹⁸

Point charges were obtained from both thiazole and oxazole fragments using the natural population analysis method incorporated in Gaussian 92.^{16,18} This method of charge partitioning uses fewer assumptions than that of the traditional Mulliken population analysis.^{19,20} Charges were obtained from optimized geometries at the RHF level of theory using a 3-21G basis set for the furanfurin model and a 3-21G* basis set for the thiophenfurin fragment.^{19,20}

Antitumor Evaluation. The following cell lines were used, L1210 and P388 murine leukemia, HL-60 human promyelocytic leukemia, K562 human myelogenous leukemia, Lewis lung carcinoma (LLC), B16 murine melanoma, and LoVo human colon adenocarcinoma.

With the exception of LoVo, cell lines were grown in RPMI 1640 medium, supplemented with 10 mM HEPES buffer and 15% new born calf serum (L1210 and P388) or 5% (LLC) or 10% (K562 and B16) or 15% (HL-60) fetal calf serum. LoVo cells were maintained in HAM's F12 medium supplemented with 10% fetal calf serum. Antibiotics were added to all cell culture media. 2-Mercaptoethanol (10 nM) was also added to P388, L1210, and HL-60, while 1% vitamins and nonessential amino acids were added to LoVo and B16. The cells were grown at 37°C in a CO_2 incubator and maintained in logarithmic phase of growth.

Various concentrations of each drug were placed with tumor cell suspension (B16, 5×10^3 cells/well; P388, L1210, and LLC, 10^4 cells/well; HL-60, K562, and LoVo, 2.5×10^5 cells/well). Cell growth was evaluated after 72 h of cell-drug contact using a modified MTT assay.^{16,21} The IC_{50} and IC_{90} (con-

centration reducing optical absorption by 50% or 90%, respectively) were calculated by the regression analysis proposed by Chou.²²

Relative Toxicity Evaluation *in Vitro*. The evaluation of *in vitro* relative toxicity as a new prescreening model for agents potentially active against tumors is described in a recent paper.²³ Briefly, the cytotoxicity of a test drug is evaluated by using an antiproliferative assay which was developed to evaluate *in vitro* antitumor activity.¹⁰ This measures [^{125}I]-5-iodo-2'-deoxyuridine ([^{125}I]UdR) incorporation in DNA after a 72 h drug-cell contact. The potential antitumor activity of new agents is estimated by comparing the cytotoxic activity on normal (human bone marrow) and tumor (human cell line HL-60) cells. In this system the ratios of the mean $\text{IC}_{50}(\text{BM})/\text{IC}_{50}(\text{HL-60})$ and the mean $\text{IC}_{90}(\text{BM})/\text{IC}_{90}(\text{HL-60})$ were greater than 1 for drugs in clinical use. In contrast, this ratio is less than 1 for compounds which have demonstrated only good cytotoxic activity toward tumor cell lines but are not clinically used. Thus, a new compound which has a ratio greater than 1 is considered to have a greater probability of becoming a clinically useful antitumor agent than a compound with a ratio lower than 1. In these sets of experiments, assays on BM samples were conducted from five different donors and three assays on HL-60 cells were performed.

Antitumor Activity *in Vivo*. The test animals used in this experiment were female BD $_2$ F $_1$ mice (8 weeks old, 20–22 g), purchased from Charles River, Italy. Each experimental group contained eight mice which were injected ip with 10^5 L1210 cells on day 0. Twenty-four hours later, drugs were solubilized in a 0.9% NaCl solution and injected ip once a day for 9 days.

Formation of NAD Analogues. NAD analogues of tiazofurin, thiophenfurin, and furanfurin were synthesized by the method cited.²⁴ Briefly, human myelogenous leukemia K562 cells (1×10^7 cells/10 mL) were incubated with [2,8- ^3H]-adenosine (185 nM, specific radioactivity 200 mCi/mmol) for 1 h at 37°C . Saline or indicated concentration of analogue was then added, incubated further for 2 h, and then centrifuged at 400g for 5 min. The cell pellet was washed once with phosphate-buffered saline, homogenized in cold 10% trichloroacetic acid, and neutralized with 0.5 M tri-*n*-octylamine in freon and an aliquot analyzed on a Partisil 10-SAX column using a RCM-10 module employing high-pressure liquid chromatography (HPLC) utilizing an ammonium phosphate buffer system.²⁵

IMPDH Assay. K562 cells (1×10^6 cells/mL, 10 mL) in logarithmic phase of growth were incubated with saline or indicated concentrations of the agents for 2 h at 37°C . The cells were then harvested by centrifugation, washed once with cold phosphate-buffered saline, and lysed in buffer containing 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 10 mM MgCl_2 , 0.5% NP-40, and 2 $\mu\text{g/mL}$ aprotinin. The lysate was kept for 20 min on ice and centrifuged at 13000g for 20 min, and the supernatant was used as a source of enzyme.

The enzyme activity was measured according to the published methodology.²⁶ Briefly, 5 μL aliquots of 0.5 M KCl containing 20 mM allopurinol were dispensed into the apex of Eppendorf tubes and dried at room temperature (25°C). For the conduct of the assay, in a total volume of 10 μL , tubes contained 5 μL of the substrate mixture containing 286 μM [2,8- ^3H]IMP (200 $\mu\text{Ci/mL}$) and 1 mM NAD. The reaction was initiated by the addition of a 5 μL aliquot of enzyme extract. After 30 min at 37°C , the reaction was terminated by heating for 1 min at 95°C . Tubes were centrifuged at 13000g for 0.5 min, 5 μL of 100% KOH was deposited on the underside of the cap, and the tubes were closed and incubated at room temperature overnight (16 h). The caps containing the droplet were cut, and the radioactivity was determined by scintillation spectrometry. IMPDH activity was expressed as nmol of XMP formed/mg of protein/h.

Determination of the Concentration of Intracellular Ribonucleotides. Cells in culture were treated with saline or agents for 2 h at 37°C ; cells were harvested by centrifugation and washed once with cold saline. Cells were extracted with cold 10% trichloroacetic acid and centrifuged for 0.5 min;

the supernatant was immediately neutralized with 0.5 M tri-*n*-octylamine in freon. An aliquot of the neutralized extract was analyzed on HPLC using a Partisil 10-SAX column as described above.

Acknowledgment. This work was supported by Ministero dell'Università e della Ricerca Scientifica e Tecnologica (40% funds), CNR, Italy, NIH Grant CA45145, and the Project Development Program, Research and Sponsored Programs, Indiana University at Indianapolis, IN.

Supporting Information Available: Coordinates, thermal parameters, and bond lengths and angles (3 pages); table of observed and calculated structure factors for thiophenfuran-4-carboxylate (**23β**) (10 pages). Ordering information is given on any current masthead page.

References

- (1) (a) Srivastava, P. C.; Pickering, M. V.; Allen, L. B.; Streeter, D. G.; Campbell, M. T.; Witkowski, J. T.; Sidwell, R. W.; Robins, R. K. Synthesis and Antiviral Activity of Certain Thiazole C-Nucleosides. *J. Med. Chem.* **1977**, *20*, 256–262. (b) Srivastava, P. C.; Robins, R. K. Synthesis and Antitumor Activity of 2-β-D-Ribofuranosylthiazole-4-carboxamide and Related Derivatives. *J. Med. Chem.* **1983**, *26*, 445–448. (c) Robins, R. K.; Srivastava, P. C.; Narayanan, V. L.; Powman, J.; Paull, K. D. 2-β-D-Ribofuranosylthiazole-4-carboxamide, a Novel Potential Antitumor Agent for Lung Tumors and Metastases. *J. Med. Chem.* **1982**, *25*, 107. (d) Jayaram, H. N.; Smith, A. L.; Glazer, R. I.; Johns, D. G.; Cooney, D. A. Studies on the mechanism of action of 2-β-D-ribofuranosylthiazole-4-carboxamide (NSC 286193): II Relationship between dose level and biochemical effects in P388 leukemia *in vivo*. *Biochem. Pharmacol.* **1982**, *31*, 3839–3845. (e) Zhen, W.; Jayaram, H. N.; Weber, G. Antitumor activity of tiazofurin in human colon carcinoma HT-29. *Cancer Invest.* **1992**, *10*, 505–511. (f) Jayaram, H. N.; Ahluwalia, G. S.; Dion, R. L.; Gebeyehu, G.; Marquez, V. E.; Kelley, J. A.; Robins, R. K.; Cooney, D. A.; Johns, D. G. Conversion of 2-β-D-ribofuranosylthiazole-4-carboxamide to an analogue of NAD with potent IMP dehydrogenase-inhibitory properties. *Biochem. Pharmacol.* **1983**, *32*, 2633–2636.
- (2) Goldstein, B. M.; Leary, J. F.; Farley, B. A.; Marquez, V. E.; Rowley, P. T. Induction of HL60 Cell Differentiation by Tiazofurin and its Analogues: Characterization and Efficacy. *Blood* **1991**, *78*, 593–598.
- (3) Parandoosh, Z.; Rubalcava, B.; Matsumoto, S. S.; Jolley, W. B.; Robins, R. K. Changes in Diacylglycerol and Cyclic GMP During the Differentiation of Human Myeloid Leukemia K562 Cells. *Life Sci.* **1990**, *46*, 315–320.
- (4) Kharbanda, S. M.; Sherman, M. L.; Springgs, D. R.; Kufe, D. W. Effects of tiazofurin on protooncogene expression during HL-60 Cells differentiation. *Cancer Res.* **1988**, *48*, 5965–5968.
- (5) Smith, R. A. In *Ribavirin: A Broad Spectrum Antiviral Agent*; Smith, R. A., Kirkpatrick, W., Eds.; Academic Press, Inc.: New York, 1980; pp 99–118.
- (6) Jayaram, H. N.; Dion, R. L.; Glazer, R. I.; Johns, D. G.; Robins, R. K.; Srivastava, P. C.; Cooney, D. A. Initial studies on the Mechanism of Action of a New Oncolytic Thiazole Nucleoside, 2-β-D-Ribofuranosylthiazole-4-carboxamide. *Biochem. Pharmacol.* **1982**, *31*, 2371–2380.
- (7) (a) Cooney, D. A.; Jayaram, H. N.; Gebeyehu, G.; Betts, C. R.; Kelley, J. A.; Marquez, V. E.; Johns, D. G. The conversion of 2-β-D-Ribofuranosyl-thiazole-4-carboxamide to an analogue of NAD with potent IMP dehydrogenase-inhibitory properties. *Biochem. Pharmacol.* **1982**, *31*, 2133–2136. (b) Kuttan, R.; Robins, R. K.; Saunders, P. P. Inhibition of inosinate dehydrogenase by metabolites of 2-β-D-Ribofuranosylthiazole-4-carboxamide. *Biochem. Biophys. Res. Commun.* **1982**, *107*, 862–868.
- (8) Burling, F. T.; Goldstein, B. M. Computational Studies of Nonbonded Sulfur-Oxygen and Selenium-Oxygen Interactions in the Thiazole and Selenazole Nucleosides. *J. Am. Chem. Soc.* **1991**, *114*, 2313–2320 and references therein.
- (9) Li, H.; Hallows, W. A.; Punzi, J. S.; Marquez, V. E.; Carrell, H. L.; Pankiewicz, K. W.; Watanabe, K. A.; Goldstein, B. M. Crystallographic Studies of Two Alcohol Dehydrogenase-bound Analogues of Thiazole-4-Carboxamide Adenine Dinucleotide (TAD), the Active Anabolite of the Antitumor Agent Tiazofurin. *Biochemistry* **1994**, *33*, 23–32.
- (10) (a) Franchetti, P.; Cristalli, G.; Grifantini, M.; Cappellacci, L.; Vittori, S.; Nocentini, G. Synthesis and Antitumor Activity of 2-β-D-Ribofuranosyl-oxazole-4-carboxamide (Oxazofurin). *J. Med. Chem.* **1990**, *33*, 2849–2852. (b) Franchetti, P.; Messini, L.; Cappellacci, L.; Grifantini, M.; Guarracino, P.; Marongiu, M. E.; Piras, G.; La Colla, P. A New Facile Synthesis and Antiviral Activity of Oxazofurin. *Nucleosides Nucleotides* **1993**, *11*, 359–368.
- (11) Goldstein, B. M.; Hallows, W.; Langs, D.; Franchetti, P.; Cappellacci, L.; Grifantini, M. C-glycosyl Bond Conformation in Oxazofurin: Crystallographic and Computational Studies of the Oxazole Analogue of Tiazofurin. *J. Med. Chem.* **1994**, *37*, 1684–1688.
- (12) Girgis, N. S.; Michael, M. A.; Smee, D. F.; Alaghamandan, H. A.; Robins, R. K.; Cottam, H. B. Direct C-Glycosylation of Guanine Analogues: The Synthesis and Antiviral Activity of Certain 7- and 9-Deazaguanine C-Nucleosides. *J. Med. Chem.* **1990**, *33*, 2750–2755 and references therein.
- (13) Rosemeyer, H.; Toth, G.; Seela, F. Assignment of Anomeric Configuration of D-Ribo-, Arabino-, 2'-Deoxyribo-, and 2',3'-Dideoxyribonucleosides by NOE Difference Spectroscopy. *Nucleosides Nucleotides* **1989**, *8*, 587–597.
- (14) Rao, S. P.; Rao, K. V. B.; Otter, B. A.; Klein, R. S.; Ren, W.-Y. C-Glycosylation of Substituted Heterocycles Under Friedel-Crafts Conditions (I): A Two-step Synthesis of the Thieno[3,4-d]pyrimidine C-Nucleoside Analog of Inosine. *Tetrahedron Lett.* **1988**, *29*, 3537–3540 and references therein.
- (15) (a) Ohri, H.; Jones, G. H.; Moffatt, J. G.; Maddox, M. L.; Christensen, A. T.; Byram, S. K. C-Glycosyl Nucleosides. V. Some Unexpected Observations on the Relative Stabilities of Compounds Containing Fused Five-Membered Rings with Epimerizable Substituents. *J. Am. Chem. Soc.* **1975**, *97*, 4602–4613. (b) Cousineau, T. J.; Secrist, J. A. III Synthesis of Certain β-D-Ribofuranosylthiazole C-Nucleosides from a Versatile Precursor. *J. Org. Chem.* **1979**, *44*, 4351–4358.
- (16) Scudiero, D. A.; Shoemaker, R. H.; Paull, K. D.; Monks, A.; Tierney, S.; Nofziger, T. H.; Cuneo, M. J.; Seniff, D.; Boyd, M. R. Evaluation of a Soluble Tetrazolium/formazan Assay for Cell Growth and Drug Sensitivity in Culture Using Human and other Tumor Cell Lines. *Cancer Res.* **1988**, *48*, 4827–4833.
- (17) Prepared from 3-thiophenecarboxylic acid. For preparation see: Campaigne, E.; LeSuer, W. M. *J. Am. Chem. Soc.* **1948**, *70*, 1555–1558. Yield: 86% (lit. 76.7%). ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, *J* = 7.1 Hz, 3H, OCH₂CH₃), 4.28 (q, *J* = 7.1 Hz, 2H, OCH₂CH₃), 7.48 (dd, *J* = 1.3, 5.1 Hz, 1H, H-4), 7.66 (dd, *J* = 2.9, 5.1 Hz, 1H, H-5), 8.33 (dd, *J* = 1.5, 2.8 Hz, 1H, H-2).
- (18) Frisch, M. J.; Trucks, G. W.; Head-Gordon, M.; Gill, P. M. W.; Wong, M. W.; Foresman, J. B.; Johnson, B. G.; Schlegel, H. B.; Robb, M. A.; Replogle, E. S.; Gomperts, R.; Andres, J. L.; Reghavarachi, K.; Binkley, J. S.; Gonzalez, C.; Martin, R. L.; Fox, D. J.; Defrees, D. J.; Baker, J.; Stewart, J. J. P.; Pople, J. A. *Gaussian 92, Revision B*; Gaussian, Inc.: Pittsburgh, PA, 1992.
- (19) Hehre, J. W.; Radom, L.; Schleyer, P. V. R.; Pople, J. A. *Ab Initio Molecular Orbital Theory*; John Wiley-Interscience Publication: New York, 1986; Chapter 4.
- (20) Reed, A. E.; Curtiss, L. A.; Weinhold, F. Intermolecular Interactions from a Natural Bond Orbital, Donor-Acceptor Viewpoint. *Chem. Rev.* **1988**, *88*, 889–926.
- (21) Mosmann, T. Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity assay. *J. Immunol. Methods* **1983**, *65*, 55–63.
- (22) Chou, T. C. On the determination of availability of ligand binding sites in steady-state system. *J. Theor. Biol.* **1977**, *65*, 345–356.
- (23) Nocentini, G.; Barzi, A. A Predictive Screening Model for *in vitro* Selection of Agents with Potential Antitumor Activity. *Arzneim.-Forsch. Drug Res.*, in press.
- (24) Gharehbaghi, K.; Sreenath, A.; Hao, Z.; Paull, K. D.; Szekeres, T.; Cooney, D. A.; Krohn, K.; Jayaram, H. N. Comparison of biochemical parameters of benzamide riboside, a new inhibitor of IMP dehydrogenase, with tiazofurin and selenazofurin. *Biochem. Pharmacol.* **1994**, *48*, 1413–1419.
- (25) Zhen, W.; Jayaram, H. N.; Wever, G. Determination of TAD (thiazole-4-carboxamide adenine dinucleotide) levels in mononuclear cells of leukemic patients treated with tiazofurin. *Biochem. Pharmacol.* **1991**, *41*, 281–286.
- (26) Cooney, D. A.; Wilson, Y.; McGee, E. A straightforward and radiometric technique for measuring IMP dehydrogenase. *Anal. Biochem.* **130**, 339–345.

JM950282R