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## Spiroidesin, a Novel Lipopeptide from the Cyanobacterium *Anabaena spiroides* That Inhibits Cell Growth of the Cyanobacterium Microcystis aeruginosa

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Spiroidesin (1), a novel D-amino acid-containing linear lipopeptide, was isolated from waterblooms of the cyanobacterium Anabaena spiroides. The structure was identified by 2D NMR and chemical degradation analyses. Spiroidesin inhibited cell growth of the toxic cyanobacterium Microcystis aeruginosa (IC<sub>50</sub>, 1.6  $\times 10^{-6} \, \mathrm{M}$ ).

Recently, cyanobacteria have been reported to produce bioactive lipopeptides. 1-6 In many cases, amino acids found in these linear peptides possess only an L- or both D- and L-configurations, and the N-terminal amino acids are combined with fatty acids. These peptides have various kinds of biological activities such as protease inhibition, 1-3 neurotoxic,4 and ichthyotoxic5 properities. During investigations into toxins from waterblooms of Anabaena spiroides, we have found a novel D- and L-homotyrosinecontaining linear lipopeptide. Herein we report the structure and biological activity of the novel linear lipopeptide, spiroidesin (1).

Spiroidesin (1) is a colorless amorphous solid:  $\lambda_{max}$  (H<sub>2</sub>O) 276 nm ( $\epsilon$  2200); [ $\alpha$ ]<sup>25</sup><sub>D</sub>  $-62^{\circ}$  (c 0.56, MeOH). In the positive HRFABMS using glycerol as the matrix, the  $[M + H]^+$  ion was observed at m/z 618.3134. From the results, the molecular formula of 1 was established to be  $C_{35}H_{43}N_3O_7$ (calcd for [M + H] 618.3179,  $\Delta$  -4.5 mmu). The spectral data (Table 1) of <sup>1</sup>H and <sup>13</sup>C NMR of 1 suggested that spiroidesin is a fatty acid-containing peptide. In the nin-

**Table 1.** <sup>1</sup>H<sup>a</sup> and <sup>13</sup>C<sup>b</sup> NMR Data for Spiroidesin (1) in

Methanol- $d_4$			
position	<sup>1</sup> H <i>J</i> (Hz)	<sup>13</sup> C	$HMBC^d$
$Hex^c$			
1		176.4	2,3, D-Htyr-2
2	2.22 (t, 7.4)	36.8	
3	1.57 (q, 7.4)	26.6	
4	1.27 (m)	32.6	
5	1.25 (m)	23.4	
6	0.82 (t, 7.0)	14.2	
D-Htyr	,		
1		174.7	2,3, L-Htyr-2
2	4.24 (dd, 4.9, 9.5)	54.2	, , ,
3	1.86 (m)	35.3	
	1.71 (m)		
4	2.36 (m)	32.0	6, 10
	2.28 (m)		-,
5		132.5	4
6, 10	6.83 (d, 8.6)	130.3	•
7, 9	6.62 (d, 8.6)	116.3	
8	0.02 (d, 0.0)	157.0	
L-Htyr		107.0	
1		173.1	2,3, D-Phe-2
2	4.21 (dd, 5.2, 9.2)	55.2	ω,5, D-1 HC-ω
3	1.97 (m)	34.9	
0	1.88 (m)	01.0	
4	2.60 (m)	32.4	6, 10
<b>T</b>	2.53 (m)	32.4	0, 10
5	2.33 (III)	132.7	4
6, 10	6.98 (d, 8.6)	130.4	4
7, 9	6.65 (d, 8.6)	116.5	
8	0.03 (d, 8.0)	157.3	
D-Phe		137.3	
1		178.2	2,3
2	4 40 (44 4 2 0 6)	57.3	۵,3
3	4.48 (dd, 4.3, 8.6)		
3	3.15 (dd, 4,3, 13.7)	39.3	
4	2.88 (dd, 8.5, 13.7)	120 6	9 9
	7 10 (4 7 0)	139.6	2, 3
5,9	7.12 (d, 7.3)	132.5	3
6, 8	7.05 (t, 7.3)	129.1	
7	6.99 (t, 7.3)	127.2	
. D. I	1 . 700 1411 (6 1	\	1 . 405 1511 (8

 $<sup>^</sup>a$  Recorded at 500 MHz ( $\delta$  values).  $^b$  Recorded at 125 MHz ( $\delta$ values). <sup>c</sup> Hex, hexanoic acid. <sup>d</sup> Proton showing HMBC correlation to indicated carbon.

hydrin spot test, sprodesin was negative. Amino acid analysis of the acid hydrolysate (6 M HCl, 110 °C for 20 h) indicated the presence of homotyrosine (Htyr) and phenylalanine (Phe). The molar ratio of Htyr and Phe was 2:1. Htyr was shown to have both of the L- and D-configurations by HPLC analysis of the Marfey's derivatives of the acid

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hydrolysate. In the case of Phe, it was shown to have the D-configuration by the HPLC analysis.

The sequence of residues in 1 was deduced by HMBC correlations (Table 1) from the α-H to the carbonyl carbon at adjacent residues and the configuration analysis of the partial methanolysis products. From the HMBC correlation, the sequence was deduced as hexanoic acid (Hex)-(Dor L-Htyr)-(L- or D-Htyr)-D-Phe, but the sequence of L- and D-Htyr could not be deduced from these data. To determine the sequence of L- and D-Htyr, spriroidin was partial methanolyzed. After the methanolysis in 2 M HCl-MeOH/ *n*-hexane (1:4, v/v) at 74 °C for 12 h, three major peptides were isolated by HPTLC using chloroform—methanol (9:1, v/v) as a solvent. Peptide-1, -2, and -3 on the HPTLC plate showed  $R_f$  0.67, 0.46, and 0.23, respectively. Peptide-1 was ninhydrin negative. In the positive HRFABMS using glycerol as the matrix, the  $[M + H]^+$  ion was observed at m/z 308.1838. From this, the molecular formula of Peptide-1 was established to be  $C_{17}H_{25}N_1O_4$  (calcd for [M + H] 308.1862,  $\Delta$  -2.4 mmu). This molecular formula indicated that Peptide-1 consisted of Htyr and hexanoic acid. After the hydrolysis of Peptide-1, the configuration of the amino acid was determined. Htyr in the peptide was shown to have the D-configuration. Peptide-2 was ninhydrin positive and consisted of D- and L-Htyr. Peptide-3 was ninhydrin positive. After the hydrolysis of Peptide-3, Htyr and Phe were detected. The configuration of the amino acid was determined. Htyr was shown to have the L-configuration, and Phe the D-configuration. From these data, the structure of spiroidesin was established as 1 [(Hex)-(D-Htyr)-(L-Htyr)-(D-Phe)].

Spiroidesin inhibited the growth of the toxic cyanobacterium Microcystis aeruginosa (NIES-88). When M. aeruginosa cells (2 × 10<sup>5</sup> cells/mL) in logarithmic growth were cultured in MA medium containing various concentrations of spiroidesin for 5 days, cell density was decreased to 50% of the control by sprodesin at a concentration of 1.6  $\times$  10<sup>-6</sup> M. Spiroidesin also inhibited chymotrypsin activity with an  $IC_{50}$  of  $1.0\,\times\,10^{-5}$  M.

#### **Experimental Section**

General Experimental Procedures. NMR spectra were recorded on a JEOL JMNA-500 spectrometer (500 MHz). <sup>1</sup>H and <sup>13</sup>C NMR chemical shifts are referenced to TMS. Homonuclear <sup>1</sup>H connectivities were determined from the COSY and HOHAHA experiments, and heteronuclear <sup>1</sup>H-<sup>13</sup>C connectivities were determined by HSQC and HMBC experiments. LRMS and HRMS were performed with a JEOL JMS-700

**Collection.** Blooms of *Anabaena spiroides* were collected from a freshwater pond in Chaing Mai, Thailand. The blooms were immediately freeze-dried, then stored at -20 °C until

Extraction and Isolation. Methanol extract from 10 g of freeze-dried cells was evaporated under reduced pressure. The remaining residue was suspended in 5% (v/v) aqueous acetic acid solution. The suspension was centrifuged at 2000 rpm for 20 min and the supernatant retained. The lipopeptide 1 was fractionated by solid-phase extraction using ODS cartridges (Sep-pak ODS).2 The fractionated lipopeptide was isolated by reversed-phase HPLC (Mightysil RP-18, 20 mm i.d.  $\times$  25 cm) with methanol (60%) containing 0.05 M phosphate buffer (pH 3.0) at 10 mL·min<sup>-1</sup>. The isolated lipopeptide was further purified by HPTLC (Merck, Si gel 60 containing fluorescent indicator) using chloroform-methanol-water (60:40:10, v/v) as the solvent. The yield of the lipopeptide 1, spirodesin, was 35 mg.

**Acid Hydrolysis.** Approximately 1–0.1 mg of spiriodesin or partial methanolysis products was hydrolyzed in 6 M HCl at 110 °C for 20 h.7

**Partial Methanolysis.** Approximately 1 mg of sprodesin was dissolved with 0.2 mL of 2 M HCl-methanol and 0.8 mL of *n*-hexane. The tube was sealed with a screw cap and was heated at 74 °C for 12 h. After heating, the hydrolysate was dried under a nitrogen stream, and the remaining residue was dissolved with a small amount of chloroform. The chloroform solution was applied to a HPTLC plate (Si gel 60 F254 (Merck), thickness 0.25 mm) and developed with chloroform-methanol (9:1, v/v) as the solvent. Three major peptides were observed at  $R_f$  0.67, 0.46, and 0.23, respectively. The peptides were collected from the plate and hydrolyzed in 6 M HCl.

**Amino Acid Analysis.** Authentic D- and L-Htyr were gifts from Dr. Mark Bradley of University of Southampton, UK. Other amino acids including D- and L-Phe were purchased from Sigma. The amino acids in the acid hydrolysate of sprodesin and partial methanolysis products were derivatized with Marfey reagent (1-fluoro-2,4-dinitrophenyl-5-L-alanine amide) and analyzed by reversed-phase HPLC on a C-18 column (NOVA-PAK C18,  $4.6 \times 150$  mm).<sup>8</sup> A linear gradient of 10 to 40% acetonitrile in 0.05 M triethylamine phosphate buffered at pH 3.0 (flow rate 2.0 mL/min; UV detection at 340 nm) was used to separate the amino acid derivatives.9 The absolute stereochemistry of each compound was determined by comparing the retention time with those for authentic L and D amino acid derivatives. The retention times (min) and configurations of the amino acids in the acid hydrolysate of spiroidesin were found to be 42.8 (L-Phe), 48.6 (D-Phe), 60.4 (L-Htyr), and 66.9 (D-Htvr).

Biological Activity. Spiroidesin was assayed for cell growth inhibition using cells of the toxic cyanobacterium Microcystis aeruginosa (NIES-88). M. aeruginosa cells (105 cells/mL) in logarithmic growth were cultured in MA medium<sup>10</sup> containing various concentrations of sprodesin  $(0, 10^{-7}, 10^{-6},$  $10^{-5}$ , and  $10^{-4}$  M) for 5 days. Cell number was counted using microscopy and expressed as an average of three replicate experiments. Spiroidesin was also assayed for chymotrypsin inhibition using α-chymotrypsin type II (Sigma C-4129) and N-benzoyl-L-tyrosine ethyl ester as a substrate. 11 When spiroidesin at the concentration of 10<sup>-5</sup> M was added in the chymotrypsin assay system using substrate at the concentration of  $2\times 10^{-5}$  M, the activity was decreased to 50% of the original activity.

**Spiroidesin:** colorless solid (35 mg, 0.35% dry wt);  $[\alpha]^{25}$ <sub>D</sub>  $-62^{\circ}$  (c 0.56, MeOH); UV  $\lambda_{\rm max}$  (H<sub>2</sub>O) 276 nm ( $\epsilon$  2200); <sup>1</sup>H and <sup>13</sup>C NMR, see Table 1; HRFABMS (pos. glycerol) m/z 618.3134  $[M + 1]^+$  (calcd for  $C_{35}H_{43}N_3O_7$ , 618.3179).

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