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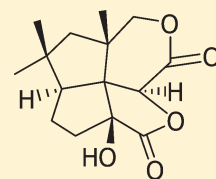
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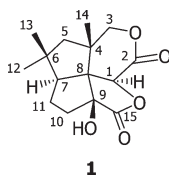
Asperaculin A, a Sesquiterpenoid from a Marine-Derived Fungus, *Aspergillus aculeatus*Nattha Ingavat,^{†,‡} Chulabhorn Mahidol,^{†,‡,§} Somsak Ruchirawat,^{†,‡,§} and Prasat Kittakoop^{*,†,‡}[†]Chulabhorn Graduate Institute and the Center for Environmental Health, Toxicology and Management of Chemicals (ETM), Chemical Biology Program, Vibhavadi-Rangsit Road, Laksi, Bangkok 10210, Thailand[‡]Chulabhorn Research Institute, Vibhavadi-Rangsit Road, Laksi, Bangkok 10210, Thailand[§]Chulabhorn Research Centre, Institute of Molecular Biosciences, Mahidol University, Bangkok 10400, Thailand Supporting Information

ABSTRACT: A novel sesquiterpenoid, asperaculin A (**1**), possessing a novel [5,5,5,6]fenestrane ring system, was isolated from the marine-derived fungus *Aspergillus aculeatus* CRI323-04. The structure of asperaculin A (**1**) was established by analysis of spectroscopic data. The name aspergillane is proposed for the sesquiterpene skeleton in asperaculin A (**1**).



Marine-derived fungi have become increasingly attractive to natural product chemists over the past decades partly because their secondary metabolites have diverse pharmaceutical activity. Fungal metabolites display various and potent biological activities,¹ and some have been used as drugs, for example, echinocandins (antifungal drugs), ergot alkaloids (for the treatment of migraine), cyclosporine (an immunosuppressive drug), and lovastatin (a cholesterol-lowering drug).^{2–4} Fungi of the genus *Aspergillus* are sources of several bioactive compounds.^{5,6} For example, *Aspergillus aculeatus* produces aculeacins A–G (antibiotics and antifungal agents)^{7,8} and CJ-15,183 (squalene synthase inhibitor and antifungal agent).⁹ Our previous investigation revealed that *A. aculeatus* CRI323-04 produced a new tyrosine-derived metabolite, aspergillusol A, an α -glucosidase inhibitor.¹⁰ We further examined the minor constituents from the mycelia of the fungus CRI323-04 and isolated a novel sesquiterpenoid, namely, asperaculin A (**1**), together with a known compound, asperparaline A (aspergillimide),^{11,12} from *A. aculeatus* CRI323-04. Herein, we report the isolation and structure elucidation of asperaculin A (**1**), whose skeleton is novel. Biosynthesis of **1** is also discussed in this paper.

The mycelial extract of *A. aculeatus* CRI323-04 was separated by size-exclusion chromatography and reversed-phase HPLC to yield asperaculin A (**1**) and a known oxindole alkaloid, asperparaline A¹¹ (also known as aspergillimide).¹² Spectral data of asperparaline A were identical in all respects to those reported.^{11,12}



Asperaculin A (**1**) was isolated as white needles. Analysis of NMR and APCITOF MS data determined the molecular formula

of asperaculin A (**1**) as C₁₅H₂₀O₅. The IR spectrum exhibited absorptions at 3461 (hydroxyl group) and 1769 cm^{−1} (carbonyl group). The ¹H NMR spectrum (CDCl₃) of asperaculin A (**1**) displayed signals of oxygenated methine (δ_{H} 4.86) and methylene (δ_{H} 4.38 and 3.88), three singlet methyls (δ_{H} 1.00, 1.15, and 1.33), and four sp³ methines and methylenes (δ_{H} 1.59–2.40) (Table 1). ¹³C and DEPT spectra revealed that asperaculin A (**1**) contained 15 carbons, attributable to three methyls, four methylenes, two methines, and six nonprotonated carbons (two of which were identified as lactone groups). A partial structure H-7/H₂-11/H₂-10 was established from ¹H–¹H COSY correlations. Two singlet methyl groups, C-12 and C-13, were located on the same quaternary carbon, C-6, which was flanked by C-5 and C-7 due to HMBC correlations from H₃-12 to C-13; H₃-13 to C-12; and H₃-12 and H₃-13 to C-5, C-6, and C-7. There were HMBC correlations (acquired in DMSO-*d*₆) from OH-9 to both C-10 and C-15, placing C-9 between C-10 and a C-15 carbonyl ester. HMBC correlations of H-1 to two carbonyl ester carbons (C-15 and C-2) implied that C-1 was between the two carbonyl groups. Since oxygenated methylene H₂-3 appeared in a low-field region, it could link either with a carbonyl ester or with an ether functional group. The HMBC correlation from H₂-3 to a C-2 carbonyl ester group indicated that C-3 was linked to C-2 via an ester bond. The HMBC correlations from H₃-14 to C-3, C-4, C-5, and C-8 and from many protons located around the ring structure, including H-1, H₂-3 α / β , H₂-5 α / β , H-7, OH-9 (acquired in DMSO-*d*₆ at 600 MHz), H-10 α , H₂-11 α / β , and H₃-14, to a quaternary C-8 established the planar structure of asperaculin A (**1**), which contained a lactonized [5,5,5,6] fenestrane skeleton, having the characteristic of C-8 locating in the middle of the molecule. The relative configuration of asperaculin A (**1**) could be determined by NOESY correlations. The NOESY correlations between H-1 and H-7 and between H₃-12 and H-7 indicated that H-1, H-7, and H₃-12 were on the same plane,

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Table 1. ^1H and ^{13}C NMR Spectroscopic Data for Asperaculin A (1)

position	δ_{C}^a	δ_{H}^a (mult, J in Hz)	δ_{C}^b	δ_{H}^b (mult, J in Hz)
1	76.5	4.86, s	76.3	5.31, s
2	167.6		168.5	
3	75.7	4.38, d (11.7, H_{α}); 3.88, d (11.7, H_{β})	74.4	4.45, d (11.6, H_{α}); 3.85, d (11.6, H_{β})
4	44.9		44.7	
5	52.5	1.86, d (14.3, H_{α}); 1.59, d (14.3, H_{β})	51.9	1.79, d (14.0, H_{α}); 1.56, d (14.0, H_{β})
6	40.2		39.0	
7	60.7	2.24, dd (7.2, 9.2)	59.3	2.03, dd (9.7, 7.7)
8	65.5		63.8	
9 (–OH)	84.3		83.1	6.43, s
10	35.9	2.40, m (H_{α}); 1.98, m (H_{β})	33.0	2.17, m (H_{α}); 1.87, m (H_{β})
11	24.0	1.74, m (H_{α}); 1.86, m (H_{β})	23.4	1.85, m (H_{α}); 1.57, m (H_{β})
12	32.2	1.15, s	31.1	1.15, s
13	27.0	1.00, s	26.8	0.94, s
14	23.1	1.33, s	23.6	1.25, s
15	177.1		177.7	

^a Acquired in CDCl_3 (600 MHz). ^b Acquired in $\text{DMSO}-d_6$ (600 MHz).

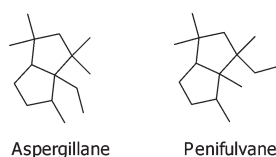


Figure 1. Two novel skeletons of fungal sesquiterpenes.

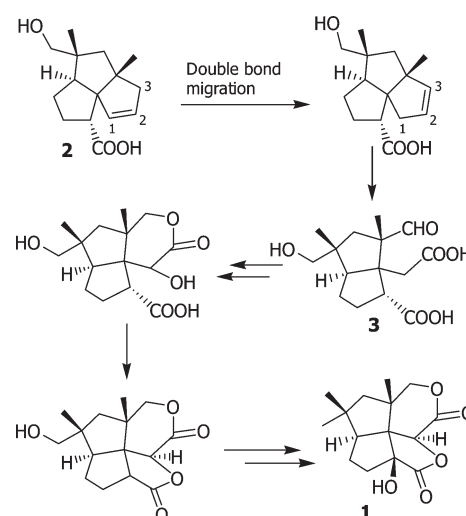
while the NOESY correlation between OH-9 and H_3 -14 established a coplanarity of these protons. Although available NMR data could not establish the configuration of a quaternary C-8, the C-8 stereochemistry in **1** should, upon biosynthetic analogy, be similar to that of penifulvins (A–E),^{13,14} closely related sesquiterpenoids derived from silphenines.

Asperaculin A (**1**) is structurally related to penifulvins (A–E).^{13,14} Penifulvin A, a novel sesquiterpenoid with activity against the fall armyworm, *Spodoptera frugiperda*, was initially separated as a major metabolite from *Penicillium griseofulvum*,¹³ while its oxidized analogues, penifulvin B–E, were discovered later in the same year.¹⁴ Herein, the skeleton of asperaculin A (**1**) was named “aspergillane”, while that of penifulvins was proposed as “penifulvane” (Figure 1). The structure of asperaculin A (**1**) is very similar to that of penifulvin D, except for the transposition of a carbonyl (C-2) and an oxygen in an ester group. Asperaculin A (**1**) is possibly biosynthesized from silphenines,¹⁴ which are metabolites of plants and fungi.^{15,16} As shown in Scheme 1, the proposed biosynthesis of asperaculin A (**1**) starts from a double-bond migration (C1/C2 to C2/C3) of the silphenine intermediate **2**. The C2/C3 double bond then undergoes oxidative cleavage to yield the intermediate **3**, which in turn undergoes sequences of oxidation and lactonizations to finally give asperaculin A (**1**). The double-bond migration is considered as a key step leading to the difference in aspergillane and penifulvane skeletons. Asperaculin A (**1**) did not exhibit cytotoxic activity (at 50 $\mu\text{g}/\text{mL}$) against HepG2, MOLT-3, A549, and HuCCA-1 cancer cell lines.

EXPERIMENTAL SECTION

General Experimental Procedures. Melting points were measured on a digital Electrothermal 9100 melting point apparatus and reported without correction. Optical rotations were measured at the

Scheme 1. Proposed Biosynthesis of Asperaculin A (1)



sodium D line (590 nm) on a JASCO DIP-370 digital polarimeter. UV–vis spectra were obtained using a Shimadzu UV-1700 PharmaSpec spectrophotometer. FTIR data were obtained using a universal attenuated total reflectance attachment on a Perkin-Elmer Spectrum One spectrometer. ^1H , ^{13}C , and 2D NMR spectra were recorded on a Bruker AVANCE 600 NMR spectrometer (operating at 600 MHz for ^1H and 150 MHz for ^{13}C). APCITOF MS spectra were obtained on a Bruker MicroTOF_{LC} spectrometer.

Fungal Material, Extraction, and Isolation. *A. aculeatus* CRI323-04 was isolated from a sponge, *Xesto-spongia testudinaria*, collected from Phi Phi Island, Krabi Province, Thailand. The strain was cultured in 10 L of potato dextrose broth, which was prepared using seawater. Mycelia were separated by filtration and were macerated in MeOH and CH_2Cl_2 , respectively. Crude MeOH extract was partitioned with hexane to remove lipid and extracted with EtOAc to yield 3.10 g of a crude extract. The extract was first fractionated by Sephadex LH-20, eluted with MeOH, producing nine fractions. Fractions 2 and 3, containing sesquiterpenoid, were further isolated by preparative reversed-phase HPLC to yield asperaculin A (**1**)

(70.8 mg) (45% MeOH in water; t_R = 22.3 min) and asperparaline A (29.3 mg) (55% MeOH in water; t_R = 12.0 min).

Asperaculin (1): white needles; mp 190–192 °C; $[\alpha]_D^{28}$ –10.2 (c 0.23, in MeOH); UV (CDCl₃) λ_{max} (log ϵ) 228.50 (2.30) nm; IR (CHCl₃) ν_{max} 3461, 2924, 1769, 1660, 1469 cm^{–1}; ¹H NMR, ¹³C NMR data, see Table 1; APCITOF MS observed m/z 281.1388 (C₁₅H₂₀O₅ + H)⁺ and 303.1225 (C₁₅H₂₀O₅ + Na)⁺, calcd 281.1389 (C₁₅H₂₀O₅ + H)⁺ and 303.1208 (C₁₅H₂₀O₅ + Na)⁺, respectively.

Cytotoxicity Assay. Cytotoxic activity for HepG2, HuCCA-1, and A549 cancer cell lines was evaluated with the MTT assay,¹⁷ while that for the MOLT-3 cell line was assessed using the XTT assay.¹⁸ Doxorubicin and etoposide were the reference drugs.

■ ASSOCIATED CONTENT

S Supporting Information. ¹H, ¹³C NMR and 2D spectra of 1. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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■ REFERENCES

- (1) Bugni, T. S.; Ireland, C. M. *Nat. Prod. Rep.* **2004**, *21*, 143–163.
- (2) Butler, M. S. *J. Nat. Prod.* **2004**, *67*, 2141–2153.
- (3) Shu, Y. Z. *J. Nat. Prod.* **1998**, *61*, 1053–1071.
- (4) Newman, D. J.; Cragg, G. M.; Snader, K. M. *J. Nat. Prod.* **2003**, *66*, 1022–1037.
- (5) Saleem, M.; Ali, M. S.; Hussain, S.; Jabbar, A.; Ashraf, M.; Lee, Y. S. *Nat. Prod. Rep.* **2007**, *24*, 1142–1152.
- (6) Larsen, T. O.; Smedsgaard, J.; Nielsen, K. F.; Hansen, M. E.; Frisvad, J. C. *Nat. Prod. Rep.* **2005**, *22*, 672–695.
- (7) Mizuno, K.; Yagi, A.; Satoi, S.; Takada, M.; Hayashi, M. *J. Antibiot.* **1977**, *30*, 297–302.
- (8) Satoi, S.; Yagi, A.; Asano, K.; Mizuno, K.; Watanabe, T. *J. Antibiot.* **1977**, *30*, 303–307.
- (9) Watanabe, S.; Hirai, H.; Ishiguro, M.; Kambara, T.; Kojima, Y.; Matsunaga, T.; Nishida, H.; Suzuki, Y.; Sugiura, A.; Harwood, H. J., Jr.; Huang, L. H.; Kojima, N. *J. Antibiot.* **2001**, *54*, 904–910.
- (10) Ingavat, N.; Dobereiner, J.; Wiyakrutta, S.; Mahidol, C.; Ruchirawat, S.; Kittakoop, P. *J. Nat. Prod.* **2009**, *72*, 2049–2052.
- (11) Hideo, H.; Yukifumi, N.; Hiroshi, N. *Tetrahedron Lett.* **1997**, *38*, 5655–5658.
- (12) Banks, R. M.; Blanchflower, S. E.; Everett, J. R.; Manger, B. R.; Reading, C. *J. Antibiot.* **1997**, *50*, 840–846.
- (13) Shim, S. H.; Swenson, D. C.; Gloer, J. B.; Dowd, P. F.; Wicklow, D. T. *Org. Lett.* **2006**, *8*, 1225–1228.
- (14) Shim, S. H.; Gloer, J. B.; Wicklow, D. T. *J. Nat. Prod.* **2006**, *69*, 1601–1605.
- (15) (a) Bohlmann, F.; Jakupovic, J. *Phytochemistry* **1980**, *19*, 259–265. (b) *Chapman & Hall/CRC Dictionary of Natural Products*; CRC Press LLC: Boca Raton, FL, 2004; Web version 2004(2).
- (16) Pedras, M. S.; Erosa-López, C. C.; Quail, J. W.; Taylor, J. L. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 3291–3294.

(17) Carmichael, J.; DeGraff, W. G.; Gazdar, A. F.; Minna, J. D.; Mitchell, J. B. *Cancer Res.* **1987**, *47*, 936–942.

(18) Doyle, A.; Griffiths, J. B. *Mammalian Cell Culture: Essential Techniques*; John Wiley & Sons: Chichester, UK, 1997.