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## Neuroprotective 2-(2-Phenylethyl)chromones of *Imperata cylindrica*

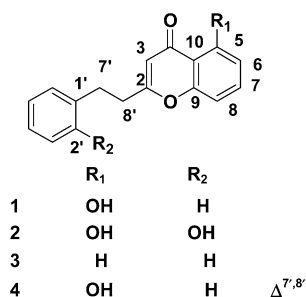
Jeong Seon Yoon, Mi Kyeong Lee, Sang Hyun Sung, and Young Choong Kim\*

College of Pharmacy and Research Institute of Pharmaceutical Science, Seoul National University, San 56-1, Sillim-Dong, Gwanak-Gu, Seoul 151-742, Korea

Received September 29, 2005

Bioactivity-guided fractionation of the methanolic extract of the rhizomes of *Imperata cylindrica* afforded a new compound, 5-hydroxy-2-(2-phenylethyl)chromone (**1**), together with three known compounds, 5-hydroxy-2-[2-(2-hydroxyphenyl)ethyl]chromone (**2**), flindersiachromone (**3**), and 5-hydroxy-2-styrylchromone (**4**). Among these four compounds, **1** and **2** showed significant neuroprotective activity against glutamate-induced neurotoxicity in primary cultures of rat cortical cells.

During our search for neuroprotective compounds from natural products, the MeOH extract of the rhizomes of *Imperata cylindrica* Beauv. (Gramineae) was found to significantly protect primary cultures of rat cortical cells from the toxicity induced by glutamate, an excitatory neurotransmitter. The rhizomes of *I. cylindrica* are widely distributed in Asia and have been described as a diuretic, anti-inflammatory, or antipyretic agent in Korean traditional herbal medicine.<sup>1</sup> Previous studies on the rhizomes of *I. cylindrica* have resulted in the isolation of various compounds such as arundoin,<sup>2</sup> cylindrin,<sup>2</sup> fernenol,<sup>2</sup> cylindol,<sup>3</sup> cylindrene,<sup>4</sup> graminones,<sup>5</sup> and imperarene.<sup>6</sup> To date, however, there has been no report related to neuroprotective constituents of this plant. Thus, we pursued the isolation of neuroprotective constituents from the MeOH extract of *I. cylindrica* rhizomes by bioactivity-guided fractionation. A new chromone, 5-hydroxy-2-(2-phenylethyl)chromone (**1**), and three known chromones (**2–4**) were obtained. Here, we report the isolation and characterization of these chromones and their neuroprotective activity against glutamate-induced neurotoxicity in primary cultures of rat cortical cells.

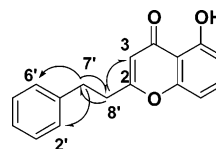


Compound **1** was obtained as a yellow powder. The positive HREIMS of **1** gave a molecular ion at  $m/z$  266.0936, corresponding to the molecular formula  $C_{17}H_{14}O_3$  (calcd  $m/z$  266.0943). The UV absorption maxima at 344 and 252 nm and the IR absorption at 1655, 1621, and 1477  $\text{cm}^{-1}$  suggested the presence of a chromone ring.<sup>7,8</sup> The  $^1\text{H}$  NMR data of **1** indicated the presence of 1,2,3-trisubstituted and monosubstituted aromatic rings from the signals at  $\delta$  7.44 (1H, t,  $J = 8.4$  Hz), 6.80 (1H, dd,  $J = 8.4$  and 0.9 Hz), and 6.71 (1H, dd,  $J = 8.4$  and 0.9 Hz) and signals at  $\delta$  7.12–7.27 (5H, m), respectively. The presence of two methylene groups was also deduced by the signals at  $\delta$  2.99 (2H, t,  $J = 7.6$  Hz) and 2.86 (2H, t,  $J = 7.6$  Hz). The other proton signals could be assigned as one hydroxyl group at  $\delta$  12.45 (1H, br s) and one olefinic proton at  $\delta$  6.00 (1H, s). The  $^{13}\text{C}$  NMR and DEPT spectra of **1** revealed the presence of two methylenes, nine methines, and six quaternary

**Table 1.** Protective Activity of Compounds **1–4** Isolated from *I. cylindrica* against Glutamate-Induced Neurotoxicity in Primary Cultures of Rat Cortical Cells

compound	concentration ( $\mu\text{M}$ )	protection (%) <sup>a</sup>
control		100.0 $\pm$ 2.1
glutamate-treated <sup>b</sup>		0.0 $\pm$ 2.2
<b>1</b>	10.0	67.0 $\pm$ 5.6***
<b>2</b>	10.0	63.6 $\pm$ 5.6***
<b>3</b>	10.0	36.1 $\pm$ 5.4*
<b>4</b>	10.0	–11.3 $\pm$ 4.9
MK-801 <sup>c</sup>	10.0	82.2 $\pm$ 6.9***

<sup>a</sup> Protection (%) was calculated as  $100 \times [\text{optical density (OD) of test compound} + \text{glutamate-treated culture} - \text{OD of glutamate-treated culture}] / [\text{OD of control culture} - \text{OD of glutamate-treated culture}]$ . The ODs of control and glutamate-injured cultures were  $1.00 \pm 0.01$  and  $0.72 \pm 0.01$ , respectively. <sup>b</sup> Glutamate-treated value differs significantly from the untreated control at a level of  $p < 0.001$ . <sup>c</sup> MK-801: dizocipine maleate, a noncompetitive antagonist of NMDA receptor. The values are expressed as mean  $\pm$  SD of triplicate experiments (\* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ ).



**Figure 1.** Key HMBC correlations for **1**.

carbons including one carbonyl carbon at  $\delta$  183.5 (s). In addition, HMBC correlations were observed between the H-7' methylene protons at  $\delta$  2.99 and C-2', C-6', C-8' and between the H-8' methylene protons at  $\delta$  2.86 and C-3, C-7'. On the basis of these findings, compound **1** was suggested as a 2-(2-phenylethyl)-chromone with one hydroxyl group. The location of the hydroxyl group at  $\delta$  12.45 was defined as C-5. Taken together, compound **1** was assigned as the new 5-hydroxy-2-(2-phenylethyl)chromone.

Three known compounds were identified from their spectroscopic data by comparison with literature values as 5-hydroxy-2-[2-(2-hydroxyphenyl)ethyl]chromone (**2**),<sup>10</sup> flindersiachromone (**3**),<sup>9</sup> and 5-hydroxy-2-styrylchromone (**4**).<sup>11</sup> Although the synthesis of compound **4** has been described,<sup>11</sup> this is the first time that it has been isolated from natural resources. In addition, compounds **2** and **3** were isolated for the first time from this plant.

The neuroprotective activity of compounds **1–4** against glutamate-induced neurotoxicity in primary cultures of rat cortical cells was evaluated by the MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) assay as described in our previous reports.<sup>12,13</sup> As shown in Table 1, compounds **1** and **2** showed significant neuroprotective activity against glutamate-induced neurotoxicity at 10.0  $\mu\text{M}$  concentration. Interestingly, however,

\* To whom correspondence should be addressed: Tel: 82-2-880-7842. Fax: 82-2-888-2933. E-mail: youngkim@snu.ac.kr.

compound **4**, which has a structure identical to compound **1** except a double bond at C-7' and C-8', showed little effect against glutamate-induced neurotoxicity. These results suggested that the absence of the double bond at C-7' and C-8' seems to be important for the neuroprotective activity of 2-(2-phenylethyl)chromone. In addition, the neuroprotective activity of compounds **1** and **2** was more potent than that of compound **3**, suggesting the importance of the 5-hydroxy group in the 2-(2-phenylethyl)chromone for neuroprotective activity. Although our present study demonstrated that the presence of the 5-hydroxy group and the absence of the double bond at C-7' and C-8' seem to be important for the neuroprotective activity of 2-(2-phenylethyl)chromone, further studies with more 2-(2-phenylethyl)chromone derivatives are required for the assessment of the relevant structure–activity relationships.

## Experimental Section

**General Experimental Procedures.** UV spectra were recorded on a Shimadzu UV-201 spectrometer using MeOH as solvent. FT-IR spectra were recorded on a Perkin-Elmer 1710 spectrometer. NMR spectra were obtained with Bruker AMX 300 and JEOL JNM-GSX 400 spectrometers. Solvent signals were used as internal references.  $^1\text{H}$ – $^1\text{H}$  COSY, HMQC, and HMBC NMR experiments were performed on the same spectrometer. EI-mass spectra were obtained on a VG Trio 2 spectrometer with a 70 eV ionizing potential. TLC and column chromatography were carried out on precoated silica gel F<sub>254</sub> plates (Merck, art. 5715), RP-18 F<sub>254</sub> plates (Merck, art. 15423), silica gel 60 (230–400 mesh, Merck), Sephadex LH-20 (18–110  $\mu\text{m}$ , Pharmacia Co. Ltd), and LiChroprep RP-18 (40–63  $\mu\text{m}$ , Merck).

**Plant Material.** The rhizomes of *I. cylindrica* were purchased from a commercial supplier in Seoul, Korea, and identified by Dr. Jong Hee Park, a professor of the College of Pharmacy, Pusan National University. A voucher specimen (SNU-0185) has been deposited in the Herbarium of the Medicinal Plant Garden, College of Pharmacy, Seoul National University.

**Extraction and Isolation.** The dried rhizomes of *I. cylindrica* (38 kg) were extracted three times with MeOH in an ultrasonic apparatus. Upon removal of the solvent under vacuum, the MeOH extract yielded 8.7 kg of material (22.9% by dry weight). The extract was suspended in H<sub>2</sub>O and partitioned repeatedly with *n*-hexane. The *n*-hexane fraction (347.5 g), which showed significant neuroprotective activity, was subjected to column chromatography (CC) over silica gel (12  $\times$  80 cm) eluted with an *n*-hexane–EtOAc–MeOH mixture (*n*-hexane; *n*-hexane–EtOAc, 50:1; 30:1; 10:1; 5:1; 3:1; 1:1; EtOAc; EtOAc–MeOH, 50:1; 30:1; 10:1; 5:1; 3:1; 1:1; MeOH; 2 L of each solvent) to afford 12 fractions (F1–F12). F3 (3.3 g) was subjected to reversed-phase (RP) CC with a MeOH–H<sub>2</sub>O step gradient (50% MeOH  $\rightarrow$  100% MeOH) to yield 10 fractions (F3-1–F3-10). Among these fractions, F3-6 (227.9 mg) was subjected to CC over Sephadex LH-20 using MeOH to yield five fractions (F3-6-1–F3-6-5). Compounds **1** (46.8 mg) and **4** (24 mg) were isolated from F3-6-2 (165.7 mg) by repeated semipreparative HPLC (YMC-Pack Pro C<sub>18</sub>, 10  $\times$  250 mm, H<sub>2</sub>O–acetonitrile = 40:60, 2 mL/min,  $t_R$  = 28.87 and 31.84, respectively) with UV detection at 254 nm. F8 (9.0 g) was subjected to RP CC with a MeOH–H<sub>2</sub>O step gradient (60% MeOH  $\rightarrow$  100% MeOH) to yield five fractions (F8-1–F8-5). Among these fractions, F8-3 (282.7 mg) was subjected to CC over Sephadex LH-20 using MeOH to yield five fractions (F8-3-1–F8-3-5). Compounds **2** (3.1 mg) and **3** (8.7 mg) were isolated from F8-3-2 (33.6 mg) by repeated semipreparative HPLC (Luna, 10  $\times$  250 mm, H<sub>2</sub>O–acetonitrile = 40:60, 2 mL/min,  $t_R$  = 19.54 and 28.98, respectively) with UV detection at 254 nm.

**5-Hydroxy-2-(2-phenylethyl)chromone (1):** yellow powder; UV (MeOH)  $\lambda_{\text{max}}$  (log  $\epsilon$ ) 344 (3.83), 252 (4.41), 220 (4.43) nm; IR  $\nu_{\text{KBr}}$  max  $\text{cm}^{-1}$  2926, 1655, 1621, 1477, 1412, 1255, 848, 805;  $^1\text{H}$  NMR

(CDCl<sub>3</sub>, 300 MHz)  $\delta$  12.45 (1H, br s, 5-OH), 7.44 (1H, t,  $J$  = 8.4 Hz, H-7), 7.12–7.27 (5H, m, H-2', 3', 4', 5', 6'), 6.80 (1H, dd,  $J$  = 8.4 and 0.9 Hz, H-8), 6.71 (1H, dd,  $J$  = 8.4 and 0.9 Hz, H-6), 6.00 (1H, s, H-3), 2.99 (2H, t,  $J$  = 7.6 Hz, H-7'), 2.86 (2H, t,  $J$  = 7.6 Hz, H-8');  $^{13}\text{C}$  NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  29.7 (C-7'), 32.8 (C-8'), 106.8 (C-8), 108.8 (C-3), 110.6 (C-10), 111.2 (C-6), 126.7 (C-4'), 128.2 (C-2', 6'), 128.7 (C-3', 5'), 135.2 (C-7), 139.4 (C-1'), 156.7 (C-9), 160.8 (C-5), 169.8 (C-2), 183.5 (C-4); HREIMS (positive)  $m/z$  266.0936 [M]<sup>+</sup> (calcd for C<sub>17</sub>H<sub>14</sub>O<sub>3</sub>, 266.0943).

**Cell Culture.** Primary cultures of rat cortical cells containing both neurons and non-neuronal cells were prepared from 17- to 19-day-old fetal rats (Sprague–Dawley) as previously reported.<sup>12</sup> Cortical cells were seeded onto a collagen-coated 48-well plate at a density of  $1 \times 10^6$  cells/mL. The cultures were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% heat-inactivated fetal calf serum, 100 IU/mL penicillin, and 100  $\mu\text{g/mL}$  streptomycin at 37 °C in a humidified atmosphere of 95% air–5% CO<sub>2</sub>. Cytosine- $\beta$ -D-arabino-furanoside (1  $\mu\text{M}$ ) was added to the culture medium 3 days after plating to inhibit the proliferation of non-neuronal cells. Cultures were allowed to mature for 15 days before being used for experiments.

**Assessment of Neuroprotective Activity.** All tested compounds were dissolved in DMSO (final culture concentration, 0.1%). Cortical cell cultures were pretreated with test compounds for 1 h and then exposed to 100  $\mu\text{M}$  glutamate. After incubation for an additional 24 h, cell viability of the cultures was assessed by the MTT assay, which reflects the mitochondrial enzyme function of cells. Protection (%) was calculated as  $100 \times [\text{optical density (OD) of test compound} + \text{glutamate-treated culture} - \text{OD of glutamate-treated culture}] / [\text{OD of control culture} - \text{OD of glutamate-treated culture}]$ .

**Statistical Analysis.** Data were evaluated for statistical significance using an analysis of variance (ANOVA) with a computerized statistical package. The data were considered to be statistically significant if the probability value was  $<0.05$ .

**Acknowledgment.** This work was supported by a Korea Research Foundation Grant (KRF-2003-015-E00216).

**Supporting Information Available:** NMR spectra for compound **1**. This information is available free of charge via the Internet at <http://pubs.acs.org>.

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NP0503808