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# Synthesis and Glycosidase Inhibition of the Enantiomer of (–)-Steviamine, the First Example of a New Class of Indolizidine Alkaloid

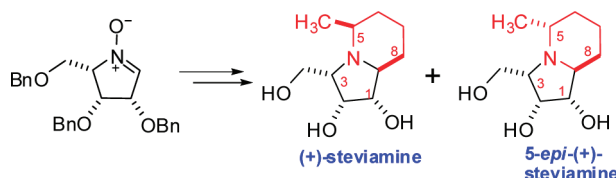
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## ABSTRACT



(+)-Steviamine, the enantiomer of the natural (–)-steviamine, and its corresponding C5 epimer have been synthesized from the d-ribose-derived cyclic nitron. (–)-Steviamine was found to be the first naturally occurring iminosugar that causes any inhibition of  $\alpha$ -galactosaminidases.

The absolute and relative stereochemistry of the five stereogenic centers in (–)-steviamine (**1**), recently isolated from *Stevia rebaudiana* (Asteraceae) leaves,<sup>1</sup> was established by X-ray crystallographic analysis of the hydrobromide salt.<sup>2</sup> This paper reports the synthesis of the enantiomer of (–)-

steviamine (**1**), (+)-steviamine (**2**), and of the corresponding C5 epimer **3** and the glycosidase inhibition profile of **1–3** (Figure 1). (–)-Steviamine (**1**) can be viewed as the indolizidine analogue of the pyrrolizidine, hyacinthacine A<sub>5</sub> (**4**), isolated from *Scilla sibirica*;<sup>3</sup> many hyacinthacines have been isolated from a range of plants.<sup>4</sup> (–)-Steviamine (**1**) is

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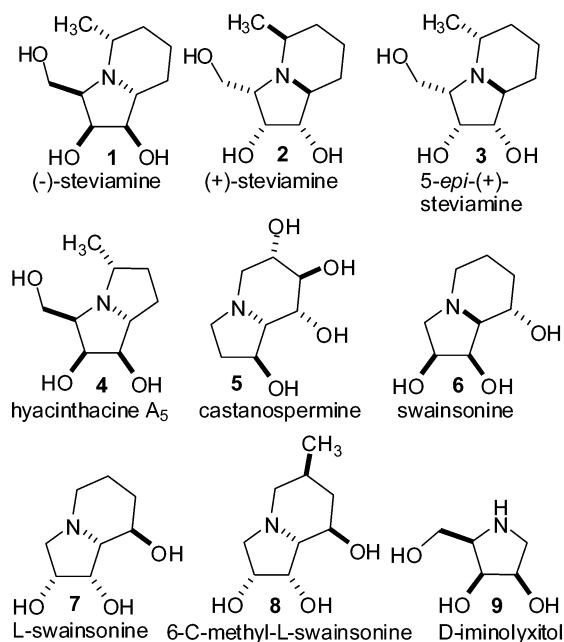
<sup>||</sup> University of Toyama.

(1) Michalik, A.; Hollinshead, J.; Jones, L.; Fleet, G. W. J.; Yu, C.-Y.; Hu, X. G.; van Well, R.; Horne, G.; Wilson, F. X.; Kato, A.; Jenkinson, S. F.; Nash, R. J. *Phytochem. Lett.* **2010**, in press.

(2) Thompson, A. L.; Michalik, A.; Nash, R. J.; Wilson, F. X.; van Well, R.; Johnson, P.; Fleet, G. W. J.; Yu, C.-Y.; Hu, X.-G.; Cooper, R. I.; Watkin, D. J. *Acta Crystallogr.* **2009**, E65, o2904.

(3) (a) Asano, N.; Kuroi, H.; Ikeda, K.; Kizu, H.; Kameda, Y.; Kato, A.; Adachi, I.; Watson, A. A.; Nash, R. J.; Fleet, G. W. J. *Tetrahedron: Asymmetry* **2000**, 11, 1–8. (b) Yamashita, T.; Yasuda, K.; Kizu, H.; Kameda, Y.; Watson, A. A.; Nash, R. J.; Fleet, G. W. J.; Asano, N. *J. Nat. Prod.* **2002**, 65, 1875–1881.

(4) (a) Kato, A.; Adachi, I.; Miyauchi, M.; Ikeda, K.; Komae, T.; Kizu, H.; Kameda, Y.; Watson, A. A.; Nash, R. J.; Wormald, M. R.; Fleet, G. W. J.; Asano, N. *Carbohydr. Res.* **1999**, 316, 95–103. (b) Kato, A.; Kato, N.; Adachi, I.; Hollinshead, J.; Fleet, G. W. J.; Kuriyama, C.; Ikeda, K.; Asano, N.; Nash, R. J. *J. Nat. Prod.* **2007**, 70, 993–997.



**Figure 1.** Iminosugars related to (–)-steviamine.

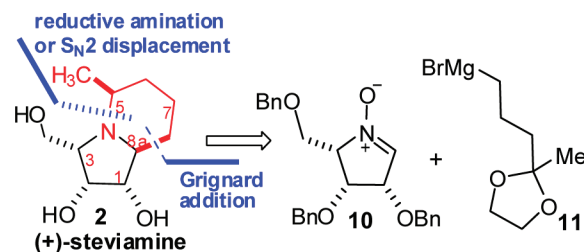
the first example of a new class of indolizidine alkaloid in which an alkyl group is attached to the piperidine ring. Polyhydroxylated indolizidines, such as castanospermine (**5**) from *Castanospermum australe*<sup>5</sup> (an inhibitor of  $\alpha$ -glucosidases) and swainsonine (**6**) from *Swainsona canescens*<sup>6</sup> (an inhibitor of  $\alpha$ -mannosidases), were among the first sugar mimics recognized. Simple derivatives of castanospermine are in development for the treatments of dengue virus<sup>7</sup> and of HCV infections;<sup>8</sup> swainsonine (**6**) has potential as a chemotherapeutic agent for the treatment of cancer.<sup>9</sup>

Nearly 200 iminosugars in which the ring oxygen of the furanose or pyranose has been replaced by nitrogen have been isolated from plants or bacteria.<sup>10</sup> Many of the enantiomers<sup>11</sup> of the naturally occurring alkaloids are themselves even more potent inhibitors of the same en-

zymes;<sup>12</sup> alternatively, they may be excellent inhibitors of different glycosidases.<sup>13</sup> For example, L-swainsonine (**7**) inhibits  $\alpha$ -rhamnosidase<sup>14</sup> rather than  $\alpha$ -mannosidases, and the C-methyl analogue **8** is an even more potent inhibitor of naringinase.<sup>15</sup> (–)-Steviamine and its enantiomer may also be viewed as bicyclic analogues of the iminolyxitols **9** and *ent*-**9**; **9** is a very potent, and *ent*-**9** a weak, competitive inhibitor of  $\alpha$ -galactosidases.<sup>16</sup>

Retrosynthesis for **2** (Scheme 1) suggested that, starting from the D-ribose-derived cyclic nitronone **10**,<sup>17</sup> (+)-steviamine

**Scheme 1.** Retrosynthesis of (+)-Steviamine (**2**)



(**2**) could be synthesized efficiently through the diastereoselective addition of Grignard reagent **11**, followed by annulation, via either intramolecular reductive amination or  $S_N2$  displacement.

According to the retrosynthetic analysis, we commenced the synthesis by making the key intermediate, ketone **13**,

(5) Hohenschütz, L. D.; Bell, E. A.; Jewess, P. J.; Leworthy, D. P.; Pryce, R. J.; Arnold, E.; Clardy, J. *Phytochemistry* **1981**, *20*, 811–814.

(6) Colegate, S. M.; Dorling, P. R.; Huxtable, C. R. *Aust. J. Chem.* **1979**, *32*, 2257–2264.

(7) Whitby, K.; Pierson, T. C.; Geiss, B.; Lane, K.; Engle, M.; Zhou, Y.; Doms, R. W.; Diamond, M. S. *J. Virol.* **2005**, *79*, 8698–8706.

(8) Durantel, D. *Curr. Opin. Invest. Drugs* **2009**, *10*, 860–870.

(9) (a) Klein, J. L. D.; Roberts, J. D.; George, M. D.; Kurtzberg, J.; Breton, P.; Chermann, J. C.; Olden, K. *Br. J. Cancer* **1999**, *80*, 87–95. (b) Lagana, A.; Goetz, J. G.; Cheung, P.; Raz, A.; Dennis, J. W.; Nabi, I. R. *Mol. Cell. Biol.* **2006**, *26*, 3181–3193.

(10) (a) Compain, P.; Martin, O. R. *Iminosugars: from synthesis to therapeutic application*; John Wiley & Sons: New York, 2007; ISBN-0-470-03391-3. (b) Asano, N.; Nash, R. J.; Molyneux, R. J.; Fleet, G. W. J. *Tetrahedron: Asymmetry* **2000**, *11*, 1645–1680. (c) Watson, A. A.; Fleet, G. W. J.; Asano, N.; Molyneux, R. J.; Nash, R. J. *Phytochemistry* **2001**, *56*, 265–295.

(11) (a) d'Alonzo, D.; Guaragna, A.; Palumbo, G. *Curr. Med. Chem.* **2009**, *16*, 473–505. (b) Blériot, Y.; Gretzke, D.; Krülle, T. M.; Butters, T. D.; Dwek, R. A.; Nash, R. J.; Asano, N.; Fleet, G. W. J. *Carbohydr. Res.* **2005**, *340*, 2713–2718. (c) Kato, A.; Kato, N.; Kano, E.; Adachi, I.; Ikeda, K.; Yu, L.; Okamoto, T.; Banba, Y.; Ouchi, H.; Takahata, H.; Asano, N. *J. Med. Chem.* **2005**, *48*, 2036–2044. (d) Asano, N.; Ikeda, K.; Yu, L.; Kato, A.; Takebayashi, K.; Adachi, I.; Kato, I.; Ouchi, H.; Takahata, H.; Fleet, G. W. J. *Tetrahedron: Asymmetry* **2005**, *16*, 223–229.

(12) (a) Scofield, A. M.; Fellows, L. E.; Nash, R. J.; Fleet, G. W. J. *Life Sci.* **1986**, *39*, 645–651. (b) Yu, C. Y.; Asano, N.; Ikeda, K.; Wang, M. X.; Butters, T. D.; Wormald, M. R.; Dwek, R. A.; Winters, A. L.; Nash, R. J.; Fleet, G. W. J. *Chem. Commun.* **2004**, 1936–1937. (c) Best, D.; Wang, C.; Weymouth-Wilson, A. C.; Clarkson, R. A.; Wilson, F. X.; Nash, R. J.; Miyauchi, S.; Kato, A.; Fleet, G. W. J. *Tetrahedron: Asymmetry* **2010**, *21*, 311–319.

(13) (a) Fleet, G. W. J.; Smith, P. W.; Evans, S. V.; Fellows, L. E. *Chem. Commun.* **1984**, 1240–1241. (b) Park, C.; Meng, L.; Stanton, L. H.; Collins, R. E.; Mast, S. W.; Yi, X.; Strachan, H.; Moremen, K. W. *J. Biol. Chem.* **2005**, *280*, 37204–37216. (c) Hakansson, A. E.; van Ameijde, J.; Guglielmini, L.; Horne, G.; Nash, R. J.; Evinson, E. L.; Kato, A.; Fleet, G. W. J. *Tetrahedron: Asymmetry* **2007**, *18*, 282–289.

(14) (a) Davis, B.; Bell, A. A.; Nash, R. J.; Watson, A. A.; Griffiths, R. C.; Jones, M. G.; Smith, C.; Fleet, G. W. J. *Tetrahedron Lett.* **1996**, *37*, 8565–8568. (b) Guo, H.; O'Doherty, G. A. *Org. Lett.* **2006**, *8*, 1609–1612.

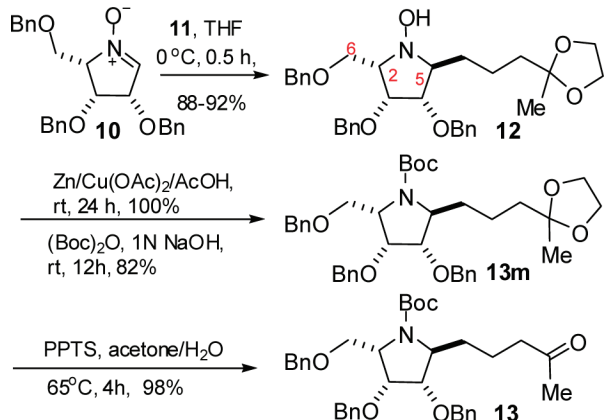
(15) Hakansson, A. E.; van Ameijde, J.; Horne, G.; Nash, R. J.; Wormald, M. R.; Kato, A.; Besra, G. S.; Gurcha, S.; Fleet, G. W. J. *Tetrahedron Lett.* **2008**, *49*, 179–184.

(16) (a) Fleet, G. W. J.; Nicholas, S. J.; Smith, P. W.; Evans, S. V.; Fellows, L. E.; Nash, R. J. *Tetrahedron Lett.* **1985**, *26*, 3127–3130. (b) Mercer, T. B.; Jenkinson, S. F.; Nash, R. J.; Miyauchi, S.; Kato, A.; Fleet, G. W. J. *Tetrahedron: Asymmetry* **2009**, *20*, 2368–2373.

(17) For the synthesis of nitronone **10** see: (a) Pillard, C.; Desvergnès, V.; Py, S. *Tetrahedron Lett.* **2007**, *48*, 6209–6213. (b) Tsou, E.-L.; Yeh, Y.-T.; Liang, P.-H.; Cheng, W.-C. *Tetrahedron* **2009**, *65*, 93–100. (c) Wang, W.-B.; Huang, M.-H.; Li, Y.-X.; Rui, P.-X.; Hu, X.-G.; Zhang, W.; Su, J.-K.; Zhang, Z.-L.; Zhu, J.-S.; Xu, W.-H.; Xie, X.-Q.; Jia, Y.-M.; Yu, C.-Y. *Synlett* **2010**, 488–492. For recent reviews of enantiopure cyclic nitronones see: (d) Revuelta, J.; Cicchi, S.; Goti, A.; Brandi, A. *Synthesis* **2007**, 485–504. (e) Brandi, A.; Cardona, F.; Cicchi, S.; Cordero, F. M.; Goti, A. *Chem.—Eur. J.* **2009**, *15*, 7808–7821. For examples on the syntheses of polyhydroxylated indolizidines via Grignard addition to nitronones, see: (f) Giovannini, R.; Marcantoni, E.; Petrini, M. *J. Org. Chem.* **1995**, *60*, 5706–5707. (g) Cardona, F.; Moreno, G.; Guarna, F.; Vogel, P.; Schuetz, C.; Merino, P.; Goti, A. *J. Org. Chem.* **2005**, *70*, 6552–6555. (h) Delsio, I.; Tejero, T.; Goti, A.; Merino, P. *Tetrahedron* **2010**, *66*, 1220–1227.

starting from the D-ribose-derived cyclic nitron **10** (Scheme 2). Thus, Grignard addition of **11**<sup>18</sup> to the *all-cis* cyclic nitron

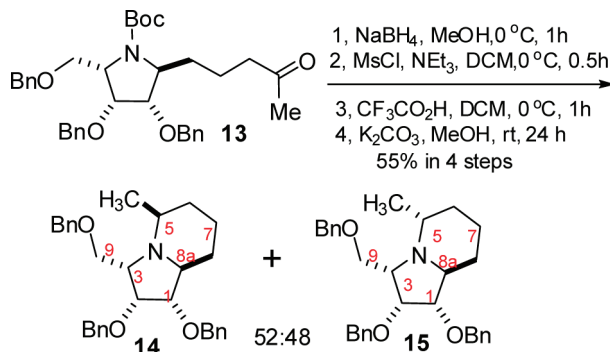
**Scheme 2.** Synthesis of Ketone **13**



**10** at 0 °C furnished the hydroxylamine **12** in excellent yields (88–92%) with high diastereoselectivity (dr >95%). The *trans*-selectivity was determined by NOESY experiment which showed H5 and H6 correlation in **12**. The gratifyingly high *trans*-selectivity can be explained by a Felkin–Anh transition-state model<sup>19</sup> and is in accordance with previous reports.<sup>20</sup> Reduction of the resulting hydroxylamine **12** by Zn–Cu(OAc)<sub>2</sub>–AcOH system gave the corresponding amine in quantitative yield, which was treated with (Boc)<sub>2</sub>O to form the *N*-Boc derivative **13m** in 82% yield. Compound **13m** was then converted to the key intermediate, *N*-Boc ketone **13**, by liberation of the carbonyl group under mild acidic condition.

With ketone **13** in hand, two parallel annulation approaches were under consideration for the construction of the second ring: (1) intramolecular nucleophilic displacement and (2) intramolecular reductive amination. The annulation by intramolecular nucleophilic displacement (Scheme 3) was first

**Scheme 3.** Construction of (+)-Steviamine (**2**) via S<sub>N</sub>2 Displacement

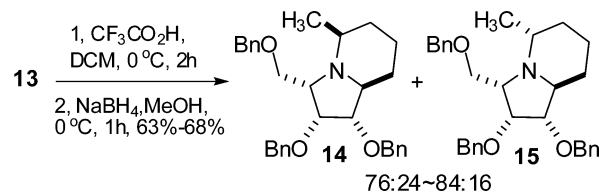


examined. Thus, reduction of *N*-Boc ketone **13** by NaBH<sub>4</sub> furnished a mixture of diastereomeric alcohols, and the

resulting alcohols were transformed into their corresponding mesylates directly without further purification. Liberation of the amino group by CF<sub>3</sub>CO<sub>2</sub>H and subsequent treatment of the resulting amine with K<sub>2</sub>CO<sub>3</sub> in methanol with catalytic water afforded the two epimeric indolizidines **14** and **15** in nearly 1:1 ratio and moderate yield.

In order to achieve better diastereoselectivity, annulation by intramolecular reductive amination was also examined (Scheme 4). Thus, reduction by NaBH<sub>4</sub> of the iminium intermediate after

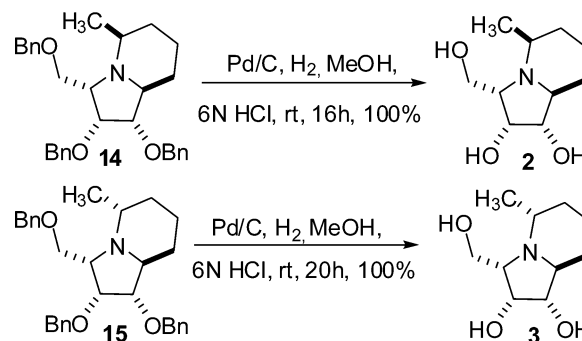
**Scheme 4.** Synthesis of (+)-Steviamine (**2**) via Reductive Amination



acidic deprotection of *N*-Boc ketone **13** generated a mixture of the two epimers **14** and **15** in good yields with relatively higher diastereoselectivity (**14/15** = 76:24–84:16) which favored amine **14**. The stereochemistry of the newly generated stereocenter was determined by the 600 MHz NOESY spectrum of **14** and **15** (**14**: H5 and H9, H5 and H8a; **15**: H3 and H5, H1 and H5; Supporting Information).

Finally, hydrogenolysis of indolizidine **14** and **15** (Scheme 5) gave (+)-steviamine (**2**) and 5-*epi*-(+)-steviamine (**3**) in

**Scheme 5.** Completion of (+)-Steviamine (**2**) and 5-*epi*-(+)-Steviamine (**3**)



quantitative yields, respectively. The <sup>1</sup>H and <sup>13</sup>C NMR spectra of compound **2** were identical (see the Supporting Informa-

(18) Bellas, T. E.; Brownlee, R. G.; Silverstein, R. M. *Tetrahedron* **1969**, 25, 5149–5153.

(19) Merino, P.; Revuelta, J.; Tejero, T.; Cicchi, S.; Goti, A. *Eur. J. Org. Chem.* **2004**, 776–782.

(20) (a) Yu, C.-Y.; Huang, M.-H. *Org. Lett.* **2006**, 8, 3021–3024. (b) Merino, P.; Delso, I.; Tejero, T.; Cardona, F.; Marradi, M.; Faggi, E.; Parmeggiani, C.; Goti, A. *Eur. J. Org. Chem.* **2008**, 2929–2947. (c) Hu, X.-G.; Jia, Y.-M.; Xiang, J.; Yu, C.-Y. *Synlett* **2010**, 982–986, and references cited therein.

tion) to those reported for the natural (–)-steviamine (**1**), and the optical rotation of **2**  $[[\alpha]^{20}_D = +34.0$  (c 1.0, MeOH)] was opposite to that of the natural (–)-steviamine  $[[\alpha]^{20}_D = -22.0$  (c 1.0, MeOH)].

(–)-Steviamine (**1**), (+)-steviamine (**2**), and 5-*epi*-(+)-steviamine (**3**) were assayed as potential glycosidase inhibitors of a range of enzymes (see the Supporting Information). (–)-Steviamine showed weak inhibition of  $\beta$ -glucosidases ( $IC_{50} = 454 \mu M$  against  $\beta$ -glucosidases from almond;  $IC_{50} = 739 \mu M$  against  $\beta$ -glucosidases from *C. saccharolyticum*) but was a good inhibitor of  $\beta$ -galactosidase (rat intestinal lactase,  $IC_{50} = 35 \mu M$ ). In spite of its structural similarity to swainsonine (**6**) and the D-iminolixitol **9**, it showed no significant inhibition of either  $\alpha$ -mannosidase or  $\alpha$ -galactosidase, respectively. The enantiomer of (–)-steviamine (**2**) and its C-5 epimer **3** were weak inhibitors of  $\alpha$ -rhamnosidase ( $IC_{50} = 484$  and  $342 \mu M$ , respectively), several orders of magnitude weaker than shown by L-swainsonine (**7**) and the C-methyl analogue **8**. The *N*-benzyl derivatives of both of the enantiomers of the iminolixitol **9** were more potent inhibitors of  $\alpha$ -rhamnosidase. However (–)-steviamine shows weak inhibition of an  $\alpha$ -galactosaminidase (GalNAcase,  $IC_{50} = 814 \mu M$ ); there has been no prior report of any natural product inhibiting any GalNAcase. GalNAcase inhibition may allow the design of chaperones for the treatment of Schindler–Kanzaki disease<sup>21</sup> and a strategy for the treatment

of cancer by the protection of macrophage activating factor.<sup>22</sup> A synthetic iminosugar analogue of GalNAc has recently been reported as a potent inhibitor of GalNAcases.<sup>23</sup> The specific inhibition GalNAcases by a natural product, and particularly one that does not contain a NAc or any amide group, is remarkable.

In conclusion, the first synthesis of (+)-steviamine (**2**), the enantiomer of the novel natural indolizidine iminosugar (–)-steviamine (**1**), has been accomplished starting from the readily available D-ribose-derived cyclic nitron **10**. (–)-Steviamine was found to be a weak inhibitor of an  $\alpha$ -galactosaminidase (GalNAcase), which is a remarkable finding and might become a starting point for the design and synthesis of more potent inhibitors of  $\alpha$ -galactosaminidase.

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**Supporting Information Available:** Experimental procedures, characterization data, and <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra for all new compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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(21) (a) Clark, N. E.; Garman, S. C. *J. Mol. Biol.* **2009**, *393*, 435–447. (b) Kanekura, T.; Sakuraba, H.; Matsuzawa, F.; Aikawa, S.; Doi, H.; Hirabayashi, Y.; Yoshii, N.; Fukushima, T.; Kanzaki, T. *J. Dermatol. Sci.* **2005**, *37*, 15–20. (c) Chabas, A.; Duque, J.; Gort, L. *J. Inher. Metab. Disease* **2007**, *30*, 108–108. (d) Staretz-Chacham, O.; Lang, T. C.; LaMarca, M. E.; Krasnewich, D.; Sidransky, E. *Pediatrics* **2009**, *123*, 1191–1207. (e) Asfaw, B.; Ledinova, J.; Dobrovolny, R.; Bakker, H. D.; Desnick, R. J.; van Diggelen, O. P.; de Jong, J. G. N.; Kanzaki, T.; Chabas, A.; Maire, I.; Conzelmann, E.; Schindler, D. *J. Lipid Res.* **2002**, *43*, 1096–1104.

(22) (a) Greco, M.; De Mitri, M.; Chiriaco, F.; Leo, G.; Brienza, E.; Maffia, M. *Cancer Lett.* **2009**, *283*, 222–229. (b) Yin, D. S.; Ge, Z. Q.; Yang, W. Y.; Liu, C. X.; Yuan, Y. J. *Cancer Lett.* **2006**, *243*, 71–79. (c) Mohamad, S. B.; Nagasawa, H.; Uto, Y.; Hori, H. *Comp. Biochem. Physiol., Part A: Mol. Integr. Physiol.* **2003**, *134*, 481–481. (d) Bin Mohamad, S.; Nagasawa, H.; Uto, Y.; Hori, H. *Anticancer Res.* **2002**, *22*, 4297–4300. (23) Best, D.; Chairatana, P.; Glawar, A. F. G.; Crabtree, E.; Butters, T. D.; Wilson, F. X.; Yu, C.-Y.; Wang, W.-B.; Jia, Y.-M.; Adachi, I.; Kato, A.; Fleet, G. W. J. *Tetrahedron Lett.* **2010**, *51*, 2222–2224.