See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/12344449

Four New Clerodane Diterpenoids from Callicarpa pentandra

ARTICLE in JOURNAL OF NATURAL PRODUCTS · SEPT	ΓEMBER 2000	
Impact Factor: 3.8 · DOI: 10.1021/np990584m · Source: PubMed		
CITATIONS	READS	
10	26	

5 AUTHORS, INCLUDING:



Yuanjian Xu Chinese Academy of Sciences

25 PUBLICATIONS 214 CITATIONS

SEE PROFILE

Four New Clerodane Diterpenoids from Callicarpa pentandra

Jin Xu, Leslie J. Harrison, Jagadese J. Vittal, Yuan-Jian Xu, and Swee-Hock Goh*

Chemistry Department, National University of Singapore, 10 Kent Ridge Crescent, Singapore 119260

Received November 19, 1999

Leaf extracts of *Callicarpa pentandra* provided four new clerodane-type diterpenoids (1–4), of which 1, 2, and 4 have ring-A-contracted structures. Their structures and stereochemistry were established by spectral data interpretation, and for 3 also by single-crystal X-ray diffraction.

Plants of the genus *Callicarpa* are known to have medicinal properties, for example, for the treatment of rheumatism, stomach disorders, and intestinal troubles.¹ Studies on piscicidal constituents have also been reported.^{2,3} The Malaysian species *Callicarpa pentandra* Roxb. (Verbenaceae), as a liquid is drunk for colds in folk medicine.⁴ We now report the isolation and structural elucidation of four clerodane diterpenoids (1–4; pentandranoic acids A–C and pentandralactone), three of which have a new contracted ring-A clerodane skeleton.

OHC
$$\frac{1}{3}$$
 $\frac{1}{10}$ $\frac{1}{1$

Results and Discussion

Pentandranoic acid A (1) was determined to have the molecular formula C20H28O4 by HREIMS, with major fragment peaks at m/z 204 [C₁₄H₂₀O]⁺, 189 [C₁₃H₁₇O]⁺, and 161 $[C_{12}H_{17}]^+$. The IR spectrum was consistent with the presence of an α,β -unsaturated aldehyde (1659 cm⁻¹), a carboxylic acid function (1710 cm⁻¹), and an α,β -unsaturated ketone (1682 cm⁻¹). The ¹H NMR spectrum showed an aldehydic proton at $\delta_{\rm H}$ 9.90 (Table 1), which correlated (HMQC) with the absorption at δ_C 188.9 (Table 2), and thus verified the presence of the α,β -unsaturated aldehyde function. This was also confirmed by a HMBC experiment showing H-3 correlating with C-1 (3*J*) and C-2 (2*J*) (Table 3). The ¹H NMR spectrum also exhibited vinylidene protons at $\delta_{\rm H}$ 6.13 and 5.90, which were found to be attached to C-16 ($\delta_{\rm C}$ 126.9) from the HMQC experiment, and they showed long-range correlations with C-12 (3J) and C-13 (2J) ($\delta_{\rm C}$ 200.8 and 143.2, respectively) in the HMBC spectrum.

Thus, another α,β -unsaturated ketone could be assumed. A weak carbon signal at δ_C 174.5 (C-15), which showed a 2J interaction with $\delta_{\rm H}$ 3.29 (H₂-14), was assigned to a carboxylic acid. The H₂-14 signal also exhibited HMBC interactions with C-12, C-13, and C-16. Finally, the positions of the four methyl groups were determined by the HMBC experiment (see Table 3), for example, Me-18 had correlations with C-2, C-4, and C-5 and Me-20 with C-8, C-9. C-10. and C-11. Thus, from NMR spectroscopy the structure of compound 1 could be deduced as an Anorclerodane derivative. Comparison of the ¹H and ¹³C NMR data with those reported^{5,6} allowed the confirmation of structure 1 as a modified clerodane skeleton. In the NOESY experiment, H-10 showed correlations with H_{β} -1, H_{β} -6, and H-8, whereas Me-19 had cross-peaks with H_{α} -1 and H_{\alpha}-6. Moreover, Me-17 and Me-19 showed correlations with each other (see Table 3). Thus, the stereochemical relationships for the methyl groups and H-10 were assigned as 17α , 19α , 20α , and 10β , and compound **1** could be determined as 2-formyl-12-oxo-A-norcleroda-2,13(16)-dien-15-oic acid. Noteworthy is that a contraction of ring A in the clerodane structure is rare.6

Pentandranoic acid B (2) was isolated from the same fraction as 1, and a norditerpene skeleton was indicated by HREIMS with the formula C₁₉H₂₈O₄, which had one carbon less than 1 and the absence of a formyl group. In support of this, the main MS fragment peak was at m/z192 ($C_{13}H_{20}O$) instead of the base peak at m/z 204 ($C_{14}H_{20}O$) for compound 1. The ¹³C NMR spectrum (Table 2) indicated that C-2 was a carbonyl carbon, showing long-range correlations with H-1, H-4, H-10, and Me-18 in the HMBC spectrum. In turn, Me-18 had long-range couplings with C-2, C-4, and C-5. The α -configuration of Me-20 was determined by NOESY correlations with H-18 and H-19. Furthermore, the fact that H_{β} -6 correlated with H-10, while H_{α} -6 showed a cross-peak with Me-19, indicated that H-10 and Me-19 are transoid. Compound 2 could also be named as 2,12-dioxo-A-norclerod-13(16)-en-15-oic acid.

Pentandranoic acid C (**3**), the most polar of the four compounds, had the formula $C_{20}H_{30}O_4$, as determined by HREIMS. A comparison of its 1H and ^{13}C NMR spectral data with those of compounds **1** and **2** suggested that the structure of **3** was similar except for the presence of two fused six-membered rings. HMQC and HMBC experiments confirmed the clerodane skeleton. However, in compound **3**, one more vinylidene carbon was present at position C-18 (δ_H 4.83, 4.78; δ_C 109.8). The much deshielded nature of H-3 (δ_H 4.30) and C-3 (δ_C 74.6) indicated the presence of a hydroxyl group, and its α configuration was determined by the NOESY spectrum and confirmed by single-crystal X-ray analysis (Figure 1). Thus, compound **3**, named as 3α -

^{*} To whom correspondence should be addressed. Tel.: 65-8743511. Fax: 65-7791691. E-mail: chmgsh@leonis.nus.edu.sg.

Table 1. ¹H NMR (500 MHz) Spectral Data of 1-4 in CDCl₃

Н	1	2	3	4	4 ^a
1α	2.15 (ddq, 14.0, 10.0, 0.8)	1.98 (dd, 17.5, 13.5)	1.47 (qd, 12.1, 3.3)	2.15 (ddq, 14.2, 12.1, 0.9)	2.22 (ddq, 14.2, 11.9, 1.0)
1β	2.3 (dd, 14.0, 5.7)	2.22 (dd, 17.5, 7.0)	1.81 (ddt 12.1, 2.0, 3.3)	2.49 (dd, 14.2, 6.0)	2.76 (dd, 14.2, 6.0)
2α			1.92 (ddt, 13.5, 3.0, 3.3)		
2β			1.53 (dddd, 13.5, 12.1, 3.3, 3.0)		
3	9.90 (s)		4.30 (t, 3.0)	9.89 (s)	10.0 (s)
4		1.94 (q, 7.0)			
6α	1.62 (dt, 12.0, 3.5)	1.68 (dt, 12.5, 3.2)	1.65 (dt, 12.2, 3.0)	1.63 (dt, 12.1, 3.3)	1.40 (dt, 12.7, 3.7)
6β	1.44 (ddd, 12.0, 8.5, 5.0)	1.34 (dt, 12.5, 5.0)	1.46 (m)	1.34 (dt, 12.1, 4.7)	1.23 (dt, 12.7, 5.3)
7	1.53 (2 H, m)	1.49 (2 H, m)	1.44 (2 H, m)	1.55 (2 H, m)	1.44 (2 H, m)
8	1.89 (m)	1.90 (m)	1.88 (m)	1.51 (m)	1.30 (m)
10	2.04 (dd, 10.0, 5.7)	2.12 (dd, 13.5, 7.0)	1.57 (dd, 12.1, 2.0)	1.81 (dd, 12.1, 6.0)	1.78 (dd, 11.9, 6.0)
11a l	2.70 (2 H, s)	2.55 (d, 15.6)	2.64 (d, 16.0)	1.44 (dd, 15.3, 2.1)	1.03 (dd, 15.3, 1.8)
11b∫	2.70 (2.11, 3)	2.73 (d, 15.6)	2.81 (d, 16.0)	1.79 (dd, 15.3, 8.6)	1.56 (dd, 15.3, 9.4)
12				4.76 (dd, 8.6, 2.1)	$4.20 \; (dd, 9.4, 1.8)^b$
14a]	3.29 (2 H, s)	3.23 (d, 16.5)	3.25 (d, 16.5)	5.88 (d, 1.3)	5.73 (d, 1.3)
14b∫		3.28 (d, 16.5)	3.30 (d, 16.5)	_	
16Z	6.13(br, s)	6.10 (br, s)	6.14 (br, s) \	4.82 (2 H, d, 1.3)	4.33 (2 H, d, 1.3)
16E	5.90 (br, s)	5.88 (br, s)	5.88 (br, s) ∫	,	, , , ,
17	0.84 (3 H, d, 6.8)	0.89 (3 H, d, 6.7)	0.85 (3 H, d, 6.8)	0.78 (3 H, d, 6.2)	0.70 (3 H, d, 6.7)
18	2.00 (3 H, d, 0.8)	0.86 (3 H, d, 7.0)	4.83 Z (br, s)	1.97 (3 H, d, 0.9)	1.62 (3 H, d, 1.0)
			4.78 E (br, s)		
19	0.91 (3 H, s)	0.72 (3 H, s)	1.22 (3 H, s)	0.88 (3 H, s)	0.76 (3 H, s)
20	0.97 (3 H, s)	0.92 (3 H, s)	0.84 (3 H, s)	0.88 (3 H, s)	0.80 (3 H, s)

^a Measured in benzene- d_6 . ^b $\delta = 5.00$ in pyridine- d_5 .

Table 2. ¹³C NMR (125 MHz) Spectral Data of **1–4** in CDCl₃

I UDIC W.	C 1 1111110 (120	I m obois		
С	1	2	3	4 ^a
1	26.8	35.9	17.8	27.3
2	137.2	219.4	34.1	137.6
3	188.9		74.6	187.7
4	172.0	59.4	160.1	169.5
5	50.8	42.7	39.9	50.8
6	33.7	37.7	37.5	34.0
7	28.3	27.8	27.0	28.5
8	38.0	38.0	36.8	38.3
9	40.2	39.8	42.3	38.8
10	53.7	49.7	48.0	55.5
11	44.9	44.8	43.1	46.0
12	200.8	200.6	201.0	65.5
13	143.2	143.1	143.2	173.2
14	37.4	37.0	37.2	114.5
15	174.5	174.5	175.7	172.9
16	126.9	127.2	126.7	70.2
17	15.8	15.9	16.6	15.4
18	9.7	17.1	109.8	9.0
19	17.0	14.2	22.8	16.9
20	17.7	17.0	17.8	17.2

^a Measured in benzene-d₆.

hydroxy-12-oxo-cleroda-4(18),13(16)-dien-15-oic acid, has the structure shown, and NMR data are given in Tables 1, 2, and 3.

Pentandralactone (4), with the molecular formula C₂₀H₂₈O₄ determined by HREIMS, had a 2-formyl-Anorclerodane structure, as in 1. However, the side chain carried an α,β -unsaturated γ -lactone whose 1H and ^{13}C NMR spectral data were consistent with those of similar compounds having this feature.8-10 In addition, in the HMBC experiment, H-14 was observed to have correlations with C-13 (²J), C-15 (²J), and C-16 (³J) and H-16 with C-13 (2*J*), C-14 (3*J*), and C-15 (3*J*). The chemical shift of C-16 ($\delta_{\rm C}$ 70.2) indicated it is an oxgenated methylene. Another hydroxyl group was assigned at C-12 according to the NMR signals observed (δ_{H} 4.74, δ_{C} 65.5). The assignment of H-12 was confirmed by the long-range correlations with C-11, C-13, C-14, and C-16. The absolute configuration of C-12 was assigned S by comparison of its 1H and 13C NMR spectra data with those reported for related diastereomeric compounds, 8-10 in particular, the NOESY H-8 to H-12

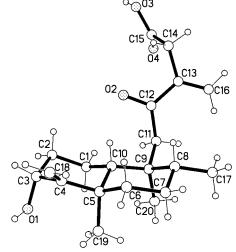


Figure 1. X-ray structure of 3.

correlation. 11 This pentandralactone also could be named as 2-formyl-12. S-hydroxy-A-norcleroda-2, 13(14)-dien-15, 16-olide.

The known compounds ursolic, corosolic, and pomolic acids, as well as 2α , 3α -dihydroxyurs-12-en-28-oic acid, were also isolated from *C. pentandra* leaves, and their structures were determined by comparison of their 1H NMR, ^{13}C NMR, and MS data with reported values. 12

Experimental Section

General Experimental Procedures. Melting points were recorded on a GALEN III hot-stage microscope and are uncorrected. Optical rotations at 25 °C were taken on a DIP-1000 polarimeter. UV spectra were recorded on a Hewlett-Packard 8452A diode array spectrometer, and IR spectra were measured on a Bio-Rad FT-IR spectrometer. NMR spectra were recorded by a Bruker AMX 500 [500 MHz (¹H) and 125 MHz (¹³C)] instrument (with standard pulse programs from Bruker pulse sequences library) in CDCl₃ solutions (unless otherwise specified) with TMS as an internal standard. EIMS were obtained on a Micromass VG 7035 mass spectrometer at 70 eV.

Liquid chromatography was carried out using Kieselgel 60 Si gel (230–400 mesh, Merck), Sephadex LH-20, Lichroprep

Table 3. Selected HMBC and NOESY Spectral Data of 1-4 in CDCl₃

position	1		2		3		4	
H^a	$HMBC^b$	NOESY ^a	$HMBC^b$	NOESY ^a	$HMBC^b$	NOESY ^a	$HMBC^b$	NOESY ^a
1α	2, 10	19, 20			5		2	19, 20
1β	2, 4, 5, 10	10	2, 5, 9, 10	4	2		2, 5	10
2α					1, 3, 4, 10			
3	1, 2	18			1, 5, 18	2α , 2β , $18Z$	1, 2	
4			2, 5, 6, 18, 19	6β , 18				
6α	5, 7, 8, 10, 19	19	5,7, 8, 10	19	5, 8, 10		8, 10	
$egin{array}{c} 6eta \ 7 \end{array}$	5, 7, 19		4, 5, 7, 19		5, 10			
	6, 8	17, 19, 20	5, 8, 9	17, 19, 20	9		5, 8	6β , 11a, 19
8	9, 17, 20	17	7, 9, 17, 20	6β , 7, 10		17		
10	1, 5, 9, 19, 20	6β , 8	1, 2, 5, 9, 11, 19	1β , 4, 6β	1, 5, 9, 19, 20	1β , $^{c}6\beta$	19, 20	6β , 8
11al	8, 9, 10, 12, 20	17, 20	8, 9, 10, 12, 20		8, 9, 10, 12, 20		9, 10, 20	
11b∫	6, 9, 10, 12, 20	1β , 8, 10	8, 9, 10, 12, 20	16 <i>Z</i> , 17, 20	8, 9, 10, 12, 20	1α, 20	8, 12	
12							11, 13, 14, 16	8
14	12, 13, 15, 16	16~E	12, 13, 16	16E	12, 13, 15, 16		13, 15, 16	
16Z	12, 14	11	12, 13		12, 13, 14	11a	13, 14	
16E	13	8	12, 13			14a, 14b	15	
17	7, 8, 9		7, 8, 9	19	7		7, 8, 9	
18	2, 4, 5	19	2, 4, 5		3, 5		2, 4, 5	
18^d					3, 5	6α , 6β		
19	4, 5, 6, 10		4, 5, 6, 10		4, 5, 6, 10	1α , 6α , 7 , 17	4, 5, 6	
20	8, 9, 10, 11	17	8, 9, 10, 11	18, 19	8, 9, 10, 11	1α, 19	8, 9, 10, 11	

^a Numbers for protons are as in Table 1. ^b Numbers refer to carbons. ^c Weak signals. ^d Refers only to 18E in compound 3.

RP $_{18}$ (40–63 $\mu m,$ Merck), and Lichroprep Diol (25–40 $\mu m,$ Merck). TLC was carried out on Si gel precoated glass plates (Merck, Kieselgel 60F $_{254}$, 250 $\mu m),$ C $_{18}$ plates (Whatman, KC $_{18}$ F, 200 $\mu m),$ and Diol Si gel precoated plates (Merck, HPTLC–Fertigplatten Diol F $_{254}$ S, 200 $\mu m).$

Plant Material. The leaves of *C. pentandra* were collected in Kinabatangan, Sabah, Malaysia, in June 1997. A voucher specimen (SAN135275) has been deposited at the herbarium of the Forest Research Centre, Sepilok, Sandakan, Sabah, Malaysia.

Extraction and Isolation. The air-dried and powdered leaves (910 g) of C. pentandra were extracted exhaustively with MeOH (20 L) and then CH₂Cl₂-MeOH (1:1). Evaporation in vacuo provided a residue that was suspended in 10% methanol in water, then extracted with *n*-hexane, CHCl₃ and n-BuOH, consecutively. The CHCl₃ fraction (70 g) was chromatographed initially over Si gel with solvent gradients using hexane, CHCl₃, and MeOH and provided two major fractions (I and II). Fraction I (about 20 g) was eluted by hexane-CHCl₃ (1:1) to pure CHCl3 and fraction II (13 g) from pure CHCl3 to MeOH-CHCl₃ (1:9). Ursolic acid (560 mg) was isolated from fraction I after chromatography on Sephadex LH-20 eluting with CHCl3-MeOH (1:1). This fraction also gave corosolic acid (6.0 mg) after flash column chromatography on Si gel and a Diol column eluting with MeOH-CHCl₃ (1:99). Two other known compounds, pomolic acid (30 mg) and 2α,3α-dihydroxyurs-12-en-28-oic acid (1.3 mg), were obtained from fraction II after repeated chromatography on Si gel eluting with MeOH-CHCl₃ (3:97). The new clerodane diterpenoids **1-4** were isolated from fraction II by flash chromatography (twice) on Si gel (1-3% MeOH in CHCl₃), followed by passage over a RP₁₈ reversed-phase column (MeOH-H₂O, 3:2) and a Diol column (CHCl₃). Further purification of compounds 1 and 2 was carried out by preparative TLC (Si gel) with Me₂CO-EtOAc-hexane (1:2:8) to obtain **1** (8.6 mg, R_f 0.25) and **2** (3.1 mg, R_f 0.30). Compound 3 (17.1 mg) was purified by Si gel preparative TLC with Me₂CO–EtOAc–hexane (1:2:8, R_f 0.20); 4 (2.6 mg) was similarly purified eluting with EtOAc-hexane-HOAc (3:7:0.05, R_f 0.25)

Pentandranoic acid A [2-formyl-12-oxo-A-norcleroda-2,13(16)-dien-15-oic acid] (1): colorless viscous solid (8.6 mg); [α]_D +44.9° (c 0.86, CHCl₃); UV (CHCl₃) $\lambda_{\rm max}$ (log ϵ) 268 (2.98), 276 (2.74) nm; IR (KBr) $\nu_{\rm max}$ 3024, 2961, 2930, 2860, 2750, 1710 (CO₂H), 1682 (C=C-C=O), 1659 (CHO), 1612 (C=C) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS m/z 332 (M⁺, 18), 317 (M⁺ - CH₃, 39), 204 (C₁₄H₂₀O⁺, 92), 189 (C₁₃H₁₇O⁺, 100), 161 (C₁₂H₁₇⁺, 98); HREIMS m/z M⁺ 332.1976 (calcd for C₂₀H₂₈O₄, 332.1988).

Pentandranoic acid B [2,12-dioxo-A-norclerod-13(16)-en-15-oic acid] (2): colorless viscous solid (3.1 mg); $[\alpha]_D$ +94.2° (c 0.31, CHCl₃); UV (CHCl₃) λ_{max} (log ϵ) 242 (2.68) nm; IR (KBr) ν_{max} 3023, 2967, 2930, 1729 (CO), 1716 (CO₂H), 1684 (C=C-C=O) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS m/z 320 (M⁺, 8), 302 (M⁺ – H₂O, 8), 192 (C₁₃H₂₀O⁺, 100), 177 (C₁₂H₁₇O⁺, 75), 121 (70), 69 (67), 41 (68); HREIMS m/z M⁺ 320.2010 (calcd for C₁₉H₂₈O₄, 320.1988).

Pentandranoic acid C [3α-hydroxy-12-oxo-cleroda-4(18),13(16)-dien-15-oic acid] (3): colorless crystals (17.1 mg); mp 135–137 °C; [α]_D +15.2° (c 0.86, CHCl₃); UV (CHCl₃) $\lambda_{\rm max}$ (log ϵ) 242 (2.69) nm; IR (KBr) $\nu_{\rm max}$ 3093 (OH, br), 3018, 2961, 2930, 1715 (CO₂H), 1682 (C=C-C=O), 1630 (C=CH₂) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS m/z 334 (M⁺, 6), 316 (M⁺ – H₂O, 12), 301 (M⁺ – H₂O – CH₃, 14), 206 (C₁₄H₂₂O⁺, 92), 173 (100), 118 (100), 55 (90); HREIMS m/z M⁺ 334.2131 (calcd for C₂₀H₃₀O₄, 334.2144).

Crystal Data for Compound 3: $C_{20}H_{30}O_4$, M = 334.44, monoclinic, $P2_12_12_1$, a = 7.2303(1), b = 11.5201(3), c = 22.5240-(5) Å, V = 1876.11(7) Å³, Z = 4, $\mu(\text{Mo } K\alpha) = 0.081 \text{ mm}^{-1}$, 9288 reflections measured, 3280 unique ($R_{int} = 0.0272$), final R_1 , wR_2 , and GOF values are 0.045, 0.0970, and 1.099 for 2771 independent reflections $[I \ge 2\sigma(I)]$ and 265 parameters. In the final difference Fourier synthesis, the electron density fluctuates in the range 0.248 to -0.138 e Å⁻³. The data collection was performed at 295 K on a Bruker SMART CCD areadetector by ω -scan method, within the limits $1.8^{\circ} \le \theta \le 25.0^{\circ}$. The data were corrected for absorption using an empirical method (SADABS), and the structure was solved by direct methods and refined by full-matrix least-squares (SHELXTL) on F^2 . Crystallographic data for 3 have been deposited with the Cambridge Crystallographic Data Centre. Copies of the data can be obtained, free of charge, on application to the Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK $[Fax: \ +44\text{-}(0)1223\text{-}336033 \ or \ E\text{-}mail: \ deposit@ccdc.cam.ac.uk].$

Pentandralactone [2-formyl-12*S***-hydroxy-A-norcleroda-2,13(14)-dien-15,16-olide] (4):** colorless solid (2.6 mg); mp 53–55 °C; [α]_D +263.5° (c 0.26, CHCl₃); UV (CHCl₃) $\lambda_{\rm max}$ (log ϵ) 260 (2.77), 268 (2.76) nm; IR (KBr) $\nu_{\rm max}$ 3400 (OH, br), 3014, 2935, 2855, 2730 and 1748 (α, β -unsaturated γ -lactone), 1650 (CHO), 1612 (C=C) cm⁻¹; ¹H and ¹³C NMR, see Tables 1 and 2; EIMS m/z 332 (M⁺, 4), 317 (M⁺ – CH₃, 48), 205 (C₁₄H₂₁O⁺, 46), 189 (C₁₄H₂₁+, 80), 107 (82), 55 (100); HREIMS m/z M⁺ 332.1976 (calcd for C₂₀H₂₈O₄, 332.1988).

Acknowledgment. We thank the National University of Singapore for a research scholarship (J.X.) and we are grateful to Ms. Wong Siew Ying and Sng Poh Tee for the NMR data

and to Mrs. Wong Lai Kwai for MS spectral measurements. We also thank Miss Tan Geok Kheng for her assistance in X-ray crystallography and Mr. L. Madani of the Forest Research Centre for the plant identification.

Supporting Information Available: NMR data for compounds **1** and **4** (Tables 1 and 2) and X-ray crystallographic data (Tables 3–7) for compound **3**. This material is available free of charge via the Internet at http://pubs.acs.org.

References and Notes

- (1) Chen, R. S.; Lai, J. S.; Wu, T. S. *J. Chin. Chem. Soc.* **1986**, *33*, 329–333
- (2) Nishino, C.; Kawazu, K.; Mitsui, T. Tetrahedron Lett. 1971, 1541– 1544.
- (3) Kawazu, K.; Inaba, M.; Mitsu, T. Agr. Biol. Chem. 1967, 31, 494–497.
- (4) Perry, L. M.; Metzger, J. Medicinal Plants of East and Southeast Asia; MIT Press: Cambridge, MA, 1980; p 425.

- (5) Habtemarian, S.; Gray, A. I.; Lavaud, C.; Massiot, G.; Skelton, B. W.; Waterman, P. G.; White, A. H. J. Chem. Soc., Perkin Trans. 1 1991, 893–896.
- (6) Bohlmann, F.; Singh, P.; Singh, R. K.; Joshi, K. C. Phytochemistry 1985, 24, 1114–1115.
- (7) Habtemarian, S.; Gray, A. I.; Halbert, G. W.; Waterman, P. G. *Planta Med.* **1990**, *56*, 187–189.
- (8) Shimomura, H.; Sashida, Y.; Ogawa, K.; Iitaka, Y. Chem. Pharm. Bull. 1983, 31, 2192–2199.
- Shimomura, H.; Sashida, Y.; Ogawa, K. Chem. Pharm. Bull. 1989, 37, 354-357.
- (10) O'Mathuna, D. P.; Doskotch, R. W. J. Nat. Prod. 1994, 57, 1382– 1390.
- (11) Shen, X. Y.; Isogai, A.; Furihata, K.; Sun, H. D.; Suzuki, A. Phytochemistry 1993, 33, 887–889.
- (12) Ahmad, V. U.; A.-ur-Rahman. Handbook of Natural Products Data; Elsevier Science: Amsterdam, 1994; Vol. 2, pp 770, 800, 802, and 816