

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/8941600>

Cytotoxic Constituents of *Aspergillus terreus* from the Rhizosphere of *Opuntia versicolor* of the Sonoran Desert

ARTICLE *in* JOURNAL OF NATURAL PRODUCTS · JANUARY 2004

Impact Factor: 3.8 · DOI: 10.1021/np030266u · Source: PubMed

CITATIONS

34

READS

27

9 AUTHORS, INCLUDING:



Kithsiri Wijeratne

The University of Arizona

63 PUBLICATIONS 1,291 CITATIONS

SEE PROFILE



Zhongge Zhang

University of California, San Diego

44 PUBLICATIONS 1,799 CITATIONS

SEE PROFILE



Leland S Pierson

Texas A&M University

64 PUBLICATIONS 3,923 CITATIONS

SEE PROFILE

Cytotoxic Constituents of *Aspergillus terreus* from the Rhizosphere of *Opuntia versicolor* of the Sonoran Desert

E. M. Kithsiri Wijeratne,[†] Thomas J. Turbyville,[†] Zhongge Zhang,[‡] Donna Bigelow,[‡] Leland S. Pierson, III,[‡] Hans D. VanEtten,[‡] Luke Whitesell,[§] Louise M. Canfield,[⊥] and A. A. Leslie Gunatilaka^{*,†}

SW Center for Natural Products Research and Commercialization, Office of Arid Lands Studies, College of Agriculture and Life Sciences, University of Arizona, 250 E. Valencia Road, Tucson, Arizona 85706-6800, Department of Plant Pathology, College of Agriculture and Life Sciences, University of Arizona, Tucson, Arizona 85721-0036, Division of Pediatric Hematology/Oncology, Steele Memorial Children's Research Center, University of Arizona, Tucson, Arizona 85724, and Department of Biochemistry and Biophysics, University of Arizona, Tucson, Arizona 85721-5042

Received June 11, 2003

A novel cyclopentenone, asterredione (**1**), two new terrecyclic acid A derivatives, (+)-5(6)-dihydro-6-methoxyterrecyclic acid A (**2**) and (+)-5(6)-dihydro-6-hydroxyterrecyclic acid A (**3**), and five known compounds, (+)-terrecyclic acid A (**4**), (–)-quadron (**5**), betulinan A (**6**), asterriquinone D (**7**), and asterriquinone C-1 (**8**), were isolated from *Aspergillus terreus* occurring in the rhizosphere of *Opuntia versicolor*, using bioassay-guided fractionation. Acid-catalyzed reaction of **2** under mild conditions afforded **4**, whereas under harsh conditions **2** yielded **5** and (–)-isoquadron (**9**). Catalytic hydrogenation and methylation of **4** afforded 5(6)-dihydro-terrecyclic acid A (**10**) and (+)-terrecyclic acid A methyl ester (**11**), respectively. The structures of **1**–**11** were elucidated by spectroscopic methods. All compounds were evaluated for cytotoxicity in a panel of three sentinel cancer cell lines, NCI-H460 (non-small cell lung cancer), MCF-7 (breast cancer), and SF-268 (CNS glioma), and were found to be moderately active. Cell cycle analysis of **2**, **4**, and **5** using the NCI-H460 cell line indicated that **4** is capable of disrupting the cell cycle through an apparent arrest to progression at the G₁ and G₂/M phases in this p53 competent cell line. A pathway for the biosynthetic origin of asterredione (**1**) from asterriquinone D (**7**) is proposed.

Soil-derived microorganisms represent a valuable resource in the search for secondary metabolites with useful therapeutic applications.¹ Rhizosphere soil, defined as the volume of soil adjacent to and influenced by plant roots (generally less than 1 mm in thickness), constitutes a region of intense microbial activity.² Because of their proximity to plant root exudates, rhizosphere microbial communities are distinct from those of nonrhizosphere (bulk) soil, and the number of microorganisms occurring in the rhizosphere is normally 10–20 times greater than that in the nonrhizosphere soil.³ This is not surprising given that each day plants release 10–40% of their photosynthate from their roots into the soil.⁴ The amount released is influenced by a number of factors including the developmental stage of the plant, soil type, stress, plant nutrition, and microbial activities.^{2a} As a consequence, the diversity of microbial species in the rhizosphere has been postulated to be influenced by different plant species,⁵ their phylogeny, and environmental factors affecting plant growth.⁶ Recent results have substantiated these hypotheses. Techniques based on the analysis of microbial DNA extracted from rhizosphere communities (such as 16S RNA sequence analysis and DNA melting hybridization) as well as analysis of fatty acid metabolic profiles indicate that the rhizosphere communities of plant species differ from each other and from nonrhizosphere communities in the surrounding soil.⁷ This includes microorganisms found in the rhizospheres of desert plants.⁸

The demonstration that rhizosphere microbial diversity is strongly influenced by the diversity of plant species and environmental factors suggests a previously unexploited

opportunity to harvest unique secondary metabolites from the rhizosphere of floristically diverse plant communities such as those in the Sonoran desert. We have recently initiated an extensive program to uncover anticancer agents in the rhizosphere microflora of Sonoran desert plants and thus far have collected over 20 000 rhizosphere bacteria and 3000 rhizosphere fungi. From this library of microorganisms, we have cultured over 1500 fungi, prepared extracts from these, and tested the extracts for cytotoxicity in a panel of three sentinel cancer cell lines [NCI-H460 (non-small cell lung cancer), MCF-7 (breast cancer), SF-268 (CNS glioma)] recently recommended by the NCI as a primary screen before submitting to the 60 cell-line assay.⁹ Several fungal extracts exhibiting ≥90% inhibition of at least one of the three cancer cell lines at a dose of 10 µg/mL were selected for bioactivity-guided fractionation, and one such extract was derived from *Aspergillus terreus* Thom. (Moniliaceae), occurring in the rhizosphere of the staghorn cholla (*Opuntia versicolor* Engelm; Cactaceae). *Aspergillus* species have been extensively investigated before and have yielded anthraquinones,¹⁰ asterriquinones,¹¹ aspothalasins,¹² aflatoxin analogues,¹³ isoquinoline alkaloids,¹⁴ *p*-terphenyls,¹⁵ xanthenes,¹⁶ bioanthrones,¹⁷ tryprostatin analogues,¹⁸ and sesquiterpenes,¹⁹ among other compounds. Some of these *Aspergillus* constituents exhibit interesting biological activities, such as the angioinhibitory effect of fumagillin from *A. fumigatus fresenius*.²⁰ In addition, reports have appeared that demonstrate in vitro and in situ inhibition of MAP2-dependent microtubule assembly by tryprostatin derivatives of *A. fumigatus* BM 939,¹⁸ HIV-reverse transcriptase inhibitory activity of asterriquinones from *A. terreus*,²¹ inhibition of binding of the Grb-2 adapter to phosphorylated EGF receptor tyrosine kinase by asterriquinones of *A. candidus*,^{11b} and oral hypoglycemic activity of some asterriquinones from *Pseudomassaria*.²²

* To whom correspondence should be addressed. Tel: (520) 741-1691. Fax: (520) 741-1468. E-mail: leslieg@ag.arizona.edu.

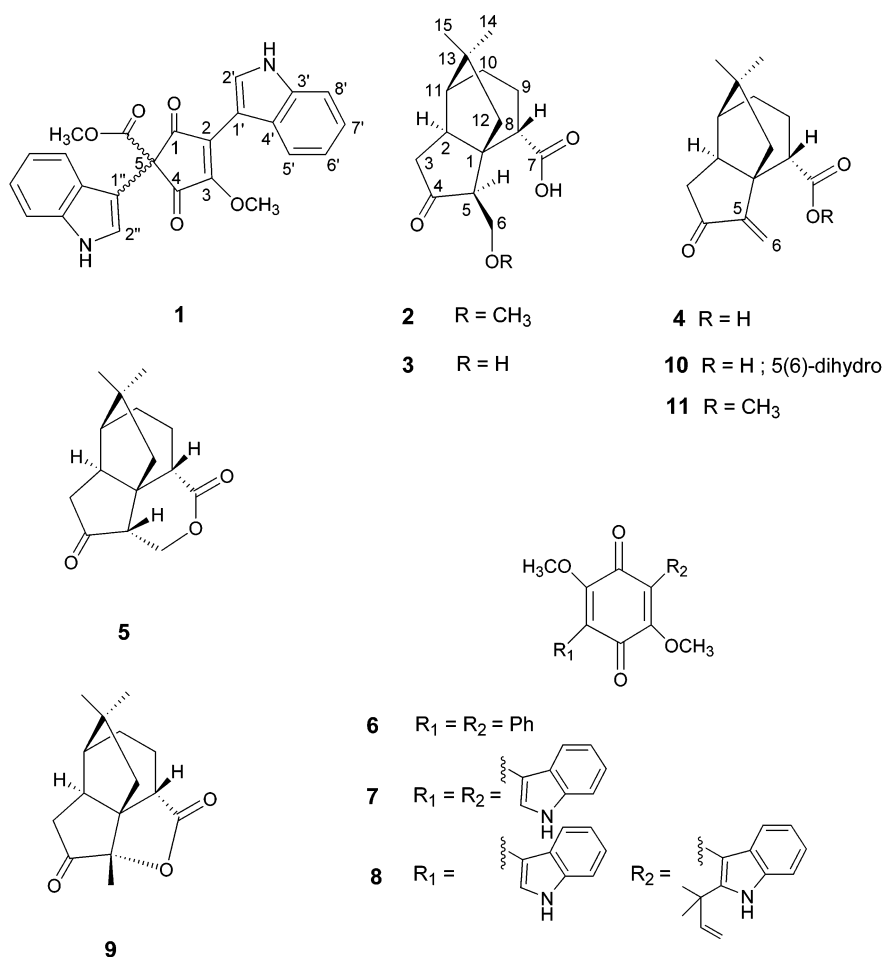
[†] SW Center for Natural Products Research and Commercialization.

[‡] Department of Plant Pathology.

[§] Steele Memorial Children's Research Center.

[⊥] Department of Biochemistry and Biophysics.

Chart 1



Using a bioassay-guided approach, we now report that the cytotoxic EtOAc extract of *A. terreus* occurring in the rhizosphere of staghorn cholla contains a novel type of cyclopentenedione, asterredione (**1**), two new sesquiterpenes, (+)-5(6)-dihydro-6-methoxyterrecyclic acid A (**2**) and (+)-5(6)-dihydro-6-hydroxyterrecyclic acid A (**3**), and the known compounds (+)-terrecyclic acid A (**4**),²³ quadrone (**5**),^{19,24} betulinan A (**6**),²⁵ asterriquinone D (**7**),²⁶ and asterriquinone C-1 (**8**).²⁶ The structures of **2**–**5** were supported by chemical interconversions, which also resulted in the formation of (–)-isoquadrone (**9**).^{27,28} Catalytic hydrogenation and methylation of **4** afforded the corresponding derivatives **10** and **11**. Herein we report the isolation of **1**–**8**, structure elucidation of **1**–**3**, cytotoxicities of **1**–**11** toward three sentinel cancer cell lines, and the effect of **2**, **4**, and **5** on the cell cycle progression of the non-small cell lung cancer cell line NCI-H460.

Results and Discussion

The bioactive CHCl₃ fraction obtained from an EtOAc extract of the rhizosphere fungus *A. terreus* on gel permeation and silica gel chromatography afforded compounds **1**–**8**. Asterredione (**1**) was obtained as a yellow solid that analyzed for C₂₄H₁₈N₂O₅ by a combination of HRMS and ¹³C NMR spectroscopy and indicated 17 degrees of unsaturation. Its IR spectrum had absorption bands at 3392, 1751, and 1718 cm^{–1}, suggesting the presence of NH/OH, ester carbonyl, and α,β-unsaturated ketone carbonyl. The ¹H NMR spectrum of **1** indicated the presence of two OCH₃ singlets at δ 4.45 and 3.78 and two 3-substituted indole residues. The ¹³C NMR spectrum, while confirming the

above functionalities in **1**, suggested the presence of two ketone carbonyls (δ 192.9 and 191.2), an ester carbonyl (δ 167.3), two olefinic carbons (δ 160.6 and 132.5), of which one is oxygenated, and a quaternary carbon (δ 64.3). In the HMBC spectrum, in addition to the expected correlations within the two indole moieties, the OCH₃ signal at δ_H 4.45 showed a correlation to the CO at δ_C 167.3, indicating the presence of a CO₂CH₃ group. The second OCH₃ signal at δ_H 3.78 had a strong correlation to the olefinic carbon at δ_C 160.6. The two indolyl moieties, CO₂CH₃ and OCH₃ groups account for C₁₉H₁₈N₂O₃ and 13 units of unsaturation. Thus, the remaining fragment of **1** has the partial formula C₅O₂ with 4 units of unsaturation and two carbonyl functions, which suggested a fully substituted cyclopentenone. The close ¹³C chemical shifts of the carbonyls (δ 192.9 and 191.2) supported a cyclopent-2-ene-1,4-dione and not a cyclopent-4-ene-1,2-dione structure. On the basis of the differences in chemical shifts of 2-H of the two indole groups, the one having 2-H at δ 8.21 was placed at the C-2/C-3 olefinic carbon of the cyclopent-2-ene-1,4-dione and the other with 2-H at δ 7.27 at the quaternary carbon, C-5. Since the OCH₃ group is attached to an olefinic carbon (C-2/C-3; see above), the CO₂CH₃ group was placed at C-5. The ¹³C NMR data for the cyclopentenedione moiety of **1** showed very close resemblance to those of the spirodione moiety of the recently reported methyl mutadione A.²⁹ Therefore, the structure of asterredione was elucidated as 2,5-di(indol-3'-yl)-3-methoxy-5-methoxycarbonylcyclopent-2-ene-1,4-dione (**1**). With the available data it was not possible to determine the stereochemical disposition of the indole and methoxy-

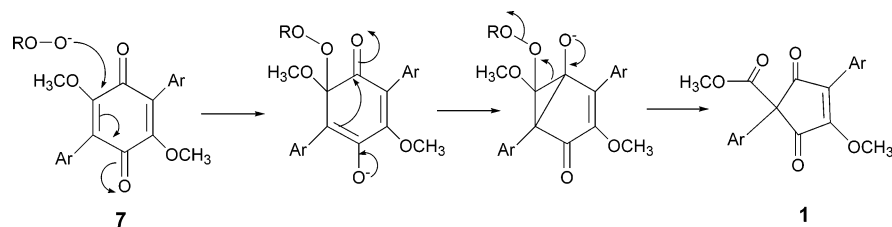


Figure 1. Proposed biosynthetic pathway to asterredione (**1**) from asterriquinone D (**7**).

Table 1. ^1H (500 MHz) and ^{13}C (125 MHz) NMR Data for **2** and **3** in CDCl_3

position	2			3	
	δ_{H}^a	δ_{C}^b	HMBC	δ_{H}^a	δ_{C}^b
1		53.8 s	H-6, H-9		53.3 s
2	2.61 dd (12.0, 6.8)	45.3 d		2.76 dd (11.9, 7.5)	45.2 d
3	2.52 ddd (19.9, 12.0, 1.0)	40.7 t		2.57 dd (20.3, 11.9)	40.9 t
	2.34 dd (19.9, 6.8)			2.39 dd (20.3, 7.5)	
4		213.8 s	H-2, H-6		218.3 s
5	2.94 dd (9.8, 3.6)	56.6 d	H-3	2.85 t (6.7)	57.9 d
6	3.44 t (9.8)	71.1 t	OCH_3 -6	3.71 dd (11.0, 7.0)	60.6 t
	3.93 dd (9.8, 3.6)			4.01 dd (11.0, 6.7)	
7		175.5 s	H-9		178.3 s
8	2.90 d (7.3)	50.5 d	H-12	2.88 t (3.0)	48.7 d
9	1.85 m	22.2 t		1.76–1.92 m	22.0 t
	1.97 t (6.3)				
10	1.76 m	28.5 t	H-8	1.76–1.92 m	28.6 t
	1.92 m				
11	1.83 m	49.3 d	H-3	1.76–1.92 m	48.8 d
12	1.46 d (14.6)	47.7 t		1.51 s	47.6 t
	1.55 d (14.6)				
13		39.9 s	H-10, H-11		40.0 s
14	1.10 s	34.1 q		1.12 s	34.2 q
15	1.15 s	27.1 q		1.15 s	27.1 q
OCH_3	3.41 s	59.5 q			

^a Multiplicities deduced from HSQC; coupling constants (J values in Hz) are in parentheses. ^b Multiplicities deduced from DEPT.

carbonyl groups at C-5. Furthermore, **1** resisted crystallization, precluding its X-ray crystal structure determination. Biogenetically, **1** may arise by a hitherto unknown hydroperoxy-mediated ring contraction of asterriquinone D (**7**) (Figure 1), which was found to co-occur in the same extract (see below).

Compound **2** crystallized as white needles. Its molecular formula, $\text{C}_{16}\text{H}_{24}\text{O}_4$, determined by a combination of HRMS and ^{13}C NMR spectroscopy, indicated 5 degrees of unsaturation. It showed IR absorption bands at 3438, 1732, and 1706 cm^{-1} and had characteristic signals in its ^{13}C NMR spectrum at δ 213.8 and 175.5, suggesting the presence of a ketone carbonyl and a carboxylic acid group. The ^1H NMR spectrum (Table 1) consisted of three 3H singlets and four independent spin systems. On the basis of their chemical shifts the 3H singlets were assigned to a OCH_3 (δ 3.41) and two CH_3 groups (δ 1.15 and 1.10) on quaternary carbons. One of the spin systems consisted of two 1H doublets ($J = 14.6\text{ Hz}$) at δ 1.46 and 1.55 due to a CH_2 sandwiched by quaternary carbons in a cyclic system. Two signals centered at δ 3.44 (t, $J = 9.8\text{ Hz}$) and 3.93 (dd, $J = 9.8$ and 3.6 Hz) together with a signal at δ 2.94 (dd, $J = 9.8$ and 3.6 Hz) suggested the presence of an ABX spin system. The chemical shifts and coupling constants of A and B were indicative of nonequivalent methylene protons adjacent to an oxygen atom. The presence of another ABX spin system in **2** was apparent from the two signals centered at δ 2.34 (dd, $J = 19.9$ and 6.8 Hz) and 2.52 (ddd, $J = 19.9$, 12.0 , and 1.0 Hz) together with a signal at δ 2.61 (dd, $J = 12.0$ and 6.8 Hz). The remaining spin system contained six protons in a complex pattern assignable to a $\text{CHCH}_2\text{CH}_2\text{CH}$ moiety. The ^{13}C NMR spectrum of **1** (Table 1) when analyzed with the help of HSQC showed the presence of two methyl, one methoxy, five methylene (one

of which is attached to an oxygen atom), four methine, and four quaternary carbons (two of which were due to a ketone and a carboxylic acid carbonyl). On the basis of the HMBC correlations (Table 1) the carbon skeleton of **2** was determined to be that of a tricyclic sesquiterpene related to quadronone (**5**).^{19,24} The 2J and 3J correlations in the HMBC spectrum were also useful in locating the carbonyl, $\text{CH}_2\text{-OCH}_3$, and CO_2H groups in the tricyclic skeleton. On the basis of the foregoing evidence, the structure of **2** was determined to be 5(6)-dihydro-6-methoxyterrecyclic acid A. The presence of (+)-terrecyclic acid A (**4**) and (–)-quadronone (**5**) in the same extract and the use of MeOH for extraction of the fungus suggested possible artifactual origin of **2** from **4**. However, both **4** and **5** failed to react with MeOH under a variety of conditions including such harsh conditions as refluxing MeOH in the presence of *p*-toluenesulfonic acid (*p*-TSA). Furthermore, when MeOH was replaced with *n*-BuOH in the extraction process, **2** was found to be present in the resulting extract, suggesting that this compound is a genuine natural product.

Compound **3** was obtained as a colorless semisolid. Its molecular formula, $\text{C}_{15}\text{H}_{22}\text{O}_4$, was determined by a combination of HRMS and ^{13}C NMR data. Comparison of the ^1H and ^{13}C NMR data of **3** with **2** indicated that they are structurally related, the major difference being the presence of an OH in **3** in place of the OCH_3 in **2**. This was further supported by the HRMS data, which showed that the molecular formula of **3** had 14 mass units less than that of **2**, identifying **3** as 5(6)-dihydro-6-hydroxyterrecyclic acid A. Compound **3** has previously been obtained as an intermediate during the total synthesis of quadronone (**5**).^{28,30} The stereochemistry at C-5 in **2** and **3** remains undetermined, though the failure to observe any spontaneous lactonization of these to quadronone (**5**) suggests that the

Table 2. Cytotoxicities of the Compounds **1–11** against a Panel of Tumor Cell Lines^a

compound	cell line ^b		
	NCI-H460	MCF-7	SF-268
1	17.4	25.2	20.7
2	19.4	20.6	21.4
3	14.5	16.5	7.8
4	10.6	24.1	14.7
5	12.4	14.3	16.9
6	58.4	8.7	28.0
7	19.9	17.7	15.8
8	24.2	4.1	25.7
9	NA ^c	NA ^c	NA ^c
10	NA ^c	NA ^c	NA ^c
11	9.4	16.7	10.2
Taxol	9.5	11.2	21.7

^a Results are expressed as IC₅₀ values in μ M except for Taxol, which is in nM. ^b Key: NCI-H460 = human non-small cell lung cancer; MCF-7 = human breast cancer; SF-268 = human CNS cancer (glioma). ^c NA = not active at 10.0 μ g/mL.

–CH₂OR groups to be of β -orientation as shown. This is further supported by the absence of NOEs between H-5 and H-8 in both **2** and **3**. A comparison of spectral and optical rotation data of compounds **4** and **5** with those reported in the literature allowed these to be identified as (+)-terrecyclic acid A^{23,31} and (–)-quadrone,¹⁹ respectively. Treatment of both **2** and **3** with *p*-TSA in toluene at 55 °C for 30 min afforded (+)-terrecyclic acid A (**4**) identical with the natural sample. However, prolonged treatment of **2**, **3**, or **4** with *p*-TSA in toluene under reflux afforded a mixture of (–)-quadrone (**5**) and (+)-isoquadrone (**9**). The former was identified by comparison with the natural sample obtained above and the latter by comparison of the spectral data with those reported in the literature for (+)-isoquadrone (**9**).^{27,28} This is the first report of the occurrence of betulinan A (**6**) in an *Aspergillus* species.

Compounds **1–11** were evaluated for in vitro cytotoxicity against a panel of three sentinel cancer cell lines, NCI-H460 (non-small cell lung), MCF-7 (breast), and SF-268 (CNS glioma). Cells were exposed to serial dilutions of test compounds for 48 h in RPMI 1640 media supplemented with 10% fetal bovine serum, and cell viability was evaluated by the MTT assay.³¹ As shown in Table 2, all except (+)-isoquadrone (**9**) and 5(6)-dihydroterrecyclic acid A (**10**) were found to be cytotoxic. The concentrations resulting in 50% inhibition of cell proliferation/survival as measured by MTT assay (IC₅₀) were found to range between 4.1 and 58.4 μ M. Compound **3** showed selective activity toward the SF-268 cell line, whereas **6** and **8** were found to be more toxic to the MCF-7 cell line. The cytotoxic activities of (+)-terrecyclic acid A (**4**),³² (–)-quadrone (**5**),^{19,33} asterriquinone D (**7**),³⁴ and asterriquinone C-1 (**8**)³⁴ have previously been reported, and a mechanism for the cytotoxic action of asterriquinones has been reported.³⁵ In the sesquiterpene series (**2–5** and **9–11**) our results suggest that the cytotoxic activity is mainly dependent on the presence of an electrophilic α -methylene carbonyl moiety, which can covalently bind to DNA or to a protein target in a Michael-type reaction. Thus, 5(6)-dihydroterrecyclic acid A (**10**), which lacks this reactive moiety, was found to be completely devoid of cytotoxicity. The cytotoxic activities of **2**, **3**, and **5** may be due to their ready conversion into **4** under the culture conditions of the bioassay. The inability of (+)-isoquadrone (**9**) to generate an α -methylene carbonyl moiety due to the absence of a proton at C-5 provides a reasonable explanation as to why this compound is inactive in the cytotoxicity assay. As a first step in elucidating the mechanism(s) of cytotoxicity of this class of compounds, **2**,

Table 3. Effect of **2**, **4**, and **5** Treatment on NCI-H460 Cell Cycle Progression

compound	G ₁ (%)	S (%)	G ₂ /M (%)
2	44.3	21.4	34.3
4	40.7	16.0	43.7
5	38.8	6.3	54.8
DMSO ^a	33.1	35.5	31.4
Taxol ^b	15.5	14.0	70.5
nocodazole ^b	13.0	6.9	80.1

^a Negative control. ^b Positive controls.

4, and **5** were subjected to flow cytometric cell cycle analysis using the non-small cell lung cancer cell line (NCI-H460), which was found to be the most sensitive to these compounds (Table 3). This cell line expresses wild-type p53 and displays an 18 h doubling time under exponential growth conditions.³⁶ Of the three compounds tested, terrecyclic acid A (**4**) and quadrone (**5**) induced the most marked alterations in the distribution of asynchronous NCI-H460 cells within the cell cycle (Figure 2). These compounds resulted in the accumulation of cells with 2n and 4n DNA content and associated depletion of S phase cells. This pattern is consistent with blocks to progression at the G₁ and G₂/M phases of the cell cycle and may underlie the cytotoxic activity observed for these compounds.³⁷ Studies to elucidate the molecular mechanism(s) of action of these compounds and animal studies to evaluate their antitumor potential are currently in progress.

Experimental Section

General Experimental Procedures. Melting points were determined on a Gallenkamp micromelting point apparatus and are uncorrected. Optical rotations were measured with a Jasco Dip-1000 polarimeter using CHCl₃ as solvent. IR spectra for KBr disks were recorded on a Shimadzu FTIR-8300 spectrometer. 1D and 2D NMR spectra were recorded in CDCl₃ with a Bruker DRX-500 instrument at 500 MHz for ¹H NMR and 125 MHz for ¹³C NMR using residual CHCl₃ as internal standard. The chemical shift values (δ) are given in parts per million (ppm), and the coupling constants are in Hz. Low-resolution and high-resolution MS were recorded respectively on Shimadzu LCMS-8000QP α and JEOL HX110A spectrometers.

Fungal Isolation, Identification, and Cultivation. *Aspergillus terreus* Thom. was isolated from the rhizosphere of a staghorn cholla (*Opuntia versicolor* Engelm.) growing in the Tucson Mountains in southern Arizona. Excised roots of *O. versicolor* [1 cm long sections (ca. 5 g)] were placed in 5 mL of phosphate-buffered saline (PBS, 0.1 M, pH = 7.4), vortexed, and sonicated three times. A serial dilution of the suspension was plated onto potato dextrose agar (PDA, Difco, Plymouth, MN) amended with chloramphenicol and streptomycin. After 4 days of incubation at 25 °C, single colonies were transferred onto water agar containing the same antibiotics, and after 3 days a pure culture of *A. terreus* was obtained by hyphal tipping. The strain is deposited in the Department of Plant Pathology and the Southwest Center for Natural Products Research and Commercialization of the University of Arizona microbial culture collection under the code name AH-00-51-F7. The strain was subcultured using Petri dishes with PDA. For long-term storage isolates were subcultured on PDA slants, overlaid with 40% glycerol, and stored at –80 °C.

Flasks (125 mL) containing 100 mL of Sabouraud medium (Difco Laboratories, Detroit, MI) were inoculated with a 5 mm mycelial plug from a pure culture and incubated with shaking (120 rpm) at 25 °C for one week. The mycelium was collected on filter paper by vacuum filtration, air-dried, frozen at –70 °C, and lyophilized for 24 h. The lyophilized mycelia were ground to a fine powder with liquid N₂ and stored at –20 °C. DNA extraction was performed using a plant DNA extraction kit (DNeasy, QIAGEN, Hilden, Germany) according to manu-

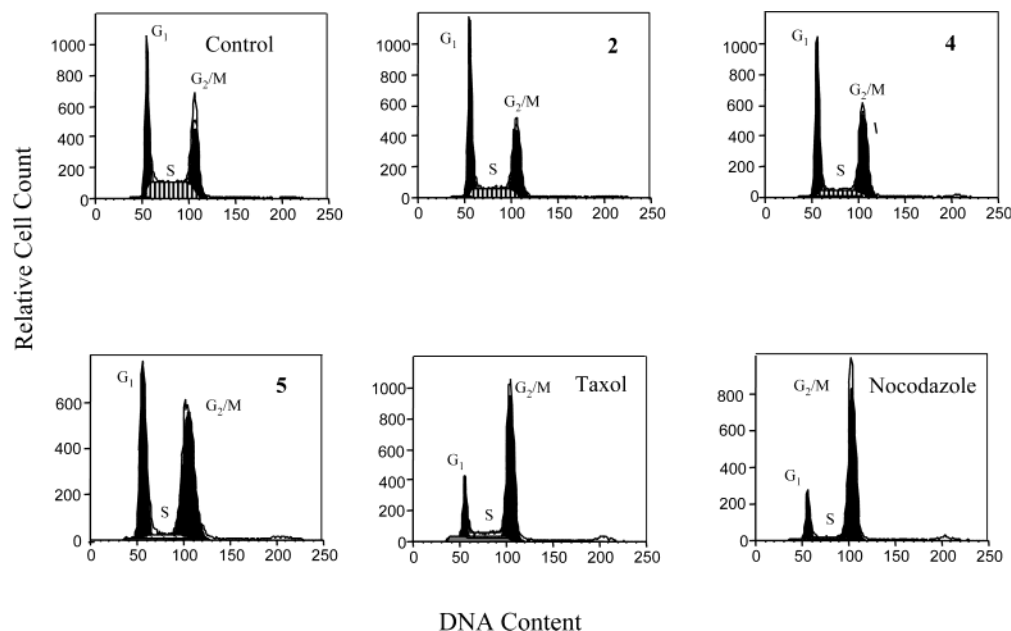


Figure 2. DNA content frequency histograms generated by flow cytometric analysis of NCI-H460 cells treated with control (DMSO), **2**, **4**, **5**, Taxol, and nocodazole.

facturer's instructions with the addition of a precipitation and wash step after the final DNA elution step. DNA was resuspended in 25 μ L of TE (10 mM Tris pH 8.0, 1 mM EDTA). The intergenic spacer (ITS) regions of the ribosomal DNA were amplified by polymerase chain reaction (PCR) using primers ITS1 and ITS4 (ITS1: TCCGTAGGTGAACCTGCGG; ITS4: TCCTCCGCTTATTGATATGC).³⁸ Each 50 μ L of reaction mixture contained 2 μ L of DNA (conc \geq 40 ng/ μ L), 5 μ L of 10 \times reaction buffer, 2 μ L each of 10 mM primer, 2 μ L of 10 mM dNTP, 3 μ L of 25 mM MgCl₂, 2 μ L of Taq polymerase, and 32 μ L of water. The reaction profile was 94 $^{\circ}$ C for 4 min, then (94 $^{\circ}$ C for 45 s, 52 $^{\circ}$ C for 45 s, and 72 $^{\circ}$ C for 4 min) for 36 cycles. The DNA product was purified using a PCR purification kit (Qiaquick, QIAGEN, Hilden, Germany) according to the manufacturer's instructions and sequenced at the University of Arizona Biotechnology Center. The DNA sequence was compared with the GenBank database (BLASTN search) and indicated 100% identity to *A. terreus*. Morphological characters consistent with known isolates of this fungus³⁹ are as follows: colonies pale grayish/brownish orange to camel in color; with a yellow to deep dirty brown reverse; yellowish soluble pigment sometimes present; conidial heads are compact, columnar pale orangish to tan with tightly packed metulae; aspergilla biserial, metulae over the upper half to two-thirds of the vesicle, vesicles (7–) 12–20 μ m wide, somewhat variable in shape, spherical or domelike; stipes smooth walled (70–) 100–250 (–300) μ m long; phialides 5–7 (–9) \times 1.5–2.5 μ m; small smooth-walled conidia (2–2.5 μ m) globose to ellipsoidal, aspergilla tightly packed, 5–7 \times 2–3 μ m (70–) 1000–250 (–300) μ m long.

To produce culture medium for isolation of secondary metabolites, the fungus was cultured in 80 T-flasks (800 mL), each containing 135 mL of PDA coated on five sides of the flasks (total surface area ca. 460 cm²), for 30 days at 27 $^{\circ}$ C.

Extraction and Isolation. Methanol (200 mL/T-flask) was added to all 80 T-flasks, sonicated, and allowed to sit for 24 h at room temperature, and the resulting extract was filtered through Whatman No. 1 filter paper and a layer of Celite 545. The filtrate was concentrated to one-fourth of its original volume and extracted with EtOAc (5 \times 600 mL). Combined EtOAc extracts were evaporated under reduced pressure to afford a dark brown semisolid (4.0 g), which was partitioned between hexane and 80% aqueous MeOH. The cytotoxic 80% aqueous MeOH fraction was diluted to 60% aqueous MeOH by the addition of H₂O and extracted with CHCl₃. Evaporation of CHCl₃ under reduced pressure yielded a dark brown

semisolid (2.75 g). A portion (2.60 g) of this was subjected to gel permeation chromatography on a column of Sephadex LH-20 (60.0 g) in hexane/CH₂Cl₂ (1:4) and eluted with hexane/CH₂Cl₂ (1:4) (450 mL) and CH₂Cl₂/acetone (3:2) (135 mL). Thirty-nine fractions (15 mL each) were collected and combined on the basis of their TLC patterns to yield seven cytotoxic fractions [A (48.5 mg), B (299.0 mg), C (584.0 mg), D (120.2 mg), E (298.5 mg), F (44.4 mg), and G (85.0 mg)]. Column chromatography of fraction A (48.5 mg) on silica gel (2.0 g) and elution with hexane/CH₂Cl₂ (1:1) afforded **6** as a yellow solid (6.1 mg). Chromatography of fraction B (100.0 mg) on silica gel (2.0 g) by elution with hexane/CH₂Cl₂ (1:1) and CH₂Cl₂ afforded two subfractions, B1 and B2. Subfraction B1 (1.2 mg) was a yellow solid identical with **6**, and subfraction B2 (52.9 mg) on crystallization with hexane/CH₂Cl₂ afforded **5** (37.9 mg). Fraction C (584.0 mg) was further fractionated on silica gel (20.0 g) by elution with hexane/CH₂Cl₂ (1:1), CH₂Cl₂, and increasing amounts of MeOH in CH₂Cl₂. Fractions eluted with 1% MeOH in CH₂Cl₂ were found to be cytotoxic, and these were combined and evaporated to yield a white solid (294.0 mg), which on recrystallization from hexane/CH₂Cl₂ afforded **2** (263.0 mg). Chromatography of the fraction D (120.2 mg) on silica gel (3.0 g) by elution with CH₂Cl₂ followed by increasing amounts of MeOH in CH₂Cl₂ afforded several fractions. Of these, the fraction (5.6 mg) eluted with 1% MeOH in CH₂Cl₂ was found to be cytotoxic, and this was further purified by preparative TLC on silica gel (CH₂Cl₂/MeOH, 99:1) to obtain **8** (2.1 mg). Fraction E (298.5 mg) was subfractionated on silica gel (7.0 g) using CH₂Cl₂ and a gradient of CH₂Cl₂ containing MeOH. The combined fractions (139.3 mg) eluted with 1% MeOH in CH₂Cl₂ was found to be cytotoxic, and this fraction was further fractionated on silica gel (3.0 g) using a gradient of hexane containing 2-propanol. The combined fractions (92.0 mg) eluted with 3% 2-propanol in hexane exhibited cytotoxicity. Purification of this fraction by reversed-phase preparative TLC on RP-18 (MeOH/H₂O, 90:10) furnished **4** (59.2 mg). Fraction F (44.4 mg) on column chromatography over silica gel (1.6 g) using a gradient of MeOH in CH₂Cl₂ afforded a cytotoxic fraction (4.3 mg) eluting with 0.5% MeOH in CH₂Cl₂. Further purification of this fraction on silica gel preparative TLC (CH₂Cl₂/MeOH, 96:4) furnished **7** as a dark brown powder (3.5 mg). The last cytotoxic fraction from the original Sephadex column, fraction G (85.0 mg), was separated by column chromatography on silica gel (2.0 g) using a gradient of MeOH in CH₂Cl₂. The cytotoxic fraction (5.2 mg) eluted with

0.5% MeOH in CH_2Cl_2 on further purification by preparative TLC (silica gel, $\text{CH}_2\text{Cl}_2/\text{MeOH}$, 96:4) yielded **1** (2.3 mg). The second cytotoxic fraction (33.7 mg) eluted with 1% MeOH in CH_2Cl_2 on further purification using silica gel preparative TLC ($\text{CH}_2\text{Cl}_2/\text{MeOH}$, 95:5) followed by RP-18 reversed-phase preparative TLC ($\text{MeOH}/\text{H}_2\text{O}$, 70:30) afforded **3** (11.3 mg).

Asterredione (1): yellow powder; mp dec $>112^\circ\text{C}$; $[\alpha]_{\text{D}}^{25} -2.26^\circ$ (c 0.5, CHCl_3); UV (EtOH) λ_{max} (log ϵ) 411 (4.92), 269 (5.23), 218.5 (5.68) nm; IR (KBr) ν_{max} 3392, 1751, 1718, 1676, 1596, 1458, 1421, 1371, 1319, 1238, 1132, 1107, 744 cm^{-1} ; ^1H NMR (CDCl_3 , 500 MHz) δ 8.67 (1H, brs, NH), 8.21 (1H, d, $J = 2.9$ Hz, H-2'), 8.18 (1H, brs, NH), 8.02 (1H, d, $J = 8.0$ Hz, H-5'), 7.79 (1H, d, $J = 8.0$ Hz, H-5''), 7.40 (1H, d, $J = 8.0$ Hz, H-8), 7.32 (1H, d, $J = 8.0$ Hz, H-8''), 7.27 (1H, d, $J = 2.8$ Hz, H-2''), 7.26 (1H, t, $J = 8$ Hz, H-7'), 7.21 (1H, t, $J = 8.0$ Hz, H-6'), 7.17 (1H, t, $J = 8.0$ Hz, H-7''), 7.12 (1H, t, $J = 8.0$ Hz, H-6''), 4.45 (3H, s, $-\text{COCH}_3$), 3.78 (3H, s, $-\text{OCH}_3$); ^{13}C NMR (CDCl_3 , 125 MHz) δ 192.9 (C, C-1), 191.2 (C, C-4), 167.3 (C, COCH_3), 160.6 (C, C-3), 136.1 (C, C-9'), 132.5 (C, C-2), 129.7 (CH, C-2'), 125.9 (C, C-4'), 125.6 (C, C-4'), 124.2 (CH, C-2''), 123.2 (CH, C-5'), 123.2 (CH, C-7'), 122.6 (CH, C-7''), 121.2 (CH, C-6'), 121.1 (CH, C-5''), 120.4 (CH, C-6''), 111.4 (CH, C-8'), 111.3 (CH, C-8''), 108.4 (C, C-3'), 105.6 (C, C-3'), 64.3 (C, C-5), 60.2 (CH₃, COCH_3), 53.4 (CH₃, OCH_3); HRFABMS m/z 414.1216 $[\text{M}]^+$ (calcd for $\text{C}_{24}\text{H}_{18}\text{N}_2\text{O}_5$, 414.1216).

(+)-5(6)-Dihydro-6-methoxyterrecyclic acid A (2): white needles; mp $141\text{--}142^\circ\text{C}$; $[\alpha]_{\text{D}}^{25} +7.70^\circ$ (c 0.5, CHCl_3); IR (KBr) ν_{max} 2935, 1732, 1706, 1454, 1388, 1263, 1209, 1170, 1107, 1091 cm^{-1} ; ^1H and ^{13}C NMR data, see Table 1; HRFABMS m/z 281.3732 $[\text{M} + 1]^+$ (calcd for $\text{C}_{16}\text{H}_{25}\text{O}_4$, 281.3721).

(+)-5(6)-Dihydro-6-hydroxyterrecyclic acid A (3): white powder; mp $158\text{--}160^\circ\text{C}$; $[\alpha]_{\text{D}}^{25} +27.30^\circ$ (c 1.0 CHCl_3); IR (KBr) ν_{max} 3257, 2898, 1728, 1473, 1389, 1218, 1186, 1167, 1149, 1055, 1029 cm^{-1} ; ^1H and ^{13}C NMR data, see Table 1; HRFABMS m/z 249.3310 $[\text{M} - \text{H}_2\text{O} + 1]^+$ (calcd for $\text{C}_{15}\text{H}_{21}\text{O}_3$, 249.3299).

(+)-Terrecyclic acid A (4): white solid; mp $122\text{--}124^\circ\text{C}$; $[\alpha]_{\text{D}}^{25} +25.7^\circ$ (c 0.5 CHCl_3); ^1H NMR and ^{13}C NMR data were consistent with literature values.²³

(-)-Quadrone (5): white needles, mp $183\text{--}184^\circ\text{C}$ (lit.²⁴ $185\text{--}186^\circ\text{C}$); $[\alpha]_{\text{D}}^{25} -60.53^\circ$ (c 0.5, CHCl_3); ^1H NMR and ^{13}C NMR data were consistent with literature values.²⁴

Betulinan A (6): yellow powder; mp $192\text{--}194^\circ\text{C}$ (lit.²⁵ $191\text{--}193^\circ\text{C}$); UV, IR, and ^1H NMR data were consistent with those reported in the literature.²⁵

Asterriquinone D (7): dark brown powder; mp $178\text{--}180^\circ\text{C}$; UV (EtOH) λ_{max} (log ϵ) 480 (4.71), 282 (5.42), 222 (5.69) nm; IR (KBr) ν_{max} 3325, 1643, 1581, 1512, 1419, 1305, 1234, 1172, 1118, 1041, 948, 732 cm^{-1} ; ^1H NMR ($\text{CDCl}_3 + \text{CD}_3\text{OD}$) δ 7.47 (2H, s, H-2' and H-2''), 7.41 (2H, d, $J = 8.1$ Hz, H-5' and H-5''), 7.31 (2H, d, $J = 8.1$ Hz, H-8' and H-8''), 7.07 (2H, t, $J = 8.1$ Hz, H-7' and H-7''), 6.99 (2H, t, $J = 8.1$ Hz, H-6' and H-6''), 3.54 (6H, s, $2 \times \text{OMe}$); APCIMS (+) m/z 399 $[\text{M} + 1]^+$; APCIMS (-) m/z 397 $[\text{M} - 1]^+$.

Asterriquinone C-1 (8): dark brown powder; mp $162\text{--}164^\circ\text{C}$; UV (EtOH) λ_{max} (log ϵ) 489 (4.19), 367 (3.97), 281 (5.12), 222 (5.49) nm; IR (KBr) ν_{max} 3394, 1720, 1637, 1596, 1460, 1301, 1263, 1060, 746 cm^{-1} ; ^1H NMR (CDCl_3) δ 8.49 (1H, brs, NH), 8.09 (1H, brs, NH), 7.60 (1H, d, $J = 2.7$ Hz, H-indole ring), 7.53 (1H, d, $J = 7.6$ Hz, Ar-H), 7.42 (1H, d, $J = 8.0$ Hz, Ar-H), 7.31 (1H, d, $J = 8.0$ Hz, Ar-H), 7.27 (1H, d, $J = 7.6$ Hz, Ar-H), 7.24 (1H, dt, $J = 7.6, 0.9$ Hz, Ar-H), 7.17 (1H, dt, $J = 8.0, 0.8$ Hz, Ar-H), 7.15 (1H, dt, $J = 7.6, 0.9$ Hz, Ar-H), 7.07 (1H, dt, $J = 8.0, 0.8$ Hz, Ar-H), 6.11 (1H, dd, $J = 17.4$ Hz, 10.5 Hz, vinyl-H), 5.19 (1H, d, $J = 17.4$ Hz, vinyl-H), 5.14 (1H, d, $J = 10.5$ Hz, vinyl-H), 1.48 (6H, s, $2 \times \text{CH}_3$); APCIMS +ve mode m/z 467 $[\text{M} + 1]^+$.

Conversion of 2 to 4. A crystal of *p*-TSA was added to a stirred solution of **2** (5.0 mg) in dry toluene (0.5 mL), and the reaction mixture was heated at 55°C for 30 min (TLC control). Toluene was removed under reduced pressure and the product purified by silica gel preparative TLC ($\text{CH}_2\text{Cl}_2/\text{MeOH}$, 92:8), giving a white solid (3.8 mg, 86%), which was identical with (+)-terrecyclic acid A (**4**) isolated above.

Conversion of 2 to 5 and 9. A crystal of *p*-TSA was added to a stirred solution of **2** (10.0 mg) in dry toluene (0.5 mL), and the reaction mixture was heated under reflux for 30 min. Toluene was removed under reduced pressure, and the crude mixture was subjected to column chromatography on silica gel with CH_2Cl_2 as eluant to obtain **5** (3.7 mg, 41%), identical with the above obtained natural sample, and (-)-isoquadrone (**9**) as a white solid (5.1 mg, 58%); mp $165\text{--}167^\circ\text{C}$ (lit.²⁸ $164\text{--}167^\circ\text{C}$); $[\alpha]_{\text{D}}^{25} -1.40^\circ$ (c 0.2, CHCl_3); IR and ^1H NMR data were consistent with literature values.²⁸

5(6)-Dihydroterrecyclic Acid A (10). Pd on carbon (10%, 1.2 mg) was added to a solution of (+)-terrecyclic acid A (**4**; 5.0 mg) in ethanol (0.5 mL) and stirred under an atmosphere of H_2 . The reaction was monitored by TLC, and after 30 min it was filtered through a cotton plug and ethanol was removed under reduced pressure to give **10** (5.0 mg, 99.2%). The product was a mixture of two diastereoisomers in the ratio 1:14 (by ^1H NMR): ^1H NMR (major isomer) δ 2.80 (1H, dd, $J = 10.6, 8.6$ Hz, H-2), 2.60 (1H, d, $J = 6.1$ Hz, H-8), 2.45–2.30 (3H, m), 1.90–1.55 (6H, m), 1.37 (1H, d, $J = 14.7$ Hz, H-12a), 1.33 (1H, d, $J = 14.7$ Hz, H-12b), 1.12 (3H, s, CH_3), 1.08 (3H, s, CH_3), 0.92 (3H, d, $J = 6.7$ Hz, CH_3); APCIMS – ve mode m/z 249 $[\text{M}^+ - 1]$.

Methyl Ester of (+)-Terrecyclic Acid A (11). CH_2N_2 (in ether) was added dropwise to a stirred solution of (+)-terrecyclic acid A (**4**; 2.0 mg) in diethyl ether (0.4 mL) and methanol (2 drops) at -12°C until the starting material disappeared (TLC control). Solvents were removed under reduced pressure, and the crude product was purified by preparative TLC (silica gel) using 5% methanol in CH_2Cl_2 as eluant to give **11** (2.1 mg, 99.0%): ^1H NMR (CDCl_3) δ 5.92 (1H, s, H-6a), 5.15 (1H, s, H-6b), 3.55 (3H, s, OMe), 3.01 (1H, dd, $J = 11.4, 9.6$ Hz, H-2), 2.99 (1H, d, $J = 8.0$ Hz, H-8), 2.61 (1H, dd, $J = 19.3, 11.4$ Hz, H-3a), 2.50 (1H, dd, $J = 19.3, 9.6$ Hz, H-3b), 2.09 (1H, m), 1.95 (1H, t, $J = 2.9$ Hz, H-9a), 1.86–1.75 (3H, m), 1.74 (2H, s, H-12a and -12b), 1.23 (3H, s, CH_3) 1.17 (3H, s, CH_3); APCIMS +ve mode m/z 263 $[\text{M}^+ + 1]$.

Cytotoxicity Bioassays. The tetrazolium-based colorimetric assay (MTT assay)³¹ was used for the in vitro assay of cytotoxicity to human non-small cell lung carcinoma (NCI-H460), human breast carcinoma (MCF-7), and human glioma (SF-268) cells.

Cell Cycle Analysis. NCI-H460 cells were harvested from flasks at 75–85% confluency, counted and plated into 60 mm dishes at a density of 2.5×10^5 cells/dish, and allowed to reattach overnight. Test compounds, **2**, **4**, and **5** were prepared in stock solutions at 8 mg/mL in DMSO. Treatment solutions were prepared at twice the desired final concentration in complete RPMI 1640 medium such that the vehicle (DMSO) did not exceed a final concentration of 0.1 vol %. The final concentrations of the compounds were 10 $\mu\text{g}/\text{mL}$, and the final concentrations of the positive controls, Taxol and nocodazole, were 0.1 and 3 μM , respectively. A control dish was treated with the vehicle control (DMSO) alone. Cells were incubated overnight in the continuous presence of the various treatments, at which point they were rinsed with PBS, harvested by trypsinization, resuspended in complete medium, pelleted by centrifugation, rinsed once with PBS, and resuspended in 1 mL of Krishan's buffer. Cells were analyzed by flow cytometric analysis on a Becton Dickinson FACScan instrument maintained by the Arizona Cancer Center as a core facility.

Acknowledgment. This work was supported by grants from the Arizona Disease Control Research Commission (ADCRC), and this support is gratefully acknowledged. We thank Dr. Annita Harlan for identification of the plant from which the rhizosphere fungus was collected, Ms. Norma Searer for the assistance in flow cytometric analysis, Mr. Arthur Kerschen, Ms. Anne Fritz, Ms. Manping Liu, and Ms. Libia Luevano for their assistance in cytotoxicity assays, and Dr. Neil Jacobsen for his assistance in interpretation of some NMR spectra. The NMR spectrometer used in this research was funded by a grant from the National Science Foundation (Grant No. 9729350).

References and Notes

- (1) Berdy, J. *Proc. 4th Int. Conf. Chem. Biotechnol. Biol. Act. Nat. Prod.* **1988**, 269–291.
- (2) (a) Curl, E. A.; Truelove, B. *The Rhizosphere*; Springer-Verlag: Berlin, 1985. (b) Metting, B. F. *Soil Microbial Ecology*; Marcel Dekker: New York, 1993.
- (3) Whipps, J. M.; Lynch, J. M. *Adv. Microbiol. Ecol.* **1986**, 9, 187–24.
- (4) (a) Whipps, J. M. In *The Rhizosphere*; Lynch, J. M., Ed.; Afric. Littlehampton: West Sussex, 1990; pp 59–97. (b) Lynch, J. M.; Whipps, J. M. In *The Rhizosphere and Plant Growth*; Keister, D. L., Gegan, P. B., Ed.; Kluwer Academic Publishers: Netherlands, 1993; pp 15–24.
- (5) Rovira, A. D. *Plant Soil* **1956**, 47, 178–194.
- (6) (a) Rovira, A. D. *Plant Soil* **1959**, 49, 53–64. (b) Rovira, A. D. In *Ecology of Soilborne Pathogens—Prelude to Biological Control*; Baker, K. F., Snyder, W. C., Eds.; University of California Press: Berkeley, 1965; pp 170–186.
- (7) (a) Smalla, K.; Wachtendorf, V.; Hever, H.; Liu, W. T.; Forney, L. *Appl. Environ. Microbiol.* **1998**, 64, 1220–1225. (b) Wang, S.; Campbell, C. D.; Edwards, A. C. *Soil Biol. Biochem.* **1998**, 30, 369–378.
- (8) Garrity, G. M.; Heimbuch, B. K.; Gagliardi, M. J. *Indust. Microbiol. Biotech.* **1996**, 17, 260–267.
- (9) <http://dtp.nci.nih.gov/branches/btb/ivclsp.html>.
- (10) Asai, A.; Yamashita, Y.; Ando, K.; Kakita, S.; Kita, K.; Suzuki, Y.; Mihara, A.; Ashizawa, T.; Mizukami, T.; Nakano, H. *J. Antibiot.* **1999**, 52, 1046–1049.
- (11) (a) Kaji, A.; Saito, R.; Nomura, M.; Miyamoto, K.-I.; Kiriya, N. *Anticancer Res.* **1997**, 17, 3675–3680. (b) Alvi, K. A.; Pu, H.; Luche, M.; Rice, A.; App, H.; McMahon, G.; Dare, H.; Margolis, B. *J. Antibiot.* **1999**, 52, 215–223.
- (12) Fang, F.; Vi, H.; Shiomi, K.; Masuma, R.; Yamaguchi, Y.; Zhang, C. G.; Zhang, X. W.; Tanak, Y.; Omura, S. *J. Antibiot.* **1997**, 50, 919–925.
- (13) (a) Bradner, W. T.; Bush, J. A.; Myllymaki, R. W.; Nettleton, D. E.; O'Herron, F. A. *Antimicrob. Agents Chemother.* **1975**, 8, 159–163. (b) Schroeder, H. W.; Verrett, M. J. *Can. J. Microbiol.* **1969**, 15, 895–898.
- (14) Kohno, J.; Hiramatsu, H.; Nihio, M.; Sakurai, M.; Okuda, T.; Komatsubara, S. *Tetrahedron* **1999**, 55, 11247–11252.
- (15) Takahashi, C.; Yoshihira, K.; Natori, S.; Umeda, M. *Chem. Pharm. Bull.* **1976**, 24, 613–620.
- (16) Hamasaki, T.; Kimura, Y. *Agric. Biol. Chem.* **1983**, 47, 163–165.
- (17) Assante, G.; Camarda, L.; Nasini, G. *Gazz. Chim. Ital.* **1980**, 110, 629–631.
- (18) Kondoh, M.; Usui, T.; Mayumi, T.; Osada, H. *J. Antibiot.* **1998**, 51, 801–804.
- (19) Calton, G. J.; Ranieri, R. L.; Espenshade, M. A. *J. Antibiot.* **1978**, 31, 38–42.
- (20) Ingber, D.; Fujita, T.; Kishimoto, S.; Sudo, K.; Kanamaru, T.; Brem, H.; Folkman, J. *Nature* **1990**, 348, 555–557.
- (21) Ono, K.; Nakane, H.; Shimizu, S.; Koshimura, S. *Biochem. Biophys. Res. Commun.* **1991**, 174, 56–62.
- (22) Zhang, B.; Salituro, G.; Szalkowski, D.; Li, Z.; Zhang, Y.; Royo, I.; Vilella, D.; Diez, M. T.; Pelaez, F.; Ruby, C.; Kendall, R. L.; Mao, X.; Griffin, P.; Calaycay, J.; Zierath, J. R.; Heck, A. V.; Smith, R. G.; Moller, D. E. *Science* **1999**, 284, 974–977.
- (23) (a) Hirota, A.; Nakagawa, M.; Sakai, H.; Isogai, A. *J. Antibiot.* **1982**, 35, 783–787. (b) Hirota, A.; Nakagawa, M.; Sakai, H.; Isogai, A. *J. Antibiot.* **1984**, 37, 475–478.
- (24) Ranieri, R. L.; Calton, G. J. *Tetrahedron Lett.* **1978**, 499–502.
- (25) Lee, I.-K.; Yun, B.-S.; Cho, S.-M.; Kim, W.-G.; Kim, J.-P.; Ryoo, I.-J.; Koshino, H.; Yoo, I.-D. *J. Nat. Prod.* **1996**, 59, 1090–1092.
- (26) Arai, K.; Masuda, K.; Kiriya, N.; Nitta, K.; Yamamoto, Y.; Shimizu, S. *Chem. Pharm. Bull.* **1981**, 29, 961–969.
- (27) Nakagawa, M.; Sakai, H.; Isogai, A.; Hirota, A. *Agric. Biol. Chem.* **1984**, 48, 2279–2283.
- (28) Danishefsky, S.; Vaughan, K.; Gadwood, R.; Tsuzuki, K. *J. Am. Chem. Soc.* **1981**, 103, 4136–4141.
- (29) Sontag, B.; Dasenbrock, J.; Arnold, N.; Steglich, W. *Eur. J. Org. Chem.* **1999**, 1051–1055.
- (30) Smith, A. B., III; Konopelski, J. P.; Wexler, B. A.; Sprengeler, P. A. *J. Am. Chem. Soc.* **1991**, 113, 3533–3542.
- (31) Rubinstein, L. V.; Shoemaker, R. H.; Paul, K. D.; Simon, R. M.; Tosini, S.; Skehan, P.; Scudiero, D. A.; Monks, A.; Boyd, M. R. *J. Nat. Cancer Inst.* **1990**, 82, 1113–1118.
- (32) (a) Nakagawa, M.; Hirota, A.; Sakai, H.; Isogai, A. *J. Antibiot.* **1982**, 35, 778–782. (b) Hirota, A.; Nakagawa, M.; Sakai, H.; Isogai, A. *J. Antibiot.* **1984**, 37, 475–478.
- (33) Bokesch, H. R.; McKee, T. C.; Cardellina, J. H., II; Boyd, M. R. *Tetrahedron Lett.* **1996**, 37, 3259–3262.
- (34) Shimizu, S.; Yamamoto, Y.; Koshimura, S. *Chem. Pharm. Bull.* **1982**, 30, 1896–1899.
- (35) (a) Kaji, A.; Saito, R.; Nomura, M.; Miyamoto, K.-I.; Kiriya, N. *Anticancer Res.* **1997**, 17, 3675–3680. (b) Shimizu, S.; Koshimura, S. *Chem. Pharm. Bull.* **1990**, 38, 2617–2619.
- (36) O'Connor, P. M.; Jackman, J.; Bae, I.; Myers, T. G.; Fan, S.; Mutoh, M.; Scudiero, D. A.; Monks, A.; Sausville, E. A.; Weinstein, J. N.; Friend, S.; Fornace, A. J., Jr.; Kohn, K. W. *Cancer Res.* **1997**, 57, 4285–4300.
- (37) Rabinovitch, P. S. *Methods Cell Biol.* **1994**, 14, 263–296.
- (38) Klich, M. A.; Pitt, J. I. *A Laboratory Guide to Common Aspergillus Species and their Teleomorphs*; Commonwealth Scientific and Industrial Research Organization: North Ryde, NSW, Australia, 1988; pp 96–97.
- (39) White, T. J.; Bruns, T. D.; Lee, S.; Taylor, J. In *PCR Protocols: A Guide to Methods and Applications*; Innis, M. A., Gelfand, D. H., Sninsky, J. J., White, T. J., Eds.; Academic Press: San Diego, 1990; pp 315–322.

NP030266U