

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/12396582>

The Structure of Sarcomejine: An Application of Long-Range 1H – 15N Correlation at Natural Abundance

ARTICLE *in* JOURNAL OF NATURAL PRODUCTS · AUGUST 2000

Impact Factor: 3.8 · DOI: 10.1021/np000083x · Source: PubMed

CITATIONS

15

READS

11

5 AUTHORS, INCLUDING:



Nikolas Fokialakis

National and Kapodistrian University of Athens

92 PUBLICATIONS 682 CITATIONS

SEE PROFILE



Prokopios Magiatis

National and Kapodistrian University of Athens

180 PUBLICATIONS 3,372 CITATIONS

SEE PROFILE

Note

The Structure of Sarcomejine: An Application of Long-Range H–N Correlation at Natural Abundance

Nikolas Fokialakis, Prokopios Magiatis, Alexios-Leandros Skaltsounis, Franois Tillequin, and Thierry Svenet

J. Nat. Prod., **2000**, 63 (7), 1004-1005 • DOI: 10.1021/np000083x • Publication Date (Web): 02 June 2000

Downloaded from <http://pubs.acs.org> on March 3, 2009

More About This Article

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Access to high resolution figures
- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article

[View the Full Text HTML](#)

The Structure of Sarcomejine: An Application of Long-Range ^1H – ^{15}N Correlation at Natural Abundance

Nikolas Fokialakis,[†] Prokopios Magiatis,[†] Alexios-Leandros Skaltsounis,^{*,†} François Tillequin,[‡] and Thierry Sévenet[§]

Laboratory of Pharmacognosy, Department of Pharmacy, University of Athens Panepistimiopolis Zografou, GR-15771 Athens, Greece, Laboratoire de Pharmacognosie de l'Université René Descartes, UMR au CNRS No. 8638, Faculté de Pharmacie, 4 Avenue de l'Observatoire, F-75006 Paris, France, and ICSN du CNRS, F-91190 Gif-sur-Yvette, France

Received February 16, 2000

A new 4(1*H*)-quinolinone alkaloid, sarcomejine (**1**), has been isolated from the bark of *Sarcomelicope megistophylla*. Its structure has been elucidated on the basis of MS and NMR data and especially with a long-range ^1H – ^{15}N correlation NMR spectrum at natural abundance.

Sarcomelicope megistophylla Hartley (Rutaceae) is a small- to medium-sized tree, 8–12 m high, easily recognized by its large (up to 34 cm long) pubescent leaves. Hartley described it as endemic to the region of Nouméa, New Caledonia.¹ Recently, we have described the chemical constituents of its leaves^{2,3} and the major alkaloids of the bark.⁴ In a continuation of our studies of the genus *Sarcomelicope*,^{5–12} we report herein the isolation and structure elucidation of a new quinolone alkaloid, from the bark of *S. megistophylla*. Sarcomejine (**1**) was isolated from the dichloromethane extract of the bark. The structure of the novel alkaloid was deduced from ^1H and ^{13}C NMR spectral data and by interpretation of its DEPT 135°, COSY 45°, HMQC, and HMBC spectra and from a gradient inverse-detected long-range ^1H – ^{15}N correlation experiment at natural abundance.

Sarcomejine (**1**) was obtained as a yellow amorphous compound, and its molecular formula was determined by HRMS as $\text{C}_{16}\text{H}_{17}\text{NO}_6$. The UV spectrum suggested a quinolone derivative. The ^1H NMR spectrum indicated four aromatic protons associated with a nonsubstituted ring A in a 4-quinolone-derived skeleton, three OCH_3 groups (two belonging to methyl ester functional groups and one placed on an sp^3 carbon), one NCH_3 group, and one deshielded aliphatic proton (δ 5.28). The ^{13}C NMR spectrum confirmed the above observations and showed the presence of three carbonyl groups. One (δ 174.7) is included in the quinolone skeleton, and the other two correspond to COOCH_3 groups (δ 168.8, 167.6). The sp^3 carbon bearing the methine proton at 5.28 ppm was observed at 78.5 ppm, suggesting that it is an oxygenated carbon attached to an aromatic system. Moreover, three OCH_3 groups and one NCH_3 group were observed at 58.9, 53.3, 52.8, and 35.9 ppm, respectively. Further information on the structure of **1** was obtained from the long-range C–H correlation in the HMBC spectrum (Figure 1). The methine proton at δ 5.28 showed a three-bond correlation with the OCH_3 carbon at δ 58.9 and a two-bond correlation with the carbonyl of the first COOCH_3 group at δ 167.6. Additionally, the methine proton showed a correlation with two quaternary aromatic carbons (145.9, 120.5 ppm). One of them (C-2; 145.9 ppm) was correlated with the protons of the NCH_3 group. Given that sarcomejine possessed an *N*-methyl-4-quinolone nucleus

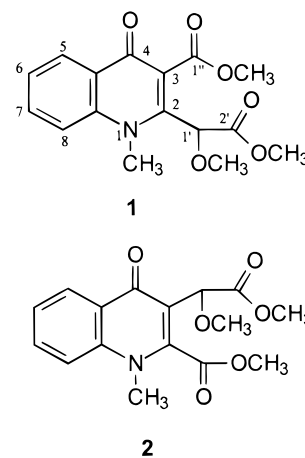


Figure 1. Sarcomejine (**1**) and the alternative structure **2**.

combined with the above-mentioned HMBC correlations, it could be assigned as either structure **1** or **2**. This problem could not be solved by a NOESY spectrum due to the overlapping of the NMe and the OMe protons of one of the COOCH_3 groups (six protons at 3.78 ppm). Therefore, the strong cross-peak observed in the NOESY spectrum (mixing time 700 ms) between the methine proton (5.28 ppm) and a methyl group at 3.78 ppm could not be used as an argument for the unequivocal structure determination of sarcomejine (**1**).

The structure elucidation of this new alkaloid as **1** and discrimination against the alternative structure **2** was provided by long-range ^1H – ^{15}N heteronuclear shift correlation studies. These represent, in the past several years, an area of active research for chemical shift assignments^{13–18} and structure elucidation of alkaloids.^{19–21} Indeed, the three-bond correlation between ^{15}N -1 (118.3 ppm) and the methine proton (5.28 ppm) (Figure 2) permitted placement of the side chain at C-2 of the 4-quinolone ring and, consequently, placement of the second COOCH_3 group at position 3. The experiment was optimized for 5 Hz 3J ^1H – ^{15}N coupling constants.

Experimental Section

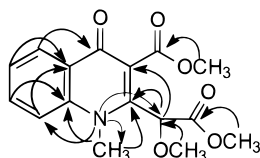
General Experimental Procedures. Optical rotations were measured with a Perkin-Elmer 341 polarimeter. UV spectra were recorded in spectroscopic grade MeOH on a Shimadzu-160A spectrophotometer. NMR spectra were recorded on Bruker DRX 400 and Bruker AC 200 spectrometers

* To whom correspondence should be addressed. Tel: +30-1-7274598. Fax: +30-1-7274594. E-mail: skaltsounis@pharm.uoa.gr.

[†] University of Athens.

[‡] Université René Descartes, Paris.

[§] ICSN du CNRS.



1

Figure 2. Selected HMBC correlations and long-range ^1H – ^{15}N correlations for sarcomejine (**1**).

^1H (400 and 200 MHz) and ^{13}C (50 MHz)]; chemical shifts are expressed in parts per million (ppm) downfield from TMS. The ^1H – ^1H and the ^1H – ^{13}C NMR experiments were performed using standard Bruker microprograms. For the ^1H – ^{15}N GHMQC spectrum, data were acquired as 3072×400 data points with a total of 290 transients accumulated/ t_1 increment. Pulse widths were 8.55 μs for ^1H and 27.7 μs for the ^{15}N at powers of 0 and –3 dB. The F_1 spectral window employed was set from 100 to 400 ppm. Pulsed field gradients, gt1–gt3, had durations of 0.8 ms. Gradient pairs were optimized as 70:30:50 for ^{15}N . EIMS were determined on a HP-6890 and HRMS on a AEI MS-902 spectrometer.

Plant Material. The plant material was collected at Nouméa (New Caledonia) in May 1984. A voucher sample (Pusset-Chauvière 261) is deposited in the herbarium of the Centre ORSTOM at Nouméa, New Caledonia.

Extraction and Isolation. Extraction of alkaloids was as described by Fokialakis et al.² The crude alkaloid mixture was chromatographed over a column containing Si gel (Merck 0.04–0.06 mm; flash), using a $\text{CH}_2\text{Cl}_2/\text{MeOH}$ gradient to give seven fractions. Fraction 1 was submitted to flash chromatography on Si gel with $\text{CH}_2\text{Cl}_2/\text{MeOH}$ (99:1) to afford sarcomejine (**1**) (6 mg).

Sarcomejine (1): $[\alpha]_D^{25} +3^\circ$ (c 0.1, CH_2Cl_2); UV (MeOH) λ_{max} (log ϵ) 290 (sh), 341 (3.73), 327 (3.67) nm; ^1H NMR (CDCl_3 , 400 MHz) δ 8.45 (1H, dd, $J = 8.8, 1.5$ Hz H-5), 7.71 (1H, td, $J = 8.8, 1.5$ Hz, H-7), 7.53 (1H, dd, $J = 8.8, 1.5$ Hz, H-8), 7.42 (1H, td, $J = 8.8, 1.5$ Hz, H-6), 5.28 (1H, s, H-1'), 3.94 (3H, s, $\text{CH}_3\text{O}-2'$), 3.78 (3H, s, N– CH_3), 3.78 (3H, s, $\text{CH}_3\text{O}-1'$), 3.55

(3H, s, $\text{CH}_3\text{O}-1'$); ^{13}C NMR (CDCl_3 , 50 MHz) δ 174.7 (C-4), 168.8 (C-1''), 167.6 (C-2'), 145.9 (C-2), 141.5 (C-8a), 133.2 (C-7), 127.0 (C-5), 126.6 (C-4a), 124.6 (C-6), 120.5 (C-3), 115.7 (C-8), 78.5 (C-1'), 58.9 ($\text{CH}_3\text{O}-1'$), 53.3 ($\text{CH}_3\text{O}-1''$), 52.8 ($\text{CH}_3\text{O}-2'$), 35.9 (N– CH_3); ^{15}N NMR (CDCl_3 , 400 MHz) δ 118.3 (N-1); EIMS m/z 319 (30), 304 (80), 272 (100); HRMS m/z 319.1052 (calcd for $\text{C}_{16}\text{H}_{17}\text{O}_6\text{N}$, 319.1056).

References and Notes

- (1) Hartley, T. G. *Bull. Mus. Natl. Hist. Nat., Sec. B: Adansonia* **1986**, 8, 183–189.
- (2) Fokialakis, N.; Mitaku, S.; Mikros, E.; Skaltsounis, A. L.; Tillequin, F.; Sévenet, T. *Phytochemistry* **1999**, 52, 1745–1748.
- (3) Papageorgiou, M.; Fokialakis, N.; Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Sévenet, T. *J. Nat. Prod.* **2000**, 63, 385–386.
- (4) Skaltsounis, A. L.; Seddrati, L.; Tillequin, F.; Koch, M.; Pusset, J.; Sévenet, T. *Nat. Prod. Lett.* **1995**, 5, 281–287.
- (5) Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J.; Sévenet, T. *Nat. Prod. Lett.* **1995**, 7, 219–225.
- (6) Mitaku, S.; Pusset, J. *Plant. Med. Phytother.* **1988**, 22, 83–87.
- (7) Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J.; Chauvière, G. *J. Nat. Prod.* **1986**, 49, 1091–1095.
- (8) Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J. *Ann. Pharm. Fr.* **1989**, 47, 149–156.
- (9) Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M.; Pusset, J.; Chauvière, G. *Heterocycles* **1987**, 26, 2057–2063.
- (10) Baudouin, G.; Tillequin, F.; Koch, M.; Dau, E. T. H.; Guilhem, J.; Pusset, J.; Chauvière, G. *J. Nat. Prod.* **1985**, 48, 260–265.
- (11) Brum-Bousquet, M.; Tillequin, F.; Koch, M.; Sévenet, T. *Planta Med.* **1985**, 51, 536–537.
- (12) Brum-Bousquet, M.; Mitaku, S.; Skaltsounis, A. L.; Tillequin, F.; Koch, M. *Planta Med.* **1988**, 55, 191–192.
- (13) Martin, G. E.; Crouch, R. C.; Sharaf, M. H. M.; Schiff, P. L. *J. Nat. Prod.* **1996**, 59, 2–4.
- (14) Hadden, C. E.; Martin, G. E. *J. Nat. Prod.* **1998**, 61, 969–972.
- (15) Crouch, R. C.; Martin, G. E. *J. Heterocycl. Chem.* **1995**, 32, 1665–1669.
- (16) Martin, G. E.; Crouch, R. C. *J. Heterocycl. Chem.* **1995**, 32, 1839–1842.
- (17) Martin, G. E.; Crouch, R. C.; Andrews, C. W. *J. Heterocycl. Chem.* **1995**, 32, 1759–1766.
- (18) Martin, G. E. *J. Heterocycl. Chem.* **1997**, 34, 695–699.
- (19) Sharaf, M. H. M.; Schiff, P. L.; Tackie, A. N.; Martin, G. E. *J. Heterocycl. Chem.* **1998**, 35, 1365–1369.
- (20) Koshino, H.; Lee, I. K.; Kim, J. P.; Kim, W. G.; Uzawa, J.; Yoo, I. D. *Tetrahedron Lett.* **1996**, 37, 4549–4540.
- (21) Shin-ya, K.; Kim, J. S.; Furihata, K.; Hayakawa, Y.; Seto, H. *Tetrahedron Lett.* **1997**, 40, 7079–7082.

NP000083X