

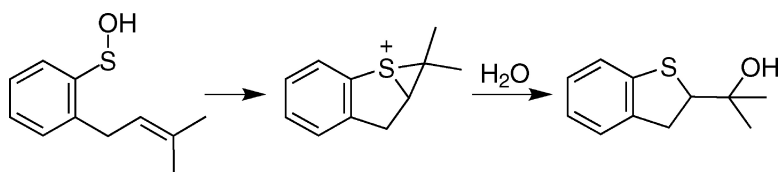
Article

Evidence for a Morin Type Intramolecular Cyclization of an Alkene with a Phenylsulfenic Acid Group in Neutral Aqueous Solution

Kripa Keerthi, Santhosh Sivaramakrishnan, and Kent S. Gates

Chem. Res. Toxicol., **2008**, 21 (7), 1368-1374 • DOI: 10.1021/tx8000187 • Publication Date (Web): 23 May 2008

Downloaded from <http://pubs.acs.org> on December 4, 2008



More About This Article

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Access to high resolution figures
- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article

[View the Full Text HTML](#)



ACS Publications
High quality. High impact.

Evidence for a Morin Type Intramolecular Cyclization of an Alkene with a Phenylsulfenic Acid Group in Neutral Aqueous Solution

Kripa Keerthi, Santhosh Sivaramakrishnan, and Kent S. Gates*

Departments of Chemistry and Biochemistry, University of Missouri—Columbia, 125 Chemistry Building, Columbia, Missouri 65211

Received January 11, 2008

Sulfenic acids (RSOH) are among the most common sulfur-centered reactive intermediates generated in biological systems. Given the biological occurrence of sulfenic acids, it is important to explore the reactivity of these intermediates under physiological conditions. The Morin rearrangement is a synthetic process developed for the conversion of penicillin derivatives into cephalosporins that proceeds via nucleophilic attack of an alkene on a sulfenic acid intermediate. In its classic form, the Morin reaction involves initial elimination of a sulfenic acid from a cyclic sulfoxide, followed by intramolecular cyclization of the resulting alkene and sulfenic acid groups to generate an episulfonium ion intermediate that undergoes further reaction to yield ring-expanded products. On the basis of the existing literature, it is difficult to assess whether the reaction between an alkene and a sulfenic group can occur under mild conditions because the conditions required to generate the sulfenic acid from the sulfoxide precursor in the Morin reaction typically involve high temperatures and strong acid. In the work described here, β -sulfinylketone precursors were used to generate a “Morin type” sulfenic acid intermediate under mild conditions. This approach made it possible to demonstrate that the intramolecular cyclization of an alkene with a phenylsulfenic acid to generate an episulfonium ion intermediate can occur in neutral aqueous solution at room temperature.

Introduction

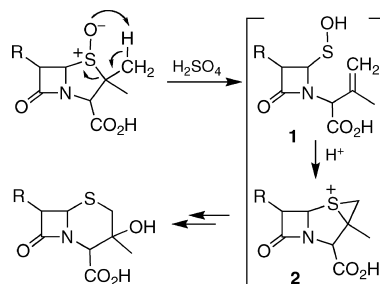
Sulfenic acids (RSOH) are among the most common sulfur-centered reactive intermediates generated in biological systems. For example, thiol groups on a wide variety of cellular proteins are converted to sulfenic acids under conditions of oxidative stress (1–14). In some cases, oxidation of key cysteine thiol residues to the corresponding sulfenic acids regulates protein function as part of normal hydrogen peroxide-mediated cell signaling processes (6–12, 14). Furthermore, sulfenic acids can be generated inside cells through the oxidation of low molecular weight thiols such as glutathione (15–17) and by the breakdown of drugs and other xenobiotics (18–32). Sulfenic acids are unstable, most commonly decomposing by reaction with nucleophiles (15, 33–36). For example, in biological systems, sulfenic acids often react with thiols to produce a disulfide linkage (eq 1) (15, 37–40).



Given the biological occurrence of sulfenic acids, it may be interesting to explore the range of nucleophiles that can react with these intermediates under physiological conditions.

The Morin rearrangement is a synthetic transformation developed for the conversion of penicillin derivatives into cephalosporins that proceeds via nucleophilic attack of an alkene on a sulfenic acid intermediate (Scheme 1) (36, 41–43). In its classic form, the Morin reaction involves initial elimination of a sulfenic acid from a cyclic sulfoxide, followed by intramolecular cyclization of the resulting alkene and sulfenic acid groups to generate an episulfonium ion intermediate (2) that undergoes further reaction to yield ring-expanded products such

Scheme 1



as that shown in Scheme 1. On the basis of existing literature, it is difficult to assess whether the reaction between an alkene and a sulfenic group can occur under mild conditions because the conditions required to generate the sulfenic acid intermediate from the sulfoxide precursor in the Morin reaction typically involve high temperatures, strong acids, or the presence of reagents such as acetic anhydride (36, 41–46). In the work described here, we employed β -sulfinylketone precursors to generate a “Morin type” sulfenic acid intermediate under mild conditions. This approach made it possible to demonstrate that the intramolecular cyclization of an alkene onto a phenylsulfenic acid moiety to generate an episulfonium ion can occur in neutral aqueous solution at room temperature.

Experimental Procedures

Materials. Reagents used were of highest purity available and were used without further purification unless otherwise noted. Materials were purchased from the following suppliers: HPLC grade solvents, Fisher; silica 60 (0.04–0.063 mm pore size) for column chromatography, Merck; and TLC plates coated with general purpose silica containing UV₂₅₄ fluorophore, Aldrich Chemical Co.; all other chemicals were purchased from Aldrich Chemical Co.

* To whom correspondence should be addressed. Tel: 573-882-6763. Fax: 573-882-2754. E-mail: gatesk@missouri.edu.

Water was distilled, deionized, and glass redistilled. All reactions were carried out under an atmosphere of nitrogen, unless otherwise noted. The oxidizing agent, dimethyl dioxirane (DMD), was freshly prepared as described (47). High-resolution mass spectrometry was performed at University of Illinois at Urbana-Champaign Mass Spectrometry facility (ESI and EI) or Washington University Mass Spectrometry facility (EI), and low-resolution mass spectrometry were performed at the University of Missouri-Columbia.

4-[2-(2-Bromo-phenylsulfanyl)ethyl]pyridine (3). This compound was prepared following the general method of Katrizky and co-workers (48). Commercially available 2-bromothiophenol (2 g, 10.6 mmol) was added to vinyl pyridine (1.6 g, 14.8 mmol) in benzene (30 mL). This mixture was refluxed overnight for 15 h. The solvent was removed under reduced pressure to give a thick, reddish brown oil. Purification of the crude compound by flash column chromatography using silica gel eluted with 2:1 hexane:ethyl acetate gave **3** (2.9 g, 95%, R_f = 0.28, in 2:1 hexane:ethyl acetate) as a thick, orange-brown oil. ^1H NMR (250 MHz, CDCl_3): δ 8.53 (d, 2H, J = 4.5 Hz), 7.57 (dd, 1H, 7.78 Hz), 7.27 (m, 2H), 7.15 (d, 2H), 7.05 (m, 1H), 3.20 (t, 2H), 2.97 (t, 2H) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 149.89, 149.03, 142.02, 134.5, 133.06, 129.34, 129.33, 126.59, 126.57, 123.87, 122.30, 34.76, 33.96, 32.56, 25.76, 17.99 ppm. HRMS (ESI) calcd for $\text{C}_{13}\text{H}_{12}\text{BrNS}$ [$\text{M} + \text{H}$] $^+$, 292.9952; found, 292.9963.

4-[2-[2-(3-Methyl-but-2-enyl)phenylsulfanyl]ethyl]pyridine (4). Compound **3** (500 mg, 1.7 mmol) in THF (20 mL) was placed in a flame-dried, argon-flushed flask. The flask was cooled to -100°C in an ether-liquid nitrogen bath. To this cooled solution, *n*-butyllithium (0.82 mL of a 2.5 M solution in hexane, 2.04 mmol) was added. The color of the solution turned dark orange yellow, and it was allowed to stir for 10 min at this temperature. Prenyl bromide (1.37 mL, 11.9 mmol) was added to this mixture, and the temperature was raised to -60°C . The mixture was allowed to stir at this temperature for 45 min. During this time, the dark orange yellow reaction mixture turned pale yellow. The reaction was quenched with 10% HCl (20 mL), and the aqueous layer was extracted with ether (3 \times 20 mL) to remove any unreacted prenyl bromide or **4**. The aqueous layer was then carefully brought to pH 7 by addition of 30% NaOH (8 mL). Care must be taken at this step because addition of too much sodium hydroxide can result in decomposition of the product. The product was extracted into ethyl acetate (3 \times 20 mL). The organic layer was washed with water (2 \times 20 mL) followed by brine (2 \times 20 mL) and then dried over anhydrous sodium sulfate to give **4** (201 mg, 42%, R_f = 0.37, in 1:1 hexane:ethyl acetate) as a pale brown oil. No further purification of this compound was necessary. ^1H NMR (250 MHz, CDCl_3): δ 8.51 (d, 2H, J = 5.43 Hz), 7.34 (m, 1H), 7.17 (m, 5H), 5.24 (m, 1H), 3.44 (d, 2H, J = 7.13 Hz), 3.15 (t, 2H), 2.91 (t, 2H), 1.72 (d, 6H, J = 6.23 Hz) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 149.71, 148.79, 141.73, 134.29, 132.88, 129.10, 129.01, 126.39, 126.32, 123.67, 122.06, 34.52, 33.69, 32.34, 25.59, 17.8 ppm. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{21}\text{NS}$ [$\text{M} + \text{H}$] $^+$, 284.1473; found, 284.1474.

3-[2-(3-Methyl-but-2-enyl)phenylsulfanyl]propionaldehyde (5a). To a solution of **4** (100 mg, 0.35 mmol) in dry, distilled acetone (5 mL) under nitrogen, methyl iodide (0.33 mL, 5.25 mmol) was added. This mixture was allowed to stir at 24°C for 3 h. The excess methyl iodide was removed by blowing nitrogen over the flask (CAUTION! Methyl iodide is carcinogenic. This must be performed in a well-ventilated hood.) Fresh dry acetone (5 mL) was added to the resulting orange yellow solid, followed by acrolein (0.046 mL, 0.70 mmol). The mixture was allowed to stir for 5 min, and then, sodium acetate (58 mg, 0.71 mmol) in water (0.2 mL) was added to the rapidly stirred mixture. The resulting green solution was allowed to stir for 18 h at room temperature. The reaction mixture was quenched by addition of water (5 mL), and the crude mixture was extracted with ether (3 \times 10 mL), water (2 \times 5 mL), and brine (2 \times 5 mL). The organic layer was dried over anhydrous sodium sulfate and evaporated to give a brown oil. This product was purified by flash column chromatography on silica gel eluted with 12:1 hexane:ethyl acetate to give **5a** (46 mg, 56%, R_f = 0.37 in 9:1 hexane:ethyl acetate) as a colorless oil. ^1H NMR (250 MHz,

CDCl_3): δ 9.77 (s, 1H), 7.34 (m, 2H), 7.17 (m, 2H), 5.24 (m, 1H), 3.45 (d, 2H, J = 7.12 Hz) 3.16 (t, 2H), 2.77 (t, 2H), 1.74 (d, 6H, J = 4.51 Hz) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 200.15, 142.12, 133.90, 133.04, 129.54, 129.26, 126.67, 126.56, 122.14, 43.05, 32.42, 26.17, 25.69, 17.91 ppm. HRMS (Cl^+) calcd for $\text{C}_{14}\text{H}_{18}\text{OS}$ [M] $^+$, 234.1078; found, 234.1080.

3-[2-(3-Methyl-but-2-enyl)phenylsulfanyl]propionic Acid Ethyl Ester (5b). To a solution of **4** (100 mg, 0.35 mmol) in acetone (5 mL) under nitrogen, excess methyl iodide (0.33 mL, 5.25 mmol) was added. This mixture was allowed to stir at 24°C for 3 h. The excess methyl iodide was removed by blowing nitrogen gas over the flask (CAUTION! Methyl iodide is carcinogenic. This must be performed in a well-ventilated hood.) Dry distilled THF (5 mL) was added to the yellow orange solid, and the mixture was cooled to -60°C using an acetone-dry ice bath. To the cold solution, sodium methoxide in methanol (0.35 mL of a 1 M solution in methanol) was added, and the mixture was stirred for 20 min. Ethyl acrylate (0.118 mL, 1.05 mmol) was added, and the reaction mixture was stirred at -60°C for an additional 45 min. [Care should be taken at this point because excess sodium methoxide will decompose the product **5b** to generate the thiophenolate. This product is stable at low temperatures (up to -30°C) but undergoes Kwart cyclization (49, 50) with the adjacent alkene at higher temperatures.] The reaction was quenched with dilute HCl (5 mL, 0.5 M) and extracted with diethyl ether (3 \times 10 mL). The organic layer was washed with water (2 \times 5 mL) and brine (2 \times 5 mL), followed by drying over anhydrous sodium sulfate. The organic solvent was evaporated under reduced pressure to give a red oil. The crude oil was purified by flash column chromatography on silica gel eluted with 9:1 hexane:ethyl acetate to give **5b** (45 mg, 44%, R_f = 0.69 in 4:1 hexane:ethyl acetate) as a colorless oil. ^1H NMR (250 MHz, CDCl_3): δ 7.35 (m, 1H), 7.17 (m, 3H), 5.25 (m, 1H), 4.13 (q, 2H), 3.46 (d, 2H, J = 7.13 Hz), 2.61 (t, 2H), 1.73 (d, 6H), 1.25 (t, 3H) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 171.73, 142.17, 134.09, 132.91, 129.74, 129.15, 126.57, 126.48, 122.24, 60.64, 34.30, 32.43, 28.85, 25.68, 17.89, 14.11 ppm. HRMS (EI) calcd for $\text{C}_{16}\text{H}_{22}\text{O}_2\text{S}$, 278.1341; found, 278.1345.

3-[2-(3-Methyl-but-2-enyl)benzenesulfinyl]propionaldehyde (6a). To a solution of **5a** (25 mg) in HPLC grade acetone (4 mL), freshly prepared DMD (2 mL, ~ 0.09 M in acetone) was added slowly. Formation of a new, more polar spot was seen on TLC. The solvent was evaporated under reduced pressure to give **6a** (24 mg, 90%, R_f = 0.19 in 1:1 hexane:ethyl acetate) as a pale brown oil. This compound was approximately 90% pure as judged by NMR; however, it was highly unstable and was used immediately in subsequent reactions. ^1H NMR (250 MHz, CDCl_3): δ 9.7 (s, 1H), 7.88 (m, 1H), 7.43 (m, 2H), 7.26 (m, 1H), 5.21 (m, 1H), 3.42 (d, 2H), 3.25 (m, 1H), 2.89 (m, 1H), 1.73 (d, 6H, J = 8.95 Hz) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 198.43, 140.89, 138.66, 134.19, 131.18, 129.74, 127.22, 123.82, 121.40, 46.91, 35.42, 30.44, 25.61, 17.99 ppm. Because of the unstable nature of this compound, mass spectrometric analysis was not possible; however, the ^1H NMR spectra clearly show a vast increase in the complexity of the splitting patterns for the protons in the propionaldehyde chain. This is indicative of introduction of a single oxygen onto the sulfur residue to generate a chiral sulfoxide that, in turn, renders hydrogens in the propionaldehyde chain diastereotopic. In contrast, in the achiral sulfide and sulfone derivatives, these proton resonances appear as simple triplets.

3-[2-(3-Methyl-but-2-enyl)benzenesulfinyl]propionic Acid Ethyl Ester (6b). This compound was prepared using the method described above for compound **6a**. Evaporation of solvent gave **6b** in 94% yield (R_f = 0.12 in 4:1 hexane:ethyl acetate) as a colorless oil. No further purification was necessary. ^1H NMR (250 MHz, CDCl_3): δ 7.90 (m, 1H), 7.43 (m, 2H), 7.23 (m, 1H), 5.22 (m, 1H), 4.13 (q, 2H), 3.40 (d, 2H, J = 7.1 Hz), 3.20 (m, 1H), 2.88 (m, 2H), 2.64 (m, 1H), 1.74 (d, 6H, J = 5.3 Hz), 1.23 (t, 3H) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 171.18, 141.02, 138.69, 134.095, 131.067, 129.58, 127.19, 123.91, 121.42, 60.99, 49.80, 30.39, 26.43, 25.60, 17.97, 14.06 ppm. HRMS (EI) calcd for $\text{C}_{16}\text{H}_{22}\text{O}_3\text{S}$, 294.1290; found, 294.1295.

Reaction of 6a in Aqueous Neutral Aqueous Solution. To a rapidly stirred solution of **6a** (20 mg, 0.08 mmol) in acetonitrile (2.5 mL), sodium phosphate buffer (0.5 mL of a 500 mM, pH 7.4) and water (2 mL) were added. The reaction mixture was stirred for 10 h (final concentrations: **6a**, 16 mM; buffer, 50 mM, pH 7.4; and acetonitrile, 50% by volume). Water (5 mL) was added, and the resulting mixture was extracted with diethyl ether (3 × 5 mL). The ether extracts were combined and washed with water (1 × 5 mL) and brine (1 × 5 mL). The solution was dried over anhydrous sodium sulfate and concentrated under reduced pressure to yield a pale yellow oil. Flash column chromatography on silica gel eluted with 9:1 hexane:ethyl acetate provided **9a** as a colorless oil (3.9 mg, 25%, R_f = 0.35 in 4:1 hexane:ethyl acetate) and **10** as a pale yellow oil (9.9 mg, 35%, R_f = 0.65 in 4:1 hexane:ethyl acetate). Compound **9a**: ^1H NMR (250 MHz, CDCl_3): δ 7.14–7.09 (m, 3H), 6.98 (t, 1H), 4.07 (dd, 1H, J = 3.85 Hz, 4.40 Hz), 3.37 (dd, 1H, J = 3.85 Hz, 8.03 Hz), 3.30 (dd, 1H, J = 4.42 Hz, 8.01 Hz), 2.196 (s, 1H), 1.26 (d, 6H, J = 11.81) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 139.97, 139.88, 127.24, 124.32, 121.68, 71.68, 61.78, 37.17, 28.80, 25.97 ppm. HRMS (EI) calcd for $\text{C}_{11}\text{H}_{14}\text{OS}$, 194.0765; found, 194.0765. Compound **10**: ^1H NMR (250 MHz, CDCl_3): δ 7.55 (m, 1H), 7.17–7.09 (m, 3H), 5.25 (m, 1H), 3.47 (d, 2H, J = 5.98 Hz), 1.73 (d, 6H, J = 6.18 Hz) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 140.87, 135.28, 133.29, 129.04, 128.89, 127.33, 126.72, 121.86, 65.78, 32.21, 25.68, 17.93, 15.20 ppm. HRMS (EI) calcd for $\text{C}_{22}\text{H}_{26}\text{S}_2$, 354.1476; found, 354.1472.

Trapping of the Sulfenic Acid Intermediate 7 by Methyl Iodide. To a stirred solution of **6a** (20 mg, 0.08 mmol) in acetonitrile (2.5 mL), sodium phosphate buffer (0.5 mL of a 500 mM, pH 7.4), excess methyl iodide (1 mL), and water (1 mL) were added. The reaction mixture was stirred for 10 h (final concentrations: **6a**, 16 mM; buffer, 50 mM, pH 7.4; and acetonitrile, 50% by volume). The methyl iodide was removed by blowing nitrogen over the reaction mixture (CAUTION! Methyl iodide is carcinogenic, and this procedure must be performed in a well-ventilated hood.) Water (5 mL) was added, and the resulting mixture was extracted with diethyl ether (3 × 5 mL). The ether extracts were combined and washed with water (1 × 5 mL) and brine (1 × 5 mL). The solution was dried over anhydrous sodium sulfate and concentrated under reduced pressure to yield a dark yellow oil. Flash column chromatography on silica gel eluted with 1:1 hexane:ethyl acetate gave **12** (3 mg, 18%, R_f = 0.18 in 1:1 hexane:ethyl acetate) as a colorless oil. IR (CHCl_3): 1021 cm^{-1} (S=O). ^1H NMR (250 MHz, CDCl_3): δ 7.98 (d, 1H), 7.47–7.22 (m, 2H), 7.23 (m, 1H), 5.18 (m, 1H), 3.40 (m, 2H), 2.67 (s, 3H), 1.73 (d, 6H, J = 2 Hz) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 143.95, 138.19, 133.83, 130.95, 129.45, 127.61, 123.12, 121.71, 42.98, 30.65, 25.60, 17.96 ppm. HRMS (EI) calcd for $\text{C}_{12}\text{H}_{16}\text{OS}$, 208.0922; found, 208.0916. In addition, the alcohol **9a** (1 mg, 7%) and the disulfide **10** (8 mg, 28%) were obtained.

Reaction of 6a in 90% Methanol–10% Aqueous Buffer. To a rapidly stirred solution of **6a** (20 mg, 0.08 mmol) in methanol (4.5 mL), sodium phosphate buffer (0.5 mL of a 500 mM, pH 7.4) was added. The reaction mixture was stirred for 20 h (final concentrations: **6a**, 16 mM; buffer, 50 mM, pH 7.4; and methanol, 90% by volume). Water (5 mL) was added, and the resulting mixture was extracted with diethyl ether (3 × 5 mL). The ether extracts were combined and washed with water (1 × 5 mL) and brine (1 × 5 mL). The solution was dried over anhydrous sodium sulfate and concentrated under reduced pressure to yield a colorless oil. Flash column chromatography on silica gel eluted with 9:1 hexane:ethyl acetate yielded **9a** (4.4 mg, 28%, 3.9 mg, 25%, R_f = 0.35 in 4:1 hexane:ethyl acetate) and **10** (8 mg, 28%). Reactions containing lithium perchlorate were conducted as described above except that LiClO_4 (26.5 mg) was added following addition of the sodium phosphate buffer. The reaction was worked up as described above. Flash column chromatography on silica gel eluted with 12:1 hexane:ethyl acetate yielded **9b** (mg, 25%, R_f = 0.55 in 4:1 hexane:ethyl acetate) as a colorless oil. ^1H NMR (250 MHz, CDCl_3): 7.16–7.11 (3H, m), 7.00 (t, 1H), 4.13 (dd, 1H), 3.32 (m, 2H), 3.29 (s, 3H), 1.27 (s, 3H), 1.21 (s, 3H) ppm. ^{13}C NMR (62.9 MHz,

CDCl_3): δ 141.81, 140.47, 127.96, 124.98, 124.75, 122.40, 77.89, 58.91, 50.39, 37.91, 23.00, 22.22 ppm. HRMS (EI) calcd for $\text{C}_{12}\text{H}_{16}\text{OS}$, 208.0907; found, 208.0922. In addition, compounds **9a** (trace) and **10** (9 mg, 33%) were also formed. A control reaction shows that a solution of **9a** (16 mg, 0.08 mmol) stirred for 15 h in methanol (4.5 mL), sodium phosphate buffer (0.5 mL of a 500 mM, pH 7.4), and lithium perchlorate (26.5 mg, 0.25 mmol) and worked up as described above returns only starting material (lithium perchlorate does not catalyze exchange of methanol into the product).

Thermolysis of 6b in CCl_4 . A solution of compound **6b** (20 mg, 0.08 mmol) in carbon tetrachloride (5 mL) was placed in a pressure tube with a Teflon screw cap (Ace Glass) equipped with a stir bar. The reaction mixture was heated at 100 °C for 3 h with rapid stirring. The solvent was then evaporated to give a pale yellow oil. Purification on silica gel eluted with 9:1 hexane ethyl acetate gave **9a** (8.5 mg, 55%) as a white waxy solid. In addition, **10** (3.4 mg, 12%) was also obtained as a pale yellow oil.

Thermolysis of 6b in CCl_4/MeOH . Compound **6b** (20 mg, 0.08 mmol) was placed in a sealed tube with a magnetic bar, carbon tetrachloride (4 mL), and methanol (1 mL). The mixture was heated at 80 °C for 10 h with rapid stirring. The reaction mixture was then transferred to a round-bottom flask, and the solvent was evaporated under reduced pressure. The crude products were purified by flash column chromatography on silica gel eluted with 9:1 hexane:ethyl acetate as an eluant, and **9b** (7.4 mg, 34%) in addition to a small amount of **9a** (1.5 mg, 9%) and **10** (5.6 mg, 16%) also formed.

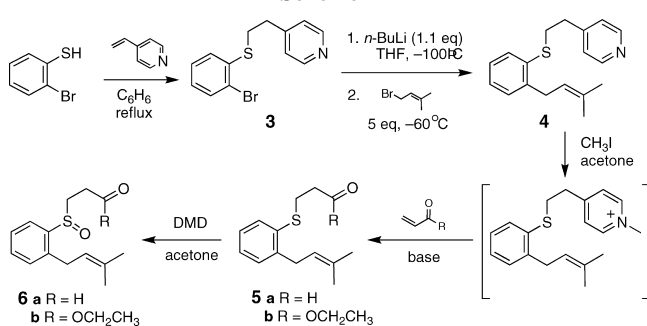
Trapping of Sulfenic Acid 7 by Methyl Propiolate in CCl_4 . Compound **6b** (20 mg, 0.08 mmol) was placed in a sealed tube with a magnetic bar, carbon tetrachloride (4 mL), and methyl propiolate (1 mL). The mixture was heated at 100 °C for 3 h with rapid stirring. The reaction mixture was transferred to a round-bottom flask, and the solvent was evaporated under reduced pressure. Flash column chromatography eluted with 6:1 hexane:ethyl acetate afforded compound **13** (17 mg, 75%, R_f = 0.37 in 4:1 hexane:ethyl acetate) as a colorless oil. ^1H NMR (250 MHz, CDCl_3): δ 7.75 (m, 1H), 7.54 (d, 2H, J = 7.1 Hz), 7.41 (m, 2H), 7.28 (m, 1H), 6.72 (d, 1H, J = 7.5 Hz), 5.21 (m, 1H), 3.77 (s, 3H), 3.62–3.45 (m, 2H), 1.74 (d, 6H, J = 6 Hz) ppm. ^{13}C NMR (62.9 MHz, CDCl_3): δ 164.54, 150.90, 139.79, 139.56, 134.25, 131.72, 130.08, 127.97, 124.66, 123.31, 121.85 ppm. HRMS (EI) calcd for $\text{C}_{15}\text{H}_{18}\text{O}_3\text{S}$, 292.1133; found, 292.1130.

Results

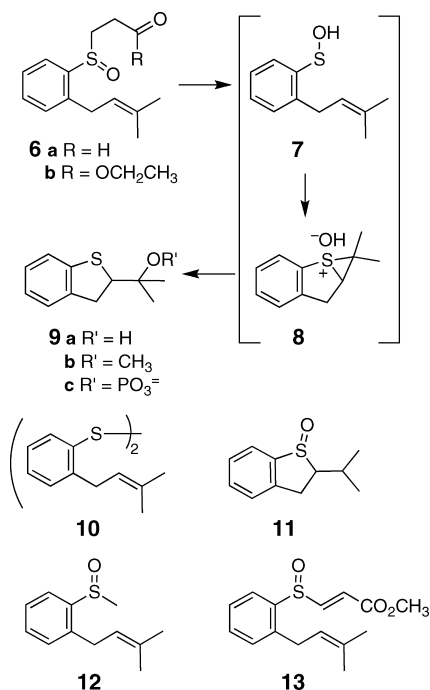
Design of Sulfenic Acid Precursors. To address the question of whether a sulfenic acid can undergo intramolecular reaction with an alkene under mild conditions to generate an alkylating intermediate, we set out to characterize the reactivity of [*ortho*-(3-methylbut-2-enyl)phenyl]sulfenic acid (**7**). Typically, it is not possible to isolate sulfenic acids due to their instability (33, 51–54); however, methods exist for the in situ generation of this functional group (12, 22, 32, 48, 55–61). In the present study, we employed a β -phenylsulfinyl aldehyde **6a** and a β -phenylsulfinyl ester derivative **6b** as phenylsulfenic acid precursors. β -Sulfinylketones such as these are known to release sulfenic acids under mild conditions via β -elimination reactions (12, 22, 55, 56).

Synthesis. We prepared the desired *ortho*-(3-methylbut-2-enyl)-substituted phenylsulfenic acid precursors **6a** and **6b** starting from commercially available 2-bromobenzenethiol (Scheme 2). The thiol substituent was masked with the ethylpyridine group by treatment with vinyl pyridine in refluxing benzene to give **3** in good yield (95%) (48). Protection of the thiol group is necessary prior to installation of the prenyl side chain because aromatic thiols have the potential to undergo intramolecular cyclization reactions with an adjacent alkene (49, 50). Lithium-halogen exchange at –100 °C, followed by treatment with prenyl bromide for 1 h at –60 °C, provided the alkene-

Scheme 2



Scheme 3



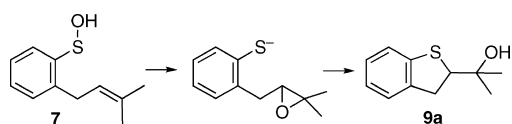
containing derivative **4** in reasonable yield (42%). The ethylpyridine group was replaced with the 3-oxopropyl side chain in a single pot by treatment of **4** with methyl iodide in acetone (**48**), followed by acrolein and sodium acetate in water to give **5a** in 56% yield. The resulting sulfide (**5a**) was oxidized with DMD in acetone (**47**) to generate the desired sulfenic acid precursor **6a**. This compound was rather unstable and was used immediately in the studies described below. We prepared an additional sulfenic acid precursor by treatment of **4** with methyl iodide in acetone, followed by sodium methoxide and ethyl acrylate in methanol at -60°C to provide **5b** in 44% yield. Oxidation with DMD provided a 94% yield of the desired β -phenylsulfinyl ester **6b**.

Reactions of the Sulfenic Acid 7. Stirring the sulfenic acid precursor **6a** (16 mM) in a solvent mixture composed of 1:1 acetonitrile:sodium phosphate buffer (50 mM, pH 7.4) at room temperature led to the formation of the 2,3-dihydrobenzo[b]thiophene derivative **9a** (25%) along with the disulfide **10** (30%). Compound **9a** is envisioned to arise via a Morin type cyclization reaction (Scheme 3), while the disulfide **10** is an expected byproduct of sulfenic acid dimerization (51, 62). It is possible that **9a** arises via hydrolysis of an initially formed product **9** resulting from reaction of phosphate with the episulfonium ion **8** (63). A related route to hydrolysis products, via a sulfate derivative, has been reported in the context of a sulfuric acid-catalyzed Morin rearrangement (see structure 56 in ref 43). Polar material that could correspond to the phosphate-

trapped episulfonium ion **9c** was observed by TLC analysis in the early stages of the reaction but was not isolated and characterized. Sulfenic acids have been observed to undergo sigmatropic addition reactions with alkenes (15, 33, 46, 52, 64–67); however, the product (**9a**) observed here is structurally (and spectroscopically) quite distinct from the compound (**11**) that would result from such a process in this molecular framework. The regiochemistry of this reaction (Scheme 3) can be rationalized as Markovnikov addition of the nucleophile to the more substituted carbon of the episulfonium ion, similar to the Morin process shown in Scheme 1. When the reaction was conducted in the presence of excess methyl iodide, the sulfoxide product (**12**) expected (53, 68) to arise from alkylation of the sulfenic acid intermediate **7** was generated in 20% yield alongside **9a** (7%) and **10** (28%). This result confirms the intermediacy of the sulfenic acid **7** in the reactions of **6a**.

Surprisingly, when **6a** was placed in a solvent mixture composed of 90:10 methanol–phosphate buffer (50 mM, pH 7), the alcohol **9a** was again obtained as the only isolable cyclized product. We had anticipated that under these reaction conditions, methanol would capture the episulfonium ion intermediate to yield **9b** (31, 32). The inability of methanol to trap the putative episulfonium ion **8** could indicate that this intermediate is generated as an ion pair that favors attack of hydroxide to yield the alcohol **9a**. Examples have been reported in which episulfonium ions are formed as ion pairs that evade reaction with bulk acetic acid (69–72); however, it should be noted that more polar solvents such as the methanol–water mixture employed here typically do not favor such ion pairing (73). Addition of salts such as lithium perchlorate can disrupt ion pairs, thus allowing reaction of the electrophilic intermediate with solvent (71, 72, 74). Indeed, addition of lithium perchlorate (50 mM) to our reaction led to the formation of the methanolysis product **9b** (25%), alongside **10** (33%) and a trace of **9a**. A control experiment confirmed that **9b** does not arise via exchange of methanol into **9a** under these reaction conditions. The origin of the lithium perchlorate salt effect, in this situation, is not completely clear. As mentioned above, the addition of LiClO_4 may disrupt ion pairing via a special salt effect. Certainly, in the case of other episulfonium ions, the addition of lithium perchlorate (~ 400 mM) can cause significant shifts away from reactions within an ion pair and toward reaction with the bulk solvent, but again, these examples were carried out in acetic acid, which is more supportive of ion pairing (71). Alternatively, the effect of added LiClO_4 on our reaction could arise through a kinetic salt effect (75). The term kinetic salt effect is used, for example, to describe the observation that reaction rates between two substrates of opposing charge decrease with increasing ionic strength of the reaction media. In our case, if the hydrolysis product **9a** arises via the reaction of oppositely charged phosphate and episulfonium ions to yield **9c**, the rate of this reaction is expected to decrease in the presence of added salt. Such a rate decrease in the pathway leading to the hydrolysis product **9a** could allow reaction of **8** with the neutral nucleophile methanol to become predominant. Finally, the inability to obtain a methanol adduct in the original reaction could reflect that the sulfenic acid **7** carries out a peroxide-like oxygenation of the alkene (this could be either intramolecular or intermolecular) followed by attack of the resulting thiol on the intermediate epoxide to yield **9a** (Scheme 4). Such peroxidic behavior of sulfenic acids is unprecedented to the best of our knowledge. Nonetheless, it is conceivable that **9a** arises via a peroxidic mechanism and that addition of LiClO_4 causes a shift in mechanism away from this route

Scheme 4



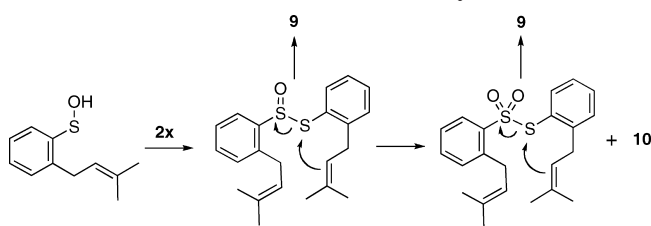
(Scheme 4) toward the Morin type reaction pathway that yields the episulfonium ion **8** (Scheme 3). While elucidation of the exact origin of the salt effect on this reaction awaits further study, the results, especially in light of those presented in the following paragraphs (which provide no evidence for peroxidic behavior of the sulfenic acid intermediate), are most consistent with the generation of an episulfonium ion **8** from the sulfenic acid **7**.

The sulfenic acid precursor **6b** is quite stable as compared to **6a**. For example, compound **6b** does not undergo decomposition even upon extended exposure (24 h) to the reaction conditions employed for the reactions of **6a**. The stability of **6b** against β -elimination is not surprising given the lower acidity of the proton α to the carbonyl in **6b** vs **6a** ($pK_a \sim 30$ vs ~ 22). A reaction was induced by heating a carbon tetrachloride solution of **6b** in a sealed tube at 100 °C for 3 h. This produced **9a** in 55% yield along with the disulfide **10** (12%). In this case, there was no difficulty in trapping the episulfonium intermediate with methanol. When the reaction was conducted in a 1:1 mixture of CCl₄/CH₃OH at 80 °C, the methanol-trapped product **9b** was isolated in 34% yield alongside the alcohol **9a** in 9% yield. The addition of methyl propiolate to the reaction led to the formation the vinyl sulfoxide **13** (75%) expected (15, 33, 66, 67) to arise from trapping of the sulfenic acid intermediate **7**. Again, this confirms the intermediacy of the sulfenic acid **7** in the reactions of **6b**.

Discussion

In the work reported here, a phenylsulfenic acid tethered to an alkene moiety (**7**) was generated from a β -phenylsulfenyl aldehyde precursor at room temperature in neutral aqueous solution. The resulting products can be explained by a mechanism involving intramolecular trapping of the sulfenic acid by the neighboring alkene to generate an episulfonium ion intermediate (**8**, Scheme 3) (76, 77).^{1,2} Sulfenic acids readily undergo dehydrative dimerization to yield thiosulfates (51, 62) that, in turn, disproportionate to the disulfide and thiosulfonate derivatives (62). In principle, generation of the electrophilic episulfonium ion **8** (or an equivalent carbocationic intermediate) could proceed via attack of the alkene on either thiosulfinate or thiosulfonate intermediates, with ejection of sulfinate or sulfonate leaving groups, respectively (Scheme 5). However, to the best of our knowledge, there is no precedent for such a reaction. On the contrary, in the context of molecules such as allicin (78) and pseudoallicin (79), thiosulfinate and thiosulfonate groups, respectively, coexist stably in the presence of an alkene moiety.

Episulfonium ions are of special relevance in the fields of medicinal chemistry and toxicology because these intermediates are highly reactive alkylating agents that often possess potent

Scheme 5. Alternatives to the Direct Cyclization of **8**

biological activities (30, 80–83). The general reaction characterized in this work might be applied to the development of biologically active episulfonium alkylating agents (30, 80, 82, 83).

Acknowledgment. We thank the National Institutes of Health (CA 83925 and CA 119131) for support of this research. In addition, we are grateful to Professor Richard Loeppky (University of Missouri), Professor Marc Greenberg (Johns Hopkins), and the reviewers for helpful discussions.

References

- (1) Saurin, A. T., Neubert, H., Brennan, J. P., and Eaton, P. (2004) Widespread sulfenic acid formation in tissues in response to hydrogen peroxide. *Proc. Nat. Acad. Sci. U.S.A.* 101, 17982–17987.
- (2) Charles, R. L., Schroder, E., May, G., Free, P., Gaffney, P. R. J., Wait, R., Begum, S., Heads, R. J., and Eaton, P. (2007) Protein sulfenylation as a redox sensor: proteomics studies using a novel biotinylated dione analogue. *Mol. Cell. Proteomics* 6, 1473–1484.
- (3) Poole, L. B., Klomsiri, C., Knaggs, S. A., Furdai, C. M., Nelson, K. J., Thomas, M. J., Fetrow, J. S., Daniel, L. W., and King, S. B. (2007) Fluorescent and affinity-based tools to detect cysteine sulfenic acid formation in proteins. *Bioconjugate Chem.* 18, 2004–2017.
- (4) Takanishi, C. L., Ma, L.-H., and Wood, M. J. (2007) A genetically encoded probe for cysteine sulfenic acid protein modification in vivo. *Biochemistry* 46, 14725–14732.
- (5) Allison, W. S. (1976) Formation and reactions of sulfenic acids in proteins. *Acc. Chem. Res.* 9, 293–299.
- (6) Claiborne, A., Yeh, J. I., Mallet, T. C., Luba, J., Crane, E. J., Charrier, V., and Parsonage, D. (1999) Protein-sulfenic acids: Diverse roles for an unlikely player in enzyme catalysis and redox regulation. *Biochemistry* 38, 15407–15416.
- (7) Poole, L. B., Karplus, P. A., and Claiborne, A. (2004) Protein sulfenic acids in redox signaling. *Annu. Rev. Pharmacol. Toxicol.* 44, 3325–347.
- (8) Stamler, J. S., and Hausladen, A. (1998) Oxidative modifications in nitrosative stress. *Nat. Struct. Biol.* 5, 247–249.
- (9) Rhee, S. G. (2006) H₂O₂, a necessary evil for cell signaling. *Science* 312, 1882–1883.
- (10) Salmeen, A., Anderson, J. N., Myers, M. P., Meng, T.-C., Hinks, J. A., Tonks, N. K., and Barford, D. (2003) Redox regulation of protein tyrosine phosphatase 1B involves a sulphenyl-amide intermediate. *Nature* 423, 769–773.
- (11) van Montfort, R. L. M., Congreve, M., Tisi, D., Carr, R., and Jhoti, H. (2003) Oxidation state of the active-site cysteine in protein tyrosine phosphatase 1B. *Nature* 423, 773–777.
- (12) Sivaramakrishnan, S., Keerthi, K., and Gates, K. S. (2005) A chemical model for the redox regulation of protein tyrosine phosphatase 1B (PTP1B). *J. Am. Chem. Soc.* 127, 10830–10831.
- (13) Sarma, B. K., and Mughes, G. (2007) Redox regulation of protein tyrosine phosphatase 1B (PTP1B): A biomimetic study on the unexpected formation of a sulphenyl amide intermediate. *J. Am. Chem. Soc.* 129, 8872–8881.
- (14) Denu, J. M., and Tanner, K. G. (1998) Specific and reversible inactivation of protein tyrosine phosphatases by hydrogen peroxide: Evidence for a sulfenic acid intermediate and implications for redox regulation. *Biochemistry* 37, 5633–5642.
- (15) Davis, F. A., and Bilmers, R. L. (1981) Chemistry of sulfenic acids. 4. The first direct evidence for the involvement of sulfenic acids in the oxidation of thiols. *J. Am. Chem. Soc.* 103, 7016–7018.
- (16) Winterbourn, C. C., and Metodiewa, D. (1999) Reactivity of biologically important thiol compounds with superoxide and hydrogen peroxide. *Free Radical Biol. Med.* 27, 322–328.
- (17) Zhang, N., Schuchmann, H.-P., and von Sonntag, C. (1991) Reaction of superoxide radical anion with dithiothreitol: A chain process. *J. Phys. Chem.* 95, 4718–4722.

¹ In the course of thiol-triggered DNA alkylation by the natural product leinamycin, a sulfenate anion is generated proximal to an alkene unit (29, 32). In this case, however, the sulfenate does not react with the alkene moiety but instead undergoes facile intramolecular cyclization onto a neighboring dithioester group.

² For two reports in which an electrophilic intermediate was generated via intramolecular reaction of a sulfenic acid with an adjacent alkene in the absence of an acid catalyst or anhydride, see refs 76 and 77.

- (18) Rose, P., Whiteman, M., Moore, P. K., and Zhu, Y. Z. (2005) Bioactive S-alk(en)yl cysteine sulfoxide metabolites in the genus *Allium*: The chemistry of potential therapeutic agents. *Nat. Prod. Rep.* 22, 351–368.
- (19) Kubec, R., Velisek, J., and Musah, R. A. (2002) The amino acid precursors and odor formation in society garlic *Tulbaghia violacea* Harv. *Phytochemistry* 60, 21–25.
- (20) Kruger, U., Senn-Bilfinger, J., Sturm, E., Figala, V., Klemm, K., Kohl, B., Rainer, G., and Schaefer, H. (1990) (H⁺, K⁺)-ATPase inhibiting 2-[(2-pyridylmethyl)sulfinyl]benzimidazoles. 3. Evidence for the involvement of a sulfenic acid in their reactions. *J. Org. Chem.* 55, 4163–4168.
- (21) Doerge, D. R., Decker, C. J., and Takazawa, R. S. (1993) Chemical and enzymatic oxidation of benzimidazoline-2-thiones: A dichotomy in the mechanism of peroxidase inhibition. *Biochemistry* 32, 58–65.
- (22) Hashmi, M., Vamvakas, S., and Anders, M. W. (1992) Bioactivation mechanism of S-(3-oxopropyl)-N-acetyl-L-cysteine, the mercapturic acid of acrolein. *Chem. Res. Toxicol.* 5, 360–365.
- (23) Kassahun, K., Pearson, P. G., Tang, W., McIntosh, I., Leung, K., Elmore, C., Dean, D., Wang, R., Doss, G., and Baillie, T. A. (2001) Studies on the metabolism of troglitazone to reactive intermediates in vitro and in vivo. Evidence for novel biotransformation pathways involving quinone methide formation and thiazolidinedione ring scission. *Chem. Res. Toxicol.* 14, 62–70.
- (24) Behrooz, S. B., Kim, W., and Gates, K. S. (1995) The reaction of *n*-propanethiol with 3H-1,2-benzodithiol-3-one 1-oxide and 5,5-dimethyl-1,2-dithiolan-3-one 1-oxide: Studies related to the reaction of antitumor antibiotic leinamycin with DNA. *J. Org. Chem.* 60, 3964–3966.
- (25) Asai, A., Hara, M., Kakita, S., Kanda, Y., Yoshida, M., Saito, H., and Saitoh, Y. (1996) Thiol-mediated DNA alkylation by the novel antitumor antibiotic leinamycin. *J. Am. Chem. Soc.* 118, 6802–6803.
- (26) Kim, W., Dannaldson, J., and Gates, K. S. (1996) Reactions of 3H-benzodithiol-3-one 1-oxide with amines and anilines. *Tetrahedron Lett.* 37, 5337–5340.
- (27) Breydo, L., Zang, H., Mitra, K., and Gates, K. S. (2001) Thiol-independent DNA alkylation by leinamycin. *J. Am. Chem. Soc.* 123, 2060–2061.
- (28) Zang, H., Breydo, L., Mitra, K., Dannaldson, J., and Gates, K. S. (2001) DNA alkylation by leinamycin can be triggered by cyanide and phosphines. *Bioorg. Med. Chem. Lett.* 11, 1511–1515.
- (29) Breydo, L., and Gates, K. S. (2002) Thiol-triggered activation of leinamycin: A theoretical study. *J. Org. Chem.* 67, 9054–9060.
- (30) Gates, K. S. (2000) Mechanisms of DNA damage by leinamycin. *Chem. Res. Toxicol.* 13, 953–956.
- (31) Chatterji, T., Kizil, M., Keerthi, K., Chowdhury, G., Posposil, T., and Gates, K. S. (2003) Small molecules that mimic the thiol-triggered alkylating properties seen in the natural product leinamycin. *J. Am. Chem. Soc.* 125, 4996–4997.
- (32) Keerthi, K., and Gates, K. S. (2007) Entering the leinamycin rearrangement via 2-(trimethylsilyl)ethyl sulfoxides. *Org. Biomol. Chem.* 5, 1595–1600.
- (33) Goto, K., Holler, M., and Okazaki, R. (1997) Synthesis, structure, and reactions of a sulfenic acid bearing a novel bowl-type substituent: The first synthesis of a stable sulfenic acid by direct oxidation of a thiol. *J. Am. Chem. Soc.* 119, 1460–1461.
- (34) Goto, K., Tokitoh, N., and Okazaki, R. (1995) Synthesis of a stable arenesulfenic acid bearing a bowl-shaped macrobicyclic cyclophane skeleton. *Angew. Chem., Int. Ed. Engl.* 34, 1124–1126.
- (35) Kice, J. L., and Cleveland, J. P. (1973) Nucleophilic substitution reactions involving sulfenic acids and sulfinyl derivatives. The nucleophile- and acid-catalyzed oxygen-18 exchange of phenyl benzenethiolsulfinate. *J. Am. Chem. Soc.* 95, 104–109.
- (36) Fekner, T., Baldwin, J. E., Adlington, R. M., and Schofield, C. J. (1998) Unusually stable azetidine sulfenic acids. *Tetrahedron Lett.* 39, 6983–6986.
- (37) Sohn, J., and Rudolph, J. (2003) Catalytic and chemical competence of regulation of Cdc25 phosphatase by oxidation/reduction. *Biochemistry* 42, 10060–10070.
- (38) Lee, S.-R., Yang, K.-S., Kwon, J., Lee, C., Jeong, W., and Rhee, S. G. (2002) Reversible inactivation of the tumor suppressor PTEN by H₂O₂. *J. Biol. Chem.* 277, 20336–20342.
- (39) Buhrman, G., Parker, B., Sohn, J., Rudolph, J., and Mattos, C. (2005) Structural mechanism of oxidative regulation of the phosphatase Cdc25B via an intramolecular disulfide bond. *Biochemistry* 44, 5307–5316.
- (40) Burgoyne, J. R., Madhani, M., Cuello, F., Charles, R. L., Brennan, J. P., Schröder, E., Browning, D. D., and Eaton, P. (2007) Cysteine redox sensor in PKG1a enables oxidant-induced activation. *Science* 317, 1393–1397.
- (41) Morin, R. B., Jackson, B. G., Mueller, R. A., Lavagnino, E. R., Scanlon, W. B., and Andrews, S. L. (1963) Chemistry of cephalosporin antibiotics. III. Chemical correlation of penicillin and cephalosporin antibiotics. *J. Am. Chem. Soc.* 85, 1896–1897.
- (42) Stoodley, R. J. (1975) Rearrangements of penicillanic acid derivatives. *Tetrahedron* 31, 2321–2345.
- (43) Sammes, P. G. (1976) Recent chemistry of the beta-lactam antibiotics. *Chem. Rev.* 76, 113–155.
- (44) Freed, J. D., Hart, D. J., and Magomedov, N. A. (2001) Trapping of the putative cationic intermediate in the Morin rearrangement with carbon nucleophiles. *J. Org. Chem.* 66, 839–852.
- (45) Hart, J. D., and Magomedov, N. (1999) Spiroquinazoline support studies: new cascade reactions based on the Morin rearrangement. *J. Org. Chem.* 64, 2990–2991.
- (46) Makisumi, Y., Takada, S., and Matasukura, Y. (1974) Thio-Claisen rearrangement of allyl aryl sulfoxides. *J. Chem. Soc. Chem. Commun.* 850.
- (47) Adam, W., Bialas, J., and Hadjirapoglou, L. (1991) A convenient preparation of acetone solutions of dimethyldioxirane. *Chem. Ber.* 124, 2377.
- (48) Katritzky, A. R., Takahashi, I., and Marson, C. M. (1986) 2-(4-Pyridyl)ethyl as a protecting group for sulfur functionality. *J. Org. Chem.* 51, 4914–4920.
- (49) Kwart, H., and Evans, R. E. (1966) The thio-Claisen rearrangement. The mechanism of thermal rearrangement of allyl aryl sulfides. *J. Org. Chem.* 31, 413–419.
- (50) Kwart, H., and Schwartz, J. L. (1974) Mechanism of the catalyzed thio-claisen reaction. Triggering of concerted rearrangement processes. *J. Org. Chem.* 39, 1575–1583.
- (51) Davis, F. A., Jenkins, L. A., and Billmers, R. L. (1986) Chemistry of sulfenic acids. 7. Reason for the high reactivity of sulfenic acids. Stabilization by intramolecular hydrogen bonding and electronegativity effects. *J. Org. Chem.* 51, 1033–1040.
- (52) Braverman, S. (1990) Rearrangements involving sulfenic acids and their derivatives. In *The Chemistry of Sulphenic Acids and Their Derivatives* (Patai, S., Ed.) pp 311–359, John Wiley and Sons, Ltd., New York.
- (53) O'Donnell, J. S., and Schwan, A. L. (2004) Generation, structure and reactions of sulfenic acid anions. *J. Sulfur Chem.* 25, 183–211.
- (54) Okuyama, T., Miyake, K., Fueno, T., Yoshimura, T., Soga, S., and Tsukurimichi, E. (1992) Equilibrium and kinetic studies of reactions of 2-methyl-2-propanesulfenic acid. *Heteroatom Chem.* 3, 577–583.
- (55) Cubbage, J. W., Guo, Y., McCulla, R. D., and Jenks, W. S. (2001) Thermolysis of alkyl sulfoxides and derivatives: A comparison of experiment and theory. *J. Org. Chem.* 66, 8722–8736.
- (56) Adams, H., Anderson, J. C., Bell, R., Neville Jones, D., Peel, M. R., and Tomkinson, N. C. O. (1998) The synthesis and Diels–Alder reactions of (*E*)- and (*Z*)-1-methoxy-3-(phenylsulfinyl)buta-1,3-dienes. *J. Chem. Soc. Perkin 1*, 3967–3973.
- (57) O'Donnell, J. S., and Schwan, A. L. (2003) beta-Sulfinyl acrylate esters as a convenient source of alkane- and arenesulfenate anions. *Tetrahedron Lett.* 44, 6293–6296.
- (58) Caupene, C., Boudou, C., Perrio, S., and Metzner, P. (2005) Remarkably mild and simple preparation of sulfenate anions from beta-sulfinylesters: A new route to enantioenriched sulfoxides. *J. Org. Chem.* 70, 2812–2815.
- (59) Nagatsugi, F., Kawasaki, T., Usui, D., Maeda, M., and Sasaki, S. (1999) Highly efficient and selective cross-linking to cytidine based on a new strategy for auto-activation within a duplex. *J. Am. Chem. Soc.* 121, 6753–6754.
- (60) Aversa, M. C., Barattucci, A., Bonaccorsi, P., and Giannetto, P. (2005) L-Cysteine, a versatile source of sulfenic acids. Synthesis of enantiopure alliin analogues. *J. Org. Chem.* 70, 1986–1992.
- (61) Foucoun, F., Caupene, C., Lohier, J.-F., Sopkava de Oliveira Santos, J., Perrio, S., and Metzner, P. (2007) 2-(Trimethylsilyl)ethyl sulfoxides as a convenient source of sulfenate ions. *Synthesis* 1315–1324.
- (62) Kice, J. L., and Rogers, T. E. (1974) Mechanism of the alkaline hydrolysis of aryl thiolsulfonates and thiolsulfonates. *J. Am. Chem. Soc.* 96, 8009–8015.
- (63) Noort, D., Platenburg, D. H. J. M., and Benschop, H. P. (1996) Solid phase synthesis of peptides containing a phosphoserine-sulfur mustard adduct. *Bioorg. Med. Chem. Lett.* 6, 2007–2012.
- (64) Barrett, A. G. M., Barton, D. H. R., and Nagubandi, S. (1980) Preparation and trapping of sulphenic acids. *J. Chem. Soc. Perkin Trans 1* 237–239.
- (65) Jones, D. N., Hill, D. R., Lewton, D. A., and Sheppard, C. (1977) Synthesis of sulfoxides by intramolecular and intermolecular addition of sulphenic acids to olefins and dienes. *J. Chem. Soc. Perkin Trans. 1* 1574–1587.
- (66) Mitra, K., Barnes, C. L., and Gates, K. S. (1999) Crystal structure of methyl *trans*-3-[(2-methoxycarbonyl)phenyl]sulfinyl]acrylate: A product resulting from trapping of a sulfenic acid by methyl propiolate. *J. Chem. Crystallogr.* 29, 1133–1136.
- (67) Ishii, A., Komiya, K., and Nakayama, J. (1996) Synthesis of a stable sulfenic acid by oxidation of a sterically hindered thiol (thiophen-

- etriptycene-8-thiol) and its characterization. *J. Am. Chem. Soc.* 118, 12836–12837.
- (68) Hogg, D. R., and Robertson, A. (1974) Sulphenate ions as ambident nucleophiles. *Tetrahedron Lett.* 43, 3783–3784.
- (69) Schmid, G. H., Strukelj, M., and Dalipi, S. (1987) The products of the reaction of thiiranium ions with competing nucleophiles. *Can. J. Chem.* 65, 1945–1950.
- (70) Balsamo, A., Benedini, P. M., Giorgi, I., Macchia, B., and Macchia, F. (1982) Interconversion of the thiazine and thiazolidine system of beta-lactam antibiotics. Electrochemical cleavage of Kamiya's disulfide promoted by bromide ion. *Tetrahedron Lett.* 23, 2991–2994.
- (71) Zefirov, N. S., Sadovaja, N. K., Novgorodtseva, L. A., Achmedova, R. S., and Baranov, S. V. (1979) New method for increasing of effective electrophilicity of weak electrophiles in addition reactions. Rearrangements and cis-addition in reactions of sulfenyl chlorides with norbornene and dimethoxybenzonorbornadiene. *Tetrahedron* 35, 2759–2765.
- (72) Gubin, A. S., Smit, V. A., Krimer, M. Z., Zefirov, N. S., Novgorodtseva, L. A., and Sadovaya, N. K. (1980) Reactivity of cyclooctene derived episulfonium ions toward nucleophiles and the course of sulfenyl halide addition to cyclooctene. *Tetrahedron* 36, 1361–1366.
- (73) Wiberg, K. B., and Österle, C. G. (1999) Deamination of *trans*-2-methyl- and *trans*-2-phenylcyclopropylamines. *J. Org. Chem.* 64, 7756–7762.
- (74) Winstein, S., Klinedinst, P. E. J., and Robinson, G. C. (1961) Salt effects and ion pairs in solvolysis and related reactions. XVII. Induced common ion rate depression and the mechanism of the special salt effect. *J. Am. Chem. Soc.* 83, 885–895.
- (75) Jonnalagadda, S. B., and Gollapalli, N. R. (2000) Kinetics of reduction of toluidine blue with sulfite - kinetic salt effect in elucidation of mechanism. *J. Chem. Educ.* 77, 506–509.
- (76) Janssen, J. W. A. M., and Kwart, H. (1977) The thermal beta-cis-elimination reaction of cyclic sulfoxides and subsequent ring expansion reactions. *J. Org. Chem.* 42, 1530–1533.
- (77) Bushby, R. J. (1976) Thermal rearrangement of the 2,2,4,4-tetramethylthietan-3-one 1-oxide; a reaction related to the ring expansion of the penicillin S-oxides. *J. Chem. Soc. Perkin Trans 1* 2590–2593.
- (78) Block, E. (1992) The organosulfur chemistry of the genus *Allium*—Implications for the organic chemistry of sulfur. *Angew. Chem., Int. Ed. Engl.* 31, 1135–1178.
- (79) Braverman, S., and Pechenick, T. (2002) Facile preparation and rearrangement of allylic dialkoxy disulfides. *Tetrahedron Lett.* 43, 499–502.
- (80) Fox, M., and Scott, D. (1980) The genetic toxicology of nitrogen and sulphur mustard. *Mutat. Res.* 75, 131–168.
- (81) Fidler, A., Moes, G. W. H., Scheffer, A. G., ver der Schans, G. P., Baan, R. A., de Jong, L. P. A., and Benschop, H. P. (1994) Synthesis, characterization, and quantitation of the major adducts formed between sulfur mustard and DNA of calf thymus and human blood. *Chem. Res. Toxicol.* 7, 199–204.
- (82) Dacre, J. C., and Goldman, M. (1996) Toxicology and pharmacology of the chemical warfare agent sulfur mustard. *Pharmacol. Rev.* 48, 289–326.
- (83) Walker, I. G., and Thatcher, C. J. (1968) Lethal effects of sulfur mustard on dividing mammalian cells. *Radiat. Res.* 34, 110–127.

TX8000187