See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/248739362

# Syntheses of Two Cytotoxic Sinapyl Alcohol Derivatives and Isolation of Four New Related Compounds from Ligularia n elumbifolia

<b>ARTICLE</b> in JOURNAL OF NATURAL PRODUCTS · JUNE :	2002
--	------

Impact Factor: 3.8 · DOI: 10.1021/np0200257

\_\_\_\_\_\_CITATIONS READS

15 14

## 7 AUTHORS, INCLUDING:



Yu Zhao

China Academy of Engineering Physics

260 PUBLICATIONS 2,489 CITATIONS

SEE PROFILE



Françoise Guéritte

French National Centre for Scientific Research

**222** PUBLICATIONS **4,984** CITATIONS

SEE PROFILE

# Syntheses of Two Cytotoxic Sinapyl Alcohol Derivatives and Isolation of Four New Related Compounds from Ligularia nelumbifolia

Yu Zhao,\*,†,‡ Xiaojiang Hao,‡ Wei Lu,§ Junchao Cai,§ Hong Yu,¹ Thierry Sevénet," and Françoise Guéritte"

Department of Traditional Chinese Medicine and Natural Drug Research, College of Pharmaceutical Sciences, Zĥejiang University, Hangzhou 310031, People's Republic of China, Laboratory of Phytochemistry, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, People's Republic of China, Shanghai Institute of Material Medica, Chinese Academy of Sciences, Shanghai 200031, People's Republic of China, Inmore Laboratory of Biotechnology, Kunming 650011, People's Republic of China, and Institut de Chimie des Substances Naturelles, CNRS, Avenue de la Terrasse, 91198 Gif-sur-Yvette, France

Received January 23, 2002

Phytochemical reinvestigation on Ligularia nelumbifolia afforded four novel sinapyl alcohol analogues named nelumols B-E (1-4) and three known sinapyl alcohol derivatives (5-7). Their structures were elucidated by NMR techniques. Total syntheses of cytotoxic geranyloxy sinapyl alcohol (6) and geranyloxy sinapyl aldehyde (7) were carried out via two different paths. The 4-O-benzyl-substituted analogues (20 and 27) as well as the 4-O-(2-methylbutenyl) derivatives (34 and 35) were also synthesized. The cytotoxicities of 6 and 7 were measured using A-549, HL-60, and KB cancer cell lines.

The genus Ligularia has been used medicinally for a long time in China. Distributed in damp shadowy regions beside brooks and sloping fields, the whole plant of Ligularia nelumbifolia [(Bur. Et Franch) Hand.-Mazz] (family Compositae, Chinese folk name Lian Ye Tuo Wu) has been used as folk medicine for pulmonary tuberculosis and apoplexy. 1 Previous phytochemical examination of *Ligularia* species revealed eremophilane derivatives.<sup>2-6</sup> Interestingly, no eremophilane derivatives were found in the species investigated by us; however, several sinapyl alcohol derivatives and aromatic components were isolated.3 Thorough examination of this species has now afforded five further sinapyl alcohol derivatives (1-5), four of which (1-4) are new compunds. In the course of our continuing search for pharmacologically active compounds, two major principles of this species, geranyloxy sinapyl alcohol (6)3,7 and geranyloxy sinapyl aldehyde (7), were found to be cytotoxic to KB cell with an IC $_{50}$  of 3.0  $\times$  10 $^{-6}$  and 2.6  $\times$  10 $^{-6}$  M, respectively. This prompted us to reinvestigate further analogues in this plant and to synthesize compounds 6 and 7 as well as several analogues for further pharmacological activity studies.

# **Results and Discussion**

Nelumol B (1) was obtained as colorless gum. EIMS and elemental analysis indicated its molecular formula to be  $C_{21}H_{30}O_5$ . Showing the molecular ion peak at m/z 362, the EIMS of 1 also exhibited a base peak due to a sinapyl alcohol fragment at m/z 210. The <sup>1</sup>H and <sup>13</sup>C NMR spectra of 1 showed close similarities with those of the geranyloxy sinapyl alcohol (**6**).<sup>3,7</sup> In the <sup>1</sup>H NMR spectrum (Table 1), the only differences were the presence in 1 of an olefinic methylene multiplet (H-9') at  $\delta$  5.00 (1H, brs) and 4.98 (1H, brs), as well as an olefinic methyl signal (H-8') at  $\delta$  1.73 (brs, 3H) in place of the olefinic H-6' and Me-9' signals of **6**. Furthermore, a signal was detected at  $\delta$  3.88 (m, 1H),

<sup>†</sup>Zhejiang University.

\*Kunming Institute of Botany, CAS.

suggesting a secondary OH group at the C-5' position. This was supported by an OH absorption band at 3399 cm<sup>-1</sup> in the IR spectrum of 1. The <sup>13</sup>C NMR spectrum of 1 was in complete accord with the proposed structure (Table 2).

Comparison of the <sup>1</sup>H and <sup>13</sup>C NMR spectra of 2 with those of 1 indicated that 2 had an oxygenated C-6', since H-6' was shifted downfield (from  $\delta$  2.06 to 4.55) when compared to 1, thus disclosing that H-6' was vicinal to the 7'(9') double bond in the case of 2. Furthermore, the <sup>1</sup>H NMR spectrum of 2 revealed the presence of an ethoxy group at C-6'. EIMS gave the molecular ion peak at m/z390, which was consistent with the molecular formula C<sub>23</sub>H<sub>34</sub>O<sub>5</sub>. Since ethanol was exclusive during the extraction and isolation procedure, compound 2 might be derived biosynthetically from precursor 6.

The <sup>1</sup>H NMR spectrum of nelumol D (3) exhibited some differences from that of geranyloxy sinapyl alcohol 6. The methylene proton (H-5') of 6 could not be found in the <sup>1</sup>H NMR spectrum of 3, while two olefinic hydrogens were observable at  $\delta$  5.58 (m, 2H). Furthermore, the methyl singlets appeared at  $\delta$  1.33 (s, 6H), somewhat higher field than those of 6 in the <sup>1</sup>H NMR spectrum, suggesting that an OH group was most likely connected to C-7', in agreement with the corresponding <sup>13</sup>C resonance appearing at  $\delta$  82.04 (s, C-7'). The olefinic carbons attributable to a trisubstituted double bond at  $\delta$  140.16 (s) and 127.88 (d)

<sup>\*</sup> To whom correspondence should be addressed. Tel: (86)-571-87217313. Fax: (86)-571-87217313. E-mail: dryuzhao@zjuem.zju.edu.cn or dryuzhao@ hotmail.com

<sup>§</sup> Shanghai Institute of Material Medica, CAS.

Inmore Laboratory of Biotechnology.
 Institut de Chimie des Substances Naturelles, CNRS.

**Table 1.** <sup>1</sup>H NMR Spectral Data [400 MHz,  $\delta_{\rm H}$  (*J*, Hz)] for Nelumols B–E (1–4) in CDCl<sub>3</sub>

position	1	2	3	4
2	6.59 s	6.60 s	6.61 s	6.61 s
6	6.59 s	6.60 s	6.61 s	6.61 s
7	6.52 dt (15.8, 1.4)	6.52 dt (15.9, 1.4)	6.55 dt (15.9, 1.4)	6.55 dt (16.0, 1.5)
8	6.28 dt (15.8, 5.8)	6.28 dt (15.9, 5.8)	6.30 dt (15.9, 6.0)	6.30 dt (16.0, 6.0)
9	4.32 dd (5.8, 1.4)	4.32 dd (5.8, 1.4)	4.32 dd (6.0, 1.4)	4.33 dd (6.0, 1.5)
1'	4.54 br d (7.2)	4.54 br d (7.1)	4.55 br d (7.0)	4.54 br d (7.2)
2'	5.66 tq (7.2, 1.0)	5.58 tq (7.1, 1.0)	5.58 m	5.61 tq (7.1, 1.0)
4'	2.06 m	2.02 m	2.74 m	5.47 dt (2.0, 1.5)
5′	3.88 m	2.00 m		
6'	2.06 m	4.55 br dt (7.0, 1.5)	5.58 m	2.74 dd (6.6, 2.0)
8′	1.73 br s	1.63 br s	1.31 s	1.26 s
9′	5.00 br s	4.92 br dd (1.5, 1.5)	1.31 s	1.26 s
	4.98 br s	4.83 br dd (1.5, 1.5)		
10'	1.65 d (1.0)	1.65 d (1.0)	1.63 d (0.9)	1.63 d (1.0)
OMe	3.86 s	3.87 s	3.87 s	3.86 s
OEt		3.65 q (7.0)	3.49 q (7.0)	3.32 q (7.0)
		1.24 t (7.0)	1.22 t (7.0)	1.14  t (7.0)

**Table 2.** <sup>13</sup>C NMR Spectral Data [100 MHz,  $\delta$  (ppm)] for Nelumols B-E (1-4) in CDCl<sub>3</sub><sup>a</sup>

	2 (2 2)	3		
C no.	<b>1</b> (mult)	<b>2</b> (mult)	<b>3</b> (mult)	<b>4</b> (mult)
1	136.5 s	136.3 s	136.6 s	138.0 s
2	103.5 d	103.3 d	103.3 d	103.4 d
3	153.7 s	153.6 s	153.7 s	153.7 s
4	139.8 s	141.0 s	139.8 s	140.0 s
5	153.7 s	153.6 s	153.7 s	153.7
6	103.5 d	103.3 d	103.3 d	103.4 s
7	131.2 d	131.1 d	131.2 d	131.2 d
8	129.0 d	127.8 d	127.9 d	127.9 d
9	63.6 t	63.5 t	63.7 t	63.7 t
1'	69.2 t	69.2 t	69.3 t	69.4 t
2'	121.4 d	120.6 d	121.2 d	121.2 d
3'	132.4 s	132.3 s	132.3 s	132.3 s
4'	39.5 t	35.4 t	42.2 t	126.9 d
5'	88.9 d	32.5 t	140.2 s	140.2 s
6'	28.7 t	75.2 d	127.9 d	42.6 t
7′	143.8 t	147.2 s	70.8s	74.8 s
8'	17.1 q	17.4 q	29.7 q	26.4 q
9'	114.1 t	111.0 t	29.7 q	26.4 q
10'	16.1 q	16.1 q	16.3 q	16.2 q
OMe	56.1 q	56.0 q	56.0 q	56.0 q
OEt	•	63.8 t	56.0 t	57.7 t

<sup>&</sup>lt;sup>a</sup> Assignment in the same column could be exchangeable.

were assigned to C-5' and C-6', respectively. This side chain is similar to that of the sinapyl alcohol derivative 5, previously isolated from Ligularia duciformis.8 However, the molecular ion peak of **3** appearing at m/z 406, i.e., 44 mass units higher than that of 5, as well as the NMR data all indicated that 3 was an C-5'-OEt derivative of 5 (Tables 1 and 2). Compound 3 might be another artifact or the enzymatic derivative of 5, as mentioned above.

Nelumol E (4) had a molecular ion peak and NMR data similar to those of 3. Elemental analysis and a DEPT spectrum revealed its molecular formula to be C<sub>23</sub>H<sub>34</sub>O<sub>6</sub>, apparently isomeric with 3. Scrutiny of its <sup>1</sup>H and <sup>13</sup>C NMR spectra with those of 3 led to the assignment of a 2'(3'),4'-(5')-diene system in compound 4 (Tables 1 and 2). A COLOC experiment on 4 exhibited correlations of olefinic H-4' with C-2' and C-10', consistent with the presence of a conjugated diene moiety in 4. This enol ether could be either an artifact or a biosynthetic derivative, as discussed above.

As 6 and 7 were cytotoxic to KB cells (Table 3) and appeared as principle metabolites in *L. nelumbifolia*, syntheses of further sinapyl alcohol derivatives become interesting. Thus, 6 and 7 were selected to be totally

The first path used commercially available sinapinic acid 8 as starting material. After esterification,9 a Mitsunobu

Table 3. IC<sub>50</sub> of 6 and 7 on Some Selected Pharmacological

	A-549 cell	HL-60 cell	KB cell
6 7	$\begin{array}{c} 3.4 \times 10^{-5} \ M \\ 2.2 \times 10^{-5} \ M \end{array}$	$\begin{array}{c} 6.7 \times 10^{-6} \ M \\ 1.2 \times 10^{-5} \ M \end{array}$	$\begin{array}{c} 3.0 \times 10^{-6}  M \\ 2.6 \times 10^{-6} M \end{array}$

reaction of the resulting methyl ester 9 with geranyl alcohol led to the geranyl derivative 10.10 Reduction of 10 by DIBAH afforded geranyloxy sinapyl alcohol 6 in an 86% yield, while oxidation of 6 by magnesium dioxide gave geranyloxy sinapyl aldehyde 7 in 92% yield (Scheme 1).

Another synthetic path started from methyl gallate (11) (Scheme 2) Acetylation led to product 12, which was subjected to a selective substitution reaction,11 during which the 4-acetoxy group was replaced by a geranyl moiety to yield compound 13b. The unexpected monodeacetylated product 13a was also formed in the reaction. The reaction time and the temperature influenced the yields of 13a and 13b. The mixture of 13a and 13b was treated with aqueous K2CO3 to give 14, which was then transformed to the methoxy derivative 15 (82% yield over two steps). Reduction of 15 by LAH afforded primary alcohol 16, which was oxidized to aldehyde 17 by pyridinium chlorochromate in 86% yield. A Knoevenagel condensation of 17 with malonic acid in the presence of piperidine afforded the *E*-form of acid **18**. Reduction of **18** by LAH afforded, apart from the 80% yield of expected target molecule 6, the 1,4-addition product 19 in 5% yield. Finally, geranyloxy sinapyl aldehyde 7 was obtained by manganese dioxide oxidation in 92% yield. The total yield of **8** was 28%. Cytotoxic screening results of synthetic **6** and 7 against A-549, HL-60, and KB cell lines are shown in Table 3.

To examine the importance of the C-4 side chain on cytotoxicity, we designed another target molecule (20) with a benzyl group attached to O-C(4). Furthermore, a fivecarbon side chain (compound 34) was also introduced to extend the SAR concept. Two paths were examined to synthesize these analogues, which are shown in Schemes 2 and 3. Cytotoxicity screening of 20, 27, 34, and 35 is shown in Table 4. It was seen that compounds 20 and 27 were less cytotoxic to KB cells than 6 and 7, while the fivecarbon side chain derivatives **34** and **35** had cytotoxicities to KB cells similar to those of 1 and 2.

# **Experimental Section**

General Experimental Procedures. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were measured on Bruker AM-400 MHz and Bruker AC-300 MHz NMR instruments, with TMS as internal

### Scheme 1a

 $^{\it a}$  (a)  $H_2SO_4$ , MeOH, reflux, 2 h, 98%; (b) geranyl alcohol, Ph<sub>3</sub>P, DEAD, 24 h, 50%; (c) DIBAH, THF, -78 °C, 2 h, 86%; (d) 1: PCC, CH<sub>2</sub>Cl<sub>2</sub>, rt, 6 h, 81%; 2: MnO<sub>2</sub>, EtOAc, rt, 92%.

#### Scheme 2<sup>a</sup>

<sup>a</sup> (a)  $H_2SO_4$ , MeOH, reflux, 2 h, 96%; (b)  $Ac_2O$ , Py, rt, 12 h, 93%; (c) geranyl bromide,  $K_2CO_3$ , DMF, 0 °C, 24 h, 50% of **13b**, 29% of **13a**; (d)  $K_2CO_3$ , MeOH $-H_2O$ , rt, 0.5 h, 90%; (e) MeI,  $K_2CO_3$ , reflux, 3 h, 91%; (f) LAH, ether, 0 °C, 90%; (g) PCC,  $CH_2Cl_2$ , rt, 6 h, 86%; (h) malonic acid, piperidine, Py, reflux, 4 h, 86%; (i) LAH, ether, 0 °C, 80% of **6**, 5% of **19**; (j) MnO<sub>2</sub>, EtOAc, rt, 2 h, 92%.

**Table 4.**  $IC_{50}$  of Compounds **20**, **27**, **34**, and **35** on KB Cells (mol/L)

20	27	34	35
8.6 × 10-4	$6.4 \times 10-4$	$7.8 \times 10-6$	5.3  imes 10-6

standard. HREIMS and EIMS were performed on a VG Auto Spec-3000 MS instrument. EIMS: direct inlet, 70 eV. Solvents and reagents were purified according to standard laboratory techniques. IR spectra were recorded on a Perkin-Elmer 577 spectrometer.

**Plant Material.** The material plant was collected in August 2000, Zhaotong County, Yunnan Province, China, and identified by Prof. Hua Peng. A voucher specimen (no. 20000806) is deposited in the Laboratory of Phytochemistry, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, Yunan Province, China.

**Extraction and Isolation.** Air-dried roots of *Ligularia nelumbifolia* [(Bur. Et Franch) Hand.-Mazz] (2.0 kg) were powdered and extracted with petroleum ether (60-90 °C)—  $Et_2O-MeOH$  (1:1:1) at room temperature (3 days × 3) to give 85 g of crude extract, which was subjected to column chromatography on 1 kg of silica gel with petroleum ether containing gradually increasing amounts of EtOAc (1:0–1:1). Ten crude fractions ( $F_1-F_{10}$ ) were obtained.  $F_1-F_7$  contained, by TLC, mainly the same products reported previously.  $^3F_8$  (2.1 g) afforded, after repeated column chromatography, 86 mg of **6** 

and 35 mg of 7.  $F_9$  (3.2 g) was chromatographed (200 g of silica gel gel, 200–300 mesh) using a CHCl<sub>3</sub>–Me<sub>2</sub>CO (20:1–1:1) step gradient. Eluates 25–28 (150 mL each) were combined and purified by PTLC (CHCl<sub>3</sub>–Me<sub>2</sub>CO, 3:1) to give 14 mg of 1 ( $R_f = 0.46$ ). Eluate 14 (120 mL) was evaporated and purified by PTLC ( $C_6H_6$ –Me<sub>2</sub>CO, 4:1) to give 21 mg of 2. Eluate 17 (80 mL) contained 26 mg of 5, which was obtained by PTLC with  $C_6H_6$ –Me<sub>2</sub>CO, 8:1 ( $R_f = 0.65$ ).  $F_{10}$  (6.6 g) was rechromatographed over silica H (200 g) with a CHCl<sub>3</sub>–EtOAc (10:1–1: 2) solvent system. Eluates 16–17 (125 mL each) were combined and evaporated, and the residue (86 mg) was purified through PTLC (CHCl<sub>3</sub>–MeOH, 8:1) to afford 17 mg of 3 ( $R_f = 0.57$ ) and 15 mg of 4 ( $R_f = 0.49$ ).

4-O-[(2E)-3,7-Dimethyl-2,7-octadien-5-ol]sinapyl alcohol (1): gum; IR (KBr)  $\nu_{\rm max}$  3399 (OH), 3349 (OH), 2977, 1659, 1583, 1504, 1459, 1420, 1332, 1241, 1127, 963 cm $^{-1}$ ; EIMS m/z (rel int) 362 [M] $^+$  (16), 347 (3), 344 (5), 329 (6), 306 (14), 277 (10), 252 (18), 238 (50), 210 (100), 182 (36), 167 (42), 154 (18);  $^1$ H NMR (CDCl $_3$ ) data, see Table 1;  $^{13}$ C NMR (CDCl $_3$ ) data, see Table 2; anal. C 69.56%, H 8.27%, calcd for C $_{21}$ H $_{30}$ O, C 69.61%, H 8.29%.

**4-***O*-[(2*E*)-3,7-Dimethyl-6-ethoxy-2,7-octadiene]-sinapyl alcohol (2): gum; IR (KBr)  $\nu_{\text{max}}$  3398 br (OH), 3072, 2939, 1653, 1583, 1504, 1456, 1418, 1333, 1241, 1128, 992, 904, 629 cm<sup>-1</sup>; EIMS m/z (rel int) 390 [M]<sup>+</sup>, (15), 375 (22), 349 (18), 344 (16), 277 (10), 210 (100), 182 (55), 167 (43), 137 (16), 121 (14), 113 (20), 69 (72), 46 (23); <sup>1</sup>H NMR (CDCl<sub>3</sub>) data, see Table

 $^a$  (a1) Benzyl bromide, DMF, 0 °C, 24 h; (a2) 2-methylbutenyl bromide,  $K_2CO_3$ , DMF, 0 °C, 10 h; (b)  $K_2CO_3$ , MeOH $-H_2O$ , rt, 0.5 h; (c) MeI,  $K_2CO_3$ , reflux, 3 h; (d) LAH, ether, 0 °C; (e) PCC,  $CH_2Cl_2$ , rt, 6 h, (f) malonic acid, piperidine, Py, reflux, 4 h; (g) LAH, ether, 0 °C; (h) MnO $_2$ , EtOAc, rt, 2 h (MB = 2-methylbutenyl).

#### Scheme 4<sup>a</sup>

 $^{a}$  (a1) Benzol, Ph<sub>3</sub>P, DEAD, 24 h, 65%; (a2) 2-methylbutenol, Ph<sub>3</sub>P, DEAD, 24 h, 60%; (b) DIBAH, THF, -78 °C, 2 h; 88% of **20**, 80% of **34**; (c) 1: PCC, CH<sub>2</sub>Cl<sub>2</sub>, rt, 6 h; 83% of **27**, 81% of **35**; 2: MnO<sub>2</sub>, EtOAc, rt, 92% of **27**, 94% of **35** (MB = 2-methylbutenyl).

1;  $^{13}$ C NMR (CDCl<sub>3</sub>) data, see Table 2; anal. C 70.73%, H 8.72%, calcd for  $C_{23}H_{34}O_5$ , C 70.77%, H 8.72%.

4-O-[(2E,5E)-3,7-Dimethyl-5-ethoxy-2,5-octadiene-7-ol]-sinapyl alcohol (3): gum; IR (KBr)  $\nu_{\rm max}$  3408 br (OH), 2967, 2926, 1665, 1582, 1504, 1459, 1417, 1332, 1240, 1127, 969, 914, 744 cm $^{-1}$ ; EIMS m/z (rel int) 406 [M] $^{+}$ , (8), 391 (2), 389 (5), 360 (6), 314 (15), 264 (3), 210 (100), 197 (3), 182 (25), 167 (42), 154 (16), 69 (18), 46 (48);  $^{1}$ H NMR (CDCl $_{3}$ ) data, see Table 1;  $^{1}$ C NMR (CDCl $_{3}$ ) data, see Table 2; anal. C 67.90%, H 8.31%, calcd for C $_{23}$ H $_{34}$ O $_{6}$ , C 67.98%, H 8.37%.

4-O-[(2E,4E)-3,7-Dimethyl-5-ethoxy-2,4-octadien-7-ol]-sinapyl alcohol (4): gum, IR (KBr)  $\nu_{\rm max}$  3402 br (OH), 3349, 2973, 2933, 1673, 1582, 1503, 1457, 1418, 1333, 1240, 1128, 969, 844 cm<sup>-1</sup>; EIMS m/z (rel int) 406 [M]<sup>+</sup>, (12), 391 (4), 389 (8), 374 (4), 360 (2), 343 (5), 210 (100), 197 (6), 182 (38), 167 (44), 154 (23), 128 (6), 69 (18), 46 (36);  $^1$ H NMR (CDCl<sub>3</sub>) data, see Table 1;  $^{13}$ C NMR (CDCl<sub>3</sub>) data, see Table 2; anal. C, 67.90%, H, 8.31%, calcd for  $C_{23}H_{34}O_6$ , C, 67.98%, H, 8.37%.

**Sinapic Acid Methyl Ester (9).** NMR and physical data were identical with a previous publication.<sup>9</sup> EIMS: m/z 238 [M]<sup>+</sup> (100), 223 (9), 207 (95), 175 (33), 163 (11), 119 (10), 91 (6). HREIMS: 238.0856 (calcd for  $C_{12}H_{14}O_5$ , 238.0841).

**Etherification of 9.** To a stirred solution of 313 mg (1.2 mmol) of Ph<sub>3</sub>P and 240 mg of **9** (1.0 mmol) in dry THF (10 mL) was added 150 mg of geraniol (1.0 mmol) and DEAD (262  $\mu$ L, 1.2 mmol) at room temperature under nitrogen. The

solution was stirred overnight and then refluxed for 0.5 h. The cooled solution was partitioned between  $H_2O$  (30 mL) and EtOAc (30 mL  $\times$  3) and dried (MgSO<sub>4</sub>). After filtration, the solvent was evaporated and the residue was subjected to CC (petroleum ether–Et<sub>2</sub>O, 5:1–2:1); 186 mg of **10** was isolated (50%).

**4-Geranyl sinapic acid methyl ester (10):** gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) 7.57 (1H, d, J=16.0 Hz, H-7), 6.72 (2H, s, H-2, H-6), 6.32 (1H, d, J=15.8 Hz, H-8), 5.53 (1H, brt, J=7.0 Hz, H-2′), 5.05 (1H, m, H-6′), 4.55 (2H, d, J=7.1 Hz, H-1′), 3.86 (6H, s, OMe-3, OMe-5), 3.79 (3H, s, CO<sub>2</sub>Me), 2.03 (4H, m, H-4′, H-5′), 1.65 (3H, s, H-8′), 1.63 (3H, s, H-9′), 1.57 (3H, s, H-10′); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ 167.3 (s, C-9), 153.82 (s, C-3), 129.7 (s, C-7), 123.92 (d, C-6′), 119.97 (d, C-2′), 116.77 (d, C-8), 105.17 (d, C-2, C-6), 69.50 (t, C-1′), 56.09 (q, OMe-3, OMe-5), 51.62 (q, CO<sub>2</sub>Me), 39.57 (t, C-4′), 26.39 (t, C-5′), 25.62 (t, C-8′), 17.61 (q, C-9′), 16.31 (q, C-10′); EIMS m/z 374 [M]<sup>+</sup> (1), 343 (1), 305 (2), 266 (1), 248 (1), 238 (100), 223 (3), 207 (8), 175 (3), 163 (2), 135 (2), 69 (13); HREIMS m/z 374.2082 (calcd for C<sub>22</sub>H<sub>30</sub>O<sub>5</sub>, 374.2093).

**Reduction of 10.** To a stirred solution of 374 mg (1.0 mmol) of **10** in dry Et<sub>2</sub>O (10 mL) was added DIBAH (1.0 mL, 1.0 M in hexane) at -78 °C under nitrogen. The solution was stirred for 0.5 h, 3 mL of H<sub>2</sub>O was added at -78 °C to quench the reaction, and the solution was allowed to warm to room

temperature. Ten milliliters of 1 M HCl was added, and the solution was extracted with EtOAc (15 mL imes 3). The organic layers were combined and dried (MgSO<sub>4</sub>). Purification by flash column afforded 299 mg of 6 (86%). Physical and NMR data for compound 6 have been reported in an earlier publication.<sup>3</sup>

Allylic Oxidation of 6 by MnO2. To a stirred suspension of 105 mg (1.2 mmol) of MnO<sub>2</sub> in EtOAc (15 mL) was added 345 mg (1.0 mmol) of **6** in EtOAc (5 mL) at room temperature, and the solution was stirred for 4 h. After filtration, the eluate was evaporated to dryness and was partitioned between H<sub>2</sub>O (20 mL) and Et<sub>2</sub>O (60 mL). The organic layer was combined and dried (MgSO<sub>4</sub>), and the solvent was evaporated to afford 7 (317 mg, 92%). Physical and NMR data for compound 7 have been reported in an earlier publication.<sup>3</sup>

**Deacetylation of 12.** To a stirred solution of 15.5 g of 12 (50 mmol) in dry DMF (150 mL) was added 20.7 g of K<sub>2</sub>CO<sub>3</sub> (150 mmol) at 0 °C. The solution was stirred for 20 min and 10.85 g (9.9 mL) of geranyl bromide (60 mmol) in dry DMF (60 mL) was added in 10 min. The solution was stirred for 10 h. After suction filtration, 300 mL of H<sub>2</sub>O was added. The mixture was extracted with EtOAc (600 mL), followed by Et<sub>2</sub>O (600 mL). The organic layers were combined, washed with brine (100 mL), and dried (MgSO<sub>4</sub>). The solution was evaporated, and the residue was subjected to CC (hexane-Et<sub>2</sub>O, 5:1) to afford 10.11 g (25 mmol) of 13b (50%) and 5.25 g (14.5 mmol) of **13a** (29%). Also, 925 mg (3.0 mmol) of **12** (6%) was recovered.

4-Geranoyl-3,5-diacetoxybenzoic acid methyl ester **(13b):** gum;  ${}^{1}$ H NMR (CDCl $_{3}$ , 300 MHz)  $\delta$  7.64 (2H, s, H-2, H-6), 5.42 (1H, brt, J = 7.0 Hz, H-2'), 5.09 (1H, m, H-6'), 4.59 (2H, d, J = 7.2 Hz, H-1'), 3.89 (3H, s,  $CO_2Me$ ), 2.36 (3H, s, OCOCH<sub>3</sub>), 2.09 (4H, m, H-4', H-5'), 1.70 (3H, s, H-8'), 1.68 (3H, s, H-9'), 1.62 (3H, s, H-10'); HREIMS m/z 404.1818 (calcd for C<sub>22</sub>H<sub>28</sub>O<sub>7</sub>, 404.1835).

4-Geranoyl-3-acetoxy-5-hydroxysinapic acid methyl **ester (13a):** gum;  ${}^{1}$ H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  7.52 (1H, brs, H-2), 7.36 (1H, brs, H-6), 5.90 (1H, brs, O*H*-5), exchanged in  $D_2O$ ), 5.48 (1H, t, J = 7.0 Hz, H-2'), 5.08 (1H, m, H-6'), 4.63 (1H, d, J = 7.1 Hz, H-1'), 3.90 (3H, s,  $CO_2Me$ ), 2.36 (3H, s, OCOCH<sub>3</sub>), 2.10 (4H, m, H-4', H-5'), 1.70 (3H, s, H-8'), 1.66 (3H, s, H-9'), 1.61 (3H, s, H-10'); HREIMS m/z 362.1709 (calcd for C<sub>20</sub>H<sub>26</sub>O<sub>6</sub>, 362.1729).

Deacetylation of 13 (13a and 13b) (e.g., 13a). To a stirred solution of 13a (2.91 g, 8.0 mmol) in MeOH (200 mL) at 0 °C was added 5.66 g (43.2 mmol) of K<sub>2</sub>CO<sub>3</sub> in H<sub>2</sub>O (60 mL) in 10 min. The solution was stirred for 20 min, and the solvent was evaporated. Then 1 M HCl was added to adjust the pH value to 2, and the aqueous solution was extracted by EtOAc (300 mL). The organic layers were combined and dried (MgSO<sub>4</sub>), and the solvent was evaporated to afford 2.32 g (7.2 mmol) of 14 (90%).

4-Geranoyl-3,5-hydroxybenzoic acid methyl ester (14): gum; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.25 (2H, s, H-2, H-6), 5.91 (1H, s, exchanged in  $D_2O$ , ArOH), 5.60 (1H, brt, J = 7.0 Hz, H-2'), 5.09 (1H, m, H-6'), 4.66 (2H, d, J = 7.1 Hz, H-1'), 3.90 (3H, s, CO<sub>2</sub>Me), 2.08 (4H, m, H-4', H-5'), 1.70 (3H, s, H-8'), 1.66 (3H, s, H-9'), 1.61 (3H, s, H-10'); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>)  $\delta$  166.9 (s, C-7), 149.2 (s, C-3, C-5), 145.2 (s, C-1), 137.4 (s, C-4), 132.1 (s, C-3'), 126.1 (s, C-7'), 123.5 (s, C-6'), 118.7 (s, C-2'), 109.5 (d, C-2, C-6), 69.9 (t, C-1'), 52.2 (q, CO<sub>2</sub>Me), 39.6 (t, C-4'), 26.2 (t, C-5'), 25.6 (q, C-8'), 17.7 (q, C-9'), 16.4 (q, C-10'); HREIMS  $\delta$  320.1622 (calcd for C<sub>18</sub>H<sub>24</sub>O<sub>5</sub>, 320.1624).

4-Geranoyl-3,5-methoxybenzoic acid methyl ester (15). To a stirred solution of 14 (320 mg, 1.0 mmol) in dry DMF (30 mL) was added 8 mg of K<sub>2</sub>CO<sub>3</sub> (6.0 mmol) at room temperature under argon, then 0.312 mL (5.0 mmol) of MeI in DMF (5 mL) was added. The solution was heated at 100 °C for 3 h and was cooled to 25 °C. After suction filtration, the filtrate was partitioned between H<sub>2</sub>O (120 mL) and EtOAc-ether (100 mL/ 100 mL). The organic layers were combined and dried (MgSO<sub>4</sub>). The solution was evaporated under reduced pressure, and the residue was subjected to PTLC; 315 mg (0.91 mmol) of 15 was obtained (91%): gum;  $^1$ H NMR (CDCl $_3$ , 300 MHz)  $\delta$  7.30 (2H, s, H-2, H-6), 5.55 (1H, brt, J = 7.0 Hz, H-2'), 5.05 (1H, m, H-6'), 4.59 (1H, d, J = 7.0 Hz, H-1'), 3.93 (3H, s, H-9'), 3.89 (6H, s, OMe-3, OMe-5), 2.04 (4H, m, H-4', H-5'), 1.66 (3H, s, C-8'), 1.64 (3H, s, C-9'), 1.60 (3H, s, C-10'); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  166.8 (s, C-7), 153.4 (s, C-3, C-5), 141.9 (s, C-1), 141.2 (s, C-4), 131.6 (s, C-3'), 125.1 (s, C-7'), 123.9 (d, C-6'), 119.9 (s, C-2'), 109.6 (d, C-2, C-6), 69.4 (t, C-1'), 56.2 (q, OMe-3, OMe-5), 52.2 (q, CO<sub>2</sub>Me), 39.6 (t, C-4'), 26.4 (t, C-5'), 25.6 (q, C-8'), 17.6 (q,  $\hat{C}$ -9'), 16.3 (q,  $\hat{C}$ -10'); HREIMS  $\delta$  348.1925 (calcd for  $C_{20}H_{28}O_5$ , 348.1937).

4-Geranoyl-3,5-dimethoxybenzyl Alcohol (16). To a stirred suspension of LAH (49 mg, 1.28 mmol) in Et<sub>2</sub>O (50 mL) at 0 °C was added a solution of 15 (280 mg, 0.8 mmol) in dry Et<sub>2</sub>O (20 mL) under argon atmosphere. The solution was stirred for 10 min and was quenched by H<sub>2</sub>O (8 mL). Then 50 mL of 1 N HCl was added, and the mixture was extracted by Et<sub>2</sub>O (150 mL). The ether layers were combined and dried (MgSO<sub>4</sub>). Evaporation of the solvent followed by PTLC afforded 230 mg of 16 (0.72 mmol, 90%): gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  6.62 (2H, brs, H-2, H-6), 5.60 (1H, brt, J = 7.0 Hz, H-2'), 5.05 (1H, m, H-6'), 4.69 (2H, brs, H-7), 4.52 (2H, d, J= 7.1 Hz, H-1'), 3.89 (6H, s, OMe-3, OMe-5), 2.10 (4H, m, H-4', H-5'), 1.70 (3H, s, H-8'), 1.68 (3H, s, H-9'), 1.63 (3H, s, H-10'); HREIMS m/z 320.1966 (calcd for C<sub>19</sub>H<sub>28</sub>O<sub>4</sub>, 320.1988).

4-Geranoyl-3,5-dimethoxybenzaldehyde (17). To a stirred suspension of PCC (225 mg, 1.04 mmol) in CH2Cl2 (30 mL) at 0 °C was added 208 mg of **16** (0.65 mmol) in  $CH_2Cl_2$  (10 mL). The solution was stirred at 0 °C for 5 h. The suspension was filtered and washed by Et<sub>2</sub>O (60 mL) and partitioned between Et<sub>2</sub>O (90 mL) and H<sub>2</sub>O (30 mL). The ether layers were combined, dried (MgSO<sub>4</sub>), and evaporated to afford a residue. PTLC of the residue afforded finally 178 mg of 17 (0.56 mmol, 86%): gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  9.86 (1H, s, H-7) 7.13 (2H, s, H-2, H-6), 5.60 (1H, brt, J = 6.9 Hz, H-2'), 5.05 (1H, m, H-6), 4.77 (2H, brd, J = 7.0 Hz, H-1'), 3.94 (6H, s, OMe-3, OMe-5), 2.04 (4H, m, H-4', H-5'), 1.64 (3H, s, H-8'), 1.63 (3H, s, H-9'), 1.57 (3H, s, H-10'); 13C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  191.2 (s, C-7), 154.2 (s, C-3, C-5), 142.5 (s, C-1), 142.3 (s, C-4), 131.7 (s, C-3'), 131.8 (s, C-7'), 123.9 (d, C-6'), 119.78 (d, C-2'), 106.6 (d, C-2, C-6), 69.6 (t, C-1'), 56.3 (q, OMe-3, OMe-5), 39.6 (t, C-4'), 26.4 (t, C-5'), 25.7 (q, C-8'), 17.7 (q, C-9'), 16.4 (q, C-10'); HREIMS m/z 318.1822 (calcd for  $C_{19}H_{26}O_4$ , 318.1831).

4-Geranoylsinapic Acid (18). To a stirred solution of malonic acid (156 mg, 1.5 mmol) in Py (15 mL) at room temperature was added 475 mg (1.5 mmol) of 17 in Py (10 mL). Piperidine (20 mg) was added to the solution. The mixture was heated at 120 °C for 4 h. The solvent was evaporated and dried (MgSO<sub>4</sub>), evaporated, and followed by CC (CHCl<sub>3</sub>-MeOH, 8:1) to afford 463 mg of 18 (1.3 mmol, 86%): gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.69 (1H, d, J = 15.9 Hz, H-7), 6.75 (2H, s, H-2, H-6), 6.34 (d, J = 15.8 Hz, H-8), 5.53 (1H, brt, J= 7.2 Hz, H-2', 5.05 (1H, m, H-6'), 4.57 (2H, brd, J = 7.1 Hz,H-1'), 3.87 (6H, s, OMe-3, OMe-5), 2.03 (4H, m, H-4', H-5'), 1.65 (3H, s, H-8'), 1.64 (3H, s, H-9'), 1.57 (3H, s, H-10'); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  172.1 (s, C-9), 154.0 (s, C-3, C-5), 147.1 (d, C-7), 141.8 (s, C-4), 139.4 (s, C-1), 131.6 (s, C-3'), 129.4 (s, C-7'), 134.0 (d, C-6'), 120.0 (d, C-2'), 116.2 (d, C-8), 105.5 (d, C-2, C-6), 69.6 (t, C-1'), 56.2 (q, OMe-3, OMe-5), 39.6 (t, C-4'), 26.4 (t, C-5'), 25.7 (q, C-8'), 17.7 (q, C-9'), 16.4 (q, C-10'); EIMS m/z 360 [M]<sup>+</sup>, (3), 345 (1), 331 (11), 316 (3), 224 (100), 209 (4), 198 (26), 181 (4), 69 (23); HREIMS m/z 360.1927 (calcd for  $C_{21}H_{28}O_5$ , 360.1937).

4-Geranoyl-7,8-dihydrosinapic Acid (19). To a stirred solution of LAH (41 mg, 1.09 mmol) in dry Et<sub>2</sub>O (15 mL) at 0 °C was added 195 mg (0.54 mmol) of **18** in dry Et<sub>2</sub>O (10 mL) under argon. The mixture was stirred for 1 h at 0 °C and was quenched by H<sub>2</sub>O (6 mL). Then 1 N HCl (10 mL) was added and extracted by Et<sub>2</sub>O (30 mL). The ether layer was dried (MgSO<sub>4</sub>), evaporated, and subjected to CC (petroleum ether-Et<sub>2</sub>O, 1:2) to afford 150 mg of 7 (0.43 mmol, 80%) and 10 mg of **19** (0.03 mmol, 5%): gum;  $^1$ H NMR (CDCl<sub>3</sub>, 300 MHz)  $^\delta$ 6.46 (2H, brs, H-2, H-6), 5.58 (1H, brt, J = 7.0 Hz, H-2'), 5.07 (1H, m, H-6'), 4.55 (2H, brd, J = 7.0 Hz, H-1'), 3.86 (2H, brt, J = 7.6 Hz, H-9), 3.90 (6H, s, OMe-3, OMe-5), 2.79 (2H, brt, J= 7.6 Hz, H-7), 2.01–1.94 (2H, m, H-8); HREIMS m/z 348.2298 (calcd for C<sub>21</sub>H<sub>32</sub>O<sub>4</sub>, 348.2300).

4-O-Benzyl-3,5-diacetoxybenzoic Acid Methyl Ester (21). The method of preparation of 21 was similar to that used for the preparation of 13. The yield 21 from 12 was 67%. This compound was identical to that reported by Pearson et al.<sup>11</sup> It was noticeable that no mono-deacetylated compound was obtained in this reaction.

- 4-O-Benzyl-3,5-dihydroxybenzoic Acid Methyl Ester (22). The method of preparation of 22 was similar to that used for the preparation of 14. The yield of 22 from 21 was 92%: gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 7.43–7.36 (5H, m, H-3'– H-7'), 7.25 (2H, s, H-2, H-6), 5.80 (brs, exchanged in D<sub>2</sub>O, ArOH), 5.16 (2H, s, H-1'), 3.90 (3H, s, CO<sub>2</sub>Me); HREIMS m/z 274.0870 (calcd for  $C_{15}H_{14}O_4$ , 274.0841). This compound was first reported by Pearson et al.<sup>16</sup>
- 4-O-Benzyl-3,5-dimethoxybenzoic Acid Methyl Ester (23). The method of preparation of 23 was similar to that used for the preparation of **15**. The yield of **23** from **22** was 90%: gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz) δ 7.25-7.5 (5H, m, H-2'-H-7'), 5.10 (2H, s, H-1'), 3.93 (3H, s, Me-8), 3.90 (3H, s, OMe-3, OMe-5);  $^{13}\mathrm{C}$  NMR (CDCl3, 75 MHz)  $\delta$  166.8 (s, C-7), 153.3 (s, C-3, C-5), 141.0 (s, C-1), 137.5 (s, C-4), 128.5 (d, C-3', C-7'), 128.27 (d, C-4', C-6'), 128.1 (d, C-5'), 125.4 (s, C-2'), 106.9 (d, C-2, C-6), 75.0 (t, C-1'), 56.3 (q, OMe-3, OMe-5), 52.3 (q,  $CO_2Me$ ); HREIMS m/z 302.1133 (calcd for  $C_{17}H_{18}O_5$ , 302.1154). This compound was first reported by Jurd et al. 14
- 4-O-Benzyl-3,5-dimethoxybenzyl Alcohol (24). The method of preparation of 24 was similar to that used for the preparation of 16. The yield of 24 from 23 is 94%. This compound was identical to that reported by Battersby et al. 15
- 4-O-Benzyl-3,5-dimethoxybenzaldehyde (25). The method of preparation of 25 was similar to that used for the preparation of 17. The yield of 25 from 24 was 88%. This compound was identical to that reported by Battersby et al. 16
- **4-***O*-**Benzylsinapic Acid (26)**. The method of preparation of 26 was similar to that used for the preparation of 18. The yield of 26 from 25 was 90%. This compound was identical to that reported by Kametani et al.17
- **4-***O*-**Benzylsinapyl Alcohol (20).** The method of preparation of **20** was similar to that used for the preparation of **7**. The yield of 20 from 26 was 87%: gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  7.51–7.20 (5H, m, H-3'–H-7'), 6.56 (2H, brs, H-2, H-6), 6.50 (1H, d, J = 15.8 Hz, H-7), 6.28 (1H, dt, J = 15.8, 5.7 Hz, H-8), 5.06 (2H, brs, H-1'), 3.88 (6H, s, OMe-3, OMe-5); HREIMS m/z 300.1341 (calcd for  $C_{18}H_{20}O_4$ , 300.1362).
- 4-O-Benzylsinapaldehyde (27). The method of preparation of 27 was similar to that used for the preparation of 7. The yield of  $\bf 27$  from  $\bf 20$  was  $\bf 94\%$ : gum;  $^1H$   $\hat{N}M\hat{R}$  (CDCl3,  $\bf 300$ MHz)  $\delta$  9.68 (1H, d, J= 7.5 Hz, H-9), 7.52–7.22 (6H, m, H-3'– H-7', H-7), 6.74 (2H, brs, H-2, H-6), 6.61 (1H, dd, J = 15.8, 7.5 Hz, H-8), 5.09 (2H, brs, H-1'), 3.90 (6H, s, OMe-3, OMe-5); HREIMS m/z 298.1229 (calcd for  $C_{18}H_{18}O_4$ , 298.1205)
- 4-O-(2-Methyl-2-butenyl)-3,5-diacetoxybenzoic Acid Methyl Ester (28). The method of preparation of 28 was similar to that used for the preparation of 13. The yield of 28 from 12 was 60%: gum;  $^1\hat{H}$  NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.65 (2H, s, H-2, H-6),  $5.\overline{37}$  (1H, t, J = 7.0 Hz, H-2'), 4.48 (2H, d, J= 7.25 Hz, H-1'), 3.86 (3H, s,  $CO_2Me$ ), 2.31 (6H, s,  $OCOCH_3$ 3, OCOCH3-5), 1.74 (3H, s, H-4'), 1.64 (3H, s, H-5'); EIMS m/z 336 [M]<sup>+</sup> (1), 321 (1), 295 (1), 281 (2), 286 (6), 253 (2), 237 (4), 226 (41), 195 (3), 184 (60), 153 (5), 121 (4), 85 (14), 69 (100); HREIMS m/z 336.1208 (calcd for  $C_{17}H_{20}O_7$ , 336.1209).
- 4-O-(2-Methyl-2-butenyl)-3,5-dihydroxybenzoic Acid Methyl Ester (29). The method of preparation of 29 was similar to that used for the preparation of 14. The yield of 29 from **28** was 91%: gum;  ${}^{1}H$  NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.20 (2H, s, H-2, H-6), 5.96 (1H, brs, exchanged in D<sub>2</sub>O, ArO*H*), 5.50 (1H, t, J = 7.0 Hz, H-2'), 4.60 (2H,  $\bar{d}$ , J = 7.0 Hz, H-1'), 3.86 (3H, s, CO<sub>2</sub>Me), 1.75 (3H, s, H-4'), 1.63 (3H, s, H-5'); EIMS m/z 252 [M]<sup>+</sup> (21), 235 (8), 226 (75), 211 (33), 205 (18), 184 (44), 167 (5), 153 (46), 149 (8), 69 (100); HREIMS m/z 252.0978 (calcd for  $C_{13}H_{16}O_5$ , 252.0998)
- 4-O-(2-Methyl-2-butenyl)-3,5-dimethoxybenzoic Acid Methyl Ester (30). The method of preparation of 30 was similar to that used for the preparation of 15. The yield of 30 from **29** was 92%: gum;  ${}^{1}H$  NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.26 (2H, s, H-2, H-6), 5.52 (1H, brt, J = 7.2 Hz, H-2'), 4.55 (2H, d, J = 7.3 Hz, H-1', 3.89 (3H, s,  $\text{CO}_2Me$ ), 3.88 (3H, s, OMe-3,

- OMe-5), 1.72 (3H, s, H-4'), 1.64 (3H, s, H-5'); HREIMS m/z 280.1300 (calcd for C<sub>15</sub>H<sub>20</sub>O<sub>5</sub>, 280.1311).
- 4-O-(2-Methyl-2-butenyl)-3,5-dimethoxybenzyl Alcohol (31). The method of preparation of 31 was similar to that used for the preparation of **16**. The yield of **31** from **30** was 89%: gum;  ${}^{1}$ H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  6.53 (2H, s, H-2, H-6), 5.51 (1H, brt, J = 7.1 Hz, H-2'), 4.56 (2H, s, H-7), 4.42 (1H, d, J =7.2 Hz, H-1'), 3.79 (6H, s, OMe-3), OMe-5), 1.69 (3H, s, H-5'), 1.62 (3H, s, H-4'); EIMS m/z 252 [M]+ (6), 239 (6), 235 (5), 226 (28), 211 (10), 205 (33), 184 (100), 182 (2), 167 (14), 155 (12), 153 (8), 127 (8), 123 (12), 109 (9), 69 (18); HREIMS  $\delta$  252.1374 (calcd for C<sub>14</sub>H<sub>20</sub>O<sub>4</sub>, 252.1362).
- 4-O-(2-Methyl-2-butenyl)-3,5-dimethoxybenzaldehyde (32). The method of preparation of 32 was similar to that used for the preparation of 17. The yield of 32 from 31 was 85%: gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  9.81 (1H, s, H-7), 7.07 (2H, s, H-2, H-6), 5.49 (1H, t, J = 7.1 Hz, H-2'), 4.56 (2H, d, J = 7.3 Hz, H-1'), 3.87 (6H, s, OMe-3, OMe-5), 1.69 (3H, s, H-4'), 1.62 (3H, s, H-5'); EIMS m/z 250 [M]<sup>+</sup> (1), 235 (1), 226 (16), 196 (2), 182 (100), 167 (8), 153 (2), 139 (4), 125 (5), 110 (6), 95 (7); HREIMS m/z 250.1199 (calcd for C<sub>14</sub>H<sub>18</sub>O<sub>4</sub>, 250.1205).
- 4-O-(2-Methyl-2-butenyl)sinapic Acid (33). The method of preparation of 33 was similar to that used for the preparation of 18. The yield of 33 from 32 was 88%: gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.68 (1H, d, J = 15.7 Hz, H-7), 6.75 (1H, s, H-2, H-6), 6.34 (1H, d, J = 15.8 Hz, H-8), 5.53 (1H, dt, J =7.2, 1.3 Hz, H-2'), 4.57 (2H, d, J = 7.2 Hz, H-1'), 3.86 (6H, s, OMe-3, OMe-5), 1.72 (3H, s, H-4'), 1.65 (3H, s, H-5'); EIMS m/z 292 [M]<sup>+</sup> (6) 277 (2), 265 (5), 250 (3), 224 (100), 209 (40), 197 (3), 195 (3), 181 (10), 163 (12), 149 (8), 135 (9), 121 (15), 69 (69); HREIMS m/z 292.1303 (calcd for  $C_{16}H_{20}O_5$ , 292.1311).
- 4-O-(2-Methyl-2-butenyl)sinapyl Alcohol (34). The method of preparation of 34 was similar to that used for the preparation of 6 from 18. The yield of 34 from 33 was 81%, while the yield of byproduct **36** was 8%: gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  6.59 (2H, s, H-2, H-C6), 6.52 (1H, d, J = 16.0, H-7), 6.27 (1H, dt, J = 15.6, 5.7 Hz, H-8), 5.54 (1H, m, H-2'), 4.47 (2H, brd, J = 7.1 Hz, H-1'), 4.30 (2H, brd, J = 5.7 Hz, H-9), 3.84 (6H. s. OMe-3, OMe-5), 1.72 (3H. H-4'), 1.65 (3H. s. H-5'): HREIMS m/z 278.1532 (calcd for  $C_{16}H_{22}O_4$ , 278.1518).
- 4-O-(2-Methyl-2-butenyl)sinapaldehyde (35). The method of preparation of 35 was similar to that used for the preparation of 7 from 6. The yield of 35 from 34 was 91%: gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  9.66 (1H, d, J = 7.6 Hz, H-9), 7.40 (1H, d, J = 15.9 Hz, H-7), 6.77 (2H, br. s, H-2, H-6), 6.60 (1H, d)dd, J = 15.9, 7.6 Hz, H-8), 5.54 (1H, brt, J = 7.2 Hz, H-2'), 4.56 (2H, brd, J = 7.2 Hz, H-1'), 3.89 (6H, s, OMe-3, OMe-5), 1.72 (3H, s, H-4'), 1.65 (3H, s, Me-5'); HREIMS m/z 276.1351 (calcd for  $C_{16}H_{20}O_4$ , 276.1362).
- 4-O-(2-Methyl-2-butenyl)-7,8-dihydrosinapyl alcohol (36): gum; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  6.48 (2H, brs, H-2, H-6), 5.53 (1H, brt, J = 7.2 Hz, H-2'), 4.51 (2H, brd, J = 7.1Hz, H-1'), 3.90 (2H, brt, J = 7.5 Hz, H-9), 3.88 (6H, s, OMe-3, OMe-5), 2.81 (2H, brt, J = 7.5 Hz, H-7), 2.03-1.96 (2H, m, H-8), 1.71 (3H, s, H-4'), 1.65 (3H, s, H-5'); HREIMS m/z 278.1509 (calcd for C<sub>16</sub>H<sub>22</sub>O<sub>4</sub>, 278.1518).

Cytotoxicity Assay. KB cells were obtained from the American type culture collection. 12 Effects of compounds on the growth of the cells were monitored at the Laboratoire de Cultures Cellulaires, ICSN, Gif-sur-Yvette, France. The IC<sub>50</sub> values refer to the concentration of drug corresponding to 50% growth inhibition after 72 h incubation. 13 The assays of A-549 and HL-60 were carried out at the Institute of Shanghai Material Medica and were performed according to published  $techniques.^{18-20}\\$ 

Acknowledgment. This work was financially supported by the Life Science Special Fund of the Chinese Academy of Sciences supported by the Ministry of Finance (STZ-00-24); the Yunnan Province Foundation of Applied and Basic Research (2000C0072M); the Foundation for Visiting Professor from the State Key Laboratory of New Drug Research at SIMM; a grant from the Dean of KIB, CAS; and the Chine-France PRA (PRA BT01-02). One of the authors (Y.Z.) expresses his thanks to the Chinese Ministry of Education as well as to Mr. Ka-Shing Lee for a "Cheung Kong Scholar Chief Professorship" at Zhejiang University, and the EGIDE foundation (France) for a fellowship at ISCN, CNRS at Gif-sur-Yvette, where this work was finally accomplished. We also thank Christiane Gaspard for conducting the cytotoxicity assays.

#### **References and Notes**

- Jiangsu New Midical College. *A Dictionary of Traditional Chinese Medicines*; Shanghai People's Press: Shanghai, 1977, p 7. Zhao, Y.; Parsons, S.; Baxter, R. L.; Tan, R. X.; Jia, Z. J.; Sun, H. D.; Rankin, D. W. H. *Tetrahedron* **1997**, *53*, 6195–6208, and references
- Zhao, Y.; Jia, Z. J.; Yang, L. Phytochemistry 1994, 37, 1149-1152.
- (4) Marco, J. A.; Sanz-Cervera, J. F.; Garcia-Sarrion, A.; Rustaiyan, A. Phytochemistry 1991, 30, 2325-2328.
- (5) Bohlmann, F.; Fritz, U. Phytochemistry 1980, 19, 2471–2472.
  (6) Ishizaki, Y.; Tanahashi, Y.; Takahashi, T.; Tori, K. Tetrahedron 1970, and to proper proper from the control of the 26. 5387-5393.
- Bohlmann, F.; Grenz, M.; Gupta, R. K.; Dhar, A. K.; Ahmed, M.;
- King, R. M.; Robinson, H. *Phytochemistry* **1980**, *19*, 2391–2397. Gao, K.; Wang, W.-S.; Jia, Z.-J. *Phytochemistry* **1998**, *47*, 269–

- (9) Fujita, M.; Yamada, M.; Nakajima, S.; Kawai, K.-I.; Nagai, M. Chem. Pharm. Bull. 1984, 32, 2622-2627.
- (10) Mitsunobu, O. Synthesis 1981, 1-28.
- (a) Pearson, A. J.; Bruhn, P. R. J. Org. Chem. 1991, 56, 7092-7097. (b) Zhu, J.; Chastanet, J.; Beugelmans, R. Syn. Commun. 1996, 26, 2479-2486.
- (12) Eagle, H. Proc. Soc. Exp. Biol. Med. 1955, 89, 362-364.
- Tempête, C.; Werner, G. H.; Farve, F.; Roja, A.; Langlois, N. Eur. J. Chem. 1995, 30, 647-650.
- Jurd, L. J. Am. Chem. Soc. 1959, 81, 4606-4610.
- (a) Battersby, A. R.; Jones, R. C. F.; Kazlauskas, R.; Thornber, C. W.; Ruchirawat, S.; Staunton, J. *J. Chem. Soc., Perkin Trans. 1* **1981**, 2016–2029. (b) Kametani, T.; Yagi, H.; Kawamura, K.; Kohno, T. Chem. Pharm. Bull. 1970, 18, 645-650.
- (16) Buttersby, A. R.; Bhatnagar, A. K.; Hachett, P.; Thornber, C. W.; Staunton, J. J. Chem. Soc., Perkin Trans. 1 1981, 2002–2009.
- Kametani, T.; Satoh, F.; Yagi, H.; Fukumoto, K. J. Org. Chem. 1968, *33*, 690-694.
- Li, Q. Personal communication.
- Mosmann, T. J. Immunol. Methods **1983**, 65, 55–63.
- Skehan, P.; Storeng, R.; Scudiero, D.; Monks, A.; McMahon, J.; Vistica, D.; Warren, J. T.; Bokesch, H.; Kenny, S.; Boyd, M. R. *J. Natl.* Cancer Inst. 1990, 82, 1107-1112.

NP0200257