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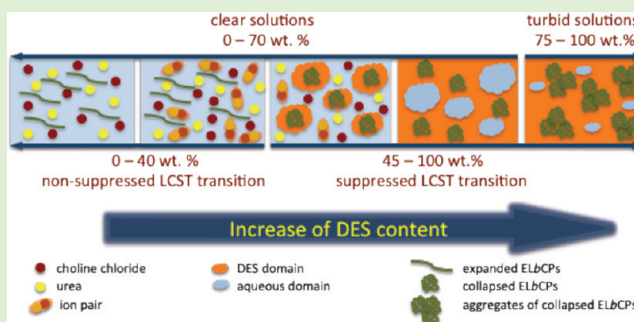
Phase Behavior of Elastin-Like Synthetic Recombinamers in Deep Eutectic Solvents

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ABSTRACT: Deep eutectic solvents promoted the stabilization of the collapsed state of elastin-like recombinamers – and the subsequent formation of aggregates – upon the loss of the structural water molecules involved in hydrophobic hydration. Cryo-etch scanning electron microscopy allowed the observation of these aggregates in neat deep eutectic solvents. The suppression of the lower critical solution temperature transition, observed by differential scanning calorimetry and dynamic light scattering, confirmed the presence of the elastin-like recombinamers in their collapsed state. Actually, the transition from the collapsed to the expanded state was suppressed even after moderate aqueous dilution – for water contents ranging from nil to ca. 45 wt % – and it was only recovered upon further addition of water – above 50 wt %. These features revealed the preferred stabilization of the collapsed state in not only neat deep eutectic solvents but also partially hydrated deep eutectic solvents. We consider that the capability to trigger the lower critical solution temperature transition by partial hydration of deep eutectic solvent may open interesting perspectives for nano(bio)technological applications of elastin-like recombinamers.



INTRODUCTION

Elastin-like polymers (ELPs) are synthetic polypeptides whose composition has been inspired by the repeating sequences found in natural elastin.^{1–8} Poly(VPGVG) is one of the most important member of the elastin-like family,⁹ where G stands for glycine, V stands for L-valine, and P stands for L-proline. Derivatives can be obtained by systematic substitutions of the valine amino acid residues of the pentamer by either natural or non-natural L-amino acids. In any case, the polymer chain is predominantly a hydrophobic polypeptide, where the only polar groups are the peptide-bond moieties themselves.¹⁰

In aqueous solutions, ELPs exhibit the capability to undergo a lower critical solution temperature (LCST) transition from a random coil to a collapsed (e.g., globular-like) structure mimicking the transition between the unfolded to the folded structure of natural elastins.^{11–13} It is widely accepted that in the collapsed state the polymer chains adopt a dynamic, regular, nonrandom structure stabilized by hydrophobic contacts that ultimately results in a phase-separated state.^{2,14–17} Mechanisms based on transition from expanded to collapsed state are actually behind the stimuli-responsive features of PNIPAM (poly(isopropyl acrylamide)), one of the most well-known thermoresponsive synthetic polymers that is often used as simple model for proteins.¹⁸

Nowadays, recombinant technology allows the production of special ELPs (the so-called elastin-like recombinamers - ELRs)

with a precision, monodisperse nature and degree of complexity that is not comparable to their chemically synthesized counterparts (e.g., bare ELPs).¹⁹ Interestingly, tailored molecular designs of ELRs allow switching the loss of hydrophobic hydration water that characterizes the LCST transition from the expanded to the collapsed state in response to different stimuli (e.g., not only different temperatures but also, under isothermal conditions, changes in pH or light).² This feature offers a tremendous versatility as compared with other synthetic stimuli-responsive polymers. For instance, dual-stimuli-responding polymers can be obtained from block copolymers (e.g., poly(*N*-isopropyl acrylamide-*co*-acrylic acid)) in which one component is temperature-sensitive and the other one is pH-sensitive (*N*-isopropyl acrylamide and acrylic acid, respectively).²⁰

Taking into account the fact that the LCST transition is related to hydrophobic association, one may wonder whether the transition from the random coil to the globular-like structure of ELRs takes place in nonaqueous solvents. This question might lack of interest for regular organic solvents if one considers that some of the most typical applications of ELRs are in biomedicine and biotechnology in which toxicity

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issues play a capital role.^{21,22} However, deep eutectic solvents (DESs)²³ are lately offering an interesting alternative to regular organic solvents for number of applications. DESs are obtained by complexation of quaternary ammonium salts with hydrogen bond donors. The charge delocalization occurring through hydrogen bonding between the halide anion with the hydrogen-donor moiety is responsible for the decrease in the freezing point of the mixture relative to the melting points of the individual components. DESs have been used as suitable solvents for dissolution/dispersion of number of biological entities (ranging from biomolecules up to microorganisms)^{24–28} and biopolymers^{29–31} but never for ELRs.

Herein, we have studied the dissolution/dispersion of ELRs in DES. The DES of choice was based on a mixture of urea and choline chloride in a 2:1 molar ratio (UCCI-DES). Since the seminal work by Abbot and coworkers describing a set of different DESs,²³ UCCI-DES has become one the most widely studied DESs and can be considered to be a model system. Actually, it has been the mixture of choice in number of papers studying the dissolution of biomolecules and biopolymers.^{24–31} With regards to ELRs, we used a linear diblock AB architecture (EL2bR) and a linear tetrablock (AB)₂ architecture (EL4bR). In both cases, one of the blocks consisted of a poly(L-valine–L-proline–L-glycine–L-valine–L-glycine) (VPGVG) in which the second valine has been regularly replaced by an L-glutamic acid (E) in one pentapeptide out of five, whereas the second block was a poly(L-valine–L-proline–L-alanine–L-valine–L-glycine) (VPAVG) in the diblock architecture and a poly(L-valine–L-glycine–L-isoleucine–L-proline–L-glycine) (VGIPG, where I stands for L-isoleucine) in the tetrablock one.^{32,33} Therefore, the polypeptide architecture of EL2bR was [(VPGVG)₂(VPGEG)(VPGVG)]₂₀-(VPAVG)₄₀ (*M_w* = 58.5 KDa), whereas that of EL4bR was {[(VPGVG)₂(VPGEG)-(VPGVG)]₁₀}[VGIPG]₆₀]₂ (*M_w* = 92.9 KDa). Both recombinamers can be considered to be simple stimuli-responsive polymers showing temperature and pH sensitiveness due to the presence of the glutamic acid residues. In practical terms, the apparent complexity of their respective block architecture just determines the temperature at which the LCST transition occurs (e.g., 33 °C for EL2bR and 16 °C for EL4bR).^{32,33} The LCST transition of elastin-like block-recombinamers (ELbRs) in UCCI-DES and aqueous dilutions of DES was studied by differential scanning calorimetry (DSC) and dynamic light scattering (DLS). The occurrence of ELbR phase segregation and aggregation in UCCI-DES was studied by cryo-etch-SEM.

■ EXPERIMENTAL PART

The preparation of UCCI-DES was accomplished by heating urea (from Aldrich) and choline chloride (from Aldrich) in a 2:1 molar ratio at 80 °C and stirring until a homogeneous liquid was formed.

The preparation of ELbRs was accomplished as described elsewhere.³² In brief, synthetic DNA duplexes encoding the oligopeptide (VPGVG)₂-(VPGEG)-(VPGVG)₂ and the oligopeptide (VPAVG)₂₀ (or (VGIPG)₆₀) were generated by polymerase chain reaction (PCR) amplification using synthetic oligonucleotides. The gene cloning, concatenation, and colony screening were performed as previously described.³⁴ Selected genes were subcloned into a modified pET-25(b) expression vector and transformed into the *E. coli* strain BLR(DE3). Expression conditions and purification protocols were as previously described.³⁴ Production yields for all of the polymers were between 80 and 200 mg/mL of bacterial culture.

The incorporation of ELbRs in UCCI-DES was accomplished by (1) dissolution of ELbR in water (1 or 5 wt %, under stirring over 24 h at 4 °C), (2) dissolution of the components that form the DES (e.g.,

urea and choline chloride in a 2:1 molar ratio) in the aqueous solution of ELbR (under stirring over further 24 h at 4 °C), and (3) freeze-drying.

Cryo-etch-SEM experiments were carried out on aqueous solutions of UCCI-DES, of ELbR and of UCCI-DES also containing ELbR. The aqueous solution of UCCI-DES was prepared by simple dissolution of the DES components in water. The UCCI-DES content in solution ranged from 0.6 to 1 wt %. The aqueous solutions of ELbR (without DES) were prepared under stirring over 24 h at 4 °C and then stored over another 24 h before analysis. The ELbR content in solution was always 2.5 wt %. The preparation of the aqueous solutions of UCCI-DES with ELbR was accomplished upon the addition of the DES components (urea and choline chloride in a 2:1 molar ratio and in the wt % mentioned above) to the aqueous solutions of ELbR described above. The solutions (any) were always stirred over 24 h at 4 °C and then stored at the same temperature over further 24 h before analysis. For comparison, aqueous solutions of ELbR (without UCCI-DES) that also contain NaCl (100 mM) were also studied. The aqueous solutions were placed in the sample holder of a cryotransfer system (Oxford CT1500), plunged into subcooled liquid nitrogen, and then transferred to a preparation unit via an air lock transfer device. The frozen specimens were cryofractured and transferred directly via a second air lock to the microscope cold stage, where they were etched for 2 min at –90 °C. After ice sublimation and in the preparation unit, the etched surfaces were sputter-coated with gold for 10 min at a sputter current of 10 mV. The thickness of the resulting gold film was within the 5–10 nm range, which allows for the undistorted observation of ELbR aggregates of ca. 100 nm diameter. Samples were subsequently transferred onto the cold stage of the scanning electron microscope chamber. Fractured surfaces were observed with a DSM 960 Zeiss scanning electron microscope at –135 °C under the following conditions: acceleration potential, 15 kV; working distance, 15 mm; and probe current, 5–10 nA.

DSC scans were carried out on aqueous solutions of ELbR (1 and 5 wt %) without and with UCCI-DES using a Mettler Toledo 822 DSC equipped with a liquid-nitrogen cooler and calibrated with a standard sample of indium. The aqueous solutions of ELbR without UCCI-DES were prepared under stirring over 24 h at 4 °C and then stored at 25 °C over another 24 h before analysis. The preparation of the aqueous solutions ELbR with DES was accomplished upon the addition of the DES components (urea and choline chloride in a 2:1 molar ratio) right after the addition of the aqueous solutions of ELbR; the solution was stirred over 24 h at 4 °C and then was stored at 25 °C over another 24 h. The final UCCI-DES content (in wt %) in the aqueous solutions ELbR was simply determined by the mass of urea and choline chloride added to the solution. After this period, 20 µL of any of these solutions (e.g., with or without UCCI-DES) was placed in 40 µL of aluminum pans hermetically sealed. An equal volume of water was placed in the reference pan. The heating program consisted of an initial isothermal stage (5 min at 5 °C), followed by heating at a constant rate of 5 °C min^{–1} from 0 to 60 °C and cooling at –5 °C min^{–1} to 0 °C.

DLS analyses were carried out on aqueous solutions of ELbR (1 wt %) that also contained UCCI-DES (45 and 55 wt %) using a Zetasizer Nano S (Malvern Instruments) equipped with a Peltier temperature controller. The aqueous solutions of ELbR either without or with UCCI-DES were prepared as described above for DSC experiments. After preparation, 400 µL of these solutions was placed into a PMMA cuvette. Data were collected within the 0 – 50 °C interval using a 1 °C min^{–1} heating ramp. The stabilization time between measurements was 10 min. The experiments were carried by quintuplicate.

■ RESULTS AND DISCUSSION

Both EL2bR and EL4bR were readily soluble in cold water (at temperatures below the LCST) but not in neat UCCI-DES. Therefore, direct addition of any of the ELbRs to neat UCCI-DES resulted in a nonhomogeneous suspension that resembled those obtained upon direct addition of any ELP to water when the temperature is above its respective LCST. Actually, the viscosity of UCCI-DES made any attempt for ELbRs

homogenization even more difficult than in water. Under these circumstances, the incorporation of ELbRs in UCCI-DES was accomplished following an indirect procedure that allows transitioning from aqueous to DES chemistry.³⁵ In brief, the procedure consisted of (1) dissolution of ELbR in water (1 wt %, under stirring over 24 h at 4 °C), (2) dissolution of the components that form the DES (e.g., urea and choline chloride) in the ELbR aqueous solution (under stirring over further 24 h at 25 °C), and (3) freeze-drying. The concentration of urea and choline chloride added in (2) was so chosen for a final ELbR content in UCCI-DES that ranged from 1 to 5 wt %. After freeze-drying, the turbid-like nature of the resulting UCCI-DES suspensions of ELbRs (Figure 1) was indicative of the formation of aggregates, the size of which was large enough to scatter visible wavelengths.

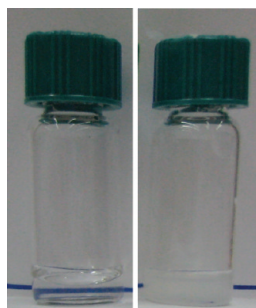


Figure 1. Left: clear solution resulting from EL2bR dissolution in water at 4 °C. Right: turbid suspension of EL2bR in DES (prepared as described in the main text). The EL2bR content was 1 wt % in both cases.

The turbidity of UCCI-DES suspensions of ELbRs resembled that of ELbRs in aqueous solutions above the LCST.^{32,33} In this latter case, the free polymer chains of ELbRs that remained disordered (e.g., in form of fully hydrated – by hydrophobic hydration – random coils) below the LCST undergo a hydrophobically driven collapse and assemble into a phase-separated state above the LCST. According to Urry's and Tamburro's models,^{14–17} the polymer chains adopt a dynamic, regular, nonrandom structure, called a β -spiral, that involves type II β -turns as the main secondary structural feature and is stabilized by intrasprial interturn and intersprial hydrophobic contacts. When the concentration of ELbR in aqueous solution is above a threshold value, hydrophobic contacts can be established not only between segments of an individual polymer chain – to form single globules – but also between segments of two or more different polymer chains and then form globular aggregates that scatter visible wavelengths.³⁶ Eventually, turbid gels are typically obtained when these aggregates are capable of forming a continuous network across the entire original solution.³³ In neat UCCI-DES, we hypothesized that the absence of water promotes ELbRs to lose essentially all of the ordered water structures resulting from hydrophobic hydration and hence the formation of globular aggregates.

The occurrence of ELbR phase segregation (to the collapsed state) and aggregation (of the resulting globules) upon freeze-drying was first studied by cryo-etch-SEM. In cryo-etch-SEM experiments, aqueous samples are plunge-frozen by immersion in subcooled liquid nitrogen (e.g., liquid nitrogen at vacuum pressure). Under these circumstances, the freezing rate is $\sim 10^4$ °C s^{−1}.³⁷ This freezing rate is far from that needed to obtain

supercooled water by freezing of pure water solutions (ca. 10^6 °C s^{−1}).³⁸ However, the presence of impurities makes freezing rates of $\sim 10^4$ °C s^{−1} fast enough to impede the formation of crystalline ice.³⁷ The frozen specimens were cryo-fractured and transferred to the microscope cold stage, where the temperature is subsequently raised to −90 °C to favor the formation of crystalline ice that, in pseudodiluted samples, readily frees itself of any solute and concentrates between adjacent ice crystals. Exposed ice vaporizes by sublimation in an etching-like process so that the resulting morphology corresponds to empty areas (where ice originally resided) surrounded by the concentrated solutes in a “fence-like” fashion. In summary, cryo-etch-SEM allows the observation of any solute originally dissolved in water after the submission of this solution to a freeze-drying process. This is why we considered that cryo-etch-SEM was a suitable technique for the observation of ELbR aggregates in UCCI-DES. Actually, cryo-etch-SEM has been previously used for observation of either the expanded or the collapsed state of ELbR upon the freezing of aqueous solutions (without further additives) that, respectively, were below or above the phase transition.^{39,40} Nonetheless, it is worth noting that the formation of “fence-like” morphologies is mainly governed by the cryo-etch process and neither the composition nor the conformation of the solutes play a capital role in this issue. Actually, “fence-like” morphologies can also be observed by cryo-etch-SEM using pseudodiluted solutions of not only seawater, polymers and silica sols^{41–44} but also of neat DESs (Figure 2).^{26,35} In our case, cryo-etch-SEM was performed on

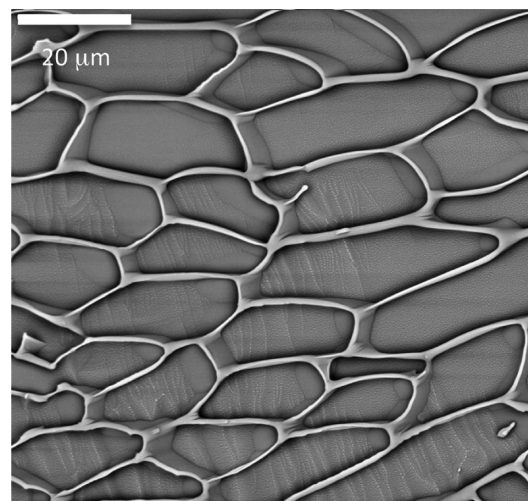


Figure 2. Cryo-etch-SEM micrograph of an aqueous solution of UCCI-DES without ELbRs. Bar is 20 μm .

clear ELbR aqueous solutions (also prepared at 4 °C and hence, below the phase transition of ELbR) without and with UCCI-DES. In this latter case, the DES components were added in the proper molar ratio but at a high dilution. (See the Experimental Part for further details.) In both cases, the ELbR aqueous solutions (maintained at 4 °C) were frozen by simple immersion in subcooled liquid nitrogen (e.g., liquid nitrogen at vacuum pressure). A close inspection to the texture of the fences revealed the differences resulting from the addition of DES components to the ELbR aqueous solutions. Thus, ELbR aqueous solutions without DES components displayed no further texture than the “fence-like” morphology (Figure 3) in agreement with previous cryo-etch-SEM studies of ELbRs in

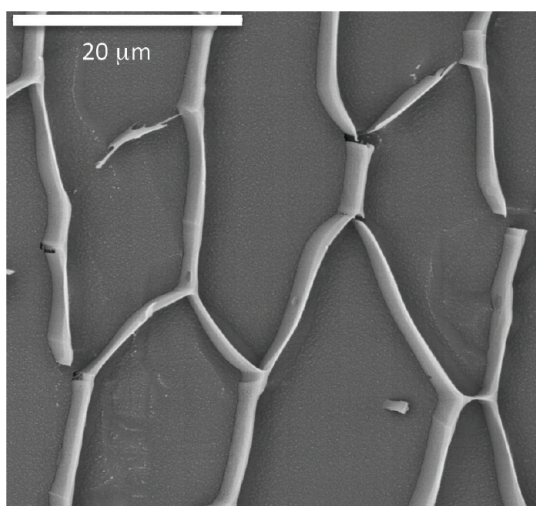


Figure 3. Cryo-etch-SEM micrograph of an aqueous solution of EL2bR without UCCI-DES representing a typical “fence-like” morphology. Bar is 20 μm . The cryo-etch-SEM micrograph of an aqueous solution of EL4bR without UCCI-DES is not shown because it did not show any significant difference.

the expanded state,^{39,40} whereas spherical micellar aggregates (characteristic of the collapsed one^{32,33}) could be observed within the “fences” of ELbRs aqueous solutions also containing DES components (Figure 4). Interestingly, micellar structures were also observed by cryo-etch-SEM in ELbR aqueous solutions that contained NaCl instead of UCCI-DES (Figure 5). It is generally assumed that in aqueous solutions of ELbR the presence of NaCl promotes the stabilization of ELbR in both the expanded and the collapsed states of ELP,⁴⁵ the former because of the better structured corona of hydrophobic hydration surrounding the nonpolar moieties and the latter due to a typical salting out effect that results in a better organization of the polymer in the collapsed state. Therefore, the formation of ELbR micelles after the freeze-drying of both UCCI-DES and NaCl aqueous solutions seems to be favored by the ionic character of the media. Actually, DESs are considered to be part of the ionic liquids family (and hence, molten salts), the ionic strength of which was eventually capable to stabilize the collapsed states of ELbRs by preventing the globular structures from self-fusion processes that typically occur in bare water (Figure 3 and 4a). Nonetheless, it is worth noting that the observation of globular structures was clearer in NaCl than in UCCI-DES because the segregation between globules and the surrounding medium (which is ultimately determining how easy the observation is) is obviously favored for solid (e.g., NaCl) versus liquid (e.g., DES) media.

The occurrence of a salting-out-like process in DESs, in which the ionic strength of the media is favoring the collapsed state of ELbRs, was corroborated when aggregation persisted upon the addition of a certain amount of water (e.g., 24 wt % in Figure 6). The UCCI-DES suspensions of ELbRs only became clear for water contents above 30 wt % (Figure 6) and, even in this case, DSC studies (see DSC plots in Figure 7 and data in Tables 1 and 2) demonstrated that ELbR yet remained in the collapsed state. Actually, water contents above 50 wt % were required for recovering of LCST transitions in both EL2bR and EL4bR.

The persistence of the collapsed state for water contents ranging from 30 to 35 to 50–55 wt % (e.g., clear ELbR

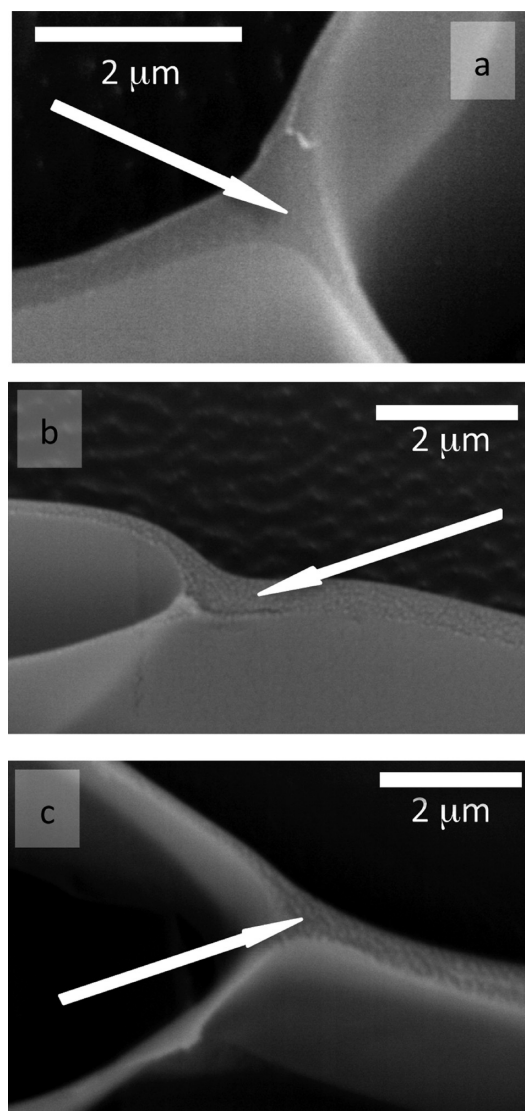


Figure 4. Detail of a cryo-etch-SEM micrograph of an aqueous solution of EL4bR without UCCI-DES (a), of an aqueous solution of EL2bR that also contains UCCI-DES (b), and of an aqueous solution of EL4bR that also contains UCCI-DES (c). In these two latter cases, arrows point to the tiny aggregates of ca. 100 nm that can be observed within the “fence-like structures” formed by UCCI-DES after ice sublimation. Bars are 2 μm . The detail of a cryo-etch-SEM micrograph of an aqueous solution of EL2bR without UCCI-DES is not shown because it did not show any significant difference as compared with that of EL4bR shown in panel a.

solutions, Figure 6) was confirmed by DLS. Figure 8 shows that water contents of 64 and 54 wt % were required for the observation of the LCST transition from the expanded (low optical density) to the collapsed (high optical density) state in EL2bR and EL4bR, respectively. Otherwise (e.g., for water contents of 54 wt % in EL2bR and 44 wt % in EL4bR), the LCST was suppressed and high optical densities were observed along the entire range of temperatures used in the experiment. It is worth noting that the ELbR diameters in the collapsed state were below 120 nm in every case as the clear character of all of these solutions anticipated. Nonetheless, it was also observed how these diameters increased along with the decrease in the water content. This feature was in agreement with the above-mentioned formation of larger globular

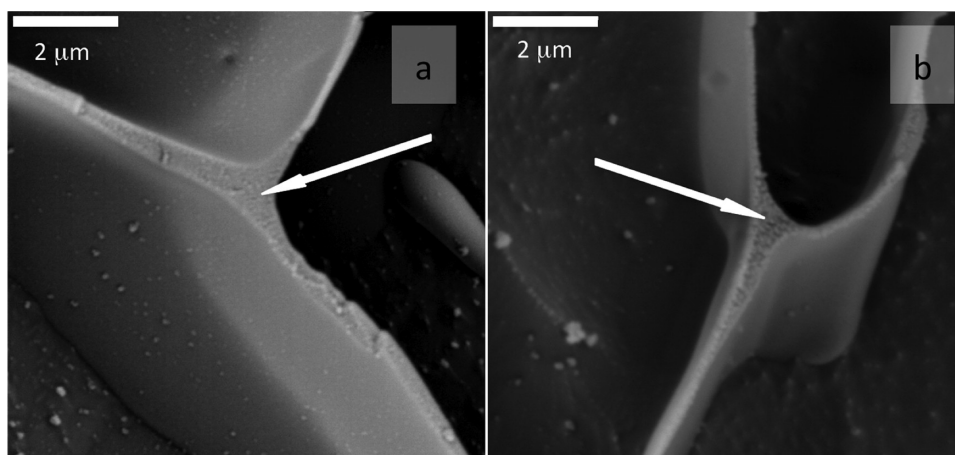


Figure 5. Cryo-etch-SEM micrographs of aqueous solutions of EL2bR (a) and EL4bR (b) containing 100 mM NaCl. Bars are 2 μm . Arrows point to the tiny aggregates of ca. 100 nm that can be observed within the “fence-like structures” formed by NaCl after ice sublimation.



Figure 6. Picture of different aqueous solutions of UCCI-DES also containing EL4bR (from left to right, the water content was 94, 64, 44, 34, 29, and 24 wt %). The EL4bR content was always 1 wt %. The picture of different aqueous solutions of UCCI-DES also containing EL2bR is not shown because it did not show any significant difference.

aggregates under conditions that are favorable for hydrophobic contacts. Whether DLS experiments could be influenced by the particular physical properties of UCCI-DES (e.g., viscosity, refractive index, density, etc.) was disregarded because, at the water contents used in the experiments, differences between UCCI-DES dilutions and bare water were not significant (Table 3).

One may also wonder what (if any) the evolution of neat DES (without ELbR) is upon dilution. It is generally assumed that DES dilution promotes the rupture of the original supramolecular complexes that are forming the DES so that for high water contents DES components become individual molecules that are solvated by the aqueous environment. We have recently used ^1H NMR spectroscopy to study UCCI-DES rupture along with the water content.³⁵ Thus, for low water contents (e.g., 15–40 wt %), the chemical shifts of the NMR spectra revealed that water molecules are solvated by UCCI-DES, whereas the opposite situation (i.e., water solvating the individual components of DES) was found for high water contents (70–85 wt %). We also found that transitioning from low to high water contents implies the growth of the water domains that, in a first stage, can be yet solvated by UCCI-DES and, later on, promote the rupture of UCCI-DES into its individual components that are solvated by water molecules.³⁵ Obviously, the opposite also applies; transitioning from high to low water contents promotes the growing of UCCI-DES

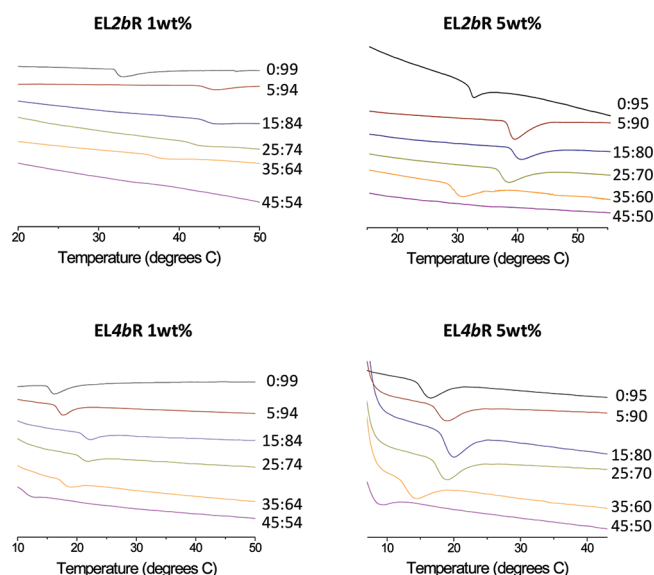


Figure 7. DSC scans of EL2bR and EL4bR suspended in aqueous solutions of DES. The ELbR contents were 1 and 5 wt %. Numbers besides scans represent the DES:water ratio in wt %. Thus, the DES contents (in wt %) ranged from 0 to 45 wt %. DSC scans of ELbRs solutions having 5 wt % were recorded for better observation of the LCSTs. It is worth noting that the absolute value of the LCST (see Tables 1 and 2 for details) slightly depends on the ELbRs content, but the evolution along with the DES content does not.

domains that, initially (i.e., while they are small enough), are water solvated, but, after further growing, the situation reverts into small water domains solvated by UCCI-DES.

In this stage, it is worth looking again at the DSC curves of ELbRs in water and in aqueous dilutions of UCCI-DES represented in Figure 7. The temperatures of the endothermic peak of EL2bR and EL4bR in water were 33 and 16 $^{\circ}\text{C}$, respectively. First addition of DES components (ca. 5–15 wt %) resulted in an increase of in the transition temperature (see the shift of the endothermic peaks in Figure 7, data can be found in Tables 1 and 2) because of the destabilization of the collapsed state.⁴⁶ This is actually the same behavior that is observed with the vast majority of proteins in which unfolding is promoted by the strong denaturing character of urea.^{47–49} The endothermic peak shifted to lower temperatures upon

Table 1. Inverse Transition Temperature of Aqueous Solutions of EL2bRs Containing Different DES Contents (from nil to 45 wt %) Obtained from DSC Scans^a

| sample | T_i by DSC (T_i by DLS) (°C) |
|---|--------------------------------------|
| EL2bR 1 wt % - H ₂ O 99 wt % | 33.0 |
| EL2bR 1 wt % - DES 5 wt % - H ₂ O 94 wt % | 44.7 |
| EL2bR 1 wt % - DES 15 wt % - H ₂ O 84 wt % | 44.3 |
| EL2bR 1 wt % - DES 25 wt % - H ₂ O 74 wt % | 42.9 |
| EL2bR 1 wt % - DES 35 wt % - H ₂ O 64 wt % | 38.2 (34.0) |
| EL2bR 1 wt % - DES 45 wt % - H ₂ O 54 wt % | |
| EL2bR 5 wt % - H ₂ O 95 wt % | 32.2 |
| EL2bR 5 wt % - DES 5 wt % - H ₂ O 90 wt % | 39.0 |
| EL2bR 5 wt % - DES 15 wt % - H ₂ O 80 wt % | 40.0 |
| EL2bR 5 wt % - DES 25 wt % - H ₂ O 70 wt % | 38.2 |
| EL2bR 5 wt % - DES 35 wt % - H ₂ O 60 wt % | 30.4 |
| EL2bR 5 wt % - DES 45 wt % - H ₂ O 50 wt % | |

^aInverse transition temperature obtained from DLS experiments is also included for DES contents of 35 wt % (between brackets).

Table 2. Inverse Transition Temperature of Aqueous Solutions of EL4bRs Containing Different DES Contents (from nil to 55 wt %) Obtained from DSC Scans^a

| sample | T_i by DSC (T_i by DLS) (°C) |
|---|--------------------------------------|
| EL4bR 1 wt % - H ₂ O 99 wt % | 16.2 |
| EL4bR 1 wt % - DES 5 wt % - H ₂ O 94 wt % | 17.6 |
| EL4bR 1 wt % - DES 15 wt % - H ₂ O 84 wt % | 22.2 |
| EL4bR 1 wt % - DES 25 wt % - H ₂ O 74 wt % | 22.0 |
| EL4bR 1 wt % - DES 35 wt % - H ₂ O 64 wt % | 17.8 |
| EL4bR 1 wt % - DES 45 wt % - H ₂ O 54 wt % | 12.6 (9.0) |
| EL4bR 1 wt % - DES 55 wt % - H ₂ O 44 wt % | |
| EL4bR 5 wt % - H ₂ O 95 wt % | 16.4 |
| EL4bR 5 wt % - DES 5 wt % - H ₂ O 90 wt % | 19.0 |
| EL4bR 5 wt % - DES 15 wt % - H ₂ O 80 wt % | 20.0 |
| EL4bR 5 wt % - DES 25 wt % - H ₂ O 70 wt % | 18.9 |
| EL4bR 5 wt % - DES 35 wt % - H ₂ O 60 wt % | 14.3 |
| EL4bR 5 wt % - DES 45 wt % - H ₂ O 50 wt % | 8.6 |
| EL2bR 5 wt % - DES 50 wt % - H ₂ O 45 wt % | |

^aInverse transition temperature obtained from DLS experiments is also included for DES contents of 45 wt % (between brackets).

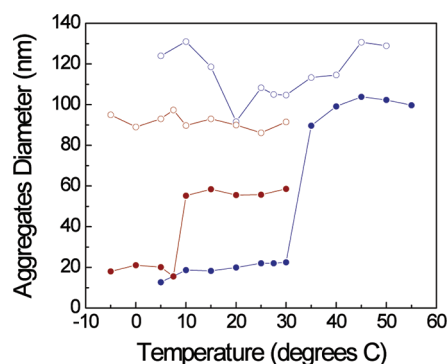


Figure 8. Hydrodynamic diameter of EL2bR (blue symbols) and EL4bR (red symbols) versus temperature in different aqueous solutions of DES. The water contents in EL2bR solutions were 64 (solid symbols) and 54 wt % (open symbol). The water contents in EL4bR solutions were 54 (solid symbols) and 44 wt % (open symbol). The ELbRs content was 1 wt %. The heating rate was 1 °C/min.

Table 3. Physical Properties of UCCI-DES and of Its Aqueous Dilutions^a

| UCCI-DES/H ₂ O (wt %) | viscosity (cP) | density (g/mL) | refractive index |
|----------------------------------|----------------|----------------|------------------|
| 100:0 | 1108.1 | 1.192 | 1.505 |
| 95:5 | 139.2 | 1.188 | 1.495 |
| 85:15 | 23.1 | 1.171 | 1.483 |
| 75:25 | 6.8 | 1.150 | 1.457 |
| 65:35 | 3.1 | 1.113 | 1.441 |
| 55:45 | 2.1 | 1.112 | 1.420 |
| 45:55 | 1.7 | 1.090 | 1.415 |
| 35:65 | 1.5 | 1.067 | 1.387 |
| 25:75 | 1.3 | 1.048 | 1.372 |
| 15:85 | 1.2 | 1.026 | 1.355 |
| 5:95 | 1.1 | 1.010 | 1.340 |
| 0:100 | 1 | 1.000 | 1.333 |

^aData were taken at 20 °C. The solutions used for DLS experiments are marked in grey.

further addition of DES components (up to intermediate contents ranging from 15 to 35 wt %). According to previous results found by ¹H NMR spectroscopy (see above),³⁵ the formation of urea-choline chloride ion pairs starts within this concentration range. The strong interaction that the DES components themselves establish to form the ion pairs restricts the availability of the individual DES components^{50–53} so that whatever their original activity/reactivity was, it tends to decrease or even vanish.²⁶ In our case, the decrease in the urea denaturing-activity would explain the stabilization of the collapsed state of ELbRs. Nonetheless, the endothermic peak shifting to lower temperatures could also be ascribed to the eventual increase in the ionic strength coming from UCCI-DES formation, as recently demonstrated by calorimetric studies on ELbR solutions containing salts (e.g., NaCl).⁴⁵ Previous ¹H NMR spectroscopy studies also revealed that upon further addition of DES components (ca. 40 wt %) UCCI-DES domains started to prevail over the aqueous ones.³⁵ For UCCI-DES contents ranging from 45 to 70 wt %, the LCST transitions of EL2bR and EL4bR were suppressed in the DSC scans over the entire range of temperatures where the LCST typically occurs. This feature was indicative of the preferred stabilization of one of the states that, according to the above-described DLS experiments, was the collapsed one. The prevalence of the collapsed state within this range of UCCI-DES content (e.g., 40–70 wt %) was confirmed when, upon further addition of UCCI-DES (e.g., UCCI-DES contents above 70 wt %), the solution became turbid, resembling the typical aspect of aqueous solutions of ELbRs above the LCST. Figure 9

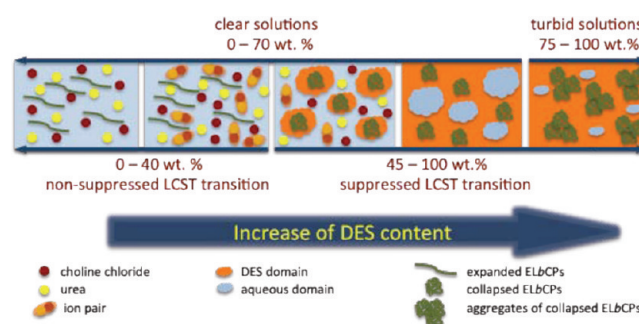


Figure 9. Scheme representing the transition from the expanded to the collapsed state of ELbRs depending on the DES/water ratio.

provides an idealized representation of the situation of ELbRs (at either the expanded or the collapsed state) and UCCI-DES (in which the components are either dissolved or forming domains) upon the transition from aqueous to poorly or nonhydrated conditions, respectively.

CONCLUSIONS

Summarizing, this manuscript reports on the study of ELbRs suspended in UCCI-DES. Cryo-etch-SEM demonstrated that the preferred conformation of ELbR in UCCI-DES is in the collapsed state. DSC and DLS confirmed that UCCI-DES provides a nonhydrated solvent of relatively high ionic strength that favors the stabilization of ELbR in the collapsed state. DSC and DLS also revealed that the collapsed state remains stable even after partial hydration (e.g., UCCI-DES aqueous solution with water contents of up to 50–55 wt %). Considering that many nano(bio)technological applications of ELP are related to the occurrence of the LCST transition,¹⁴ the capability to trigger this transition from the expanded to the collapsed state of ELbR in UCCI-DES by partial hydration is of interest not only from a fundamental point of view but also from a practical one because it may open novel and interesting fields where ELbRs can be applied.

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Notes

The authors declare no competing financial interest.

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REFERENCES

- (1) Qu, Y.; Payne, S. C.; Apkarian, R. P.; Conticello, V. P. *J. Am. Chem. Soc.* **2000**, *122*, 5014–5015.
- (2) Urry, D. W.; Luan, C. H.; Parker, T. M.; Gowda, D. C.; Prasad, K. U.; Reid, M. C.; Safavy, A. J. *Am. Chem. Soc.* **1991**, *113*, 4345–4346.
- (3) Meyer, D. E.; Chilkoti, A. *Nat. Biotechnol.* **1999**, *17*, 1112–1115.
- (4) Rodríguez-Cabello, J. C.; Reguera, J.; Girotti, A.; Alonso, M.; Testera, A. M. *Prog. Polym. Sci.* **2005**, *30*, 1119–1145.
- (5) Cappello, J.; Crissman, J. W.; Crissman, M.; Ferrari, F. A.; Textor, G.; Wallis, O.; Whitley, J. R.; Zhou, X.; Burman, D.; Aukerman, L.; Stedronsky, E. R. *J. Controlled Release* **1998**, *53*, 105–117.
- (6) Spezzacatena, C.; Pepe, A.; Green, L. M.; Sandberg, L. B.; Bochicchio, B.; Tamburro, A. M. *Eur. J. Org. Chem.* **2005**, *8*, 1644–1651.
- (7) Nuhn, H.; Klok, H.-A. *Biomacromolecules* **2008**, *9*, 2755–2763.
- (8) Winkler, S.; Wilson, D.; Kaplan, D. L. *Biochemistry* **2000**, *39*, 12739–12746.
- (9) Urry, D. W. *Angew. Chem., Int. Ed.* **1993**, *32*, 819–841.
- (10) Urry, D. W. *J. Phys. Chem.* **1997**, *101*, 11007–11028.
- (11) San Biagio, P. L.; Madonia, F.; Trapane, T. L.; Urry, D. W. *Chem. Phys. Lett.* **1988**, *145*, 571–574.
- (12) Rodríguez-Cabello, J. C.; Alonso, M.; Pérez, T.; Herguedas, M. M. *Biopolymers* **2000**, *54*, 282–288.
- (13) Valiaev, A.; Lim, D. W.; Schmidler, S.; Clark, R. L.; Chilkoti, A.; Zauscher, S. J. *Am. Chem. Soc.* **2008**, *130*, 10939–10946.
- (14) Urry, D. W. In *What Sustains Life? Consilient Mechanisms for Protein-Based Machines and Materials*; Springer: Singapore, 2006.

- (15) San Biagio, P. L.; Madonia, F.; Trapane, T. L.; Urry, D. W. *Chem. Phys. Lett.* **1988**, *145*, 571–574.
- (16) Tamburro, A. M.; Guantieri, V.; Pandolfo, L.; Scopa, A. *Biopolymers* **1990**, *29*, 855–870.
- (17) Lelj, F.; Tamburro, A. M.; Villani, V.; Grimaldi, P.; Guantieri, V. *Biopolymers* **1992**, *32*, 159–170.
- (18) Schild, H. G.; Tirrell, D. A. *J. Phys. Chem.* **1990**, *94*, 4352–4356.
- (19) Rodríguez-Cabello, J. C.; Martín, L.; Alonso, M.; Arias, F. J.; Testera, A. M. *Polymer* **2009**, *50*, 5159–5169.
- (20) Xia, F.; Feng, L.; Wang, S.; Sun, T.; Song, W.; Jiang, W.; Jiang, L. *Adv. Mater.* **2006**, *18*, 432–436.
- (21) Daamen, W. F.; Veerkamp, J. H.; van Hest, J. C. M.; van Kuppevelt, T. H. *Biomaterials* **2007**, *28*, 4378–4398.
- (22) Woodhouse, K. A.; Klement, P.; Chen, V.; Gorbet, M. B.; Keeley, F. W.; Stahl, R.; Fromstein, J. D.; Bellingham, C. M. *Biomaterials* **2004**, *25*, 4543–4553.
- (23) Abbott, A. P.; Capper, G.; Davies, D. L.; Rasheed, R. K.; Tambyrajah, V. *Chem. Commun.* **2003**, 70–71.
- (24) Byrne, N.; Angell, C. A. *Chem. Commun.* **2009**, 1046.
- (25) Meli, L.; Miao, J.; Dordick, J. S.; Linhardt, R. J. *Green Chem.* **2010**, *12*, 1883–1892.
- (26) Gutiérrez, M. C.; Ferrer, M. L.; Yuste, L.; Rojo, F.; del Monte, F. *Angew. Chem., Int. Ed.* **2010**, *49*, 2158–2162.
- (27) Perriman, A. W.; Cölfen, H.; Hughes, R. W.; Barrie, C. L.; Mann, S. *Angew. Chem., Int. Ed.* **2009**, *48*, 6242–6246.
- (28) Perriman, A. W.; Brogan, A. P. S.; Cölfen, H.; Tsoureas, N.; Owen, G. R.; Mann, S. *Nat. Chem.* **2010**, *2*, 622–626.
- (29) Phillips, D. M.; Drummy, L. F.; Conrady, D. G.; Fox, D. M.; Naik, R. R.; Stone, M. O.; Trulove, P. C.; DeLong, H. C.; Mantz, R. A. *J. Am. Chem. Soc.* **2004**, *126*, 14350–14351.
- (30) Murugesan, S.; Wiencek, J. M.; Ren, R. X.; Linhardt, R. J. *Carbohydr. Polym.* **2006**, *63*, 268–271.
- (31) Vijayaraghavan, R.; Thompson, B. C.; MacFarlane, D. R.; Kumar, R.; Surianarayanan, M.; Aishwarya, S.; Sehgal, P. K. *Chem. Commun.* **2010**, *46*, 294–296.
- (32) Ribeiro, A.; Arias, F. J.; Reguera, J.; Alonso, M.; Rodríguez-Cabello, J. C. *Biophys. J.* **2009**, *97*, 312–320.
- (33) Martín, L.; Arias, F. J.; Alonso, M.; García-Arévalo, C.; Rodríguez-Cabello, J. C. *Soft Matter* **2010**, *6*, 1121–1124.
- (34) Girotti, A.; Reguera, J.; Rodríguez-Cabello, J. C.; Arias, F. J.; Alonso, M.; Matestera, A. J. *Mater. Sci., Mater. Med.* **2004**, *15*, 479–484.
- (35) Gutiérrez, M. C.; Ferrer, M. L.; Mateo, C. R.; del Monte, F. *Langmuir* **2009**, *25*, 5509–5515.
- (36) Dreher, M. R.; Simnick, A. J.; Fischer, K.; Smith, R. J.; Patel, A.; Schmidt, M.; Chilkoti, A. J. *Am. Chem. Soc.* **2008**, *130*, 687–694.
- (37) Kirsop, B. E.; Doyle, A. *Maintenance of Microorganism and Cultured Cells*; Academic Press: London, 1991.
- (38) Velikov, V.; Borick, S.; Angell, C. A. *Science* **2001**, *294*, 2335.
- (39) McMillan, R. A.; Caran, K. L.; Apkarian, R. P.; Conticello, V. P. *Macromolecules* **1999**, *32*, 9067–9070.
- (40) Wright, E. R.; McMillan, R. A.; Cooper, A.; Apkarian, R. P.; Conticello, V. P. *Adv. Funct. Mater.* **2002**, *12*, 149–154.
- (41) Menger, F. M.; Zhang, H.; Caran, K. L.; Seredyuk, V. A.; Apkarian, R. P. *J. Am. Chem. Soc.* **2002**, *124*, 1140–1141.
- (42) Menger, F. M.; Galloway, A. L.; Chlebowski, M. E.; Apkarian, R. P. *J. Am. Chem. Soc.* **2004**, *126*, 5987–5989.
- (43) Ferrer, M. L.; Esquembre, R.; Ortega, I.; Mateo, C. R.; del Monte, F. *Chem. Mater.* **2006**, *18*, 554–559.
- (44) Gutiérrez, M. C.; García-Carvajal, Z. Y.; Jobbágy, M.; Yuste, L.; Rojo, F.; Ferrer, M. L.; del Monte, F. *Adv. Funct. Mater.* **2007**, *17*, 3505–3513.
- (45) Reguera, J.; Urry, D. W.; Parker, T. M.; McPherson, D. T.; Rodríguez-Cabello, J. C. *Biomacromolecules* **2007**, *8*, 354–358.
- (46) Sagie, L. B.; Zhang, Y.; Litosh, V. A.; Chen, X.; Cho, Y.; Cremer, P. S. *J. Am. Chem. Soc.* **2009**, *131*, 9304–9310.
- (47) Bennion, B. J.; Daggett, V. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 5142–5147.
- (48) Schellman, J. A. *Biophys. Chem.* **2002**, *96*, 91–101.

- (49) Pace, C. N.; Marshall, H. F. *Arch. Biochem. Biophys.* **1980**, *199*, 270–276.
- (50) Carriazo, D.; Gutiérrez, M. C.; Ferrer, M. L.; del Monte, F. *Chem. Mater.* **2010**, *22*, 6146–6152.
- (51) Gutiérrez, M. C.; Carriazo, D.; Ania, C.; Parra, J.; Ferrer, M. L.; del Monte, F. *Energy Environ. Sci.* **2011**, *4*, 3535–3544.
- (52) Gutiérrez, M. C.; Carriazo, D.; Tamayo, A.; Jiménez, R.; Picó, F.; Rojo, J. M.; Ferrer, M. L.; del Monte, F. *Chem.—Eur. J.* **2011**, *17*, 10533–10537.
- (53) Serrano, M. C.; Gutiérrez, M. C.; Ferrer, M. L.; del Monte, F. *Chem. Commun.* **2012**, *48*, 579–581.