

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/14511679>

# 2-Phenyl-4-quinolinecarboxamides: A Novel Class of Potent and Selective Non-Peptide Competitive Antagonists for the Human Neurokinin-3 Receptor

ARTICLE in JOURNAL OF MEDICINAL CHEMISTRY · JULY 1996

Impact Factor: 5.45 · DOI: 10.1021/jm9602423 · Source: PubMed

CITATIONS

54

READS

46

12 AUTHORS, INCLUDING:



**Giuseppe A.M. Giardina**

Istituto Italiano di Tecnologia

116 PUBLICATIONS 1,199 CITATIONS

SEE PROFILE



**Carlo Farina**

Metys Pharmaceuticals AG

130 PUBLICATIONS 1,170 CITATIONS

SEE PROFILE



**Luca Raveglia**

Aptuit

53 PUBLICATIONS 858 CITATIONS

SEE PROFILE



**Douglas Hay**

Hay DDC, LLC

143 PUBLICATIONS 5,530 CITATIONS

SEE PROFILE

# JOURNAL OF MEDICINAL CHEMISTRY

© Copyright 1996 by the American Chemical Society

Volume 39, Number 12

June 7, 1996

## *Communications to the Editor*

### **2-Phenyl-4-quinolinecarboxamides: A Novel Class of Potent and Selective Non-Peptide Competitive Antagonists for the Human Neurokinin-3 Receptor**

Giuseppe A. M. Giardina,<sup>\*,†</sup> Henry M. Sarau,<sup>‡</sup> Carlo Farina,<sup>†</sup> Andrew D. Medhurst,<sup>§</sup> Mario Grugni,<sup>†</sup> James J. Foley,<sup>‡</sup> Luca F. Raveglia,<sup>†</sup> Dulcie B. Schmidt,<sup>‡</sup> Roberto Rigolio,<sup>†</sup> Marco Vassallo,<sup>†</sup> Vittorio Vecchietti,<sup>†</sup> and Douglas W. P. Hay<sup>‡</sup>

*Department of Chemistry, SmithKline Beecham S.p.A.,  
Via Zambeletti, 20021 Baranzate, Milano, Italy,*

*Department of Pulmonary Pharmacology,*

*SmithKline Beecham Pharmaceuticals, 709 Swedeland*

*Road, King of Prussia, Pennsylvania 19406-0939, and*

*Department of Neurology Research, SmithKline Beecham  
Pharmaceuticals, New Frontiers Science Park,*

*Third Avenue, Harlow, Essex, England CM19 5AW*

*Received March 27, 1996*

Pharmacological and molecular biological studies indicate the existence of at least three human tachykinin receptor subtypes, designated neurokinin-1 (NK-1), neurokinin-2 (NK-2), and neurokinin-3 (NK-3),<sup>1–3</sup> which belong to the superfamily of G-protein-coupled receptors possessing seven transmembrane domains.<sup>4</sup> The endogenous ligands for these receptors constitute a family of small neuropeptides, named tachykinins or neurokinins, which share the common carboxy-terminal region Phe-X-Gly-Leu-MetNH<sub>2</sub>. The main mammalian tachykinins, substance P, neurokinin A (NKA), and neurokinin B (NKB), interact with all three tachykinin receptors, although there is a defined agonist rank order of potency for NK-1, NK-2, and NK-3 receptors, respectively; for example, for the NK-3 receptor the rank potency order is NKB > NKA > substance P.

Over the past few years potent and selective peptide and non-peptide antagonists for the NK-1 and NK-2 receptors have been identified.<sup>5–8</sup> These pharmacological tools accelerated the clarification of physiological and pathophysiological roles of these receptors<sup>9</sup> and the

potential therapeutic indications for NK-1 and NK-2 receptor antagonists.<sup>10–13</sup> In contrast to the NK-1 and NK-2 receptor research area, there is limited information on the biology and potential pathophysiological significance of the NK-3 receptor. This was, to a large extent, due to the lack of sufficient potency and selectivity of the peptide NK-3 antagonists described thus far<sup>14</sup> and to the absence of non-peptide NK-3 receptor ligands until recently. However, the recent disclosure of the "peptoid" NK-3 antagonist PD 157672<sup>15</sup> and the peptide-derived PD 161182<sup>16</sup> has provided improved reagents for these studies and stimulated the search for more potent and metabolically stable non-peptide NK-3 receptor antagonists.

The human NK-3 (hNK-3) receptor mRNA has been detected, using polymerase chain reaction (PCR), to various regions in the central nervous system (CNS) and also, albeit to a lesser extent, in some peripheral tissues, including kidney, placenta, lung, and colon.<sup>17</sup> Activation of NK-3 receptors modulates the release of various transmitters in the CNS and periphery,<sup>18–20</sup> suggesting that they may have a neuromodulatory role.

Recently, (*S*)-(+)-*N*-{3-[1-benzoyl-3-(3,4-dichlorophenyl)piperidin-3-yl]prop-1-yl}-4-phenylpiperidin-4-yl}-*N*-methylacetamide, SR 142801 (**1**, Chart 1), was reported as the first potent non-peptide NK-3 receptor antagonist.<sup>21,22</sup> Chemically, SR 142801 derives from a constrained analog of SR 48968, a potent NK-2 receptor antagonist (*K*<sub>i</sub> = 0.51 nM, for displacement of [<sup>125</sup>I]-NKA) in rat duodenum membranes,<sup>7</sup> which has moderate affinity for NK-3 receptors (IC<sub>50</sub> = 320 nM, for displacement of [<sup>3</sup>H]senktide {succinyl-[Asp<sup>9</sup>MePhe<sup>8</sup>]-SP(6–13)}) in guinea pig cerebral cortex membranes.<sup>23</sup>

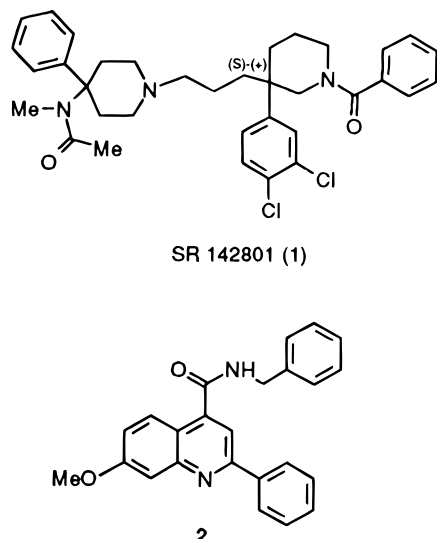
We now report on the discovery of a novel class of potent and selective non-peptide NK-3 receptor antagonists—structurally unrelated to the piperidine derivatives SR 142801 and SR 48968—which are based on the 2-phenylquinoline backbone.<sup>24</sup> Chemical synthesis (Scheme 1), radioligand binding affinities for the cloned human neurokinin receptors stably expressed in CHO cell lines (hNKs-CHO),<sup>17,25,26</sup> *in vitro* functional activity in the rabbit isolated iris sphincter muscle preparation (antagonism of senktide-induced contraction),<sup>27</sup> and structure–activity relationships (SARs) of the novel

<sup>†</sup> Italy.

<sup>‡</sup> USA.

<sup>§</sup> England.

Chart 1

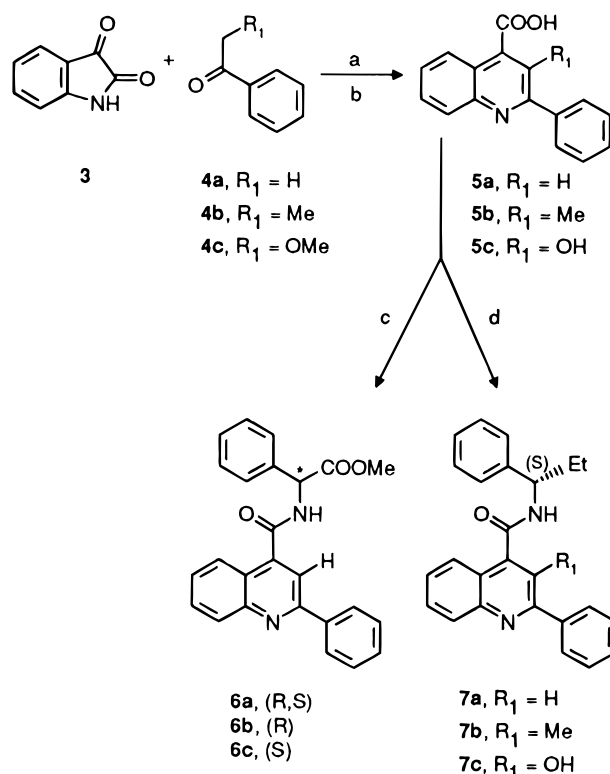


potent NK-3 receptor antagonists **6a–c** and **7a–c** (Table 1) will be summarized; a comparison to SR 142801 will be presented. For compound **7c**, functional activity in HEK 293 cells expressing the hNK-3 receptor (NKB-induced  $\text{Ca}^{2+}$  mobilization)<sup>22</sup> will be described.

**Results and Discussion.** The 4-quinolinecarboxamide derivative **2**, specifically designed to arrange the phenyl ring in position 2 and the benzene ring of the carboxamide side chain in a relative space position suitable for a potential interaction with NK (particularly NK-1) receptors,<sup>28</sup> was shown to possess no affinity for hNK-1 receptors ( $K_i > 10 \mu\text{M}$ , for displacement of [ $^3\text{H}$ ]-substance P binding from hNK-1-CHO membranes) and moderate affinity for the hNK-3 receptor ( $K_i = 533 \text{ nM}$ , for displacement of [ $^{125}\text{I}$ ]MePhe<sup>7</sup>-NKB binding from hNK-3-CHO membranes). Thus, compound **2** was utilized as a structural lead, suitable for chemical modifications aimed at optimizing the NK-3 receptor binding affinity of the compound, by variation of the carboxamide side chain in position 4 and of the substitution pattern on the quinoline ring system.

The synthesis of compounds **6a–c** and **7a–c** was accomplished as described in Scheme 1. Isatin **3** was submitted to the Pfitzinger reaction, by refluxing with phenyl ketones **4a–c** in the presence of an excess of potassium hydroxide to obtain the 2-phenylquinoline-4-carboxylic acids **5a,b** in high yields; 3-hydroxy-2-phenylquinoline-4-carboxylic acid (**5c**) was obtained by refluxing the corresponding methoxy intermediate in 48% hydrobromic acid. Secondary carboxamides **6a–c** and **7a–c** were obtained by coupling the carboxylic acids **5a–c** with the appropriate primary amines in a 7:3 mixture of THF and MeCN in the presence of dicyclohexylcarbodiimide (DCC) and 1-hydroxybenzotriazole (HOBT) to maximize yields and avoid racemization of the resulting compounds.

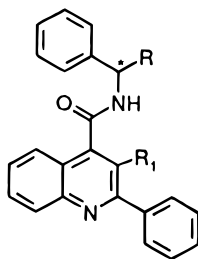
Incorporation of the methoxycarbonyl substituent at the benzyl position of compound **2** and removal of the 7-methoxy group produced a 17-fold increase in hNK-3 receptor binding affinity (Table 1, cf. **6a**, hNK-3-CHO binding,  $K_i = 30.7 \text{ nM}$ , with compound **2**, hNK-3-CHO binding,  $K_i = 533 \text{ nM}$ ). Further studies with enantiomerically pure stereoisomers **6b** and **6c** revealed that hNK-3 receptor binding affinity and functional activity in rabbit isolated iris sphincter muscle (antagonism of

Scheme 1.<sup>a</sup> Synthesis of Compounds of General Formula **6a–c** and **7a–c**

<sup>a</sup> Reagents: (a) KOH, EtOH, reflux; (b)  $\text{R}_1 = \text{OMe}$ , 48% HBr, reflux; (c) methyl phenylglycinate hydrochloride, DCC, HOBT, Et<sub>3</sub>N, THF/MeCN, room temperature; (d) (S)-1-phenylpropylamine, DCC, HOBT, THF/MeCN, room temperature.

senktide-induced contractions) are enantioselective properties of this chemical series, the (R)-stereoisomer **6b** (hNK-3-CHO binding,  $K_i = 13.3 \text{ nM}$ ) being about 2-fold more potent than racemic **6a** and almost 2 orders of magnitude more potent than the (S)-enantiomer **6c** (hNK-3-CHO binding,  $K_i = 925 \text{ nM}$ ) as shown in Table 1. Neurokinin receptor selectivity studies using radioligand competition binding with hNK-2 and hNK-1 receptors expressed in CHO cell membranes demonstrated that compound **6b** has an exquisite selectivity for hNK-3 with respect to the hNK-1 receptor (hNK-1/hNK-3  $K_i$  ratio  $> 7500$ ) and a good selectivity versus the hNK-2 receptor (hNK-2/hNK-3  $K_i$  ratio = 91).

Further chemical efforts were aimed at (a) replacing the ester functionality to circumvent potential metabolic instability problems and (b) optimizing the NK-3 receptor binding affinity and selectivity. Incorporation of a variety of substituents at the benzyl position (including alkyl, aralkyl, aryl, aminoalkyl, hydroxyalkyl, alkoxyalkyl, acyl, acid, and amide groups) revealed that linear two or three carbon atom chains maintained high and enantioselective NK-3 binding affinity and *in vitro* functional antagonist activity (data not shown). The ethyl derivative **7a** (hNK-3-CHO binding,  $K_i = 18.0 \text{ nM}$ ) was therefore selected to investigate the effect of the quinoline substitution pattern on biological activities. Whereas substitution at position 5, 6, 7, and 8 of the quinoline ring system produced compounds with potencies lower or similar to that of the unsubstituted analog, incorporation of alkyl groups at position 3 of the quinoline ring resulted, in general, in an increased binding affinity for the hNK-3 receptor. For example, incorporation of a methyl group, as in compound **7b** (R

**Table 1.** Chemical Structures, Binding Affinities at Cloned Human Neurokinin Receptors (hNK-1, hNK-2, and hNK-3) Expressed in CHO Cells, and *in Vitro* Functional Activities (Antagonism of Senktide-Induced Contractions) in Rabbit Isolated Iris Sphincter Muscle (RISM) Preparation for Compounds **6a–c**, **7a–c**, and SR 142801

compound	R	R <sub>1</sub>	config (*)	binding affinities, K <sub>i</sub> mean ± SEM (nM) <sup>a</sup>			antagonism of senktide-induced contractions in RISM, <sup>b</sup> K <sub>b</sub> <sup>f</sup> mean ± SEM (nM) <sup>a</sup>
				hNK-3 <sup>c</sup>	hNK-2 <sup>d</sup>	hNK-1 <sup>e</sup>	
<b>6a</b>	COOMe	H	( <i>R,S</i> )	30.7 ± 3.8	2947 ± 585	>100000	42 ± 12
<b>6b</b>	COOMe	H	( <i>R</i> )	13.3 ± 2.7	1221 ± 189	>100000	43 ± 6.2
<b>6c</b>	COOMe	H	( <i>S</i> )	925 ± 250	8333 ± 926	>100000	3982 (2)
<b>7a</b>	Et	H	( <i>S</i> )	18.0 ± 1.0	237 (2)	>100000	9.7 ± 1.7
<b>7b</b>	Et	Me	( <i>S</i> )	4.2 ± 0.6	277 ± 57	>100000	7.7 ± 1.0
<b>7c</b>	Et	OH	( <i>S</i> )	1.0 ± 0.1	144 ± 22	>100000	5.4 ± 3.4
SR 142801 ( <b>1</b> ) <sup>g</sup>				1.2 ± 0.2	40.3 ± 5.5	1300 (2)	2.0 ± 0.5

<sup>a</sup> Average of three to eight independent determinations ( $n = 3-8$ ), unless otherwise indicated in parentheses. <sup>b</sup> According to ref 27. <sup>c</sup> Inhibition of [<sup>125</sup>I]MePhe<sup>7</sup>-NKB binding in hNK-3-CHO cell membranes, according to ref 17. <sup>d</sup> Inhibition of [<sup>125</sup>I]NKA binding in hNK-2-CHO cell membranes, according to ref 25. <sup>e</sup> Inhibition of [<sup>3</sup>H]substance P binding in hNK-1-CHO cell membranes, according to ref 26. <sup>f</sup> The equilibrium-dissociation constant K<sub>b</sub> for the antagonist–NK-3 receptor complex was calculated from the equation: K<sub>b</sub> = [B]/CR – 1 where CR is the concentration ratio of agonist used in the presence and absence of antagonist B. <sup>g</sup> In-house data.

= Et, R<sub>1</sub> = Me), produced a 4-fold increase of the hNK-3 receptor binding affinity (hNK-3-CHO, K<sub>i</sub> = 4.2 nM), possibly due to a limitation of the conformational space of the 4-carboxamide side chain.

This finding led to a comprehensive investigation of the optimal substituent at position 3, aimed at improving NK-3 binding affinity, *in vitro* functional antagonist activity, and selectivity for the hNK-3 versus hNK-2 receptors (a feature apparently reduced in the α-Et series, cf. compound **7a** with **6b**). (*S*)-(–)-*N*-(α-Ethylbenzyl)-3-hydroxy-2-phenylquinoline-4-carboxamide (**7c**, SB 223412), featuring a hydroxy group at position 3 of the quinoline nucleus, possessed the best overall profile identified among compounds synthesized in this class thus far. Compound **7c** is, in fact, as potent (hNK-3-CHO binding, K<sub>i</sub> = 1.0 nM; antagonism of senktide-induced contraction in rabbit isolated iris sphincter muscle, K<sub>b</sub> = 5.4 nM) as the reference compound SR 142801 (hNK-3-CHO binding, K<sub>i</sub> = 1.2 nM; antagonism of senktide-induced contraction in rabbit isolated iris sphincter muscle, K<sub>b</sub> = 2.0 nM), but is more selective for hNK-3 versus the other tachykinin receptors: it has a hNK-2/hNK-3 K<sub>i</sub> ratio of 144 (vs 13 determined for the 3-H parent, **7a**, and 34 determined for SR 142801) and a hNK-1/hNK-3 K<sub>i</sub> ratio > 100 000 (vs 1000 for SR 142801).

Moreover, compound **7c** (at concentrations of 1 or 10 μM) was without effect in a multitude of assays for various receptors, ion channels, and enzymes (data not shown), including sodium channel (site 2) and opioid (μ, κ, δ) receptors for which SR 142801 was described to possess binding affinities in the 0.1 to 1 μM concentration range.<sup>21</sup>

The cellular functional activity of compound **7c** was also evaluated utilizing HEK 293 cells expressing the hNK-3 receptor; compound **7c**, at 10–1000 nM concentration range, produced a concentration dependent antagonism of NKB-induced Ca<sup>2+</sup> mobilization with a K<sub>b</sub> of 3.0 nM. Schild plot analysis of these data revealed

a slope of the regression line of 1.2, consistent with competitive antagonism. Unlike SR 142801,<sup>29</sup> compound **7c** displayed no time dependence for activity and its effects were rapidly reversed by washing, providing further evidence that it is acting as a reversible, competitive antagonist.

In summary, through the use of cloned human neurokinin receptor assays, we have discovered a novel chemical class of potent and selective non-peptide competitive antagonists for the hNK-3 receptor based on the quinoline structure; the most prominent member is compound **7c**, which has equivalent potency and greater selectivity for the hNK-3 versus the hNK-2 and hNK-1 receptors than SR 142801 (**1**), the only potent non-peptide NK-3 receptor antagonist described to date. We have also shown that these novel NK-3 receptor antagonists have potent functional activity in the rabbit isolated iris sphincter muscle preparation. Compounds **7b** and **7c** should prove to be extremely useful pharmacological tools for anatomical localization and *in vitro* and *in vivo* functional characterization of hNK-3 receptors. Information gained from such studies will help to elucidate the most appropriate therapeutic indications for selective NK-3 receptor antagonists.

A more detailed description of our investigations within this chemical class, including SARs for the ester and the 2-phenyl ring replacements, and for the quinoline ring substitution will be reported in subsequent manuscripts, along with the results of our studies aimed at the replacement of the quinoline ring by other (hetero)aromatic systems.

**Acknowledgment.** We are grateful to Joseph Weinstein, Fabrizio Bellucci, Geoffrey D. Clarke, and Paola Zaratini for the helpful discussions; we acknowledge Davide Graziani and Stefano Cavagnera for their chemical technical expertise and Karl Erhard for the production of SR 142801 via chiral HPLC separation of the racemic mixture (in-house produced); and we also thank

Renzo Mena and Alberto Cerri for mass and NMR spectroscopic determinations. [Medicinal Chemistry, SmithKline Beecham Pharmaceuticals, King of Prussia, Pennsylvania, 19406-0939 (J.W., K.E.); Department of Chemistry (F.B., D.G., S.C., R.M., A.C.) and Department of Biology (G.D.C., P.Z.), SmithKline Beecham SpA Via Zambelletti, 20021 Baranzate, Milan, Italy.]

**Supporting Information Available:** Detailed synthetic procedures and spectroscopic and analytical data for compounds **6a–c** and **7a–c** (5 pages). Ordering information is given on any current masthead page.

## References

- Maggio, J. E. Tachykinins. *Annu. Rev. Neurosci.* **1988**, *11*, 13–28.
- Regoli, D.; Drapeau, G.; Dion, S.; Couture, R. New Selective Agonists for Neurokinin Receptors: Pharmacological Tools for Receptor Characterization. *Trends Pharmacol. Sci.* **1988**, *9*, 290–295.
- Maggi, C. A.; Patachini, R.; Rovero, P.; Giachetti, A. Tachykinin Receptors and Tachykinin Receptor Antagonists. *J. Auton. Pharmacol.* **1993**, *13*, 23–93.
- Nakanishi, S. Mammalian Tachykinin Receptors. *Annu. Rev. Neurosci.* **1991**, *14*, 123–136.
- Snider, R. M.; Constantine, S. J. W.; Lowe, J. A.; Longo, K. P.; Lebel, W. S.; Woody, H. A.; Drozda, S. E.; Desai, M. C.; Vinick, F. J.; Spencer, R. W.; Hess, H. J. A Potent Nonpeptide Antagonist of the Substance P (NK-1) Receptor. *Science* **1991**, *251*, 435–437.
- Emonds-Alt, X.; Vilain, P.; Goulaouic, P.; Proietto, V.; Van Broeck, D.; Advenier, C.; Naline, E.; Neliat, G.; Le Fur, G.; Breliere, J. C. A Potent and Selective Non-Peptide Antagonist of the Neurokinin A (NK-2) Receptor. *Life Sci.* **1992**, *50*, 101–106.
- McElroy, A. B. Neurokinin Receptor Antagonists. *Curr. Opin. Ther. Pat.* **1993**, *3*, 1157–1172.
- Desai, M. C. Recent Advances in the Discovery and Characterization of Substance P Antagonists. *Expert Opin. Ther. Pat.* **1994**, *4*, 315–321.
- Mantyh, P. W.; Vigna, S. R.; Maggio, J. E. Receptor Involvement in Pathology and Disease. In *The Tachykinin Receptors*; Buck, S. H., Ed.; Humana Press: Totowa, NJ, 1994; pp 581–610.
- Lowe, J. A.; Snider, R. M. The Role of Tachykinins in Pulmonary Disease. In *Annual Reports in Medicinal Chemistry*; Bristol, J. A., Ed.; Academic Press, Inc.: New York, 1993; Vol. 28, pp 99–107.
- Tattersall, F. D.; Rycroft, W.; Hill, R. G.; Hargreaves, R. J. Enantioselective Inhibition of Apomorphine-Induced Emesis in the Ferret by the Neurokinin-1 Receptor Antagonist CP-99,994. *Neuropharmacology* **1994**, *33*, 259–260.
- Walsh, D. M.; Stratton, S. C.; Harvey, F. J.; Beresford, I. J. M.; Hagan, R. M. The Anxiolytic-Like Activity of GR159897, a Non-Peptide NK-2 Receptor Antagonist, in Rodent and Primate Models of Anxiety. *Psychopharmacology* **1995**, *121*, 186–191.
- Ishizuka, O.; Mattiasson, A.; Andersson, K.-E. Tachykinin Effects on Bladder Activity in Conscious Normal Rats. *J. Urol.* **1995**, *154*, 257–261.
- Drapeau, G.; Rouissi, N.; Nantel, F.; Rhaleb, N.-E.; Tousignant, C.; Regoli, D. Antagonists for the Neurokinin NK-3 Receptor Evaluated in Selective Receptor Systems. *Regul. Pept.* **1990**, *31*, 125–135.
- Boden, P.; Eden, J. M.; Hodgson, J.; Horwell, D. C.; Howson, W.; Hughes, J.; McKnight, A. T.; Meecham, K.; Pritchard, M. C.; Raphy, J.; Ratcliffe, G. S.; Suman-Chauhan, N.; Woodruff, G. N.; The Rational Development of Small Molecule Tachykinin NK-3 Receptor Selective Antagonists—The Utilization of a Dipeptide Chemical Library in Drug Design. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 1679–1684.
- Boden, P.; Eden, J. M.; Hodgson, J.; Horwell, D. C.; Pritchard, M. C.; Raphy, J.; Suman-Chauhan, N. The Development of a Novel Series of Non-Peptide Tachykinin NK-3 Receptor Selective Antagonists. *Bioorg. Med. Chem. Lett.* **1995**, *16*, 1773–1778.
- Buell, G.; Schulz, M. F.; Arkinstall, S. J.; Maury, K.; Missotten, M.; Adami, N.; Talabot, F.; Kawashima, E. Molecular Characterization, Expression and Localization of Human Neurokinin-3 Receptor. *FEBS* **1992**, *299*, 90–95.
- Stoessl, A. J.; Dourish, C. T.; Iversen, S. D. Pharmacological Characterization of the Behavioural Syndrome Induced by the NK-3 Tachykinin Agonist Senktide in Rodents: Evidence for Mediation by Endogenous 5-HT. *Brain Res.* **1990**, *517*, 111–116.
- Stoessl, A. J.; Szczutkowski, E.; Glenn, B.; Watson, I. Behavioural Effects of Selective Tachykinin Agonists in Midbrain Dopamine Regions. *Brain Res.* **1991**, *565*, 254–263.
- Arenas, E.; Alberch, J.; Perez-Navarro, E.; Solsona, C.; Marsal, J. Neurokinin Receptors Differentially Mediate Endogenous Acetylcholine Release Evoked by Tachykinins in the Neostriatum. *J. Neurosci.* **1991**, *11*, 2332–2338.
- Emonds-Alt, X.; Bichon, D.; Ducoux, J. P.; Heaulme, M.; Miloux, B.; Poncelet, M.; Proietto, V.; Van Broeck, D.; Vilain, P.; Neliat, G.; Soubrie, P.; Le Fur, G.; Breliere, J. C. SR 142801, the First Potent Non-Peptide Antagonist of the Tachykinin NK-3 Receptor. *Life Sci.* **1995**, *56*, PL 27–32.
- Oury-Donat, F.; Carayon, P.; Thurneyssen, O.; Pailhon, V.; Emonds-Alt, X.; Soubrie, P.; Le Fur, G. Functional Characterization of the Non-Peptide Neurokinin-3 (NK-3) Receptor Antagonist, SR 142801 on the Human NK-3 Receptor Expressed in Chinese Hamster Ovary Cells. *J. Pharmacol. Exp. Ther.* **1995**, *274*, 148–154.
- Petitot, F.; Beaujouan, J.-C.; Saffroy, M.; Torrens, Y.; Glowinski, J. The Nonpeptide NK-2 Antagonist SR 48968 is also a NK-3 Antagonist in the Guinea Pig but not in the Rat. *Biochem. Biophys. Res. Commun.* **1993**, *191*, 180–187.
- Farina, C.; Giardina, G. A. M.; Grugni, M.; Raveglia, L. F. International Patent Application WO 95/32948 (07.12.95).
- Aharony, D.; Conner, G. E.; Woodhouse, D. P. Pharmacologic Characterization of the Novel Ligand [4,5-<sup>3</sup>H-LEU<sup>9</sup>] Neurokinin-A Binding to NK-2 Receptors on Hamster Urinary Bladder Membranes. *Neuropeptides* **1992**, *23*, 121–130.
- Payan, D. G.; McGillis, J. P.; Organist, M. L. Binding Characteristics and Affinity Labeling of Protein Constituents of the Human IM-9 Lymphoblast Receptor for Substance P. *J. Biol. Chem.* **1986**, *261*, 14321–14329.
- Hall, J. M.; Mitchell, D.; Morton, I. K. M. Neurokinin Receptors in the Rabbit Iris Sphincter Characterised by Novel Agonist Ligands. *Eur. J. Pharmacol.* **1991**, *199*, 9–14.
- Desai, M. C.; Lefkowitz, S. L.; Thadeio, P. F.; Longo, K. P.; Snider, R. M. Discovery of a Potent Substance P Antagonists: Recognition of the Key Molecular Determinant. *J. Med. Chem.* **1992**, *35*, 4911–4913.
- Patachini, R.; Barthò, L.; Holzer, P.; Maggi, C. A. Activity of SR 142801 at Peripheral Tachykinin Receptors. *Eur. J. Pharmacol.* **1995**, *278*, 17–25.

JM9602423