See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/6055256

Phytochemicals for Health, the Role of Pulses

| ARTICLE in JOURNAL OF AGRICULTURAL AND Impact Factor: 2.91 · DOI: 10.1021/jf071704w · Source: PubMed |) FOOD CHEMISTRY · NOVEMBER 2007 | |
|--|----------------------------------|--|
| | | |
| CITATIONS | READS | |
| 105 | 263 | |
| | | |
| | | |
| | | |
| 2 AUTHORS: | | |



Simone Rochfort

Department of Environment and Primary I...

68 PUBLICATIONS **1,266** CITATIONS

SEE PROFILE



Joe Panozzo

Department of Environment and Primary I...

96 PUBLICATIONS 1,231 CITATIONS

SEE PROFILE

REVIEWS

Phytochemicals for Health, the Role of Pulses

SIMONE ROCHFORT*,† AND JOE PANOZZO‡

Department of Primary Industries, Werribee Centre, 621 Sneydes Rd, Werribee 3030, Victoria, Australia, and Department of Primary Industries, Horsham Centre, 110 Natimuk Road, Horsham 3400, Victoria, Australia

Pulses are the seeds of legumes that are used for human consumption and include peas, beans, lentils, chickpeas, and fava beans. Pulses are an important source of macronutrients, containing almost twice the amount of protein compared to cereal grains. In addition to being a source of macronutrients and minerals, pulses also contain plant secondary metabolites that are increasingly being recognised for their potential benefits for human health. The best-studied legume is the soybean, traditionally regarded as an oilseed crop rather than a pulse. The potential health benefits of soy, particularly with respect to isoflavone content, have been the subject of much research and the focus of several reviews. By comparison, less is known about pulses. This review investigates the health potential of pulses, examining the bioactivity of pulse isoflavones, phytosterols, resistant starch, bioactive carbohydrates, alkaloids and saponins. The evidence for health properties is considered, as is the effect of processing and cooking on these potentially beneficial phytochemicals.

KEYWORDS: Pulses; phytochemicals; macronutrients

INTRODUCTION

Pulses have traditionally played a major role in providing food nutrition particularly in the Indian subcontinent and other developing countries, while in western countries, the staple diet has been based on animal-derived protein.

Traditionally, pulses were consumed with minimal processing, and consumers were interested primarily with size, shape, and color characteristics. The markets were driven by price and availability. As the countries of the Indian subcontinent developed, a greater emphasis was placed on processing characteristics, which included hydration and cooking times as well as dehulling and splitting efficiency. This represents the current status for the majority of markets that consume pulse grains as a staple diet. While these market traits represent basic quality characteristics, the underlying chemical characteristics are based on protein and starch composition and phenolic compounds that affect the taste and color of the seed coat and cotyledon.

The nutritional properties of pulses have been investigated extensively and have been reported to impart physiologically beneficial effects in humans. Pulse grains are high in protein, carbohydrates, and dietary fiber and are a rich source of other nutritional components (1). The chemical composition and nutritive value of Australian pulses has been collated by Peterson et al. (2).

The value of pulses can be enhanced by physically fractionating the grain into basic constituents such as protein, starch, and fiber and using these products to supplement other food ingredients to enhance the nutritive value of food. There is now an increased awareness of the health-associated value of pulses in western countries. Pulse grains contain a large number of bioactive compounds which have a metabolic benefit when consumed on a regular basis (3).

Demand has increased regarding the use of pulses for human consumption either to extract a functional compound (e.g., starch protein or fiber) to incorporate this into cereal-based products or to extract phytocompounds which are bioactive and can be used as nutraceutical products.

Figure 1 represents the changing emphasis for plant breeding and consumer demand. There is a need to increase the knowledge base for pulses by understanding more of the functional and bioactive properties of pulse grains (4).

Considerable genetic variation has been reported in the chemical composition of pulses both between and within species. In addition, chemical composition is modified by environmental factors during plant development, and many of the phytocompounds are secondary metabolites produced during seed development and seed maturation.

This paper reviews the current knowledge around certain classes of pulse phytochemicals, including starch, phytosterols,

^{*} To whom correspondence should be addressed. Tel.: +61 3 9742 8704. Fax: +61 3 9742 8700. E-mail: simone.rochfort@dpi.vic.gov.au.

[†] Werribee Centre.

[‡] Horsham Centre.

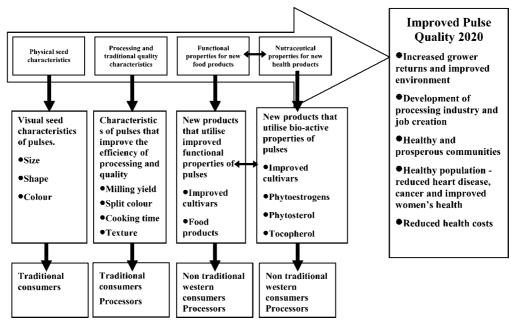


Figure 1. Developing quality pulses for a sustainable environment, population, and community. (4)

isoflavones, saponins, alkaloids, and bioactive carbohydrates. The potential for these metabolites to influence human health is discussed as are processing methods and agricultural practices that influence the levels of these compounds in food.

RESISTANT STARCH

Starch is a major carbohydrate in pulse grains, and due to its high concentration of amylose, the process of digestion and metabolism is therefore of interest, particularly as there is a strong negative correlation between the intake of starch and the risk of colorectal cancer (5).

Starch can be classified according to digestibility as soluble, insoluble, or resistant starch (RS). Until recently, starch was thought to undergo complete breakdown and absorption upon digestion. In 1992, Englyst et al. referred to RS as the proportion of starch that is not hydrolyzed or digested as it passes through the gastrointestinal tract (6). Resistant starch that reaches the large intestine has a physiological function similar to that of dietary fiber. Resistant starch can be considered a probiotic and acts as a substrate for microbiological fermentation, producing short-chain fatty acids (SCFAs), methane, and carbon dioxide, conferring benefits to human colonic health, and to a lesser extent can impact lipid and glucose metabolism. The production of these fermentation products from the consumption of RS is less than that from the consumption of nondigestible oligosaccharides (7). It is believed that the SCFAs produced mediate the benefits of RS rather than RS exerting a physical bulking effect (8).

Short-chain fatty acids consist principally of butyrate, propionate, and acetate and are metabolic products of anaerobic bacterial fermentation (9) and are the preferred respiratory fuel of the colonocytes lining the colon. These cells serve to increase blood flow, lower luminal pH, and help prevent abnormal colonic cell populations (10). Human feeding studies have shown that RS consumption in a diet results in an increase of SCFA in the colon (11).

Pulse grains are high in RS (**Table 1**) and retain their functionality even after cooking (12).

Worldwide, the dietary intake of RS varies considerably. In developing countries, the intake is between 30 and 40 g/day (13). In the EU, the intake of RS is between 3 and 6 g/day

Table 1. RS Composition in Pulse Grains and Wheat Bran (12)

| grain product | raw (% RS) | cooked (% RS) |
|-----------------|------------|---------------|
| field pea (229) | 2.4 | 1.9 |
| lentil (229) | 3.3 | 2.5 |
| chickpea (229) | 3.4 | 2.3 |
| wheat (bran) | 0.4 | not reported |

(14), and in Australia, a similar intake has been reported by Baghurst et al. (13) These figures represent the total amount from all sources including fruit and vegetables.

Heat treatment or cooking of pulses increases hydrolysis; however, incomplete starch gelatinization and the formation of RS induced by high amylose starch results in lower digestibility (15) and may contribute to low glycemic responses in humans (6). As a food ingredient, RS has a lower calorific (8 kJ/g) value compared with fully digestible starch (15 kJ/g); however, it can be incorporated into a wide range of mainstream food products such as baked products without affecting the processing properties or the overall appearance and taste of the product (16). This may represent an opportunity to increase the consumption of pulse grains by fractionating pulses and incorporating these products into cereal-based products.

PHYTOSTEROLS

Epidemiological data indicates that the consumption of grains, including pulses, lowers the mortality rates from cardiovascular disease (17). Elevated levels of serum low-density lipoprotein (LDL) cholesterol is a major cause of cardiovascular disease, and studies have shown that every reduction of 1% in LDL cholesterol results in a 1% reduction in cardiovascular mortality (18). These and other studies have led to the promotion of lifestyle changes, which have resulted in a reduction in LDL cholesterol.

The consumption of pulse grains has been reported to lower serum cholesterol and increase the saturation levels of cholesterol in the bile. A dietary study conducted by Duane on humans over a seven week period showed that serum LDL cholesterol was significantly lower during the consumption of a diet consisting of beans, lentils, and field peas (19). The study showed that consumption of pulses lowers LDL cholesterol by

Figure 2. Common phytosterols.

Figure 3. Generic structure of flavone and isoflavone classes.

partially interrupting the entrohepatic circulation of the bile acids and increasing the cholesterol saturation by increasing the hepatic secretion of cholesterol. The study also concluded that other pulse components in the diet may also have contributed to the observed effect; in particular, saponins, which are hydrolyzed by intestinal bacteria to diosgenin, may have exerted a positive effect (20, 21). Several studies have demonstrated the efficacy of plant sterols and stanols in the reduction of blood cholesterol levels, and plant sterols are increasingly incorporated into foods for this purpose (22, 23).

Phytosterols are structural components of the plant-cell membranes. In pulses, they are present in small quantities, and the most common phytosterols are β -sitosterol (1), campersterol (2), and stigmasterol (3), **Figure 2** (24). These compounds are also abundant as sterol glucosides and esterified sterol glucosides, with β -sitosterol representing 83% of the glycolipids in defatted chickpea flour (25).

ISOFLAVONES

Flavones and isoflavones have been isolated from a wide variety of plants, though the isoflavones are largely reported from the Fabaceae/Leguminosae family. There has been enormous interest in their biological activity. Chemically, they are based on phenylchromen-4-one and have the general structures shown in **Figure 3**. The increased interest in the biological effects of these molecules can be demonstrated by the increase in published literature in this area. **Figure 4** graphs the number of references citing biological activity of flavones or isoflavones since 1940. The number of publications in the past few years (2000 on) is more than double that published in the entire preceding 60 years (1263 vs 619). Of these, 570 papers deal specifically with compounds from legumes.

In terms of specific compounds studied, genistein and daidzein are the most cited (275 and 189 references, respectively) with the related glycosides, genistin and daidzin (54 and 46 references, respectively) also the subject of considerable interest. Five of the most reported isoflavones are genistein (4), daidzein (5), coumestrol (6), formononetin (7), and biochanin A (8) (Figure 5). Genistin and daidzin are the seven-glucose derivatives of 4 and 5, respectively.

According to the USDA survey on isoflavone content, lentils do not contain significant amounts of these isoflavones (26).

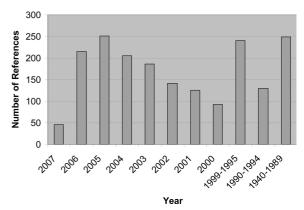


Figure 4. References reporting biological activity for flavones or isoflavones (search terms: Scifinder (all databases selected) using the phrase "activity of isoflavone or isoflavonol or isoflavanol" on April 23, 2007).

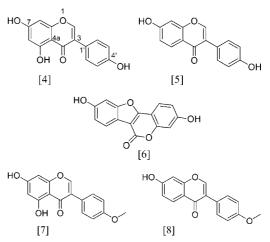


Figure 5. Important isoflavones.

Chickpeas contain 0.04 mg/100 g daidzein, 0.06 mg/100 g genistein, 0.14 mg/100 g formononetin, and approximately 1.7 mg/100 g biochanin A. Soybeans have significantly higher levels of daidzein (47 mg/100 g) and genistein (74 mg/100 g) but contain less formononetin and biochanin A compared to chickpeas, 0.03 mg/100 g and 0.07 mg/100 g, respectively. No figures are given for lupins.

Recently, there has been attention focused on a different class of isoflavones, the glyceollins, which have been reported from soy. These are biosynthetically related to pterocarpan and probably derive from the condensation of pterocarpan and a C5-terpene, **Figure 6** (27). The glyceollin types of isoflavone have not been reported from chickpeas, lupins, or lentils, despite the fact that chickpeas are capable of synthesizing the related, upstream metabolite daidzein.

Activities and Bioavailability. There are many biological activities associated with the isoflavones, including a reduction in osteoporosis and the prevention of cancer and cardiovascular disease, and they can be used for the treatment of symptoms of menopause. The potential health benefits of isoflavones for humans have been the subject of several reviews that have analyzed clinical, animal, and in vitro evidence for biological activity (28–41).

Since the early 1990s, a significant research effort has focused on the putative anticancer effects of isoflavones, in particular, the effect on breast cancer. Initial interest in this area was due to epidemiological observations of low breast cancer occurrence in Asian populations where the intake of soy and associated isoflavones is high. There has been sufficient research in this area that, in November 2005, a workshop was held to review

Figure 6. Biosynthetic relationship between isoflavones found in soy.

the literature and make research recommendations (32). The recent meta-analysis by Trock et al. (28) highlights the difficulties of comparing literature clinical studies. The authors studied 12 case-control and six cohort studies (with the number of subjects varying from 88 to 1459) but were forced to make several assumptions regarding the quantities of isoflavones ingested. Trock et al. conclude there is a small inverse correlation between soy intake and breast cancer but note that data limitations cannot rule out the possibility that this result is an artifact of the analysis. One of the recommendations of the conference was to encourage future studies to reduce the heterogeneity of soy exposure data in the literature and provide more detail regarding not only total soy food but also nutrient content, such as the levels of isoflavones (29).

The intake of isoflavones has been recommended for menopausal women to relieve symptoms of menopause (instead of hormonal replacement therapy, HRT) (42). A recent review by Cassidy et al. (34) concluded that, although additional studies are required, there was limited evidence for the ability of isoflavones to relieve the symptoms of menopause such as hot flashes. The HRT-like actions of the isoflavones are thought to be due to the estrogenic effects of the metabolites. Recently, the wisdom of the recommendation to increase isoflavones for their HRT effect has been challenged due to the potential increased risk of breast cancer in those using HRT, with some in vivo and vitro work suggesting genistein and daidzein may stimulate estrogen-dependent human breast tumor growth (43–45). The isoflavones display both estrogen agonist and antagonist activity (46). Again, the evidence here is confounding, with conflicting results in the literature, but the recent study by Wood et al. (46) suggests that, in part, this may be due to differences in endogenous estrogen levels in the reported studies. The authors found no estrogenic effect for isoflavones at low levels of estrogen, as would be the case in menopausal women, a different situation to that of the various rat models where there is no basal level of estrogen. Studies also show that the isoflavones bind preferentially to the β form of the estrogen receptor (ER β) rather than ER α (47). It is generally thought that the estrogen-associated risk of breast cancer is modulated through the $ER\alpha$ isoform. The ERα isoform promotes epithelial proliferation in the breast, while $ER\beta$ does not (48). Although genistein and daidzein have received the most attention in this area, there is also evidence that the glyceollins act through the ER pathways (49, 50).

Interestingly, there is also some evidence that the degree of processing may have an impact on the ability to stimulate tumorogenisis, with highly processed products more likely to be problematic (45, 51, 52). This conclusion would seem to correlate with the epidemiological evidence of a reduced risk of breast cancer in Asian populations, where the soy products are minimally processed. It has also been suggested that early and prolonged exposure to isoflavones, through a high-legume diet, is more beneficial than a later, higher consumption of isoflavone supplements (45).

Mechanistic studies suggest that the isoflavones may promote cancer cell death not only through $ER\beta$ but also more directly through the down-regulation of cell survival enzymes such as

NF-kappaB (53–56), the activation of apoptosis via ER stress pathways (m-calpain, GADD153, GRP78, and caspase-12) (57, 58), and mitochondrial apoptotic pathways (Mcl-1 down-regulation and Bad cleavage) (57). These observations suggest that the isoflavones have the potential to prevent other cancers as well, and there have been studies around prostate cancer (53, 59–67), colorectal cancer (68, 69), and head and neck cancer (54). The research includes in vitro, in vivo, and clinical studies, and as with breast cancer, the results of these studies are mixed, but indicative of some protective effect due to isoflavones.

One of the potential confounding factors in cohort studies is the possibility that studies of single nutrients and food may be inconsistent because they do not account for related foods or the potential synergistic interaction of food combinations and other factors that may effect bioavailability (including cooking). In an effort to address these, Velie et al. (70) undertook a large diet-based cohort study (40 559 postmenopausal women). They found three diet groups across the U.S.A.; the only diet with significant negative correlation with invasive breast cancer was the traditional southern diet, which correlated to high legume intake, low mayonnaise intake, and potentially cabbage intake. This "whole of diet" or "whole of food" approach may indeed be a very important consideration since, as discussed later, the legumes that contain these isoflavones also contain other metabolites (in particular, saponins and sugar derivatives) which also possess anticancer activity.

Pharmacokinetic studies indicate that the plasma levels of the isoflavones can reach biologically significant levels (low micromolar) (46, 71). The glycosides are hydrolyzed to produce the aglycones, which have a half-life in the plasma of 4–8 h (71). Interestingly, equol (11) a human metabolite of daidzein, which is also highly bioactive, is not found ubiquitously. In a recent study, this metabolite was found in only 30% of women studied. Equol is likely to be a product of action by intestinal microflora, and it has been suggested that it may be more bioactive than the parental isoflavone (36). This observation introduces yet another potential source of variability in clinical studies—since an individual's microflora may be highly specific, the products of gut bacteria will vary between subjects in a study, and this may have large contributing effects on the intersubject variability.

There is growing evidence that the isoflavones may have a role to play in the treatment of metabolic disorders. A metaanalysis of 38 different controlled clinical trials concluded that soy protein intake led to decreased serum concentrations of total cholesterol, LDL cholesterol, and triglycerides (35). Studies in rats have shown that chickpea consumption can normalize triacylglyceride levels in hypercholesterolemic rats (72). This area of research is in its infancy compared to the enormous focus that isoflavones and cancer have received; however, there have been several mechanistic studies which suggest potential efficiency. Some of this action is related to the compounds' ability to act in a similar way to estrogen. Genistein, daidzein, and biochanin A have been reported as estrogen-related receptor α (ERR α) agonists. The orphan ERRs comprising ERR α , ERR β , and ERR γ bind and regulate transcription via estrogen response elements but do not bind endogenous estrogens. ERR α is involved in energy homeostasis and so is a likely target for the treatment of metabolic disorders (73). PPAR α and PPAR γ (the peroxisome proliferator-activated receptors) active compounds are used to correct dyslipidemia and to restore glycaemia balance, respectively. Formononetin, biochanin A, genistein, and daidzein act as PPAR α and PPAR γ activators. Biochanin A and formononetin, in particular, are of interest in that they are active at low doses (1–4 μ mol/L) (74). These compounds are both present in greater amounts in chickpeas compared to soy beans, and for conditions such as type II diabetes, the intake of pulses such as chickpeas may be of greater benefit.

Even less well-studied is the potential protective effect of isoflavones against neurodegenerative diseases. Although much more experimental evidence is required to investigate this hypothesis, the initial reports are intriguing. Interest in this area again arose from epidemiological studies that suggested postmenopausal women using estrogen replacement therapy had a decreased risk of developing dementia (75, 76). Genistein, daidzein, and glycitein (12) were examined in a transgenic nematode model for their ability to alleviate β amyloid expression-induced paralysis. Only glycitein demonstrated significant activity, and this at a relatively high concentration (100 μ g/mL) (77). However, the ability of this isoflavone to reduce the formation of the β amyloid is nonetheless fascinating and surely warrants further investigation. More recently, an investigation of biochanin A suggested it may be protective against Parkinson's disease through its ability to protect dopaminergic neurons (78). These intriguing results suggest that pulses may have a role to play in healthy aging strategies, though clearly much additional research is required.

Cooking/Processing Effects. Although there have been several studies on pulses investigating the effect of cooking techniques on the removal of compounds such as phytate, oligosaccharides, and saponins (as discussed later), these studies have not, in general, addressed the stability of the isoflavones. There have been several studies on the isoflavone content of foods (26, 79-83) but few tracing their stability from the legume to the processed product. One of the more detailed reports in this area demonstrated that processing does have a significant effect on isoflavone content and indeed causes chemical modification of the isoflavones. The most common observation was the loss of the esterified malonate to form the glycoside of the isoflavones under any heat conditions (including baking and frying) (84). These authors also noted the almost complete absence of any isoflavones in the "low-fat" soy products. On the basis of the published literature, it seems the greater the degree of processing, the lower the amounts of isoflavones in the resulting product, but it remains unclear actually how much of the bioactives are lost during different processing methods.

Agricultural Studies. There is good evidence that farming practice can directly influence the levels of isoflavones in crops. The majority of this work has focused on soy, and it is likely that the results would transfer to pulses, though this hypothesis requires testing. One field study on soy beans demonstrated that irrigation enhances isoflavone content by as much as 2.5 fold (85). The application of potassium-rich fertilizer also results in an increase in the desirable bioactives (86). A 3-year breeding study demonstrated that levels of isoflavones are related to both environmental and genetic elements that would be susceptible to selection (83), and these genomic regions have been identified (87). This is an active area of research; for example, the U.S. Agricultural Research

Service has initiated research projects working to understand the elicitation mechanism of flavones to enhance content in plants (88).

The isoflavones of edible legumes are not well-characterized for their natural role in plants, but there is evidence that many are antimicrobial and may have a role in plant protection. For example, the antifungal activity of lupin isoflavones has been demonstrated (89). In a study on the soybean cultivar response to fungal attack, it was noted that of two saprophytic fungi (Mucor ramosissimus and Rhizopus sp.) only M. ramosissimus induced an accumulation of metabolites including isoflavones. In the strains resistant to the fungus, a greater number of isoflavones (including glyceollins I, II, and III; glycinol; glyceocarpin; genistein; isoformononetin; and N-acetyltyramine) were induced, and with the exception of genistein, the compounds were demonstrated to possess antifungal activity (90). The cell-wall glucan from another fungus (Phytophthora megasperma Drechs. f. sp. glycinea, Kuan and Erwin) is also an elicitor of protective isoflavones (including the glycoside conjugates) in the soybean (91). An investigation of the fungal elicititor from Diaporthe phaseolorum f. sp. Meridionalis, the causal agent of stem canker, suggested that the elicitation of isoflavones may be mediated by the nitric oxide synthase pathway (92).

Interestingly, there is some evidence that specific symbiotic interactions have evolved to take advantage of the isoflavones' chemistry. It has been demonstrated that the symbiotic relationship between the fungus *Rhizobium lupini* and *Lupinus albus* stimulates an increase in production of prenylated isoflavones in the root nodules (93). Theses prenylated isoflavones possess in vitro activity against a number of other *Rhizobium* species. Tahara et al. (94) have described the isolation and testing of several flavones, isoflavones (including the novel compound isolupalbigenin, 13), and chalcone metabolites from the yellow lupin and demonstrated that many of them possess antifungal activity against *Cladosporium herbarum*. Indeed, many novel isoflavones were described for the first time from lupins and reportedly possess antifungal activity (89, 95–103).

These studies suggest not only that isoflavones have potential benefits for both human and plant health but also that these traits are subject to manipulation through both farm management and breeding strategies.

SAPONINS

Saponins are secondary metabolites of mixed biosynthesis. They consist of a triterpene or steroid nucleus (the aglycone) with mono- or oligosaccharides attached to this core. Saponins have long been considered undesirable due to toxicity and their haemolytic activity. However, there is enormous structural diversity within this chemical class, and only a few are toxic (104). The most common saponins in legumes include the soyasaponins, which are classified into group A, B, and E saponins on the basis of the chemical structure of the aglycone. These have the general structure shown in **Figure 7**.

Soyasaponins do not have reported toxicity in monkeys, humans, rats, or chicks, although high levels do impart a bitter taste (104). This is not a universal trait of the structure class, and the potential for the use of sweet saponins has been the subject of recent literature (105, 106).

Saponins have been reported in many edible legumes, although the detailed structures were not always established. They have been found in lupins (107, 108), lentils (109, 110), and chickpeas (104, 110–113), as well as soy, various beans, and peas (104).

Activities and Bioavailability. The spectrum of biological activity of saponins is as broad as the structure class. The literature suggests that legumous saponins may possess anti-

Group B saponins R₁=H, R₂=OH Group E saponins R₂=O

Figure 7. Chemical structure of saponins.

cancer activity (104, 114-119) and be beneficial for hyperlipidemia (72, 104, 113). The adjuvant properties of certain saponins has also been utilized in vaccines for many years (120). The best studied are the soyasaponins both in terms of epidemiology and in vitro and in vivo systems.

Group A saponins R₁=OH, R₂=OH

Epidemiologic studies suggest that saponins may play a role in protection from cancer (104), and there are a number of hypothesized modes of action. Mechanistic experiments that give some insight into the potential mode of action of saponins have attracted recent research attention. Metastatic events are critical in cancer proliferation, and there is evidence that glycosylation is an important event in this process (121–126). Chang et al. (115) have recently demonstrated that soyasaponin I decreases the expression of α -2,3-linked sialic acid on the cell surface, which in turn suppresses the metastatic potential of melanoma cells. The observed anticancer activity may therefore in part be due to this observed sialyltransferase inhibition activity.

Additional mechanistic studies indicate that there is evidence for saponin regulation of the apoptosis pathway enzymes (AKT, Bcl, and ERK1/2), leading to programmed cell death of cancer cells (116, 127-129). Research on colon cancer cells suggests that it is the lipophilic saponin cores that may be responsible for the biological activity. The in vitro fermentations carried out by these authors also suggest that colonic microflora hydrolyze soyasaponins to the aglycones, potentially enhancing the activity of the soyasaponins (119). This proposed hydrolyzation process is supported by later in vivo and in vitro research that demonstrated that group B soyasaponins were not detected from urine or fecal samples but that the metabolite, soyasapogenol B, was detected in fecal samples. Hu et al. (118) showed that uptake by Caco-2 cells was limited, indicating poor intestinal absorption. Studies on saponins from other sources suggest that intestinal uptake is largely by diffusion mechanisms (130–132). There is some suggestion that microbial and hepatic modification (esterification with fatty acids) may enhance bioavailability (133), but saponins are generally thought to have low bioavailability. There is some evidence that certain materials enhance the absorption of saponins, for example, chitosan and sodium deoxycholate (131), and so further research could increase the understanding of additional dietary factors (within or external to the legume) that may enhance uptake. Bioavailability is also influenced by individual metabolism, food processing methods, and interaction with bile acids (104), further complicating research in this area.

The hyperlipidemic action of saponins has not been wellstudied, and the results can be conflicting (134), but some studies suggest that saponins may reduce cholesterol through the formation of an insoluble complex with cholesterol, thus preventing absorption in the intestine. Additionally, some saponins increase the excretion of bile acids—an indirect method of decreasing cholesterol (135).

Cooking/Processing Effects. Cooking and processing can have a significant effect on the levels of available saponins in legumes. Interestingly, the results are not necessarily the same for all legumes. Soaking and cooking studies on chickpeas and lentils suggest that 2–5% of saponin content can be lost from chickpeas during cooking, but a much larger 6–14% can be lost from lentils (110). The method of cooking has a significant effect on saponin loss, with autoclaving having a large effect (136). Some saponins are thermolabile and may interconvert or degrade (e.g., soyasaponin VI forms soyasaponin I with increased cooking times) (104, 137). In terms of human health, it is unclear what the biological significance of such interconversions may

Agricultural Studies. The role of saponins in the plant is not clear. It is suggested that they play a role in chemical defense. Studies of some lupin saponins show that they possess moderate antifungal activity (108), and it is possible that the bitter taste of some saponins, particularly the acetyl derivatives, may act as a deterrent to herbivores (104). The saponins of edible legumes are not well-characterized for their natural role in plants. Studies of other plant saponins suggest that many are antimicrobial and may have a role in the protection of plants from microbial infection, a suggestion which is supported by the observation that saponin-deficient strains are often less disease-resistant (138–148).

ALKALOIDS

In general, the majority of alkaloids from edible legumes have been reported from lupins. Lupins have a relatively short history of use as a grain crop, and it is only recently (the past 20-30 years) that cultivars have been developed with a reduced alkaloid content (149). These cultivars are often referred to as "sweet" lupins since the alkaloids often impart a bitter taste. This is not to say that lupins are the only edible legumes from which alkaloids have been reported. The alkaloid trigonelline (14) has been reported from peas (150), and it is possible that targeted studies of chickpeas and lentils would also reveal low levels of alkaloids as well.

In particular, it would be interesting to examine some of the less-cultivated landrace varieties of chickpeas and lentils to investigate alkaloid chemistry. Over centuries of cultivated use, alkaloids may well have been bred out of the now-accepted varieties due to toxicity, as well as to enhance palatability. However, such compounds may have biological activity of value in certain circumstances. For example, although toxic to some individuals, hydroxypyrimidine glucoside alkaloids, which are

the antimalarial principals of fava beans, have beneficial properties for human health (151). Lupins have produced a large range of interesting alkaloid chemicals, and both edible and related lupin species continue to be the subject of literature reporting novel chemistry (152–158).

Activities and Bioavailability. As a broad chemical class, alkaloids demonstrate a diversity of biological activity. One of the most intriguing recent reports discusses the enhancement of insulin secretion by lupin quinolizidine alkaloids (159). The authors note that this increased secretion only occurs in the presence of relatively high glucose levels and so may be of relevance for managing type II diabetes.

Cooking/Processing Effects. Although there may be trace amounts of alkaloids present in legumes cultivated for human consumption, preparation often removes these chemicals. The alkaloid concentration may be enhanced in the seed (158); however, in general, soaking removes a significant proportion of alkaloids from the lupin seed (160).

Agricultural Studies. Although the alkaloid content of the legume seeds may be undesirable for human or animal consumption, there is evidence that the alkaloid content is protective for the plant. For instance, it has been noted that "sweet" lupins are susceptible to a large number of insect herbivores to which the wild-type plants are resistant (161). Alkaloids are not the only bitter principals that may be responsible for such activity. As has already been discussed, saponins may affect both palatability and disease resistance.

BIOACTIVE CARBOHYDRATES

Activities and Bioavailability. For many years, the focus on legume sugars and oligosaccharides has centered on the minimization of the raffinose sugars. The link between the raffinose sugars and flatulence has long been understood (162, 163) and can be an important impediment to the increased consumption of legumes (164). Flatulence and related disorders, including bloating and diarrhea, are due to a lack of α-galactosidases in the upper gut. The oligosaccharides then enter the lower gut where they are metabolized by the bacteria, resulting in an accumulation of carbon dioxide and hydrogen (165). The raffinose sugars include raffinose (15), stachyose (16), and verbascose (17) (Figure 8). There are appreciable levels of these oligosaccharides in chickpeas, lentils, lupins, and field peas (1–12% of dry weight) with considerable variation between the different pulses (166). Studies on the quantities of these sugars in chickpeas show a distinct difference between desi and kabuli types. On average, the desi-type chickpeas are 16% higher in the three oligosaccharides, but with very low levels of verbascose found in either. Interestingly, the kabuli type also has 47% more sucrose than the desi chickpeas (165), a likely indication that these are factors subject to breeding (since the kabuli type has been bred for thousands of years, primarily for human consumption).

Oligosaccharides have also been demonstrated to be of potential value for immune health (167–170). Evidence for the mode of action is scant, but there are tantalizing hints that such immune modulation may occur through the enhancement of the innate immune system (in particular, through the Tol-like receptors) (171). Evidence for immunomodulating oligosaccharides or sugar components for gut health is currently focused more on milk sugars (172–176) than those from plants. However, it has been noted that the oligofructose metabolites from plants may play an important role in these functions, and additional research in this area is warranted (177, 178).

Phytic acid, or inositol hexaphosphate, has also been identified as an antinutritional, this compound acting through its ability

Figure 8. Raffinose family of oligosaccharides.

to inhibit mineral uptake. The uptake of zinc, calcium, and iron has been studied, and though the findings are mixed, there is evidence that zinc uptake, in particular, may be inhibited by phytic acid (179–186).

Pulse grains are a dietary source of minerals, although their bioavailability is considered lower because of the concentration of phytates (187). Phytic acid (myo-inositol-(1,2,3,4,5,6) hexakisphosphate; IP6) is the major source of phosphorous in pulses (188), although the concentration is less than 2%; for chickpeas, phytic acid constituents make up 52% of the total available phosphorous (189).

Phytic acid has the ability to chelate multivalent ions, those of alkaline metals being soluble in water, while divalent metals such as zinc, calcium, and iron salts are insoluble. This limits the bioavailability of minerals (190).

During food processing and digestion, phytase dephosphorylates phytic acid [IP6] to *myo*-inositol pentaphosphate [IP5], *myo*-inositol tetraphosphate [IP4], *myo*-inositol triphosphate [IP3], *myo*-inositol diphosphate [IP2], and *myo*-inositol monophosphate [IP1]; however, only IP5 and IP6 have a negative effect on the bioavailabity of divalent metal ions (191).

Of the various forms of phytates, IP6 is present in the greatest quantity and the most stable during cooking, followed by IP5, IP4, and IP3 (**Table 2**).

The ability to chelate with metal ions has associated IP6 with antinutritional properties, although the presence of IP5 and -6 in virtually all mammalian cells (192) contradicts this view (193), and reports indicate that there may be some protective effects such as decreasing the risk of iron-mediated colon cancer

Table 2. Inositol Phosphate Content of Raw and Cooked Pulses^a

| | | IP3 % | IP4 % | IP5 % | IP6 % |
|---------------------------|--------|-------|-------|-------|-------|
| chickpea (K) ^b | raw | nd | nd | 0.10 | 0.40 |
| | cooked | nd | 0.03 | 0.12 | 0.34 |
| chickpea (K) ^c | raw | 0.03 | nd | 0.03 | 0.38 |
| chickpea (D) ^c | raw | 0.03 | nd | 0.03 | 0.38 |
| field pea | raw | 0.00 | 0.01 | 0.08 | 0.43 |
| · | cooked | 0.01 | 0.02 | 0.10 | 0.33 |
| lentil | raw | 0.02 | 0.01 | 0.08 | 0.55 |
| | cooked | 0.05 | 0.05 | 0.21 | 0.47 |
| lentil ^c | raw | 0.02 | 0.02 | 0.05 | 0.24 |

^a Expressed as a percentage. ^b Reported values multiplied by 420, 500, 580, or 660 to convert from millimoles to milligrams per kilogram for IP3, IP4, IP5, or IP6, respectively (230). ^c Adapted from ref 231; K, Kabuli; D, Desi; nd, not detected.

and lowering serum cholesterol and triglycerides in experimental animals (194). Phytic acid has also been shown to have a positive effect due to its antioxidant (195) and anticarcinogenic effects (190, 196).

IP6 was demonstrated to be effective in controlling cancer in experimental mammary tumours (197) and in human prostate carcinoma cells (198). Using rats, Shamsuddin demonstrated that IP6 is quickly absorbed through the stomach and upper intestine and is distributed as inositol and IP1, and in in vitro studies, IP6 is taken up by malignant cells undergoing invariable dephosphorylation to IP1–5 and inositol (199).

Recently, focus has turned to the potentially beneficial effects of some of these antinutritionals. Phytic acid, in particular, has been the focus of research into its anticancer and hypocholesterolaemic action (135, 200–205). Phytic acid has been studied in relation to prostate cancer (201, 206, 207), breast cancer (204), colon cancer (208-211), and leukaemia (212-214). The mechanism of the anticancer activity is still not well-understood, though cell arrest at the G1 phase is indicated, potentially acting through the cyclin-dependent kinase pathway (206, 207). There is also some indication that phytic acid affects the immune response and that immunomodulating activity may be, in part, responsible for anticancer activity through the activation of natural killer cells (210). Regardless of the precise mode of action, there is sufficient excitement around the anticancer activity of phytic acid that some researchers are calling for extensive clinical trials of the phytochemical (200, 205).

Cooking/Processing Effects. As mentioned earlier, the removal of oligosaccharides, in particular, of the raffinose family, and undesirable sugars such as phytic acid has been studied for some time. Many of the undesirable oligosaccharides can be dramatically reduced by soaking or cooking (166, 215-218). Treatment with α -galactosidases (219), germination (220-223), and even irradiation (224) have also been found to be effective methods to reduce these constituents.

Agricultural Studies. The biological functions of these sugars are not certain, but it seems likely that phytic acid acts as a store of phosphorous within the seed (225). In addition, there is evidence that some of the raffinose sugars are protective against cell damage under dehydration conditions (226, 227). Genetic studies suggest that these metabolite levels are subject to manipulation (225, 228).

Conclusion. The overall benefits of pulses have historically been associated with their macronutrient composition, such as protein and starch, although there is now sufficient evidence to suggest that non-nutritional bioactive compounds play a significant role. Despite a large volume of literature, there is still the need for additional work around the substantiation of the health benefits of different pulse metabolites and the potential synergistic effects between the different classes of bioactives.

There is good evidence that levels of these bioactives in the plant are subject to manipulation. There appears to be significant natural variation both between and within the different species of pulses and the opportunity to enhance the concentrations through plant breeding or the application of agronomic practices.

LITERATURE CITED

- Tharanathan, R. N.; Mahadevamma, S. Grain Legumes- a boon to human nutrition. *Trends Food Sci. Technol.* 2003, 14, 507– 518
- (2) Peterson, D. S.; Sipsa, S.; Mackintosh, J. B. *The Chemical Composition and Nutritive Value of Australian Pulses*; Grains Research and Development Corporation: Kingston ACT, Australia. 1997.
- (3) Guillon, F.; Champ, M. M. Carbohydrate fractions of legumes: uses in human nutrition and potential for health. *Br. J. Nutr.* **2002**, *88* (Suppl. 3), S293–306.
- (4) Panozzo, J.; Materne, M. Developing Quality Pulses for a Sustainable Environment, Population and Community. *Personal Communication*, 2007.
- (5) Cassidy, A.; Bingham, S. A.; Cummings, J. H. Starch intake and colorectal cancer risk: an international comparison. *Br. J. Cancer* 1994, 69, 937–942.
- (6) Englyst, H. N.; Kingman, S. M.; Cummings, J. H. Classification and measurement of nutritionally important starch fractions. *Eur. J. Clin. Nutr.* 1992, 46 (Suppl. 2), S33–50.
- (7) Christl, S. U.; Mursgatroyd, P. R.; Gibson, G. R.; Cummings, J. H. Production, metabolism, and excretion of hydrogen in the large intestine. *Gastroenterology* 1992, 102, 1269–1277.
- (8) Topping, D. L.; Fukushima, M.; Bird, A. R. Resistant starch as a prebiotic and symbiotic: state of art. *Proc. Nutr. Soc.* 2003, 62, 171–176.
- (9) Andoh, A.; Tsujikawa, T.; Fujiyama, Y. Role of dietary fiber and short-chain fatty acids in the colon. *Curr. Pharm. Des.* 2003, 9, 347–358.
- (10) Topping, D. L.; Clifton, P. M. Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. *Physiol. Rev.* 2001, 81, 1031–1064.
- (11) Cummings, J. H.; Beatty, E. R.; Kingman, S. M.; Bingham, S. A.; Englyst, H. N. Digestion and physiological properties of resistant starch in the human large bowel. *Br. J. Nutr.* **1996**, *75*, 733–747
- (12) McCleary, B. V.; Rossiter, P. Measurement of novel dietary fibers. J. AOAC Int. 2004, 87, 707–717.
- (13) Baghurst, K.; Baghurst, P. A.; J, R. S. Dietary fibre, non-starch polysaccharide and resistant starch intakes in Australia. In *CRC Handbook of Dietary Fibre in Fibre in Human Health*; Spiller, G. A., Ed.; CRC Press LLc: Boca Raton, FL, 2001; pp583–591.
- (14) Dyssler, P.; Hoffmann, D. In Estimation of resistant starch intake in Europe; Asp, N.-G., Amelsvoort, J. M. M. v., Hautvat, J. G. A. J., Eds.; EURESTA: Wageningen, The Netherlands, 1994; pp 84–86.
- (15) Hoover, R.; Zhou, Y. In-vitro and in-vivo hydrolysis of legume starches by alpha amylase and resistant starch formation in legumes- a review. *Carbohydr. Polym.* 2003, 54, 401–417.
- (16) Sajilata, M. G.; Singhal, R. S.; Kulkarni, P. Resistant starch A review. Compr. Rev. Food Sci. Food Saf. 2006, 5, 1–17.
- (17) Kushi, L. H.; Meyer, K. A.; Jacobs, D. R., Jr. Cereals, legumes, and chronic disease risk reduction: evidence from epidemiologic studies. Am. J. Clin. Nutr. 1999, 70, 451S–458S.
- (18) Shepherd, J.; Codde, S. M.; Ford, I.; Isles, C. G.; Lorimer, A. R.; McFarlane, P. W.; Mckillop, J. H.; Packhard, C. J. Prevention of coronary heart disease with pravastatin in men with hypercholesterolemia. N. Engl. J. Med. 1995, 333, 1301–1307.
- (19) Duane, W. C. Effects of legume consumption on serum cholesterol, biliary lipids, and sterol metabolism in humans. *J. Lipid Res.* 1997, 38, 1120–1128.
- (20) Fenwick, D. E.; Oakenfull, D. Saponin content of food plants and some prepared foods. J. Sci. Food Agric. 1983, 34, 186– 191.

- (21) Thewles, A.; Parslow, R. A.; Coleman, R. Effect of diosgenin on biliary cholesterol transport in the rat. *Biochem. J.* 1993, 291 (Pt 3), 793–798.
- (22) Thompson, G. R.; Grundy, S. M. History and development of plant sterol and stanol esters for cholesterol-lowering purposes. Am. J. Cardiol. 2005, 96, 3D–9D.
- (23) Gylling, H.; Miettinen, T. A. The effect of plant stanol- and sterol-enriched foods on lipid metabolism, serum lipids and coronary heart disease. Ann. Clin. Biochem. 2005, 42, 254–263.
- (24) Benveniste, P. Sterol Biosynthesis. Annu. Rev. Plant Physiol. 1986, 37, 275–308.
- (25) Sanchez-Voique, R.; Clemente, A.; Vioque, J.; Bautista, J.; Millan, F. Polar lipids of defatted chickpea (Cicer arietinum L.) flour and protein isolates. *Food Chem.* 1998, 63, 357–361.
- (26) USDA-Iowa State University Database on the Isoflavone Content of Foods, Release 1.3. In Agricultural Research Service; U.S. Department of Agriculture: Washington, DC, 2002.
- (27) Burow, M. E.; Boue, S. M.; Collins-Burow, B. M.; Melnik, L. I.; Duong, B. N.; Carter-Wientjes, C. H.; Li, S.; Wiese, T. E.; Cleveland, T. E.; McLachlan, J. A. Phytochemical glyceollins, isolated from soy, mediate antihormonal effects through estrogen receptor alpha and beta. *J. Clin. Endocrinol. Metab.* 2001, 86, 1750–1758.
- (28) Trock, B. J.; Hilakivi-Clarke, L.; Clarke, R. Meta-analysis of soy intake and breast cancer risk. J. Natl. Cancer Inst. 2006, 98, 459–471.
- (29) Martinez, M. E.; Thomson, C. A.; Smith-Warner, S. A. Soy and breast cancer: the controversy continues. *J. Natl. Cancer Inst.* 2006, 98, 430–431.
- (30) Yan, L.; Spitznagel, E. L. Meta-analysis of soy food and risk of prostate cancer in men. *Int. J. Cancer* 2005, 117, 667–669.
- (31) Lephart, E. D.; Setchell, K. D.; Handa, R. J.; Lund, T. D. Behavioral effects of endocrine-disrupting substances: phytoestrogens. *ILAR J.* 2004, 45, 443–454.
- (32) Messina, M.; McCaskill-Stevens, W.; Lampe, J. W. Addressing the soy and breast cancer relationship: review, commentary, and workshop proceedings. J. Natl. Cancer Inst. 2006, 98, 1275– 1284.
- (33) Cooke, G. M. A review of the animal models used to investigate the health benefits of soy isoflavones. *J. AOAC Int.* **2006**, *89*, 1215–1227.
- (34) Cassidy, A.; Albertazzi, P.; Lise Nielsen, I.; Hall, W.; Williamson, G.; Tetens, I.; Atkins, S.; Cross, H.; Manios, Y.; Wolk, A.; Steiner, C.; Branca, F. Critical review of health effects of soyabean phyto-oestrogens in post-menopausal women. *Proc. Nutr. Soc.* 2006, 65, 76–92.
- (35) Ricketts, M. L.; Moore, D. D.; Banz, W. J.; Mezei, O.; Shay, N. F. Molecular mechanisms of action of the soy isoflavones includes activation of promiscuous nuclear receptors. A review. *J. Nutr. Biochem.* 2005, 16, 321–330.
- (36) Atkinson, C.; Frankenfeld, C. L.; Lampe, J. W. Gut bacterial metabolism of the soy isoflavone daidzein: exploring the relevance to human health. *Exp. Biol. Med. (Maywood, NJ, U.S.)* 2005, 230, 155–170.
- (37) Marin, F. R.; Perez-Alvarez, J. A.; Soler-Rivas, C. Isoflavones as functional food components. *Stud. Nat. Prod. Chem.* 2005, 32, 1177–1207.
- (38) Polkowski, K.; Mazurek, A. P. Biological properties of genistein. A review of in vitro and in vivo data. *Acta Pol. Pharm.* 2000, 57, 135–155.
- (39) Williamson, G.; Manach, C. Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. Am. J. Clin. Nutr. 2005, 81, 243S–255S.
- (40) Manach, C.; Williamson, G.; Morand, C.; Scalbert, A.; Remesy, C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am. J. Clin. Nutr.* 2005, 81, 230S–242S.
- (41) Moon, Y. J.; Wang, X.; Morris, M. E. Dietary flavonoids: effects on xenobiotic and carcinogen metabolism. *Toxicol. in Vitro* 2006, 20, 187–210.
- (42) Beck, V.; Rohr, U.; Jungbauer, A. Phytoestrogens derived from

- red clover: an alternative to estrogen replacement therapy. *J. Steroid Biochem. Mol. Biol.* **2005**, *94*, 499–518.
- (43) Ju, Y. H.; Fultz, J.; Allred, K. F.; Doerge, D. R.; Helferich, W. G. Effects of dietary daidzein and its metabolite, equol, at physiological concentrations on the growth of estrogen-dependent human breast cancer (MCF-7) tumors implanted in ovariectomized athymic mice. *Carcinogenesis* 2006, 27, 856–863.
- (44) Ju, Y. H.; Allred, C. D.; Allred, K. F.; Karko, K. L.; Doerge, D. R.; Helferich, W. G. Physiological concentrations of dietary genistein dose-dependently stimulate growth of estrogen-dependent human breast cancer (MCF-7) tumors implanted in athymic nude mice. J. Nutr. 2001, 131, 2957–2962.
- (45) Allred, C. D.; Allred, K. F.; Ju, Y. H.; Virant, S. M.; Helferich, W. G. Soy diets containing varying amounts of genistein stimulate growth of estrogen-dependent (MCF-7) tumors in a dose-dependent manner. *Cancer Res.* 2001, 61, 5045–5050.
- (46) Wood, C. E.; Register, T. C.; Franke, A. A.; Anthony, M. S.; Cline, J. M. Dietary soy isoflavones inhibit estrogen effects in the postmenopausal breast. *Cancer Res.* 2006, 66, 1241–1249.
- (47) Kostelac, D.; Rechkemmer, G.; Briviba, K. Phytoestrogens modulate binding response of estrogen receptors alpha and beta to the estrogen response element. J. Agric. Food Chem. 2003, 51, 7632–7635.
- (48) McCarty, M. F. Isoflavones made simple genistein's agonist activity for the beta-type estrogen receptor mediates their health benefits. *Med. Hypotheses* 2006, 66, 1093–1114.
- (49) Salvo, V. A.; Boue, S. M.; Fonseca, J. P.; Elliott, S.; Corbitt, C.; Collins-Burow, B. M.; Curiel, T. J.; Srivastav, S. K.; Shih, B. Y.; Carter-Wientjes, C.; Wood, C. E.; Erhardt, P. W.; Beckman, B. S.; McLachlan, J. A.; Cleveland, T. E.; Burow, M. E. Antiestrogenic glyceollins suppress human breast and ovarian carcinoma tumorigenesis. *Clin. Cancer Res.* 2006, 12, 7159–7164.
- (50) Wood, C. E.; Clarkson, T. B.; Appt, S. E.; Franke, A. A.; Boue, S. M.; Burow, M. E.; McCoy, T.; Cline, J. M. Effects of soybean glyceollins and estradiol in postmenopausal female monkeys. *Nutr. Cancer* 2006, 56, 74–81.
- (51) Allred, C. D.; Allred, K. F.; Ju, Y. H.; Goeppinger, T. S.; Doerge, D. R.; Helferich, W. G. Soy processing influences growth of estrogen-dependent breast cancer tumors. *Carcinogenesis* 2004, 25, 1649–1657.
- (52) Allred, C. D.; Twaddle, N. C.; Allred, K. F.; Goeppinger, T. S.; Churchwell, M. I.; Ju, Y. H.; Helferich, W. G.; Doerge, D. R. Soy processing affects metabolism and disposition of dietary isoflavones in ovariectomized BALB/c mice. *J. Agric. Food Chem.* 2005, 53, 8542–8550.
- (53) Davis, J. N.; Kucuk, O.; Sarkar, F. H. Genistein inhibits NFkappa B activation in prostate cancer cells. *Nutr. Cancer* 1999, 35, 167–174.
- (54) Alhasan, S. A.; Aranha, O.; Sarkar, F. H. Genistein elicits pleiotropic molecular effects on head and neck cancer cells. *Clin. Cancer Res.* 2001, 7, 4174–4181.
- (55) Sarkar, F. H.; Adsule, S.; Padhye, S.; Kulkarni, S.; Li, Y. The role of genistein and synthetic derivatives of isoflavone in cancer prevention and therapy. *Mini Rev. Med. Chem.* 2006, 6, 401–407.
- (56) Singh, A. V.; Franke, A. A.; Blackburn, G. L.; Zhou, J. R. Soy phytochemicals prevent orthotopic growth and metastasis of bladder cancer in mice by alterations of cancer cell proliferation and apoptosis and tumor angiogenesis. *Cancer Res.* 2006, 66, 1851–1858.
- (57) Yeh, T. C.; Chiang, P. C.; Li, T. K.; Hsu, J. L.; Lin, C. J.; Wang, S. W.; Peng, C. Y.; Guh, J. H. Genistein induces apoptosis in human hepatocellular carcinomas via interaction of endoplasmic reticulum stress and mitochondrial insult. *Biochem. Pharmacol.* 2007, 73, 782–792.
- (58) Sergeev, I. N. Genistein induces Ca2+ -mediated, calpain/ caspase-12-dependent apoptosis in breast cancer cells. *Biochem. Biophys. Res. Commun.* 2004, 321, 462–467.

- (59) Raffoul, J. J.; Banerjee, S.; Singh-Gupta, V.; Knoll, Z. E.; Fite, A.; Zhang, H.; Abrams, J.; Sarkar, F. H.; Hillman, G. G. Down-regulation of apurinic/apyrimidinic endonuclease 1/redox factor-1 expression by soy isoflavones enhances prostate cancer radio-therapy in vitro and in vivo. *Cancer Res.* 2007, 67, 2141–2149.
- (60) Messina, M.; Kucuk, O.; Lampe, J. W. An overview of the health effects of isoflavones with an emphasis on prostate cancer risk and prostate-specific antigen levels. J. AOAC Int. 2006, 89, 1121– 1134.
- (61) Atherton, K. M.; Mutch, E.; Ford, D. Metabolism of the soyabean isoflavone daidzein by CYP1A2 and the extra-hepatic CYPs 1A1 and 1B1 affects biological activity. *Biochem. Pharmacol.* 2006, 72, 624–631.
- (62) Maskarinec, G.; Morimoto, Y.; Hebshi, S.; Sharma, S.; Franke, A. A.; Stanczyk, F. Z. Serum prostate-specific antigen but not testosterone levels decrease in a randomized soy intervention among men. Eur. J. Clin. Nutr. 2006, 60, 1423–1429.
- (63) Raschke, M.; Rowland, I. R.; Magee, P. J.; Pool-Zobel, B. L. Genistein protects prostate cells against hydrogen peroxideinduced DNA damage and induces expression of genes involved in the defence against oxidative stress. *Carcinogenesis* 2006, 27, 2322–2330.
- (64) Handayani, R.; Rice, L.; Cui, Y.; Medrano, T. A.; Samedi, V. G.; Baker, H. V.; Szabo, N. J.; Shiverick, K. T. Soy isoflavones alter expression of genes associated with cancer progression, including interleukin-8, in androgen-independent PC-3 human prostate cancer cells. J. Nutr. 2006, 136, 75–82.
- (65) Hedlund, T. E.; Maroni, P. D.; Ferucci, P. G.; Dayton, R.; Barnes, S.; Jones, K.; Moore, R.; Ogden, L. G.; Wahala, K.; Sackett, H. M.; Gray, K. J. Long-term dietary habits affect soy isoflavone metabolism and accumulation in prostatic fluid in caucasian men. *J. Nutr.* 2005, 135, 1400–1406.
- (66) Mentor-Marcel, R.; Lamartiniere, C. A.; Eltoum, I. A.; Greenberg, N. M.; Elgavish, A. Dietary genistein improves survival and reduces expression of osteopontin in the prostate of transgenic mice with prostatic adenocarcinoma (TRAMP). *J. Nutr.* 2005, *135*, 989–995.
- (67) Suzuki, K.; Koike, H.; Matsui, H.; Ono, Y.; Hasumi, M.; Nakazato, H.; Okugi, H.; Sekine, Y.; Oki, K.; Ito, K.; Yamamoto, T.; Fukabori, Y.; Kurokawa, K.; Yamanaka, H. Genistein, a soy isoflavone, induces glutathione peroxidase in the human prostate cancer cell lines LNCaP and PC-3. *Int. J. Cancer* 2002, 99, 846– 852.
- (68) Cotterchio, M.; Boucher, B. A.; Manno, M.; Gallinger, S.; Okey, A.; Harper, P. Dietary phytoestrogen intake is associated with reduced colorectal cancer risk. J. Nutr. 2006, 136, 3046–3053.
- (69) Adams, K. F.; Lampe, P. D.; Newton, K. M.; Ylvisaker, J. T.; Feld, A.; Myerson, D.; Emerson, S. S.; White, E.; Potter, J. D.; Lampe, J. W. Soy protein containing isoflavones does not decrease colorectal epithelial cell proliferation in a randomized controlled trial. Am. J. Clin. Nutr. 2005, 82, 620–626.
- (70) Velie, E. M.; Schairer, C.; Flood, A.; He, J. P.; Khattree, R.; Schatzkin, A. Empirically derived dietary patterns and risk of postmenopausal breast cancer in a large prospective cohort study. Am. J. Clin. Nutr. 2005, 82, 1308–1319.
- (71) Setchell, K. D.; Brown, N. M.; Desai, P. B.; Zimmer-Nechimias, L.; Wolfe, B.; Jakate, A. S.; Creutzinger, V.; Heubi, J. E. Bioavailability, disposition, and dose-response effects of soy isoflavones when consumed by healthy women at physiologically typical dietary intakes. J. Nutr. 2003, 133, 1027–1035.
- (72) Zulet, M. A.; Macarulla, M. T.; Portillo, M. P.; Noel-Suberville, C.; Higueret, P.; Martinez, J. A. Lipid and glucose utilization in hypercholesterolemic rats fed a diet containing heated chickpea (Cicer aretinum L.): a potential functional food. *Int. J. Vitam. Nutr. Res.* 1999, 69, 403–411.
- (73) Ariazi, E. A.; Jordan, V. C. Estrogen-related receptors as emerging targets in cancer and metabolic disorders. *Curr. Top. Med. Chem.* 2006, 6, 203–215.
- (74) Shen, P.; Liu, M. H.; Ng, T. Y.; Chan, Y. H.; Yong, E. L. Differential effects of isoflavones, from Astragalus membranaceus and Pueraria thomsonii, on the activation of PPARalpha,

- PPARgamma, and adipocyte differentiation in vitro. *J. Nutr.* **2006**, *136*, 899–905.
- (75) Zec, R. F.; Trivedi, M. A. The effects of estrogen replacement therapy on neuropsychological functioning in postmenopausal women with and without dementia: a critical and theoretical review. *Neuropsychol. Rev.* 2002, 12, 65–109.
- (76) Yoon, B. K.; Kim, D. K.; Kang, Y.; Kim, J. W.; Shin, M. H.; Na, D. L. Hormone replacement therapy in postmenopausal women with Alzheimer's disease: a randomized, prospective study. *Fertil. Steril.* 2003, 79, 274–280.
- (77) Gutierrez-Zepeda, A.; Santell, R.; Wu, Z.; Brown, M.; Wu, Y.; Khan, I.; Link, C. D.; Zhao, B.; Luo, Y. Soy isoflavone glycitein protects against beta amyloid-induced toxicity and oxidative stress in transgenic Caenorhabditis elegans. *BMC Neurosci.* 2005, 6, 54.
- (78) Chen, H. Q.; Jin, Z. Y.; Li, G. H. Biochanin A protects dopaminergic neurons against lipopolysaccharide-induced damage through inhibition of microglia activation and proinflammatory factors generation. *Neurosci. Lett.* 2007, 417, 112–117.
- (79) Penalvo, J. L.; Heinonen, S. M.; Nurmi, T.; Deyama, T.; Nishibe, S.; Adlercreutz, H. Plant lignans in soy-based health supplements. J. Agric. Food Chem. 2004, 52, 4133–4138.
- (80) Mulligan, A. A.; Welch, A. A.; McTaggart, A. A.; Bhaniani, A.; Bingham, S. A. Intakes and sources of soya foods and isoflavones in a UK population cohort study (EPIC-Norfolk). Eur. J. Clin. Nutr. 2007, 61, 248–254.
- (81) Ritchie, M. R.; Cummings, J. H.; Morton, M. S.; Michael Steel, C.; Bolton-Smith, C.; Riches, A. C. A newly constructed and validated isoflavone database for the assessment of total genistein and daidzein intake. *Br. J. Nutr.* 2006, 95, 204–213.
- (82) Clarke, D. B.; Barnes, K. A.; Lloyd, A. S. Determination of unusual soya and non-soya phytoestrogen sources in beer, fish products and other foods. *Food Addit. Contam.* 2004, 21, 949– 962.
- (83) Mebrahtu, T.; Mohamed, A.; Wang, C. Y.; Andebrhan, T. Analysis of isoflavone contents in vegetable soybeans. *Plant Foods Hum. Nutr.* 2004, 59, 55–61.
- (84) Coward, L.; Smith, M.; Kirk, M.; Barnes, S. Chemical modification of isoflavones in soyfoods during cooking and processing. Am. J. Clin. Nutr. 1998, 68, 1486S–1491S.
- (85) Bennett, J. O.; Yu, O.; Heatherly, L. G.; Krishnan, H. B. Accumulation of genistein and daidzein, soybean isoflavones implicated in promoting human health, is significantly elevated by irrigation. J. Agric. Food Chem. 2004, 52, 7574–7579.
- (86) Vyn, T. J.; Yin, X.; Bruulsema, T. W.; Jackson, C. J.; Rajcan, I.; Brouder, S. M. Potassium fertilization effects on isoflavone concentrations in soybean [Glycine max (L.) Merr.]. *J. Agric. Food Chem.* 2002, 50, 3501–3506.
- (87) Kassem, M. A.; Meksem, K.; Iqbal, M. J.; Njiti, V. N.; Banz, W. J.; Winters, T. A.; Wood, A.; Lightfoot, D. A. Definition of Soybean Genomic Regions That Control Seed Phytoestrogen Amounts. J. Biomed. Biotechnol. 2004, 2004, 52–60.
- (88) Agriculture, U. D. Research Project: Determine Isoflavonoid Induction in Legumes and Their Phytoestrogenic Effects in Animal Systems. http://www.ars.usda.gov/research/projects/ projects.htm?ACCN_NO=405260 (Accessed April 20, 2007).
- (89) Tahara, S.; Ingham, J. L.; Nakahara, S.; Mizutani, J.; Harborne, J. B. Antifungal isoflavones in lupines. Part 2. Fungitoxic dihydrofuranoisoflavones and related compounds in white lupine, Lupinus albus. *Phytochemistry* 1984, 23, 1889–1900.
- (90) Garcez, W. S.; Martins, D.; Garcez, F. R.; Marques, M. R.; Pereira, A. A.; Oliveira, L. A.; Rondon, J. N.; Peruca, A. D. Effect of spores of saprophytic fungi on phytoalexin accumulation in seeds of frog-eye leaf spot and stem canker-resistant and -susceptible soybean (Glycine max L.) cultivars. *J. Agric. Food Chem.* 2000, 48, 3662–3665.
- (91) Graham, M. Y.; Graham, T. L. Rapid Accumulation of Anionic Peroxidases and Phenolic Polymers in Soybean Cotyledon Tissues following Treatment with Phytophthora megasperma f. sp. Glycinea Wall Glucan. *Plant Physiol.* 1991, 97, 1445–1455.

- (92) Modolo, L. V.; Cunha, F. Q.; Braga, M. R.; Salgado, I. Nitric oxide synthase-mediated phytoalexin accumulation in soybean cotyledons in response to the Diaporthe phaseolorum f. sp. meridionalis elicitor. *Plant Physiol.* 2002, 130, 1288–1297.
- (93) Gagnon, H.; Grandmaison, J.; Ibrahim, R. K. Phytochemical and immunocytochemical evidence for the accumulation of 2'hydroxylupalbigenin in lupin nodules and bacteroids. *Mol. Plant-Microbe Interact.* 1995, 8, 131–137.
- (94) Tahara, S.; Katagiri, Y.; Ingham, J. L.; Mizutani, J. Prenylated flavonoids in the roots of yellow lupin. *Phytochemistry* 1994, 36, 1261–1271.
- (95) Tahara, S.; Moriyama, M.; Orihara, S.; Ingham, J. L.; Kawabata, J.; Mizutani, J. Naturally occurring coumaranochroman-4-ones: a new class of isoflavonoids from lupines and Jamaican dogwood. Z. Naturforsch., C: J. Biosci. 1991, 46, 331–340.
- (96) Tahara, S.; Shibaki, S.; Ingham, J. L.; Mizutani, J. Further isoflavonoids from white lupine roots. Z. Naturforsch., C: J. Biosci. 1990, 45, 147–153.
- (97) Tahara, S.; Orihara, S.; Ingham, J. L.; Mizutani, J. Seventeen isoflavonoids from Lupinus albus roots. *Phytochemistry* 1989, 28, 901–911.
- (98) Lane, G. A.; Newman, R. H. Isoflavones from Lupinus angustifolius root. *Phytochemistry* 1986, 26, 295–300.
- (99) Tahara, S.; Hashidoko, Y.; Ingham, J. L.; Mizutani, J. Isoflavonoids of yellow lupin. Part II. New 5-O-methylisoflavones in the roots of yellow lupine (Lupinus luteus L. cv. Barpine). Agric. Biol. Chem. 1986, 50, 1809–1819.
- (100) Hashidoko, Y.; Tahara, S.; Mizutani, J. Isoflavonoids of yellow lupine. Part I. New complex isoflavones in the root of yellow lupine (Lupinus luteus L., cv. Barpine). Agric. Biol. Chem. 1986, 50, 1797–1807.
- (101) Tahara, S.; Ingham, J. L.; Mizutani, J. New coumaronochromones from white lupine, Lupinus albus L. (Leguminosae). Agric. Biol. Chem. 1985, 49, 1775–1783.
- (102) Ingham, J. L.; Tahara, S.; Harborne, J. B. Fungitoxic isoflavones from Lupinus albus and other Lupinus species. Z. Naturforsch., C: J. Biosci. 1983, 38, 194–200.
- (103) Fukui, H.; Egawa, H.; Koshimizu, K.; Mitsui, T. isoflavone with antifungal activity from immature fruits of Lupinus luteus. *Agric. Biol. Chem.* 1973, 37, 417–421.
- (104) Shi, J.; Arunasalam, K.; Yeung, D.; Kakuda, Y.; Mittal, G.; Jiang, Y. Saponins from edible legumes: chemistry, processing, and health benefits. *J. Med. Food* 2004, 7, 67–78.
- (105) Kennelly, E. J.; Suttisri, R.; Kinghorn, A. D. Novel sweet-tasting saponins of the cycloartane, oleanane, secodammarane, and steroidal types. Adv. Exp. Med. Biol. 1996, 405, 13–24.
- (106) Uher, M. In Natural and Synthetic Nonsaccharide Sweeteners, Chemical and Functional Properties of Food Saccharides 2004; Tomasik, P., Ed.; CRC Press LLC: Boca Raton, FL, 2004; pp 387–403.
- (107) Woldemichael, G. M.; Montenegro, G.; Timmermann, B. N. Triterpenoidal lupin saponins from the Chilean legume Lupinus oreophilus Phil. *Phytochemistry* 2003, 63, 853–857.
- (108) Woldemichael, G. M.; Wink, M. Triterpene glycosides of Lupinus angustifolius. *Phytochemistry* 2002, 60, 323–327.
- (109) Morcos, S. R.; Gabrial, G. N.; El-Hafez, M. A. Nutritive studies on some raw and prepared leguminous seeds commonly used in the Arab Republic of Syria. Z. Ernahrungswiss. 1976, 15, 378– 386
- (110) Ruiz, R. G.; Price, K. R.; Arthur, A. E.; Rose, M. E.; Rhodes, M. J. C.; Fenwick, R. G. Effect of Soaking and Cooking on the Saponin Content and Composition of Chickpeas (Cicer arietinum) and Lentils (Lens culinaris). J. Agric. Food Chem. 1996, 44, 1526–1530.
- (111) Kerem, Z.; German-Shashoua, H.; Yarden, O. Microwave-assisted extraction of bioactive saponins from chickpea (Cicer arietinum L). J. Sci. Food. Agric. 2005, 85, 406–412.
- (112) El-Adawy, T. A. Nutritional composition and antinutritional factors of chickpeas (Cicer arietinum L.) undergoing different cooking methods and germination. *Plant Foods Hum. Nutr.* 2002, 57, 83–89.

- (113) Alzorriz, M. A. Z.; Hernandez, J. A. M. Possible beneficial role of dietary chickpea Cicer aretinum L. var. macrocarpum or pharmacological treatment with β 3-adrenergic agonist. *Anal. Real Acad. Farm.* 1999, 65, 327–349.
- (114) Zou, K.; Zhao, Y. Y.; Zhang, R. Y. A cytotoxic saponin from Albizia julibrissin. Chem. Pharm. Bull. (Tokyo) 2006, 54, 1211– 1212.
- (115) Chang, W. W.; Yu, C. Y.; Lin, T. W.; Wang, P. H.; Tsai, Y. C. Soyasaponin I decreases the expression of alpha2,3-linked sialic acid on the cell surface and suppresses the metastatic potential of B16F10 melanoma cells. *Biochem. Biophys. Res. Commun.* 2006, 341, 614–619.
- (116) Ellington, A. A.; Berhow, M. A.; Singletary, K. W. Inhibition of Akt signaling and enhanced ERK1/2 activity are involved in induction of macroautophagy by triterpenoid B-group soyasaponins in colon cancer cells. *Carcinogenesis* 2006, 27, 298–306.
- (117) MacDonald, R. S.; Guo, J.; Copeland, J.; Browning, J. D., Jr.; Sleper, D.; Rottinghaus, G. E.; Berhow, M. A. Environmental influences on isoflavones and saponins in soybeans and their role in colon cancer. *J. Nutr.* 2005, 135, 1239–1242.
- (118) Hu, J.; Reddy, M. B.; Hendrich, S.; Murphy, P. A. Soyasaponin I and sapongenol B have limited absorption by Caco-2 intestinal cells and limited bioavailability in women. *J. Nutr.* **2004**, *134*, 1867–1873.
- (119) Gurfinkel, D. M.; Rao, A. V. Soyasaponins: the relationship between chemical structure and colon anticarcinogenic activity. *Nutr. Cancer* 2003, 47, 24–33.
- (120) Rajput, Z. I.; Hu, S. H.; Xiao, C. W.; Arijo, A. G. Adjuvant effects of saponins on animal immune responses. J. Zhejiang Univ., Sci. B 2007, 8, 153–161.
- (121) Bieberich, E.; Tencomnao, T.; Kapitonov, D.; Yu, R. K. Effect of N-glycosylation on turnover and subcellular distribution of N-acetylgalactosaminyltransferase I and sialyltransferase II in neuroblastoma cells. J. Neurochem. 2000, 74, 2359–2364.
- (122) Aubert, M.; Panicot, L.; Crotte, C.; Gibier, P.; Lombardo, D.; Sadoulet, M. O.; Mas, E. Restoration of alpha(1,2) fucosyltransferase activity decreases adhesive and metastatic properties of human pancreatic cancer cells. *Cancer Res.* 2000, 60, 1449–1456.
- (123) Nakamura, M.; Kudo, T.; Narimatsu, H.; Furukawa, Y.; Kikuchi, J.; Asakura, S.; Yang, W.; Iwase, S.; Hatake, K.; Miura, Y. Single glycosyltransferase, core 2 beta 1→6-N-acetylglucosaminyltransferase, regulates cell surface sialyl-Lex expression level in human pre-B lymphocytic leukemia cell line KM3 treated with phorbolester. J. Biol. Chem. 1998, 273, 26779–26789.
- (124) Yamamoto, H.; Kaneko, Y.; Rebbaa, A.; Bremer, E. G.; Moskal, J. R. alpha2,6-Sialyltransferase gene transfection into a human glioma cell line (U373 MG) results in decreased invasivity. J. Neurochem. 1997, 68, 2566–2576.
- (125) Gorelik, E.; Duty, L.; Anaraki, F.; Galili, U. Alterations of cell surface carbohydrates and inhibition of metastatic property of murine melanomas by alpha 1,3 galactosyltransferase gene transfection. *Cancer Res.* 1995, 55, 4168–4173.
- (126) Harvey, B. E.; Thomas, P. Inhibition of CMP-sialic acid transport in human liver and colorectal cancer cell lines by a sialic acid nucleoside conjugate (KI-8110). *Biochem. Biophys. Res. Commun.* 1993, 190, 571–575.
- (127) Xiao, J. X.; Huang, G. Q.; Zhu, C. P.; Ren, D. D.; Zhang, S. H. Morphological study on apoptosis Hela cells induced by soyasaponins. *Toxicol. in Vitro* 2007.
- (128) Godlewski, M. M.; Slazak, P.; Zabielski, R.; Piastowska, A.; Gralak, M. A. Quantitative study of soybean-induced changes in proliferation and programmed cell death in the intestinal mucosa of young rats. *J. Physiol. Pharmacol.* 2006, 57 (Suppl. 7), 125–133.
- (129) Zhu, J.; Xiong, L.; Yu, B.; Wu, J. Apoptosis induced by a new member of saponin family is mediated through caspase-8dependent cleavage of Bcl-2. *Mol. Pharmacol.* 2005, 68, 1831– 1838.
- (130) Han, M.; Han, L. M.; Wang, Q. S.; Bai, Z. H.; Fang, X. L. [Mechanism of oral absorption of panaxnotoginseng saponins]. Yao Xue Xue Bao 2006, 41, 498–505.

- (131) Huang, C. R.; Wang, G. J.; Wu, X. L.; Li, H.; Xie, H. T.; Lv, H.; Sun, J. G. Absorption enhancement study of astragaloside IV based on its transport mechanism in caco-2 cells. *Eur. J. Drug Metab. Pharmacokinet.* 2006, 31, 5–10.
- (132) Han, M.; Fang, X. L. Difference in oral absorption of ginsenoside Rg1 between in vitro and in vivo models. *Acta Pharmacol. Sin.* 2006, 27, 499–505.
- (133) Paek, I. B.; Moon, Y.; Kim, J.; Ji, H. Y.; Kim, S. A.; Sohn, D. H.; Kim, J. B.; Lee, H. S. Pharmacokinetics of a ginseng saponin metabolite compound K in rats. *Biopharm. Drug Dispos.* 2006, 27, 39–45.
- (134) Calvert, G. D.; Blight, L.; Illman, R. J.; Topping, D. L.; Potter, J. D. A trial of the effects of soya-bean flour and soya-bean saponins on plasma lipids, faecal bile acids and neutral sterols in hypercholesterolaemic men. *Br. J. Nutr.* 1981, 45, 277–281.
- (135) Sidhu, G. S.; Oakenfull, D. G. A mechanism for the hypocholesterolaemic activity of saponins. *Br. J. Nutr.* 1986, 55, 643–649.
- (136) Jood, S.; Chauhan, B. M.; Kapoor, A. C. Saponin content of chickpea and black gram: Varietal differences and effects of processing and cooking methods. J. Sci. Food Agric. 2006, 37, 1121–1124
- (137) Price, K. R.; Fenwick, G. R.; Jurzysta, M. Soyasapogenols separation, analysis and interconversions. *J. Sci. Food Agric*. 1986, 37, 1027–1034.
- (138) Barile, E.; Bonanomi, G.; Antignani, V.; Zolfaghari, B.; Sajjadi, S. E.; Scala, F.; Lanzotti, V. Saponins from Allium minutiflorum with antifungal activity. *Phytochemistry* 2007, 68, 596–603.
- (139) Simons, V.; Morrissey, J. P.; Latijnhouwers, M.; Csukai, M.; Cleaver, A.; Yarrow, C.; Osbourn, A. Dual effects of plant steroidal alkaloids on Saccharomyces cerevisiae. *Antimicrob. Agents Chemother.* 2006, 50, 2732–2740.
- (140) Alabdul Magid, A.; Voutquenne, L.; Harakat, D.; Pouny, I.; Caron, C.; Moretti, C.; Lavaud, C. Triterpenoid saponins from the fruits of Caryocar villosum. J. Nat. Prod. 2006, 69, 919– 926
- (141) Mandal, P.; Sinha Babu, S. P.; Mandal, N. C. Antimicrobial activity of saponins from Acacia auriculiformis. *Fitoterapia* 2005, 76, 462–465.
- (142) Plaza, A.; Cinco, M.; Tubaro, A.; Pizza, C.; Piacente, S. New triterpene glycosides from the stems of Anomospermum grandifolium. J. Nat. Prod. 2003, 66, 1606–1610.
- (143) Pistelli, L.; Bertoli, A.; Lepori, E.; Morelli, I.; Panizzi, L. Antimicrobial and antifungal activity of crude extracts and isolated saponins from Astragalus verrucosus. *Fitoterapia* 2002, 73, 336–339.
- (144) Bouarab, K.; Melton, R.; Peart, J.; Baulcombe, D.; Osbourn, A. A saponin-detoxifying enzyme mediates suppression of plant defences. *Nature* 2002, 418, 889–892.
- (145) Bedir, E.; Khan, I. A.; Walker, L. A. Biologically active steroidal glycosides from Tribulus terrestris. *Pharmazie* 2002, 57, 491– 493.
- (146) Iorizzi, M.; Lanzotti, V.; Ranalli, G.; De Marino, S.; Zollo, F. Antimicrobial furostanol saponins from the seeds of Capsicum annuum L. var. acuminatum. J. Agric. Food Chem. 2002, 50, 4310–4316.
- (147) Yadava, R. N. A new biologically active triterpenoid saponin from the leaves of Lepidagathis hyalina Nees. *Nat. Prod. Lett.* **2001**, *15*, 315–322.
- (148) Papadopoulou, K.; Melton, R. E.; Leggett, M.; Daniels, M. J.; Osbourn, A. E. Compromised disease resistance in saponindeficient plants. *Proc. Natl. Acad. Sci. U.S.A.* 1999, 96, 12923– 12928
- (149) Hymowitz, T. Grain legumes. In Advances in new crops; Janick, J., Simon, J. E., Eds.; Timber Press: Portland, OR, 1990; pp 54–57
- (150) Hammouda, I. F. M.; Ahmed, Z. F. A phytochemical study of the seeds of Pisum sativum. I. Congr. Sci. Farm. Conf. Commun. 1962, 21, 551–564.

- (151) Ginsburg, H.; Atamna, H.; Shalmiev, G.; Kanaani, J.; Krugliak, M. Resistance of glucose-6-phosphate dehydrogenase deficiency to malaria: effects of fava bean hydroxypyrimidine glucosides on Plasmodium falciparum growth in culture and on the phagocytosis of infected cells. *Parasitology* 1996, 113 (Pt 1), 7–18
- (152) Mohamed, M. H. New alkaloids from Lupinus albus L. seeds. Bull. Pharm. Sci. 1998, 21, 55–60.
- (153) Dini, I.; Schettino, O.; Dini, A. Studies on the Constituents of Lupinus mutabilis (Fabaceae). Isolation and Characterization of Two New Isoflavonoid Derivatives. J. Agric. Food Chem. 1998, 46, 5089–5092.
- (154) Asres, K. The alkaloids of Lupinus princei. *Egypt. J. Pharm. Sci.* **1996**, *37*, 1–10.
- (155) Mohamed, M. H.; Koskinen, A. M. Termine, a new lupin alkaloid from the seeds of Lupinus termis. *Bull. Pharm. Sci., Assiut Univ.* 1995, 18, 55–58.
- (156) Mohamed, M. H.; El-Shorbagi, A. N. A. (±)-Termisine, a novel lupine alkaloid from the seeds of Lupinus termis. *J. Nat. Prod.* **1993**, *56*, 1999–2002.
- (157) Strack, D.; Becher, A.; Brall, S.; Witte, L. Quinolizidine alkaloids and the enzymic syntheses of their cinnamic and hydroxycinnamic acid esters in Lupinus angustifolius and L. luteus. *Phytochemistry* 1991, 30, 1493–1498.
- (158) Khafagy, S. M.; El-Masry, S.; Saleh, M. R. I.; Dabbas, S. W. A. Phytochemical investigation of Lupinus termis grown in Egypt. *Pharmazie* **1974**, *29*, 65–66.
- (159) Lopez, G.; Pedro, M.; Garzon de la Mora, P.; Wysocka, W.; Maiztegui, B.; Alzugaray, M. E.; Del Zotto, H.; Borelli, M. I. Quinolizidine alkaloids isolated from Lupinus species enhance insulin secretion. *Eur. J. Pharmacol.* 2004, 504, 139–142.
- (160) Gabrial, G. N.; Morcos, S. R. The use of Lupinus termis L. cultivated in Egypt, as a food protein supplement. Z. Ernahrungswiss. 1976, 15, 333–339.
- (161) Wink, M. Plant breeding: importance of plant secondary metabolites for protection against pathogens and herbivores. *Theor. Appl. Genet.* 1988, 75, 225–233.
- (162) Patwardhan, V. N.; White, J. W., Jr. Problems associated with particular foods. *Toxicants Occurring Nat. Food, 2nd Ed.* 1973, 477–507.
- (163) Cristofaro, E.; Mottu, F.; Wuhrmann, J. J. Involvement of the raffinose family of oligosaccharides in flatulence. *Sugars Nutr.*, [Pap. Int. Conf.] 1974, 313–336.
- (164) Schneider, A. V. Overview of the market and consumption of pulses in Europe. *Br. J. Nutr.* **2002**, 88 (Suppl. 3), S243–250.
- (165) Saini, H. S.; Knights, E. J. Chemical constitution of starch and oligosaccharide components of 'desi' and 'kabuli' chickpea (Cicer arietinum) seed types. *J. Agric. Food Chem.* 1984, 32, 940–944.
- (166) Saini, H. S. In Legume seed oligosaccharides; Recent Adv. Res. Antinutr. Factors Legume Seeds, Proc. Int. Workshop, 1988; Huisman, J., Van der Poel, T. F. B., Liener, I. E., Eds.; Pudoc: Wageningen, Neth, 1989; pp 329–341.
- (167) Eiwegger, T.; Stahl, B.; Schmitt, J.; Boehm, G.; Gerstmayr, M.; Pichler, J.; Dehlink, E.; Loibichler, C.; Urbanek, R.; Szepfalusi, Z. Human milk-derived oligosaccharides and plant-derived oligosaccharides stimulate cytokine production of cord blood T-cells in vitro. *Pediatr. Res.* 2004, 56, 536–540.
- (168) Swanson, K. S.; Grieshop, C. M.; Flickinger, E. A.; Healy, H. P.; Dawson, K. A.; Merchen, N. R.; Fahey, G. C., Jr. Effects of supplemental fructooligosaccharides plus mannanoligosaccharides on immune function and ileal and fecal microbial populations in adult dogs. *Arch. Tierernahr.* 2002, 56, 309–318.
- (169) Schley, P. D.; Field, C. J. The immune-enhancing effects of dietary fibres and prebiotics. *Br. J. Nutr.* 2002, 87 (Suppl. 2), S221–230.
- (170) Buddington, R. K.; Kelly-Quagliana, K.; Buddington, K. K.; Kimura, Y. Non-digestible oligosaccharides and defense functions: lessons learned from animal models. *Br. J. Nutr.* 2002, 87, S231–S239.

- (171) Lindsay, J. O.; Whelan, K.; Stagg, A. J.; Gobin, P.; Al-Hassi, H. O.; Rayment, N.; Kamm, M. A.; Knight, S. C.; Forbes, A. Clinical, microbiological, and immunological effects of fructooligosaccharide in patients with Crohn's disease. *Gut* 2006, 55, 348–355
- (172) Bruzzese, E.; Volpicelli, M.; Squaglia, M.; Tartaglione, A.; Guarino, A. Impact of prebiotics on human health. *Dig. Liver Dis.* 2006, 38 (Suppl. 2), S283–287.
- (173) Oddy, W. H. The impact of breastmilk on infant and child health. Breastfeed. Rev. 2002, 10, 5–18.
- (174) Walzem, R. L.; Dillard, C. J.; German, J. B. Whey components: millennia of evolution create functionalities for mammalian nutrition: what we know and what we may be overlooking. *Crit. Rev. Food Sci. Nutr.* 2002, 42, 353–375.
- (175) Chierici, R.; Fanaro, S.; Saccomandi, D.; Vigi, V. Advances in the modulation of the microbial ecology of the gut in early infancy. *Acta Paediatr. Suppl.* 2003, 91, 56–63.
- (176) Mountzouris, K. C.; McCartney, A. L.; Gibson, G. R. Intestinal microflora of human infants and current trends for its nutritional modulation. *Br. J. Nutr.* 2002, 87, 405–420.
- (177) Stahl, B.; M'Rabet, L.; Vos, A. P.; Garssen, J.; Boehm, G. Immunomodulating oligosaccharides, and therapeutic use thereof. 2004-NL750 2005039597, 20041025, 2005.
- (178) Kaur, N.; Gupta, A. K. Applications of inulin and oligofructose in health and nutrition. J. Biosci. 2002, 27, 703–714.
- (179) Adebowale, Y. A.; Adeyemi, A.; Oshodi, A. A. Variability in the physicochemical, nutritional and antinutritional attributes of six Mucuna species. *Food Chem.* 2004, 89, 37–48.
- (180) Juliano, B. O. Comparative nutritive value of various staple foods. Food Rev. Int. 1999, 15, 399–434.
- (181) Manary, M. J.; Hotz, C.; Krebs, N. F.; Gibson, R. S.; Westcott, J. E.; Broadhead, R. L.; Hambidge, K. M. Zinc homeostasis in Malawian children consuming a high-phytate, maize-based diet. *Am. J. Clin. Nutr.* 2002, 75, 1057–1061.
- (182) Hemalatha, S.; Platel, K.; Srinivasan, K. Influence of germination and fermentation on bioaccessibility of zinc and iron from food grains. Eur. J. Clin. Nutr. 2007, 61, 342–348.
- (183) Lind, T.; Lonnerdal, B.; Persson, L. A.; Stenlund, H.; Tennefors, C.; Hernell, O. Effects of weaning cereals with different phytate contents on hemoglobin, iron stores, and serum zinc: a randomized intervention in infants from 6 to 12 mo of age. Am. J. Clin. Nutr. 2003, 78, 168–175.
- (184) Adams, C. L.; Hambidge, M.; Raboy, V.; Dorsch, J. A.; Sian, L.; Westcott, J. L.; Krebs, N. F. Zinc absorption from a lowphytic acid maize. Am. J. Clin. Nutr. 2002, 76, 556–559.
- (185) Manary, M. J.; Krebs, N. F.; Gibson, R. S.; Broadhead, R. L.; Hambidge, K. M. Community-based dietary phytate reduction and its effect on iron status in Malawian children. *Ann. Trop. Paediatr.* 2002, 22, 133–136.
- (186) Manary, M. J.; Hotz, C.; Krebs, N. F.; Gibson, R. S.; Westcott, J. E.; Arnold, T.; Broadhead, R. L.; Hambidge, K. M. Dietary phytate reduction improves zinc absorption in Malawian children recovering from tuberculosis but not in well children. *J. Nutr.* 2000, 130, 2959–2964.
- (187) Sandberg, A. S. Bioavailability of minerals in legumes. Br. J. Nutr. 2002, 88 (Suppl. 3), S281–285.
- (188) Cheryan, M. Phytic acid interactions in food systems. Crit. Rev. Food Sci. Nutr. 1980, 13, 297–335.
- (189) Ravindran, V.; Ravindran, G.; Silvalogan, S. Total and phytate phosphorous contents of various food and feed stuffs of plant origin. *Food Chem.* 1994, 50, 113–136.
- (190) Thompson, L. U. Potential health benefits and problems associated with anti-nutrients in foods. *Food Res. Int.* 1993, 26, 131–149.
- (191) Sandberg, A. S.; Carlsson, N. G.; Svanberg, U. Effects of inositol tri-, tetra-, penta-, and hexaphosphates on in vitro estimation of iron availability. J. Food Sci. 1989, 54, 159–161.
- (192) Szwergold, B. S.; Graham, R. A.; Brown, T. R. Observation on inositol pentakis- and hexakis-phosphates in mammalian tissues by ¹³P NMR. *Biochem. Biophys. Res. Commun.* **1987**, 264, 874– 881.

- (193) Menniti, F. S.; Oliver, K. G.; Putney, J. W.; Shears, S. B. Inositol phosphates and cell signalling: new views of InsP₅ and InsP₆. *TIBS* 1993, 18, 53–56.
- (194) Champ, M. M. Non-nutrient bioactive substances of pulses. Br. J. Nutr. 2002, 88 (Suppl. 3), S307–319.
- (195) Graf, E.; Eaton, J. W. Antioxidant functions of phytic acid. Free Radic. Biol. Med. 1990, 8, 61–69.
- (196) Thompson, L. U.; Zhang, L. Phytic acid and minerals: effect on early markers of risk for mammary and colon carcinogenesis. *Carcinogenesis* 1991, 12, 2041–2045.
- (197) Shamsuddin, A. M.; Vucenik, I. Mammary tumor inhibition by IP6: a review. Anticancer Res. 1999, 19, 3671–3674.
- (198) Zi, X.; Singh, R. P.; Agarwal, R. Impairment of erbB1 receptor and fluid-phase endocytosis and associated mitogenic signaling by inositol hexaphosphate in human prostate carcinoma DU145 cells. *Carcinogenesis* 2000, 21, 2225–2235.
- (199) Shamsuddin, A. M. Anti-cancer function of phytic acid. Int. J. Food Sci. Technol. 2002, 37, 769–768.
- (200) Vucenik, I.; Shamsuddin, A. M. Protection against cancer by dietary IP6 and inositol. *Nutr. Cancer* 2006, 55, 109–125.
- (201) Singh, R. P.; Agarwal, R. Prostate cancer and inositol hexaphosphate: efficacy and mechanisms. *Anticancer Res.* 2005, 25, 2891– 2903.
- (202) Shamsuddin, A. M. Anti-cancer function of phytic acid. Int. J. Food Sci. Technol. 2002, 37, 769–782.
- (203) Urbano, G.; Lopez-Jurado, M.; Aranda, P.; Vidal-Valverde, C.; Tenorio, E.; Porres, J. The role of phytic acid in legumes: Antinutrient or beneficial function. J. Physiol. Biochem. 2000, 56, 283–294.
- (204) Shamsuddin, A. K. M.; Vucenik, I.; Yang, G. Y. Anti-cancer function of phytic acid from cereals and legumes. *Proc. Int. Cancer Congr., Free Pap. Posters*, 16th 1994 1994, 1, 647–650.
- (205) Shamsuddin, A. M. Inositol phosphates have novel anticancer function. J. Nutr. 1995, 125, 7258–7328.
- (206) Agarwal, C.; Dhanalakshmi, S.; Singh, R. P.; Agarwal, R. Inositol hexaphosphate inhibits growth and induces G1 arrest and apoptotic death of androgen-dependent human prostate carcinoma LNCaP cells. *Neoplasia* 2004, 6, 646–659.
- (207) Singh, R. P.; Agarwal, C.; Agarwal, R. Inositol hexaphosphate inhibits growth, and induces G1 arrest and apoptotic death of prostate carcinoma DU145 cells: modulation of CDKI-CDKcyclin and pRb-related protein-E2F complexes. *Carcinogenesis* 2003, 24, 555–563.
- (208) Tian, Y.; Song, Y. Effects of inositol hexaphosphate on proliferation of HT-29 human colon carcinoma cell line. World J. Gastroenterol. 2006, 12, 4137–4142.
- (209) Weglarz, L.; Molin, I.; Orchel, A.; Parfiniewicz, B.; Dzierzewicz, Z. Quantitative analysis of the level of p53 and p21(WAF1) mRNA in human colon cancer HT-29 cells treated with inositol hexaphosphate. Acta Biochim. Pol. 2006, 53, 349–356.
- (210) Zhang, Z.; Song, Y.; Wang, X. L. Inositol hexaphosphate-induced enhancement of natural killer cell activity correlates with suppression of colon carcinogenesis in rats. World J. Gastroenterol. 2005, 11, 5044–5046.
- (211) Jenab, M.; Thompson, L. U. Phytic acid in wheat bran affects colon morphology, cell differentiation and apoptosis. *Carcinogenesis* **2000**, *21*, 1547–1552.
- (212) Fox, C. H.; Eberl, M. Phytic acid (IP6), novel broad spectrum anti-neoplastic agent: a systematic review. *Complement. Ther. Med.* **2002**, *10*, 229–234.
- (213) Deliliers, G. L.; Servida, F.; Fracchiolla, N. S.; Ricci, C.; Borsotti, C.; Colombo, G.; Soligo, D. Effect of inositol hexaphosphate (IP(6)) on human normal and leukaemic haematopoietic cells. *Br. J. Haematol.* 2002, 117, 577–587.
- (214) Shamsuddin, A. M.; Baten, A.; Lalwani, N. D. Effects of inositol hexaphosphate on growth and differentiation in K-562 erythroleukemia cell line. *Cancer Lett.* 1992, 64, 195–202.
- (215) Iyengar, A. K.; Kulkarni, P. R. Oligosaccharide levels of processed legumes. J. Food Sci. Technol. 1977, 14, 222–223.

- (216) Rose, S.; Bush, C. S.; Bettle, G., III; Rutzinski, J. L.; Malnati, L. Process for reducing flatulence in legumes by soaking. 2000-489745 6355291, 20000121, 2002.
- (217) Wilson, J. R.; Payne, M. A.; Rakijian, D. R.; Zane, R. S. O. Process for removing flatulence-associated oligosaccharides in legumes. 2006-365002 2006198934, 20060228, 2006.
- (218) Han, I. H.; Baik, B.-K. Oligosaccharide content and composition of legumes and their reduction by soaking, cooking, ultrasound, and high hydrostatic pressure. *Cereal Chem.* 2006, 83, 428–433.
- (219) Frias, J.; Doblado, R.; Vidal-Valverde, C. Kinetics of soluble carbohydrates by action of endo/exo a-galactosidase enzyme in lentils and peas. Eur. Food Res. Technol. 2003, 216, 199–203.
- (220) Savitri, A.; Desikachar, H. S. R. A comparative study of flatus production in relation to the oligosaccharide content of some legumes. *Nutr. Rep. Int.* 1985, 31, 337–344.
- (221) Garg, S. K.; Adlakha, G.; Mital, B. K. Incidence of flatulence upon ingestion of pulses and legumes. *Microbiol., Aliments, Nutr.* 1992, 10, 11–22.
- (222) Khokhar, S.; Frias, J.; Price, K. R.; Fenwock, G. R.; Hedley, C. L. Physico-chemical characteristics of khesari dhal (Lathyrus sativus): changes in a-galactosides, monosaccharides and disaccharides during food processing. *J. Sci. Food. Agric.* 1996, 70, 487–492.
- (223) Jaya, T. V.; Naik, H. S.; Venkatraman, L. V. Effect of germinated legumes on the rate of in vitro gas production by Clostridium perfringens. *Nutr. Rep. Int.* 1979, 20, 393–401.
- (224) Machaiah, J. P.; Pednekar, M. D. Carbohydrate composition of low dose radiation-processed legumes and reduction in flatulence factors. *Food Chem.* 2002, 79, 293–301.

- (225) Stevenson-Paulik, J.; Bastidas, R. J.; Chiou, S. T.; Frye, R. A.; York, J. D. Generation of phytate-free seeds in Arabidopsis through disruption of inositol polyphosphate kinases. *Proc. Natl. Acad. Sci. U.S.A.* 2005, 102, 12612–12617.
- (226) Jones, D. A.; DuPont, M. S.; Ambrose, M. J.; Frias, J.; Hedley, C. L. The discovery of compositional variation for the raffinose family of oligosaccharides in pea seeds. *Seed Sci. Res.* 1999, 9, 305–310.
- (227) Caffrey, M.; Fonseca, V.; Leopold, A. C. Lipid-Sugar Interactions: Relevance to Anhydrous Biology. *Plant Physiol.* 1988, 86, 754–758.
- (228) De Lumen, B. O. Molecular strategies to improve protein quality and reduce flatulence in legumes: a review. *Food Struct.* 1992, 11, 33–46.
- (229) de Almeida Costa, G. E.; Queiroz-Monici, K. d. S.; Reis, S. M. P. M; Costa de Oliveira, A. Chemical composition, dietary fibre and resistant starch contents of raw and cooked pea, common bean, chickpea and lentil legumes. *Food Chem.* 2006, 94, 327–330.
- (230) Morris, E. R.; Hill, A. D. Inositol phosphate content of selected dry beans, peas and lentils, raw and cooked. *J. Food Compos. Anal.* 1996, 9, 2–12.
- (231) Bhatty, R. S.; Slinkard, A. E. Relationship between phytic acid and cooking quality in lentil. J. Can. Inst. Food Sci. Technol. 1989, 22, 137–142.

Received for review June 11, 2007. Revised manuscript received July 21, 2007. Accepted July 24, 2007.

JF071704W