Journal of Medicinal Chemistry

© Copyright 2000 by the American Chemical Society

Volume 43, Number 23

November 16, 2000

Perspective

Therapeutic Opportunities for Muscarinic Receptors in the Central Nervous System

Christian C. Felder,* Frank P. Bymaster, John Ward, and Neil DeLapp

Eli Lilly Research Laboratories, Indianapolis, Indiana 46285

Received December 14, 1999

Introduction

The neurotransmitter acetylcholine binds to two types of cholinergic receptors: the ionotropic family of nicotinic receptors and the metabotropic family of muscarinic receptors (Figure 1). Nicotinic receptors are ligandgated ion channels that modulate cell membrane potentials. Muscarinic receptors belong to the large superfamily of plasma membrane-bound G proteincoupled receptors (GPCRs). To date, five subtypes of muscarinic receptors have been cloned and sequenced from a variety of species including mammalian (human, cow, pig, rat, mouse) and nonmammalian (chicken, Drosophila melanogaster, frog) and show a remarkably high degree of sequence similarity across species as well as across receptor subtype. The M₁-M₅ muscarinic receptors are expressed predominantly within the parasympathetic nervous system which exerts excitatory and inhibitory control over central and peripheral tissues and participate in a number of physiologic functions including heart rate, arousal, cognition, sensory processing, and motor control. Availability of molecular probes and receptor subtype-selective antibodies has provided detailed knowledge of receptor distribution and shown significant overlap in a wide variety of brain regions and peripheral tissues. Muscarinic agonists such as muscarine and pilocarpine and antagonists such as atropine have been known for over a century, but little progress has been made in the discovery of receptor subtype-selective compounds making it difficult to assign specific functions to the individual receptors. Recently, muscarinic receptor knockout mice have been developed and some improvement has been made in the selectivity of ligands that will hopefully generate a new understanding of the role of each receptor in physiological processes and especially brain function.

This review will focus on the pharmacology of the muscarinic receptor family and therapeutic opportunities for muscarinic intervention in disorders of the central nervous system. Discussion of the recent advances in the pharmacology and molecular biochemistry of muscarinic receptors will be followed by summaries of the involvement of this family of receptors in psychosis, Alzheimer's disease, and Parkinson's disease. Due to their broad central and peripheral distribution, muscarinic receptors have shown promise as targets for the treatment of such diverse diseases such as glaucoma, bradycardia, gastric acid secretion, and asthma and as antispasmotics for GI disturbances. Additional relevant reviews have discussed these topics including specific discussion of muscarinic receptor characterization, 1-3 smooth muscle function, 4 molecular analysis, 5 ion channel function,6 subtype-specific toxins,7 signal transduction,8 and pharmacology.9

Muscarinic Receptor Structure and Function

 M_1 – M_5 Receptor Genes. The first pharmacological evidence for the presence of multiple subtypes of muscarinic receptors appeared in the early $1950s^{10}$ and became more evident as differences in tissue responses to various muscarinic ligands were observed. Before cloning revealed at least five receptor subtypes, pharmacological evidence was able to resolve at least three

 $^{^{\}ast}$ Corresponding author address: Eli Lilly Research Laboratories, Drop 0510, Lilly Corporate Center, Indianapolis, IN 46285. Phone: 317-276-1380. Fax: 317-276-7600, alt fax: 317-276-5546. E-mail: felder@lilly.com.

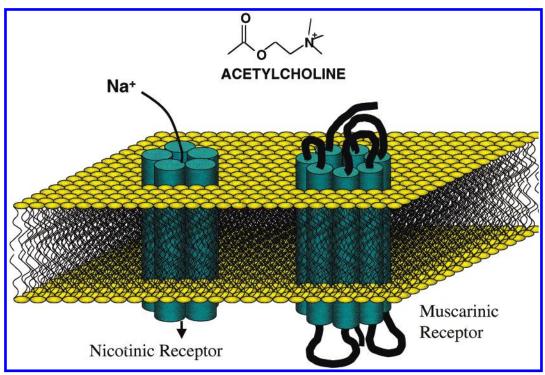


Figure 1. Cartoon of the two types of acetylcholine receptors. Acetylcholine receptors are integral membrane proteins that occur as either nicotinic-sensitive ionotropic or muscarinic-sensitive metabotropic receptors. Nicotinic receptors are ion channels created by the assembly of five protein subunits. Muscarinic receptors belong to the superfamily of GPCRs which are characterized as a single peptide which weaves in and out of the plasma membrane creating seven hydrophobic membrane spanning domains and three extracellular and cytoplasmic loops.

pharmacologically defined subtypes classified by capital M₁-M₃. Genes encoding the M₁ and M₂ receptors were cloned first by Numa and colleagues¹¹ followed by genes encoding the M₃-M₅ receptors by Bonner et al. ^{12,13} No convincing pharmacological or molecular evidence suggesting that additional members of this family exist has been provided to date. A recent meeting of the IUPHAR panel on GPCR nomenclature has suggested a common designation for the five muscarinic receptor subtypes denoted by capital M₁-M₅ to reflect their order of discovery.³ The five known muscarinic receptor subtypes share common characteristics with the superfamily of GPCRs and especially those that bind small molecules or neurotransmitters. 14 The muscarinic receptors are single integral plasma membrane glycoproteins coded by genes that lack introns. Their topology upon folding has recently been shown to be similar to the wellcharacterized photoreceptor rhodopsin which is believed to have seven hydrophobic α-helical domains spanning the plasma membrane seven times connected by hydrophilic intracellular and extracellular loops. 15,16

Progress in developing therapeutically useful muscarinic agents has been slow based on the current inventory of relatively nonselective ligands for the five receptor subtypes. This difficulty may be due to the significant amino acid sequence identity between the five receptors possibly creating similarity across all five binding domains. One must keep in mind, however, that examples exist of single amino acid differences in GPCR subtypes that yield significant differences in ligand binding affinity.¹⁷ X-ray crystallography of the prototypic GPCR, bacteriorhodopsin, has been used to predict determinants of ligand-receptor interactions through binding site modeling. However, a high-resolution crystal structure does not currently exist for the muscarinic

or other related GPCRs. Therefore, receptors have been modeled based on the low-resolution structure of rhodopsin¹⁸ and computer modeling of the sequences of known GPCRs. 19-22 These models predict that the seven transmembrane spanning domains form a pocket that defines the ligand-binding domain. A two-dimensional electron crystallographic map of bovine rhodopsin confirmed the expectation that the seven hydrophobic helical bundles form a cylindrical type core with four of the bundles almost perpendicular to the membrane and three take on a more tilted orientation.²⁰ A recent 2.8 Å threedimensional crystal structure of rhodopsin was completed that will no doubt help refine our predictions about muscarinic receptor structure and function. 16

The critical amino acid residues involved in acetylcholine binding have been identified by covalent affinitylabeling and mutagenesis experiments. Acetylcholine is predicted to interact with amino acids present in the outer regions of the binding pocket about 10−15 Å from the surface. In the M₁ receptor, a critical asparagine residue (Asp105) is involved in binding of the positively charged amino headgroup of acetylcholine and most equivalent muscarinic ligands based on binding of radiolabeled acetylcholine mustard²³ and propylbenzylcholine mustard. 19,24 Mutagenesis of this site resulted in a reduction in ligand affinities. 25,26 Asp105 is conserved in most GPCRs that bind biogenic amine type neurotransmitters. Six additional residues critical for acetylcholine binding (Thr231, Thr234, Tyr148, Tyr506, Tyr529, Tyr533) are conserved across the five muscarinic receptors but not other GPCRs. Two tryptophans in the M₃ receptor (Trp503, Trp143) which are conserved across many GPCRs were found to be critically involved in high-affinity ligand binding and are predicted to face the ligand-binding cavity.²⁷

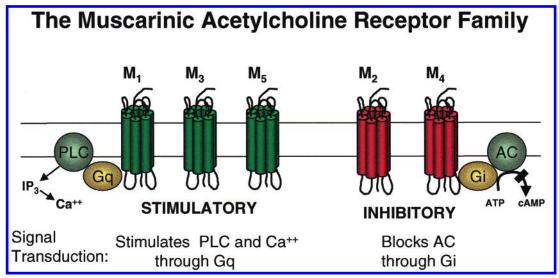


Figure 2. Family of five muscarinic acetylcholine receptors. The muscarinic receptor family has five known members designated M_1-M_5 based on the order in which they were cloned. The M_1 , M_3 , and M_5 stimulatory receptors couple primarily to the mobilization of intracellular calcium. The increase in cytoplasmic calcium occurs following stimulation of a signaling cascade which begins with coupling to the heterotrimeric G protein, Gq, followed by stimulation of the effector enzyme, phospholipase C, which releases inositol-1,4,5-trisphosphate (IP₃). The M_2 and M_4 inhibitory receptors negatively modulate adenylate cyclase to reduce cytoplasmic levels of cAMP.

Competitive antagonists are thought to bind to the same critical residues facing the binding pocket but appear to also interact with additional determinants which influence their affinity for each receptor. For example, mutation of Ser120 to Ala resulted in a loss of antagonist binding affinity with little effect on agonist binding.²⁸ Moreover, replacement of Asn507 with Asp, Ser, or Ala resulted in a differential loss of antagonist affinity based on the structural class of compound.²⁹

Signal Transduction. The physiological response to muscarinic receptor stimulation is mediated through a cascade of protein interactions including G protein activation followed by stimulation or inhibition of an effector channel or enzyme.8 Tissue localization, pre- or postsynaptic location, complement of signal transduction machinery contained in the cell, and receptor subtype expression pattern influence the biological consequence of muscarinic receptor stimulation. The functional activity of M₁, M₃, and M₅ receptors is most commonly associated with their ability to mobilize intracellular calcium through stimulation of phospholipase C β (Figure 2). Phospholipase C β activation releases IP₃ which stimulates the IP₃ ion channel receptor to release Ca2+ from intracellular compartments resulting in a transient cytosolic calcium flux. The M2 and M₄ receptors inhibit adenylate cyclase activity reducing the local concentrations of cAMP (Figure 2). They also block the activity of voltage-operated calcium channels involved in neurotransmitter release. More recently, the signal transduction pathways associated with muscarinic receptors have expanded to include more effectors and to include pathways normally thought to interact exclusively with growth factors.8

In addition to their ability to stimulate phospholipase C through Gq, the M₁, M₃, and M₅ receptors have also been shown to stimulate phospholipase A₂³⁰⁻³³ and phospholipase D³⁴⁻³⁸ in various cell lines and primary cultured cells. Phospholipase A2 or D activation has a common requirement for extracellular calcium. A cation channel mediating muscarinic receptor-dependent calcium influx has been initially characterized and may provide the regulatory step essential for phospholipase activation.^{8,36} These studies showed that the muscarinic receptor-operated calcium channel is voltage-independent, has a low conductance, and in the cell lines studied may be under the regulation of a small-molecularweight G protein. Expression studies with a mammalian homologue of the *Drosophila* Trp/Trpl cation channels indicated that the Trp6 channel was under the regulation of the M₅ receptor.³⁹ Receptor-operated calcium channels may be a key signaling pathway for other calcium-mobilizing receptors. Additional signal transduction activation has been associated with the M_1 , M_3 , and M₅ receptors including the inefficient stimulation of adenylate cyclase through both Gs and G proteinindependent pathways. 40-43

The M₂ and M₄ receptors couple primarily to the inhibition of adenylate cyclase but also interact with Gs to stimulate adenylate cyclase. 40-42,44 Pertussis toxin treatment, which blocks Gi/Go function, increases the likelihood of Gs and muscarinic receptor interaction suggesting that G protein stoichiometry within the cell plays a key role in directing the signaling potential of each receptor. M₂ and M₄ receptors stimulate phospholipase C in some cell lines which is thought to be mediated in part by $\beta\gamma$ -subunits released following heterotrimeric G protein activation. 45 A novel signaling pathway involving the activation of phospholipase A2 by Gi-coupled receptors, including the M₂ and M₄ receptors, is stimulated when phospholipase A₂ is preactivated either directly or with calcium-mobilizing agonists.⁴⁶ In this study, M₂ and M₄ receptors augmented purinergic receptor-stimulated phospholipase A2 activation. This may be physiologically relevant since the purinergic receptor agonist, ATP, is co-released with many neurotransmitters, including acetylcholine, and may serve to prime arachidonic acid mobilizing or other signal transduction pathways.

The regulation of muscarinic receptor function and trafficking by kinase enzymes has been extensively studied and has been recently reviewed. 47,48 Muscarinic receptors regulate kinase and phosphatase activity either through heterotrimeric G protein activation and subsequent second-messenger production or through the direct activation of small-molecular-weight G proteins. 49-53 Hence, pathways normally thought to be mediated by growth factor receptors have been linked to GPCRs in ways that are just beginning to be appreciated. Many of the recent discoveries of muscarinic receptor signaling have been initially observed in ectopic expression models using mammalian cell lines. For the most part, however, signal transduction observed in cell lines has also been observed when receptors are studied in primary culture or tissue slice preparations. Due to the large overlap in expression patterns between the five muscarinic receptors in brain, it has been difficult to ascribe a specific signaling pathway to a single receptor subtype without selective pharmacological agents.

Pharmacology

Sources of Muscarinic Receptor Subtypes. Before the cloning of the muscarinic receptors, isolated tissues served as sources of putative pure muscarinic receptor subtype responses for many decades. Superior cervical ganglion has been used as a predominant M₁ receptor functional model^{54,55} as well as vas deferens.⁵⁶ Guinea pig bladder has served for M3 receptor responses, ⁵⁷ and cardiac tissue, especially isolated atria, as a source of M₂ receptor.⁵⁸ The M₅ receptor is localized primarily in the substantia nigra and VTA.59 However, localization of the M₅ receptor in other brain regions and periphery has been controversial.⁶⁰ This is due to its low expression and lack of tools for its positive identification other than by PCR, which provides identification of mRNA but no quantitative measure of protein levels. Only one source of discrete M₅ receptor expression has been described to date in a human melanoma cell line, A2058.⁶¹ In this cell, M₅ receptors uncharacteristically coupled to the inhibition of adenylate cyclase. Pharmacological approaches have also been applied to M₅ receptor characterization and localization and suggest that the distribution may be wider than previously thought. 60 Cloning and ectopic expression of human and rat M₁-M₅ muscarinic receptor cDNA in cell lines has provided a source of pure receptor subtype for binding and functional studies. Considerable information has accumulated using these cells about the molecular structure, pharmacology, and signal transduction of each subtype. The recent challenge has been to correlate data derived from the molecular studies to tissue and in vivo pharmacology.

The recent creation of mice bearing a genetic deletion for the M_1 , M_2 , and M_4 receptors has provided an opportunity to investigate physiological responses to ligand challenge in animals and tissues devoid of one or more receptor subtypes. For example, M_1 receptor knockout mice lost their seizure response to pilocarpine suggesting that M_1 receptors play a role in the etiology of epilepsy. Esuperior cervical ganglion cells isolated from M_1 knockout mice no longer displayed M current inhibition following application of a muscarinic agonist verifying its predominant functional M_1 receptor content. Spontaneously beating isolated atria normally

show a reduction in the number of beats per unit time following addition of a muscarinic agonist to the bath media. M₂ knockout mice did not respond to muscarinic agonist application indicating that regulation of heart rate is predominantly regulated through the M2 receptor. 63 However, a more complex regulation is likely to occur in vivo and includes central and ganglionic involvement. M₂ receptors were also shown to play a major role in the regulation hypothermia, tremor, and analgesia in mice. Remarkably, M₄ knockout mice respond similarly to wild-type mice following muscarinic agonist-induced analgesia, tremor, salivation, or hypothermia. M₄ knockout mice were hyperactive and hyperresponsive to D₁ dopamine agonists, amphetamine, and apomorphine suggesting a role in regulating dopamine release and dopamine-mediated locomotor activity. 64 Using M₁, M₂, and M₄ knockout mice, Hille and colleagues were able to show that N and L type voltagegated calcium channels were differentially regulated by the muscarinic subtypes in mouse superior cervical ganglion neurons. 65 They found that the fast regulatory pathway was M₂ receptor-mediated, the slow secondmessenger-dependent pathway was M₁-mediated, and neither was regulated by M₄ receptors. Additional studies with M₃ and M₅ single knockout and combinations of double-muscarinic receptor knockout mice will no doubt enhance our understanding of the physiological role of each member of this receptor family.

Subtype-Selective Muscarinic Agonists. The development of muscarinic receptor subtype-selective compounds has been challenging, and on detailed examination the relative selectivity has been limited. This review will concentrate on compounds that have appeared more recently in the literature and for which some claim for muscarinic subtype selectivity has been mentioned. ^{66,67}

A popular strategy for improving both metabolic stability and muscarinic subtype selectivity has been to replace the ester group of the muscarinic agonist arecoline (1) or conformationally more rigid azabicyclic analogues with bioisosteric heterocycles. For instance, evaluation of pyrazole analogues 2 detected compounds that show M_3 agonist selectivity compared to M_1 agonist activity in tissues. ⁶⁸ By contrast, Lu 25-109 (3) in which a tetrazole replaces the ester group of 1 shows partial M_1 agonist activity in tissues and in an M_1 cell line but M_2 and M_3 antagonist activity in tissues. ⁶⁹

Among a series of azabicyclic pyrazines $\bf 4$ is a partial M_1 -selective agonist that has 5-fold greater M_1 efficacy than the selective M_1/M_3 agonist L 689-660^{70,71} (5), less M_3 agonist activity than $\bf 5$, and no M_2 agonist activity in the guinea pig atrium. Among a series of 1,2,5-thiadiazolyl azabicyclics that have muscarinic activity, LY297802 (6) is found to produce potent muscarinically mediated antinociceptive activity. While $\bf 6$ is a M_1 agonist in the rabbit vas deferens, M_2 and M_3 antago-

nism or weak partial agonism was demonstrated in other tissues. Subsequently, studies using other muscarinic agonists lacking M_1 agonist activity in tissues and M_1 cell lines showed that M_1 agonist activity is not required for antinociception. However, structurally related PTAC (7) shows muscarinic antagonist activity in M_1 , M_3 , and M_5 cell lines but partial agonist effects in M_2 and M_4 cell lines. Functional dopamine antagonist activity and antipsychotic-like activity could be demonstrated with 7 in several different tests, but 7 has no affinity for dopamine receptors suggesting a potentially new therapeutic use for muscarinic agonists that may be mediated through M_2 or M_4 receptor subtypes.

Restricting the rotation of the heterocyclic ester bioisostere within this structural class has not dramatically improved muscarinic subtype selectivity. Rigid arecoline analogues $\bf 8a-c$ show some M_2/M_4 selectivity when compared across muscarinic receptor subtype cell lines using R-SAT technology, 78 but the selectivity appears to be an artifact of the test system since standard muscarinic agonists show similar selectivity. 79 In tissues, the 8-methyl derivative $\bf 8d$ shows some functionally selective partial M_1 agonist activity. 80 The fused heterocyclic analogues of the muscarinic analgesics such as $\bf 9$ and $\bf 10$ do not show improved muscarinic subtype selectivity compared to $\bf 11a$, nor is functional selectivity observed in receptor subtype clones. 81

Replacement of the tetrahydropyridine ring of arecoline analogues with a tetrahydropyrimidine as in $\mathbf{12a,b}$ and $\mathbf{13}$ provides selective M_1 efficacy in cell lines compared to efficacy in M_2 and M_3 cell lines.⁸²

Because oximes have electrostatic properties similar to esters, an alternative strategy of replacing an ester group with an oxime has been frequently used in muscarinic agonist research. On the basis of binding studies in tissues, tropinone oxime 14 appears to have some selectivity for M_1 receptors over M_2 and M_3 receptors, but the compound did show functional ago-

nism in an M_3 preparation but not in an M_2 assay.⁸³ CI-1017 (**15**) has 8-fold greater affinity for M_1 receptors compared to M_2 receptors as well as functional selectivity at M_1 and M_4 receptors compared to M_2 , M_3 , and M_5 receptors using both second-messenger assays and R-SAT technology.⁸⁴ Additional promising oximes from this series have also been described.⁸⁵ Sabcomeline (SB 202026, **16**) does not show selective affinity for muscarinic subtypes, but it is a potent and efficacious M_1 agonist in the rat cervical ganglion while in other tissues and in vivo assays it is a much less potent and efficacious M_2 and M_3 agonist.^{86,87}

Rigid analogues of acetylcholine have also been pursued as a strategy for obtaining muscarinic subtype selectivity. A relatively older example that has seen continued development until very recently is talsaclidine (17), an analogue of aceclidine (18).88 Talsaclidine shows only minor differences in affinity among the M₁-M₅ receptor subtypes. However, in functional assays, 17 is most potent and efficacious in cells expressing M1 receptors, slightly less in M3 cells, and inactive in M2 cells. Tissue assays also suggest reasonable M1 selectivity. Nortropane (19), has 12-fold higher affinity for M2 receptors compared to M4 receptors but much higher selectivity for M₂ receptors compared to M₁ and M₃ receptors.⁸⁹ Functional assays demonstrate that 19 is an agonist at M₁-M₄ receptors but only a partial agonist at M₃ receptors. WAY-131256 (20), a rigid carbamate analogue of acetylcholine, does not show selective affinity among the M_1-M_5 receptor subtypes, and **20** is comparably potent agonist at both M1 and M2 receptors. 90 By contrast, in vivo studies suggested that 20 is selective for M₁ receptors compared to M₂ and M₃ receptors. Thiadiazole ester bioisosteres of aceclidine (21a,b) are as comparably efficacious and potent as the functionally selective M₁ agonist xanomeline (11b) in an M₁ cell line and have very little M₃ activity in vivo.⁹¹

Oxotremorine analogues $\bf 22a,b$ have selective affinity for M_1 receptors in rat brain compared to M_2 receptors in rat heart, and binding index studies suggest that $\bf 22a$ is a partial muscarinic agonist while $\bf 22b$ is a full agonist. Where, subtype-selective analogues of pilocarpine ($\bf 23$) have not been revealed but thiopilocarpine (SDZ-ENS 163, $\bf 24$) continued into clinical trials. SDZ-ENS 163 acts in vitro and in vivo as a partial agonist at M_1 and M_3 receptors but as an antagonist at M_2 muscarinic receptors. However, $\bf 24$ did not distinguish between m_1 and m_3 receptors in radioligand binding studies.

Some compounds seemingly unrelated to classical muscarinic agonists have shown some hint of muscarinic subtype selectivity. For instance, certain small-molecular-weight proteins in snake venoms appear to have high selective affinity for M₁ receptors compared to M₂ receptors, and they also stimulate M₁ receptors containing tissues without producing effects on M2 and M3 receptor-mediated tissues. 95,96 The antipsychotic clozapine, which has high muscarinic affinity but only modest M₁ subtype selectivity, was found to be a potent agonist in M₄ cell lines but an antagonist in the other muscarinic receptor subtype cell lines.⁹⁷ Subsequently, a few, but not all, antipsychotic agents have also been found to have M₄ agonist activity in cells expressing the receptor, but they did not modulate cAMP levels in tissue.98 This result is consistent with a weak partial agonism that is most evident in cell lines expressing high levels of receptor. This is supported by more recent evidence showing that clozapine behaved as a weak partial agonist at M₁-M₄ receptors expressed in cell lines. 99 Although perhaps inspired by the M₁ agonist R46559,¹⁰⁰ SK-946 (**25**) is a structurally unique diamine that has selective affinity for M₁ receptors compared to M₂ receptors in rat brain and which stimulates M₁ receptors in rat fetal hippocampal neuronal cells. 101,102 Two seemingly unique classes of amidinoquanidines **26**¹⁰³ and amidines **27**¹⁰⁴ have also been recently disclosed that may have some functional muscarinic subtype selectivity.

The cholinergic hypothesis of dementia and Alzheimer's disease and the marginal clinical success of acetylcholinesterase inhibitors continue to support the development of selective M₁ agonists for the treatment of these diseases. However, none of the functionally selective M₁ agonists reported to be in clinical trials have emerged into clinical practice suggesting that considerably more effort is needed to adequately test the M₁ cholinergic hypothesis. More recently, the potential use of selective agonists in the management of pain and in the treatment of schizophrenia has also gained attention. Although the muscarinic subtypes associated with these treatments in humans is not yet well-resolved, the M_2 and M_4 subtypes seem to be highly implicated. The therapeutic uses of M₃-selective muscarinic agonists remain unrealized although the predominance of M₃ receptors on smooth muscle and

exocrine glands suggests treatments for dysfunction in these tissues. The potential therapeutic uses of M₅selective agonists also remains an unknown especially because of the low abundance of this receptor in the brain and the absence of a pharmacological defined test for this subtype. In all cases, the development of potent and efficacious, molecularly defined, subtype-selective agonists would be of great benefit in clarifying and realizing the therapeutic potential of muscarinic ago-

Subtype-Selective Muscarinic Antagonists. The methods used to determine the muscarinic subtype selectivity of antagonists have been no more consistent than for muscarinic agonists. Most of the caveats used in assessing muscarinic agonist subtype selectivity should be equally applied to claims for muscarinic antagonist subtype selectivity. Additional factors to consider with muscarinic antagonists are their potentials to not only act as antagonists but to also act as allosteric modulators of muscarinic receptors. 105 Among the therapeutic uses suggested for subtype-selective antagonists, the treatment of peripheral smooth muscle disorders such as bladder, airway, and bowel disorders with M₃ antagonists has been particularly of interest, while antagonists of M2 and M4 receptors have been suggested as treatments for movement disorders, dementia, cardiac disorders, and pain.

Certain arylalkylamines have shown selectivity for M₃ receptors in tissues, and three of these (28-30) have been evaluated clinically as potential treatments for irritable bowel syndrome, obstructive airway diseases, and urinary incontinence, respectively. Radioligand binding studies in tissues suggests that 1 has higher affinity for M₃ receptors than for M₁, M₂, or M₄ receptors. 106 Functional tissue assays confirm that **28** is selective for M₃ receptors compared to M₁ and M₂ receptors, but the magnitude of the selectivity is dependent on the particular M₃ tissue preparation used. While 29 shows 4-fold functional selectivity for inhibition of muscarinic receptors in the urinary bladder compared to muscarinic receptors in the parotid glands in vivo, this selectivity cannot be attributed to activity at a single muscarinic receptor subtype. 107-109 In cloned human muscarinic (M_1-M_5) receptors, **29** does not discriminate among the subtypes in binding experiments nor are there large differences in affinity for muscarinic receptors in specific tissues (M₁-M₃). A similar profile is presented by the major metabolite of **29**. 109 Comparisons of muscarinic antagonists with different in vivo and in vitro selectivities led to the conclusion that a balance between M2 and M3 affinity was required for bladder selectivity. Darifenacin (30) not only is a more potent antagonist in tissues controlled by M₃ receptor subtypes compared to tissues controlled by M₁ and M₂ receptor subtypes but also appears to be selective for M₃ receptors in smooth muscle preparations compared to M₃ receptors in glandular tissue. ¹⁰⁷ More recently, the M₃ selectivity of **30** was confirmed in binding experiments with cloned human muscarinic receptor subtypes where 30 shows a 47-fold separation between M_2 and M_3 affinity. 106,110

Arylalkylamine NPC-14695 (31) also shows higher affinity for tissue M3 receptors than for the M2 or M1 receptor.¹¹¹ The observed 11- and 37-fold separation

between bladder function and mydriasis and salivation, respectively, also suggests selectivity for smooth muscle M₃ receptors in vivo. High selectivity for bronchial smooth muscle muscarinic receptors compared to muscarinic receptors in the heart and salivary gland is also seen with **31**. Vamicamide (**32**) shows similar potency as an M₁ and M₃ antagonist in tissue preparations but is less potent in M2 tissue. 112 The ability of 32 to inhibit bladder contractions yet have little effect on heart rate and poorly penetrate the blood-brain barrier suggests that 32 might be useful for the treatment of bladder hyperactivity. KRP-197 (33a) has subnanomolar affinity for M₃ receptors in the ileum, comparable affinity for M₁ receptors, but 13-fold lower affinity for M₂ receptors in the atrium and is also suggested to be a potential treatment for urinary incontinence. 113,114 Structurally related imidazole 33b has somewhat lower M3 affinity than 33a but has higher selectivity for M3 receptors than 33a. In contrast to other compounds within this structural class, the S-isomer of commercially available antihistamine dimethindine (34) exhibits higher affinity for M2 receptors than for M1, M3, or M4 receptors in tissues in both functional and binding assays. 115

Complex esters of amino alcohols have been shown to have extremely varied muscarinic subtype-selectivity profiles. For instance, like many of the arylalkylamines, quinuclidinyl tropate 35 shows selectivity for M₃ receptors in functional assays in tissues with 4-fold selectivity over M₄ receptors but greater separations between M₃ and M₁ or M₂ activity. The M₃ subtype selectivity of **35** is also supported by binding experiments in cloned receptors (M_1-M_4) where **35** has 10-fold higher affinity for M₃ receptors than for M₄ receptors. Tissue binding assays suggest that LG50643 (36) has greatest affinity for M₃ receptors, 2.7-fold greater than for M₁ receptors, and 15-fold greater than for M_2 receptors. ¹¹⁷ In trachea

preparations, functional M₃ versus M₂ selectivity was demonstrated by showing a 7-fold separation between postjunctional and prejunctional potency. In contrast, S-ET-126 (37) is functionally an M₁ antagonist in tissues with 8-fold selectivity over M₂ activity and even less M₃ antagonist activity. 118 Extensive molecular modification of 2-ethylthio diphenylacetate 38 did not lead to improved M₂ selectivity in functional tissue assays. 119 Highest M₂ selectivity was usually found among quaternary derivatives. Binding studies in cloned muscarinic receptor subtypes have not confirm the M₂ subtype selectivity of 38.120 Tropane ester PG-9 (39) produces antinociception in several standard tests and is highly potent as an antagonist in the putative M₄-selective guinea pig uterus assay. 121 In tissues selectively controlled by M₁-M₃ receptors, **39** is at least a 10-fold less potent antagonist. Microdialysis experiments show that **39** increases release of acetylcholine in the rat cerebral cortex within the antinociceptive dose range suggesting that nociception produced by 39 may be mediated through inhibition of presynaptic M₄ receptors.

Carbamates of many of these structurally related amino alcohols also show M₃ selectivity. YM-58790 (40) has high affinity for both M₁ and M₃ receptors in tissue binding assays but 11-fold lower affinity for M2 receptors. 122 In vivo, **40** is distinguished from other members of a series by the 6.7-fold separation between antagonism of M₃-mediated responses in the bladder and salivary glands as well as a lack of blockade of M2mediated bradycardia. YM-46303 (41) has higher muscarinic affinity than 40 but is somewhat less selective in vivo. 123 YM- 53705 (42) has M_3 subtype affinity comparable to that of **40** but greater separation (10-fold) between effects on the bladder and on salivary glands. 124

Tripitramine (43) incorporates structural elements of the two M₂-selective muscarinic antagonists methoctramine¹²⁵ and AQ-RA 741. 126,127 In functional assays in tissues, 43 blocks M2 receptors 1600-fold better than M₃ receptors and in tissue binding assays is equally selective for M₂ over M₃ receptors. The order of affinity for the muscarinic subtypes is $M_2 > M_1 > M_4 > M_3$ with only a 2-fold selectivity between M₁ and M₄ receptors. In cloned human muscarinic receptors, 43 is selective for M₂ receptors, but the magnitude of the selectivity between M₂ and M₁ receptors (6-fold) is significantly less than that seen in tissue binding experiments. 128 By contrast, dipitramine (44) has highest affinity for M₁ receptors in the M₁-M₄ tissue binding assays but there is only a 5-fold separation between M₁ and M₂ affinity. 127 Spirotramine (45) has lower affinity for M₁ receptors than **44**, but **45** does show higher M₁ selectivity than **44** in tissue binding assays. 129 Recently, additional structure-activity studies were reported in this structural class, but clearly superior subtype selectivity was not obtained. 130

Dibenzodiazapenes continue to provoke interest as potential subtype-selective muscarinic antagonists, particularly as M2 antagonists. Both YM59981 (46) and YM55758 (47) have much higher M₂ affinity than the prototypical M2-selective antagonist AF-DX 116 (48) in rat heart binding assays, and they also have greater separation (>40-fold) between M₂ and M₃ affinity. ¹³¹ Modest separation between M₂ and M₁ affinity (4- and 6.3-fold, respectively) is also seen. Both 46 and 47 are distinguished within their chemical series by high oral activity and poor brain penetration, advantageous properties for peripherally acting cardiac agents. Similarly, succinamides YM43571 (49) and YM47244 (50) also have higher M₂ affinity and selectivity than **48** and better M_2/M_1 and M_2/M_3 selectivity (>10 and >79, respectively) than 46 and 47.132 A large separation between M₂- and M₃-mediated effects is also seen in vivo with **49** and **50**, but the M₂ effects are noncompetitive and there is some concern about the oral bioavailability of the compounds. An earlier succinamide **51** has somewhat lower M₂ affinity and M₂/M₃ selectivity than **49** and **50** and comparable M_2/M_1 selectivity (16-fold), but the in vivo M2 antagonist activity is also noncompetitive. 133 Attempts to develop selective M_2 antagonists that cross the blood-brain barrier as potential dementia treatments led to phenylacetamides 52a,b.134 Both **52a,b** have higher affinity for M₂ receptors than **48** in tissue binding assays, and both show 7-fold higher affinity for M_2 receptors than for M_1 receptors. In contrast to earlier M2-selective compounds, 52b does penetrate the blood-brain barrier. Desaza analogues

53 and **54** of the M_1 -selective antagonist pirenzepine (**55**) that have a greater potential for blood—brain barrier penetration have also been prepared.¹³⁵ While **53** has higher affinity for cloned M_1 receptors than **55**, the M_1 selectivity of **53** is no better than that for **55**. Both **54** and **55** had comparable M_1 affinity, but the M_1/M_4 selectivity of **54** (10- fold) was higher than for **55** (4-fold).

Both **56a**,**b**, silicon-containing analogues of the relatively M_1 -selective antagonist piperiden (**57**), have essentially the same subtype affinity and order of subtype selectivity ($M_1 > M_4 > M_3 > M_2$) as **57**. 136 Among a series of stable, enantiomerically pure homologues of silatricyclamol iodide **58**, (R)-**59** has the highest M_1 affinity and selectivity in functional assays but the M_1 affinity and selectivity in binding assays is less. 137 Unsaturated analogues **60a**,**b** of the relatively M_3 -selective antagonists hexahydrosiladifenidols **61a**,**b** have higher affinities but lower receptor subtype selectivity than the parent compounds. 138

A variety of miscellaneous structures have also been found to have muscarinic antagonist subtype selectivity. The tetrahydropyridine **62** that is a heterocyclic ana-

logue of a number of muscarinic agonists has modest selectivity for M₁ receptors compared to M₂ receptors in tissue binding assays. 139 A number of similar heterocyclic analogues have been described as muscarinic antagonists, but there is insufficient data for determining if they have subtype selectivity. 140 Reduction of the furanone ring of himbacine (63) gives compounds with diminished M2 subtype affinity but which retain some selectivity compared to M₁ subtype affinity.¹⁴¹ A number of other modifications of himbacine that may lead to subtype selectivity have appeared in abstract form. 142 Liriodenine (64) is 10-fold more potent in inhibiting muscarinic-induced contractions in trachea and ileum than in atrial preparations suggesting some potential for M₃ subtype selectivity. ¹⁴³ 3-α-Chloroimperialine (**65**) is a potent muscarinic antagonist that has 6-fold higher affinity for cloned M₂ receptors than M₄ receptors and even greater selectivity compared to M1 and M3 receptors. 144 Isoquinoline 66 does not have particularly high affinity for cloned muscarinic receptors but did show 10fold selectivity for M4 receptors compared to M3 receptors and even greater selectivity compared to the other muscarinic subtypes. 145 Functional assays in muscarinic subtype cell lines confirmed that 66 shows highest potency in blocking muscarinic agonist-induced responses in an M₄ cell line, but **66** is only 10-fold less potent in blocking similar effects in an M₂ cell line. Certain snake toxins have also been demonstrated to selectively bind to and antagonize responses at the M₄ receptor subtype. 146-149

Subtype-Selective Allosteric Modulators. The development of muscarinic agonists and antagonists with high degrees of subtype selectivity has remained relatively elusive probably because of the close sequence homology within the ligand-binding pocket among the muscarinic receptor subtypes. The development of subtype-selective allosteric modulators may be a more promising prospect because of the much more varied sequences in the extracellular domains of the muscarinic subtypes where the allosteric modulators are presumed to bind to the receptor. 150 Another potential advantage of allosteric modulators is the potential for either enhancement or diminution of a particular subtype modulatory effect by acetylcholine or an exogenous muscarinic agonist or antagonist. The magnitude of any increase or decrease in subtype activity could also be controlled by the magnitude of cooperativity produced by a particular allosteric modulator. Regardless of these potential advantages, the number of reports of allosteric modulators conferring muscarinic subtype selectivity remain relatively sparse and the potencies and additional activities of most of these modulators suggest that they are only suitable as lead structures.

The rank order of allosteric-mediated negative cooperativity for N-methylscopolamine (NMS) binding in M₁-M₃ receptors with the cardiac-selective muscarinic antagonists gallamine (67), methoctramine (68), AF-DX116 (**69**), and himbacine (**70**) is $M_2 > M_1 > M_3$. ¹⁵¹ A similar rank order of negative cooperativity is seen with the ion channel blockers verapamil (71), PCP (72), quinidine (73), and secoverine (74). 151

A number of neuromuscular blocking agents have been found to produce differential modulation of muscarinic subtypes through allosteric interactions. Alcuronium (75) increases the binding of NMS to M_2 and M₄ receptors but inhibits the binding to M₁, M₃, and M₅ receptors. 152 Brucine (76) increases the affinity of acetylcholine for M₁ and M₃ receptors but produces different patterns of affinity augmentation at receptor subtypes with other muscarinic agonists. 153 Other workers also found that acetylcholine affinity for M₁ receptors is increased almost 2-fold by 76, while M3 receptor affinity is increased 3-fold by the brucine derivative 77 and 2.6-fold by 78.154 These compounds produce no increases or less of an increase in affinity for acetylcholine at the other muscarinic receptor subtypes. By contrast, all three compounds and strychnine (79)¹⁵⁵ increase NMS binding at M₂ receptors by at least 2-fold while smaller increases in affinity are seen at the other receptor subtypes. N-Substituted strychnine analogues show various patterns of positive and negative cooperativity with NMS binding, but none of these derivatives increase acetylcholine binding. 154

Eburnamonine (80) augments the binding of acetylcholine to M₂ and M₄ receptors and produces positive cooperativity at M2 receptors with other muscarinic agonists. ¹⁵³ Vincamine (**81**) produces positive allosteric effects with some agonists at some subtypes. 153 Recently, a second distinct allosteric site sensitive to staurosporine and related indolcarbazoles has been described for M_1-M_4 receptors which is distinct from the site which binds strychnine-related compounds. 156 This opens up additional avenues for drug development based on allosteric modulation of muscarinic receptors.

The therapeutic potential of allosteric modulators should span all of the potential uses of selective agonists and antagonists since the therapy would only depend on the subtype being addressed and the degree and direction of the cooperativity produced by the modulator. Clearly, the work on allosteric modulators of muscarinic receptor subtypes remains in relative infancy compared to work toward subtype-selective muscarinic agonists and antagonists. Progress in the development of subtype-selective modulators should benefit from the application of broad screening techniques that are now pervasive in the pharmaceutical industry.

Schizophrenia

Schizophrenia is a severe psychosis afflicting more than 1% of the population worldwide, and about 100 000 new cases are diagnosed each year. The predominant symptoms of schizophrenia are classified as positive (hallucinations, delusions, vocal outbursts, hearing of voices), negative (attention deficits, social withdrawal, poverty of thought and speech, anhedonia, anergia), and cognitive (deficits in short term and executive memory). Classical neuroleptics, such as haloperidol (Haldol) and chlorpromazine (Thorazine), are effective in treating the positive symptoms of the disease and were discovered based on their ability to block dopamine receptors. 157 However, these compounds induce side effects such as extrapyramidal symptoms (EPS) and hyperprolactinemia. Newer "atypical" antipsychotics, such as clozapine and olanzapine, are effective against both positive and negative symptoms, have fewer tendencies to cause EPS and hyperprolactinemia, and therefore provide a significant improvement over previous treatments. However, these agents require several weeks before their therapeutic effects are fully realized and are relatively ineffective against cognitive deficits. Moreover, clozapine

can induce a potentially fatal blood disease, agranulocytosis, in a small percentage of patients and therefore requires constant blood monitoring during treatment. Alternative targets for therapeutic intervention have been proposed including serotonin, dopamine, glutamate, and muscarinic receptors in an attempt to provide drug therapies for treatment-resistant patients, increase the speed of therapeutic benefit, and improve the cognitive symptoms.

Muscarinic receptors are particularly compelling as therapeutic targets for the treatment of schizophrenia based on evidence that muscarinic antagonists produce a psychosis in humans with symptoms similar to the positive and negative behaviors and cognitive deficits associated with the disease. 158-162 Hallucinations are one of the most debilitating components of schizophrenia and degenerative neurological diseases. Certain drugs and substances of abuse can induce hallucinations in humans suggesting that receptors for these compounds may play a role in generating hallucinations and may be attractive therapeutic targets to treat these debilitating symptoms. For example, ketamine and phencyclidine (NMDA receptor ligands), muscimol and β -carbolines (GABA receptor ligands), LSD (serotonin receptor ligand), and cocaine (dopamine transporter ligand) induce hallucinations that resemble those seen in neurological diseases. Cholinergic antagonists induce a psychosis that includes delusions and hallucinations which are most likely mediated through muscarinic receptors since nicotinic receptor antagonists, such as mecamylamine, are not inherently hallucinogenic. In particular, the muscarinic antagonists, quinuclidinyl benzilate (QNB), scopolamine, and atropine induce a psychosis in humans that includes schizophrenic-like auditory and visual hallucinations. 158,159,161,162 At low doses, the antagonists induce visual hallucinations that involve clearly defined objects or people and auditory hallucinations that include music and voices that inspire dialogue. At higher doses of muscarinic antagonists, however, the hallucinations progress to severe delusions and loss of awareness. Patients with a previous history of psychosis, those suffering schizophrenia, and Parkinson's patients being treated with anticholinergics are particularly susceptible to muscarinic antagonistinduced hallucinations. 161,163,164

The pathophysiology of schizophrenia is thought to be related to a relative increased dopaminergic neurotransmission.¹⁶⁵ This hypothesis is supported by pharmacological evidence that agents that stimulate the dopamine system induce the positive symptoms of schizophrenia and antidopaminergics relieve these symptoms. 157,166 Interest in cholinergic treatment of schizophrenia predates the dopamine hypothesis.¹⁶⁷ Early clinical studies with cholinergic agonists 168,169 and the acetylcholinesterase inhibitor physostigmine^{170,171} suggested that cholinergic receptor stimulation improved the psychotic symptoms of schizophrenics. However, this approach had limited acceptance due to the poorly tolerated cholinergic side effects associated with these nonselective agents. An understanding of the relationship between the cholinergic and dopaminergic system was emerging which suggested that they were normally in balance and reciprocally regulated each other. It was speculated that in schizophrenia, an imbalance existed between the dopamine and acetylcholine system that could possibly be corrected with muscarinic agonist therapy. ^{171,172} Since the dopamine and muscarinic systems reciprocally modulate each other, and since muscarinic antagonists induce the positive symptoms of schizophrenia, it follows that muscarinic agonists may be useful in reducing the positive symptoms associated with schizophrenia and other psychosis.

Additional evidence has accumulated supporting the hypothesis that muscarinic agents may be useful in the treatment of psychosis based on an interaction with the dopamine system. Pharmacological evidence showed that the dopamine and muscarinic systems reciprocally modulate each other. 173-175 Muscarinic agonists or cholinesterase inhibitors were shown to depolarize neurons expressing dopamine receptors¹⁷⁶ and increase dopamine release in the nucleus accumbens when injected near the VTA.¹⁷⁷ Dopamine release may be modulated through a muscarinic heteroreceptor on dopaminergic neurons with pharmacological properties of an M₁ receptor. 178-180 More recently, xanomeline, which has a mixed partial agonist and antagonist profile at the muscarinic receptor subtypes, reduced psychotic symptoms of patients with Alzheimer's disease including hallucinations and delusions. 181 A chemically related muscarinic compound, PTAC, with partial M₂/M₄ agonist and M₁/M₃/M
₅ antagonist properties was suggested as a potential treatment for schizophrenia based on preclinical data with similarities to atypical neuroleptics such as clozapine and olanzapine. 76 PTAC inhibited conditioned avoidance responding in rats without inducing catalepsy seen with similar doses of haloperidol suggesting that its action is not due to nonspecific motor effects. PTAC also blocked apomorphine-induced climbing in mice suggesting that its mechanism of action involved modulation of the dopamine system. Moreover, in in vivo electrophysiological experiments, PTAC inhibited the number of spontaneous firing A10 dopamine cells in the VTA relative to the substantia nigra but had little if any effect on the firing of substantia nigra A9 dopamine cells suggesting low EPS liability. In contrast, haloperidol blocks the spontaneous firing of both A9 and A10 cells which may contribute to the EPS seen with this compound. Atypical neuroleptics with little motor side effects such as clozapine and olanzapine also do not appreciably affect A9 cell-firing rates. PTAC, however, inhibited A10 cell firing after both acute and chronic treatment suggesting that it may have a rapid onset of action. PTAC had no affinity for either dopamine or serotonin receptors suggesting it has a unique mode of action through muscarinic receptors.

Identifying the muscarinic receptor subtype(s) associated with dopamine regulation is a challenging problem since all five muscarinic receptors are found in brain areas rich in dopamine neurons. However, some differences in receptor distribution have been observed for both mRNA and protein. Of the five known muscarinic receptor subtypes, M_1 and M_4 receptors are most abundant in forebrain areas thought to be involved in schizophrenia (cerebral cortex, basal ganglia, hippocampus). 182 M_1 and M_4 receptors in particular are colocalized with dopamine receptors in caudate-putamen and cerebral cortex. 183,184 More recently, mice lacking the M_4 receptor were hyperactive and hyperresponsive

to DA_1 agonists suggesting a role for the M_4 receptor in the tonic suppression of the dopamine system. 64

The M₂ receptor follows an opposite distribution pattern with highest densities in brainstem and in lower abundance than M₁ and M₄ in the forebrain. M₂ receptors are located on cholinergic interneurons of the striatum and tegmental cholinergic neurons and are considered to be the predominant autoreceptor regulating acetylcholine release. 184,185 It has been suggested that the psychotogenic actions of antimuscarinic agents may be mediated predominantly through the M2 autoreceptors located on Ch5 and Ch6 cholinergic tegmental neurons. 186 The Ch5 and Ch6 neurons project to dopamine-rich areas of the VTA and substantia nigra and are the only known cholinergic inputs to these areas. $^{187-190}$ In support of this proposal, postmortem brain tissue obtained from schizophrenics was found to have a higher density of Ch5 and Ch6 cholinergic fibers. 191 It should be noted that some investigators have not found changes in cholinergic neurons of schizophrenic brain tissue. 192 M₂ receptors are also widely distributed on noncholinergic cells, 183,193 and their function on these cells is unknown.

The M₅ receptors are predominantly localized to the substantia nigra pars compacta and VTA and are thought to be the receptor responsible for acetylcholinemediated dopamine release. It has been suggested that it is the only muscarinic receptor subtype co-localized with D₂ receptor mRNA in the substantia nigra and VTA.183 Recent studies using RT-PCR and a pharmacological labeling approach suggest that the M₅ receptor is more widely distributed than previously considered. 60,194 M_{5} receptor distribution is distinct from the other four subtypes with significant labeling in outer layers of the cortex, hippocampus, caudate putamen, olfactory tubercle, and highest in substantia nigra. M₃ receptors are found in cerebral cortex, thalamus, and hippocampus but not in the basal ganglia¹⁹⁵ and may have similar functions as the M_1 receptor in these areas.

As described above, the molecular mechanism of antimuscarinic-induced psychosis has not been clearly defined and would benefit from receptor subtype-selective pharmacological agents. Treatment of schizophrenia with muscarinic agents may provide significant therapeutic utility when treating the positive and negative symptoms with the added benefit of improving cognitive function. A muscarinic ligand might also be useful in treating the psychosis and dementia associated with neurodegenerative diseases such as Alzheimer's disease, Lewybody disease, and Parkinson's disease. The complexity of the cholinergic nervous system and the overlapping distribution of the five muscarinic receptor subtypes within the brain present a significant challenge to the development of muscarinic-based therapeutics for psychosis and other indications.

Alzheimer's Disease

Alzheimer's disease (AD) is a disorder characterized by progressive age-related impairment of neurological function. The clinical manifestations include progressive loss of cognition (dementia) as well as psychosis, loss of reasoning ability, agitation, reduction in personal care, and other disturbances of affect. 196–198 Patients are usually impaired in two or more areas of cognition

including, memory, language, visiospatial function, and attention. Postmortem brain tissue samples from AD patients consistently show a neuropathology characterized by progressive accumulation of amyloid plaques and neurofibrillary tangles, shrinkage and death of neurons, and inflammatory changes marked by accumulation of activated microglia. 199–202 These changes are associated with degeneration of neurons expressing cholinergic, serotonergic, glutamanergic, and neuropeptide neurotransmitter systems. 203

The loss of cholinergic neurons in particular may contribute to the etiology of AD due to their localization in memory-forming areas of the brain and participation in cognitive function. Anatomical evidence has shown that cholinergic innervation projects from the nucleus Basalis of Meynart to hippocampus, amygdala, and neocortex, ^{204–207} which are considered to be neural substrates for learning and memory ^{208–210} and for visual representation and judgment. ²¹¹ Cerebral cortex, hippocampus, and amygdala show extensive AD pathology, ^{212,213} reduction in acetylcholine levels, ^{205,214–216} and loss of cholinergic innervation. ^{209,210,217,218} The loss of the acetylcholine-synthesizing enzyme, choline acetyltransferase, was observed in cerebral cortex and hippocampus from postmortem AD tissue. ^{214,215,219}

Cholinesterase Inhibitors. The cholinergic hypothesis assumes that replacement of acetylcholine, which is progressively lost during the course of AD, would potentially delay and possibly restore losses in cognitive abilities. Two approaches have been used to test this hypothesis including using cholinesterase inhibitors to maintain synaptic levels of acetylcholine^{220,221} or alternatively administration of synthetic agonists to stimulate acetylcholine receptors of either the muscarinic or nicotinic family.^{222,223} Cholinesterase inhibitors were shown to reverse cognitive loss in rodents following lesions of the cholinergic system projecting to hippocampus and cortex.^{224,225} Previous studies showed cognitive loss in rats after lesions were made in cholinergic memory circuits.²²⁶⁻²²⁹ Using another approach, spatial memory impairment in rats resulting from lesions of the basal forebrain cholinergic projection system were partially reversed by brain implantation of acetylcholineproducing cells.²³⁰ On the basis of a plethora of preclinical and clinical data, cholinesterase inhibitors are currently the only approved treatment for cognitive deficits associated with AD. Donepezil (Aricept), tacrine (Cognex), and rivastigmine (Exelon) are approved for use in the United States, and three compounds have received European approval, tacrine being the first and, more recently, donepezil and rivastigmine (ENA-713). Other compounds under investigation include galantamine, physostigmine, eptastigmine, and metrifonate. Currently it is believed that cholinesterase inhibitors act primarily by increasing available acetylcholine within the active synapse thereby restoring deficient cholinergic neurotransmission. Mechanistically, tacrine, donepezil, galantamine, and physostigmine are reversible inhibitors of acetylcholinesterase and butyrylcholinesterase, rivastigmine is a pseudoirreversible inhibitor of these enzymes, while metrifonate is considered to be an irreversible inhibitor. Early clinical trials with cholinesterase inhibitors, such as tacrine, showed modest but statistically insignificant effects on cognitive

performance even when combined with the acetylcholine precursor lecithin.²³¹ Subsequent trials allowing increased dosing showed significant improvement in measures of cognitive performance but were still modest in outcome due in part to dose limiting side effects including GI disturbances and hepatotoxicity. 232,233 Donepezil was shown to be potentially more effective than tacrine due to a lower side effect profile and possibly due to its higher selectivity for acetylcholinesterases over butyrylcholinesterases. However, only modest improvements in cognitive function were observed in early stage AD patients with this compound.²³⁴ Physostigmine has undergone clinical testing and consistent with this class of compounds has shown marginal effectiveness with both statistically significant^{235,236} and insignificant outcomes.237

Cholinesterase inhibition has several obvious drawbacks. These compounds depend on an intact cholinergic system still capable of synthesizing acetylcholine; therefore, they may be most useful in early stages of AD and lose effectiveness over time. The presence of M_2 autoreceptors would potentially respond to acetylcholine and have opposing actions on acetylcholine release. Selective cholinesterase inhibitors, free of dose-limiting side effects, are not currently available, and current compounds may not allow sufficient modulation of acetylcholine levels to elicit the full therapeutic response. For this reason alternative approaches have been considered including direct stimulation of either muscarinic or nicotinic acetylcholine receptors.

Muscarinic Receptor Agonists. Pharmacological evidence suggests that muscarinic and possibly nicotinic receptors play an important role in learning and memory. For example, muscarinic antagonists disrupt short-term memory in rats^{238–240} and mice.²⁴¹ The muscarinic receptor antagonist scopolamine disrupts memory in humans²⁴² which can be reversed with the cholinesterase inhibitor physostigmine.²³⁵ Nicotinic receptors may also offer a viable therapeutic target for cognitive enhancement due to their location in memory-forming areas of the brain. The $\alpha 4\beta 2$ receptor subtype may be the most interesting target of the nicotinic receptor family due to its higher abundance in brain, though levels of this receptor are several orders of magnitude lower in density compared to muscarinic receptors. Nicotinic receptors are located on presynaptic cholinergic terminals, and nicotinic agonists increase acetylcholine release resulting in a feed-forward effect on cholinergic transmission. However, nicotinic antagonists do not induce cognitive impairment seen with muscarinic antagonists suggesting a less direct involvement in the memory process.

Determination of which of the five muscarinic receptor subtypes are involved in memory formation and storage has been hampered by the lack of receptor subtype-selective ligands. Due to its high abundance in hippocampus and cortex, the M_1 receptor may be largely involved in memory formation. Moreover, pirenzepine, which is an M_1 receptor-preferring antagonist, disrupts memory in rats and mice antagonist, disrupts memory in rats and mice cannot be completely ruled out. A recent study found that muscarinic agonist potentiation of N-methyl-D-aspartate receptor currents in rat hippocampal neurons, believed to be a predomi-

nant mechanism for cholinergic modulation of hippocampal function, was mediated by M₁ receptors, supporting the role of this subtype in learning and memory.²⁴⁴ These findings suggest that the M₁ receptor may be an attractive target for cognitive enhancement in AD. Radioligand binding studies on postmortem brain tissue from AD patients demonstrated a loss of M₂ receptors that are thought to be predominantly presynaptic and a preservation of postsynaptic M₁ receptors.²⁴⁵ M₁ agonist replacement therapy has been considered by several groups as potentially more promising than development of cholinesterase inhibitors, since the latter approach depends on acetylcholine release from presynaptic neurons that are progressively lost during the course of the disease. The first attempts at muscarinic replacement therapy were unsuccessful due to the lack of selectivity of the muscarinic agonists and associated muscarinic side effects. Cholinergic side effects include salivation, lacrimation, and GI disturbances. This is supported by the lack of effectiveness of bethanecol and arecoline in the treatment of AD.^{246,247} Development of muscarinic compounds that are truly receptor subtype selective has not been achieved leaving the hypothesis partly untested. The first clinical evidence that a direct muscarinic agonist with significant M₁ activity would improve cognitive function in patients with AD was shown with xanomeline.¹⁸¹ The xanomeline trial resulted in significant improvements in cognition, despite dose-limiting adverse events. Its favorable effects on psychotic-like behaviors of AD suggest that muscarinic agonists may be useful for the treatment of schizophrenia or other psychosis.

Use of direct-acting agonists for the M₁ receptor in the treatment of AD may have limitations. A recent report demonstrated an apparent decrease in the levels of M₁ receptor protein in AD brain tissue as determined by immunoprecipitation with subtype-selective antibodies, although the same study showed no change in M₁ receptors as determined by radioligand binding in agreement with previous work.248 Although not demonstrated directly, this finding suggests altered M1 receptor structure in AD brain, a suggestion further supported by studies demonstrating impaired M₁ receptor functional activity in the disease. 249-251 More recently reduction in high-affinity binding of M₁ receptors in AD brain was noted suggesting reduced efficiency of receptor-G protein interactions resulting in loss of signal transduction efficiency.²⁵² It is not clear if sufficient M₁ receptor reserve exists in AD brain areas regulating cognitive function to compensate for these partial reductions in muscarinic receptor function. It may also be possible to delay or greatly reduce the loss of M₁ receptor function if M₁ agonist replacement is initiated early in the disease.

In addition to the involvement of the cholinergic system in cognitive function, muscarinic receptors have been implicated in the control of amyloid precursor protein (APP) processing. The etiology of AD is in part due to the abnormal processing of APP leading to the formation and deposition of a neurotoxic fragment, amyloid- β peptide (A β). The senile plaques of AD consist predominantly of a 39–43-residue of A β . A β is produced by proteolysis of the much larger APP. Nitsch and colleagues first demonstrated that muscarinic agonist

stimulation of HEK293 cells transfected with M₁ and M₃ receptors (but not M₂ or M₄) resulted in increased release of a soluble N-terminal derivative of APP (APPs).²⁵³ AAPs is derived from the secretory pathway whose activation is believed to reduce the formation of amyloidogenic Aβ.²⁵⁴ Subsequently, it was shown that activation of M₁ receptors on cells containing either normal or Swedish mutant forms of APP resulted in significant reductions in A β release into the medium.²⁵⁵ In addition to the muscarinic receptors, other GPCRs linked to the stimulation of phosphatidylinositol hydrolysis (i.e. $5HT_{2a}$, $5HT_{2c}$, $mGluR_{1\alpha}$, bradykinin) have also been shown to stimulate nonamyloidogenic APPs release from transfected cells.²⁵⁶ In this study, release of APPs could be stimulated in superfused brain slices by electrical depolarization and a significant component of this effect could be blocked by atropine indicating involvement of muscarinic receptors. These workers also demonstrated that APPs release from brain slices could be stimulated by agonists with partial selectivity for M₁ receptors, but not by nonselective agonists. Important observations on APP processing in vivo have been made by Haroutunian and colleagues who demonstrated that excitotoxic lesions of the basal forebrain cholinergic projection system in rats resulted in upregulation of APP mRNA in the cortex accompanied by increased levels of a soluble APP derivative in the CSF.²⁵⁷ Lesions of rat basal forebrain cholinergic neurons in vivo with 192IgG-saporin have been shown to increase the amount of membrane-associated APP and reduce the amount of soluble (presumably secreted) APP in the cortex, changes that were reversed by restoring cholinergic function via implantation of fibroblasts secreting nerve growth factor. 258 Signal transduction associated with M1 receptor activation leads to modulation of APP release. 254 These studies indicate that M_1 receptors in vivo may affect APP processing in the same way as observed in cell systems in vitro. In addition, muscarinic agonists have been shown to reduce the phosphorylation state of tau protein in cultured cells as well as in vivo. 259,260 Hyperphosphorylated tau is the major component of neurofibrillary tangles in AD brain.²⁶¹ Therefore, muscarinic receptor-mediated regulation of APP processing and tau phosphorylation suggest the possibility that muscarinic agonist replacement may have the added benefit of affecting the progression of AD by inhibiting those processes which contribute to accumulation of plagues and tangles.

AD like other neurodegenerative diseases is the result of multiple abnormalities, all of which have not been fully recognized. The development of a therapeutic designed to help a patient repair or recover from a genetic defect may be beneficial but may only impact a fraction of the total patient population. A full understanding of this disease could easily require decades of challenging research that may do little to help a rapidly aging population of middle-aged adults. It has been shown that palliative treatments, though far from ideal, can significantly reduce the emotional and financial burdens of both patient and family. An approach focused on reducing or delaying cognitive decline and difficult behaviors through agonist replacement therapy has yet to be adequately evaluated.

Parkinson's Disease

Parkinson's disease is characterized by four predominant symptoms: bradykinesia (slow and difficult movement), muscular rigidity, resting tremor that abates during a voluntary movement, and problems maintaining balance which is seen as a disturbed gait and possible falling. The symptoms result from a loss of dopaminergic neurons that project to the striatum (caudate and putamen) from the substantia nigra pars compacta beyond what is normally expected during the aging process. Late stage disease progresses from akinesia to immobility. A parkinsonism-like state can be induced by dopamine-blocking drugs such as the neuroleptics, haloperidol and chlorpromazine, and the antiemetics, prochloperazine and metoclopromide. Other neurological diseases also can induce these symptoms such as less common neurodegenerative diseases and stroke. Current treatments include muscarinic antagonists such as benztropine and trihexyphenidyl and dopamine replacement regiments such as the dopamine precursor levodopa or the dopamine agonists bromocriptine and pergolide.

As in schizophrenia, the predominant neurochemical abnormality in parkinsonism results from disturbances in the balance between the cholinergic and dopaminergic systems. Loss of dopaminergic neurons results in a predominance of the cholinergic system resulting in the tremor and movement disorders. This hypothesis is supported by clinical observations showing that cholinergic agonists and cholinesterase inhibitors induce or exacerbate parkinsonian tremors. 262-266 Muscarinic antagonists, although effective at treating movement disorders associated with parkinsonism, have a tendency to induce undesirable side effects including psychosis as described above. Anticholinergics currently prescribed are relatively nonselective for the individual muscarinic receptor subtypes providing little information at the mechanistic level as to their sites of action. Animal models have helped investigators understand the neurochemical basis for Parkinson's disease in relationship to the dopamine system, but little information is available about which muscarinic receptor subtypes modulate the dopamine system in the basal ganglia.

Recent studies have focused on vacuous jaw tremors in rodents as a model of human resting parkinsonian tremor. Muscarinic agonists such as pilocarpine induce a parkinsonian jaw tremor in rodents that oscillates at a similar frequency to jaw and hand tremors observed in human Parkinson's patients.²⁶⁷ Pharmacological characterization of the muscarinic subtypes involved in the jaw tremor is consistent with regulation by the M₄ receptor.²⁶⁸ Dopamine depletion studies and direct injections of muscarinic agonists and antagonists into brain regions indicated that regulation of tremor is initiated predominantly in the ventrolateral striatal area and that the substantia nigra also plays an important role.²⁶⁹ The distribution of muscarinic receptors in basal ganglia is described above and indicates that all five receptor subtypes, in particular the M_1 , M_4 , and M₅ receptors, are expressed in significant levels in the neostriatum and on dopamine cells. Selective muscarinic ligands and muscarinic receptor knockout mice should help support the hypothesis that the M₄ receptor

plays a key role in tremor control. These data suggest that a selective M_4 antagonist might offer an improvement over current muscarinic antagonist therapies for parkinsonian tremor.

Conclusion and Future Directions

As described in this review, muscarinic ligands have had a long history in the treatment of central nervous system disorders. Previous compounds, including those more recently developed such as xanomeline and milameline for AD-related cognitive decline, offer limited therapeutic benefit due to their lack of selectivity resulting in dose-limiting side effects. Advances in molecular biochemistry and genetics have provided a new collection of research tools creating new therapeutic development opportunities for muscarinic and other receptor families. The discovery and development of more selective ligands will occur through screening of larger and more chemically diverse compound libraries with the help of sensitive in vitro assays run in parallel across receptor families. Knockout mice and other forms of genetically manipulated animal models will deepen our understanding of receptor subtype function and help unravel their complex interactions with other proteins. Hopefully the genetic revolution will help physicians improve their detection, and possibly prediction of neural disease, allowing earlier therapeutic intervention during and before the disease process. Even with improved diagnostics and knowledge of disease mechanism, the ultimate goal of halting or preventing disease progression may be years away. For this reason, palliative treatments for AD and other diseases still remain very attractive as primary and adjunctive therapies.

The cholinergic hypothesis predicts that loss of cholinergic function plays a significant role in the cognitive decline observed in AD.²⁷⁰ Two or more decades of effort leave cholinesterase inhibitors as the only FDA-approved treatments currently available for AD-related cognitive decline. The marginal efficacy of cholinesterase inhibitors and difficulty of developing selective muscarinic agonist-based therapeutics has softened the enthusiasm for the muscarinic agonist approach to the treatment of cognitive decline, psychosis, and other CNS diseases. The new challenge is to provide solid evidence for the role of the M₁ receptor in modulating postsynaptic function and demonstrating that an M₁ agonist alone is sufficient to restore cognitive acuity. It will also be important to understand the relative contribution of M₂ receptor blockade and if this activity alone is sufficient to achieve robust therapeutic effects superior to current therapies. Most likely, the progressive basal forebrain cholinergic deafferentation that occurs in AD will quickly obviate this mechanistic approach. Combinations of M₁ agonism, M₂ antagonism, and cholinesterase inhibition will be investigated for efficacy and will be optimized based on patient response and genetic profile. Preclinical models of cognitive performance are now providing better information about efficacy at the various cognitive domains such as attention, recent and long-term memory, spatial learning, and possibly executive function. It will be important to determine if therapeutic balance of nicotinic receptor versus muscarinic receptor modulation has the potential to provide even greater benefit and which cognitive domains are most impacted.

Treatments for schizophrenia and psychosis have centered on modulation of the dopamine system, and remarkable improvements have been made in treating both positive and negative symptoms associated with the disease using atypical antipsychotic medications. However, atypical antipsychotics such as clozapine are based on complex mechanisms that are not fully understood and have limited usefulness in treating cognitive deficits associated with psychosis. Reciprocal modulation between the dopamine and acetylcholine systems suggests that muscarinic receptor-based medications would offer an alternative approach that could also restore cognitive deficits. The challenge is to understand the relative contributions of each muscarinic receptor subtype, and possibly nicotinic receptor subtypes, to the positive, negative, and cognitive symptoms of psychosis.

Pharmacological and behavioral data support a role for the M₂ and M₃ receptors in mediating many of the parasympathomimetic cholinergic symptoms associated with muscarinic ligands. Recent studies using muscarinic receptor knockout mice have largely supported previous pharmacological findings. Therefore, stimulation of M₂ and M₃ receptors appears to be responsible for many of the side effects observed with previous muscarinic agonists. If M₂ and M₃ receptor activity is avoided in future compounds, will they retain their usefulness as therapeutics? The answer to this key question will determine the success of muscarinic-based efforts since M2 and M3 receptors are located in areas of the brain involved in cognition, psychosis, and movement control. Peripheral cholinergic-regulated physiology is regulated in part through central mechanisms. The role of each muscarinic receptor subtype in central versus peripheral physiology has yet to be explored in detail. Successful design of novel selective ligands devoid of cholinergic side effects will require a deeper understanding of these interactions especially as they affect the cardiovascular and gastrointestinal systems. Although the use of receptor knockout animals has already been helpful in this regard, there are concerns about the limitations of this approach. The nervous systems of receptor knockout animals may undergo significant developmental changes in brain architecture to compensate for loss of the gene target. These potential rearrangements may confound interpretation of data derived from these animals. Alternative genetic approaches are possible which provide conditional and localized regulation of receptors in adult animals. Hopefully these models will be available soon. The slow decline in interest in cholinergic-based therapeutics that has occurred over the past decade is beginning to be revitalized through the influence of modern molecular biochemistry. In general, GPCRs have proven to be useful therapeutic targets for many historical and modern pharmaceuticals. On the basis of recent learning, the next decade should prove particularly exciting and productive for the muscarinic receptor family.

Biographies

Christian Felder received his B.S. in chemistry from the College of William and Mary in 1977, his M.S. in biochemistry from the University of Maryland in 1980, and his Ph.D. in biochemistry from Georgetown University in 1987. He then spent three years with Julius Axelrod as a staff fellow at the NIMH. Following his postdoctoral training, he established his own laboratory as Chief of the Unit on Molecular and Cellular Signaling at the NIMH. In 1997, he joined Eli Lilly Research Laboratories where he is currently Senior Research Scientist and Chair of the Cholinergic Strategy Group.

Frank Bymaster received his B.S. in pharmacy from Butler University in 1968 and his M.S. in pharmacology from Indiana University. He joined Eli Lilly Research Laboratories in 1970 and currently is Senior Research Scientist and Biological Pharmacology Scientific Leader in the Neuroscience Research Division.

John Ward received his B.S. in chemistry and his Ph.D. in organic chemistry from the University of Texas at Austin in 1967 and 1971, respectively. Following NIH sponsored postdoctoral training with Leo Paquette at The Ohio State University, he joined the discovery research effort at Eli Lilly Research Laboratories in 1972. In 1999, he moved to Protein Design Labs, Inc. as Senior Scientist.

Neil DeLapp received his B.S. in chemistry from Phillips University (Enid Oklahoma) in 1966 and his Ph.D. in biochemistry from Florida State in 1972. He spent three years in postdoctoral research with Dr. Marvin Karasek in the Department of Dermatology at Stanford University and joined Eli Lilly Research Laboratories in the Physiology Department in 1975. He is currently Senior Research Scientist in the Lead Optimization Biology Division.

References

- (1) Hulme, E. C.; Birdsall, N. J.; Buckley, N. J. Muscarinic receptor subtypes. Ann. Rev. Pharmacol. Toxicol. 1990, 30, 633-73.
- Caulfield, M. P. Muscarinic receptors-characterization, coupling and function. Pharmacol. Ther. 1993, 58, 319-79.
- (3) Caulfield, M. P.; Birdsall, N. J. International Union of Pharmacology. XVII. Classification of muscarinic acetylcholine receptors. Pharmacol. Rev. 1998, 50, 279-90.
- (4) Eglen, R. M.; Hegde, S. S.; Watson, N. Muscarinic receptor subtypes and smooth muscle function. Pharmacol. Rev. 1996, 48, 531-65.
- Wess, J. Molecular biology of muscarinic acetylcholine receptors. Crit. Rev. Neurobiol. 1996, 10, 69-99.
- (6) Brown, D. A.; Abogadie, F. C.; Allen, T. G.; Buckley, N. J.; Caulfield, M. P.; Delmas, P.; Haley, J. E.; Lamas, J. A.; Selyanko, A. A. Muscarinic mechanisms in nerve cells. Life Sci. 1997, 60, 1137 - 44
- (7) Jerusalinsky, D.; Harvey, A. L. Toxins from mamba venoms: small proteins with selectivities for different subtypes of muscarinic acetylcholine receptors. Trends Pharmacol. Sci. 1994, 15, 424 - 30.
- (8) Felder, C. C. Muscarinic acetylcholine receptors: signal transduction through multiple effectors. *FASEB J.* **1995**, *9*, 619–25.
- Eglen, R. M.; Watson, N. Selective muscarinic receptor agonists and antagonists. Pharmacol. Toxicol. 1996, 78, 59-68.
- (10) Riker, W. F.; Wescoe, W. C. The pharmacology of flexedil with observations on certain analogues. Ann. N. Y. Acad. Sci. 1951,
- (11) Kubo, T.; Fukuda, K.; Mikami, A.; Maeda, A.; Takahashi, H.; Mishina, M.; Haga, T.; Haga, K.; Ichiyama, A.; Kangawa, K.; et al. Cloning, sequencing and expression of complementary DNA encoding the muscarinic acetylcholine receptor. Nature 1986,
- (12) Bonner, T. I.; Buckley, N. J.; Young, A. C.; Brann, M. R. Identification of a family of muscarinic acetylcholine receptor genes. *Science* **1987**, *237*, 527–32; published erratum appears in Science 1987, 237, 4822.
- (13) Bonner, T. I.; Young, A. C.; Brann, M. R.; Buckley, N. J. Cloning and expression of the human and rat m5 muscarinic acetylcholine receptor genes. Neuron 1988, 1, 403-10.
- (14) Hulme, E. C.; Birdsall, N. J.; Buckley, N. J. Muscarinic receptor subtypes. Ann. Rev. Pharmacol. Toxicol. 1990, 30, 633-73.
- (15) Schertler, G. F.; Villa, C.; Henderson, R. Projection structure of rhodopsin. Nature 1993, 362, 770-2.
- Palczewski, K.; Kumasaka, T.; Hori, T.; Behnke, C. A.; Motoshima, H.; Fox, B. A.; Le Trong, I.; Teller, D. C.; Okada, T.; Stenkamp, R. E.; Yamamoto, M.; Miyano, M. Crystal structure of rhodopsin: A G protein-coupled receptor [see comments]. *Science* **2000**, *289*, 739–45.
- (17) Kenakin, T. P. The classification of seven transmembrane receptors in recombinant expression systems. Pharmacol. Rev. **1996**, 48, 413-63.
- (18) Schertler, G. F. Structure of rhodopsin. Eye 1998, 12, 504-10.

- (19) Kurtenbach, E.; Curtis, C. A.; Pedder, E. K.; Aitken, A.; Harris, A. C.; Hulme, E. C. Muscarinic acetylcholine receptors. Peptide sequencing identifies residues involved in antagonist binding and disulfide bond formation. *J. Biol. Chem.* 1990, 265, 13702–8
- (20) Baldwin, J. M. The probable arrangement of the helices in G protein-coupled receptors. *EMBO J.* **1993**, *12*, 1693–703.
- (21) Baldwin, J. M. Structure and function of receptors coupled to G proteins. Curr. Opin. Cell Biol. 1994, 6, 180–90.
- (22) Schwartz, T. W. Locating ligand-binding sites in 7TM receptors by protein engineering. Curr. Opin. Biotechnol. 1994, 5, 434– 44.
- (23) Spalding, T. A.; Birdsall, N. J.; Curtis, C. A.; Hulme, E. C. Acetylcholine mustard labels the binding site aspartate in muscarinic acetylcholine receptors. *J. Biol. Chem.* **1994**, *269*, 4092–7.
- (24) Curtis, C. A.; Wheatley, M.; Bansal, S.; Birdsall, N. J.; Eveleigh, P.; Pedder, E. K.; Poyner, D.; Hulme, E. C. Propylbenzilylcholine mustard labels an acidic residue in transmembrane helix 3 of the muscarinic receptor. *J. Biol. Chem.* 1989, 264, 489–95.
- (25) Hulme, E. C.; Curtis, C. A.; Page, K. M.; Jones, P. G. The role of charge interactions in muscarinic agonist binding, and receptorresponse coupling. *Life Sci.* 1995, 56, 891–8.
- response coupling. *Life Sci.* **1995**, *56*, 891–8.

 (26) Schwarz, R. D.; Spencer, C. J.; Jaen, J. C.; Mirzadegan, T.; Moreland, D.; Tecle, H.; Thomas, A. J. Mutations of aspartate 103 in the Hm2 receptor and alterations in receptor binding properties of muscarinic agonists. *Life Sci.* **1995**, *56*, 923–9.
- (27) Wess, J.; Maggio, R.; Palmer, J. R.; Vogel, Z. Role of conserved threonine and tyrosine residues in acetylcholine binding and muscarinic receptor activation. A study with m3 muscarinic receptor point mutants. J. Biol. Chem. 1992, 267, 19313–9.
- (28) Wess, J.; Gdula, D.; Brann, M. R. Site-directed mutagenesis of the m3 muscarinic receptor: identification of a series of threonine and tyrosine residues involved in agonist but not antagonist binding. *EMBO J.* 1991, 10, 3729–34.
 (29) Bluml, K.; Mutschler, E.; Wess, J. Functional role in ligand
- (29) Bluml, K.; Mutschler, E.; Wess, J. Functional role in ligand binding and receptor activation of an asparagine residue present in the sixth transmembrane domain of all muscarinic acetylcholine receptors. J. Biol. Chem. 1994, 269, 18870–6.
- (30) Tence, M.; Cordier, J.; Premont, J.; Glowinski, J. Muscarinic cholinergic agonists stimulate arachidonic acid release from mouse striatal neurons in primary culture. *J. Pharmacol. Exp. Ther.* 1994, 269, 646–53.
- (31) Strosznajder, J.; Samochocki, M. Carbachol-stimulated release of arachidonic acid and eicosanoids from brain cortex synaptoneurosome lipids of adult and aged rats. *Adv. Exp. Med. Biol.* **1992**, *318*, 251–8.
- (32) Unno, T.; Komori, S.; Ohashi, H. Some evidence against the involvement of arachidonic acid in muscarinic suppression of voltage-gated calcium channel current in guinea-pig ileal smooth muscle cells. Br. J. Pharmacol. 1996, 119, 213–22.
- (33) Conklin, B. R.; Brann, M. R.; Buckley, N. J.; Ma, A. L.; Bonner, T. I.; Axelrod, J. Stimulation of arachidonic acid release and inhibition of mitogenesis by cloned genes for muscarinic receptor subtypes stably expressed in A9 L cells. *Proc. Natl. Acad. Sci. U.S.A.* 1988, 85, 8698–702.
- (34) Schurmann, A.; Schmidt, M.; Asmus, M.; Bayer, S.; Fliegert, F.; Koling, S.; Massmann, S.; Schilf, C.; Subauste, M. C.; Voss, M.; Jakobs, K. H.; Joost, H. G. The ADP-ribosylation factor (ARF)-related GTPase ARF-related protein binds to the ARF-specific guanine nucleotide exchange factor cytohesin and inhibits the ARF-dependent activation of phospholipase D. J. Biol. Chem. 1999, 274, 9744-51.
- (35) Dolezal, V.; Lisa, V.; Tucek, S. Differential effects of the M1-M5 muscarinic acetylcholine receptor subtypes on intracellular calcium and on the incorporation of choline into membrane lipids in genetically modified Chinese hamster ovary cell lines. *Brain Res. Bull.* 1997, 42, 71-8.
- Res. Bull. **1997**, 42, 71–8.

 (36) Singer-Lahat, D.; Rojas, E.; Felder, C. C. Muscarinic receptor activated Ca2+ channels in nonexcitable cells. *Prog. Brain Res.* **1996**, 109, 195–9.
- (37) Gustavsson, L.; Lundqvist, C.; Hansson, E. Receptor-mediated phospholipase D activity in primary astroglial cultures. *Glia* 1993, 8, 249-55.
- (38) Sandmann, J.; Peralta, E. G.; Wurtman, R. J. Coupling of transfected muscarinic acetylcholine receptor subtypes to phospholipase D. J. Biol. Chem. 1991, 266, 6031–4.
- (39) Boulay, G.; Zhu, X.; Peyton, M.; Jiang, M.; Hurst, R.; Stefani, E.; Birnbaumer, L. Cloning and expression of a novel mammalian homologue of Drosophila transient receptor potential (Trp) involved in calcium entry secondary to activation of receptors coupled by the Gq class of G protein. J. Biol. Chem. 1997, 272, 29672–80.
- (40) Singer-Lahat, D.; Ma, A. L.; Felder, C. C. Independent induction of morphological transformation of CHO cells by receptoractivated cyclic AMP synthesis or by receptor-operated calcium influx. *Biochem. Pharmacol.* 1996, 51, 495-502.

- (41) Tachado, S. D.; Virdee, K.; Akhtar, R. A.; Abdel-Latif, A. A. M3 muscarinic receptors mediate an increase in both inositol trisphosphate production and cyclic AMP formation in dog iris sphincter smooth muscle. J. Ocular Pharmacol. 1994, 10, 137– 47.
- (42) Johnson, J. A.; Friedman, J.; Halligan, R. D.; Birnbaumer, M.; Clark, R. B. Sensitization of adenylyl cyclase by P2 purinergic and M5 muscarinic receptor agonists in L cells. *Mol. Pharmacol.* 1991, 40, 539–46.
- (43) Heldman, E.; Barg, J.; Fisher, A.; Levy, R.; Pittel, Z.; Zimlichman, R.; Kushnir, M.; Vogel, Z. Pharmacological basis for functional selectivity of partial muscarinic receptor agonists. *Eur. J. Pharmacol.* 1996, 297, 283–91.
- (44) Felder, C. C.; Kanterman, R. Y.; Ma, A. L.; Axelrod, J. A transfected m1 muscarinic acetylcholine receptor stimulates adenylate cyclase via phosphatidylinositol hydrolysis. *J. Biol. Chem.* 1989, 264, 20356–62.
- (45) Zhu, X.; Birnbaumer, L. G protein subunits and the stimulation of phospholipase C by Gs-and Gi-coupled receptors: Lack of receptor selectivity of Galpha(16) and evidence for a synergic interaction between Gbeta gamma and the alpha subunit of a receptor activated G protein. Proc. Natl. Acad. Sci. U.S.A. 1996, 93, 2827-31.
- (46) Felder, C. C.; Williams, H. L.; Axelrod, J. A transduction pathway associated with receptors coupled to the inhibitory guanine nucleotide binding protein Gi that amplifies ATPmediated arachidonic acid release. *Proc. Natl. Acad. Sci. U.S.A.* 1991, 88, 6477–80.
- (47) Nadler, L. S.; Rosoff, M. L.; Hamilton, S. E.; Kalaydjian, A. E.; McKinnon, L. A.; Nathanson, N. M. Molecular analysis of the regulation of muscarinic receptor expression and function. *Life* Sci. 1999, 64, 375–9.
- (48) Bunemann, M.; Hosey, M. M. G-protein coupled receptor kinases as modulators of G-protein signaling. *J. Physiol.* **1999**, *517*, 5–23.
- (49) Kim, J. Y.; Yang, M. S.; Oh, C. D.; Kim, K. T.; Ha, M. J.; Kang, S. S.; Chun, J. S. Signaling pathway leading to an activation of mitogen-activated protein kinase by stimulating M3 muscarinic receptor. *Biochem. J.* **1999**, *337*, 275–80.
- (50) Tsai, W.; Morielli, A. D.; Cachero, T. G.; Peralta, E. G. Receptor protein tyrosine phosphatase alpha participates in the m1 muscarinic acetylcholine receptor-dependent regulation of Kv1.2 channel activity. *EMBO J.* **1999**, *18*, 109–18.
- (51) Gusovsky, F.; Lueders, J. E.; Kohn, E. C.; Felder, C. C. Muscarinic receptor-mediated tyrosine phosphorylation of phospholipase C-gamma. An alternative mechanism for cholinergic-induced phosphoinositide breakdown. J. Biol. Chem. 1993, 268, 7768–72.
- (52) Gutkind, J. S.; Robbins, K. C. Activation of transforming G protein-coupled receptors induces rapid tyrosine phosphorylation of cellular proteins, including p125FAK and the p130 v-src substrate. *Biochem. Biophys. Res. Commun.* 1992, 188, 155– 61.
- (53) Inoue, R.; Waniishi, Y.; Yamada, K.; Ito, Y. A possible role of tyrosine kinases in the regulation of muscarinic receptoractivated cation channels in guinea pig ileum. *Biochem. Biophys. Res. Commun.* 1994, 203, 1392–7.
- (54) Boddeke, H. W. Different effects of muscarinic agonists in rat superior cervical ganglion and hippocampal slices. Eur. J. Pharmacol. 1991, 201, 191–7.
- (55) Buckley, N. J.; Burnstock, G. Autoradiographic localization of peripheral M1 muscarinic receptors using [3H]pirenzepine. *Brain Res.* 1986, 375, 83–91.
- (56) Shannon, H. E.; Sawyer, B. D.; Bemis, K. G.; Bymaster, F. P.; Health, I.; Mitch, C. H.; Ward, J. S. Muscarinic M1 receptor agonist actions of muscarinic receptor agonists in rabbit vas deferens. Eur. J. Pharmacol. 1993, 232, 47–57.
- (57) Nilvebrant, L. On the muscarinic receptors in the urinary bladder and the putative subclassification of muscarinic receptors. *Acta Pharmacol. Toxicol.* **1986**, *59* (Suppl. 1), 1–45.
- (58) Eglen, R. M. Muscarinic M2 and M3 receptor function in smooth muscle. Proc. Western Pharmacol. Soc. 1996, 39, 57–60.
- (59) Yasuda, R. P.; Ciesla, W.; Flores, L. R.; Wall, S. J.; Li, M.; Satkus, S. A.; Weisstein, J. S.; Spagnola, B. V.; Wolfe, B. B. Development of antisera selective for m4 and m5 muscarinic cholinergic receptors: distribution of m4 and m5 receptors in rat brain. *Mol. Pharmacol.* 1993, 43, 149–57.
- (60) Reever, C. M.; Ferrari-DiLeo, G.; Flynn, D. D. The M5 (m5) receptor subtype: fact or fiction? Life Sci. 1997, 60, 1105-12.
- (61) Kohn, E. C.; Alessandro, R.; Probst, J.; Jacobs, W.; Brilley, E.; Felder, C. C. Identification and molecular characterization of a m5 muscarinic receptor in A2058 human melanoma cells. Coupling to inhibition of adenylyl cyclase and stimulation of phospholipase A2. J. Biol. Chem. 1996, 271, 17476-84.

- (62) Hamilton, S. E.; Loose, M. D.; Qi, M.; Levey, A. I.; Hille, B.; McKnight, G. S.; Idzerda, R. L.; Nathanson, N. M. Disruption of the m1 receptor gene ablates muscarinic receptor-dependent M current regulation and seizure activity in mice. *Proc. Natl. Acad. Sci. U.S.A.* 1997, 94, 13311–6.
- (63) Gomeza, J.; Shannon, H.; Kostenis, E.; Felder, C.; Zhang, L.; Brodkin, J.; Grinberg, A.; Sheng, H.; Wess, J. Pronounced pharmacologic deficits in M2 muscarinic acetylcholine receptor knockout mice. *Proc. Natl. Acad. Sci. U.S.A.* 1999, 96, 1692–7.
- (64) Gomeza, J.; Kostenis, E.; Felder, C.; Bymaster, F.; Brodkin, J.; Shannon, H.; Xia, B.; Deng, C. X.; Wess, J. Enhancement of D1 dopamine receptor-mediated locomotor stimulation in M-4 muscarinic acetylcholine receptor knockout mice. *Proc. Natl. Acad.* Sci. U.S.A. 1999, 96, 10483–8.
- (65) Shapiro, M. S.; Loose, M. D.; Hamilton, S. E.; Nathanson, N. M.; Gomeza, J.; Wess, J.; Hille, B. Assignment of muscarinic receptor subtypes mediating G-protein modulation of Ca2+chanels by knockout mice. *Proc. Natl. Acad. Sci. U.S.A.* 1999, 96, 10899–904.
- (66) Jean, J. C.; Davis, R. E. Recent advances in the design and characterization of muscarinic agonists and antagonists. *Annu. Rep. Med. Chem.* 1994, 29, 23–32.
- (67) Moltzen, E. K.; Bjornholm, B. Medicinal chemistry of muscarinic agonists: developments since 1990. *Drugs Future* 1995, 20, 37– 54.
- (68) Plate, R.; Plaum, M. J.; de Boer, T.; Andrews, J. S.; Rae, D. R.; Gibson, S. Synthesis and muscarinic activities of 3-(pyrazolyl)-1,2,5,6-tetrahydropyridine derivatives. *Bioorg. Med. Chem.* 1996, 4, 227–37.
- (69) Meier, E.; Frederiksen, D.; Nielsen, M.; Lemboel, H. L.; Pedersen, H.; Hyttel, J. Pharmacological in vitro characterization of the arecoline bioisostere, Lu 25-109-T, a muscarinic compound with M1-agonist and M2/M3-antagonist properties. *Drug Dev. Res.* 1997, 40, 1–16.
- (70) Hargreaves, R. J.; McKnight, A. T.; Scholey, K.; Newberry, N. R.; Street, L. J.; Hutson, P. H.; Semark, J. E.; Harley, E. A.; Patel, S.; Freedman, S. L-689,660, a novel cholinomimetic with functional selectivity for M1 and M3 muscarinic receptors. Br. J. Pharmacol. 1992, 107, 494-501.
- (71) Aagaard, P.; McKinney, M. Pharmacological characterization of the novel cholinomimetic L-689,660 at cloned and native brain muscarinic receptors. J. Pharmacol. Exp. Ther. 1993, 267, 1478– 83.
- (72) Ward, J. S.; Merritt, L.; Calligaro, D. O.; Bymaster, F. P.; Shannon, H. E.; Sawyer, B. D.; Mitch, C. H.; Deeter, J. B.; Peters, S. C.; Sheardown, M. J.; et al. Functionally selective M1 muscarinic agonists. 3. Side chains and azacycles contributing to functional muscarinic selectivity among pyrazinylazacycles. *J. Med. Chem.* 1995, 38, 3469–81.
- (73) Olesen, P. H.; Sauerberg, P.; Treppendahl, S.; Larsson, O.; Sheardown, M. J.; Suzdak, P. D.; Mitch, C. H.; Ward, J. S.; Bymaster, F. P.; Shannon, H. E.; Swedberg, M. D. B. 3-Alkylthio-1,2,5-thiadiazol-1-azabicycles, structure activity relationships for antinociception mediated by central muscarinic receptors. Eur. J. Med. Chem. 1996, 31, 221–230.
- (74) Shannon, H. E.; Sheardown, M. J.; Bymaster, F. P.; Calligaro, D. O.; Delapp, N. W.; Gidda, J.; Mitch, C. H.; Sawyer, B. D.; Stengel, P. W.; Ward, J. S.; Wong, D. T.; Olesen, P. H.; Suzdak, P. D.; Sauerberg, P.; Swedberg, M. D. Pharmacology of butylthio-[2.2.2] (LY297802/NNC11-1053): a novel analgesic with mixed muscarinic receptor agonist and antagonist activity. *J. Pharmacol. Exp. Ther.* 1997, 281, 884–94.
- (75) Sheardown, M. J.; Shannon, H. E.; Swedberg, M. D.; Suzdak, P. D.; Bymaster, F. P.; Olesen, P. H.; Mitch, C. H.; Ward, J. S.; Sauerberg, P. M1 receptor agonist activity is not a requirement for muscarinic antinociception. *J. Pharmacol. Exp. Ther.* 1997, 281, 868-75.
- (76) Bymaster, F. P.; Shannon, H. E.; Rasmussen, K.; Delapp, N. W.; Mitch, C. H.; Ward, J. S.; Calligaro, D. O.; Ludvigsen, T. S.; Sheardown, M. J.; Olesen, P. H.; Swedberg, M. D.; Sauerberg, P.; Fink-Jensen, A. Unexpected antipsychotic-like activity with the muscarinic receptor ligand (5R,6R)6-(3-propylthio-1,2,5-thiadiazol-4-yl)-1-azabicyclo[3.2.1]octane. Eur. J. Pharmacol. 1998, 356, 109-19.
- (77) Sauerberg, P.; Jeppesen, L.; Olesen, P. H.; Rasmussen, T.; Swedberg, M. D.; Sheardown, M. J.; Fink-Jensen, A.; Thomsen, C.; Thogersen, H.; Rimvall, K.; Ward, J. S.; Calligaro, D. O.; DeLapp, N. W.; Bymaster, F. P.; Shannon, H. E. Muscarinic agonists with antipsychotic-like activity: structure—activity relationships of 1,2,5-thiadiazole analogues with functional dopamine antagonist activity. *J. Med. Chem.* 1998, 41, 4378—84.
- (78) Brauner-Osborne, H.; Brann, M. R. Pharmacology of muscarinic acetylcholine receptor subtypes (m1-m5): high throughput assays in mammalian cells. Eur. J. Pharmacol. 1996, 295, 93– 102

- (79) Brauner-Osborne, H.; Ebert, B.; Brann, M. R.; Falch, E.; Krogsgaard-Larsen, P. Annulated heterocyclic bioisosteres of norarecoline. Synthesis and molecular pharmacology at five recombinant human muscarinic acetylcholine receptors. *J. Med. Chem.* 1995, 38, 2188–95.
- (80) Lenz, S. M.; Meier, E.; Pedersen, H.; Frederiksen, K.; Boegesoe, K. P.; Krogsgaard-Larsen, P. Muscarinic agonists. Syntheses and structure—activity relationships of bicyclic isoxazole ester bioisosteres of norarecoline. Eur. J. Med. Chem. 1995, 30, 263-70.
- (81) Sauerberg, P.; Olesen, P. H.; Sheardown, M. J.; Rimvall, K.; Thogersen, H.; Shannon, H. E.; Sawyer, B. D.; Ward, J. S.; Bymaster, F. P.; DeLapp, N. W.; Calligaro, D. O.; Swedberg, M. D. Conformationally constrained analogues of the muscarinic agonist 3-(4-(methylthio)-1,2,5-thiadiazol-3-yl)-1,2,5,6-tetrahydro-1-methylpyridine. Synthesis, receptor affinity, and antinociceptive activity. J. Med. Chem. 1998, 41, 109–16.
- ceptive activity. *J. Med. Chem.* **1998**, *41*, 109–16.

 (82) Messer, W. S.; Abuh, Y. F.; Ryan, K.; Shepheard, M. A.; Schroeder, M.; Abunada, S.; Sehgal, R.; El-Assadi, A. A. Tetrahydropyrimidine derivatives display functional selectivity for M1 muscarinic receptors in brain. *Drug Dev. Res.* **1997**, *40*, 171–84.
- (83) Xu, R.; Sim, M. K.; Go, M. L. Synthesis and pharmacological characterization of O-alkynyloximes of tropinone and N-methylpiperidinone as muscarinic agonists. J. Med. Chem. 1998, 41, 3220–31.
- (84) Tecle, H.; Barrett, S. D.; Lauffer, D. J.; Augelli-Szafran, C.; Brann, M. R.; Callahan, M. J.; Caprathe, B. W.; Davis, R. E.; Doyle, P. D.; Eubanks, D.; Lipiniski, W.; Mirzadegan, T.; Moos, W. H.; Moreland, D. W.; Nelson, C. B.; Pavia, M. R.; Raby, C.; Schwarz, R. D.; Spencer, C. J.; Thomas, A. J.; Jaen, J. C. Design and synthesis of m1-selective muscarinic agonists: (R)-(-)-(Z)-1-Azabicyclo[2.2.1]heptan-3-one, O-(3-(3'-methoxyphenyl)-2-propynyl)oxime maleate (CI-1017), a functionally m1-selective muscarinic agonist. J. Med. Chem. 1998, 41, 2524-36.
- (85) Schwarz, R.; et al. Development of M1 subtype selective muscarinic agonists for Alzheimer's disease: Translation of in vitro selectivity into in vivo efficacy. *Drug Dev. Res.* 1997, 40, 133– 143
- (86) Bromidge, S. M.; Brown, F.; Cassidy, F.; Clark, M. S. G.; Dabbs, S.; Hadley, M. S.; Hawkins, J.; Loudon, J. M.; Naylor, C. B.; Orlek, B. S.; Riley, G. J. Design of (R(Z))-(+)-alpha-(methoxy-imino)-1-azabicyclo(2.2.2)octane-3-acetonitrile (SB-202026), a functionally selective azabicyclic muscarinic M1 agonist incorporating the N-methoxy imidoyl nitrile group as a novel ester bioisostere. *J. Med. Chem,* 1997, 4265–80.
 (87) Loudon, J. M.; Bromidge, S. M.; Brown, F.; Clark, M. S.; Hatcher,
- (87) Loudon, J. M.; Bromidge, S. M.; Brown, F.; Clark, M. S.; Hatcher, J. P.; Hawkins, J.; Riley, G. J.; Noy, G.; Orlek, B. S. SB 202026: a novel muscarinic partial agonist with functional selectivity for M1 receptors. J. Pharmacol. Exp. Ther. 1997, 283, 1059–68.
- (88) Ensinger, H. A.; Doods, H. N.; Immel-Sehr, A. R.; Kuhn, F. J.; Lambrecht, G.; Mendla, K. D.; Muller, R. E.; Mutschler, E.; Sagrada, A.; Walther, G.; et al. WAL 2014 – a muscarinic agonist with preferential neuron-stimulating properties. *Life Sci.* 1993, 52, 473–80.
- (89) Pei, X. F.; Gupta, T. H.; Badio, B.; Padgett, W. L.; Daly, J. W. 6beta-Acetoxynortropane: a potent muscarinic agonist with apparent selectivity toward M2-receptors. J. Med. Chem. 1998, 41, 2047–55.
- (90) Sabb, A. L.; Stein, R. P.; Vogel, R. L.; Tasse, R.; Amburn, S.; Fairman, D. K.; Kowal, D.; Malhortra, D.; Boast, C. A.; Bartolomeo, D.; Morris, H.; Sailer, T.; Moyer, J. A.; Abou-Gharbia, M.; Ho, D. M. WAY-131256 is an orally active, efficacious, and in vivo functionally selective M1 agonist. *Drug Dev. Res.* 1997, 40, 185–92.
- (91) Ward, J. S.; Merritt, L.; Calligaro, D. O.; Bymaster, F. P.; Shannon, H. E.; Mitch, C. H.; Whitesitt, C.; Brunsting, D.; Sheardown, M. J.; Olesen, P. H.; Swedberg, M. D.; Jeppesen, L.; Sauerberg, P. 1,2,5-Thiadiazole analogues of aceclidine as potent m1 muscarinic agonists. *J. Med. Chem.* 1998, 41, 379– 92
- (92) Conti, P.; Dallanoce, C.; De Amici, M.; De Micheli, C.; Ebert, B. Synthesis and binding affinity of new muscarinic ligands structurally related to oxotremorine. *Bioorg. Med. Chem. Lett.* 1907, 7, 1033—6.
- 1997, 7, 1033-6.
 (93) Brass, E. P.; Polinsky, R.; Sramek, J. J.; Moore, M.; Jones, D.; Veroff, A. E.; Wardle, T. S.; Cutler, N. R. Effects of the cholinomimetic SDZ ENS-163 on scopolamine-induced cognitive impairment in humans. *J. Clin. Psychopharmacol.* 1995, *15*, 58-62.
- (94) Enz, A.; Boddeke, H.; Sauter, A.; Rudin, M.; Shapiro, G. SDZ ENS 163 a novel pilocarpine like drug: pharmacological in vitro and in vivo profile. *Life Sci.* **1993**, *52*, 513–20.
- (95) Jerusalinsky, D.; Harvey, A.; Karlsson, E.; Potter, L. Workshop: the use of muscarinic toxins in the study of muscarinic receptors. *Life Sci.* 1997, 60, 1161–2.
- (96) Jerusalinsky, D.; Kornisiuk, E.; Bernabeu, R.; Izquierdo, I.; Cervenansky, C. Muscarinic toxins from the venom of Dendroaspis snakes with agonist-like actions. *Toxicon* 1995, 33, 389–97.

- (97) Zorn, S. H.; Jones, S. B.; Ward, K. M.; Liston, D. R. Clozapine is a potent and selective muscarinic M4 receptor agonist. *Eur. J. Pharmacol.* **1994**, *269*, R1–2.
- (98) Zeng, X. P.; Le, F.; Richelson, E. Muscarinic m4 receptor activation by some atypical antipsychotic drugs. *Eur. J. Pharmacol.* **1997**, *321*, 349–54.

 (99) Olianas, M. C.; Maullu, C.; Onali, P. Mixed agonist-antagonist
- properties of clozapine at different human cloned muscarinic receptor subtypes expressed in Chinese hamster ovary cells. Neuropsychopharmacology **1999**, *20*, 263–70.
- (100) Wermuth, C. G.; Bourguignon, J. J.; Hohhmann, R.; Boigegrain, R.; Brodin, R.; Kan, J. P. SR 46559 A and related aminopyridazines are potent muscarinic agonists with no cholinergic syndrome. *Biomed. Med. Chem. Lett.* **1992**, *2*, 833–8.
- (101) Suzuki, T.; Oka, M.; Maeda, K.; Furusawa, K.; Mitani, T.; Kataoka, T. N-[2-(1-Azabicyclo[3.3.0]octan-5-yl)ethyl]-2-nitroanilime, a potent muscarinic agonist. Chem. Pharmacol. Bull. **1997**, 1218-20.
- (102) Suzuki, T.; Usui, T.; Oka, M.; Suzuki, T.; Kataoka, T. Synthesis and muscarinic activity of a series of quinolines and naphthalenes with a 1-azabicyclo[3.3.0]octane moiety. Chem. Pharmacol. Bull. **1998**, 46, 1265–73.
- Nason, D. M.; Lee, M. H.; Biggers, C. K.; Ward, K. M.; Nowakowski, J. T.; Yohannes, D.; Liston, D. R.; Villalobos, A. A novel structurally unique class of muscarinic agonists. Book of Abstracts; 212th ACS National Meeting, Orlando, FL, Aug 25 29, 1996; MEDI 081
- (104) Hollinshead, S. P.; Huff, B. E.; Hughes, P. F.; Mendoza, J. S.; Mitch, C. H.; Staszak, M. A.; Ward, J. S.; Wilson, J. W. Preparation of indanes as antipsychotics. Patent application: WO 97-US997 970122; Chem. Abstr. 1997, 127, 176278.
- (105) Trankle, C.; Andresen, I.; Lambrecht, G.; Mohr, K. M2 receptor binding of the selective antagonist AF-DX 384: possible involvement of the common allosteric site. Mol. Pharmacol. 1998, 53, 304 - 12.
- (106) Watson, N.; Reddy, H.; Stefanich, E.; Eglen, R. M. Characterization of the interaction of zamifenacin at muscarinic receptors in vitro. Eur. J. Pharmacol. 1995, 285, 135-42.
- (107) Gillberg, P. G.; Sundquist, S.; Nilvebrant, L. Comparison of the in vitro and in vivo profiles of tolterodine with those of subtypeselective muscarinic receptor antagonists. Eur. J. Pharmacol. **1998**, *349*, 285–92.
- (108) Nilvebrant, L.; Gillberg, P. G.; Sparf, B. Antimuscarinic potency and bladder selectivity of PNU-200577, a major metabolite of tolterodine. *Pharmacol. Toxicol.* **1997**, *81*, 169–72.
- (109) Nilvebrant, L.; Andersson, K. E.; Gillberg, P. G.; Stahl, M.; Sparf, B. Tolterodine - a new bladder-selective antimuscarinic agent. Eur. J. Pharmacol. **1997**, 327, 195–207.
- (110) Smith, C. M.; Wallis, R. M. Characterization of [3H]-darifenacin as a novel radioligand for the study of muscarinic M3 receptors. I. Recept. Signal. Transduct. Res. 1997, 17, 1−3.
- (111) Kaiser, C.; Audia, V. H.; Carter, J. P.; McPherson, D. W.; Waid, P. P.; Lowe, V. C.; Noronha-Blob, L. Synthesis and antimuscarinic activity of some 1-cycloalkyl-1-hydroxy-1-phenyl-3-(4substituted piperazinyl)-2-propanones and related compounds. *J. Med. Chem.* **1993**, *36*, 610–6.
- (112) Oyasu, H.; Yamamoto, T.; Sato, N.; Sawada, T.; Ozaki, R.; Mukai, T.; Ozaki, T.; Nishii, M.; Sato, H.; Fujiwara, T.; et al. Urinary bladder-selective action of the new antimuscarinic compound vamicamide. *Arzneim.-Forsch.* **1994**, *44*, 1242–9.
- (113) Miyachi, H.; Kiyota, H.; Segawa, M. Novel imidazole derivatives with subtype-selective antimuscarinic activity (1). Bioorg. Med. Chem. Lett. 1998, 8, 1807–12.
- (114) Miyachi, H.; Kiyota, H.; Segawa, M. Novel imidazole derivatives with subtype-selective antimuscarinic activity (2). Bioorg. Med. Chem. Lett. **1998**, *8*, 2163–8.
- (115) Pfaff, O.; Hildebrandt, C.; Waelbroeck, M.; Hou, X.; Moser, U.; Mutschler, E.; Lambrecht, G. The (S)-(+)-enantiomer of dimethindene: a novel M2-selective muscarinic receptor antagonist. Eur. J. Pharmacol. 1995, 286, 229-40.
- (116) Ghelardini, C.; Bartolini, A.; Galeotti, N.; Bellucci, C.; Dei, S.; Gualtieri, F. In vitro characterization of a novel, potent and selective M3 antagonist. Life Sci. 1997, 61, 1217-26.
- (117) D'Agostino, G.; Renzetti, A. R.; Zonta, F.; Subissi, A. Selectivity of LG50643 for postjunctional muscarinic-receptor subtype in the guinea-pig trachea. *J. Pharm. Pharmacol.* **1994**, *46*, 332–6. (118) Ghelardini, C.; Bartolini, A.; Galeotti, N.; Teodori, E.; Gualtieri,
- F. S-(-)-ET 126: a potent and selective M1 antagonist in vitro and in vivo. *Life Sci.* **1996**, *58*, 991–1000.
- (119) Scapecchi, S.; Angeli, P.; Dei, S.; Ghelardini, C.; Gualtieri, F.; Marucci, G.; Paparelli, F.; Romanelli, M. N.; Teodori, E. SAR studies on the potent and selective muscarinic antagonist 2-ethylthio-2,2-diphenylacetic acid N,N-diethylaminoethyl ester. Arch. Pharm. 1997, 330, 122–8. (120) Angeli, P.; Gualthieri, F.; Maggio, R.; Fiorella, P.; Scapecchi, S.
- Affinity profiles at five cloned human muscarinic receptors (m1m5) of a new series of antimuscarinic drugs. Pharm. Pharmacol. Lett. 1993, 3, 84-7.

- (121) Ghelardini, C.; Galeotti, N.; Gualtieri, F.; Marchese, V.; Bellucci, C.; Bartolini, A. Antinociceptive and antiamnesic properties of the presynaptic cholinergic amplifier PG-9. J. Pharmacol. Exp. Ther. 1998, 284, 806-16.
- Naito, R.; Takeuchi, M.; Morihira, K.; Hayakawa, M.; Ikeda, K.; Shibanuma, T.; Isomura, Y. Selective muscarinic antagonists. Synthesis and antimuscarinic properties of 4-piperidyl benzhydrylcarbamate derivatives. Chem. Pharm. Bull. 1998, 46, 1274 - 85
- (123) Naito, R.; Takeuchi, M.; Morihira, K.; Hayakawa, M.; Ikeda, K.; Shibanuma, T.; Isomura, Y. Selective muscarinic antagonists. II. Synthesis and antimuscarinic properties of biphenylylcarbamate derivatives. Chem. Pharm. Bull. 1998, 46, 1286-94.
- (124) Takeuchi, M.; Naito, R.; Yonetoku, Y.; Ikeda, K.; Isomura, Y. 1,2,3,4-Tetrahydro-2-isoquinoline carboxylate derivatives: a novel class of selective muscarinic antagonists. III. Abstr. Pap. Am. Chem. Soc. 1997, 213, 7722-7.
- (125) Melchiorre, C. Polymethylene tetraamines: a novel class of cardioselective M2-antagonists. Med. Res. Rev. 1990, 10, 327-
- Eberlein, W.; Engel, W.; Mihm, G.; Rudolf, K.; Wetzel, B.; Entzeroth, M.; Mayer, N.; Doods, H. Structure—activity relation-(126)ships and pharmacological profile of selective tricyclic antimuscarinics. Trends Pharmacol. Sci. 1989 (Suppl.), 50-4.
- Minarini, A.; Bolognesi, M. L.; Budriesi, R.; Canossa, M.; Chiarini, A.; Spampinato, S.; Melchiorre, C. Design, synthesis, and biological activity of methoctramine-related tetraamines bearing an 11-acetyl-5,11-dihydro-6H-pyrido[2,3-b][1,4]benzodiazepin-6-one moiety: structural requirements for optimum occupancy of muscarinic receptor subtypes as revealed by symmetrical and unsymmetrical polyamines. J. Med. Chem.
- 1994, *37*, 3363–72. (128) Maggio, R.; Barbier, P.; Bolognesi, M. L.; Minarini, A.; Tedeschi, receptor antagonist tripitramine. *Eur. J. Pharmacol.* **1994**, *268*, 459–62.
- (129) Melchiorre, C.; Minarini, A.; Budriesi, R.; Chiarini, A.; Spampinato, S.; Tumiatti, V. The design of novel methoctraminerelated tetraamines as muscarinic receptor subtype selective antagonists. *Life Sci.* **1995**, *56*, 837–44.

 (130) Bolognesi, M. L.; Minarini, A.; Budriesi, R.; Cacciaguerra, S.; Chiarini, A.; Spampinato, S.; Tumiatti, V.; Melchiorre, C.
- Universal template approach to drug design: polyamines as selective muscarinic receptor antagonists. *J. Med. Chem.* **1998**, *41*, 4150-60
- (131) Watanabe, T.; Kakefuda, A.; Tanaka, A.; Takizawa, K.; Hirano, S.; Shibata, H.; Yamagiwa, Y.; Yanagisawa, I. Synthesis and biological evaluation of phenylacetyl derivatives having low central nervous system permeability as potent and selective M2 muscarinic receptor antagonists. Chem. Pharm. Bull. 1998, 46,
- (132) Watanabe, T.; Kakefuda, A.; Kinoyama, I.; Takizawa, K.; Hirano, S.; Shibata, H.; Yanagisawa, I. Synthesis of novel succinamide derivatives having a 5,11-dihydro-6H-pyrido[2,3-b][1,4]benzodiazepin-6-one skeleton as potent and selective M2 muscarinic receptor antagonists. II. *Chem. Pharm. Bull.* **1997**, *45*, 1458–
- (133) Watanabe, T.; Kinoyama, I.; Kakefuda, A.; Okazaki, T.; Takizawa, K.; Hirano, S.; Shibata, H.; Yanagisawa, I. Synthesis of novel succinamide derivatives having the 5,11-dihydro-6Hpyrido[2,3-b][1,4]benzodiazepin-6-one skeleton as potent and selective M2 muscarinic receptor antagonists. I. Chem. Pharm. Bull. **1997**, 45, 996–1007. (134) Gitler, M. S.; Cohen, V. I.; De la Cruz, R.; Boulay, S. F.; Jin, B.;
- Zeeberg, B. R.; Reba, R. C. A novel muscarinic receptor ligand which penetrates the blood brain barrier and displays in vivo selectivity for the m2 subtype. *Life Sci.* **1993**, *53*, 1743–51. Van Hijfte, L.; Zerr, V.; Richards, M. H.; Moser, P.; Hibert, M.
- F.; van Giersbergen, P. L. M. Discovery of a new class of centrally active M1 selective muscarinic antagonists related to pirenzepine. Med. Chem. Res. 1996, 6, 190-6.
- (136) Tacke, R.; Pikies, J.; Wiesenberger, F.; Ernst, L.; Schomburg, D.; Waelbroeck, M.; Christophe, J.; Lambrecht, G.; Gross, J.; et al. Sila-biperiden and endo-sila-biperiden: syntheses, crystal structure, and antimuscarinic properties. *J. Organomet. Chem.*
- 1994, 466, 15-27.
 Tacke, R.; Reichel, D.; Kropfgans, M.; Jones, P. G.; Mutschler, E.; Gross, J.; Hou, X.; Waelbroeck, M.; Lambrecht, G. Biological Recognition of Enantiomeric Silanes: Syntheses and Antimuscarinic Properties of Optically Active (2-Aminoethyl)cyclohexyl-(hydroxymethyl)phenylsilanes and Related Quaternary Ammonium Derivatives. *Organometallics* **1995**, *14*, 251–62. Tacke, R.; Forth, B.; Waelbroeck, M.; Gross, J.; Mutschler, E.;
- Lambrecht, G. Unsaturated derivatives of the muscarinic antagonists hexahydro-sila- difenidol (HHSiD) and p-fluorohexahydro-sila-difenidol (p-F-HHSiD) with an (E)-Si-CH:CH-CH2-N moiety: syntheses and binding affinities at muscarinic receptor subtypes. J. Organomet. Chem. 1995, 505, 73-9.

- (139) Shannon, H. E.; Bymaster, F. P.; Hendrix, J. C.; Quimby, S. J.; Mitch, C. H. Novel alkoxy-oxazolyl-tetrahydropyridine muscarinic cholinergic receptor antagonists. *Psychopharmacology* **1995**, *117*, 208–15.
- (140) Johansson, G.; Sundquist, S.; Nordvall, G.; Nilsson, B. M.; Brisander, M.; Nilvebrant, L.; Hacksell, U. Antimuscarinic 3-(2-furanyl)quinuclidin-2-ene derivatives: synthesis and structure—activity relationships. *J. Med. Chem.* 1997, 40, 3804–19.
- (141) Malaska, M. J.; Fauq, A. H.; Kozikowski, A. P.; Aagaard, P. J.; McKinney, M. Chemical modification of ring C of himbacine: Discovery of a pharmacophoric element for M2-selectivity. *Bioorg. Med. Chem. Lett.* 1995, 5, 61–6.
- (142) Chackalamannil, S.; Davies, R. J.; Wang, Y.; Asberom, T.; Doller, D.; Wong, J.; Leone, D.; McPhail, A. T. Total synthesis of (+)-himbacine and (+)-himbeline. *J. Org. Chem.* **1999**, *64*, 1932–40.
- (143) Lin, C. H.; Chang, G. J.; Su, M. J.; Wu, Y. C.; Teng, C. M.; Ko, F. N. Pharmacological characteristics of liriodenine, isolated from Fissistigma glaucescens, a novel muscarinic receptor antagonist in guinea-pigs. Br. J. Pharmacol. 1994, 113, 275–81.
- (144) Baumgold, J.; Pryzbyc, R. L.; Reba, R. C. 3-alpha-Chlorimperialine: An M-2-selective muscarinic receptor antagonist that penetrates into brain. *Eur. J. Pharmacol.* **1994**, *251*, 2–3.
- (145) Augelli-Szafran, C. E.; Jaen, J. C.; Moreland, D. W.; Nelson, C. B.; Penvose-Yi, J. R.; Schwarz, R. D. Identification and characterization of m4 selective muscarinic antagonists. *Bioorg. Med. Chem. Lett.* 1998, 8, 1991–6.
- (146) Karlsson, E.; Jolkkonen, M.; Satyapan, N.; Adem, A.; Kumlin, E.; Hellman, U.; Wernstedt, C. Protein toxins that bind to muscarinic acetylcholine receptors. *Ann. N. Y. Acad. Sci.* 1994, 9, 153–61.
- (147) Jolkkonen, M.; van Giersbergen, P. L.; Hellman, U.; Wernstedt, C.; Karlsson, E. A toxin from the green mamba *Dendroaspis* angusticeps: amino acid sequence and selectivity for muscarinic m4 receptors. *FEBS Lett.* **1994**, *352*, 91–4.
- (148) Olianas, M. C.; Adem, A.; Karlsson, E.; Onali, P. Identification of rat brain muscarinic M4 receptors coupled to cyclic AMP using the selective antagonist muscarinic toxin 3. *Eur. J. Pharmacol.* **1998**, *357*, 235–42.
- (149) Purkerson, S. L.; Potter, L. T. Use of antimuscarinic toxins to facilitate studies of striatal m4 muscarinic receptors. *J. Pharmacol. Exp. Ther.* **1998**, *284*, 707–13.
- (150) Ellis, J. Állosteric binding sites on muscarinic receptors. *Drug Dev. Res.* 1997, 40, 193–204.
- (151) Lee, N. H.; el-Fakahany, E. E. Allosteric interactions at the m1, m2 and m3 muscarinic receptor subtypes. *J. Pharmacol. Exp. Ther.* 1991, 256, 468-79.
- (152) Jakubik, J.; Bacakova, L.; el-Fakahany, E. E.; Tucek, S. Subtype selectivity of the positive allosteric action of alcuronium at cloned M1-M5 muscarinic acetylcholine receptors. *J. Pharmacol. Exp. Ther.* 1995, 274, 1077–83.
- (153) Jakubik, J.; Bacakova, L.; El-Fakahany, E. E.; Tucek, S. Positive cooperativity of acetylcholine and other agonists with allosteric ligands on muscarinic acetylcholine receptors. *Mol. Pharmacol.* 1997, 52, 172–9.
- (154) Lazareno, S.; Gharagozloo, P.; Kuonen, D.; Popham, A.; Birdsall, N. J. Subtype-selective positive cooperative interactions between brucine analogues and acetylcholine at muscarinic receptors: radioligand binding studies. *Mol. Pharmacol.* 1998, 53, 573–89; published erratum appears in *Mol. Pharmacol.* 1999, 55, 194.
- 89; published erratum appears in *Mol. Pharmacol.* **1999**, *55*, 194. (155) Lazareno, S.; Birdsall, N. J. Detection, quantitation, and verification of allosteric interactions of agents with labeled and unlabeled ligands at G protein-coupled receptors: interactions of strychnine and acetylcholine at muscarinic receptors. *Mol. Pharmacol.* **1995**, *48*, 362–78.
- (156) Lazareno, S.; Popham, A.; Birdsall, N. J. Allosteric interactions of staurosporine and other indolocarbazoles with N-[methyl-(3)H]scopolamine and acetylcholine at muscarinic receptor subtypes: identification of a second allosteric site. *Mol. Pharmacol.* 2000, 58, 194–207.
- (157) Creese, I.; Burt, D. R.; Snyder, S. H. Dopamine receptor binding predicts clinical and pharmacological potencies of antischizophrenic drugs. *Science* 1976, 192, 481–3.
- (158) Abood, L. G., Biel, J. H. Anticholinergic psychotomimetic agents. Int. Rev. Neurobiol. 1962, 4, 217–73.
- (159) Granacher, R. P.; Baldessarini, R. J. Physostigmine. Its use in acute anticholinergic syndrome with antidepressant and antiparkinson drugs. *Arch. Gen. Psychiatry* 1975, 32, 375–80.
- (160) Baumgold, J.; Abood, L. G.; Aronstam, R. Studies on the relationship of binding affinity to psychoactive and anticholingergic potency of a group of psychotomimetic glycolates. *Brain Res.* **1977**, *124*, 331–40.
- (161) Mego, D. M.; Omori, J. M.; Hanley, J. F. Transdermal scopolamine as a cause of transient psychosis in two elderly patients. S. Med. J. 1988, 81, 394–5.

- (162) Fisher, C. M. Visual hallucinations on eye closure associated with atropine toxicity. A neurological analysis and comparison with other visual hallucinations. Can. J. Neurol. Sci. 1991, 18, 18– 27
- (163) Smith, J. M. Abuse of antiparkinson drugs: A review of the literature. J. Clin. Psychiatry 1980, 41, 351-4.
- literature. *J. Clin. Psychiatry* **1980**, *41*, 351–4. (164) Fisch, R. Z. Trihexyphenidyl abuse: Therapeutic implications for negative symptoms of schizophrenia? *Acta Psychiatric Scand.* **1987**, *75*, 91–94.
- (165) Seeman, P. Dopamine receptors and the dopamine hypothesis of schizophrenia. *Synapse* **1987**, *1*, 133–52.
- (166) Carlsson, A. Antipsychotic drugs, neurotransmitters, and schizophrenia. Am. J. Psychiatry 1978, 135, 165-73.
 (167) Cohen, L. H.; Thale, T.; Tissenbaum, M. J. Acetylcholine
- (167) Cohen, L. H.; Thale, T.; Tissenbaum, M. J. Acetylcholine treatment of schizophrenia. Arch. Neurol. Psychiatry 1944, 51, 171–5
- (168) Pfeiffer, C. C.; Jenney, E. H. The inhibition of conditional response and counter action of schizophrenia by muscarinic stimulation of the brain. Ann. N. Y. Acad. Sci. 1957, 66, 753– 64
- (169) Collard, J.; Lecog, R.; Demaret, A. Un assai de therapeutique, pathogenique de la schizophrenie par un acetylcholonique l'oxotremorine. *Acta Neurol. Belg.* 1965, 65, 122-7.
 (170) Rosenthal, R.; Bigelow, L. B. The effects of physostigmine in
- (170) Rosenthal, R.; Bigelow, L. B. The effects of physostigmine in phenothiazine resistant chronic schizophrenic patients: preliminary observations. *Comp. Psychiatry* 1973, *14*, 489–94.
 (171) Janowsky, D. S.; el-Yousef, M. K.; Davis, J. M.; Sekerke, H. J.
- (171) Janowsky, D. S.; el-Yousef, M. K.; Davis, J. M.; Sekerke, H. J. Antagonistic effects of physostigmine and methylphenidate in man. Am. J. Psychiatry 1973, 130, 1370–6.
- (172) Friedhoff, A. J.; Alpert, M. A dopaminergic-cholinergic mechanism in the production of psychotic symptoms. *Biol. Psychiatry* 1973, 6, 165–9.
- (173) McGeer, P. L.; Grewaal, D. S.; McGeer, E. G. Influence of noncholinergic drugs on rat striatal acetylcholine levels. *Brain Res.* 1974, 80, 211-7.
- (174) Bymaster, F. P.; Reid, L. R.; Nichols, C. L.; Kornfeld, E. C.; Wong, D. T. Elevation of acetylcholine levels in striatum of rat brain by LY163502, trans-(-)-5,5a,6,7,8,9a,10-octahydro-6-propylpyrimido less than 4,5-g greater than quinolin-2-amine dihydrochloride, a potent and stereospecific dopamine (D2) agonist. *Life Sci.* 1986, *38*, 317–22.
- (175) Bymaster, F. P.; Wong, D. T.; Mitch, C. H.; Ward, J. S.; Calligaro, D. O.; Schoepp, D. D.; Shannon, H. E.; Sheardown, M. J.; Olesen, P. H.; Suzdak, P. D.; et al. Neurochemical effects of the M1 muscarinic agonist xanomeline (LY246708/NNC11-0232). *J. Pharmacol. Exp. Ther.* 1994, 269, 282-9.
 (176) Lacey, M. G.; Mercuri, N. B.; North, R. A. Actions of cocaine on
- (176) Lacey, M. G.; Mercuri, N. B.; North, R. A. Actions of cocaine on rat dopaminergic neurones in vitro. Br. J. Pharmacol. 1990, 99, 731–5
- (177) Blaha, C. D.; Allen, L. F.; Das, S.; Inglis, W. L.; Latimer, M. P.; Vincent, S. R.; Winn, P. Modulation of dopamine efflux in the nucleus accumbens after cholinergic stimulation of the ventral tegmental area in intact, pedunculopontine tegmental nucleuslesioned, and laterodorsal tegmental nucleus-lesioned rats. *J. Neurosci.* 1996, 16, 714–22.
- (178) Marchi, M.; Raiteri, M. On the presence in the cerebral cortex of muscarinic receptor subtypes which differ in neuronal localization, function and pharmacological properties. *J. Pharmacol. Exp. Ther.* **1985**, 230–3.
- (179) Niedzwiecki, D. M.; Cubeddu, L. X.; Mailman, R. B. Comparative antidopaminergic properties of thioridazine, mesoridazine and sulforidazine on the corpus striatum. *J. Pharmacol. Exp. Ther.* **1989**, *250*, 117–25.
- (180) Xu, M.; Mizobe, F.; Yamamoto, T.; Kato, T. Differential effects of M1- and M2-muscarinic drugs on striatal dopamine release and metabolism in freely moving rats. *Brain Res.* 1989, 495, 232–42.
- (181) Bodick, N. C.; Offen, W. W.; Levey, A. I.; Cutler, N. R.; Gauthier, S. G.; Satlin, A.; Shannon, H. E.; Tollefson, G. D.; Rasmussen, K.; Bymaster, F. P.; Hurley, D. J.; Potter, W. Z.; Paul, S. M. Effects of xanomeline, a selective muscarinic receptor agonist, on cognitive function and behavioral symptoms in Alzheimer disease. Arch. Neurol. 1997, 54, 465-73.
- (182) Levey, A. I. Immunological localization of m1-m5 muscarinic acetylcholine receptors in peripheral tissues and brain. *Life Sci.* 1993, 52, 441-8.
- (183) Weiner, D. M.; Levey, A. I.; Brann, M. R. Expression of muscarinic acetylcholine and dopamine receptor mRNAs in rat basal ganglia. *Proc. Natl. Acad. Sci. U.S.A.* 1990, 87, 7050–4.
- (184) Bernard, V.; Normand, E.; Bloch, B. Phenotypical characterization of the rat striatal neurons expressing muscarinic receptor genes. J. Neurosci. 1992, 12, 3591–600.
- (185) Vilaro, M. T.; Wiederhold, K. H.; Palacios, J. M.; Mengod, G. Muscarinic cholinergic receptors in the rat caudate-putamen and olfactory tubercle belong predominantly to the m4 class: in situ hybridization and receptor autoradiography evidence. Neuroscience 1991, 40, 159–67.

- (186) Yeomans, J. S. Role of tegmental cholinergic neurons in dopaminergic activation, antimuscarinic psychosis and schizophrenia. Neuropsychopharmacology 1995, 12, 3–16.
- (187) Clarke, P. B.; Hommer, D. W.; Pert, A.; Skirboll, L. R. Innervation of substantia nigra neurons by cholinergic afferents from pedunculopontine nucleus in the rat: neuroanatomical and electrophysiological evidence. *Neuroscience* 1987, 23, 1011–9.
- (188) Gould, E.; Woolf, N. J.; Butcher, L. L. Cholinergic projections to the substantia nigra from the pedunculopontine and laterodorsal tegmental nuclei. *Neuroscience* **1989**, *28*, 611–23.
- (189) Woolf, N. J.; Harrison, J. B.; Buchwald, J. S. Cholinergic neurons of the feline pontomesencephalon. II. Ascending anatomical projections. *Brain Res.* **1990**, *520*, 55–72.
- (190) Bolam, J. P.; Francis, C. M.; Henderson, Z. Cholinergic input to dopaminergic neurons in the substantia nigra: a double immunocytochemical study. *Neuroscience* 1991, 41, 483–94.
- (191) Karson, C. N.; Garcia-Rill, E.; Biedermann, J.; Mrak, R. E.; Husain, M. M.; Skinner, R. D. The brain stem reticular formation in schizophrenia. *Psychiatry Res.* 1991, 40, 31–48.
- (192) Haroutunian, V.; Davidson, M.; Kanof, P. D.; Perl, D. P.; Powchik, P.; Losonczy, M.; McCrystal, J.; Purohit, D. P.; Bierer, L. M.; Davis, K. L. Cortical cholinergic markers in schizophrenia. Schizophrenia Res. 1994, 12, 137–44.
- (193) Levey, A. I.; Kitt, C. A.; Simonds, W. F.; Price, D. L.; Brann, M. R. Identification and localization of muscarinic acetylcholine receptor proteins in brain with subtype-specific antibodies. *J. Neurosci.* 1991, 11, 3218–26.
- (194) Wei, J.; Walton, E. A.; Milici, A.; Buccafusco, J. J. m1-m5 muscarinic receptor distribution in rat CNS by RT-PCR and HPLC. J. Neurochem., 1994, 63, 815-21.
- (195) Brann, M. R.; Ellis, J.; Jorgensen, H.; Hill-Eubanks, D.; Jones, S. V. Muscarinic acetylcholine receptor subtypes: localization and structure/function. *Prog. Brain Res.* 1993, 98, 121-7.
- (196) Drevets, W. C.; Rubin, E. H. Psychotic symptoms and the longitudinal course of senile dementia of the Alzheimer type. *Biol. Psychiatry* 1989, 25, 39–48.
- (197) Bartus, R. T.; Dean, R. L. d.; Beer, B.; Lippa, A. S. The cholinergic hypothesis of geriatric memory dysfunction. *Science* 1982, 217, 408–14.
- (198) Teri, L.; Borson, S.; Kiyak, H. A.; Yamagishi, M. Behavioral disturbance, cognitive dysfunction, and functional skill. Prevalence and relationship in Alzheimer's disease. *J. Am. Geriatrics Soc.* 1989, 37, 109–16.
- (199) Hardy, J.; Allsop, D. Amyloid deposition as the central event in the aetiology of Alzheimer's disease. *Trends Pharmacol. Sci.* **1991**, *12*, 383–8.
- (200) Selkoe, D. J. The molecular pathology of Alzheimer's disease. Neuron 1991, 6, 487–98.
- (201) Price, D. L.; Sisodia, S. S. Cellular and molecular biology of Alzheimer's disease and animal models. *Annu. Rev. Med.* 1994, 45, 435–46.
- (202) Aisen, P. S.; Davis, K. L. Inflammatory mechanisms in Alzheimer's disease: implications for therapy. Am. J. Psychiatry 1994, 151, 1105–13.
- (203) Nitsch, R. M. From acetylcholine to amyloid: neurotransmitters and the pathology of Alzheimer's disease. *Neurodegeneration* 1996, 5, 477–82.
- (204) Mesulam, M. M.; Mufson, E. J.; Levey, A. I.; Wainer, B. H. Cholinergic innervation of cortex by the basal forebrain: cytochemistry and cortical connections of the septal area, diagonal band nuclei, nucleus basalis (substantia innominata), and hypothalamus in the rhesus monkey. *J. Comp. Neurol.* 1983, 214, 170–97.
- (205) Mesulam, M. M.; Geula, C. Nucleus basalis (Ch4) and cortical cholinergic innervation in the human brain: observations based on the distribution of acetylcholinesterase and choline acetyltransferase. *J. Comp. Neurol.* **1988**, *275*, 216–40.
- (206) Mesulam, M. M.; Hersh, L. B.; Mash, D. C.; Geula, C. Differential cholinergic innervation within functional subdivisions of the human cerebral cortex: a choline acetyltransferase study. *J. Comp. Neurol.* **1992**, *318*, 316–28.
- (207) Hedreen, J. C.; Struble, R. G.; Whitehouse, P. J.; Price, D. L. Topography of the magnocellular basal forebrain system in human brain. *J. Neuropathol. Exp. Neurol.* **1984**, 43, 1–21.
- human brain. *J. Neuropathol. Exp. Neurol.* **1984**, *43*, 1–21. (208) Hyman, B. T.; Van Hoesen, G. W.; Damasio, A. R. Memory-related neural systems in Alzheimer's disease: an anatomic study. *Neurology* **1990**, *40*, 1721–30.
- study. Neurology 1990, 40, 1721–30.

 (209) Benzing, W. C.; Mufson, E. J.; Armstrong, D. M. Immunocytochemical distribution of peptidergic and cholinergic fibers in the human amygdala: their depletion in Alzheimer's disease and morphologic alteration in nondemented elderly with numerous senile plaques. Brain Res. 1993, 625, 125–38.
- (210) Emre, M.; Heckers, S.; Mash, D. C.; Geula, C.; Mesulam, M. M. Cholinergic innervation of the amygdaloid complex in the human brain and its alterations in old age and Alzheimer's disease. *J. Comp. Neurol.* 1993, 336, 117–34.

- (211) Hof, P. R.; Cox, K.; Morrison, J. H. Quantitative analysis of a vulnerable subset of pyramidal neurons in Alzheimer's disease: I. Superior frontal and inferior temporal cortex. *J. Comp. Neurol.* 1990, 301, 44–54.
- (212) Masliah, E.; Miller, A.; Terry, R. D. The synaptic organization of the neocortex in Alzheimer's disease. *Med. Hypoth.* 1993, 41, 334–40.
- (213) Samuel, W.; Terry, R. D.; DeTeresa, R.; Butters, N.; Masliah, E. Clinical correlates of cortical and nucleus basalis pathology in Alzheimer dementia. *Arch. Neurol.* 1994, 51, 772–8.
- (214) Davies, P.; Maloney, A. J. Selective loss of central cholinergic neurons in Alzheimer's disease [letter]. Lancet 1976, 2, 1403.
- (215) Whitehouse, P. J.; Price, D. L.; Struble, R. G.; Clark, A. W.; Coyle, J. T.; Delon, M. R. Alzheimer's disease and senile dementia: loss of neurons in the basal forebrain. *Science* 1982, 215, 1237–9.
- (216) Araujo, D. M.; Lapchak, P. A.; Robitaille, Y.; Gauthier, S.; Quirion, R. Differential alteration of various cholinergic markers in cortical and subcortical regions of human brain in Alzheimer's disease. J. Neurochemistry 1988, 50, 1914–23.
- (217) Nagai, T.; McGeer, P. L.; Peng, J. H.; McGeer, E. G.; Dolman, C. E. Choline acetyltransferase immunohistochemistry in brains of Alzheimer's disease patients and controls. *Neuroscience Lett.* **1983**, *36*, 195–9.
- (218) Coyle, J. T.; Price, D. L.; DeLong, M. R. Alzheimer's disease: a disorder of cortical cholinergic innervation. *Science* 1983, 219, 1184–90.
- (219) Bierer, L. M.; Haroutunian, V.; Gabriel, S.; Knott, P. J.; Carlin, L. S.; Purohit, D. P.; Perl, D. P.; Schmeidler, J.; Kanof, P.; Davis, K. L. Neurochemical correlates of dementia severity in Alzheimer's disease: relative importance of the cholinergic deficits. J. Neurochem. 1995, 64, 749-60.
- (220) Giacobini, E. Invited review: Cholinesterase inhibitors for Alzheimer's disease therapy: from tacrine to future applications. *Neurochem. Int.* 1998, 32, 413–9.
- (221) Krall, W. J.; Sramek, J. J.; Cutler, N. R. Cholinesterase inhibitors: a therapeutic strategy for Alzheimer disease. *Ann. Pharmacother.* **1999**, *33*, 441–50.
- (222) Ladner, C. J.; Lee, J. M. Pharmacological drug treatment of Alzheimer disease: the cholinergic hypothesis revisited [see comments]. J. Neuropathol. Exp. Neurol. 1998, 57, 719–31.
- (223) Fisher, A. Muscarinic receptor agonists in Alzheimer's disease more than just symptomatic treatment? CNS Drugs 1999, 12, 197–214
- (224) Murray, C. L.; Fibiger, H. C. Learning and memory deficits after lesions of the nucleus basalis magnocellularis: reversal by physostigmine. *Neuroscience* 1985, 14, 1025–32.
 (225) Mandel, R. J.; Thal, L. J. Physostigmine improves water maze
- (225) Mandel, R. J.; Thal, L. J. Physostigmine improves water maze performance following nucleus basalis magnocellularis lesions in rats. *Psychopharmacology* 1988, 96, 421–5.
- (226) Flicker, C.; Dean, R. L.; Watkins, D. L.; Fisher, S. K.; Bartus, R. T. Behavioral and neurochemical effects following neurotoxic lesions of a major cholinergic input to the cerebral cortex in the rat. *Pharmacol. Biochem. Behav.* **1983**, *18*, 973–81.
- (227) Walsh, T. J.; Tilson, H. A.; DeHaven, D. L.; Mailman, R. B.; Fisher, A.; Hanin, I. AF64A, a cholinergic neurotoxin, selectively depletes acetylcholine in hippocampus and cortex, and produces long-term passive avoidance and radial-arm maze deficits in the rat. *Brain Res.* 1984, 321, 91–102.
- (228) Kesner, R. P.; Adelstein, T.; Crutcher, K. A. Rats with nucleus basalis magnocellularis lesions mimic mnemonic symptomatology observed in patients with dementia of the Alzheimer's type. *Behav. Neurosci.* **1987**, *101*, 451–6.
- (229) Mallet, P. E.; Beninger, R. J.; Flesher, S. N.; Jhamandas, K.; Boegman, R. J. Nucleus basalis lesions: implication of basoamygdaloid cholinergic pathways in memory. *Brain Res. Bull.* 1995, 36, 51–6.
- (230) Winkler, J.; Suhr, S. T.; Gage, F. H.; Thal, L. J.; Fisher, L. J. Essential role of neocortical acetylcholine in spatial memory [see comments]. *Nature* 1995, 375, 484-7.
- (231) Gauthier, S.; Bouchard, R.; Lamontagne, A.; Bailey, P.; Bergman, H.; Ratner, J.; Tesfaye, Y.; Saint-Martin, M.; Bacher, Y.; Carrier, L.; et al. Tetrahydroaminoacridine-lecithin combination treatment in patients with intermediate-stage Alzheimer's disease. Results of a Canadian double-blind, crossover, multicenter study [see comments]. N. Engl. J. Med. 1990, 322, 1272-6.
- (232) Eagger, S. A.; Levy, R.; Sahakian, B. J. Tacrine in Alzheimer's disease. Acta Neurol. Scand. Suppl. 1992, 139, 75–80.
- (233) Davis, R. E.; Doyle, P. D.; Carroll, R. T.; Emmerling, M. R.; Jaen, J. Cholinergic therapies for Alzheimer's disease. Palliative or disease altering? Arzneim.-Forsch. 1995, 45, 425–31.
- (234) Rogers, S. L. Perspectives in the management of Alzheimer's disease: clinical profile of donepezil. *Dementia Geriatr. Cognit. Disord.* 1998, 9 (Suppl. 3), 29–42.
 (235) Davis, K. L.; Mohs, R. C.; Tinklenberg, J. R.; Pfefferbaum, A.;
- (235) Davis, K. L.; Mohs, R. C.; Tinklenberg, J. R.; Pfefferbaum, A.; Hollister, L. E.; Kopell, B. S. Physostigmine: improvement of long-term memory processes in normal humans. *Science* 1978, 201, 272–4.

- (236) Stern, Y.; Sano, M.; Mayeux, R. Long-term administration of oral physostigmine in Alzheimer's disease. Neurology 1988, 38,
- (237) Jotkowitz, S. Lack of clinical efficacy of chronic oral physostig-
- mine in Alzheimer's disease. *Ann.Neurol.*, **1983**, *14*, 690–1. (238) Bymaster, F. P.; Heath, I.; Hendrix, J. C.; Shannon, H. E. Comparative behavioral and neurochemical activities of cholinergic antagonists in rats. J. Pharmacol. Exp. Ther. 1993, 267, 16-24.
- (239) Andrews, J. S.; Jansen, J. H.; Linders, S.; Princen, A. Effects of disrupting the cholinergic system on short-term spatial memory in rats. Psychopharmacology 1994, 115, 485-94.
- (240) Ohno, M.; Yamamoto, T.; Watanabe, S. Blockade of hippocampal M1 muscarinic receptors impairs working memory performance of rats. *Brain Res.* **1994**, *650*, 260–6.
- (241) Ukai, M.; Shinkai, N.; Kameyama, T. Cholinergic receptor agonists inhibit pirenzepine-induced dysfunction of spontaneous alternation performance in the mouse. Gen. Pharmacol. 1995, 26, 1529-32
- (242) Drachman, D. A.; Leavitt, J. Human memory and the cholinergic system. A relationship to aging? Arch. Neurol. 1974, 30, 113-
- (243) Levey, A. I. Muscarinic acetylcholine receptor expression in memory circuits: implications for treatment of Alzheimer disease. *Proc. Natl. Acad. Sci. U.S.A.* **1996**, *93*, 13541–6.
- (244) Marino, M. J.; Rouse, S. T.; Levey, A. I.; Potter, L. T.; Conn, P. J. Activation of the genetically defined m1 muscarinic receptor potentiates N-methyl-D-aspartate (NMDA) receptor currents in hippocampal pyramidal cells. Proc. Natl. Acad. Sci. U.S.A. 1998, 95, 11465-70.
- (245) Mash, D. C.; Flynn, D. D.; Potter, L. T. Loss of M2 muscarine receptors in the cerebral cortex in Alzheimer's disease and experimental cholinergic denervation. Science 1985, 228, 1115-
- (246) Penn, R. D.; Martin, E. M.; Wilson, R. S.; Fox, J. H.; Savoy, S. M. Intraventricular bethanechol infusion for Alzheimer's disease: results of double-blind and escalating-dose trials. Neurology 1988, 38, 219-22.
- (247) Tariot, P. N.; Cohen, R. M.; Welkowitz, J. A.; Sunderland, T.; Newhouse, P. A.; Murphy, D. L.; Weingartner, H. Multiple-dose arecoline infusions in Alzheimer's disease. Arch. Gen. Psychiatry **1988**, 45, 901-5.
- (248) Flynn, D. D.; Ferrari-DiLeo, G.; Mash, D. C.; Levey, A. I. Differential regulation of molecular subtypes of muscarinic receptors in Alzheimer's disease. J. Neurochem. 1995, 64, 1888-
- (249) Jope, R. S.; Song, L.; Powers, R. E. Cholinergic activation of phosphoinositide signaling is impaired in Alzheimer's disease brain. *Neurobiol. Aging* 1997, *18*, 111–20.
 (250) Crews, F. T.; Kurian, P.; Freund, G. Cholinergic and serotonergic
- stimulation of phosphoinositide hydrolysis is decreased in Alzheimer's disease. *Life Sci.* **1994**, *55*, 1993–2002.

 (251) Ferrari-DiLeo, G.; Mash, D. C.; Flynn, D. D. Attenuation of
- muscarinic receptor-G-protein interaction in Alzheimer disease.
- *Mol. Chem. Neuropathol.* **1995**, *24*, 69–91. (252) Ladner, C. J.; Lee, J. M. Reduced high-affinity agonist binding at the M(1) muscarinic receptor in Alzheimer's disease brain: differential sensitivity to agonists and divalent cations. Exp. Neurol. **1999**, 158, 451–8.
- (253) Nitsch, R. M.; Slack, B. E.; Wurtman, R. J.; Growdon, J. H. Release of Alzheimer amyloid precursor derivatives stimulated by activation of muscarinic acetylcholine receptors. Science 1992, *258*, 304–
- (254) Hung, A. Y.; Haass, C.; Nitsch, R. M.; Qiu, W. Q.; Citron, M.; Wurtman, R. J.; Growdon, J. H.; Selkoe, D. J. Activation of protein kinase C inhibits cellular production of the amyloid betaprotein. J. Biol. Chem. 1993, 268, 22959-62.

- (255) DeLapp, N.; Wu, S.; Belagaje, R.; Johnstone, E.; Little, S.; Shannon, H.; Bymaster, F.; Calligaro, D.; Mitch, C.; Whitesitt, C.; Ward, J.; Sheardown, M.; Fink-Jensen, A.; Jeppesen, L.; Thomsen, C.; Sauerberg, P. Effects of the M1 agonist xanomeline on processing of human beta-amyloid precursor protein (FAD, Swedish mutant) transfected into Chinese hamster ovary-m1 cells. Biochem. Biophys. Res. Commun. 1998, 244, 156-60.
- (256) Farber, S. A.; Nitsch, R. M.; Schulz, J. G.; Wurtman, R. J. Regulated secretion of beta-amyloid precursor protein in rat brain. J. Neurosci. 1995, 15, 7442-51.
- Haroutunian, V.; Greig, N.; Pei, X. F.; Utsuki, T.; Gluck, R.; Acevedo, L. D.; Davis, K. L.; Wallace, W. C. Pharmacological modulation of Alzheimer's beta-amyloid precursor protein levels in the CSF of rats with forebrain cholinergic system lesions. Brain Res. Mol. Brain Res. 1997, 46, 161-8.
- Rossner, S.; Ueberham, U.; Yu, J.; Kirazov, L.; Schliebs, R.; Perez-Polo, J. R.; Bigl, V. In vivo regulation of amyloid precursor protein secretion in rat neocortex by cholinergic activity. Eur. J. Neurosci. **1997**, *9*, 2125–34.
- (259) Sadot, E.; Gurwitz, D.; Barg, J.; Behar, L.; Ginzburg, I.; Fisher, A. Activation of m1 muscarinic acetylcholine receptor regulates tau phosphorylation in transfected PC12 cells. J. Neurochem. **1996**, 66, 877–80.
- (260) Genis, I.; Fisher, A.; Michaelson, D. M. Site-specific dephosphorylation of tau of apolipoprotein E-deficient and control mice by M1 muscarinic agonist treatment. J. Neurochem. 1999, 72, 206-
- (261) Grundke-Iqbal, I.; Iqbal, K.; Tung, Y. C.; Quinlan, M.; Wisniewski, H. M.; Binder, L. I. Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. Proc. Natl. Acad. Sci. U.S.A. 1986, 83, 4913-
- (262) Duvoisin, R. C. Cholinergic-anticholinergic antagonism in parkinsonism. Arch. Neurol. 1967, 17, 124-36.
- (263) Harbaugh, R. E.; Roberts, D. W.; Coombs, D. W.; Saunders, R. L.; Reeder, T. M. Preliminary report: intracranial cholinergic drug infusion in patients with Alzheimer's disease. Neurosurgery **1984**, 15, 514-8.
- (264) Noring, U.; Povlsen, U. J.; Casey, D. E.; Gerlach, J. Effect of a cholinomimetic drug (RS 86) in tardive dyskinesia and drugrelated parkinsonism. Psychopharmacology 1984, 84, 569-71.
- Ott, B. R.; Lannon, M. C. Exacerbation of parkinsonism by tacrine. Clin. Neuropharmacol. 1992, 15, 322-5.
- (266) Keltner, N. L. Tacrine: a pharmacological approach to Alzheimer's disease. J. Psychosoc. Nursing Mental Health Serv. 1994, 32, 37-9
- (267) Hunker, C. J.; Abbs, J. H. Uniform frequency of parkinsonian resting tremor in the lips, jaw, tongue, and index finger. Movement Disord. **1990**, 5, 71–7.
- Mayorga, A. J.; Cousins, M. S.; Trevitt, J. T.; Conlan, A.; Gianutsos, G.; Salamone, J. D. Characterization of the muscarinic receptor subtype mediating pilocarpine-induced tremulous jaw movements in rats. Eur. J. Pharmacol. 1999, 364, 7–11.
- Salamone, J. D.; Mayorga, A. J.; Trevitt, J. T.; Cousins, M. S.; Conlan, A.; Nawab, A. Tremulous jaw movements in rats: a model of parkinsonian tremor. Prog. Neurobiol. 1998, 56, 591-611.
- (270) Bartus, R. T. On neurodegenerative diseases, models, and treatment strategies: lessons learned and lessons forgotten a generation following the cholinergic hypothesis. Exp. Neurol. **2000**, 163, 495-529.

JM990607U