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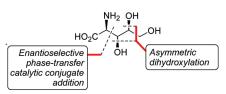
An Enantioselective Synthesis of (+)-Polyoxamic Acid via Phase-Transfer Catalytic Conjugate Addition and Asymmetric Dihydroxylation

Yeon-Ju Lee, † Yohan Park, † Mi-hyun Kim, † Sang-sup Jew, † and Hyeung-geun Park*,‡

†Marine Natural Products Chemistry Laboratory, Korea Ocean Research and Development Institute, Ansan 426-744, Korea, and [‡]Research Institute of Pharmaceutical Science and College of Pharmacy, Seoul National University, Seoul 151-742, Korea

hgpk@snu.ac.kr

Received November 16, 2010



(+)-Polyoxamic acid

A new enantioselective synthetic method of (+)-polyoxamic acid is reported. (+)-Polyoxamic acid could be obtained in 7 steps with 46% overall yield from diphenylmethyl-glycineimine tert-butyl ester via an enantioselective phase-transfer conjugate addition (99% yield, 96% ee) and an asymmetric dihydroxylation (98% yield, 94% de) as the key reactions.

Polyoxamic acid (1) is an amino acid bearing three contiguous hydroxyl groups, two of which are attached to stereogenic centers. It is a key component of peptidyl nucleoside antibiotics called polyoxins, which inhibit chitin synthetase of Candida albicans, a human fungi pathogen, and of various phytopathogenic fungi as well.²

On the other hand, 3,4-diepipolyoxamic acid (2) constitutes the structures of sphingofungins A-D, potent antifungal agents which inhibit serinepalmitoyl transferase to block the biosynthesis of sphingolipids.³ In both cases of polyoxins and sphingofungins, it is well-known that 3,4-dihydroxyamino acid moieties (1 and 2) are very important pharmacophores and their configurations are closely related to their biological activities.4

Due to their significance on the biological activities of polyoxins and sphingofungins along with their structural uniqueness, a variety of methods for the synthesis of polyoxamic acid and its stereoisomers have been developed over the past several years. Most commonly, they could be synthesized from chiral carbohydrates⁵ or amino acids⁶ depending on the stereogenic centers; however, only two methods were reported for the enantioselective synthesis of 1.7

In continuation of our studies on the synthesis of nonproteinogenic amino acids of pharmaceutical interest through phase-transfer reactions, we tried to develop an enantioselective synthesis of 1, which would later enable the synthesis of various stereoisomers of 1, including 2.

As shown in the retrosynthetic strategy (Scheme 1), the C2(S) chirality can, in principle, be induced by the enantioselective phase-transfer catalytic conjugate addition of diphenylmethylglycineimine *tert*-butyl ester (4).⁸ Both C(3S) and C(4S) configurations of the dihydroxy group can be derived from asymmetric dihydroxylation of 2, which can be obtained by olefination of 3.

First, the phase-transfer catalytic conjugate addition was carried out with 4 and methyl acrylate to introduce an α-carbomethoxyethyl moiety of 3 under phase-transfer catalytic reaction conditions. But, much to our disappointment, only moderate enantioselectivity (68% ee) was observed.⁹ In addition, significant racemization might be possible during the α -phenylselenylation of 3 in basic condition for olefination.

Address correspondence to this author. Phone: 82-2-880-8264. Fax: 82-2-872-9129.

⁽¹⁾ Isono, K; Suzuki, S. Heterocycles 1979, 13, 333.

^{(2) (}a) Naider, F.; Shenbagamurthi, P.; Steinfeld, A. S.; Smith, H. A.; Boney, C.; Becker, J. M. Antimicrob. Agents Chemother. 1983, 24, 787. (b) Mehta, R. J.; Kingsbury, W. D.; Valenta, J.; Actor, P. Antimicrob. Agents Chemother. 1984, 25, 373.

Naider, F. J. Med. Chem. 1983, 26, 1518. (c) Kobayashi, S.; Furuta, T.; Hayashi, T.; Nishijima, M.; Hanada, K. J. Am. Chem. Soc. 1998, 120, 908. (5) For recent examples, see: (a) Kim., I. S.; Li., Q. R.; Dong., G. R.; Woo., S. H.; Park, H.-j.; Zee, O. P.; Jung., Y. H. Synlett 2008, 19, 2958. (b) Falentin, C.; Beaupère, D.; Denailly, G.; Stasik, I. Tetrahedron 2008, 64, 9989. (c) Li, S.; Hui, X.-P.; Yang, S.-B.; Jia, Z.-J.; Xu, P.-F.; Lu, T.-J. Tetrahedron: Asymmetry 2005, 16, 1729. (d) Ichikawa, Y.; Ito, T.; Isobe, M. Chem.—Eur. J. 2005, 11, 1949. (e) Ulgheri, F.; Orrù, G.; Crisma, M.; Spanu, P. Tetrahedron Lett. 2004, 45, 1047. (f) Soengas, R. G.; Estévez, J. C.;

Estévez, R. J. Teterahedron: Asymmetry 2003, 14, 3955.

(6) For selected examples, see: (a) Verenea, G.; Datta, A. Tetrahedron Lett. 1998, 39, 119, (b) Harwood, L. M.; Robertson, S. M. Chem. Commun. 1998, 2641. (c) Bandini, E.; Martelli, G.; Spunta, G.; Bongini, A.; Panunzio, M.; Piersanti, G. Tetrahedron: Asymmetry 1997, 8, 3717. (d) Matsuura, F.; Hamada, Y.; Shioiri, T. Tetrahedron Lett. 1994, 35, 733. (e) Paz, M. M.; Sardina, J. J. Org. Chem. 1993, 58, 6990. (f) Ikota, N. Chem. Pharm. Bull. 1989, 37, 3399. (g) Garner, P.; Park., J. M. J. Org. Chem. 1988, 53, 2979.

^{(7) (}a) Trost, B. M.; Krueger, A. C.; Bunt, R. C.; Zambrano, J. J. Am. Chem. Soc. **1996**, 118, 6520. (b) Enders, D.; Vrettou, M. Synthesis **2006**, 2155.

⁽⁸⁾ For recent reviews on the phase-transfer catalysis, see: (a) Maruoka, K.; Ooi, T. Chem. Rev. 2003, 103, 3013. (b) O'Donnell, M. J. Acc. Chem. Res. 2004, 37, 506. (c) Lygo, B.; Andrews, B. I. Acc. Chem. Res. 2004, 37, 518. (d) Hashimoto, T.; Maruoka, K. Chem. Rev. 2007, 107, 5656. (e) Jew, S.-s.; Park, H.-g. Chem. Commun. 2009, 7090.

⁽⁹⁾ A Michael reaction between 5 and ethyl acrylate under the same condition (12, 50% KOH, CH₂Cl₂, 0°C) as that depicted in Table 1 afforded the alkylated product with 68% ee in 87% yield.

SCHEME 1. Retrosynthesis of (+)-Polyoxamic Acid (1)

SCHEME 2. Synthesis of 3,4-Didehydroamino Acid 3a

So our strategy needed modification and we chose ethyl α -phenylselenylacrylate (5) 10 as an alternate Michael acceptor, which already contains a phenylselenyl group. The phase-transfer catalytic Michael addition was performed with 4 and ethyl α -phenylselenylacrylate (5) under PTC conditions with 50% aqueous KOH in CH₂Cl₂ (Scheme 2).

To find an optimal catalyst for the conjugate addition, five representative catalysts¹¹ (PTCs, **10**–**14**, Table 1), which exhibited excellent catalytic efficiencies in the enantioselective catalytic conjugate addition of **4**, were examined. As shown in Table 1, all of the catalysts provided **6** as 1:1 mixtures of diastereomers in almost quantitative chemical yields, but the enantioselectivities were dramatically dependent on the PTC catalysts. Among the catalysts used, catalyst **12** yielded the highest enantioselectivity at 0 °C (entry 3, 90% ee) and even higher enantioselectivity was observed at –20 °C (entry 4, 96% ee). The stereoselectivities were determined by the diastereomer ratios of **6** as well as enantiomer ratios of **9** using chiral column chromatography. ¹² In all cases examined, the ratio of

TABLE 1. Enantioselective Phase-Transfer Catalytic Conjugate Addition of 5^a

$$\begin{array}{c} \text{CO}_2\text{Et} \\ \text{SePh} \quad \textbf{5} \\ \text{Ph} \\ \text{Ph} \\ \end{array} \begin{array}{c} \text{So} \\ \text{CO}_2\text{Et} \\ \\ \text{SePh} \quad \textbf{5} \\ \\ \text{CH}_2\text{CI}_2 \\ \end{array} \begin{array}{c} \text{Ph} \\ \text{N} \\ \text{SePh} \\ \\ \text{t-BuO}_2\text{C} \\ \end{array} \begin{array}{c} \text{SePh} \\ \text{O}_2\text{Et} \\ \\ \text{O}_2\text{Et} \\ \end{array}$$

entry	chiral PTC	temp (°C)	time (h)	yield of 6^b (%)	% ee of 9 ^{c,d}
1	10	0	1	99	48
2	11	0	2	99	59
3	12	0	1	99	90
4	12	-20	1	99	96
5	13	0	1	98	78
6	14	0	2	99	69

^aThe reaction was carried out with 3.0 equiv of 5 and 10.0 equiv of 50% KOH in the presence of chiral catalysts (10 mol %) in methylene chloride. ^bIsolated yields after purification by column chromatography. ^cEnantiopurity was determined by HPLC analysis, using a chiral column (DAICEL Chiralcel AD-H) with hexanes/2-propanol (volume ratio = 99:1) as a solvent. In this case, it was established by analysis of the racemate, of which enantiomers were fully resolved. ^dDiastereomer ratios of 6 correspond to enantiomer ratios of 9. See the Supporting Infomation for details.

the two major diasteromers to the two minor diastereomers of **6** was in accordance with the enantiomer ratio of **9**. The absolute stereochemistry of C(2) in the conjugate addition adduct (**6**) was confirmed by comparison of the optical rotation of the final product **1** with the reported value. ^{6f}

Next, the direct oxidation of selenyl ester **6** was avoided due to the tendency of isomerization of the olefinated product **2** to the more conjugated benzophenone imine 2,3-didehydroglutamate. To prevent the isomerization, a benzophenone imine group was hydrolyzed to **7** in acidic conditions. The resulting amino ester was protected with a 9-phenylfluorenyl (Pf) group by treatment with 9-bromo-9-phenylfluorene in the presence of K₃PO₄ and PbNO₂, which proved to prevent the isomerization by stereoelectronic hindrance. A-9-Phenylfluorenyl-glutamate **8** was then converted to 3,4-didehydroglutamate **9** by using NaIO₄ and NaHCO₃ at an ambient temperature with 98% yield without isomerization.

With 3,4-dihydroglutamate **9** in hand, efforts have been made to find the optimal conditions for asymmetric dihydroxylation (Table 2). Without any ligand, less than 2% conversion of **9** to **15** was observed with quantitative recovery of the starting material after 24 h (entry 1). In the presence of

⁽¹⁰⁾ For the preparation, see: Berlin, S.; Engman, L. *Tetrahedron Lett.* **2000**, *41*, 3701.

^{(11) (}a) O'Donnell, M. J.; Bennett, W. D.; Wu, S. J. Am. Chem. Soc. 1989, 111, 2353. (b) Corey, E. J.; Xu, F.; Noe, M. C. J. Am. Chem. Soc. 1997, 119, 12414. (c) Jew, S.-s.; Yoo, M.-S.; Jeong, B.-S.; Park, H.-g. Org. Lett. 2002, 4, 4245. (d) Park, H.-g.; Jeong, B.-S.; Yoo, M.-S.; Lee, J.-H.; Park, M.-K.; Lee, Y.-J.; Kim, M.-J.; Jew, S.-s. Angew. Chem., Int. Ed. 2002, 41, 3036.

⁽¹²⁾ The diastereomer ratio of 6 corresponds to the enantiomer ratio of 9. See the Supporting Infomation for details.

⁽¹³⁾ Rubio, A.; Ezquerra, J. Tetrahedron Lett. 1995, 36, 5823

⁽¹⁴⁾ Gmeiner, P.; Feldman, P. L.; Chu-moyer, M. Y.; Rapport, H. J. Org. Chem. 1990, 55, 3068 and references cited therein.

TABLE 2. Asymmetric Dihydroxylation of 9^a

entry	ligand	time (h)	yield ^b (%)	% de ^c
1	none	24	< 2	
2	DABCO	24	84	< 2
3	16	3.5	91	75
4	17	4.0	88	83
5	18	2.5	91	87
6	19	5.0	93	88
7	20	3.5	99	94

^aSee the Supporting Infomation for detailed reaction conditions. ^bIsolated yields after purification by column chromatography. ^cDiaster-eopurity was determined by HPLC analysis, using a chiral column (DAICEL Chiralcel AD-H) with hexanes/2-propanol (volume ratio = 92:8) as a solvent.

the achiral ligand DABCO, the reaction proceeded efficiently (94% yield) to yield 15 with no stereoselectivity (<2% de), indicating that the stereogenecity of C(2) does not affect the facial selectivities of 3,4-dihydroxylations (entry 2). As chiral ligands, five quinine derivatives (16-20) were examined. The absolute configuration of the products was determined by the application of the mnemonic devices for predicting facial selectivity in asymmetric dihydroxylations¹⁵ and confirmed later by comparison of the optical rotation of final product 1 with the reported value. 6 In the case of dimeric quinine derivative 16 and 17, which usually provide highly enantioselective dihydroxylations, 15 moderate facial selectivities were observed (entries 3 and 4, 75% de and 83% de). Monomer ligands 18 and 19 were also examined and surprisingly, products were obtained with higher diastereoselectivities (entries 5 and 6, 87% de and 88% de). When 20 (HQN-CLB) was used as a chiral ligand based upon the expectation that reducing the steric hindrance of the chiral ligand might be beneficial, the highest diastereoselectivity (94% de) was obtained providing 15 in a quantitative yield. The successful results prompted us to apply HQD-CLB under the optimal conditions for the preparation of 3,4-diepipolyoxamic acid (2). However, much to our disappointment, only moderate enantioselectivity of 3,4-diepi-15 with comparable chemical yield was obtained (42% de, 93% yield).

Treatment of 15 with NaBH₄/LiCl in methanol at 0 °C afforded aminotriol 21 with 88% yield, which upon treat-

SCHEME 3. Completion of the Synthesis of 1

ment with trifluoroacetic acid provided (+)-polyoxamic acid (1) after purification by Dowex ion-exchange resin (Scheme 3). The physical characteristics of 1 were in full agreement with the reported values in the literature $\{([\alpha]^{23}_{D} 2.5 (c\ 0.2, H_2O) (lit.^{6f} [\alpha]^{23}_{D}\ 2.8 (c\ 1.0, H_2O)\}.$

In conclusion, we have developed an efficient and enantioselective synthesis of (+)-polyoxamic acid from a commercially available diphenylmethyl glycineimine *tert*-butyl ester (4) in 7 steps (46% overall yield, 96% ee) by an enantioselective phase-transfer catalytic conjugate addition and asymmetric dihydroxylation. It is worthwhile to point out that all three stereogenic centers were constructed by the use of cinchona derived catalysts (12, 20) which are easily prepared from inexpensive materials. Further applications to the synthesis of similar polyoxamic acid-type compounds with an aminotriol core structure including 2 are now under investigation.

Experimental Section

Representative Procedure for the Enantioselective Phase-Transfer Catalytic Alkylation of 4 (6). A solution of ethyl 2-(phenylselanyl)acrylate 5 (383 mg, 1.5 mmol) in dichloromethane (0.3 mL) was added to a solution of N-(diphenylmethylene)glycine tert-butyl ester 4 (148 mg, 0.50 mmol) and chiral catalyst 12 (30.3 mg, 0.05 mmol) in dichloromethane (1.2 mL). The reaction mixture was then cooled to -20 °C, 50% aqueous KOH (0.56 mL) was added, and the resulting mixture was allowed to stir at -20 °C for 1.0 h. The resulting mixture was diluted with dichloromethane (20 mL), washed with water (2 \times 5 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The resulting brown oil was purified by silica gel column chromatography to afford 6 (273 mg, 0.50 mmol, 99% yield) as a pale yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 7.60 (d, J = 7.0 Hz, 1H), 7.55 (d, J = 7.5 Hz, 1H), 7.53 (d, J = 7.0 Hz, 1H), 7.42 - 7.33 (m, 5H),7.29 - 7.22 (m, 4H), 7.19 - 7.15 (m, 2H), 7.11 (dd, J = 2.5, 6.0 Hz, 1H), 4.10 (dd, J = 4.2, 9.5 Hz, 0.5H), 3.97–3.86 (m, 2H), 3.82-3.75 (m, 1H), 3.60 (dd, J = 6.5, 8.8 Hz, 0.5H), 2.54-2.48(m, 1H), 2.44-2.40 (m, 0.5H), 2.30-2.24 (m, 0.5H), 1.36 (s, 4.5H),1.35 (s, 4.5H), 1.04 (t, J = 7.0 Hz, 1.5H), 1.01 (t, J = 7.5 Hz, 1.5H) ppm; ¹³C NMR (100 MHz, CDCl₃) δ 172.8, 172.5, 171.9, 171.4, 170.9, 170.5, 139.6, 139.5, 136.6, 136.4, 136.0, 135.8, 130.6, 130.5, 129.2, 129.1, 128.5, 128.5, 128.2, 128.1, 128.1, 128.0, 81.6, 81.5, 64.5, 64.1, 61.1, 61.0, 40.5, 40.3, 36.3, 35.1, 28.2, 28.2, 14.1, 14.0 ppm; IR (KBr) 3348, 2970, 1467, 1379, 1305, 1160, 1129, 952, 817, 760 cm^{-1} ; LRMS (FAB⁺) m/z 552.0 [M + H]⁺; HRMS (ESI) $[M_3 + H]^+$ calcd for $C_{30}H_{33}NO_4Se$ 552.1648, found 552.1641. $[\alpha]^{23}$ _D -10.7 (c 1.0, CHCl₃). The diasteromeric ratio was determined by HPLC analysis in comparison with authentic racemic materials. {DAICEL chiralpak AD-H, hexane:2-propanol = 99:1,

⁽¹⁵⁾ Kolb, H. C.; VanNieuwenhze, M. S.; Sharpless, K. B. *Chem. Rev.* **1994**, *94*, 2483 and references cited therein.

flow rate = 1.0 mL/min, 23 °C, λ = 254 nm, retention times: 14.71 (minor), 18.79 (minor), 19.86 (major), 28.17 (major), 1.3:1.7:40:53 dr}.

Representative Procedure for the Asymmetric Dihydroxylation of 9 (15). To a solution of 9 (117 mg, 0.25 mmol) and methanesulfonamide (35 mg, 0.37 mmol) in tert-butanol (2.0 mL) and water (2.0 mL) was added K₂CO₃ (104 mg, 0.75 mmol) and chiral ligand 20 (23 mg, 0.05 mmol). The mixture was then cooled to 0 °C, then K₃Fe(CN)₆ (247 mg, 0.75 mmol) was added followed by K₂OsO₂(OH)₄ (9.2 mg, 0.025 mmol). The mixture was allowed to stir at 0 °C for 3.5 h, after which time the reaction was quenched by dropwise addition of saturated NaHSO₃ aqueous solution. The mixture was then diluted with ethyl acetate (20 mL), washed with water (2 × 5 mL), dried over anhydrous MgSO₄, filtered, and concentrated in vacuo. The resulting yellow oil was purified by silica gel column chromatography to afford 15 (125 mg, 0.25 mmol, 99% yield) as a colorless oil. ¹H NMR (300 MHz, CDCl₃) δ 7.85 (d, J = 7.5 Hz, 2H), 7.38-7.29 (m, 5H), 7.21-7.14 (m, 6H), 4.17 (dd. J=7.1 Hz, 2H), 4.02 (s, 1H), 3.72 (d, J = 5.5 Hz, 1H), 2.76 (d, J = 6.0 Hz, 1H), 1.23 (t, J = 7.1 Hz, 3H), 1.17 (s, 9H) ppm; ¹³C NMR (100 MHz, CDCl₃) δ 172.3, 171.8, 148.0, 147.8, 143.5, 141.1, 128.8, 128.5, 128.5, 128.2, 127.8, 127.4, 126.4, 125.8, 125.2, 120.3,

119.9, 82.4, 72.8, 72.3, 71.2, 61.6, 57.5, 27.8, 14.1 ppm; IR (KBr) 3492, 2979, 1734, 1451, 1394, 1369, 1213, 1155, 1073, 844, 754, 699 cm $^{-1}$; LRMS (FAB $^+$) m/z 504 [M + H] $^+$; HRMS (ESI) $[M + H]^+$ calcd for $C_{30}H_{34}O_6N$ 504.2386, found 504.2389. $[\alpha]^{23}_D$ 5.3 (c 1.0, CHCl₃). The diasteromeric ratio was determined by HPLC analysis in comparison with authentic racemic materials. {DAICEL chiralpak AD-H, hexane:2-propanol = 92:8, flow rate = 1.0 mL/min, 23 °C, λ = 254 nm, retention times: 13.92 (minor), 14.85 (minor), 18.48 (major), 32.66 (minor), 2.4:0.6:95.5:1.5 dr, 94% de from **9** (96% ee)}.

Acknowledgment. This work was supported by the Midcareer Researcher Support Programs of the National Research Foundation of Korea (2009-0078814) and by the National Research Foundation of Korea Grant funded by the Korean Government (MEST) (NRF-C1ABA001-2010-0020428)

Supporting Information Available: Characterizations of all compounds, additional experimental procedures, and analytical data. This material is available free of charge via the Internet at http://pubs.acs.org.