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Synthesis, DNA Affinity, and Antiprotozoal Activity of Linear Dications: Terphenyl Diamidines and Analogues

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Diamidines 10a-g and 18a,b were obtained from dinitriles 9a-g and 15a,b by treatment with lithium trimethylsilylamide or upon hydrogenation of bis-O-acetoxyamidoximes. Dinitriles 9a-g were prepared via Suzuki reactions between arylboronic acids and arylnitriles. Potential prodrugs 12a-f and 17 were prepared via methylation of the diamidoximes 11a-f and 16a. Significant DNA affinities for rigid-rod molecules were observed. Compounds 10a, 10b, 10d, 18a, and 18b show IC₅₀ values of 5 nM or less against Trypanosoma brucei rhodesiense (T. b. r.) and 10a, 10b, 10e, 18a, and 18b gave similar ones against Plasmodium falciparum (P.f.). The dications, 10a, 10d, 10f, and 10g are more active than furamidine in vivo. The prodrugs are only moderately effective on oral administration. Mouse liver microsome bioconversion of the methamidoxime prodrugs is significantly reduced from that of pafuramidine and suggests that the in vivo efficacy of these prodrugs is, in part, due to poor bioconversion.

Introduction

Pentamidine (1) has been known for its antimicrobial activity for over 50 years, and it remains the only aromatic diamidine to have seen significant human use. 1,2 Pentamidine is used against antimony-resistant leishmaniasis, initial stage human African trypanosomiasis (HAT), and also as a secondary drug for AIDS-related *P. jiroveci* (formerly *P. carinii*) pneumonia.¹ A prodrug of furamidine (2), which is administered orally, is currently in Phase II clinical trials against malaria and Phase III trials against pneumocystis pneumonia and HAT.^{1,3-7} These dicationic molecules bind in the minor groove of DNA at AT rich sites, and this interaction is believed to be key to their antimicrobial activity. The molecules are concentrated in the kinetoplast, which is thought to be the first step in their antitrypanosomal action.1b Minor groove binding is postulated to lead to the inhibition of DNA dependent enzymes or the direct inhibition of transcription.^{1,8-11} For the trypanosomes, the selectivity of the dications is likely to include uptake by amidine transporters. 12 For about two decades, a fundamental element in the design of new minor groove binding molecules has been that the molecular scaffold bearing the amidine units should present crescent shape geometry complementary to the curve of the minor groove of DNA.¹³ In addition to the shape complementary component of these molecules, van der Waals contacts with the walls of the groove have been shown to be an important contributor to binding affinity. 14-16 A recent theoretical review of the binding interactions of over 20 minor groove binders concluded that the curvature of the small molecule was important to provide energetically favorable van der Waals contacts.¹⁷ Pentamidine, furamidine, and many analogues have been shown to meet this crescent shape requirement. 1,13,18,19 In contrast, recent reports have shown that diamidine (3) (CGP 3), which despite the fact that it is a nearly linear molecule, binds quite strongly to the AT rich minor groove.^{20–22}

A careful analysis of binding data, crystal structure, and molecular dynamic simulations leads to the conclusion that water mediated interactions between 3 and the DNA minor groove, in effect, provide the needed curvature. ^{21,22} Recently, we reported strong DNA affinities and excellent antiprotozoan activity for the near linear biphenylbenzimidazoles (4).²³ A detailed study of the DNA binding of 4, including a crystal structure of its cocrystal with the Dickerson dodecamer, shows that the linear molecule binds quite strongly to the minor groove.²⁴ As with CGP 40215, a bound water molecule provides a key interaction between 4 and the floor of the groove to, in effect, provide curvature.²⁴ A review of the literature reveals that other linear dicationic molecules exhibit marked antiprotozoan activity. For example, bis-4,4'-amidinobiphenyl (5) and bis-4,4'-amidinophenylethyne (6) have been reported to show significant in vivo activity in a trypanosomal mouse model for Trypanosoma rhodesiense.25 In view of these several examples of linear dicationic molecules that show promising antimicrobial activity and significant DNA minor groove affinity, we have undertaken a study of linear systems based upon the rigid-rod terphenyl scaffold. Rigid-rod molecules, which cannot be compressed or bent, have received considerable attention in material science²⁶ and are receiving growing attention in the biological sciences.²⁷ Such compounds, which do not match the curvature of the minor groove, may represent a new class of dicationic antimicrobial agents that exhibit a new type of antiparasitic effect. We report the synthesis of novel dicationic terphenyls and their evaluation as potential minor groove binders and antiprotozoan agents.

The pK of aromatic amidines is greater than 10, and consequently, such molecules are charged at physiological pH.²⁸ The lack of oral bioavailability of diamidines such as pentamidine, furamidine and analogues is well known.^{19,29} In addition, several analogues of furamidine show excellent activity on intravenous dosing but are ineffective on oral administration.^{18,19} Generally, oral administration is the preferred dosing regime, and hence, prodrug strategies for diamidines that have the potential to overcome their limited oral bioavailability merit attention. We have reported that bis-amidoximes and bis-*O*-alkylamidoximes

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Figure 1. Structures of key dicationic antiprotozoan agents.

Scheme 1a

^a Reagents and conditions: (i) Pd(PPh₃)₄, Na₂CO₃, toluene, 80 °C; (ii) (a) LiN(TMS)₂, THF, r.t., overnight; (b) HCl(gas), dry ethanol, r.t., overnight; (iii) NH₂OH.HCl/KO-*t*-Bu, DMSO; (iv) (a) AcOH/Ac₂O; (b) H₂/Pd−C, AcOH; (v) LiOH/(CH₃)₂SO₄.

of a number of diamidine systems are effective prodrugs.^{30–32} As part of our continuing effort to develop orally effective antimicrobial agents, we include the synthesis of examples of these two types of potential prodrugs in the terphenyl series.

Chemistry. As shown in Scheme 1, a series of linear terphenyl diamidines and their aza-analogues was obtained from the respective dinitriles 9a-g either by direct reaction using lithium trimethylsilylamide (cf. 10b, c, e-g) or from the bis-O-acetoxyamidoxime followed by hydrogenation in glacial acetic acid (cf. 10a, d). The dinitriles 9a-g were prepared via a Suzuki coupling reaction either employing bis-1,4-phenyleneboronic acid with 4-bromobenzonitrile or 6-chloronicotinonitrile or by employing 4-cyanophenylboronic acid with 1,4-dibromobenzene and its derivatives. The potential prodrugs 12a-f were prepared via methylation of the respective diamidoxime 11a-f with dimethyl sulfate in DMF solution and using Li(OH) as a base. The hydrochloride salts of the amidoximes 11a-f and 12a-f were obtained by passing hydrogen chloride gas into an ethanolic solution of their free bases.

As outlined in Scheme 2, dinitrile compound 15a was obtained in two steps starting with a Suzuki coupling reaction of 5-(4-bromophenyl)furan-2-carboxaldehyde with 4-formylphenylboronic acid to yield dialdehyde 14a. Reaction of 14a with hydroxylamine hydrochloride in the presence of Na₂CO₃, followed by acetic anhydride induced dehydration of the oxime furnished 15a. The acetate salt of 18a was obtained from dinitrile **15a** through bis-O-acetoxyamidoxime followed by hydrogenation in glacial acetic acid. In a similar way, diamidine 18b was prepared starting from compound 15b. Dinitrile 15b was obtained employing a selective Suzuki coupling reaction of 2,5-dibromopyridine with 4-formylphenylboronic acid to give bromoaldehyde 13. A Stille coupling reaction between 13 and 5-(diethoxymethyl-furan-2-yl)-tributyltin, followed by acid hydrolysis of the acetal gave 14b, a key precursor of 15b. The potential prodrug, N-methoxy-5-[4'-(N-methoxyamidino)-biphenyl-4-yl]furan-2-carboxamidine (17) was prepared via methylation of the respective diamidoxime **16a**. The hydrochloride salts of the amidoximes, 16a,b and 17, were made by passing hydrogen chloride gas into an ethanolic solution of their free bases.

Biology. Table 1 contains the results from DNA binding studies and in vitro antiparasitic evaluations for the new linear dications. The $\Delta T_{\rm m}$ values reflect the DNA affinities of these compounds. Although the $\Delta T_{\rm m}$ value for ${\bf 10a}$ is about 20% less than that of furamidine, it is significantly greater than that of pentamidine. Despite the fact that ${\bf 10a}$ is a linear rigid-rod it shows impressive DNA affinity, perhaps using a water molecule to effectively provide curvature as has been shown for other linear dicationic molecules. 21,22,24

Although aromatic diamidines have been universally observed to be minor groove binding compounds in AT base pair sequences of DNA, the compounds have generally been crescent shaped systems that fit into the double helix groove and directly interact with the A and T bases at the floor of the groove. The linear compounds in this study are more similar to 3 and 4, which are also minor groove binding compounds, but it is important to test the binding mode of the new linear compounds. A characteristic feature of groove binding diamidines is that they have a strong positive induced CD signal when they bind to AT DNA sequences. Intercalation binding gives weak induced CD signals that are generally negative. Consistent with a minor groove binding mode, the linear diamidines in this article give a positive induced CD signal when added to polydA.polydT, and the spectra are similar to those obtained for 3 and 4. An example set of CD spectra for 10a are shown in Supporting Information Figure 1, and the strong, positive induced CD signals at the maximum absorption wavelength of 10a are clearly seen.

Scheme 2^a

$$B_{r}$$
 $N=$
 B_{r}
 $N=$
 A_{r}
 A_{r}

^a Reagents and conditions: (i) 4-Formylphenylboronic acid, Pd(PPh₃)₄; (ii) (a) 5-(diethoxymethyl-furan-2-yl)-tributyltin, Pd(PPh₃)₄; (b) Hydrolysis with 2M HCl; (iii) (a) NH₂OH.HCl; (b) Ac₂O; (iv) NH₂OH.HCl/KO-*t*-Bu, DMSO; (v) LiOH/(CH₃)₂SO₄; (vi) (a) AcOH/Ac₂O; (b) H₂/Pd-C, AcOH.

Table 1. DNA Affinities and in Vitro Antiprotozoan Data for Linear Dications

$$RN$$
 H_2N
 R_1
 R_1
 R_2
 R_3
 R_4
 R_5
 R_6

	R	R_1	R_2	X	Y	M	ΔT m a (° C)	$\frac{T. b. r. b}{IC_{50}}$ (nM)	P. f. b IC ₅₀ (nM)	$\frac{\text{cytotoxicity}^c}{\text{IC}_{50}} \\ (\mu \text{M})$
code										
1	NA	NA		NA	NA	NA	12.6	2.2	NT	2.1
2	NA	NA		NA	NA	NA	25	4.5	15.5	6.4
10a	H	H	nil	CH	CH	CH	18.2	5	1	22.1
11a	OH	H	nil	CH	CH	CH	ND	22.8K	>11.8K	>211
12a	OMe	H	nil	CH	CH	CH	ND	13.8K	371	118.3
10b	H	F	nil	CH	CH	CH	15.1	2	1	6.4
11b	OH	F	nil	CH	CH	CH	ND	> 201K	925	29.1
12b	OMe	F	nil	CH	CH	CH	ND	51.5K	4.2K	8.5
10c	H	CF_3	nil	CH	CH	CH	10.1	48	15	9.4
11c	OH	CF_3	nil	CH	CH	CH	ND	29.9K	3.1K	13.3
12c	OMe	CF_3	nil	CH	CH	CH	ND	36.9K	3.8K	42.2
10d	H	Н	nil	CH	N	CH	18.7	2	10	49.9
11d	OH	H	nil	CH	N	CH	ND	9.3K	7.6K	> 184
12d	OMe	H	nil	CH	N	CH	ND	34.7K	3.2K	> 166
10e	H	H	nil	N	N	CH	8.1	16	0.5	40.9
12e	OMe	Н	nil	N	N	CH	ND	49K	5.7K	> 177
10f	H	Н	nil	CH	CH	N	9.2	47	10	26.6
12f	OMe	H	nil	CH	CH	N	ND	82.9K	1.2K	> 158
10g	Н	OMe	OMe	CH	C	CH	5.2	15	37	> 100
18a	Н	NA	NA	NA	NA	NA	19.1	5	1	56.1
16a	OH	NA	NA	NA	NA	NA	ND	67K	6.6K	106
17	OMe	NA	NA	NA	NA	NA	ND	24K	1.6K	> 205
18b	Н	NA	NA	NA	NA	NA	17.6	2	10	30.9

^a Poly(d(A-T)₂ in MES10 buffer; ratio compound/DNA is 0.3. ^b The *T. b. r.*(*Trypanosoma brucei rhodesiense*) strain was STIB900, and the *P. f.* (*Plasmodium falciparum*) strain was K1. The values are duplicate determinations; see refs 23 and 31. ^c Cytotoxicity was evaluated using cultured L6 rat myoblast cells using the same assay procedure for *T. b. r*.

The central ring substituted terphenyldiamidines (10a-c, 10g) show a progressive decrease (from 18.2 to 5.2 ΔT m) in binding affinity as larger and more substituents are added to the aryl

rings. The substituents can cause an increase in the torsion angles between the aryl rings because of increased repulsive van der Waals interactions. A second important feature of the substit-

uents is that they increase the energy required to reduce the angle between directly bonded aryl systems. Reduced twist appears to be an important feature of unfused compound complexes with the DNA minor groove. The biphenyl torsional angle in 4, for example, is 28° relative to the unbound value of 44°.24 Large twists in the compound molecular shape are expected to decrease favorable interactions of the compounds with the DNA minor groove and increase steric clash with the walls of the groove. We conducted molecular mechanics calculations to compare the equilibrium torsional angles in the substituted compounds (angle between the substituted phenyl and the unsubstituted phenyl bonded adjacent to the substituent) as well as the energies of the compounds when the angle was reduced to 20°. The values are 44° and 17 kcal/mol for 10a; 46° and 26 kcal/mol for 10b; and 55° and 92 kcal/mol for 10c, in agreement with the decreasing order of their $\Delta T_{\rm m}$ values. For 10g, the values are 50° and 41 kcal/mol, but presumably because it has two aryl ring junctions, 10g has the lowest $\Delta T_{\rm m}$ value in this set of compounds. These results suggest that a near planar array enhances the fit of linear rigid-rod molecules in the DNA minor groove.

The aza analogues (10d-f) do not show a recognizable pattern for the effect of the introduction of a heterocyclic nitrogen atom into the various phenyl rings. The $\Delta T_{\rm m}$ for 10d with one N-atom and the parent molecule 10a are equivalent, and thus, essentially, there is no effect of the introduction of one heterocyclic nitrogen in the central ring. Unexpectedly, the introduction of two nitrogen atoms in the central ring as exemplified by 10e dramatically reduces the $\Delta T_{\rm m}$ value. The origin of this effect is unclear. Similarly, the introduction of one nitrogen atom in each of the terminal rings also significantly lowers the $\Delta T_{\rm m}$ value. Further biophysical studies are underway to attempt to understand the effect of N-atom introduction on DNA affinities of these linear rigid-rods. Replacement of one of the benzamidine units with a furamidine one (18a, 18b) gives $\Delta T_{\rm m}$ values quite close to that of the parent 10a, showing that replacing a terminal phenyl with furan has little effect on DNA affinity. A more detailed analysis of the DNA binding properties of these linear systems will be forthcoming in due course.

These linear dications also exhibit significant in vitro antiparasitic activity (Table 1). Five dications (**10a**, **10b**, **10d**, **18a**, **18b**) show IC₅₀ values of 5 nM or less against T. b. r. In addition, five molecules (**10a**, **10b**, **10e**, **18a**, **18b**) exhibit IC₅₀ values of near 5nM or less against P.f. The diaza analogue **10e** is of particular interest because it shows a 30-fold selectivity for P. f. As expected, the prodrugs do not show significant in vitro activity.

The promising in vitro activities of these analogues lead us to evaluate them in the STIB900 acute mouse model for *T. b. r.* (Table 2). Most of the dications on intraperitoneal dosing show a significant increase in survival time of treated animals compared to untreated controls. Four of the linear dications (**10a**, **10d**, **10f**, **10g**, **18b**) gave results superior to that of furamidine in this model.

Given the encouraging in vivo activities found for several of these dications, the amidoxime and methamidoxime potential prodrugs were evaluated on oral administration in the same animal model. Although several of the prodrugs (12b, 11d, 12d) significantly increased the survival time of treated animals versus that of untreated controls, none of the compounds approximated the efficacy of pafuramidine (Table 2). This disappointing result clearly suggests significant differences in pharmacodynamics and/or metabolism of the linear prodrugs.

Because we have found that methoximes generally show greater in vivo efficacy than amidoximes,² we sought to gain

Table 2. Antitrypanosomal Activity in Vivo of Linear Dications in the STIB900 Mouse Model^a

compd	dosage mg/kg	cures^b	survival (days) ^c	
1	20	0/4	>42.75	
•	ip	0.44		
2	20	0/4	> 52.5	
	5	0/4	35.5	
pafuramidine d	ip 25	1/4	>60	
paruramume	po	1/4	- 00	
10a	20	2/4	>47.75	
	ip			
11a	100	0/4	24.75	
	po			
12a	100	0/4	20.75	
	po			
10b	5	0/4	>51.75	
111	ip	0/4	22.25	
11b	25	0/4	22.25	
12b	po 25	0/4	37.25	
120	po	0/4	31.23	
10c	5	0/4	17.25	
100	ip	٠,٠	17,120	
11c	25	0/4	9	
	po			
12c	25	0/4	7.25	
	po			
10d	20	4/4	>60	
	5	2/4	>42	
11d	ip 100	1/4	>35.75	
110	po	1/4	- 33.13	
12d	25	0/4	>42.75	
	po			
10e	5	0/4	23.5	
	ip			
12e	25	0/4	12.5	
	po			
10f	5	2/4	>60	
12f	ip 25	0/4	12	
121	po	0/4	12	
10g	5	2/4	>51.75	
108	ip	27 1	51.75	
18a	20	0/4	21.25	
	ip			
16a	25	0/4	16	
	po			
17	25	0/4	6.25	
101	po	2/4	> 50.5	
18b	20	3/4	>53.5	
	5 ip	2/4	>41.75	

^a See refs 23 and 31 for details of the STIB900 mouse model. Dosage was for 4 days and was either intraperitoneal (ip) or oral (po) as noted. ^b The number of mice that survive 60 days and are parasite free. ^c Average days of survival; untreated controls died between day 7 and 8 post infection. ^d Pafuramidine = 2,5-bis(4-methoxyamidinophenyl)furan.

some insight into the low efficacy of these linear prodrugs by comparing the rates of metabolic transformation of selected methamidoximes (**12a**, **12b**, **12d**–**12f**) to that of pafuramidine when exposed to mouse liver microsomes (Table 3). The substrate depletion data presented in Table 3 shows that pafuramidine rapidly disappears on exposure to mouse liver microsomes with a $t_{1/2}$ value <15 min. In contrast, the linear methamidoximes are much more metabolically stable with $t_{1/2}$ values greater than 90 min for all compounds studied except for **12d**, which gave a $t_{1/2}$ value of 55 min. The results from the mouse liver microsome studies are consistent with the in vivo efficacy results from the STIB900 mouse model and suggest that the slow metabolism of the linear prodrugs in mice

Table 3. Results from an in Vitro Metabolic Transformation Investigation of Selected Methoxamidines

compd	$t_{1/2}^{a}$		
pafuramidine	<15		
12a	>90		
12b	>90		
12d	<15 >90 >90 55 >90 >90		
12e	>90		
12f	>90		

a $t_{1/2}$ in minutes for methoxamidine disappearance.

significantly contributes to their low efficacy. The bioconversion of these linear prodrugs needs to be investigated in other species.

We have shown that linear rigid-rod dications effectively bind to DNA, exhibit low nanomolar IC $_{50}$ values against $T.\ b.\ r.$ and $P.\ f.$, and show promising activity on intraperitoneal administration in the STIB900 mouse model. However, the prodrugs show only modest efficacy on oral dosing in the same mouse model, and they are significantly more slowly metabolized than pafuramidine by mouse liver microsomes. To capitalize on the efficacy of these potent dications, other prodrugs that rely on different bioconversion pathways need to be developed.

Experimental Section

Molecular Mechanics Calculations. Calculations were performed with the Spartan 04 software package (Wavefunction, Inc.) and the SYBYL force field. The compounds were first energy minimized with geometry optimization, and the phenyl—phenyl (or substituted phenyl) torsion angle was determined. The torsion was then locked into a 20° angle and a single-point energy calculated with the same software to give an idea of the energy cost of making the aromatic system more planar, as is usually required for minor groove binding (see ref 23, for example).

Efficacy Evaluations. In vitro assays with T. b. r. STIB 900 and the P. f. K1 strain as well as the efficacy study in an acute mouse model for T. b. r. STIB 900 were carried out as previously reported. 23,31

Tm Measurements. Thermal melting experiments were conducted with a Cary 300 spectrophotometer. Cuvettes for the experiment were mounted in a thermal block, and the solution temperatures were monitored by a thermistor in the reference cuvette. Temperatures were maintained under computer control and were increased at 0.5 C/min. The experiments were conducted in 1 cm path length quartz cuvettes in MES 10 buffer (MES 10mM, EDTA 1mM, NaCl 100mM). The concentrations of DNA were determined by measuring the absorbance at 260 nm. A ratio of 0.3 compound per base was used for the complex, and DNA with no compound was used as a control.

Metabolic Stability Experiments. ³³ Dimethyl sulfoxide, magnesium chloride, potassium dihydrogen phosphate, anhydrous disodium hydrogen phosphate, and β-nicotinamide adenine dinucleotide 2'-phosphate tetrasodium salt were purchased from Sigma (St. Louis, MO). Microsomes from pooled male mice B6C3F1 (n = 126) were purchased from XenoTech LLC (Lenexa, KS).

To determine the metabolic stability of prodrugs, **12a**, **12b**, **12d**, **12e**, **12f**, and pafuramidine were incubated with microsomal protein from mice (0.5 mg/mL final target concentration), phosphate buffer (100 mM, pH 7.4), and MgCl₂ (5 mM) in a total incubation volume of 0.5 mL. Incubations were performed in duplicate. The prodrugs were prepared as 10 mM stock solutions in DMSO and were added to the incubation mixtures in 2.5 μ L of DMSO to keep the organic solvent at 0.5% (v/v) in the final incubations. Samples were preincubated at 37 °C for 3 min, and the reactions were started by the addition of 50 μ L of β -NADPH (2 mM final target concentration). Reactions were stopped at 0, 5, 10, 15, 20, 25, and 30 min by quenching with 100 μ L of acetonitrile. The samples were centrifuged at 3000 rpm for 5 min. Pafuramidine was used as the internal standard for all other compounds. **12b** was used as the internal standard for pafuramidine. Samples were analyzed im-

mediately after centrifugation using LC-MS/MS systems as described below.

Automated sample analysis was performed using Analyst software (version 1.3.1, Applied Biosystems, Foster City, CA). The Analyst controlled HPLC-MS/MS system consisted of two Shimadzu Scientific (Columbia, MD) solvent delivery pumps, a thermostated (6 °C) LEAP HTC autosampler (Carrboro, NC), and an Applied Biosystems API4000 triple quadrupole mass spectrometer. Reversed-phase gradient chromatography was used to elute the prodrugs from an Aquasil (C_{18} 5 μ , 50 \times 2.1 mm) analytical column at a flow rate of 1 mL/min, following a 3μ L injection. Starting conditions for each injection were 90:10, water/methanol with 0.1% formic acid in each. The relative amounts of water/ methanol were held constant for 0.5 min while the column eluted to waste. After 0.5 min, the eluent was directed to the mass spectrometer and the relative amount of methanol increased linearly to 90% at 3 min postinjection. This amount of methanol was held for 0.5 min to wash the column. The column was reequilibrated under the starting conditions for the final 0.5 min. Total run time was 4 min.

The mass spectrometer was connected to the HPLC system by a TurboIonSpray interface. Nitrogen, from a Peak Scientific (Bedford, MA) nitrogen generator, was used as the curtain, nebulizer, and collision gas. User controlled voltages, gas pressures, and source temperature were optimized for the detection of the parent and product ions of the prodrugs. All compounds were analyzed in positive ion mode using multiple reaction monitoring. Care was taken to not inject compounds with similar transitions in succession.

Microsoft Excel 2003 for Windows (Seattle, WA) was used for the calculation of the percent substrate loss and to calculate half-lives. Ratios of peak areas obtained at different times were compared to that at zero time point and were multiplied by 100 to get the percent parent drug left intact. The in vitro half-lives were determined by plotting $\ln \%$ remaining versus time, then measuring the slope (k) of this plot by using the first-order decay formula ($C = C_0 e^{-kt}$): $t_{1/2} = 0.693/\text{slope}$

Synthetic Protocols. Melting points were recorded using a Thomas-Hoover (Uni-Melt) capillary melting point apparatus and are uncorrected. TLC analysis was carried out on silica gel 60 F₂₅₄ precoated aluminum sheets and detected under UV light. 1H and ¹³C NMR spectra were recorded by employing a Varian Unity Plus 300 spectrometer (Varian, Inc., Palo Alto, California), and chemical shifts (δ) are in ppm relative to TMS as the internal standard. Mass spectra were recorded on a VG analytical 70-SE spectrometer (VG Analytical, Ltd., Manchester, United Kingdom). Elemental analyses were obtained from Atlantic Microlab Inc. (Norcross, GA) and are within ± 0.4 of the theoretical values, except for 12a and 18a, which were validated by HRMS. The compounds reported as salts frequently analyzed correctly for fractional moles of water and/or ethanol of solvation. In each case, proton NMR showed the presence of indicated solvents. All chemicals and solvents were purchased from Aldrich Chemical Co., Fisher Scientific, Frontier Scientific, or Lancaster Synthesis, Inc.

4,4"-Bis-cyano[1,1';4',1"]terphenyl (9a). To a stirred solution of 4-bromobenzonitrile (0.91 g, 5 mmol), and tetrakis(triphenylphosphine) palladium (200 mg) in toluene (10 mL) under a nitrogen atmosphere was added 5 mL of a 2 M aqueous solution of Na₂CO₃ followed by 1,4-phenylenebisboronic acid (0.41 g, 2.5 mmol) in 5 mL of methanol. The vigorously stirred mixture was warmed to 80 °C for 12 h. The solvent was evaporated, and the precipitate was partitioned between methylene chloride (200 mL) and 2 M aqueous Na₂CO₃ (15 mL) containing 3 mL of concentrated ammonia. The organic layer was dried (Na₂SO₄) and then concentrated to dryness under reduced pressure to afford **9a** as a white solid in 73% yield; mp 299–300 °C (DMF) (Lit.³⁴ mp 272 °C). ¹H NMR (DMSO- d_6): δ 7.89 (s, 4H), 7.94–7.96 (m, 8H). MS (ESI) m/e (rel int.): 280 (M⁺, 100).

[1,1';4',1']Terphenyl-4,4"-bis-N-hydroxyamidine (11a). A mixture of hydroxylamine hydrochloride (1.83 g, 26.3 mmol, 10 equiv) in anhydrous DMSO (30 mL) was cooled to 5 °C, and then

potassium *t*-butoxide (2.95 g, 26.3 mmol, 10 equiv) was added portion wise. To the mixture was added dinitrile **9a** (728 mg, 2.6 mmol), and the reaction was kept stirring overnight at room temperature. The reaction mixture was poured onto ice/water, whereupon a white precipitate of the diamidoxime formed. The product was collected by filtration and washed with water to afford **11a** (free base) in 68% yield; mp > 300 °C. ¹H NMR (DMSO-*d*₆): δ 5.67 (br s, 4H), 7.71–7.79 (m, 12H), 9.60 (br s, 2H). MS (ESI) *m/e* (rel int.): 347 (M⁺+1, 40), 279 (100). High-resolution Calc. for C₂₀H₁₉N₄O₂ ms 347.1508. Observed: 347.1500.

Hydrochloride Salt of 11a. The hydrochloride salt of **11a** was prepared by suspending the free base in dry ethanol, cooling the mixture in an ice bath, and passing HCl gas for about 10 min; mp \geq 300 °C. ¹H NMR (D₂O/DMSO- d_6): δ 7.28-7.35 (m, 4H), 7.77-7.80 (m, 8H). ¹³C NMR (DMSO- d_6): δ 156.1, 138.3, 137.3, 134.9, 127.7, 127.1, 124.7. Anal. Calc. for C₂₀H₁₈N₄O₂-2.0HCl-0.25H₂O-0.25C₂H₅OH: C, 56.56; H, 5.09; N, 12.87. Found: C, 56.56; H, 5.12; N, 12.55.

[1,1';4',1"]Terphenyl-4,4"-bis-amidine Acetate Salt (10a). To a solution of 11a (347 mg, 1 mmol) in glacial acetic acid (10 mL) was slowly added acetic anhydride (0.35 mL). After stirring overnight and TLC indicated complete acylation of the starting material, 10% palladium on carbon (80 mg) was then added. The mixture was placed on a Parr hydrogenation apparatus at 50 psi for 4 h at room temperature. The mixture was filtered through Hyflo and the filter pad washed with water. The filtrate was evaporated under reduced pressure, and the precipitate was collected and washed with ether to give 10a; mp >300 °C. ¹H NMR (D₂O/DMSO- d_6): δ 1.78 (s, 2 × CH₃), 7.52–7.54 (m, 4H), 7.58–7.61 (m, 2H), 7.76–7.90 (m, 6H). MS (ESI) m/e (rel int.): 315 (M⁺+1, 33), 158 (100), 121 (40). Anal. Calc. For C₂₀H₁₈N₄-2.0CH₃CO₂H-0.5H₂O: C, 64.99; H, 6.13; N, 12.63. Found: C, 64.97; H, 6.01; N. 12.60.

[1,1';4',1"]Terphenyl-4,4"-bis-N-methoxyamidine (12a). To a suspension of the amidoxime 11a (347 mg, 1 mmol) in DMF (15 mL) was added LiOH·H₂O (252 mg, 6 mmol, in 3 mL of H₂O), which was followed by dimethyl sulfate (630 mg, 5 mmol). The reaction mixture was kept stirring overnight, after which it was poured onto ice/water, and the precipitate was filtered, washed with water, and dried to give the desired compound in 72% yield; mp 270–272 °C. ¹H NMR (DMSO- d_6): δ 3.75 (s, 6H), 6.10 (br s, 4H), 7.75–7.80 (m, 12H).

Hydrochloride Salt of 12a. Mp 240–242 °C. ¹H NMR (DMSO- d_6): δ 3.86 (s, 6H), 7.95–7.90 (m, 12H), 8.45 (br s, 4H). MS (ESI) m/e (rel int.): 375 (M⁺ +1, 100), 346 (25). High-resolution Calc. for $C_{22}H_{23}N_4O_2$ ms 375.1821. Observed: 375.1819. Anal. Calc. for $C_{22}H_{22}N_4O_2$ -2.0HCl: C, 59.07; H, 5.41; N, 12.52. Found: C, 59.54; H, 5.44; N, 12.10.

2'-Fluoro-[1,1';4',1'']terphenyl-4,4''-bis-carbonitrile (9b). Adopting the same procedure used for the preparation of **9a**, a Suzuki coupling reaction was performed using 2,5-dibromo-1-flourobenzene (3.50 g, 13.78 mmol) and 4-cyanophenylboronic acid (4.45 g, 30.32 mmol) to yield the target bis-cyano derivative in 89% yield; mp 289–291 °C. 1 H NMR (DMSO- d_6): δ 7.72–7.83 (m, 5H) 7.90–7.99 (m, 6H). Anal. Calc. for $C_{20}H_{11}FN_2$: C, 80.52; H, 3.71. Found: C, 80.24; H, 3.95.

2'-Fluoro-[1,1';4',1'']terphenyl-4,4"-bis-amidine Hydrochloride Salt (10b). Dinitrile **9b** (498 mg, 1.67 mmol), suspended in freshly distilled THF (5 mL), was treated with lithium trimethylsilylamide (1 M solution in THF, 4 mL, 4 mmol), and the reaction was allowed to stir overnight. The reaction mixture was then cooled to 0 °C and to which was added HCl saturated ethanol (100 mL), whereupon a precipitate started forming. The mixture was left to run overnight, after which it was diluted with ether, and the resultant solid was collected by filtration. The diamidine was purified by neutralization with 1 N NaOH followed by filtration of the resultant solid and washing with water (3×). Finally, the free base was stirred with ethanolic HCl overnight and diluted with ether, and the solid formed was filtered and dried to give diamidine salt **10b**; mp > 300 °C. ¹H NMR (DMSO- d_6): δ 7.72–7.89 (m, 5H), 7.97–8.07 (m, 6H), 9.34 (br s, 4H), 9.54 (br s, 4H). MS (ESI) m/e (rel int.): 333

(M⁺, 100), 316 (23), 299 (27). Anal. Calc. for $C_{20}H_{17}FN_4$ -2.0HCl-0.75H₂O: C, 57.35; H, 4.93; N, 13.37. Found: C, 57.54; H, 4.88; H, 13.35.

2'-Fluoro-[1,1';4',1"]terphenyl-4,4"-bis-*N***-hydroxyamidine (11b).** The same procedure described for the preparation of **11a** was used starting with **9b**; yield 96%; mp > 300 °C. 1 H NMR (DMSO- d_6): δ 5.83 (br s, 4H), 7.62-7.73 (m, 5H), 7.79-7.81 (m, 6H), 9.60 (br s, 2H). MS (ESI) m/e (rel int.): 364 (M⁺, 100), 183 (42).

Hydrochloride Salt of 11b. Mp 282–284 °C. ¹H NMR (D₂O/DMSO- d_6): δ 7.74–7.86 (m, 9H), 8.02–8.05 (m, 2H). Anal. Calc for C₂₀H₁₇FN₄O₂-2.0HCl-0.5H₂O: C, 53.82; H, 4.51; N, 12.55. Found: C, 53.88; H, 4.43; N, 12.29.

2'-Fluoro-[1,1';4',1'']terphenyl-4,4''-bis-methoxyamidine (12b). The target compound was prepared by adopting the same procedure mentioned for **12a** starting with **11b**; yield 74%; mp 172–174 °C. 1 H NMR (DMSO- 4 6): δ 3.76 (s, 6H), 6.11 (br s, 4H), 7.60–7.78 (m, 11H). MS (ESI) m/e (rel int.): 393 (M⁺ + 1, 100), 197 (28).

Hydrochloride Salt of 12b. Mp 250–252 °C. ¹H NMR (D₂O/DMSO- d_6): δ 3.85 (s, 6H), 7.71–7.88 (m, 9H), 7.96 (d, J=7.8 Hz, 2H). Anal. Calc. for C₂₂H₂₁FN₄O₂-2.0HCl-0.25H₂O-0.25C₂H₅-OH: C, 56.14; H, 5.23; N, 11.63. Found: C, 56.13; H, 4.93; N, 11.43

2'-Trifluoromethyl-[1,1';4',1'']terphenyl-4,4''-bis-carbonitrile (9c). Following the same synthetic procedure employed for **9a** using 1,4-dibromo-2- triflouromethylbenzene (3.33 g, 10 mmol) and 4-cyanophenylboronic acid (3.23 g, 22 mmol) yielded dinitrile **9c** as a white solid (87%); mp 181–183 °C. ¹H NMR (DMSO- d_6): δ 7.55–7.58 (m, 3H), 7.94–8.02 (m, 6H), 8.04–8.16 (m, 2H). Anal. Calc. for C₂₁H₁₁F₃N₂: C, 72.41; H, 3.18. Found: C, 72.68; H, 3.34.

2'-Trifluoromethyl-[1,1';4',1'']terphenyl-4,4''-bis-amidine Hydrochloride Salt (10c). The diamidine was prepared following the procedure used for **10b**; mp > 300 °C. ^{1}H NMR (DMSO- d_{6}): δ 7.56–7.64 (m, 3H), 7.96–8.18 (m, 8H), 9.36 (br s, 4H), 9.57 (br s, 4H). MS (ESI) m/e (rel int.): 383 (M+ + 1, 23), 192 (100). Anal. Calc. for C₂₁H₁₇F₃N₄-2.0HCl-0.5H₂O-0.1C₂H₅OH: C, 54.30; H, 4.42; N, 11.94. Found: C, 54.11; H, 4.24; N, 11.66.

2'-Trifluoromethyl-[1,1';4',1"]terphenyl-4,4"-bis-*N***-hydroxyamidine (11c).** The same procedure described for the preparation of **11a** was used starting with **9c**; yield 94%; mp 204–206 °C. ¹H NMR (DMSO- d_6): δ 5.96 (br s, 4H), 7.37 (d, J = 7.8 Hz, 2H), 7.50 (d, J = 7.8 Hz, 1H), 7.75 (d, J = 7.8 Hz, 2H), 7.80–7.82 (m, 4H), 8.03–8.07 (m, 2H), 9.76 (br s, 2H). MS (ESI) m/e (rel int.): 415 (M⁺ + 1, 26), 208 (100).

Hydrochloride Salt of 11c. Mp 229-231 °C. ¹H NMR (D₂O/DMSO- d_6): δ 7.55-7.61 (m, 3H), 7.84-7.92 (m, 4H), 8.07-8.18 (m, 4H). Anal. Calc. for C₂₁H₁₇F₃N₄O₂-2.0HCl-1.25H₂O-0.25C₂H₅OH: C, 49.53; H, 4.44; N, 10.74. Found: C, 49.55; H, 4.40; N, 10.55.

2'-Trifluoromethyl-[1,1';4',1'']terphenyl-4,4''-bis-methoxyamidine (12c). Amidoxime **11c** was used to prepare the corresponding methoxime using the above-mentioned method in 72% yield; mp 186–188 °C. ¹H NMR (DMSO- d_6): δ 3.75 (s, 3H), 3.76 (s, 3H), 6.14 (br s, 4H), 7.36 (d, J = 8.4 Hz, 2H), 7.50 (d, J = 8.1 Hz, 1H), 7.73 (d, J = 8.1 Hz, 2H), 7.80–7.82 (m, 4H), 8.03–8.07 (m, 2H). MS (ESI) m/e (rel int.): 443 (M⁺ + 1, 59), 222 (100).

Hydrochloride Salt of 12c. Mp 214-216 °C. ¹H NMR (D₂O/DMSO- d_6): δ 3.84 (s, 6H), 7.51 (m, 3H), 7.83 (d, J=8.1 Hz, 2H), 7.89 (d, J=8.4 Hz, 2H), 7.98 (d, J=8.1 Hz, 2H), 8.11-8.15 (m, 2H). Anal. Calc. for C₂₃H₂₁F₃N₄O₂-2.0HCl-1.25H₂O-0.5C₂H₅OH: C, 51.39; H, 5.12; N, 9.98. Found: C, 51.54; H, 4.84; N, 9.94.

Phenyl[1,1']pyridyl[4',1"]phenyl-4,4"-bis-carbonitrile (9d). 2,5-Dibromopyridine and 4-cyanophenylboronic acid were reacted under the above-mentioned Suzuki coupling conditions to give the target dinitrile 9d, which was purified by column chromatography (EtOAc/Hexane, 80:20); yield 84%; mp 270–272 °C (Lit.³⁵ mp 264–266 °C). ¹H NMR (DMSO- d_6): δ 7.97–8.05 (m, 6H), 8.25 (dd, J=1.8, 8.1 Hz, 1H), 8.33–8.38 (m, 3H), 9.13 (d, J=1.8 Hz, 1H). MS (ESI) m/e (rel int.): 281 (M⁺, 100).

Phenyl[1,1']pyridyl[4',1'']phenyl-4,4''-bis-*N***-hydroxyamidine (11d).** The diamidoxime was obtained from dinitrile **9d** using the standard procedure in 97% yield; mp > 300 °C. ¹H NMR (DMSO- d_6): δ 5.89 (br s, 4H), 7.79-7.81 (m, 6H), 8.08-8.23 (m, 4H), 9.03 (d, J = 1.5 Hz, 1H), 9.74 (br s, 2H). ¹³C NMR (DMSO- d_6): δ 154.3, 150.4, 150.3, 147.2, 138.3, 136.8, 134.6, 133.7, 133.4, 132.8, 126.1, 125.9, 125.8, 125.5, 119.9. MS (ESI) m/e (rel int.): 348 (M⁺ + 1, 52), 174 (100).

Hydrochloride Salt of 11d. Mp 292–294 °C. ¹H NMR (D₂O/DMSO- d_6): δ 7.89–7.91 (m, 4H), 8.09 (d, J = 8.7 Hz, 2H), 8.27 (d, J = 8.7 Hz, 1H), 8.36–8.40 (m, 3H), 9.15 (d, J = 1.8 Hz, 1H). Anal. Calc. for C₁₉H₁₇N₅O₂-3.0HCl-1.0H₂O-0.3C₂H₅OH: C, 48.18; H, 4.90; N, 14.33. Found: C, 48.18; H, 4.74; N, 14.18.

Phenyl[1,1']pyridyl[4',1'']phenyl-4,4''-bis-amidine Acetate Salt (10d). Diamidine 10d was synthesized in two consecutive steps as described for 10a. First by acetylation of amidoxime 11d to give the bis-acetoxime intermediate and second by direct reduction; mp 273–275 °C. ¹H NMR (D₂O/DMSO- d_6): δ 1.70 (s, 2.8 x CH₃), 7.84–7.89 (m, 4H), 8.06–8.25 (m, 6H), 8.95 (s, 1H). MS (ESI) m/e (rel int.): 316 (M⁺ + 1, 100), 158 (98). 13 C NMR (D₂O/DMSO- d_6 , of hydrochloride salt): δ 165.9, 154.8, 148.4, 143.5, 142.2, 136.5, 134.2, 129.3, 129.0, 128.5, 127.9, 127.8, 127.7, 122.0. Anal. Calc. for C₁₉H₁₇N₅-2.8CH₃CO₂H-0.75H₂O: C, 59.44; H, 6.02; N, 14.09. Found: C, 59.27; H, 5.92; N, 14.16.

Phenyl[1,1']pyridyl[4',1'']phenyl-4,4''-bis-N-methoxyamidine (12d). Diamidoxime **11d** was used to prepare the target compound using the standard procedure in 89% yield; mp 245–247 °C. ¹H NMR (DMSO- d_6): δ 3.76 (s, 6H), 6.13 (br s, 4H), 7.78–7.84 (m, 6H), 8.08–8.23 (m, 4H), 9.03 (d, J=2.4 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 154.3, 150.6, 150.5, 147.5, 138.8, 137.4, 134.9, 133.5, 133.0, 132.2, 126.4, 126.3, 126.1, 126.0, 120.2, 60.6. MS (ESI) m/e (rel int.): 376 (M⁺ + 1, 100).

Hydrochloride Salt of 12d. Mp 241–242 °C. ¹H NMR (D₂O/DMSO- d_6): δ 3.88 (s, 6H), 7.94 (d, J = 8.7 Hz, 4H), 8.05 (d, J = 8.7 Hz, 2H), 8.27 (d, J = 8.7 Hz, 1H), 8.34–8.41 (m, 3H), 9.14 (d, J = 2.4 Hz, 1H). Anal. Calc. for C₂₁H₂₁N₅O₂-3.0HCl-1.75H₂O-0.5C₂H₅OH: C, 48.98; H, 5.69; N, 12.98. Found: C, 48.99; H, 5.56; N, 12.75.

- **2,5-Bis-(4'-cyanophenyl)-pyrimidine (9e).** 2-Chloro-5-bromopyrimidine³⁶ and 4-cyanophenylboronic acid were reacted under the above-mentioned Suzuki coupling conditions to give target dinitrile **9e**; 91% yield; mp 305-306.5 °C (DMF). ¹H NMR (DMSO- d_6): δ 7.98-8.07 (m, 6H), 8.58 (s, 2H), 9.34 (s, 2H). ¹³C NMR (DMSO- d_6): δ 161.1, 155.5, 140.4, 137.8, 132.6, 132.3, 129.9, 128.0, 127.4, 118.1, 118.0, 113.0, 111.3. MS (ESI) m/e (rel int.): 282 (M⁺, 100), 141 (10), 127 (80). Anal. Calc. for C₁₈H₁₀N₄: C, 76.58; H, 3.57. Found: C, 76.25; H, 3.82.
- **2,5-Bis-(4'-amidinophenyl)-pyrimidine (10e).** The same procedure described for the preparation of **10b** was used starting with **9e**; 86% yield; mp > 300 °C. ¹H NMR (D₂O/DMSO- d_6): δ 7.91 8.00 (m, 6H), 8.54 (s, 2H), 9.26 (s, 2H). ¹³C NMR (D₂O/DMSO- d_6): δ 166.1, 166.0, 162.4, 156.3, 142.0, 139.3, 131.1, 130.3, 129.4, 129.0, 128.5, 128.0, 122.2. MS (ESI) m/e (rel int.): 317 (M⁺ + 1, 100), 159 (45). Anal. Calc. for C₁₈H₁₆N₆-3.0HCl-0.25H₂O: C, 50.24; H, 4.57; N, 19.53. Found: C, 50.33; H, 4.80; N, 19.47.
- **2,5-Bis-[4'-(N-hydroxyamidino)phenyl]-pyrimidine (11e).** The same procedure described for the preparation of **11a** was used starting with **9e**; 97% yield; mp 290–292 °C. ¹H NMR (DMSO- d_6): δ 5.71 (s, 4H), 7.84–7.87 (m, 6H), 8.42 (s, 2H), 9.23 (s, 2H), 9.58 (s, 2H). ¹³C NMR (DMSO- d_6): δ 161.6, 154.7, 150.2, 150.1, 136.9, 135.2, 133.7, 133.4, 130.2, 127.0, 126.0, 125.8, 125.3. MS (ESI) m/e (rel int.): 349 (M⁺ + 1, 100), 332 (15), 315 (10), 282 (60).
- **2,5-Bis-[4'-(N-methoxyamidino)phenyl]-pyrimidine (12e).** The same procedure described for the preparation of **12a** was used starting with **11e**; free base yield 64%; mp 217–218 °C (DMF). ¹H NMR (DMSO- d_6): δ 3.76 (s, 6H), 6.19 (s, 4H), 7.83–7.88 (m, 6H), 8.43 (s, 2H), 9.28 (s, 2H). ¹³C NMR (DMSO- d_6): δ 161.6, 155.0, 150.5, 150.4, 137.4, 134.5, 134.3, 132.6, 130.3, 127.2, 126.3, 126.2, 125.9, 60.5. MS (ESI) m/e (rel int.): 377 (M⁺ + 1, 100), 347 (30), 330 (25).

- **Hydrochloride Salt of 12e.** Mp 242-243 °C. ¹H NMR (D₂O/DMSO- d_6): δ 3.81 (s, 6H), 7.81-7.84 (m, 4H), 7.99 (s, 2H), 8.49 (s, 2H), 9.26 (s, 2H). Anal. Calc. for C₂₀H₂₀N₆O₂-2.6HCl-0.75C₂H₅-OH): C, 51.05; H, 5.40; N, 16.61. Found: C, 51.35; H, 5.46; N, 16.44.
- **1,4-Bis-(5'-cyanopyridin-2'-yl)phenylene (9f).** The same procedure described for **9a** was used by employing 6-chloronicotinonitrile (2 equiv) and 1,4-phenylenebisboronic acid (1 equiv) to furnish **9f** in 94% yield; mp >300 °C (DMF). ¹H NMR (DMSO- d_6): δ 8.10–8.40 (m, 8H), 9.18 (s, 2H). MS (ESI) m/e (rel int.): 282 (M⁺, 100), 254 (10), 179 (20). Anal. Calc. for C₁₈H₁₀N₄: C, 76.58; H, 3.57. Found: C, 76.33; H, 3.71.
- **1,4-Bis-(5'-amidinopyridin-2'-yl)-phenylene** (**10f).** The same procedure described for the preparation of **10b** was used starting with **9f**; 90% yield; mp > 300 °C. ¹H NMR (D₂O/DMSO- d_6): δ 8.42 (m, 8H), 9.15 (s, 2H). ¹³C NMR (D₂O/DMSO- d_6): δ 164.5, 160.3, 149.1, 139.3, 138.0, 128.5, 123.6, 121.3. MS (ESI) m/e (rel int.): 317 (M⁺ + 1, 100), 300 (70), 283 (50), 273 (15), 246 (35). Anal. Calc. for C₁₈H₁₆N₆-4.0HCl: C, 46.77; H, 4.36; N, 18.18. Found: C, 46.98; H, 4.55; N, 17.89.
- **1,4-Bis-[5'-(N-hydroxyamidinopyridin-2'-yl)]-phenylene (11f).** The same procedure described for the preparation of **11a** was used starting with **9f**; 96% yield; mp > 300 °C. ¹H NMR (DMSO- d_6): δ 5.88 (s, 4H), 8.03 (d, J = 8.1 Hz, 2H), 8.13 (d, J = 8.1 Hz, 2H), 8.23-8.36 (m, 4H), 8.99 (s, 2H), 9.75 (s, 2H). 13 C NMR (DMSO- d_6): δ 155.3, 148.7, 146.4, 138.6, 133.6, 127.6, 126.6, 119.3. MS (ESI) m/e (rel int.): 349 (M⁺ + 1, 100), 334 (30), 282 (20).
- **1,4-Bis-[5'-(N-methoxyamidinopyridin-2'-yl)]-phenylene (12f).** The same procedure described for the preparation of **12a** was used starting with **11f** to furnish the free base of **12f** in 70% yield; mp 218–219 °C (DMF). ¹H NMR (DMSO- d_6): δ 3.78 (s, 6H), 6.29 (s, 4H), 8.05–8.15 (m, 4H), 8.26 (s, 4H), 8.95 (s, 2H). ¹³C NMR (DMSO- d_6): δ 155.7, 149.0, 146.8, 138.7, 134.3, 127.1, 126.9, 119.7, 60.8. MS (ESI) m/e (rel int.): 377 (M⁺ + 1, 100), 330 (10), 189 (25).
- **Hydrochloride Salt of 12f.** Mp 251–253 °C. ¹H NMR (D₂O/DMSO- d_6): δ 3.82 (s, 6H), 8.20 (s, 4H), 8.26 (s, 4H), 8.94 (s, 2H). Anal. Calc. for C₂₀H₂₀N₆O₂-4.0HCl-2.0H₂O-0.2C₂H₅OH): C, 43.17; H, 5.18; N, 14.80. Found: C, 43.32; H, 5.04; N, 14.42.
- **2,5-Bis(4-cyanophenyl)-1,4-dimethoxybenzene** (**9g**). The same procedure described for **9a** was used by employing 1,4-dibromo-2,5-dimethoxybenzene (1 equiv) and 4-cyanophenylboronic acid (2 equiv) to furnish **9g** in 87% yield; mp 299–301 °C (DMF), Lit.³⁷ mp 278 °C. ¹H NMR (DMSO- d_6): δ 3.78 (s, 6H), 7.13 (s, 2H), 7.78 (d, J=8.4 Hz, 4H), 7.91 (d, J=8.4 Hz, 4H).
- **2,5-Bis(4-amidinophenyl)-1,4-dimethoxybenzene** (**10g**). The same procedure described for the preparation of **10b** was used starting with **9g**; yield 90%; mp > 300 °C. ¹H NMR (D₂O/DMSO- d_6): δ 3.80 (s, 6H), 7.13 (s, 2H), 7.81 (d, J=7.2 Hz, 4H), 7.91 (d, J=7.2 Hz, 4H). ¹³C NMR (D₂O/DMSO- d_6): δ 165.8, 150.9, 143.4, 130.1, 129.6, 128.0, 126.7, 115.2, 56.9. MS (ESI) m/e (rel int.): 375 (M⁺ + 1, 20), 358 (25), 345 (40), 328 (100). Anal. Calc. for C₂₂H₂₂N₄O₂-2.0HCl-2.25H₂O: C, 54.15; H, 5.88; N, 11.48. Found: C, 54.05; H, 5.81; N, 11.16.
- **5-(4'-Formylbiphenyl-4-yl)-furan-2-carboxaldehyde (14a)**. The same procedure described for **9a** was used employing 5-(4-bromophenyl)-furan-2-carboxaldehyde (1.25 g, 5 mmol), and 4-formylphenyl-boronic acid (894 mg, 6 mmol) to afford **14a** in 85% yield; mp 173–174 °C (SiO₂, hexanes/EtOAc, 70:30). ¹H NMR (DMSO- d_6); δ 7.42 (d, J=3.6 Hz, 1H), 7.70 (d, J=3.6 Hz, 1H), 7.93–8.01 (m, 8H), 9.64 (s, 1H), 10.07 (s, 1H). ¹³C NMR (DMSO- d_6); δ 192.7, 177.9, 157.6, 151.8, 144.6, 139.6, 135.3, 130.2, 128.6, 127.9, 127.3, 125.7, 109.5. MS (ESI) m/e (rel int.): 276 (M⁺, 100), 247 (5), 219 (25), 189 (25). Anal. Calc. for $C_{18}H_{12}O_3$: C, 78.24; H, 4.37. Found: C, 77.99; H, 4.44.
- **5-(4'-Cyanobiphenyl-4-yl)-furan-2-carbonitrile (15a)**. To a stirred solution of **14a** (552 mg, 2 mmol) in 10 mL of methanol was slowly added an aqueous solution (8 mL) of hydroxylamine hydrochloride (280 mg, 4 mmol) and sodium carbonate (424 mg, 4 mmol). The reaction mixture was allowed to reflux for 6 h. The solvent was evaporated, the precipitate was partitioned between

water and ethyl acetate (150 mL), and the organic layer was dried (Na₂SO₄) and then concentrated to dryness under reduced pressure. The crude oxime was allowed to reflux in acetic anhydride (8 mL) for 4 h. The reaction mixture was poured slowly onto ice—water and the precipitate was filtered and washed with water to afford **15a** in 59% yield; mp 216–218 °C. ¹H NMR (DMSO- d_6); δ 7.38 (d, J = 3.6 Hz, 1H), 7.75 (d, J = 3.6 Hz, 1H), 7.88–7.96 (m, 8H). ¹³C NMR ((DMSO- d_6); δ 157.4, 143.4, 138.9, 132.9, 128.3, 127.8, 127.5, 125.8, 125.4, 124.3, 118.7, 112.0, 110.4, 108.3. MS (ESI) m/e (rel int.): 270 (M⁺, 100), 241 (5), 214 (10). Anal. Calc. for $C_{18}H_{10}N_2O$: C, 79.99; H, 3.73. Found: C, 79.85; H, 3.91.

5-[4'-(N-Hydroxyamidino)-biphenyl-4-yl]-furan-2-N-hydroxyamidine (16a). The same procedure described for **11a** was used starting with **15a** to furnish **16a** (free base) in 92% yield; mp 229–230 °C. 1 H NMR (DMSO- 4 G); δ 5.87 (s, 4H), 6.85 (d, J = 3.6 Hz, 1H), 7.06 (d, J = 3.6 Hz, 1H), 7.71–7.87 (m, 8H), 9.70 (s, 2H).

Hydrochloride Salt of 16a. Mp >300 °C. ¹H NMR (D₂O/DMSO- d_6); 7.38 (d, J=3.9 Hz, 1H), 7.82 (d, J=3.9 Hz, 1H), 7.87 (d, J=8.4 Hz, 2H), 7.92 (d, J=8.4 Hz, 2H), 8.01 (d, J=8.4 Hz, 2H), 8.15 (d, J=8.4 Hz, 2H). 13 C NMR (DMSO- d_6); δ 158.7, 158.5, 148.7, 143.5, 138.8, 138.4, 128.6, 128.5, 127.4, 126.9, 125.5, 124.6, 119.2, 108.8. MS (ESI) m/e (rel int.): 337 (M⁺ + 1, 65), 322 (50), 307 (100), 272 (40). Anal. Calc. for $C_{18}H_{16}N_4O_{3-2.0}HCl$: C, 52.82; H, 4.43; N, 13.68. Found: C, 52.60; H, 4.45; N, 13.30.

5-[4'-(*N*-Methoxyamidino)-biphenyl-4-yl]-furan-2-*N*-methoxyamidine (17). The same procedure described for **12a** was used starting with **16a** to give free base of **17** in 92% yield; mp 214–215 °C. ¹H NMR (DMSO- d_6); δ 3.75 (s, 6H), 6.12 (s, 2H), 6.17 (s, 2H), 6.91 (d, J = 3.6 Hz, 1H), 7.07 (d, J = 3.6 Hz, 1H), 7.76–8.00 (m, 8H).

Hydrochloride Salt of 17. Mp 194–196 °C. ¹H NMR (D₂O/DMSO- d_6): δ 3.88 (s, 6H), 7.20 (d, J=3.6 Hz, 1H), 7.34 (d, J=3.6 Hz, 1H), 7.77–8.13 (m, 8H). Anal. Calc. for C₂₀H₂₀N₄O₃-2.0HCl: C, 54.92; H, 5.07; N, 12.81. Found: C, 54.88; H, 5.29; N, 12.99.

5-(4'-Amidinobiphenyl-4-yl)-furan-2-amidine Acetate Salt (18a). The same procedure described for 10a was used starting with 16a to give 18a in 71% yield; mp 236–238 °C. ¹H NMR (D₂O/DMSO- d_6); δ 1.78 (s, 2 x CH₃), 7.31 (d, J = 3.6 Hz, 1H), 7.61 (d, J = 3.6 Hz, 1H), 7.85–8.11 (m, 8H). EIMS m/e (rel int.); 304 (M⁺, 20), 287 (100), 270 (50), 216 (30), 190 (10). High-resolution Calc. for C₁₈H₁₆N₄O ms 304.1324. Observed: 304.1320. Anal. Calc. for C₁₈H₁₆N₄O-2.0AcOH-1.75H₂O-0.25EtOH: C, 57.81; H, 6.25; N, 11.98. Found: C, 58.18; H, 5.90; N, 11.56.

4-(5-Bromopyridin-2-yl)-benzaldehyde (13). The same procedure described for **10a** was used by employing 2,5-dibromopyridine (1 equiv) and 4-formylphenylboronic acid (1 equiv); yield 71%; mp 120–121 °C (SiO₂, hexanes/EtOAc, 80:20). ¹H NMR (DMSO- d_6); δ 7.97 (d, J = 8.4 Hz, 2H), 8.04 (d, J = 8.7 Hz, 1H), 8.16 (dd, J = 2.4, 8.7 Hz, 1H), 8.26 (d, J = 8.4 Hz, 2H), 8.80 (d, J = 2.4 Hz, 1H), 10.08 (s, 1H). MS (ESI) m/e (rel int.): 262 (M⁺, 100), 232 (30), 153 (55). Anal. Calc. for C₁₂H₈BrNO: C, 54.99; H, 3.08. Found: C, 54.75; H, 3.14.

5-[6-(4-Formylphenyl)-pyridin-3-yl]-furan-2-carboxaldehvde (14b). To a stirred solution of 13 (2.62 g, 10 mmol) in 40 mL of dry 1,4-dioxane was added palladium tetrakis-triphenylphosphine (400 mg), followed by 5-(diethoxymethylfuran-2-yl)-tributyltin (4.59 g, 10 mmol), and the reaction mixture was refluxed at 100 °C for 24 h. The solvent was then evaporated to dryness to give a dark brown residue, which was suspended in water and extracted with CH₂Cl₂. The organic layer was passed over Hyflo, dried (Na₂SO₄), and evaporated to dryness under reduced pressure, followed by acid hydrolysis with 2 M HCl to furnish 14b; yield 79%; mp 204–206 °C (EtOH). ¹H NMR (DMSO- d_6); δ 7.55 (d, J = 3.6 Hz, 1H, 7.73 (d, J = 3.6 Hz, 1H), 8.06 (d, J = 8.4 Hz, 2H),8.27 (d, J = 8.4 Hz, 1H), 8.37 - 8.41 (m, 3H), 9.24 (d, J = 2.1 Hz,1H), 9.67 (s, 1H), 10.09 (s, 1H). ¹³C NMR (DMSO- d_6); δ 192.8, 178.1, 155.2, 154.7, 152.3, 146.3, 142.9, 136.5, 133.3, 130.0, 127.2, 125.1, 124.3, 121.4, 110.6. Anal. Calc. for C₁₇H₁₁NO₃: C, 73.63; H, 3.99. Found: C, 73.75; H, 3.81.

5-[6-(4-Cyanophenyl)-pyridin-3-yl]-furan-2-carbonitrile (15b). The same procedure described for **15a** was used starting with **14b**; yield 40%; mp 203–205 °C. 1 H NMR (DMSO- d_{6}); δ 7.53 (d, J = 3.6 Hz, 1H), 7.82 (d, J = 3.6 Hz, 1H), 8.00 (d, J = 8.4 Hz, 2H), 8.27 (d, J = 8.4 Hz, 1H), 8.35–8.37 (m, 3H), 9.21 (d, J = 2.1 Hz, 1H). MS (ESI) m/e (rel int.): 272 (M⁺ + 1, 100), 182 (15). Anal. Calc. for $C_{17}H_{9}N_{3}O$: C, 75.26; H, 3.34. Found: C, 75.44; H, 3.54.

5-{6-[4-(N-Hydroxyamidino)-phenyl]-pyridin-3-yl}-furan-2-*N***-hydroxyamidine (16b).** The same procedure described for **11a** was used starting with **15b**; yield 90%; mp 194–195 °C. ¹H NMR (DMSO- d_6); δ 5.89 (s, 2H), 5.95 (s, 2H), 6.90 (d, J = 3.6 Hz, 1H), 7.22 (d, J = 3.6 Hz, 1H), 8.00 (d, J = 8.4 Hz, 2H), 8.07–8.26 (m, 4H), 9.11 (d, J = 2.1 Hz, 1H), 9.76 (s, 2H). MS (ESI) m/e (rel int.): 338 (M⁺ + 1, 100), 323 (20). Anal. Calc. for $C_{17}H_{15}N_5O_3$: C, 60.52; H, 4.48. Found: C, 60.73; H, 4.22.

5-[6-(4-Amidinophenyl)-pyridin-3-yl]-furan-2-carboxamidine acetate salt (18b). The same procedure described for **10a** was used starting with **16b**; yield 57%; mp 239–241 °C. ¹H NMR (D₂O/DMSO- d_6); δ 1.93 (s, 3 x CH₃), 7.31 (d, J = 3.6 Hz, 1H), 7.42 (d, J = 3.6 Hz, 1H), 7.92 (d, J = 8.4 Hz, 2H), 8.15–8.37 (m, 4H), 9.24 (s, 1H). EIMS m/e (rel int.); 306 (M⁺ +1, 55), 289 (10), 273 (15), 237 (100). High-resolution Calc. for C₁₇H₁₆N₅O ms 306.1354. Observed: 306.1349. Anal. Calc. for C₁₇H₁₅N₅O-3.0AcOH-2.4H₂O: C, 52.25; H, 6.02; N, 13.25. Found: C, 51.95; H, 5.70; N, 12.90.

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Supporting Information Available: Elemental analysis for new compounds and CD spectra titration of **10a** into poly(dA).poly(dT). This material is available free of charge via the Internet at http://pubs.acs.org

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