Synthesis and Structure-Activity Relationships of a New Set of 2-Arylpyrazolo[3,4-c]quinoline Derivatives as Adenosine Receptor Antagonists

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Received March 10, 2000

In a recent paper (Colotta et al. J. Med. Chem. 2000, 43, 1158–1164) we reported the synthesis and adenosine receptor binding activity of two sets of 2-aryl-1,2,4-triazolo[4,3-a]quinoxalines (A and B) some of which were potent and selective A_1 or A_3 antagonists. In this paper the synthesis of a set of 2-arylpyrazolo [3,4-c] quinolin-4-ones 1-10, 4-amines 11-18, and 4-aminosubstituted derivatives 19-35 are reported. The binding activity at bovine A_1 and A_{2A} and human cloned A₃ adenosine receptors showed that (i) the substituent on the appended 2-phenyl ring could be used to modulate A₁ and A₃ affinity, (ii) the 4-amino group was necessary for A₁ and A_{2A} binding activity, and (iii) a nuclear or extranuclear C=O proton acceptor at position 4 yielded potent and selective A₃ antagonists. These results are in agreement with those of the previously reported series A and B suggesting a similar adenosine receptor binding mode. In particular, the A_3 nanomolar affinity of 1-8, 31-33, and 35 confirms the hypothesis of the presence in the N-6 region of the adenosine A₃ subtype of a proton donor able to bind to a C=O proton acceptor at position 4.

Introduction

Adenosine is thought to mediate a wide variety of effects by interacting with four subtypes of adenosine receptors (AR): A₁, A_{2A}, A_{2B}, and A₃. All four AR subtypes are coupled via a G-protein to the enzyme adenylcyclase in either an inhibitory (A1 and A3 subtypes) or stimulatory manner (A2A and A2B subtypes).1 For A₁, A_{2A}, and A₃ AR, selective agonists and antagonists are now available, $^{2-7}$ while the A_{2B} subtype still lacks selective ligands.

In recent years, the potential therapeutic use of selective AR subtype antagonists as renal protective, 8,9 anti-Parkinson, ¹⁰ antiinflammatory, antiasthmatic, and antiischemic agents^{11–14} has attracted great attention. Since most of the AR antagonists are nitrogen-containing heterocyclic compounds, some research in our laboratory has been directed toward the synthesis of tricyclic heteroaromatic systems as AR antagonists. 15-19

In a recent paper²⁰ we reported the synthesis and binding activity at bovine A₁ and A_{2A} AR and at human cloned A₃ AR of two sets of 2-aryl-1,2,4-triazolo[4,3-a]quinoxalines (A and B). Some of these A and B compounds were potent and selective A₁ or A₃ antagonists. A structure—activity relationship (SAR) study on the 1,4-dione and 4-amino-1-one series (see A and B, respectively, Chart 1) showed that the 4-NH₂ proton donor group was essential for A1 and A2A receptorligand interaction while it was not necessary for A₃ recognition. The binding results indicated also that the presence of a 4-oxo function (series A) or of an acyl substituent on the 4-amino group (series **B**) afforded

some potent and selective A₃ receptor antagonists. Moreover, in both series the nature and the position of the substituent on the 2-phenyl ring could be used to modulate the A_1/A_3 selectivity. To verify whether this structural requirement could be applied to tricyclic systems of similar size and shape, we report in this paper the synthesis and A₁, A_{2A}, and A₃ AR binding activity of some 2-arylpyrazolo[3,4-c]quinolin-4-ones **1−10** and of their corresponding 4-amines **11−18** and 4-amino-substituted derivatives **19–35**, which can be considered the 1-decarbonyl analogues of series A and **B**, respectively.

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Chart 1. Previously and Hereby Reported Adenosine Receptor Antagonists

Scheme 1a

^a (a) NaH, R₁X, DMF; (b) PCl₅/POCl₃, pyridine; (c) PCl₅/POCl₃; (d) method A: cyclohexylamine; (e) method B: NH₃(g), absolute

Scheme 2^a

^a (a) Method A: excess of R₂NH₂; (b) method B: R₂NH₂, Et₃N, absolute EtOH.

Chemistry

The synthetic pathways which yielded compounds **1**−**37** are illustrated in Schemes 1−3.

The synthesis of 1, 2, 4-7, and 9, which were originally prepared as benzodiazepine receptor ligands. has already been reported.²¹ The 2-(2-methylphenyl) derivative 3 and its 2-(4-chlorophenyl) analogue 8 were obtained by reacting the 3-ethoxalylindole²² with arylhydrazine hydrochlorides as described to prepare 1, 2,

Scheme 3a

^a (a) RCOCl, pyridine, CH₂Cl₂; (b) PhCH₂COOH, 1-hydroxybenzotriazole, Et₃N, 4-(dimethylamino)pyridine, 1-(3-(dimethylamino)propyl)-3-ethylcarbodiimide hydrochloride, DMF; (c) RNCO, THF.

and 4-7.21 The 5-N-n-propyl derivative 10 ensued by the reaction of 1 with n-propyl bromide following the procedure described to prepare 9.21 Reaction of 1 and 5-7 with a mixture of PCl₅/POCl₃ and pyridine afforded the 1-(2-aryl-2*H*-pyrazolo[3,4-*c*]quinolin-4-yl)pyridinium chlorides 36-39, while the reaction of 1-6 and 8 with a neat mixture of PCl₅/POCl₃ gave the 2-aryl-4-chloro-2H-pyrazolo[3,4-c]quinolines **40–46**. It must be noted that both the pyridinium salts **36–39** and the 4-chloro derivatives **40–46** were unstable; nevertheless they were pure enough to be spectroscopically characterized and used without further purification. Refluxing 36-**39** with an excess of cyclohexylamine gave the 2-aryl-2H-pyrazolo[3,4-c]quinolin-4-amines **11** and **15**–**17**. Compound 11 was also obtained with more satisfactory yields from its corresponding 4-chloro derivative 40 and ammonia. Thus, the other 4-amino derivatives 12–14 and 18 were prepared following this pathway, i.e., from the corresponding 4-chloro intermediates **41–43** and **46** and ammonia (Scheme 1).

Allowing the 4-chloro intermediates **40–46** to react with suitable amines gave the 4-N-cycloalkylamines **19–28** and 4-*N*-aralkylamines **29** and **30** (Scheme 2).

Finally, Scheme 3 depicts the reaction of 2-phenyl-2H-pyrazolo[3,4-c]quinolin-4-amine **11** with suitable acyl chlorides or phenylacetic acid, or with suitable isocyanates, to afford the 4-amido 31-33 and 4-ureido derivatives 34 and 35, respectively.

Biochemistry

Compounds 1-35 were tested for their ability to displace [${}^{3}H$] \mathcal{N}^{6} -cyclohexyladenosine ([${}^{3}H$]CHA) from A₁ AR in bovine cerebral cortical membranes, [3H]-2-[[4-(2-carboxyethyl)phenethyl]amino]-5'-(N-ethylcarbamoyl)adenosine ([³H]CGS 21680) from A_{2A} AR in bovine striatal membranes, and $[^{125}I]N^6$ -(4-amino-3-iodobenzyl)-5'-N-methylcarbamoyladenosine ([125I]AB-MECA) from human cloned A₃ AR stably expressed in CHO cells. In fact, due to the species differences in A₃ primary amino acid sequence, new A3 AR ligands have to be tested on cloned human A_3 receptors.^{23–25} On the contrary, for A_1 and A_{2A} AR subtypes there is a good amino acid sequence homology, since standard antagonists, as theophylline and 1,3-dipropyl-8-cyclopentylxanthine (DPCPX), showed an affinity at bovine A_1 and A_{2A} receptors comparable to those reported at the cloned human ones.26-28

Table 1. Adenosine Receptor Binding Activity

				K _i (nM) or % inhibition ^a			
	R	R_1	R_2	A ₁ ^b	A _{2A} ^c	$\mathbf{A_3}^{\mathrm{d}}$	
1	Н	Н	Н	2100 ± 170	8600 ± 710	30.8 ± 2.6	
2	Н	Cl	Н	284 ± 23	52%	44.1 ± 3.2	
3	2-Me	Н	Н	3200 ± 260	0%	79.2 ± 6.3	
4	3-Me	Н	Н	830 ± 75	17%	5.0 ± 0.4	
5	4-Me	Н	Н	3900 ± 290	22%	3.2 ± 0.2	
6	3-F	Н	Н	583 ± 49	0%	45.3 ± 3.9	
7	4-OMe	Н	Н	1078 ± 29	18%	3.2 ± 0.2	
8	4-Cl	Н	Н	464 ± 39	35%	2.9 ± 0.1	
9	Н	Н	Me	1230 ± 110	47%	118 ± 10	
10	Н	Н	n-C ₃ H ₇	950 ± 86	50%	68.4 ± 5.3	
11	Н	Н	Н	69.2 ± 4.8	331 ± 26	551 ± 34	
12	Н	Cl	Н	86.1 ± 6.3	5750 ± 480	309 ± 23	
13	2-Me	Н	Н	520 ± 38	1600 ± 140	3600 ± 280	
14	3-Me	Н	Н	129 ± 9.5	3750 ± 310	99.3 ± 7.8	
15	4-Me	Н	Н	215 ± 14	21%	188 ± 15	
16	3-F	Н	Н	110 ± 8.9	49.5 ± 3.2	788 ± 69	
17	4-OMe	Н	Н	238 ± 20	3700 ± 310	90.2 ± 7.3	
18	4-Cl	Н	Н	2290 ± 180	11200 ± 960	150 ± 12	
19	Н	Н	√ >	8.6 ± 0.6	3800 ± 320	707 ± 53	
20	Н	Cl	\sim	27.1 ± 2.2	4140 ± 380	2128 ± 170	
21	3-Me	Н	\Leftrightarrow	22.2 ± 1.8	11400 ± 980	44.1 ± 3.7	
22	4-Me	Н	\sim	71.9 ± 5.6	13%	115 ± 9.1	
23	3-F	Н	$ \sim $	54.7 ± 4.1	5750 ± 510	155 ± 12	
24	4-Cl	Н	\Leftrightarrow	478 ± 39	8%	56.5 ± 4.5	
25	Н	Н	\bigcirc	3.2 ± 0.24	849 ± 75	60.5 ± 4.8	
26	2-Me	Н	\triangle	148 ± 13	329 ± 26	152 ± 13	
27	3-Me	Н	\triangle	40.7 ± 31	1200 ± 94	22.3 ± 1.6	
28	3-F	Н	\triangle	9.3 ± 0.8	2210 ± 180	34%	
29	Н	Н	CH_2Ph	132 ± 11	5400 ± 380	35.8 ± 2.8	
30	Н	Н	$(CH_2)_2Ph$	60.1 ± 5.2	5051 ± 460	32.9 ± 2.3	
31	Н	Н	COMe	75.9 ± 6.4	2400 ± 190	48.2 ± 3.7	
32	Н	Н	COPh	42%	3%	2.1 ± 0.1	
33	Н	Н	$COCH_2Ph$	107 ± 9.3	13%	9.9 ± 0.8	
34	Н	Н	CONHPh	1500 ± 138	32%	108 ± 9.6	
35	Н	Н	CONHCH ₂ Ph	186 ± 15	5%	8.3 ± 0.7	
Theophylline				3800 ± 340	21000 ± 1800	86000 ± 7800	
DPCPX				0.5 ± 0.03	337 ± 28	1300 ± 125	

 $[^]a$ The $K_{\rm i}$ values are means \pm SEM of four separate assays, each performed in triplicate. b Displacement of specific [$^3{\rm H}$]CHA binding in bovine brain membranes or percentage of inhibition (P_0) of specific binding at 20 $\mu{\rm M}$ concentration. c Displacement of specific [$^3{\rm H}$]CGS 21680 in bovine striatal membranes or percentage of inhibition (P_0) of specific binding at 20 $\mu{\rm M}$ concentration. d Displacement of specific [$^1{\rm E}^2{\rm H}$]AB-MECA binding at human A $_3$ receptors expressed in CHO cells or percentage of inhibition (P_0) of specific binding at 1 $\mu{\rm M}$ concentration.

The binding results of 1-35, together with those of the ophilline and DPCPX included as antagonist reference compounds, are shown in Table 1.

Results and Discussion

The A_1 , A_{2A} , and A_3 binding results of compounds **1–35** displayed in Table 1 show that the syntheses of these 2-arylpyrazolo[3,4-c]quinolines have produced some potent and selective A_1 (**19**, **25**, **28**) and A_3 (**4**, **5**, **7**, **8**, **32**) antagonists and only one compound (**16**) showed good A_{2A} affinity.

The A_3 affinity and selectivity of the 2-arylpyrazolo-[3,4-c]quinolin-4-ones **1**–**10** is noteworthy. In fact, compounds **1**–**8** showed low A_1 affinity, were inactive at the A_{2A} , and displayed nanomolar adenosine A_3 receptor affinity. Alkylation at position 5 of the parent compound **1** afforded compounds **9** and **10** which showed slightly increased A_1 and decreased A_3 affinity with respect to **1**.

The influence of the 8-chloro substituent on the benzofused moiety and that of simple substituents at different positions of the 2-phenyl ring was investigated in the 4-ones 1-8, 4-amines 11-18, and 4-N-cycloalkylamino derivatives 19-28. It is well-known^{5,17,29-31} that the AR affinity of ligands of similar size and shape can be enhanced by the presence of chlorine atom on the benzofused moiety in a position corresponding to our 8 position. However, in our pyrazoloquinolines a chlorine atom at position 8 (compounds 2, 12, 20) did not elicit the expected beneficial effect on either AR subtypes. In fact, the A_1 , A_{2A} , and A_3 affinities of **2**, **12**, and **20** were either unaffected or decreased by the presence of the 8-chloro substituents with the exception of the lone 8-chloro-2-phenylpyrazoloquinolin-4-one 2 which showed an A₁ affinity 7-fold higher than that of the 8-unsubstituted compound 1.

The presence of a substituent on the 2-phenyl ring affected AR subtype affinities differently. In general, the presence of substituent on the appended 2-phenyl ring negatively affected A_1 and A_{2A} potency, while the A_3 affinity was dependent on the nature and position of the substituent.

A methyl group in the *ortho*-position (compounds **3**, **13**, **26**) decreased the affinity at all three AR subtypes, with the only exception being the A_{2A} affinity of the 4-N-cyclopentylamino derivative **26** ($K_i = 329.6$ nM) which was 2.5-fold more active than its corresponding 4-N-cyclopentyl-2-phenyl parent compound **25** ($K_i = 849$ nM).

The electron-donating 3-methyl substituent (compounds **4**, **14**, **21**, **27**) increased the A_3 affinity, while it decreased the A_1 and A_{2A} ones, with the only exception being compound **4** which was 2.5-fold more active at the A_1 receptor than its parent compound **1**. The electron-withdrawing fluorine atom at the *meta*-position of the 2-phenyl ring (compounds **6**, **16**, **23**, **28**) had contrasting effects on the affinity at all three AR subtypes. In fact, these 2-(3-fluorophenyl) derivatives showed a decreased affinity at all three AR subtypes with respect to those of the corresponding 2-phenyl-unsubstituted compounds (**1**, **11**, **19**, **25**). There were three exceptions: the A_1 affinity of **6**, the A_{2A} of **16**, and the A_3 of **23** which were on the contrary enhanced. It must be noted that

compound 16 is the only one reported in this paper that showed A_{2A} activity ($K_i = 49.5$ nM). We should also like to point out the A₁ affinity of the 4-N-cyclopentyl-2-(3fluorophenyl) **28** ($K_i = 9.3$ nM) which, although less active at this subtype than its corresponding parent compound **25** ($K_i = 3.2$ nM), is a highly A_1/A_3 -selective antagonist.

The electron-donating 4-methyl group (compounds 5, 15, 22) and 4-methoxy group (compounds 7, 17) and the electron-withdrawing 4-chloro substituent (compounds 8, 18, 24) on the appended 2-phenyl moiety enhanced the A_3 affinity in particular in the 4-one series (see 5. 7, 8 vs 1). On the contrary, all the 2-(4-phenylsubstituted) derivatives were inactive at the A_{2A} subtype. A similar negative effect of the p-phenyl substituent can be observed in the A_1 affinity, with the only exception being the *para*-substituted 4-one derivatives 7 and 8 which showed a 2- and 4.5-fold enhancement, respectively, in A₁ affinity with respect to that of the parent compound 1.

We would like to highlight the contrasting effect of the substituent on the 2-phenyl ring toward A₁ affinity in the 4-one series 1-8 and 4-amino series 11-28: in compounds 4 and 6-8 the effect was positive while in amines 11–28 the effect was always negative. Instead, the A_3 affinity in both the 4-one (1-8) and 4-amino (11-28) series was enhanced by the presence of a parasubstituent, of whatever nature, or by the electrondonating 3-methyl substituent on the appended 2-phenyl

Replacement of the 4-oxo function with the 4-amino group yielded nonselective AR ligands. In fact, the pyrazologuinolin-4-amines 11–18, as a whole, displayed higher A₁ and A_{2A} receptor affinities than the corresponding 4-one derivatives 1-8. These latter, on the contrary, were more active than 11-18 on the A₃ subtype. These SAR are in accordance with those of the previously reported series A and B^{20} and confirm the importance of the 4-amino proton donor group in A₁ and A_{2A}^{17,20} and the 4-carbonyl group in A₃²⁰ receptor—ligand interactions.

Finally, a hydrogen atom of the 4-amino group of the parent 2-phenyl-2*H*-pyrazolo[3,4-*c*]quinolin-4-amine **11** was replaced by an aralkyl (compounds 29, 30), an acyl (compounds 31-33), and a carbamoyl residue (compounds 34, 35). All these 4-N-substituted compounds **29–35** displayed low or null A_{2A} affinity. The aralkyl derivatives **29** and **30** displayed nanomolar A₁ and A₃ affinities and, as a consequence, were not A₁/A₃-selective antagonists. Among the 4-amido derivatives **31–33**, the 4-acetamido **31** was an A₁/A₃ nonselective ligand while the 4-benzoylamido 32 was the most potent and selective A_3 antagonist among those tested in this study. Homologation of 32 gave the 4-phenylacetamido 33, which although still A_3 potent ($K_i = 9.9$ nM) was also active at the A_1 subtype ($K_i = 107.2$ nM) and as a consequence was less A_3/A_1 -selective than **32**. The importance of the C=O amide group at position 4 in A₃ receptor-ligand interaction is shown by comparing the A_3 affinity of the 4-N-benzovlamido **32** ($K_i = 2.1 \text{ nM}$) vs the 4-N-benzylamino **29** ($K_i = 35.8$ nM) and the 4-Nphenylacetamido **33** ($K_i = 9.9 \text{ nM}$) vs the 4-N-phenethylamino **30** ($K_i = 32.9$ nM). By replacing one hydrogen atom of the 4-amino group of 11 with a carbamoyl residue the ureido derivatives **34** and **35** were prepared. Compounds **34** and **35** were both less active at the A_1 and A_{2A} receptors while they were more active at the A_3 subtype than **11**, further confirming the importance of the C=O group at position 4 for A₃ affinity.

In conclusion, the SAR of compounds 1-35 were in accordance with those of the previously reported series **A** and \mathbf{B}^{20} and confirmed some different structural requirements of each AR subtype recognition site. Similarly as in the **A** and **B** series compounds **1–35** were little active at the A_{2A} receptor and the substituent on the 2-phenyl ring affected differently the A₁ and A₃ affinities. The comparison of A_1 and A_{2A} affinities of the 4-amino derivatives 11–18 with those of the 4-ones 1–8 confirmed that in these tricyclic ligands the 4-amino group is essential for A₁ and A_{2A} receptor-ligand interaction.²⁰ The nanomolar A₃ affinity of compounds 1-8, 31-33, and 35 stressed the importance for A_3 receptor recognition of the presence at position 4 of either a nuclear (as in 4-ones 1-8) or extranuclear (as in amides **32** and **33** or ureide **35**) carbonyl group.²⁰ Finally, the similarity of the SAR of 1–35, A, and B suggests a similar AR binding mode. Since we have hypothesized that in compounds **B** the N-4 region corresponds to that of the N-6 of adenosine,²⁰ we may hypothesize that in compounds 11-35 as well the N-4 region corresponds to the adenosine N-6 one. It follows that the A_3 nanomolar affinity of 1-8, 31-33, and 35could be due to the presence in the N-6 region of the adenosine A₃ subtype of a proton donor able to bind to the C=O proton acceptor, which should explain the A₃ activity of 1-8, 31-33, and 35 and of the other A_3 antagonists reported in the literature. 5-7,32

Experimental Section

(A) Chemistry. Silica gel plates (Merck F₂₅₄) and silica gel 60 (Merck, 70-230 mesh) were used for analytical and column chromatography, respectively. All melting points were determined on a Gallenkamp melting point apparatus. Microanalyses were performed with a Perkin-Elmer 260 elemental analyzer for C, H, N, and the results were within $\pm 0.4\%$ of the theoretical values. The IR spectra were recorded with a Perkin-Elmer 1420 spectrometer in Nujol mulls and are expressed in cm^{-1} . The 1H NMR spectra were obtained with a Varian Gemini 200 instrument at 200 MHz. The chemical shifts are reported in δ (ppm) and are relative to the central peak of the solvent that was always DMSO-d₆. The following abbreviations are used: s = singlet, d = doublet, dd = doubledoublet, t = triplet, m = multiplet, br = broad, and ar = aromatic protons. Physical data of the newly synthesized compounds are listed in Table 2.

General Procedure for the Synthesis of 4,5-Dihydro-2-(2-methylphenyl)-2*H*-pyrazolo[3,4-*c*]quinolin-4-one (3) and 2-(4-Chlorophenyl)-4,5-dihydro-2H-pyrazolo[3,4-c]**quinolin-4-one (8).** The title compounds were obtained from 3-ethoxalylindole²² (1.0 g, 4.6 mmol) and the suitable arylhydrazine hydrochloride (10.1 mmol) as described in ref 21 to prepare 1, 2, and 4-7. The title compounds displayed the following spectral data.

- 3: ¹H NMR 2.27 (s, 3H, CH₃), 7.23-7.57 (m, 7H, ar), 7.98 (d, 1H, ar, J = 7.6 Hz), 9.04 (s, 1H, H-1), 11.48 (s, 1H, NH); IR 3140, 1670.
- **8**: ¹H NMR 7.24–7.40 (m, 3H, ar), 7.72 (d, 2H, ar, J = 8.9Hz), 7.95 (d, 1H, ar, J = 7.7 Hz), 8.07 (d, 2H, ar, J = 8.9 Hz), 9.51 (s, 1H, H-1), 11.51 (s, 1H, NH); IR 3160, 1680.

4,5-Dihydro-2-phenyl-5-*N-n*-propyl-2*H*-pyrazolo[3,4-*c*]**quinolin-4-one (10).** The title compound was obtained from **1** (0.250 g, 0.96 mmol) and *n*-propyl bromide (0.124 mL, 1.44 mmol) as described in ref 21 to prepare 9. The title compound

Table 2. Physical Data of Newly Synthesized Compounds

	R	\mathbf{R}_1	R_2	mp, °C	solv ^a	% yield
3	2-Me	Н	Н	> 300	Α	60
8	4-Cl	Н	Н	> 300	В	43
10	Н	Н	n-C ₃ H ₇	140-143	С	62
11	Н	Н	Н	197-198	D	56 ^b , 90 ^c
12	Н	C1	Н	235-237	Е	80
13	2-Me	Н	H	186-188	F	50
14	3-Me	Н	Н	218-220	E	60
15	4-Me	Н	Н	235-237	D	35
16	3-F	Н	Н	217-220	F	45
17	4-OMe	Н	Н	193-194	E	40
18	4-Cl	Н	Н	243-246	F	64
19	Н	Н	47	138-140	G	58
20	Н	Cl	~~~	165-168	Е	57
21	3-Me	Н	~~	146-148	Е	35
22	4-Me	Н	~>	147-149	F	50
23	3-F	Н	\Rightarrow	152-155	E	88
24	4-C1	Н	~~~	180-183	F	83
25	Н	Н	\bigcirc	104-106	Н	35
26	2-Me	Н	٥	100-102	I	40
27	3-Me	Н	\bigcirc	96-98	I	45
28	3-F	Н	Ď	143-145	G	77
29	Н	Н	CH₂Ph	128-130	Е	45
30	Н	Н	$(CH_2)_2Ph$	125-127	G	60
31	Н	Н	СОМе	231-233	Е	98
32	Н	Н	COPh	223-225	J	98
33	Н	Н	COCH ₂ Ph	240-242	K	62
34	Н	Н	CONHPh	255-257	В	47
35	Н	Н	CONHCH₂Ph	242-243	В	70

 a Recrystallization solvents: A = 1-ethoxyethanol; B = glacial acetic acid; C = cyclohexane/ethyl acetate; D = ethyl acetate; E = ethanol; F = methanol; G = cyclohexane; H = n-hexane; I = cyclohexane/petroleum ether; J = acetonitrile; K = 2-butanone. b From pyridinium chloride **36**. c From 4-chloro derivative **40**.

displayed the following 1 H NMR data: 0.99 (t, 3H, CH₃, J = 7.2 Hz), 1.65–1.80 (m, 2H, CH₂), 4.30 (t, 2H, N–CH₂, J = 7.5 Hz), 7.29–7.68 (m, 6H, ar), 8.03–8.07 (m, 3H, ar), 9.51 (s, 1H, H-1).

General Procedure for the Synthesis of 1-(2-Aryl-2*H*-pyrazolo[3,4-*c*]quinolin-4-yl)pyridinium Chlorides 36–39. A mixture of 1, 5–7 (1.38 mmol) and phosphorus pentachloride (0.085 g, 0.41 mmol) in phosphorus oxychloride (8 mL) and anhydrous pyridine (0.3 mL, 3.71 mmol) was heated at reflux for 30 min. The cooled mixture was quenched with ce/water (about 50 mL) to yield a solid which was collected and washed with water. These pyridinium chlorides were very unstable; nevertheless they were pure enough to be characterized and used without further purification. The ¹H NMR data of the title compounds are as follows.

36: 7.56–7.77 (m, 3H, ar), 7.82–7.98 (m, 2H, pyridinium H-3 and H-5), 8.24–8.30 (m, 3H, ar), 8.50–8.57 (m, 3H, ar), 9.02 (t, 1H, pyridinium H-4, J = 7.7 Hz), 10.19–10.23 (m, 3H, H-1 + pyridinium H-2 and H-6).

37: 2.44 (s, 3H, CH₃), 7.53 (d, 2H, ar, J = 8.6 Hz), 7.82–7.98 (m, 2H, pyridinium H-3 and H-5), 8.16 (d, 2H, ar, J = 8.6 Hz), 8.27 (d, 1H, ar, J = 8.2 Hz), 8.49–8.56 (m, 3H, ar), 9.02 (t, 1H, pyridinium H-4, J = 7.6 Hz), 10.12 (s, 1H, H-1), 10.21 (d, 2H, pyridinium H-2 and H-6, J = 6.8 Hz).

38: 7.40–7.96 (m, 4H, 2 ar + pyridinium H-3 and H-5), 8.15–8.29 (m, 3H, ar), 8.45–8.57 (m, 3H, ar), 9.02 (t, 1H,

pyridinium H-4, J = 7.6 Hz), 10.21-10.30 (m, 3H, H-1 + pyridinium H-2 and H-6).

39: 3.90 (s, 3H, OCH₃), 7.26 (d, 2H, ar, J = 8.9 Hz), 7.82–7.94 (m, 2H, pyridinium H-3 and H-5), 8.17–8.29 (m, 3H, ar), 8.49–8.56 (m, 3H, ar), 9.01 (t, 1H, pyridinium H-4, J = 7.9 Hz), 10.09 (s, 1H, H-1), 10.21 (d, 2H, pyridinium H-2 and H-6, J = 6.7 Hz).

General Procedure for the Synthesis of 2-Aryl-4-chloro-2*H*-pyrazolo[3,4-*c*]quinolines 40–46. A mixture of 1–6, 8 (5.35 mmol) and phosphorus pentachloride (0.335 g, 1.61 mmol) in phosphorus oxychloride (50 mL) was heated at reflux for 2 h. Evaporation at reduced pressure of the excess of phosphorus oxychloride yielded a residue which was treated with cold water (50 mL) and quickly collected. These 4-chloro derivatives were very unstable; nevertheless they were pure enough to be characterized and used without further purification. The ¹H NMR data of the title compounds are as follows.

40: 7.54-7.80 (m, 5H, ar), 8.01 (d, 1H, ar, J=7.0 Hz), 8.14 (d, 2H, ar, J=7.3 Hz), 8.33 (d, 1H, ar, J=7.7 Hz), 9.86 (s, 1H, H-1).

41: 7.56–7.75 (m, 4H, ar), 8.00–8.12 (m, 3H, ar), 8.46 (d, 1H, H-9, J = 2.4 Hz), 9.89 (s, 1H, H-1).

42: 2.23 (s, 3H, CH₃), 7.47–7.70 (m, 6H, ar), 7.97–8.02 (m, 1H, ar), 8.30–8.34 (m, 1H, ar), 9.36 (s, 1H, H-1).

43: 2.49 (s, 3H, CH₃), 7.38 (d, 1H, ar, J = 8.1 Hz), 7.55 (t, 1H, ar, J = 7.9 Hz), 7.60–7.77 (m, 3H, ar), 8.33 (d, 1H, ar, J = 8.9 Hz), 9.84 (s, 1H, H-1).

44: 2.42 (s, 3H, CH₃), 7.48 (d, 2H, ar, J = 8.0 Hz), 7.68–7.74 (m, 2H, ar), 8.01–8.04 (m, 3H, ar), 8.32 (d, 1H, ar, J = 7.4 Hz), 9.81 (s, 1H, H-1).

45: 7.37-7.45 (m, 1H, ar), 7.62-7.80 (m, 3H, ar), 7.98-8.05 (m, 3H, ar), 8.29 (dd, 1H, ar, J=7.6, 2.3 Hz), 9.89 (s, 1H, H-1)

46: 7.70–7.79 (m, 4H, ar), 8.02 (d, 1H, ar, J= 7.7 Hz), 8.19 (d, 2H, ar, J= 8.9 Hz), 8.31 (d, 1H, ar, J= 8.3 Hz), 9.89 (s, 1H, H-1).

General Procedure for the Synthesis of 2-Aryl-2H-pyrazolo[3,4-c]quinolin-4-amines 11–18. Method A (11, 15–17). A mixture of 36–39 (0.75 mmol) in cyclohexylamine (0.85 mL) was heated at reflux for 30 min. The resulting oil was treated with water (200 mL) and extracted with diethyl ether (200 mL). The organic phase was washed with water (2 \times 100 mL) and dried (Na₂SO₄). Evaporation at reduced pressure of the solvent yielded a residue which was treated with diethyl ether and collected. Compounds 11 and 16 were directly recrystallized, while compounds 15 and 17, before recrystallization, were further purified by column chromatography, eluting system chloroform/methanol (9:1).

Method B (11–14, 18). A mixture of **40–43, 46** (2 mmol) in absolute ethanol (20 mL) saturated with ammonia was heated overnight at 120 °C in a sealed tube. Upon cooling, the mixture yielded solid crude **11–14** and **18**, which were collected, washed with water and recrystallized. The spectral data of the title compounds are as follows.

11: ¹H NMR 6.93 (br s, 2H, NH₂), 7.19–7.61 (m, 6H, ar), 8.01 (d, 1H, ar, J= 7.8 Hz), 8.12 (d, 2H, ar, J= 8.0 Hz), 9.50 (s, 1H, H-1); IR 3500, 3300, 3100, 1690.

12: ¹H NMR 7.08 (br s, 2H, NH₂), 7.41–7.70 (m, 5H, ar), 8.06–8.10 (m, 3H, ar), 9.57 (s, 1H, H-1); IR 3490, 3310, 1660.

13: ¹H NMR 2.26 (s, 3H, CH₃), 6.92 (br s, 2H, NH₂), 7.25 (t, 1H, ar, J = 7.8 Hz), 7.35–7.59 (m, 6H, ar), 8.00 (d, 1H, ar, J = 7.9 Hz), 9.03 (s, 1H, H-1); IR 3360, 3220, 3180, 1650.

14: ¹H NMR 2.46 (s, 3H, CH₃), 6.95 (br s, 2H, NH₂), 7.20–7.56 (m, 5H, ar), 7.88–8.02 (m, 3H, ar), 9.48 (s, 1H, H-1); IR 3490, 3310, 1650.

15: ¹H NMR 2.39 (s, 3H, CH₃), 6.93 (br s, 2H, NH₂), 7.22–7.53 (m, 5H, ar), 7.97–8.01 (m, 3H, ar), 9.44 (s, 1H, H-1); IR 3470, 3300, 1650.

16: ¹H NMR 6.99 (br s, 2H, NH₂), 7.24–7.72 (m, 5H, ar), 7.95–8.02 (m, 3H, ar), 9.56 (s, 1H, H-1); IR 3460, 3290, 1640.

17: ¹H NMR 3.85 (s, 3H, OCH₃), 6.93 (br s, 2H, NH₂), 7.15–7.25 (m, 3H, ar), 7.38–7.52 (m, 2H, ar), 7.98–8.02 (m, 3H, ar), 9.37 (s, 1H, H-1); IR 3450, 3300, 1640.

18: ¹H NMR 6.97 (br s, 2H, NH₂), 7.24–7.50 (m, 3H, ar), 7.78 (d, 2H, ar, J = 9.0 Hz), 8.00 (d, 1H, ar, J = 7.4 Hz), 8.16 (d, 2H, ar, J = 8.9 Hz), 9.54 (s, 1H, H-1); IR 3470, 3310, 1650.

General Procedure for the Synthesis of 4-N-Aminosubstituted-2-aryl-2*H*-pyrazolo[3,4-*c*]quinolines 19–30. Method A (19, 20, 23, 25, 28-30). A mixture of 40-41, 45 (2 mmol) and the suitable cycloalkyl- or aralkylamine (2 mL) was heated overnight at 120 °C in a sealed tube. The cooled mixture was treated with diethyl ether (20 mL) affording a solid which was eliminated. Evaporation at reduced pressure of the clear solution gave a solid which was collected and treated with cyclohexane to yield the crude title compounds. All crude compounds but one (25) were directly recrystallized from suitable solvent. Compound 25, before recrystallization, was purified by column chromatography, eluting system chloroform/ ethyl acetate (9:1). Evaporation of the central eluates gave an oil which became solid by treatment with petroleum ether.

Method B (21, 22, 24, 26, 27). A mixture of 42-44, 46 (2 mmol), the suitable cycloalkylamine (2.4 mmol) and triethylamine (0.55 mL, 4 mmol) in absolute ethanol (20 mL) was heated overnight at 120 °C in a sealed tube. Evaporation of the solvent at reduced pressure yielded a solid which was treated with petroleum ether, collected and washed with water. Crude compounds 22 and 24 were directly recrystallized, while crude compounds 21 and 26-27, before recrystallization, were purified by column chromatography, eluting system chloroform/ethyl acetate (9:1). Evaporation of the central eluates gave an oil which became solid by treatment with petroleum ether. The spectral data of the title compounds are as follows.

19: ¹H NMR 1.19–2.06 (m, 10H, cyclohexyl protons), 4.23– 4.38 (m, 1H, cyclohexyl proton), 6.98 (d, 1H, NH, J = 8.7 Hz), 7.17-7.69 (m, 6H, ar), 8.00 (d, 1H, ar, J = 7.7 Hz), 8.13 (d, 2H, ar, J = 7.2 Hz), 9.48 (s, 1H, H-1); IR 3430.

20: ¹H NMR 1.07–2.09 (m, 10H, cyclohexyl protons), 4.15– 4.35 (m, 1H cyclohexyl proton), 7.18 (d, 1H, NH, J = 8.3 Hz), 7.37 (dd, 1H, ar, J = 8.7, 2.4 Hz), 7.46–7.58 (m, 2H, ar), 7.66 (t, 2H, ar, J = 7.3 Hz), 8.07 - 8.17 (m, 3H, ar), 9.60 (s, 1H, H-1);

21: ¹H NMR 1.10-1.85 (m, 8H, cyclohexyl protons), 2.00-2.08 (m, 2H, cyclohexyl protons), 2.45 (s, 3H, CH₃), 4.18-4.35 (m, 1H, cyclohexyl proton), 6.97 (d, 1H, NH, J = 8.1 Hz), 7.16– 7.56 (m, 5H, ar), 7.88-7.99 (m, 3H, ar), 9.45 (s, 1H, H-1); IR 3440

22: ¹H NMR 1.10–1.83 (m, 8H, cyclohexyl protons), 2.00– 2.10 (m, 2H, cyclohexyl protons), 2.42 (s, 3H, CH₃), 4.18-4.48 (m, 1H, cyclohexyl proton), 6.94 (d, 1H, NH, J = 7.9 Hz), 7.21 (t, 1H, ar, J = 7.4 Hz), 7.33-7.56 (m, 4H, ar), 7.96-8.04 (m, 3H, ar), 9.45 (s, 1H, H-1); IR 3440.

23: ¹H NMR 1.10–2.02 (m, 10H, cyclohexyl protons), 4.22– 4.34 (m, 1H, cyclohexyl proton), 7.06 (d, 1H, NH, J = 8.6 Hz), 7.17-7.45 (m, 3H, ar), 7.52-7.75 (m, 2H, ar), 7.92-8.09 (m, 3H, ar), 9.55 (s, 1H, H-1); IR 3440.

24: ¹H NMR 1.10-1.81 (m, 8H, cyclohexyl protons), 2.00-2.01 (m, 2H, cyclohexyl protons), 4.20-4.29 (m, 1H, cyclohexyl proton), 7.01 (d, 1H, NĤ, J = 8.9 Hz), 7.17-7.55 (m, 3H, ar), 7.72 (d, 2H, ar, J = 8.9 Hz), 7.95 (d, 1H, ar, J = 7.0 Hz), 8.16 (d, 2H, ar, J = 8.9 Hz), 9.50 (s, 1H, H-1); IR 3440.

25: ¹H NMR 1.60–1.74 (m, 6H, cyclopentyl protons), 2.02– 2.10 (m, 2H, cyclopentyl protons), 4.68-4.71 (m, 1H, cyclopentyl protons) pentyl proton), 7.16-7.25 (m, 2H, 1 ar + NH), 7.35-7.68 (m, 5H, ar), 7.99 (d, 1H, ar, J = 7.7 Hz), 8.13 (d, 2H, ar, J = 7.5Hz), 9.49 (s, 1H, H-1); IR 3430.

26: ¹H NMR 1.50–1.88 (m, 6H, cyclopentyl protons), 1.92– 2.16 (m, 2H, cyclopentyl protons), 2.26 (s, 3H, CH₃), 4.60–4.79 (m, 1H, cyclopentyl proton), 7.16-7.59 (m, 8H, 7 ar + NH), 7.98 (d, 1H, ar, J = 7.6 Hz), 9.02 (s, 1H, H-1); IR 3360.

27: ¹H NMR 1.56-1.78 (m, 6H, cyclopentyl protons), 2.00-2.09 (m, 2H, cyclopentyl protons), 2.51 (s, 3H, $\dot{\rm CH_3}$), 4.58–4.76 (m, 1H, cyclopentyl proton), 7.13–7.57 (m, 6H, 5 ar + NH), 7.85-7.98 (m, 3H, ar), 9.45 (s, 1H, H-1); IR 3420.

28: ¹H NMR 1.55–1.79 (m, 6H, cyclopentyl protons), 2.05– 2.09 (m, 2H, cyclopentyl protons), 4.50-4.70 (m, 1H, cyclopentyl proton), 7.18-7.43 (m, 4H, 3 ar + NH), 7.53-7.74 (m, 2H, ar), 7.93-8.09 (m, 3H, ar), 9.54 (s, 1H, H-1); IR 3440.

29: 4.84 (d, 2H, CH₂, J = 6.2 Hz), 7.22–7.67 (m, 11H, ar), 7.98-8.02 (m, 4H, 3 ar + NH), 9.50 (s, 1H, H-1); IR 3430.

30: ¹H NMR 3.07 (t, 2H, CH₂, J = 7.7 Hz), 3.80–3.87 (m, 2H, CH₂), 7.21-7.69 (m, 12H, 11 ar + NH), 8.01 (d, 1H, ar, J = 7.6 Hz), 8.11 (d, 2H, ar, J = 7.6 Hz), 9.50 (s, 1H, H-1); IR

General Procedure for the Synthesis of 4-N-Acetamido-2-phenyl-2*H*-pyrazolo[3,4-*c*]quinoline (31) and 4-*N*-Benzamido-2-phenyl-2*H*-pyrazolo[3,4-*c*]quinoline (32). A solution of acetyl chloride or benzoyl chloride (4 mmol) in anhydrous dichloromethane (3 mL) was slowly added at 0 °C to a suspension of 11 (0.520 g, 2 mmol) in anhydrous dichloromethane (10 mL) and anhydrous pyridine (1.61 mL, 20 mmol). The mixture was stirred at room temperature for 24 h. Evaporation at reduced pressure of the solvent yielded a residue which was treated with ethanol/water (10 mL), collected and recrystallized. The title compounds displayed the following spectral data.

31: ¹H NMR 2.39 (s, 3H, CH₃), 7.51–7.70 (m, 5H, ar), 7.84– 7.89 (m, 1H, ar), 8.13-8.24 (m, 3H, ar), 9.70 (s, 1H, H-1), 10.31 (br s, 1H, NH); IR 3400, 3140, 1680.

32: ¹H NMR 7.42-7.90 (m, 12H, 11 ar + NH), 8.03 (d, 2H, ar, J = 7.5 Hz), 8.29 (d, 1H, ar, J = 7.5 Hz), 9.80 (s, 1H, H-1); IR 3160, 1695.

4-N-Phenylacetamido-2-phenyl-2H-pyrazolo[3,4-c]quin**oline (33).** A mixture of **11** (0.260 g, 1 mmol), phenylacetic acid (0.816 g, 6 mmol), 1-(3-(dimethylamino)propyl)-3-ethylcarbodiimide hydrochloride (1.150 g, 6 mmol), 1-hydroxybenzotriazole (0.810 g, 6 mmol), triethylamine (2.08 mL, 15 mmol) and 4-(dimethylamino)pyridine (0.012 g, 0.1 mmol) in anhydrous DMF (5 mL) was stirred at room temperature for 2 h. The resulting solid was collected, washed with water (10 mL) and recrystallized. The title compound displayed the following spectral data: ¹H NMR 4.06 (s, 2H, CH₂), 7.27-7.71 (m, 10H, ar), 7.85-7.90 (m, 1H, ar), 8.12-8.23 (m, 3H, ar), 9.71 (s, 1H, H-1), 10.57 (br s, 1H, NH); IR 3380, 3140, 1680.

4-Phenylureido-2-phenyl-2*H*-pyrazolo[3,4-*c*]quinoline (34) and 4-Benzylureido-2-phenyl-2H-pyrazolo[3,4**cquinoline (35).** The suitable isocyanate (1.7 mmol) was added to a suspension of 11 (0.300 g, 1.15 mmol) in anhydrous THF (20 mL). The mixture was refluxed for 2 h under nitrogen atmosphere. The resulting solid was collected and recrystallized. The title compounds displayed the following ¹H NMR

34: 7.12 (t, 1H, ar, J = 8.4 Hz), 7.36–7.98 (m, 9H, ar), 8.00 (d, 1H, ar, J = 6.2 Hz), 8.17 - 8.24 (m, 3H, ar), 9.68 - 9.73 (m, 2H, H-1 + NH), 12.73 (s, 1H, NH).

35: 4.60 (d, 2H, CH₂, J = 5.8 Hz), 7.34–7.78 (m, 11H, ar), 8.13-8.22 (m, 3H, ar), 9.22 (br s, 1H, NH), 9.68 (s, 1H, H-1), 10.48 (t, 1H, NH, J = 5.8 Hz).

(B) Biochemistry. A_1 and A_{2A} Receptor binding. Displacement of [3H]CHA from A₁ AR in bovine cortical membranes and [3H]CGS 21680 from A_{2A} AR in bovine striatal membranes was performed as described. $^{\rm 33}$

A₃ Receptor binding. The displacement of [125I]AB-MECA in membranes prepared from CHO cells stably expressing the human A₃ receptor was performed as described. ³⁴ The assay medium consisted of a buffer containing 50 mM Tris-HCl, 10 mM MgCl₂, and 1 mM EDTA at pH 8.12. The glass incubation tubes, containing 20 μ L of the membrane suspension (0.2 mg of protein/mL, stored at -80 °C in the same buffer), 20 μ L of [125 I]AB-MECA (final concentration 0.2 nM), and 10 μ L of the tested ligand, were incubated for 60 min at 25 °C in a total volume of 100 μ L. After incubation, the samples were filtered on Whatman $\overline{\text{GF/C}}$ filters presoaked for $1\ h$ in 0.5% poly-(ethylenimine) followed by three washes with 5 mL of ice-cold incubation buffer. Nonspecific binding was determined in the presence of 200 μ M NECA. Specific binding was obtained by subtracting nonspecific binding from total binding

Compounds were dissolved in DMSO (buffer/concentration of 2%) and added to the assay mixture. Blank experiments were carried out to determine the effect of solvent on binding.

Protein estimation was based on a reported method, 35 after solubilization with 0.75 N sodium hydroxide, using bovine serum albumine as standard.

The concentration of the tested compound that produced 50% inhibition of specific [3 H]CHA, [3 H]CGS 21680 or [125 I]-AB-MECA binding (IC $_5$ 0) was calculated using a nonlinear regression method implemented in the InPlot program (Graph-Pad, San Diego, CA) with five concentrations of displacer, each performed in triplicate. Inhibition constants (K_i) were calculated according to the Cheng–Prusoff equation. The dissociation constants (K_d) of [3 H]CHA, [3 H]CGS 21680, and [1 25I]AB-MECA are 1.2, 14, and 1.4 nM, respectively.

Acknowledgment. We thank Dr. Karl-Norbert Klotz of the University of Wüzburg, Germany, for providing cloned human A₃ receptors expressed in CHO cells.

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