Novel Synthetic Oleanane and Ursane Triterpenoids with Various Enone Functionalities in Ring A as Inhibitors of Nitric Oxide Production in Mouse **Macrophages**[†]

Tadashi Honda,[‡] Gordon W. Gribble,*,[‡] Nanjoo Suh,[§] Heather J. Finlay,^{‡,||} BarbieAnn V. Rounds,[‡] Lothar Bore,[‡] Frank G. Favaloro, Jr., *Yongping Wang, *§ and Michael B. Sporn*, *§

Department of Chemistry, Dartmouth College, and Department of Pharmacology and Toxicology, Dartmouth Medical School, Hanover, New Hampshire 03755

Received January 7, 2000

We initially randomly synthesized about 60 oleanane and ursane triterpenoids as potential anti-inflammatory and cancer chemopreventive agents. Preliminary screening of these derivatives for inhibition of production of nitric oxide induced by interferon- γ in mouse macrophages revealed that 3-oxooleana-1,12-dien-28-oic acid (**B-15**) showed significant activity ($IC_{50} = 5.6$ μ M). On the basis of the structure of **B-15**, 19 novel olean- and urs-12-ene triterpenoids with a 1-en-3-one functionality having a substituent at C-2 in ring A have been designed and synthesized. Among them, 3-oxooleana-1,12-diene derivatives with carboxyl, methoxycarbonyl, and nitrile groups at C-2 showed higher activity than the lead compound **B-15**. In particular, 2-carboxy-3-oxooleana-1,12-dien-28-oic acid (3) had the highest activity (IC₅₀ = 0.07 μ M) in this group of triterpenoids. The potency of 3 was similar to that of hydrocortisone ($IC_{50} = 0.01$ μ M), although 3 does not act through the glucocorticoid receptor. Interesting structure—activity relationships of these novel synthetic triterpenoids are also discussed.

Introduction

Oleanane and ursane triterpenoids are pentacyclic compounds with 30 carbon atoms, which are derived biosynthetically by the cyclization of squalene. The group includes a very large number of naturally occurring members that cover an impressive variety of functional groups.² Many compounds of this group are reported to have interesting biological, pharmacological, or medicinal activities similar to those of retinoids and steroids, such as anti-inflammatory activity, suppression of tumor promotion, suppression of immunoglobulin synthesis, protection of the liver against toxic injury, induction of collagen synthesis, and induction of differentiation in leukemia or teratocarcinoma cells.³ However, the potency of these triterpenoids is relatively weak. There are no systematic studies of structureactivity relationships based on chemical modification of oleanane and ursane triterpenoids.4 We have therefore considered that bioassay-directed systematic drug design and synthesis of derivatives of oleanolic acid (1) and ursolic acid (2), which are commercially available, could be of great value in discovering novel structures with high biological potency.

The high output of nitric oxide (NO) produced by inducible nitric oxide synthase (iNOS), which is expressed in activated macrophages, plays an important role in host defense. However, excessive production of NO also can destroy functional normal tissues during acute and chronic inflammation.⁵ This phenomenon is also closely related mechanistically to carcinogenesis.⁶ Thus, inhibitors of NO production in macrophages are potential anti-inflammatory and cancer chemopreventive drugs. Because oleanolic and ursolic acids are already known to have weak anti-inflammatory and anticarcinogenic activity, 3a,3b,3e,3f we focused our attention on therapeutic agents of these diseases. For this purpose, we have adopted an assay system that measures inhibition of NO production induced by interferon-γ (IFN- γ) in mouse macrophages⁷ as a preliminary screening assay system. We synthesized various oleanolic and ursolic acid derivatives and tested them as inhibitors of NO production. As a result, we have identified a series of novel olean-12-ene triterpenoids with a 1-en-3-one functionality having carboxyl, methoxycarbonyl, and nitrile groups at C-2 in ring A that show significant inhibitory activity (IC₅₀ = $0.01-0.1 \mu M$ level) against production of NO induced by IFN-γ in mouse macrophages. In particular, 2-carboxy-3-oxooleana-1,12-dien-28-oic acid (3) had the highest activity (IC₅₀ = 0.07 μ M) in this group of compounds. The potency of 3 was similar to that of hydrocortisone (IC₅₀ = 0.01 μ M), although **3** does not act through the glucocorticoid receptor. We report here the synthesis, inhibitory activity, and structure-activity relationships of these novel triterpenoids in detail.

Chemistry

Discovery of Lead Compound. When we started this project, we had no information about a lead

 $^{^\}dagger$ Part of this work has been reported in preliminary form: (a) Honda, T.; Finlay, H. J.; Gribble, G. W.; Suh, N.; Sporn, M. B. New enone derivatives of oleanolic acid and ursolic acid as inhibitors of nitric oxide production in mouse macrophages. *Bioorg. Med. Chem. Lett.* **1997**, 7, 1623–1628. (b) Honda, T.; Rounds, B. V.; Bore, L.; Favaloro, F. G., Jr.; Gribble, G. W.; Suh, N.; Wang, Y.; Sporn, M. B. Novel synthetic oleanane triterpenoids: a series of highly active inhibitors of nitric oxide production in mouse macrophages. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 3429–3434.

^{*} Address correspondence to either author. For G.W.G.: phone, 603-646-3118; fax, 603-646-3946; e-mail, Grib@dartmouth.edu. For M.B.S.: phone, 603-650-6557; fax, 603-650-1129; e-mail, Michael.Sporn@dartmouth.edu.

[‡] Department of Chemistry.

§ Department of Pharmacology and Toxicology

"Present address: The Bristol-Myers Squibb Pharmaceutical Research Institute, P.O. Box 5400, Princeton, NJ 08543-5400.

Table 1. Preliminary Screening Results of Synthetic Oleanane and Ursane Triterpenoids

ompd	skeleton	C-3	C-12	C-13	C-17	inhibition (%) at $10 \mu \mathrm{M}^b$	ref
1	olean-12-ene	β-ОН	Н		CO ₂ H	38	10
2	urs-12-ene	β -OH	Н		CO_2H	0	15
A-1	olean-12-ene	β -OH	Н		CO_2Me	0	10
A-2	urs-12-ene	β -OH	Н		CO_2Me	0	15
4 -3	olean-12-ene	β -OAc	Н		CO_2Me	10	10
\-4	urs-12-ene	β -OAc	Н		CO_2Me	15	15
A -5	olean-12-ene	β -OAc	Н		CO_2H	0	10
A-6	urs-12-ene	β -OAc	Н		CO_2H	0	15
A -7	olean-12-ene	β -OH	Н		CH_2OH	0	28
A-8	urs-12-ene	β -OH	Н		CH_2OH	8	29
A -9	olean-12-ene	β -OAc	Н		CH_2OAc	4	28
A-10	urs-12-ene	β -OAc	Н		CH ₂ OAc	0	29
A-11	oleanane	β -OAc	α-OH	β -H	CO_2Me	0	c
A-12	oleanane	β -OAc	β -OH	β -H	CO_2Me	0	c
A-13	oleanane	β -OAc	β -OAc	β-H	CH ₂ OAc	0	c
A-14	oleanane	β -OAc	α-OH	β-H	CH ₂ OAc	0	c
A-15	oleanane	΄β- Ο Η	α-ОН	΄β- H	CH ₂ OH	48	c
A-16	oleanane	β -OH	β -OH	β -H	CH ₂ OH	20	c
A-17	oleanane	β -OH	=0	β -H	CO ₂ Me	0	10
A-18	oleanane	β -OAc	=0	β -H	CO ₂ Me	0	10
A-19	olean-12-ene	α-ΟΗ	Н	P	CO ₂ H	18	30
A-20	urs-12-ene	α-OH	H		CO ₂ H	48	31
4-21 ^a	oleanane	β -OH	α-ОН	-0-	-CH(OH)-	21	32
A -22 ^a	oleanane	β-OH	α-OH	-0-	-CO-	13	32
A-23	oleanane	β-OAc		α-epoxy-	CO ₂ Me	0	10
1-23 1-24	oleanane	<i>β</i> -OHC <i>β</i> -OH	α-ΟΗ	β -OH	CH ₂ OH	22	32
B-1	olean-12-ene	=0	Н	ρ -OH	CO ₂ H	16	10
3-1 3-2	urs-12-ene	=0	H		CO ₂ H	22	33
B-3	olean-12-ene	=0	H		CO ₂ Me	24	10
3-3 3-4	urs-12-ene	=0	п Н		CO ₂ Me	16	15
B-4 B-5	olean-12-ene	=0	H		CHO	11	34
		=0	п Н		CHO	21	
B-6	urs-12-ene	=0	н Н				34
B-7	oleana-11,13(18)-diene			0.11	CO ₂ H	47	<i>c</i>
B-8	oleanane	=0	=0	β -H	CO ₂ Me	3	10
B-9	oleanane	=0	=0	β- H	CO ₂ H	37	c
B-10	oleanane	=O	=0	β -H	CHO	38	c
B-11 ^a	oleanane	=O	α-Br	-0-	-CO-	4	10
B-12 ^a	oleanane	=O	=O	-O-	-CO-	0	10
B-13	oleana-1,12-diene	=O	Н		CO_2Me	19	9
3-14	ursa-1,12-diene	=O	Н		CO_2Me	0	d
B-15	oleana-1,12-diene	=O	Н		CO_2H	85	d
B-16	ursa-1,12-diene	=O	Н		CO_2H	41	14
C- 1 ^a	urs-12-ene	=O	Н		CO_2H	55	c
C- 2	olean-12-ene	α-Cl	Н		CO_2Me	2	c
C- 3	olean-12-ene	α-Cl	Н		CO_2H	0	c
D-1	oleana-2,12-diene	Н	Н		CO_2Me	3	35
D-2	oleana-2,12-diene	Н	Н		CO_2H	0	c
D-3 ^a	olean-12-ene		Н		CO_2H	0	c
E-1 ^a	A-ring cleaved olean-12-ene		Н		CO_2Me	21	36
E- 2 a	A-ring cleaved olean-12-ene		Н		CO_2H	33	37
E-3 ^a	A-ring cleaved urs-12-ene		Н		CO_2H	39	37
E- 4 a	A-ring cleaved olean-12-ene		Н		CO_2H	22	37
E- 5 a	A-ring cleaved urs-12-ene		Н		CO ₂ H	55	37
E- 6 ^a	A-ring cleaved urs-12-ene		H		CO ₂ H	10	37
E-1 ^a	C-ring cleaved oleanane	β -OAc			CH ₂ OAc	52	c
F- 2 ^a	C-ring cleaved oleanane	β-OAc			CH ₂ OAc	12	c
F- 3 ª	C-ring cleaved oleanane	β-OAc			CH ₂ OAc	52	c
	- Imp dicarca dicananc	P 3/10			01120110	<i>5</i> ≈	·

Table 1 (Continued)

					inhibition (%)			
compd	skeleton	C-3	C-12	C-13	C-17	at 10 μ M b	ref	
G-1 ^a	olean-12-ene		Н		CO ₂ Me	0	36	
G-2 ^a	olean-12-ene		Н		CO_2H	51	37	
hydrocortisone						80		

^a Structure shown below this table. ^b Details of the evaluation method are described in the Experimental Section. ^c Unknown compound (synthesis and spectral data will be published elsewhere). ^d Unknown compound (synthesis and spectral data are shown in this paper).

Scheme 1^a

^a Reagents: (a) PhSeCl, EtOAc; mCPBA, pyr, EtOAc; (b) LiI, DMF.

compound. Therefore, about 60 oleanolic and ursolic acid derivatives were initially randomly synthesized. They are divided into seven categories: 3-hydroxy derivatives, **A**; 3-oxo derivatives, **B**; chloro derivatives, **C**; dehydroxy-oleanane derivatives, **D**; A-ring cleaved derivatives, **E**; C-ring cleaved oleanane derivatives, **F**; and lactams, **G** (see Table 1). In the preliminary screen of these derivatives for inhibition of production of NO induced by IFN- γ in mouse macrophages, 3-oxooleana-1,12-dien-28-oic acid (**B**-15) was found to show significant activity (inhibition: 85% at 10 μ M, IC₅₀ = 5.6 μ M). (See Tables 1 and 2.)

Design and Synthesis of New Derivatives. When **B-15** is compared with the other derivatives, it has the following features: first, it is an oleanane; second, it has a 1-en-3-one functionality in ring A; third, it has a carboxyl group at C-17. We focused our attention on the 1-en-3-one functionality in ring A among these features. We therefore designed novel olean- and urs-12-ene triterpenoids with a 1-en-3-one functionality having a substituent at C-2 in ring A, **3-19**, and novel triter-

penoid—steroid hybrid compounds, **20** and **21** 8 (see Table 2). The syntheses of these newly designed derivatives and compounds **B-13**–**B-16** are illustrated in Schemes 1–6.

Ester **B-13**⁹ was synthesized in 62% yield by introduction of a double bond at C-1 of methyl oleanonate (**B-3**)¹⁰ with phenylselenenyl chloride (PhSeCl) in ethyl acetate and sequential addition of pyridine and mchloroperbenzoic acid. 11,12 Acid B-15 was synthesized in 85% yield by halogenolysis of **B-13** with lithium iodide in N.N-dimethylformamide (DMF).¹³ Similarly, acid $\mathbf{B} ext{-}\mathbf{16}^{14}$ was synthesized in 58% yield via ester $\mathbf{B} ext{-}\mathbf{14}$ from methyl ursonate (B-4).15 Epoxide 229 was prepared in 99% yield by epoxidation of **B-13** with alkaline hydrogen peroxide. Treatment of **22** with sodium methoxide¹⁶ gave enone 23 (yield, 87%; 98% based on recovered 22). Diosphenol **24** was synthesized by demethylation of the methyl enol ether at C-2 of 23 with hydrochloric acid in acetic acid (yield, 81%). Halogenolysis of 24 gave acid 4 (yield, 18%). Halogenolysis of 23 gave a desired partial demethylated product 5 in 28% (41% based on recovered

Scheme 2a

B-13
$$\xrightarrow{Q}$$
 CO_2Me
 ECO_2Me
 ECO_2Me

^a Reagents: (a) 30% H₂O₂, NaOH(aq), THF; (b) NaOMe, MeOH; (c) HCl, AcOH; (d) LiI, DMF.

Scheme 3^a

22 a
$$X \longrightarrow H$$
 CO_2Me b $X \longrightarrow H$ CO_2H CO_2

^a Reagents: (a) HX, AcOH, CHCl₃; (b) LiI, DMF.

23) yield.¹⁷ Chloride 6 was synthesized in 81% yield from 22 with hydrogen chloride in acetic acid and chloroform. 18 Halogenolysis of 6 gave chloride 7 in 77% yield. Similarly, bromides 8 and 9 were prepared from 22 and 8 (yield, 96% and 76%), respectively. Hydroxymethylene 25^{19,20} was prepared in 95% yield by formylation of B-3 with ethyl formate in the presence of sodium methoxide in benzene.²¹ Isoxazole **26** was prepared in 86% yield by condensation of 25 with hydroxylamine.²² Cleavage of the isoxazole moiety of **26** with sodium methoxide gave nitrile **27** in 99% yield.²² ¹H NMR showed that 27 is a mixture of three tautomers [27a, 27b (2 α -cyano), and 27c (2 β -cyano)] and that 27a is the major one in CDCl₃. Enone 10 was prepared in 88% yield by 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) oxidation of **27** in benzene, although the same method as for B-13 gave 10 in only 35% yield. Halogenolysis of 10 gave acid 11 in 71% (91% based on recovered 10) yield. Similarly, ursane derivative 12 was synthesized in 52% yield via 28,20,23 29, and 30 from **B-4**. Acid **13** was prepared in 74% yield by halogenolysis of **12**. Enal **14** was prepared from **25** by PhSeClpyridine in methylene chloride and sequential addition of 30% hydrogen peroxide²⁴ (yield, 71%; 79% based on recovered 25). Halogenolysis of 14 did not give acid 15 but a complex mixture. Therefore, the synthesis of acid **15** from oleanonic acid (\mathbf{B} - $\mathbf{1}$)¹⁰ was attempted. Formylation of **B-1** with ethyl formate in the presence of sodium methoxide in tetrahydrofuran gave **32**²⁰ (yield, 45%; 66% based on recovered **B-1**). Acid **15** was prepared from 32 according to the same method as for 14 (yield, 71%; 84% based on recovered 32). Jones oxidation of 14 gave acid 16 in 30% (39% based on recovered 14)

yield. Because this yield was not enough to synthesize derivatives **3** and **17–19** from **16**, an alternative route was adopted. Ester **31** was prepared in 74% (89% based on recovered B-3) yield from B-3 by Stiles' reagent (methoxymagnesium methyl carbonate) in DMF,²⁵ followed by methylation with diazomethane. ¹H NMR showed that **31** is the single tautomer in CDCl₃ as depicted in Scheme 5. Enone 17 was prepared from 31 according to the same method as for 14 (yield, 83%; 90% based on recovered 31). Hydrolysis of 17 with potassium hydroxide in aqueous methanol gave acid **16** selectively in 97% yield because the methoxycarbonyl group at C-17 of 17 is sterically hindered. Halogenolysis of 16 gave dicarboxylic acid 3 in 58% yield. Methylation of 3 with methanol under acidic conditions gave ester 18 selectively in 78% yield because of the steric hindrance of the carboxylic acid at C-17 of 3. Amide 19 was prepared selectively in 96% yield from 17 with saturated ammonia-methanol.

Biological Results and Discussion

The inhibitory activities [IC₅₀ (μ M) value] of compounds **B-1**, **B-13**, **B-15**, **B-16**, **1–21**, and hydrocortisone (a positive control) on NO production induced by IFN-γ in mouse macrophages are shown in Table 2. These derivatives are arranged according to the strength of Taft's σ^* values²⁶ of substituents at C-2. These results provide the following interesting structure-activity relationships:

(1) In the A ring, a 1-en-3-one functionality is important for significant activity. The lead compound **B-15** is much more potent than the C-3 ketone **B-1** and the

Scheme 4a

B-3 a HO
$$\frac{1}{H}$$
 HO $\frac{1}{H}$ HO $\frac{1}{H$

^a Reagents: (a) HCO₂Et, NaOMe, PhH; (b) NH₂OH·HCl, aq EtOH; (c) NaOMe, Et₂O, MeOH; (d) DDQ, PhH; (e) LiI, DMF.

Scheme 5^a

25 a OHC
$$\stackrel{\stackrel{\circ}{H}}{\stackrel{\circ}{H}}$$
 14 16 $\stackrel{\circ}{\stackrel{\circ}{H}}$ 3 $\stackrel{\circ}{\stackrel{\circ}{H}}$ 18 $\stackrel{\circ}{\stackrel{\circ}{H}}$ 3 $\stackrel{\circ}{\stackrel{\circ}{H}}$ 18 $\stackrel{\circ}{\stackrel{\circ}{H}}$ 3 1 17 19

^a Reagents: (a) PhSeCl, pyr, CH₂Cl₂; 30% H₂O₂, CH₂Cl₂; (b) Jones; (c) Stiles' reagent, DMF; (d) CH₂N₂, Et₂O, THF; (e) KOH, aq MeOH; (f) LiI, DMF; (g) H₂SO₄, MeOH; (h) NH₃, MeOH.

Scheme 6a

^a Reagents: (a) HCO₂Et, NaOMe, THF; (b) PhSeCl, pyr, CH₂Cl₂; 30% H₂O₂, CH₂Cl₂.

C-3 alcohol **1** (oleanolic acid). Also, the ursane derivative **B-16** is more potent than the C-3 alcohol **2** (ursolic acid).

- (2) A correlation between Taft's σ^* values of substituents at C-2 and biological activity is not observed. This result shows that the activity does not depend on the strength of electron-withdrawing effect of a substituent at C-2.
- (3) Carboxyl, methoxycarbonyl, and nitrile groups at C-2 enhance activity. Compounds **3**, **10**, **11**, **16**, and **17** are about 10-100 times more potent than **B-15**. In
- particular, **3** showed the highest activity (IC₅₀ = 0.07 μ M) in this series of compounds. The potency of **3** was similar to that of hydrocortisone (IC₅₀ = 0.01 μ M).
- (4) Hydroxyl, aminocarbonyl, methoxy, chloride, and bromide groups decrease activity. Compounds 4-9 and 19 are much less potent than B-15.
- (5) A formyl group does not confer activity but only toxicity.
 - (6) 23,24-Dimethyl groups are important for signifi-

Table 2. Activity of Olean- and Urs-12-ene Triterpenoids with Various 1-En-3-one Functionalities

$$R_1$$
 R_2 R_1 R_2 R_3 R_4 R_5 R_6 R_7 R_8 R_9 R_9

				Taft's σ^*			activity c
compd	skeleton ^a	R ₁ at C-2	R ₂ at C-17	value of R ₁	formula	analyses b	$IC_{50} (\mu M)$
B-13	O	Н	CO ₂ Me		$C_{31}H_{46}O_3$	ref 9	31
B-15	O	H	CO_2H		$C_{30}H_{44}O_{3} \cdot 3/4H_{2}O$	C, H	5.6
20	D	H	CO_2Me		$C_{29}H_{40}O_3 \cdot 1/4H_2O$	C, H	>40
21	D	Н	CO_2H		$C_{28}H_{38}O_{3} \cdot 1/3H_{2}O$	C, H	13
B-16	U	H	CO_2H		$C_{30}H_{44}O_3$	ref 14	13
5	O	OH	CO_2H	1.34	$C_{30}H_{44}O_4 \cdot 1/2H_2O$	C, H	27
19	O	$CONH_2$	CO_2Me	1.68	$C_{32}H_{47}O_4N \cdot 3/4H_2O$	C, H, N	14
4	O	OMe	CO_2H	1.81	$C_{31}H_{46}O_4 \cdot 1/2H_2O$	C, H	30
17	O	CO_2Me	CO_2Me	2.00	$C_{33}H_{48}O_5$	C, H	0.9
18	O	CO_2Me	CO_2H	2.00	$C_{32}H_{46}O_5$	C, H	2.2
16	O	CO_2H	CO_2Me	2.08	$C_{32}H_{46}O_5 \cdot 1/2H_2O$	C, H	0.8
3	O	CO_2H	CO_2H	2.08	$C_{31}H_{44}O_5$	C, H	0.07
14	O	CHO	CO_2Me	2.15	$C_{32}H_{46}O_4$	C, H	$toxic^d$
15	O	CHO	CO_2H	2.15	$C_{31}H_{44}O_4 \cdot 1/2H_2O$	C, H	$toxic^d$
8	O	Br	CO_2Me	2.84	$C_{31}H_{45}O_3Br$	C, H	>40
9	O	Br	CO_2H	2.84	$C_{30}H_{43}O_3Br\cdot H_2O$	C, H	7.3
6	O	Cl	CO_2Me	2.96	$C_{31}H_{45}O_3Cl$	C, H	>40
7	O	Cl	CO_2H	2.96	$C_{30}H_{43}O_3Cl \cdot 1/4H_2O$	C, H	>40
10	O	CN	CO_2Me	3.30	$C_{32}H_{45}O_3N \cdot 1/4H_2O$	C, H, N	0.7
11	O	CN	CO_2H	3.30	$C_{31}H_{43}O_3N \cdot 1/2H_2O$	C, H, N	0.6
12	U	CN	CO_2Me	3.30	$C_{32}H_{45}O_3N \cdot 3/4H_2O$	C, H, N	5.1
13	U	CN	CO_2H	3.30	$C_{31}H_{43}O_3N \cdot H_2O$	C, H, N	6.2
B-1		oleanonic acid			$C_{30}H_{46}O_3$	ref 10	37
1		oleanolic acid			$C_{30}H_{48}O_3$	ref 10	>40
2		ursolic acid			$C_{30}H_{48}O_3$	ref 15	$toxic^e$
		hydrocortisone					0.01

^a O, 3-oxooleana-1,12-diene; D, 23,24-dinor-3-oxooleana-1,4,12-triene; U, 3-oxoursa-1,12-diene. ^b C, H, and N analyses were within ±0.4% of the theoretical values. ^c Details of the evaluation method are described in the Experimental Section. IC₅₀ values of **3** and hydrocortisone were determined in the range of 0.1 pM-1 μ M (10-fold dilutions). The other compounds were assayed in the range of 0.01-40 μ M (4-fold dilutions). Values are an average of two separate experiments. d Compounds 14 and 15 were toxic to cells above $I\mu M$ and were not active below 1 μ M. e Ursolic acid (2) was toxic to cells above 10 μ M and was not active below 10 μ M.

cant activity. B-15 is more potent than 23,24-dinorolean-1-en-3-one derivative 21.

(7) The oleanane skeleton is more potent than the ursane skeleton. **B-15**, **10**, and **11** are more potent than **B-16**, **12**, and **13**, respectively.

(8) The role of methoxycarbonyl and carboxyl groups at C-17 is ambiguous. In some analogues, the carboxyl group is more potent than the methoxycarbonyl group: acids **B-15**, **3**, **9**, and **21** are more potent than esters **B-13**, **16**, **8**, and **20**, respectively. For other analogues, the carboxyl and methoxycarbonyl groups show similar activity: acids 11 and 13 show similar activity to esters 10 and 12, respectively. Lastly, acid 18 is less potent than ester 17.

The inhibitory activity of new triterpenoids 3 and 11 was not blocked by the glucocorticoid antagonist, RU-486,²⁷ which reverses the action of hydrocortisone (see Figure 1). These data strongly suggest that the actions of these triterpenoids on the iNOS system are not mediated by their interaction with the glucocorticoid receptor.

On the basis of these structure—activity relationships, further lead optimization is in progress. Further biological evaluation and studies on the mechanism of action of **3** are also in progress.

Experimental Section

General Experimental Procedures. Melting points were determined on a Thomas-Hoover capillary melting point apparatus and are uncorrected. Optical rotations were measured with a Jasco DIP-181 digital polarimeter. UV and IR spectra were recorded on a Hewlett-Packard 8451A UV/VIS spectrophotometer and a Perkin-Elmer 600 series FTIR spectrophotometer, respectively. ¹H (300 MHz) and ¹³C (75 MHz) NMR spectra were recorded on a Varian XL-300 Fourier transform spectrometer. The chemical shifts are reported in δ (ppm) using the δ 7.27 signal of CHCl₃ (¹H NMR) and the δ 77.23 signal of CDCl₃ (13C NMR) as internal standards. Lowresolution mass spectra and high-resolution MS data were obtained on a Micromass 70-VSE unless otherwise stated. Elemental microanalysis was performed by Atlantic Microlab Inc. TLC and preparative TLC (prep-TLC) were performed with Merck precoated TLC plates silica gel 60 \hat{F}_{254} . Flash column chromatography was done with Select Scientific silica gel (230-400 mesh). The standard work up method was as follows: an organic extract was washed with saturated aqueous NaHCO3 solution (three times) followed by saturated aqueous NaCl solution (three times), then dried over anhydrous MgSO₄, and filtered. The filtrate was evaporated in

Methyl 3-Oxooleana-1,12-dien-28-oate (B-13).9 A solution of methyl oleanonate (B-3)10 (2.00 g, 4.27 mmol) and phenylselenenyl chloride (98%) (1.00 g, 5.12 mmol) in EtOAc (85 mL) was stirred at room temperature for 3 h. To the stirred

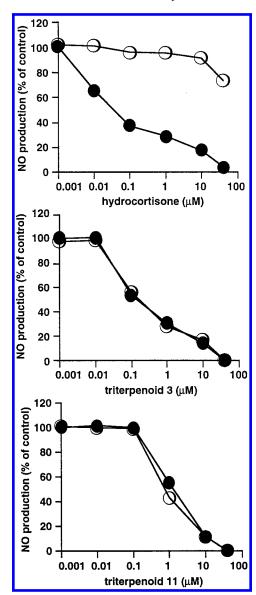


Figure 1. Blockage by glucocorticoid antagonist RU486 of hydrocortisone-inhibited NO production but not of triterpenoid (3 and 11) inhibited NO production in primary mouse macrophages. Macrophage cells were incubated with IFN-γ (20 ng/ mL) together with hydrocortisone or triterpenoids without RU486 (\bullet); in some cases RU486 (1 μ M) was added simultaneously to both hydrocortisone- and triterpenoid-treated cell wells (O). RU486 itself does not interfere with NO production at the concentration tested.

mixture, saturated aqueous NaHCO₃ solution was added. After most of the aqueous layer was removed, pyridine (844 mg, 10.7 mmol) and m-chloroperbenzoic acid (50-60%) (3.68 g, 10.7 mmol) were added to the organic layer. The mixture was stirred at room temperature for 1 h. The mixture was washed with 5% aqueous NaOH solution (three times), saturated aqueous NH₄Cl solution (three times), and saturated aqueous NaCl solution (three times); dried over anhydrous MgSO₄; and filtered. The filtrate was evaporated in vacuo to give a solid. The solid was subjected to flash column chromatography [hexanes-EtOAc (5:1)] to give **B-13** as a crystalline solid (1.23 g, 62%): mp 159–161 °C; $[\alpha]^{25}_D$ +103° (\dot{c} 0.64, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 230 (3.92) nm. IR (KBr): 2946, 2867, 1728, 1660 cm⁻¹. ¹H NMR (CDCl₃): δ 7.04 (1H, d, J= 10.1 Hz), 5.81 (1H, d, J = 10.1 Hz), 5.36 (1H, t, J = 3.7 Hz), 3.64, (3H, s), 2.90 (1H, dd, J = 4.6, 13.9 Hz), 1.17 (3H, s), 1.16 (6H, s), 1.10, 0.94, 0.91, 0.83 (each 3H, s). 13 C NMR (CDCl₃): δ 205.5, 178.4, 159.3, 144.5, 125.2, 121.9, 53.6, 51.8, 47.0, 45.9, 44.7, 42.2, 42.0, 41.7, 40.3, 39.7, 34.1, 33.3, 32.7, 32.5, 30.9, 28.0, 27.9, 26.0,

23.8, 23.5, 23.2, 21.8, 19.1, 18.8, 17.5. EIMS (70 eV) m/z. 466 [M]⁺ (73), 451 (11), 407 (31), 262 (57), 203 (100). HREIMS: Calcd for C₃₁H₄₆O₃: 466.3447. Found: 466.3446.

Methyl 3-Oxoursa-1,12-dien-28-oate (B-14). B-14 was prepared from methyl ursonate (**B-4**)¹⁵ according to the same method as for **B-13** to give an amorphous solid (66%): $[\alpha]^{26}$ _D +93° (c 0.77, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 232 (3.95) nm. IR (KBr): 2974, 2935, 2871, 1725, 1669 cm⁻¹. ¹H NMR (CDCl₃): δ 7.06 (1H, d, J = 10.1 Hz), 5.81 (1H, d, J = 10.1Hz), 5.33 (1H, t, J = 3.8 Hz), 3.63 (3H, s), 2.28 (1H, d, J =11.5 Hz), 1.17, 1.15 (each 3H, s), 1.10 (6H, s), 0.95 (3H, d, J =5.4 Hz), 0.87 (3H, d, J = 6.3 Hz), 0.85 (3H, s). ¹³C NMR (CDCl₃): δ 205.5, 178.2, 159.5, 139.0, 125.2, 125.0, 53.7, 53.3, 51.7, 48.4, 44.7, 42.6, 41.9, 40.5, 39.5, 39.2, 39.1, 36.8, 33.0, 30.8, 28.2, 28.1, 24.4, 23.7, 23.5, 21.8, 21.4, 19.1, 19.0, 17.7, 17.2. EIMS (70 eV) m/z. 466 [M]⁺ (14), 406 (12), 262 (74), 203 (100). HREIMS: Calcd for C₃₁H₄₆O₃: 466.3447. Found: 466.3442.

3-Oxooleana-1,12-dien-28-oic Acid (B-15). A mixture of B-13 (100 mg, 0.21 mmol) and LiI (500 mg) in dry DMF (2 mL) was heated under reflux for 6 h. The mixture was acidified with 5% aqueous HCl solution and then extracted with a mixture of $\hat{C}H_2Cl_2$ and Et_2O (1:2) three times. The extract was worked up according to the standard method to give a solid (110 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (5:1) followed by hexanes-EtOAc (2: 1)] to give **B-15** as an amorphous solid (82 mg, 85%): $[\alpha]^{26}D$ +103° (c 0.45, CHCl₃). UV (ÉtOH) $\lambda_{\rm max}$ ($\log \epsilon$): 230 (3.75) nm. IR (KBr): 2941, 2866, 1732, 1695, 1671 cm⁻¹. ¹H NMR (CDCl₃): δ 7.04 (1H, d, J = 10.2 Hz), 5.81 (1H, d, J = 10.2Hz), 5.35 (1H, t, J = 3.3 Hz), 2.86 (1H, dd, J = 4.2, 13.4 Hz), 1.16, 1.152, 1.147, 1.07, 0.94, 0.91, 0.84 (each 3H, s). ¹³C NMR $(CDCl_3)$: δ 205.5, 184.5, 159.2, 144.2, 125.3, 122.1, 53.5, 46.8, 45.8, 44.7, 42.1, 41.9, 41.3, 40.2, 39.7, 34.0, 33.3, 32.6, 32.5, 30.9, 28.0, 27.8, 26.0, 23.7, 23.5, 23.0, 21.8, 19.0, 18.9, 17.7. EIMS (70 eV) m/z. 452 [M]⁺ (8.8), 437 (3.8), 406 (6.8), 248 (80), 233 (14), 203 (100). HREIMS: Calcd for C₃₀H₄₄O₃: 452.3290. Found: 452.3289. Anal. (Table 2).

3-Oxoursa-1,12-dien-28-oic Acid (B-16).14 B-16 was prepared from **B-14** according to the same method as for **B-15** to give an amorphous solid (88%): $[\alpha]^{26}_D$ +91° (c 0.84, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 230 (3.99) nm. IR (KBr): 3306, 2973, 2930, 2870, 1729, 1695, 1669 cm $^{-1}$. ¹H NMR (CDCl₃): δ 7.07 (1H, d, J = 10.1 Hz), 5.82 (1H, d, J = 10.1 Hz), 5.33 (1H, t, J)= 3.7 Hz), 2.24 (1H, d, J = 11.2 Hz), 1.18, 1.16, 1.11, 1.09 (each 3H, s), 0.96 (3H, d, J = 6.1 Hz), 0.88 (3H, d, J = 6.4 Hz), 0.88 (3H, s). ¹³C NMR (CDCl₃): δ 205.5, 183.9, 159.4, 138.8, 125.3, 53.6, 52.9, 48.3, 44.7, 42.5, 41.9, 40.5, 39.6, 39.2, 39.0, 36.8, 32.9, 30.8, 28.2, 28.1, 24.2, 23.7, 23.4, 21.8, 21.3, 19.0, 17.8, 17.2. FABMS (NBA) m/z. 453 [M + H]⁺ (100) (by a Micromass ZAB-SE). HRFABMS: Calcd for $C_{30}H_{44}O_3 + H$: 453.3369. Found: 453.3335 (by a Micromass 70-SE-4F).

2-Carboxy-3-oxooleana-1,12-dien-28-oic Acid (3). A mixture of 16 (109 mg, 0.21 mmol) and LiI (520 mg) in dry DMF (1.5 mL) was heated under reflux for 1 h. After 5% aqueous HCl solution was added, the acidic mixture was extracted with EtOAc three times. The extract was washed with water (three times) and saturated aqueous NaCl solution (three times), dried over anhydrous MgSO₄, and filtered. The filtrate was evaporated in vacuo to give a residue (108 mg). The residue was subjected to flash column chromatography [CH₂Cl₂-MeOH (15:1) followed by CH₂Cl₂-MeOH (10:1)] to afford 3 as a crystalline solid (61 mg, 58%): mp > 250 °C dec; $[\alpha]^{26}$ _D +81° (c 0.53, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 234 (3.88) nm. IR (KBr): 3389, 2943, 2872, 1752, 1696, 1637 cm⁻¹. ¹H NMR (CDCl₃): δ 8.43 (1H, s), 5.37 (1H, t, J = 3.5 Hz), 2.87 (1H, dd, J = 3.8, 13.9 Hz), 1.25, 1.22, 1.18, 1.15, 0.95, 0.93, 0.88 (each 3H, s). ¹³C NMR (CDCl₃): δ 209.0, 183.9, 173.2, 165.2, 144.2, 123.4, 121.7, 52.4, 46.8, 45.7, 45.5, 42.3, 41.4, 41.1, 40.6, 40.4, 34.0, 33.2, 32.5, 32.3, 30.9, 28.4, 27.8, 26.0, 23.7, 23.5, 23.0, 22.0, 19.0, 18.4, 17.8. EIMS (70 eV) m/z. 496 [M]+ (3.0), 478 (3.4), 452 (7.6), 248 (56), 231 (35), 203 (100). HREIMS: Calcd for C₃₁H₄₄O₅: 496.3189. Found: 496.3196. Anal. (Table 2).

2-Hydroxy-3-oxooleana-1,12-dien-28-oic Acid (4). 4 was prepared from 24 according to the same method as for B-15

except that the reaction time was 2 h. The reaction mixture was subjected to flash column chromatography [hexanes-EtOAc (5:1) followed by hexanes-EtOAc (4:1)] to give 4 as an amorphous solid (18%): $[\alpha]^{25}_D + 99^\circ$ (c 0.46, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 272 (3.71) nm. IR (KBr): 3434, 2938, 1698, 1667, $^{1649}~{\rm cm}^{-1}$. $^{1}{\rm H}~{\rm NMR}~({\rm CDCl_3})$: $\delta~6.35~(1{\rm H,~s}),~5.96~(1{\rm H,~brs}),$ 5.34 (1H, t, J = 3.5 Hz), 2.86 (1H, dd, J = 3.8, 13.9 Hz), 1.23, 1.22, 1.14, 1.11, 0.94, 0.92, 0.83 (each 3H, s). ¹³C NMR (CDCl₃): δ 201.2, 184.0, 144.1, 143.9, 128.4, 122.3, 54.0, 46.8, 45.8, 44.1, 43.3, 42.2, 41.3, 40.2, 38.7, 34.0, 33.3, 32.6, 30.9, 27.8, 27.4, 26.1, 23.8, 23.6, 23.0, 22.0, 19.9, 18.9, 17.7. EIMS (70 eV) m/z. 468 [M]⁺ (3.2), 248 (13), 203 (23), 149 (42), 84 (100). HREIMS: Calcdfor C₃₀H₄₄O₄: 468.3240. Found: 468.3222. Anal. (Table 2).

2-Methoxy-3-oxooleana-1,12-dien-28-oic Acid (5). A mixture of 23 (230 mg, 0.46 mmol) and LiI (1045 mg) in dry DMF (3.5 mL) was heated under reflux for 4 h. The reaction mixture was worked up according to the same method as for B-15 to give a solid (230 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (3:1) followed by hexanes-EtOAc (2:1), then hexanes-EtOAc (1:1)] to give 24 (35 mg; 16%, 23% based on recovered **23**), **23** (74 mg), **4** (27 mg; 12%, 18% based on recovered 23), and 5 as an amorphous solid (63 mg; 28%, 41% based on recovered **23**): $[\alpha]^{26}_{D} + 96^{\circ}$ (c 0.29, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 266 (3.84) nm. IR (KBr): 3307, 2947, 2862, 1732, 1693, 1622 cm⁻¹. ¹H NMR (CDCl₃): δ 5.96 (1H, s), 5.36 (1H, t, J = 3.3 Hz), 3.56 (3H, s), 2.87 (1H, dd, J)= 4.2, 13.9 Hz), 1.17 (9H, s), 1.11, 0.94, 0.91, 0.84 (each 3H, s). ¹³C NMR (CDCl₃): δ 200.0, 184.4, 149.1, 144.4, 126.1, 122.1, 55.0, 53.2, 46.8, 45.9, 45.4, 43.3, 42.2, 41.3, 40.2, 38.5, 34.0, 33.3, 32.5, 30.9, 28.6, 27.8, 26.1, 23.8, 23.0, 22.0, 20.4, 19.2, 17.6. EIMS (70 eV) m/z. 482 [M]+ (11), 415 (6.5), 245 (18), 203 (33), 157 (100). HREIMS: Calcd for C₃₁H₄₆O₄: 482.3396. Found: 482.3375. Anal. (Table 2).

Methyl 2-Chloro-3-oxooleana-1,12-dien-28-oate (6). A solution of 22 (99 mg, 0.21 mmol) in AcOH including 1 M HCl (2.5 mL) and CHCl₃ (2.5 mL) was stirred at room temperature overnight. The mixture was diluted with CH2Cl2. After it was washed with water three times, it was worked up according to the standard method to give a solid (96 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (6:1)] to afford **6** as an amorphous solid (84 mg, 81%): $[\alpha]^{26}$ _D +98° (c 0.26, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 250 (3.91) nm. IR (KBr): 2943, 2866, 1727, 1689 cm $^{-1}$. 1 H NMR (CDCl $_{3}$): δ 7.22 (1H, s), 5.34 (1H, t, J = 3.5 Hz), 3.62 (3H, s), 2.89 (1H, dd, J = 4.2, 13.7 Hz), 1.203, 1.197 (each 3H, s), 1.14 (6H, s), 0.93, 0.90, 0.80 (each 3H, s). ^{13}C NMR (CDCl3): $\,\delta$ 197.4, 178.3, 155.0, 144.5, 129.8, 121.5, 53.3, 51.8, 46.9, 46.3, 45.8, 42.2, 42.1, 41.64, 41.57, 40.3, 34.0, 33.3, 32.4, 30.9, 28.4, 27.8, 26.0, 23.8, 23.5, 23.1, 22.1, 19.1, 18.8, 17.5. EIMS (70 eV) m/z. 500 [M]⁺ (21), 262 (27), 247 (96), 203 (100). HREIMS: Calcd for C₃₁H₄₅O₃Cl: 500.3057. Found: 500.3060. Anal. (Table 2).

2-Chloro-3-oxooleana-1,12-dien-28-oic Acid (7). 7 was prepared from 6 according to the same method as for B-15 except that the reaction time was 4 h. The reaction mixture was subjected to flash column chromatography [hexanes-EtOAc (4:1) followed by hexanes-EtOAc (3:1)] to give 7 as an amorphous solid (77%): $[\alpha]^{26}D + 88^{\circ}$ (c 0.50, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 252 (3.20) nm. IR (KBr): 3297, 2943, 2870, 1733, 1691, 1601 cm⁻¹. ¹H NMR (CDCl₃): δ 7.23 (1H, s), 5.35 (1H, t, J = 3.3 Hz), 2.86 (1H, dd, J = 4.3, 13.8 Hz), 1.22, 1.21, 1.16, 1.13, 0.94, 0.92, 0.84 (each 3H, s). 13 C NMR (CDCl₃): δ 197,4, 184.4, 154.9, 144.3, 129.9, 121.8, 53.2, 46.8, 46.4, 45.8, 42.21, 42.16, 41.6, 41.3, 40.3, 34.0, 33.3, 32.5, 32.4, 30.9, 28.5, 27.8, 26.0, 23.7, 23.5, 23.0, 22.1, 19.0, 18.9, 17.7. EIMS (70 eV) m/z. 486 [M]+ (25), 248 (100), 203 (96). HREIMS: Calcd for C₃₀H₄₃O₃Cl: 486.2901. Found: 486.2898. Anal. (Table 2).

Methyl 2-Bromo-3-oxooleana-1,12-dien-28-oate (8). A solution of 22 (220 mg, 0.46 mmol) in AcOH including 1 M HBr (4.9 mL) and CHCl₃ (6.1 mL) was stirred at room temperature for 1 h. The mixture was diluted with CH₂Cl₂. After it was washed with water three times, it was worked up according to the standard method to give a solid (260 mg). The solid was subjected to flash column chromatography [hexanes-

EtOAc (6:1)] to afford 8 as an amorphous solid (238 mg, 96%): $[\alpha]^{26}_{D} + 88^{\circ} (c \ 0.51, CHCl_{3})$. UV (EtOH) $\lambda_{max} (\log \epsilon)$: 260 (3.69) nm. IR (KBr): 2943, 2870, 1733, 1691, 1601 cm⁻¹. ¹H NMR (CDCl₃): δ 7.49 (1H, s), 5.35 (1H, t, J = 3.5 Hz), 3.63 (3H, s), 2.90 (1H, dd, J = 4.0, 13.8 Hz), 1.20, 1.15 (each 6H, s), 0.94, 0.91, 0.81 (each 3H, s). 13 C NMR (CDCl₃): δ 197.3, 178.3, 159.5, 144.6, 121.8, 121.5, 53.3, 51.8, 46.9, 46.5, 45.8, 43.1, 42.3, 42.1, 41.7, 40.3, 34.0, 33.3, 32.4, 30.9, 28.7, 27.8, 26.0, 23.8, 23.6, 23.2, 22.3, 19.1, 18.7, 17.5. EIMS (70 eV) m/z. 546 (5.0) and $544 \ (5.2) \ [M]^+, \ 262 \ (8.5), \ 203 \ (24), \ 118 \ (100), \ 116 \ (100).$ HREIMS: Calcd for C₃₁H₄₅O₃Br: 544.2552. Found: 544.2553. Anal. (Table 2).

2-Bromo-3-oxooleana-1,12-dien-28-oic Acid (9). 9 was prepared from 8 according to the same method as for B-15 except that the reaction time was 4 h. The reaction mixture was subjected to flash column chromatography [hexanes-EtOAc (4:1) followed by hexanes-EtOAc (3:1)] to give 9 as an amorphous solid (76%): $[\alpha]^{26}_D$ +82° (c 0.31, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 260 (3.52) nm. IR (KBr): 3434, 2939, 2870, 1727, 1686, 1601 cm $^{-1}$. ¹H NMR (CDCl₃): δ 7.49 (1H, s), 5.35 (1H, t, J = 3.4 Hz), 2.86 (1H, dd, J = 4.2, 13.7 Hz), 1.21 (6H, s), 1.16, 1.14, 0.94, 0.92, 0.83 (each 3H, s). 13 C NMR (CDCl₃): δ 197.2, 184.4, 159.3, 144.3, 121.84, 121.79, 53.3, 46.8, 46.5, 45.8, 43.1, 42.2, 42.0, 41.3, 40.3, 34.0, 33.2, 32.5, 32.4, 30.9, 28.7, 27.8, 26.0, 23.7, 23.5, 23.0, 22.2, 19.1, 18.7, 17.7. EIMS (70 eV) m/z. 532 (13) and 530 (14) [M]+, 285 (5.6), 283 (6.2), 248 (100), 235 (10), 233 (11), 203 (84). HREIMS: Calcd for C₃₀H₄₃O₃Br: 530.2396. Found: 530.2383. Anal. (Table 2).

Methyl 2-Cyano-3-oxooleana-1,12-dien-28-oate (10). A solution of 27 (141 mg, 0.28 mmol) and DDQ (98%) (79 mg, 0.34 mmol) in benzene (10 mL) was heated under reflux for 4 h. After insoluble matter was removed by filtration, the filtrate was evaporated in vacuo to give a solid. The solid was subjected to flash column chromatography [benzene-acetone (20:1)] to give a crystalline solid (123 mg, 88%): mp 201–202 °C; $[\alpha]^{26}$ _D +67° (c 0.53, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 240 (3.65) nm. IR (KBr): 2945, 2874, 2232, 1724, 1686 cm⁻¹. ¹H NMR (CDCl₃): δ 7.75 (1H, s), 5.36 (1H, t, J = 3.5 Hz), 3.64 (3H, s), 2.91 (1H, dd, J = 3.9, 13.9 Hz), 1.22, 1.21, 1.15, 1.14, 0.94, 0.92, 0.83 (each 3H, s). 13 C NMR (CDCl₃): δ 198.3, 178.3, 170.2, 144.8, 121.1, 115.2, 114.0, 52.8, 51.8, 46.9, 45.8, 45.1, 42.3, 41.7, 41.3, 40.8, 40.5, 34.0, 33.3, 32.4, 32.3, 30.9, 27.9, 27.8, 26.0, 23.8, 23.4, 23.1, 21.8, 18.9, 18.1, 17.6. EIMS (70 eV) m/z. 491 $[M]^+$ (35), 459 (13), 432 (27), 262 (22), 247 (24), 203 (100). HREIMS: Calcd for $C_{32}H_{45}O_3N$: 491.3399. Found: 491.3391. Anal. (Table 2).

2-Cyano-3-oxooleana-1,12-dien-28-oic Acid (11). 11 was prepared from 10 according to the same method as for B-15 except that the reaction time was 3 h. The reaction mixture was subjected to flash column chromatography [hexanes-EtOAc (3:1) followed by hexanes-EtOAc (2:1), then hexanes-EtOAc (1:1)] to give 11 as an amorphous solid (71%, 91% based on recovered **10**): $[\alpha]^{26}_D + 61^\circ$ (c 0.66, CHCl₃). UV (EtOH) λ_{max} $(\log \epsilon)$: 238 (3.87) nm. IR (KBr): 3387, 2947, 2870, 2233, 1729, 1691, 1609 cm⁻¹. 1 H NMR (CDCl₃): δ 7.75 (1H, s), 5.35 (1H, t, J = 3.3 Hz), 2.86 (1H, dd, J = 4.0, 13.6 Hz), 1.22, 1.21, 1.15, 1.12, 0.94, 0.92, 0.85 (each 3H, s). 13 C NMR (CDCl₃): δ 198.2, 184.3, 170.1, 144.4, 121.4, 115.1, 114.1, 52.7, 46.8, 45.7, 45.0, 42.2, 41.3, 40.8, 40.5, 33.9, 33.2, 32.5, 32.2, 30.9, 27.9, 27.7, 25.9, 23.7, 23.4, 22.9, 21.8, 18.9, 18.1, 17.7. EIMS (70 eV) m/z. 477 [M]⁺ (18), 462 (5.6), 431 (16), 416 (10), 248 (76), 235 (25), 203 (100). HREIMS: Calcd for C₃₁H₄₃O₃N: 477.3243. Found: 477.3240. Anal. (Table 2).

Methyl 2-Cyano-3-oxoursa-1,12-dien-28-oate (12). 12 was prepared from 30 according to the same method as for 10 to give an amorphous solid (62%): $[\alpha]^{26}_D +53^{\circ} (c \ 0.35, CHCl_3).$ UV (EtOH) λ_{max} (log ϵ): 240 (3.74) nm. IR (KBr): 2973, 2926, 2870, 2229, 1723, 1686 cm⁻¹. ¹H NMR (CDCl₃): δ 7.77 (1H, s), 5.33 (1H, t, J = 3.7 Hz), 3.62 (3H, s), 2.29 (1H, d, J = 11.2Hz), 1.23, 1.21, 1.14, 1.11 (each 3H, s), 0.96, 0.88 (each 3H, d, J = 6.3 Hz), 0.86 (3H, s). ¹³C NMR (CDCl₃): δ 198.3, 178.1, 170.4, 139.3, 124.2, 115.2, 114.0, 53.2, 52.8, 51.7, 48.3, 45.1, 42.7, 41.2, 40.70, 40.65, 39.1, 39.0, 36.7, 32.6, 30.8, 28.1, 28.0, 24.3, 23.6, 23.4, 21.8, 21.3, 18.9, 18.2, 17.8, 17.2. EIMS (70 eV) m/z. 491 [M]⁺ (38), 431 (35), 262 (46), 249 (82), 203 (65), 84 (100). HREIMS: Calcd for $C_{32}H_{45}O_3N$: 491.3399. Found: 491.3395. Anal. (Table 2).

2-Cyano-3-oxoursa-1,12-dien-28-oic Acid (13). 13 was prepared from 12 according to the same method as for **B-15** except that the reaction time was 4 h. The reaction mixture was subjected to prep-TLC [hexanes—EtOAc (1.5:1)] to give 13 as an amorphous solid (74%): $[\alpha]^{26}_{\rm D} + 48^{\circ}$ (c 0.50, CHCl₃). UV (EtOH) $\lambda_{\rm max}$ ($\log \epsilon$): 238 (3.86) nm. IR (KBr): 3417, 2973, 2926, 2870, 2233, 1731, 1689 cm⁻¹. ¹H NMR (CDCl₃): δ 7.77 (1H, s), 5.31 (1H, t, J = 3.2 Hz), 2.24 (1H, d, J = 11.0 Hz), 1.22, 1.20, 1.12, 1.11 (each 3H, s), 0.95, 0.88 (each 3H, d, J = 5.7 Hz), 0.87 (3H, s). ¹³C NMR (CDCl₃): δ 198.2, 184.2, 170.2, 139.0, 124.4, 115.1, 114.1, 52.8, 52.7, 48.2, 45.0, 42.6, 41.2, 40.68, 40.65, 39.1, 39.0, 36.7, 32.5, 30.7, 28.1, 28.0, 24.1, 23.6, 23.3, 21.8, 21.3, 18.9, 18.2, 17.7, 17.2. EIMS (70 eV) m/z. 477 [M]⁺ (22), 431 (23), 248 (100), 203 (48). HREIMS: Calcd for $C_{31}H_{43}O_{3}N$: 477.3243. Found: 477.3240. Anal. (Table 2).

Methyl 2-Formyl-3-oxooleana-1,12-dien-28-oate (14). To a stirred solution of phenylselenenyl chloride (98%) (161 mg, 0.82 mmol) in CH₂Cl₂ (7.2 mL) was added a solution of pyridine (75 mg, 0.95 mmol) in CH₂Cl₂ (1.0 mL) in an ice bath. After 15 min, a solution of **25** (204 mg, 0.41 mmol) in CH₂Cl₂ (2.0 mL) was added, and the mixture was stirred an additional 1 h. After the mixture was washed with 10% agueous HCl solution (3 mL) twice, 30% H₂O₂ (0.4 mL) was added to the stirred mixture in the ice bath. After an additional 40 min, the mixture was worked up according to the standard method to give a solid (199 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (5:1)] to afford 25 (20 mg) and 14 as an amorphous solid (144 mg; 71%, 79% based on recovered 25): $[\alpha]^{26}_D + 12^\circ$ (c 0.60, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 238 (3.85) nm. IR (KBr): 2946, 2867, 1724, 1703, 1673, 1610 cm $^{-1}$. ¹H NMR (CDCl₃): δ 10.00 (1H, s), 7.79 (1H, s), 5.37 (1H, t, J = 3.6 Hz), 3.63 (3H, s), 2.90 (1H, dd, J = 4.2, 13.9 Hz), 1.18, 1.17, 1.16, 1.14, 0.94, 0.91, 0.85 (each 3H, s). 13 C NMR (CDCl₃): δ 203.7, 190.7, 178.3, 165.2, 144.5, 131.2, 121.6, 52.8, 51.8, 47.0, 45.8, 45.1, 42.3, 41.7, 41.3, 40.5, 39.8, $34.0,\,33.3,\,32.44,\,32.38,\,30.9,\,28.2,\,27.8,\,26.0,\,23.8,\,23.5,\,23.2,\,23$ 21.7, 19.2, 18.2, 17.6. EIMS (70 eV) m/z. 494 [M]+ (95), 435 (87), 262 (40), 203 (100). HREIMS: Calcd for C₃₂H₄₆O₄: 494.3396. Found: 494.3398. Anal. (Table 2).

 $\textbf{2-Formyl-3-oxooleana-1,12-dien-28-oic Acid (15)}. \ \textbf{15} \ was$ prepared from **32** according to the same method as for **14**. The reaction mixture was subjected to flash column chromatography [hexanes-EtOAc (3:1) followed by hexanes-EtOAc (2:1)] to give 15 as an amorphous solid (71%, 84% based on recovered **32**): $[\alpha]^{26}_D$ +26° (c 0.95, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 240 (3.82) nm. IR (KBr): 2948, 2866, 1725, 1701, 1674, 1608 cm⁻¹. ¹H NMR (CDCl₃): δ 10.00 (1H, s), 7.79 (1H, s), 5.36 (1H, t, J = 3.3 Hz), 2.86 (1H, dd, J = 3.8, 13.9 Hz), 1.18, 1.17, 1.15, 1.14, 0.94, 0.92, 0.87 (each 3H, s). 13 C NMR (CDCl₃): δ 203.7, 190.7, 184.3, 165.0, 144.2, 131.2, 121.8, 52.8, 46.8, 45.7, 45.1, 42.3, 41.4, 41.3, 40.5, 39.8, 34.0, 33.2, 32.5, 32.3, 30.9, 28.2, 27.8, 26.0, 23.7, 23.5, 23.0, 21.6, 19.2, 18.2, 17.8. EIMS (70 eV) m/z: 480 [M]⁺ (5.5), 434 (3.1), 419 (3.4), 248 (56), 233 (27), 203 (100). HREIMS: Calcd for C₃₁H₄₄O₄: 480.3240. Found: 480.3237. Anal. (Table 2).

Methyl 2-Carboxy-3-oxooleana-1,12-dien-28-oate (16). (1) From 14: To a solution of 14 (357 mg, 0.72 mmol) in acetone (71 mL) was added Jones reagent (0.5 mL) dropwise in an ice bath. The mixture was stirred in the ice bath for 20 min. After excess Jones reagent was decomposed with MeOH, the acetone was evaporated in vacuo. After water was added to the resultant mixture, the aqueous mixture was extracted with EtOAc three times. The extract was washed with water (three times) and saturated aqueous NaCl solution (three times), dried over anhydrous MgSO₄, and filtered. The filtrate was evaporated in vacuo to give a residue (294 mg). The residue was subjected to flash column chromatography [hexanes-EtOAc (1:1) followed by EtOAc to afford 14 (89 mg) and 16 as a crystalline solid (109 mg; 30%, 39% based on recovered **14**): mp 230-231 °C; $[\alpha]^{26}_D + 85^\circ$ (c 0.61, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 234 (3.78) nm. IR (KBr): 3436, 2946, 2876, 1756,

1722, 1633 cm⁻¹. ¹H NMR (CDCl₃): δ 8.43 (1H, s), 5.36 (1H, t, J = 3.5 Hz), 3.64 (3H, s), 2.90 (1H, dd, J = 3.9, 13.7 Hz), 1.24, 1.21, 1.19, 1.13, 0.94, 0.91, 0.85 (each 3H, s). ¹³C NMR (CDCl₃): δ 209.2, 178.4, 173.4, 165.2, 144.5, 123.3, 121.4, 52.4, 51.8, 47.0, 45.7, 45.5, 42.3, 41.7, 41.1, 40.6, 40.4, 34.0, 33.3, 32.4, 32.3, 30.9, 28.3, 27.8, 26.0, 23.8, 23.5, 23.1, 22.0, 19.0, 18.3, 17.7. EIMS (70 eV) m/z. 510 [M]+ (16), 492 (15), 451 (14), 433 (14), 262 (27), 203 (100). HREIMS: Calcd for C₃₂H₄₆O₅: 510.3345. Found: 510.3347. Anal. (Table 2).

(2) From 17: A solution of 17 (500 mg, 0.95 mmol) in MeOH (29 mL) and aqueous KOH solution (KOH, 2.9 g; water, 10 mL) was heated under reflux for 15 min. After removal of MeOH in vacuo, the mixture was acidified with 5% aqueous HCl solution. It was extracted with EtOAc (three times). The extract was washed with water and saturated aqueous NaCl solution (each three times), dried over MgSO₄, and filtered. The filtrate gave 16 as a crystalline solid (470 mg, 97%). It was used for the next reaction without further purification.

Methyl 2-Methoxycarbonyl-3-oxooleana-1,12-dien-28oate (17). 17 was prepared from 31 by the similar method as for 14. The reaction mixture was subjected to flash column chromatography [hexanes-EtOAc (4:1)] to give 17 as an amorphous solid (83%, 90% based on recovered 31): $[\alpha]^{26}D$ +63° (c 0.78, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 230 (3.97) nm. IR (KBr): 2947, 2866, 1727, 1684, 1624 cm⁻¹. ¹H NMR (CDCl₃): δ 7.73 (1H, s), 5.37 (1H, t, J = 3.5 Hz), 3.79, 3.64 (each 3H, s), 2.90 (1H, dd, J = 3.9, 13.7 Hz), 1.16 (6H, s), 1.15, 1.12, 0.94, 0.91, 0.84 (each 3H, s). 13 C NMR (CDCl₃): δ 201.2, 178.4, 166.0, 164.3, 144.5, 129.2, 121.7, 52.7, 52.4, 51.8, 47.0, 45.9, 45.8, 42.3, 41.8, 41.5, 40.3, 39.5, 34.1, 33.3, 32.4, 32.3, $30.9,\ 28.7,\ 27.8,\ 25.9,\ 23.8,\ 23.6,\ 23.2,\ 21.5,\ 19.4,\ 18.0,\ 17.5.$ EIMS (70 eV) m/z: 524 [M]+ (24), 492 (23), 465 (13), 262 (35), 203 (100). HREIMS: Calcd for C₃₃H₄₈O₅: 524.3502. Found: 524.3494. Anal. (Table 2).

2-Methoxycarbonyl-3-oxooleana-1,12-dien-28-oic Acid (18). A solution of 3 (52 mg, 0.10 mmol) in MeOH (5.2 mL) containing concentrated H₂SO₄ (0.15 mL) was heated under reflux for 30 min. After saturated aqueous NaCl solution was added to the mixture, it was extracted with EtOAc three times. The extract was worked up according to the standard method to give a residue (53 mg). The residue was subjected to flash column chromatography [hexanes-EtOAc (2:1)] to give 18 as an amorphous solid (42 mg, 78%): $[\alpha]^{26}_D +61^{\circ}$ ($c 0.5\bar{6}$, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 230 (3.83) nm. IR (KBr): 3323, 2947, 2866, 1733, 1695, 1622 cm $^{-1}$. ¹H NMR (CDCl₃): δ 7.73 (1H, s), 5.37 (1H, t, J = 3.4 Hz), 3.79 (3H, s), 2.86 (1H, dd, J = 4.1, 13.7 Hz), 1.16, 1.15, 1.14, 1.12, 0.94, 0.92, 0.86 (each 3H, s). ¹³C NMR (CDCl₃): δ 201.1, 184.2, 166.0, 164.2, 144.2, 129.2, $122.0,\ 52.7,\ 52.4,\ 46.9,\ 45.9,\ 45.8,\ 42.2,\ 41.5,\ 41.4,\ 40.3,\ 39.5,$ 34.0, 33.3, 32.5, 32.3, 30.9, 28.7, 27.8, 26.0, 23.7, 23.6, 23.0, 21.4, 19.4, 18.0, 17.7. EIMS (70 eV) m/z: 510 [M]⁺ (2.6), 495 (2.0), 478 (2.5), 432 (3.0), 263 (29), 248 (58), 231 (37), 203 (100). HREIMS: Calcd for C₃₂H₄₆O₅: 510.3345. Found: 510.3344. Anal. (Table 2).

Methyl 2-Aminocarbonyl-3-oxooleana-1,12-dien-28oate (19). A solution of 17 (100 mg, 0.19 mmol) in saturated ammonia MeOH (10 mL) was kept at room temperature overnight. The mixture was evaporated in vacuo to give a solid (108 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (1.5:1)] to give **19** as an amorphous solid (94 mg, 96%): $[\alpha]^{26}_D + 77^{\circ}$ (c 0.60, CHCl₃). UV (EtOH) λ_{max} (log ϵ): 236 (3.91) nm. IR (KBr): 3413, 2943, 2866, 1727, 1689 cm^{-1} . ¹H NMR (CDCl₃): δ 8.45 (1H, brs), 8.27 (1H, s), 5.72 (1H, brs), 5.37 (1H, t, J = 3.4 Hz), 3.64 (3H, s), 2.90 (1H, dd, J = 4.2, 13.9 Hz), 1.17, 1.16, 1.15, 1.14, 0.94, 0.92, 0.84 (each 3H, s). ¹³C NMR (CDCl₃): δ 205.8, 178.4, 169.0, 165.8, 144.3, 121.8, 52.2, 51.8, 47.0, 46.0, 45.7, 42.3, 41.8, 41.2, 40.4, 39.6, 34.1, 33.3, 32.5, 32.3, 30.9, 29.1, 27.8, 26.0, 23.8, 23.6, 23.2, 21.9, 19.4, 18.6, 17.6. EIMS (70 eV) m/z: 509 [M]⁺ (34), 492 (23), 450 (100), 262 (19), 203 (56). HREIMS: Calcd for C₃₂H₄₇O₄N: 509.3505. Found: 509.3500. Anal. (Table 2).

Methyl 1α , 2α -Epoxy-3-oxoolean-12-en-28-oate (22). To a solution of B-13 (223 mg, 0.48 mmol) in 2 N aqueous NaOH solution (1.7 mL) and THF (11 mL) was added a solution of

 $30\%~H_2O_2~(1.4~mL)$ in MeOH (2.8 mL) in an ice bath. The mixture was stirred at room temperature for 4 h. To the mixture were added saturated aqueous NaHSO3 and 5% aqueous NaOH solutions, successively. After removal of THF and MeOH, the resultant mixture was acidified with 6 M aqueous HCl solution. The acidic layer was extracted with CH₂Cl₂ three times. The extract was worked up according to the standard method to give 22 as a crystalline solid (228 mg, 99%). This material was used for the next reaction without further purification. An analytically pure sample was obtained by recrystallization from MeOH as colorless needles: mp 212-213 °C; $[\alpha]^{26}_D$ +157° (c 0.80, CHCl₃). IR (KBr): 2943, 2866, 1727, 1699 cm⁻¹. ¹H NMR (CDCl₃): δ 5.36 (1H, t, J= 3.3 Hz), 3.64 (3H, s), 3.50 (1H, d, J = 4.5 Hz), 3.37 (1H, d, J = 4.5 Hz),2.90 (1H, dd, J = 4.2, 13.9 Hz), 1.21, 1.11, 1.01, 0.97, 0.94, 0.92, 0.80 (each 3H, s). 13 C NMR (CDCl₃): δ 213.0, 178.4, 144.5, 121.8, 64.1, 57.1, 51.8, 47.0, 46.3, 45.9, 45.0, 42.1, 41.7, 40.8, 39.7, 38.8, 34.1, 33.3, 32.5, 32.3, 30.9, 28.2, 28.0, 26.0, 24.0, 23.8, 23.3, 21.1, 19.1, 17.4, 15.1. EIMS (70 eV) m/z. 482 [M] (7.7), 422 (13), 262 (31), 249 (11), 203 (100). HREIMS: Calcd for C₃₁H₄₆O₄: 482.3396. Found: 482.3391.

Methyl 2-Methoxy-3-oxooleana-1,12-dien-28-oate (23). A mixture of 22 (300 mg, 0.62 mmol) and Na (360 mg) in MeOH (36 mL) was heated under reflux for 48 h. After removal of MeOH in vacuo, the resultant mixture was diluted with water and then acidified with 6 M aqueous HCl solution. The aqueous mixture was extracted with a mixture of CH2Cl2 and Et₂O (1:2) three times. The extract was worked up according to the standard method to give a solid (320 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (4:1)] to afford 22 (31 mg) and 23 as an amorphous solid (270 mg; 87%, 98% based on recovered 22): UV (EtOH) λ_{max} (log ε): 266 (3.77) nm. IR (KBr): 2946, 2866, 1727, 1682, 1621 cm⁻¹. ¹H NMR (CDCl₃): δ 5.96 (1H, s), 5.36 (1H, t, J = 3.5Hz), 3.64, 3.55 (each 3H, s), 2.90 (1H, dd, J = 4.1, 13.7 Hz), 1.17 (6H, s), 1.16, 1.13, 0.93, 0.90, 0.81 (each 3H, s). ¹³C NMR (CDCl₃): δ 200.1, 178.4, 149.0, 144.6, 126.3, 121.9, 54.9, 53.2, 51.8, 47.0, 45.9, 45.4, 43.3, 42.3, 41.7, 40.2, 38.4, 34.0, 33.3, 32.6, 32.5, 30.9, 28.5, 27.8, 26.0, 23.81, 23.76, 23.2, 22.0, 20.4, 19.2, 17.4. EIMS (70 eV) m/z. 496 [M]⁺ (80), 436 (21), 328 (19), 262 (36), 203 (100). HREIMS: Calcd for C₃₂H₄₈O₄: 496.3553. Found: 496.3544.

Methyl 2-Hydroxy-3-oxooleana-1,12-dien-28-oate (24). A suspension of 23 (100 mg, 0.20 mmol) in 3 M aqueous HCl solution (3 mL) and AcOH (3 mL) was heated under reflux for 5 h. The mixture was neutralized with saturated aqueous Na₂CO₃ solution. The mixture was extracted with CH₂Cl₂ three times. The extract was worked up according to the standard method to give a solid (90 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (5:1)] to afford 24 as an amorphous solid (78 mg, 81%): UV (EtOH) λ_{max} (log ϵ): 272 (3.63) nm. IR (KBr): 3426, 2939, 2870, 1725, 1667, 1648 cm $^{-1}$. ¹H NMR (CDCl₃): δ 6.35 (1H, s), 5.93 (1H, brs), 5.34 (1H, t, J = 3.5 Hz), 3.63 (3H, s), 2.89 (1H, dd, J = 4.0, 13.7 Hz), 1.22 (6H, s), 1.13, 1.12, 0.94, 0.91, 0.80 (each 3H, s). ¹³C NMR (CDCl₃): δ 201.3, 178.4, 144.4, 143.9, 128.4, 122.0, 54.1, 51.8, 47.0, 45.9, 44.1, 43.3, 42.2, 41.6, 40.2, 38.7, 34.1, 33.3, 32.7, 32.5, 30.9, 27.8, 27.4, 26.1, 23.8, 23.6, 23.2, 22.0, 19.8, 18.9, 17.5. EIMS (70 eV) m/z. 482 [M]+ (26), 446 (68), 422 (25), 262 (35), 203 (100). HREIMS: Calcd for C₃₁H₄₆O₄: 482.3396. Found: 482.3387.

Methyl 2-Hydroxymethylene-3-oxoolean-12-en-28-oate (25). 19 To a stirred mixture of **B-3** (1084 mg, 2.31 mmol) and ethyl formate (97%) (707 mg, 9.26 mmol) in benzene (12 mL) was added NaOMe (501 mg, 9.27 mmol). The mixture was stirred at room temperature for 1 h. After the mixture was washed with 5% aqueous HCl solution twice, it was worked up according to the standard method to give 25 as an amorphous solid (1095 mg, 95%). This material was used for the next reaction without further purification. An analytically pure sample was obtained by flash column chromatography [hexanes-EtOAc (7:1)] and subsequent recrystallization from MeOH as colorless needles: mp 199–201 °C. UV (EtOH) λ_{max} $(\log \epsilon)$: 296 (3.94) nm. IR (KBr): 3426, 2943, 2862, 1725, 1637,

1588 cm⁻¹. ¹H NMR (CDCl₃): δ 14.92 (1H, d, J= 3.1 Hz), 8.58 (1H, d, J = 3.1 Hz), 5.35 (1H, t, J = 3.7 Hz), 3.64 (3H, s), 2.90(1H, dd, J = 4.2, 13.6 Hz), 2.29 (1H, d, J = 14.4 Hz), 1.92 (1H, d, J = 14.4 Hz), 1.20, 1.16, 1.12, 0.94 (each 3H, s), 0.91 (6H, s), 0.80 (3H, s). 13 C NMR (CDCl₃): δ 190.9, 188.6, 178.4, 144.0, 122.3, 106.0, 52.3, 51.8, 47.0, 46.0, 45.9, 42.0, 41.6, 40.3, 39.4, 39.3, 36.5, 34.1, 33.3, 32.5, 32.1, 30.9, 28.6, 27.9, 25.9, 23.8, 23.6, 23.3, 21.1, 19.7, 16.8, 14.7. EIMS (70 eV) m/z: 496 [M] (4.4), 437 (23), 262 (38), 233 (20), 203 (100). HREIMS: Calcd for C₃₂H₄₈O₄: 496.3553. Found: 496.3550.

Methyl Isoxazolo[4,5-b]olean-12-en-28-oate (26). A mixture of 25 (994 mg, 2.0 mmol), hydroxylamine hydrochloride (1391 mg, 20 mmol) in water (1.8 mL) and EtOH (48 mL) was heated under reflux for 1 h. After EtOH was removed in vacuo, EtOAc was added to the resultant mixture. The EtOAc layer was washed with water (three times) and saturated aqueous NaCl solution (three times), dried over MgSO₄, and filtered. The filtrate gave a solid (1086 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (6:1) followed by hexanes-EtOAc (5:1)] to give 26 as an amorphous solid (934 mg, 86%): UV (EtOH) $\bar{\lambda}_{max}$ (log ϵ): 228 (3.65) nm. IR (KBr): 2940, 2864, 1725 cm⁻¹. ¹H NMR (CDCl₃): δ 7.98 (1H, s), 5.34 (1H, t, J = 3.5 Hz), 3.63 (3H, s), 2.89 (1H, dd, J = 4.4, 13.7 Hz), 2.42 (1H, d, J = 15.1 Hz), 1.30, 1.21, 1.15, 0.93, 0.90, 0.88, 0.79 (each 3H, s). 13 C NMR (CDCl₃): δ 178.4, 173.2, 150.4, 144.0, 122.3, 109.0, 53.7, 51.7, 46.9, 46.3, 46.0, 42.0, 41.6, 39.5, 38.9, 35.5, 34.9, 34.0, 33.3, 32.5, 32.1, 30.9, 29.0, 27.9, 25.9, 23.8, 23.5, 23.2, 21.6, 19.0, 16.7, 15.4. EIMS (70 eV) m/z. 493 [M]⁺ (11), 434 (18), 262 (28), 249 (16), 203 (100). HREIMS: Calcd for C₃₂H₄₇O₃N: 493.3556. Found: 493.3556.

Methyl 2-Cyano-3-oxoolean-12-en-28-oate (27). To a stirred solution of 26 (887 mg, 1.80 mmol) in Et₂O (50 mL) and MeOH (25 mL) was added NaOMe (3.2 g) in an ice bath. The mixture was stirred at room temperature for 1 h. The mixture was diluted with a mixture of CH₂Cl₂ and Et₂O (1:2) (50 mL). After the extract was washed with 5% aqueous HCl solution, it was worked up according to the standard method to afford 27 as an amorphous solid (879 mg, 99%). This material was used for the next reaction without further purification. An analytically pure sample was obtained by flash column chromatography [hexanes-EtOAc (5:1)] as an amorphous solid: UV (EtOH) $\lambda_{\rm max}$ (log ϵ): 238 (3.88) nm. IR (KBr): 2946, 2870, 2202, 1724, 1633 cm $^{-1}$. ¹H NMR of major tautomer **27a** (CDCl₃): δ 6.15 (1H, brs), 5.31 (1H, t, J = 3.6 Hz), 3.63 (3H, s), 2.88 (1H, dd, J = 4.0, 13.6 Hz), 2.09 (1H, d, J = 15.0)Hz), 1.16, 1.13, 1.07, 0.95, 0.93, 0.90, 0.76 (each 3H, s). EIMS (70 eV) m/z. 493 [M]⁺ (6.3), 434 (17), 262 (19), 249 (20), 203 (100). HREIMS: Calcd for $C_{32}H_{47}O_3N$: 493.3556. Found: 493,3548

Methyl 2-Hydroxymethylene-3-oxours-12-en-28-oate (28). 23 28 was prepared from B-4 according to the same method as for 25 to give an amorphous solid (quantitative). This material was used for the next reaction without further purification. An analytically pure sample was obtained by flash column chromatography [hexanes-EtOAc (7:1)] and subsequent recrystallization from MeOH as colorless needles: mp 170–171 °C. UV (EtOH) λ_{max} (log ϵ): 294 (3.86) nm. IR (KBr): 3426, 2947, 2921, 2866, 1727, 1637, 1590 cm⁻¹. ¹H NMR (CDCl₃): δ 14.91 (1H, brs), 8.57 (1H, s), 5.31 (1H, t, J = 3.7Hz), 3.62 (3H, s), 2.31 (1H, d, J = 14.4 Hz), 2.27 (1H, d, J = 14.4 Hz) 12.5 Hz), 1.95 (1H, d, J = 14.4 Hz), 1.19, 1.12, 1.10 (each 3H, s), 0.96 (3H, d, J = 6.0 Hz), 0.92 (3H, s), 0.87 (3H, d, J = 6.6Hz), 0.81 (3H, s). 13 C NMR (CDCl₃): δ 191.0, 188.5, 178.2, 138.4, 125.6, 106.0, 53.2, 52.3, 51.7, 48.4, 45.7, 42.4, 40.3, 39.7, $39.5,\ 39.3,\ 39.1,\ 36.8,\ 36.4,\ 32.4,\ 30.9,\ 28.7,\ 28.2,\ 24.4,\ 23.7,\ 28.2,\ 24.4,\$ 23.6, 21.4, 21.1, 19.7, 17.2, 17.0, 14.8. EIMS (70 eV) m/z. 496 [M]⁺ (11), 437 (15), 262 (80), 233 (41), 203 (100). HREIMS: Calcd for C₃₂H₄₈O₄: 496.3553. Found: 496.3547.

Methyl Isoxazolo[4,5-b]urs-12-en-28-oate (29). 29 was prepared from 28 according to the same method as for 26 to give an amorphous solid (84%): UV (EtOH) λ_{max} (log ϵ): 228 (3.70) nm. IR (KBr): 2969, 2922, 2870, 1725 cm⁻¹. ¹H NMR (CDCl₃): δ 7.98 (1H, s), 5.31 (1H, t, J = 3.4 Hz), 3.62 (3H, s), 2.46 (1H, d, J = 15.0 Hz), 2.27 (1H, d, J = 11.1 Hz), 1.31, 1.22, 1.10 (each 3H, s), 0.96 (3H, d, J = 6.3 Hz), 0.90 (3H, s), 0.88 (3H, d, J = 6.3 Hz), 0.81 (3H, s). ¹³C NMR (CDCl₃): δ 178.2, 173.2, 150.4, 138.4, 125.5, 109.1, 53.7, 53.2, 51.7, 48.3, 46.3, 42.3, 39.7, 39.3, 39.1, 38.8, 36.8, 35.8, 34.9, 32.4, 30.9, 29.1, 28.3, 24.4, 23.7, 23.5, 21.6, 21.4, 19.0, 17.2, 16.9, 15.6. EIMS (70 eV) m/z: 493 [M]⁺ (9.1), 434 (20), 262 (65), 249 (33), 203 (100). HREIMS: Calcd for $C_{32}H_{47}O_3N$: 493.3556. Found:

Methyl 2-Cyano-3-oxours-12-en-28-oate (30). 30 was prepared from 29 according to the same method as for 27 to give an amorphous solid (quantitative). This material was used for the next reaction without further purification. An analytically pure sample was obtained by flash column chromatography [hexanes-EtOAc (5:1)] as an amorphous solid: UV (EtOH) λ_{max} (log ϵ): 238 (3.93) nm. IR (KBr): 2947, 2870, 2203, 1724, 1631 cm⁻¹. ¹H NMR of major tautomer **30a** (CDCl₃): δ 5.92 (1H, brs), 5.28 (1H, t, J = 3.5 Hz), 3.61 (3H, s), 2.26 (1H, d, J = 11.0 Hz), 2.13 (1H, d, J = 15.0 Hz), 1.16, 1.13, 1.08, 1.07, 0.96 (each 3H, s), 0.95, 0.77 (each 3H, d, J = 6.3 Hz). EIMS (70 eV) m/z: 493 [M]+ (6.8), 434 (19), 262 (62), 249 (44), 203 (100). HREIMS: Calcd for C₃₂H₄₇O₃N: 493.3556. Found: 493.3558

Methyl 3-Hydroxy-2-methoxycarbonyloleana-2,12-dien-**28-oate** (**31**). A mixture of **B-3** (2.0 g, 4.27 mmol) and 1.8 M DMF solution of methoxymagnesium methyl carbonate (Stiles' reagent) (20 mL, 36 mmol) was heated under reflux for 2 h while a slow stream of N2 was bubbled through the mixture with a pipet. To the mixture were added 5% aqueous HCl solution and EtOAc. The aqueous layer was extracted with EtOAc (three times). The combined organic layers were washed with water (three times) and saturated aqueous NaCl solution (three times), dried over $MgSO_4$, and filtered. The filtrate was evaporated in vacuo to give a solid (2.26 g). To a solution of the solid in THF (30 mL) was added excessive amount of ethereal diazomethane. The mixture was kept at room temperature for 10 min. The mixture was evaporated in vacuo to give a solid (2.38 g). The solid was subjected to flash column chromatography [hexanes-EtOAc (7:1)] to give B-3 (330 mg) and **31** as crystals (1.66 g; 74%, 89% based on recovered **B-3**): mp 160–162 °C. UV (EťOH) $\lambda_{\rm max}$ (log ϵ): 262 (4.01) nm. IR (KBr): 2948, 2858, 1737, 1660, 1615 cm⁻¹. ¹H NMR (CDCl₃): δ 12.51 (1H, s), 5.33 (1H, t, J = 3.7 Hz), 3.74, 3.63 (each 3H, s), 2.89 (1H, dd, J = 4.2, 13.9 Hz), 2.35 (1H, d, J = 15.7 Hz), 1.18, 1.14, 1.10, 0.94 (each 3H, s), 0.91 (6H, s), 0.78 (3H, s). ¹³C NMR (CDCl₃): δ 178.5, 177.9, 174.2, 143.8, 122.6, 94.3, 52.5, 51.8, 51.7, 47.0, 46.13, 46.09, 42.0, 41.7, 39.4, 38.6, 38.4, 35.7, 34.1, 33.3, 32.6, 32.1, 31.0, 28.8, 27.9, 26.0, 23.8, 23.6, 23.3, 20.4, 19.8, 16.8, 15.1. EIMS (70 eV) m/z. 526 [M]⁺ (0.6), 494 (5.6), 479 (2.5), 466 (1.6), 435 (13), 262 (28), 203 (100). HREIMS: Calcd for C₃₃H₅₀O₅: 526.3658. Found: 526.3658.

2-Hydroxymethylene-3-oxoolean-12-en-28-oic Acid (32). To a stirred mixture of oleanonic acid (**B-1**)¹⁰ (540 mg, 1.19 mmol) and ethyl formate (97%) (357 mg, 4.66 mmol) in THF (12 mL) was added NaOMe (258 mg, 4.78 mmol). The mixture was stirred at room temperature overnight. The mixture was acidified with 10% aqueous HCl solution. The mixture was extracted with EtOAc three times. The extract was worked up according to the standard method to give a solid (600 mg). The solid was subjected to flash column chromatography [hexanes-EtOAc (5:1) followed by hexanes-EtOAc (4:1)] to afford **B-1** (168 mg) and **32** as a crystalline solid (260 mg; 45%, 66% based on recovered **B-1**): mp 200-203 °C dec. UV (EtOH) λ_{max} (log ϵ): 292 (3.93) nm. IR (KBr): 2946, 2654, 1732, 1694, 1644, 1587 cm⁻¹. 1 H NMR (CDCl₃): δ 14.91 (1H, brs), 8.59 (1H, s), 5.34 (1H, t, J = 3.5 Hz), 2.86 (1H, dd, J = 4.5, 13.9 Hz), 2.29 (1H, d, J = 14.6 Hz), 1.93 (1H, d, J = 14.6 Hz), 1.19, 1.16, 1.10, 0.94, 0.92, 0.91, 0.82 (each 3H, s). 13 C NMR (CDCl₃): δ 190.7, 188.8, 184.7, 143.8, 122.6, 105.9, 52.2, 46.8, 46.0, 45.9, 41.9, 41.2, 40.2, 39.34, 39.30, 36.5, 34.0, 33.3, 32.6, 32.0, 30.9, 28.6, 27.8, 25.9, 23.7, 23.5, 23.1, 21.0, 19.6, 17.0, 14.6. EIMS (70 eV) m/z. 482 [M]⁺ (1.8), 438 (2.7), 436 (3.6), 248 (77), 203 (100). HREIMS: Calcdfor C₃₁H₄₆O₄: 482.3396. Found: 482.3392.

Evaluation Methods. 1. Reagents. Recombinant mouse IFN-γ (LPS content, <10 pg/mL) was purchased from Genzyme

(Cambridge, MA). All other chemicals were purchased from Sigma Chemical Co. (St. Louis, MO). Inhibitory test compounds were dissolved in DMSO before addition to cell cultures; final concentrations of DMSO were 0.1% or less. Controls with DMSO alone were run in all cases.

- 2. Cell Culture. To obtain primary macrophages, female CD-1 mice, 5-10 weeks of age (Charles River Breeding Laboratories, Wilmington, MA), were injected intraperitoneally with 2 mL of 4% thioglycollate broth (Difco Laboratories, Detroit, MI). Four days after injection, peritoneal macrophages were harvested and processed according to Nathan's procedure. 7b Cells were seeded in 96-well plates at 2×10^5 cells/well and incubated for 48 h with 20 ng/mL IFN- γ in the presence or absence of inhibitory test compounds.
- 3. Measurement of NO Production in Mouse Macrophages. Nitrite accumulation was used as an indicator of NO production in the medium and was assayed by the Griess reaction. Ta Griess reagent (100 μ L) was added to 100 μ L of each supernatant from IFN- γ or inhibitory test compound-treated cells in triplicate. The protein determination was performed by Bradford protein assay. The plates were read at 550 nm against a standard curve of sodium nitrite.

Acknowledgment. We thank Drs. Carl Nathan and Qiao-wen Xie for expert advice on the preparation of macrophages and the nitric oxide assay. We also thank Dr. Steven Mullen (University of Illinois) for the mass spectra and Professor David A. Evans and Mr. Brett D. Allison (Harvard University) for the optical rotation measurements. This investigation was supported by funds from NIH Grant 1 R01-CA78814; the Norris Cotton Cancer Center; U.S. Department of Defense Grants DAMD17-96-1-6163, DAMD17-98-1-8604, and DAMD17-99-1-9168; the Oliver and Jennie Donaldson Charitable Trust: the National Foundation for Cancer Research; and a Zenith Award from the Alzheimer's Association. M.B.S. is an Oscar M. Cohn Professor, F.G.F. is an Oscar M. Cohn Scholar, and Y.W. is a Howard Hughes Medical Institute Predoctoral Fellow.

References

(1) (a) Connolly, J. D.; Overton, K. H. The Triterpenoids. In Chemistry of Terpenes and Terpenoids, Newman, A. A., Ed.; Academic Press: New York, 1972; pp 207–287. (b) Nakanishi, K.; Ito, S. Biosynthesis of oleanene and ursene triterpenes in tissue culture of *Isodon japonicus*. In *Natural Products Chemistry*, Vol. 3; Nakanishi, K., Goto, T., Ito, S., Natori, S., Nozoe, S., Eds.; Kodansha: Tokyo, 1983; pp 185–187.

(2) Devon, T. K.; Scott, A. I. The Terpenes. In *Handbook of Naturally*

Occurring Compounds, Vol. 2; Academic Press: New York, 1972; pp 281–384.

(a) Nishino, H.; Nishino, A.; Takayasu, J.; Hasegawa, T.; Iwashima, A.; Hirabayashi, K.; Iwata, S.; Shibata, S. Inhibition of the tumor-promoting action of 12-O-tetradecanoylphorbol-13acetate by some oleanane-type triterpenoid compounds. Cancer Res. 1988, 48, 5210-5215. (b) Hirota, M.; Mori, T.; Yoshio, M.; Iriye, R. Suppression of tumor promoter-induced inflammation of mouse ear by ursolic acid and 4,4-dimethylcholestane derivatives. *Agric. Biol. Chem.* **1990**, *54*, 1073–1075. (c) Yu, L.; Ma, R.; Wang, Y.; Nishino, H.; Takayasu, J.; He, W.; Chang, M.; Zhen, J.; Liu, W.; Fan, S. Potent anti-tumorigenic effect of tubeimoside 1 isolated from the bulb of Bolbostemma paniculatum (Maxim) Franquet. Int. J. Cancer 1992, 50, 635-638. (d) Umehara, K.; Takagi, R.; Kuroyanagi, M.; Ueno, A.; Taki, T.; Chen, Y.-J. Studies on differentiation-inducing activities of terpenes. *Chem. Pharm. Bull.* **1992**, *40*, 401–405. (e) Singh, G. B.; Singh, S.; Bani, S.; Gupta, B. D.; Banerjee, S. K. Antiinflammatory activity of oleanolic acid in rats and mice. J. Pharm. Pharmacol. 1992, 44, 456-458. (f) Huang, M.-T.; Ho, C.-T.; Wang, Z. Y.; Ferrao, T.; Lou, Y.-R.; Stauber, K.; Ma, W.; Georgiadis, C.; Laskin, J. D.; Conney, A. H. Inhibition of skin tumorigenesis by rosemary and its constituents carnosol and ursolic acid. Cancer Res. 1994, 54, 701-708. (g) Liu, J.; Liu, Y.; Mao, Q.; Klaassen, C. D. The effects of 10 triterpenoid compounds on experimental liver injury in mice. Fundam. Appl. Toxicol. **1994**, 22, 34–40. (h) Liu, J. Pharmacology of oleanolic acid and ursolic acid. J. Ethnopharmacol. 1995, 49, 57-68 and references therein.

- (4) After we started this project, the following systematic studies of structure-activity relationships based on chemical modification of oleanane triterpenoids were reported: (a) Sakurawi, K.; Yasuda, F.; Tozyo, T.; Nakamura, M.; Sato, T.; Kikuchi, J.; Terui, Y.; Ikenishi, Y.; Iwata, T.; Takahashi, K.; Konoike, T.; Mihara, S.; Fujimoto, M. Endothelin receptor antagonist triterpenoid, myriceric acid A, isolated from *Myrica cerifera*, and structure activity relationships of its derivatives. Chem. Pharm. Bull. 1996, 44, 343–351. (b) Kashiwada, Y.; Wang, H.-K.; Nagao, T.; Kitanaka, S.; Yasuda, I.; Fujioka, T.; Yamagishi, T.; Cosentino, L. M.; Kozuka, M.; Okabe, H.; Ikeshiro, Y.; Hu, C.-Q.; Yeh, E.; Lee, K.-H. Anti-AIDS agents. 30. Anti-HIV activity of oleanolic acid, pomolic acid, and structurally related triterpenoids. J. Nat.
- *Prod.* **1998**, *61*, 1090–1095. (a) Moncada, S.; Palmer, R. M. J.; Higgs, E. A. Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol. Rev.* **1991**, *43*, 109–141. (b) Nathan, C. F.; Xie, Q.-W. Regulation of biosynthesis of nitric oxide. J. Biol. Chem. 1994, 269, 13725 13728. (c) Anggard, E. Nitric oxide: mediator, murderer, and medicine. *Lancet* **1994**, *343*, 1199–1206. (d) Nathan, C. F.; Xie, Q.-W. Nitric oxide synthases: roles, tolls, and controls. Cell 1994, 78, 915-918.
- (a) Sporn, M. B.; Roberts, A. B. Peptide growth factors and inflammation, tissue repair, and cancer. *J. Clin. Invest.* **1986**, *78*, 329–332. (b) Ohshima, H.; Bartsch, H. Chronic infections and inflammatory processes as cancer risk factors: possible role of nitric oxide in carcinogenesis. Mutat. Res. 1994, 305, 253-
- (a) Ding, A.; Nathan, C. F.; Graycar, J.; Derynck, R.; Stuehr, D. J.; Srimal, S. Macrophage deactivating factor and transforming growth factors- β 1, - β 2, and - β 3 inhibit induction of macrophage nitrogen oxide synthesis by IFN-γ. J. Immunol. 1990, 145, 940-944. (b) Bogdan, C.; Paik, J.; Vodovotz, Y.; Nathan, C. Contrasting mechanisms for suppression of macrophage cytokine release by transforming growth factor- β and interleukin-10. *J. Biol. Chem.* **1992**, *267*, 23301–23308.
- (8) Synthesis of 20 and 21 was already reported: Honda, T.; Gribble, G. W. Design and synthesis of 23,24-dinoroleanolic acid derivatives, novel triterpenoid-steroid hybrid molecules. J. Org. Chem. **1998**, *63*, 4846–4849.
- (9) Brieskorn, C. H.; Seifert, M. Methylgruppen-Umlagerungen an Triterpenoiden, 2. Mitt. Aromatisierung des Ringes A. Arch. Pharm. **1982**, 315, 846–851.
 (10) Simonsen, J.; Ross, W. C. J. Oleanolic acid. In *The Terpenes*,
- Cambridge University: Cambridge, 1957; Vol. 5, pp 221–285.
 (11) Sharpless, K. B.; Lauer, R. F.; Teranishi, A. Y. Electrophilic and nucleophilic organoselenium reagents. New routes to α,βunsaturated carbonyl compounds. J. Am. Chem. Soc. 1973, 95, 6137 - 6139
- (12) DDQ oxidation of B-3 in benzene (reflux) did not give B-13 at
- (13) Dean, P. D. G. Halogenolysis of methyl glycyrrhetate with
- lithium iodide-dimethylformamide. *J. Chem. Soc. (C)* **1965**, 6655.

 (14) Begum, S.; Adil, Q.; Siddiqui, B. S.; Siddiqui, S. Symptomics from preprint 2β -hydroxyursolic acid and other ursane analogues from ursonic acid. *Aust. J. Chem.* **1993**, *46*, 1067–1071.
- (15) Simonsen, J.; Ross, W. C. J. Ursolic acid. In *The Terpenes*;
 Cambridge University: Cambridge, 1957; Vol. 5, pp 114–135.
 (16) Kurata, Y.; Hirota, H.; Honda, T.; Takahashi, T. Preparation of
- a tricyclic A-ring analog of quassin. Chem. Pharm. Bull. 1987, *35*, 837–840.
- (17) In addition to 5, these conditions also gave the completely demethylated product 4 (yield, 12%; 18% based on recovered 23) and the other partially demethylated product 24 (yield, 16%; 23% based on recovered 23) from 23.
- (18) Shaw, J. I.; Stevenson, R. 4-Bromo- and 4-chloro-cholest-4-en-3-one. J. Chem. Soc. 1955, 3549-3551.

- (19) Zaprutko, L. Triterpenoids. Part 9. [1] Structure elucidation of a new sodium dichromate oxidation product of methyl oleanolate and some of its derivatives. Pol. J. Chem. 1995, 69, 1003-1012.
- ¹H and ¹³C NMR of 25, 28, and 32 in CDCl₃ showed that they are the single tautomer as depicted in Schemes 4 and 6 (see Experimental Section).
- (21) Clinton, R. O.; Manson, A. J.; Stonner, F. W.; Neumann, H. C.; Christiansen, R. G.; Clarke, R. L.; Ackerman, J. H.; Page, D. F.; Dean, J. W.; Dickinson W. B.; Carabateas, C. Steroidal[3,2-c]pyrazoles. II. Androstanes, 19-norandrostanes and their unsaturated analogs. J. Am. Chem. Soc. 1961, 83, 1478-1491.
- Johnson, W. S.; Shelberg, W. E. A plan for distinguishing between some five- and six-membered ring ketones. J. Am. Chem. Soc. 1945, 67, 1745-1754.
- (23) Glen, A. T.; Lawrie, W.; McLean, J.; Younes, M. E.-G. Triterpenoid constituents of rose-bay willow-herb. J. Chem. Soc. (C) **1967**, 510-515.
- Liotta, D.; Barnum, C.; Puleo, R.; Zima, G.; Bayer, C.; Kezar, H. S., III. A simple method for the efficient synthesis of unsaturated β -dicarbonyl compounds. J. Org. Chem. 1981, 46, 2920-2923.
- (25) Finkbeiner, H. L.; Stiles, M. Chelation as a driving force in organic reactions. IV. Synthesis of α -nitro acids by control of the carboxylation-decarboxylation equilibrium. J. Am. Chem. Soc.
- **1963**, *85*, 616–622. (26) Albert, A. Table and discussion: electronic effects in molecules (Hammett and Taft sigma values). Selective Toxicity, 7th ed.; Chapman and Hall: London, 1985; pp 644-649.
- Gagne, D.; Pons, M.; Philibert, D. RU38486: a potent antiglucocorticoid in vitro and in vivo. J. Steroid Biochem. 1985, 23, 247-
- Narayanan, C. R.; Natu, A. A. Synthesis of some bridged triterpene ethers. J. Org. Chem. 1974, 39, 2639-2641 and references therein.
- Younes, M. E.-G. Chemical examination of local plants. XIV. triterpenoids from the leaves of Egyptian Callistemon lanceolatus. Aust. J. Chem. 1975, 28, 221-224.
- Huneck, S. Triterpene-IV. Die Triterpensäuren des Balsams von Liquidambar orientalis Miller. Tetrahedron 1963, 19, 479–482.
- (31) Huneck, S.; Snatzke, G. Triterpene, IX. Über die Triterpene aus der Rinde von Sambucus nigra L. und die Darstellung von 3-epi-Ursolsäure. Chem. Ber. 1965, 98, 120-125.
- Kitagawa, I.; Kitazawa, K.; Yoshioka, I. Photochemical transformation leading to eupteleogenin-I. Introduction of epoxylactone system. Tetrahedron 1972, 28, 907-921.
- Kuwada, S.; Matsukawa, T. Ursolic acid. II. Oxidation of ursolic acid and its derivatives with chromic acid. J. Pharm. Soc. Jpn. (Yakugaku Zasshi) 1933, 53, 593-612.
- (34) Sahu, N. P.; Mahato, S. B.; Chakravarti, R. N. Effect of Raney nickel on some dihydroxy triterpenoids in high boiling solvents. J. Indian Chem. Soc. 1973, 50, 771–773.
- (35) Djerassi, C.; Thomas, D. B.; Livingston, A. L.; Thompson, C. R. Terpenoids. XXXI. The structure and stereochemistry of medicagenic acid. J. Am. Chem. Soc. 1957, 79, 5292-5297.
- Sundararamaiah, T.; Ramraj, S. K.; Rao, K. L.; Bai, V. V. Synthesis of A-aza triterpenes-I: A-aza triterpenes from methyl oleanonate, methyl betulonate and lupenone. J. Indian Chem. Soc. 1976, 53, 664-665.
- Finlay, H. J.; Honda, T.; Gribble, G. W.; Danielpour, D.; Benoit, N. E.; Suh, N.; Williams, C.; Sporn, M. B. Novel A-ring cleaved analogs of oleanolic and ursolic acids which affect growth regulation in NRP.152 prostate cells. Bioorg. Med. Chem. Lett. **1997**, 7, 1769-1772.

JM000008J