See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/44625033

## Rough around the Edges: The Inflammatory Response of Microglial Cells to Spiky Nanoparticles

**ARTICLE** in ACS NANO · MAY 2010

Impact Factor: 12.88 · DOI: 10.1021/nn100776z · Source: PubMed

**CITATIONS** 

29

READS

55

### **3 AUTHORS**, INCLUDING:



Alexandre Albanese

Massachusetts Institute of Technology

11 PUBLICATIONS 979 CITATIONS

SEE PROFILE



Edward A. Sykes

University of Toronto

13 PUBLICATIONS 330 CITATIONS

SEE PROFILE

# Rough around the Edges: The **Inflammatory Response of Microglial** Cells to Spiky Nanoparticles

Alexandre Albanese, Edward A. Sykes, and Warren C. W. Chan\*

Institute of Biomaterials and Biomedical Engineering, Donnelly Centre for Cellular and Biomolecular Research, Department of Chemical Engineering, Materials Science and Engineering, and Chemistry, University of Toronto, Toronto, Ontario M5S 3G9, Canada

**ABSTRACT** The versatility of nanoparticle design has established nanotechnology as a potential "onestop solution" to many biological and medical applications. The capacity to control nanoparticle size, shape, and surface chemistry has enabled their use as imaging contrast agents or carriers for drugs and other compounds. However, concerns of nanoparticle toxicity have surfaced that could limit their clinical translation. In order to overcome this challenge, researchers are starting to characterize how particle properties influence their interactions with biological systems. By identifying the specific nanoparticle parameters responsible for toxicity, it may be possible to engineer safer and nontoxic nanoparticles.

t the most basic level, a nanoparticle's geometric and surface design can change its cellular uptake, accumulation, biological response, and toxicity. Over the past two decades, substantial work has been conducted to understand the mechanism and capacity of cellnanoparticle interactions. Arising from this research, it is now generally accepted that spherical nanoparticles with sub-100-nm diameters can be readily internalized by cells and will preferentially accumulate within the cell in the 40-60 nm size range.<sup>1,2</sup> The signaling response triggered by nanoparticles is usually related to the recognition of ligands on the nanoparticle surface by the cell's receptors. These preliminary studies have also determined that rod and spherical morphologies can enhance the rate of nanoparticle uptake,3 while cationic surface chemistries yield enhanced cellular uptake over their neutral and anionic counterparts.4,5

Researchers have moved beyond in vitro cell culture into animal models in order to study the details of nanoparticle interactions. Studying the consequences of nanoparticle presence in each recipient animal will be more intricate and time-

Studying the consequences of nanoparticle presence in each recipient animal will be more intricate and timeconsuming than current in vitro cell studies.

© 2010 American Chemical Society

\*Address correspondence to warren.chan@utoronto.ca.

Published online May 25, 2010.

See the accompanying Article by Hutter

10.1021/nn100776z

et al. on p 2595.

of cancer cell-specific surface markers (e.g., EGFR, TfR, HER2) have provided nanotechnologists with specific objectives for their design parameters.8,9 Such discoveries could not have occurred using cell culture models alone. These tumor-targeting nanoparticles have improved diagnostic sensitivity and therapeutic efficiency and have spawned a number of products that are now FDA-approved or pending approval.8 Nanotoxicity. An important but controversial issue regarding nanotechnology developments is the potential toxicity of nanoparticles. Heavy metals and other compounds of known toxicity are typically used to achieve the desirable optical, electrical, and magnetic properties of nanoparticles. The bundling of these compounds into a compact nanoparticle could affect their delivery into organs and cells. Researchers have shown that in vivo biodistri-

bution is dependent on size and surface

chemistry<sup>10</sup> and that nanoparticle clearance via the kidney occurs when their size

consuming than current in vitro cell stud-

ies. It has already become apparent that

ties in vitro and in vivo. For example, the

breakdown of CdSe quantum dots led to

significant cell death in cultured liver hepa-

tocytes<sup>6</sup> while showing no signs of toxicity

in rat models.<sup>7</sup> Under these circumstances,

animal models are likely more relevant in

determining toxicity and optimizing design

of nanostructures for human applications.

To illustrate, one of the most studied nano-

particle applications is for targeting and de-

agents into tumors. The discovery of unique

properties within the tumor microenviron-

ment (i.e., poor lymphatic drainage and im-

mature vasculature) and the identification

livering chemotherapeutic or contrast

nanomaterials may exhibit different proper-

is below 6 nm.11 Elucidating the complex nanoparticle-biological interactions requires characterization of transport kinetics, clearance, and variations in gene expression in response to nanoparticle exposure in cell culture and animal models. Generally, nanoparticles may cause cell death in a number of ways including the production of reactive oxygen species that could modify protein function or DNA structure, which could ultimately alter the state, function, and response of cells.12 While there have been significant research activities on this topic, it has been somewhat frustrating for researchers not to be able to say conclusively, "nanoparticle parameter(s) x causes effect y." Unlike small organic molecules, nanoparticles can be engineered controllably with welldefined geometric and chemical properties. By systematically evaluating a single parameter for each experiment, one should be able to establish precise correlations between nanoparticle design and biological interactions.

Gold Nanoparticles as a Model. Spherical gold nanoparticles are perhaps the most studied nanoparticles for biological applications. They have been used as probes for imaging and in vitro diagnostics 13 and hyperthermia therapy<sup>14</sup> and can be used as carriers for a variety of smallmolecule drugs such as siRNA<sup>15</sup> or doxorobucin.16 Studies on gold nanoparticle toxicity and their interactions with blood components, cells, and tissues are a crucial step in their translation into the clinic. Beyond their potential use in medicine, gold nanoparticles are excellent models for characterizing how the size, shape, and surface chemistry of nanostructures impact cell behavior. These nanoparticles are easy to synthesize in a wide variety of sizes and shapes, their surface can be modified with a wide range of chemistries, and they can be quantified using conventional techniques such as atomic emission

While there have been significant research activities on the potential toxicity of nanoparticles, it has been frustrating for researchers not to be able to say conclusively, "nanoparticle parameter(s) x causes effect y."

spectroscopy and ultraviolet/visible spectrophotometry.

In Vivo Transport and Immune Response of Metal Nanoparticles. It has become apparent that nanoparticles do not diffuse freely throughout tissues and organs. Instead, their accumulation appears to be governed by their size and functional surface coating.10 While blood-filtering organs such as the liver and spleen typically sequester the majority of administered nanoparticles, penetration into organs such as the brain can be difficult on account of the highly selective blood-brain barrier (BBB). Phagocytic cells such as macrophages, dendritic cells, and monocytes are located in various tissues and will nonspecifically take up nanoparticles. Nanoparticles are first engulfed by the cellular membrane and then internalized during the phagocytosis process. Current studies have provided evidence that increases in diameter<sup>17,18</sup> and aspect ratio of nanoparticles can impact phagocytic kinetics. 19,20 Of note, phagocytosis appears to increase when the diameter of a nanoparticle is increased or when the tangent angle at the point of contact between the cell membrane and nanoparticle is decreased. 19,20

Macrophages are an important part of the reticuloendothelial system, are involved in the clearance and metabolism of foreign particles such as bacteria and viruses, and can stimulate lymphocytes and other immune cells in response to foreign particles. These processes could significantly influence the inflammatory response, where chemical mediators called cytokines released by the macrophages induce accumulation of fluid and immune cells. The intensity and location of the inflammation will determine if cell death and tissue damage occurs in a specific organ. By profiling the cytokines released after nanoparticle exposure, one can determine whether a process is pro- or antiinflammatory. Examples of inflammatory nanoparticles include cobalt and nickel, which induce the release of tumor necrosis factor (TNF)- $\alpha$  and macrophage inflammatory protein (MIP)-2 cytokines subsequent to phagocytosis in cell culture.21 In contrast, cerium oxide nanoparticles suppress macrophage production of free radical nitric oxide and dampen the inflammatory response.<sup>22</sup> Gold nanoparticles are neither immunosuppressive nor inflammatory.<sup>21</sup> In vivo experiments also reveal a wide range of immune responses when alveolar macrophages are exposed to inflammatory ferric oxide nanoparticles<sup>23</sup> or immunosuppressive carbon nanotubes.24 It will be necessary in future studies to single out specific nanoparticle properties that alter immune response in a desirable manner.

**Interactions of Non-uniform** Nanoparticles with Brain Cells. In this issue's article by Hutter et al.,25 the authors attempt to understand the effect of nanoparticle geometry on biological response in the central nervous system (CNS). The CNS is difficult to study mainly because of the restricted transport of nanoparticles across the BBB. The researchers introduced one of three

different nanoparticle geometries (spheres, rods, or urchins) to multiple CNS-associated cells. Although a diverse number of cell types reside in the brain, the researchers characterized uptake in two cell types: microglia and neurons. The authors demonstrated that nanoparticles with thin protrusions, known as nanourchins, are preferentially taken up by microglia, while only rods were internalized by neurons, suggesting that cells display selectivity toward certain particle geometries. Since all nanoparticles had the same surface chemistry, they propose that morphological differences in nanourchins promoted phagocytosis by microglia. Neurons relay chemical and electrical signals to and from the CNS, while microglia cells are the resident macrophages of the brain and spine. Neurons typically internalize small molecules such as proteins via endocytic mechanisms, whereas microglia phagocytose irregularly shaped particulates. Since nanoparticle morphology preference may correlate with cell function, the differences in uptake between these cell types should not be surprising.

Whether in the CNS or in other tissues, it is important to address whether accumulated nanoparticles will eventually activate the tissue's resident phagocytes causing inflammation. Hutter et al. investigated whether nanoparticle uptake up-regulated inflammatory molecules such as Toll-like receptor-2 (TLR-2) on microglia. This receptor is commonly expressed on phagocytes and recognizes multiple pathogen-associated molecular patterns found on the surface of bacteria, yeast, or parasites.26 Binding of this receptor enhances phagocytosis rates and triggers the release of several proinflammatory cytokines. In vitro experiments revealed that nanourchins and nanorods both upregulated TLR-2 expression in microglia, whereas nanospheres did not (Figure 1). More detailed analysis re-

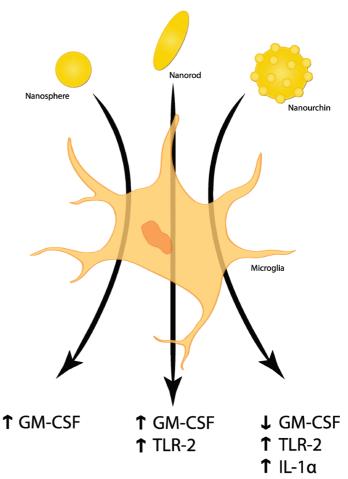


Figure 1. Differential release of chemical mediators called cytokines in response to nanoparticle geometry.

vealed differential inflammatory cytokine expression in response to each nanoparticle. Nanourchins provoked increased interleukin- $1\alpha$ (IL-1α) production while suppress-

The differences observed by Hutter et al. in nanoparticle-induced cytokines suggest a unique microgliamediated immune response for each morphology in vivo.

ing granulocyte macrophage colony stimulating factor (GM-CSF). In contrast, poly(ethylene glycol)coated (PEG) gold spheres and rods both increased the production of GM-CSF. These differences in nanoparticle-induced cytokines suggest a unique microglia-mediated immune response for each morphology in vivo.

#### CONCLUSIONS

In the last 5 years, there has been a significant interest in elucidating the interactions of nanoparticles with biological systems.<sup>27</sup> While the task is quite daunting considering the number of nanoparticle chemical compositions, sizes, shapes, and surface chemistries and the overall complexity and intricacy of physiological systems, these studies are indispensable. To be able to evaluate completely the importance of each parameter in a

biological system can take several decades. Hutter et al. described in their conclusions that many other nanoparticle parameters must be studied in order for them to develop guidelines for engineering nanoparticles as contrast agents for imaging the morphology and cells of the CNS. The success of such a large-scale endeavor will be beneficial to the field of nanotechnology since the outcome of these studies will provide important design parameters to engineer safe nanomaterials for a wide variety of medical applications. As an example, rapid clearance of nanoparticles by uptake into macrophages will lead to reduced accumulation in tumors, possibly rendering nanoparticles ineffective as imaging agents or drug carriers. This problem was solved by the discovery of the antifouling properties of the polymer PEG for coating nanoparticles.28 The PEGylation chemistry is now an important part of the nanotoolbox. Fundamental studies on the interactions of nanoparticles with biological systems are important to the advancement of nanotechnology for medical applications.

Acknowledgment. W.C.W.C. acknowledges the Canadian Institute of Health Research (RMF-72551, MOP-93532), Natural Sciences and Engineering Research Council (NETGP 35015-07, RGPIN 288231-09), Canadian Foundation for Innovation, and Ministry of Research and Innovation in Ontario for research support. A.A. acknowledges the Ontario Graduate Scholarship and the Barbara and Frank Milligan Graduate Fellowship.

### **REFERENCES AND NOTES**

- 1. Zhang, S. L.; Li, J.; Lykotrafitis, G.; Bao, G.; Suresh, S. Size-Dependent Endocytosis of Nanoparticles. Adv. Mater. 2009, 21, 419-424.
- 2. Chithrani, B. D.; Chan, W. C. W. Elucidating the Mechanism of Cellular Uptake and Removal of **Protein-Coated Gold Nanoparticles** of Different Sizes and Shapes. Nano Lett. 2007, 7, 1542-1550.
- 3. Gratton, S. E.; Ropp, P. A.; Pohlhaus, P. D.; Luft, J. C.; Madden, V. J.; Napier, M. E.; DeSimone, J. M. The Effect of Particle Design on Cellular Internalization Pathways. Proc. Natl. Acad. Sci. U.S.A. 2008, 105, 11613-11618.

- 4. Hauck, T. S.; Ghazani, A. A.; Chan, W. C. W. Assessing the Effect of Surface Chemistry on Gold Nanorod Uptake, Toxicity, and Gene Expression in Mammalian Cells. Small 2008, 4, 153-159.
- Slowing, I.; Trewyn, B. G.; Lin, V. S. Y. Effect of Surface Functionalization of MCM-41-Type Mesoporous Silica Nanoparticles on the Endocytosis by Human Cancer Cells. J. Am. Chem. Soc. 2006, 128, 14792-14793.
- 6. Derfus, A. M.; Chan, W. C. W.; Bhatia, S. N. Probing the Cytotoxicity of Semiconductor Quantum Dots. Nano Lett. **2004**, 4, 11–18.
- 7. Hauck, T. S.; Anderson, R. E.; Fischer, H. C.; Newbigging, S.; Chan, W. C. W. In Vivo Quantum-Dot Toxicity Assessment. Small 2010, 6, 138 - 144.
- 8. Peer, D.; Karp, J. M.; Hong, S.; FaroKhzad, O. C.; Margalit, R.; Langer, R. Nanocarriers as an **Emerging Platform for Cancer** Therapy. Nat. Nanotechnol. 2007, 2, 751-760.
- 9. Matsumura, Y.; Maeda, H. A New Concept for Macromolecular Therapeutics in Cancer Chemotherapy: Mechanism of Tumoritropic Accumulation of Proteins and the Antitumor Agent Smancs. Cancer Res. 1986, 46, 6387-6392.
- 10. Perrault, S. D.; Walkey, C.; Jennings, T.; Fischer, H. C.; Chan, W. C. W. **Mediating Tumor Targeting** Efficiency of Nanoparticles through Design. Nano Lett. 2009, 9, 1909-1915.
- 11. Choi, H. S.; Liu, W.; Misra, P.; Tanaka, E.; Zimmer, J. P.; Itty Ipe, B.; Bawendi, M. G.; Frangioni, J. V. Renal Clearance of Quantum Dots. Nat. Biotechnol. 2007, 25, 1165-1170.
- 12. Nel, A.; Xia, T.; Madler, L.; Li, N. Toxic Potential of Materials at the Nanolevel. Science 2006, 311, 622-627.
- 13. Nam, J. M.; Thaxton, C. S.; Mirkin, C. A. Nanoparticle-Based Bio-Bar Codes for the Ultrasensitive Detection of Proteins. Science 2003, 301, 1884-1886.
- 14. Hirsch, L. R.; Stafford, R. J.; Bankson, J. A.; Sershen, S. R.; Rivera, B.; Price, R. E.; Hazle, J. D.; Halas, N. J.; West, J. L. Nanoshell-Mediated Near-Infrared Thermal Therapy of Tumors under Magnetic Resonance Guidance. Proc. Natl. Acad. Sci. U.S.A. 2003, 100, 13549-13554.
- 15. Lee, J. S.; Green, J. J.; Love, K. T.; Sunshine, J.; Langer, R.; Anderson, D. G. Gold, Poly(β-amino ester) Nanoparticles for Small Interfering RNA Delivery. Nano Lett. 2009, 9, 2402-2406.
- 16. Park, J. H.; von Maltzahn, G.; Xu, M. J.; Fogal, V.; Kotamraju, V. R.; Ruoslahti, E.; Bhatia, S. N.; Sailor, M. J. Cooperative Nanomaterial System to Sensitize, Target, and

- Treat Tumors. Proc. Natl. Acad. Sci. U.S.A. **2010**, 107, 981-986.
- 17. Kreyling, W. G.; Semmler, M.; Erbe, F.; Mayer, P.; Takenaka, S.; Schulz, H.; Oberdorster, G.; Ziesenis, A. Translocation of Ultrafine Insoluble Iridium Particles from Lung Epithelium to Extrapulmonary Organs Is Size Dependent but Very Low. J. Toxicol. Environ. Health A **2002**, 65, 1513-1530.
- 18. Fang, C.; Shi, B.; Pei, Y. Y.; Hong, M. H.; Wu, J.; Chen, H. Z. In Vivo **Tumor Targeting of Tumor Necrosis** Factor-α-Loaded Stealth Nanoparticles: Effect of MePEG Molecular Weight and Particle Size. Eur. J. Pharm. Sci. 2006, 27, 27-36.
- 19. Champion, J. A.; Mitragotri, S. Shape Induced Inhibition of Phagocytosis of Polymer Particles. Pharm. Res. **2009**, 26, 244-249.
- 20. Champion, J. A.; Mitragotri, S. Role of Target Geometry in Phagocytosis. Proc. Natl. Acad. Sci. U.S.A. 2006, 103, 4930-4934.
- 21. Mo, Y.; Zhu, X.; Hu, X.; Tollerud, D. J.; Zhang, O. Cytokine and NO Release from Peripheral Blood Neutrophils after Exposure to Metal Nanoparticles: In Vitro and Ex Vivo Studies. Nanotoxicology 2008, 2, 79-87
- 22. Hirst, S. M.; Karakoti, A. S.; Tyler, R. D.; Sriranganathan, N.; Seal, S.; Reilly, C. M. Anti-inflammatory **Properties of Cerium Oxide** Nanoparticles. Small 2009, 5, 2848-2856.
- 23. Zhu, M. T.; Feng, W. Y.; Wang, B.; Wang, T. C.; Gu, Y. Q.; Wang, M.; Wang, Y.; Ouyang, H.; Zhao, Y. L.; Chai, Z. F. Comparative Study of Pulmonary Responses to Nano- and Submicron-Sized Ferric Oxide in Rats. Toxicology 2008, 247, 102-111.
- 24. Mitchell, L. A.; Lauer, F. T.; Burchiel, S. W.; McDonald, J. D. Mechanisms for How Inhaled Multiwalled **Carbon Nanotubes Suppress** Systemic Immune Function in Mice. Nat. Nanotechnol. 2009, 4, 451-456.
- 25. Hutter, E.; Boridy, S.; Labrecque, S.; Lalancette-He bert, M.; Kriz, J.; Winnik, F.; Maysinger, D. Microglial Response to Gold Nanoparticles. ACS Nano 2010, 4, 2595-2606.
- 26. Medzhitov, R. Toll-like Receptors and Innate Immunity. Nat. Rev. Immunol. 2001, 1, 135-145.
- 27. Nel, A. E.; Madler, L.; Velegol, D.; Xia, T.; Hoek, E. M.; Somasundaran, P.; Klaessig, F.; Castranova, V.; Thompson, M. Understanding Biophysicochemical Interactions at the Nano-Bio Interface. Nat. Mater. **2009**, *8*, 543–557.
- 28. Akerman, M. E.; Chan, W. C. W.; Laakkonen, P.; Bhatia, S. N.; Ruoslahti, E. Nanocrystal Targeting In Vivo. Proc. Natl. Acad. Sci. U.S.A. **2002**, 99, 12617-12621.

