See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/223466100

Formal α -Vinylation of Amino Acids. Use of a New Benzeneselenolate Equivalent.

ARTICLE in THE JOURNAL OF ORGANIC CHEMISTRY · DECEMBER 1993

Impact Factor: 4.72 · DOI: 10.1021/jo00077a012

CITATIONS	READS
28	90

2 AUTHORS, INCLUDING:



David B Berkowitz

University of Nebraska at Lincoln

104 PUBLICATIONS 1,772 CITATIONS

SEE PROFILE

Articles

Formal α -Vinylation of Amino Acids. Use of a New Benzeneselenolate Equivalent

Michelle L. Pedersen and David B. Berkowitz*

Department of Chemistry, University of Nebraska-Lincoln, Lincoln, Nebraska 68588-0304

Received July 23, 1993®

A new synthetic approach to the formal α -vinylation of α -amino acids is described, in which the readily available electrophile, ethylene oxide, serves as the vinyl cation equivalent. N-Benzoyl α -amino esters bearing appropriate side-chain protecting groups are deprotonated with lithium diisopropylamide in THF/TMEDA at -78 °C to generate the corresponding dianions. Exposure of these to ethylene oxide results in C-alkylation and lactonization to give the corresponding racemic, α -substituted homoserine lactones 2a-j in 61-85% yield. Next, reduction of diphenyl diselenide with sodium trimethoxyborohydride generates a benzeneselenolate anion equivalent which efficiently cleaves the α-substituted homoserine lactones without competing lactone reduction (69–97% yields for 3a-j following diazomethane workup). The protected α -[2-(phenylseleno)ethyl] amino acids thereby obtained are oxidized to the corresponding selenoxides through the agency of ozone at -78 °C. Pyrolysis of these in refluxing benzene or carbon tetrachloride gives the protected α -vinyl amino acids 4a-c and 4e-j in 95-100% yield. In the case of methyl N^{a} -benzoyl-2-[2'-(phenylseleno)ethyl]- N^{τ} -tritylhistidinate (3d), oxidation and pyrolysis (80%) are carried out in one pot by refluxing with tetrabutylammonium periodate in chloroform. Finally, deprotection is achieved by acidic hydrolysis. This methodology has been successfully applied to the synthesis, in racemic form, of the α -vinyl amino acids derived from phenylalanine (5a), DOPA (5b), histidine (5c), lysine (5d), ornithine (5e), valine (5f), alanine (5g), and homoserine (5h). In addition, α -vinylaspartic acid (9) and α -vinylarginine (10) could be obtained from α -vinylhomoserine derivative 4j and α -vinylornithine (5f), respectively.

Introduction

 α -Vinyl amino acids¹⁻⁵ are an important class of mechanism based inhibitors for enzymes of the amino acid decarboxylase class. For example, DOPA decarboxylase, ornithine decarboxylase, and glutamate decarboxylase, all important medicinal targets, are irreversibly inhibited by the corresponding α -vinyl amino acids. α -Vinylhistidine reversibly inhibits histidine decarboxylase. In Indeed, a large number of amino acid decarboxylases are known, for which the corresponding α -vinyl amino acids might be expected to act as inhibitors. In the case of α -vinyl-DOPA, the "mechanism-based" nature of the inhibition has been borne out, as inactivation is known to require enzymatic decarboxylation. Hence, there is considerable interest in α -vinyl amino acids, both from a pharmaceutical and from a mechanistic point of view.

Abstract published in Advance ACS Abstracts, November 1, 1993.
(1) (a) Maycock, A. L.; Aster, S. D.; Patchett, A. A. Dev. Biochem.
1979, 6, 115-129. (b) Ribereau-Gayon, G.; Danzin, C.; Palfreyman, M. G.; Aubry, M.; Wagner, J.; Metcalf, B. W.; Jung, M. J. Biochem. Pharm.
1979, 28, 1331-1335. (c) Maycock, A. L.; Aster, S. D.; Patchett, A. A. In Enzyme-Activated Irreversible Inhibitors; Seiler, N., Jung, M. J., Koch-Weser, J., Eds.; Elsevier: North Holland, 1978; pp 211-220.

(2) Danzin, C.; Casara, P.; Claverie, N.; Metcalf, B. W. J. Med. Chem. 1981, 24, 16-20.

(3) Metcalf, B.; Jung, M. U. S. Patent 4,147,873, April 3, 1979.
(4) Tendler, S. J. B.; Threadgill, M. D.; Tisdale, M. J. J. Chem. Soc., Perkin Trans. I 1987, 2617-2623.

(5) (a) Cho, C.; Ishii, R.; Hyeon, S.; Suzuki, A. Agric. Biol. Chem. 1987, 51, 2597–2598. (b) Soper, T. S.; Manning, J. M.; Marcotte, P. A.; Walsh, C. T. J. Biol. Chem. 1977, 252, 1571–1575. (c) Rando, R. R.; Relyea, N.; Cheng, L. Ibid. 1976, 251, 3306–3312.

(6) For a review article on amino acid decarboxylases see: Boeker, E. A.; Snell, E. E. *The Enzymes*; Academic: New York, 1972; Vol 6, pp 217-253.

Among the synthetic approaches to α -vinyl amino acids, the α -vinylation of protected amino acids is arguably the most direct. This approach begins with the amino acid side chain in place, as provided by Nature or a Strecker synthesis, and utilizes the intrinsic acidity of the α -proton to access an α -carbanion, which is then alkylated with a vinyl cation equivalent. There are several examples of this strategy in the literature, in which an amino acid derivative is deprotonated and then alkylated with either a phenylsulfonyl-activated Michael acceptor or an α -silyl accetaldehyde as vinyl cation equivalent. However, these

(7) For α-vinylation approaches, see: (a) Sawada, S.; Nakayama, T.;
 Esaki, N.; Tanaka, H.; Soda, K.; Hill, R. K. J. Org. Chem. 1986, 51, 3384–3386. (b) Steglich, W.; Wegmann, H. Synthesis 1980, 481–483. (c) Huldrik, P. F.; Kulkarni, A. S. J. Am. Chem. Soc. 1981, 103, 6251–6253. (d) Metcalf, B. W.; Bonilavri, E. J. Chem. Soc., Chem. Commun. 1978, 914–915.

(8) For alternative approaches, see: (a) Castelhano, A. L.; Horne, S.; Taylor, G. J.; Billedeau, R.; Krantz, A. Tetrahedron, 1988 44, 5451-5466. (b) Münster, P.; Steglich, W. Synthesis 1987, 223-225. (c) Weber, T.; Aeschimann, R.; Maetzke, T.; Seebach, D. Helv. Chim. Acta 1986, 69, 1365-1377. (d) Castelhano, A. L.; Horne, S.; Billedeau, R.; Krantz, A. Tetrahedron Lett. 1986, 27, 2435-2438. (e) Greenlee, W. J.; Taub, D.; Patchett, A. A. Tetrahedron Lett. 1978, 3999-4002. (f) Metcalf, B. W.; Jund, K. Ibid. 1978, 3689-3692. (g) Taub, D.; Patchett, A. A. Ibid. 1977, 2745-2748.

(9) For syntheses of (±)-α-vinylglycine see: (a) Fitzner, J. F.; Pratt,
D. V.; Hopkins, P. B. Tetrahedron Lett. 1985, 26, 1959–1962. (b) Vyas,
D. M.; Chiang, Y.; Doyle, T. W. J. Org. Chem. 1984, 49, 2037–2039. (c)
Greenlee, W. J. J. Org. Chem. 1984, 49, 2632–2634. (d) Baldwin, J. E.;
Haber, S. B.; Hoskins, C.; Kruse, L. I. J. Org. Chem. 1977, 42, 1239–1241.
(e) Friis, P.; Helboe, P.; Larsen, P. O. Acta Chem. Scand., Ser. B 1974, 28, 317–321 and ref 7c.

(10) For syntheses of α-vinylglycine in enantiomerically enriched form see: (a) Pellicciari, R.; Natalini, B.; Marinozzi Synth. Commun. 1988, 18, 1715–1721. (b) Hanessian, S.; Sahoo, S. P. Tetrahedron Lett. 1984, 25, 1425–1428. (c) Schollkopf, U.; Nozulak, J.; Groth, U. Tetrahedron 1984, 40, 1409–1417. (d) Afzali-Ardakani, A.; Rapoport, H. J. Org. Chem. 1980, 45, 4817 and ref 19.

Scheme I

Scheme I

$$H_3CO$$
 Ph
 H_3CO
 Ph
 H_3CO

vinyl cation equivalents are not commercially available and the scope of these methods, with regard to side chain compatibility, remains to be demonstrated.

We have taken a conceptually different approach to the formal α -vinylation of amino acids, in which a readily available electrophile, ethylene oxide, serves as the vinyl cation equivalent (Scheme I). Alkylation of a suitable α -carbanion with ethylene oxide is expected to proceed with facile 5-exo-trig cyclization to give an α -substituted homoserine lactone directly. The necessary two-carbon fragment thereby becomes attached to the α -carbon. Its elaboration to a vinyl group follows from the sequence benzeneselenolate-mediated lactone cleavage, selenide oxidation and selenoxide pyrolysis. Herein, we report on the workability of this scheme and its compatibility with various amino acid side chains.

Results and Discussion

We initially observed that the α -monoanion of N-benzylidene-protected amino acid esters¹² is not sufficiently nucleophilic to react with ethylene oxide at an acceptable rate at room temperature. Rather, conversion to the homoserine lactone is inefficient and requires heating a solution of this monoanion with excess ethylene oxide in a sealed vessel overnight. On the other hand, the corresponding dianions, generated from N-benzoyl-protected α -amino esters, ¹³ react readily with ethylene oxide at or below room temperature. The desired homoserine lactones 2, in racemic form, were obtained in good to very good yield in all cases studied (Table I). Evidently then, under these conditions, C-alkylation of the benzamido ester dianions is favored over N-alkylation and, even in the presence of excess ethylene oxide, monoalkylation predominates. Also noteworthy here is the ability to generate and effectively use dianions bearing silyl aryl ether (1b), N-tritylimidazole (1d), and imine (1e, 1f) functional groups on the side chains.

Having developed conditions for the introduction of a two carbon fragment at the α -position, it remained to convert that fragment into a vinyl group. Both Smith¹⁴

Table I. Dianion Alkylation with Ethylene Oxide

Table 1. Dianion Alkylation with Ethylene Oxide							
protected amino acid	t (h)		product	yield			
a O MeO	1	(±)	NHBz	85%			
BZHN OTBS OTBS	1	(±)	NHBz	84% S			
BZHN O	1	(±)	NHBz OTBS	82%			
BZHN NCPh3	2	(±)	NHBz NCPh3	80%			
BZHN e O N=CHP	h 2	(±)	NHBz NHE	3z 61%*			
BzHN N=CHPh MeO	3	(±)	NHBz	78%			
BzHN g O Me MeO Me	1	(±)	NHBz Me	84%			
BzHN O Me MeO	0.5	(±)	NHBz Me	72% ^b			
i O NHB2	8	(±)	NHBz OSiPh ₂ t-	52%° Bu (62%) ^d			
j O NHBz	8	(±)	NHBz Ot-Bu	52%° (62%) ^d			

^a Overall yield for 3 steps: (i) LDA, $(CH_2)_2O$; (ii) 3 N HCl, THF; (iii) PhCOCl, NEt₃, CH_2Cl_2 . ^b Reference 11. ^c Overall yield for 2 steps: (i) LDA, $(CH_2)_2O$; (ii) Ph-t-BuSiCl, imidazole, DMF. ^d Yield based on recovered 1i in the first step. ^e Overall yield for 2 steps: (i) LDA, $(CH_2)_2O$; (ii) isobutylene, CH_2Cl_2 , H_2SO_4 .

and Liotta¹⁵ had reported nucleophilic benzeneselenolate equivalents capable of efficiently cleaving simple γ -lactones.¹⁶ Smith and co-workers reduced diphenyl diselenide with sodium borohydride in dimethylformamide. Liotta and co-workers pointed out that Smith actually must have a sodium benzeneselenolate—diborane complex (NaPh-SeBH₃) and reported lactone cleavage with sodium benzeneselenolate generated from sodium metal and diphenyl diselenide or from sodium hydride and benzeneselenol.

Screening of these benzeneselenolate reagents indicated that, for cleaving N-benzamido- α -substituted homoserine lactones, the Smith conditions were superior to the Liotta conditions in terms of both rate and yield. So, for example, in the case of α -methylhomoserine lactone 2h, we obtained only 18-30% of the desired phenyl selenide using the Liotta conditions as compared with 86% under the conditions of Smith.¹¹ In each case, the intermediate carboxylate was

⁽¹¹⁾ A preliminary communication of this α -vinylation methodology (one example) has appeared: Berkowitz, D. B. Synth. Commun. 1990, 20, 1819–1829.

⁽¹²⁾ Stork, G.; Leong, A. Y. W.; Touzin, A. M. J. Org. Chem. 1976, 41, 3491-3493.

^{(13) (}a) McIntosh, J. M.; Thangarasa, R.; Ager, D. J.; Zhi, B. Tetrahedron 1992, 48, 6219-6224. (b) Krapcho, A. P.; Dundulis, E. A. Tetrahedron Lett. 1976, 26, 2205-2208.

^{(14) (}a) Scarborough, R. M., Jr.; Toder, B. H.; Smith, A. B., III. J. Am. Chem. Soc. 1980, 102, 3904-3913. (b) Scarborough, R. M., Jr.; Smith, A. B., III. Tetrahedron Lett. 1977, 4361-4364.

^{(15) (}a) Ley, S. V.; O'Neill, I. A.; Low, C. M. R. Tetrahedron 1986, 46, 5363-5368. (b) Liotta, D.; Sunay, U.; Santiesteban, H.; Markiewicz, W. Yoneda, F.; Kuroda, K. J. Org. Chem. 1981, 46, 2605-2610. (c) Liotta, D.; Santiesteban, H. Tetrahedron Lett. 1977, 4369-4372.

⁽¹⁶⁾ For other examples of the cleavage of lactones with benzenese-lenolate anion, see: (a) Soucy, F.; Wernic, D.; Beaulieu, P. J. Chem. Soc., Perkin Trans. I 1991, 2885-2887. (b) Dowd, P.; Kennedy, P. Synth. Commun. 1981, 11, 935-941. (c) Hoye, T. R.; Caruso, A. J. Tetrahedron Lett. 1978, 4611-4614.

Table II. Lactone Cleavage with "NaPhSeB(OMe);"

protonated and esterified with diazomethane to give an α -[2-(phenylseleno)ethyl] α -amino ester. Unfortunately, attempts to apply this procedure to homoserine lactones bearing bulky α-substituents met with unsatisfactory results. Thus, with homoserine lactone 2i, bearing an α -[2-[(tert-butyldiphenylsilyl)oxy]ethyl] side chain, lactone reduction (49%) was seen to compete with lactone cleavage (51%) under the Smith conditions. This problem was solved by developing a new benzeneselenolate equivalent.¹⁷

We reasoned that a phenyl selenide reagent possessing a potentially Lewis acidic, oxophilic boron component as in the Smith reagent, 14,15 but lacking reducing equivalents, might display both the reactivity and selectivity required. Accordingly, diphenyl diselenide was reduced with 2 equiv of sodium trimethoxyborohydride in DMF (60 °C, 1 h). Treatment of lactone 2h with this reagent gave the desired α -[2-(phenylseleno)ethyl] amino ester 3i in 90% yield. These conditions have been successfully applied to the cleavage of a variety of α -substituted N-benzoylhomoserine lactones (Table II).17 These results suggest that the

Table III. Selenide Oxidation/Pyrolysis and Deprotection

17	55 (1/3 (1)4			(±/3
		yi		
	selenide -	step 1	step 2	product
a (±)	BzHN SePh MeO OTBS	99%	70%	α-vinyl- phenylalanine
b (±)	BzHN OTBS SePh	97%	77%	α-vinyl- DOPA*
c (±)	BzHN O SePh	95%	b	
d (±)	BzHN N SePh	80%°	73%	α-vinyl- histidine
e (±)	BzHN NHBz O SePh MeO NHBz	99%	60%	α-vinyl- lysine
f (±)	BzHN O SePh	100%	84%	α-vinyl- omithine
g (±)	BzHN Me O SePh	99%	85%	α-vinyl- valine
h (±)	BzHN Me O SePh MeO OSiPh ₂ t-Bu	100% ^d	89%	α-vinyl- alanine
i (±)	SePh MeO Ot-Bu	100%	78%	α-vinyl- homoserine
j (±)	BzHN SePh	99%	88%	α-vinyl- homoserine

^a Isolated as the HCl salt. ^b Side-chain deprotection unsuccessful. ^c Bu₄NIO₄ (1 equiv), CHCl₃, reflux. ^d Reference 11.

benzeneselenolate equivalent generated by the reduction of diphenyl diselenide with sodium trimethoxyborohydride may well be the reagent of choice of the cleavage of α -substituted homoserine lactones.

Oxidation of the selenides 3 to the required selenoxides was next undertaken. Several oxidants, including hydrogen peroxide, sodium periodate, Chloramine T,18 and ozone were examined. Oxidation with ozone at -78 °C in methylene chloride proved to be the best method in all but one case. The intermediate selenoxides were not isolated but rather were pyrolyzed directly to the protected α -vinyl amino acids. Addition of 1-hexene to the selenoxide solution prior to pyrolysis, as described by Barton and Crich, 19 resulted in nearly quantitative yields for the oxidation/pyrolysis sequence (Table III). Failure to include the 1-hexene gave markedly reduced yields.

In the case of 3d, however, all attempts at ozonemediated selenoxide oxidation resulted in destruction of

Tetrahedron 1985, 41, 4347-4357.

⁽¹⁷⁾ Pedersen, M. L.; Berkowitz, D. B. Tetrahedron Lett. 1992, 33, 7315-7318.

⁽¹⁸⁾ Sharpless, K. B.; Gordon, K. M.; Lauer, P. F.; Patrick, D. W.;
Singer, S. P.; Young, M. W. Chem. Scr. 1975, 8A, 9-13.
(19) Barton, D. H. R.; Crich, D.; Herve, Y.; Potier, P.; Thiery, J.

the imidazole ring of the side chain. A convenient onepot oxidation/pyrolysis procedure was found to solve this problem. Simply refluxing 3d with 1 equiv of tetrabutylammonium periodate in chloroform gave 4d directly in 80% isolated yield.

Finally, amino group, carboxyl group and side-chain deprotection were achieved simultaneously by refluxing the protected α-vinyl amino acids 4 with 6 N HCl (Table III). Only the methylenedioxy side chain of α -vinyl-DOPA derivative 4c resisted cleavage under these conditions. This case was not pursued further as the alternative α -vinyl-DOPA derivative 4b, bearing tert-butyldimethylsilyl (TBS) ether catechol protecting groups, was readily deprotected. We note that installation of the side chain protecting groups was also much more facile for 1b (TBS ethers) than for 1c (methylenedioxy, vide infra), giving the b series considerable advantage over the c series for the α -vinylation of DOPA. On the other hand, in the case of homoserine, both tert-butyl ether (j series) and tertbutyldiphenylsilyl ether (i series) side-chain protecting groups were nicely compatible with this chemistry and easily hydrolytically cleaved in the final step.

In addition, α -vinylaspartic acid (9) could be synthesized in four steps from the protected α -vinylhomoserine derivative 4j (Scheme II). Thus, tert-butyl ether deprotection readily gave γ -lactone 6. The pendant hydroxyethyl side chain could be released from lactone 6 by treatment with methylamine. The hydroxy amide 7 then underwent smooth four-electron oxidation to imide 8. Acidic hydrolysis provided α -vinylaspartic acid.

Finally, α -vinylarginine 10 could be obtained from α -vinylornithine 5f, in a single step, employing S-ethylisothiouronium bromide as formal amidine donor (Scheme III).²⁰

Conclusions

A new procedure for the formal α -vinylation of α -amino acids has been developed. Key features include the use of the readily available electrophile ethylene oxide as a vinyl cation equivalent and the introduction of a new, chemoselective benzeneselenolate equivalent for α -substituted homoserine lactone cleavage. The overall yields obtained for this four step sequence [(i) lactone formation

with ethylene oxide, (ii) lactone cleavage with benzene-selenolate anion, (iii) oxidation/pyrolysis, and (iv) deprotection] compare favorably to previously reported yields for the α -vinylation of protected amino acids. However, the need to synthesize a vinyl cation equivalent is circumvented here. Furthermore, this methodology has been shown to be compatible with a considerable variety of side-chain functionality (β -branched alkyl, aryl, imine, benzamide, N-tritylimidazole, tert-butyl ether, alkyl silyl ether, aryl silyl ether, and acetal) and to provide ready access to the α -vinyl amino acids derived from alanine, phenylalanine, valine, homoserine, DOPA, ornithine, lysine, histidine, arginine, and aspartate.

Experimental Section

General. All reactions were conducted under an argon atmosphere using oven-dried glassware unless otherwise noted. Methylene chloride, pyridine, and diisopropylamine were distilled from CaH₂. THF, Et₂O and benzene were distilled from sodium benzophenone ketyl. Methanol was distilled from Mg. Other reagents were obtained from commercial sources and used without further purification. Flash chromatography was performed using Merck silica gel 60 (230-400 mesh). ¹H NMR spectra were recorded on a GE Omega-500, GE Omega-300, NT-360, or VXR-200 instrument. Proton-decoupled ¹³C NMR spectra were acquired on a GE Omega-500, GE Omega-300, or VXR-200 instrument. Infrared spectra were obtained using an Analect RFX-65 FTIR spectrometer. Mass spectra were acquired at the Midwest Center for Mass Spectrometry (University of Nebraska—Lincoln) and are reported as m/z (relative intensity). Elemental analyses were carried out by M-H-W Labs (Phoenix, AZ). Melting points were determined using a Meltemp II (Laboratory Devices) apparatus and are uncorrected.

Methyl N-Benzoyl-3',4'-bis(benzoyloxy)phenylalaninate. To a solution of (D,L)-DOPA methyl ester hydrochloride²¹ (22.0 g. 88.8 mmol) and 4-(dimethylamino)pyridine (1.08 g, 8.88 mmol) in pyridine (440 mL) at 0 °C was added benzoyl chloride (30.9 mL, 266 mmol). After being stirred at rt for 1 h, the reaction mixture was filtered and then evaporated in vacuo. Flash chromatography [EtOAc/hexane/CH₂Cl₂ (50:50:5)] afforded the title compound (41.0 g, 88%) as a white solid: mp 173-175 °C; ¹H NMR (300 MHz, CDCl₃) δ 3.32-3.34 (app d, J = 5 Hz, 2 H), 3.80 (s, 3 H), 5.11-5.17 (app dt, J = 5, 7 Hz, 1 H), 6.71-6.73 (d, J = 7 Hz, 1 H, 7.10-7.20 (m, 3 H), 7.30-7.57 (m, 9 H), 7.78-7.82 $(m, 2 H), 8.00-8.05 (m, 4 H); {}^{13}C NMR (75 MHz, CDCl₃) \delta 37.3,$ 52.6, 53.5, 123.6, 124.7, 127.2, 127.2, 127.5, 128.5, 128.7, 128.7, 128.7, 130.1, 131.8, 133.7, 134.8, 141.6, 142.5, 164.1, 164.2, 166.9, 171.7; IR (film) 3420-3200, 1740, 1645 cm⁻¹; MS (FAB, 3-NOBA) 524 (20, MH+), 464 (6), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₃₁H₂₆NO₇ (MH⁺) 524.1709, obsd 524.1718. Anal. Calcd for C₃₁H₂₅NO₇: C, 71.12; H, 4.81; N, 2.67. Found: C, 71.38; H, 4.88; N, 2.71.

Methyl N-Benzoyl-3',4'-dihydroxyphenylalaninate. To a suspension containing methyl N-benzoyl-3',4'-bis(benzoyloxy)phenylalaninate (20.1 g, 38.4 mmol) in MeOH (425 mL) and THF (85 mL) at rt was added K₂CO₃ (5.31 g, 38.4 mmol). After 2 h at rt, Dowex 50 × 8 (7.70 g, ca. 40 molar equiv) was added and the reaction mixture stirred for 1 h. Following filtration and evaporation of the solvent, flash chromatography (20-50% EtOAc/hexane) yielded the title compound (9.80 g, 81%) as a white solid: mp 142-143 °C; ¹H NMR (500 MHz, DMSO-d₆) δ 2.88-2.98 (m, 2 H), 3.62 (s, 3 H), 4.52-4.57 (m, 1 H), 6.51-6.66 (m, 3 H), 7.43-7.54 (m, 3 H), 7.79-7.81 (m, 2 H), 8.65-8.69 (br s, 1 H), 8.71 (s, 1 H), 8.73 (s, 1 H); ¹³C NMR (125 MHz, CDCl₃) & 35.7, 51.7, 54.6, 115.3, 116.3, 119.7, 127.3, 128.2, 128.3, 131.3, 133.7, 143.8, 144.9, 166.4, 172.3; IR (film) 3345-3200, 1735, 1635 cm⁻¹ MS (FAB, 3-NOBA) 316 (100, MH+), 256 (13), 105 (58); HRMS (FAB, 3-NOBA) calcd for C₁₇H₁₈NO₅ (MH⁺) 316.1185, obsd 316.1190. Anal. Calcd for C₁₇H₁₇NO₅: C, 64.76; H, 5.43; N, 4.44. Found: C, 64.86; H, 5.51; N, 4.26.

Methyl N-Benzoyl-3',4'-bis[(tert-butyldimethylsilyl)oxy]**phenylalaninate** (1b). To a solution of methyl N-benzoyl-3', 4'. dihydroxyphenylalaninate (9.80 g, 31.1 mmol), imidazole (8.50 g, 124 mmol) and 4-(dimethylamino)pyridine (380 mg, 3.11 mmol) in DMF (200 mL) was added tert-butyldimethylsilyl chloride (11.7 g, 77.8 mmol). The resulting reaction mixture was heated at 50 °C for 3 h and then poured into NaHCO₃ (aq, 300 mL) and Et₂O (200 mL). The organic layer was further extracted with Et₂O ($2 \times 200 \text{ mL}$) and the combined extracts were dried (MgSO₄) and evaporated. Chromatrography (33% EtOAc/hexane) afforded 1b (12.5 g, 74%) as a white solid: mp 72-73 °C; ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3) \delta 0.11 \text{ (s, 6 H)}, 0.16 \text{ (s, 3 H)}, 0.17 \text{ (s, 3 H)}, 0.92$ (s, 9 H), 0.95 (s, 9 H), 3.07-3.13 (dd, J = 5.5, 13 Hz, 1 H), 3.12-3.19(dd, J = 6, 14 Hz, 1 H), 3.74 (s, 3 H), 4.97-5.04 (app dt, J = 6,8 Hz, 1 H), 6.53-6.60 (m, 3 H), 6.72-6.75 (d, J = 8 Hz, 1 H), 7.37-7.51 (m, 3 H), 7.69-7.71 (m, 2 H); ¹³C NMR (75 MHz, CDCl₃) δ -4.2 (2 C), -4.14, -4.11, 18.3, 18.4, 25.8, 25.9, 37.1, 52.3, 53.4, 121.0, 122.1, 122.3, 127.0, 128.5, 128.7, 131.7, 133.9, 146.1, 146.9 166.8, 172.1; IR (film) 3350-3310, 1745, 1645 cm⁻¹; MS (FAB, 3-NOBA) 544 (48, MH⁺), 486 (75), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₉H₄₈NO₅Si₂ (MH⁺) 544.2914, obsd 544.2908. Anal. Calcd for C₂₉H₄₅NO₅Si₂: C, 64.03; H, 8.34; N, 2.58. Found: C, 64.02; H, 8.20; N, 2.55.

Methyl N-Benzoyl-3',4'-(methylenedioxy)phenylalaninate (1c). A solution of methyl N-benzoyl-3',4'-dihydroxyphenylalaninate (10.3 g, 32.7 mmol) and CsF (24.8 g, 164 mmol) in DMF (20 mL) was shaken at rt for 1 h. Then CH₂Cl₂ (9.40 mL, 147 mmol) was added and the resulting reaction mixture heated at 115 °C for 2 h. The cooled (rt) reaction mixture was diluted with Et₂O (300 mL) and extracted with cold NaHCO₃ (3 × 200 mL). The organic layer was dried (MgSO₄), filtered, and evaporated. Flash chromatography (40% EtOAc/hexane) gave 1c (4.20 g, 40%) as a white solid: mp 126-128 °C; ¹H NMR (300 MHz, CDCl₃) δ 3.09-3.16 (dd, J = 5, 14 Hz, 1 H), 3.17-3.23 (dd, J = 5, 14 Hz, 1 H), 3.76 (s, 3 H), 4.99-5.05 (app dt, J = 5, 7 Hz, 1 H), 5.92 (s, 2 H), 6.55-6.57 (d, J = 7 Hz, 1 H), 6.55-6.72 (m, 3 H), 7.39-7.53 (m, 3 H), 7.72-7.75 (m, 2 H); ¹³C NMR (75 MHz, CDCl₃) & 37.6, 52.4, 53.6, 101.0, 108.3, 109.5, 122.4, 127.0, 128.6, 129.4, 131.8, 133.8, 146.7, 147.8, 166.8, 172.0; IR (film) 3330, 1740, 1640 cm⁻¹; MS (FAB, 3-NOBA) 328 (100, MH⁺), 268 (22), 206 (76); HRMS (FAB, 3-NOBA) calcd for C₁₈H₁₈NO₅ (MH⁺) 328.1185, obsd 328.1183. Anal. Calcd for C₁₈H₁₇NO₅: C, 66.05; H, 5.23; N, 4.28. Found: C, 66.08; H, 5.30; N, 4.22

Methyl Na-Benzoyl-Na-tritylhistidinate (1d). To a solution of methyl N-benzoylhistidinate²² (7.77 g, 28.4 mmol) and triphenylmethyl chloride (15.7 g, 56.9 mmol) in CH₂Cl₂ (120 mL) at rt was added NEt₃ (17.0 mL, 122 mmol). After 4.5 h, the volatiles were removed in vacuo and the residue was purified by column chromatography (30% acetone/CHCl₃) to yield 1d (14.3 g, 98%) as a light yellow solid: mp 74-76 °C; ¹H NMR (300 MHz, DMSO d_6) δ 3.07-3.14 (dd, J = 4, 14 Hz, 1 H), 3.15-3.21 (dd, J = 5, 14 Hz, 1 H), 3.63 (s, 3 H), 4.98-5.04 (app dt, J = 5, 8 Hz, 1 H), 6.60-6.60 (m, 1 H), 7.08-7.14 (m, 6 H), 7.27-7.51 (m, 13 H), 7.89-7.93 (m, 2 H), 8.38–8.41 (d, J = 8 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 26.6, 52.0, 52.9, 75.2, 119.6, 127.2, 128.0 (2 C), 128.3, 129.6, 131.3, 134.0, 136.6, 138.7, 142.2, 166.8, 171.7; IR (film) 3380-3260, 1745, 1660 cm⁻¹; MS (FAB, 3-NOBA) 516 (50, MH⁺), 105 (100); HRMS (FAB, 3-NOBA) calcd for $C_{33}H_{30}N_3O_3$ (MH⁺) 516.2287, obsd 516.2282. Anal. Calcd for C₃₃H₂₉N₃O₃: C, 76.87; H, 5.67; N, 8.15. Found: C, 77.00; H, 5.82; N, 7.96.

Methyl N^{α} -Benzoyl-N-benzylidenelysinate (1e). To a suspension of N^{α} -benzoyllysine²³ (4.00 g, 16.0 mmol) in MeOH (14 mL) at 0 °C was added, dropwise, thionyl chloride (1.28 mL, 17.6 mmol). After heating at 40 °C for 1 h and evaporation of the volatiles in vacuo, methyl N^{α} -benzoyllysinate hydrochloride salt (4.81 g) was obtained of sufficient purity to be used directly for the next step: ¹H NMR (300 MHz, DMSO- d_{θ}) δ 1.38–1.82 (m, 6 H), 2.74 (m, 2 H), 3.62 (s, 3 H), 4.05–4.05 (m, 1 H), 7.45–7.52 (m, 3 H), 7.93–8.07 (m, 2 H), 8.15 (br s, 3 H), 8.82–8.84 (d, 1 H); ¹³C NMR (75 MHz, DMSO- d_{θ}) δ 22.6, 26.3, 29.7, 38.2, 51.8, 52.6, 127.5, 128.1, 131.4, 133.5, 166.6, 172.6. To a suspension of this hydrochloride salt and benzaldehyde (1.54 mL, 15.1 mmol) in

Methyl N^α-Benzoyl-N^δ-benzylideneornithinate (1f). From N^{α} -benzoylornithine²³ (5.00 g, 21.2 mmol) and thionyl chloride (1.7 mL, 23.3mmol), according to the procedure for 1e, was obtained methyl N^{α} -benzoylornithinate hydrochloride salt (6.07 g) of sufficient purity to be used directly for the next step: 1H NMR (300 MHz, DMSO- d_6) δ 1.64-1.71 (m, 2 H), 1.85-1.91 (m, 2 H), 2.77-2.78 (m, 2 H), 3.63 (s, 3 H), 4.39-4.47 (m, 1 H), 7.44-7.57 (m, 3 H), 7.91–7.93 (m, 2 H), 8.11 (s, br), 8.87–8.90 (d, 1 H); ¹³C NMR (75 MHz, DMSO-d₆): δ 22.4, 25.9, 36.8, 51.0, 51.1, 126.0, 127.1, 130.5, 131.7, 166.6, 171.3. Treatment of this hydrochloride salt with benzaldehyde (1.91 mL, 18.8 mmol) and triethylamine (2.92 mL, 20.9 mmol), as for 1e, gave 1f (6.02 g, 84% for 2 steps)as an off-white solid: mp 80-81 °C; ¹H NMR (500 MHz, CDCl₂): δ 1.78-1.84 (app quintet, J = 7 Hz, 2 H), 1.93-2.08 (m, 2 H), 3.63-3.68 (app t, J = 7 Hz, 2 H), 3.75 (s, 3 H), 4.86-4.90 (app dt, J = 5, 7 Hz, 1 H), 7.21–7.22 (d, J = 7 Hz, 1 H), 7.35–7.48 (m, 6) H), 7.68-7.81 (m, 4 H), 8.28 (s, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 26.8, 30.1, 52.4, 52.4, 60.6, 127.1, 128.1, 128.5, 128.6, 130.7, 131.6, 133.9, 136.0, 161.5, 167.1, 172.9; IR (film) 3330, 1740, 1640 cm ⁻¹; MS (FAB, 3-NOBA) 339 (100, MH+), 105 (75); HRMS (FAB, 3-NOBA) calcd for C₂₀H₂₃N₂O₃ (MH⁺) 339.1709, obsd 339.1709. Anal. Calcd for $C_{20}H_{22}N_2O_3$: C, 70.99; H, 6.55; N, 8.27. Found: C, 70.91; H, 6.71; N, 8.40.

General Procedure A. N-Benzoyl-2-benzylhomoserine Lactone (2a). All solutions were deoxygenated by freezing (liquid N_2), six cycles of evacuation, and purging with Ar. To a solution of diisopropylamine (22.4 mL, 160 mmol) and TMEDA (36.2 mL, 240 mmol) in THF (800 mL) at -78 °C was added n-butyllithium (100 mL, 1.6 M in n-hexane), and the resulting solution was stirred for 30 min at 0 °C. The LDA solution thereby generated was cooled to -78 °C and a solution of 1a24 (14.6 g, 52.0 mmol) in THF (350 mL) at -78 °C was added via cannula. The resulting bright orange solution was stirred for 1 h at -78 °C. Then the cooling bath was removed and excess ethylene oxide (ca. 25 g) was bubbled into the solution with stirring over a period of 45 min. The reaction mixture was poured into ether (500 mL) and NH₄Cl (aq, 500 mL) and vigorously stirred for 15 min. After further extraction with ether $(3 \times 300 \,\mathrm{mL})$, the combined organics were dried (MgSO₄), filtered, evaporated, and chromatographed (50% EtOAc/hexane) to give 2a (13.0 g, 85%) as a white solid: mp 170–172 °C; ¹H NMR (200 MHz, CDCl₃) δ 2.66–2.77 (m, 2 H), 3.12-3.19 (d, J = 13 Hz, 1 H), 3.22-3.29 (d, J = 13 Hz, 1 H), 3.44-3.57 (app dt, J = 8, 9 Hz, 1 H), 4.22-4.32 (app dt, J = 3, 9 Hz, 1 H), 6.61 (s, 1 H), 7.15-7.68 (m, 10 H); ¹³C NMR (50 MHz, CDCl₃) δ 33.5, 41.6, 66.0, 127.0, 127.9, 128.7, 128.9, 130.0, 132.1, 176.7, 178.1; IR (film) 3320, 1760, 1650 cm⁻¹; MS (EI) 295 (8.6, M⁺), 204 (100); HRMS (EI) calcd for C₁₈H₁₇NO₃ (M⁺) 295.1208, obsd 295.1203. Anal. Calcd for C₁₈H₁₇NO₃: C, 73.21; H, 5.80; N, 4.74. Found: C, 73.33; H, 5.95; N, 4.66.

N-Benzoyl-2-[3',4'-bis'(tert-butyldimethylsilyl)oxy]benzyl]homoserine Lactone (2b). From 1b (4.88 g, 8.98 mmol), following General Procedure A, was obtained 2b (4.18 g, 84%), as a white solid, after flash chromatography (0-50% EtOAc/hexane): mp 138-140 °C; ¹H NMR (500 MHz, CDCl₈) δ 0.17 (s, 6 H), 0.18 (s, 3 H), 0.19 (s, 3 H), 0.97 (s, 18 H), 2.73-2.80 (m, 2 H), 3.11-3.13 (d, J = 13 Hz,1 H), 3.14-3.17 (d, J = 13 Hz, 1 H), 3.52-3.55 (app dt, J = 8, 9 Hz, 1 H), 4.30-4.35 (app dt, J = 3, 9 Hz, 1 H), 6.62 (s, 1 H), 6.71-6.80 (m, 3 H), 7.40-7.43 (m,

CH₂Cl₂ (23 mL) at 0 °C was added NEt₃ (2.22 mL, 15.94 mmol). The reaction mixture was stirred for 21 h at rt and then diluted with Et₂O (150 mL) and washed with H₂O (2 × 100 mL) and brine (100 mL). The organic layer was dried (MgSO₄) and evaporated to give 1e (4.58 g, 81 % for 2 steps) as an oil: ¹H NMR (500 MHz, CDCl₃) δ 1.41–1.59 (m, 2 H), 1.63–2.07 (m, 4 H), 3.58–3.63 (m, 2 H), 3.74 (s, 3 H), 4.82–4.86 (m, 1 H), 6.68-6.70 (d, J = 8 Hz, 1 H), 7.34–7.51 (m, 6 H), 7.67–7.78 (m, 4 H), 8.25 (s, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 22.9, 30.2, 32.2, 52.3, 52.5, 61.0, 127.0, 128.0, 128.4, 128.5, 130.4, 131.6, 134.0, 136.2, 161.0, 167.0, 173.1; IR (film) 3320, 1740, 1640 cm⁻¹; MS (FAB, 3-NOBA) calcd for C₂₁H₂₆N₂O₃ (MH+) 353.1865, obsd 353.1861. Anal. Calcd for C₂₁H₂₆N₂O₃: C, 71.57; H, 6.86; N, 7.95. Found: C, 71.44; H, 6.90; N, 7.80.

 ⁽²²⁾ Campbell, J. B. J. Chem. Soc., Perkin Trans. I 1983, 1213-1217.
 (23) Nosho, Y.; Seki, T.; Kondo, M.; Ohfuji, T.; Tamura, M.; Okai, H. J. Agric. Food Chem. 1990, 38, 1368-1373.

⁽²⁴⁾ Schnyder, J.; Rothenberg, M. Helv. Chim. Acta 1975, 58, 521-523.

2 H), 7.49–7.52 (m, 1 H), 7.71–7.73 (m, 2 H); ¹³C NMR (75 MHz. CDCl₃) δ -4.17, -4.13 (2 C), -4.07, 18.4, 18.5, 25.87, 25.90, 33.2, 41.5, 60.1, 65.9, 121.3, 122.7, 122.9, 126.7, 127.0, 128.6, 132.0, 133.4, 147.2, 166.6, 177.3; IR (film) 3340, 1775, 1640 cm⁻¹; MS (FAB, 3-NOBA) 556 (58, MH+), 351 (96), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₃₀H₄₆NO₅Si₂ (MH+) 556.2914, obsd 556.2914. Anal. Calcd for C₈₀H₄₅NO₅Si₂: C, 64.83; H, 8.16; N, 2.52. Found: C, 64.64; H, 8.29; N, 2.50.

N-Benzoyl-2-[3',4'-(methylenedioxy)benzyl]homoserine Lactone (2c). From 1c (1.80 g, 5.50 mmol), following General Procedure A, was obtained 2c (1.53 g, 82%), as a white solid, following chromatography (50% EtOAc/hexane): mp 185-187 °C; ¹H NMR (300 MHz, CDCl₃) δ 2.72-2.85 (m, 2 H), 3.12-3.16 (d, J = 13 Hz, 1 H), 3.20-3.24 (d, J = 13 Hz, 1 H), 3.65-3.73 (app)dt, J = 7, 9 Hz, 1 H), 4.33-4.40 (app dt, J = 3, 9 Hz, 1 H), 5.94 (s, 2 H), 6.66 (s, 1 H), 6.68-6.77 (m, 3 H), 7.39-7.54 (m, 3 H), 7.71-7.78 (m, 2 H); ¹⁸C NMR (75 MHz, CDCl₃) δ 33.4, 41.3, 60.1, 66.1, 101.2, 108.5, 110.2, 123.3, 127.0, 127.3, 128.7, 132.1, 133.4, 147.3, 148.0, 166.8, 177.1; IR (film) 3410-3340, 1770, 1650 cm⁻¹; MS (FAB, 3-NOBA) 340 (100, MH+), 218 (33), 105 (89); HRMS (FAB, 3-NOBA) calcd for C₁₉H₁₈NO₅ (MH⁺) 340.1185, obsd 340.1185. Anal. Calcd for C₁₉H₁₇NO₅: C, 67.25; H, 5.05; N, 4.13. Found: C, 67.00; H, 5.20; N, 4.14.

N-Benzoyl-2-[(1'-trityl-4'-imidazolyl)methyl]homoserine Lactone (2d). General Procedure A was followed. except that NaHCO₃ (aq) was substituted for NH₄Cl (aq) in the workup. From 1d (1.80 g, 5.50 mmol) was obtained 2d (1.48 g, 80%), as a white solid, after flash chromatography (45:45:10 CHCl₃/hexane/acetone): mp 104-106 °C; ¹H NMR (500 MHz, CDCl₃) δ 2.18-2.23 (ddd, J = 4, 8, 13 Hz, 1 H), 2.67-2.73 (app dt, J = 9, 13 Hz, 1 H), 2.78-2.81 (d, J = 14 Hz, 1 H), 3.15-3.18 (d, J = 14 Hz, 1 H, 4.19-4.24 (app q, J = 9 Hz, 1 H), 4.56-4.61 (app q, J = 9 Hz, 1 H)dt, J = 4, 9 Hz, 1 H), 6.75 (s, 1 H), 7.08–7.18 (m, 6 H), 7.22–7.49 $(m, 13 H), 7.90-7.92 (m, 2 H), 9.18 (s, 1 H); {}^{13}C NMR (125 MHz,$ CDCl₃) δ 31.5, 33.6, 58.9, 64.9, 75.5, 120.7, 127.3, 128.1 (2 C), 128.2, 128.4, 129.7, 131.6, 133.2, 135.4, 139.1, 142.1, 166.1, 176.0; IR (film) 3380-3200, 1770, 1660 cm⁻¹; MS (FAB, 3-NOBA) 528 (100, MH+), 243 (100); HRMS (FAB, 3-NOBA) calcd for C₃₄H₃₀N₃O₃ (100, MH⁺) 528.2287, obsd 528.2298. Anal. Calcd for $C_{34}H_{29}N_3O_3$: C, 77.40; H, 5.54; N, 7.96. Found: C, 77.20; H, 5.35; N, 7.65.

N-Benzoyl-2-(4'-benzamidobutyl)homoserine Lactone (2e). Starting from 1e (4.11 g, 11.7 mmol), General Procedure A was followed, except that NaHCO3 (aq) was substituted for NH4Cl (aq) in the workup. The crude N^{ϵ} -benzylidene lactone was taken up in THF (48 mL) and 3 N HCl (48 mL), stirred at rt for 23 h, and then evaporated. The residue was taken up in CH₂Cl₂ (230 mL) and benzovi chloride (3.38 mL, 29.1 mmol) and cooled to 0 °C. After addition of NEt₃ (4.06 mL, 29.1 mmol), the reaction mixture was stirred for 15 h at rt and then poured into NaHCO₃ (300 mL) and EtOAc (300 mL). The organic layer was further washed with 1 N HCl (300 mL) and brine (300 mL), then dried (MgSO₄), filtered, and evaporated. Flash chromatography (50-75% EtOAc/hexane) yielded 2e (2.70 g, 61%) as a white solid: mp 154-155 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.51-1.69 (m, 4 H), 1.86-1.96 (m, 1 H), 2.14-2.24 (m, 1 H), 2.31-2.38 (ddd, J =3, 8, 13 Hz, 1 H), 2.82-2.93 (app dt, J = 9, 13 Hz, 1 H), 3.33-3.41(m, 1 H), 3.55-3.65 (m, 1 H), 4.24-4.32 (app dt, J = 8, 9 Hz, 1)H), 4.51-4.59 (app dt, J = 3, 9 Hz, 1 H), 6.57-6.61 (m, 1 H), 7.27-7.50 (m, 6 H), 7.55 (s, 1 H), 7.67-7.90 (m, 4 H); ¹³C NMR (75 MHz, CDCl₃) δ 19.8, 29.6, 33.4, 34.2, 38.1, 58.7, 65.2, 126.9, 127.5, 128.3, 128.5, 131.5, 131.6, 133.2, 134.3, 167.3, 168.8, 176.5; IR (film): 3390-3185, 1765, 1640 cm⁻¹; MS (FAB, 3-NOBA) 381 (61, MH+), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₂H₂₅N₂O₄ (MH⁺) 381.1814, obsd 381.1819. Anal. Calcd for $C_{22}H_{24}N_2O_4$: C, 69.46; H, 6.36; N, 7.36. Found: C, 69.24; H, 6.37;

N-Benzoyl-2-(3'-benzamidopropyl)homoserine Lactone (2f). From 1f (5.80 g, 17.1 mmol), according to the procedure for the synthesis of 1e, was obtained 2f (4.91 g, 78%), as a white solid, following flash chromatography (50-75% EtOAc/hexane): mp 179–181°C; ¹H NMR (300 MHz, CDCl₃) δ 1.77–1.98 (m, 3 H), 2.14-2.23 (m, 1 H), 2.48-2.56 (ddd, J = 3, 8, 13 Hz, 1 H), 2.76-2.87(app dt, J = 9, 13 Hz, 1 H), 3.42-3.51 (m, 1 H), 3.54-3.63 (m, 1 H), 4.26-4.35 (app dt, J = 8, 9 Hz, 1 H), 4.53-4.60 (app dt, J =3, 9 Hz, 1 H), 6.39-6.43 (m, 1 H), 7.27-7.53 (m, 6 H), 7.74-7.88

(m, 4 H); 13 C NMR [75 MHz, DMSO- d_6 /CDCl₃ (1:1)] δ 23.1, 31.9, 32.9, 39.1, 58.4, 64.5, 127.1, 127.5, 128.0 (2 C), 130.8, 131.4, 133.3, 134.5, 166.3, 166.4, 176.1; IR (film) 3360, 3290, 1750, 1640 cm⁻¹; MS (FAB, 3-NOBA) 367 (54, MH+), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₁H₂₃N₂O₄ (MH⁺) 367.1658, obsd 367.1654. Anal. Calcd for C₂₁H₂₂N₂O₄: C, 68.84; H, 6.05; N, 7.64. Found: C, 69.02; H, 6.06; N, 7.43.

N-Benzoyl-2-isopropylhomoserine Lactone (2g). From 1g25 (10.2 g, 43.2 mmol), following General Procedure A, was obtained 2g (8.97 g, 84%), as a white solid, following chromatography (20-60% EtOAc/hexane): mp 125-127 °C; ¹H NMR (200 MHz, CDCl₃) δ 0.93-0.96 (d, J = 7 Hz, 3 H), 1.08-1.11 (d, J = 7 Hz, 3 H, 2.19-2.26 (app heptet, J = 7 Hz, 1 H), 2.45-2.64(m, 2 H), 4.18-4.31 (app dt, J = 7, 9 Hz, 1 H), 4.57-4.68 (ddd, J = 4, 9, 10 Hz, 1 H), 6.60 (s, 1 H), 7.36-7.50 (m, 3 H), 7.72-7.76 (m, 2 H); ¹³C NMR (50 MHz, CDCl₃) δ 16.6 (2 C), 28.4, 33.5, 62.7, 66.0, 127.1, 128.4, 131.8, 133.1, 167.2, 177.4; IR (film) 3340, 1760, 1660 cm⁻¹; MS (EI) 247 (1.5, M⁺), 105 (100); HRMS (EI) calcd for C₁₄H₁₇NO₃ (M⁺) 247.1208, obsd 247.1203. Anal. Calcd for C₁₄H₁₇NO₃: C, 68.00; H, 6.93; N, 5.66. Found: C, 68.12; H, 6.86; N. 5.39.

N-Benzoyl-2-(2'-hydroxyethyl)homoserine Lactone. General Procedure A was followed except that, after bubbling excess ethylene oxide into the solution of the dianion, the reaction mixture was capped and stirred at rt overnight. From 1i26 (14.0 g 68.2 mmol) were obtained (recovered) 1i (2.20 g) and the title compound [9.80 g, 58% (68% based on recovered 1i)], as a white solid, following flash chromatography (50-75% EtOAc/hexane): mp 103-105 °C; ¹H NMR (500 MHz, CDCl₃) δ 1.98-2.03 (ddd, J = 2, 7, 15 Hz, 1 H), 2.10-2.15 (ddd, <math>J = 3, 8, 15 Hz, 1 H),2.32-2.37 (ddd, J = 3, 8, 13 Hz, 1 H), 2.88 (s, 1 H), 2.90-2.94 (m,1 H), 3.89-3.93 (m, 1 H), 4.08-4.12 (m, 1 H), 4.23-4.29 (m, 1 H), 4.57-4.61 (app dt, J = 3, 9, 12 Hz, 1 H), 7.37-7.40 (m, 2 H), 7.42-7.49 (m, 1 H), 7.77-7.81 (m, 2 H), 8.41 (s, 1 H); ¹⁸C NMR (125 MHz, CDCl₃) δ 33.0, 37.0, 59.08 59.13, 65.3, 127.1, 128.6, 132.0, 133.1, 166.8, 176.8; IR (film) 3670-3300, 1771, 1653 cm⁻¹; MS (EI): 249 (13, M⁺), 105 (100); HRMS (EI) calcd for C₁₈H₁₅-NO₄ (M⁺) 249.1001, obsd 249.0993. Anal. Calcd for C₁₈H₁₅NO₄: C, 62.64; H, 6.06; N, 5.62. Found: C, 62.63; H, 6.17; N, 5.50.

N-Benzoyl-2-[2'-[(tert-butyldiphenylsilyl)oxy]ethyl]homoserine Lactone (2i). To a solution of N-benzoyl-2-(2'hydroxyethyl)homoserine lactone (1.60 g, 6.42 mmol) and imidazole (960 mg, 14.1 mmol) in DMF (25 mL) was added a solution of tert-butyldiphenylsilyl chloride (1.84 mL, 7.06 mmol) in DMF (5mL). After being stirred 16 h at rt, the reaction mixture was poured into NaHCO₃ (20 mL) and extracted with Et₂O (4 × 150 mL). The combined extracts were dried (MgSO₄), filtered, evaporated, and chromatographed to give 2i (2.79 g, 89%) as a white solid: mp 132-134 °C; ¹H NMR (500 MHz, CDCl₃) δ 1.04 (s. 9 H), 1.99-2.04 (ddd, J = 3, 7, 11 Hz, 1 H), 2.22-2.27 (ddd, J = 3, 8, 11 Hz, 1 H, 2.31-2.35 (ddd, J = 3, 8, 11 Hz, 1 H),2.86-2.93 (m, 1 H), 3.92-3.96 (ddd, J = 3, 8, 11 Hz, 1 H), 4.11-4.15(ddd, J = 3, 7,11 Hz, 1 H), 4.20-4.25 (app q, J = 9 Hz, 1 H),4.57-4.62 (app dt, J = 3, 9 Hz, 1 H), 7.22-7.25 (m, 2 H), 7.32-7.45(m, 7 H), 7.61-7.71 (m, 6 H), 7.99 (s, 1 H); ¹⁸C NMR (125 MHz, $CDCl_3$) δ 19.1, 26.8, 32.5, 37.6, 59.0, 60.8, 65.0, 127.3, 127.9 (2 C), 128.4, 130.1 (2 C), 131.7, 132.4, 132.7, 133.3, 135.4, 135.4, 167.2, 176.3; IR (film) 3375, 1780, 1660 cm⁻¹; MS (FAB, 3-NOBA) 488 (45, MH+), 232 (90), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₉H₃₄NO₄Si (MH⁺) 488.2257, obsd 488.2237. Anal. Calcd for C₂₉H₃₃NO₄Si: C, 71.43; H, 6.82; N 2.87. Found: C, 71.43; H, 6.87; N, 2.71.

N-Benzoyl-2-[2'-(tert-butyloxy)ethyl]homoserine Lactone (2i). Isobutylene was bubbled into a solution of N-benzovl-2-(2'-hydroxyethyl)homoserine lactone (2.00 g, 8.02 mmol) and H₂SO₄ (0.52mL) in CH₂Cl₂ (65 mL) for 1 h. The flask was stoppered and the reaction mixture stirred at rt for 10 h. After dilution (50 mL CH₂Cl₂), the reaction was quenched with NaHCO. (aq, 100 mL). The organic layer was further extracted with H₂O (100 mL), 1 N HCl (100 mL), and H_2O (100 mL) and then dried (Na_2SO_4) , filtered, and evaporated to provide 2j (2.17 g, 89%) as an off white solid: mp 106-108 °C; ¹H NMR (500 MHz, CDCl₃)

⁽²⁵⁾ Applewhite, T. H.; Waite, H.; Niemann, C. J. Am. Chem. Soc. 1958, 80, 1465-1469

⁽²⁶⁾ Knobler, Y.; Frankel, M. J. Chem. Soc. 1958, 1629-1631.

δ 1.22 (s, 9 H), 1.95–2.00 (app dd, J = 7, 15 Hz, 1 H), 2.17–2.27 (m, 2 H), 2.90–2.97 (app dt, J = 9, 12 Hz, 1 H), 3.62–3.66 (m, 1 H), 3.84–3.87 (m, 1 H), 4.24–4.29 (app q, J = 8 Hz, 1 H), 4.59–4.64 (app dt, J = 3, 9 Hz, 1 H), 7.25–7.41 (m, 2 H), 7.47–7.50 (m, 1 H), 7.79–7.81 (m, 2 H), 8.53 (s, 1 H); 13 C NMR (75 MHz, CDCl₃) δ 27.4, 32.4, 35.7, 58.1, 58.9, 64.8, 74.4, 127.1, 128.3, 131.7, 133.3, 166.4, 176.3; IR (film) 3350, 1770, 1663 cm⁻¹; MS (EI) 305 (0.2, M⁺), 232 (28), 105 (100); HRMS (EI) calcd for C₁₇H₂₃NO₄ (M⁺) 305.1627, obsd 305.1617. Anal. Calcd for C₁₇H₂₃NO₄: C, 66.87; H, 7.59; N 4.58. Found: C, 66.83; H, 7.48; N, 4.34.

General Procedure B. Methyl N-Benzoyl-2-[2'-(phenylseleno)ethyl]phenylalaninate (3a). All solutions were deoxygenated by freezing (liquid N2), six cycles of evacuation, and purging with Ar. To an argon-purged flask containing sodium trimethoxyborohydride (563 mg, 4.41 mmol) was added a solution of diphenyl diselenide (687 mg, 2.20 mmol) in DMF (30 mL). The resulting orange suspension was heated to 65 °C for 1 h. whereby a clear, colorless solution resulted. To this solution at rt was added a solution of lactone 2a (1.00 g, 3.39 mmol) in DMF (30 mL), and the reaction mixture was heated at 110 °C (preheated oil bath) for 2 h. After the solution was cooled to rt, the solvent was removed in vacuo, and the residue was diluted with Et2O (200 mL) and brought to pH 4 with 1 N HCl (aq). The aqueous layer was extracted twice more with Et₂O (200 mL) and the combined organics were dried (MgSO₄), filtered, and esterified with diazomethane. Evaporation of the solvent and chromatography (0-10% EtOAc/hexane) yielded 3a (1.52 g, 97%) as a white solid. In another run, lactone 2a (1.23 g, 9.59 mmol) gave selenide 3a (3.10 g, 90%): mp 124-126 °C; ¹H NMR (360 MHz, CDCl₂) δ 2.24–2.32 (ddd, J = 6, 11, 14 Hz, 1 H), 2.43–2.51 (ddd, J = 6, 11, 12 Hz, 1 H), 2.73-2.79 (ddd, J = 5, 11, 12 Hz, 1 H), 2.97-3.01 (d, J = 13 Hz, 1 H), 3.14-3.21 (ddd, J = 5, 11, 14 Hz, 1 H), 3.64 (s, 3 H), 3.76-3.80 (d, J = 13 Hz, 1 H), 6.84-6.86 (m, 3 H), 7.01-7.14 (m, 6 H), 7.24-7.39 (m, 5 H), 7.51-7.55 (m, 2 H); ¹³C NMR (50 MHz, CDCl₈) δ 21.9, 36.1, 40.6, 52.8, 66.8, 126.9, 126.9, 127.0, 128.3, 128.6, 129.1, 129.3, 129.6, 131.6, 132.4, 134.9, 135.9, 166.7, 173.4; IR (film) 3410, 1735, 1663 cm⁻¹; MS (EI) 467 $(0.4, M^+)$, 310 (33), 105 (100); HRMS (EI) calcd for $C_{25}H_{25}NO_8Se$ (M+) 467.0999, obsd 467.0980. Anal. Calcd for C₂₅H₂₅NO₃Se: C, 64.38; H, 5.40; N, 3.00. Found: C, 64.42; H, 5.30; N, 2.91.

Methyl N-Benzoyl-2-[2"-(phenylseleno)ethyl]-3',4'-bis-[(tert-butyldimethylsilyl)oxy]phenylalaninate(3b). From lactone 2b (4.00 g, 720 mmol), following General Procedure B. was obtained 3b (4.24 g, 81 %) as an oil, after flash chromatography (0-10% EtOAc/hexane): ¹H NMR (300 MHz, CDCl₃) δ 0.028 (s, 3 H), 0.031 (s, 3 H), 0.16 (s, 6 H), 0.88 (s, 9 H), 0.95 (s, 9 H), 2.33-2.43 (ddd, J = 5, 11, 14 Hz, 1 H), 2.55-2.65 (ddd, J = 5, 11, 12 Hz, 1 H), 2.85-2.94 (ddd, J = 5, 11, 12 Hz, 1 H), 2.99-3.04 (d,J = 14 Hz, 1 H), 3.24-3.34 (ddd, J = 5, 11, 14 Hz, 1 H), 3.77 (s, 3 H), 3.76-3.80 (d, J = 14 Hz, 1 H), 6.43-6.65 (m, 3 H), 7.06 (s, 1 H), 7.21-7.28 (m, 3 H), 7.38-7.53 (m, 5 H), 7.67-7.71 (m, 2 H); ¹³C NMR (75 MHz, CDCl₃) δ -4.31, -4.26, -4.1 (2 C), 18.2, 18.4, 21.8, 25.7, 25.9, 36.0, 40.0, 52.7, 66.7, 120.6, 122.3, 122.6, 126.8, 126.9, 128.5, 128.9, 129.0, 130.1, 131.5, 132.2, 134.8, 145.9, 146.4, 166.4,.173.4; IR (film) 3410, 1735, 1670 cm⁻¹; MS (FAB, 3-NOBA) 728 (6, MH+), 351 (72), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₃₇H₅₄NO₅Si₂Se (MH⁺)'728.2705, obsd 728.2681. Anal. Calcd for C₃₇H₅₃NO₅Si₂Se: C, 61.13; H, 7.35; N, 1.93. Found: C, 61.07; H, 7.38; N, 1.88.

Methyl N-Benzoyl-2-[2"-(phenylseleno)ethyl]-3',4'-(methylenedioxy)phenylalaninate (3c). From lactone 2c (500 mg, 1.47 mmol), following General Procedure B, was obtained 3c (621 mg, 83%), as a white solid, after flash chromatography (0-10% EtOAc/hexane): mp 110-111 °C; ¹H NMR (300 MHz, CDCl₃) δ 2.31-2.41 (ddd, J = 6, 11, 14 Hz, 1 H), 2.52-2.62 (ddd, J = 6, 11, 12 Hz, 1 H), 2.83-2.92 (ddd, J = 5, 11, 12 Hz, 1 H), 2.98-3.03 (d, J = 14 Hz, 1 H), 3.22-3.32 (ddd, J = 5, 11, 14 Hz, 1 H), 3.77 (s, 3 H), 3.81-3.86 (d, J = 14 Hz, 1 H), 5.86 (s, 2 H), 6.42-6.61 (m, 3 H), 7.02 (s, 1 H), 7.21-7.27 (m, 3 H), 7.39-7.53 (m, 5 H), 7.67-7.71 (m, 2 H); 13 C NMR (75 MHz, CDCl₃) δ 21.9, 36.0, 40.3, 52.9, 67.0, 100.9, 108.1, 109.7, 122.8, 126.8, 126.9, 128.6, 129.0 (2 C), 129.4, 131.6, 132.3, 134.9, 146.5, 147.5, 166.7, 173.4; IR (film) 3410, 1730, 1660 cm⁻¹; MS (FAB, 3-NOBA) 512 (14, MH+), 354 (47), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₆H₂₆NO₅Se (MH⁺) 512.0979, obsd 512.0977. Anal. Calcd for

C₂₆H₂₆NO₆Se: C, 61.18; H, 4.94; N, 2.74. Found: C, 61.06; H, 4.98; N, 2.72.

Methyl Na-Benzoyl-2-[2'-(phenylseleno)ethyl]-Na-tritylhistidinate (3d). General Procedure B was followed except that 100 mM NaOAc/HOAc buffer (aq, pH 5) was substituted for 1 N HCl in the acidification step. From lactone 2d (1.44 g, 2.73 mmol) was obtained 3d (1.32 g, 69%) as a white solid, after flash chromatography (0-50% EtOAc/hexane): mp 66-68 °C; 1H NMR (500 MHz, CDCl₃) δ 2.46-2.51 (m, 1 H), 2.68-2.74 (m, 1 H), 2.85-2.96 (m, 2 H), 3.10-3.13 (d, J = 14 Hz, 1 H), 3.62-3.65 (d, J = 14 Hz, 1 Hz), 3.62-3.65 (d, J = 14 Hz), 3.62-3.6514 Hz, 1 H), 3.73 (s, 3 H), 6.50 (s, 1 H), 6.97-6.98 (m, 6 H), 7.14-7.29 (m, 12 H), 7.34-7.49 (m, 6 H), 7.63-7.67 (m, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 21.9, 34.1, 36.1, 52.8, 64.8, 76.3, 120.5, 126.7, 127.0, 128.0 (2 C), 128.03, 128.5, 129.0, 129.6, 130.4, 131.4, 132.1, 134.5, 138.4, 142.1, 166.2, 173.6; IR (film) 3415-3240, 1735, 1665 cm⁻¹; MS (FAB, 3-NOBA) 700 (4, MH⁺), 243 (100); HRMS (FAB, 3-NOBA) calcd for $C_{41}H_{38}N_3O_3Se700.2078$ (MH+), obsd 700.2062. Anal. Calcd for C41H37N3O3Se: C, 70.48; H, 5.34; N, 6.01. Found: C, 70.34; H, 5.40; N, 5.86.

Methyl N^{α} , N^{ϵ} -Dibenzoyl-2-[2'-(phenylseleno)ethyl]lysinate (3e). From lactone 2e (421 mg, 1.11 mmol), following General Procedure B, was obtained 3e (539 mg, 88%), as a white solid, after flash chromatography (0-25% EtOAc/hexane). In another run, lactone 2e (8.00 g, 21.0 mmol) gave selenide 3e (9.90 g, 85%): mp 48-49 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.04-1.10 (m, 1 H), 1.32-1.68 (m, 3 H), 1.78-1.88 (m, 1 H), 2.21-2.31 (ddd, J = 6, 11, 14 Hz, 1 H), 2.53-2.63 (ddd, J = 6, 11, 12 Hz, 1 H),2.70-2.94 (m, 2 H), 2.97-3.06 (ddd, J = 5, 11, 14 Hz, 1 H), 3.32-3.46 (m, 2 H), 3.76 (s, 3 H), 6.21-6.25 (m, 1 H), 7.19-7.24 (m, 3 H), 7.30 (s, 1 H), 7.31-7.53 (m, 8 H), 7.64-7.77 (m, 4 H); 18 C NMR (75 MHz, CDCl₃) δ 21.0, 21.8, 28.8, 34.3, 36.3, 39.0, 53.1, 65.4, 126.8, 126.9, 127.0, 128.4, 128.6, 128.9, 129.0, 131.2, 131.7, 132.3, 134.5, 134.6, 166.3, 167.5, 174.3; IR (film) 3410-3300, 1735, 1640 cm⁻¹; MS (FAB, 3-NOBA) 553 (16, MH⁺), 395 (50), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₉H₃₃N₂O₄Se (MH⁺) 553.1606, obsd 553.1613. Anal. Calcd for C₂₉H₃₂N₂O₄Se: C, 63.15; H, 5.85; N, 5.08. Found: C, 63.20; H, 5.75; N, 4.98.

Methyl N°,N°-Dibenzoyl-2-[2'-(phenylseleno)ethyl]ornithinate (3f). From lactone 2f (600 mg, 1.64 mmol), following General Procedure B, was obtained 3f (695 mg, 79%), as a white solid, after flash chromatography (0-25% EtOAc/hexane). In another run, lactone 2f (4.19 g, 11.4 mmol) gave selenide 3f (4.43 g, 72%): mp 180-182 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.40-1.53 (m, 2 H), 1.88-1.98 (ddd, J = 6, 11, 14 Hz, 1 H), 2.20-2.30 (ddd, J = 6, 11, 14 Hz, 1 H)J = 6, 11, 14 Hz, 1 H), 2.53-2.63 (ddd, <math>J = 6, 11, 12 Hz, 1 H),2.76-2.89 (m, 2 H), 2.98-3.08 (ddd, J = 5, 11, 14 Hz, 1 H), 3.28-3.083.48 (m, 2 H), 3.77 (s, 3 H), 6.27-6.31 (m, 1 H), 7.18-7.24 (m, 3 H), 7.33 (s, 1 H), 7.35-7.54 (m, 8 H), 7.73-7.79 (m, 4 H); 13 C NMR (75 MHz, CDCl₃) δ 21.7, 23.9, 32.2, 36.2, 39.3, 53.2, 65.2, 126.8, 126.86, 126.91, 128.4, 128.6, 129.0, 129.9, 131.3, 131.8, 132.2, 134.2, 134.4, 166.3, 167.5, 174.1; IR (film) 3330, 1735, 1640 cm⁻¹; MS (FAB, 3-NOBA) 539 (MH+, 10), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₈H₃₁N₂O₄Se 539.1452, obsd 539.1436. Anal. Calcd for C₂₈H₃₀N₂O₄Se: C, 62.57; H, 5.62; N, 5.21. Found: C, 62.70; H, 5.80; N, 5.37.

Methyl N-Benzoyl-2-[2'-(phenylseleno)ethyl]valinate (3g). From lactone 2g (500 mg, 2.02 mmol), following General Procedure B, was obtained 3g (814 mg, 96%) as an oil after flash chromatography (0-10% EtOAc/hexane). In another run, lactone 2g (2.40 g, 9.71 mmol) gave selenide 3g (3.57 g, 88%): 1H NMR $(500 \text{ MHz}, \text{CDCl}_3) \delta 0.89-0.90 \text{ (d, } J = 7 \text{ Hz, } 3 \text{ H), } 0.99-1.00 \text{ (d, }$ J = 7 Hz, 3 H, 2.44-2.50 (ddd, J = 5, 11, 14 Hz, 1 H, 2.55-2.61(ddd, J = 5, 11, 12 Hz, 1 H), 2.75-2.81 (m, 1 H), 2.88-2.93 (ddd, J)J = 5, 11, 12 Hz, 1 H), 3.22-3.28 (ddd, <math>J = 5, 11, 14 Hz, 1 H), 3.75(s, 3 H), 7.18-7.24 (m, 3 H), 7.29 (s, 1 H), 7.42-7.52 (m, 5 H), 7.77-7.78 (m, 2 H); ¹³C NMR (50 MHz, CDCl₃) δ 17.7, 18.1, 22.7, 33.2, 33.8, 52.7, 69.0, 126.7, 126.8, 128.6, 129.0, 130.5, 131.6, 132.1,135.0, 166.0, 173.7; IR (film) 3400, 1730, 1660 cm⁻¹; MS (EI) 419 (1.3, M⁺), 262 (100); HRMS (EI) calcd for C₂₁H₂₅NO₃Se (M⁺) 419.0999, obsd 419.1001. Anal. Calcd for C₂₁H₂₅NO₈Se: C, 60.29; H, 6.02; N, 3.35. Found: C, 60.31; H, 6.11; N, 3.10.

Methyl N-Benzoyl-4-O-(tert-butyldiphenylsilyl)-2-[2'-(phenylseleno)ethyl]homoserinate (3i). From lactone 2i (3.15 g, 6.46 mmol), following General Procedure B, was obtained 3i (3.83 g, 90%), as a colorless oil, after flash chromatography (0–10% EtOAc/hexane): ¹H NMR (500 MHz, CDCl₃) δ 0.99 (s, 9 H),

2.28–2.38 (m, 2 H), 2.58–2.64 (app dt, J=5, 12 Hz, 1 H), 2.71–2.75 (app dt, J=4, 8 Hz, 1 H), 2.83–2.88 (app dt, J=5, 12 Hz, 1 H), 3.04–3.11 (ddd, J=5, 11, 14 Hz, 1 H), 3.62–3.66 (m, 2 H), 3.68 (s, 3 H), 7.21–7.59 (m, 19 H), 7.76–7.78 (m, 2 H); ¹³C NMR (125 MHz, CDCl₃) δ 19.1, 21.5, 26.8, 36.6, 37.4, 52.9, 60.1, 63.3, 126.8, 126.9, 127.6 (2 C), 127.7, 128.6, 129.0, 129.6, 131.6, 132.2 (2 C), 133.16, 133.21, 134.5, 135.4, 135.5, 166.2, 174.3; IR (film) 3410, 1730, 1665 cm⁻¹; MS (EI) 659 (1.0, M⁺), 602 (41), 105 (100); HRMS (EI) calcd for C₃₆H₄₁NO₄SiSe (M⁺) 659.1970, obsd 659.1941. Anal. Calcd for C₃₆H₄₁NO₄SiSe: C, 65.64; H, 6.27; N, 2.13. Found: C, 65.79; H, 6.46; N, 1.88.

Methyl N-Benzoyl-4-O-tert-butyl-2-[2'-(phenylseleno)ethyl]homoserinate (3j). From lactone 2j (3.80 g, 12.4 mmol), following General Procedure B, was obtained 3j (5.52 g, 93%), as a white solid, after flash chromatography (0-10% EtOAc/hexane): mp. 65-67 °C; IR (film) 3410, 1730, 1665 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.07 (s, 9 H), 2.13-2.19 (ddd, J =5, 9, 14 Hz, 1 H), 2.29-2.35 (ddd, J = 5, 12, 14 Hz, 1 H), 2.60-2.65(app dt, J = 5, 12 Hz, 1 H), 2.66-2.71 (app dt, J = 3, 7 Hz, 1 H),2.81-2.87 (app dt, J = 5, 12 Hz, 1 H), 2.92-2.99 (ddd, J = 5, 12, 14 Hz, 1 H), 3.32-3.37 (m, 2 H), 3.74 (s, 3 H), 7.18-7.22 (m, 3 H), 7.40-7.45 (m, 5 H), 7.63 (s, 1 H), 7.80-7.82 (m, 2 H); ¹⁸C NMR (125 MHz, CDCl₃) δ 22.2, 27.8, 36.1, 37.1, 53.1, 57.7, 63.9, 73.6, 127.4, 127.5, 129.3, 129.7, 132.2, 132.79, 132.83, 135.2, 166.6, 175.0; MS (EI) 477 (3, M+), 320 (68), 105 (100); HRMS (EI) calcd for C24H31NO4Se 477.1418, obsd 477.1409. Anal. Calcd for C24H31NO4Se: C, 60.50; H, 6.56; N, 2.94. Found: C, 60.28; H, 6.59; N, 2.83.

Methyl N-Benzoyl-2-[2'-(phenylseleno)ethyl]glycinate (3k). From $1i^{23}$ (400 mg, 1.95 mmol), according to General Procedure B, was obtained 3k (623 mg, 85%) as a white crystalline solid after flash chromatography (0–30% EtOAc/hexane): mp 96–98 °C; ¹H NMR (300 MHz, CDCl₃) δ 2.14–2.23 (m, 1 H), 2.32-2.43 (m, 1 H), 2.87–2.99 (m, 2 H), 3.75 (s, 3 H), 4.89–4.96 (app dt, J = 5, 8 Hz, 1 H), 6.85–6.87 (d, J = 7 Hz, 1 H), 7.23–7.25 (m, 3 H), 7.40–7.49 (m, 5 H), 7.77–7.80 (m, 2 H); 13 C NMR (125 MHz, CDCl₃) δ 23.1, 33.1, 52.6, 52.7, 127.0, 127.2, 128.6, 129.1, 129.5, 131.9, 132.8, 133.6, 167.1, 172.4; IR (film) 3240–3390, 1740, 1640 cm⁻¹; MS (EI) 377 (6, M⁺); 220 (99); 105 (100); HRMS (EI) calcd for $C_{18}H_{19}NO_3$ Se 377.0530, obsd 377.0527. Anal. Calcd for $C_{18}H_{19}NO_3$ Se: C, 57.45 H, 5.09; N, 3.72. Found: C, 57.62; H, 5.15; N, 3.61.

Methyl 4-(Phenylseleno)butanoate (31). From diphenyl diselenide (1.27 g, 4.07 mmol), sodium trimethoxyborohydride (1.04 g, 8.14 mmol), and γ -butyrolactone (447 μ L, 5.82 mmol), according to General Procedure B, was obtained 31^{14a} (1.37 g, 92%) after flash chromatography (100% hexane).

General Procedure C. Methyl N-Benzoyl-2-vinylphenylalaninate (4a). Ozone was bubbled into a solution of selenide 3a (6.84 g, 14.7 mmol) in CH₂Cl₂ (150 mL) at -78 °C until a light blue color persisted. After addition of 1-hexene (31.7 mL, 255 mmol), this cold solution was added dropwise to refluxing CCl4 (or PhH) (500 mL) and refluxing was continued for 30 min. Evaporation and flash chromatography (0-10% EtOAc/hexane) yielded 4a (4.48g, 99%) as a white solid: mp 116-117 °C; IR (film) 3410-3265, 1740, 1650 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 3.40–3.45 (d, J = 14 Hz, 1 H), 3.82 (s, 3 H), 3.89–3.93 (d, J = 14 Hz, 1 H), 5.28-5.34 (d, J = 17 Hz, 1 H), 5.32-5.35 (d, J = 11Hz, 1 H), 6.14-6.24 (dd, J = 11, 17 Hz, 1 H), 6.99 (s, 1 H), 7.06-7.09 (m, 2 H), 7.19-7.22 (m, 3 H), 7.39-7.50 (m, 3 H), 7.69-7.72 (m, 2 H); ¹³C NMR (50 MHz, CDCl₃) δ 40.0, 53.0, 65.9, 116.3, 126.9, 127.1, 128.3, 128.6, 130.0, 131.6, 134.8, 135.7, 136.3, 166.4. 172.3; MS (EI) 309 (3, M+), 218 (53), 105 (100); HRMS (EI) calcd for C₁₈H₁₉NO₃ (M⁺) 309.1365, obsd 309.1362. Anal. Calcd for C₁₉H₁₉NO₃: C, 73.77; H, 6.19; N, 4.52. Found: C, 73.89; H, 6.24;

Methyl N-Benzoyl-2-vinyl-3',4'-bis[(tert-butyldimethyl-silyl)oxy]phenylalaninate (4b). From selenide 3b (825 mg, 1.14 mmol), following General Procedure C, was obtained 4b (627 mg, 97%), as a white solid, after flash chromatography (10% EtoAc/hexane): mp 77–78 °C; ¹H NMR (300 MHz, CDCl₃) δ 0.05 (s, 3 H), 0.06 (s, 3 H), 0.14 (s, 3 H), 0.15 (s, 3 H), 0.89 (s, 9 H), 0.94 (s, 9 H), 3.27–3.32 (d, J=14 Hz, 1 H), 3.68–3.73 (d, 14 Hz, 1 H), 3.81 (s, 3 H), 5.23–5.29 (d, J=17 Hz, 1 H), 5.32 (d, J=11 Hz, 1 H), 6.12–6.22 (dd, J=11, 17 Hz, 1 H), 6.52–6.68 (m, 3 H), 6.98 (s, 1 H), 7.36–7.48 (m, 3 H), 7.68–7.71 (m, 2 H); 13 C

NMR (75 MHz, CDCl₃) δ -4.24, -4.2, -4.14, -4.1, 18.2, 18.4, 25.8, 25.9, 39.8, 52.9, 65.6, 116.0, 120.7, 122.7, 123.0, 127.0, 128.5, 128.6, 131.5, 134.6, 136.4, 146.0, 146.5, 166.1, 172.4; IR (film) 3415–3250, 1740, 1660 cm⁻¹; MS (FAB, 3-NOBA) 570 (21, MH+), 351 (100); HRMS (FAB, 3-NOBA) calcd for $C_{31}H_{48}NO_5Si_2$ (MH+) 570.3071, obsd 570.3060. Anal. Calcd for $C_{31}H_{47}NO_5Si_2$: C, 65.34; H, 8.31; N, 2.45. Found: C, 65.12; H, 8.15; N, 2.31.

Methyl N-Benzoyl-2-vinyl-3',4'-(methylenedioxy)phenylalaninate (4c). From selenide 3c (500 mg, 0.979 mmol), following General Procedure C, was obtained 4c (328 mg, 95%), as a white solid, after flash chromatography (10% EtOAc/ hexane): mp 89-90 °C; IR (film) 3400-3320, 1740, 1650 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 3.31–3.34 (d, J = 14 Hz, 1 H), 3.82 (s, 3 H), 3.82-3.85 (d, J = 14 Hz, 1 H), 5.28-5.32 (d, J = 17 Hz, 1 H), 5.31-5.33 (d, J = 11 Hz, 1 H), 5.88 (s, 2 H), 6.13-6.19 (dd, J = 11, 17 Hz, 1 H, 6.54-6.65 (m, 3 H), 7.03 (s, 1 H), 7.40-7.50(m, 3 H), 7.72-7.74 (m, 2 H); ¹³C NMR (75 MHz, CDCl₈) & 39.7, 53.0, 65.8, 100.8, 108.0, 110.0, 116.2, 123.0, 126.8, 128.5, 129.1, 131.6, 134.6, 136.1, 146.5, 147.4, 166.3, 172.2; MS (FAB, 3-NO-BA): 354 (53, MH+), 232 (18), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₀H₂₀NO₅ (MH⁺) 354.1341, obsd 354.1331. Anal. Calcd for C₂₀H₁₉NO₅: C, 67.98; H, 5.42; N, 3.96. Found: C, 67.92; H, 5.52; N, 3.76.

Methyl N^α-Benzoyl-N⁷-trityl-2-vinylhistidinate (4d). A solution containing 3d (995 mg, 1.43 mmol) and tetrabutylammonium periodate (803 mg, 1.85 mmol) in CHCl₃ (25 mL) was refluxed for 5 h and then evaporated and chromatographed (0-50% EtOAc/hexane) to give 4d (618 mg, 80%) as a white solid: mp 73-75 °C; ¹H NMR (500 MHz, CDCl₃) δ 3.09-3.12 (d, J = 14Hz, 1 H), 3.25-3.27 (d, J = 14 Hz, 1 H), 3.75 (s, 3 H), 4.98-5.01(d, J = 17 Hz, 1 H), 5.12-5.15 (d, J = 11 Hz, 1 H), 6.29-6.34 (dd, J = 11 Hz, 1 H)J = 11, 17 Hz, 1 H, 6.55 (s, 1 H), 7.05-7.09 (m, 6 H), 7.24-7.34(m, 9 H), 7.39-7.49 (m, 4 H), 7.89-7.91 (m, 2 H), 8.79 (s, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 35.4, 52.7, 64.4, 75.3, 114.5, 120.8, 127.3, 128.0, 128.1, 128.4, 129.7, 131.4, 134.2, 135.7, 135.9, 138.5, 142.2, 165.8, 172.6; IR (film) 3360-3270, 1740, 1670 cm⁻¹; MS (FAB, 3-NOBA) 542 (2, MH+), 243 (100); HRMS (FAB, 3-NOBA) calcd for C₃₅H₃₂N₃O₃ (MH⁺) 542.2443, obsd 542.2424. Anal. Calcd for C₃₈H₃₁N₃O₃: C, 77.61; H, 5.77; N, 7.76. Found: C, 77.48; H, 5.65; N. 7.65.

Methyl N^{α} , N^{α} . Dibenzoyl-2-vinyllysinate (4e). From selenide 3e (9.50 g, 17.2 mmol), following General Procedure C, was obtained 4e (6.70 g, 99%), as a white solid, after flash chromatography (10-50% EtOAc/hexane): mp 57-58 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.16–1.27 (m, 1 H), 1.38–1.74 (m, 3 H), 2.08-2.18 (app dt, J = 5, 13 Hz, 1 H), 2.62-2.72 (app dt, J = 4, 13 Hz, 1 H), 3.37-3.54 (m, 2 H), 3.79 (s, 3 H), 5.22-5.26 (d, J =11 Hz, 1 H), 5.23-5.29 (d, J = 17 Hz, 1 H), 6.08-6.17 (dd, J = 11, 17 Hz, 1 H), 6.40-6.43 (m, 1 H), 7.25 (s, 1 H), 7.26-7.52 (m, 6 H), 7.67-7.83 (m, 4 H); ¹⁸C NMR (75 MHz, CDCl₃) δ 20.8, 28.9, 34.0, 38.7, 53.1, 65.0, 115.2, 126.9, 127.0, 128.4, 128.6, 131.2, 131.6, 134.3, 134.5, 136.6, 166.4, 167.7, 172.9; IR (film) 3410-3220, 1740, 1640 cm-1; MS (FAB, 3-NOBA) 395 (37, MH+), 105 (100); HRMS (FAB, 3-NOBA) calcd for $C_{23}H_{27}N_2O_4$ (MH⁺) 395.1971, obsd 395.1970. Anal. Calcd for C23H26N2O4: C, 70.04; H, 6.64; N, 7.10. Found: C, 69.90; H, 6.59; N, 6.91.

Methyl N°,N°-Dibenzoyl-2-vinylornithinate (4f). From selenide 3f (3.85 g, 7.16 mmol), according to General Procedure C, was obtained 4f (2.71 g, 100%) as a white solid, after flash chromatography (10–50% EtOAc/hexane): mp 148–149 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.53–1.61 (m, 2 H), 2.19–2.29 (ddd, $J=7,\ 10,\ 14$ Hz, 1 H), 2.76–2.86 (ddd, $J=6,\ 10,\ 14$ Hz, 1 H), 3.36–3.56 (m, 2 H), 3.81 (s, 3 H), 5.25–5.29 (d, J=11 Hz, 1 H), 5.27–5.32 (d, J=17 Hz, 1 H), 6.03–6.12 (dd, $J=11,\ 17$ Hz, 1 H), 6.39–6.43 (m, 1 H), 7.26–7.55 (m, 7 H), 7.77–7.84 (m, 4 H); ¹³C NMR (75 MHz, CDCl₃) δ 23.8, 32.1, 39.4, 53.1, 64.7, 115.5, 126.9, 126.9, 128.3, 128.5, 131.1, 131.7, 133.9, 134.3, 136.2, 166.4, 167.6, 172.6; IR (film) 3380–3270, 1735, 1635 cm⁻¹; MS (FAB, 3-NOBA) 381 (39, MH+), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₂₂H₂₅N₂O₄ (MH+) 381.1814, obsd 381.1820. Anal. Calcd for C₂₂H₂₄N₂O₄: C, 69.46; H, 6.36; N, 7.36. Found: C, 69.25; H, 6.30; N, 7.26

Methyl N-Benzoyl-2-vinylvalinate (4g). From selenide 3g (1.50 g, 3.59 mmol), according to General Procedure C, was obtained 4g (926 mg, 99%) as a white solid, after flash chromatography (10% EtOAc/hexane): mp 80-81 °C; ¹H NMR

(500 MHz, CDCl₃) δ 0.96–0.98 (d, J = 7 Hz, 3 H), 1.00–1.01 (d, J = 7 Hz, 3 H), 2.37–2.39 (app heptet, J = 7 Hz, 1 H), 3.77 (s, 3 H), 5.12–5.15 (d, J = 17 Hz, 1 H), 5.26–5.28 (d, J = 11 Hz, 1 H), 6.33–6.39 (dd, J = 11, 17 Hz, 1 H), 6.62 (s, 1 H), 7.41–7.44 (m, 2 H), 7.49–7.50 (m, 1 H), 7.79–7.80 (m, 2 H); ¹³C NMR (50 MHz, CDCl₃) δ 17.2, 17.8, 35.1, 52.5, 67.3, 115.3, 127.0, 128.6, 131.7, 133.2, 134.3, 166.3, 172.4; IR (film) 3370–3270, 1740, 1650 cm⁻¹; MS (FAB, 3-NOBA) 262 (61, MH⁺), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₁₅H₁₉NO₃: C, 68.94; H, 7.33; N, 5.36. Found: C, 68.76; H, 7.47; N, 5.28.

Methyl N-Benzoyl-4-O-(tert-butyldiphenylsilyl)-2-vinylhomoserinate (4i). From selenide 3i (2.10 g, 3.19 mmol), according to General Procedure C, was obtained 4i (1.57 g, 99%) as a white solid, after flash chromatography (100% hexane): mp 83–85 °C; ¹H NMR (500 MHz, CDCl₃) δ 1.01 (s, 9 H), 2.37–2.41 (app dt, J = 4, 14.5 Hz, 1 H), 2.48-2.53 (m, 1 H), 3.74 (s, 3 H), 3.75-3.81 (m, 2 H), 5.24-5.28 (d, J = 17 Hz, 1 H), 5.26-5.28 (d, $J = 11 \text{ Hz}, 1 \text{ H}, 6.16-6.21 (dd, } J = 11, 17 \text{ Hz}, 1 \text{ H}, 7.27-7.48$ (m, 9 H), 7.58-7.63 (m, 4 H), 7.76-7.77 (m, 2 H), 7.88 (s, 1 H); ¹³C NMR (75 MHz, CDCl₃) δ 19.1, 26.9, 37.6, 52.9, 60.6, 63.7, 115.1, 127.1, 127.7, 127.7, 128.5, 129.8 (2 C), 131.5, 132.8, 132.9, 134.2, 135.4, 135.4, 136.0, 166.3, 173.0; IR (film) 3410, 1740, 1670 cm-1; MS (FAB, 3-NOBA) 502 (11, MH+), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₈₀H₃₅NO₄Si (MH⁺) 502.2413, obsd 502.2395. Anal. Calcd for C₃₀H₃₅NO₄Si: C, 71.82; H, 7.03; N, 2.79. Found: C, 71.60; H, 7.14; N, 2.60.

Methyl N-Benzoyl-4-O-tert-butyl-2-vinylhomoserinate (4j). From selenide 3j (1.50 g, 3.15 mmol), according to General Procedure C, was obtained 4j (1.00 g, 100%) as a white solid, after flash chromatography (0–10% EtOAc/hexane): mp 74–76 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.16 (s, 9 H), 2.15–2.22 (m, 1 H), 2.30–2.40 (m, 1 H), 3.48–3.54 (m, 2 H), 3.76 (s, 3 H), 5.16–5.22 (d, J=17 Hz, 1 H), 5.24–5.27 (d, J=11 Hz, 1 H), 6.17–6.27 (dd, J=11, 17 Hz, 1 H), 7.39–7.52 (m, 3 H), 7.84–7.87 (m, 2 H), 8.31 (s, 1 H); ¹³C NMR (125 MHz, CDCl₃) δ 27.3, 36.1, 52.6, 57.7, 63.9, 73.6, 114.8, 127.1, 128.4, 131.5, 134.2, 135.6, 165.8, 173.0; IR (film) 3370, 1740, 1670 cm⁻¹; MS (FAB, 3-NOBA) 320 (52, MH⁺), 246 (53), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₁₈H₂₆NO₄: C, 67.69; H, 7.89; N, 4.39. Found: C, 67.69; H, 7.95; N, 4.29.

α-Vinylphenylalanine (5a). A suspension of 4a (500 mg, 1.62 mmol) in 6 N HCl (13 mL) was refluxed for 4 h. After extraction with CH₂Cl₂ (25 mL), the aqueous layer was evaporated in vacuo with mild heating (T < 50 °C). The residue was applied to a Dowex 50 × 8 ion exchange column. After the column was washed with several volumes of H₂O, elution with 1.3 M NH₄OH afforded 5a (247 mg, 80%): ¹H NMR (500 MHz, D₂O) δ 2.99–3.04 (d, J = 14 Hz, 1 H), 5.32–5.36 (d, J = 14 Hz, 1 H), 5.19–5.25 (d, J = 17 Hz, 1 H), 5.32–5.36 (d, J = 11 Hz, 1 H), 6.05–6.15 (dd, J = 11, 17 Hz, 1 H), 7.17–7.20 (m, 2 H), 7.27–7.31 (m, 3 H); ¹³C NMR (50 MHz, D₂O) δ 41.1, 65.9, 116.3, 127.6, 128.6, 129.9, 133.3, 134.9, 173.3; MS (EI) 191 (1, M⁺), 146 (33), 100 (100); HRMS (EI) calcd for C₁₁H₁₃NO₂ (M⁺) 191.0946, obsd 191.0942. Anal. Calcd for C₁₁H₁₃NO₂: C, 69.09; H, 6.85; N, 7.32. Found: C, 68.87; H, 6.84; N, 7.10.

α-Vinyl-3',4'-dihydroxyphenylalanine Hydrochloride Salt (5b). A suspension of 4b (100 mg, 0.175 mmol) in 6 N HCl (3 mL) was heated at 150 °C in a sealed glass vessel for 4 h. The cooled (rt) reaction mixture was extracted with CH₂Cl₂ (50 mL) and the aqueous layer evaporated in vacuo to provide 5b (34.8 mg, 77%) as a tan solid [$R_f = 0.31, n$ -BuOH/AcOH/H₂O (4:1:1)]: ¹H NMR (500 MHz, D₂O) δ 3.06–3.08 (d, J = 14 Hz, 1 H), 3.39–3.42 (d, J = 14 Hz, 1 H), 5.40–5.44 (d, J = 18 Hz, 1 H), 5.53–5.55 (d, J = 11 Hz, 1 H), 6.20–6.26 (dd, J = 11, 18 Hz, 1 H), 6.74–6.94 (m, 3 H); ¹³C NMR (125 MHz, D₂O): δ 41.5, 66.8, 117.3, 118.2, 118.8, 123.6, 126.7, 135.4, 144.6, 145.0, 174.0; HRMS (FAB, 3-NOBA) calcd for C₁₁H₁₄NO₄ (HCl salt - Cl⁻) 224.0923, obsd 224.0920. Anal. Calcd for C₁₁H₁₄NO₄Cl: C, 50.88; H, 5.43; N, 5.39. Found: C, 50.70; H, 5.46; N, 5.15.

α-Vinylhistidine (5d). A suspension of 4d (200 mg, 0.369 mmol) in 6 N HCl (4 mL) was heated at 150 °C in a sealed glass vessel for 5 h and purified as for 5a to give 5d (48.7 mg, 73%) as a white solid: mp 179–181 °C; ¹H NMR (300 MHz, D_2O) δ 3.06–3.12 (d, J=15 Hz, 1 H), 3.32–3.37 (d, J=15 Hz, 1 H), 5.31–5.37 (d, J=18 Hz, 1 H), 5.40–5.43 (d, J=11 Hz, 1 H),

6.10–6.19 (dd, J = 11, 18 Hz, 1 H), 7.01 (s, 1 H), 7.70 (s, 1 H); 13 C NMR (75 MHz, D₂O) δ 33.9, 66.4, 117.6, 118.2, 131.9, 135.8, 136.8, 174.7; MS (FAB, 3-NOBA) 182 (100, MH⁺); HRMS (FAB, 3-NOBA) calcd for C₈H₁₂N₃O₂ (MH⁺) 182.0929, obsd 182.0932. Anal. Calcd for C₈H₁₁N₃O₂: C, 53.04; H, 6.12; N, 23.18. Found: C, 53.20; H, 6.30; N, 23.00.

α-Vinyllysine (5e). A suspension of 4e (4.00 g, 10.1 mmol) in 6 N HCl (80 mL) was heated at 150 °C in a sealed glass vessel for 4.5 h. Purification was as for 5a, except that elution from the Dowex 50 column was with a linear gradient of NH₄OH (0–1.3 M) to provide 5e (1.05 g, 60%) as a white solid: mp 130–132 °C; ¹H NMR (300 MHz, D_2O) δ 1.22–1.32 (m, 1 H), 1.37–1.48 (m, 1 H), 1.59–1.72 (m, 3 H), 1.84–1.94 (app dt, J=5, 14 Hz, 1 H), 2.95–3.00 (app t, J=7 Hz, 2 H), 5.17–5.21 (d, J=11 Hz, 1 H), 5.19–5.25 (d, J=18 Hz, 1 H), 6.02–6.11 (dd, J=11, 18 Hz, 1 H); ¹³C NMR (125 MHz, D_2O) δ 20.1, 26.6, 36.6, 38.8, 63.7, 113.6, 139.3, 178.3; MS (FAB, 3-NOBA) 173 (MH+, 100); HRMS (FAB, 3-NOBA) calcd for $C_8H_{17}N_2O_2$ (MH+) 173.1290, obsd 173.1294.

α-Vinylornithine (5f). From 4f (96.2 mg, 0.263mmol) in 6 N HCl (3 mL), according to the procedure for 5e, was obtained 5f (33.8 mg, 84%): mp 149–150 °C; ¹H NMR (500 MHz, D₂O) δ 1.63–1.69 (m, 1 H), 1.74–1.80 (m, 2 H), 1.96–2.01 (m, 1 H), 3.03–3.06 (app t, J=7 Hz, 2 H), 5.28–5.30 (d, J=11 Hz, 1 H), 5.29–5.33 (d, J=18 Hz, 1 H), 6.10–6.16 (dd, J=11, 18 Hz, 1 H); 13 C NMR (75 MHz, D₂O) δ 21.9, 34.5, 39.1, 63.3, 113.8, 139.6, 178.9; MS (FAB, 3-NOBA) 159 (100, MH+); HRMS (FAB, 3-NOBA) calcd for C₇H₁₆N₂O₂ (MH+) 159.1133, obsd 159.1128. Anal. Calcd for C₇H₁₄N₂O₂: C, 53.15; H, 8.92; N, 17.70. Found: C, 52.99; H, 8.74; N, 17.53.

α-Vinylvaline (5g). From 4g (2.30 g, 8.80 mmol) in 6 N HCl (70 mL), according to the procedure for 5a, was obtained 5g (1.08 g, 85%) as a white solid: mp >245 °C; ¹H NMR (500 MHz, D₂O) δ 0.99–1.01 (d, J=7 Hz, 3 H), 1.01–1.03 (d, J=7 Hz, 3 H), 2.33–2.42 (app heptet, J=7 Hz, 1 H), 5.27–5.31 (d, J=18 Hz, 1 H), 5.43–5.45 (d, J=11 Hz, 1 H), 6.10–6.16 (dd, J=11 Hz, 18 Hz, 1 H); ¹³C NMR (50 MHz, D₂O) δ 17.7, 18.6, 35.3, 72.0, 117.9, 137.9, 176.7; MS (EI) 143 (0.2, M+), 100 (100); HRMS (EI) calcd for C₇H₁₃NO₂ (M+) 143.0946, obsd 143.0942. Anal. Calcd for C₇H₁₃NO₂: C, 58.72; H, 9.15; N, 9.78. Found: C, 58.62; H, 9.06; N, 9.57.

 α -Vinylalanine (5h). From 4h¹¹ (500 mg, 2.14 mmol) in 6 N HCl (17 mL), according to the procedure for 5a, was obtained 5h^{7b} (219 mg, 89%) as a white solid.

α-Vinylhomoserine (5i). From 4i (500 mg, 0.997 mmol) in 6 N HCl (9 mL), according to the procedure for 5a, was obtained 5i (114 mg, 78%) as a white solid: mp 175 °C; ¹H NMR (500 MHz, D_2O) δ 2.16–2.25 (m, 2 H), 3.77–3.85 (m, 2 H), 5.38–5.41 (d, J=17 Hz, 1 H), 5.45–5.47 (d, J=11 Hz, 1 H), 6.08–6.14 (dd, J=11, 17 Hz, 1 H); ¹³C NMR (75 MHz, D_2O) δ 36.8, 58.6, 65.9, 117.5, 135.7, 174.6; MS (FAB, 3-NOBA) 146 (100, MH+), 120 (15), 100 (26); HRMS (FAB, 3-NOBA) calcd for $C_6H_{12}NO_3$ (MH+) 146.0817, obsd 146.0817. Anal. Calcd for $C_6H_{11}NO_3$: C, 49.65; H, 7.64; N, 9.64. Found: C, 49.53; H, 7.64; N, 9.60.

 α -Vinylhomoserine (5i). From 4j (500 mg, 1.57 mmol) in 6 N HCl (12 mL), according to the procedure for 5a, was obtained 5i (200 mg, 88%) identical in all aspects with the material obtained from 4i.

N-Benzoyl- α -vinylhomoserine Lactone (6). To a solution of 4j (1.38 g, 4.32 mmol) in CHCl₃ (28 mL) at 0 °C was added CF₃CO₂H (33.0 mL, 432 mmol) and stirring continued for 30 h at rt. After dilution with CH₂Cl₂ (200 mL), NaHCO₃ (aq, 200 mL) was added carefully. After further extraction with CH₂Cl₂ (100 mL), the combined organics were dried (MgSO₄), filtered, and evaporated to yield 6 (837 mg, 84%) as a white solid: mp 145-146 °C; ¹H NMR (500 MHz, CDCl₃) δ 2.82-2.88 (ddd, J = 9, 11, 13 Hz, 1 H), 2.93-2.97 (ddd, J = 1, 6, 13 Hz, 1 H), 4.27-4.32(ddd, J = 6, 9, 11 Hz, 1 H), 4.54-4.58 (app dt, J = 1, 9 Hz, 1 H),5.43-5.45 (d, J = 11 Hz, 1 H), 5.45-5.48 (d, J = 17 Hz, 1 H), 6.12-6.18 (dd, J = 11, 17 Hz, 1 H), 6.73-6.76 (m, 1 H), 7.44-7.47(m, 2 H), 7.52-7.56 (m, 1 H), 7.79-7.85 (m, 2 H); ¹⁸C NMR (75 MHz, CDCl₃) δ 34.3, 61.1, 65.9, 118.3, 127.0, 128.6, 132.1, 133.3 (2 C), 166.6, 175.2; IR (film) 3410-3220, 1770, 1655 cm⁻¹; MS (FAB, 3-NOBA) 232 (98, MH+), 105 (100); HRMS (FAB, 3-NOBA) calcd for C₁₃H₁₄NO₃ (MH⁺) 232.0974, obsd 232.0974. Anal. Calcd for C₁₃H₁₃NO₃: C, 67.52; H, 5.66; N, 6.06. Found: C, 67.34; H, 5.74; N, 5.91.

2-Benzamido-2-(hydroxyethyl)-N-methyl-3-butenamide (7). Lactone 6 (820 mg, 3.54 mmol) was taken up in ethanolic methylamine (4 ml, 8.0 M) and the reaction flask sealed. After 6 h of stirring at rt, the volatiles were removed in vacuo to provide 7 (897 mg, 96%) as a white solid: mp 140–141 °C; ${}^{1}H$ NMR (500 MHz, CDCl₃) δ 2.25–2.37 (m, 2 H), 2.84 (s, 3 H), 2.85 (s, 3 H), 3.86-3.94 (m, 2 H), 5.30-5.34 (d, J = 17 Hz, 1 H), 5.35-5.37 (d, J = 11 Hz, 1 H, 6.16-6.22 (dd, J = 11, 17 Hz, 1 H), 6.87-6.87(br s, 1 H), 7.41-7.44 (m, 2 H), 7.49-7.52 (m, 1 H), 7.83-7.85 (m, 2 H), 8.42 (s. 1 H); ¹³C NMR (75 MHz, CDCl₃) δ 26.7, 38.3, 59.4, 64.9, 116.1, 127.1, 128.6, 131.8, 133, 137.8, 166.8, 172.8; IR (film) 3320, 1775, 1650 cm⁻¹; MS (FAB, 3-NOBA): 263 (100, MH+), 204 (62); HRMS (FAB, 3-NOBA) calcd for $C_{14}H_{19}N_2O_3$ (MH⁺) 263.1396, obsd 263.1404. Anal. Calcd for C₁₄H₁₈N₂O₃: C, 64.11; H, 6.92; N, 10.67. Found: C, 64.30; H, 6.86; N, 10.45.

2-Benzamido-N-methyl-2-vinylsuccinimide (8). A solution of 7 (856 mg, 3.05 mmol) and pyridinium dichromate (PDC) (4.01 g, 10.7 mmol) in DMF (8 mL) was stirred for 4 h at rt. Workup consisted of filtration through Celite, dilution with Et₂O (50 mL). and extraction with H_2O (10 × 30 mL). Flash chromatography (50-75% EtOAc/hexane) afforded 8 (654 mg, 77%) as a white solid: mp 114-115 °C; ¹H NMR (500 MHz, CDCl₃) δ 3.05-3.09 (d, J = 18 Hz, 1 H), 3.09 (s, 3 H), 3.30-3.33 (d, J = 18 Hz, 1 H),5.47-5.49 (d, J = 11 Hz, 1 H), 5.52-5.56 (d, J = 17 Hz, 1 H), 6.02-6.08 (dd, J = 11, 17 Hz, 1 H), 6.54 (s, 1 H), 7.43-7.46 (m, 2 H), 7.52-7.55 (m, 1 H), 7.75-7.77 (m, 2 H); ¹³C NMR (75 MHz, CDCl₃) δ 25.2, 40.3, 61.0, 119.1, 127.1, 128.7, 132.4, 132.6, 134.7, 167.0, 173.9, 175.8; IR (film) 3340, 1780, 1705, 1660 cm⁻¹; MS (FAB, 3-NOBA) 259 (49, MH+), 105 (COPh, 100); HRMS (FAB, 3-NOBA) calcd for C₁₄H₁₅N₂O₃ (MH⁺) 259.1083, obsd 259.1083. Anal. Calcd for C₁₄H₁₄N₂O₃: C, 65.11; H, 5.46; N, 10.84. Found: C, 65.25; H, 5.60; N, 10.90.

 α -Vinylaspartic Acid (9). A suspension of 8 (45.0 mg, 0.174 mmol) in 10 N HCl (2.5 mL) was heated in a sealed glass vessel at 150 °C for 5 h. After extraction with CH₂Cl₂ (50 mL), the aqueous layer was evaporated in vacuo. Purification by chromatography on polyethyleneimine-(PEI)-cellulose, washing with H₂O and elution with BuOH/H₂O/AcOH (4:1:1), afforded 9 (18.8 mg, 68%) as a white solid: ${}^{1}H$ NMR (500 MHz, $D_{2}O$) δ 2.98-3.02 (d, J = 18 Hz, 1 H), 3.24-3.28 (d, J = 18 Hz, 1 H), 5.49-5.52 (d, J = 18 Hz, 1 Hz, 1 Hz), 5.49-5.52 (d, J = 18 Hz, 1 Hz), 5.49-5.52 (d

J = 18 Hz, 1 H), 5.52-5.55 (d, J = 11 Hz, 1 H), 6.04-6.09 (dd, J= 11, 18 Hz, 1 H); 13 C NMR (125 MHz, D_2 O) δ 39.7, 63.3, 119.5, 134.1. 173.5, 174.6; HRMS (FAB, glycerol) calcd for $C_6H_{10}NO_4$ (MH+) 160.0610, obsd 160.0604.

 α -Vinylarginine (10). To a solution of α -vinylornithine 5f (482 mg, 3.05 mmol) in 2 M NaOH (1.22 mL) was added S-ethylisothiouronium bromide (5.64 g, 30.5 mmol). The reaction mixture was maintained at ca. pH 10.5 by addition of 2 M NaOH and stirred at 45 °C for 36 h [TLC ($R_f = 0.13$; 20% NH₄OH/ EtOH)]. The reaction mixture was then brought to pH 8 with 1 N HCl and the volatiles were removed in vacuo. Silica gel chromatography (20% NH₄OH/EtOH) gave 10 (360 mg, 59%) as a white solid: mp 179-181 °C; ¹H NMR (500 MHz, D_2O) δ 1.52-1.58 (m, 1 H), 1.66-1.82 (m, 2 H), 1.95-2.01 (m, 1 H), 3.24-3.27 (m, 2 H), 5.18-5.22 (d, J = 11 Hz, 1 H), 5.20-5.26 (d, J = 12 Hz, 1 H) $17.6 \,\mathrm{Hz}, 1 \,\mathrm{H}), 6.01-6.11 \,(\mathrm{dd}, J = 11, 17.6 \,\mathrm{Hz}, 1 \,\mathrm{H}); {}^{19}\mathrm{C}\,\mathrm{NMR}\,(125)$ MHz, D₂O) δ 23.6, 33.6, 41.5, 65.8, 117.6, 136.4, 157.7, 175.2; MS (FAB, 3-NOBA): 201 (100, MH+); HRMS (FAB, 3-NOBA) calcd for C₈H₁₇N₄O₂ (MH⁺) 201.1351, obsd 201.1350.

Acknowledgment. Financial support from the donors of the Petroleum Research Fund, administered by the American Chemical Society, and the University of Nebraska Research Council is gratefully acknowledged. The authors wish to thank Craig L. Semerad (NSF REU Program) for assistance in synthesis of the protected α -amino acids. High resolution mass spectra were obtained from the Midwest Center for Mass Spectrometry, supported by NSF DIR9017262. M.L.P. is the recipient of GAANN (P200A100067) and Patricia Roberts Harris Fellowships (P094B20126) from the U.S. Department of Education.

Supplementary Material Available: ¹H NMR spectra for compounds 5a, 5b, 5d-i, 9 and 10 (10 pages). This material is contained in libraries on microfiche, immediately follows this article on the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.