Indirect determination of trace amounts of fluoride in natural waters by ion chromatography: a comparison of on-line post-column fluorimetry and ICP-MS detectors



María Montes Bayón, Ana Rodríguez Garcia,† J. Ignacio García Alonso* and Alfredo Sanz-Medel

Department of Physical and Analytical Chemistry, University of Oviedo, c/Julián Clavería 8, 33006 Oviedo, Spain

Received 10th September 1998, Accepted 20th November 1998

An alternative method for the determination of trace levels of fluoride in drinking and sea-water samples is presented. It is based on the formation of the aluminium monofluoride complex in excess of Al³+ and separation of the two species formed (AlF²+ and Al³+) in a small (5 cm long, CG²) ion exchange guard column. The final determination is accomplished by both ICP-MS specific detection and post column derivatisation with fluorimetric detection. Fundamental studies on the formation kinetics of the complex, ion chromatographic separation and optimum aluminium concentration were carried out using spectrofluorimetric detection by post-column reaction of the species with 8-hydroxyquinoline-5-sulfonic acid in a micellar medium of cetyltrimethylammonium bromide. Fluorimetric detection showed good detection limits, but interferences from cations such as Mg²+ and Zn²+ required the use of the longer CS² ion exchange column. Iron interfered in relatively large amounts but adding EDTA to the sample solution eliminated the interference. A similar separation methodology was applied using ICP-MS detection for the indirect determination of fluoride, by monitoring aluminium at mass 27. In this case, a detection limit of 0.1 ng ml⁻¹ was obtained using 0.45 m HNO₃ as eluent and no interference caused by high concentrations of iron was observed. The proposed method was applied to the determination of very low levels of fluoride in natural waters.

Introduction

During the last decade, the majority of fluoride determinations have been performed using techniques such as potentiometry with fluoride ion selective electrodes (ISE), 1,2 ion-exchange chromatography with conductivity detection,^{3–4} spectrophotometry⁵ and most recently capillary electrophoresis.^{6,7} The use of ISEs has been the preferred technique for this determination, but their sensitivity is insufficient to measure fluoride at ng ml⁻¹ levels. A spectrophotometric method using SPADNS has been applied to drinking waters.⁵ In the case of ion chromatography, the weak binding affinity of fluoride to the ion exchangers used to perform the separation process causes its early elution from the column, too close to the so-called injection peak, containing non-retained compounds and also the sample solvent. Most interferences in fluoride determination come from the presence of high levels of iron or aluminium in the sample. In these cases distillation of fluoride as HF can be performed.8

Atomic spectrometric techniques have not been used so far for direct fluoride determinations. The high excitation and ionisation potentials presented by this halogen resulted in poor sensitivity for atomic emission spectrometric (AES) detection even using powerful spectrochemical sources such as helium-based plasmas (*e.g.*, He MIP or He ICP). In this respect, good detection limits have been achieved for other halogens such as chloride or bromide, ^{9,10} but no results have been reported so far on fluoride determinations by AES. One interesting and recently developed alternative involves fluoride determination by electrospray mass spectrometry, with promising results. ¹¹

Several groups have investigated indirect fluoride determination. Marco *et al.*¹² determined fluoride by measuring the Previous work in our laboratory¹⁵ showed that aluminium could be better detected in the presence of cationic micelles of cetyltrimethylammonium bromide (CTAB) and it was also observed that the aluminium monofluoride complex could be detected by this fluorimetric reaction after ion chromatographic separation from Al^{3+,16,17} Here, the optimum conditions for the formation of the AlF²⁺ complex were studied using ion chromatography and post-column fluorimetric detection. Two types of elution conditions were evaluated, using K₂SO₄^{14,16} and HNO₃ as eluents. The analytical characteristics using fluorimetric detection were compared with ICP-MS as a specific detection method monitoring aluminium at *m/z* 27. Examples of the application of the proposed method are presented for the determination of low fluoride levels in drinking and sea-water samples.

Experimental

Instrumentation

The chromatographic system used consisted of a Pharmacia (Uppsala, Sweden) Model P-500 medium pressure pump and an Model 5M PA inert valve from Pharmacia fitted with a 100 μ l sample loop and a 5 cm long Dionex (Camberley, UK) Ion Pac

molecular absorption of the AlF²⁺ complex in a graphite tube using a Pt hollow cathode lamp. The chromatographic separation of Al–fluoride species was first described by Bertsch and Anderson, which is who determined the stability constants of the several possible AlF_x species. Later, Jones determined fluoride as AlF²⁺ after chromatographic separation from the excess of Al³⁺ using indirect fluorimetric detection with 8-hydroxyquinoline-5-sulfonic acid, obtaining detection limits in the low ppb range.

[†] Present address: Ingenieros Asesores SA, Polígono Silvota, Llanera, Asturias, Spain.

HPIC-CG2 ion exchange column. For the experiments using the K_2SO_4 as eluent, the column was immersed in a water-bath at 50 °C.

The spectrofluorimetric detector was a Shimadzu (Kyoto, Japan) R-F5000 equipped with a 12 μ l flow cell. The excitation and emission wavelengths were 390 and 500 nm, respectively, and the chromatograms obtained were recorded using a Shimadzu Chromatopac C-R3A integrator. The post-column reagent was pumped using a Scharlau (Barcelona, Spain) HP4 peristaltic pump.

Al-specific ICP-MS detection was carried out using an HP 4500 instrument (Hewlett-Packard, Yokogawa Analytical Systems, Tokyo, Japan) fitted with a concentric nebuliser and a Peltier cooled (2 °C) spray chamber. All chromatograms were obtained monitoring aluminium at *m/z* 27 using time resolved analysis.

Fig. 1 shows the instrumental set-up of the system using alternatively fluorimetric or ICP-MS detection. Neither detection system could be used on-line because of incompatibility of the eluents and post-column reaction. For fluorimetric detection, the eluent from the column was mixed with the post-column reagent using a T-piece and a 2 m \times 0.5 mm id PTFE reaction coil. For ICP-MS detection, the eluent leaving the column was connected directly to the nebuliser.

Reagents

All chemicals were of analytical-reagent grade unless stated otherwise and water obtained from a Milli-Q system (Millipore, Molsheim, France) was used to prepare stock standard solutions of all the reagents. Aluminium standard solution (1000 $\mu g \ ml^{-1})$ was obtained from Merck (Darmstadt, Germany). Stock standard solutions of F^- (1000 and 1 $\mu g \ ml^{-1})$ were prepared by dissolving solid NaF (Merck) in water.

For fluorimetric detection, K_2SO_4 (Merck) was used to prepare the mobile phase. The post-column reagents, 8-hydroxy-quinoline-5-sulfonic acid (HQS) and CTAB, were obtained from Sigma Aldrich, (St. Louis, MO, USA). The pH of the post-column reagent was adjusted using acetic acid–sodium acetate buffer (Merck). To prevent interferences from other metals present in the sample such as Fe, a standard solution of EDTA (Sigma Aldrich) was also used.

For ICP-MS determination, no post-column reagent was necessary and the mobile phase used was prepared from 65% HNO₃ (Suprapur, Merck) and diluted with Milli-Q water.

Preparation of AlF²⁺ complex

Samples and standard solutions were adjusted to pH 3 with nitric acid and spiked with Al³⁺, at least a five-fold mass excess of Al to fluoride being required to ensure that only AlF²⁺ was formed. The samples were diluted by volume (fluorimetric detection) or mass (ICP-MS detection), transferred into 10 ml

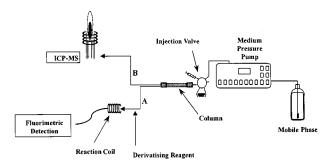


Fig. 1 Instrumental set-up of the system using (a) fluorimetric and (b) ICP-MS detection.

polypropylene test-tubes and immersed in a water-bath at 50 $^{\circ}$ C for 60 min. This ensured quantitative formation of the AIF²⁺ complex. For natural water samples and fluorimetric detection, EDTA was added at 1.6×10^{-5} M.

Chromatographic separation

For fluorimetric detection, the mobile phase was $0.1~{\rm M~K_2SO_4}$ in water adjusted to pH 3 with nitric acid. A flow rate of 1 ml min $^{-1}$ was used. The CG2 HPLC column was immersed in a water-bath at 50 °C following the recommendations of Jones $\it et~al.^{18}$ For ICP-MS detection, the mobile phase was 0.45 M HNO3 at a flow rate of 0.5 ml min $^{-1}$ and the column was kept at room temperature.

Spectrofluorimetric detection

The post-column reagent contained HQS and CTAB at optimum concentrations of 1×10^{-3} and 2×10^{-3} M, respectively and the optimum pH of 6 was adjusted with 0.25 M acetic acid–sodium acetate buffer. The optimum flow rate was 0.43 ml min $^{-1}$. The excitation and emission wavelengths selected were 390 and 500 nm respectively, providing the highest analytical signals for AlF²+ determination. The excitation and emission slits were both 5 nm.

ICP-MS detection

The ICP-MS operating conditions are summarised in Table 1. The output from the column was fed directly to the inlet of the concentric nebuliser and the *m/z* value monitored was 27 using the time resolved analysis mode, a 0.5 s integration time and 1 point per mass unit.

Results and discussion

Kinetic studies

The initial aim of this work was to obtain chromatographic conditions that allowed the separation and detection of the AlF²⁺ complex from the excess Al³⁺, to provide a sensitive method for indirect fluoride determination. First, some studies on the formation kinetics were performed and the determination

Table 1 Typical operating conditions

Instrument	HP 4500
Rf power	1300 W
Nebuliser	Meinhard
Spray chamber	Scott type, double pass, cooled at 2 °C
Sampling depth	5.7 mm
Gas flow rates—	
External	15 l min ⁻¹
Intermediate	1 l min ⁻¹
Carrier	1.17 l min ⁻¹
Ion lens settings—	
Extract 1	−221 V
Extract 2	-106 V
Einzel 1, 3	-144V
Einzel 2	39.3V
Omega bias	-48V
Omega (+)	6 V
Omega (–)	-7 V
QP focus	8 V
Ion deflector	39 V
Oxide level (CeO+/Ce+)	< 0.5%
Double charged level (Ce ²⁺ /Ce ⁺)	<1%

of the complex was carried out using spectrofluorimetric detection.

In aqueous acidic solution, aluminium ions are present as $[Al(H_2O)_6]^{3+}$, which can react with F^- to form the AlF^{2+} complex. It has been demonstrated that in a highly acidic medium, F^- reacts with H^+ to form HF, leading to a decrease in the rate of complexation of Al^{3+} with F^- . At pH>3.0, the hydrolysis reaction of Al^{3+} can take place with the formation of $Al(OH)_t^{(3-t)+}$, which reduces free aluminium and therefore the concentration of the complex. The optimum pH for the complex formation seems to be between 2 and 4, 1.14 and therefore in this work the pH selected was 2.6-3, where the complex AlF^{2+} proved to be stable.

Under these conditions, several parameters were evaluated in order to determine the formation kinetics of the complex, such as temperature and excess of aluminium necessary to obtain the quantitative formation of the AlF²⁺.

Fig. 2 shows the fluorimetric peak heights obtained for the AlF²⁺ complex as a function of solution temperature (15, 22 and 50 °C with solutions containing 100 ng ml⁻¹ fluoride and 1 µg ml⁻¹ aluminium) and complexation time. As can be appreciated, on heating the solution containing fluoride and aluminium at 50 °C, the formation of the complex can be considered quantitative after 50 min. At room temperature, more than 5 h are necessary to obtain stable signals for the complex. The slow reaction kinetics of the AlF²⁺ complex formed at room temperature could allow its separation from the excess of Al³⁺ without any decomposition or formation of alternative species during passage through the chromatographic column.

In order to optimise the aluminium concentration to be added for complete formation of the complex, solutions containing 200 ng ml $^{-1}$ fluoride were tested. Increasing amounts of aluminium were added to each sample and the solutions were heated at 50 °C for 1 h. The results obtained are shown in Fig. 3 and were measured fluorimetrically as peak height. As can be observed, a plateau was reached when using an aluminium concentration of 1 μg ml $^{-1}$ or higher. In order to increase the linear range as much as possible, a concentration of 10 μg g $^{-1}$ of aluminium was used in subsequent studies using fluorimetric detection.

Ion exchange separation: study of the mobile phase

Fig. 4 shows the typical chromatograms obtained for the separation of AlF²⁺ and Al³⁺ using 0.1 m $\rm K_2SO_4$ as eluent and increasing amounts of fluoride from 0 to 400 ng ml⁻¹. As can be observed, using 0.1 m $\rm K_2SO_4$ the AlF²⁺ peak increases with increase in fluoride concentration and the excess Al³⁺ elutes after 2.5 min. In order to optimise the concentration of $\rm K_2SO_4$ in

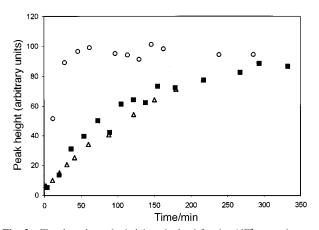


Fig. 2 Fluorimetric peaks heights obtained for the AlF²⁺ complex as a function of solution temperature (Δ , 15; \blacksquare , 22; and \bigcirc , 50 °C) containing 100 ng ml⁻¹ fluoride and 1 μ g ml⁻¹ aluminium.

the eluent, several concentrations were evaluated and Fig. 5 shows the standard representation of the logarithm of the capacity factor ($\log k'$) *versus* the negative logarithm of potassium concentration ($-\log [K^+]$).¹⁹ As can be observed, the slopes of the lines are 1.90 and 3.12 for AlF²⁺ and Al³⁺, respectively, and therefore it seems clear that the charge of the compounds is +2 and +3, respectively, and the structure of the complex is the one proposed.

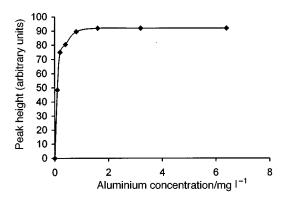


Fig. 3 Optimisation of the aluminium concentration required for complete formation of the AlF²⁺ complex in a solution containing 200 ng ml⁻¹ fluoride.

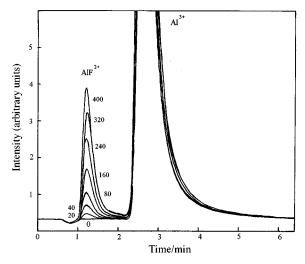


Fig. 4 Typical chromatograms obtained for the separation of AlF²⁺ and Al³⁺ using 0.1 M $\rm K_2SO_4$ as eluent and increasing amounts of fluoride from 0 to 400 ng ml⁻¹.

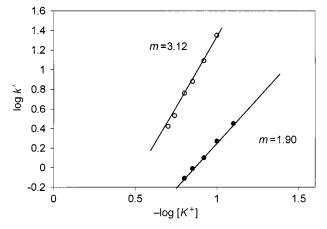


Fig. 5 Representation of the logarithm of the capacity factor ($\log k'$) obtained both for AlF²⁺ and Al³⁺ using different K₂SO₄ eluents. The slope of the log–log plot provides the charge of the species.¹⁹

Because saline solutions, such as K_2SO_4 , are not recommended in ICP-MS (possible clogging of the central channel of the torch and deposits on the cones), alternative mobile phases were tested. HNO₃ was found to be a good eluent for the separation of AlF²⁺ and Al³⁺ and detection by ICP-MS. Different concentrations of nitric acid, from 0.15 to 0.75 M, were tested for the above mentioned separation. The conditions chosen for subsequent studies were 0.45 M nitric acid at a flow rate of 0.5 ml min⁻¹ and detection at m/z = 27. The chromatogram obtained under these conditions for 20 ng g⁻¹ F⁻ in the presence of 100 ng g⁻¹ of Al is shown in Fig. 6. As can be observed, two aluminium containing peaks are detected. The first peak could be ascribed to the AlF²⁺ complex as its peak height/area was found to be proportional to the concentration of fluoride in the sample.

Analytical performance characteristics

Analytical performance characteristics for both detection modes are summarised in Table 2. The linear dynamic range for fluoride determination depends on the excess of aluminium added to the sample. It was observed with both detection modes that, for a given aluminium concentration, the upper linear limit for fluoride determinations was about one fifth of the total aluminium concentration. For ICP-MS, aluminium concentrations higher than 500 ng g⁻¹ were not tested. Using fluorimetric detection, linear upper limits up to 2000 ng ml F⁻ were obtained (using $10 \mu g ml^{-1}$ excess Al^{3+}).

The detection limits obtained were 0.6 ng ml⁻¹and 0.1 ng g⁻¹ for fluorimetry and ICP-MS, respectively, calculated as three times the standard deviation of the blank divided by the slope of the linear calibration graph between 0 and 5 ng g⁻¹. The detection limit using ICP-MS detection is one of the lowest ever reported for the determination of fluoride.

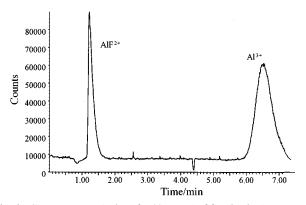


Fig. 6 Chromatogram obtained for 20 ng $\rm g^{-1}$ of fluoride in the presence of 100 ng $\rm g^{-1}$ of aluminium using ICP-MS detection. Eluent, 0.45 M nitric acid.

Table 2 Comparative analytical performance characteristics of spectrofluorimetric and ICP-MS detection

Analytical characteristics	Spectrofluorimetric detection	ICP-MS detection
Detection limit Precision Linear range Regression coefficient (r) (n = 7 points)	0.6 ng ml ⁻¹ 2.3% ^a Up to 2000 ^c ng ml ⁻¹ 0.9995	0.1 ng g^{-1} $4\%^b$ Up to 100^d ng g^{-1} 0.9993

 $[^]a$ For five injections of 20 ng ml $^{-1}$ fluoride. b For five injections of 20 ng g $^{-1}$ fluoride. c Using 10 µg g $^{-1}$ aluminium. d Using 500 ng g $^{-1}$ aluminium.

Interference studies

An exhaustive study of possible interferences from other anions and cations was carried out first using spectrofluorimetric detection. The results are summarised in Table 3. As can be observed, typical anions present in natural waters such as HCO_3^- and Cl^- do not interfere at the maximum concentration tested (200 μg ml⁻¹). Other anions such as $H_2PO_4^-$ and BO_3^{3-} can be present at 100 and 200 μg ml⁻¹, respectively, without causing interference.

The main interferences were observed from trace metals, which can be present in natural waters. Fe³⁺ at concentrations higher than $0.5 \,\mu g \, ml^{-1}$ decreased considerably the peak height from the AlF²⁺ peak owing to competition with Al³⁺ for fluoride or quenching of the fluorimetric reaction. However, in the presence of 1.6×10^{-5} M EDTA this interference was eliminated. Ca²⁺ and Sr²⁺ up to 100 and 50 μ g ml⁻¹, respectively, did not show any interference effect. Cu2+ and Pb²⁺ at concentrations higher than those in Table 3 produced a small decrease in the fluorimetric signal of AlF²⁺, which can be ascribed to co-elution with A1F2+ and the formation of competing chelates with the post-column reagent. On the other hand, Zn²⁺ and Mg²⁺ formed fluorescent chelates with the postcolumn reagent and co-eluted with AlF2+ using the 5 cm CG2 column. The use of a 25 cm CS2 column and a lower eluent concentration of 0.05 M K_2SO_4 resulted in the separation of the AlF2+ peak from Mg2+ and Zn2+. However, in this case the retention time for Al3+ increased to 30 min, as shown in Fig. 7 for a real water sample.

Using ICP-MS aluminium specific detection, no effect on detection from co-eluting divalent cations should be expected. The only possible interference could be with the formation and retention of AlF²⁺ complex. No effect of Fe³⁺ on the height or area of the AlF²⁺ peak was obtained for Fe³⁺ concentrations up to 100 ng g⁻¹ at the same aluminium concentration and 20 ng g⁻¹ of fluoride. Also, when monitoring at m/z 57,

Table 3 Effect of foreign ions on the determination of fluoride with spectrofluorimetric detection (200 ppb F^- , 10 ppm Al^{3+})

Interference	Maximum concentration allowed/ μ g ml ⁻¹ (recovery 100 ± 5%)
Fe ³⁺	0.5
	(10, in the presence of 1.6×10^{-5} M EDTA)
Zn^{2+}	Interferes
Mg ²⁺	10
Cu ²⁺	1.6
Ca ²⁺	100^{a}
Sr ²⁺	50^a
Pb ²⁺	2
HCO ⁻ (as NaHCO ₃)	200^{a}
Cl- (as KCl)	200^{a}
H_2PO_4 (as $NH_4H_2PO_4$)	100^{a}
BO ₃ ³⁻ (as Na ₃ BO ₃)	200^{a}
a Maximum concentration	tested.

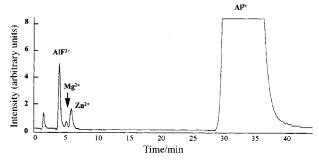


Fig. 7 Chromatogram obtained for a real water sample using the CS2 column (25 cm long) and 0.05 M $\rm K_2SO_4$ as eluent with fluorimetric detection.

representing a minor iron isotope, no elution of Fe from the column could be detected owing to insufficient sensitivity. Wang *et al.*¹ have reported that the formation of the Fe–fluoride complex is strongly pH dependent. The optimum pH for ironfluoride complex formation is 1.5 but at pH 2.5 complex formation decreased significantly.¹ Our results on pH effects agreed well with such observations.

Determination of fluoride in fresh and sea-water samples

Fluorimetric detection. Natural water samples may contain a large range of fluoride concentration from a few ng ml⁻¹ to several µg ml⁻¹. The concentrations of interfering compounds should be well below those indicated in Table 3 except, perhaps, for Fe, Mg and Zn in some samples. The chromatogram of a mineral water sample diluted 1 + 9 with Milli-Q water and containing 3.1 µg ml⁻¹ of fluoride determined by ISE is shown in Fig. 7; 0.05 M H₂SO₄ as eluent and the CS2 column were used. As can be observed, the AlF²⁺ peak is well separated from Mg²⁺ and Zn²⁺. The concentration found in this sample by reference to a calibration graph was $3.1 \pm 0.1 \,\mu g \, ml^{-1}$ (n = 5) and the recovery for spiked fluoride was 97%. Unfortunately, under these conditions the retention time for Al^{3+} was > 30 min, increasing dramatically the time required for each measurement as Al3+ must be eluted from the column before the next injection.

ICP-MS detection. Under the optimum separation conditions using HNO $_3$ for elution, the retention time for Al $^{3+}$ is < 8 min, allowing a sampling rate of 6–7 h $^{-1}$, and the detection is much more selective. Therefore, the proposed ICP-MS method was applied to the determination of fluoride in natural and drinking waters from a variety of sources and with different saline concentrations. As no stable aqueous fluoride reference material was available, it was decided to compare the proposed methodology with the fluoride ion selective electrode (FISE) and spiking the samples with fluoride to calculate the recoveries.

In order to minimise aluminium addition to the samples and contamination of the ICP-MS system, up to 200-fold dilution of some drinking and sea-water samples was necessary. The results obtained are summarised in Table 4. As some of the concentrations found were around 150 ng g $^{-1}$, and sometimes lower, the determination using FISE was adequate in only a few cases. As can be observed, the results obtained were in good agreement with the values found by FISE, when this determination was possible. In the other cases tested, recoveries of $100 \pm 10\%$ were obtained, showing the applicability of the proposed methodology to perform fluoride determinations at extremely low levels.

Table 4 Results obtained for the determination of fluoride in water samples using ICP-MS after dilution of the samples

Water sample (dilution factor)	Concentration found ($n = 3$) by ICP-MS/ ng g ⁻¹	Concentration found by FISE/ng g ⁻¹	Spiked amount/ ng g ⁻¹	Recovery (%)		
Fontecelta (200)	8050 ± 80	7700	4300	104		
Font-Vella (10)	182 ± 2	_	205	97.8		
Tap water (10)	161 ± 1	_	210	90		
Sea-water a (100)	1030 ± 60	1080	1080	97.5		
^a Collected at Gijon, Asturias, Spain.						

Conclusions

The formation of the AlF²⁺ complex in excess of Al³⁺ can be considered quantitative after thermal treatment of the sample for 1 h at 50 °C. Once formed, the complex is stable and can be separated from the excess of aluminium by cation exchange chromatography without decomposition even using highly acidic eluents (0.45 M HNO₃).

The detection of aluminium can be accomplished either by a post-column fluorimetric reaction, with interferences from other cations such as Fe^{3+} , Zn^{2+} and Mg^{2+} , or by ICP-MS. The latter detection method proved to be extremely sensitive, with a detection limit of 0.1 ng g^{-1} of F^- , and free from interferences from other cations and anions in natural water samples.

In comparison with the fluoride ISE, the proposed indirect ICP-MS method is at least two orders of magnitude more sensitive, it is not affected by interferences from aluminium and iron (in fact, the formation of the AIF²⁺ complex is the basis of the method) and it can be applied without modification to a large range of water samples of different salinity.

Acknowledgements

We thank Hewlett-Packard for the loan of the HP 4500 instrument and the DGCYT (Madrid) for financial support through Project DG-94-PB-1331.

References

- H. Wang, Z. Zhang, A. Sun, D. Liu and R. Liu, *Talanta*, 1996, 43, 2067.
- R. W. Kahama, J. J. M. Damen and J. M. ten Cate, *Analyst*, 1997, 122, 855.
- 3 T. A. Biemer, N. Asral and A. Sippy, J. Chromatogr. A, 1997, 771, 355
- 4 J. M. Talmage and T. A. Biemer, J. Chromatogr. A, 1987, 410, 494.
- 5 S. A. Sen, K. Kesava Rao, M. A. Frizzell and G. Rao, Field Anal. Chem. Technol., 1998, 2(1), 51.
- P. Wang, S. F. Y. Li and H. K. Lee, J. Chromatogr. A., 1997, 765, 353.
- 7 S. A. Shamsi and N. D. Danielson, Anal. Chem., 1995, 37, 1845.
- 8 Standard Methods for the Examination of Water and Waste Water, American Public Health Association, New York, 15th edn., 1980, p. 337
- F. Camuña, J. E. Sánchez-Uría and A. Sanz-Medel, Spectrochim. Acta, Part B, 1993, 48, 1115.
- A. H. Mohammed, J. T. Creed, T. M. Davidson and J. A. Caruso, *Appl. Spectrosc.*, 1989, 43, 1127.
- 11 D. A. Barnett and G. Horlick, J. Anal. At. Spectrom., 1997, 12, 497.
- 12 V. Marco, F. Carrillo, C. Pérez-Conde and C. Cámara, *Anal. Chim. Acta*, 1993, **283**, 489.
- 13 P. M. Bertsch and M. A. Anderson, Anal. Chem., 1989, 339, 535.
- 14 P. Jones, Anal. Chim. Acta., 1992, **258**, 123.
- J. I. Garcia Alonso, A. López García, E. Blanco González and A. Sanz-Medel, Anal. Chim. Acta, 1989, 225, 339.
- 16 J. I. Garcia Alonso, A. Rodriguez Garcia and A. Sanz-Medel, paper presented at Euroanalysis VII, Vienna, 1990.
- 17 A. Rodriguez Garcia, Master's Degree, Faculty of Chemistry, University of Oviedo (1991).
- 18 P. Jones, L. Ebdon and T. Williams, *Analyst*, 1988, **113**, 641.
- 19 R. Rosset, H. Conde and A. Jordy, Manuel Pratique de Chromatographie en Phase Liquide, Masson, Paris, 2nd edn., 1982.

Paper 8/07079B