

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/231438428>

# Modeling Copper–Dioxygen Reactivity in Proteins: Aliphatic C–H Bond Activation by a New Dicopper(II)–Peroxo Complex

ARTICLE *in* JOURNAL OF THE AMERICAN CHEMICAL SOCIETY · OCTOBER 1994

Impact Factor: 12.11 · DOI: 10.1021/ja00100a068

CITATIONS

99

READS

21

5 AUTHORS, INCLUDING:



**Samiran Mahapatra**

Unilever

42 PUBLICATIONS 2,218 CITATIONS

SEE PROFILE



**Jason Halfen**

University of Wisconsin - Eau Claire

73 PUBLICATIONS 3,480 CITATIONS

SEE PROFILE



**Lawrence Que**

University of Minnesota Twin Cities

451 PUBLICATIONS 25,695 CITATIONS

SEE PROFILE

# Modeling Copper–Dioxygen Reactivity in Proteins: Aliphatic C–H Bond Activation by a New Dicopper(II)–Peroxo Complex

Samiran Mahapatra, Jason A. Halfen,  
Elizabeth C. Wilkinson, Lawrence Que, Jr., and  
William B. Tolman\*

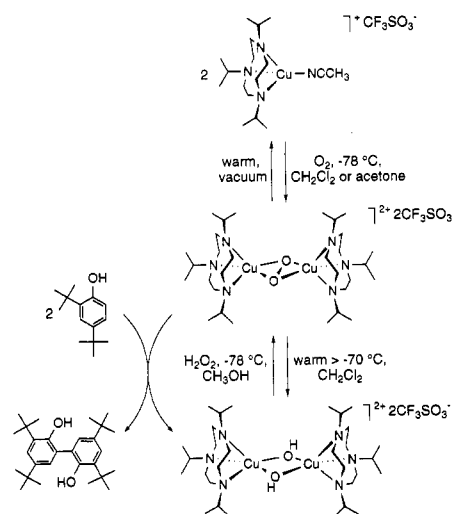
Department of Chemistry, University of Minnesota  
207 Pleasant Street SE, Minneapolis, Minnesota 55455

Received July 5, 1994

The central role of the binding and/or activation of dioxygen by copper(I) ions in a wide range of important processes in biology and catalysis has led to intense interest in the synthesis, characterization, and examination of the reactivity of  $\text{Cu}_2\text{O}_2$  complexes.<sup>1</sup> Relatively few such complexes have been definitively identified, however, and only two,  $[(\text{TPMPA})\text{Cu}]_2(\text{O}_2)(\text{PF}_6)_2$ <sup>2,3</sup> and  $[(\text{Tp}^{\text{Pr}}\text{Cu})_2(\text{O}_2)]$ ,<sup>2,4</sup> have been characterized structurally by X-ray crystallography. The former complex contains a *trans*- $\mu$ -1,2-peroxo ligand, while the latter has a planar dicopper(II)- $\mu$ - $\eta^2$ : $\eta^2$ -peroxo core that accurately models the dioxygen adduct of hemocyanin (Hc).<sup>5</sup> Despite extensive studies of the reaction pathways followed by these and other less well-defined  $\text{Cu}_2\text{O}_2$  adducts in synthetic and biological systems, our understanding of the relationship between their structural features and their reactivity with substrates remains rudimentary.<sup>1,6</sup> In particular, insight into the mechanistic details of aliphatic hydrocarbon oxidations that involve  $\text{Cu}_2\text{O}_2$  species in proteins, such as those catalyzed by dopamine  $\beta$ -monooxygenase (DBM),<sup>7</sup> peptidylglycine  $\alpha$ -amidating enzyme (PAM),<sup>8</sup> and particulate methane monooxygenase (pMMO),<sup>9</sup> has been limited by the relatively small number of synthetic copper–dioxygen complexes available for study and their general lack of reactivity with aliphatic C–H bonds. Here we report the preparation of a new dicopper(II)- $\mu$ - $\eta^2$ : $\eta^2$ -peroxo complex and demonstrate unequivocally the cleavage of an  $\text{sp}^3$  C–H bond by the  $\text{Cu}_2\text{O}_2$  unit.<sup>10</sup>

Treatment of solutions of  $[\text{LCu}(\text{CH}_3\text{CN})]\text{CF}_3\text{SO}_3$  ( $\text{L} = 1,4,7$ -triisopropyl-1,4,7-triazacyclononane)<sup>11,12</sup> in  $\text{CH}_2\text{Cl}_2$  or acetone with dry  $\text{O}_2$  at  $-78^\circ\text{C}$  resulted in the formation of red-brown  $[(\text{LCu})_2(\text{O}_2)](\text{CF}_3\text{SO}_3)_2$  (Scheme 1). The adduct was not perturbed by an  $\text{N}_2$  purge or application of vacuum at  $-78^\circ\text{C}$ , suggesting that  $\text{O}_2$  binding is essentially irreversible at this temperature. Assignment of a dicopper(II)- $\mu$ - $\eta^2$ : $\eta^2$ -peroxo structure to the product is based on (i) manometric data  $[\text{Cu}:\text{O}_2 = 2.2(2):1]$ ; (ii) its UV–vis spectrum, which contains  $\text{O}_2^{2-} \rightarrow \text{Cu}(\text{II})$  charge transfer bands [Figure 1;  $\lambda_{\text{max}} = 365$  ( $\epsilon \sim 11\,000\text{ M}^{-1}$

Scheme 1



$\text{cm}^{-1}$ , 510 (1000 nm)<sup>13</sup> analogous to those of  $[(\text{Tp}^{\text{Pr}}\text{Cu})_2(\text{O}_2)]$ <sup>14</sup> and oxyHc;<sup>15</sup> (iii) its EPR silence at 77 K, suggestive of antiferromagnetic coupling between  $\text{Cu}(\text{II})$  ions; and (iv) its resonance Raman spectrum, which contains  $\nu_{\text{O-O}}$  at  $722\text{ cm}^{-1}$  [ $\nu_{\text{O-O}}(^{18}\text{O}_2) = 680\text{ cm}^{-1}$ ;  $\Delta\nu(^{16}\text{O}_2-^{18}\text{O}_2) = 42\text{ cm}^{-1} = \Delta\nu_{\text{calc}}$ ]. This low  $\nu_{\text{O-O}}$  value is consistent with a weak O–O bond<sup>6,15</sup> and is similar to those of  $[(\text{Tp}^{\text{R}^2}\text{Cu})_2(\text{O}_2)]$  ( $\text{R} = \text{Ph}$ ,  $759\text{ cm}^{-1}$ ;  $\text{R} = \text{iPr}$ ,  $741\text{ cm}^{-1}$ ;  $\text{R} = \text{Me}$ ,  $731\text{ cm}^{-1}$ )<sup>4</sup> and oxyHc ( $750\text{ cm}^{-1}$ ).<sup>16</sup> Additional evidence in favor of the presence of a peroxo ligand in the  $\text{O}_2$  adduct includes (i) identification of excess  $\text{H}_2\text{O}_2$  (77% yield) by iodometric titration after addition of excess  $\text{HBF}_4\cdot\text{Et}_2\text{O}$  to the complex and (ii) generation of a UV–vis spectrum identical to that obtained upon addition of  $\text{O}_2$  to the  $\text{Cu}(\text{I})$  starting material

(1) Selected reviews: (a) Karlin, K. D.; Gulteh, Y. *Prog. Inorg. Chem.* **1987**, *35*, 219–328. (b) Sorrell, T. N. *Tetrahedron* **1989**, *40*, 3–68. (c) Karlin, K. D.; Tyeklár, Z.; Zuberbühler, A. D. In *Bioinorganic Catalysis*; Reedijk, J., Ed.; Marcel Dekker, Inc.: New York, 1993; pp 261–315. (d) Kitajima, N.; Moro-oka, Y. *Chem. Rev.* **1994**, *94*, 737–758.

(2) Abbreviations used: TmpA, tris(2-pyridylmethyl)amine;  $\text{Tp}^{\text{Pr}}$ , tris(3,5-diisopropylpyrazolyl)hydroborate.

(3) Tyeklár, Z.; Jacobson, R. R.; Wei, N. N.; Murthy, N. N.; Zubieta, J.; Karlin, K. D. *J. Am. Chem. Soc.* **1993**, *115*, 2677–2689.

(4) Kitajima, N.; Fujisawa, K.; Fujimoto, C.; Moro-oka, Y.; Hashimoto, S.; Kitagawa, T.; Toriumi, K.; Tatsumi, K.; Nakamura, A. *J. Am. Chem. Soc.* **1992**, *114*, 1277–1291.

(5) Magnus, K. A.; Ton-That, H.; Carpenter, J. E. *Chem. Rev.* **1994**, *94*, 727–735.

(6) Solomon, E. I.; Baldwin, M. J.; Lowery, M. D. *Chem. Rev.* **1992**, *92*, 521–542.

(7) (a) Stewart, L. C.; Klinman, J. P. *Annu. Rev. Biochem.* **1988**, *57*, 551–592. (b) Tian, G.; Berry, J. A.; Klinman, J. P. *Biochemistry* **1994**, *33*, 226–234 and references therein.

(8) (a) Merkle, D. J.; Kulathila, R.; Young, S. D.; Freeman, J.; Villafranca, J. J. In *Bioinorganic Chemistry of Copper*; Karlin, K. D., Tyeklár, Z., Eds.; Chapman & Hall: New York, 1993; pp 196–209. (b) Kulathila, R.; Consalvo, A. P.; Fitzpatrick, P. F.; Freeman, J. C.; Synder, L. M.; Villafranca, J. J.; Merkle, D. J. *Arch. Biochem. Biophys.* **1994**, *311*, 191–195 and references therein.

(9) (a) Nguyen, H.-H. T.; Shiemke, A. K.; Jacobs, S. J.; Hales, B. J.; Lindstrom, M. E.; Chan, S. I. *J. Biol. Chem.* **1994**, *269*, 14995–15005. (b) Chan, S. I.; Nguyen, H.-H. T.; Shiemke, A. K.; Lindstrom, M. E. In *Bioinorganic Chemistry of Copper*; Karlin, K. D., Tyeklár, Z., Eds.; Chapman & Hall: New York, 1993; pp 184–195.

(10) Examples of attack of  $\text{Cu}_2\text{O}_2$  species on arenes: (a) Nasir, M. S.; Cohen, B. I.; Karlin, K. D. *J. Am. Chem. Soc.* **1992**, *114*, 2482–2494. (b) Karlin, K. D.; Nasir, M. S.; Cohen, B. I.; Cruse, R. W.; Kaderli, S.; Zuberbühler, A. D. *J. Am. Chem. Soc.* **1994**, *116*, 1324–1336. (c) Sayre, L. M.; Nadkarni, D. V. *J. Am. Chem. Soc.* **1994**, *116*, 3157–3158 and references therein. (d) Casella, L.; Gullotti, M. In *Bioinorganic Chemistry of Copper*; Karlin, K. D., Tyeklár, Z., Eds.; Chapman & Hall: New York, 1993; pp 292–305. (e) Feringa, B. L. In *Bioinorganic Chemistry of Copper*; Karlin, K. D., Tyeklár, Z., Eds.; Chapman & Hall: New York, 1993; pp 306–324. (f) Martell, A. E.; Menif, R.; Ngwenya, P. M.; Rockcliffe, D. A. In *Bioinorganic Chemistry of Copper*; Karlin, K. D., Tyeklár, Z., Eds.; Chapman & Hall: New York, 1993; pp 325–337. (g) Réglie, M.; Amadé, E.; Alilou, E. H.; Eydoux, F.; Pierrot, M.; Waegell, B. In *Bioinorganic Chemistry of Copper*; Karlin, K. D., Tyeklár, Z., Eds.; Chapman & Hall: New York, 1993; pp 348–362.

(11) Haselhorst, G.; Stoetzel, S.; Strassburger, A.; Walz, W.; Wieghardt, K.; Nuber, B. *J. Chem. Soc., Dalton Trans.* **1993**, 83–90.

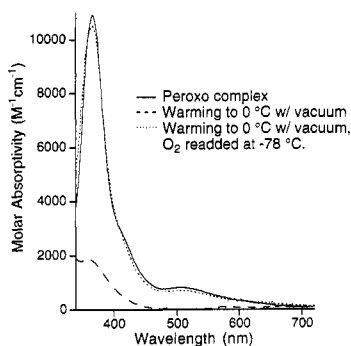
(12)  $[(\text{LCu}(\text{CH}_3\text{CN}))_2(\text{CF}_3\text{SO}_3)_2]$   $^1\text{H NMR}$  ( $\text{CD}_3\text{CN}$ , 300 MHz)  $\delta$  3.07 (septet,  $J = 6.6\text{ Hz}$ , 3H), 2.84–2.74 (m, 6H), 2.50–2.59 (m, 6H), 1.96 (s, 3H), 1.20 (d,  $J = 6.6\text{ Hz}$ , 18H). Anal. Calcd. for  $\text{C}_{118}\text{H}_{136}\text{Cu}_2\text{F}_6\text{N}_6\text{O}_3\text{S}_2$ : C, 42.47; H, 7.13; N, 11.00. Found: C, 42.86; H, 7.43; N, 10.18.  $[(\text{LCu})_2(\text{OH})_2](\text{CF}_3\text{SO}_3)_2$ : FTIR (KBr,  $\text{cm}^{-1}$ ) 3590 ( $\nu_{\text{OH}}$ ), 2653 ( $\nu_{\text{OD}}$ ), 3440 ( $\text{H}_2\text{O}$ ), 2977, 2879, 1626 ( $\text{H}_2\text{O}$ ), 1498, 1471, 1390, 1370, 1351, 1260, 1224 ( $\text{CF}_3\text{SO}_3^-$ ), 1149 ( $\text{CF}_3\text{SO}_3^-$ ), 1074, 1030 ( $\text{CF}_3\text{SO}_3^-$ ), 967, 765, 720, 637 ( $\text{CF}_3\text{SO}_3^-$ ), 572, 517; UV–vis ( $\text{CH}_2\text{Cl}_2$ ) [ $\lambda_{\text{max}}$ , nm ( $\epsilon$ ,  $\text{M}^{-1}\text{cm}^{-1}$ )] 360 (2540), 664 (200); EPR (9.1 GHz, 77 K) silent; electrospray MS ( $\text{CH}_2\text{Cl}_2$ )  $m/z$  (relative abundance) 821.2  $[(\text{LCu})_2(\text{OH})_2(\text{CF}_3\text{SO}_3)]^+$ , 100, 335.1  $[(\text{LCu}(\text{OH}))]^+$ , 40. Anal. Calcd. for  $\text{C}_{122}\text{H}_{168}\text{Cu}_2\text{F}_6\text{N}_6\text{O}_8\text{S}_2$ : C, 39.62; H, 7.07; N, 8.66. Found: C, 38.88; H, 7.01; N, 8.44. The complex as isolated is extremely hygroscopic; inclusion of one water molecule, the presence of which is supported by the IR spectrum, significantly improves the elemental analysis. Anal. Calcd. for  $\text{C}_{122}\text{H}_{170}\text{Cu}_2\text{F}_6\text{N}_6\text{O}_9\text{S}_2$ : C, 38.90; H, 7.14; N, 8.50.

(13) The low ratio of  $\epsilon$  values for the charge transfer bands ( $\sim 11:1$  vs  $\sim 20:1$  in oxyHc) has been observed for other proposed  $\text{Cu}_2\text{O}_2$  adducts. See: (a) Sorrell, T. N.; Garrity, M. L.; Richards, J. L.; White, P. S. *Inorg. Chim. Acta* **1994**, *218*, 103–108. (b) Nasir, M. S.; Karlin, K. D.; McGowty, D.; Zubieta, J. *J. Am. Chem. Soc.* **1991**, *113*, 698–700.

(14) Baldwin, M. J.; Root, D. E.; Pate, J. E.; Fujisawa, K.; Kitajima, N.; Solomon, E. I. *J. Am. Chem. Soc.* **1992**, *114*, 10421–10431.

(15) (a) Solomon, E. I.; Tuzek, F.; Root, D. E.; Brown, C. A. *Chem. Rev.* **1994**, *94*, 827–856. (b) Baldwin, M. J.; Root, D. E.; Pate, J. E.; Fujisawa, K.; Kitajima, N.; Solomon, E. I. *J. Am. Chem. Soc.* **1992**, *114*, 10421–10431.

(16) Ling, J.; Nestor, L. P.; Czernuszewicz, R. S.; Spiro, T. G.; Frackiewicz, R.; Sharma, K. D.; Loehr, T. M.; Sanders-Loehr, J. *J. Am. Chem. Soc.* **1994**, *116*, 7682–7691.

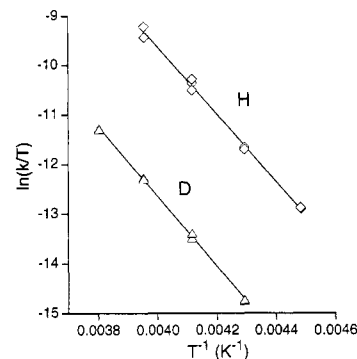


**Figure 1.** UV-vis spectrum of  $[(\text{LCu})_2(\text{O}_2)](\text{CF}_3\text{SO}_3)_2$  and spectroscopic demonstration of the reversible nature of  $\text{O}_2$  binding.

by adding aqueous  $\text{H}_2\text{O}_2$  to a solution of the dicopper(II)-bis-( $\mu$ -hydroxo) complex  $[(\text{LCu})_2(\text{OH})_2](\text{CF}_3\text{SO}_3)_2$ <sup>12</sup> in  $\text{CH}_3\text{OH}$  at  $-78^\circ\text{C}$ .

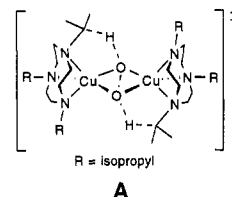
Although it appears on the basis of the data collected so far that  $[(\text{LCu})_2(\text{O}_2)](\text{CF}_3\text{SO}_3)_2$  and  $[(\text{Tp}^{\text{R}2}\text{Cu})_2(\text{O}_2)]$  contain structurally similar  $\mu$ - $\eta^2$ - $\eta^2$ -peroxo units, key aspects of their reactivity differ. For example, in contrast to the  $\text{Tp}^{\text{R}2}$  peroxo complexes,<sup>17</sup>  $\text{O}_2$  binding to  $[(\text{LCu})(\text{CH}_3\text{CN})](\text{CF}_3\text{SO}_3)$  is reversible upon warming under vacuum, as shown by the UV-vis cycling data shown in Figure 1. In addition, quantitative radical coupling of 2,4-di-*tert*-butylphenol was effected by the peroxo compound at  $-78^\circ\text{C}$  in the absence of uncoordinated  $\text{O}_2$  to cleanly afford  $[(\text{LCu})_2(\text{OH})_2](\text{CF}_3\text{SO}_3)_2$  (Scheme 1). This efficient radical generation and coupling differs from the reactivity observed for  $[(\text{Tp}^{\text{Me}2}\text{Cu})_2(\text{O}_2)]$ , which yields diphenoquinones under similar conditions,<sup>17</sup> but is analogous to that reported for a putative "bent"  $\mu$ - $\eta^2$ - $\eta^2$ -peroxo complex.<sup>18</sup>

Most intriguing were the results of experiments in which  $[(\text{LCu})_2(\text{O}_2)](\text{CF}_3\text{SO}_3)_2$  was warmed without application of vacuum in the absence of external reagents, conditions which led to the formation of the hydroxo-bridged compound  $[(\text{LCu})_2(\text{OH})_2](\text{CF}_3\text{SO}_3)_2$  as the major product (85% yield, Scheme 1).<sup>19</sup> Decomposition of the peroxo complex perdeuterated at the isopropyl substituents yielded OD-bridged product (FTIR;  $\nu_{\text{OD}} = 2653\text{ cm}^{-1}$ ,  $\nu_{\text{OH}}/\nu_{\text{OD}} = 1.35$ ; calcd = 1.37), demonstrating conclusively that the H(D) atoms of the bridges are derived from the isopropyl groups of the triazacyclononane ligand. Analysis of kinetic data obtained by monitoring the decrease of the 510 nm band of the dicopper(II)-peroxo complexes containing H- and D-substituted isopropyl groups as a function of time over the temperature range 223–263 K revealed that the reaction to form  $[(\text{LCu})_2(\text{OH})_2](\text{CF}_3\text{SO}_3)_2$  was first-order with respect to the peroxo complex and that it exhibited a large primary isotope effect [ $k_{\text{H}}/k_{\text{D}} = 18(1)$  at 298 K]<sup>20</sup> with a minor temperature dependence described by  $\Delta H^\ddagger_{\text{H}} = 13.5(5)\text{ kcal mol}^{-1}$ ,  $\Delta S^\ddagger_{\text{H}} = -12(1)\text{ eu}$ ,  $\Delta H^\ddagger_{\text{D}} = 14.0(5)\text{ kcal mol}^{-1}$ , and  $\Delta S^\ddagger_{\text{D}} = -16(1)\text{ eu}$  (Figure 2). These data conclusively demonstrate that the rate-determining step in the decomposition of the peroxo species involves cleavage of a C–H(D) bond of the isopropyl group(s), a transformation relevant to C–H activation processes mediated by  $\text{Cu}/\text{O}_2$  species in proteins (cf. D $\beta$ M, PAM, and pMMO) that, to our knowledge, has not been demonstrated previously for a synthetic copper–peroxo complex.<sup>21</sup> Reinaud and Theopold observed similar attack at the ligand isopropyl groups of a dicobalt–peroxo complex  $[(\text{Tp}^{\text{Pr}2}\text{Co})_2(\text{O}_2)]$ <sup>22</sup> but with significantly different kinetic parameters [ $\Delta H^\ddagger_{\text{H}} = 16.4(5)\text{ kcal mol}^{-1}$ ,  $\Delta S^\ddagger_{\text{H}} = -12(1)\text{ eu}$ ,  $\Delta H^\ddagger_{\text{D}} = 19.2(5)\text{ kcal mol}^{-1}$ , and  $\Delta S^\ddagger_{\text{D}} = -8(1)\text{ eu}$ ]. These kinetic data were interpreted to indicate a substantial tunneling contribution to the reaction rate that is not



**Figure 2.** Eyring plot for the decompositions of  $[(\text{LCu})_2(\text{O}_2)](\text{CF}_3\text{SO}_3)_2$  (H) and  $[(d_{21}\text{-LCu})_2(\text{O}_2)](\text{CF}_3\text{SO}_3)_2$  (D) in  $\text{CH}_2\text{Cl}_2$  (see text for derived activation parameters).

evident in our system.<sup>23</sup> Nonetheless, a rate-determining step involving peroxo O–O bond cleavage in concert with (not prior to)<sup>24</sup> two intramolecular H atom abstractions analogous to that proposed for the  $\text{Tp}^{\text{Pr}2}\text{Co}$  case (cf. transition state A) is consistent with the large  $k_{\text{H}}/k_{\text{D}}$  and negative  $\Delta S^\ddagger$  values we have measured.<sup>25</sup> Subsequent rapid trapping of the resulting tertiary alkyl radicals by solvent or  $\text{CH}_3\text{CN}$  hydrogen atoms would account for the final product isolated.



Important aspects of biological copper–dioxygen chemistry are modeled by the work described here. Both structural and functional mimicry of Hc are demonstrated by the reversible binding of  $\text{O}_2$  to yield a dicopper(II)- $\mu$ - $\eta^2$ - $\eta^2$ -peroxo species. Most significantly, the definitive characterization of an aliphatic C–H bond activation reaction by the  $\text{Cu}_2\text{O}_2$  unit suggests that such moieties (or related species) in biological systems may be capable of directly attacking nonaromatic hydrocarbons, a mechanistic possibility for copper protein-catalyzed oxidations that should be considered in future synthetic and biochemical research.

**Acknowledgment.** Funding in support of this research was provided by the Searle Scholars Program/Chicago Community Trust (W.B.T.), the National Science Foundation (National Young Investigator Award to W.B.T.), and the National Institutes of Health (GM33162 to L.Q.). We thank Professor Klaus Theopold (Delaware) for providing a preprint of ref 22 and the reviewers for helpful comments.

**Supplementary Material Available:** Representative first-order kinetic plot (233 K) for the peroxo complex decomposition reactions (1 page). This material is contained in many libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

(23) (a) Bell, R. P. *Chem. Soc. Rev.* **1974**, 3, 513–544. (b) Caldin, E. F. *Chem. Rev.* **1969**, 69, 135–156. (c) Kwart, H. *Acc. Chem. Res.* **1982**, 15, 401–408.

(24) In contrast, the pathway proposed for the decomposition of  $[(\text{Tp}^{\text{Me}2}\text{Cu})_2(\text{O}_2)]$  involves rate-determining peroxo cleavage to monomeric  $\text{Tp}^{\text{Me}2}\text{CuO}^\bullet$  species that are subsequently trapped (e.g., by a  $\text{Tp}^{\text{Me}2}\text{Cu}(\text{I})$  fragment to yield  $[(\text{Tp}^{\text{Me}2}\text{Cu})_2(\text{O})]$ ).<sup>17</sup>

(25) A reviewer has suggested that A "is extremely ordered and is in many ways similar to a ternary complex if one considers the two isopropyl hydrogens independently. Such a ternary complex is usually formed via two sequential binary events and the intermediate complex AB is stabilized pending its interaction with C. A possible pathway here might be such a sequential reaction where the peroxide undergoes a slow one-electron event followed by a fast radical abstraction at the second isopropyl group. The kinetics of such a process would be indistinguishable from the current proposal." We concur, as long as the "slow one-electron event" involves C–H(D) bond breaking (to explain the kinetic isotope effect).

(17) Kitajima, N.; Koda, T.; Iwata, Y.; Moro-oka, Y., *J. Am. Chem. Soc.* **1990**, 112, 8833–8839.

(18) Paul, P. P.; Tyeklár, Z.; Jacobson, R. R.; Karlin, K. D. *J. Am. Chem. Soc.* **1991**, 113, 5322–5332.

(19) Another minor (<15%) coproduct is formed that will be discussed in a future report.

(20) Calculated at this temperature from the activation parameters.

(21) Hydroxylation of one ethyl group of *N,N,N',N'*-tetraethylethylenediamine ligated to Cu(I) in the presence of excess  $\text{O}_2$  has been reported, but the direct involvement of a copper–peroxo complex as the oxidant in this reaction was not verified. See: Thompson, J. S. *J. Am. Chem. Soc.* **1984**, 106, 8308–8309 and references therein.

(22) Reinaud, O. M.; Theopold, K. H. *J. Am. Chem. Soc.* **1994**, 116, 6979–6980.