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Highly Effective PQQ Inhibition by Alkynyl and Aryl Monoand Diiodonium Salts[†]

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Abstract: PQQ (methoxatin), a bis(quinone) tricarboxylic acid, is an organic cofactor in a variety of biological redox processes. It is effectively inhibited on a micromolar scale by alkynyl and aryl monoiodonium salts, whereas bis-(iodonium)triflates 5 and 8 are PQQ inhibitors at nanomolar levels.

PQQ (methoxatin, 1), a bis(quinone) tricarboxylic acid, is an organic cofactor in an increasing number of biological redox processes. As a consequence, there is considerable current interest and enhanced research activity in the biochemical roles of PQQmediated redox cycling in biological processes.1 Recent evidence suggests that PQQ is an essential nutrient for mouse pups.2 Furthermore, PQQ, given to animals in pharmacological amounts, affords protection against (1) hepatotoxicity caused by liver poisons in rats,3a (2) acetaldehyde accumulation following ethanol loading in rats,3b (3) oxidative-stress-induced cataract formation in hydrocortisone-treated chick embryos,3c (4) inflammation induced by carrageenin in rat paws,3d and (5) neuroexcitatory agents like N-methyl-D-aspartic acid that target the glutamate receptor redox site in neurons.3e Recent evidence indicates that PQQ, first found as a bacterial cofactor for alcohol dehydrogenases, 3f is widely distributed in animal cells, tissues, and fluids and also functions as a redox cofactor in mitochondrial complex I.3g,h PQQ also diminishes brain necrosis in a rat model of a

A common way of investigating the role of PQQ in biological processes is to inhibit these processes with agents that effectively sequester PQQ. PQQ is a trianionic quinoid compound with three carboxyl groups that are ionized at physiological pH. PQQ chelates metal ions and forms charge-transfer complexes with aromatic amino acids.^{3j} We have found that organic cations like

† Dedicated to Prof. E. J. Corey on the occasion of his 65th birthday.

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N-methylphenazonium (phenazine methosulfate), berberine, and MPP+ (N-methylphenylpyridinium) sequester PQQ and inhibit its ability to catalyze glycinate-fueled redox cycling. PQQ is also sensitive to metal ions, especially indium, manganese, lead, vanadyl and trialkyltin. An especially useful organic cation with profound physiological actions is the diphenyleneiodonium cation, DPI, 2, a member of the family of polycoordinated iodine species. We found that DPI sequesters PQQ and expect that this accounts for the physiological actions of DPI that include (1) its induction of hypoglycemia and lactic acidosis caused by blockage of gluconeogenesis following mitochondrial toxicity, b (2) the inhibition of respiratory burst in stimulated neutrophils, c (3) the inhibition of nitric oxide synthase in endothelial cells, and (4) the induction of myopathy in rats, chronically treated with sublethal amounts of DPI. Se.f

Likewise, there is a renaissance in polycoordinated iodine chemistry⁶ and, in particular, in new iodonium species.^{7,8} With the known⁶ wide-ranging biological activity of iodonium

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Scheme I

Scheme II

salts and the specific inhibition of PQQ by DPI in mind, we decided to examine the inhibition of PQQ by new types of iodonium salts. Two major classes of new iodonium salts were investigated monoiodonium salts, as exemplified by 3 and 4, and bis(iodonium) species 5-8. The preparation of 3,9c 4,10 6,11a and 711b has been previously described. Bis(iodonium) salts 5 were prepared12 as described in Scheme I.

Commercial diiodobenzene 9 was oxidized to 10 by CF₃CO₃H in 85% yield. Reaction of 10 with 2 equiv of the appropriate RSiMe₃ 11 and Me₃SiOSO₂CF₃ (12), respectively, in CH₂Cl₂ gave 5a-d in 66-97% isolated yields. 13 Likewise, 5h was prepared as outlined in Scheme I using 2,5-bis(trimethylsilyl)thiophene instead of 11. Compounds 8 were prepared by the iodoniumtransfer reaction⁹ of 2,5-bis(tributyltin)thiophene (13) with arylcyanoiodonium triflates 14.96

All new compounds had both HRMS and spectral properties in accord with the proposed structures and expectations^{6,7} for iodonium salts as detailed in the experimental procedure.

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(13) See the Experimental Section for individual compounds and yields. The isolated yield of the deactivated p-FC₆H₄ 5a was only 6%.

Table I. Inhibition of PQQ-Catalyzed Redox Cycling by Iodonium

entry	compound	IC ₅₀ a
1	BPI, Ph ₂ I+	10.0 μM
2	DPI, 2	1.5 µM
3	3a	6.0 µM
4	3b	3.0 µM
5	3c	19 μ M
6	. 3d	437 nM
7	4	667 nM
8	5a	7 nM
9	5b	13 nM
10	5c	33 nM
11	5d	77 nM
12	5e	333 nM
13	5f	667 nM
14	5g	63 nM
15	5h	36 nM
16	6a	1.3 μM
17	6b	667 μM
18	7	2.0 μM
19	8a	29 nM
20	8b	13 nM

^a Using 13.3 nM PQQ.

PQQ inhibition studies were carried out via the PQQ-catalyzed redox-cycling assay4a,b using 13.3 nM PQQ and adding various concentrations of the inhibitor to determine the IC50. A stock solution of the inhibitor was prepared in DMSO and then further diluted in water to the desired concentration. PQQ without inhibitor and the various concentrations of the inhibitors were run in parallel, the latter to control for any direct reduction of NBT.

In this assay, glycine present in large excess is oxidized at pH 10 in air in a reaction catalyzed by small amounts of PQQ. As reduced PQQ is generated, it is oxidized by dioxygen back to PQQ and superoxide is generated. Superoxide then reduces nitroblue tetrazolium (T+) to formazan (TH). In 20 min at 37 °C, about 2000 redox cycles occur such that nanomolar concentrations of PQQ generate micromolar amounts of formazan dve. Agents that sequester PQQ like the iodonium compounds inhibit this redox-cycling assay.

$$\begin{array}{c} {\rm NH_2\text{-}CH_2\text{-}COO^-} \\ {\rm NH=CH\text{-}COO^-} \end{array} \\ \begin{array}{c} {\rm PQQH_2} \\ {\rm PQQ} \end{array} \\ \end{array} \\ \begin{array}{c} {\rm PQQ} \\ {\rm 2\ O_2} \end{array} \\ \end{array} \\ \begin{array}{c} {\rm TH} \\ {\rm Th} \\ \end{array}$$

The results are summarized in Table I. Perusal of the data in Table I reveals that as a group the bis(iodonium) compounds are clearly better inhibitors of PQQ than the monoiodonium compounds, in the redox-cycling assay for PQQ. Two of the bis(iodonium) compounds (5a and 5b) have been tested in mitochondria, and both compounds strongly inhibit NADH-fueled mitochondrial electron transport.3h The inhibition is reversed by the addition of PQQ. Physiologically, the new iodonium compounds may also show selective permeability for certain cells. For instance, while DPI is especially useful for preparing animal models of myopathies,5e bis(iodonium) compounds may show a selective toxicity for other cells and tissues. Inhibition by the new type of alkynyl monocations 3a-c (entries 3-5) is comparable to inhibition of PQQ by diphenyliodonium (entry 1) and DPI (entry 2), whereas inhibition of PQQ by the most active bis-(iodonium) salts 5a-c (entries 8-10) is 102-103 better than that of BPI and DPI. Likewise, bis(iodonium) salts 8 are nearly 1000 times better inhibitors of PQQ than DPI. Hence, we expect widespread use of these bis(iodonium) species as highly effective inhibitors of biological processes involving PQQ.

Experimental Section

General Methods. Melting points (uncorrected) were obtained with a Mel-Temp capillary melting-point apparatus. Infrared spectra were recorded on a Mattson FT-IR spectrophotometer. NMR spectra were

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⁽¹²⁾ Select members of 5 as a tosylate, p-CH₃C₆H₄SO₃-, have previously been prepared by a different procedure in generally low yields; see: Koser, G. F.; Carman, C. S. U. S. Patent 4,513,137, 1985. Likewise, for 8a as a tosylate see: Jezic, Z. U.S. Patent 3,712,920, 1973.

recorded on a Varian XL 300 spectrometer at 300 MHz (¹H NMR), 75 MHz (¹³C NMR), and 282 MHz (¹⁹F NMR). Chemical shifts for ¹H and ¹³C NMR are reported in parts per million (ppm) relative to internal tetramethylsilane or the proton resonance due to the residual protons in the deuterated NMR solvent; the chemical shifts for ¹⁹F NMR are relative to external CFCl₃. Mass spectra were obtained with a VG Micromass 7050E double-focusing high-resolution mass spectrometer with the VG data system 2000 under positive ion fast atom bombardment (FAB) conditions at 8 keV. 3-Nitrobenzyl alcohol was used as a matrix in CH₂-Cl₂ or CHCl₃ as the solvent, and polypropyleneglycol was used as a reference for peak matching. Microanalyses were performed by Atlantic Microlab Inc., Norcross, GA.

Materials. All commercial reagents were ACS reagent grade and used without further purification. Arylcyanoiodonium triflates 14a,b were prepared from (bis(trifluoroacetoxy)iodo)arenes, trimethylsilyl triflate, and cyanotrimethylsilane. ^{9b} Iodonium salts 3, ^{9c} 4, ¹⁰ 6, ^{11a} and 7^{11b} were prepared by known methods. All solvents used were dried by distillation over CaH₂. The reaction flasks were flame-dried and flushed with nitrogen.

p-Bis[bis(trifluoroacetoxy)iodo]benzene (10). p-Diiodobenzene (9) (3.3 g, 10 mmol) was added by small portions during 30 min to a stirred mixture of CF₃CO₃H (prepared from trifluoroacetic anhydride (10 mL, 71 mmol) and 80% hydrogen peroxide (2 mL, 47 mmol) by a known procedure¹⁴). The reaction mixture was stirred for 0.5 h at -78 °C and then for 2 h at -20 °C and left overnight at room temperature. Concentration of the resulting clear solution and crystallization of the product by addition of ether afforded analytically pure 10 as a white microcrystalline solid, yield 6.64 g (85%); mp 195-197 °C dec. IR (CCl₄, cm⁻¹): 3084, 3061 (C₆H₄), 1664, 1146, 982 (all CF₃CO₂). ¹H NMR (CF₃CO₂H/CDCl₃ 1/10): δ -78.9 (s, CF₃CO₂). ¹³C [¹H] NMR (CF₃CO₂H/CDCl₃ 1/10): δ -78.9 (s, CF₃CO₂). ¹³C [¹H] NMR (CF₃CO₂H/CDCl₃ 1/10): δ 118.7 (q, CF₃), 126.0 (s, C_{ipsoAr}), 137.9 (s, CH_{Ar}), 162.4 (q, C=O). Anal. Calcd for C₁₄H₄I₂O₈F₁₂: C, 21.50; H, 0.52. Found: C, 21.16; H, 0.78.

General Procedure for the Preparation of (p-Phenylene)bis(iodonium) Salts 5. To a stirred solution of 10 (0.78 g, 1 mmol) in CH_2Cl_2 (20 mL) were added the corresponding silylated arene 11 (2.5–3 mmol) and Me₃-SiOTf (12) (0.5 mL, 2.5 mmol) at -78 °C under N_2 . The reaction mixture was allowed to warm to room temperature and additionally stirred for 3–5 h. Colorless microcrystalline products 5a–f precipitated in analytically pure form.

5a: yield 0.05 g (6%); mp 252-260 °C dec. IR (CCl₄, cm⁻¹): 3086 (Ar), 1245, 1167, 1028 (all CF₃SO₃). ¹H NMR (DMSO- d_6): δ 7.25 (dd, 4H, J = 7.0 Hz), 8.12 (m, 8H). ¹⁹F NMR (DMSO- d_6): δ -78.5 (s, CF₃SO₃), -106.1 (s, ArF). ¹³C [¹H] NMR (DMSO- d_6): δ 121.3 (q, J = 318 Hz, CF₃SO₃-), 120.1, 120.4, 137.3, 138.4, 138.7, 139.2 (all Ar). 5b: yield 0.7 g (90%); mp 280-290 °C dec. ¹⁵

5e: yield 0.95 g (92%); mp 270–275 °C dec. IR (CCl₄, cm⁻¹): 3082 (Ar), 1265, 1170, 1024 (all CF₃SO₃). ¹H NMR (DMSO- d_6): δ 7.82 (d, 4H, J = 7.3 Hz), 7.87 (d, 4H, J = 7.4 Hz), 8.12 (s, 4H). ¹⁹F NMR (DMSO- d_6): δ -77.8 (s, CF₃SO₃). ¹³C [¹H] NMR (DMSO- d_6): δ 121.3 (q, J = 318 Hz, CF₃SO₃⁻), 96.5, 120.1, 120.5, 138.4, 138.9, 141.2 (all s, Ar). HRMS (FAB) for C₁₉H₁₂Sl₄O₃F₃ (M - TfO⁻)+ calcd 884.663334, found 884.661143. Anal. Calcd for C₂₀H₁₂I₄O₆F₆S₂: C, 23.23; H, 1.17; I, 49.09. Found: C, 23.40; H, 1.20; I, 49.22.

5d: yield 0.79 g (66%); mp 275–278 °C dec. IR (CCl₄, cm⁻¹): 3082 (Ar), 1247, 1168, 1026 (all CF₃SO₃). ¹H NMR (DMSO- d_6): δ 7.36 (d, 4H, J = 7.5 Hz), 7.70 (d, 4H, J = 7.6 Hz), 7.80 (d, 4H, J = 7.5 Hz), 8.12 (s, 4H), 8.14 (d, 4H, J = 7.5 Hz). ¹⁹F NMR (DMSO- d_6): δ -78.4 (s, CF₃SO₃). ¹³C [¹H] NMR (DMSO- d_6): δ 121.0 (q, J = 318 Hz, CF₃SO₃-), 95.2, 120.3, 120.6, 131.4, 136.5, 136.8, 136.9, 137.5, 138.4, 138.6 (all Ar). HRMS (FAB) for C₃₁H₂₀SL₄O₃F₃ (M - TfO⁻)+ calcd 1036.725934, found 1036.723128. Anal. Calcd for C₃₂H₂₀L₄O₅F₆S₂: C, 32.40; H, 1.70; I, 42.79. Found: C, 32.24; H, 1.69; I, 42.61.

5e: yield 0.9 g (97%); mp 255-257 °C dec.15

5f: yield 0.7 g (85%); mp 183-185 °C dec. 16

5g: yield 0.93 g (86%); mp 256–258 °C dec. IR (CCl₄, cm⁻¹): 3082 (Ar), 2957 (Me₃Si), 1245, 1167, 1027 (all CF₃SO₃). ¹H NMR (DMSO- d_6): δ 0.21 (s, 18H), 7.62 (m, 8H), 7.74 (d, 4H, J = 7.6 Hz), 8.12 (s, 4H), 8.15 (d, 4H, J = 7.5 Hz). ¹⁹F NMR (DMSO- d_6): δ -78.2 (s, CF₃SO₃). ¹³C [¹H] NMR (DMSO- d_6): δ -1.5 (s), 121.0 (q, J = 318 Hz, CF₃SO₃-), 120.1, 120.7, 131.4, 136.5, 136.8, 136.9, 137.5, 138.4, 138.6, 141.5 (all Ar). HRMS (FAB) for C₃₇H₃₈SSi₂I₂O₃F₃ (M - TfO⁻)+ calcd 929.011939, found 929.012254.

5h: yield 0.8 g (85%); mp 225–230 °C dec. IR (CCl₄, cm⁻¹): 3075, 2965, 1474, 1384, 1282, 1235, 1165, 1026, 977. ¹H MMR (CD₃CN): δ 8.03 (s, 4H), 7.90 (d, 2H, J = 8.0 Hz), 7.24 (d, 4H, J = 8.0 Hz), 0.21 (s, 18H, 2Me₃Si). ¹9F NMR (CD₃CN): δ -78.3 (s, CF₃SO₃). HRMS (FAB) for C₂₁H₂₆S₃Si₂I₂O₃F₃ (M - TfO⁻)⁺ calcd 788.862184, found 788.861207.

General Procedure for the Reaction of 2,5-Bis(tributyltin)thiophene (13) with Aryleyanolodonium Triflates 14. To a stirred solution of reagent 14 (1 mmol) was added a solution of the appropriate tributyltin derivative 13 (1-1.5 equiv) in CH_2Cl_2 (15 mL) at -40 °C. The mixture was warmed to room temperature and stirred until the formation of a clear solution. The product was precipitated from the reaction mixture by the addition of dry hexane (20-30 mL). The microcrystalline iodonium triflate salt was filtered under nitrogen, washed with dry hexane (30 mL) and dried in vacuo. Analytically pure materials were obtained by recrystallization from a concentrated solution of the iodonium salt in CH_2Cl_2 by the addition of hexane and ether.

8a: yield 0.39 g (49%); mp 193–194 °C dec. IR (CCl₄, cm⁻¹): 3097, 3066, 1241, 1159, 1023. ¹H NMR (DMSO- d_6 /CD₃CN): δ 7.55 (t, 4H, J = 8.0 Hz), 7.71 (t, 2H, J = 8.0 Hz), 7.83 (s, 2H), 8.19 (d, 4H, J = 8.0 Hz). ¹⁹F NMR (DMSO- d_6 /CD₃CN): δ –78.74 (s, CF₃SO₃⁻¹). ¹³C [¹H] NMR (DMSO- d_6 /CD₃CN): δ 111.0, 119.4, 121.2 (q, J = 320.7 Hz, CF₃SO₃⁻¹), 132.3, 132.9, 135.1, 140.9. Anal. Calcd for C₁₈H₁₂I₂O₆F₆S₃: C, 27.43; H, 1.53; S, 12.20. Found: C, 27.35; H, 1.56; S, 12.12.

8b: yield 0.21 g (25%); mp 170 °C dec. IR (CCl₄, cm⁻¹): 3098, 1575, 1246, 1170, 1027. ¹H NMR (CD₃CN): δ 8.15 (dd, 4H), 7.85 (s, 2H), 7.3 (dd, 4H). ¹⁹F NMR (CD₃CN): δ -78.1 (CF₃SO₃⁻), -105.0 (FC₆H₄). ¹³C MMR (CD₃CN): δ 162.5 (d), 138.7, 136.3, 136.2, 121.0 (q, CF₃SO₃), 117.7, 110.5. HRMS (FAB) m/z 674.804664 (M – CF₃SO₃⁻)⁺, calcd for C₁₇H₁₀I₂S₂F₅O₃ 675.807858.

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