Combinatorial Synthesis of Natural Product-like Molecules Using a First-Generation Spiroketal Scaffold

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Recently, significant attention has been focused on the synthesis small-molecule libraries based on natural product or natural product-like structures. In this paper, we report our initial studies on the use of the 1,7-dioxaspiro[5,5]undecane (spiroketal) moiety as a rigid-core template for elaboration using parallel synthesis techniques. The synthesis of a spiroketal scaffold that is reminiscent of the spiroketal subunits found in the spiroketal macrolide antibiotics will be described. Elaboration of three independently addressable functional groups on the scaffold using solution-phase parallel synthesis techniques led to the preparation of a small library of natural product-like compounds. These studies pave the way for evaluation of highly functionalized spiroketals in phenotypic assays and as prospective antagonists of protein—protein interactions.

Introduction

The synthesis of novel and rigid scaffolds for diversification using combinatorial chemistry techniques has been an important goal in the discovery of new biologically active molecules.1 A number of literature examples have been reported wherein rigid architectures have been employed, including substituted glycouril compounds,2 calix[4]arene derivatives,3 spirodiamino acid scaffolds,4 tri- and tetracarboxylate cores,⁵ cyclopeptides,⁶ carbohydrate scaffolds,⁷ and steroid nuclei.8 Recently, significant attention has also been focused on the synthesis of small-molecule libraries based on natural product or natural product-like structures.⁹ We have initiated studies in this area, employing the 1,7dioxaspiro[5,5]undecane moiety as a rigid-core template for elaboration using parallel synthesis techniques. The spiroketal moiety is a prevalent structure in a wide number of biologically important natural products¹⁰ and has also been employed as a scaffold in the synthesis of conformationally restrained glycomimetics¹¹ and gyrase inhibitors¹² (parts a and b of Figure 1, respectively). The use of spiroketals as lead structures is a potentially promising area for investigation because recent literature reports have shown that simplified compounds exhibit interesting pharmacological effects, such as tubulin modulation and cytotoxicity against tumor cell lines (cf. parts c and d of Figure 1).¹³ Herein, we report our studies on the synthesis of a spiroketal scaffold containing three independently addressable functional groups and the use of parallel solution-phase synthesis¹⁴ methods (extractive workups, polymer reagent, scavenger techniques,

and column-based extractions) for elaboration of the scaffold. We have focused our initial efforts on the preparation of highly functionalized spiroketals with general structure 1 (Figure 2), using a scaffold that is reminiscent of spiroketal subunits found in the spiroketal macrolide antibiotics cytovaricin, 15 spongistatins (altohyrtins), 16 and the rutamycins/ oligomycins.¹⁷ In addition to further development of methodologies for streamlined synthesis (a descriptor for methodology that lends itself to efficient application to parallel organic synthesis), our motivation to prepare libraries of spiroketals is also based on planned evaluation of these molecules in phenotypic assays18 and as antagonists of protein—protein interactions. 19 Functionalized spiroketals 1 are ultimately derived from spiroketal ketone 2, which may be prepared from condensation of chiral ketone 3 and aldehyde 4 using established reaction sequences.²⁰

Results and Discussion

The synthesis of protected hydroxy ketone fragment **3** (Scheme 1) was initiated by hydrolytic kinetic resolution $(HKR)^{21}$ of (\pm) -1,2-epoxy-5-hexene to afford *S*-epoxide **5** (85%). Epoxide ring opening using 2-methyl-1,3-dithiane²² provided hydroxydithiane **6**, which was silylated to afford **7**. Although treatment of **7** with mercury(II) perchlorate afforded low yields of **3**, presumably due to competitive oxymercuration of the terminal olefin, dithiane cleavage using 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ)²³ afforded methyl ketone **3** in high yield.²⁴ Preparation of the requisite aldehyde fragment **4** was initiated by protection of commercially available 5-hexen-1-ol as a *p*-methoxybenzyl ether to afford **8** (Scheme 2). Sharpless asymmetric dihydroxylation²⁵ of **8** using AD-mix- β provided diol **9** (80% ee), which was converted to acetonide **10**. PMB deprotection

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(a) Sialyl Lewis X Mimic Reference 11

(b) Novobiocin (Gyrase Inhibitor) Mimic Reference 12

(c) Causes tubulin depolymerization and exhibits potent cytotoxicity against taxol-and vincristine-resistant human cancer cell lines Reference 13b-e

OBn 3

(d) Causes cell death of Jurkat cells by apoptosis Reference 13a

Figure 1. Non-natural spiroketal derivatives.

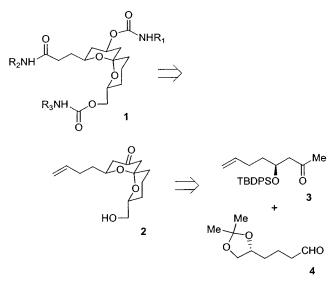


Figure 2. General structure and retrosynthesis of spiroketals derived from elaboration of a three-vector scaffold.

(DDQ)²⁶ of 10 followed by Swern oxidation²⁷ provided aldehyde 4 (83%, two steps). Union of 3 with aldehyde 4 was effected using the Mukaiyama reaction²⁸ of the derived silyl enol ether 11 to afford 12 as a 1:1 mixture of diastereomers (Scheme 3). Oxidation of 12 with Collin's reagent^{16a} afforded β -diketone 13 (64%). Global deprotection—spiroketalization of 13 was accomplished by treatment with 48% aqueous HF/CH₃CN/CH₂Cl₂ (6:9:50)²⁹ to produce the target spiroketal scaffold 2 as a single diastereomer (80%). Stereochemical assignment of scaffold 2 was supported by nuclear Overhauser effect (NOE) experiments (Scheme 3, 5% NOE between H₁ and H₂) and subsequently confirmed by X-ray crystallographic analysis of a functionalized spiroketal (cf. Figure 4).

Initial studies for scaffold modification focused on oximation of the spiroketal ketone functionality (Scheme 4).9g,30 Treatment of 2 with methoxylamine hydrochloride (NaOAc, 2:1 EtOH/H₂O) provided oxime **14** in quantitative yield. Reaction of 14 with 1,1'-carbonyldiimidazole (CDI) in THF followed by treatment of the intermediate imidazolide³¹ with excess L-phenylalanine ethyl ester afforded a carbamate ethyl ester, which after hydrolysis gave spiroketal acid 15. To evaluate the possible use of a "catch and release" protocol for product purification, 15 was incubated with PS-Trisamine resin³³ in CH₂Cl₂, the resin was washed, and products were released by treatment with 20% AcOH in CH₂Cl₂. Unfortunately, the released product 15 was contaminated with elimination product 16, which suggested the need for reduction of the spiroketal ketone to permit more robust elaboration chemistries (Scheme 5). Silvl protection of 2 afforded silvl ether 17, which was selectively reduced (NaBH₄, CeCl₃•7H₂O, MeOH, 0 °C)³⁴ to equatorial alcohol 18. We next implemented a plan to convert the secondary alcohol into a series of carbamates for the first diversity step. Hydroxyspiroketal 18 was converted to p-nitrophenyl carbonate 19 (4-nitrochloroformate, pyridine), which was treated with 1-naphthalenemethylamine to produce carbamate 20 in near-quantitative yield. For library synthesis, three primary amines were employed for the R₁ diversity step (cf. Figure 3). Terminal olefin **20** was converted to acid **21** in three steps by osmylation, oxidative cleavage using Pb(OAc)4, and chlorite oxidation.³⁵ In these reactions, only extractive workups were required for product purification.

R₂ and R₃ diversity elements were next incorporated onto the spiroketal scaffold using a series of streamlined solutionphase reactions. The R₂ diversity step was accomplished using amide bond formation (Scheme 6). Coupling of acid 21 with L-phenylalanine ethyl ester hydrochloride (EDC, HOBt, DIEA, CH₂Cl₂) gave amide **22** in near-quantitative yield (99%) after liquid-liquid extraction. 14b,c For library synthesis, three amino acid ethyl esters were employed (cf., Figure 3), in which case nine parallel reactions were effectively conducted on the Quest 210 (Argonaut Technologies) using 10 mL Teflon reaction vessels to facilitate reactions and subsequent liquid-liquid extractions directly in the same vessel. Introduction of the R₃ diversity element required silyl deprotection and carbamate formation of the pendant hydroxymethyl group. For example, treatment of 22 with excess HF pyridine in THF effected cleavage of the

Scheme 1a

$$(\pm) \qquad \qquad a \qquad \qquad b \qquad \qquad b \qquad \qquad Me \qquad \qquad Me$$

^a Reagents and conditions: (a) (*S*,*S*)-salen-Co, H₂O (0.55 equiv), AcOH (5 mol %), THF, 0 °C to room temp, 85%; (b) *n*-BuLi, THF, HMPA, −35 °C, 2h, −78 °C, 5, 73%; (c) TBDPSCl, imidazole, room temp, 24 h, 90%; (d) DDQ, CH₃CN/H₂O (9:1), room temp, 4 h, 81%.

Scheme 2^a

^a Reagents and conditions: (a) NaH, DMF, 0 °C to room temp, 3 h, p-MeOC₆H₄CH₂Br, 0 °C to room temp, 12 h, 98%; (b) AD-mix-β, t-BuOH/H₂O (1:1), 0 °C, 18 h, 90%; (c) acetone, TsOH, room temp, 30 min, 92%; (d) DDQ, CH₂Cl₂/H₂O (18:1), 0 °C, 5 h, 86%; (e) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, -70 °C, 96%.

Scheme 3^a

^a Reagents and conditions: (a) (i) KHMDS, THF, −78 °C, 1.5 h, (ii) TMSCl, −78 °C, 2 h, 98%; (b) **4**, −78 °C, BF₃OEt₂, 1.5 h, 65%; (c) CrO₃, Celite, pyridine, CH₂Cl₂, 3.5 h, 64%; (d) HF/CH₃CN/CH₂Cl₂ (6:9:50), 24 h, 80%.

TBS ether. After completion of the reaction, residual HF was neutralized using a liquid scavenger, methoxytrimethylsilane (MeOSiMe₃), which affords volatile side products. ³⁶ Pyridine was removed by directly passing reaction mixtures through a Waters Oasis MCX (mixed-mode cation exchange) cartridge (6 cm³) attached to Luer fittings below the reaction vessels to afford alcohol 23. Nine such parallel TBS deprotection reactions were carried out in this manner on the Quest 210. 23 was further converted to imidazolide 24 using a slight excess of CDI in CH₂Cl₂. After completion of the reaction, excess CDI and the byproduct imidazole were removed from the reaction mixture using solid-supported liquid extraction (SLE)³⁷ (1 M aqueous NaH₂PO₄³⁸ preloaded onto ChemElut Plus liquid-liquid extraction cartridges).³³ For carbamate formation, imidazolide 24 was reacted with isoamylamine (9.0 equiv) in toluene (50 °C, 12 h) to yield fully functionalized spiroketal 25 (98% yield, 88% HPLC purity). Excess amine and imidazole byproducts were removed by gravity elution of the reaction mixtures through a ChemElut Plus liquid-liquid extraction cartridge pretreated with 10% HCl. The choice of toluene for carbamate formation is derived from the requirement for use of waterimmiscible solvents in the SLE procedure.³⁷ For the R₃ diversity step, parallel carbamate formation reactions were carried out in 10 mL (13 mm × 150 mm) reactors on the Argonaut FirstMate synthesizer. Using the methodology described in Scheme 6, we synthesized a small library of 90 highly functionalized spiroketals (3 \times 3 \times 10) in yields ranging from 77 to 97% with HPLC purities from 80 to 96% (Table 1). Figure 4 also shows a single X-ray crystal structure analysis³⁹ of a representative highly functionalized spiroketal (33), which both confirms the absolute stereochemistry of

Scheme 4^a

a Reagents and conditions: (a) methoxylamine hydrochloride, EtOH/H₂O (2:1), room temp, 2 h, 100%; (b) (i) CDI, THF, room temp, 5h, (ii) L-phenylalanine ethyl ester hydrochloride, pyridine, 55 °C, 12 h, (iii) LiOH, H₂O, THF, room temp, 5 h, 95% (three steps); (c) (i) PS-Trisamine (4.0 equiv), CH₂Cl₂, (ii) 20% AcOH/CH2Cl2.

Scheme 5^a

a Reagents and conditions: (a) TBSCl, imidazole, DMF, room temp, 12 h, 90%; (b) CeCl₃·7H₂O, NaBH₄, MeOH, 0 °C, 70%; (c) 4-nitrophenylchloroformate, pyridine, THF, room temp, 20 min, 100%; (d) 1-naphthalenemethylamine, DMF, room temp, 4 h, 98%; (e) (i) OsO₄, NMO, t-BuOH/H₂O (2:1), (ii) Pb(OAc)₄, THF, (iii) NaClO₂, NaH₂PO₄, t-BuOH/2-methyl-2-butene (2:1), 97% (three steps).

the spiroketal scaffold and illustrates the interesting projection of functionality from the spiroketal core.

As an alternative to extractive workups, we have also investigated use of polymer reagent and scavengers for R₂ and R₃ diversity steps (Scheme 7). For example, treatment of spiroketal acid 21 (1.0 equiv) with PS-HOBt resin (DIC, catalyst DMAP, DMF/CH2Cl2 (1:2)) followed by resin washing led to the resin-bound HOBt ester 42. Release of product from resin with L-leucine ethyl ester hydrochloride (1.5 equiv, DIEA, CH₂Cl₂) afforded amide 43, in which case excess amino acid ester was removed by passing the reaction mixture through 12 mL ChemElut Plus cartridges pretreated with 10% HCl. Conversion of 43 to imidazolide 44 followed the earlier sequence (cf. $22 \rightarrow 24$, Scheme 6). Further conversion of 44 to carbamate 45 was accomplished by treatment of 44 with 4-(aminomethyl)pyridine (9 equiv, 50 °C, toluene). In this and other cases involving use of primary amines containing basic sites for carbamate formation, consecutive amine scavenging using PS-isocyanate resin^{33,40}

and removal of imidazole using SLE (ChemElut Plus cartridge, 1 M aqueous NaH₂PO₄) were found to produce spiroketal products in high yield and purities (cf. 45: 81% yield, 90% HPLC purity).

Spiroketal functionalization was also extended to a fourth diversity element as described in Scheme 8. Reaction of imidazolide 44 with benzylamine as previously described and workup using SLE afforded spiroketal 46. Ester hydrolysis of 46 (LiOH, THF) followed by acidification and extraction led to acid 47. Coupling of acid 47 with 2-diphenylethylamine (EDC, HOBt, DIEA, CH₂Cl₂) gave functionalized spiroketal product 48 in 95% overall yield and 86% HPLC purity after liquid-liquid extraction.

Finally, we have briefly examined an alternative method for spiroketal scaffold diversification involving direct reaction of the pendant alkene (Scheme 9). Nitrophenyl carbonate 19 was converted to carbamate 49 using previously described methods. Nitrile oxide 1,3-dipolar cycloaddition of 49 (1nitropropane, PhNCO, catalyst Et₃N, benzene, reflux)⁴¹

Figure 3. Building blocks used for the spiroketal library.

Scheme 6^a

^a Reagents and conditions: (a) L-phenylalanine ethyl ester hydrochloride, EDC, HOBt, DIEA, CH₂Cl₂, room temp, 12 h, 99%; (b) (i) HF•pyridine, THF, room temp, 4 h, (ii) MeOSiMe₃ (2 equiv), room temp, 2 h, (iii) Waters MCX cartridge, 100%; (c) (i) CDI (1.1 equiv), CH₂Cl₂, room temp, 4 h, (ii) ChemElut Plus cartridge (1 M NaH₂PO₄), 100%; (d) (i) isoamylamine, toluene, 50 °C, (ii) ChemElut Plus cartridge (10% HCl), 98%.

afforded spiroketal isoxazole **50** as a mixture of diastereomers. Streamlined removal of the TBS protecting group of **50** and imidazolide formation as described previously gave spiroketal imidazolide **51** in 71% (three steps) yield. Fully functionalized spiroketal **52** was obtained in 95% yield (96% HPLC purity) by reaction of **51** with 2-methoxyphenylamine and reaction workup using solid-supported liquid extraction. Such alternative functionalization protocols illustrate the versatility of the spiroketal scaffold and the possibility for preparation of a number of diverse structures.

Conclusion

We have synthesized a rigid spiroketal scaffold containing three independently addressable vectors for the functionalization and have used the scaffold to generate a small combinatorial library of natural product-like molecules. Synthesis of the compounds employed streamlined solution-phase synthesis methods including polymer-supported reagents and scavengers and solid-supported liquid extraction cartridges. Preliminary studies have shown that up to four diversity elements may be installed on the spiroketal scaffold to produce highly functionalized molecules. Additional and diverse functionalization reactions of the spiroketal scaffold may be performed to prepare other spiroketal chemotypes. We are currently examining the biological activity of these compounds as prospective antagonists of protein—protein interactions, as well as the preparation of alternative spiroketal scaffolds in an effort to produce larger and more diverse

Table 1. Representative Fully Functionalized Spiroketals

compound	R ₁	R ₂	R ₃	yield (%) ^a	purity (%) ^b
25	HN-	Ph EtO NH O Ph	HN Me	97	88
26 27	HN-S	EtO NHO O Me Me	HN—Ph	98 92	94
28	HN-	O Me H₃CO M NH O	HN-	93	89
29	HN O	EtO Ph	HN	77	86 0 R ₁
30	HN CO	Me Me EtO NH O	HN-_CI	82	87 R ₂
31	HN TO	Ö <u>M</u> e EtO √NH O Me	HN~_O~	74	87 R ₃ O
32	HN-\F	EtO NH	HNO	96	84
33	HN-_F	H₃CO Me O NH ○ Ph	HN Ph	79	85
34	HN-_F	EtO √ NH	N.	89	80
35	HN	O Me Me Me NH O	HN_OMe	80	92
36	HN-CO	EtO NH O Me	HN∽Ph	95	90
37	HN-	EtO NH	HN-	93	89
38	HN-	ä	HN—CF ₃	87	83
39	HN-	H ₃ CO Me NH O Me Me	HN-\CI	86	89
40	HN-	EtO NH	HN—Ph Ph	84	96
41	HN-O	H₃CO Me NH	H N ─Ph	82	94

^a Chemical yield based on weight of crude product obtained from purified spiroketal carbamates (cf. compound 20). ^b HPLC analysis: 20% CH₃CN/H₂O (integral at 230 nm).

libraries. The results of these and related studies will be reported in due course.

Experimental Section

General Information. ¹H NMR spectra were recorded on a 400 MHz spectrometer at ambient temperature with CDCl₃ as the solvent unless otherwise stated. ¹³C NMR spectra were recorded on a 75.0 MHz spectrometer at ambient temperature. Chemical shifts are reported in parts per million relative to chloroform (1 H, δ 7.24; 13 C, δ 77.0). Data for 1 H NMR are reported as follows: chemical shift, integration, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, b = broad), and coupling constants. All ¹³C NMR spectra were recorded with complete proton decou-

$$= \frac{\text{MeO} + \text{MeO} + \text{MeO}$$

Figure 4. Chem3D representation of the X-ray structure of spiroketal 33.

Scheme 7^a

^a Reagents and conditions: (a) PS-HOBt resin, DIC (1.0 equiv), DMAP (0.1 equiv), DMF/CH₂Cl₂ (1:3), 7 h; (b) (i) L-leucine ethyl ester hydrochloride (1.5 equiv), DIEA (3.0 equiv), CH₂Cl₂, 5 h, (ii) Chem-Elut cartridge (10% HCl), 78% (two steps); (c) (i) HF•pyridine, room temp, 4 h, (ii) MeOSiMe₃ (2 equiv), room temp, 2 h, (iii) Waters MCX cartridge; (d) (i) CDI, CH₂Cl₂, room temp, 4 h, (ii) ChemElut Plus cartridge (1 M NaH₂PO₄) (98%, two steps); (e) (i) 4-(aminomethyl)pyridine, toluene, 50 °C, 12 h, (ii) PS-isocyanate resin, (iii) ChemElut Plus cartridge (1 M NaH₂PO₄), 81%.

pling. Infrared spectra were recorded on a Nicolet Nexus 670 FT-IR. Chiral HPLC analysis was performed on an Agilent 1100 series (CHIRALCEL OD, column no. OD00CE-AI015). Achiral HPLC analysis was performed on an Agilent 1100 series (reverse-phase C-18 column) using water and CH₃CN as eluents. Optical rotations were recorded on an AUTOPOL III digital polarimeter at 589 nm and are reported as $[\alpha]^{22}$ _D (concentration in grams per 100 mL of solvent). High-resolution mass spectra were obtained in the Boston University mass spectrometry laboratory using a Finnegan MAT-90 spectrometer. Methylene chloride (CH₂Cl₂) and toluene were distilled from calcium hydride. Tetrahydrofuran was distilled from sodium and benzophenone. Analytical thin-layer chromatography was performed on 0.25 mm silica gel 60-F plates. Flash chromatography was performed using 200-400 mesh silica gel (Natland International Corporation). All other reagents were used as supplied by Sigma-Aldrich, Lancaster, TCI, and Alfa Aesar unless otherwise noted. ChemElut Plus cartridges (3.0 mL aqueous capacity (12 mL)) were obtained from Argonaut Technologies (part no. 900252). Waters Oasis MCX 6 cm³ cartridges (part no. 186000776) were obtained from Waters, Inc. PS-HOBt and PS-isocyanate resins were obtained from Argonaut Technologies (San Carlos, CA). Parallel reactions were carried out using the Quest-210 and FirstMate synthesizers (Argonaut Technologies, San Carlos, CA).

(*S*)-1,2-Epoxy-5-Hexene (5). To a stirred solution of (\pm)-1,2-epoxy-5-hexene (20 g, 0.20408 mol) and *S*,*S*-salen-Co catalyst (489 mg, 5 mol %) at 0 °C were added consecutively THF (1.5 mL), AcOH (175 μ L, 2.0 mol %), and water (1.5 mL, 0.55 equiv) at the same temperature. The reaction was warmed to room temperature and stirred for 16 h, at which

Scheme 8a

a Reagents and conditions: (a) (i) benzylamine, toluene, 50 °C, 12 h, (ii) ChemElut Plus cartridge (10% HCl), 98%; (b) (i) 1M aqueous LiOH, THF, room temp, 5 h, (ii) 5% HCl, 98%; (c) 2-diphenylethylamine, EDC, HOBt, DIEA, CH₂Cl₂, room temp,12 h, 95%.

Scheme 9^a

^a Reagents and conditions: (a) piperonylamine, DMF, 98%; (b) 1-nitropropane, phenyl isocyanate, Et₃N, benzene, reflux, 24 h; (c) (i) HF•pyridine, room temp, 4 h, (ii) MeOSiMe₃ (2 equiv), room temp, 2 h, (iii) Waters MCX cartridge; (d) (i) CDI, THF, room temp, 5 h, (ii) Chem-Elut cartridge (1 M NaH₂PO₄), 71% (three steps); (e) (i) 2-methoxyphenethylamine, toluene, 50 °C, 12 h, (ii) Chem-Elut cartridge (10% HCl), 95%.

time volatiles (epoxide) were vacuum-transferred (0.025 Torr) into a cooled (-78 °C) receiving flask. The recovered epoxide was passed through a small pad of silica gel to remove the residual water, and THF was removed using distillation at ambient pressure to afford (S)-1,2-epoxy-5hexene 5 (8.5 g, 85%) as a clear oil. For enantiomeric excess determination, the epoxide was ring-opened with 2-naphthalenethiol. To a stirred solution of epoxide (50 mg, 0.51 mmol) in MeOH (1.0 mL) were added at 0 °C 2-naphthalenethiol (90 mg, 0.56 mmol, 1.1 equiv) and Et₃N (57 mg, 0.56 mmol). After the solution was stirred for 2 h, water was added. The reaction mixture was extracted with EtOAc, and the organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The

crude product was purified by silica gel chromatography (10-20% EtOAc/hexanes) to yield the 2-naphthylsulfide derivative (125 mg, 95%) as a clear oil, 90% ee [chiral HPLC, 10% 2-propanol/hexane, Rt = 14.18 min (major), 15.45 min (minor)].

Hydroxydithiane (6).

A solution of 2-methyl-1,3-dithiane (8.2 g, 0.061 mol) in THF (30.0 mL) and HMPA (3.0 mL) was cooled to -30°C, and a solution of *n*-BuLi (2.6 M solution in hexane 25.90 mL, 0.0674 mol) was added slowly. The reaction mixture was stirred at -30 °C for 2 h. A solution of epoxide 5 (6.0 g, 0.061 mol) in THF (10 mL) was added dropwise to the reaction mixture at -78 °C. After the solution was stirred for 2 h at -78 °C, the reaction mixture was warmed to -40°C and stirred for a further 1 h. The reaction mixture was quenched with saturated NH₄Cl and extracted with EtOAc. The organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The crude product was purified by silica gel chromatography (2-10% EtOAc/hexanes) to yield compound 6 (9.6 g, 73%) as a clear oil. $[\alpha]^{22}_{D}$ -41.7° (c 2.0, CHCl₃). IR (neat): \tilde{v}_{max} = 3440, 3074, 1639, 1276, 995 cm⁻¹. ¹H NMR: δ 5.85–5.75 (m, 1H), 5.04-4.92 (m, 2H), 3.94-3.89 (m, 1H), 3.41 (s, 1H), 3.02-2.93 (m, 2H), 2.78-2.73 (m, 2H), 2.36 (dd, J =9.2, 14.8 Hz, 1H), 2.22–1.99 (m, 3H), 1.90–1.80 (m, 2H), 1.64-1.55 (m, 5H), 1.50-1.41 (m, 1H) ppm. ¹³C NMR: δ 138.6, 115.0, 68.42, 47.9, 47.7, 37.1, 30.0, 28.7, 27.0, 26.8, 24.8 ppm. HR-MS (CI) calcd for $C_{11}H_{20}OS_2$ [M + H]⁺, 233.0956; found, 233.1018.

TBDPS Dithiane (7).

To a solution of alcohol 6 (17. 0 g, 0.0787 mol) in DMF (17.0 mL) were added imidazole (13.4 g, 0.197 mol) and TBDPSC1 (24.96 g, 0.094 mol). After being stirred for 16 h at 25 °C, the reaction mixture was diluted with water and extracted with EtOAc. The organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The crude product was purified by silica gel chromatography (0-5% EtOAc/hexanes) to yield **7** (32.2 g, 90%) as a clear oil. $[\alpha]^{22}_D$ -4.0° (c 1.0, CHCl₃). IR (neat): $\tilde{v}_{\text{max}} = 3074$, 3048, 1639, 1276, 997 cm⁻¹. ¹H NMR: δ 7.73–7.69 (m, 4H), 7.42–7.34 (m, 6H), 5.59– 5.49 (m, 1H), 4.86–4.80 (m, 2H), 4.13–4.03 (m, 1H), 2.70– 2.48 (m, 4H), 2.32–2.17 (m, 2H), 2.09–1.97 (m, 2H), 1.87– 1.77 (m, 2H), 1.75-1.53 (m, 4H), 1.02 (bs, 9H) ppm. ¹³C NMR: δ 138.7, 136.3, 136.2, 134.8, 134.5, 129.8, 129.7, 127.8 114.5, 71.2, 47.8, 37.3, 29.2, 28.8, 27.3, 26.8, 25.2, 19.6 ppm. HR-MS (CI) calcd for $C_{27}H_{38}OS_2Si [M + H]^+$, 471.2133; found, 471.2208.

Methyl Ketone (3).

To a stirred solution of compound 7 (20 g, 0.044 mol) in CH₃CN/H₂O (9:1) (150 mL) was added DDQ (15.0 g, 0.066 mol, 1.5 equiv) dissolved in CH₃CN/H₂O (9:1) (150 mL) dropwise. After the mixture was stirred for 3 h, saturated NaHCO₃ was added and the reaction mixture was extracted with EtOAc. The organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography

(0–5% EtOAc/hexanes) to yield compound **3** (13.6 g, 81%) as a clear oil. [α]²²_D +17.0° (c 1.0, CHCl₃). IR (neat): $\tilde{\nu}_{\rm max}$ = 3071, 1715, 1640, 998 cm⁻¹. ¹H NMR: δ 7.69–7.66 (m, 4H), 7.44–7.36 (m, 6H), 5.64–5.54 (m, 1H), 4.90–4.83 (m, 2H), 4.26–4.20 (m, 1H), 2.53 (ABq, J = 6.0, 15.6 Hz, 2H), 2.04–2.00 (m, 2H), 1.97 (s, 3H), 1.56–1.51 (m, 2H), 1.04 (bs, 9H) ppm. ¹³C NMR: δ 207.3, 138.3, 136.1, 134.3, 134.2, 129.9, 127.8, 114.8, 69.8, 50.9, 36.5, 31.1, 29.4, 27.2, 19.6 ppm. HR-MS (CI) calcd for C₂₄H₃₂O₂Si [M⁺], 380.2172; found, 380.2197.

1-[(4-Methoxyphenyl)methoxy]hex-5-ene (8).

5-Hexenol (10.0 g, 0.1 mol, 1.0 equiv) was added at 0 °C to a flask containing NaH [5.28 g, 0.11 mol, 1.1 equiv (60%) in mineral oil, prewashed with 3×40 mL hexane)] and DMF (40 mL). The mixture was brought to room temperature, stirred for 1 h, and recooled to 0 °C. 4-Methoxybenzyl chloride (13.56 mL, 2.85 mmol, 1.0 equiv) was then added dropwise. After the mixture was stirred at room temperature for 12 h, the reaction was quenched with saturated NH₄Cl, diluted with water, and extracted with CH₂Cl₂. The organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The crude product was purified by silica gel chromatography (2-5% EtOAc/ hexanes) to yield compound 8 (21.8 g, 99%) as a clear oil. IR (neat): $\tilde{\nu}_{\text{max}} = 3075, 3048, 1036, 994 \text{ cm}^{-1}$. ¹H NMR: δ 7.25 (d, J = 8.0 Hz, 2H), 6.86 (d, J = 8.0 Hz, 2H), 5.85 5.71 (m, 1H), 5.02–4.93 (m, 2H), 4.42 (s, 2H), 3.76 (s, 3H), 3.44 (t, J = 6.8 Hz, 2H), 2.09-2.03 (m, 2H), 1.65-1.58(m, 2H), 1.50–1.43 (m, 2H) ppm. 13 C NMR: δ 159.2, 138.8, 130.9, 129.2, 128.4, 114.5, 113.8, 113.6, 72.5, 70.0, 55.2, 33.6, 29.3, 25.6 ppm. HR-MS (CI) calcd for $C_{14}H_{20}O_{2}$ [M⁺], 220.1463; found, 220.1471.

Diol (9).

To a stirred and cooled (0 °C) solution of AD-mix- β (50.0 g,) in t-BuOH/H₂O (1:1) (360 mL) was added compound 8 (12.0 g, 0.0545 mol), and the reaction mixture was stirred at 0 °C for 24 h. The reaction mixture was brought to room temperature and filtered, and the residue was washed with t-BuOH. The combined organic layers were concentrated in vacuo. After extraction of concentrated residue with EtOAc, the organic layer was dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (60-100% EtOAc/hexanes) to afford compound **9** (14.0 g, 90%) as a white solid. $[\alpha]^{22}D - 1.8^{\circ}$ (c 1.0, CHCl₃); 80% ee [chiral HPLC 20% 2-propanol/hexane, Rt = 17.79 min (major), 19.29 min (minor)]. IR (neat): $\tilde{\nu}_{\text{max}}$ = 327, 3074, 1652, 1170 cm⁻¹. ¹H NMR: δ 7.25 (d, J = 8.4 Hz, 2H), 6.87 (d, J = 8.4 Hz, 2H), 4.42 (s, 2H), 3.79 (s, 3H), 3.68-3.52 (m, 2H), 3.46-3.36 (m, 5H), 1.68-1.33 (m, 6H) ppm. ¹³C NMR: δ 159.3, 130.7, 129.4, 113.9, 72.7, 72.2, 70.1, 66.8, 55.4, 33.0, 29.7, 22.4 ppm. HR-MS (CI) calcd for C₁₄H₂₂O₄ [M⁺], 254.1518; found, 254.1496.

Acetonide (10).

To a stirred solution of compound 9 (14.0 g, 0.055 mol) in acetone (30 mL) was added p-toluenesulfonic acid monohydrate (100 mg, 0.525 mmol). After the mixture was stirred for 2 h at 25 °C, the solvent was concentrated in vacuo. The product was directly purified by silica gel chromatography (20% EtOAc/hexanes) to yield compound **10** (14.9 g, 92%) as a clear oil. $[\alpha]^{22}_D$ -8.3° (c 1.0, CHCl₃). IR (neat): $\tilde{\nu}_{max}$ = 3075, 1715, 1612, 1171 cm⁻¹. ¹H NMR: δ 7.26–7.23 (m, 2H), 6.88-6.85 (m, 2H), 4.42 (s, 2H), 4.09-4.00 (m, 2H), 3.78 (s, 3H), 3.50-3.42 (m, 2H), 1.67-1.60 (m, 4H), 1.51-1.45 (m, 2H), 1.40 (s, 3H), 1.38-1.37 (m, 1H), 1.35 (S, 3H) ppm. 13 C NMR: δ 159.2, 130.8, 129.2, 113.8, 108.6, 76.1, 72.6, 69.9, 69.5, 55.2, 33.5, 29.8, 27.0, 25.8, 22.6 ppm. HR-MS (CI) calcd for $C_{17}H_{26}O_4$ [M + H]⁺, 295.1831; found, 295.1873.

Acetonide Alcohol (53).

To a stirred and cooled (0 °C) solution of compound 10 (13.4 g, 0.046 mol) in CH₂Cl₂/H₂O (18:1) (304 mL), DDQ (11.4 g, 0.050 mol, 1.1 equiv) was added. After stirring for 4 h at 0 °C, the reaction was quenched with saturated NaHCO₃, diluted with water, and extracted with CH₂Cl₂. The organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (30-80% EtOAc/ hexanes) to afford compound 53 (6.8 g, 90%) as a clear oil. $[\alpha]^{22}_{\rm D} - 12.3^{\circ} (c \ 1.0, \text{CHCl}_3)$. IR (neat): $\tilde{\nu}_{\rm max} = 3411 \text{ cm}^{-1}$. ¹H NMR: δ 4.05–3.95 (m, 2H), 3.56 (t, J = 6.4 Hz, 2H), 3.44 (t, J = 7.2 Hz, 1H), 2.16 (bs, 1H), 1.65-1.38 (m, 5H), 1.36 (s, 3H), 1.32–1.31 (m, 1H), 1.35 (S, 3H) ppm. ¹³C NMR: δ 108.9, 76.8, 76.2, 69.6, 62.6, 33.4, 32.8, 27.1, 25.9, 22.2 ppm. HR-MS (CI) calcd for $C_9H_{18}O_3$ [M⁺], 174.1256; found, 174.1237.

Acetonide Aldehyde (4).

To a solution of oxalyl chloride (2.65 g, 0.021 mol, 1.4 equiv) in CH₂Cl₂ (30 mL) was added dropwise DMSO (3.73 g, 0.048 mol, 3.2 equiv) in CH_2Cl_2 (10 mL) at -70 °C. After the mixture was stirred for 10 min, a solution of alcohol 53 (2.6 g, 0.015 mol) in CH₂Cl₂ (10 mL) was added. After the mixture was stirred for 30 min, Et₃N (7.5 g, 0.075 mol, 5.0 equiv) was added. After 10 min, the cooling bath was removed and reaction was quenched with water and extracted with CH₂Cl₂. The organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (30% EtOAc/hexanes) to afford aldehyde 4 (2.47 g, 96%) as a clear oil. IR (neat): $\tilde{\nu}_{\text{max}} = 2723$, 1724 cm⁻¹. ¹H NMR: δ 9.54 (d, J = 4.8 Hz, 1H), 3.88-3.79 (m, 2H), 3.31-3.26 (m, 1H), 2.31-2.26 (m, 2H), 1.56-1.32 (m, 4H), 1.32-1.10 (m, 6H) ppm. 13 C NMR: δ 201.6, 108.6, 75.5, 69.1, 43.4, 32.8, 26.8, 25.5, 18.3 ppm.

Silyl Enol Ether (11).

To a stirred and cooled (-78 °C) solution of methyl ketone 3 (2.8 g, of 7.34 mmol) in THF (25 mL) was added KHMDS (0.66 M solution in toluene (15.7 mL, 10.315 mmol) dropwise over 15 min. After the mixture was stirred for 1.5 h at -78 °C, TMSCl (1.04 g, 9.5789 mmol) was added and stirring was continued for an additional 2 h at -78 °C. The reaction was quenched with saturated NaHCO3 and extracted with ethyl acetate, and the organic layer was washed with brine, dried over Na₂SO₄, and filtered and concentrated in vacuo. The product was passed through a small pad of silica gel pretreated with 2% Et₃N + 2% ethyl acetate/hexanes to yield silyl enol ether 11 (2.2 g, 93%) as a clear oil, which was used without further purification.

Aldol Product (12).

To a stirred and cooled (-78 °C) solution of silyl enol ether 11 (2.2 g, 4.87 mmol) in CH₂Cl₂ (2.5 mL) aldehyde 4 (0.837 g, 4.87 mmol) in CH₂Cl₂ (2.5 mL) was added and stirred for 5 min. BF₃•OEt₂ (0.829 g, 5.84 mmol, 1.2 equiv) was added dropwise, and stirring continued at −78 °C for an additional 1.5 h. The reaction mixture was quenched with saturated NaHCO₃ and extracted with CH₂Cl₂, the organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (10-35% EtOAc/ hexanes) to yield aldol product 12 (1.7 g, 65%) as a clear oil. [α]²²_D +7.0° (c 1.0, CHCl₃). IR (neat): $\tilde{\nu}_{max} = 3467$, 3071, 1708, 1640, 998 cm⁻¹. 1 H NMR: δ 7.67–7.63 (m, 4H), 7.43-7.35 (m, 6H), 5.62-5.52 (m, 1H), 4.88-4.83 (m, 2H), 4.26–4.17 (m, 1H), 4.10–3.99 (m, 2H), 3.95–3.80 (m, 1H), 3.50–3.46 (m, 1H), 2.93–2.91 (m, 1H), 2.59–2.34 (m, 4H), 2.15–1.95 (m, 2H), 1.62–1.47 (m, 6H), 1.43–1.39 (m, 4H), 1.33–1.27 (m, 4H), 1.01 (bs, 9H) ppm. 13 C NMR: δ 210.5, 138.2, 136.1, 134.1, 130.0, 127.8, 114.9, 108.9, 76.8, 76.2, 76.1, 69.7, 69.6, 69.5, 67.4, 67.2, 50.7, 50.4, 50.1, 36.6, 36.5, 36.3, 33.7, 33.6, 29.4, 27.2, 26.0, 22.1, 21.8, 19.6 ppm. HR-MS (CI) calcd for $C_{33}H_{48}O_5Si$ [M + H]⁺, 553.3271; found, 553.3415.

β -Diketone (13).

To a slurry of Celite (30 g) in CH₂Cl₂ (150 mL) was added pyridine (8.8 mL, 108.9 mmol, 21 equiv) followed by CrO₃

(4.2 g, 41.5 mmol, 10 equiv, dried in vacuo prior to use for 12 h). The resulting heterogeneous mixture was stirred for 2 h at room temperature. Aldol product 12 (2.8 g, 5.2 mmol) in CH₂Cl₂ (50 mL) was added slowly, and the reaction mixture was stirred vigorously for an additional 1.5 h. The dark-red reaction mixture was poured into saturated aqueous NH₄Cl and filtered, the residue was washed with ethyl acetate, and the filtrate was extracted with ethyl acetate. The combined reddish organic layer was passed through a small pad of Florisil, and the crude product was further eluted with EtOAc (200 mL). The resulting colorless filtrate was dried over MgSO₄, filtered, and concentrated in vacuo. The product was purified by silica gel chromatography (15-30% EtOAc/ hexanes) to yield β -diketone 13 (1.78 g, 64%) as a clear oil. $[\alpha]^{22}_{\rm D}$ +8.9° (c 1.0, CHCl₃). IR (neat): $\tilde{\nu}_{\rm max}$ = 3071, 1652, 1616, 997 cm⁻¹. 1 H NMR: δ 7.71–7.63 (m, 4H), 7.44– 7.34 (m, 6H), 5.63-5.53 (m, 1H), 5.31 (s, 1H), 4.88-4.82 (m, 2H), 4.17-4.00 (m, 3H), 3.48 (t, J = 7.2 Hz, 1H), 2.43-2.33 (m, 2H), 2.24 (t, J = 7.2 Hz, 1H), 2.03–1.97 (m, 2H), 1.71–1.47 (m, 6H), 1.39 (s, 3H), 1.34 (s, 3H), 1.02 (bs, 9H) ppm. 13 C NMR: δ 194.5, 191.0, 138.3, 136.1, 135.0, 134.3, 134.1, 129.9, 127.8, 114.9, 114.8, 109.0, 100.9, 75.9, 70.8, 69.6, 57.7, 50.7, 45.8, 43.6, 38.4, 36.4, 33.3, 29.25, 27.2, 26.8, 25.9, 22.0, 19.6 ppm. HR-MS (CI) calcd for C₃₃H₄₆O₅-Si $[M + H]^+$, 551.3115; found, 551.3189.

Spiroketal (2).

To a mixture of 48% aqueous HF/CH₃CN/CH₂Cl₂ (6:9:50) (72 mL) was added β -diketone 13 (2.4 g, 4.45 mmol), and the reaction was stirred at room temperature for 24 h. The mixture was poured carefully into an ice-cold saturated NaHCO₃ solution and extracted with EtOAc. The organic layer was washed with brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (10-35% EtOAc/hexanes) to yield spiroketal 2 (0.9 g, 80%) as a clear oil. IR (neat): $\tilde{\nu}_{max} =$ 3445, 3076, 1717, 981 cm⁻¹. ¹H NMR: δ 5.85–5.75 (m, 1H), 5.03-4.94 (m, 2H), 3.92-3.85 (m, 1H), 3.66-3.61 (m, 1H), 3.51-3.39 (m, 2H), 2.41-2.24 (m, 4H), 2.21-2.08 (m, 3H), 2.00–1.54 (m, 5H), 1.49–1.39 (m, 2H), 1.31–1.21 (m, 1H) ppm. 13 C NMR: δ 206.1, 138.1, 115.2, 99.5, 71.2, 68.6, 66.0, 52.0, 47.2, 35.6, 34.9, 30.0, 25.9, 18.6 ppm. HR-MS (CI) calcd for $C_{14}H_{22}O_4$ [M + H]⁺: 255.1518; found, 255.1587.

Spiroketal TBS Ether (17).

To a stirred solution of spiroketal **2** (2.1 g, 8.77 mmol) in DMF (4.0 mL) were added consecutively imidazole (1.6 g, 23.2 mmol) and TBSCl (1.6 g, 10.8 mmol). After being

stirred for 24 h, the reaction mixture was diluted with water and extracted with EtOAc. The organic layer was washed with water and brine, dried over MgSO4, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (5-20% EtOAc/hexanes) to yield spiroketal 17 (2.7 g, 90%) as a light-yellow oil. IR (neat): $\tilde{\nu}_{\text{max}} = 3077, 1726, 1641, 983 \text{ cm}^{-1}. {}^{1}\text{H NMR}: \delta 5.88-$ 5.78 (m, 1H), 5.05–4.95 (m, 2H), 3.97–3.91 (m, 1H), 3.62– 3.56 (m, 1H), 3.50 (dd, J = 6.4, 10.4 Hz, 1H), 3.42 (dd, J= 4.8, 10.8 Hz, 1H), 2.40-2.24 (m, 4H), 2.18-2.10 (m, 2H), 1.95-1.57 (m, 6H), 1.42 (ddd, J = 4.0, 4.4, 13.2 Hz, 1H), 1.20–1.11 (m, 1H), 0.84 (s, 9H), 0.01 (s, 3H), 0.00 (s, 3H) ppm. 13 C NMR: δ 205.9, 138.2, 115.2, 99.3, 71.6, 68.1, 66.6, 52.1, 47.2, 35.7, 34.9, 30.1, 26.6, 26.1, 18.9, 18.5, -5.1,-5.2 ppm. HR-MS (CI) calcd for $C_{20}H_{36}O_4Si$ [M + H]⁺, 369.2383; found, 369.2439.

Spiroketal Alcohol (18).

To a stirred and cooled (0 °C) solution of spiroketal 17 (1.9 g, 5.1630 mmol) in MeOH (10 mL) was added CeCl₃•7H₂O (2.3 g, 6.2 mmol, 1.2 equiv), and the reaction mixture was stirred for 10 min. To the resulting clear solution was added NaBH₄ (235 mg, 6.2 mmol, 1.2 equiv). After being stirred for 20 min, the reaction mixture was diluted with water and extracted with EtOAc, and the organic layer washed with water and brine, dried over MgSO4, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (10-30% EtOAc/hexanes) to afford alcohol 18 (1.34 g, 70%) as a clear oil. IR (neat): $\tilde{\nu}_{\text{max}} =$ 3354, 3077, 1652, 977 cm⁻¹. 1 H NMR: δ 5.88–5.78 (m, 1H), 5.03-4.92 (m, 2H), 4.16-4.08 (m, 1H), 3.67-3.50 (m, 3H), 3.46-3.42 (m, 1H), 2.31-2.22 (m, 1H), 2.14-2.05 (m, 1H), 2.01–1.96 (m, 1H), 1.92–1.81 (m, 2H), 1.68–1.37 (m, 7H), 1.26–1.07 (m, 3H), 0.86 (s, 9H), 0.02 (s, 6H) ppm. ¹³C NMR: δ 138.9, 114.7, 97.7, 70.7, 67.5, 66.9, 65.0, 45.2, 41.2, 35.6, 35.5, 30.4, 27.3, 26.1, 18.9, 18.5, -5.0, -5.1 ppm. HR-MS (CI) calcd for C₂₀H₃₈O₄Si [M⁺], 370.2539; found, 370.2494.

Spiroketal Nitrophenyl Carbamate (19).

4-Nitrophenyl chloroformate (1.06 g, 5.270 mmol, 1.5 equiv) was added to a stirred solution of alcohol **18** (1.3 g, 3.51 mmol) in THF (10 mL) and pyridine (0.89 g, 11.24 mmol, 3.2 equiv). After the mixture was stirred for 20 min, water was added and the reaction mixture was extracted with EtOAc. The organic layer was washed with 5% HCl, water, saturated NaHCO₃, water, and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified

by silica gel chromatography (10% EtOAc/hexanes) to yield **19** (1.78 g, 100%) as a clear oil. ¹H NMR: δ 8.25 (d, J =9.2 Hz, 2H), 7.35 (d, J = 9.2 Hz, 2H), 5.87-5.79 (m, 1H), 5.23-5.17 (m, 1H), 5.05-4.94 (m, 2H), 3.80-3.68 (m, 1H), 3.63-3.51 (m, 2H), 3.50-3.44 (m, 1H), 2.30-2.23 (m, 1H), 2.18-2.09 (m, 2H), 1.94-1.82 (m, 1H), 1.70-1.30 (m, 10H), 0.87 (s, 9H), 0.04 (s, 6H) ppm.

Representative Procedure for Carbamate Preparation: Spiroketal Carbamate (20).

To a stirred solution of 19 (480 mg, 0.943 mmol) in DMF (2.0 mL) was added 1-naphthalenemethylamine (222 mg, 1.42 mmol, 1.5 equiv). After being stirred for 4 h, the reaction mixture was diluted with water and extracted with EtOAc. The organic layer was washed with 5% HCl, water, saturated NaHCO₃, water, and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was purified by silica gel chromatography (10–15% EtOAc/hexanes) to yield carbamate 20 (515 mg, 98%) as a clear oil. IR (neat): $\tilde{\nu}_{\text{max}} = 3336, 3077, 1716, 1652, 978 \text{ cm}^{-1}$. ¹H NMR: $\delta 8.02$ (d, J = 8.0 Hz, 1H), 7.86 (d, J = 8.0 Hz, 1H), 7.80-7.77(m, 1H), 7.55-7.48 (m, 2H), 7.41-7.38 (m, 2H), 5.88-5.78 (m, 1H), 5.18–5.12 (m, 1H), 5.04–4.93 (m, 2H), 4.85– 4.75 (m, 3H), 3.72–3.68 (m, 1H), 3.61–3.56 (m, 2H), 3.47– 3.42 (m, 1H), 2.29–2.22 (m, 1H), 2.15–2.00 (m, 3H), 1.95– 1.79 (m, 1H), 1.65–1.11 (m, 9H), 0.89 (s, 9H), 0.07(s, 6H) ppm. ¹³C NMR: δ 155.9, 38.8, 134.1, 131.6, 129.0, 128.7, 126.4, 126.1, 125.6, 123.6, 114.8, 97.7, 70.7, 68.8, 67.2, 66.9, 43.4, 41.6, 37.9, 35.5, 35.4, 30.3, 27.5, 26.2, 18.8, -5.0,-5.1 ppm.

Representative Procedure for Carboxylic Acid Formation: Spiroketal Carbamate Acid (21).

To a stirred solution of carbamate 20 (515 mg, 0.93 mmol) in t-BuOH/H₂O (2:1) (3.0 mL) was added 4-methylmorpholine N-oxide (120 mg, 1.02 mmol, 1.1 equiv) followed by OsO_4 (4 wt % solution in water, 50 μ L). After the mixture was stirred for 12 h, the solvent was evaporated in vacuo, brine was added, and the reaction mixture was extracted with ethyl acetate. The organic layer was dried over MgSO4 and filtered and concentrated in vacuo to yield the diol intermediate (546 mg, 100%) as a light-brown oil, which was used without further purification.

To a stirred solution of the above diol (540 mg, 0.9199 mmol) in THF (10 mL) was added at room temperature Pb(OAc)₄ (530 mg, 1.1959 mmol, 1.3 equiv). After the mixture was stirred for 30 min, water was added and the reaction mixture was extracted with ethyl acetate. The organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo to yield the spiroketal aldehyde (509 mg, 100%) as a light-yellow oil that was used without further purification.

To a solution of the above aldehyde (500 mg, 0.9 mmol) in tert-butyl alcohol (3.0 mL) was added 2-methyl-2-butene (1.5 mL). A solution of sodium chlorite (162 mg, 1.8 mmol, 2.0 equiv, 80% NaClO₂) and sodium dihydrogen phosphate (216 mg, 1.8 mmol, 2.0 equiv) in H₂O (1.5 mL) was added slowly to the solution, and the resulting solution was stirred for 5 h at room temperature. The solvent was concentrated in vacuo, water (10 mL) was added, and the reaction acidified to pH 4.0 with 5% HCl. The reaction was extracted with EtOAc. The organic layer was washed with brine, dried over MgSO₄, and filtered and concentrated in vacuo to yield 21 (500 mg, 97%) as a white solid. IR (neat): $\tilde{\nu}_{\text{max}} = 3338$, 3077, 1711, 1649, 997 cm⁻¹. ¹H NMR: δ 8.00 (d, J = 7.2. Hz, 1H), 7.85 (d, J = 8.0 Hz, 1H), 7.79 - 7.77 (m, 1H), 7.54 -7.47 (m, 2H), 7.40–7.39 (m, 2H), 5.22–5.09 (m, 1H), 4.88– 4.70 (m, 3H), 3.79–3.62 (m, 1H), 3.50–3.44 (m, 3H), 2.62– 2.54 (m, 1H), 2.45–2.37 (m, 1H), 2.07–1.97 (m, 2H), 1.88– 1.72 (m, 3H), 1.68-1.50 (m, 3H), 1.44-1.15 (m, 5H), 0.89 (s, 9H), 0.0(s, 6H) ppm. 13 C NMR: δ 179.4, 156.7, 134.9, 134.8, 132.4, 129.8, 129.5, 127.5, 127.2, 127.0, 126.4, 124.4, 98.7, 71.6, 69.3, 67.8, 67.7, 44.1, 42.3, 38.4, 36.0, 31.7, 28.0, 27.0, 19.5, 19.4, -4.1, -4.2 ppm. HR-MS (CI) calcd for $C_{31}H_{45}NO_7Si [M + H]^+$, 572.2965; found, 572.3049.

Representative Procedure for Amide Formation: Spiroketal Amide (22).

To a 10 mL Quest 210 Teflon reaction vessel was added a solution of acid 21 (100 mg, 0.175 mmol) in CH₂Cl₂ (2.0 mL). To the above acid were added consecutively Lphenylalanine ethyl ester hydrochloride (56.0 mg, 0.25 mmol), 1-hydroxybenzotriazole (HOBt) (33.0 mg, 0.25 mmol), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC) (47 mg, 0.25 mmol), and diisopropylethylamine (DIEA) (91 mg, 0.77 mmol). After the mixture was agitated for 12 h, water was added and the reaction mixture was extracted with EtOAc. The organic layer was washed consecutively with 5% HCl, water, and brine, dried over MgSO₄, and filtered and concentrated in vacuo to afford amide 22 (129 mg, 99%) as a white solid that was used without further purification. IR (neat): $\tilde{\nu}_{\text{max}} = 3326, 3076,$ 1733, 1711, 1635 cm⁻¹. ¹H NMR: δ 7.84 (d, J = 8.0. Hz, 1H), 7.68 (d, J = 8.0 Hz, 1H), 7.62–7.60 (m, 1H), 7.39– 7.30 (m, 2H), 7.24–7.21 (m, 2H), 7.12–7.03 (m, 3H), 6.93– 6.91 (m, 2H), 5.82 (d, J = 8.0 Hz, 1H), 5.01–4.95 (m, 1H), 4.76-4.58 (m, 4H), 3.96 (q, J = 7.2, 14.0 Hz, 2H), 3.54-3.44 (m, 1H), 3.44-3.40 (m, 1H), 3.35-3.26 (m, 2H), 2.982.89 (m, 2H), 2.33–2.25 (m, 1H), 2.07–1.99 (m, 1H), 1.90–1.80 (m, 2H), 1.64–1.56 (m, 3H), 1.46–1.33 (m, 3H), 1.26–0.95 (m, 7H), 0.73 (s, 9H), -0.09 (s, 3H), -0.1 (s, 3H) ppm. $^{13}\mathrm{C}$ NMR: δ 172.4, 171.9, 155.8, 136.2, 134.1, 131.5, 129.5, 129.0, 128.7, 127.2, 126.7, 126.4, 126.1, 125.6, 123.6, 97.7, 70.8, 68.5, 67.1, 66.9, 61.6, 53.3, 43.3, 41.5, 38.2, 37.6, 35.2, 33.0, 31.6, 27.3, 26.1, 18.7, 18.6, 14.3, -5.0, -5.1 ppm. HR-MS (CI) calcd for $\mathrm{C_{42}H_{58}N_2O_8Si}$ [M + H]+, 747.3962; found, 747.4073.

Representative Procedure for TBS Group Deprotection: Spiroketal Alcohol (23).

To a 10 mL Quest 210 Teflon reaction vessel was added amide 22 (129 mg, 0.173 mmol) dissolved in THF (0.5 mL) followed by 1.0 M solution of HF/pyridine (3.5 mL, 20 equiv) in THF. After the mixture was agitated for 4 h, MeOSiMe₃ (962 µL, 7.0 mmol, 2.0 equiv) was added to neutralize HF and the reaction mixture was agitated for a further 2 h. The reaction mixture was directly passed through a Waters MCX cartridge attached to the Quest 210 lower Luer manifold into a collection vial. The cartridge was washed further with THF, and the combined filtrate was concentrated in vacuo to afford alcohol 23 (109 mg, 100%) as a white solid, which was used without further purification. IR (neat): $\tilde{v}_{\text{max}} = 3400$, 3330, 3061, 1710, 1659 cm⁻¹. ¹H NMR: δ 7.98 (d, J = 8.0. Hz, 1H), 7.84 (d, J = 7.6 Hz, 1H), 7.78–7.76 (m, 1H), 7.54–7.46 (m, 2H), 7.42–7.37 (m, 2H), 7.28-7.19 (m, 3H), 7.08-7.07 (m, 2H), 6.0 (d, J =7.6 Hz, 1H), 5.19-5.07 (m, 1H), 4.99-4.87 (m, 1H), 4.85-4.66 (m, 3H), 4.12 (q, J = 7.2, 14.0 Hz, 2H), 3.70-3.40 (m, 4H), 3.12-3.04 (m, 2H), 2.45-2.37 (m, 1H), 2.27-2.19 (m, 1H), 2.06-1.99 (m, 3H), 1.99-1.72 (m, 4H), 1.64-1.50 (m, 2H), 1.49–1.24 (m, 4H), 1.21 (t, J = 7.2 Hz, 3H) ppm. ¹³C NMR: δ 172.4, 172.0, 155.9, 136.2, 134.1, 133.9, 131.5, 129.5, 129.0, 128.7, 127.3, 126.8, 126.5, 126.1, 125.6, 123.5, 97.9, 70.7, 68.4, 67.2, 66.3, 61.7, 53.4, 43.3, 41.5, 38.2, 37.6, 35.2, 32.9, 31.6, 26.4, 18.5, 14.4, 14.3 ppm. HR-MS (CI) calcd for $C_{36}H_{44}N_2O_8$ [M⁺], 632.3098; found, 632.3040.

Representative Procedure for Imidazolide Formation: Spiroketal Imidazolide (24).

To a 10 mL (13 mm \times 150 mm) reactor on the FirstMate synthesizer, a solution of alcohol **23** (109 mg, 0.17 mmol) in CH₂Cl₂ (2.0 mL) and 1,1'-carbonyldiimidazole (CDI) (31

mg, 0.19 mmol, 1.1 equiv) were added. After being stirred for 4 h, the reaction mixture was eluted through a 12 mL ChemElut Plus cartridge pretreated with 3.0 mL of 1 M NaH₂PO₄. The cartridge was washed with 1.0 mL of CH₂-Cl₂, and the combined organic layers were concentrated in vacuo to afford imidazolide 24 (125 mg, 100%) as a white solid. IR (neat): $\tilde{v}_{\text{max}} = 3317, 3061, 1765, 1719 1665 \text{ cm}^{-1}$. ¹H NMR: δ 8.15 (bs, 1H), 7.98 (d, J = 8.0. Hz, 1H), 7.86 (d, J = 7.6 Hz, 1H), 7.80-7.77 (m, 1H), 7.57-7.47 (m, 1H)2H), 7.44-7.38 (m, 3H), 7.28-7.21 (m, 3H), 7.10-7.02 (m, 3H), 5.96 (d, J = 7.6 Hz, 1H), 5.16-5.02 (m, 1H), 4.92-4.86 (m, 1H), 4.85-4.70 (m, 3H), 4.35-4.26 (m, 2H), 4.12 (q, J = 7.2, 14.0 Hz, 2H), 3.90-3.80 (m, 1H), 3.68-3.60(m, 1H), 3.15-3.04 (m, 2H), 2.38-2.32 (m, 1H), 2.25-2.17 (m, 1H), 2.06–1.99 (m, 2H), 1.99–1.74 (m, 3H), 1.66– 1.54 (m, 3H), 1.47-1.40 (m, 1H), 1.36-1.16 (m, 6H) ppm. ¹³C NMR: δ 172.1, 171.9, 155.8, 148.9, 136.2, 134.1, 133.9, 131.5, 131.0, 129.5, 129.0, 128.7, 127.3, 126.8, 126.5, 126.2, 125.6, 123.5, 98.1, 70.6, 68.2, 67.6, 67.5, 61.7, 53.3, 43.3, 41.3, 38.2, 37.5, 34.9, 32.7, 31.5, 29.9, 26.8, 18.4, 14.3 ppm. HR-MS (CI) calcd for C₄₀H₄₆N₄O₉ [M⁺], 726.3265; found, 726.3227.

Representative Procedure for Imidazolide Formation: Trifunctional Spiroketal (25).

To a 10 mL 13 mm × 150 mm reactor on the FirstMate synthesizer, a solution of imidazolide 24 (10.0 mg, 0.014 mmol) in toluene (0.5 mL) was added followed by isoamylamine (11.0 mg, 0.124 mmol, 9.0 equiv), and the reaction mixture was stirred at 50 °C for 12 h. The reaction mixture was brought to room temperature and eluted through a 12 mL ChemElut Plus cartridge pretreated with 3.0 mL of 10% HCl. The cartridge was further washed with toluene (2.5 mL), and the combined organic layers were concentrated to afford compound 25 (10.1 mg, 98%) as a waxy solid. ¹H NMR: δ 7.98 (d, J = 7.6 Hz, 1H), 7.86 (d, J = 8.0 Hz, 1H), 7.80-7.78 (m, 1H), 7.59–7.47 (m, 2H), 7.44–7.38 (m, 2H), 7.29– 7.20 (m, 3H), 7.14–7.06 (m, 2H), 5.96 (d, J = 6.4 Hz, 1H), 5.20-5.05 (m, 1H), 4.90-4.70 (m, 4H), 4.12 (q, J = 7.2, 14.0 Hz, 2H), 4.02–3.89 (m, 2H), 3.71–3.59 (m, 2H), 3.25– 3.05 (m, 4H), 2.48–2.36 (m, 1H), 2.32–2.22 (m, 1H), 2.12– 1.94 (m, 2H), 1.84-1.72 (m, 3H), 1.68-1.11 (m, 14H), 0.96 (d, J = 6.4 Hz, 6H) ppm. ¹³C NMR: δ 172.4, 172.0, 156.8, 155.8, 136.2, 134.1, 134.0, 131.5, 129.5, 129.0, 128.7, 127.3, 126.8, 126.5, 126.2, 125.6, 123.6, 97.9, 68.6, 68.5, 67.7, 67.2, 61.7, 53.4, 43.4, 41.4, 39.6, 39.1, 38.3, 37.7, 35.0, 32.7, 31.4, 29.9, 26.9, 25.9, 22.7, 18.7, 14.3 ppm. HR-MS (CI) calcd for $C_{42}H_{55}N_3O_9$ [M + H]⁺, 746.3938; found, 746.4019.

Procedure for Amide Formation Using PS-HOBt Resin: Spiroketal Amide (43). To a 10 mL Quest 210 reaction vessel was added PS-HOBt resin (1.12 mmol/g, 110

mg, 0.123 mmol) and a mixture of DMF/CH₂Cl₂ (1:3) (1.2 mL). To the above resin were added consecutively acid 21 (70 mg, 0.123 mmol 1.0 equiv), DMAP (1.5 mg, 10 mol %), and 1,3-diisopropylcarbodiimide (DIC) (16.0 mg, 0.123 mmol 1.0 equiv). After the mixture was agitated for 7 h, the resin was filtered, washed with DMF, THF, DMF, CH₂Cl₂ $(3 \times 5 \text{ mL each})$, and dried under high vacuum for 2 h to yield PS-HOBt ester resin 42. To a 10 mL Quest 210 reaction vessel was added dried PS-HOBt ester resin 42 (110 mg, 0.123 mmol) and CH₂Cl₂ (1.0 mL). To the above resin were added consecutively L-leucine ethyl ester hydrochloride (36.0 mg, 0.185 mmol, 1.5 equiv) and disopropylethylamine (DIEA) (48 mg, 0.77 mmol). After the mixture was agitated for 6 h, the solvent was removed and the resin was washed with CH_2Cl_2 (2 × 2 mL). The combined organic solvents were directly eluted through a 12 mL ChemElut Plus cartridge pretreated with 3.0 mL of 10% HCl. The cartridge was washed with a further 1.0 mL of CH₂Cl₂, and the combined organic layers were concentrated in vacuo to afford amide 43 (68.2 mg, 78%) as a waxy solid. ¹H NMR: δ 8.0 (d, J = 8.0. Hz, 1H), 7.86 (d, J = 7.6 Hz, 1H), 7.99-7.78(m, 1H), 7.53–7.48 (m, 2H), 7.42–7.40 (m, 1H), 5.92 (d, J = 8.8 Hz, 1H), 5.21-5.09 (m, 1H), 4.89-4.68 (m, 3H), 4.64-4.49 (m, 1H), 4.14 (q, J = 6.8, 14.0 Hz, 2H), 3.73-3.65 (m, 1H), 3.62–3.56 (m, 1H), 3.54–3.42 (m, 2H), 2.54– 2.45 (m, 1H), 2.28–2.19 (m, 1H), 2.10–1.90 (m, 2H), 1.89– 1.74 (m, 2H), 1.50–1.40 (m, 8H), 1.39–1.10 (m, 7H), 0.98– 0.90 (m, 6H), 0.089 (s, 9H), 0.07 (s, 6H) ppm.

Trifunctional Spiroketal (45).

To a 10 mL (13 mm × 150 mm) reactor on the FirstMate synthesizer containing a solution of imidazolide 44 (10.0 mg, 0.014 mmol) in toluene (0.5 mL) was added 4-(aminomethyl)pyridine (14.0 mg, 0.043 mmol, 9.0 equiv), and the reaction mixture was stirred at 50 °C for 12 h. The reaction mixture was brought to room temperature, PS-isocyanate resin (1.25 mmol/g, 0.220 g, 0.275 mmol, 2.1 equiv) and toluene (1.5 mL) were added, and the mixture was agitated for 5 h. The reaction mixture was added directly to a 12 mL ChemElut cartridge was pretreated with 3.0 mL of 1 M NaH₂-PO₄ and was gravity-eluted. The cartridge was further washed with toluene (1.0 mL), and the combined organic layers were concentrated to afford compound 45 (8.5 mg, 81%). ¹H NMR: δ 8.60–8.53 (m, 2H), 8.01 (d, J = 7.6 Hz, 1H), 7.86 (d, J = 8.0 Hz, 1H), 7.80-7.77 (m, 1H), 7.56-7.48 (m, 1H)2H), 7.42-7.38 (m, 2H), 5.90 (d, J = 8.0 Hz, 1H) 5.70-5.62 (m, 1H), 5.55–5.38 (m, 1H), 4.92–4.86 (m, 1H), 4.82– 4.70 (m, 2H), 4.61-4.52 (m, 1H), 4.41 (t, J = 6.8 Hz, 2H),4.15 (q, J = 6.8, 14.0 Hz, 2H), 4.10-4.07 (m, 1H), 3.99-3.94 (m, 1H), 3.76–3.45 (m, 2H), 2.56–2.37 (m, 1H), 2.32– 2.20 (m, 1H), 2.14-1.95 (m, 2H), 1.86-1.75 (m, 2H), 1.53-

1.40 (m, 4H), 1.38–1.15 (m, 10H), 0.96–0.53 (m, 8H) ppm. HR-MS (CI) calcd for $C_{40}H_{52}N_4O_9$ [M + H]⁺, 733.3734; found, 733.3767.

Compound (48).

To a stirred solution of ester 46 (17.0 mg, 0.0237 mmol) in THF (1.0 mL) was added 1.0 M aqueous solution of LiOH $(236 \mu L, 10 \text{ equiv})$. After being stirred for 5 h, the reaction mixture was acidified with 10% HCl to pH 3.0 and extracted with EtOAc. The organic layer was washed with brine, dried over MgSO₄, and concentrated in vacuo to afford acid 47 (15 mg, 92%) as a waxy solid.

To a stirred solution of the above acid 47 (14.0 mg, 0.0203 mmol) in CH₂Cl₂ (1.0 mL) were added consecutively 2,2diphenylethylamine (6.0 mg, 0.029 mmol), 1-hydroxybenzotriazole (HOBt) (4.0 mg, 0.029 mmol), 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC) (6.0 mg, 0.029 mmol), and diisopropylethylamine (DIEA) (8.0 mg, 0.061 mmol). After the mixture was stirred for 12 h, water was added and the reaction mixture was extracted with EtOAc. The organic layer was washed with 5% HCl, water, and brine, dried over MgSO₄, and filtered and concentrated in vacuo to afford amide 48 (17.2 mg, 96%) as a waxy solid. ¹H NMR: δ 8.01 (d, J = 7.6 Hz, 1H), 7.85 (d, J = 8.4 Hz, 1H), 7.80–7.77 (m, 1H), 7.55–7.47 (m, 2H), 7.43–7.39 (m, 2H), 7.34–7.19 (m, 15H), 6.02–5.86 (m, 2H), 5.34 (bs, 1H), 5.18-5.05 (m, 1H), 4.90-470 (m, 3H), 4.42-4.28 (m, 2H), 4.18-4.12 (m, 2H), 4.02-3.90 (m, 3H), 3.79-3.59 (m, 3H), 2.34-2.24 (m, 2H), 2.14-1.92 (m, 3H), 1.90-1.78 (m, 2H), 1.74-1.38 (m, 6H), 1.33-1.13 (m, 4H), 1.01-0.72 (m, 6H) ppm. HR-MS (CI) calcd for $C_{53}H_{62}N_4O_8$ [M + H]⁺, 883.4568; found, 883.4584.

Spiroketal Isoxazole (52). To a stirred solution of alkene 49 (30.0 mg, 0.055 mmol) in benzene were added consecutively 1-nitropropane (7.0 mg, 0.083 mmol, 1.5 equiv), phenyl isocyanate (20.0 mg, 0.166 mmol, 3.0 equiv), and Et₃N (20 μ L, catalytic amount), and the reaction mixture was stirred at 25 °C for 30 min. The resultant white solid suspension was refluxed for 24 h. After the mixture was cooled, water was added and the mixture was extracted with ethyl acetate, the organic layer was washed with water and brine, dried over MgSO₄, and filtered and concentrated in vacuo. The product was passed through small pad of silica gel, eluting with EtOAc to yield isoxazole **50** (34 mg, 100%), which was used without further purification.

To a 10 mL Quest 210 reaction vessel was added the above isoxazole 50 (34 mg, 0.0552 mmol) dissolved in THF (0.5 mL), followed by HF•pyridine (1.0 M in THF, 2.76 mL, 2.76 mmol, 50 equiv). After the mixture was mixed for 4 h, Me-OSiMe₃ (760 μ L, 5.52 mmol, 2.0 equiv) was added to neutralize the HF and the reaction mixture wasagitated for a further 2 h. The reaction mixture was directly passed through a Waters MCX cartridge attached to the Quest 210 lower Luer manifold and collected directly into a scintillation vial. The cartridge was washed further with THF, and the combined filtrates were concentrated in vacuo to afford an alcohol (28.0 mg, 100%), which was used without further purification.

To a 10 mL (13 mm \times 150 mm) reactor on the FirstMate synthesizer, a solution of the above alcohol (28 mg, 0.056 mmol) in CH₂Cl₂ (1.5 mL) was added followed by 1,1'carbonyldiimidazole (CDI) (140 mg, 0.084 mmol, 1.5 equiv). After the mixture was stirred for 4 h, the reaction mixture was eluted through a 12 mL ChemElut Plus cartridge pretreated with 3.0 mL of 1 M NaH₂PO₄. The cartridge was washed with 1.0 mL of CH₂Cl₂, and the combined organic layers were concentrated in vacuo. The product was purified by silica gel chromatography (50-80% EtOAc/hexanes) to yield imidazolide **51** (23.8 mg, 71%). ¹H NMR: δ 8.12 (s, 1H), 7.42 (s, 1H), 7.06 (s, 1H), 6.75–6.68 (m, 3H), 5.92 (s, 2H), 5.07-5.02 (m, 1H), 4.90-4.80 (m, 1H), 4.56-4.45 (m, 1H), 4.32-4.31 (m, 2H), 4.21-4.20 (m, 2H), 3.86-3.82 (m, 1H), 3.62-3.50 (m, 1H), 2.95 (dd, J = 10.0, 16.4 Hz, 1H), 2.52 (dd, J = 8.0, 16.8 Hz, 1H), 2.36-2.29 (m, 2H), 2.19-1.99 (m, 2H), 1.92–1.80 (m, 2H), 1.73–1.55 (m, 5H), 1.50– 1.40 (m, 2H), 1.39-1.11 (m, 6H) ppm.

To a stirred solution of imidazolide **51** (14.0 mg, 0.023 mmol) in toluene (0.5 mL) was added 2-methoxyphenethylamine (32.0 mg, 0.211 mmol, 9.0 equiv), and the reaction mixture was stirred at 50 °C for 12 h. The reaction was brought to room temperature and eluted through a 12 mL ChemElut Plus cartridge pretreated with 3.0 mL of 10% HCl. The cartridge was further washed with toluene (2.5 mL), and the combined organic layers were concentrated to afford 52 (14.0 mg 95%) as a waxy solid. ¹H NMR: δ 7.20–7.16 (m, 1H), 7.12-7.11 (m, 1H), 6.89-6.83 (m, 2H), 6.76-6.69 (m, 3H), 5.92 (s, 2H), 5.17-5.02 (m, 1H), 4.98-4.92 (m, 1H), 4.90-4.80 (m, 1H), 4.58-4.46 (m, 1H), 4.28-4.10 (m, 2H), 4.09-3.98 (m, 1H), 3.93-3.88 (m, 1H), 3.82 (s, 3H), 3.72-3.52 (m, 2H), 3.44-3.32 (m, 2H), 2.95 (dd, J = 10.4, 16.8Hz, 1H), 2.80 (t, J = 6.8 Hz, 2H), 2.52 (dd, J = 7.6, 16.8 Hz, 1H), 2.32 (q, J = 7.2, 14.8 Hz, 2H), 2.10-1.98 (m, 2H), 1.87-1.71 (m, 2H), 1.70-1.38 (m, 7H), 1.34-1.11 (m, 6H). HR-MS (CI) calcd for $C_{36}H_{47}N_3O_{10}$ [M + H]⁺, 682.3261; found, 682.3271.

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Note Added in Proof. An approach to the synthesis of symmetrically substituted spiroketals using a polyethylene glycol-based ketal protecting group was recently reported. See: Haag, R.; Leach, A. G.; Ley, S. V.; Nettekoven, M.; Schnaubelt, J. *Synth. Commun.* **2001**, *31*, 2965–2977.

Supporting Information Available. ¹H NMR and mass spectral data for spiroketal compounds, X-ray structure coordinates of **33**, and X-ray crystallographic data for **33**. This material is available free of charge via the Internet at http://pubs.acs.org.

References and Notes

- (a) Dolle, R. E. Comprehensive survey of chemical libraries yielding enzyme inhibitors, receptor agonists and antagonists, and other biologically active agents: 1992 through 1997. *Mol. Diversity* 1998, 3, 199–233. (b) Dolle, R. E.; Nelson, K. H., Jr. Comprehensive Survey of Combinatorial Library Synthesis: 1998. *J. Comb. Chem.* 1999, 1, 235–282. (c) Dolle, R. E. Comprehensive Survey of Combinatorial Library Synthesis: 1999. *J. Comb. Chem.* 2000, 2, 383–433. (d) Dolle, R. E. Comprehensive survey of combinatorial libraries with undisclosed biological activity: 1992–1997. *Mol. Diversity* 2000, 4, 233–256.
- (2) Pryor, K. E.; Rebek, J., Jr. Multifunctionalized Glycolurils. Org. Lett. 1999, 1, 39–42.
- (3) Park, H. S.; Lin, Q.; Hamilton, A. D. Protein Surface Recognition by Synthetic Receptors: A Route to Novel Submicromolar Inhibitors for α-Chymotrypsin. J. Am. Chem. Soc. 1999, 121, 8–13.
- (4) Dolle, R. E.; Barden, M. C.; Brennan, P. E.; Ahmed, G.; Tran, V.; Ho, D. M. Application of the intramolecular azomethine imine cycloaddition to the construction of a novel, orthogonally protected spirodiamino acid scaffold. *Tetrahedron Lett.* 1999, 40, 2907–2908.
- (5) Pryor, K. E.; Shipps, W., Jr.; Skyler, D. A.; Rebek, J., Jr. The activated core approach to combinatorial chemistry: a selection of new core molecules. *Tetrahedron* 1998, 54, 4107–4124.
- (6) (a) Mink, D.; Mecozzi, S.; Rebek, J., Jr. Natural products analogs as scaffolds for supramolecular and combinatorial chemistry. *Tetrahedron Lett.* 1998, 39, 5709-5712. (b) Rasmussen, P. H.; Rebek, J., Jr. A pseudopeptide platform with side chains addressable for combinatorial applications. *Tetrahedron Lett.* 1999, 40, 3511-3514.
- (7) (a) Hirschmann, R.; Ducry, L.; Smith, A. B., III. Development of an Efficient, Regio- and Stereoselective Route to Libraries Based on the β-D-Glucose Scaffold. *J. Org. Chem.* 2000, 65, 8307–8316. (b) Ghosh, M.; Dulina, R. G.; Kakarla, R.; Sofia, M. J. Efficient Synthesis of a Stereochemically Defined Carbohydrate Scaffold: Carboxymethyl 2-Acetamido-6-azido-4-*O*-benzyl-2-deoxy-α-D-glucopyranoside. *J. Org. Chem.* 2000, 6, 8387–8390. (c) Moitessier, N.; Dufour, S.; Chretien, F.; Thiery, J. P.; Maigret, B.; Chapleur, Y. Design, synthesis and preliminary biological evaluation of a focused combinatorial library of stereo-diverse carbohydrate-scaffold-based peptidomimetics. *Bioorg. Med. Chem.* 2001, 9, 511–523.
- (8) Barry, J. F.; Davis, A. P.; Perez-Payan, M. N.; Elsegood, M. R. J.; Jackson, R. F. W.; Gennari, C.; Piarulli, U.; Gude, M. A Trifunctional Steroid-Based Scaffold for Combinatorial Chemistry. *Tetrahedron Lett.* 1999, 40, 2849–2852.
- (9) For representative examples, see: (a) Dragoli, D. R.; Thompson, L. A.; O'Brien, J.; Ellman, J. A. Parallel Synthesis of Prostaglandin E1 Analogues. *J. Comb. Chem.* 1999, *I*, 534–539. (b) Tan, D. S.; Foley, M. A.; Stockwell, B. R.; Shair, M. D.; Schreiber, S. L. Synthesis and Preliminary Evaluation of a Library of Polycyclic Small Molecules for Use in Chemical Genetic Assays. *J. Am. Chem.*

Soc. 1999, 121, 9073-9087. (c) Nicolaou, K. C.; Pfefferkorn, J. A.; Roecker, A. J.; Cao, G.-Q.; Barluenga, S.; Mitchell, H. J. Natural Product-like Combinatorial Libraries Based on Privileged Structures. 1. General Principles and Solid-Phase Synthesis of Benzopyrans. J. Am. Chem. Soc. 2000, 122, 9939-9953. (d) Lindsley, C. W.; Chan, L. K.; Goess, B. C.; Joseph, R.; Shair, M. D. Solid-Phase Biomimetic Synthesis of Carpanone-like Molecules. J. Am. Chem. Soc. 2000, 122, 422-423. (e) Wipf, P.; Reeves, J. T.; Balachandran, R.; Giuliano, K. A.; Hamel, E.; Day, B. W. Synthesis and Biological Evaluation of a Focused Mixture Library of Analogues of the Antimitotic Marine Natural Product Curacin A. J. Am. Chem. Soc. 2000, 122, 9391-9395. (f) Hijikuro, I.; Doi, T.; Takahashi, T. Parallel Synthesis of a Vitamin D₃ Library in the Solid-Phase. J. Am. Chem. Soc. 2001, 123, 3716-3722. (g) Pelish, H. E.; Westwood, N. J.; Feng, Y.; Kirchhausen, T.; Shair, M. D. Use of Biomimetic Diversity-Oriented Synthesis To Discover Galanthamine-like Molecules with Biological Properties beyond Those of the Natural Product. J. Am. Chem. Soc. 2001, 123, 6740-6741. (h) Recent reviews: Wessjohann, L. A. Synthesis of naturalproduct-based compound libraries. Curr. Opin. Chem. Biol. 2000, 4, 303-309. (i) Hall, D. G.; Manku, S.; Wang, F. Solution- and solid-phase strategies for the design, synthesis, and screening of libraries based on natural product templates: a comprehensive survey. J. Comb. Chem. 2001, 3, 125-150.

- (10) For reviews on the synthesis of spiroketals, see: (a) Kluge, A. F. Synthesis of 1,7-dioxaspiro[5.5]undecanes. Heterocycles 1986, 24, 1699-1740. (b) Perron, F.; Albizati, K. F. Chemistry of spiroketals. Chem. Rev. 1989, 89, 1617-1661. (c) Jacobs, M. F.; Kitching, W. Spiroacetals of marine origin. Curr. Org. Chem. 1998, 2, 395-436.
- (11) Birbeck, A. A.; Ley, S. V.; Prodger, J. C. Spiroketal Glycomimetics: The Synthesis of a Conformationally Restrained Sialyl Lewis X Mimic. Bioorg. Med. Chem. Lett. **1995**, *5*, 2637.
- (12) Bell, W.; Block, M. H.; Cook, C.; Grant, J. A.; Timms, D. Design, synthesis, and evaluation of a novel series of spiroketals based on the structure of the antibacterial gyrase inhibitor novobiocin. J. Chem. Soc., Perkin Trans. 1 1997, 18, 2789.
- (13) (a) Mitsuhashi, S.; Shima, H.; Kawamura, T.; Kikuchi, K.; Oikawa, M.; Ichihara, A.; Oikawa, H. The spiroketals containing a benzyloxymethyl moiety at C8 position showed the most potent apoptosis-inducing activity. Bioorg. Med. Chem. Lett. 1999, 9, 2007-2012. (b) Huang, H.; Mao, C.; Jan, S.-T.; Uckun, F. M. Stereocontrolled synthesis of a novel pharmacophore of the tubulin-depolymerizing marine natural product spongistatin. Tetrahedron Lett. 2000, 41, 1699-1702. (c) Uckun, F. M.; Mao, C.; Jan, S.-T.; Huang, H.; Vassilev, A. O.; Sudbeck, E. A.; Navara, C. S.; Narla, R. K. SPIKET and COBRA compounds as novel tubulin modulators with potent anticancer activity. Curr. Opin. Invest. Drugs (PharmaPress Ltd.) 2000, 1, 252-256. (d) Uckun, F. M.; Mao, C.; Vassilev, A. O.; Huang, H.; Jan, S.-T. Structurebased design of a novel synthetic spiroketal pyran as a pharmacophore for the marine natural product spongistatin 1. Bioorg. Med. Chem. Lett. 2000, 10, 541-545. (e) Uckun, F. M.; Sudbeck, E. A.; Mao, C.; Ghosh, S.; Liu, X.-P.; Vassilev, A. O.; Navara, C. S.; Narla, R. K. Structure-based design of novel anticancer agents. Curr. Cancer Drug Targets 2001, 1, 59–71. For efforts to produce simplified spiroketal analogues of the HIV protease inhibitor didemnaketal A, see: (f) Fan, X.; Flentke, G. R.; Rich, D. H. Inhibition of HIV-1 protease by a Subunit of Didemnaketal A. J. Am. Chem. Soc. 1998, 120, 8893-8894.
- (14) (a) Han, H.; Wolfe, M. M.; Brenner, S.; Janda, K. D. Liquidphase combinatorial synthesis. Proc. Natl. Acad. Sci. U.S.A. 1995, 92, 6419-6423. (b) Boger, D. L.; Tarby, C. M.; Myers, P. L.; Caporale, L. H. Generalized Dipeptidomimetic Tem-

- plate: Solution Phase Parallel Synthesis of Combinatorial Libraries. J. Am. Chem. Soc. 1996, 118, 2109-2110. (c) Cheng, S.; Comer, D. D.; Williams, J. P.; Myers, P. L.; Boger, D. L. Novel Solution Phase Strategy for the Synthesis of Chemical Libraries Containing Small Organic Molecules. J. Am. Chem. Soc. 1996, 118, 2567-2573. (d) For recent reviews on parallel solution-phase synthesis using polymer reagents and scavengers, see: Parlow, J. J.; Devraj, R. V.; South, M. S. Solution-phase chemical library synthesis using polymer-assisted purification techniques. Curr. Opin. Chem. *Biol.* **1999**. *3*. 320–336. (e) Flynn, D. L. Phase-trafficking reagents and phase-switching strategies for parallel synthesis. Med. Res. Rev. 1999, 19, 408-431 (f) Brummer, O.; Clapham, B.; Janda, K. D. Recent developments and applications of polymer-supported reagents in synthetic organic chemistry. Curr. Opin. Drug Discovery Dev. 2000, 3, 462-473.
- (15) Total synthesis: Evans, D. A.; Kaldor, S. W.; Jones, T. K.; Clardy, J.; Stout, T. J. Total synthesis of the macrolide antibiotic cytovaricin J. Am. Chem. Soc. 1990, 112, 7001-31.
- (16) Recent synthetic studies: (a) Evans, D. A.; Trotter, B. W.; Coleman, P. J.; Cote, B.; Dias, L. C.; Rajapakse, H. A.; Tyler, A. N. Enantioselective total synthesis of altohyrtin C (spongistatin 2). Tetrahedron 1999, 55, 8671-8726. (b) Crimmins, M. T.; Katz, J. D. Synthesis of the C16-C28 Spiroketal Subunit of Spongistatin 1 (Altohyrtin A): The Pyrone Approach. Org. Lett. 2000, 2, 957-960. (c) Zuev, D.; Paquette, L. A. A Modular Approach to Marine Macrolide Construction. 3. Enantioselective Synthesis of the C1-C28 Sector of Spongistatin 1 (Altohyrtin A). Org. Lett. 2000, 2, 679-682. (d) Wallace, G. A.; Scott, R. W.; Heathcock, C. H. Synthesis of the C29-C44 Portion of Spongistatin 1 (Altohyrtin A). J. Org. Chem. 2000, 65, 4145-4152. (e) Smith, A. B., III.; Lin, Q.; Doughty, V. A.; Zhuang, L.; McBriar, M. D.; Kerns, J. K.; Brook, C. S.; Murase, N.; Nakayama, K. The spongistatins: architecturally complex natural products. Part two. Synthesis of the C(29-51) subunit, fragment assembly, and final elaboration to (+)spongistatin 2. Angew. Chem., Int. Ed. 2001, 40, 196–199.
- (17) Synthetic studies: (a) Evans, D. A.; Ng, H. P.; Rieger, D. L. Total synthesis of the macrolide antibiotic rutamycin B. J. Am. Chem. Soc. 1993, 115, 11446-11459. (b) Jain, N. F.; Panek, J. S. Application of chiral (E)-crotylsilanes in synthesis: the asymmetric synthesis of the C19-C34 spiroketal fragment of rutamycin B. Tetrahedron Lett. 1997, 38, 1349-1352. (c) Synthesis of the spiroketal segment (C19-C34) of the rutamycins, antifungal metabolites of Streptomyces species. White, J. D.; Ohba, Y.; Porter, W. J.; Wang, S. *Tetrahedron Lett.* **1997**, *38*, 3167–3170. (d) Panek, J. S.; Jain, N. F. Total Synthesis of Rutamycin B and Oligomycin C. J. Org. Chem. 2001, 66, 2747-2756.
- (18) For example, see: Mayer, T. U.; Kapoor, T. M.; Haggarty, S. J.; King, R. W.; Schreiber, S. L.; Mitchison, T. J. Small molecule inhibitor of mitotic spindle bipolarity identified in a phenotype-based screen. Science 1999, 286, 971-974.
- (19) For a review on antagonists of protein-protein interactions, see: Cochran, A. G. Antagonists of protein-protein interactions. Chem. Biol. 2000, 7, R85-R94.
- (20) For representative syntheses of spiroketal ketones from methyl ketone aldol or acylation reactions, see refs 12, 13a, 14a, and: (a) Evans, D. A.; Gage, J. R.; Leighton, J. L. Total Synthesis of (+)-Calyculin A. J. Am. Chem. Soc. 1992, 114, 9434-9453. (b) Claffey, M. M.; Heathcock, C. H. A Method for Constructing the C2-C12 Dispiroacetal Moiety of Altohyrtin A. J. Org. Chem. 1996, 61, 7646-7647. (c) Paquette, L. A.; Zuev, D. A Modular Approach to Marine Macrolide Construction. 1. An Enantiocontrolled Route to

- the C1–C12 (AB) Spiroacetal Sector. *Tetrahedron Lett.* **1997**, *38*, 5115–5118. (d) Eppley, A. W.; Totah, N. I. A New Chiral Titanium Species for the Ring Opening Reactions of Meso Epoxides. *Tetrahedron* **1997**, *48*, 16545–16552.
- (21) (a) Tokunaga, M.; Larrow, J. F.; Kakiuchi, F.; Jacobsen, E. N. Asymmetric catalysis with water: efficient kinetic resolution of terminal epoxides by means of catalytic hydrolysis. *Science* **1997**, *277*, 936–938. (b) Hinterding, K.; Jacobsen, E. N. Regioselective Carbomethoxylation of Chiral Epoxides: A New Route to Enantiomerically Pure β-Hydroxy Esters. *J. Org. Chem.* **1999**, *64*, 2164–2165.
- (22) (a) Smith, A. B., III; Friestad, G. K.; Barbosa, J.; Bertounesque, E.; Hull, K. G.; Iwashima, M.; Qiu, Y.; Salvatore, B. A.; Spoors, P. G.; Duan, J. J.-W. Total Synthesis of (+)-Calyculin A and (-)-Calyculin B: Asymmetric Synthesis of the C(9–25) Spiroketal Dipropionate Subunit. *J. Am. Chem. Soc.* **1999**, *121*, 10468–10477. (b) For examples of epoxide ring opening with 2-methyl-1,3-dithiane, see: Schnurrenberger, P.; Hungerbuehler, E.; Seebach, D. Total synthesis of (+)-colletodiol from (*S*,*S*)-tartrate and (*R*)-3-hydroxybutanoate. *Liebigs Ann. Chem.* **1987**, 733–744. (c) Davey, A. E.; Parsons, A. F.; Taylor, R. J. K. Reactions of dithiane derived alcohols: promotion of the Thorpe—Ingold effect by trimethylsilyl and *tert*-butyl substituents. *J. Chem. Soc.*, *Perkin Trans. 1* **1989**, 1853–1858.
- (23) Tanemura, K.; Dohya, H.; Imamura, M.; Suzuki, T.; Horaguchi, T. Deprotection of 1,3-dithianes by 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ). *Chem. Lett.* **1994**, 965–968.
- (24) Compound 6 was treated with DDQ to effect dithiane cleavage to produce a β-hydroxy ketone which was compared with a known compound, see: Silverman, I. R.; Edington, C.; Elliott, J. D.; Johnson, W. S. Asymmetric synthesis via acetal templates. 13. Preparation of aldol compounds from butane-1,3-diol acetals. J. Org. Chem. 1987, 52, 180–183.
- (25) Kolb, H. C.; VanNieuwenhze, M. S.; Sharpless, K. B. Catalytic Asymmetric Dihydroxylation. *Chem. Rev.* 1994, 94, 2483–2547.
- (26) Oikawa, Y.; Yoshioka, T.; Yonemitsu, O. Specific removal of *O*-methoxybenzyl protection by DDQ oxidation. *Tetra-hedron Lett.* 1982, 23, 885–888.
- (27) Mancuso, A. J.; Swern, D. Activated dimethyl sulfoxide: useful reagents for synthesis. *Synthesis* **1981**, 165.
- (28) Mukaiyama, T. The directed aldol reaction. *Org. React.* (N.Y.) 1982, 28, 203–331.
- (29) Smith, A. B., III; Friestad, G. K.; Barbosa, J.; Bertounesque, E.; Hull, K. G.; Iwashima, M.; Qiu, Y.; Salvatore, B. A.; Spoors, P. G.; Duan, J. J.-W. Total Synthesis of (+)-Calyculin A and (-)-Calyculin B: Asymmetric Synthesis of the C(9-25) Spiroketal Dipropionate Subunit. *J. Am. Chem. Soc.* 1999, 121, 10468-10477.
- (30) For the synthesis of spiroketal oximes of milbemycin, see: Andrews, I. P.; Dorgan, R. J. J.; Harvey, T.; Hudner, J. F.; Hussain, N.; Lathbury, D. C.; Lewis, N. J.; Macaulay, G. S.; Morgan, D. O.; Stockman, R.; White, C. R. A Practical Synthesis of the Milbemycin SB-201561. *Tetrahedron Lett.* 1996, 37, 4811–4814.
- (31) For a review of chemistry using azolides, see: Staab, H. A. Syntheses with heterocyclic amides (azolides). *Angew. Chem., Int. Ed. Engl.* **1962**, *7*, 351–367.
- (32) For examples of resin capture and "catch and release", see:
 (a) Keating, T. A.; Armstrong, R. W. Postcondensation Modifications of Ugi Four-Component Condensation Products: 1-Isocyanocyclohexene as a Convertible Isocyanide. Mechanism of Conversion, Synthesis of Diverse Structures, and Demonstration of Resin Capture. J. Am. Chem. Soc. 1996, 118, 2574-2583. (b) Siegel, M. G.; Hahn, P. J.; Dressman, B. A.; Fritz, J. E.; Grunwell, J. R.; Kaldor, S. W. Rapid purification of small molecule libraries by ion exchange chromatography. Tetrahedron Lett. 1997, 38, 3357-3360. (c) Kulkarni, B. A.; Ganesan, A. Ion-exchange

- resins for combinatorial synthesis: 2,4-pyrrolidinediones by Dieckmann condensation. *Angew. Chem., Int. Ed. Engl.* **1997**, *36*, 2454–2455. (d) Liu, Y.; Zhao, C.; Bergbreiter, D. E.; Romo, D. Simultaneous Deprotection and Purification of BOC-Amines Based on Ionic Resin Capture. *J. Org. Chem.* **1998**, *63*, 3471–3473. (e) Hu, Y.; Baudart, S.; Porco, J. A., Jr. Parallel synthesis of 1,2,3-thiadiazoles employing a "catch and release" strategy. *J. Org. Chem.* **1999**, *64*, 1049–1051.
- (33) Commercially available from Argonaut Technologies (San Carlos, CA).
- (34) (a) Gemal, A. L.; Luche, J.-L. Lanthanoids in organic synthesis. 6. Reduction of α-enones by sodium borohydride in the presence of lanthanoid chlorides: synthetic and mechanistic aspects. J. Am. Chem. Soc. 1981, 103, 5454–5459. (b) For a recent reduction of a spiroketal ketone using this method, see: Taniguchi, T.; Ogasawara, K. A. Diastereocontrolled Synthesis of (+)-Febrifugine: A Potent Antimalarial Piperidine Alkaloid. Org. Lett. 2000, 2, 3193–3195.
- (35) Bal, B. S.; Childers, W. E., Jr.; Pinnick, H. W. Oxidation of α,β-unsaturated aldehydes. *Tetrahedron* 1981, 37, 2091– 2096.
- (36) Hu, Y.; Porco, J. A., Jr. Alcoholysis and carbonyl hydrosilylation reactions using a polymer-supported trialkylsilane. *Tetrahedron Lett.* **1998**, *39*, 2711–2714.
- (37) For examples of parallel workups employing SLE and liquid—liquid extraction cartridges, see: (a) Johnson, C. R.; Zhang, B.; Fantauzzi, P.; Hocker, M.; Yager, K. M. Libraries of N-alkylamino heterocycles from nucleophilic aromatic substitution with purification by solid supported liquid extraction. *Tetrahedron* 1998, 54, 4097—4106. (b) Breitenbucher, J. G.; Johnson, C. R.; Haight, M.; Phelan, J. C. Generation of a piperazine-2-carboxamide library: a practical application of the phenol—sulfide react and release linker. *Tetrahedron Lett.* 1998, 39, 1295—1298. (c) Johnson, C. R.; Zhang, B.; Fantauzzi, P.; Hocker, M.; Yager, K. Solution synthesis of heterocyclic libraries: nucleophilic aromatic substitution and automated solid supported liquid—liquid extraction. *Innovation Perspect. Solid Phase Synth. Comb. Libr., Collect. Pap., Int. Symp.* 1999, 209—210.
- (38) Coull, J. M.; Carlson, D. V.; Weith, H. L. Synthesis and characterization of a carbamate linked oligonucleoside. *Tetrahedron Lett.* 1987, 28, 745–748.
- (39) Crystals of **33** suitable for X-ray analysis were obtained by slow evaporation from hexanes/ethyl acetate. The structure was solved and refined (*R* factor = 6.4%). Crystallographic data have been deposited with the Cambridge Crystallographic Data Centre (CCDC-167385). Copies of the data can be obtained free of charge by contacting the CCDC, 12 Union Road, Cambridge CB21EZ, U.K. (fax (+44)-1223-336-033; e-mail deposit@ccdc.cam.ac.uk. See Supporting Information for further details on the X-ray structure and coordinates.
- (40) (a) Kaldor, S. W.; Siegel, M. G.; Fritz, J. E.; Dressman, B. A.; Hahn, P. J. Use of solid supported nucleophiles and electrophiles for the purification of non-peptide small molecule libraries. *Tetrahedron Lett.* 1996, 37, 7193–7196.
 (b) Kaldor, S. W.; Fritz, J. E.; Tang, J.; McKenney, E. R. Discovery of antirhinoviral leads by screening a combinatorial library of ureas prepared using covalent scavengers. *Bioorg. Med. Chem. Lett.* 1996, 6, 3041–3044. (c) Booth, J. R.; Hodges, J. C. Polymer-Supported Quenching Reagents for Parallel Purification. *J. Am. Chem. Soc.* 1997, 119, 4882–4886. (d) Booth, R. J. Hodges, J. C. Solid-Supported Reagent Strategies for Rapid Purification of Combinatorial Synthesis Products. *Acc. Chem. Res.* 1999, 32, 18–26.
- (41) For representative reaction conditions, see: Kurth, M. J.; Ahlberg Randall, L. A.; Takenouchi, K. Solid-Phase Combinatorial Synthesis of Polyisoxazolines: A Two-Reaction Iterative Protocol. J. Org. Chem. 1996, 61, 8755–8761.

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