

Supervised Learning with Conditional Inference Tree

Cardiotocograms, also known as CTGs, have been instrumental within clinical medicine for a long time. Obstetricians use these measurements and classifications to obtain detailed information and intelligence about newborns and their mother prior and during labor. In 2018, an article presented through the Journal of Clinical Medicine detailed the practicality of CTGs. The same article noted that interpretations of these sensorial readings is mainly attributed to the observer; which creates challenges of consistency of interpretations and defies the human naked-eye. Questions like what happens if /when the interpreter misses a key detail, or what could be the meaning of a combination of diagnostic signals, furthermore, what time-sensitive conditions may the measurements expose, requiring immediate actions are few examples of concerns posed by the continuous practice of merely optical assessments of a CTG (Zhao, Zhang & Deng, 2018).

The following exploration presents an assessment of CTGs using the conditional inference tree (ctree) model. The same shows how the algorithm expedites and enhances the interpretation of CTG readings while appraising multiple fetal readings simultaneously. Moreover, the study aims to identify potential hidden patterns which may require further attention.

The dataframe to be analyzed comes for the UCI Machine Learning Repository, and it consists of measurements of fetal heart rate (FHR) and uterine contraction (UC) as identified and recorded by cardiotocograms. The multivariate dataframe contains 2126 observations and 23 variables. Each diagnostic attribute within these CTGs were automatically processed and measured. Ultimately, all CTGs were classified by three subject matter experts, and under unanimity, assigned with response-labels based on the fetal state and/or morphologic detected patterns (Dua, & Graff, 2019).

The following list, obtained from the data dictionary, explains the meaning of the involved codes across these readings.

LB - FHR baseline (beats per minute)
AC - Number of accelerations per second
FM - Number of fetal movements per second
UC - Number of uterine contractions per second
DL - Number of light decelerations per second
DS - Number of severe decelerations per second
DP - Number of prolonged decelerations per second
ASTV - Percentage of time with abnormal short-term variability
MSTV - Mean value of short term variability
ALTV - Percentage of time with abnormal long-term variability
MLTV - Mean value of long -erm variability
Width - Width of FHR histogram
Min - Minimum of FHR histogram
Max - Maximum of FHR histogram
Nmax - Number of histogram peaks
Nzeros - Number of histogram zeros
Mode - Histogram mode
Mean - Histogram mean
Median - Histogram median
Variance - Histogram variance
Tendency - Histogram tendency
CLASS - FHR pattern class code (1 to 10)
NSP - Fetal state class code (N=normal; S = suspect; P=pathologic)

As observed, the above list includes unique CTG measurements, statistical attributes as well as observations from some of the recorded variables. The last two variables, CLASS and NSP, represent the previously mentioned classification and response-labeling conducted by the obstetricians.

Exploratory Analysis. The given cardiotocography.csv file was loaded and vectored as `ctg`. A look into the structure of the dataframe confirms some of the variables and information obtained from the repository and .csv file itself. The dataframe contains 2126 observations and 23

variables, formatted as integers and numeric values, as illustrated on figure 1.0. A glimpse over these represented values highlight few transformation options. Case in point, the targeted variable, which is the NSP , will need to get converted to a factor. Others like FM, DP, or ALTV may be representative of asymmetrical distributions. Furthermore, a variable like DS could have only one time of value, making it unusable for classification purposes.

```
> str(ctg)
'data.frame': 2126 obs. of 23 variables:
 $ LB      : int  120 132 133 134 132 134 134 122 122 122 ...
 $ AC      : num  0 0.006 0.003 0.003 0.007 0.001 0.001 0 0 0 .
 $ FM      : num  0 0 0 0 0 0 0 0 0 0 ...
 $ UC      : num  0 0.006 0.008 0.008 0.008 0.01 0.013 0 0.002 ...
 $ DL      : num  0 0.003 0.003 0.003 0 0.009 0.008 0 0 0 ...
 $ DS      : num  0 0 0 0 0 0 0 0 0 0 ...
 $ DP      : num  0 0 0 0 0 0.002 0.003 0 0 0 ...
 $ ASTV    : int  73 17 16 16 16 26 29 83 84 86 ...
 $ MSTV    : num  0.5 2.1 2.1 2.4 2.4 5.9 6.3 0.5 0.5 0.3 ...
 $ ALTV    : int  43 0 0 0 0 0 0 6 5 6 ...
 $ MLTV    : num  2.4 10.4 13.4 23 19.9 0 0 15.6 13.6 10.6 ...
 $ Width   : int  64 130 130 117 117 150 150 68 68 68 ...
 $ Min     : int  62 68 68 53 53 50 50 62 62 62 ...
 $ Max     : int  126 198 198 170 170 200 200 130 130 130 ...
 $ Nmax    : int  2 6 5 11 9 5 6 0 0 1 ...
 $ Nzeros  : int  0 1 1 0 0 3 3 0 0 0 ...
 $ Mode    : int  120 141 141 137 137 76 71 122 122 122 ...
 $ Mean    : int  137 136 135 134 136 107 107 122 122 122 ...
 $ Median  : int  121 140 138 137 138 107 106 123 123 123 ...
 $ Variance: int  73 12 13 13 11 170 215 3 3 1 ...
 $ Tendency: int  1 0 0 1 1 0 0 1 1 1 ...
 $ CLASS   : int  9 6 6 6 2 8 8 9 9 9 ...
 $ NSP     : int  2 1 1 1 1 3 3 3 3 3 ...
> |
```

Figure 1.0 – Possible irregular distributions across the variables.

Followed the dataframe structure assessment, the `summary(ctg)` command was used to expand and corroborates other aspects within the data.

```
> summary(ctg)
```

LB		AC		FM		UC		DL		DS	
Min.	:106.0	Min.	:0.000000	Min.	:0.000000	Min.	:0.000000	Min.	:0.000000	Min.	:0.000e+00
1st Qu.	:126.0	1st Qu.	:0.000000	1st Qu.	:0.000000	1st Qu.	:0.002000	1st Qu.	:0.000000	1st Qu.	:0.000e+00
Median	:133.0	Median	:0.002000	Median	:0.000000	Median	:0.004000	Median	:0.000000	Median	:0.000e+00
Mean	:133.3	Mean	:0.003178	Mean	:0.009481	Mean	:0.004366	Mean	:0.001889	Mean	:3.293e-06
3rd Qu.	:140.0	3rd Qu.	:0.006000	3rd Qu.	:0.003000	3rd Qu.	:0.007000	3rd Qu.	:0.003000	3rd Qu.	:0.000e+00
Max.	:160.0	Max.	:0.019000	Max.	:0.481000	Max.	:0.015000	Max.	:0.015000	Max.	:1.000e-03

DP		ASTV		MSTV		ALTV		MLTV		Width	
Min.	:0.0000000	Min.	:12.00	Min.	:0.200	Min.	:0.000	Min.	:0.000	Min.	:3.00
1st Qu.	:0.0000000	1st Qu.	:32.00	1st Qu.	:0.700	1st Qu.	:0.000	1st Qu.	:4.600	1st Qu.	:37.00
Median	:0.0000000	Median	:49.00	Median	:1.200	Median	:0.000	Median	:7.400	Median	:67.50
Mean	:0.0001585	Mean	:46.99	Mean	:1.333	Mean	:9.847	Mean	:8.188	Mean	:70.45
3rd Qu.	:0.0000000	3rd Qu.	:61.00	3rd Qu.	:1.700	3rd Qu.	:11.000	3rd Qu.	:10.800	3rd Qu.	:100.00
Max.	:0.0050000	Max.	:87.00	Max.	:7.000	Max.	:91.000	Max.	:50.700	Max.	:180.00

Min		Max		Nmax		Nzeros		Mode		Mean		Median	
Min.	:50.00	Min.	:122	Min.	:0.000	Min.	:0.0000	Min.	:60.0	Min.	:73.0	Min.	:77.0
1st Qu.	:67.00	1st Qu.	:152	1st Qu.	:2.000	1st Qu.	:0.0000	1st Qu.	:129.0	1st Qu.	:125.0	1st Qu.	:129.0
Median	:93.00	Median	:162	Median	:3.000	Median	:0.0000	Median	:139.0	Median	:136.0	Median	:139.0
Mean	:93.58	Mean	:164	Mean	:4.068	Mean	:0.3236	Mean	:137.5	Mean	:134.6	Mean	:138.1
3rd Qu.	:120.00	3rd Qu.	:174	3rd Qu.	:6.000	3rd Qu.	:0.0000	3rd Qu.	:148.0	3rd Qu.	:145.0	3rd Qu.	:148.0
Max.	:159.00	Max.	:238	Max.	:18.000	Max.	:10.0000	Max.	:187.0	Max.	:182.0	Max.	:186.0

Variance		Tendency		CLASS		NSP	
Min.	:0.00	Min.	: -1.0000	Min.	:1.00	Min.	:1.000
1st Qu.	:2.00	1st Qu.	:0.0000	1st Qu.	:2.00	1st Qu.	:1.000
Median	:7.00	Median	:0.0000	Median	:4.00	Median	:1.000
Mean	:18.81	Mean	:0.3203	Mean	:4.51	Mean	:1.304
3rd Qu.	:24.00	3rd Qu.	:1.0000	3rd Qu.	:7.00	3rd Qu.	:1.000
Max.	:269.00	Max.	:1.0000	Max.	:10.00	Max.	:3.000

```
> |
```

Figure 1.1 – NSP, as the response variable, will be converted to a factor

Case in point, judging by the numbers, Width, Min, Max, Nmax, Nzeros, Mode, Mean, Median, Variance, and Tendency appear to be statistical results of a measurement. Also, the variance across distributions is evident, thus, extra steps will have to be taken during the pre-processing phase. Lastly, no NA values were represented to this point, and using the `>colSums(is.na(ctg))` confirms there are no missing values.

```
> colSums(is.na(ctg))
```

LB	AC	FM	UC	DL	DS	DP	ASTV	MSTV	ALTV	MLTV	Width	Min
0	0	0	0	0	0	0	0	0	0	0	0	0
Max	Nmax	Nzeros	Mode	Mean	Median	Variance	Tendency	CLASS	NSP			
0	0	0	0	0	0	0	0	0	0			

```
> |
```

Figure 1.2 – No identified missing values across the dataset.

After reviewing the statistical attributes of the data set, a histogram was built to represent the LB variable. Per the data dictionary, this particular attribute is the most influential attribute of the set. The image in figure 1.3 shows how equally distributed the LB attribute is, and by the featured frequency, the mean will most likely fall between 130-135 beats per minute.

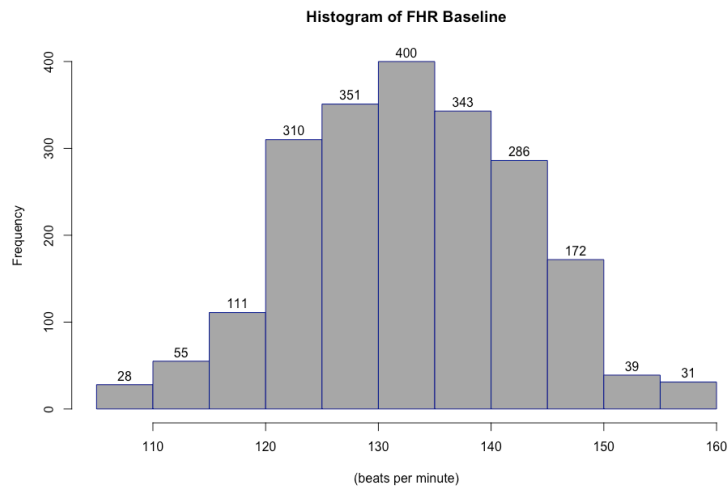


Figure 1.3 – Fetal heart rate baseline (LB variable) fairly distributed.

Further exploration of the LB variable (see figure 1.4) underscores some readings across the measurement particularly distanced from the mean. See the Appendix section for figure 1.4 code.

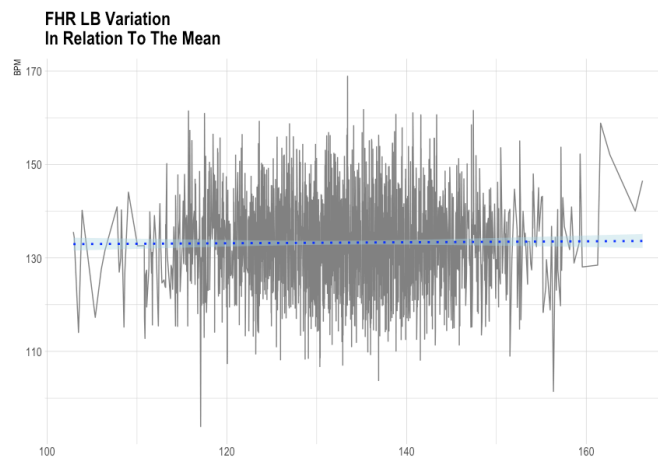


Figure 1.4 – Baseline variation across the dataset.

Motivated by the variability of the line graph, a deeper look was taken to identify out of the represented observations which ones were outside the 2-standard deviation (s.d.) ranges.

Figure 1.5, portrays what the developed code (see appendix A2) captured. It shows the values associated to 1 or 2 s.d.s' boundaries, and how some of the readings exceed such limits.

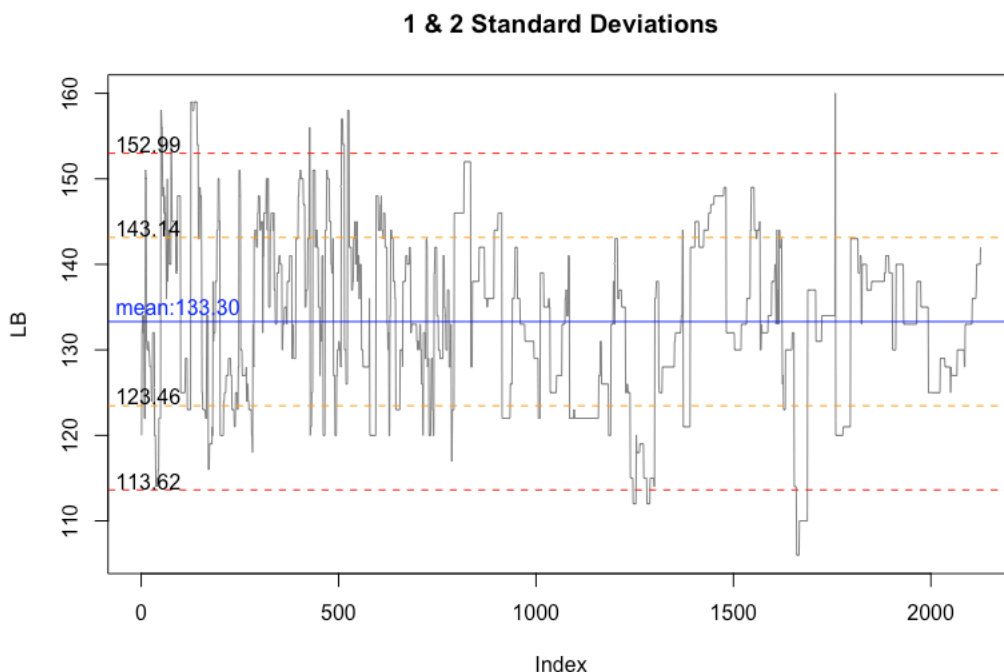


Figure 1.5 – Graphical display of LB's 1 & 2 standard deviations from the mean

Further calculation, like the one below, shows how critical this reading resulted, given that those readings within 2 s.d. account for over 96% of the studied observations. That said, this does not imply the contained readings are one way or the other (normal, suspect, or pathologic), but instead shows the usability of these quantified observations.

```
> # LB Observations higher than 2-s.d.
> lba<-(sum(ctg$LB >152.99)) #39
> # LB Observations lower than 2-s.d.
> lbb<-(sum(ctg$LB <113.62)) #44
> lba+lbb #=83 obs outside of 2-s.d.
[1] 83
> sum(between(ctg$LB, 113.62, 152.99))/nrow(ctg) # of obs within 2-s.d.
[1] 0.9609595
>
```

Figure 1.6 – Observations within 2 s.d. of the LB variable account for 96% of the dataset.

Considering the findings through the EDA process, and explanatory details from the data source, variables that were not part of the original measurements were excluded. Specifically, these variables were: width, min, max, nmax, nzeros, mode, mean, median, variance, and tendency. After further inquiry, these characteristics and calculations were related to the FHR baseline histogram made by the analytical software used by the dataset originators (Zhao, Zhang & Deng, 2018). Initially, the CLASS variable was converted to a factor and all of its 10 levels renamed. Unfortunately, this variable appears to be also a classification by the medical professionals, and contributed to overfitting of the model, thus forcing to reassess the modeling approach. That said, the CLASS variable was used as a guide for interpretation, but not included on the final set of variables to test within the model. Besides, the NSP variable was transformed to a factor, and its levels renamed to *normal*, *suspect*, and *pathologic*.

```
> # b. Exclude non-original measurements, rename targeted values
> ctg[12:22] <- NULL
> ctg$NSP<-as.numeric(ctg$NSP)
> ctg$NSP<-factor(ctg$NSP, levels= 1:3, labels = c("Normal", "Suspect", "Pathologic"))
> # ctg$CLASS<-as.numeric(ctg$CLASS)
> # ctg$CLASS<-factor(ctg$CLASS, levels=1:10, labels= c('A', 'B', 'C', 'D', 'SH', 'AD', 'DE', 'LD', 'FS', 'SP'))
```

Figure 1.7 – Transformation steps prior to creating a model.

After transforming the mentioned variables, a graphical representation of the target variable was created. Figure 1.8 displays a basic barplot and tabulation of the number of observations, including the NSP renamed values and their respective counts.

```
# Visualization of original NSP
plot(ctg$NSP, main="Original NSP Distribution",
     xlab="Fetal State Classification",
     ylab="Frequency", col=c(3, 7, 2))
text(ctg$NSP, labels=as.character(tabulate(ctg$NSP)), adj=3, pos=3)
```

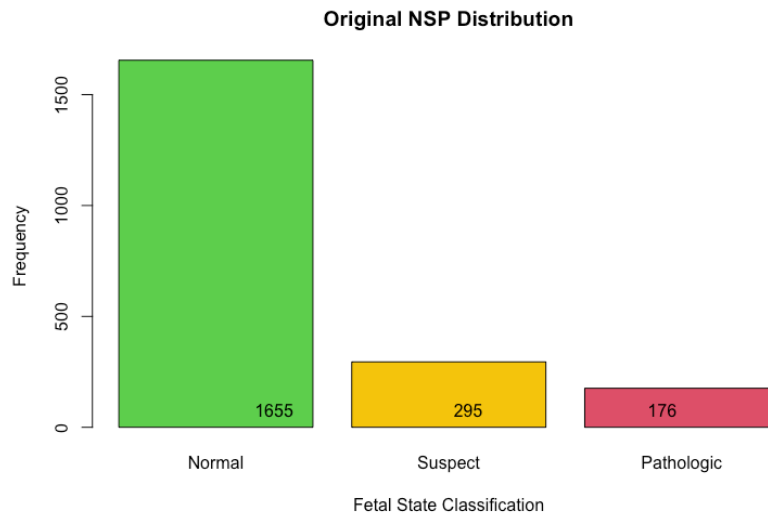


Figure 1.8 – Original NSP labeled observations.

At this point, the dataset was better organized and ready for the model portion. Bellow there is a representation of the final selected variables and their respective distributions.

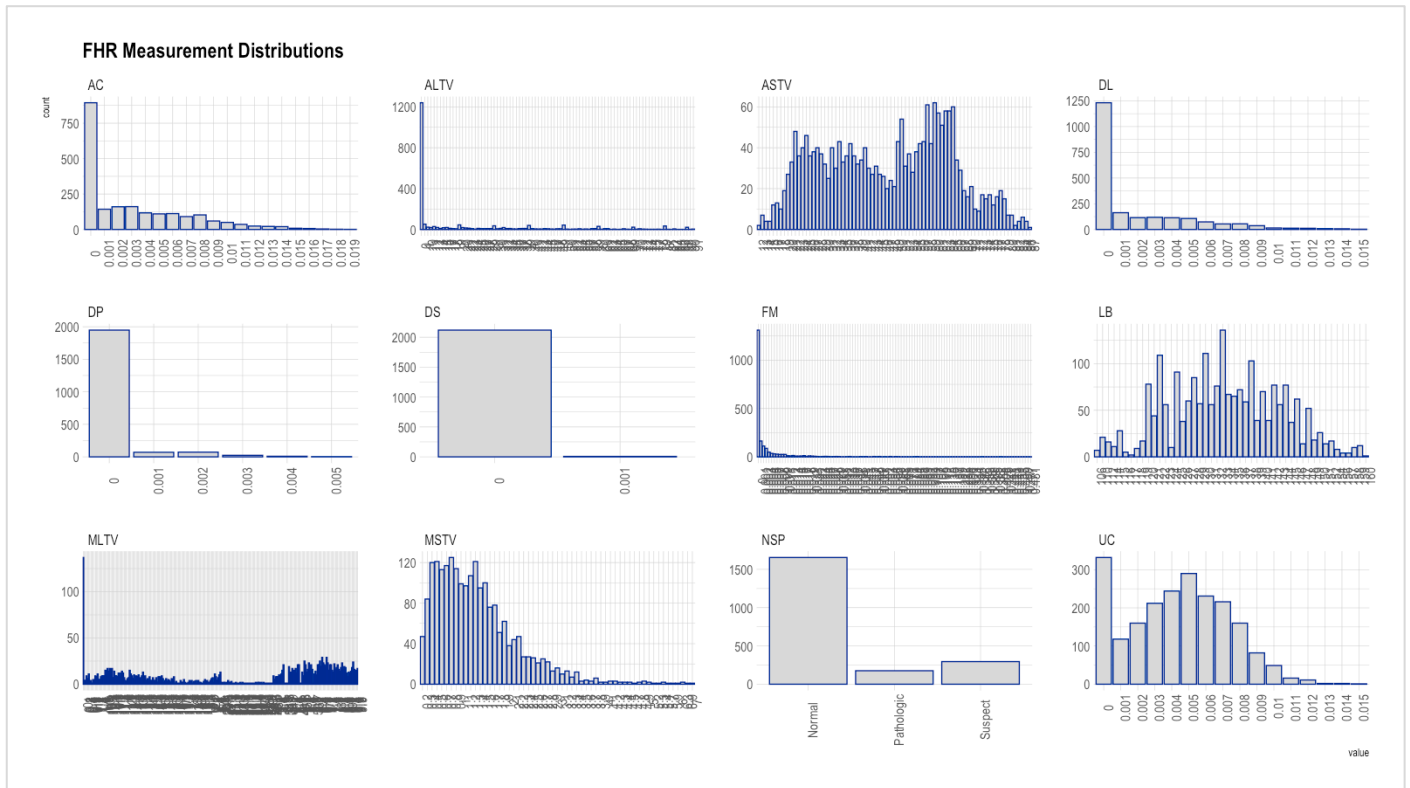


Figure 1.9 – Dataset transformed.

Algorithm Intuition: As previously stated, the conditional inference tree (ctree) is the algorithm to employ for this dataset classification. The objective is to use the model for identification of those independent variables with the greatest influence against the response variable. Author, Torsten Hothorn, summarizes the algorithm behind the ctree as follow, “A conditional inference trees estimate a regression relationship by binary recursive partitioning in a conditional inference framework (2006). Hothorn explains that first “the algorithm tests the sample for hypothesis of independence between the independent variables and the response”. If the hypothesis cannot be rejected the process stops. Conversely, it selects the variable with the strongest measured association (measured by the corresponding p-value). Secondly, it implements a binary decision and split in the selected variable, and recursively continues this process until all observations get evaluated (Hothorn, 2006).

The first step developing the model was setting a seed and splitting the dataset into different samples to train and test the model. After numerous evaluations, comparison of different simulations, and accuracy appraisals, the dataset splitting proportions were set to .70 and .30. Per figure 2.0, the probability weights for these subsets were set to 70%, and 30% for the training and testing subsets, respectively.

```
> #5. Split the data into a training and test set
> set.seed(1234)
> ind <- sample(2, nrow(ctg), replace = T, prob = c(0.70, 0.30))
> train.data <- ctg[ind == 1, ]
> test.data <- ctg[ind == 2, ]
```

Figure 2.0 – Subsets probability weights.

Appendix A4 portrays the structure of the developed training model. It consist of 25 inner nodes and 26 terminal nodes for a total of 51 nodes. The first split or branching to the

right occurs at the root where driven by DP “prolonged deceleration” > 0.001 , it groups 149 observations, with a majority of pathologic cases distributed through nodes 47, 48, & 49. To the left of the root, DP readings of < 0.001 decelerations are further split by ALTV “percentage of time with abnormal long term variability” of < 68 to the left, and > 68 to the right to node 44. The footprint of the tree is significantly large, illustrated on figure 2.1 Also, although the greater concentration of pathologic cases are located at the right side of the tree, additional pathologic cases are noticed on the left limbs, namely node 22, 21, and 15. As an observation, this is an example of irregular behaviors potentially hidden to the human sight.

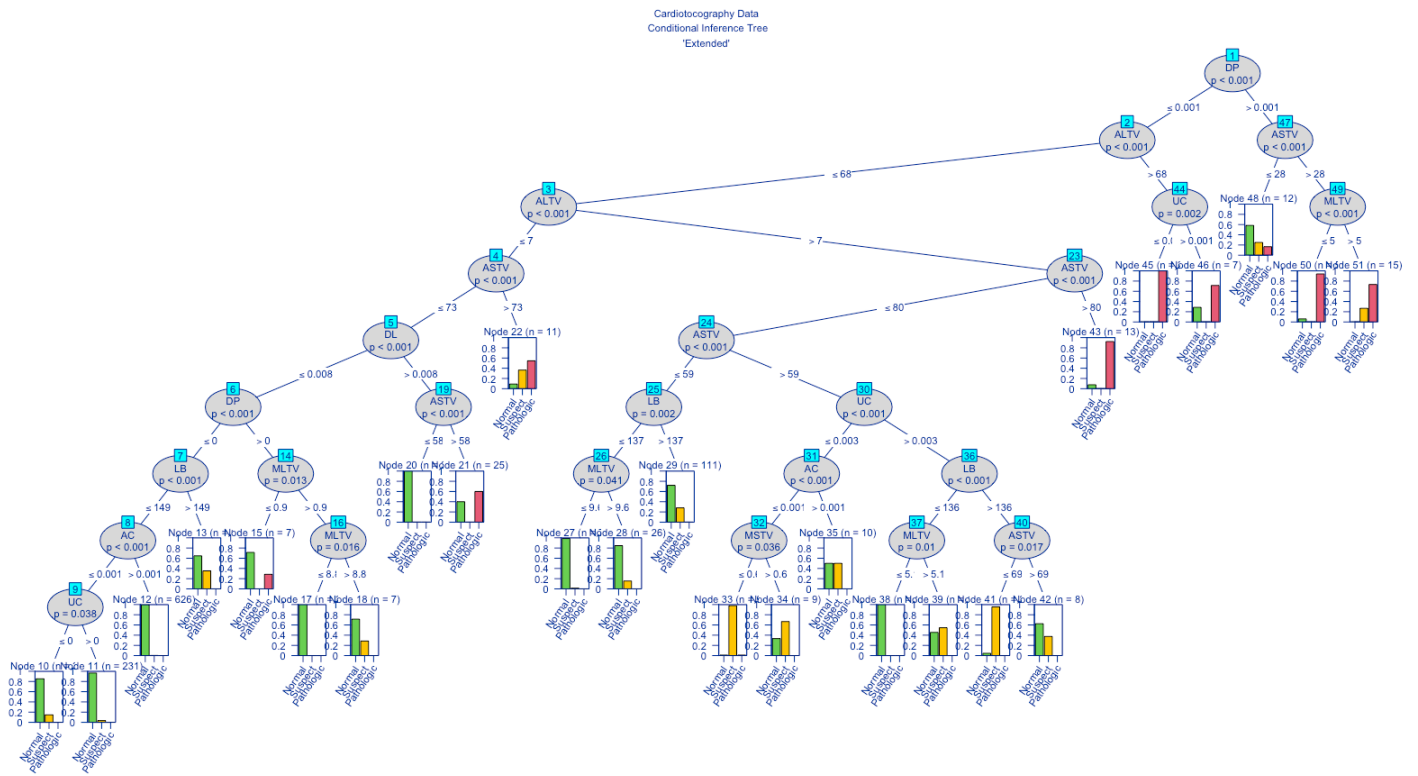


Figure 2.1 – Structure of training dataset model.

The following confusion matrix illustrates how the model performed against the training data. Out of 1523 total observations, the model properly categorized 1413 for a predicted classification accuracy of ~93%.

```
> table(predict(model), train.data$NSP, dnn=c("PREDICTED", "ACTUAL"))
```

	ACTUAL		
PREDICTED	Normal	Suspect	Pathologic
Normal	1168	70	4
Suspect	10	123	1
Pathologic	17	8	122

Figure 2.2 – Training model confusion matrix.

Output. Completed the training portion, the test sample was evaluated against the model. As mentioned before, this set contained the remaining 30% of the sampled population. The same branched out to 16 inner nodes and 17 leaf nodes, for a total of 33. Nodes 33, 32, 30, 28, 18, 17, and 15 were distinguished by containing observations identified as *pathologic*. Alternatively, node 27, 24, and 14 predominant classification was *suspect*. Refer to appendix A5 for a full depiction of this model in “simple” mode.

```
Model formula:
NSP ~ LB + AC + FM + UC + DL + DS + DP + ASTV + MSTV + ALTV +
      MLTV

Fitted party:
[1] root
| [2] DP <= 0.001
| | [3] ALTV <= 68
| | | [4] ALTV <= 15
| | | | [5] DP <= 0
| | | | | [6] DL <= 0.01
| | | | | | [7] ALTV <= 1: Normal (n = 291, err = 1.0%)
| | | | | | [8] ALTV > 1
| | | | | | [9] LB <= 143
| | | | | | | [10] ASTV <= 59: Normal (n = 78, err = 0.0%)
| | | | | | | [11] ASTV > 59: Normal (n = 14, err = 42.9%)
| | | | | | | [12] LB > 143
| | | | | | | [13] ASTV <= 45: Normal (n = 7, err = 14.3%)
| | | | | | | [14] ASTV > 45: Suspect (n = 13, err = 15.4%)
| | | | | | [15] DL > 0.01: Normal (n = 10, err = 30.0%)
| | | | | [16] DP > 0
| | | | | [17] MSTV <= 1.7: Normal (n = 12, err = 25.0%)
| | | | | [18] MSTV > 1.7: Normal (n = 12, err = 50.0%)
| | | | [19] ALTV > 15
| | | | [20] ASTV <= 59
| | | | [21] LB <= 147
| | | | | [22] FM <= 0: Normal (n = 30, err = 3.3%)
| | | | | [23] FM > 0: Normal (n = 7, err = 28.6%)
| | | | | [24] LB > 147: Suspect (n = 13, err = 46.2%)
| | | | [25] ASTV > 59
| | | | [26] MSTV <= 0.5
| | | | | [27] ASTV <= 77: Suspect (n = 51, err = 2.0%)
| | | | | [28] ASTV > 77: Suspect (n = 9, err = 44.4%)
| | | | | [29] MSTV > 0.5: Normal (n = 11, err = 27.3%)
| | | [30] ALTV > 68: Pathologic (n = 14, err = 14.3%)
| | [31] DP > 0.001
| | | [32] ASTV <= 57: Normal (n = 10, err = 50.0%)
| | | [33] ASTV > 57: Pathologic (n = 21, err = 0.0%)

Number of inner nodes: 16
Number of terminal nodes: 17
```

Figure 2.4 – Structure of test data model.

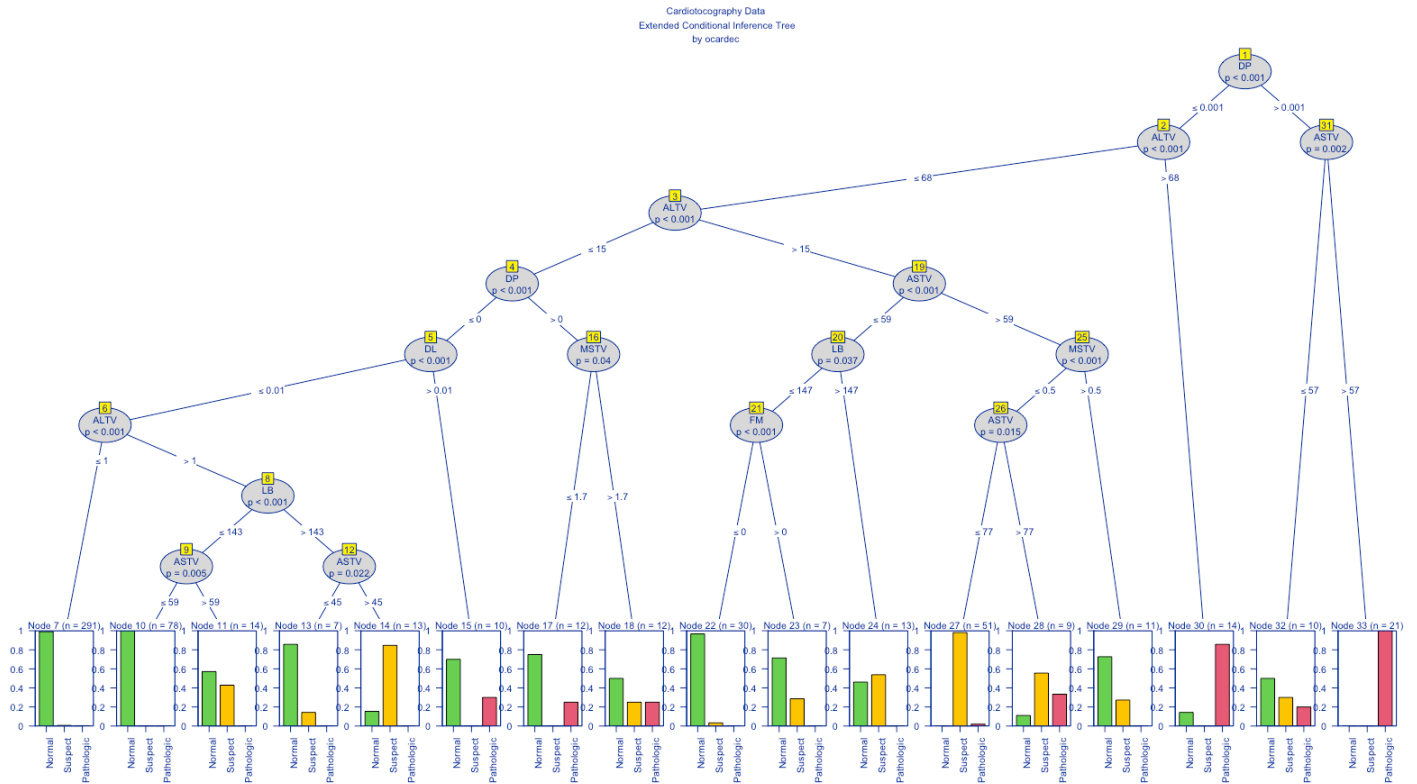


Figure 2.5 – Test data conditional inference tree.

Per the above graphic (figure 2.5), variables prolonged decelerations (DP), percentage of time with abnormal long term variability (ALTV), and percentage of time with abnormal short term variability (ASTV) were key splitting nodes affecting all other child-nodes. The data does not provide the causes of these irregularities, but it does highly the important of such percentages as triggers to constantly observe. Importantly, node 5 portrays a split based on FHR baseline changes (DL). It segregates readings based on those bellow or above the 1% variation. Those greater than or equal to 0.01 branch out to terminal node 15, including *pathologic* readings. On the other side, those less than or equal to 0.01 branch to the left. This node 5 splitting point, captures 403 out of the 637 sample for a total of 64.5% (including node 15) based on the DL reading, highlighting the previously mentioned assumption on the importance of those DL readings within 2 standard deviations.

The confusion matrix for the test.data model captures a total of 555 correct predictions for a classification accuracy rate of 92% with confidence intervals between 89.5% and 94%.

```
> # Confusion matrix and stats
> testPred2 <- predict(model2, newdata = test.data, method="NSP")
> confusionMatrix(testPred2, test.data$NSP)
Confusion Matrix and Statistics
```

	Reference		
Prediction	Normal	Suspect	Pathologic
Normal	449	21	12
Suspect	9	73	4
Pathologic	2	0	33

```
Overall Statistics

      Accuracy : 0.9204
      95% CI : (0.8958, 0.9407)
    No Information Rate : 0.7629
    P-Value [Acc > NIR] : < 2.2e-16

      Kappa : 0.7809

    McNemar's Test P-Value : 0.001165

Statistics by Class:
```

	Class: Normal	Class: Suspect	Class: Pathologic
Sensitivity	0.9761	0.7766	0.67347
Specificity	0.7692	0.9745	0.99639
Pos Pred Value	0.9315	0.8488	0.94286
Neg Pred Value	0.9091	0.9594	0.97183
Prevalence	0.7629	0.1559	0.08126
Detection Rate	0.7446	0.1211	0.05473
Detection Prevalence	0.7993	0.1426	0.05804
Balanced Accuracy	0.8727	0.8755	0.83493

Figure 2.6 – Test data sample confusion matrix.

Conclusion. Based on the above model’s descriptions, and inferred findings, one may argue and advocate for the use of supervised learning algorithms, like the ctree. This can definitely serve as supporting mechanisms through the medical industry, especially, as evaluated here neonatal, or labor and delivery medical departments. Per the described intended goals, this study employed the ctree methodology to assess and classified independent variables and their influence toward the response variable of NSP. As previously alluded, the model enhances

uncovering inclinations, branches, or tree nodes not typically perceivable by the human naked-eye, thus discourages the practice of CTG interpretation solely based on the medical practitioner experience.

Although this is a basic assessment on the usability of the ctree algorithm, supplemented with other data mining techniques including, but not limited to, clustering, classification and dimensionality reduction, could boost the performance accuracy of the model while reducing error variances. Some encountered challenges while conducting the study were related to identifying best transformation techniques or interpretation of the data, given the limited background of the medial domain. As a recommendation, any future development of this algorithm in relation to CTGs would highly benefit from having sustainable knowledge of this medial area. For best accuracy outcomes, better transformation techniques and variable selection process is essential.

References

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<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6111566/>

Appendix A

```
# Stats of LB
t.test(ctg$LB)
m<-mean(ctg$LB)
std<-sd(ctg$LB)
upr=m+std
lwr=m-std

lbdf <- data.frame(ctg,my_x = 0 + rnorm(length(ctg$LB), mean=m,
sd=std),my_y = 0 + rnorm(length(ctg$LB), mean=m, sd=std))
# Show LB Variation
print(pltlb<-ggplot(lbdf, xlab=FALSE, aes(x=(my_x), y=my_y))+
geom_line(col="grey51",linemitre=1)+
geom_smooth(method=lm,color="blue",lty=3,fill="lightblue",se=T)+
labs(x=NULL,y="BPM",title="FHR LB Variation\nIn Relation To The Mean")+
theme_ipsum())
```

Appendix A1. Figure 1.4 Code.

```
# What is 2-standard deviations from the mean?
upr2=m+(std*2)
lwr2=m-(std*2)
# Plot LB distribution boundaries
plot.new()
plot(ctg$LB, type="l", col="grey51", ylab="LB", main="1 & 2 Standard
Distributions")
abline(h = m, col = "blue")
abline(h = upr, col = "orange", lty=2)
abline(h = lwr, col = "orange", lty=2)
abline(h = upr2, col = "red", lty=2)
abline(h = lwr2, col = "red", lty=2)
text(-65,134, "mean:133.30", col = "blue", adj = c(0, -.1))
text(-65,upr, round(upr, 2), col = "black", adj = c(0, -.1))
text(-65,lwr, round(lwr, 2), col = "black", adj = c(0, -.1))
text(-65,upr2, round(upr2, 2), col = "black", adj = c(0, -.1))
text(-65,lwr2, round(lwr2, 2), col = "black", adj = c(0, -.1))
```

Appendix A2. Figure 1.5 Code.


```

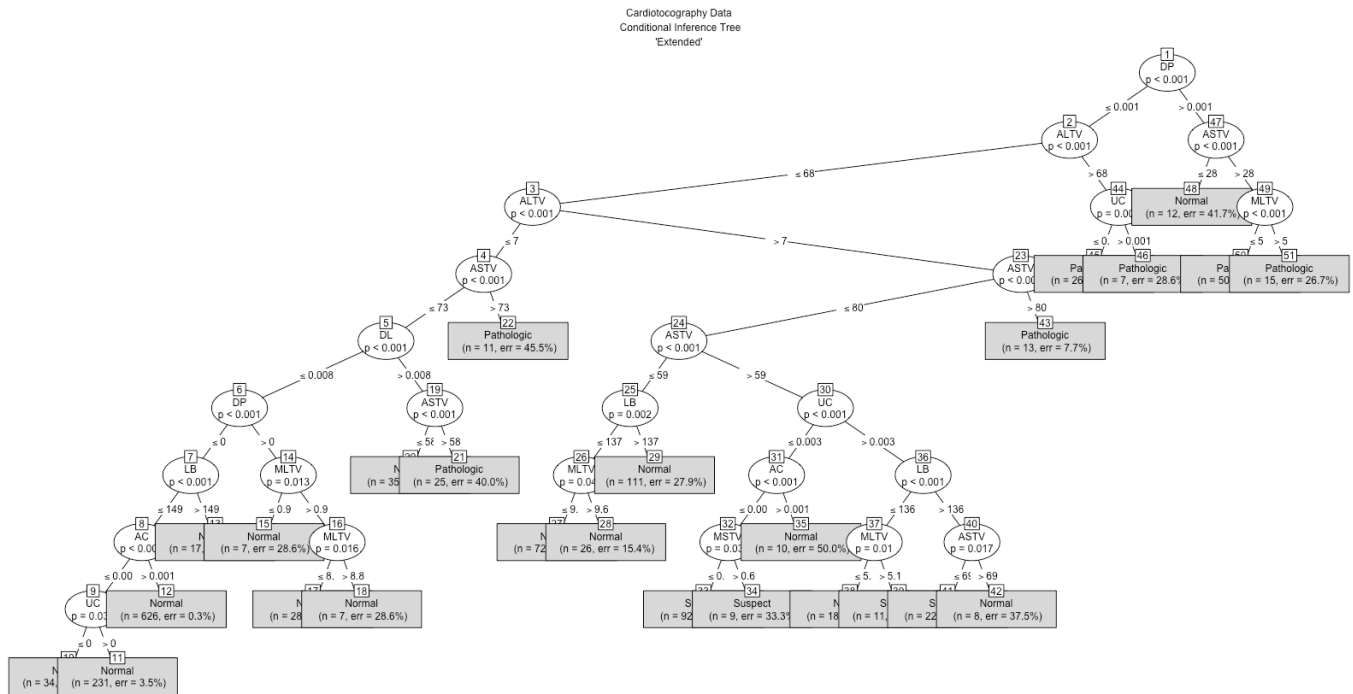
| | | | | [12] DP > 0: Normal (n = 30, err = 13.3%)
| | | | | [13] LB > 143
| | | | | [14] AC <= 0.003: Normal (n = 56, err = 25.0%)
| | | | | [15] AC > 0.003: Normal (n = 54, err = 0.0%)
| | | | | [16] ASTV > 58
| | | | | [17] ALTV <= 6
| | | | | [18] DL <= 0.008
| | | | | [19] ASTV <= 76
| | | | | [20] DP <= 0
| | | | | [21] UC <= 0: Normal (n = 8, err = 37.5%)
| | | | | [22] UC > 0
| | | | | [23] MLTV <= 7.7: Normal (n = 90, err = 0.0%)
| | | | | [24] MLTV > 7.7: Normal (n = 49, err = 14.3%)
| | | | | [25] DP > 0: Normal (n = 12, err = 16.7%)
| | | | | [26] ASTV > 76: Pathologic (n = 7, err = 42.9%)
| | | | | [27] DL > 0.008: Pathologic (n = 27, err = 44.4%)
| | | | | [28] ALTV > 6
| | | | | [29] ASTV <= 69: Suspect (n = 27, err = 22.2%)
| | | | | [30] ASTV > 69: Normal (n = 11, err = 9.1%)
| | | | | [31] ALTV > 15
| | | | | [32] ASTV <= 79
| | | | | [33] ASTV <= 59
| | | | | [34] MLTV <= 9.3
| | | | | [35] FM <= 0.003: Normal (n = 97, err = 10.3%)
| | | | | [36] FM > 0.003: Normal (n = 7, err = 42.9%)
| | | | | [37] MLTV > 9.3: Suspect (n = 19, err = 42.1%)
| | | | | [38] ASTV > 59
| | | | | [39] MSTV <= 0.6
| | | | | [40] UC <= 0.006: Suspect (n = 114, err = 4.4%)
| | | | | [41] UC > 0.006: Suspect (n = 10, err = 40.0%)
| | | | | [42] MSTV > 0.6
| | | | | [43] LB <= 139: Normal (n = 15, err = 13.3%)
| | | | | [44] LB > 139: Suspect (n = 7, err = 14.3%)
| | | | | [45] ASTV > 79: Pathologic (n = 7, err = 14.3%)
| | | | | [46] ALTV > 68
| | | | | [47] UC <= 0.001: Pathologic (n = 20, err = 0.0%)
| | | | | [48] UC > 0.001: Pathologic (n = 7, err = 28.6%)
| | | | | [49] DP > 0.001
| | | | | [50] ASTV <= 24: Normal (n = 11, err = 36.4%)
| | | | | [51] ASTV > 24: Pathologic (n = 59, err = 8.5%)

```

Number of inner nodes: 25

Number of terminal nodes: 26

Appendix A4. Training set model formula and fitted structure.



Appendix A5. Conditional Inference Tree, training dataset, "simple" mode.

```
> #8. visualize the tree
> plot(model, main="Cardiotocography Data\n Conditional Inference Tree\n'Extended'",
+       type="simple", ep_args = list(justmin = 8), drop_terminal = F,
+       gp = gpar(fontsize = 9), margins = c(4,4, 4, 4))
> plot(model, type="extended", ep_args = list(justmin = 8), drop_terminal=F, tnex=1.5,
+       gp=gpar(fontsize = 8, col="dark blue"),
+       inner_panel = node_inner(model, fill=c("light grey","cyan"), pval=T),
+       terminal_panel=node_barplot(model, fill=c(3,7,2), beside=T, ymax=1, rot = 75,
+       just = c(.95,.5), ylines=T, widths = 2, gap=0.05, reverse=F, id=T),
+       margins = c(5,3, 4, 3),
+       main = "Cardiotocography Data\n Conditional Inference Tree\n'Extended'")
```

Appendix A6. Figure 2.1 code.