# Simulation framework for chromatin architecture post UV-C irradiation

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## 1 The simulation framework

To representing the dynamics of the chromatin in the UV damage microenvironment, simulations are performed in a three-dimensional spherical domain with reflecting boundaries. Our simulation describe the dynamics of DNA and nucleosome signal loss from a circular region of interest (ROI) 15 minutes post UV-C irradiation. The experimental measurements are reported as the maximal z-projected values of GFP signals. Accordingly, simulation result will be obtained by projecting the three-dimensional polymer dynamics onto a two dimensional plane parallel to the x-y plane and passing through the center of the spherical domain.

## 1.1 The polymer model

We use a cross-linked Rouse chain of N monomers connected by harmonic springs [1] to represent a coarse-grained model of the chromatin. Springs connecting adjacent monomers in the linear chain are assigned a spring constant  $2k_BT/b^2$  and a minimal length  $L_0 > 0$ . Cross-links are represented by connecting non nearest-neighbor (NN) monomers by a harmonic springs with a spring constant  $2k_BT/b^2$  and zero minimal length. The measure of cross-linking is given by the percentage  $0 \le \alpha \le 100$  of non-NN monomers connected out of the total. For each realization of the polymer, cross-links are added between pairs of monomers chosen uniformly at random. The dynamics of the cross-linked polymer is then simulated up to its relaxation time, determined by be the slowest mode of the linear chain [1].

#### 1.2 UV irradiation

At the end of relaxation steps an instantaneous UV beam is shot with its focal point placed at the polymer's center-of-mass (CM). Small fluctuations of monomers are not of interest at the scale of our simulation, therefore the diffusion force acting on the polymer is stopped at this point. Damages caused by UV are restricted to be contained within a fixed two-dimensional circular region of area  $A_0$  centered at laser's focal point. Damages to DNA are represented by monomers labeled as damaged. For each UV dose u, damages caused by UV are uniformly distributed between the monomers in  $A_0$  at the time of beam shot. With increase of UV dose, we increase the probability of damages linearly as  $k_t u$ , with  $k_t$  in units of bp/msec.

#### 1.2.1 The repair stage

To simulate the affect of repair proteins crowding at sites of DNA damage, a circular exclusion region is centered at each damaged monomer. An elastic pushing force originating from the damaged monomer and having affect radius  $r_p$  is applied on any monomers entering the exclusion region. In addition, all cross-links from and to damaged monomers are removed to simulate chromatin de-compaction caused by the activity of chromatin remodlers and repair proteins [REF].

The polymer will evolve into a new steady spatial configuration which represents the chromatin 15 minutes post UV-C. At which point, the region of interest (ROI) is defined as the circle containing 90% of the damaged monomers and centered at their CM. The ROI remains a fixed region used to track the number of damaged and undamaged monomers within it. Measurements are done off-line, such that the ROI's position is retrospectively updated to be centered at the CM of the monomers tagged damaged.

## 1.3 Post repair stage

As damaged monomers are repaired, the exclusion region is removed from damaged monomers. Cross-links are reintroduced by linking any two monomers located within an encounter distance of  $\epsilon$  from one another. Cross-links post repair are allowed to be added up to initial cross-linking percentage  $\alpha$  is not exceeded.

## 1.4 Measuring structural similarity

A measure of similarity between the polymer spatial organization before UV and post repair is based on the comparison of their encounter probability matrices.

### 1.5 Parameter Values

## References

[1] Masao Doi and Sam F Edwards. *The theory of polymer dynamics*, volume 73. oxford university press, 1988.