

testing

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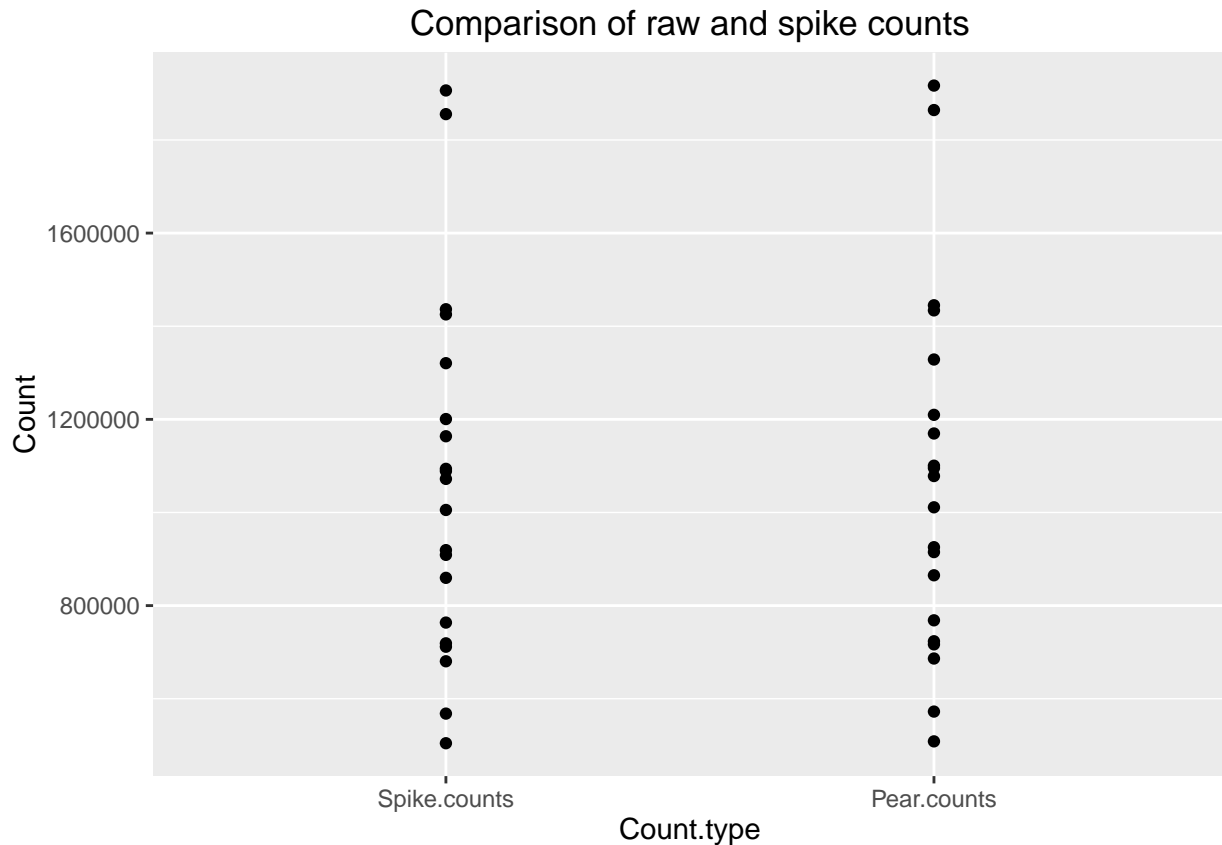
Overview

When testing primer independence using samples 1-20 from DNA160609LC, Burcu noticed a four-fold range in spike counts between samples. For example, the spike corresponding to V1/J1-1 in sample 1 could have a count of 10, whereas the same spike has a count of 40 in sample 2. We usually give the sequencing core unequal amounts of DNA in our samples, but these 20 samples have the same initial concentration of spikes. We would expect them to then have the same final concentration as well, assuming that they amplified similarly.

We can compare the 9-bp spike count totals with the PEAR fastq read counts and determine if the distributions are similar.

Compare distributions

##	Sample	Spike.counts	Pear.counts
## 1	1	680413	686374
## 2	2	1200647	1209673
## 3	3	859639	865117
## 4	4	1088957	1095477
## 5	5	1436461	1445022
## 6	6	1093733	1100437
## 7	7	1425387	1433990
## 8	8	1855464	1864230
## 9	9	1320599	1328501
## 10	10	1072110	1078247
## 11	11	1906327	1916805
## 12	12	1005399	1011097
## 13	13	763504	768413
## 14	14	504436	508421
## 15	15	919145	925440
## 16	16	711713	716742
## 17	17	568063	572435
## 18	18	908939	914924
## 19	19	718989	723413
## 20	20	1163698	1169669



We can see that the distributions are almost identical. These results suggest that the variation came from any of the following (or a combination of them):

1. Sample preparation - variable amounts of spikes were pipetted into the samples
2. PCR amplification - samples have same starting material, but individual reactions amplified differently in the thermocycler
3. Sequencing - Samples seeded unevenly on the sequencing machine

Next Steps

Do we have quantitative data of sample concentration prior to PCR (nanodrop or cubit?), if so, we should check that distribution. We can also ask Bob if he saw a difference in seeds.