
Introduction to structural equation modeling and mixed models in

Day 7

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- Categorical Variables in SEM
-

Categorical Variables in SEM

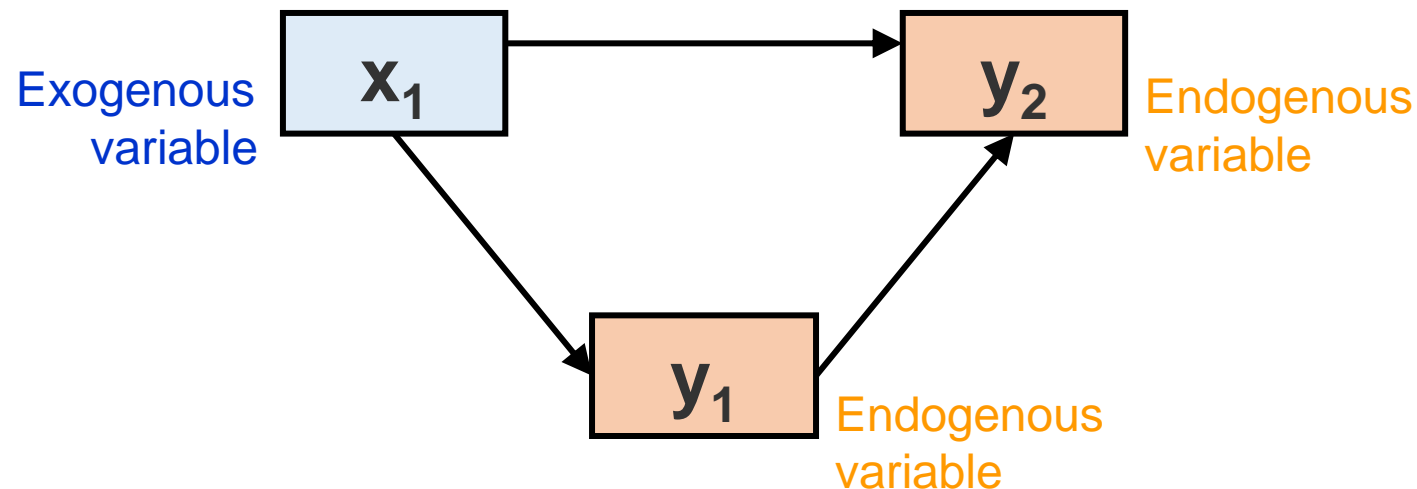
Categorical / discrete data

- binary (yes/no, failure/success, dead/alive, male/female),
- nominal (site 1, site 2, site 3)
- ordinal levels (small < medium < large; young < middle < old).

Categorical Variables in SEM

Categorical / discrete data

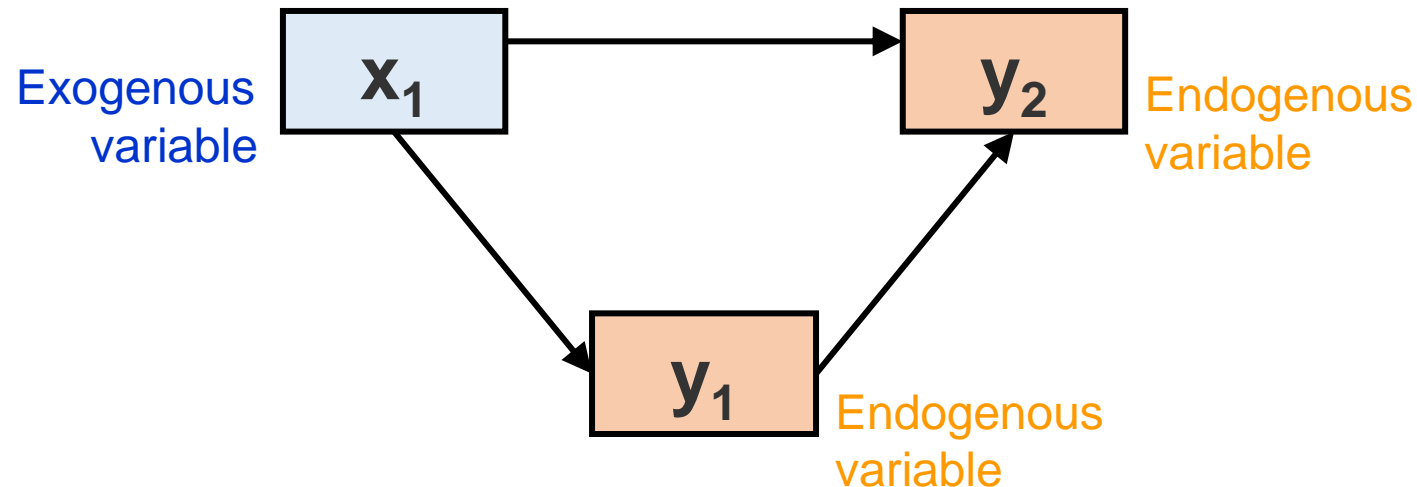
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Categorical Variables in SEM

Categorical / discrete data

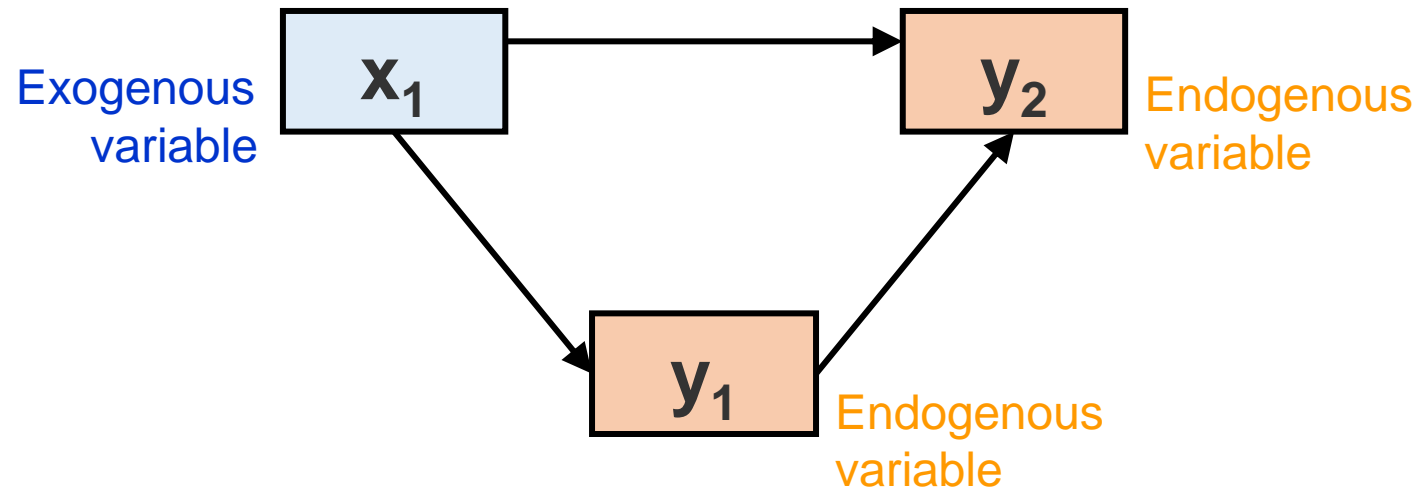
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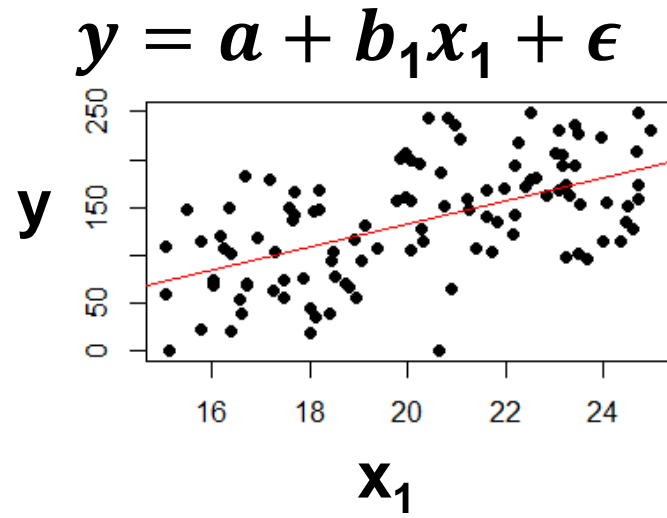
Categorical Variables in SEM

Categorical / discrete data

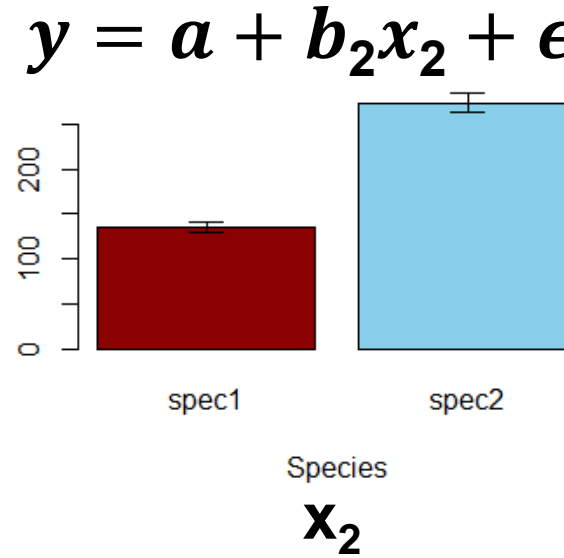
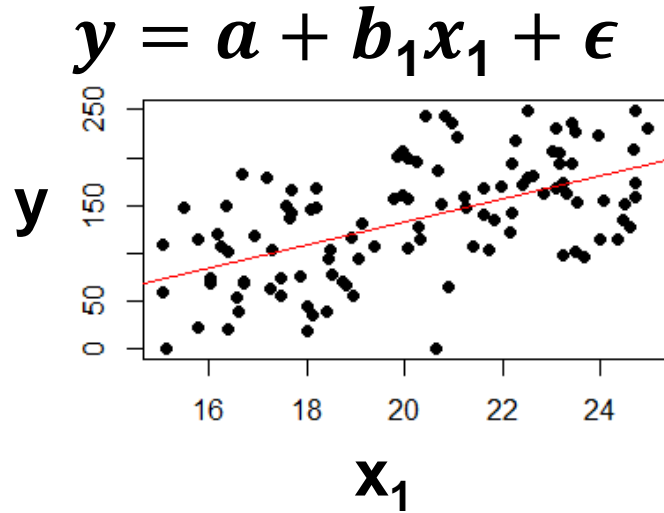
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Categorical Variables in SEM

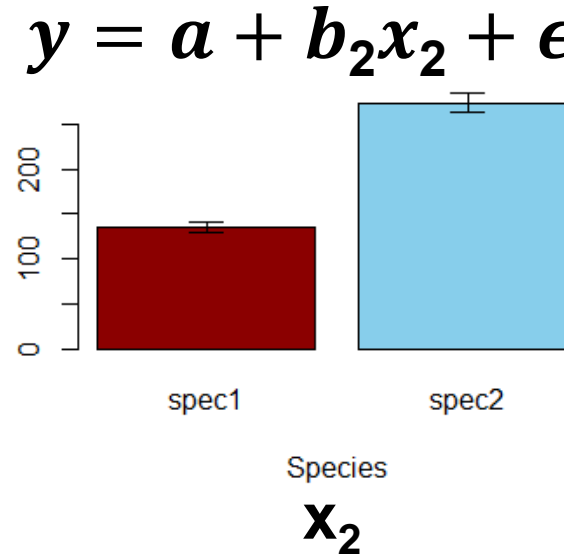
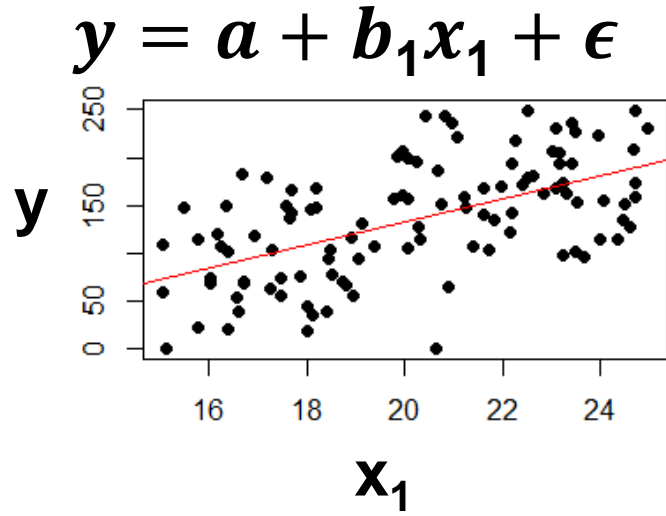


Categorical Variables in SEM



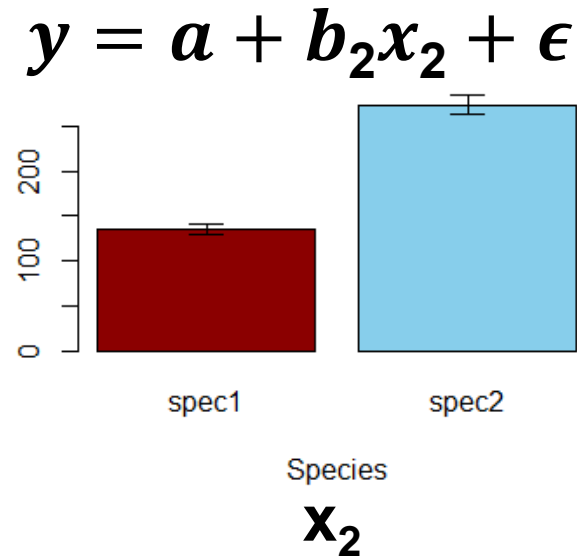
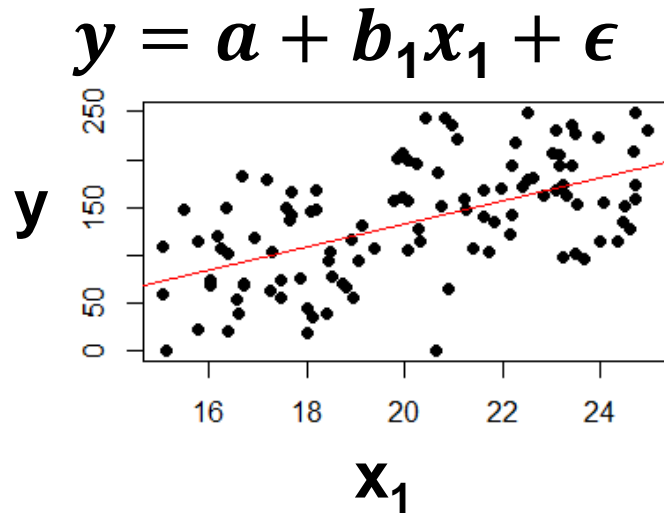
x_2	
Species	
spec1	
spec1	
spec2	
spec1	
spec2	

Categorical Variables in SEM



x_2	
Species	
spec1	
spec1	
spec2	
spec1	
spec2	

Categorical Variables in SEM



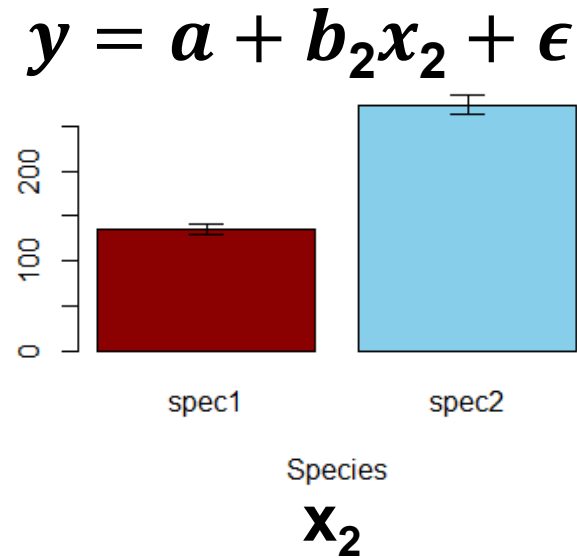
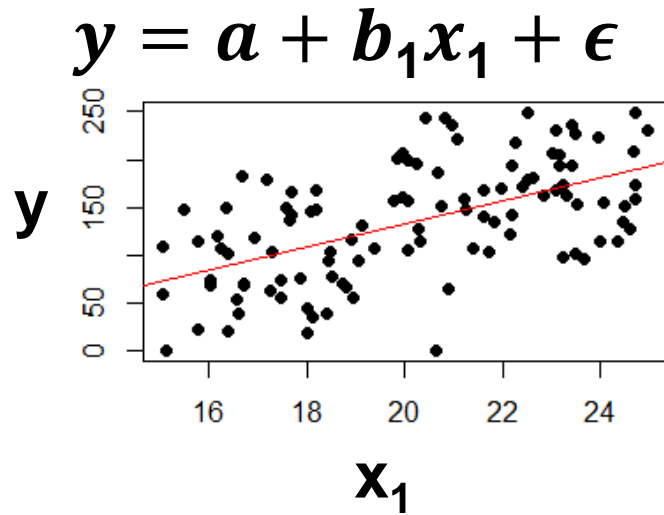
x_2

Species
spec1
spec1
spec2
spec1
spec2



spec1	spec2
1	0
1	0
0	1
1	0
0	1

Categorical Variables in SEM



x_2

Species
spec1
spec1
spec2
spec1
spec2



spec1	spec2
1	0
1	0
0	1
1	0
0	1

Exogenous Categorical Variables

Approaches when we have Exogenous Categorical Variables:

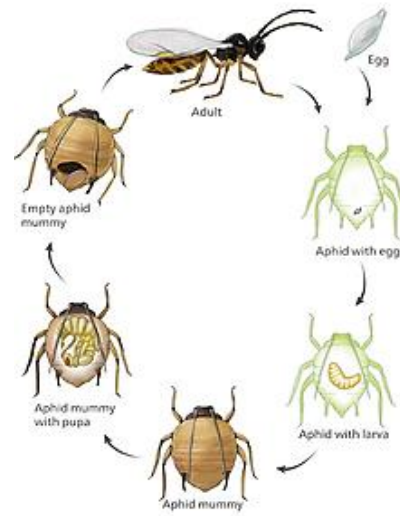
- 1) for nominal, binary, or ordinal variables, create separate dummy variables for each factor levels (treat them as absent “0” or present “1”).
 - The key: for the factor with k levels use $k-1$ dummy variables (to avoid singularity)
- 2) for binary variables, set the values as 0 or 1 and model as numeric (yields a single coefficient).
- 3) for ordinal variables, set the values depending on the order of the factor, e.g., small = 1 < medium = 2 < large = 3, and then model as numeric (yields a single coefficient).
- 4) Use `piecewiseSEM`

Biocontrol agents of crop-pests (aphids)

Lacewing larva



Parasitic wasp

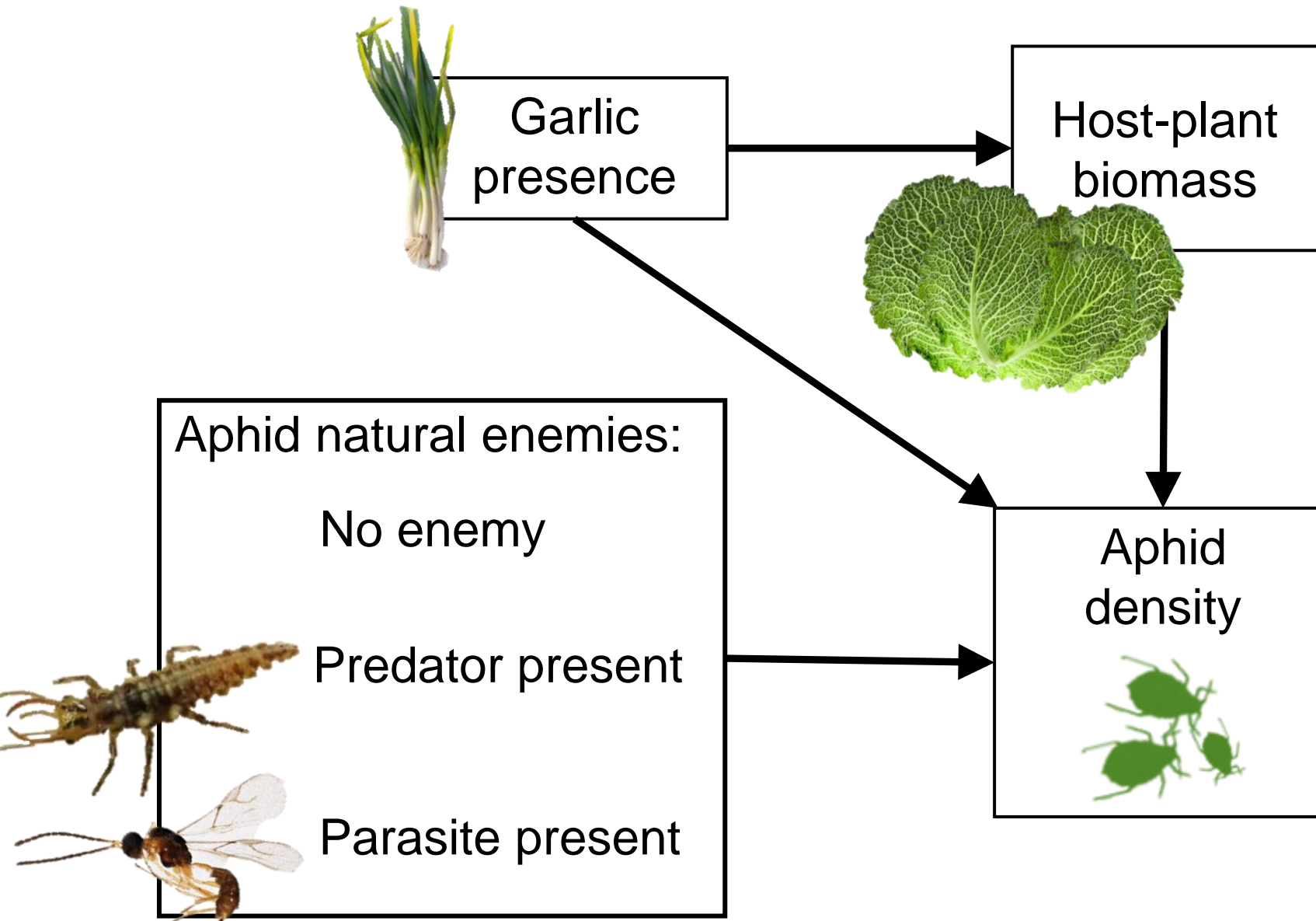


Intercropping with repellent plants



Example

Categorical Exogenous Variable



150 experimental microcosms

Example

Categorical Exogenous Variable

```
# Read and check the data
```

```
aphid_data <- read_csv("Data/Aphid_data.csv")
```

```
> str(aphid_data)
```

```
spec_tbl_ [150 × 4] (S3: spec_tbl_df/tbl_df/tbl/data.frame)
```

```
$ aphid      : num [1:150] 14.9 35.6 43.8 2.1 36.7 ...
```

```
$ host_plant: num [1:150] 38.8 40.7 46.9 35.2 50.9 ...
```

```
$ garlic_ef : chr [1:150] "present" "absent" "absent" "present" ...
```

```
$ enemy      : chr [1:150] "predator" "predator" "no_enemy" "parasite" ..
```

binary variable



nominal variable



Example

Categorical Exogenous Variable

```
# Create dummy variables-----  
# convert "enemy" in 3 binary dummy variables  
# and convert garlic_ef into 1 binary variable called garlic  
  
aphid_data <- aphid_data %>%  
  mutate(n = 1) %>%  
  pivot_wider(names_from = enemy, values_from = n,  
              values_fill = list(n = 0)) %>% # convert "enemy"  
  mutate(garlic = case_when(garlic_ef == "present" ~ 1, # convert " garlic_ef"  
                           garlic_ef == "absent" ~ 0))
```


Example

Categorical Exogenous Variable

```
$ aphid      : num [1:150] 14.9 35.6 43.8 2.1 36.7 ...
$ host_plant: num [1:150] 38.8 40.7 46.9 35.2 50.9 ...
$ garlic_ef  : chr [1:150] "present" "absent" "absent" "present" .
$ garlic     : num [1:150] 1 0 0 1 0 1 1 1 0 0 ...
$ predator   : num [1:150] 1 1 0 0 0 0 1 0 1 0 ...
$ no_enemy   : num [1:150] 0 0 1 0 1 1 0 0 0 0 ...
$ parasite   : num [1:150] 0 0 0 1 0 0 0 1 0 1 ...
```

**(0/1) - dummy
variable for
binary**

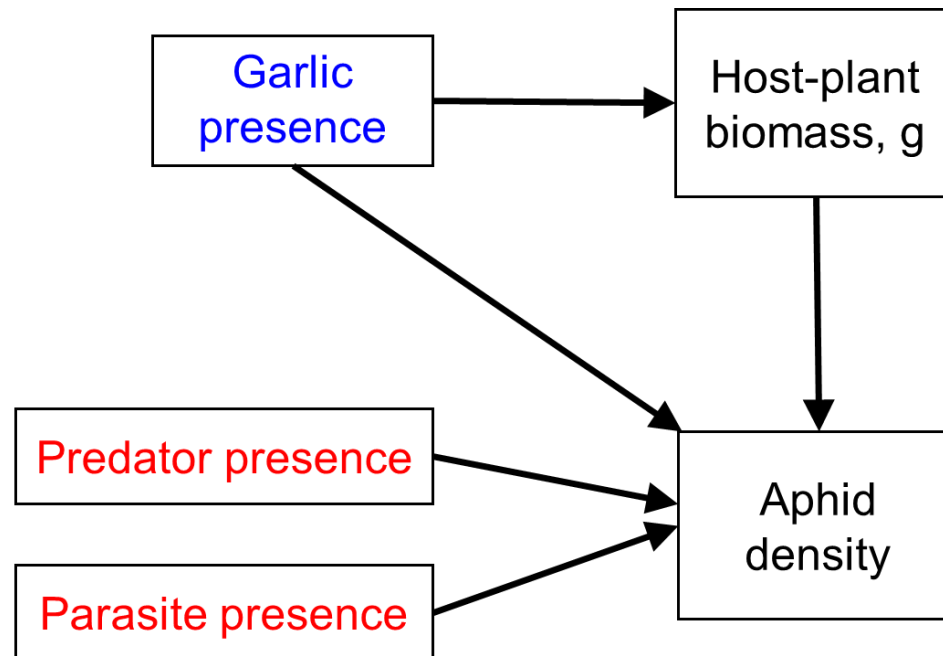
**dummy
variables
created for
each factor
level from the
nominal
variable**

Example

Categorical Exogenous Variable

```
# specify and fit the model in lavaan  
sem_mod <- ' aphid ~ host_plant + garlic + predator + parasite  
             host_plant ~ garlic  
'
```

Only 2 out of 3
dummy variables
are included



Example

Categorical Exogenous Variable

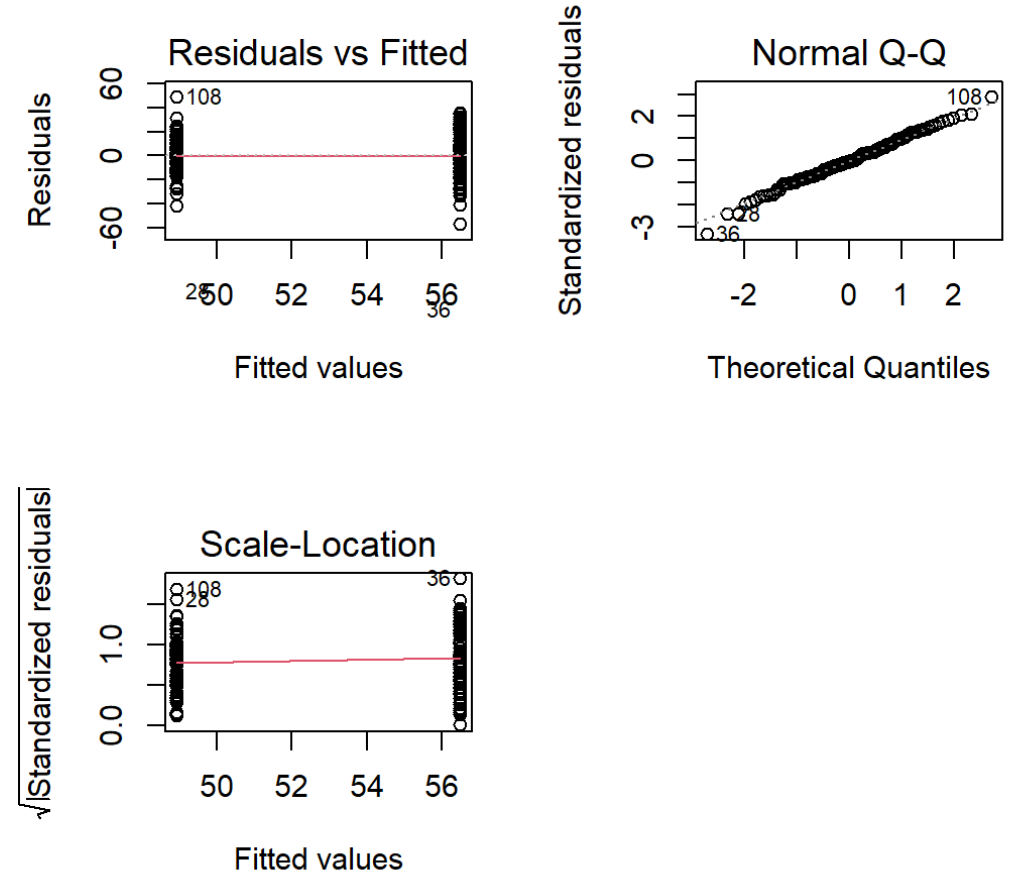
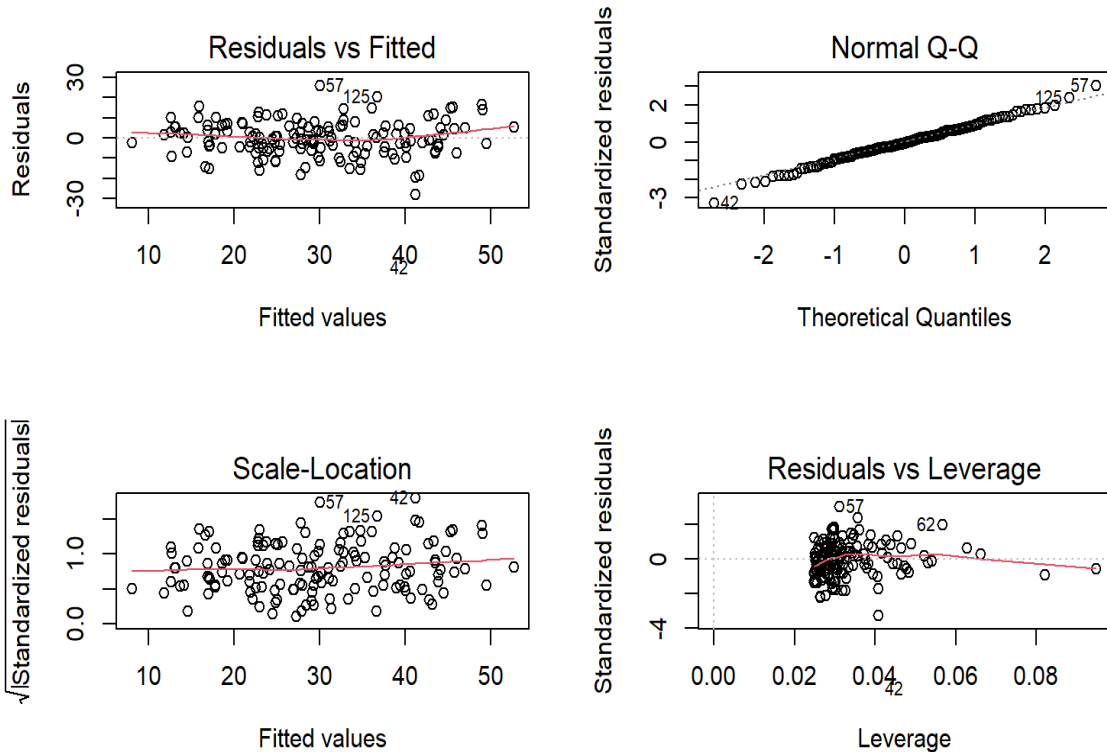
```
#Check the assumptions:
# Normality of residuals
mod1 <- lm(aphid ~ host_plant + garlic + predator + parasite, aphid_data)
car::vif(mod1) # check for correlation among predictors
>
host_plant      garlic      predator      parasite
    1.061508    1.066580    1.343933    1.355159

mod2 <- lm(host_plant ~ garlic, aphid_data)
```

Example

Categorical Exogenous Variable

```
par(mfrow=c(2,2))  
plot(mod1)  
plot(mod2)  
par(mfrow=c(1,1))
```



Example

Categorical Exogenous Variable

```
# Normality of data
library(MVN)
mvn(aphid_data %>%
    select(-enemy_cat, -garlic_ef, -no_enemy),
    mvnTest="mardia", univariateTest="SW")
```

```
>
$multivariateNormality
```

	Test	Statistic	p value	Result
1	Mardia Skewness	71.3789094435619	0.000273949726176333	NO
2	Mardia Kurtosis	-3.9601958323228	7.48883249854781e-05	NO
3	MVN	<NA>	<NA>	NO

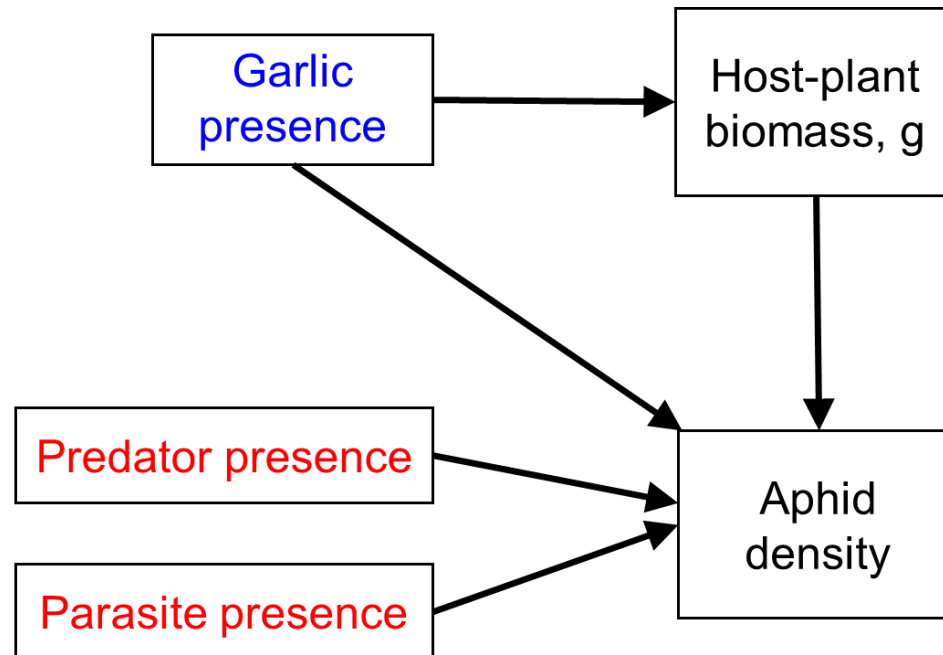
Recap: Protocol for violated assumptions of covariance-based SEM

Violated assumptions	Steps for Corrections
Non-normality of Residuals	Data transformation: e.g. <i>log</i> , <i>square root</i>
	Local estimation with GLM: package <code>piecewiseSEM</code>
Data are not multivariate normal	MLM estimation with robust SE & test statistic: <code>library(lavaan) # Always report results for 'robust' test statistics</code> <code>sem(..., estimator="MLM", se="robust"</code> <code> #or test="Satorra-Bentler")</code>
	Bootstrapping: <code># Always report results for 'robust' test statistics</code> <code>library(lavaan)</code> <code>sem(..., test="bollen.stine", se="bootstrap")</code>
Missing data	Full information maximum likelihood: <code>library(lavaan)</code> <code>sem(..., missing="fiml") #for normal data</code> <code>sem(..., missing="fiml", estimator="MLR") #for non-normal data</code>
Positive definite S matrix	Check for multicollinearity in each single regression model: <code>library(car)</code> <code>vif(m2) # vif ≤ 2 (no collinearity)</code>
Dependant samples (hierarchical)	Local estimation with LMM or GLMM: package <code>piecewiseSEM</code>
Not sufficient sample size	Local estimation: package <code>piecewiseSEM</code>

Example

Categorical Exogenous Variable

```
# specify and fit the model in lavaan  
  
sem_mod <- ' aphid ~ host_plant + garlic + predator +  
parasite  
  
host_plant ~ garlic  
'
```



Example

Categorical Exogenous Variable

```
# specify and fit the model in lavaan

sem_mod <- ' aphid ~ host_plant + garlic + predator +
parasite

host_plant ~ garlic
'

fit <- sem(sem_mod,
           test="Satorra-Bentler", data=aphid_data)

summary(fit, standardize = T, rsq=T, fit.measures=TRUE)
```


Example

Categorical Exogenous Variable

```
> summary(fit, standardize = T, rsq=T, fit.measures=TRUE)
```

Model Test User Model:

	Standard	Scaled
Test Statistic	1.658	1.655
Degrees of freedom	2	2
P-value (Chi-square)	0.436	0.437
Scaling correction factor		1.002
Satorra-Bentler correction		
...		
Robust Comparative Fit Index (CFI)		1.000
...		
RMSEA	0.000	0.000
90 Percent confidence interval - upper	0.153	0.153
P-value H ₀ : RMSEA ≤ 0.050	0.558	0.559
...		
SRMR	0.025	0.025

How to
present fit
statistics?

$\chi^2 = 1.65$, DF=2,
n=150, p = 0.43

RMSEA=0,
(CI = 0, 0.15) ,
p_{RMSEA}=0.55,

CFI=1.00;

SRMR=0.025

Example

Categorical Exogenous Variable

```
> summary(fit, standardize = T, rsq=T, fit.measures=TRUE)
```

...

Regressions:

	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
aphid ~						
host_plant	0.408	0.041	9.925	0.000	0.408	0.534
garlic	-8.506	1.437	-5.921	0.000	-8.506	-0.321
predator	-11.372	1.707	-6.663	0.000	-11.372	-0.405
parasite	-7.375	1.712	-4.309	0.000	-7.375	-0.262
host_plant ~						
garlic	-7.570	2.769	-2.734	0.006	-7.570	-0.218

Variances:

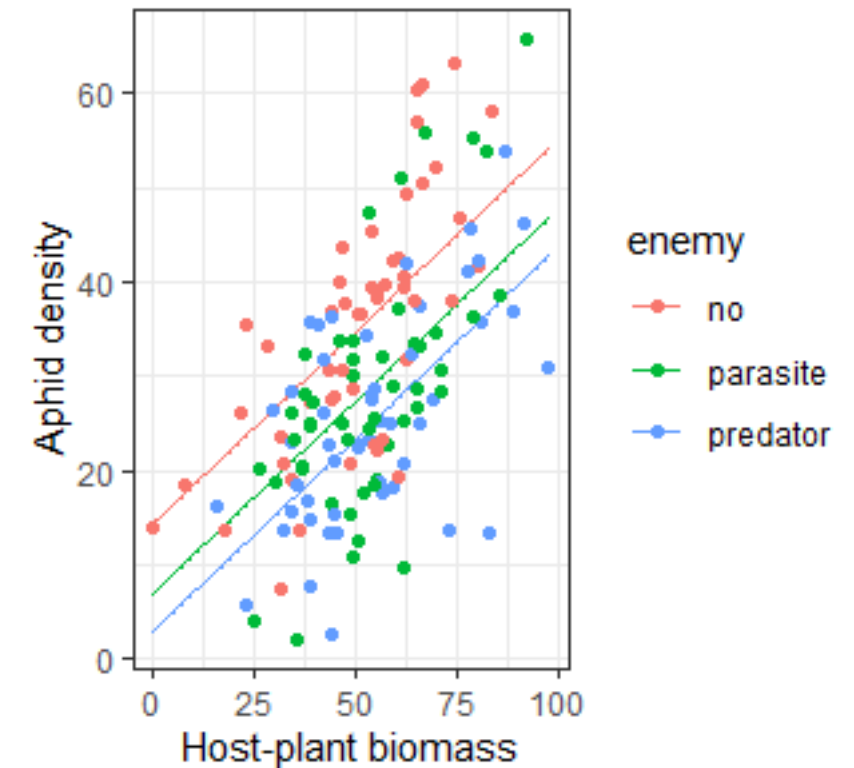
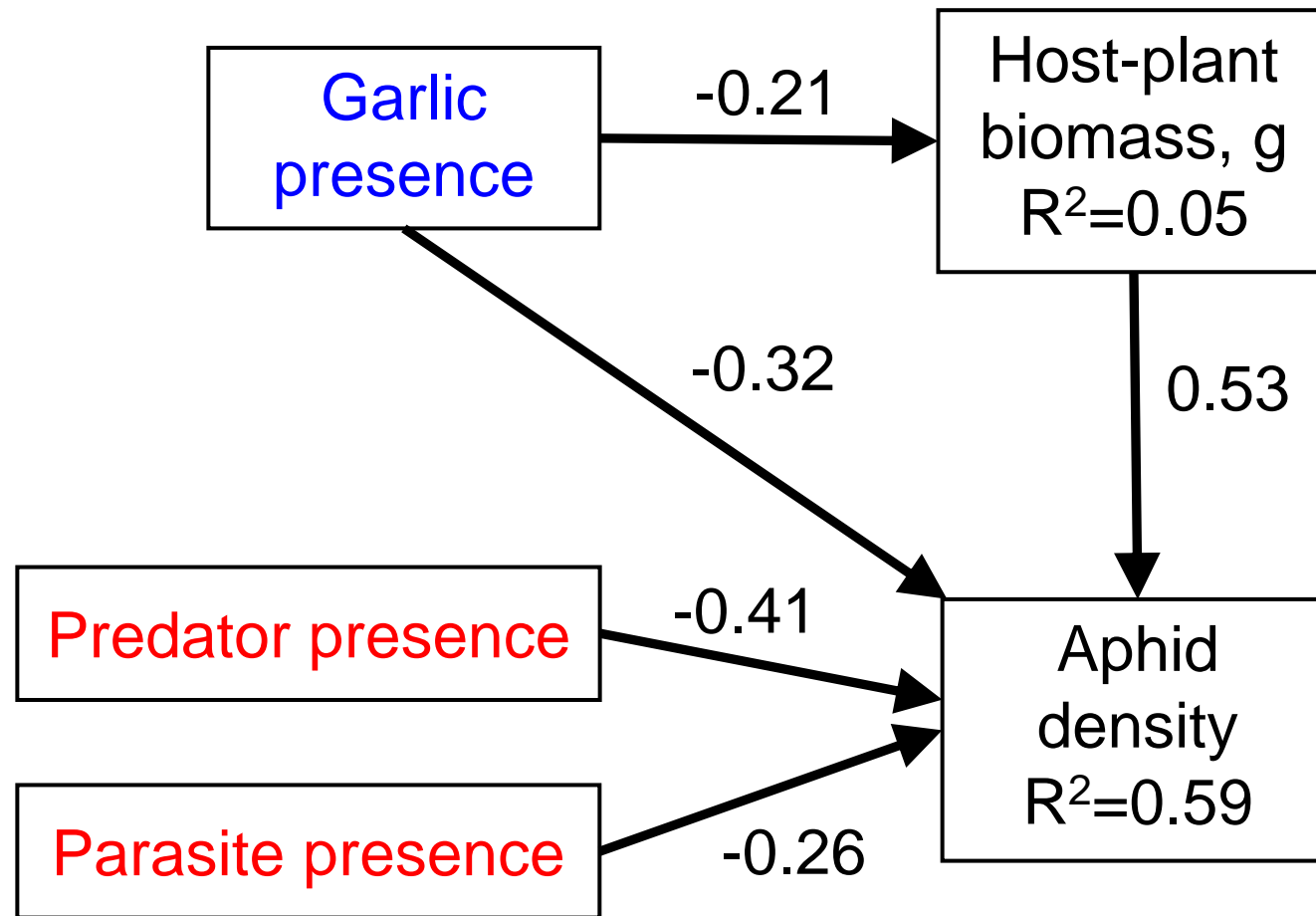
	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
.aphid	72.753	8.401	8.660	0.000	72.753	0.414
.host_plant	287.445	33.191	8.660	0.000	287.445	0.953

R-Square:

	Estimate
aphid	0.586
host_plant	0.047

Example

Categorical Exogenous Variable



$\chi^2 = 1.65$, $DF=2$, $n=150$, $p = 0.43$ $RMSEA=0$, $(CI = 0, 0.15)$, $p_{RMSEA}=0.55$, $CFI=1.00$; $SRMR=0.025$

Example

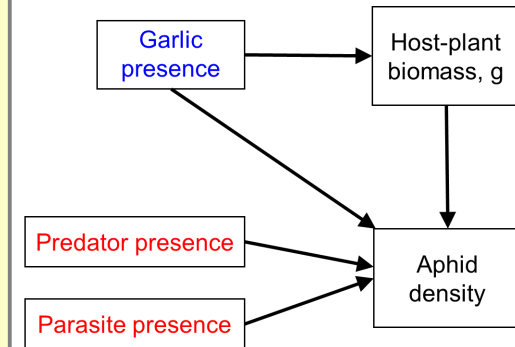
Categorical Exogenous Variable

```
# calculate indirect effects
sem_mod <- ' aphid ~ a1*host_plant + a2*garlic + predator + parasite
            host_plant ~ a3*garlic
            # define indirect and total effect
            direct := a2
            indirect := a3*a1
            total := direct + indirect
            '

fit <- sem(sem_mod, data=aphid_data)
summary(fit, standardize = T, rsq = T, fit.measures=T)
>
```

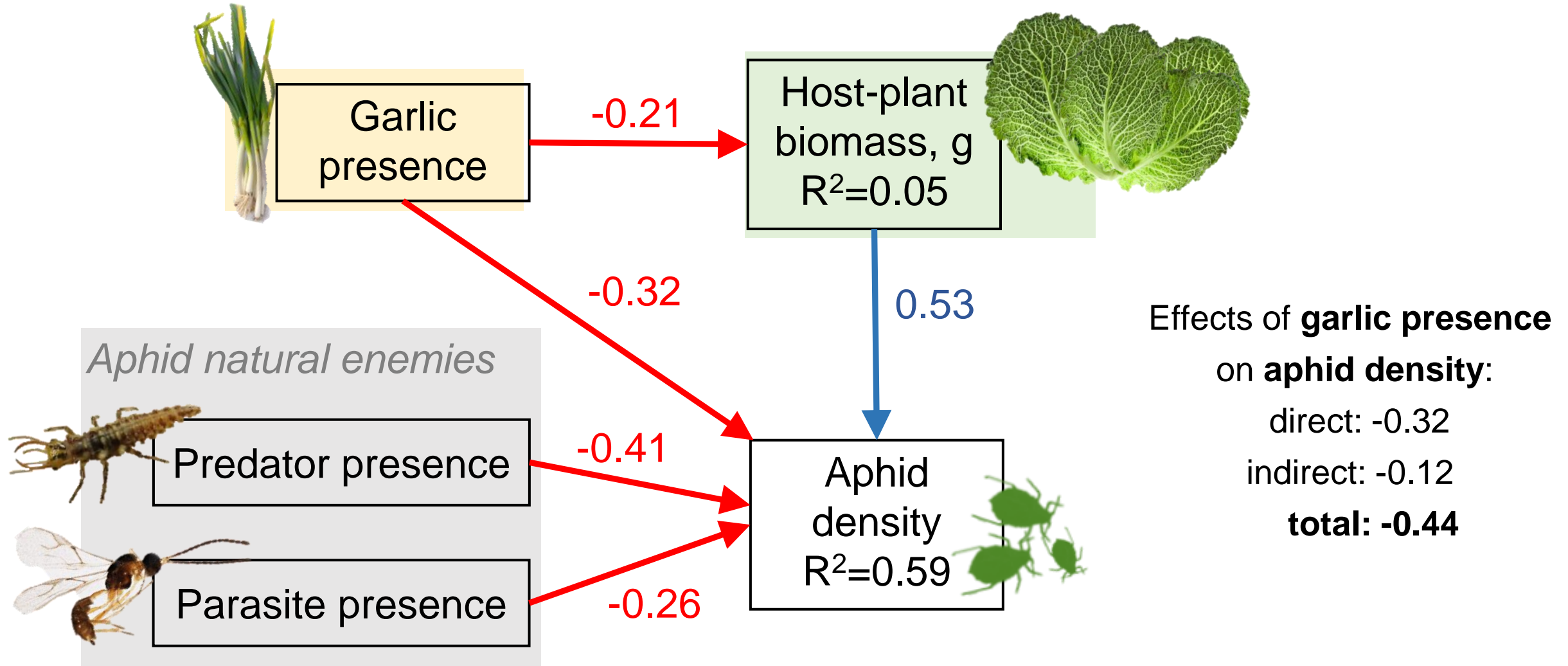
Defined Parameters:

	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
direct	-8.506	1.437	-5.921	0.000	-8.506	-0.321
indirect	-3.086	1.171	-2.636	0.008	-3.086	-0.116
total	-11.592	1.800	-6.439	0.000	-11.592	-0.437



Example

Categorical Exogenous Variable



$\chi^2 = 1.65$, $DF=2$, $n=150$, $p = 0.43$ $RMSEA=0$, $(CI = 0, 0.15)$, $p_{RMSEA}=0.55$, $CFI=1.00$; $SRMR=0.025$

Endogenous Categorical Variables

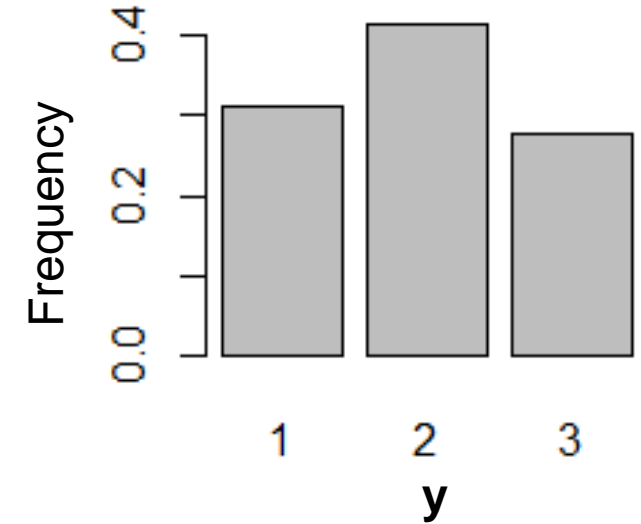
Approaches when we have Endogenous Categorical Variables:

- 1) for binary and ordinal variables use the argument 'ordered' in ***lavaan*** with fitting function 'sem'
- 2) for nominal variables (i.e., levels are not ordered) use the factor levels to construct a composite variable.

Endogenous Categorical Variables

- Normal distribution means continuous data
- Ordinal data can not be assumed normal

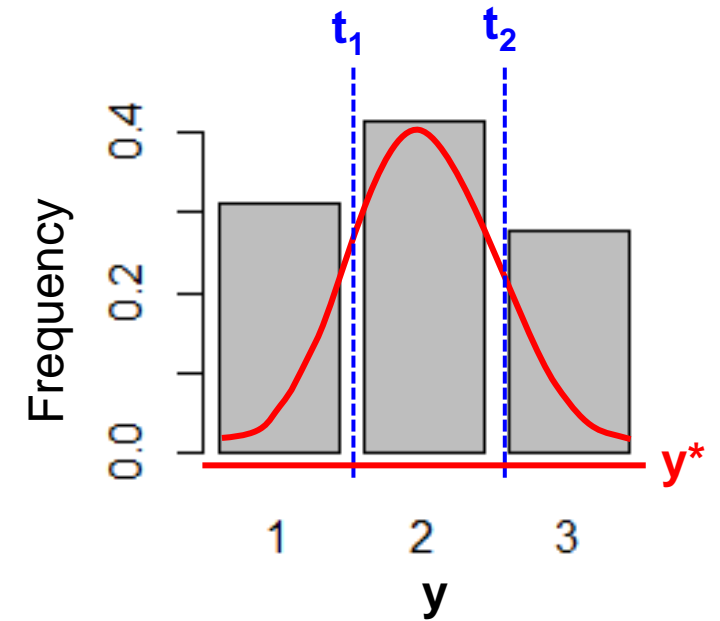
Solution: to use the threshold models



Endogenous Categorical Variables

- Normal distribution means continuous data
- Ordinal data can not be assumed normal

Solution: to use the threshold models

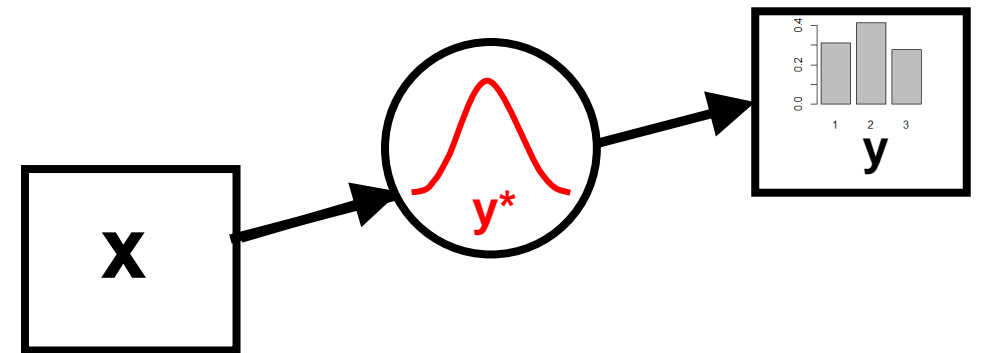
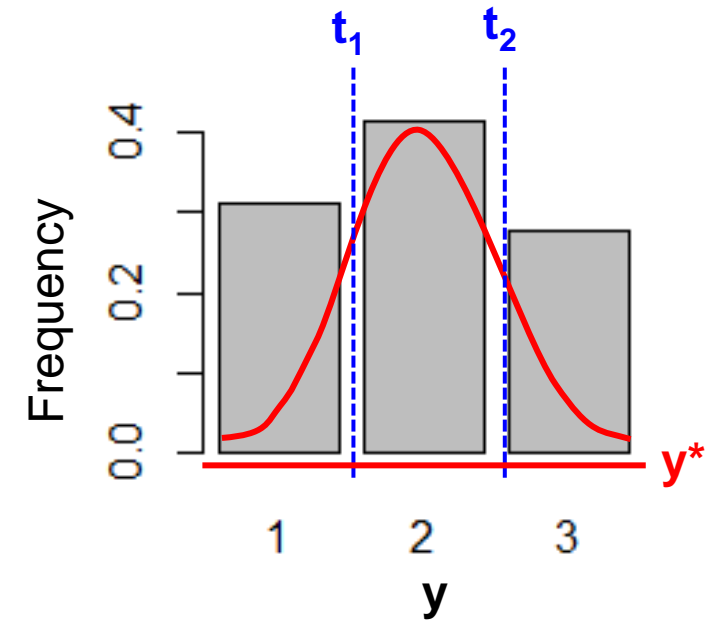


Threshold, for instance t_1 is a match between the probability of $y=1$ and actual percent that the observed data =1.

Endogenous Categorical Variables

- Normal distribution means continuous data
- Ordinal data can not be assumed normal

Solution: to use the threshold models



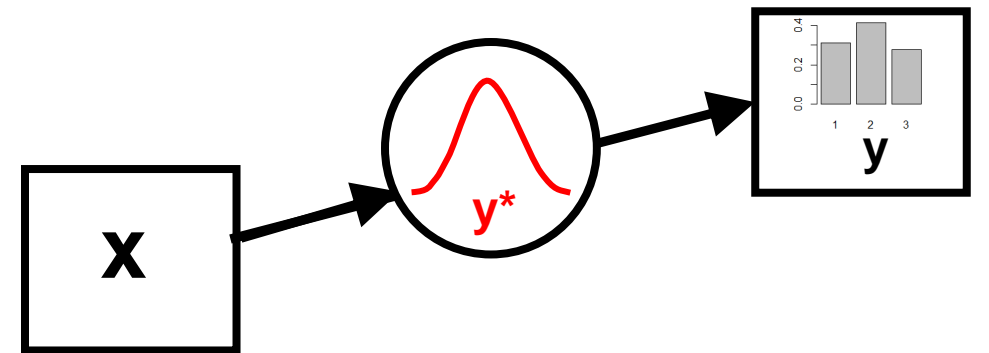
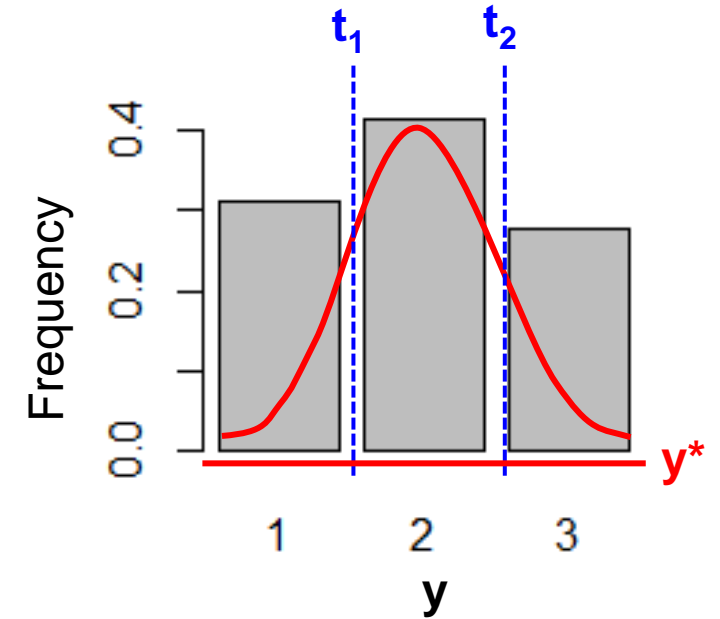
Endogenous Categorical Variables

- Normal distribution means continuous data
- Ordinal data can not be assumed normal

Solution: to use the threshold models

Estimation not via ML but via
(diagonally) weighted least squares (D)WLS

$$F_{WLS} = (\mathbf{s} - \boldsymbol{\sigma})^\top \mathbf{W}^{-1} (\mathbf{s} - \boldsymbol{\sigma})$$



Example

Categorical Endogenous Variable

Human activities affect fish communities in ponds

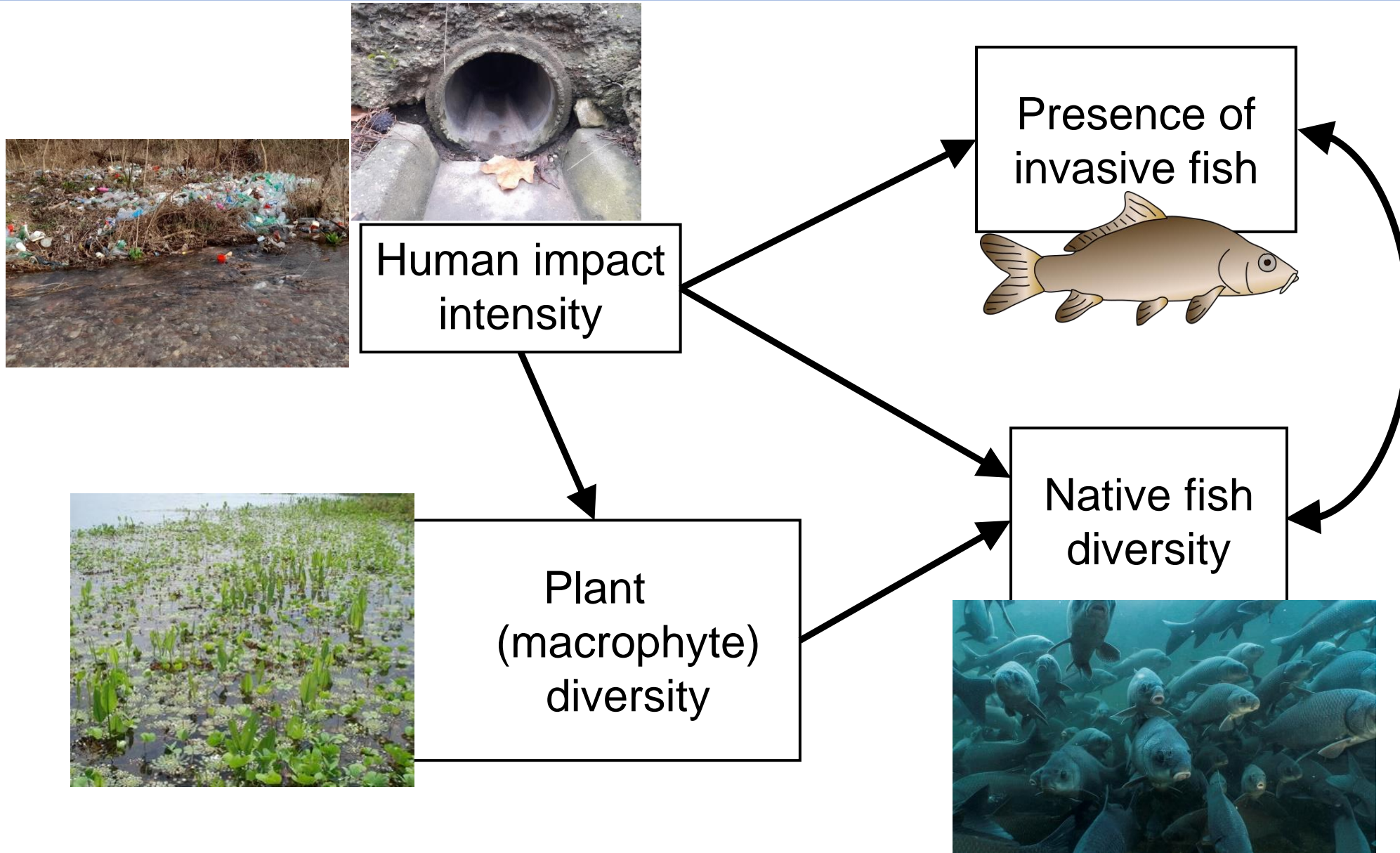


120 ponds



Example

Categorical Endogenous Variable



Example

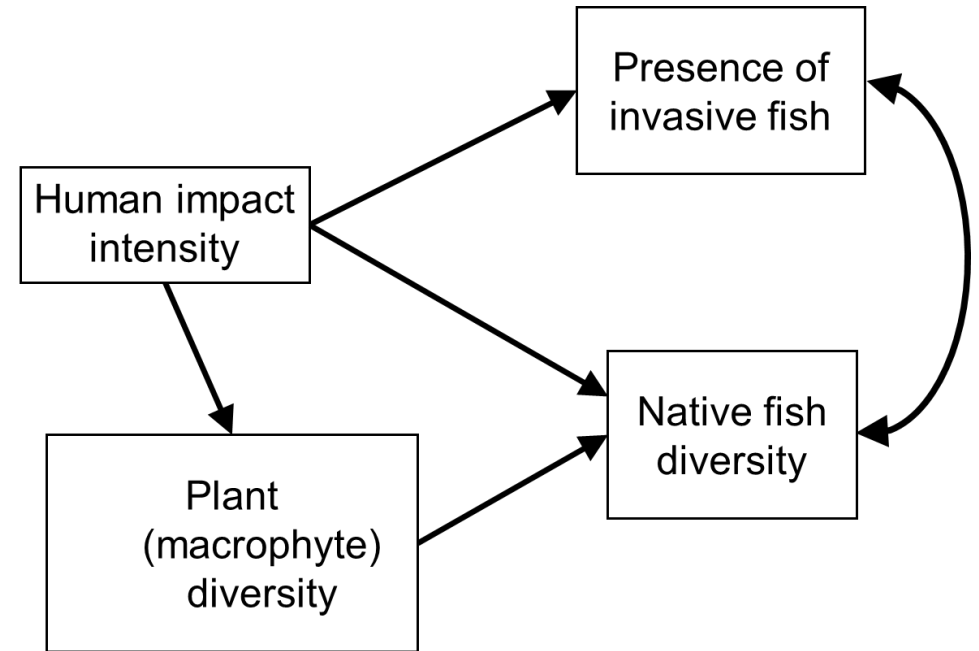
Categorical Endogenous Variable

```
# Read and check the data
fish_data <- read_csv("Data/Fish_data.csv")
str(fish_data)

sem_mod2 <- ' inv_fish ~ HII
              native_fish ~ plant_div + HII
              plant_div ~ HII
              native_fish ~~ inv_fish
            '

fit2 <- sem(sem_mod2, data=fish_data,
            ordered = c("inv_fish"))

summary(fit2, standardize = T, rsq = T)
```



Example

Categorical Endogenous Variable

```
# Read and check the data
```

Estimator	DWLS
Optimization method	NLMINB
Number of model parameters	10
Number of observations	120

Model Test User Model:

	Standard	Scaled
Test Statistic	0.022	0.022
Degrees of freedom	1	1
P-value (Chi-square)	0.882	0.882
Scaling correction factor		1.000
Shift parameter		0.000
simple second-order correction		

Parameter Estimates:

Standard errors	Robust.sem
-----------------	------------

Example

Categorical Endogenous Variable

```
# Read and check the data
```

...	Standard	Scaled
Comparative Fit Index (CFI)	1.000	1.000
Tucker-Lewis Index (TLI)	1.085	1.098
Robust Comparative Fit Index (CFI)		NA
Robust Tucker-Lewis Index (TLI)		NA
...		
RMSEA	0.000	0.000
90 Percent confidence interval - lower	0.000	0.000
90 Percent confidence interval - upper	0.121	0.121
P-value H_0: RMSEA <= 0.050	0.898	0.898
P-value H_0: RMSEA >= 0.080	0.081	0.081
Robust RMSEA		NA
...		
SRMR	0.007	0.007

robust RMSA and other fit measures are not calculated in DWLS

Use standard measures

Example

Categorical Endogenous Variable

Regressions:

	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
inv_fish ~						
HII	0.308	0.128	2.411	0.016	0.308	0.268
native_fish ~						
plant_div	0.475	0.059	7.994	0.000	0.475	0.576
HII	-1.186	0.424	-2.797	0.005	-1.186	-0.210
plant_div ~						
HII	-1.785	0.695	-2.569	0.010	-1.785	-0.261

Covariances:

	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
.inv_fish ~~						
.native_fish	-1.466	0.572	-2.561	0.010	-1.466	-0.383

Thresholds:

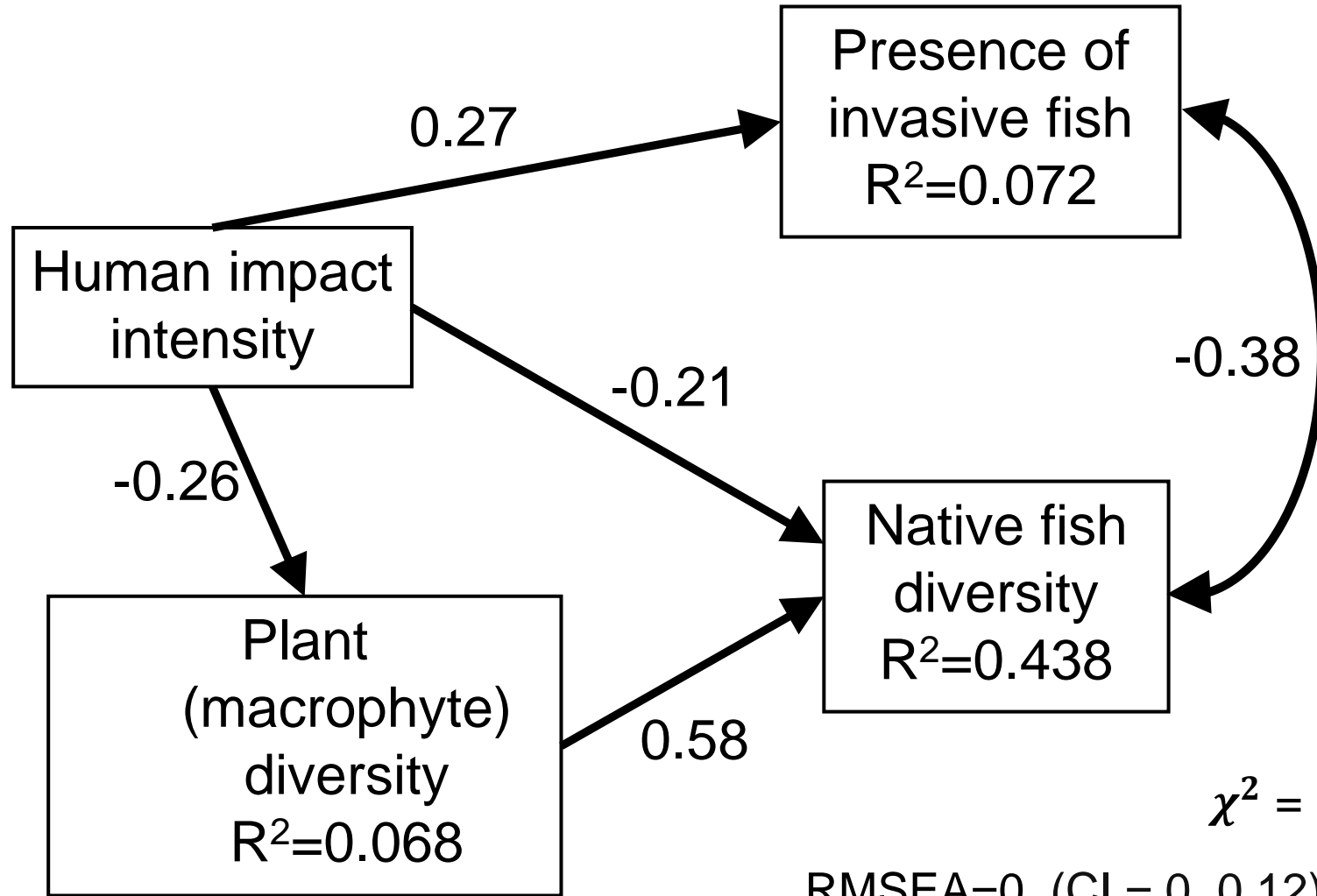
	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
inv_fish t1	0.567	0.288	1.969	0.049	0.567	0.546

R-Square:

	Estimate
inv_fish	0.072
native_fish	0.438
plant_div	0.068

Example

Categorical Exogenous Variable



$$\chi^2 = 0.022, DF=1, n=120, p = 0.88$$

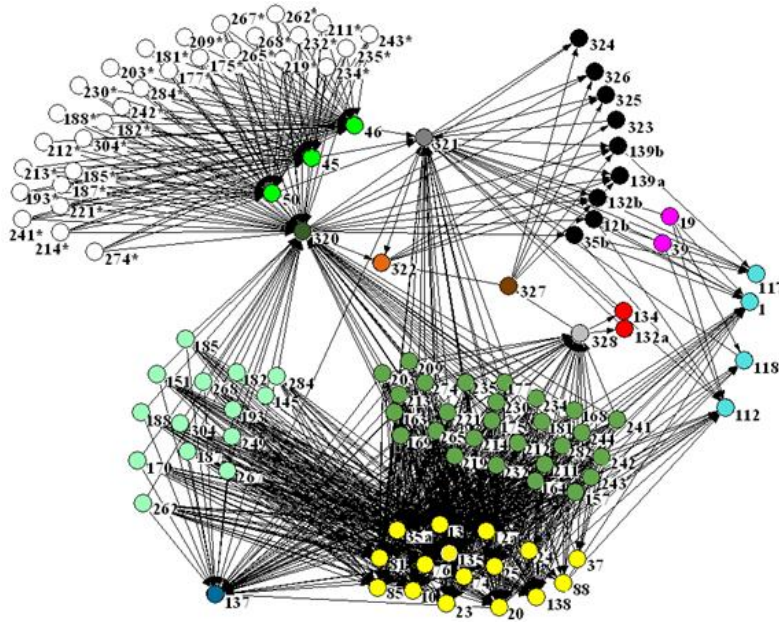
$$RMSEA=0, (CI = 0, 0.12), p_{RMSEA}=0.89, CFI=1.00; SRMR=0.007$$

Protocol for treating categorical variables in SEM

Categorical Variables	Exogenous Categorical Variables	Endogenous Categorical Variables
Binary variables yes/no; presence/absence; failure/success; dead/alive; male/female	1. Set the values as 0 or 1 and model as numeric (yields a single path coefficient).	<pre>library(lavaan)</pre> <pre>sem(..., ordered=c("categ_varibl"))</pre> <ul style="list-style-type: none"> Take care that the levels of your variable have the correct order (e.g. small < medium < large) DWMS estimator is used, which corrects for non-normal data and for ordered data. Report 'robust' test statistics for χ^2. But report 'scaled' RMSA, CFI, SRMR; as no clear suggestions exist regarding the application of these fit indices for non-ML estimators.
	2. Create separate dummy variables for each factor levels with values 0, 1 each. Rule: for the factor with k levels use k-1 dummy variables (to avoid singularity).	
	3. Use package <code>piecewiseSEM</code>	
Ordinal variables: small < medium < large; yang < middle < old	1. Set the values depending on the order of the factor, e.g., small = 1 < medium = 2 < large = 3, and then model as numeric.	<pre>library(piecewiseSEM)</pre> Endogenous categorical variables are not implemented in <code>piecewiseSEM</code> . Treat binary and ordinal variables as numerical (follow step 1 shown for 'Endogenous Categorical Variables')
	2. Create separate dummy variables for each factor levels with values 0, 1 each. Rule: for the factor with k levels use k-1 dummy variables (to avoid singularity).	
	3. Use package <code>piecewiseSEM</code>	
Nominal variables study sites (e.g., site 1, site 2, site 3); countries; sampling campaigns	1. Create separate dummy variables for each factor levels with values 0, 1 each. Rule: for the factor with k levels use k-1 dummy variables (to avoid singularity).	Use the factor levels to construct a composite variable.
	2. Use package <code>piecewiseSEM</code>	Nominal endogenous categorical variables are not implemented in <code>piecewiseSEM</code>

Day 7 Task 1

Effects of land use on arthropod food webs in grasslands



Food webs



Net sampling of arthropods in grasslands

235 grasslands

Food-web length

“1 level”: only herbivores and decomposers,

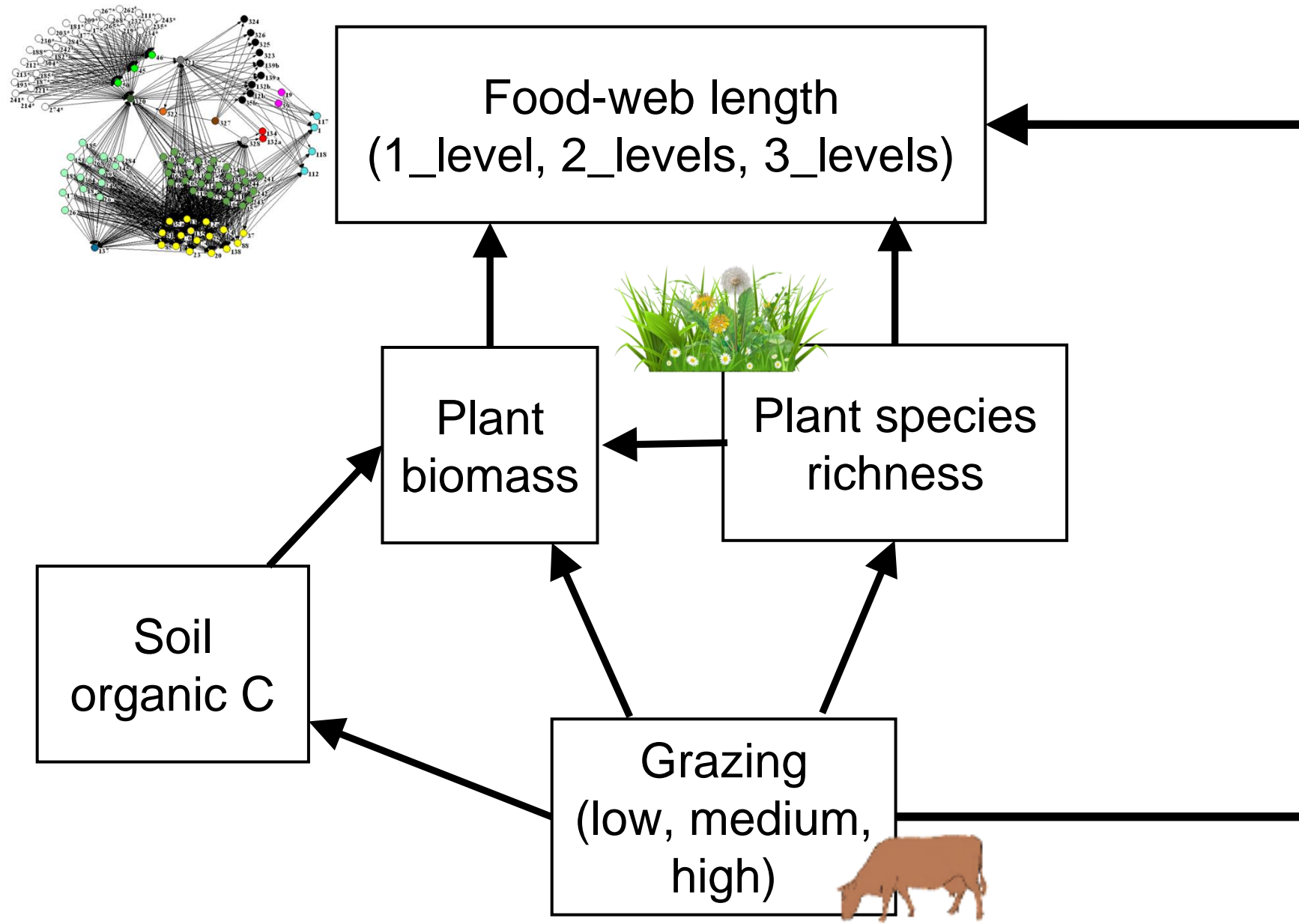
“2 levels”: carnivores present in addition to level 1,

“3 levels”: omnivores present in addition to level 1 and level 2.

Grazing intensity
 (“low”, “medium”, or “high”)

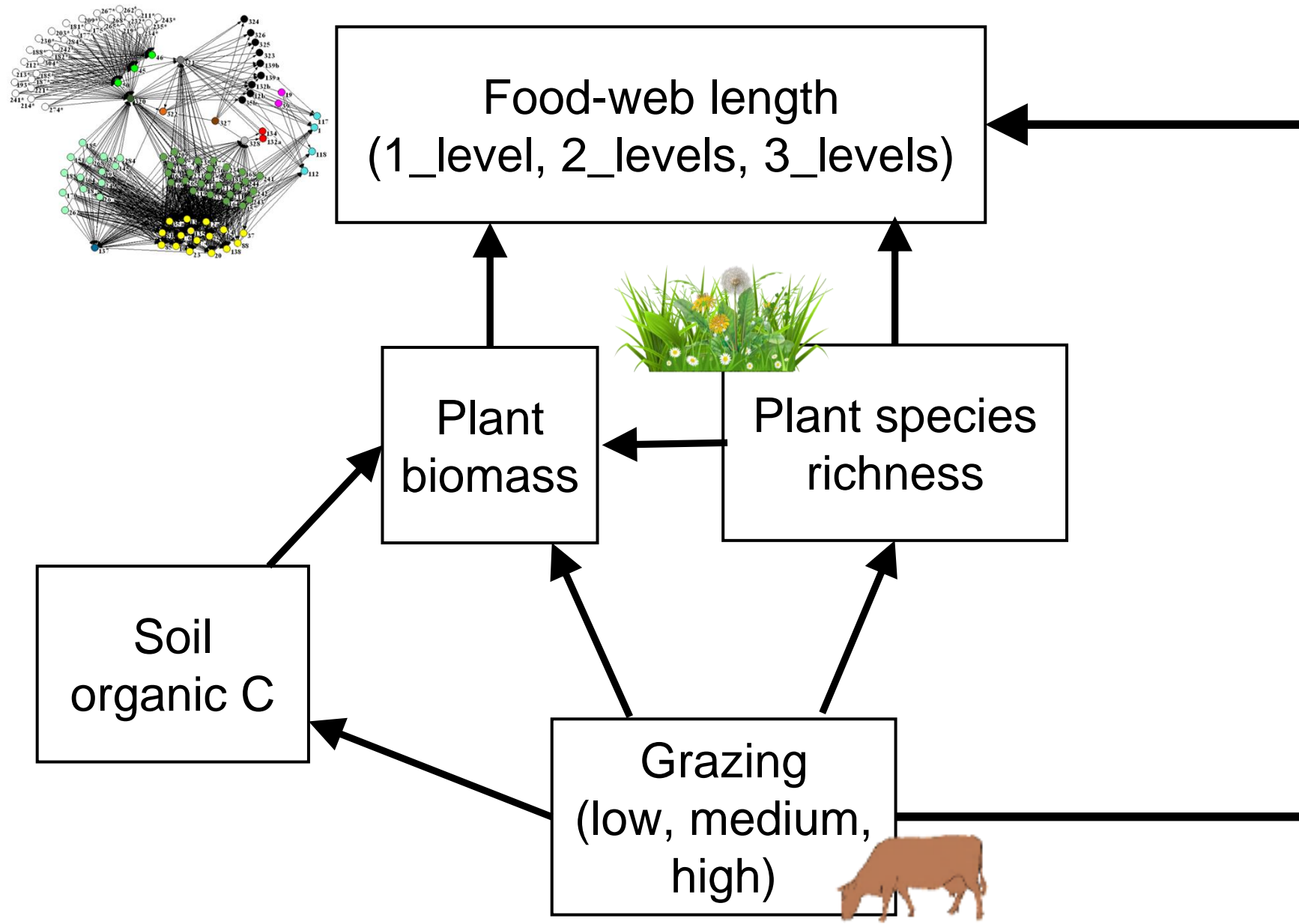
Day 7 Task 1

Effects of land use on food webs in grasslands



Day 7 Task 1

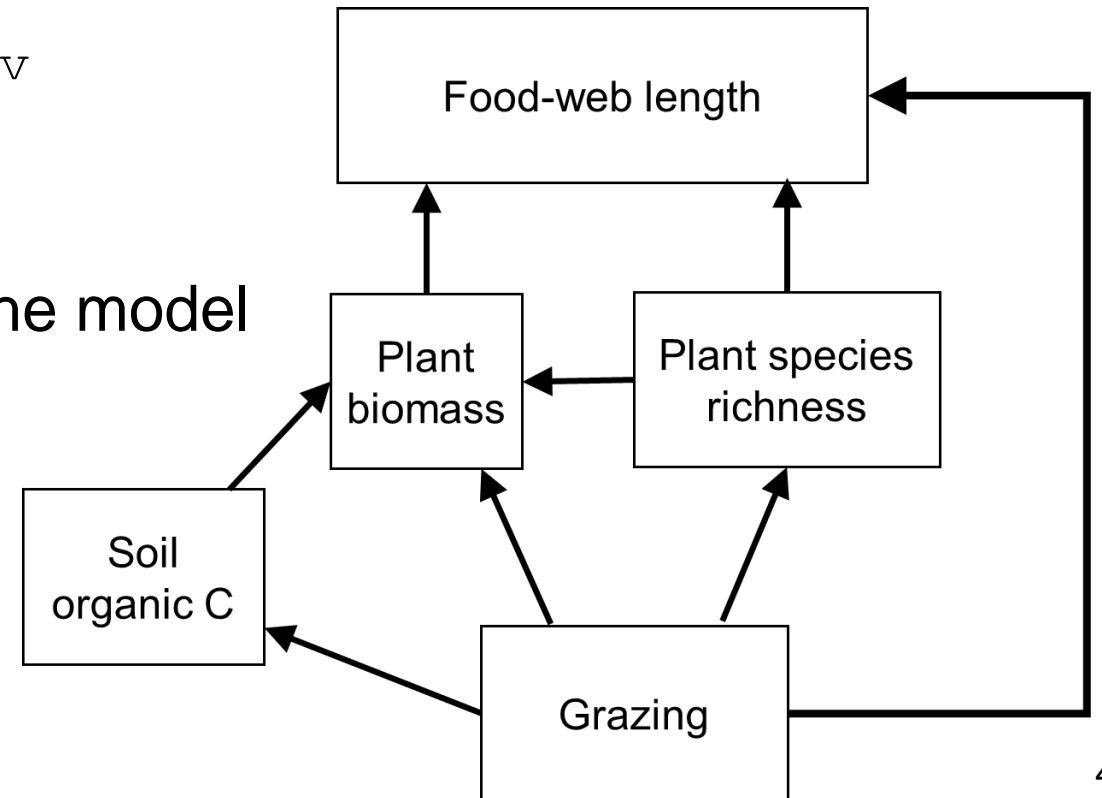
Effects of land use on food webs in grasslands



Day 7 Task 1

Effects of land use on food webs in grasslands

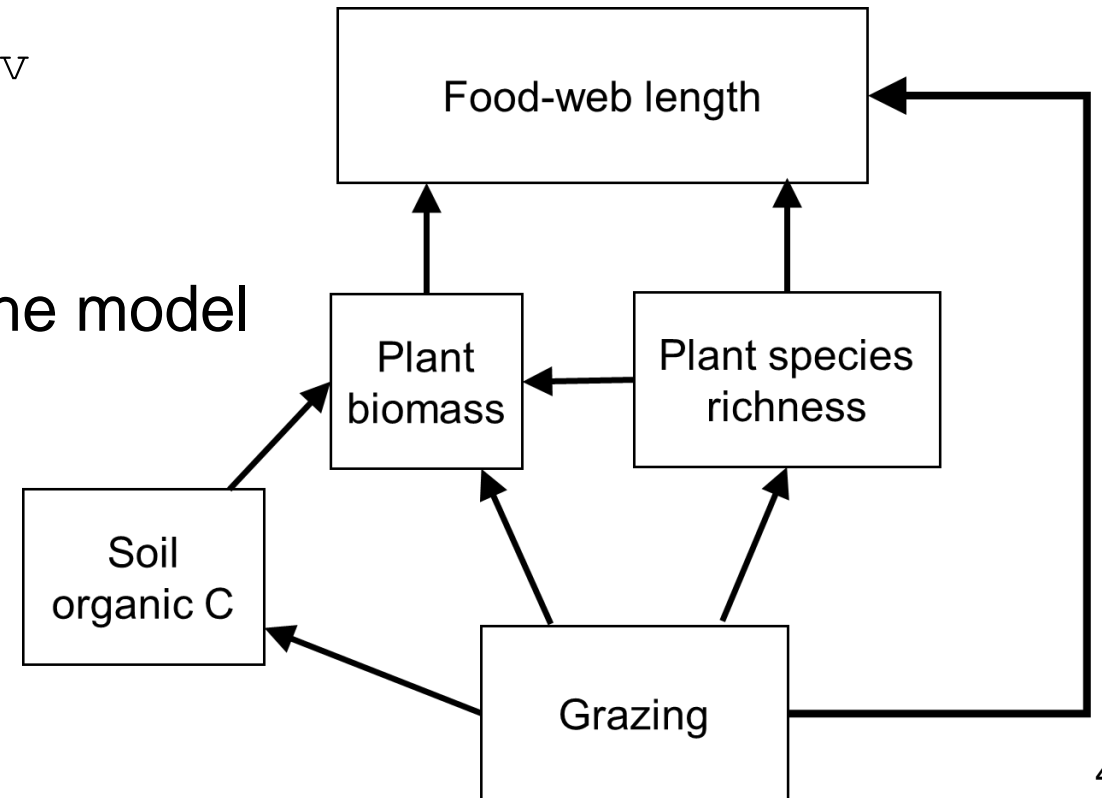
1. Specify the following model in lavaan
 - For this, if needed, recode the categorical variables in a way appropriate for the analysis
3. Fit the model using data `Food-web_data.csv`
4. Get the fit indices
5. Fill in Standardized Coefficients and R^2 for the model
6. Think about how to interpret the results



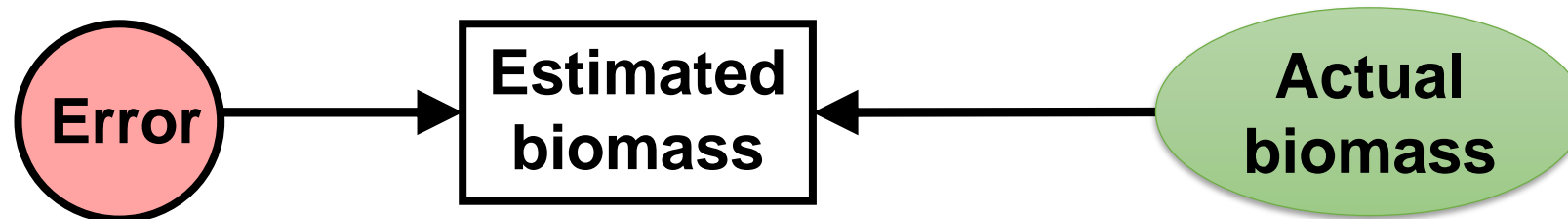
Day 7 Task 1

Effects of land use on food webs in grasslands

1. Specify the following model in lavaan
 - For this, if needed, recode the categorical variables in a way appropriate for the analysis
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- Latent Variables in SEM



Latent Variables in SEM

- What are Latent Variables? Why to use them?
- Multi-indicator Latent Variables
- Fitting Latent Variables
(Confirmatory Factor Analysis)

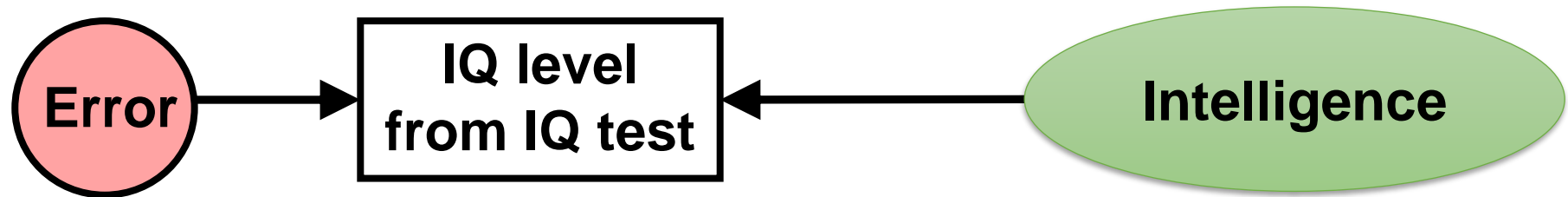
Latent Variables in SEM

- **What are Latent Variables? Why to use them?**
- Multi-indicator Latent Variables
- Fitting Latent Variables
(Confirmatory Factor Analysis)

What is Latent Variable?

Latent – hypothetical, hidden

- a variable that is **unmeasured**, but is **hypothesized to exist**
- scientific concept that is **not directly observed**, but is hypothetical **construct**
- can be **approximated using observable indicators**

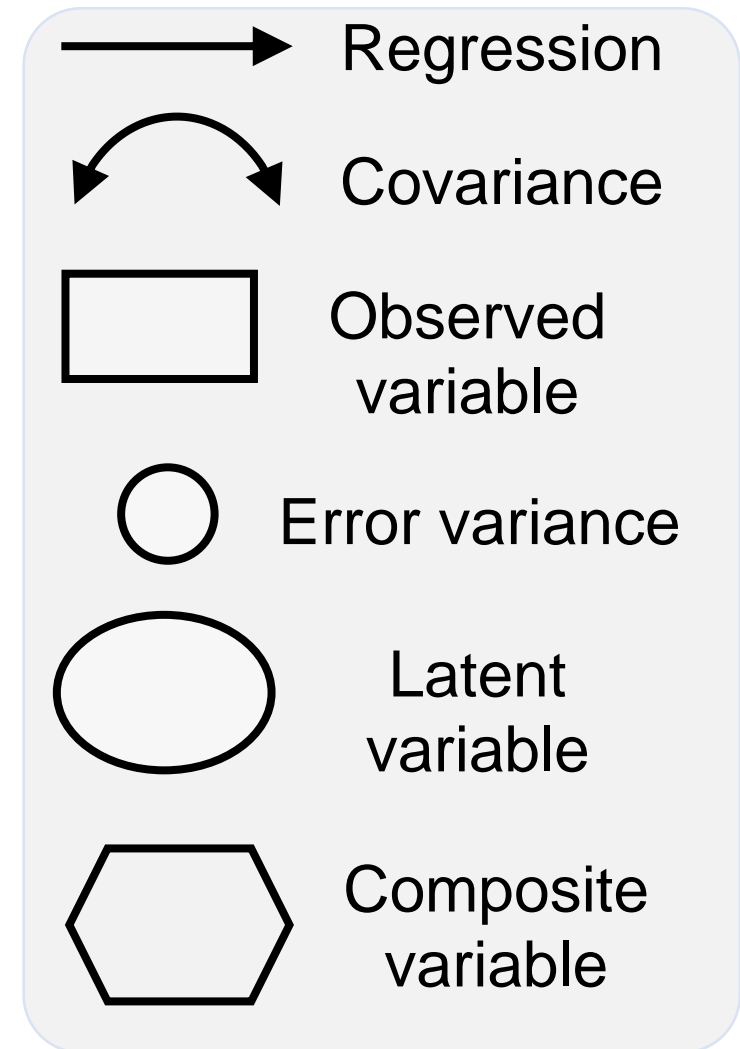


What is Latent Variable?

Specification operators in 'lavaan'

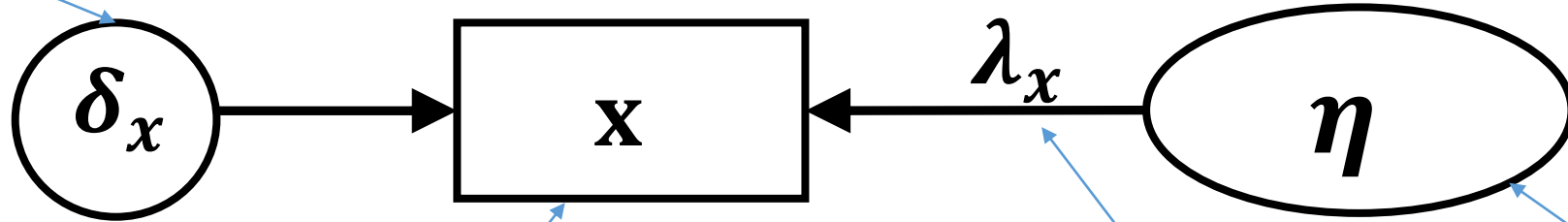
formula type	operator	meaning
Regression	~	"regressed on"
Correlation	~~	"correlated with"
Intercept	~ 1	"estimates intercept"
Latent variable	=~	"is measured by"
Composite	<~	"is caused by"

Path Diagram Notations:



What is Latent Variable?

The error in the measurement of \mathbf{x} by η



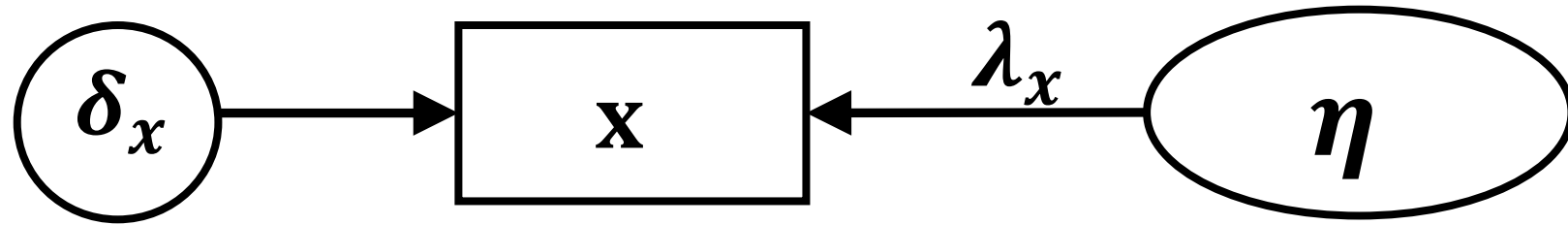
Observed variable
“manifest indicator”

“factors” or “latent traits”

The relationship between a latent variable and its observed indicator

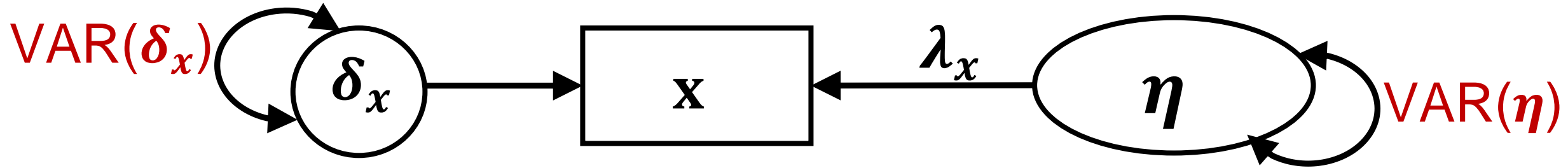
Latent variable

What is Latent Variable?



$$\mathbf{x} = \lambda_x \eta + \delta_x$$

What is Latent Variable?

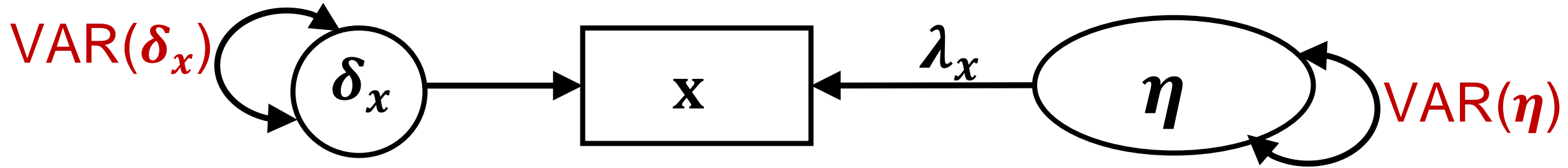


$$\mathbf{x} = \lambda_x \eta + \delta_x$$

$$\eta \sim N(0, \text{SD}(\eta))$$

$$\delta \sim N(0, \text{SD}(\delta))$$

What is Latent Variable?



$$\mathbf{x} = \lambda_x \eta + \delta_x$$

$$\eta \sim N(0, \text{SD}(\eta))$$

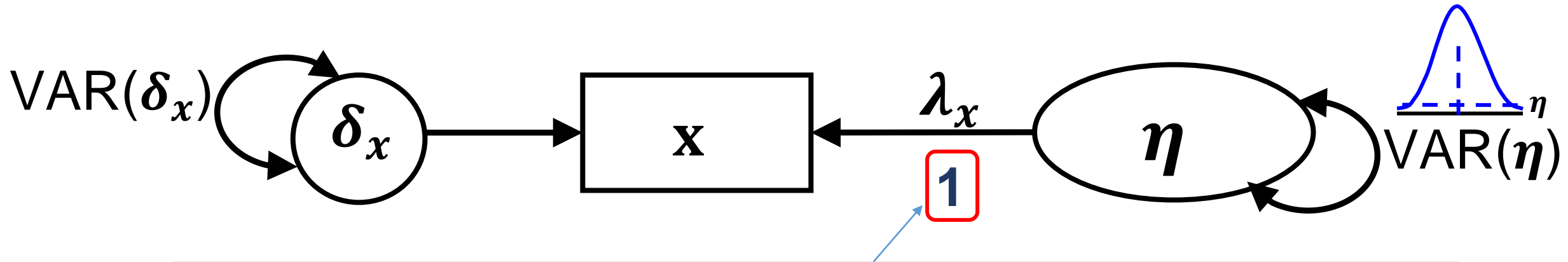
$$\delta \sim N(0, \text{SD}(\delta))$$

$$\text{VAR}(\mathbf{x}) = \lambda_x^2 \text{VAR}(\eta) + \text{VAR}(\delta)$$

How much variance does the LV explain?

$$\frac{\lambda_x^2 \text{VAR}(\eta)}{\lambda_x^2 \text{VAR}(\eta) + \text{VAR}(\delta)}$$

What is Latent Variable?



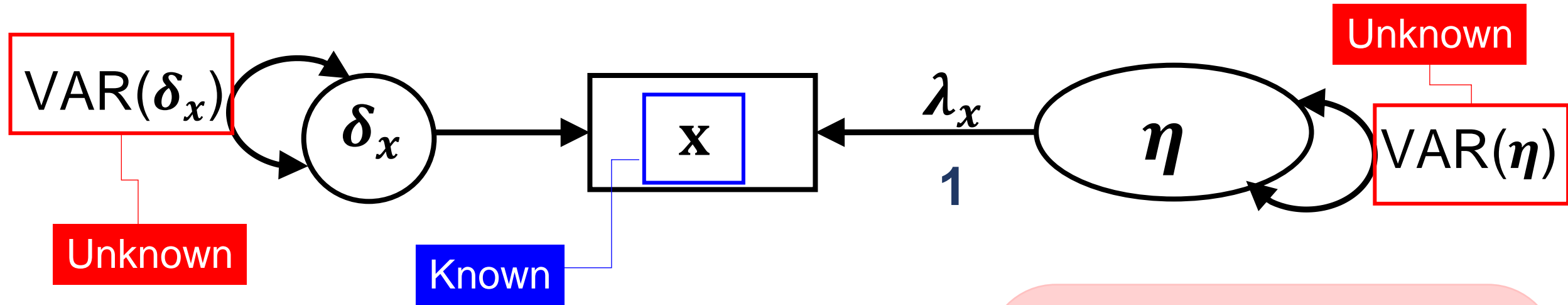
Raw scale coefficient: matches observed (co)variances to parameters $\text{VAR}(\delta)$ and $\text{VAR}(\eta)$

We explain the data well if:

$$\text{VAR}(\mathbf{x}) = \text{VAR}(\eta) + \text{VAR}(\delta)$$

- What is the scale/unit of our LV?
It needs to be defined to get the regression weights.

What is Latent Variable?



$$\text{DF} = t_{\max} - t = -1$$

- Model is not identified

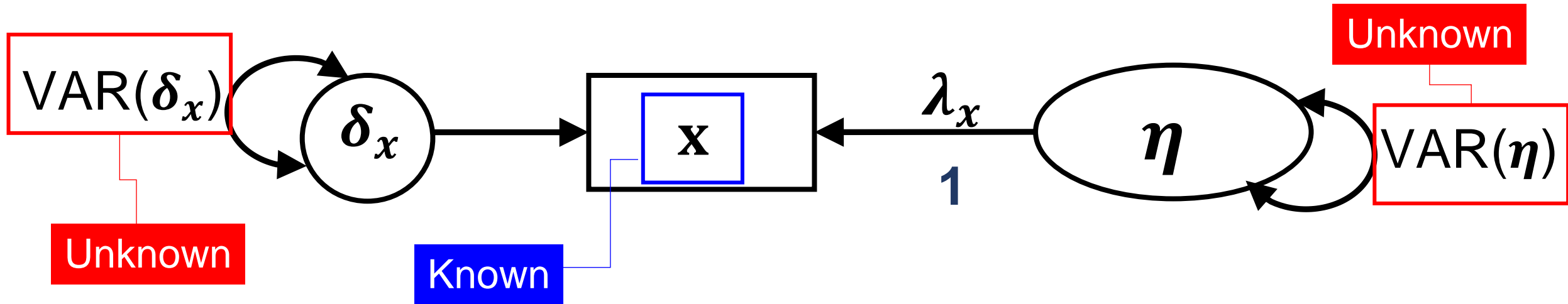
$$t \leq t_{\max}$$
$$t_{\max} = \frac{s(s+1)}{2} = 1$$

$s = 1$ known

$t = 2$ unknowns

What is Latent Variable?

New



Rules for LV models:

- Scaling of LV
- Non-negative DF

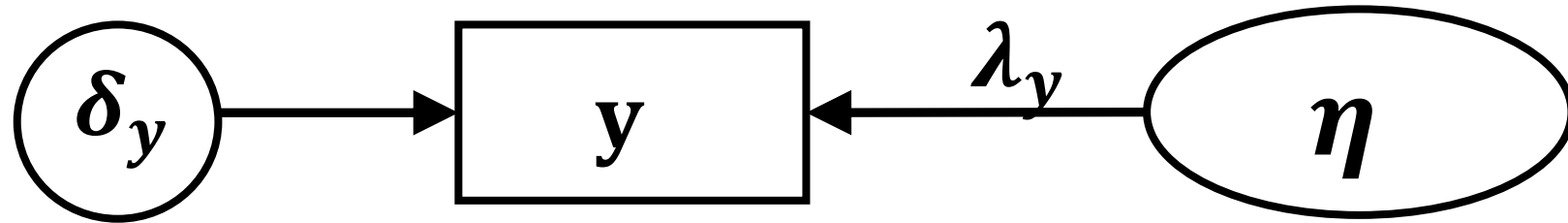


We need at least:

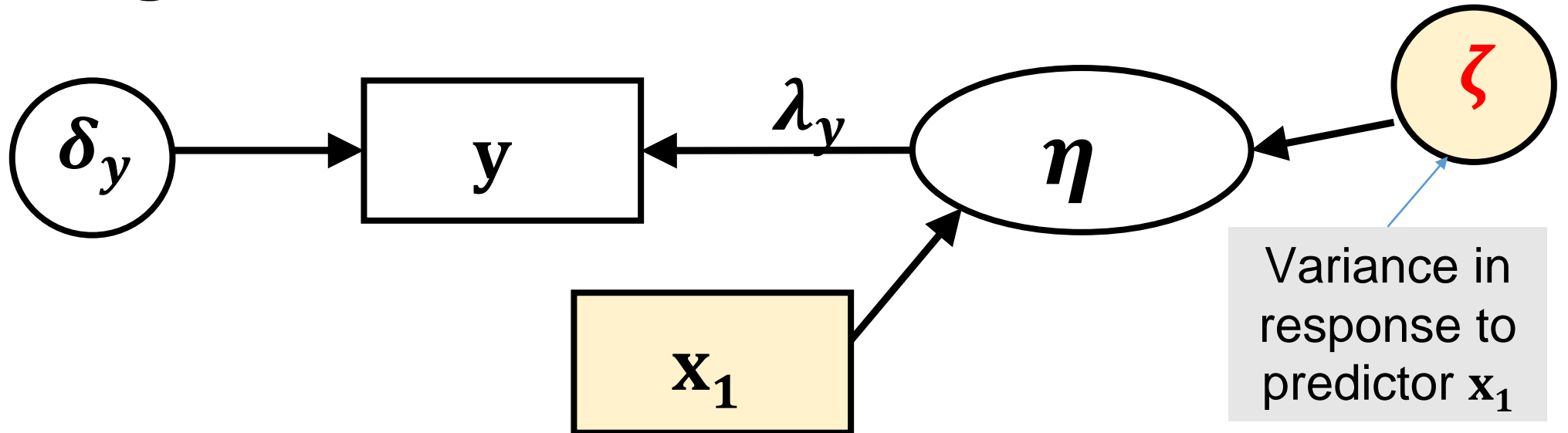
- 3 indicators for a single LV
- 2 indicators per LV for models with multiple (correlated) LVs

What is Latent Variable?

Latent **Exogenous** Variable



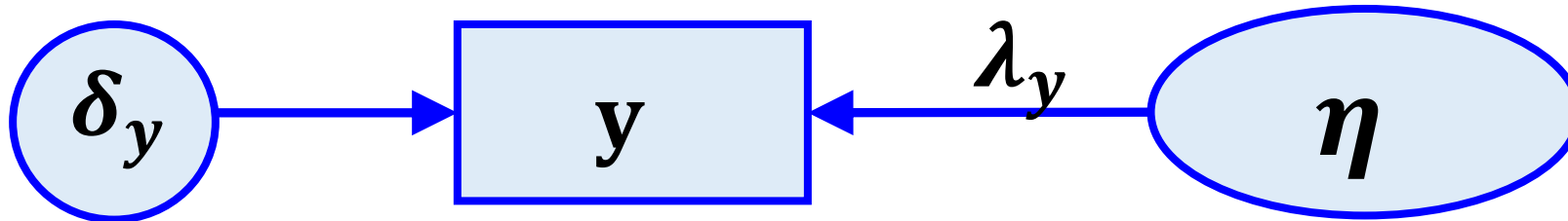
Latent **Endogenous** Variable



What is Latent Variable?

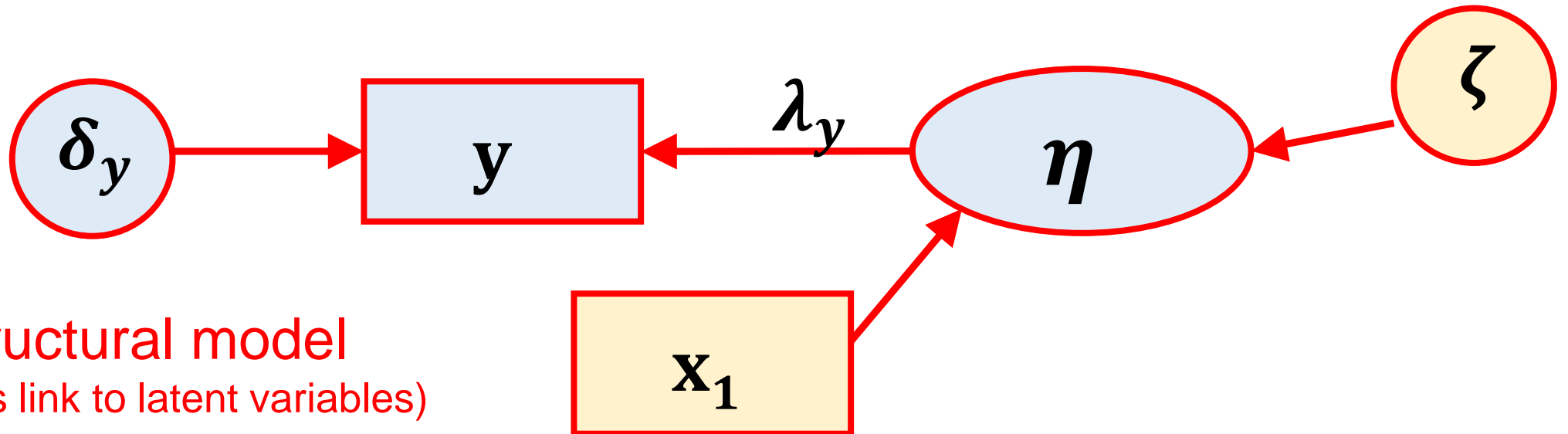
Measurement model

(solely relates indicators to latent variables)

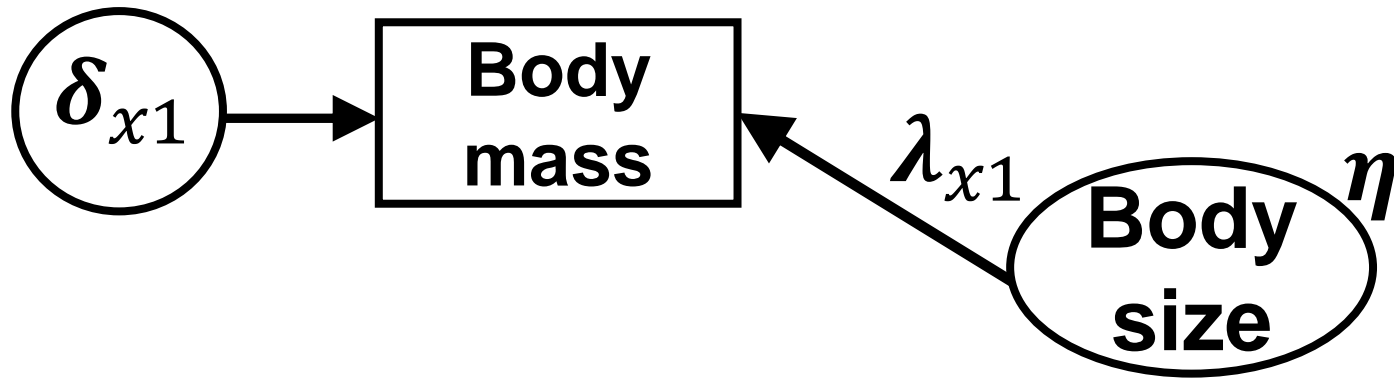


Structural model

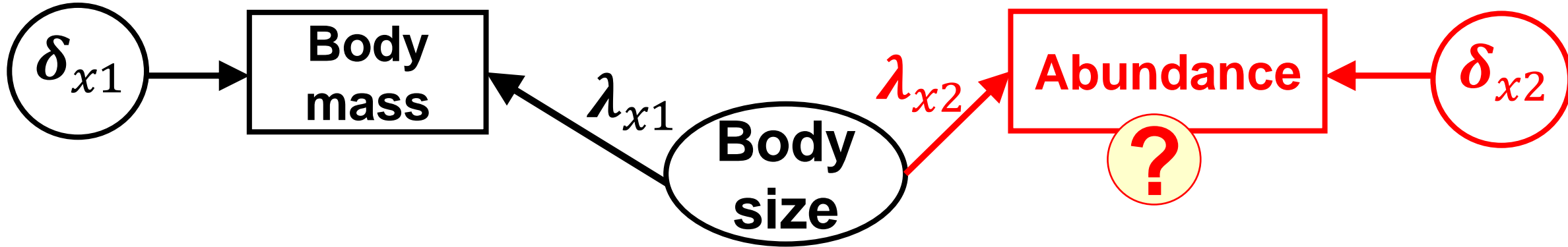
(has link to latent variables)



Latent Variables



Latent Variables



- Be sure that the latent variable reflects the actual properties captured by the indicator variables!

Why use Latent Variables?

- Allows estimating complex and **multifaceted concepts**
- **Reduces random error** in construct (latent variable)

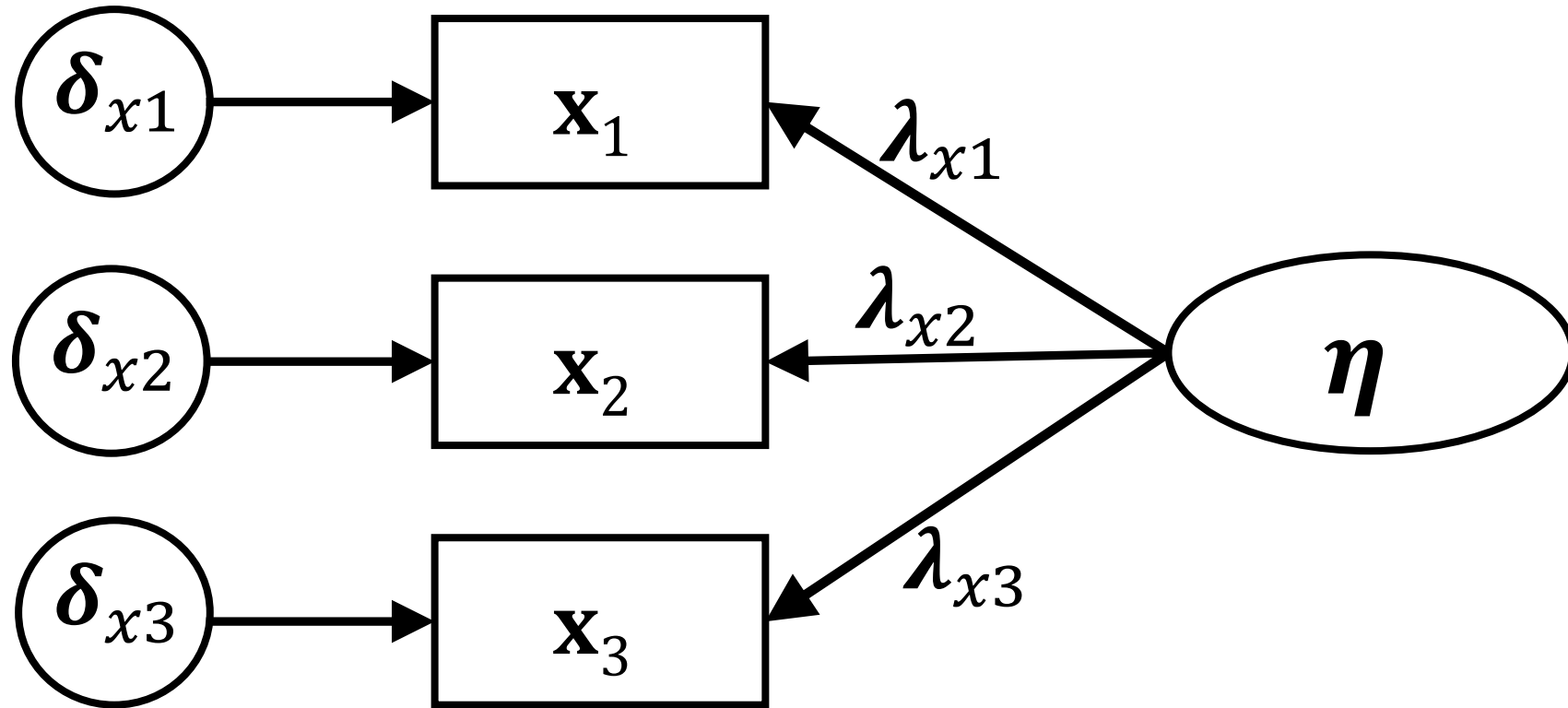
random error in dependent variables
→ less precisely measured estimates

random error in independent variables
→ underestimated regression coefficients

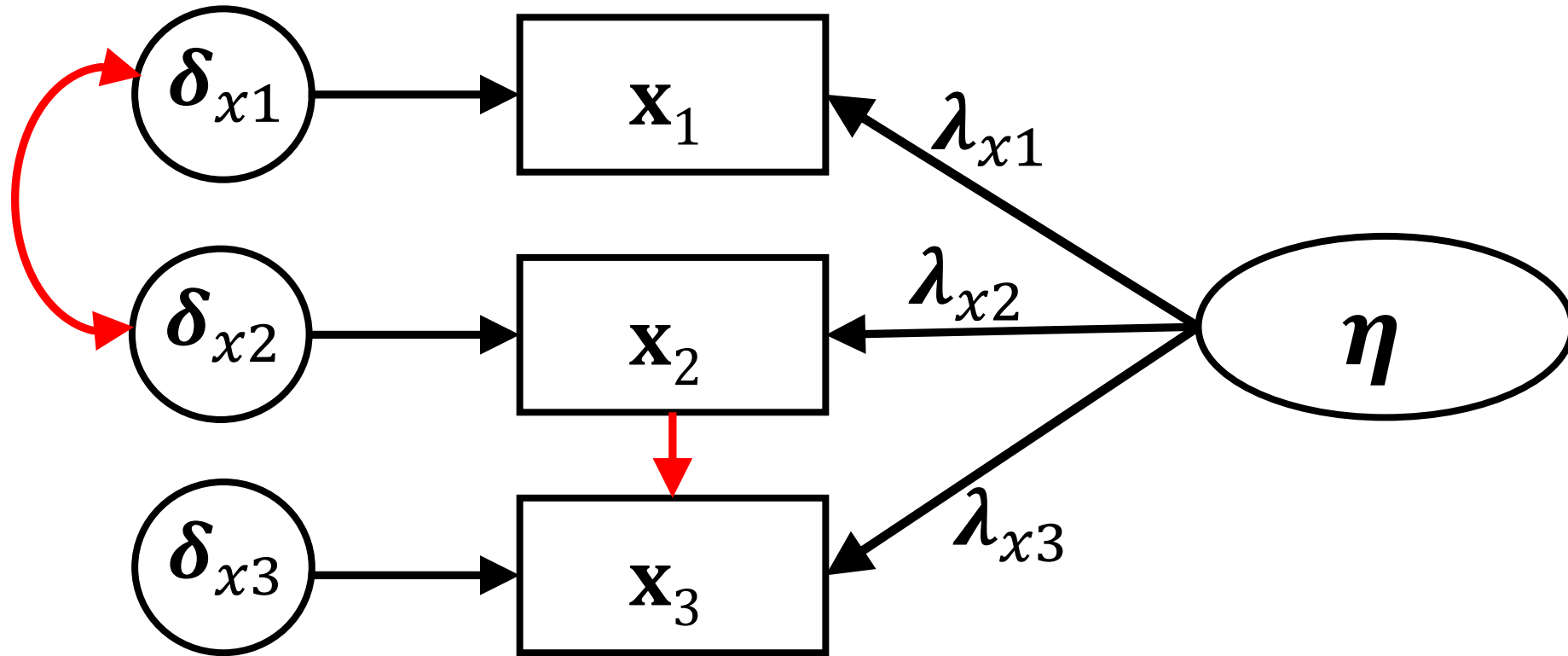
Latent Variables in SEM

- What are Latent Variables? Why to use them?
- **Multi-indicator Latent Variables**
- Fitting Latent Variables
(Confirmatory Factor Analysis)

Multi-indicator Latent Variables

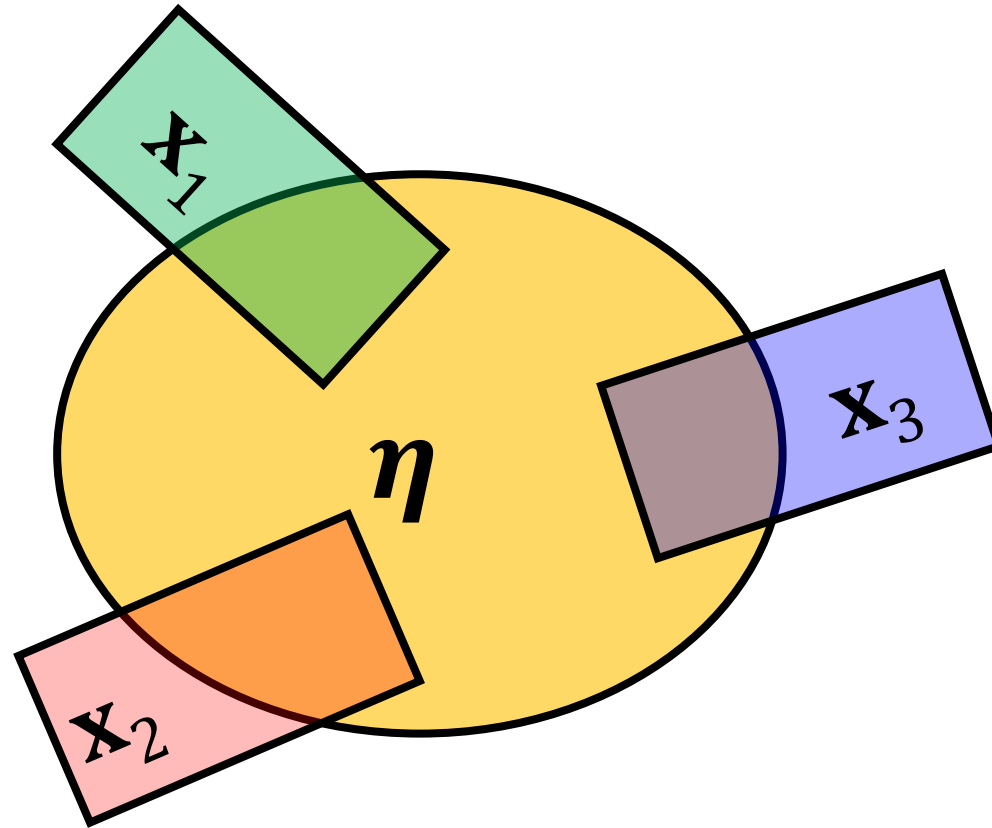


Multi-indicator Latent Variables



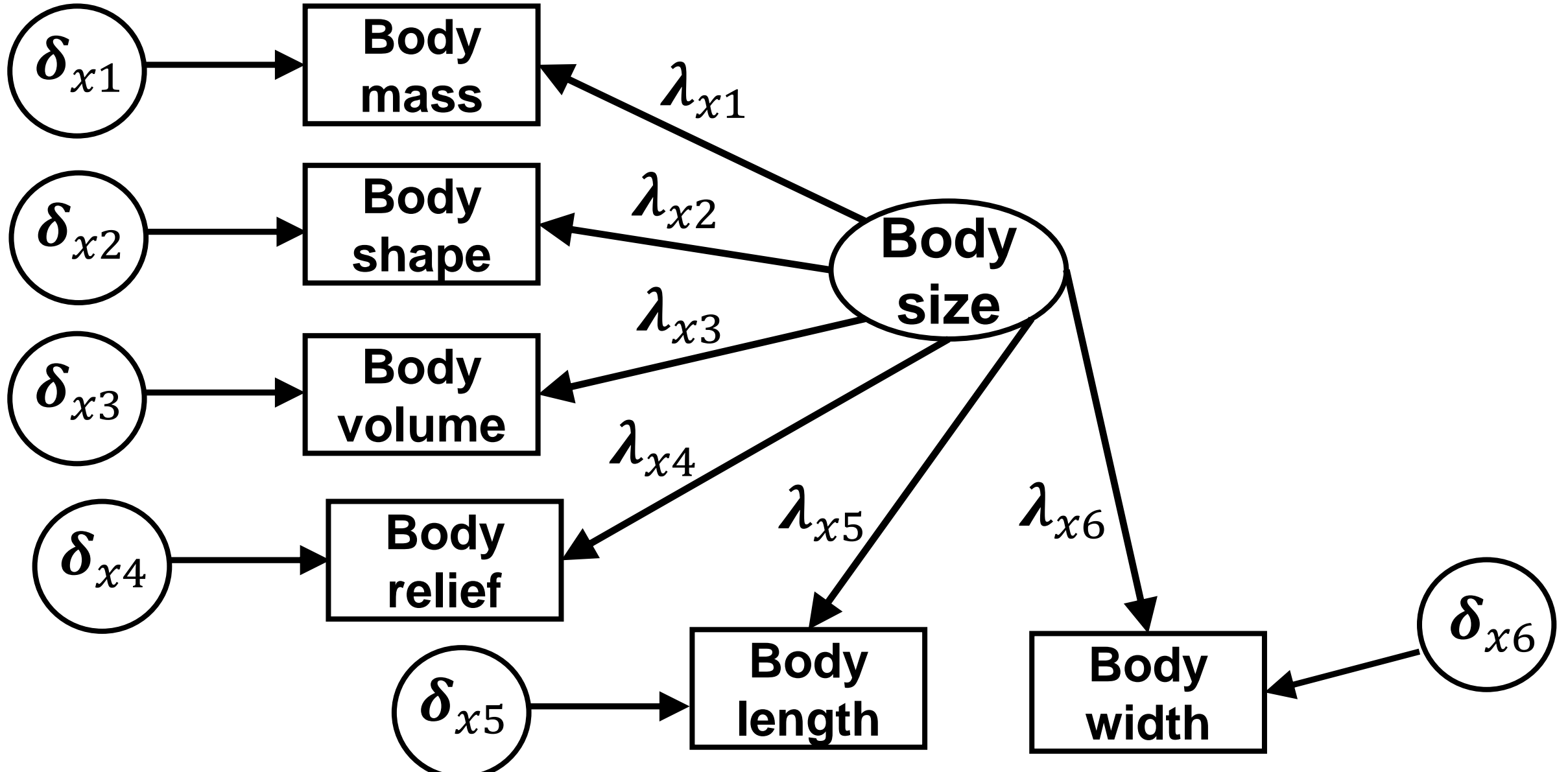
- Indicators may have causal links
- Indicators may covary for other reasons

Multi-indicator Latent Variables



Latent variable η represents shared information of observed indicators x

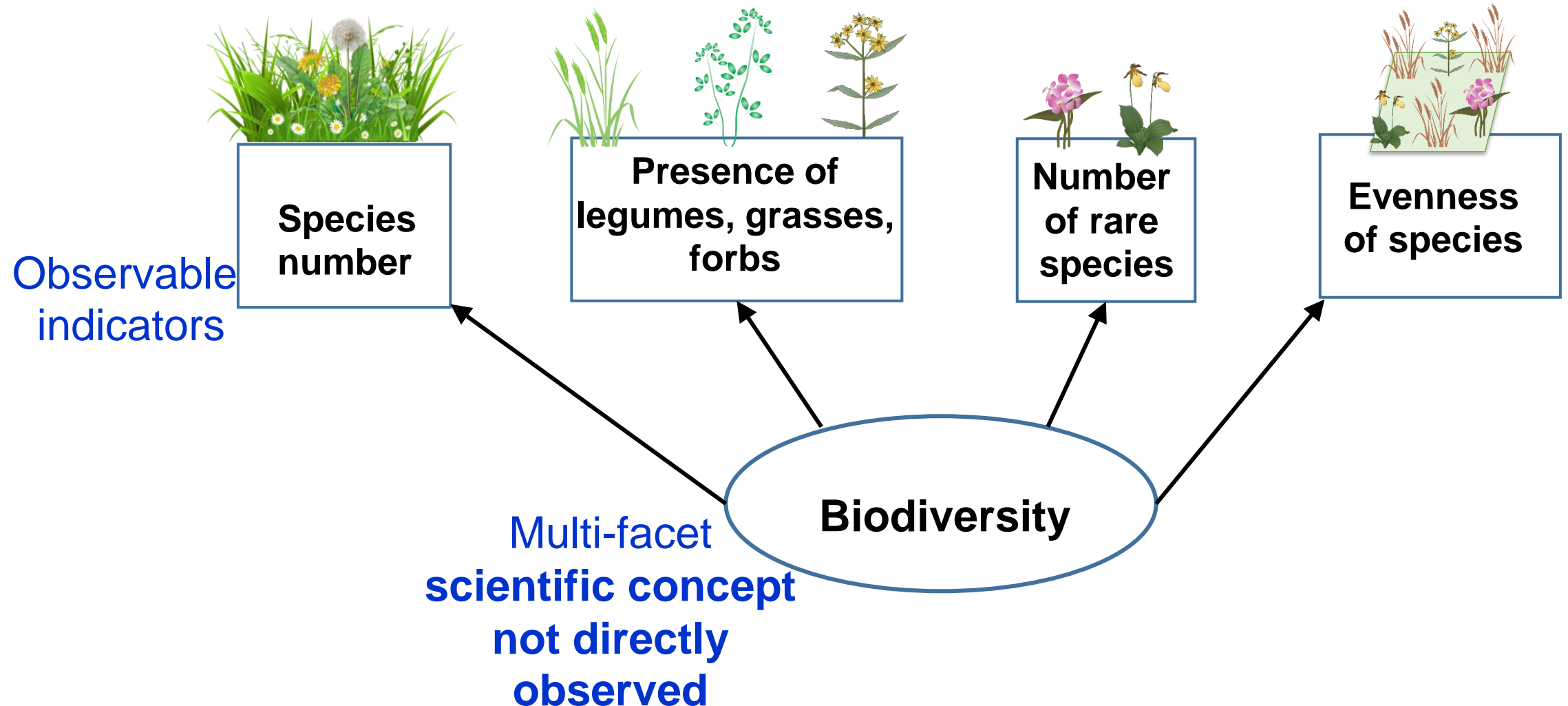
Multi-indicator Latent Variables



What is Latent Variable?

Example

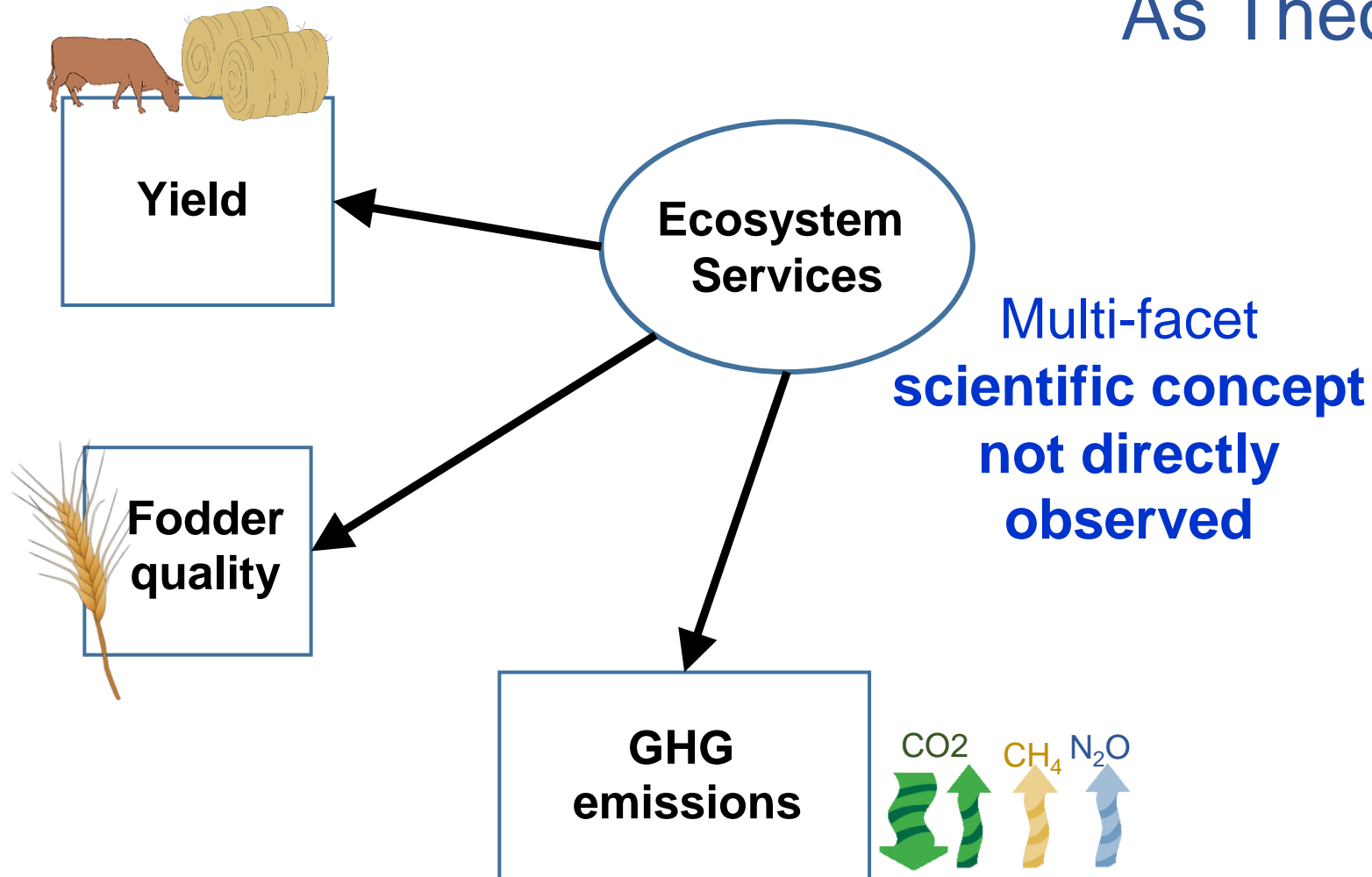
As Theoretical Constructs



What is Latent Variable?

Example

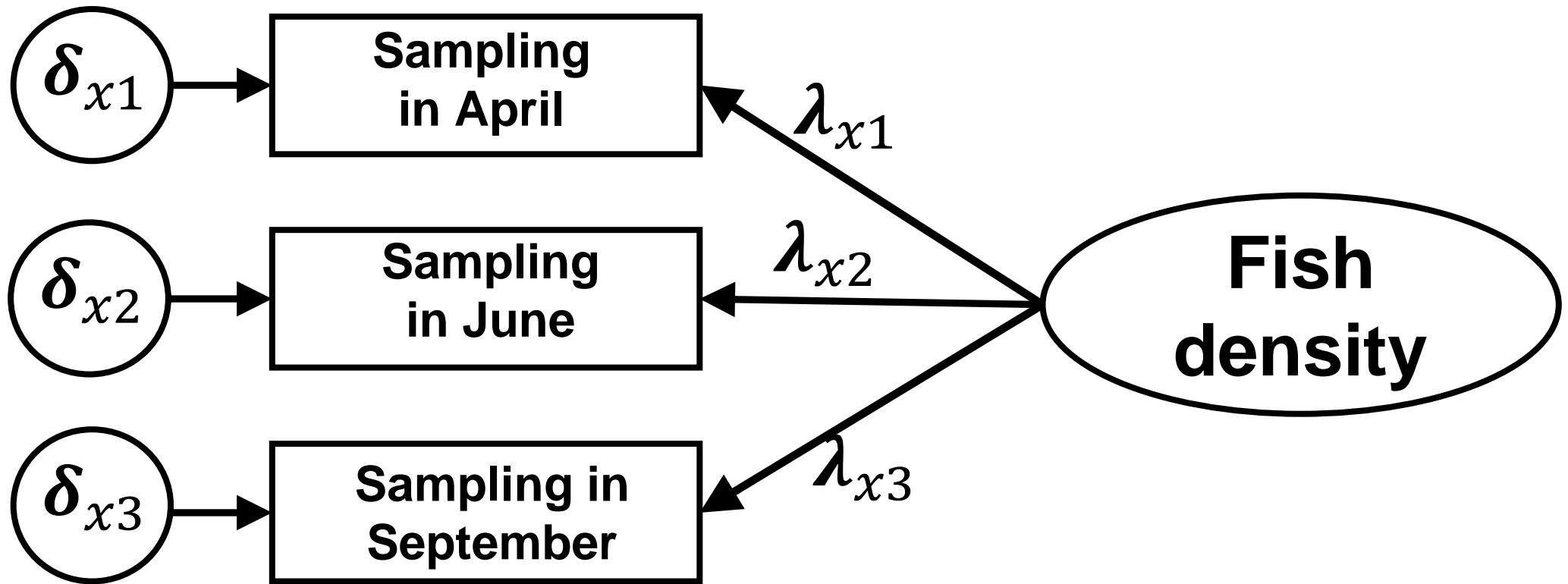
As Theoretical Constructs



Multi-indicator Latent Variables

Example

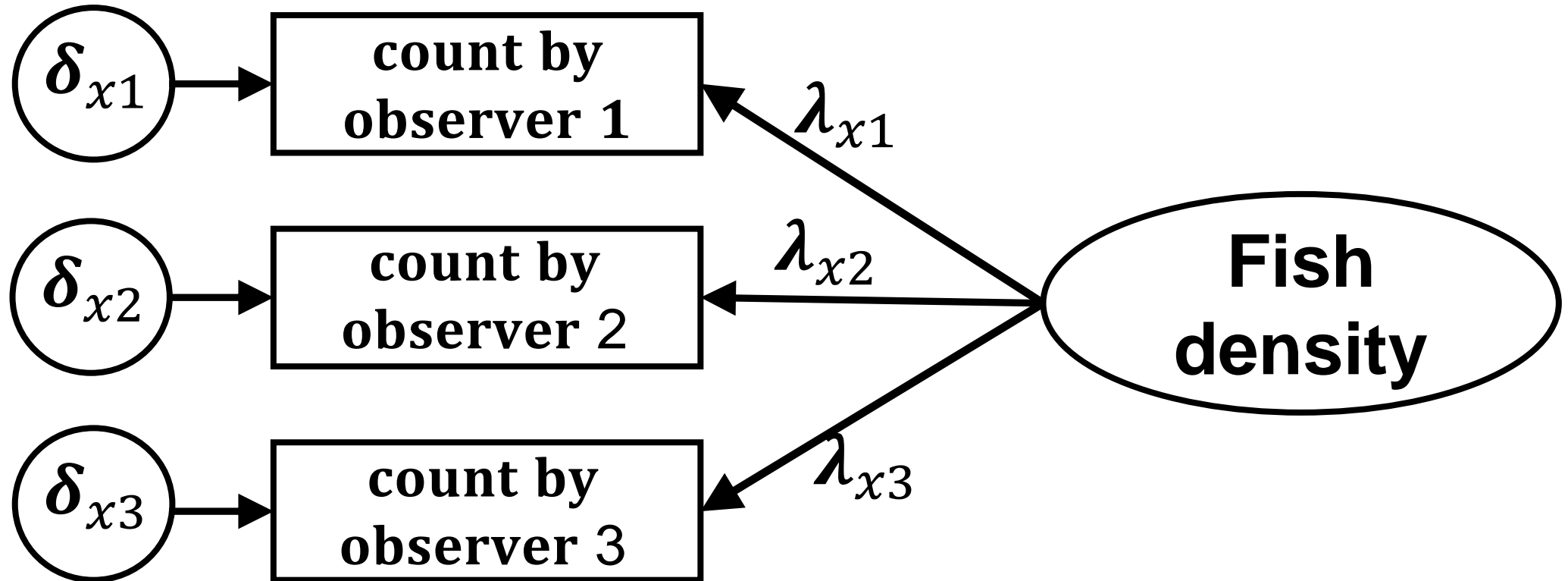
Repeated Measurements



Multi-indicator Latent Variables

Example

Multi-sampling



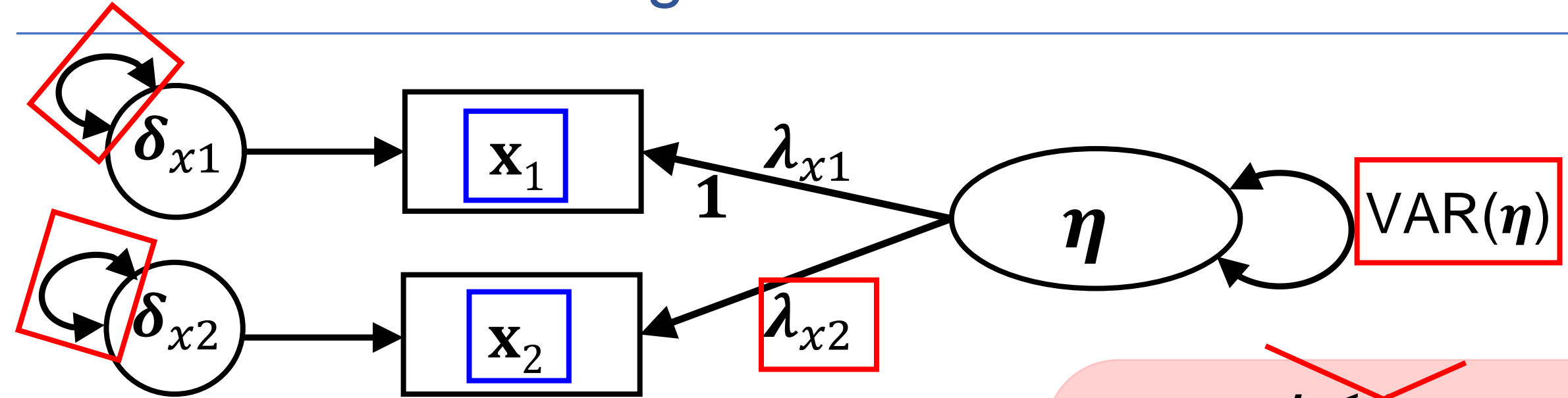
Why use Latent Variables with Multiple Indicators?

- Allows estimating complex and multifaceted concepts
- Reduces random error in construct (latent variable)
- Better accuracy in measurement of relationships due to shared variation between observed indicators.

Latent Variables in SEM

- What are Latent Variables? Why to use them?
- Multi-indicator Latent Variables
- **Fitting Latent Variables**
(Confirmatory Factor Analysis)

Fitting Latent Variables



Rules for LV models:

- Scaling of LV
- Non-negative DF

$$\text{DF} = -1$$

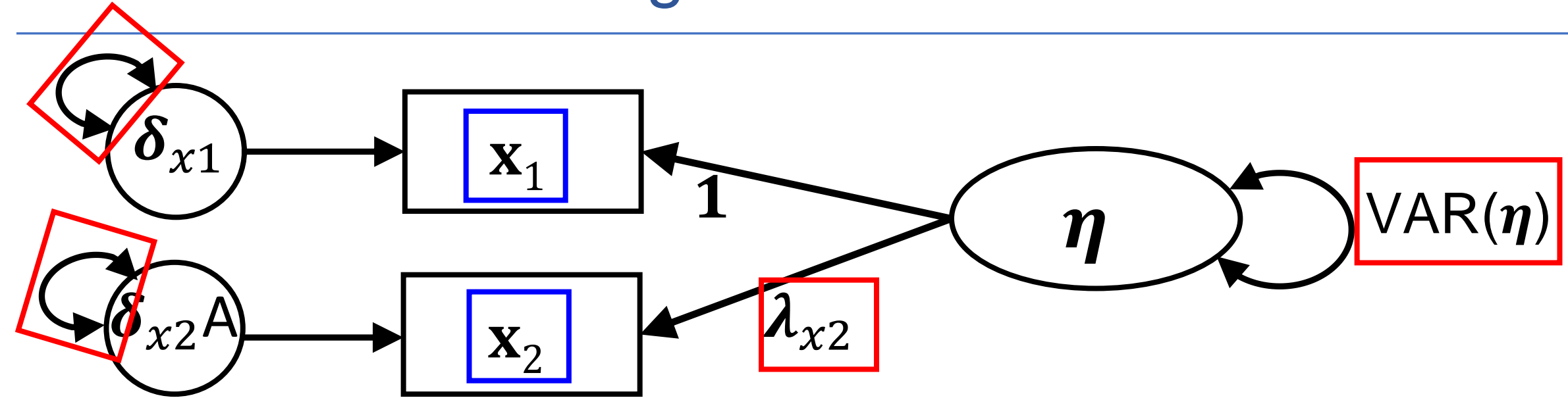
~~$t \leq t_{max}$~~

$$t_{max} = \frac{s(s+1)}{2} = 3$$

$s = 2$ knowns

$t = 4$ unknowns

Fitting Latent Variables



Rules for LV models:

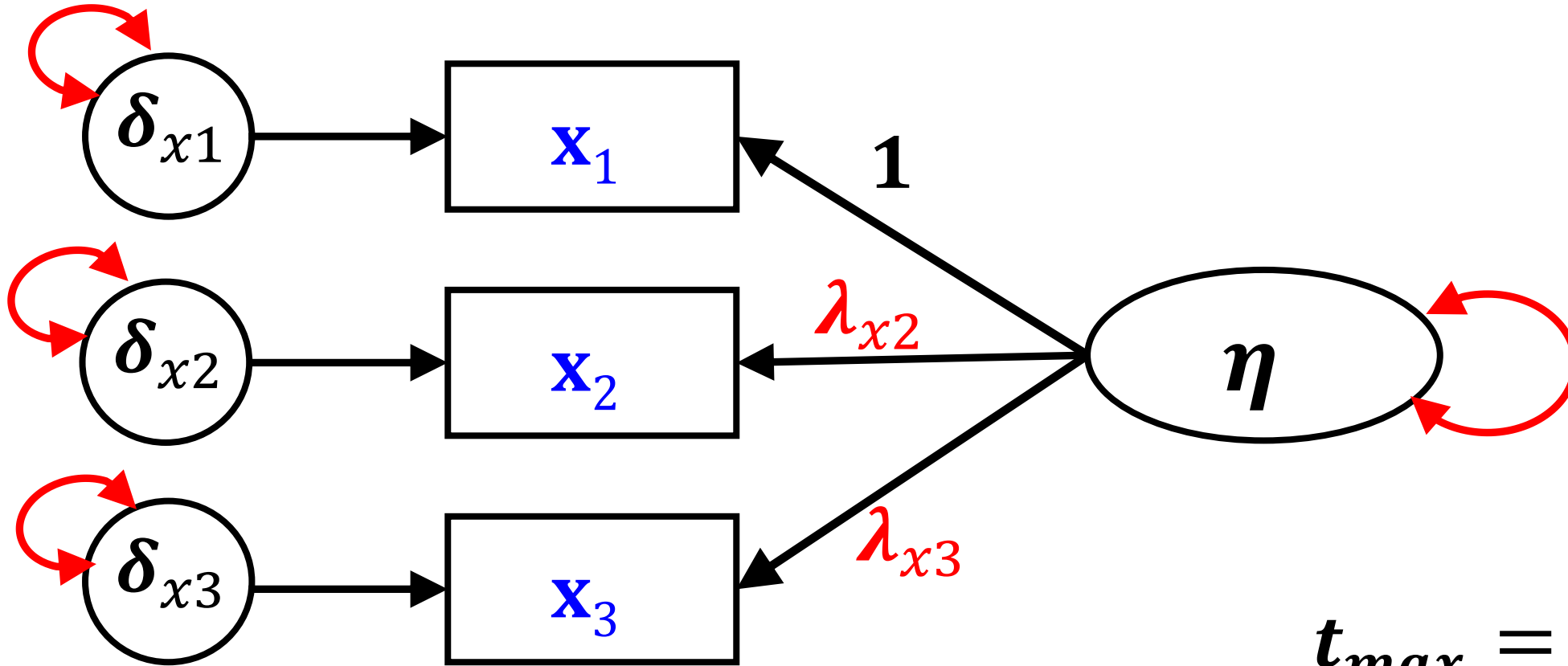
- Scaling of LV
- Non-negative DF



We need at least:

- 3 indicators for a single LV
- 2 indicators per LV for models with multiple (correlated) LVs

Fitting Latent Variables



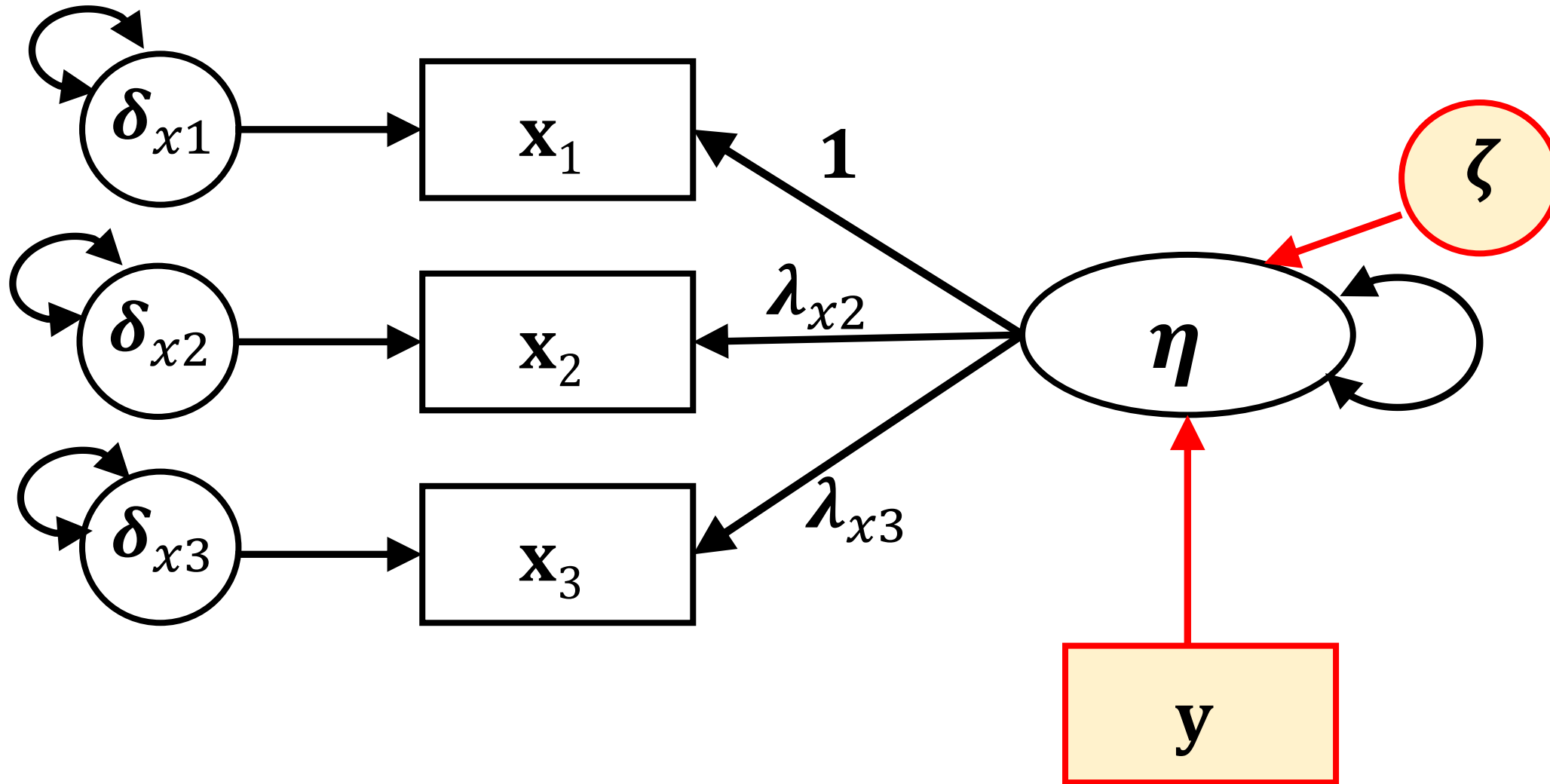
$$t_{max} = \frac{s(s+1)}{2} = 6$$

$s = 3$ knowns

$t = 6$ unknowns

$$\text{DF} = t_{max} - t = 0$$

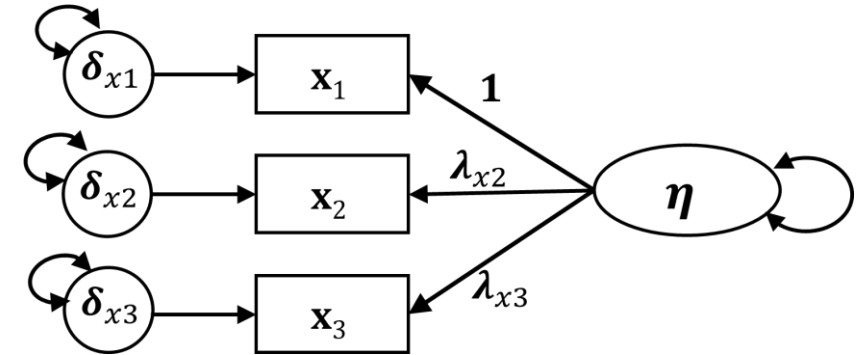
Fitting Latent Variables



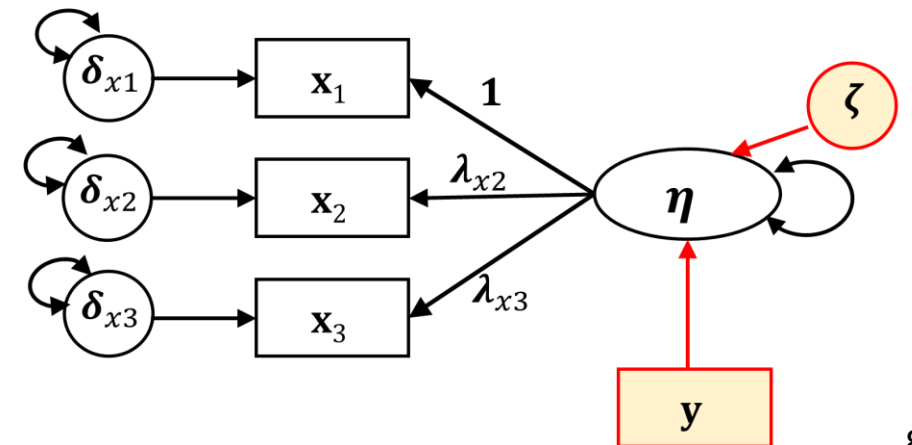
Fitting Latent Variables

1) Evaluate the latent relationships among variables (**Confirmatory Factor Analysis**).

- Do our indicators make a Good Latent Variable?



2) Use Latent Variables as a Response or a Predictor



Population-based ecological restoration

Aim: understand the performance of transplanted plants as a function of their dissimilarity to local conditions

Sabine National
Wildlife
Refuge, Louisiana,
USA



```
# Read and check the data
```

```
travis <- read.csv(" Travis_data.csv")
```

Travis, S. E., & Grace, J. B. (2010). Predicting performance for ecological restoration: a case study using *Spartina alterniflora*. *Ecological Applications*, 20(1), 192-204.

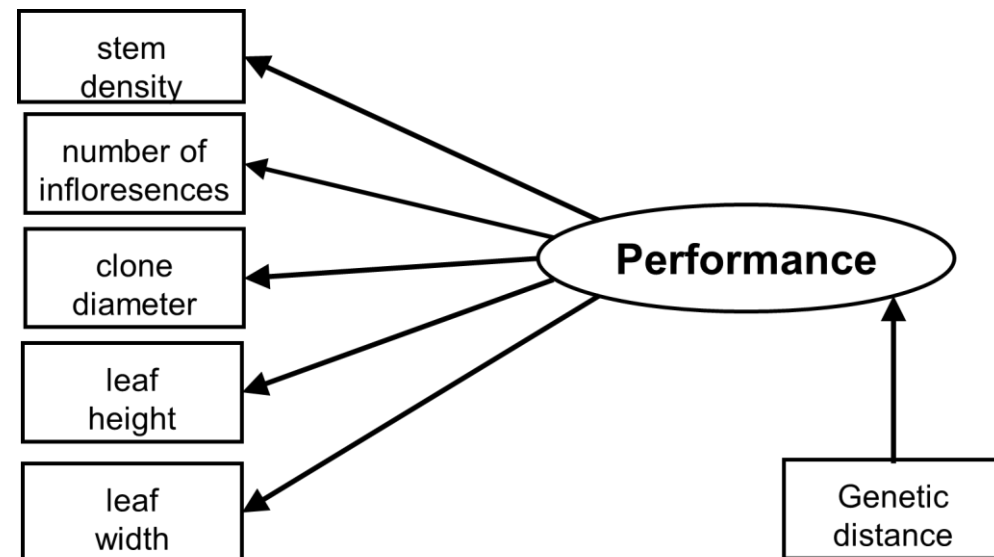
Confirmatory Factor Analysis

Exercise

Sabine National Wildlife
Refuge, Louisiana, USA

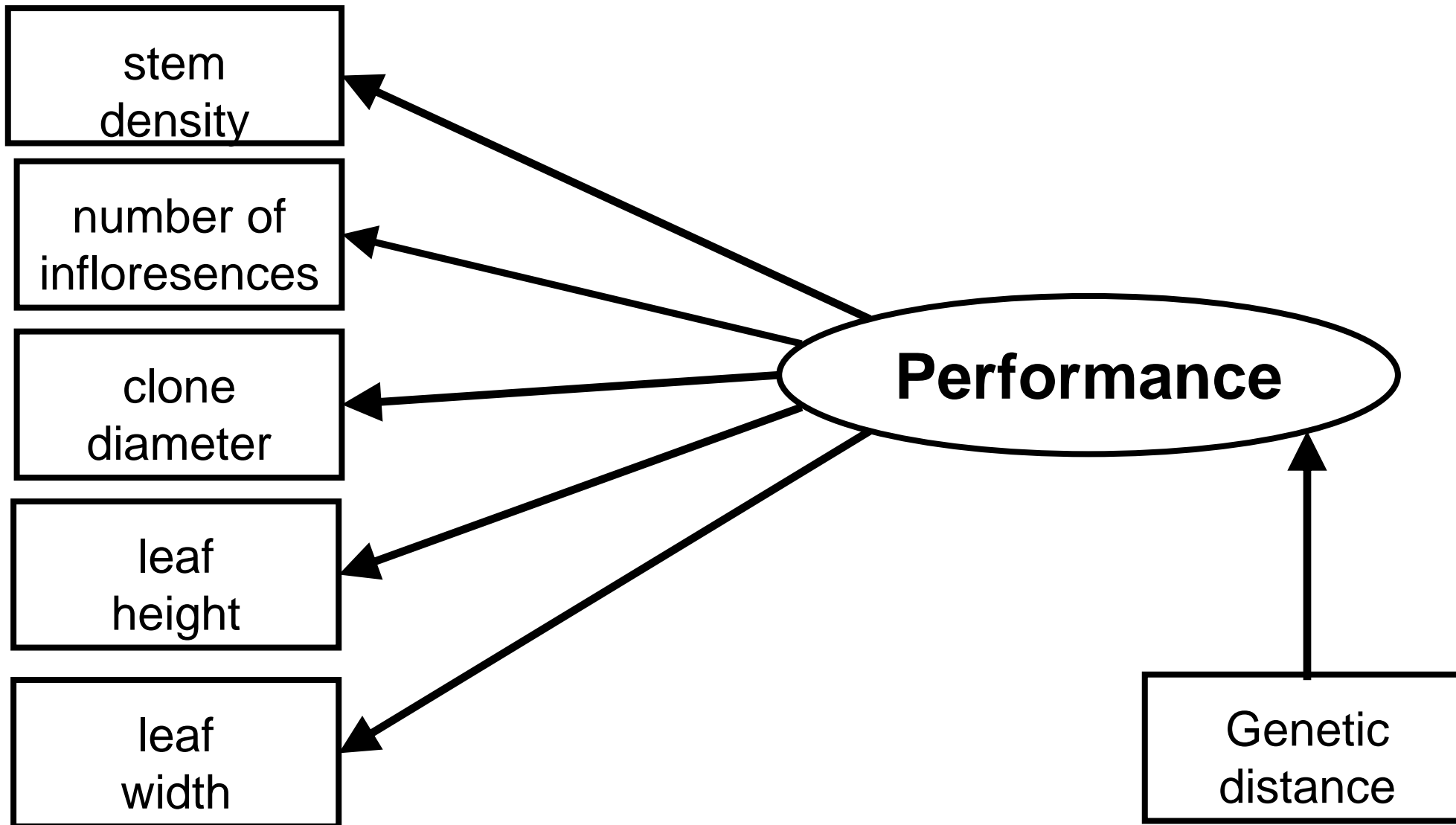


- Collected individuals of the salt marsh plant *Spartina alterniflora* eight clones each from 23 populations
- Transplanted individuals and measured their performance relative to local populations.
- Performance was approximated with stem density, the number of inflorescences, clone diameter, leaf height, and leaf width

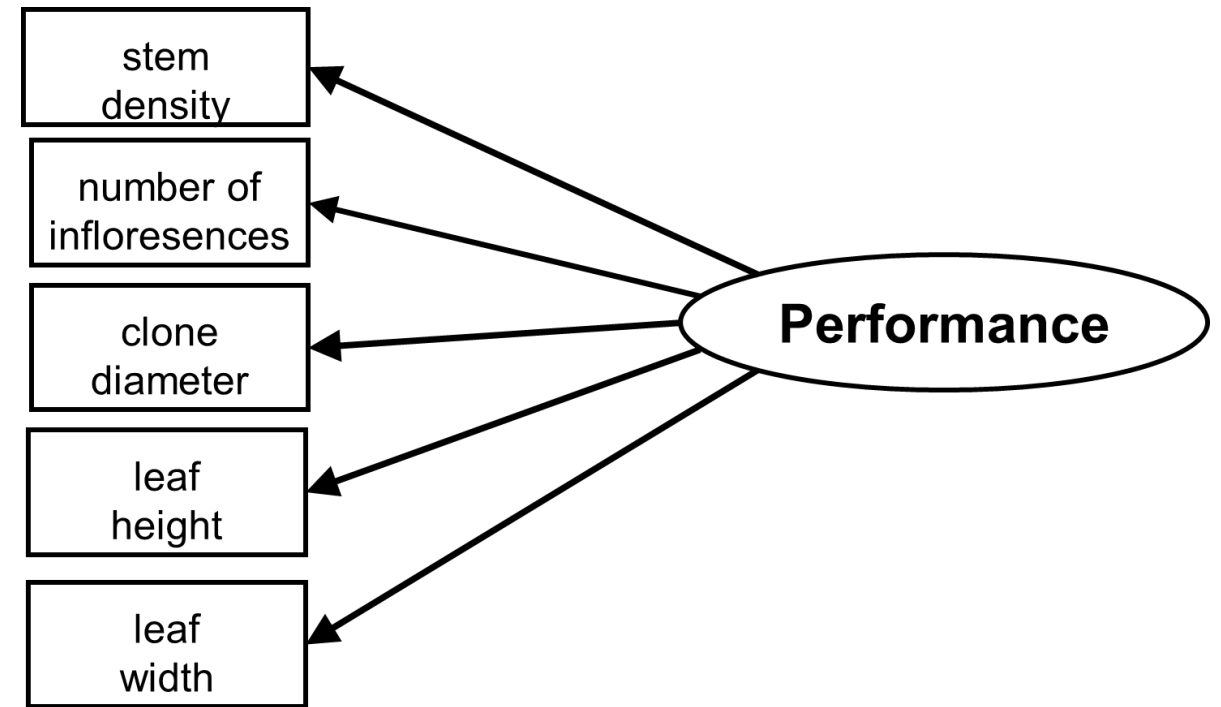


Confirmatory Factor Analysis

Exercise



- 1) Evaluate the latent relationships among variables
(**Confirmatory Factor Analysis**).
- Do our indicators make a Good Latent Variable?



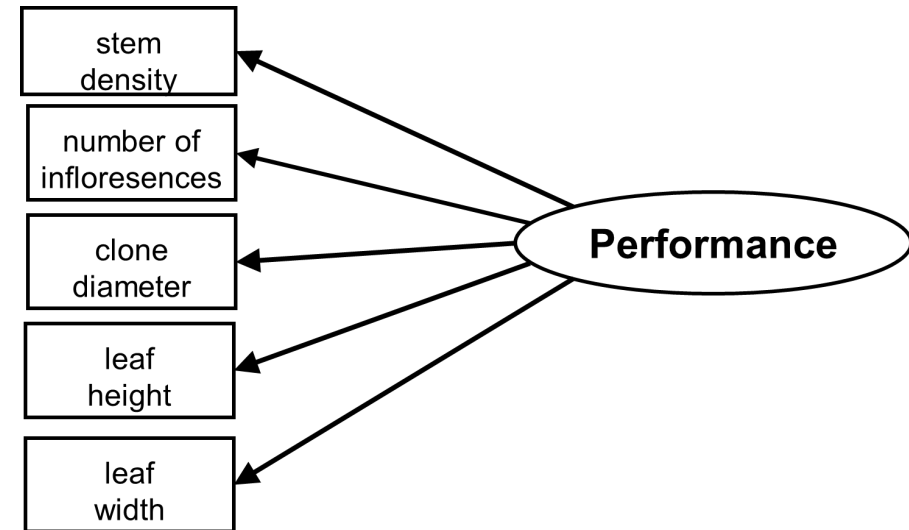
A first step is to analyze the “measurement model” using CFA.

Confirmatory Factor Analysis

Exercise

```
# Read and check the data
travis <- read_csv("Travis_data.csv")

str(travis)
# correlations
cor(travis[, 4:8])
```



```
> round(cor(travis[, 4:8]),2)
```

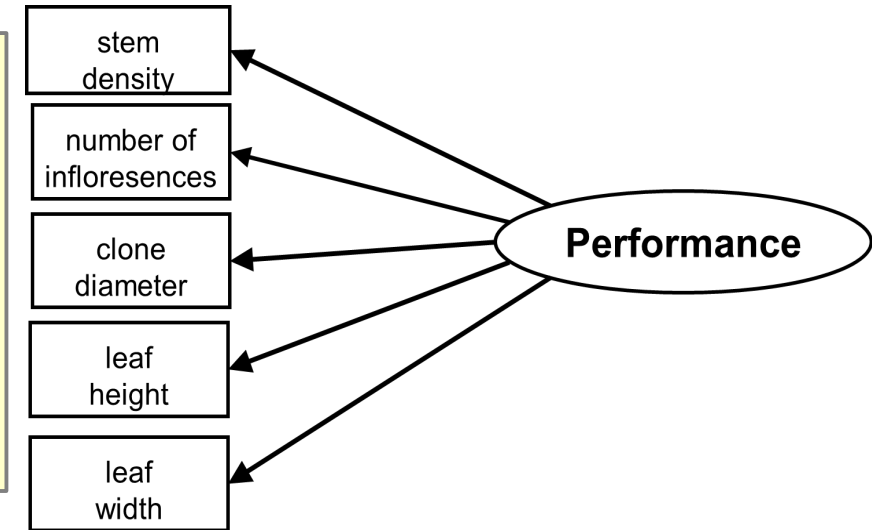
	stems	infls	clonediam	leafht	leafwidth
stems	1.00	0.83	0.93	0.73	0.65
infls	0.83	1.00	0.81	0.69	0.60
clonediam	0.93	0.81	1.00	0.77	0.73
leafht	0.73	0.69	0.77	1.00	0.97
leafwidth	0.65	0.60	0.73	0.97	1.00

Confirmatory Factor Analysis

Exercise

```
# specify the model
cfa_mod <- `
performance =~ stems + infls + clonediam + leafht + leafwidth
`

# fit the model
cfa_fit <- sem(cfa_mod, travis)
```



Warning message:

In lav_object_post_check(object) :

lavaan WARNING: some estimated ov variances are negative

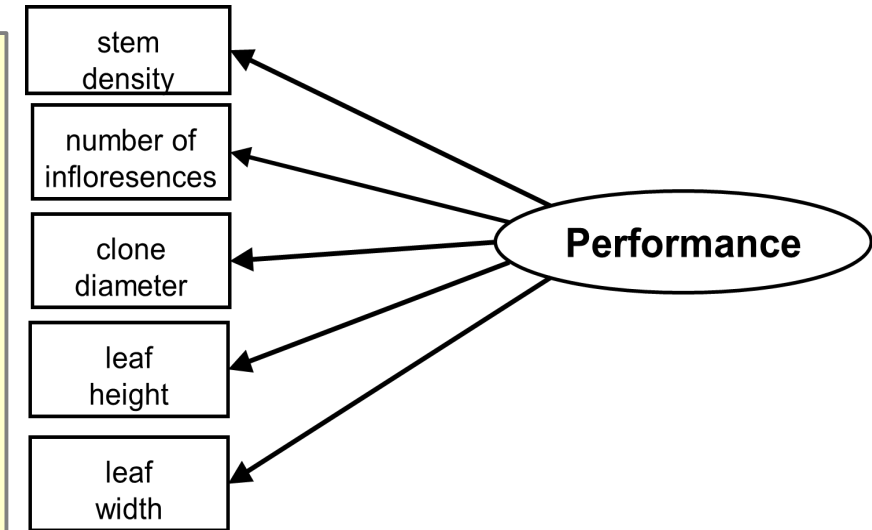
Confirmatory Factor Analysis

Exercise

```
> summary(cfa_fit)
```

```
lavaan 0.6-9 ended normally after 82 iterations
```

Estimator	ML
Optimization method	NLMINB
Number of model parameters	10
Number of observations	23
Model Test User Model:	
Test statistic	51.106
Degrees of freedom	5
P-value (Chi-square)	0.000



Confirmatory Factor Analysis

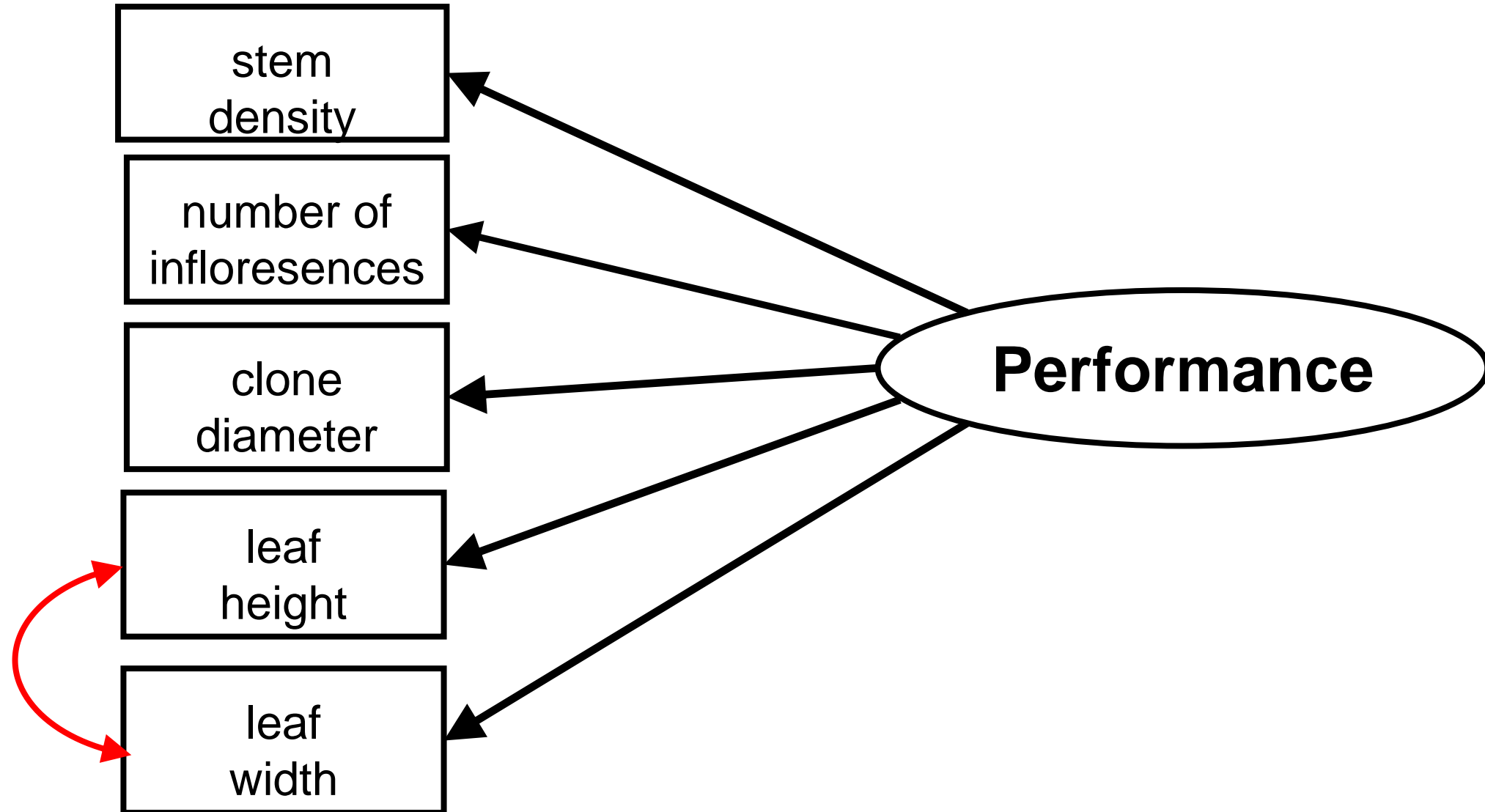
Exercise

```
> modindices(cfa_fit)
```

	lhs	op	rhs	mi	epc	sepc.lv	sepc.all	sepc.nox
12	stems	~~	infls	10.470	11.784	11.784	0.677	0.677
13	stems	~~	clonediam	17.152	112.521	112.521	0.871	0.871
14	stems	~~	leafht	0.693	-7.889	-7.889	-0.517	-0.517
15	stems	~~	leafwidth	2.214	-1.836	-1.836	-0.346	-0.346
16	infls	~~	clonediam	8.773	11.092	11.092	0.621	0.621
17	infls	~~	leafht	0.062	-0.312	-0.312	-0.148	-0.148
18	infls	~~	leafwidth	2.906	-0.281	-0.281	-0.383	-0.383
19	clonediam	~~	leafht	4.028	-21.233	-21.233	-1.357	-1.357
20	clonediam	~~	leafwidth	0.037	-0.261	-0.261	-0.048	-0.048
21	leafht	~~	leafwidth	37.862	17.177	17.177	26.752	26.752

Confirmatory Factor Analysis

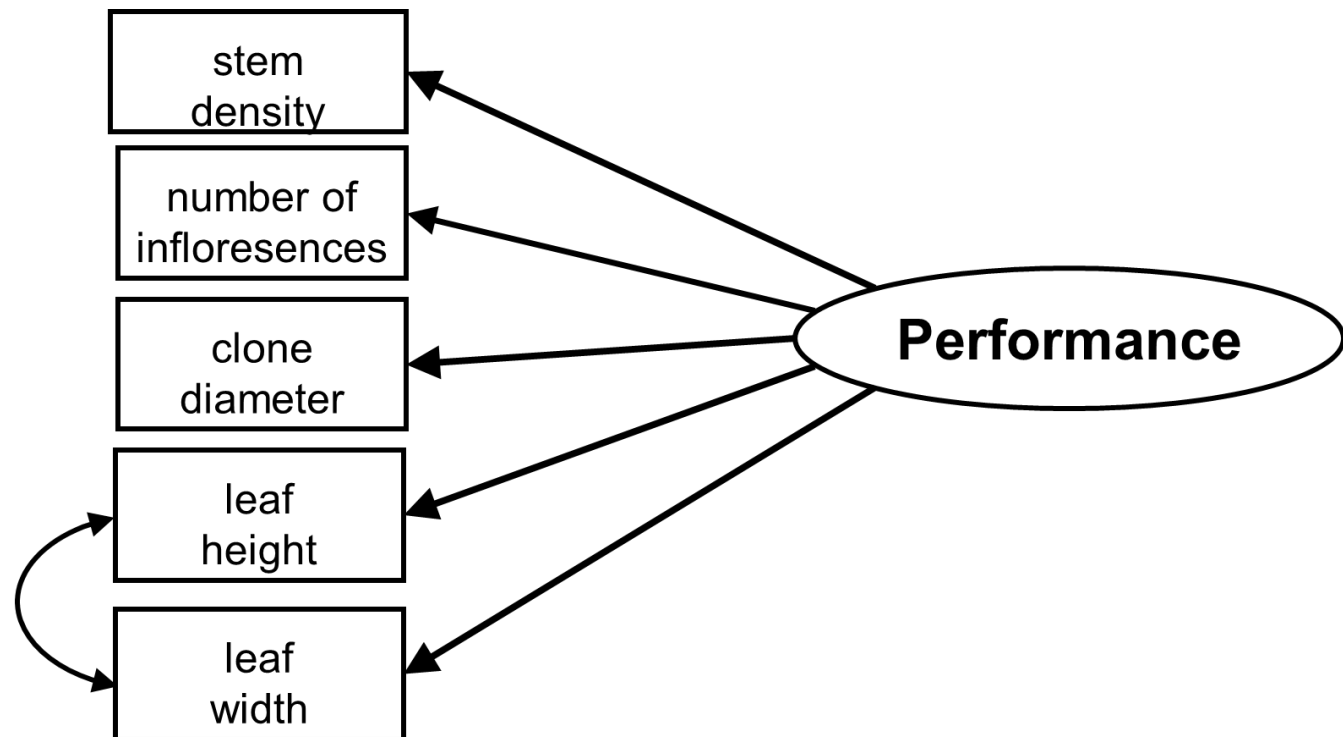
Exercise



Confirmatory Factor Analysis

Exercise

```
cfa_mod2 <- '  
performance =~ stems + infls + clonediam + leafht + leafwidth  
leafht ~~ leafwidth  
'  
  
cfa_fit2 <- sem(cfa_mod2, travis)  
summary(cfa_fit2)
```



Confirmatory Factor Analysis

Exercise

Estimator	ML
Optimization method	NLMINB
Number of model parameters	11
Number of observations	23
Model Test User Model:	
Test statistic	7.410
Degrees of freedom	4
P-value (Chi-square)	0.116

Confirmatory Factor Analysis

Exercise

...

Latent Variables:

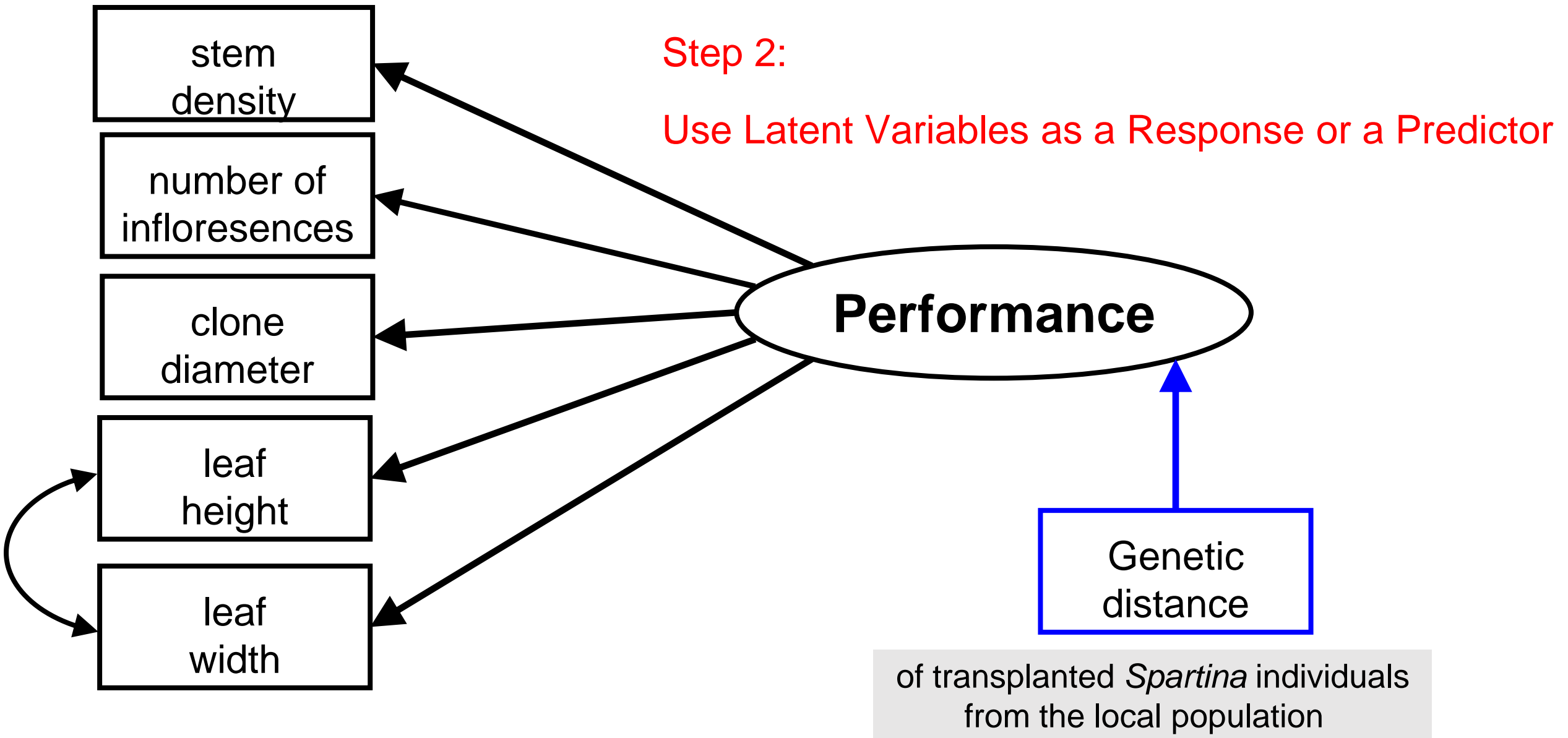
	Estimate	Std.Err	z-value	P(> z)
performance =~				
stems	1.000			
infls	0.117	0.016	7.173	0.000
clonediam	1.086	0.096	11.319	0.000
leafht	0.697	0.127	5.509	0.000
leafwidth	0.082	0.018	4.529	0.000

Covariances:

	Estimate	Std.Err	z-value	P(> z)
.leafht ~~				
.leafwidth	10.831	3.432	3.156	0.002

CFA as a part of structural model

Exercise



CFA as a part of structural model

Exercise

```
SEM_latent_mod <- `
  # latent
performance =~ stems + infls + clonedia + leafht + leafwdth

  # structural paths
performance ~ geneticdist

  # correlated errors
leafht ~~ leafwdth
`

SEM_latent_fit <- sem(SEM_latent_mod , travis)

summary(SEM_latent_fit, standardize = T, rsq = T, fit.measures=T)
```

CFA as a part of structural model

Exercise

Estimator

ML

Optimization method

NLMINB

Number of model parameters

12

Number of observations

23

Model Test User Model:

Test statistic

12.237

Degrees of freedom

8

P-value (Chi-square)

0.141

CFA as a part of structural model

Exercise

Latent Variables:

	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
performance =~						
stems	1.000				15.555	0.962
infls	0.117	0.017	6.929	0.000	1.822	0.853
clonediam	1.106	0.096	11.508	0.000	17.199	0.969
leafht	0.711	0.127	5.601	0.000	11.066	0.785
leafwidth	0.084	0.018	4.650	0.000	1.308	0.718

Regressions:

	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
performance ~						
geneticdist	-51.673	11.365	-4.547	0.000	-3.322	-0.708

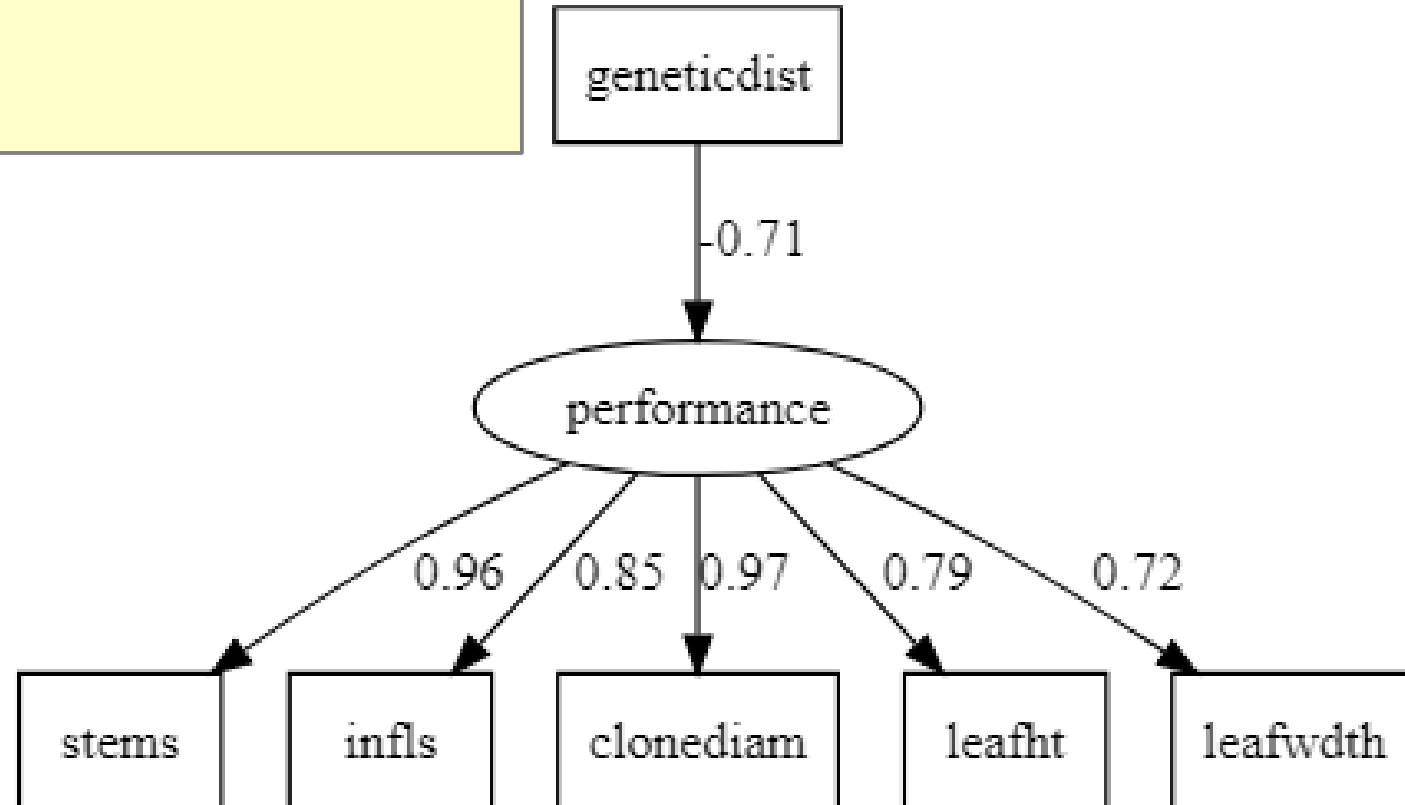
Covariances:

	Estimate	Std.Err	z-value	P(> z)	Std.lv	Std.all
.leafht ~~						
.leafwidth	10.416	3.312	3.145	0.002	10.416	0.940

CFA as a part of structural model

Exercise

```
library(lavaanPlot)
lavaanPlot(model = SEM_latent_fit,
  coefs = TRUE, stand=TRUE,
  # graph_options = list(layout = "circo"),
  # stars = 'regress', # shows stars for regr coef
  digits = 2)
```

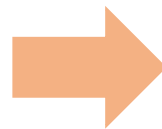


Day 7 Task 2

Human impact on macroinvertebrate body size in ponds



**Human
Impact Intensity**



Macroinvertebrate body size



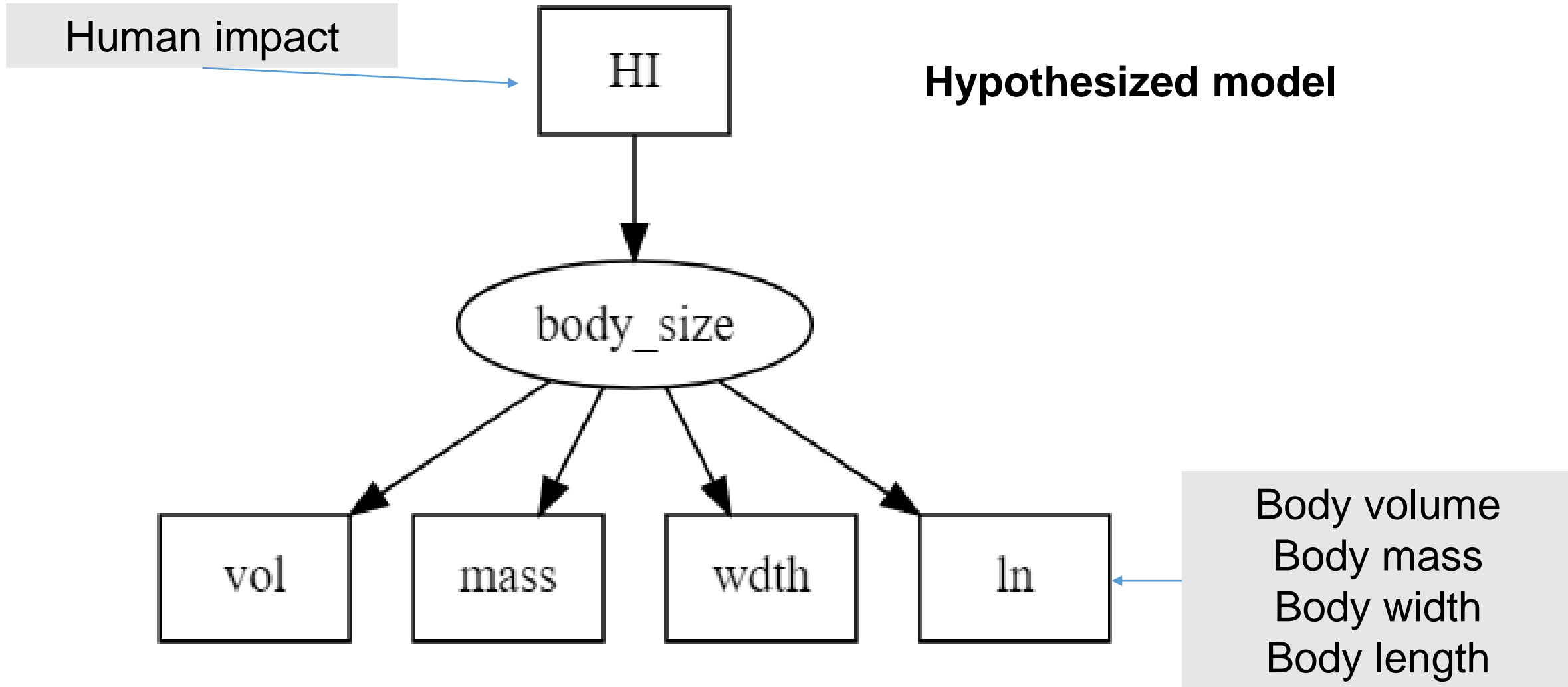
Body size traits

Body mass
Body volume
Body length
Body width

```
# Read and check the data  
read.csv(" Bodysize_data.csv")
```

Day 5 Task 2

Human impact on
macroinvertebrate body size in ponds



1. Perform the confirmatory factor analysis for the latent variable “body size”
2. Use the results from step 1 and perform the SEM by adding human impact variable
3. Fill in Standardized Coefficients and R^2 for the model, add the fit indices
4. Think about how to interpret the results