Lactate transport inhibition therapeutically reprograms fibroblast metabolism in experimental pulmonary fibrosis

David R. Ziehr1,2,3, Nathan M. Krah4, Kevin Leahy2, K. Mark Parnell5, Jack Varon2,3, Rebecca M. Baron2,3, Nancy J. Philp6, Lida P. Hariri3,7, Edy Y. Kim2,3, Rachel S. Knipe1,3, Jared Rutter8,9, and William M. Oldham2,3,‡

1 Department of Medicine, Massachusetts General Hospital, Boston, MA  
2 Department of Medicine, Brigham and Women’s Hospital and Harvard Medical School, Boston, MA  
3 Department of Medicine, Harvard Medical School, Boston, MA  
4 Department of Human Genetics, University of Utah, Salt Lake City, UT  
5 Vettore Biosciences, San Francisco, CA  
6 Department of Pathology, Anatomy & Cell Biology, Thomas Jefferson University, Philadelphia, PA  
7 Department of Pathology, Massachusetts General Hospital, Boston, MA  
8 Department of Biochemistry, University of Utah, Salt Lake City, UT  
9 Howard Hughes Medical Institute, University of Utah School of Medicine, Salt Lake City, UT

‡ Correspondence: [William M. Oldham <woldham@bwh.harvard.edu>](mailto:woldham@bwh.harvard.edu)

# Abstract

# Keywords

# Introduction

Idiopathic pulmonary fibrosis (IPF) is a chronic and progressive lung disease with high mortality and limited therapeutic options. IPF affects approximately 150,000 patients in the U.S. with a median survival of 3-5 years (1–3). Currently approved pharmacotherapies for IPF are limited to the antifibrotics pirfenidone and nintedanib that slow, but do not stop, disease progression (4, 5), leaving lung transplantation as the only option available to eligible patients with progressive disease. The limited efficacy of antifibrotic therapies emphasizes the need for novel therapeutic approaches targeting different features of IPF pathobiology.

Accumulating evidence suggests that metabolic reprogramming may be one such therapeutic vulnerability in IPF (6, 7). Lung fibrosis is driven by the excessive deposition of extracellular matrix by myofibroblasts (3). Fundamental changes in myofibroblast metabolism support myofibroblast differentiation and extracellular matrix production (8–12). In particular, increased glycolysis and lactate production have been observed in IPF myofibroblasts *ex vivo* and following TGF-β1 (TGFβ)-induced myofibroblast differentiation *in vitro* (8, 9, 13, 14). These metabolic changes are critical for fibrogenesis, as small molecule inhibitors of glucose uptake, glycolysis, and lactate fermentation prevent myofibroblast differentiation *in vitro* and attenuate pulmonary fibrosis in animal models (8–10, 13, 15, 16). Unfortunately, low target affinities, poor specificity, narrow therapeutic indices, and common genetic resistance have all hampered the translation of these investigational compounds for clinical use (17–19). Moreover, the molecular mechanisms by which these metabolic inhibitors attenuate the myofibroblast differentiation transcriptional program remain unclear. In order to leverage metabolic therapies for IPF, more targeted and better characterized drugs must be developed.

Toward this end, we sought to characterize the effects of a novel metabolic approach - inhibition of lactate transport - on myofibroblast differentiation and experimental pulmonary fibrosis. Lactate must be secreted to sustain glycolysis in myofibroblasts. A family of monocarboxylate transporters (MCT1-4) maintains lactate homeostasis and small molecule inhibitors of these transporters have been under active clinical development for oncologic applications where glycolytic reprogramming also features prominently in disease pathobiology (20, 21). Importantly, compared to previously investigated inhibitors of glycolysis, MCT inhibitors exhibit favorable pharmacologic profiles and have successfully translated to human clinic trials (22). Before the promise of this therapeutic approach in IPF may be realized, however, the preclinical efficacy and molecular mechanisms-of-action of lactate transport inhibitors must be demonstrated experimentally.

In this work, we evaluated the contribution of lactate transporters to experimental pulmonary fibrosis. We found increased expression of the lactate transporters MCT1 and MCT4 IPF patient lungs, consistent with increased glycolysis previously observed. Inhibition of these transporters attenuated bleomycin-induced lung fibrosis *in vivo* and TGFβ-induced myofibroblast differentiation *in vitro*, where MCT4 inhibition demonstrated increased therapeutic efficacy. *Using stable isotope tracing, high-resolution spatial metabolomics, we find…* Based on these findings, we report a novel MCT4 inhibitor suitable for human clinical studies. Together, these data illuminate a novel approach to disrupt the metabolic program fueling the exuberant fibrosis at the core of IPF and provide proof-of-concept to target lactate transporters in the treatment of this deadly disease.

# Results

## MCT expression increases in human pulmonary fibrosis and experimental models

Of the four lactate transporters, MCT1 and MCT4 are the most highly expressed in the lung (23). Based on this observation, we characterized MCT1 and MCT4 expression in human IPF lung explants obtained at the time of transplantation. Consistent with a pathologic role for MCT1 and MCT4 in IPF, we found significantly increased expression of these proteins in human IPF lung compared to non-fibrotic control samples (**Figure 1A-B**). These findings were recapitulated in experimental pulmonary fibrosis where intratracheal bleomycin increased both MCT1 and MCT4 expression (**Figure 1C-D**).

A defining feature of IPF is the activation of tissue myofibroblasts, which are characterized by the *de novo* expression of smooth muscle α-actin (α-SMA); stress fiber formation; and increased migration, contraction, and extracellular matrix production (7, 9, 24). TGFβ is the most potent inducer of myofibroblast activation *in vitro* and *in vivo*. TGFβ-dependent α-SMA expression is a well-accepted and widely utilized model of myofibroblast activation relevant to pulmonary fibrosis (10, 25–27). Consistent with our findings in human IPF lungs, we observed increased expression of MCT1 and MCT4 in normal human lung fibroblasts following TGFβ treatment (**Figure 1E-F**). These findings are consistent with increased expression of other glycolytic enzymes and the associated metabolic changes that have been previously observed in these cells (9). Together, these data demonstrate that pulmonary fibrosis is associated with increased expression of lactate transporters in the lung generally and in myofibroblasts specifically.

## Myofibroblast differentiation *in vitro* requires lactate transport

Using RNA interference and pharmacologic approaches, we next tested whether MCT expression and activity were required for myofibroblast differentiation *in vitro*. Lung fibroblasts were transfected with siRNA targeting MCT1 and MCT4 separately and together. After 24 h, the cells were treated with TGFβ for 48 h to induce myofibroblast differentiation. The siRNAs reduced lactate transporter protein levels > 80% (**Figure 2A-B**). Decreasing MCT1 or MCT4 expression caused a marked reduction in TGFβ-stimulated α-SMA expression. Notably, siMCT1 also decreased MCT4 expression. Interestingly, the mRNA sequence homology of these transporters is low and the MCT1 siRNA sequences do not match the MCT4 mRNA sequence, perhaps suggesting a post-translational explanation for this phenomenon. No adverse effects of lactate transporter knockdown on cell viability were observed, and siMCT1 significantly increased cell count in both control and TGFβ-treated cells (**Supplemental Figure 2A**).

Given the reduction of MCT4 expression following siMCT1 treatment, we next evaluated the effects of pharmacologic MCT inhibitors on myofibroblast differentiation. AZD3965 is a high-affinity (*Ki* 1.6 nM) inhibitor of MCT1 (28). VB124 is a recently developed high-affinity (IC50 19 nM) inhibitor of MCT4 (21). IPF lung fibroblasts were differentiated with TGFβ in the presence of these MCT inhibitors (**Figure 2C-D**). MCT4 inhibition by VB124 alone, or in combination with AZD3965, decreased Col1a1 and α-SMA expression. These effects were also observed in normal lung fibroblasts where AZD3965 as well as VB124, separately or together, decreased α-SMA expression (**Figure 2E-F**). AR-C155858, which is a high-affinity(*Ki* 2 nM) inhibitor of both MCT1 and MCT2 (29), also decreased α-SMA expression alone and in combination with VB124 (**Supplemental Figure 2B-C**). Pharmacologic MCT inhibition did not affect cell counts over 48 h of treatment (**Supplemental Figure 2D-E**). As anticipated, decreased Col1a1 and α-SMA expression was associated with decreased myofibroblast contractility as assessed by gel contraction assay (**Figure 2G-H**). Together, these data indicate that MCT expression and activity are required for myofibroblast differentiation *in vitro*.

## MCT inhibition reprograms myofibroblast metabolism

MCTs are critical regulators of cellular lactate homeostasis. MCT1 is considered to be the main lactate importer in cells that use lactate to fuel oxidative phosphorylation or gluconeogenesis and is ubiquitously expressed. MCT1 also mediates lactate export in glycolytic cells (30, 31). MCT4 is considered to be the main lactate exporter in glycolytic cells and is up-regulated by hypoxia-inducible transcription factors (30, 32). MCT4 can also function as a lactate importer with a KM ~1 mM (33).

To evaluate the metabolic consequences of lactate transporter inhibition, we measured extracellular lactate in the conditioned medium from cells treated with MCT siRNA or pharmacologic inhibitors (**Figure 3A-B**). Silencing MCT1 and MCT4, separately or together, decreased TGFβ-stimulated lactate efflux in lung fibroblasts (**Figure 3A**). Silencing both transporters was required to block TGFβ-stimulated lactate efflux completely. By contrast, pharmacologic inhibition of either MCT1 or MCT4 did not decrease TGFβ-stimulated lactate efflux (**Figure 3B**), and inhibition of both transporters was required to decrease extracellular lactate. Similarly, inhibition of MCT1, MCT2, and MCT4 with the combination of AR-C155858 and VB124 was required to prevent increases in lactate efflux (**Supplemental Figure 3A**). Compared to lactate production, the effect of MCT inhibitors on extracellular glucose was less pronounced, although we did observe a trend toward decreased glucose consumption by cells treated with both AZD3965 and VB124 (**Supplemental Figure 3B**). These findings are consistent with prior studies suggesting compensatory roles for MCT1 and MCT4 in lactate export (34). Moreover, these data also suggest that inhibition of glycolysis is not the mechanism by which lactate transport inhibition attenuates myofibroblast differentiation.

To further interrogate the metabolic consequences of lactate transport inhibition, we next measured proton efflux (PER) and oxygen consumption (OCR) rates of lung fibroblasts treated with TGFβ in combination with MCT inhibitors (**Figure 3C, Supplemental Figure 3C**). Since MCTs co-transport protons with lactate, PER correlates with lactate efflux. Consistent with prior reports (8–10, 16, 35), we observed increases in both PER CR following TGFβ stimulation for 48 h, indicating increases in both glycolysis and oxidative phosphorylation. Similar to our direct measurements of extracellular lactate and glucose, simultaneous treatment with AZD3965 and VB124 was required to reduce PER. As anticipated, this decrease in PER was associated with a commensurate increase in OCR as myofibroblasts shifted their metabolism from glycolysis to oxidative phosphorylation. Unexpectedly, an increase in OCR was observed with AZD3965 and VB124 when administered individually. Indeed, the primary consequence of MCT1 or MCT4 inhibition alone was an increase in cellular ATP production rates driven by an increase in oxidative phosphorylation (**Figure 3D-E**). MCT inhibition decreased spare respiratory capacity, indicating that the basal respiratory rate of treated cells was closer to their maximal oxidative capacity (**Supplemental Figure 3D**). No significant differences in glycolytic capacity or electron transport chain coupling efficiency were observed (**Supplemental Figure 3D**). Together, these data suggest that the primary metabolic consequence of MCT inhibition is the activation of oxidative phosphorylation rather than inhibition of glycolysis.

To develop additional support for this hypothesis, we performed liquid chromatography-mass spectrometry-based profiling of extracellular and intracellular metabolites from cells treated with AZD3965 and VB124. As suggested by the Seahorse analysis, inhibiting a single lactate transporter had modest effects on extracellular metabolite levels (**Figure 4A-D, Supplemental Figure 4**). This analysis confirmed the results of extracellular lactate measurements by enzyme assay, which showed dual inhibition was required to decrease lactate efflux (**Figure 4B**). In addition to lactate, dual lactate transporter inhibition also had broader impacts on amino acid fluxes (**Figure 4C-D**). Several metabolites were differentially regulated by TGFβ treatment (**Supplemental Figure 4A**) and dual lactate transport inhibition, including leucine, alanine, ornithine, and ketoleucine.

Similar effects on intracellular metabolites were observed. Principal components (PC) analysis demonstrated excellent clustering of treatment groups with the drug effect corresponding to PC1 and the TGFβ treatment effect corresponding to PC2. Similar to the extracellular flux results, the magnitude of drug-induced perturbations were increased from MCT1 to MCT4 to combined inhibition (**Figure 4E**), as demonstrated by intracellular lactate levels (**Figure 4F**). As expected from measures of extracellular lactate following MCT inhibition, intracellular lactate accumulation increases modestly with MCT4 inhibition and markedly with MCT1/4 inhibition. TGFβ treatment was associated with diverse changes in the intracellular metabolomic profile of treated fibroblasts (**Supplemental Figure 4C-D**). The effect of AZD3965 alone on intracellular metabolite levels was modest (**Supplemental Figure 4G-H**). By contrast, MCT4 inhibition by VB124 alone (**Supplemental Figure 4K-L**) or combined with AZD3965 (**Figure 4G-H**) caused substantial perturbations to intracellular metabolism. Specifically, we observed enrichment of the glycolysis and tricarboxylic acid (TCA) cycle metabolite sets with MCT4 inhibition. Together, these data indicate that inhibition of lactate export leads to accumulation of upstream glycolytic intermediates that are rerouted to mitochondrial oxidative metabolic pathways.

To test this hypothesis, we next labeled lung fibroblasts with [U-13C6]-glucose (8 mM) in medium containing lactate (2 mM), glutamine (1 mM), and pyruvate (1 mM) during TGFβ stimulation and treatment with MCT inhibitors (**Figure 4I**). TGFβ increases 13C incorporation from glucose into pyruvate, lactate, citrate, succinate (SUC), and malate (MAL), consistent with increased flux from glucose into the TCA cycle. While AZD3965 had little impact on these labeling patterns, MCT4 inhibition significantly increased the fractions of these metabolites labeled by glucose, providing direct evidence for a proportional increase in glucose oxidation following MCT4 inhibition, consistent with the bioenergetic and steady-state metabolomics experiments described above.

Increased metabolite labeling by glucose must by offset by decreased labeling from other substrates. Since recent data suggest that lactate is a major oxidative fuel source in the lung (36–38), we reasoned that MCT inhibition would decrease oxidation of exogenous lactate. To test this, lung fibroblasts were cultured with [U-13C3]-lactate (2 mM) in medium containing naturally labeled glucose, glutamine, and pyruvate (**Figure 4J**). Extracellular [U-13C3]-lactate labeled ~ 50% of intracellular pyruvate and lactate at baseline with significant downstream incorporation into TCA metabolites. This labeling was decreased following TGFβ treatment, mirroring increased fractional labeling from glucose. MCT inhibition had no impact on fractional labeling of TCA intermediates by [U-13C]-glutamine (**Supplemental Figure 4M**). Interestingly, MCT4 inhibition alone or with AZD3965 decreased 13C labeling of intracellular metabolites by lactate, emphasizing the importance of bi-directional lactate transport mediated by MCTs. Similar to our measurements of extracellular lactate, the effects of MCT inhibition were more pronounced when both inhibitors were used simultaneously, again demonstrating some functional redundancy of MCT1 and MCT4 in lung fibroblasts.

## MCT inhibition attenuates pro-fibrotic transcriptional programs

To identify anti-fibrotic transcriptional programs mediated by MCT inhibition, we performed RNA-seq on lung fibroblasts treated with TGFβ in combination with AZD3965 or VB124 (**Figure 5**). Principal components analysis (PCA) of the transcriptional changes mirrored the metabolomic findings. The first principal component corresponded to TGFβ treatment, accounting for 71% of the overall variance, while MCT4 inhibition, either alone or in combination with MCT1 inhibition aligned with the second principal component (**Figure 5A**). Samples treated with the MCT1 inhibitor AZD3965 were similar to vehicle-treated controls. Differential expression analysis of TGFβ-treated cells identified the expected increased expression of extracellular matrix proteins (**Supplemental Figure 5A**) and enrichment of the epithelial-to-mesenchymal (EMT) gene set, among others (**Figure 5B**). Consistent with the PCA results, only GRIK4 and BRI3 were differentially expressed following AZD3965 (**Supplemental Figure 5B**). By contrast, VB124, alone (**Supplemental Figure 5C**) or in combination with AZD3965 (**Figure 5C**), demonstrated more substantial reprogramming of myofibroblast transcription with 443 of 24,902 genes (2%) differentially expressed at an FDR < 0.05.

Co-administration of either AZD3965 or VB124 reversed TGFβ-dependent enrichment of the EMT gene set, which is the Hallmark gene set containing genes related to fibrosis (**Figure 5B**). Leading edge analysis of the EMT gene set identified seven genes shared among all three comparisons (*i.e.*, genes increased by TGFβ and decreased by both AZD3965 and VB124) (**Figure 5D**). These genes include: biglycan (BGN), COL6A3, Frizzled 8 (FZD8), matrix Gla protein (MGP), Prostate Transmembrane Protein, Androgen Induced 1 (PMEPA1), TIMP metallopeptidase inhibitor 1 (TIMP1), and tenascin-C (TNC). Many of these genes contribute to the pathobiology of pulmonary fibrosis or serve as biomarkers of disease or treatment response (39–43). Together, these data suggest lactate transport inhibitors attenuate the pro-fibrotic transcriptional program in TGFβ-treated LFs.

## Lactate transport inhibition may signal through histone lactylation

We next sought to identify the signaling mechanism(s) linking lactate transport inhibition to anti-fibrotic transcriptional programs. Since increasing intracellular lactate was associated with more potent inhibition of α-SMA expression, we first treated cells with TGFβ in combination with extracellular lactate (10 mM). A prior study demonstrated that extracellular lactate modestly increased α-SMA expression in the absence of TGFβ (13). We found that extracellular lactate did not impact α-SMA expression in TGFβ-treated LFs (**Figure 6A-B**), suggesting that accumulation of intracellular lactate alone does not mediate the anti-fibrotic effects of MCT inhibition.

Prior work has also linked inhibition of glycolysis and lactate production to hypoxia-inducible factor 1α activation (HIF-1α), which contributes to myofibroblast differentiation (10, 13). We found maximal HIF-1α activation 6 h following TGFβ stimulation. At this time point, HIF-1α protein levels were similar across vehicle-, AZD3965-, and VB124-treated cells (**Figure 6C-D**). In LFs treated with dual inhibitors, HIF-1α protein was increased compared to vehicle, consistent with enrichment of the “Hypoxia” gene set in our RNA-seq data (**Figure 5B**). These findings, in light of prior studies, indicate that lactate transport inhibition acts downstream of HIF-1α-dependent transcriptional programs to inhibit myofibroblast differentiation.

Next, we examined TGFβ-dependent signaling. TGFβ activates both SMAD and non-SMAD signaling pathways through a cascade of protein phosphorylation events. MCT inhibitors did not decrease Smad3 or ERK phosphorylation after 48 h of TGFβ stimulation (**Figure 6E-H**).

Post-translational protein modification of histone lysine residues by lactylation has been identified as a novel mechanism of metabolic regulation of gene expression (44). Given the increase in intracellular lactate following MCT4 inhibition, we measured H3 histone lactylation by immunoblot (**Figure 6I-J**). We found that inhibition of both MCT1 and MCT4 increased histone lactylation and may account for some of the transcriptional changes observed following these treatments.

## Lactate transport inhibition contributes to antioxidant defense

Lactate metabolism is closely coupled to cellular redox homeostasis through its metabolism by lactate dehydrogenases, which couple lactate oxidation to NADH production. Since prior studies have suggested that reactive oxygen species (ROS) are important for TGFβ-mediated gene expression, we next examined the impact of lactate transport inhibition on cellular redox homeostasis. Consistent with an increase in intracellular lactate with MCT4 inhibition, we observed a concomitant increase in intracellular NADH/NAD+ (**Figure 7A**), while the NADPH/NADP+ ratio was unchanged (**Figure 7B**). Treatment-dependent increases in NADH/NAD+ corresponded to reductions in total ROS as measured by CellROX (**Figure 7C**). Notably, we did not observe TGFβ-dependent increases in total ROS production nor an impact of TGFβ or inhibitors on mitochondrial superoxide production (**Supplemental Figure 7A**).

Proline biosynthesis has been previousyl suggested as a mechanism by which NIH-3T3 fibroblasts prevent excessive ROS accumulation following TGFβ stimulation (45).

# Discussion

Craig Thompson paper: Lactate activates ETC independently of its metabolism

Craig Thompson paper: Isotope labeling in lung fibroblasts

# Methods

# Acknowledgements

# Conflicts of Interest

# References

1. Esposito DB et al. [Idiopathic Pulmonary Fibrosis in United States Automated Claims. Incidence, Prevalence, and Algorithm Validation](https://doi.org/10.1164/rccm.201504-0818OC). *Am J Respir Crit Care Med* 2015;192(10):1200–1207.

2. Raghu G, Chen S-Y, Hou Q, Yeh W-S, Collard HR. [Incidence and prevalence of idiopathic pulmonary fibrosis in US adults 18-64years old](https://doi.org/10.1183/13993003.01653-2015). *Eur Respir J* 2016;48(1):179–186.

3. Martinez FJ et al. [Idiopathic pulmonary fibrosis](https://doi.org/10.1038/nrdp.2017.74). *Nat Rev Dis Primers* 2017;3:17074.

4. King TE et al. [A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis](https://doi.org/10.1056/NEJMoa1402582). *N Engl J Med* 2014;370(22):2083–2092.

5. Richeldi L et al. [Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis](https://doi.org/10.1056/NEJMoa1402584). *N Engl J Med* 2014;370(22):2071–2082.

6. Bueno M, Calyeca J, Rojas M, Mora AL. [Mitochondria dysfunction and metabolic reprogramming as drivers of idiopathic pulmonary fibrosis](https://doi.org/10.1016/j.redox.2020.101509). *Redox Biol* 2020;33:101509.

7. Selvarajah B, Azuelos I, Anastasiou D, Chambers RC. [Fibrometabolism-An emerging therapeutic frontier in pulmonary fibrosis](https://doi.org/10.1126/scisignal.aay1027). *Sci Signal* 2021;14(697):eaay1027.

8. Bernard K et al. [Metabolic Reprogramming Is Required for Myofibroblast Contractility and Differentiation](https://doi.org/10.1074/jbc.M115.646984). *J Biol Chem* 2015;290(42):25427–25438.

9. Xie N et al. [Glycolytic Reprogramming in Myofibroblast Differentiation and Lung Fibrosis](https://doi.org/10.1164/rccm.201504-0780OC). *Am J Respir Crit Care Med* 2015;192(12):1462–1474.

10. Goodwin J et al. [Targeting Hypoxia-Inducible Factor-1α/Pyruvate Dehydrogenase Kinase 1 Axis by Dichloroacetate Suppresses Bleomycin-induced Pulmonary Fibrosis](https://doi.org/10.1165/rcmb.2016-0186OC). *Am J Respir Cell Mol Biol* 2018;58(2):216–231.

11. Hamanaka RB et al. [Glutamine Metabolism Is Required for Collagen Protein Synthesis in Lung Fibroblasts](https://doi.org/10.1165/rcmb.2019-0008OC). *Am J Respir Cell Mol Biol* 2019;61(5):597–606.

12. Selvarajah B et al. [mTORC1 amplifies the ATF4-dependent de novo serine-glycine pathway to supply glycine during TGF-β1-induced collagen biosynthesis](https://doi.org/10.1126/scisignal.aav3048). *Sci Signal* 2019;12(582):eaav3048.

13. Kottmann RM et al. [Lactic acid is elevated in idiopathic pulmonary fibrosis and induces myofibroblast differentiation via pH-dependent activation of transforming growth factor-β](https://doi.org/10.1164/rccm.201201-0084OC). *Am J Respir Crit Care Med* 2012;186(8):740–751.

14. Kottmann RM et al. [Pharmacologic inhibition of lactate production prevents myofibroblast differentiation](https://doi.org/10.1152/ajplung.00058.2015). *Am J Physiol Lung Cell Mol Physiol* 2015;309(11):L1305–1312.

15. Judge JL et al. [Prevention and treatment of bleomycin-induced pulmonary fibrosis with the lactate dehydrogenase inhibitor gossypol](https://doi.org/10.1371/journal.pone.0197936). *PLoS One* 2018;13(5):e0197936.

16. Cho SJ, Moon J-S, Lee C-M, Choi AMK, Stout-Delgado HW. [Glucose Transporter 1-Dependent Glycolysis Is Increased during Aging-Related Lung Fibrosis, and Phloretin Inhibits Lung Fibrosis](https://doi.org/10.1165/rcmb.2016-0225OC). *Am J Respir Cell Mol Biol* 2017;56(4):521–531.

17. Rodríguez-Enríquez S, Marín-Hernández A, Gallardo-Pérez JC, Carreño-Fuentes L, Moreno-Sánchez R. [Targeting of cancer energy metabolism](https://doi.org/10.1002/mnfr.200700470). *Mol Nutr Food Res* 2009;53(1):29–48.

18. Michelakis ED et al. [Inhibition of pyruvate dehydrogenase kinase improves pulmonary arterial hypertension in genetically susceptible patients](https://doi.org/10.1126/scitranslmed.aao4583). *Sci Transl Med* 2017;9(413):eaao4583.

19. Pelicano H, Martin DS, Xu R-H, Huang P. [Glycolysis inhibition for anticancer treatment](https://doi.org/10.1038/sj.onc.1209597). *Oncogene* 2006;25(34):4633–4646.

20. Puri S, Juvale K. [Monocarboxylate transporter 1 and 4 inhibitors as potential therapeutics for treating solid tumours: A review with structure-activity relationship insights](https://doi.org/10.1016/j.ejmech.2020.112393). *Eur J Med Chem* 2020;199:112393.

21. Cluntun AA et al. [The pyruvate-lactate axis modulates cardiac hypertrophy and heart failure](https://doi.org/10.1016/j.cmet.2020.12.003). *Cell Metab* 2021;33(3):629–648.e10.

22. Halford S et al. [A Phase I Dose-escalation Study of AZD3965, an Oral Monocarboxylate Transporter 1 Inhibitor, in Patients with Advanced Cancer](https://doi.org/10.1158/1078-0432.CCR-22-2263). *Clin Cancer Res* 2023;29(8):1429–1439.

23. Frangogiannis N. [Transforming growth factor-β in tissue fibrosis](https://doi.org/10.1084/jem.20190103). *J Exp Med* 2020;217(3):e20190103.

24. Tomasek JJ, Gabbiani G, Hinz B, Chaponnier C, Brown RA. [Myofibroblasts and mechano-regulation of connective tissue remodelling](https://doi.org/10.1038/nrm809). *Nat Rev Mol Cell Biol* 2002;3(5):349–363.

25. Cui H et al. [Inhibition of Glutaminase 1 Attenuates Experimental Pulmonary Fibrosis](https://doi.org/10.1165/rcmb.2019-0051OC). *Am J Respir Cell Mol Biol* 2019;61(4):492–500.

26. Rangarajan S et al. [Metformin reverses established lung fibrosis in a bleomycin model](https://doi.org/10.1038/s41591-018-0087-6). *Nat Med* 2018;24(8):1121–1127.

27. Locy ML et al. [Oxidative cross-linking of fibronectin confers protease resistance and inhibits cellular migration](https://doi.org/10.1126/scisignal.aau2803). *Sci Signal* 2020;13(644):eaau2803.

28. Curtis NJ et al. [Pre-clinical pharmacology of AZD3965, a selective inhibitor of MCT1: DLBCL, NHL and Burkitt’s lymphoma anti-tumor activity](https://doi.org/10.18632/oncotarget.18215). *Oncotarget* 2017;8(41):69219–69236.

29. Ovens MJ, Davies AJ, Wilson MC, Murray CM, Halestrap AP. [AR-C155858 is a potent inhibitor of monocarboxylate transporters MCT1 and MCT2 that binds to an intracellular site involving transmembrane helices 7-10](https://doi.org/10.1042/BJ20091515). *Biochem J* 2010;425(3):523–530.

30. Halestrap AP. [The SLC16 gene family - structure, role and regulation in health and disease](https://doi.org/10.1016/j.mam.2012.05.003). *Mol Aspects Med* 2013;34(2-3):337–349.

31. Brooks GA. [The Science and Translation of Lactate Shuttle Theory](https://doi.org/10.1016/j.cmet.2018.03.008). *Cell Metab* 2018;27(4):757–785.

32. Ullah MS, Davies AJ, Halestrap AP. [The plasma membrane lactate transporter MCT4, but not MCT1, is up-regulated by hypoxia through a HIF-1alpha-dependent mechanism](https://doi.org/10.1074/jbc.M511397200). *J Biol Chem* 2006;281(14):9030–9037.

33. Contreras-Baeza Y et al. [Monocarboxylate transporter 4 (MCT4) is a high affinity transporter capable of exporting lactate in high-lactate microenvironments](https://doi.org/10.1074/jbc.RA119.009093). *J Biol Chem* 2019;294(52):20135–20147.

34. Benjamin D et al. [Dual Inhibition of the Lactate Transporters MCT1 and MCT4 Is Synthetic Lethal with Metformin due to NAD+ Depletion in Cancer Cells](https://doi.org/10.1016/j.celrep.2018.11.043). *Cell Rep* 2018;25(11):3047–3058.e4.

35. Schruf E et al. [Human lung fibroblast-to-myofibroblast transformation is not driven by an LDH5-dependent metabolic shift towards aerobic glycolysis](https://doi.org/10.1186/s12931-019-1058-2). *Respir Res* 2019;20(1):87.

36. Hui S et al. [Glucose feeds the TCA cycle via circulating lactate](https://doi.org/10.1038/nature24057). *Nature* 2017;551(7678):115–118.

37. Hui S et al. [Quantitative Fluxomics of Circulating Metabolites](https://doi.org/10.1016/j.cmet.2020.07.013). *Cell Metab* 2020;32(4):676–688.e4.

38. Copeland CA et al. [MYC overrides HIF-1α to regulate proliferating primary cell metabolism in hypoxia](https://doi.org/10.7554/eLife.82597). *Elife* 2023;12:e82597.

39. Kwapiszewska G et al. [Transcriptome profiling reveals the complexity of pirfenidone effects in idiopathic pulmonary fibrosis](https://doi.org/10.1183/13993003.00564-2018). *Eur Respir J* 2018;52(5):1800564.

40. Williams LM et al. [Identifying collagen VI as a target of fibrotic diseases regulated by CREBBP/EP300](https://doi.org/10.1073/pnas.2004281117). *Proc Natl Acad Sci U S A* 2020;117(34):20753–20763.

41. Baarsma HA, Königshoff M. [’WNT-er is coming’: WNT signalling in chronic lung diseases](https://doi.org/10.1136/thoraxjnl-2016-209753). *Thorax* 2017;72(8):746–759.

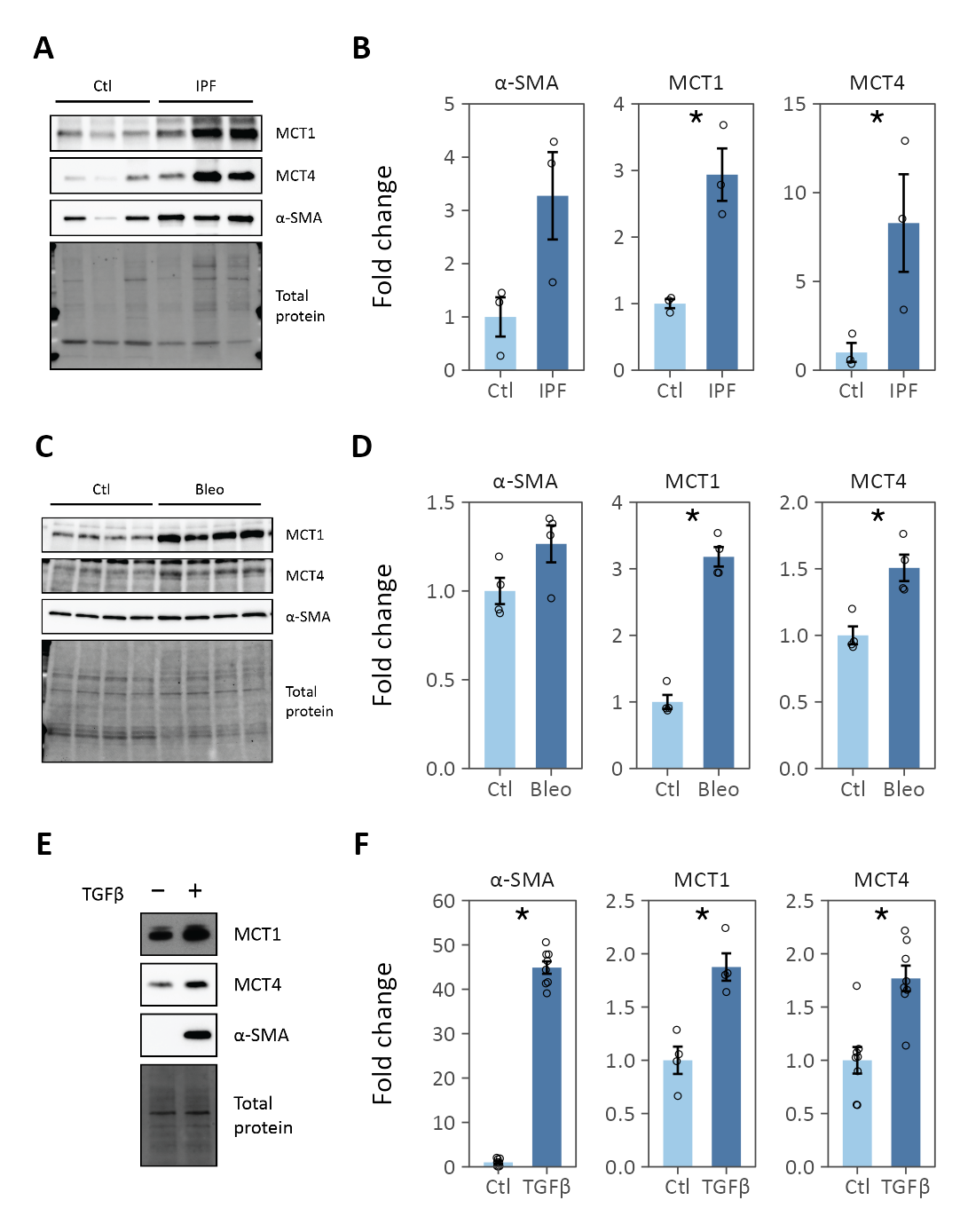
42. Todd JL et al. [Circulating matrix metalloproteinases and tissue metalloproteinase inhibitors in patients with idiopathic pulmonary fibrosis in the multicenter IPF-PRO Registry cohort](https://doi.org/10.1186/s12890-020-1103-4). *BMC Pulm Med* 2020;20(1):64.

43. Bhattacharyya S et al. [Tenascin-C drives persistence of organ fibrosis](https://doi.org/10.1038/ncomms11703). *Nat Commun* 2016;7:11703.

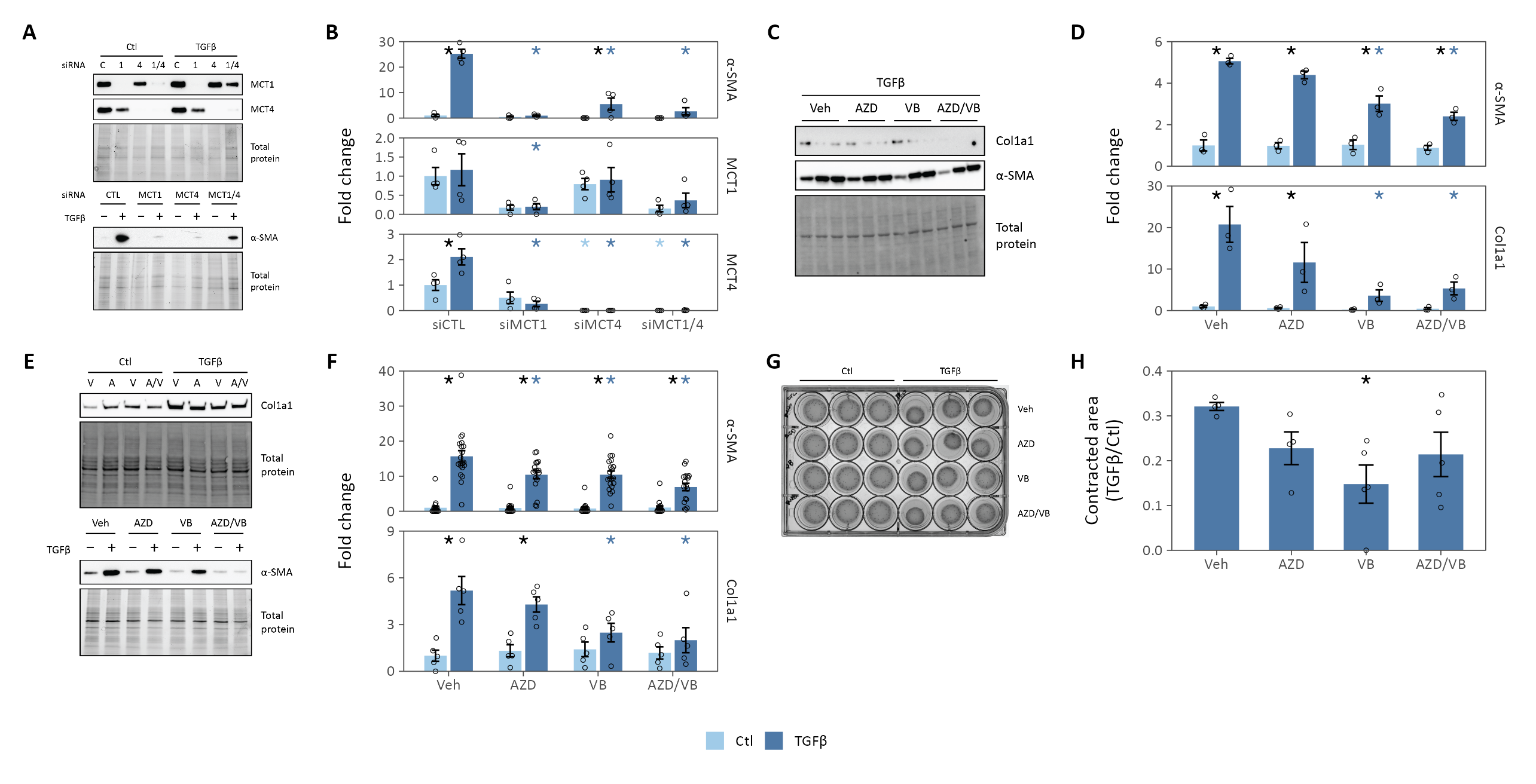
44. Zhang D et al. [Metabolic regulation of gene expression by histone lactylation](https://doi.org/10.1038/s41586-019-1678-1). *Nature* 2019;574(7779):575–580.

45. Schwörer S et al. [Proline biosynthesis is a vent for TGFβ-induced mitochondrial redox stress](https://doi.org/10.15252/embj.2019103334). *EMBO J* 2020;39(8):e103334.

# Figures



**Figure 1 - Lactate transporter expression increases in human IPF and experimental models.** (**A**) Immunoblot of whole lung homogenates from explanted IPF lungs and controls (Ctl). (**B**) Quantification of band densities from (A). (**C**) Immunoblot of whole lung homogenates from bleomycin (bleo)- and vehicle (Ctl)-treated mice. (**D**) Quantification of band densities from (C). (**E**) Representative immunoblot of cell lysates from normal human lung fibroblasts treated with TGFβ to induce myofibroblast differentiation. (**F**) Quantification of band densities from (E). Individual data points are biological replicates. Summary data are mean ± SEM (\* p < 0.05).



**Figure 2 - Lactate transport inhibition decreases myofibroblast differentiation *in vitro*.** (**A**) Representative immunoblot of normal lung fibroblast cell lysates after treatment with siRNA targeting MCT1 or MCT4. (**B**) Quantification of protein expression levels following MCT1 and MCT4 knockdown. (**C**) Immunoblot of IPF lung fibroblast lysates from three separate donors after treatment with TGFβ and MCT inhibitors AZD3965 (AZD) or VB124 (VB). (**D**) Quantification of band densities from (C). (**E**) Representative immunoblot of normal lung fibroblasts treated with TGFβ and MCT inhibitors. (**F**) Quantification of α-SMA expression following MCT inhibition. (**G**) Representative image of a gel contraction assay after 24 h treatment with TGFβ and MCT inhibitors. (**H**) Quantification of contracted areas. Individual data points are biological replicates. Summary data are mean ± SEM (\* p < 0.05; *black* compares TGFβ to control within a given treatment, *colored* compares the treatment effect to control for a given condition).