Class 7: Machine Learning I

Olivia Chu

In this class, we will explore clustering and dimensionality reduction methods.

K-means

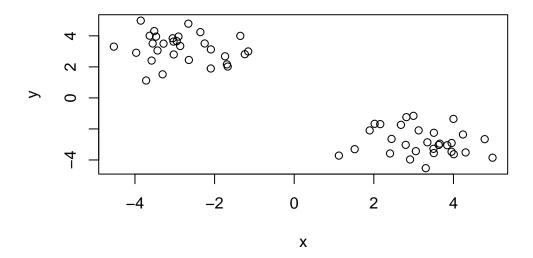
Make up some input data where we know what the answer should be.

```
tmp <- c( rnorm(30, -3), rnorm(30, +3) )
x <- cbind(x=tmp, y=rev(tmp))
head(x)</pre>
```

```
x y
[1,] -3.305802 1.519537
[2,] -3.969310 2.910768
[3,] -3.630350 4.008798
[4,] -4.529918 3.304849
[5,] -2.093125 3.125892
[6,] -3.032065 3.629303
```

Quick plot of x to see the two groups at -3,+3 and +3,-3.

```
plot(x)
```



Use the kmeans() function, setting k to 2 and nstart=20.

```
km <- kmeans(x, centers = 2, nstart=20)
km</pre>
```

K-means clustering with 2 clusters of sizes 30, 30

Cluster means:

Clustering vector:

Within cluster sum of squares by cluster: [1] 47.39788 47.39788

```
(between_SS / total_SS = 92.0 %)
```

Available components:

[1] "cluster" "centers" "totss" "withinss" "tot.withinss"

[6] "betweenss" "size" "iter" "ifault"

Q. How many points are in each cluster?

km\$size

[1] 30 30

Each cluster has 30 points.

Q. What 'component' of your result object details... - cluster assignment/membership?

- cluster center?

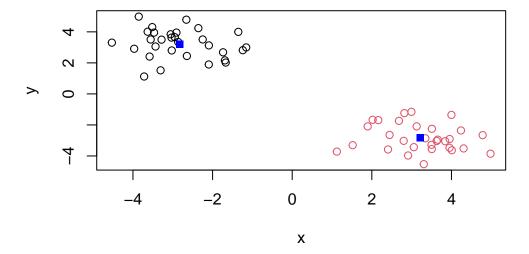
km\$cluster

km\$centers

x y 1 -2.820738 3.215811 2 3.215811 -2.820738

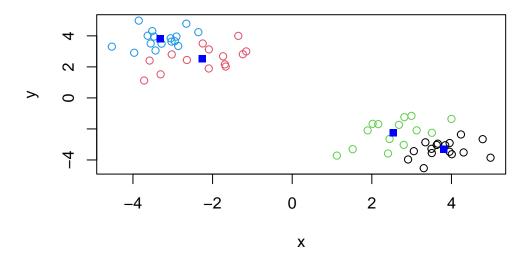
Q. Plot x colored by the kmeans cluster assignment and add cluster centers as blue points.

plot(x, col=km\$cluster)
points(km\$centers, col="blue", pch=15)



Play with kmeans and ask for different numbers of clusters.

```
km <- kmeans(x, centers = 4, nstart=20)
plot(x, col=km$cluster)
points(km$centers, col="blue", pch=15)</pre>
```



Hierarchial Clustering

This is another very useful and widely employed clustering method which has the advantage over kmeans in that it can help reveal the something of the true grouping in your data.

The hclust() function wants a distance matrix as input. We can get this from the dist() function.

```
d <- dist(x)
hc <- hclust(d)
hc</pre>
```

Call:

hclust(d = d)

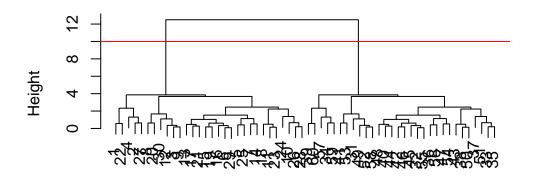
Cluster method : complete
Distance : euclidean

Number of objects: 60

There is a plot method for hclust results:

```
plot(hc)
abline(h=10, col="red")
```

Cluster Dendrogram

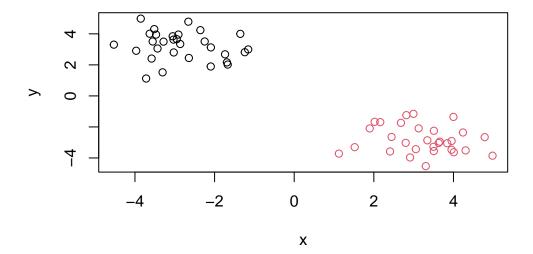


d hclust (*, "complete")

To get my cluster membership vector, I need to "cut" my tree to yield sub-trees or branches with all the members of a given cluster residing on the same cut branch. The function to do this is called cutree().

```
grps <- cutree(hc, h=10)
grps</pre>
```

```
plot(x, col=grps)
```



It is often helpful to use the k= argument to cutree rather than the h= height of cutting with cutree(). This will cut the tree to yield the number of clusters you want.

Principal Component Analysis (PCA)

The base R function for PCA is called prcomp().

PCA of UK food data

Import the data.

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url)
head(x)</pre>
```

X England Wales Scotland N.Ireland Cheese Carcass_meat Other_meat Fish 5 Fats_and_oils Sugars

Q1. How many rows and columns are in your new data frame named x? What R functions could you use to answer this questions?

Complete the following code to find out how many rows and columns are in x? $\dim(x)$

[1] 17 5

Using the dim() function, I found that there are 17 rows and 5 columns in my data frame x. I can also use the functions nrow() and ncol() to find the number of rows and columns.

```
# This is the minus indexing approach that does not work
# rownames(x) <- x[,1]
# x <- x[,-1]
# head(x)

# Here, I am removing the first column in order to make sure my first column is "England"
x <- read.csv(url, row.names=1)
head(x)</pre>
```

	England	Wales	Scotland	${\tt N.Ireland}$
Cheese	105	103	103	66
Carcass_meat	245	227	242	267
Other_meat	685	803	750	586
Fish	147	160	122	93
Fats_and_oils	193	235	184	209
Sugars	156	175	147	139

dim(x)

[1] 17 4

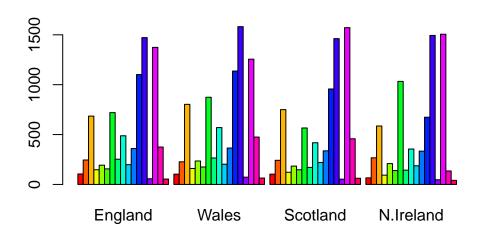
Using the dim() function, there are now 17 rows and 4 columns in data frame 'x'.

Q2. Which approach to solving the 'row-names problem' mentioned above do you prefer and why? Is one approach more robust than another under certain circumstances?

I prefer to use the approach that sets the correct row names as opposed to removing the first column. This is because if we were to write the code to delete the first row, every time to run the code, a column will be deleted until we run into an 'Error' message.

With the second approach, this does not happen. Once the rows are set correctly, they remain that way.

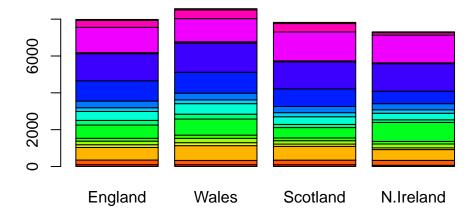
```
barplot(as.matrix(x), beside=T, col=rainbow(nrow(x)))
```



Q3. Changing what optional argument in the above barplot() function results in the following plot?

Changing the 'beside' argument to 'beside=F' will result in the following plot (containing 4 bars). This tells the function to stack the sub-bars.

```
barplot(as.matrix(x), beside=F, col=rainbow(nrow(x)))
```



Q5. Generating all pairwise plots may help somewhat. Can you make sense of the following code and resulting figure? What does it mean if a given point lies on the diagonal for a given plot?

pairs(x, col=rainbow(10), pch=16)



These plots show the comparison of food type consumption between two respective countries.

If a given point lies on the diagonal for a given plot, that means there is a positive correlation between the two countries and that they consume similar amounts of that specific food type.

Q6. What are the main differences between N. Ireland and the other countries of the UK in terms of this data-set?

N. Ireland consumes vastly different amounts of fresh potatoes, fresh fruit, and alcoholic drinks compared to the other countries of this data-set.

```
# Use the prcomp() PCA function
pca <- prcomp( t(x) )
summary(pca)</pre>
```

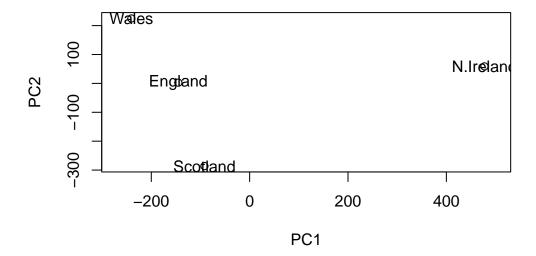
Importance of components:

```
PC1 PC2 PC3 PC4
Standard deviation 324.1502 212.7478 73.87622 4.189e-14
Proportion of Variance 0.6744 0.2905 0.03503 0.000e+00
Cumulative Proportion 0.6744 0.9650 1.00000 1.000e+00
```

A "PCA plot" (a.k.a "Score plot", PC1vsPC2 plot, etc.)

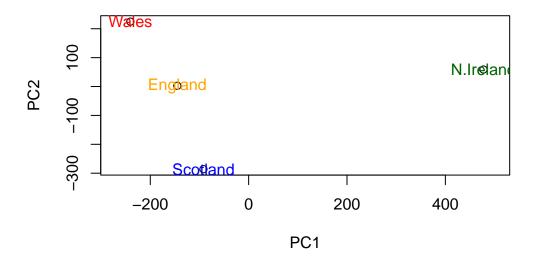
Q7. Complete the code below to generate a plot of PC1 vs PC2. The second line adds text labels over the data points.

```
# Plot PC1 vs PC2
plot( pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2", xlim=c(-270,500) )
text(pca$x[,1], pca$x[,2], colnames(x))
```



Q8. Customize your plot so that the colors of the country names match the colors in our UK and Ireland map and table at start of this document.

```
plot( pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2", xlim=c(-270,500) )
text(pca$x[,1], pca$x[,2], colnames(x), col=c("orange", "red", "blue", "darkgreen"))
```



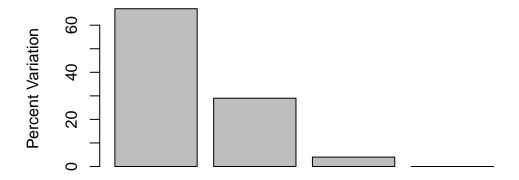
```
# Calculate how much variation in the original data each PC accounts for v <- round( pca\ensuremath{^{\circ}}2/sum(pca\ensuremath{^{\circ}}3 + 100 ) v
```

[1] 67 29 4 0

```
# or the second row here...
z <- summary(pca)
z$importance</pre>
```

```
PC1 PC2 PC3 PC4
Standard deviation 324.15019 212.74780 73.87622 4.188568e-14
Proportion of Variance 0.67444 0.29052 0.03503 0.000000e+00
Cumulative Proportion 0.67444 0.96497 1.00000 1.0000000e+00
```

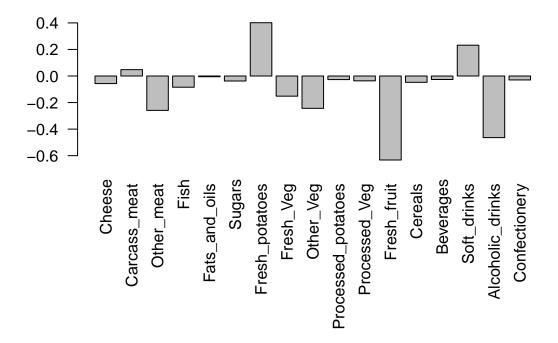
```
#Plot of variances
barplot(v, xlab="Principal Component", ylab="Percent Variation")
```



Principal Component

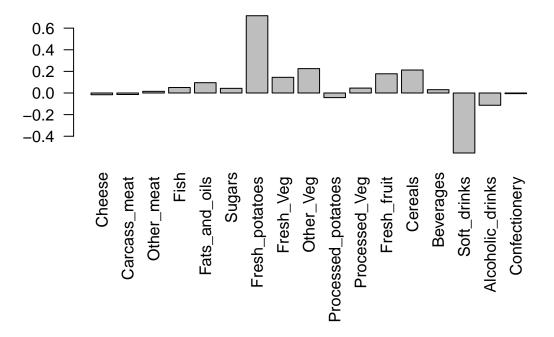
Let's focus on PC1 as it accounts for > 90% of variance.

```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,1], las=2 )
```



Q9: Generate a similar 'loadings plot' for PC2. What two food groups feature prominantely and what does PC2 maniply tell us about?

```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,2], las=2 )
```



The two food groups that feature predominantly are fresh potatoes and soft drinks.