
Bontera Research

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1

Potatoes

1.1 Soft Rot

1.1.1 Description

Bacterial soft rot affects a number of fruits and vegetables, including potatoes. It is a post-harvest disease, occurring while the crop is stored or in transit (Rich, 2013). It is characterized by a watery soft spot on the side of the crop and an strong odor.

1.1.2 Geography and Soil Type

Soft rot arises worldwide, from South Africa (Ngadze et al., 2012) to the northeastern United States (Ge et al., 2021). Pérombelon notes that the soil has to be both nutritionally deficient and over-watered for soft rot to spread. Some bacterium are specific to potato soft rot because of the cool, temperate climate where they are grown (Pérombelon, 2002). Soft rot favors warm temperatures and high moisture levels in the soil and during storage (Rich, 2013).

1.1.3 Cause

Two main genera cause soft rot: *Pectobacterium* and *Dickeya* (Youdkes et al., 2020). Bacterial growth is accelerated by any wound or puncture of the potato skin, especially a tender spot formed by standing water or insect bites breaking the outer layer (Rich, 2013).

1.1.4 Biocontrol

Historically, farmers have used water-management and sanitation to control soft rot, but the bacterial predator *Bdellovibrio bacteriovorus* and similar organisms have shown considerable promise as well, according to a recent study by Youdkes et al. (2020). All the strains tested were effective in reducing *Pectobacterium* and *Dickeya* greatly. Strains introduced in the tubers before the onset of soft rot were much more effective in fighting it later on, possibly because glucose consumption by the prey was not inhibiting growth (Youdkes et al., 2020). A second study also determined that a mixture of *Pseudomonas putida* and *Pseudomonas fluorescens* decreased the severity of the disease and prevented transmission to child potatoes.

A third study found that certain varieties of rhizosphere bacteria could also be used to control soft rot caused by *Pectobacterium* strains (Krzyzanowska et al., 2012). The results indicate that 18 various rhizobacteria were effective in inhibiting the spread of soft rot, out of the 1165 tested. The study noted, however, that the bacteria's ability to control *Pectobacterium* *in vitro* may not reflect the true results *in vivo*.

1.2 Golden Nematode Disease

1.2.1 Description

Nematodes are nearly invisible worms that attack potato plants and tubers (Rich, 2013). Rich also reports that some nematodes cause disease directly, while others act as vectors or catalysts for viral and fungal illnesses. Plants affected by golden nematode disease have necrotic and wilting leaves with stunted growth; many do not recover, resulting in severely reduced crop yields (Rich, 2013).

1.2.2 Geography and Soil Type

The golden nematode is native to Peru, but has spread all throughout the world (Rich, 2013). They can survive in any climate that potatoes can grow, but very strict quarantine and sanitation guidelines have prevented further spread (Evans & Brodie, 1980).

Mimee et al. (2015) found that optimal nematode egg hatching occurs when soil temperatures are between $59^{\circ}F$ and $80^{\circ}F$. They determined that

increasing temperatures in cooler climates could lead to accelerated spread of golden nematode disease. Both dry weather and light soil favor the disease (Rich, 2013).

1.2.3 Cause

The golden nematode (*Globodera rostochiensis*) is the main cause of Golden Nematode Disease (Rich, 2013). The disease gets its name from the golden or brown cysts containing nematode eggs present after an infestation (Rich, 2013). This is how the the disease spreads: the cysts cling to containers, equipment, and tubers and are transferred between fields or farms.

1.2.4 Biocontrol

Some farmers have reported the natural decline of nematode populations because of fungi parasitism (Evans, 1993). 10 species were isolated from the soil, *Cylindrocarpon destructans* being the most promising (Evans, 1993). That same study found that “when an inoculum of straw colonised by *C. destructans* was placed around potato seed tubers planted in [nematode] infested soil ... the numbers of juvenile stages of *G. rostochiensis* ... decreased by 62%.”

1.3 Brown Rot

1.3.1 Description

Brown rot, also known as bacterial wilt, is a very destructive bacterial disease (Rich, 2013). It was first reported it in the United States in 1896. Potatoes afflicted with brown rot excrete a “slimy ooze” at the base of the stem and eyes, although infected plants may also produce healthy tubers (Rich, 2013). In the early stages of the disease, the shoot system may wilt and the tubers will turn brown (Kabeil et al., 2008).

1.3.2 Geography and Soil Type

Brown rot heavily favors warmer climates; so much so that it is virtually eradicated in Canada and the northern United States (Rich, 2013). If it does

infect potatoes in cooler climates, brown rot is harder to detect early on (Kabeil et al., 2008). The plants may not show any signs of infection until later stages of infection.

In a study by van Elsat et al. (2000), a steady decline of brown rot was observed in loamy sand soil. They also found that severe drought drastically reduced brown rot infections across the board. The disease prefers moist, slightly acid, netral, and alkaline soil (Rich, 2013).

1.3.3 Cause

Pseudomonas solanacearum is the bacterium that causes brown rot (van Elsas et al., 2000). There are three races, each of which attack different crops and have different optimal climates (Rich, 2013):

Race 1: Attacks eggplant, tobacco, tomato, and potatoes

Race 2: Attacks banana

Race 3: Attacks mainly potato, but is weakly pathogenic to tobacco

P. solanacearum usually enters a host through a wound on the skin or roots (Rich, 2013). Rich (2013) also reports that the afflicted crops may infect the soil, transferring brown rot to any new plants grown afterwards.

1.3.4 Biocontrol

One effective way to combat brown rot is to add sulfur to sandy soil, followed by limestone in the summer (Rich, 2013). Furthermore, a number of potato variants are resistant to *P. solanacearum*; planting these cultivars will help minimize losses (Rich, 2013).

A study was also conducted by Fujiwara et al. in 2011 that experimented with using bacteriophages to control brown rot. While many of the viruses they tried were effective, application of ϕ RSL1 (from family *Myoviridae*) was the best method. Although it did not kill all the bacterial cells, it was successful in preventing the onset of disease. ϕ RSL1 worked much better when it was introduced before *P. solanacearum* as a preventative measure, although it did also have an effect when it was introduced after (Fujiwara et al., 2011).

1.4 Common Scab

1.4.1 Description

Common scab is a fungal disease that shows no symptoms above ground in potato plants (Rich, 2013). While not effecting yeild, common scab makes the infected tuber less appealing to look at, effecting marketability to consumers (Rich, 2013). Symptoms include scabs and lesions of varying shapes and sizes on the skin of the tuber (Dees & Wanner, 2012).

1.4.2 Geography and Soil Type

Common scab occurs on every continent except Antarctica (Rich, 2013). In the United States, it is concentrated around the northwest, midwest, and northeastern portions (Braun et al., 2017). The disease prefers “a pH higher than 5.2, temperatures of 20–22 °C, and a soil moisture below field capacity during early tuberization.” (Archuleta & Easton, 1981; Braun et al., 2017)

1.4.3 Cause

The main cause of common scab in potatoes is various species of *Streptomyces* (Flores-González et al., 2008). It is disputed whether *Streptomyces* is a fungus or bacterium because it demonstrates characteristics of both (the USDA considers it a fungus) (Rich, 2013). The responsible strains synthesize thaxtomins and phytotoxins, chemicals required for the scabs to form (Flores-González et al., 2008). Flores-González et al. (2008) also report that the toxins exhibit properties similar to those of an infection.

1.4.4 Biocontrol

The usual technique for controlling common scab has been seed treatment with pentachloronitrobenzene or maneb-zinc dust, but these show minimal effect on certain causal species (Lee et al., 2004). Therefore, a more effective solution is needed for the control of common scab. Four *Streptomyces* isolates, A020645, A010321, A010564, and A020973, showed antagonistic behavior against the common scab in a study by Lee et al. (2004). They lowered the formation of scabs and lesions by >60% and 55%, respectively. Additionally, these four isolates were also resistant against 10 commonly used chemicals in potato fields, further improving their effectiveness.

2

Corn (Maize)

2.1 Dry Rot

2.1.1 Description

Dry rot is the most common ear rot disease (Ullstrup, 1961). Corn infected early in the growing season becomes grayish brown with fungal growth in between the kernels (Ullstrup, 1961). If the ear is infected later, then it shows no superficial signs, the only symptom being a white mold forming in the cob and slightly discolored kernels (Ullstrup, 1961).

2.1.2 Geography and Soil Type

Dry rot can occur at anywhere from 40° to 90°F, the best temperature being about 80°F (Melhus & Durrell, 1922). The disease favors a dry spell in the summer followed by wet weather while the corn silks (Melhus & Durrell, 1922; Ullstrup, 1961). The optimal soil pH for dry rot growth is between 3.7 and 5.9, with a small amount of growth occurring in slightly basic soil (Eddins et al., 1930). Dry rot is especially widespread in soil with low potassium and high nitrogen (Ullstrup, 1961).

2.1.3 Cause

Scientists determined in 1834 that the primary cause of dry rot is the fungus *Stenocarpella maydis* (Durrell, 1923; Ullstrup, 1961). Mature spores are carried by the wind in warm, moist weather to healthy plants (Ullstrup,

1961). It enters into the ear through wounds in the husk, but may also infect the stalk and roots. Spores can lie dormant in soil for potentially several years while waiting for a host to infect (Eddins et al., 1930). *S. maydis* infects the center of the ear first, only coming to the surface to release its spores (Melhus & Durrell, 1922). Some hybrid species of corn have mild resistance, but no species is completely immune (Ullstrup, 1961).

2.1.4 Biocontrol

Two *Streptomyces* isolates, DAUFPE 11470 and DAUFPE 14632, were very effective in the control of *S. maydis*; they reduced the pathogen in seedlings by 87.3% and 85.6%, respectively (Bressan & Figueiredo, 2005; Bressan, 2003). Seed germination was also increased by about 30% when *Streptomyces* was introduced in the same study.

Furthermore, several strains of *Bacillus subtilis* were found to be very effective in the control of *S. maydis*, as well as able to withstand extreme temperature and pH values (Petatán-Sagahón et al., 2011). They inhibited dry rot by nearly 100%. In a recent study by Petatán-Sagahón et al. (2011), it was reported that “the strains of *B. subtilis* are the most promising due to their ability to produce endospores that can remain in the soil during long periods and can be produced easily at an industrial level with a low cost (Stein, 2005).”

2.2 Stewart's Bacterial Wilt

2.2.1 Description

Bacterial wilt is a bacterial illness that can very quickly lead to the loss of sweet corn (Robert, 1967). Infected plants resemble corn with a low water supply and may not produce ears if they survive the disease (Ullstrup, 1961). When the stalks are cut, yellow cysts of bacteria ooze out from the wound (Ullstrup, 1961). Streaks of dead tissue form on the leaves before they wilt (Robert, 1967). The disease is much less deadly on varieties of dent corn than it is on sweet corn (Rand & Cash, 1933).

2.2.2 Geography and Soil Type

The eastern United States experiences the worst effects of bacterial wilt in corn (Robert, 1967; Ullstrup, 1961). While one harsh winter is enough to reduce the disease in the north, a series of cold winters is needed further south (Robert, 1967). However, multiple mild winters can build up the disease greatly (Robert, 1967). Soil type has little effect on bacterial wilt, although plants are more likely to die in unfertile soil (Thomas, 1924; Ullstrup, 1961). Thomas reported in 1924 that, while there was evidence of infection through seeds, no plants were infected through the soil in a greenhouse study.

2.2.3 Cause

Erwinia stewartii is the cause of Stewart's bacterial wilt in corn (Ullstrup, 1961). Ullstrup (1961) found that *E. stewartii* is carried through the winter on the bodies of corn flea beetles: "they feed on young corn plants and start infections on the leaves ... [and] spread the disease from infected to healthy plants." Streaks on the leaves of infected plants start from the beetle feeding on it, and then spread along the veins and up into the stalk (Robert, 1967). Lesions along the stalk of the corn plant can be covered with fungi, which is often mistaken to be the cause of the wilting and infection (Ullstrup, 1961).

2.2.4 Biocontrol

Wicklow and Poling found in 2009 that *Acremonium zeae* produces natural antibiotics that can protect corn against a number of pathogens that cause bacterial wilt, stalk rot, and leaf blight in corn. It was, however, ineffective against *E. stewartii* (Wicklow & Poling, 2009). Currently, the best control against Stewart's bacterial wilt is the cultivation of resistant hybrids (Golden Cross Bantam, Country Gentleman, Evergreen, etc.) and the use of insecticides to kill beetles that carry the disease (Ullstrup, 1961). Correct amounts of potassium minimize bacterial wilt, while high nitrogen predispose corn to infection (Ullstrup, 1961).

2.3 Northern Corn Leaf Blight (NCLB)

2.3.1 Description

This leaf disease is a very devastating fungal illness (Zhang et al., 2021). If it becomes well established a couple weeks following silking, crop yield can be reduced by over 30% (Ullstrup, 1961). Symptoms include long spots on leaves that spread until the entire plant (sans kernels) is taken over late in the growing season (Ullstrup, 1961). The symptoms resemble those of frost damage once northern corn leaf blight has progressed to a late stage.

2.3.2 Geography and Soil Type

NCBL is found in most moist environments where corn can be grown (Ullstrup, 1961). When conditions are perfect, such as in Florida, the disease can appear very early in the growing season and infect seedlings (Ullstrup, 1961). Ullstrup reports in his book, *Corn Diseases in the United States and Their Control*, that “germination and penetration take place within 6 to 18 hours when free water is present and the temperature is within 65° and 80°F.” He found that it is prevalent in the eastern half of the United States, extending into the central area when conditions are favorable.

2.3.3 Cause

NCBL is caused by the heterothallic fungus *Setosphaeria turcica* (Leonard et al., 1989). It overwinters on corn leaves until its spores form on old lesions (Ullstrup, 1961). Then the wind can carry it a great distance during the summer. The races of *S. turcica* are classified based on their virulence against specific corn genes:

Race 0: All *Ht* genes are resistant to it

Race 1: Can attack *Ht1*

Race 23: Can attack *Ht23*

2.3.4 Biocontrol

Klebsiella jilinsis 2N3 is a bacterium that can degrade some herbicides, but also promotes corn growth and NCLB resistance (Zhang et al., 2021). Treated plants showed a 67.44% decrease in *S. turcica* in pot experiments, as well as increased production of defence enzymes (Zhang et al., 2021).

Furthermore, Zhang et al. determined in 2021 that *K. jilinsis* 2N3 can “grow in a nitrogen-free environment, dissolve inorganic phosphorus and potassium, and produce indoleacetic acid and a siderophore,” as well as prevent the onset of NCLB.

2.4 Yellow Leaf Blight

2.4.1 Description

Yellow leaf blight was first detected in Wisconsin in 1967 (Comstock et al., 1973). It usually develops in large, local outbreaks (Arny & Nelson, 1971). Infected plants develop spots of dead tissue on their leaves, surrounded by a yellowish-tan tint (Ullstrup, 1961). When the infection has progressed, brownish-black sacks of fungus spores form in the center of the lesions on older leaves, eventually breaking and oozing under wet weather (Ullstrup, 1961).

2.4.2 Geography and Soil Type

This illness is common in the northern portion of the corn belt (Arny & Nelson, 1971). It is most severe in cool, wet weather, being a fungal disease (Ullstrup, 1961). 90-95% of spores germinate within 5 hours at 12, 15, or 18°C (Castor et al., 1977). Insufficient tilling accelerates yellow leaf blight because the trash and debris from last season’s crop may infect corn growing near it (Castor et al., 1977; Ullstrup, 1961).

2.4.3 Cause

Phyllosticta maydis is the causal agent of yellow leaf blight (Arny & Nelson, 1971). It cannot infect seeds, instead surviving in the post-harvest debris of previously infected plants (Ullstrup, 1961). Texas male-sterile corn is the most susceptible to this illness. In wet and windy weather, the pycnidia break and spread *P. maydis* spores to the leaves of healthy plants (Ullstrup, 1961). Pycnidia are tiny, black pockets of spores that usually develop on older leaves, but may form on younger leaves in moist conditions (Ullstrup, 1961).

2.4.4 Biocontrol

The most effective solution for the control of yellow leaf blight is the use of resistant hybrids, as well as clean plowing and tilling to bury debris (Ullstrup, 1961). Furthermore, growth of *P. maydis* can be reduced by using natural fungicides or crop rotation.

3

Soybean

3.1 Charcoal rot

3.1.1 Description

Charcoal rot is a very devastating disease; it led to the loss of over 30 million bushels of soybean in 2007 (Wrather & Koenning, 2009). It effects the roots and stem of plants, causing the bark to peel back and turn a grayish-black (Dunleavy et al., 1966). The disease effects a number of commercial crops in addition to soybean.

3.1.2 Geography and Soil Type

This illness is widespread in the warmer parts of the United States and Europe (Dunleavy et al., 1966). It is reported to have caused 50% of all soybean losses in Yugoslavia (Acimovic, 1963) and \$173.80 million in damages in the United States during 2002 (Islam et al., 2012). Meyer et al. (1974) report that temperature only has an effect when charcoal rot infects seeds, not mature plants. Seedling infection was highest at 30 and 35°C (Meyer et al., 1974). Charcoal rot causes severe disease when plant growth is already stunted by hot weather or poor soil (Dunleavy et al., 1966).

3.1.3 Cause

Charcoal rot is a soilborne fungal illness caused by *Macrophomina phaseolina* (Dunleavy et al., 1966). This fungus can infect over 500 plant species

worldwide, using a wide range of enzymes and toxins to do so (Islam et al., 2012). *M. phaseolina* secretes a cocktail of carbohydrate-degrading enzymes to break apart soybean tissue and enter the plant (Islam et al., 2012). It then produces a large amount of mycelium that plug up pores and blood vessels in the host plant, forcing it to wilt and turn grayish-black (Islam et al., 2012).

3.1.4 Biocontrol

Khaledi and Taheri reported in 2016 that *Trichoderma harzianum* isolates T7 and T14 inhibited the growth of *M. phaseolina*. *In vitro*, *T. harzianum* T7 inhibited mycelial growth by 58.67%. Additionally, T7 reduced disease more effectively when applied to seeds or soil, not directly to the plant in live experiments (Khaledi & Taheri, 2016).

Rhizobium japonicum was also found to be a biocontrol agent against charcoal rot (Al-Ani et al., 2012). It increased seed germination percentages and lowered the chance of infection in pot tests, as well as reducing the severity of infection when it did occur (Al-Ani et al., 2012). Seeds should be coated in *R. japonicum* before they are exposed to *M. phaseolina* for the best protection (Al-Ani et al., 2012).

3.2 Anthracnose

3.2.1 Description

Anthrachnose is a minor soybean disease that caused the loss of over 10 million bushels in 2007 (Wrather & Koenning, 2009). Infected seeds usually never germinate and infected plants are killed by numerous dark brown lesions and cankers (Dunleavy et al., 1966).

3.2.2 Geography and Soil Type

Dunleavy reports in his book *Soybean Diseases* that anthracnose is present throughout the major soybean-growing areas of the United States. He found that the disease is most destructive during rainy periods in late summer. Additionally, he reports that the disease survives “from one growing season to the next in diseased stems left in the field ... [and] overwinter[s] on infected seeds and ... [is] spread by planting diseased seeds in new localities.”

3.2.3 Cause

Two fungi cause anthracnose: *Glomerella glycines* and *Colletotrichum truncatum* (Dunleavy et al., 1966). Of the two, the latter is much more common (Dunleavy et al., 1966). While growing, the disease shows no external signs until it sporulates and invades the host plant's outer tissue (Dunleavy et al., 1966). Lower branches and leaves are killed by the fungus, as well as younger pods; seeds from pods higher up are shriveled and moldy (Dunleavy et al., 1966).

3.2.4 Biocontrol

A study in 2003 by Saber et al. found that *Trichoderma lignorum* and *Trichoderma viride* reduced disease severity in soybean plants infected with anthracnose. The bacterium also increased fresh weight, dry weight, number of pods and seeds per plant, and weight of seeds per plant (Saber et al., 2003). Another study testing various common biocontrol agents against *C. truncatum* found that *T. viride* inhibited 79.4% of the pathogen, followed by *T. hamatum* and *Pseudomonas fluorescens* with 73.74% and 69.31% pathogen inhibition, respectively (Jagtap et al., 2012).

3.3 Frogeye Leaf Spot (FLS)

3.3.1 Description

Frogeye leaf spot is primarily a fungal leaf disease (Dunleavy et al., 1966). Eyespot shaped lesions with a light tan or gray center area and a reddish-brown border form on the leaves of infected plants (Dunleavy et al., 1966). Under optimal growing conditions, the disease can also effect seeds, pods, and stems (Mian et al., 2008).

3.3.2 Geography and Soil Type

FLS prefers hot and humid growing conditions (Mian et al., 2008). It is usually contained to the southern United States, although it may spread north when certain conditions are met (Dunleavy et al., 1966). Lesions caused by FLS are reduced in soil that has been irrigated with acid with a pH of 2 (Walker et al., 1994). The disease overwinters in the seeds, leaves,

and stems of infected plants (Dunleavy et al., 1966). The occurrence of lesions peaks at 25-30°C and 90% relative humidity (Mian et al., 2008).

3.3.3 Cause

Cercospora sojina Hara is the fungus responsible for frog-eye leaf spot (Mian et al., 2008). Two races of the fungus are capable of causing the illness (Dunleavy et al., 1966). Seed infection initially occurs through direct contact with lesions or through seed to seed during the growing season (Mian et al., 2008). *C. sojina*'s spores can be carried short distances by wind, leading to increased infection under favorable conditions (Mian et al., 2008). Lesions do not appear for up to 2 weeks after infection, although the time lessens as temperature increases (Mian et al., 2008).

3.3.4 Biosolution

Two *Bacillus* strains (BNM340, and BNM122) were found to inhibit *C. sojina* growth (Simonetti et al., 2012). They inhibited growth by 52-53% *in vitro* and were similarly effective when applied to seedlings (Simonetti et al., 2012). In a separate study, strain CHERP5 also enhanced soybean resistance to FLS and reduced disease severity (Tonelli & Fabra, 2014).

3.4 Phytophthora rot

3.4.1 Description

Phytophthora rot is a fungal disease that is particularly dangerous because it affects soybean at any stage of its development (Schmitthenner, 2000). It led to the loss of over 50 million bushels of soybean each year in the United States from 2003 to 2006 (Wrather & Koenning, 2009). If seedlings are infected, then the plant is quickly killed by soft rot of the stem (Dunleavy et al., 1966). The first symptoms are wilting leaves and a dark brown lesion on the stem in older plants (Schmitthenner, 2000).

3.4.2 Geography and Soil Type

This illness is prevalent in poorly drained soil and during wet growing seasons (Dunleavy et al., 1966). It can also occur in well-drained soil if

plants were flooded when they were young (Schmitthenner, 2000). It has been reported throughout the United States, as well as Argentina, Australia, Brazil, Canada, China, Hungary, Italy, Japan, and Russia (Schmitthenner, 2000). Phytophthora rot prefers temperatures at about 25°C for germination and 25-28°C for mycelium growth (Schmitthenner, 2000). Schmitthenner reported in 2000 that “phytophthora root rot is most common in heavy, tightly compacted, fine (clay) soil or soils with an impervious, shallow hardpan that are subject to saturation and flooding.”

3.4.3 Cause

Phytophthora sojae is the fungus that causes phytophthora rot (Dunleavy et al., 1966). There are 37 races of the pathogen and a number of raceless isolates (Schmitthenner, 2000). Over 80 species of *Phytophthora* can rot many commercial crops, although *P. sojae* is limited to soybean (Tyler, 2007).

3.4.4 Biocontrol

Several *Bacillus* races have been tested for antagonistic behavior against phytophthora rot. In greenhouse tests, *Bacillus altitudinis* JSCX-1 was about 63.94% effective at controlling *P. sojae* (Lu et al., 2017). Lesions on leaves were reduced by 61.11% in the same study. When applied to seedlings, JSCX-1 protected against infection with an efficiency of 75.94%. The *Bacillus* isolate promoted genetic resistance in soybean plants and increased yields (Lu et al., 2017).

4

Peanuts

4.1 Blackhull

4.1.1 Description

Blackhull is a superficial, black discoloration of peanut shells (Kokalis-Burelle et al., 1997). Smaller black spots usually combine into larger, darker lesions that eventually envelop the entire shell (Kokalis-Burelle et al., 1997). These spots may also be found on the pegs and roots in later stages of the disease (Kokalis-Burelle et al., 1997).

4.1.2 Geography and Soil Type

Blackhull can survive for a long time in soil by producing resistant spores (Damicone et al., 2014). It is common in the southern United States, especially in New Mexico. This disease heavily favors low temperatures late in the growing season, high soil moisture, high-density plantings, and the rotation to peanuts after cotton or sweet potatoes (Kokalis-Burelle et al., 1997).

4.1.3 Cause

Chalara elegans (*Thielaviopsis basicola*) causes blackhull and black rot in a number of crops, including peanuts (Kokalis-Burelle et al., 1997). peanuts are infected immediately following spore germination, and new spore production often occurs in the following 72 hours (Kokalis-Burelle et al., 1997). The

disease can very quickly spread between crops when the correct conditions for germination are met. Kokalis-Burelle et al. report that the “initial inoculum arises from infected hulls and other plant parts left in the field from previous seasons.”

4.1.4 Biocontrol

The fungicides benomyl and thiophanate-methyl have been shown to reduce blackhull occurrence in peanuts (Kokalis-Burelle et al., 1997). Furthermore, crop rotations with sorghum have been found to reduce blackhull as well (Damicone et al., 2014). There are no commercial control methods specifically targeting *C. elegans*, however.

In cotton, *Paenibacillus alvei* strain K-165 inhibited *C. elegans* growth and reduced lesions on cotton seedlings (Schoina et al., 2011). Quantity of the blackhull fungus was reduced by 25% (Schoina et al., 2011). In the study, “It was shown that a seed coating treatment consisting of K-165 xanthan gum and talc was the most effective in reducing disease symptoms and increasing plant height and fresh weight”.

4.2 Crown Rot

4.2.1 Description

Crown rot is a particularly devastating disease in peanuts. In some fields, losses have been reported to be as high as 50% (Melouk & Damicone, 1997), although incidence is usually 2-14% (Kishore et al., 2001). It mainly causes wilting of several branches and the death of peanut seedlings following heavy progression of the rot (Suzui & Makino, 1980). It is most prevalent in the lower portion of the plant, right above the roots but below the branches (Suzui & Makino, 1980). In later stages, it can also affect the lower branches and upper roots, forming yellow lesions before turning black (Suzui & Makino, 1980).

4.2.2 Geography and Soil Type

Crown rot of peanuts was first reported in Sumatra in 1926 (Anderson, 1926). Since then, it has spread to all major peanut-growing areas worldwide (Melouk & Damicone, 1997). Gibson reported in 1953 that “seedlings raised

at 30-37°C were most disposed to severe development of crown rot, and the progress of the disease was most rapid at these temperatures.” It was also found that the effects of the disease were most severe in light textured soil (Gibson, 1953).

4.2.3 Cause

The fungus *Aspergillus niger* is the main cause of crown rot (Gibson, 1953), although several other fungi have also been isolated from infected plants (Suzui & Makino, 1980). From these, however, *A. niger* was the most common cause, affecting the vast majority of the peanuts studied.

4.2.4 Biocontrol

Bacteria of the family *Trichoderma* was found to be widely antagonistic towards *A. niger* by Kishore et al. in 2001. Of the 16 isolates they studied, *T. harzianum* was the most effective, inhibiting crown rot 38.6% of the time pre-emergence in groundnut seedlings. Isolate *T. viride* was also very effective in controlling *A. niger* infection.

In another study, *Paenibacillus polymyxa* was found to greatly reduce the incidence of crown rot in the field (Haggag & Timmusk, 2008). By cultivating it in the soil around the seedlings’ roots, researchers were able to prevent the growth of *A. niger* and the onset of crown rot.

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