

Caitlin Kramer

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Education

- **Northeastern University**
College of Science, B.S./M.S. Chemistry

Boston, MA
2011 - 2016

Experience

- **DNA Damage Recognition and Tolerance Laboratory**

MS Student, previously Undergraduate Researcher

Supervised by Dr. Penny Beuning

Northeastern University

May 2012 – Present

- Developed protocols for expression and purification of multiple proteins.
- Led research on the uncharacterized *E. coli* DNA damage response gene *ybfE* and confirmed a computational prediction of DNA binding using *in vitro* methods. Structurally characterized the protein using site-directed mutagenesis, circular dichroism, and molecular modeling. Conducted *in vivo* assays to elucidate the gene's functional role.
- Investigated the putative Y-family DNA polymerase DinB from plant symbiont *Sinorhizobium meliloti*. Cloned gene into pET expression vector. Troubleshoot problems with protein solubility to acquire viable protein for characterization. Assayed protein activity by primer extension.
- Designed experimental protocols for damaging DNA *in vivo* and *in vitro* with nitrofurazone and benzyl bromide. Prepared samples for LC-MS characterization of resulting adducts.
- Mentored and trained undergraduate students in the lab.

- **GlaxoSmithKline**

Library Chemistry Co-op

Supervised by Dr. Josephine Yuen

Boston MDR

January 2013 – June 2013

- Participated in development work, production, and quality control of a 49 million member purine-scaffold library using ELT (Encoded Library Technology).
- Ensured general reaction scheme was valid prior to library production.
- Checked that ligation of DNA tags was not impeded by other reactants.
- Determined the scope of reactive chemical building blocks to be used in production.

- **InfoCommons Service Desk**

Student Staff

Supervised by Mike Horgan

Northeastern University

September 2011 – Present

- Assisted customers in person and on the phone with addressing computer problems on personal and lab computers.
 - Kept printers and lab computers in working order.
 - Communicated with managers to provide the most up-to-date information to customers and other staff.
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Skills and Qualifications

- **Chemical biology:** PCR, site-directed mutagenesis, molecular cloning, gel electrophoresis, Western Blot, protein expression (including pET system) and purification (AKTA), primer extension (³²P), differential scanning fluorimetry, DNA purification, cell culture, sterile technique, bacterial transformation, sequencing data analysis
 - **Analytical:** Circular dichroism, NMR (Varian), LC-MS, HPLC, UV/Vis, IR, TLC, data analysis
 - **Organic:** Crystallization, separation, extraction, distillation, TLC, anhydrous technique, purification, characterization, library construction, quality control
 - **Computer:** Linux, PC, Mac, Office suite, SQL, Racket, Python, emacs, LaTeX, Pymol, YASARA, Entrez Direct, EndNote
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Conferences and Presentations

- Matz Research Symposium 2015 ◇ 20 minute oral presentation given to Northeastern alumni, faculty and students.
 - ASBMB 2015 Annual Meeting ◇ Boston. Presented poster: “Expression and purification of putative Y-family polymerase DinB from *Sinorhizobium meliloti*”
 - ASBMB 2016 Annual Meeting ◇ San Diego. Presenting poster: “Characterization of the *E. coli* SOS response protein YbfE”
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Awards

- Matz Research Scholarship ◇ 2014. Competitive grant given to undergraduates used to fund 6 month full-time research co-op.
 - Provost Thesis Grant ◇ 2015. Funding awarded selectively to graduate students based on written proposals.
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Affiliations

- American Chemical Society, since 2015
- American Society for Biochemistry and Molecular Biology, since 2015.