

Browse Networks in the Matlab Command Window Using surfNet

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INTRODUCTION

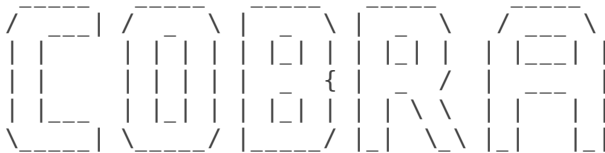
In this tutorial, we will demonstrate how to browse a COBRA model in verbal format in the Matlab command window through an initial call and interactive mouse clicking.

MATERIALS

EQUIPMENT SETUP

Start CobraToolbox

```
initCobraToolbox;
```



COntstraint-Based Reconstruction and Analysis
The COBRA Toolbox - 2017

Documentation:
<http://opencobra.github.io/cobratoolbox>

```
> Checking if git is installed ... Done.
> Checking if the repository is tracked using git ... Done.
> Checking if curl is installed ... Done.
> Checking if remote can be reached ... Done.
> Initializing and updating submodules ... Done.
> Adding all the files of The COBRA Toolbox ... Done.
> Define CB map output... set to svg.
> Retrieving models ... Done.
> TranslateSBML is installed and working properly.
> Configuring solver environment variables ...
- [-*--] ILOG_CPLEX_PATH: /Users/sxc554/Applications/IBM/ILOG/CPLEX_Studio127/cplex/matlab/x86-64_osx
- [*--] GUROBI_PATH: /Library/gurobi700/mac64/matlab
- [----] TOMLAB_PATH : --> set this path manually after installing the solver ( see instructions )
- [-*--] MOSEK_PATH: /Users/sxc554/mosek/7/toolbox/r2013aom
Done.
> Checking available solvers and solver interfaces ... Done.
> Setting default solvers ... Done.
> Saving the MATLAB path ... Done.
- The MATLAB path was saved in the default location.
```

```
> Summary of available solvers and solver interfaces
```

Support	LP	MILP	QP	MIQP	NLP	
cplex_direct	full			0	0	0
dqqMinos	full			1	-	-
glpk	full			1	1	-
gurobi	full			1	1	1
ibm_cplex	full			0	0	-
matlab	full			1	-	-

mosek	full	1	1	1	-	-
pdco	full	1	-	1	-	-
quadMinos	full	1	-	-	-	1
tomlab_cplex	full	0	0	0	0	-
qpng	experimental	-	-	1	-	-
tomlab_snopt	experimental	-	-	-	-	0
gurobi_mex	legacy	0	0	0	0	-
lindo_old	legacy	0	-	-	-	-
lindo_legacy	legacy	0	-	-	-	-
lp_solve	legacy	1	-	-	-	-
opti	legacy	0	0	0	0	0

Total	-	8	3	4	1	2

+ Legend: - = not applicable, 0 = solver not compatible or not installed, 1 = solver installed.

```
> You can solve LP problems using: 'dqqMinos' - 'glpk' - 'gurobi' - 'matlab' - 'mosek' - 'pdco' - 'quadMinos'
> You can solve MILP problems using: 'glpk' - 'gurobi' - 'mosek'
> You can solve QP problems using: 'gurobi' - 'mosek' - 'pdco' - 'qpng'
> You can solve MIQP problems using: 'gurobi'
> You can solve NLP problems using: 'matlab' - 'quadMinos'

> Checking for available updates ...
--> You cannot update your fork using updateCobraToolbox(). [b1da0e @ add-tutorial-browseNetwork].
Please use the MATLAB.devTools (https://github.com/opencobra/MATLAB.devTools).
```

PROCEDURE

Load the *E. coli* iJO1366 model as an example model.

```
load('iJO1366.mat')
```

Browse a network

Browse the network by starting from an initial metabolite, e.g., D-glucose in the extracellular compartment.

```
surfNet(iJO1366, 'glc__D_e')
```

```
Met #1195 glc__D_e, D-Glucose, C6H12O6
Consuming reactions:
#164 EX_glc__D_e, Bd: -10 / 1000, D-Glucose exchange
glc__D_e <=>
#1355 GLCtex_copy1, Bd: -1000 / 1000, Glucose transport via diffusion (extracellular to periplasm)
glc__D_e <=> glc__D_p
#1356 GLCtex_copy2, Bd: 0 / 1000, Glucose transport via diffusion (extracellular to periplasm)
glc__D_e -> glc__D_p
Producing reactions: none

Show previous steps...
```

All reactions producing or consuming '**glc__D_e**' will have their reaction indices (#xxx), ids (.rxns), bounds (.lb/.ub), names (.rxnNames) and formulae printed on the command window. All reactions and the participating metabolites are hyperlinked. For example, **click** on the reaction '**GLCtex_copy1**'. (This is equivalent to run the following command.)

```
surfNet([], 'GLCtex_copy1', 0, 'none', 0, 1, [], 0) % called by clicking 'GLCtex_copy1'
```

```
Rxn #1355 GLCtex_copy1, Bd: -1000 / 1000, Glucose transport via diffusion (extracellular to periplasm)
```

	glc__D_e	glc__D_p	
	id	Met	Stoich metNames, metFormulas
Reactant:			
#1195	glc__D_e	-1	D-Glucose, C6H12O6
Product:			
#1587	glc__D_p	1	D-Glucose, C6H12O6

[Show previous steps...](#)

Details for the metabolites will appear, e.g., indeices, ids, stoichiometric coefficients, names and chemical formulae. By iteratively clicking on the reactions and metabolites that you are interested in, you can browse through the metabolic network.

Now, say you have gone through a series of metabolites and reactions (glc__D_e, GLCtex_copy1, glc__D_p, GLCptspp, g6p_c):

Click glc__D_p:

```
surfNet([], 'glc__D_p', 0, 'none', 0, 1, [], 0) % called by clicking 'glc__D_p'
```

```
Met #1587 glc__D_p, D-Glucose, C6H12O6
Consuming reactions:
#1336 GLCDpp, Bd: 0 / 1000, Glucose dehydrogenase (ubiquinone-8 as acceptor) (periplasm)
q8_c + glc__D_p + h2o_p -> q8h2_c + glcn_p + h_p
#1352 GLCabcpp, Bd: 0 / 1000, D-glucose transport via ABC system (periplasm)
atp_c + h2o_c + glc__D_p -> adp_c + glc__D_c + h_c + pi_c
#1353 GLCptspp, Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)
pep_c + glc__D_p -> g6p_c + pyr_c
#1354 GLCt2pp, Bd: 0 / 1000, D-glucose transport in via proton symport (periplasm)
glc__D_p + h_p -> glc__D_c + h_c
Producing reactions:
#1252 G1PPpp, Bd: 0 / 1000, Glucose-1-phosphatase
glp_p + h2o_p -> glc__D_p + pi_p
#1355 GLCtex_copy1, Bd: -1000 / 1000, Glucose transport via diffusion (extracellular to periplasm)
glc__D_e <=> glc__D_p
#1356 GLCtex_copy2, Bd: 0 / 1000, Glucose transport via diffusion (extracellular to periplasm)
glc__D_e -> glc__D_p
#1607 LACZpp, Bd: 0 / 1000, B-galactosidase
h2o_p + lcts_p -> gal_p + glc__D_p
#2463 TREHpp, Bd: 0 / 1000, Alpha,alpha-trehalase (periplasm)
h2o_p + tre_p -> 2 glc__D_p
```

[Show previous steps...](#)

Click GLCptspp:

```
surfNet([], 'GLCptspp', 0, 'none', 0, 1, [], 0) % called by clicking 'GLCptspp'
```

```
Rxn #1353 GLCptspp, Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)
pep_c + glc__D_p -> g6p_c + pyr_c
id Met Stoich metNames, metFormulas
Reactant:
#784 pep_c -1 Phosphoenolpyruvate, C3H2O6P
#1587 glc__D_p -1 D-Glucose, C6H12O6
Product:
#508 g6p_c 1 D-Glucose 6-phosphate, C6H11O9P
#853 pyr_c 1 Pyruvate, C3H3O3
```

[Show previous steps...](#)

Click g6p_c:

```
surfNet([], 'g6p_c', 0, 'none', 0, 1, [], 0) % called by clicking 'g6p_c'
```

Met #508 g6p_c, D-Glucose 6-phosphate, C6H11O9P

Consuming reactions:

```
#1283 G6PDH2r, Bd: -1000 / 1000, Glucose 6-phosphate dehydrogenase
g6p_c + nadp_c <=> 6pgl_c + h_c + nadph_c
#1284 G6PP, Bd: 0 / 1000, Glucose-6-phosphate phosphatase
g6p_c + h2o_c -> glc__D_c + pi_c
#2077 PGI, Bd: -1000 / 1000, Glucose-6-phosphate isomerase
g6p_c <=> f6p_c
#2461 TRE6PS, Bd: 0 / 1000, Alpha,alpha-trehalose-phosphate synthase (UDP-forming)
g6p_c + udpg_c -> h_c + tre6p_c + udp_c
```

Producing reactions:

```
#477 AB6PGH, Bd: 0 / 1000, Arbutin 6-phosphate glucohydrolase
arbt6p_c + h2o_c -> g6p_c + hqn_c
#1214 FFSD, Bd: 0 / 1000, Beta-fructofuranosidase
h2o_c + suc6p_c -> fru_c + g6p_c
#1231 FRULYSDG, Bd: -1000 / 1000, Fructoselysine phosphate deglycase
frulysp_c + h2o_c <=> g6p_c + lys__L_c
#1285 G6Pt6_2pp, Bd: 0 / 1000, Glucose-6-phosphate transport via phosphate antiport (periplasm)
2 pi_c + g6p_p -> g6p_c + 2 pi_p
#1353 GLCptspp, Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)
pep_c + glc__D_p -> g6p_c + pyr_c
#1500 HEX1, Bd: 0 / 1000, Hexokinase (D-glucose:ATP)
atp_c + glc__D_c -> adp_c + g6p_c + h_c
#2082 PGMt, Bd: -1000 / 1000, Phosphoglucomutase
glp_c <=> g6p_c
#2459 TRE6PH, Bd: 0 / 1000, Trehalose-6-phosphate hydrolase
h2o_c + tre6p_c -> g6p_c + glc__D_c
```

[Show previous steps...](#)

In each click, there is also a button '**Show previous steps...**' at the bottom. Clicking on it will show the metabolites and reactions that you have visited in order. This is equivalent to calling:

```
surfNet([], [], 0, 'none', 0, 1, [], 0, struct('showPrev', true)) % called by clicking 'Show
```

```
glc__D_e>>GLCtex_copy1>>glc__D_p>>GLCptspp>>g6p_c>>
```

You can go back to any of the intermediate metabolites/reactions by clicking the hyperlinked mets/rxns shown.

Call options

Shown below are various call options for including flux vectors and customizing display. All call options are preserved during the interactive browsing by mouse clicking.

Show objective reactions

Omit the 'metrxn' (2nd) argument to print objective reactions:

```
surfNet(iJ01366)
```

```
Rxn #8 BIOMASS_Ec_iJ01366_core_53p95M, Bd: 0 / 1000, E. coli biomass objective function (iJ01366) - co
0.000223 10fthf_c + 2.6e-05 2fe2s_c + 0.000223 2ohph_c + 0.00026 4fe4s_c + 0.51369 ala__L_c + 0.000223 a
0.29579 arg__L_c + 0.24105 asn__L_c + 0.24105 asp__L_c + 54.1248 atp_c + 0.000122 bmocogdp_c + 2e-06 b
0.005205 ca2_c + 0.005205 cl_c + 0.000576 coa_c + 2.5e-05 cobalt2_c + 0.13351 ctp_c + 0.000709 cu2_c +
0.09158 cys__L_c + 0.026166 datp_c + 0.027017 dctp_c + 0.027017 dgtp_c + 0.026166 dttp_c + 0.000223 fa
0.006715 fe2_c + 0.007808 fe3_c + 0.26316 gln__L_c + 0.26316 glu__L_c + 0.61264 gly_c + 0.2151 gtp_c +
```

```

48.6015 h2o_c + 0.094738 his__L_c + 0.29053 ile__L_c + 0.19519 k_c + 0.45053 leu__L_c + 0.34316 lys__L_c
0.15369 met__L_c + 0.008675 mg2_c + 0.000223 mlthf_c + 0.000691 mn2_c + 7e-06 mobd_c + 0.001831 nad_c
0.000447 nadp_c + 0.013013 nh4_c + 0.000323 ni2_c + 0.017868 pel60_c + 0.054154 pel61_c + 0.18527 phe
0.000223 pheme_c + 0.22106 pro__L_c + 0.000223 pydx5p_c + 0.000223 ribflv_c + 0.21579 ser__L_c + 0.000
+ 0.004338 so4_c + 0.000223 thf_c + 0.000223 thmpp_c + 0.25369 thr__L_c + 0.056843 trp__L_c + 0.1379 t
5.5e-05 udcpgdp_c + 0.1441 utp_c + 0.42316 val__L_c + 0.000341 zn2_c + 0.019456 kdo2lipid4_e +
0.013894 murein5px4p_p + 0.045946 pel60_p + 0.02106 pel61_p -> 53.95 adp_c + 53.95 h_c + 53.9457 pi_c
0.7739 ppi_c

```

id	Met	Stoich	metNames, metFormulas
Reactant:			
#1	10fthf_c	-0.000223	10-Formyltetrahydrofolate, C20H21N7O7
#69	2fe2s_c	-0.000026	[2Fe-2S] iron-sulfur cluster, S2Fe2
#82	2ohph_c	-0.000223	2-Octaprenyl-6-hydroxyphenol, C46H70O2
#167	4fe4s_c	-0.00026	[4Fe-4S] iron-sulfur cluster, S4Fe4
#255	ala__L_c	-0.513689	L-Alanine, C3H7NO2
#265	amet_c	-0.000223	S-Adenosyl-L-methionine, C15H23N6O5S
#294	arg__L_c	-0.295792	L-Arginine, C6H15N4O2
#298	asn__L_c	-0.241055	L-Asparagine, C4H8N2O3
#302	asp__L_c	-0.241055	L-Aspartate, C4H6NO4
#307	atp_c	-54.124831	ATP, C10H12N5O13P3
#314	bmocogdp_c	-0.000122	Bis-molybdopterin guanine dinucleotide, C40H44N20O27P4S4Mo
#317	btn_c	-0.000002	Biotin, C10H15N2O3S
#326	ca2_c	-0.005205	Calcium, Ca
#355	cl_c	-0.005205	Chloride, Cl
#358	coa_c	-0.000576	Coenzyme A, C21H32N7O16P3S
#359	cobalt2_c	-0.000025	Co2+, Co
#377	ctp_c	-0.133508	CTP, C9H12N3O14P3
#379	cu2_c	-0.000709	Cu2+, Cu
#383	cys__L_c	-0.09158	L-Cysteine, C3H7NO2S
#392	datp_c	-0.026166	DATP, C10H12N5O12P3
#401	dctp_c	-0.027017	DCTP, C9H12N3O13P3
#412	dgtc_c	-0.027017	DGTP, C10H12N5O13P3
#451	dtcp_c	-0.026166	DTTP, C10H13N2O14P3
#468	fad_c	-0.000223	Flavin adenine dinucleotide oxidized, C27H31N9O15P2
#474	fe2_c	-0.006715	Fe2+, Fe
#475	fe3_c	-0.007808	Fe3+, Fe

Call with a list of mets/rxns

The 'metrxn' argument can be a string of id for a metabolite or reaction. It can also be a cell array of ids, e.g.,

```
surfNet(iJ01366, {'glc__D_p'; 'GLCptspp'; 'g6p_c'})
```

Met #1587 glc__D_p, D-Glucose, C6H12O6

Consuming reactions:

```

#1336 GLCDpp, Bd: 0 / 1000, Glucose dehydrogenase (ubiquinone-8 as acceptor) (periplasm)
q8_c + glc__D_p + h2o_p -> q8h2_c + glcn_p + h_p
#1352 GLCabcpp, Bd: 0 / 1000, D-glucose transport via ABC system (periplasm)
atp_c + h2o_c + glc__D_p -> adp_c + glc__D_c + h_c + pi_c
#1353 GLCptspp, Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)
pep_c + glc__D_p -> g6p_c + pyr_c
#1354 GLCt2pp, Bd: 0 / 1000, D-glucose transport in via proton symport (periplasm)
glc__D_p + h_p -> glc__D_c + h_c

```

Producing reactions:

```

#1252 G1PPpp, Bd: 0 / 1000, Glucose-1-phosphatase
glp_p + h2o_p -> glc__D_p + pi_p
#1355 GLCtex_copy1, Bd: -1000 / 1000, Glucose transport via diffusion (extracellular to periplasm)
glc__D_e <=> glc__D_p
#1356 GLCtex_copy2, Bd: 0 / 1000, Glucose transport via diffusion (extracellular to periplasm)
glc__D_e -> glc__D_p
#1607 LACZpp, Bd: 0 / 1000, B-galactosidase
h2o_p + lcts_p -> gal_p + glc__D_p
#2463 TREHpp, Bd: 0 / 1000, Alpha,alpha-trehalase (periplasm)

```

`h2o_p + tre_p -> 2 glc__D_p`

Rxn #1353 GLCptspp, Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)

`pep_c + glc__D_p -> g6p_c + pyr_c`

id Met Stoich metNames, metFormulas

Reactant:

#784 pep_c -1 Phosphoenolpyruvate, C3H206P

#1587 glc__D_p -1 D-Glucose, C6H1206

Product:

#508 g6p_c 1 D-Glucose 6-phosphate, C6H1109P

#853 pyr_c 1 Pyruvate, C3H303

Met #508 g6p_c, D-Glucose 6-phosphate, C6H1109P

Consuming reactions:

#1283 G6PDH2r, Bd: -1000 / 1000, Glucose 6-phosphate dehydrogenase

`g6p_c + nadp_c <=> 6pgl_c + h_c + nadph_c`

#1284 G6PP, Bd: 0 / 1000, Glucose-6-phosphate phosphatase

`g6p_c + h2o_c -> glc__D_c + pi_c`

#2077 PGI, Bd: -1000 / 1000, Glucose-6-phosphate isomerase

`g6p_c <=> f6p_c`

#2461 TRE6PS, Bd: 0 / 1000, Alpha,alpha-trehalose-phosphate synthase (UDP-forming)

`g6p_c + udpg_c -> h_c + tre6p_c + udp_c`

Producing reactions:

#477 AB6PGH, Bd: 0 / 1000, Arbutin 6-phosphate glucohydrolase

`arbt6p_c + h2o_c -> g6p_c + hqn_c`

#1214 FFSD, Bd: 0 / 1000, Beta-fructofuranosidase

`h2o_c + suc6p_c -> fru_c + g6p_c`

#1231 FRULYSDG, Bd: -1000 / 1000, Fructoselysine phosphate deglycase

`frulysp_c + h2o_c <=> g6p_c + lys__L_c`

#1285 G6Pt6_2pp, Bd: 0 / 1000, Glucose-6-phosphate transport via phosphate antiport (periplasm)

`2 pi_c + g6p_p -> g6p_c + 2 pi_p`

#1353 GLCptspp, Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)

`pep_c + glc__D_p -> g6p_c + pyr_c`

#1500 HEX1, Bd: 0 / 1000, Hexokinase (D-glucose:ATP)

`atp_c + glc__D_c -> adp_c + g6p_c + h_c`

Show metabolite names in reaction formulae

Some models may use generic ids for mets/rxns. In this case, call surfNet with the 'metNameFlag' (3rd) argument turned on to show the names for metabolites (.metNames) in the reaction formulae, e.g.,

```
surfNet(iJ01366, 'fgam_c', 1)
```

Met #484 fgam_c, N2-Formyl-N1-(5-phospho-D-ribosyl)glycinamide, C8H13N209P

Consuming reactions:

#2207 PRFGS, Bd: 0 / 1000, Phosphoribosylformylglycinamide synthase

`ATP + N2-Formyl-N1-(5-phospho-D-ribosyl)glycinamide + L-Glutamine + H2O -> ADP + 2-(Formamido)-N1-(5-phospho-D-ribosyl)glycinamide + L-Glutamate + H+ + Phosphate`

Producing reactions:

#1316 GARFT, Bd: -1000 / 1000, Phosphoribosylglycinamide formyltransferase

`10-Formyltetrahydrofolate + N1-(5-Phospho-D-ribosyl)glycinamide <=> N2-Formyl-N1-(5-phospho-D-ribosyl)glycinamide + 5,6,7,8-Tetrahydrofolate`

#1317 GART, Bd: 0 / 1000, GAR transformylase-T

`ATP + Formate + N1-(5-Phospho-D-ribosyl)glycinamide -> ADP + N2-Formyl-N1-(5-phospho-D-ribosyl)glycinamide`

[Show previous steps...](#)

Hide reaction details

Turn off the 'showMets' (6th) argument to suppress details for reactions, e.g.,

```
surfNet(iJ01366, iJ01366.rxns(1001:1010), [], [], [], 0)
```

```
Rxn #1001 DHPPDA2, Bd: 0 / 1000, Diaminohydroxyphosphoribosylaminopryrimidine deaminase (25drapp)  
25drapp_c + h_c + h2o_c -> 5apru_c + nh4_c
```

```
Rxn #1002 DHPS2, Bd: 0 / 1000, Dihydropteroate synthase  
4abz_c + 6hnhptpp_c -> dhpt_c + ppi_c
```

```
Rxn #1003 DHPTDCs2, Bd: 0 / 1000, 4,5-dihydroxy-2,3-pentanedione cyclization (spontaneous)  
dhptd_c -> mdhdf_c
```

```
Rxn #1004 DHPTDNR, Bd: 0 / 0, Dihydropteridine reductase  
dhptdn_c + 3 h_c + nadph_c -> nadp_c + thptdn_c
```

```
Rxn #1005 DHPTDNRN, Bd: 0 / 0, Dihydropteridine reductase (NADH)  
dhptdn_c + 3 h_c + nadh_c -> nad_c + thptdn_c
```

```
Rxn #1006 DHPTPE, Bd: -1000 / 1000, Dihydroneopterin triphosphate 2'-epimerase  
ahdt_c <=> dhmptp_c
```

```
Rxn #1007 DHQS, Bd: 0 / 1000, 3-dehydroquininate synthase  
2dda7p_c -> 3dhq_c + pi_c
```

```
Rxn #1008 DHQTi, Bd: 0 / 1000, 3-dehydroquininate dehydratase, irreversible  
3dhq_c -> 3dhsk_c + h2o_c
```

```
Rxn #1009 DIMPTex, Bd: -1000 / 1000, DIMP transport via diffusion (extracellular to periplasm)  
dimp_e <=> dimp_p
```

```
Rxn #1010 DINSt2pp, Bd: 0 / 1000, Deoxyinosine transport in via proton symport (periplasm)  
din_p + h_p -> din_c + h_c
```

[Show previous steps...](#)

Look at one or more flux distributions

First, get a flux distribution by optimizing the biomass production of the model (the standard flux balance analysis¹). Then call surfNet with the flux distribution (4th argument) to look at how the flux through pyruvate is distributed:

```
s = optimizeCbModel(iJ01366, 'max', 'one');  
surfNet(iJ01366, 'pyr_c', [], s.x)
```

Met #853 pyr_c, Pyruvate, C3H3O3

Consuming reactions with non-zero fluxes :

#511 ACHBS (0.28541), Bd: 0 / 1000, 2-aceto-2-hydroxybutanoate synthase
2obut_c + h_c + pyr_c -> 2ahbut_c + co2_c

#513 ACLS (0.85886), Bd: 0 / 1000, Acetolactate synthase
h_c + 2 pyr_c -> alac_S_c + co2_c

#618 ALATA_L (-0.57111), Bd: -1000 / 1000, L-alanine transaminase
akg_c + ala_L_c <=> glu_L_c + pyr_c

#987 DHDPs (0.36441), Bd: 0 / 1000, Dihydrodipicolinate synthase
aspsa_c + pyr_c -> 23dhdp_c + h_c + 2 h2o_c

#1053 DXPS (0.00279), Bd: 0 / 1000, 1-deoxy-D-xylulose 5-phosphate synthase
g3p_c + h_c + pyr_c -> co2_c + dxyl5p_c

#2047 PDH (7.96919), Bd: 0 / 1000, Pyruvate dehydrogenase
coa_c + nad_c + pyr_c -> accoa_c + co2_c + nadh_c

#2171 POR5 (0.10684), Bd: -1000 / 1000, Pyruvate synthase
coa_c + 2 flxso_c + pyr_c <=> accoa_c + co2_c + 2 flxr_c + h_c

#2466 TRPAS2 (-0.05584), Bd: -1000 / 1000, Tryptophanase (L-tryptophan)
h2o_c + trp_L_c <=> indole_c + nh4_c + pyr_c

Producing reactions with non-zero fluxes :

#554 ADCL (0.00066), Bd: 0 / 1000, 4-aminobenzoate synthase


```

4adcho_c -> 4abz_c + h_c + pyr_c
#666 ANS (0.05584), Bd: 0 / 1000, Anthranilate synthase
chor_c + gln_L_c -> anth_c + glu_L_c + h_c + pyr_c
#813 CHRPL (0.00022), Bd: 0 / 1000, Chorismate pyruvate lyase
chor_c -> 4hbz_c + pyr_c
#908 CYSTL (0.1512), Bd: 0 / 1000, Cystathionine b-lyase
cyst_L_c + h2o_c -> hcys_L_c + nh4_c + pyr_c
#978 DHAPT (0.86538), Bd: 0 / 1000, Dihydroxyacetone phosphotransferase
dha_c + pep_c -> dhap_c + pyr_c
#1353 GLCptspp (10), Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)
pep_c + glc_D_p -> g6p_c + pyr_c

```

[Show previous steps...](#)

All reactions involving pyruvate with non-zero fluxes are printed. The flux values are in the parentheses following the reaction ids. Note that reactions stated as consuming or producing the metabolite have taken the directions of the fluxes into account. Therefore, supplying a different flux distribution or not supplying may give different display. By default, only reactions with non-zero fluxes are printed if a flux distribution is supplied. Turn the 'nonzeroFluxFlag' (5th) argument off to show all reactions:

```
surfNet(iJ01366, 'pyr_c', [], s.x, 0)
```

Met #853 pyr_c, Pyruvate, C3H3O3

Consuming reactions:

```

#511 ACHBS (0.28541), Bd: 0 / 1000, 2-aceto-2-hydroxybutanoate synthase
2obut_c + h_c + pyr_c -> 2ahbut_c + co2_c
#513 ACLS (0.85886), Bd: 0 / 1000, Acetolactate synthase
h_c + 2 pyr_c -> alac_S_c + co2_c
#618 ALATA_L (-0.57111), Bd: -1000 / 1000, L-alanine transaminase
akg_c + ala_L_c <=> glu_L_c + pyr_c
#987 DHDPs (0.36441), Bd: 0 / 1000, Dihydrodipicolinate synthase
aspsa_c + pyr_c -> 23dhdp_c + h_c + 2 h2o_c
#1053 DXPS (0.00279), Bd: 0 / 1000, 1-deoxy-D-xylulose 5-phosphate synthase
g3p_c + h_c + pyr_c -> co2_c + dxyl5p_c
#2047 PDH (7.96919), Bd: 0 / 1000, Pyruvate dehydrogenase
coa_c + nad_c + pyr_c -> accoa_c + co2_c + nadh_c
#2067 PFL (0), Bd: 0 / 1000, Pyruvate formate lyase
coa_c + pyr_c -> accoa_c + for_c
#2171 POR5 (0.10684), Bd: -1000 / 1000, Pyruvate synthase
coa_c + 2 flxso_c + pyr_c <=> accoa_c + co2_c + 2 flxr_c + h_c
#2172 POX (0), Bd: 0 / 1000, Pyruvate oxidase
h2o_c + pyr_c + q8_c -> ac_c + co2_c + q8h2_c
#2198 PPS (0), Bd: 0 / 1000, Phosphoenolpyruvate synthase
atp_c + h2o_c + pyr_c -> amp_c + 2 h_c + pep_c + pi_c
#2466 TRPAS2 (-0.05584), Bd: -1000 / 1000, Tryptophanase (L-tryptophan)
h2o_c + trp_L_c <=> indole_c + nh4_c + pyr_c

```

Producing reactions:

```

#507 ACGAptspp (0), Bd: 0 / 1000, N-Acetyl-D-glucosamine transport via PEP:Pyr PTS (periplasm)
pep_c + acgam_p -> acgam6p_c + pyr_c
#516 ACMANAptspp (0), Bd: 0 / 1000, N-acetyl-D-mannosamine transport via PTS (periplasm)
pep_c + acmana_p -> acmanap_c + pyr_c
#518 ACMUMptspp (0), Bd: 0 / 1000, N-acetylmuramate transport via PEP:Pyr PTS (periplasm)
pep_c + acmum_p -> acmum6p_c + pyr_c
#522 ACNML (0), Bd: 0 / 1000, N-Acetylneuraminate lyase
acnam_c -> acmana_c + pyr_c
#554 ADCL (0.00066), Bd: 0 / 1000, 4-aminobenzoate synthase
4adcho_c -> 4abz_c + h_c + pyr_c
#617 ALATA_D2 (0), Bd: 0 / 1000, D-alanine transaminase
ala_D_c + pydx5p_c -> pyam5p_c + pyr_c
#619 ALATA_L2 (0), Bd: 0 / 1000, Alanine transaminase
ala_L_c + pydx5p_c -> pyam5p_c + pyr_c
#666 ANS (0.05584), Bd: 0 / 1000, Anthranilate synthase
chor_c + gln_L_c -> anth_c + glu_L_c + h_c + pyr_c
#698 ARBTptspp (0), Bd: 0 / 1000, Arbutin transport via PEP:Pyr PTS (periplasm)
pep_c + arbt_p -> arbt6p_c + pyr_c

```


You can also compare multiple flux distributions by supplying them in a matrix format, each column being a flux distribution. For example, get another flux distribution maximizing the biomass production using D-fructose instead of glucose as substrate. Then call `surfNet` to look at reactions with different fluxes.

Original uptake rates:

```
printUptakeBound(iJ01366);
```

```
EX_ca2_e -1000
EX_cbl1_e -0.01
EX_cl_e -1000
EX_co2_e -1000
EX_cobalt2_e -1000
EX_cu2_e -1000
EX_fe2_e -1000
EX_fe3_e -1000
EX_glc__D_e -10
EX_h_e -1000
EX_h2o_e -1000
EX_k_e -1000
EX_mg2_e -1000
EX_mn2_e -1000
EX_mobd_e -1000
EX_na1_e -1000
EX_nh4_e -1000
EX_ni2_e -1000
EX_o2_e -1000
EX_pi_e -1000
EX_sel_e -1000
EX_slnt_e -1000
EX_so4_e -1000
EX_tungs_e -1000
EX_zn2_e -1000
```

Use fructose instead of glucose as substrate:

```
iJ01366 = changeRxnBounds(iJ01366, {'EX_glc__D_e'; 'EX_fru_e'}, [0; -10], {'L'; 'L'});
printUptakeBound(iJ01366);
```

```
EX_ca2_e -1000
EX_cbl1_e -0.01
EX_cl_e -1000
EX_co2_e -1000
EX_cobalt2_e -1000
EX_cu2_e -1000
EX_fe2_e -1000
EX_fe3_e -1000
EX_fru_e -10
EX_h_e -1000
EX_h2o_e -1000
EX_k_e -1000
EX_mg2_e -1000
EX_mn2_e -1000
EX_mobd_e -1000
EX_na1_e -1000
EX_nh4_e -1000
EX_ni2_e -1000
EX_o2_e -1000
```

```
EX_pi_e -1000
EX_sel_e -1000
EX_slnt_e -1000
EX_so4_e -1000
EX_tungs_e -1000
EX_zn2_e -1000
```

Run FBA again to get a flux distribution using fructose as substrate. Then look at reactions with different fluxes in the glucose and fructose cases using surfNet.

```
sFru = optimizeCbModel(iJ01366, 'max', 'one'); % FBA
fluxMatrix = [s.x, sFru.x]; % put two flux vectors in a matrix
rxnDiff = abs(fluxMatrix(:, 1) - fluxMatrix(:, 2)) > 1e-6; % reactions with different fluxes
surfNet(iJ01366, iJ01366.rxns(rxnDiff), [], fluxMatrix, [], 0)
```

```
Rxn #139 EX_fru_e (0, -10), Bd: -10 / 1000, D-Fructose exchange
fru_e <=>
```

```
Rxn #164 EX_glc__D_e (-10, 0), Bd: 0 / 1000, D-Glucose exchange
glc__D_e ->
```

```
Rxn #623 ALAt2pp_copy2 (-0.00511, 0), Bd: -1000 / 1000, L-alanine transport in via proton symport (periplasm)
ala__L_p + h_p <=> ala__L_c + h_c
```

```
Rxn #624 ALAt4pp (0.00511, 0), Bd: 0 / 1000, L-alanine transport in via sodium symport (periplasm)
ala__L_p + na1_p -> ala__L_c + na1_c
```

```
Rxn #1230 FRUK (0, 5.75203), Bd: 0 / 1000, Fructose-1-phosphate kinase
atp_c + flp_c -> adp_c + fdp_c + h_c
```

```
Rxn #1238 FRUpts2pp (0, 4.24797), Bd: 0 / 1000, Fructose transport via PEP:Pyr PTS (f6p generating) (periplasm)
pep_c + fru_p -> f6p_c + pyr_c
```

```
Rxn #1239 FRUptspp (0, 5.75203), Bd: 0 / 1000, D-fructose transport via PEP:Pyr PTS (periplasm)
pep_c + fru_p -> flp_c + pyr_c
```

```
Rxn #1240 FRUtex (-0, 10), Bd: -1000 / 1000, D-fructose transport via diffusion (extracellular to periplasm)
fru_e <=> fru_p
```

```
Rxn #1353 GLCptspp (10, 0), Bd: 0 / 1000, D-glucose transport via PEP:Pyr PTS (periplasm)
pep_c + glc__D_p -> g6p_c + pyr_c
```

```
Rxn #1356 GLCtex_copy2 (10, 0), Bd: 0 / 1000, Glucose transport via diffusion (extracellular to periplasm)
glc__D_e -> glc__D_p
```

```
Rxn #1377 GLUt2rpp (0, -0.00511), Bd: -1000 / 1000, L-glutamate transport via proton symport, reversible
glu__L_p + h_p <=> glu__L_c + h_c
```

```
Rxn #1378 GLUt4pp (0, 0.00511), Bd: 0 / 1000, Na+/glutamate symport (periplasm)
glu__L_p + na1_p -> glu__L_c + na1_c
```

```
Rxn #1758 MDH (4.82506, 4.82528), Bd: -1000 / 1000, Malate dehydrogenase
mal__L_c + nad_c <=> h_c + nadh_c + oaa_c
```

```
Rxn #1837 MOX (0.0016, 0.00138), Bd: -1000 / 1000, Malate oxidase
mal__L_c + o2_c <=> h2o2_c + oaa_c
```

```
Rxn #2048 PDX5P02 (0.00022, 0), Bd: 0 / 1000, Pyridoxine 5'-phosphate oxidase (anaerobic)
nad_c + pdx5p_c -> h_c + nadh_c + pydx5p_c
```

```
Rxn #2049 PDX5P0i (0, 0.00022), Bd: 0 / 1000, Pyridoxine 5'-phosphate oxidase
o2_c + pdx5p_c -> h2o2_c + pydx5p_c
```

```
Rxn #2064 PFK (5.75203, 0), Bd: 0 / 1000, Phosphofructokinase
atp_c + f6p_c -> adp_c + fdp_c + h_c
```

```
Rxn #2077 PGI (5.91807, -4.08193), Bd: -1000 / 1000, Glucose-6-phosphate isomerase  
g6p_c <=> f6p_c
```

[Show previous steps...](#)

Customize model data to be displayed

Customize the fields for metabolites and reactions to be printed by supplying the 'field2print' (7th) argument. It is defaulted to be:

```
{{'metNames','metFormulas'}, {'rxnNames','lb','ub'}}
```

The first cell contains the metabolite-related fields to be printed and the second cell contains the reaction-related fields to be printed. It can also be inputted as a single cell array of strings, as long as from the size (equal to #mets or #rxns) or from the name of the field (starting with 'met' or 'rxn'), the fields are recognizable to be met- or rxn-related. For example, show the grRules for rxns but omit the bounds and show the constraint sense (csense) associated with each metabolite. Note the difference from the original call:

```
surfNet(iJ01366, 'fdp_c', [], [], [], [], {'metNames','metFormulas','rxnNames','grRules','csense'})
```

```
Met #473 fdp_c, D-Fructose 1,6-bisphosphate, C6H10O12P2, csense: E  
Consuming reactions with non-zero fluxes :  
#1151 FBA (5.75203, 5.75203), Fructose-bisphosphate aldolase, grRules: b2097 or b1773 or b2925  
fdp_c <=> dhap_c + g3p_c  
Producing reactions with non-zero fluxes :  
#1230 FRUK (0, 5.75203), Fructose-1-phosphate kinase, grRules: b2168  
atp_c + flp_c -> adp_c + fdp_c + h_c  
#2064 PFK (5.75203, 0), Phosphofructokinase, grRules: b3916 or b1723  
atp_c + f6p_c -> adp_c + fdp_c + h_c
```

[Show previous steps...](#)

```
surfNet(iJ01366, 'fdp_c')
```

```
Met #473 fdp_c, D-Fructose 1,6-bisphosphate, C6H10O12P2  
Consuming reactions:  
#1151 FBA, Bd: -1000 / 1000, Fructose-bisphosphate aldolase  
fdp_c <=> dhap_c + g3p_c  
#1153 FBP, Bd: 0 / 1000, Fructose-bisphosphatase  
fdp_c + h2o_c -> f6p_c + pi_c  
Producing reactions:  
#1230 FRUK, Bd: 0 / 1000, Fructose-1-phosphate kinase  
atp_c + flp_c -> adp_c + fdp_c + h_c  
#2064 PFK, Bd: 0 / 1000, Phosphofructokinase  
atp_c + f6p_c -> adp_c + fdp_c + h_c
```

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The last argument (8th) 'nCharBreak' sets the number of characters printed per line. By default, it is equal to the width of the Matlab command window. Note the difference:

Characters per line = width of the command window (default):

```
surfNet(iJ01366, [], [], [], [], [], 0)
```

```
Rxn #8 BIOMASS_Ec_iJ01366_core_53p95M, Bd: 0 / 1000, E. coli biomass objective function (iJ01366) - cor  
0.000223 10fthf_c + 2.6e-05 2fe2s_c + 0.000223 2ohph_c + 0.00026 4fe4s_c + 0.51369 ala_L_c + 0.000223 a  
0.29579 arg_L_c + 0.24105 asn_L_c + 0.24105 asp_L_c + 54.1248 atp_c + 0.000122 bmocogdp_c + 2e-06 b  
0.005205 ca2_c + 0.005205 cl_c + 0.000576 coa_c + 2.5e-05 cobalt2_c + 0.13351 ctp_c + 0.000709 cu2_c +
```

```

0.09158 cys__L_c + 0.026166 datp_c + 0.027017 dctp_c + 0.027017 dgtp_c + 0.026166 dttp_c + 0.000223 fa
0.006715 fe2_c + 0.007808 fe3_c + 0.26316 gln__L_c + 0.26316 glu__L_c + 0.61264 gly_c + 0.2151 gtp_c +
48.6015 h2o_c + 0.094738 his__L_c + 0.29053 ile__L_c + 0.19519 k_c + 0.45053 leu__L_c + 0.34316 lys__L
0.15369 met__L_c + 0.008675 mg2_c + 0.000223 mlthf_c + 0.000691 mn2_c + 7e-06 mobd_c + 0.001831 nad_c
0.000447 nadp_c + 0.013013 nh4_c + 0.000323 ni2_c + 0.017868 pel60_c + 0.054154 pel61_c + 0.18527 phe
0.000223 pheme_c + 0.22106 pro__L_c + 0.000223 pydx5p_c + 0.000223 ribflv_c + 0.21579 ser__L_c + 0.00
+ 0.004338 so4_c + 0.000223 thf_c + 0.000223 thmpp_c + 0.25369 thr__L_c + 0.056843 trp__L_c + 0.1379 t
5.5e-05 udcpdp_c + 0.1441 utp_c + 0.42316 val__L_c + 0.000341 zn2_c + 0.019456 kdo2lipid4_e +
0.013894 murein5px4p_p + 0.045946 pel60_p + 0.02106 pel61_p -> 53.95 adp_c + 53.95 h_c + 53.9457 pi_c
0.7739 ppi_c

```

[Show previous steps...](#)

40 characters per line:

```
surfNet(iJ01366, [], [], [], [], 0, [], 40)
```

```

Rxn #8 BIOMASS_Ec_iJ01366_core_53p95M, Bd: 0 / 1000, E. coli biomass objective function (iJ01366) - co
0.000223 10fthf_c + 2.6e-05 2fe2s_c +
0.000223 2ohph_c + 0.00026 4fe4s_c +
0.51369 ala__L_c + 0.000223 amet_c +
0.29579 arg__L_c + 0.24105 asn__L_c +
0.24105 asp__L_c + 54.1248 atp_c +
0.000122 bmocogdp_c + 2e-06 btn_c +
0.005205 ca2_c + 0.005205 cl_c +
0.000576 coa_c + 2.5e-05 cobalt2_c +
0.13351 ctp_c + 0.000709 cu2_c +
0.09158 cys__L_c + 0.026166 datp_c +
0.027017 dctp_c + 0.027017 dgtp_c +
0.026166 dttp_c + 0.000223 fad_c +
0.006715 fe2_c + 0.007808 fe3_c +
0.26316 gln__L_c + 0.26316 glu__L_c +
0.61264 gly_c + 0.2151 gtp_c +
48.6015 h2o_c + 0.094738 his__L_c +
0.29053 ile__L_c + 0.19519 k_c +
0.45053 leu__L_c + 0.34316 lys__L_c +
0.15369 met__L_c + 0.008675 mg2_c +
0.000223 mlthf_c + 0.000691 mn2_c +
7e-06 mobd_c + 0.001831 nad_c +
0.000447 nadp_c + 0.013013 nh4_c +
0.000323 ni2_c + 0.017868 pel60_c +
0.054154 pel61_c + 0.18527 phe__L_c +
0.000223 pheme_c + 0.22106 pro__L_c +
0.000223 pydx5p_c + 0.000223 ribflv_c
+ 0.21579 ser__L_c + 0.000223 sheme_c
+ 0.004338 so4_c + 0.000223 thf_c +
0.000223 thmpp_c + 0.25369 thr__L_c +
0.056843 trp__L_c + 0.1379 tyr__L_c +
5.5e-05 udcpdp_c + 0.1441 utp_c +
0.42316 val__L_c + 0.000341 zn2_c +
0.019456 kdo2lipid4_e +
0.013894 murein5px4p_p +
0.045946 pel60_p + 0.02106 pel61_p ->
53.95 adp_c + 53.95 h_c + 53.9457 pi_c
+ 0.7739 ppi_c

```

[Show previous steps...](#)

80 characters per line:

```
surfNet(iJ01366, [], [], [], [], 0, [], 80)
```

```

Rxn #8 BIOMASS_Ec_iJ01366_core_53p95M, Bd: 0 / 1000, E. coli biomass objective function (iJ01366) - co

```

0.000223 10fthf_c + 2.6e-05 2fe2s_c + 0.000223 2ohph_c + 0.00026 4fe4s_c +
 0.51369 ala__L_c + 0.000223 amet_c + 0.29579 arg__L_c + 0.24105 asn__L_c +
 0.24105 asp__L_c + 54.1248 atp_c + 0.000122 bmocogdp_c + 2e-06 btn_c +
 0.005205 ca2_c + 0.005205 cl_c + 0.000576 coa_c + 2.5e-05 cobalt2_c +
 0.13351 ctp_c + 0.000709 cu2_c + 0.09158 cys__L_c + 0.026166 datp_c +
 0.027017 dctp_c + 0.027017 dgtp_c + 0.026166 dttp_c + 0.000223 fad_c +
 0.006715 fe2_c + 0.007808 fe3_c + 0.26316 gln__L_c + 0.26316 glu__L_c +
 0.61264 gly_c + 0.2151 gtp_c + 48.6015 h2o_c + 0.094738 his__L_c +
 0.29053 ile__L_c + 0.19519 k_c + 0.45053 leu__L_c + 0.34316 lys__L_c +
 0.15369 met__L_c + 0.008675 mg2_c + 0.000223 mlthf_c + 0.000691 mn2_c +
 7e-06 mobd_c + 0.001831 nad_c + 0.000447 nadp_c + 0.013013 nh4_c +
 0.000323 ni2_c + 0.017868 pel60_c + 0.054154 pel61_c + 0.18527 phe__L_c +
 0.000223 pheme_c + 0.22106 pro__L_c + 0.000223 pydx5p_c + 0.000223 ribflv_c +
 0.21579 ser__L_c + 0.000223 sheme_c + 0.004338 so4_c + 0.000223 thf_c +
 0.000223 thmpp_c + 0.25369 thr__L_c + 0.056843 trp__L_c + 0.1379 tyr__L_c +
 5.5e-05 udcpdp_c + 0.1441 utp_c + 0.42316 val__L_c + 0.000341 zn2_c +
 0.019456 kdo2lipid4_e + 0.013894 murein5px4p_p + 0.045946 pel60_p +
 0.02106 pel61_p -> 53.95 adp_c + 53.95 h_c + 53.9457 pi_c + 0.7739 ppi_c

[Show previous steps...](#)

REFERENCES

[1] Orth, J. D., Thiele I., and Palsson, B. Ø. What is flux balance analysis? *Nat. Biotechnol.*, 28(3), 245–248 (2010).