# Model manipulation

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#### INTRODUCTION

In this tutorial, we will do a manipulation with the simple model of the first few reactions of the glycolisis metabolic pathway as created in the "Model Creation" tutorial.

Glycolysis is the metabolic pathway that occurs in most organisms in the cytosol of the cell. First, we will use the beginning of that pathway to create a simple constraint-based metabolic network (Figure 1).

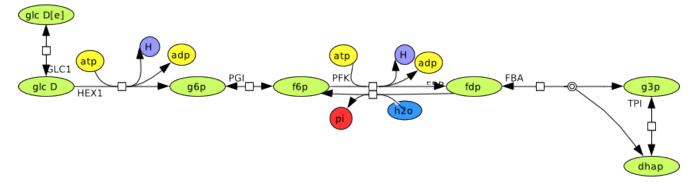


Figure 1: A small metabolic network consisting of the seven reactions in the glycolisis pathway.

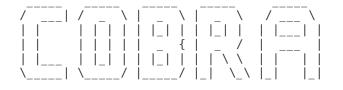
At the beginning of the reconstruction, an integrity of the draft reconstruction must be initially assessed. The accuracy of the stochiometry, necessity of each reaction and metabolite, direction and reversibility of the reactions needed to be evaluated.

After creating or loading the model, the model can be modified to simulate different conditions, such as:

- · Creating, adding and handling reactions;
- Adding Exchange, Sink and Demand reactions;
- Altering reaction bounds;
- · Altering Reactions;
- · Remove reactions and metabolites:
- · Search for duplicates and comparison of two models;
- Changing the model objective;
- Changing the direction of reaction(s);

#### Start CobraToolbox

# initCobraToolbox;



COnstraint-Based Reconstruction and Analysis The COBRA Toolbox - 2017

Documentation:

http://opencobra.github.io/cobratoolbox

> Checking if git is installed ... Done.

```
> Checking if the repository is tracked using git ... Done.
> Checking if curl is installed ... Done.
> Checking if remote can be reached ... Done.
> Initializing and updating submodules ... Done.
> Adding all the files of The COBRA Toolbox ... Done.
> Define CB map output... set to svg.
> Retrieving models ...
                          Done.
Warning: > TranslateSBML is installed but is not working properly.
> Try running
  >> TranslateSBML('/home/vanja/fork-cobratoolbox/test/models/Ec iAF1260 flux1.xml');
  in order to debug.
> Configuring solver environment variables ...
   - [----] ILOG_CPLEX_PATH : --> set this path manually after installing the solver ( see instructions
   - [-*--] GUROBI PATH: /opt/gurobi702/linux64/matlab
   - [----] TOMLAB_PATH : --> set this path manually after installing the solver ( see instructions )
   - [----] MOSEK PATH : --> set this path manually after installing the solver ( see instructions )
  Done.
> Checking available solvers and solver interfaces ... Done.
> Setting default solvers ... Done.
> Saving the MATLAB path ... Done.
   - The MATLAB path was saved as ~/pathdef.m.
> Summary of available solvers and solver interfaces
            LP MILP
                           OP MIOP
   Support
                                         NI P
```

Support	LI IIILI	Q1	TITQI	IVLI			
cplex_direct	full		0	0	0	0	-
dqqMinos	full		1	-	-	-	-
glpk	full		1	1	-	-	-
gurobi	full		1	1	1	1	-
ibm_cplex	full		0	0	0	0	-
matlab	full		1	-	-	-	1
mosek	full		0	0	0	-	-
pdco	full		1	-	1	-	1
quadMinos	full		1	-	-	-	1
tomlab cplex	full		0	0	0	0	-
qpng	experimental		-	-	1	-	-
tomlab_snopt	experimental		-	-	-	-	0
gurobi_mex	legacy		0	0	0	0	-
lindo_old	legacy		0	-	-	-	-
lindo_legacy	legacy		0	-	-	-	-
lp_solve	legacy		1	-	-	-	-
opti	legacy		0	0	0	0	0
Total	-		7	2	3	1	3

```
> You can solve LP problems using: 'dqqMinos' - 'glpk' - 'gurobi' - 'matlab' - 'pdco' - 'quadMinos' -
> You can solve MILP problems using: 'glpk' - 'gurobi'
> You can solve QP problems using: 'gurobi' - 'pdco' - 'qpng'
> You can solve MIQP problems using: 'gurobi'
> You can solve NLP problems using: 'matlab' - 'pdco' - 'quadMinos'
```

+ Legend: - = not applicable, 0 = solver not compatible or not installed, 1 = solver installed.

```
> Checking for available updates ...
```

#### Generate a network

A constraint-based metabolic model contains the stoichiometric matrix with reactions and metabolites [1].

<sup>--&</sup>gt; You cannot update your fork using updateCobraToolbox(). [ed9593 @ revised-tutorial]. Please use the MATLAB.devTools (https://github.com/opencobra/MATLAB.devTools).

*S* is stoichiometric representation of metabolic networks corresponding to the reactions in the biochemical pathway. In an each column of the *S* is a biochemical reaction and in each row is a precise metabolite. There is a stoichiometric coefficient of zero, which means that metabolite not participate in that distinct reaction. The coefficient also can be positive when the appropriate metabolite is produced, or negative for every metabolite depleted [1].

```
ReactionFormulas = {'glc-D[e] -> glc-D[c]',...
    'glc-D[c] + atp[c] -> H[c] + adp[c] + g6p[c]',...
    'g6p[c] <=> f6p[c]',...
    'atp[c] + f6p[c] -> H[c] + adp[c] + fdp[c]',...
    'fdp[c] + h2o[c] -> f6p[c] + pi[c]',...
    'fdp[c] -> g3p[c] + dhap[c]',...
    'dhap[c] -> g3p[c]'};
ReactionNames = {'GLCt1', 'HEX1', 'PGI', 'PFK', 'FBP', 'FBA', 'TPI'};
lowerbounds = [-20, 0, -20, 0, 0, -20, -20];
upperbounds = [20, 20, 20, 20, 20, 20, 20];
model = createModel(ReactionNames, ReactionNames, ReactionFormulas,...
    'lowerBoundList', lowerbounds, 'upperBoundList', upperbounds);
```

```
Warning: Metabolite glc-D[e] not in model - added to the model
Warning: Metabolite glc-D[c] not in model - added to the model
GLCt1 glc-D[e] <=> glc-D[c]
Warning: Metabolite atp[c] not in model - added to the model
Warning: Metabolite H[c] not in model - added to the model
Warning: Metabolite adp[c] not in model - added to the model
Warning: Metabolite g6p[c] not in model - added to the model
HEX1 glc-D[c] + atp[c] \rightarrow H[c] + adp[c] + g6p[c]
Warning: Metabolite f6p[c] not in model - added to the model
PGI g6p[c] \iff f6p[c]
Warning: Metabolite fdp[c] not in model - added to the model
PFK atp[c] + f6p[c] -> H[c] + adp[c] + fdp[c]
Warning: Metabolite h2o[c] not in model - added to the model
Warning: Metabolite pi[c] not in model - added to the model
FBP fdp[c] + h2o[c] \rightarrow f6p[c] + pi[c]
Warning: Metabolite g3p[c] not in model - added to the model
Warning: Metabolite dhap[c] not in model - added to the model
FBA fdp[c] \iff g3p[c] + dhap[c]
TPI dhap[c] \ll g3p[c]
```

We can now have a look at the different model fields created. The stoichiometry is stored in the S field of the model, which was described above. Since this is commonly a sparse matrix (i.e. it does contain a lot of zeros, to display it it is useful to use the full representation)

```
full(model.S)
ans =
     - 1
             0
      1
                    0
                           0
                                   0
                                                 0
            - 1
      0
                    0
                                   0
                                                 0
            - 1
                           - 1
      0
                    0
             1
                           1
                                   0
                                          0
                                                 0
      0
             1
                    0
                           1
                                   0
                                          0
                                                 0
      0
             1
                   -1
                           0
                                  0
                                          0
                                                 0
      0
             0
                    1
                           - 1
                                  1
                                          0
                                                 0
      0
             0
                    0
                           1
                                 - 1
                                         - 1
                                                 0
      0
             0
                    0
                           0
                                 - 1
                                          0
                                                 0
```

Some descriptive fields always present are model.mets and model.rxns which represent the metabolites and the reactions respectively.

```
model.mets
ans =
     'glc-D[e]'
     'glc-D[c]'
     'atp[c]'
     'H[c]'
     'adp[c]'
     'q6p[c]'
     'f6p[c]'
     'fdp[c]'
     'h2o[c]'
     'pi[c]'
     'g3p[c]'
     'dhap[c]'
model.rxns
ans =
     'GLCt1'
     'HEX1'
     'PGI'
     'PFK'
     'FBP'
     'FBA'
     'TPI'
```

Fields in the COBRA model are commonly column vectors, which can be an important detail when writing functions manipulating these fields.

There are a few more fields present in each COBRA model:

model.lb, indicating the lower bounds of each reaction, and model.ub indicating the upper bound of a reaction.

```
[{'Reaction ID', 'Lower Bound', 'Upper Bound'};... % this displays an array with
model.rxns,num2cell(model.lb), num2cell(model.ub)] % reaction names and flux bounds.
ans =
    'Reaction ID'
                    'Lower Bound'
                                     'Upper Bound'
    'GLCt1'
                    [
                             -20]
                                    [
                                              20]
    'HEX1'
                              0]
                                  [
                                              20]
                    [
    'PGI'
                             -201
                                              201
                    ſ
                                   [
    'PFK'
                             0]
                                              201
                    [
                                    [
    'FBP'
                              01
                                              201
                                    [
    'FBA'
                    [
                             -20]
                                    [
                                              201
    'TPI'
                             -20]
                                              20]
```

%This is a convenience function which does pretty much the same as the line above printFluxBounds(model);

```
Reaction ID
              Lower Bound
                              Upper Bound
      GLCt1
                  -20.000
                                   20.000
       HEX1
                    0.000
                                   20.000
                                   20.000
       PGI
                   -20.000
                                   20.000
       PFK
                    0.000
       FBP
                                   20.000
                    0.000
                                   20.000
       FBA
                   -20.000
       TPI
                  -20.000
                                   20.000
```

Before we start to modify the model, it might be useful to store some of the current properties of the model

```
mets_length = length(model.mets);
rxns_length = length(model.rxns);
```

# Creating, adding and handling reactions

If we want to add a reaction to the model or modify an existing reaction we are using function addReaction.

We will add some more reactions from glycolysis.

• The formula approach

```
model = addReaction(model, 'GAPDH',...
    'reactionFormula', 'g3p[c] + NAD[c] + 2 pi[c] -> NADH[c] + H[c] + 13bpg[c]');

Warning: Metabolite NAD[c] not in model - added to the model
Warning: Metabolite 13bpg[c] not in model - added to the model
Warning: Metabolite 13bpg[c] not in model - added to the model
GAPDH 2 pi[c] + g3p[c] + NAD[c] -> H[c] + NADH[c] + 13bpg[c]

model = addReaction(model, 'PGK',...
    'reactionFormula', '13bpg[c] + adp[c] -> atp[c] + 3pg[c]');

Warning: Metabolite 3pg[c] not in model - added to the model
PGK adp[c] + 13bpg[c] -> atp[c] + 3pg[c]

model = addReaction(model, 'PGM', 'reactionFormula', '3pg[c] <=> 2pg[c]' );

Warning: Metabolite 2pg[c] not in model - added to the model
PGM 3pg[c] <=> 2pg[c]
```

Display of stoichiometric matrix after adding reactions. Note the enlarge link when you move your mouse over the output to display the full matrix:

```
full(model.S)
ans =
    - 1
          0
                0
                     0
                           0
                                0
                                                       0
                0
                          0
                                                       0
    1
         - 1
                    0
                                0
                                      0
                                           0
                                                 0
                0
                           0
                                0
                                      0
                                           0
                                                       0
    0
         - 1
                    - 1
                                                 1
                     1
                                           1
                                                       0
```

```
0
          0
               1
                                         - 1
                                               0
0
     1
         - 1
               0
                     0
                          0
                                               0
          1
0
     0
               - 1
                    1
                          0
                               0
                                     0
                                          0
                                               0
0
     0
         0
              1
                    - 1
                         - 1
                               0
                                    0
                                          0
                                               0
0
     0
          0
               0
                    - 1
                          0
                               0
                                    0
                                          0
                                               0
                                               0
```

```
% one extra column is added(for added reaction) and 5 new % rows(for NADH, NAD, 13bpg, 2pg and 3pg metabolites)
```

The following functions are used when we want to search reactions sequence in the model and change the order of the selected reaction.

```
rxnID = findRxnIDs(model, model.rxns);
model = moveRxn(model, 8, 1);
```

While the latter does not modify the structure as such it can help in keeping a model tidy.

• The list approach

The addReaction function has ability to recognize duplicate reactions when an order of metabolites and an abbreviation of the reaction are different.

```
model = addReaction(model, 'GAPDH2',...
   'metaboliteList', {'g3p[c]', 'NAD[c]', 'pi[c]', '13bpg[c]', 'NADH[c]','H[c]' },...
   'stoichCoeffList', [-1; -1; -2; 1; 1; 1], 'reversible',false);
```

Warning: Model already has the same reaction you tried to add: GAPDH

Since the second call should not have added anything we will check if the number of the reaction increased by the three reactions we added (and not by the one duplicated) and the number of metabolites was incremented by five (13bpg, NAD, NADH, 23bpg and 2pg).

```
assert(length(model.rxns) == rxns_length + 3);
assert(length(model.mets) == mets_length + 5);
```

# Adding Exchange, Sink and Demand reactions

Specific type of reactions in the constraint-based models are reactions that are using and recycling accumulated metabolites, or producing required metabolites in the model.

- 1. *Exchange reactions* Reactions added to the model to move metabolites across the created *in silico* compartments. Those compartments represent intra- and intercellular membranes.
- 2. Sink reactions The metabolites, produced in reactions that are outside of an ambit of the system or in unknown reactions, are supplied to the network with reversible sink reactions.
- 3. *Demand reactions* Irreversible reactions added to the model to consume metabolites that are deposited in the system.

There are two ways to implement that kind of reactions:

#### 1. Use addReaction with the documented function call:

In the bigger networks we can find our exchange reactions with the following functions:

```
[selExc,selUpt] = findExcRxns(model, 0, 1) % determines whether a reaction is a
selExc =
   0
   0
   0
   0
   0
   0
   0
   0
   0
   0
   1
selUpt =
   0
   0
   0
   0
   0
   0
   0
   0
   0
   0
                                               % general exchange reaction and whether
```

% its an uptake.

2. Use a utility function to create a particular reaction type: addExchangeRxn,

addSinkReactions, addDemandReaction.

```
rxnNames: {12×1 cell}
    subSystems: {12×1 cell}
      metNames: {17×1 cell}
       grRules: {12×1 cell}
model = addSinkReactions(model, {'13bpg[c]', 'NAD[c]'})
sink 13bpg[c] 13bpg[c] <=>
sink NAD[c] NAD[c] <=>
model =
          rxns: {14×1 cell}
             S: [17×14 double]
            lb: [14×1 double]
            ub: [14×1 double]
             c: [14×1 double]
          mets: {17×1 cell}
             b: [17×1 double]
         rules: {14×1 cell}
         genes: {0×1 cell}
        osense: -1
        csense: [17×1 char]
    rxnGeneMat: [14×0 double]
      rxnNames: {14×1 cell}
    subSystems: {14×1 cell}
      metNames: {17×1 cell}
       grRules: {14×1 cell}
 model = addDemandReaction(model, {'dhap[c]', 'q3p[c]'})
DM dhap[c] dhap[c] ->
DM_g3p[c] g3p[c] \rightarrow
model =
          rxns: {16×1 cell}
             S: [17×16 double]
            lb: [16×1 double]
```

osense: -1

csense: [17×1 char]
rxnGeneMat: [12×0 double]

ub: [16×1 double]
 c: [16×1 double]
mets: {17×1 cell}
 b: [17×1 double]
rules: {16×1 cell}
genes: {0×1 cell}

osense: -1

csense: [17×1 char]
rxnGeneMat: [16×0 double]
 rxnNames: {16×1 cell}
subSystems: {16×1 cell}
 metNames: {17×1 cell}
 grRules: {16×1 cell}

It is important to emphasize that previous knowledge base informations should be taken into account. Most of them could disrupt future analysis of the model.

For instance, if it is familiar that flux through one reaction is *X* times the flux through another reaction, it is recommended to specify that in your model.

# **Altering Reaction bounds**

In order to respect the transport and exchange potential of a particular metabolite, or to resemble the different conditions in the model, we frequently need to set appropriate limits of the reactions.

```
model = changeRxnBounds(model, 'EX_glc-D[e]', -18.5, 'l');
```

# **Modifiying Reactions**

The addReaction function also is a good choice when modifying reactions. By supplying a new stoichiometry, the old will be overwritten. For example further up, we added awrong stoichiometry for the GAP-Dehydrogenase with a phosphate coefficient of 2. (easily visiple by printing the reaction)

```
printRxnFormula(model,'rxnAbbrList','GAPDH');

GAPDH 2 pi[c] + g3p[c] + NAD[c] -> H[c] + NADH[c] + 13bpg[c]
```

We can correct this by simply calling ddReaction again with the corrected stoichiometry. In essence parts which are not provided are taken from the old reaction, and only the new ones overwrite the existing data

```
model = addReaction(model, 'GAPDH',...
    'metaboliteList', {'g3p[c]', 'NAD[c]', 'pi[c]', '13bpg[c]', 'NADH[c]', 'H[c]' },...
    'stoichCoeffList', [-1; -1; 1; 1; 1]);
Warning: Reaction with the same name already exists in the model, updating the reaction
GAPDH pi[c] + g3p[c] + NAD[c] -> H[c] + NADH[c] + 13bpg[c]
```

We might also want to add a gene rule to the reaction. This can either be done using

```
model = changeGeneAssociation(model,'GAPDH','G1 and G2');

New gene G1 added to model
New gene G2 added to model

printRxnFormula(model,'rxnAbbrList',{'GAPDH'},'gprFlag',true);

GAPDH pi[c] + g3p[c] + NAD[c] -> H[c] + NADH[c] + 13bpg[c] G1 and G2
```

The other option to achieve this is to use addReaction and the geneRule parameter

```
model = addReaction(model, 'PGK', 'geneRule', 'G2 or G3', 'printLevel', 0);
```

Warning: Reaction with the same name already exists in the model, updating the reaction New gene G3 added to model

```
printRxnFormula(model,'gprFlag',true);
```

```
GAPDH pi[c] + g3p[c] + NAD[c] -> H[c] + NADH[c] + 13bpg[c] G1 and G2
GLCt1 glc-D[e] <=> glc-D[c]
HEX1 glc-D[c] + atp[c]
                        -> H[c] + adp[c] + g6p[c]
PGI q6p[c] \iff f6p[c]
PFK atp[c] + f6p[c] -> H[c] + adp[c] + fdp[c]
FBP fdp[c] + h2o[c] \rightarrow f6p[c] + pi[c]
FBA fdp[c] \iff g3p[c] + dhap[c]
TPI dhap[c] \ll g3p[c]
PGK adp[c] + 13bpg[c] \rightarrow atp[c] + 3pg[c] G2 or G3
PGM 3pg[c] <=> 2pg[c]
EX glc-D[e] glc-D[e] <=> 2 Ratio EX glc-D[c] EX glc-D[e]
EX glc-D[c] glc-D[c] + Ratio_EX_glc-D[c]_EX_glc-D[e] <=>
sink 13bpg[c] 13bpg[c] <=>
sink NAD[c] NAD[c] <=>
DM dhap[c] dhap[c] ->
DM_g3p[c] g3p[c] \rightarrow
```

#### Remove reactions and metabolites

In order to detach reactions from the model, the following function has been used:

#### Remove metabolites

```
model = removeMetabolites(model, {'3pg[c]', '2pg[c]'}, false);
```

For instance, in previous code the many metabolites from 'GAPDH' were deleted, but the reaction is still present in the model (since there are more metabolites left). The false indicates, that empty reactions should not be removed.

To delete metabolites and reactions with zero rows and columns, the following function can be used:

```
model = removeTrivialStoichiometry(model)
```

```
model =
    rxns: {9×1 cell}
        S: [15×9 double]
        lb: [9×1 double]
        ub: [9×1 double]
        c: [9×1 double]
        mets: {15×1 cell}
        b: [15×1 double]
        rules: {9×1 cell}
        genes: {3×1 cell}
        osense: -1
        csense: [15×1 char]
```

```
rxnNames: {9×1 cell}
  subSystems: {9×1 cell}
    metNames: {15×1 cell}
     grRules: {9×1 cell}
        note: 'EX glc-D[c] andEX glc-D[e]are set to have a ratio of1:2.'
model = removeRxns(model, {'GAPDH', 'PGK'});
```

# Search for duplicates and comparison of two models

rxnGeneMat: [9×3 double]

Since genome-scale metabolic models are expanding every day [2], the need for comparison and merge of them is also spreading.

The elementary functions for the model manipulation, besides main actions, simultaneously perform the structural analysis and comparison (e.g. addReaction). Likewise, there are additional functions that are only dealing with analysing similarities and differences within and between the models.

Checking for reaction duplicates by reaction abbreviation, by using method 's' that will not detect reverse reactions, and method 'FR' that will neglect reactions direction:

```
[model, removedRxn, rxnRelationship] = checkDuplicateRxn(model, 'S', 1, 1);
Checking for reaction duplicates by stoichiometry ...
 no duplicates found.
```

```
Adding duplicate reaction to the model:
 model = addReaction(model, 'GLCt1 duplicate reverse',...
      'metaboliteList', {'glc-D[e]','glc-D[c]'},...
      'stoichCoeffList', [1 -1], 'lowerBound',0,'upperBound', 20,'checkDuplicate', 0);
  GLCt1 duplicate reverse glc-D[c] -> glc-D[e]
 fprintf('>> Detecting duplicates using S method\n');
  >> Detecting duplicates using S method
 method = 'S';
 %will not be removed as does not detect reverse reaction
  [model,removedRxn, rxnRelationship] = checkDuplicateRxn(model, method, 1, 1);
  Checking for reaction duplicates by stoichiometry ...
   no duplicates found.
 assert(rxns length + 1 == length(model.rxns));
 % The reaction length has been reevaluated
 fprintf('>> Detecting duplicates with using FR method\n');
  >> Detecting duplicates with using FR method
```

```
method = 'FR';
%will be removed as detects reverse reaction
```

```
[model, removedRxn, rxnRelationship] = checkDuplicateRxn(model, method, 1, 1);

Checking for reaction duplicates by stoichiometry (up to orientation) ...
   Keep: GLCt1 glc-D[e] <=> glc-D[c]
Duplicate: GLCt1_duplicate_reverse glc-D[c] -> glc-D[e]

assert(rxns_length == length(model.rxns));
```

 Function checkCobraModelUnique marks the reactions and metabolites that are not unique in the model.

```
model = checkCobraModelUnique(model, false)
```

```
model =
          rxns: {7×1 cell}
             S: [12×7 double]
            lb: [7×1 double]
            ub: [7×1 double]
             c: [7×1 double]
          mets: {12×1 cell}
             b: [12×1 double]
         rules: {7×1 cell}
         genes: {3×1 cell}
        osense: -1
        csense: [12×1 char]
    rxnGeneMat: [7×3 double]
      rxnNames: {7×1 cell}
    subSystems: {7×1 cell}
      metNames: {12×1 cell}
       grRules: {7×1 cell}
          note: 'EX glc-D[c] and EX glc-D[e] are set to have a ratio of 1:2.'
```

# Changing the model's objective

Simulating different conditions in the model is often necessery in favor of performing calculations that investigate a specific objective. One of the elementary objectives is optimal growth [3]. Model can be modified to get different conditions with changing the model objective:

```
modelNew = changeObjective(model, 'GLCt1', 0.5);
% multiple rxns, default coefficient (1)
modelNew = changeObjective(model, {'PGI'; 'PFK'; 'FBP'});
```

#### The direction of reactions

For some purposes, it is important to only have irreversible reactions in a model, i.e. only allowing positive flux in all reactions. This can be important if e.g. absolute flux values are of interest and negative flux would reduce an objective while it should actually increase it. The COBRA toolbox offers functionality to change a model to an irreversible format, by splitting all reversible reactions and adjusting the respective lower and upper bounds, such that the model capacities stay the same.

Lets see, how the glycolysis model currently looks:

```
printRxnFormula(model);
```

```
GLCt1 glc-D[e] <=> glc-D[c]

HEX1 glc-D[c] + atp[c] -> H[c] + adp[c] + g6p[c]

PGI g6p[c] <=> f6p[c]

PFK atp[c] + f6p[c] -> H[c] + adp[c] + fdp[c]

FBP fdp[c] + h2o[c] -> f6p[c] + pi[c]

FBA fdp[c] <=> g3p[c] + dhap[c]

TPI dhap[c] <=> g3p[c]
```

To convert a model to an irreversible model use the following command:

```
[modelIrrev, matchRev, rev2irrev, irrev2rev] = convertToIrreversible(model);
```

Lets compare the irreversible model with the original model:

```
printRxnFormula(modelIrrev);
```

```
GLCt1_f glc-D[e] -> glc-D[c]
GLCt1_b glc-D[c] -> glc-D[e]
HEX1 glc-D[c] + atp[c] -> H[c] + adp[c] + g6p[c]
PGI_f g6p[c] -> f6p[c]
PGI_b f6p[c] -> g6p[c]
PFK atp[c] + f6p[c] -> H[c] + adp[c] + fdp[c]
FBP fdp[c] + h2o[c] -> f6p[c] + pi[c]
FBA_f fdp[c] -> g3p[c] + dhap[c]
FBA_b g3p[c] + dhap[c] -> fdp[c]
TPI_f dhap[c] -> g3p[c]
TPI_b g3p[c] -> dhap[c]
```

You will notice, that there are more reactions in this model, and that all reactions which have a lower bound < 0 are split in two.

There is also a function to convert an irreversible model to a reversible model:

```
modelRev = convertToReversible(modelIrrev);
```

If we now compare the reactions of this model with those from the original model, they should look the same.

### printRxnFormula(modelRev);

```
GLCt1 glc-D[e] <=> glc-D[c]

HEX1 glc-D[c] + atp[c] -> H[c] + adp[c] + g6p[c]

PGI g6p[c] <=> f6p[c]

PFK atp[c] + f6p[c] -> H[c] + adp[c] + fdp[c]

FBP fdp[c] + h2o[c] -> f6p[c] + pi[c]

FBA fdp[c] <=> g3p[c] + dhap[c]

TPI dhap[c] <=> g3p[c]
```

#### References

- [1] Orth, J. D., Thiele I., and Palsson, B. Ø. (2010). What is flux balance analysis? *Nat. Biotechnol.*, 28(3), 245–248.
- [2] Feist, A. M., Palsson, B. (2008). The growing scope of applications of genome-scale metabolic reconstructions: the case of *E. coli. Nature Biotechnology*, *26*(6), 659–667.

[3] Feist, A. M., Palsson, B. O. (2010). The Biomass Objective Function. *Current Opinion in Microbiology*, *13*(3), 344–349.