

¹ `ecodive`: Parallel and Memory-Efficient R Package for Ecological Diversity Analysis

³ Daniel P Smith  ^{1,2}¶, Sara J Javornik Cregeen  ^{1,2}, and Joseph F Petrosino  ^{1,2}

⁵ 1 The Alkek Center for Metagenomics and Microbiome Research, Department of Molecular Virology and
⁶ Microbiology, Baylor College of Medicine, Houston, TX 77030, USA ² Department of Molecular Virology
⁷ and Microbiology, Baylor College of Medicine, Houston, TX, USA  ¶ Corresponding author

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⁸ Summary

⁹ Understanding the complexity of biological communities - whether bacteria in the human
¹⁰ gut, trees in a forest, or plankton in the ocean - is a central goal of ecology. Researchers
¹¹ quantify this complexity using "diversity metrics," which describe the variety of species within a
¹² single site (alpha diversity) or the differences in composition between two sites (beta diversity).
¹³ `ecodive` is an R package designed to calculate these metrics efficiently. It bridges the gap
¹⁴ between complex ecological theory and practical data analysis, providing researchers with a
¹⁵ unified toolset to process large-scale datasets that were previously computationally prohibitive.
¹⁶ By leveraging parallel processing and optimized memory management, `ecodive` enables rapid,
¹⁷ high-throughput analysis of microbial and macro-ecological communities.

¹⁸ Statement of Need

¹⁹ A primary challenge in modern ecological analysis is the management of high-dimensional data.
²⁰ As sequencing technologies improve, datasets are growing to include thousands of samples
²¹ and tens of thousands of unique taxa. Beta diversity calculations, which involve comparing
²² every sample to every other sample, exhibit $O(n^2)$ complexity. This quadratic scaling creates a
²³ significant bottleneck; a dataset that doubles in size requires four times the processing power,
²⁴ often overwhelming standard desktop computers.

²⁵ Furthermore, the software landscape for ecological metrics is fragmented. A researcher needing
²⁶ to calculate a specific set of indices - for example, Faith's Phylogenetic Diversity, Bray-Curtis
²⁷ dissimilarity, and UniFrac distances - often must install and manage multiple R packages
²⁸ (`picante`, `vegan`, `GUniFrac`), each with different dependencies, input formats, and performance
²⁹ limitations.

³⁰ `ecodive` solves these problems by providing a centralized, high-performance library. It targets
³¹ ecologists, microbiologists, and bioinformaticians who require a robust, dependency-free solution
³² for diversity analysis. By consolidating 50 standard metrics into a single, optimized framework,
³³ it eliminates the need for "package hopping" and enables the analysis of massive datasets on
³⁴ standard hardware.

³⁵ State of the Field

³⁶ Several R packages exist for diversity analysis, but `ecodive` offers a unique contribution through
³⁷ its scope and performance. The standard package for community ecology, `vegan` ([Oksanen et](#)
³⁸ [al., 2001](#)), provides excellent implementations of non-phylogenetic metrics (e.g., Bray-Curtis)

39 but lacks phylogenetic awareness (e.g., UniFrac). Conversely, packages like picante (Kembel et
40 al., 2010) and GUniFrac (Chen et al., 2023) specialize in phylogenetic metrics but do not offer
41 a comprehensive suite of general-purpose indices. The phyloseq (McMurdie & Holmes, 2013)
42 package wraps many of these tools but relies on their underlying, often serial, implementations.
43 More generalized packages like phlentropy (Drost, 2018) offer an impressive breadth of 46
44 distinct distance measures. However, phlentropy focuses on information theory and general
45 probability distributions rather than ecology. Critically, it includes many asymmetric divergences
46 (where distance $A \rightarrow B \neq B \rightarrow A$) which, while mathematically valuable, are unsuitable for
47 standard ecological ordination methods like PCoA that require symmetric distance matrices.
48 Furthermore, phlentropy lacks critical domain-specific metrics such as the UniFrac family
49 and alpha diversity richness estimators like Chao1.
50 ecodive builds upon this landscape by unifying these distinct domains. It implements 50
51 symmetric metrics chosen specifically for their relevance to ecological ordination and analysis.
52 Crucially, unlike the serial R or C implementations found in most peer packages (see **Research
53 Impact**), ecodive is built entirely on a parallelized C engine, providing orders-of-magnitude
54 faster performance while ensuring numerical identity with established tools.

55 Software Design

56 The architecture of ecodive balances the user-friendly conventions of R with the raw
57 performance of C. A critical design trade-off centered on data representation.

58 Most R users work with dense matrices where samples are rows and features are columns.
59 Standard R functions like `dist()` expect this format. However, ecological matrices are typically
60 90-99% zeros (sparse). Storing them as dense matrices wastes gigabytes of RAM, and
61 processing them row-by-row is cache-inefficient for many distance algorithms.

62 To address this, ecodive maintains the standard R interface (samples-as-rows) but
63 fundamentally alters the backend data structure:

- 64 1. **Transparent Transformation:** When a standard matrix is passed to ecodive, it is internally
65 converted into a column-compressed sparse matrix (`dgCMatrix`) with samples transposed
66 to columns. This incurs a one-time overhead but allows the C engine to skip zeros
67 entirely and access memory in a cache-friendly, column-major pattern.
- 68 2. **Power User Bypass:** For extremely large datasets where the overhead of this
69 transformation is non-trivial, users can manually provide data in the native `dgCMatrix`
70 format (samples as columns). ecodive detects this optimized state and bypasses the
71 transformation step, operating directly on the existing C pointers. This allows for
72 “zero-copy” analysis of massive datasets.
- 73 3. **Parallelization Strategy:** ecodive employs a direct implementation using the standard
74 POSIX threads (`pthreads`) library, avoiding the memory duplication overhead of forking
75 processes found in R’s `parallel` package. This design enables fine-grained, dynamic load
76 balancing, ensuring efficient execution even when calculating partial distance matrices.

77 Research Impact Statement

78 ecodive has demonstrated immediate utility in high-dimensional microbiome studies. The
79 core C algorithms in ecodive were originally developed for and deployed in the `rbiom` package
80 (Smith, 2020). As part of `rbiom`, these optimized metrics have already been utilized in diverse
81 microbial ecology studies, including research on preterm infant microbiomes (Ahearn-Ford et
82 al., 2025), dietary interventions (DiMattia et al., 2025), and relationship satisfaction (Cheng
83 et al., 2023). ecodive extracts these proven, high-performance components into a standalone,
84 lightweight library to make them accessible to the broader R ecosystem without `rbiom`’s specific
85 visualization and data structure dependencies.

86 In benchmarks comparing 15 ecological R packages, ecodive consistently ranked as the fastest
 87 and most memory-efficient solution:

- 88 **Speed:** For the widely-used Unweighted UniFrac metric ($N = 50$), ecodive completed
 89 calculations in 6.4ms, compared to 2.5s for picante (396x faster) and 297ms for phyloseq
 90 (46x faster).
- 91 **Scalability:** For standard Bray-Curtis dissimilarity ($N = 1006$), ecodive processed the
 92 matrix in ~20ms, whereas vegan required 1.68s.
- 93 **Memory:** ecodive's sparse architecture reduced memory allocation for large operations
 94 from gigabytes (in abdiv or tabula) to megabytes, enabling analyses on laptops that
 95 previously required clusters.

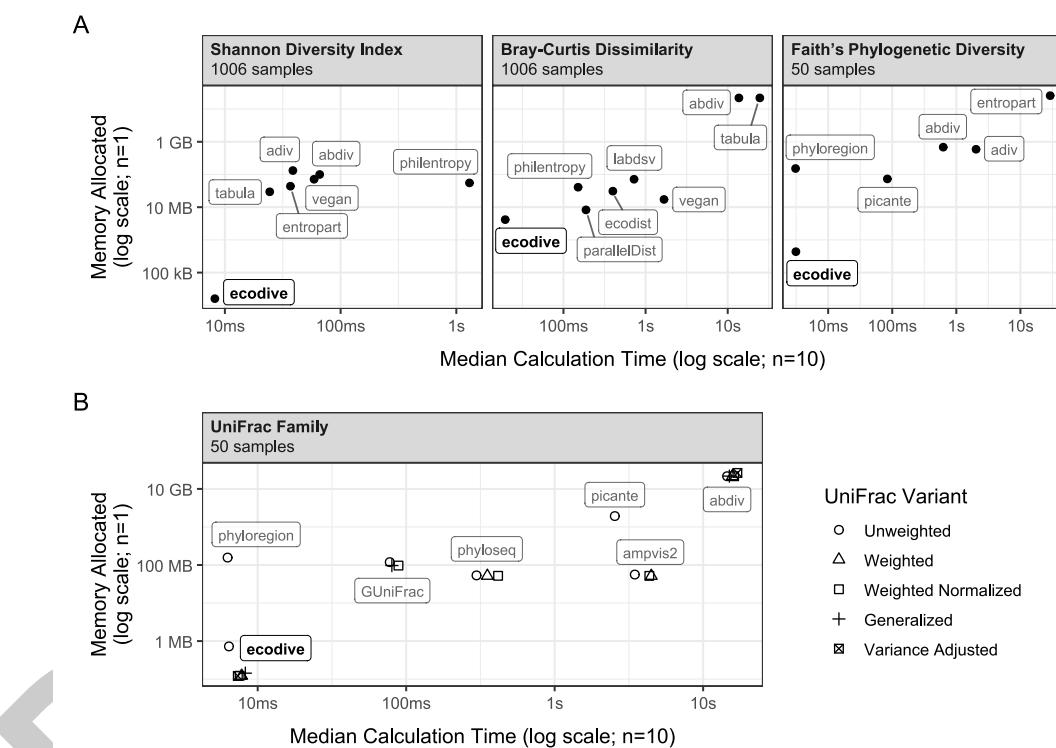


Figure 1: Benchmarking results. Execution time (x-axis) vs. peak memory usage (y-axis) for various diversity metrics across 15 R packages. ecodive (highlighted) consistently occupies the bottom-left quadrant, indicating high speed and low memory footprint. Note the log scale on both axes.

96 The package is fully documented with vignettes covering performance tuning and metric
 97 selection, and is available for installation with zero external R dependencies, ensuring high
 98 community readiness and long-term stability.

99 Example Usage

100 ecodive is designed for ease of use and integrates seamlessly with existing bioinformatics
 101 workflows, such as those using phyloseq objects. For example, calculating weighted UniFrac
 102 distances is straightforward:

```
103 data(esophagus, package = 'phyloseq')
  104 ecodive::weighted_unifrac(esophagus)
  105 #>       B          C
```

```
#> C 0.1050480
#> D 0.1401124 0.1422409
```

103 AI Usage Disclosure

104 Generative AI tools (Google Gemini) were used to assist in the drafting and revision of this
105 manuscript and the generation of documentation. No AI tools were used to write the functional
106 source code (R or C) of the software. All AI-generated text was critically reviewed, verified for
107 accuracy, and edited by the authors.

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