

1 lickcalc: Easy analysis of lick microstructure in 2 experiments of rodent ingestive behaviour

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Software

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6 Summary

7 Lick microstructure is a term used in behavioural neuroscience to describe the information that
8 can be obtained from a detailed examination of rodent drinking behaviour. Rather than simply
9 recording total intake (volume consumed), lick microstructure examines how licks are grouped,
10 and the spacing of these groups of licks. This type of analysis can provide important insights
11 into why an animal is drinking, for example, whether it is influenced by taste or affected
12 by consequences of consumption (e.g., feeling “full”). The simplicity of using lickcalc, a
13 browser-based application hosted at <https://lickcalc.uit.no>, will make microstructural analysis
14 accessible to any researchers who wish to employ it while providing sophisticated analyses with
high scientific value.

16 Statement of need

17 lickcalc is a software suite that performs microstructural lick analysis on timestamps of lick
18 onsets and/or offsets. Microstructural analysis was first described in Davis & Smith (1992)
19 and has since then been used to understand diverse phenomena. In-depth reviews on many
20 of these, and microstructural parameters used to study them, are available (Johnson, 2018;
21 Naneix et al., 2020; Smith, 2001). Briefly, although much of the foundational work on drinking
22 microstructure was on licking for nutritive solutions (e.g., sucrose solutions), microstructural
23 analysis can also be used to study intake of water (McKay & Daniels, 2013; Santollo et
24 al., 2021), ethanol (Patwell et al., 2021), and other tastants such as non-caloric artificial
25 sweeteners, sodium, and quinine (Lin et al., 2012; Spector & St. John, 1998; Verharen et al.,
26 2019). Lick microstructure has been used to shed light on, for example, how licking is affected
27 by neuropeptides (McKay & Daniels, 2013), enzymes in the mouth (Chometton et al., 2022),
28 ovarian hormones (Santollo et al., 2021), nutrient restriction (Naneix et al., 2020), response
29 to alcohol (Patwell et al., 2021), and diet (Johnson, 2012).

30 Lick microstructure can provide nuanced information about why an animal is drinking. Briefly,
31 by considering the distribution on interlick intervals (ILIs), individual licks can be grouped into
32 bursts (also sometimes referred to as clusters or bouts) based on a minimum ILI that separates
33 each burst. The number of lick bursts during a session is thought to reflect post-ingestive
34 feedback from the consumed fluid, whereas the average number of licks per burst is thought
35 to reflect palatability of the solution. Often, changes in microstructure are accompanied
36 by changes in total intake, but this is not always the case: sometimes, equal intake will be
37 achieved by quite different licking patterns that indicate changes in orosensory and post-ingestive
38 feedback (Johnson et al., 2010; Volcko et al., 2020). Analyzing lick microstructure is therefore
39 highly valuable when trying to understand how a manipulation, X , affects appetite; if X causes
40 an animal to feel more sated after drinking, that may lead to a different interpretation than
41 if X were to reduce the palatability of the solution. Because of the value of microstructural
42 data, many labs habitually record and analyze it. There are many others, however, that have

43 not yet begun collecting and/or analysing these data. Investing in lickometers can be costly,
44 but there are an increasing number of cost-effective alternatives to commercial products. As
45 such, several open-source lickometer designs are now available (e.g. Frie & Khokhar (2024);
46 Monfared et al. (2024); Petersen et al. (2024); Silva et al. (2024); Raymond et al. (2018)).

47 Recording individual licks with high temporal resolution is necessary for microstructural analysis
48 of drinking behavior, but another barrier to reporting microstructure is its analysis. This
49 problem is now easily solved by lickcalc. lickcalc does not require any special software
50 or coding knowledge: all the user has to do is load a file with timestamps of lick onsets
51 (and, ideally, offsets) into the application and lickcalc will generate detailed microstructural
52 analysis, with a high degree of user control over key parameters. Resulting data provide values
53 for number of licks, number of bursts, and burst size (among others) – the values that are
54 often reported and used to draw inferences about postigestive and orosensory feedback of
55 the solution. But importantly, several plots are also displayed that show information that
56 helps with quality control of the data and challenges the user to think critically about which
57 parameters they have chosen. In short, lickcalc makes microstructural analysis accessible
58 to any with appropriate data, while providing detailed information needed for appropriate
59 parameter selection and quality control.

60 Key features

61 lickcalc has several features that make it exceptionally user-friendly while providing sophisticated
62 and detailed microstructural analysis. Some of these features include:

- 63 ■ **Ease of use:** Files of various formats can simply be dragged into the lickcalc software
64 to trigger analyses. Parameters can be set manually using sliders, and results exported
65 to Excel with the push of a button.
- 66 ■ **Flexibility:** The user sets key parameters appropriate for their experimental setup and
67 data. Data can be analysed across the whole-session, within different epochs, or based
68 on a trial structure.
- 69 ■ **Customization:** By using the configuration file, users can change default settings to
70 match their preferences and avoid manually changing settings for each file loaded.
- 71 ■ **Results compilation:** Data from multiple sessions and/or individuals can be exported into
72 a single Excel file, which streamlines analysis. A batch mode is also included allowing
73 multiple files to be analysed simultaneously.
- 74 ■ **Detail of analysis:** One of the benefits of using lickcalc is the level of detail it provides.
75 In addition to the properties often reported (e.g., burst number, burst size), lickcalc
76 computes and displays attributes of the data that are important in establishing the quality
77 of the data and determining appropriate parameters for its microstructural analysis. Four
78 charts are:
 - 79 1) *intraburst lick frequency*, or how often certain interlick intervals within a burst of
80 licking occur. While a rodent is licking, its tongue makes rhythmic protrusions that
81 are under the control of a central pattern generator (Travers et al., 1997). Rats
82 typically lick 6–7 times per second (Davis & Smith, 1992), while mice lick at a
83 slightly higher rate (Johnson et al., 2010). In addition to these species differences,
84 there are also strain differences (Johnson et al., 2010; St. John et al., 2017).
85 Because intraburst lick rate is under the control of the central pattern generator, it
86 should remain relatively stable across mice and conditions (unless a manipulation
87 is expected to cause changes in the central pattern generator). A typical chart
88 for a mouse might show a sharp peak around an intraburst ILI of ~129 ms, which
89 corresponds to a lick rate of 7.75 Hz. Much smaller peaks are often present at the
90 harmonics of the intraburst ILI (e.g., a primary peak at 129 ms will have smaller
91 peaks at 258 ms, 387 ms, and so on), often because of “missed licks” in which the

- 92 mouse attempts to lick but its tongue misses the spout. A large number of these,
 93 or other differences from the expected pattern of results, may indicate problems
 94 with the experimental setup (e.g., if the animal fails to reach the spout frequently,
 95 then perhaps the spout is too far away).
- 96 2) *lick length* is only available when lick offsets are included in the data file. As with
 97 intraburst lick frequency, lick length should show little variability and the graph
 98 will have a sharp peak. Occasionally a lickometer will register longer licks than
 99 normal. A common cause of this is formation of a fluid bridge between the tongue
 100 and the spout during periods of high frequency licking. This can be prevented by
 101 moving the bottle further from the animal. In addition, other causes are if a rodent
 102 grabs the spout with its paws, or if a fluid droplet hangs between the spout and
 103 the cage and thus completes the electrical circuit. Concerns about data quality may
 104 be warranted with increasing number and duration of long licks. `lickcalc` displays
 105 both the number and maximum duration of licks above the threshold that the user
 106 has set. There is also an option to remove these problematic licks from the dataset.
- 107 3) *burst frequency*, or how often certain burst sizes occurred. This is informative
 108 because burst size, by virtue of being a mean (mean licks per burst), does not
 109 take into account potentially relevant information about the distribution of licks in
 110 each burst. For example, a burst size of 80 could result from bursts all containing
 111 between 70 and 90 licks, or from many single licks and one or two bursts with a lot
 112 of licks. The latter case might raise some questions about how reliable the burst
 113 size value is. Although single licks occur, they can also be caused by non-tongue
 114 contact with the lickometer. Changing the minimum licks/burst parameter can
 115 filter out some of these suspect “licks.”
- 116 4) *Weibull probability*. The Weibull analysis, as described in Davis (1996), uses a
 117 mathematical equation to fit data to a survival function. Although used by some
 118 (Aja et al., 2001; Moran et al., 1998; Spector & St. John, 1998), it is still relatively
 119 rare to find Weibull probabilities in microstructural analyses. The Weibull function
 120 can be used on several aspects of data, such as lick rate across a session, but in
 121 the `lickcalc` program the Weibull probability is calculated for burst size. It plots
 122 the probability that, given n licks, the mouse will continue to lick. This makes it
 123 sensitive to the licks per burst parameter that is set by the user. The Weibull α
 124 and β values reflect the slope and shape parameters, respectively. Slope (α) has
 125 been shown to vary with palatability.

126 Design and usage

127 `lickcalc` is hosted by UiT The Arctic University of Norway and can be accessed at <https://lickcalc.uit.no>. Alternatively, it can be installed locally following instructions in the repository.
 128 To use `lickcalc`, the user drags a file into the application, changes file format if necessary,
 129 and indicates which column contains the lick onsets (and, if applicable, the lick offsets). A
 130 plot is automatically generated that displays a histogram of licks across the session. Session
 131 length defaults to the time of the last lick but can be manually changed, or set in the optional
 132 config file. Session length can be set in seconds, minutes, or hours. The bin size (licks per
 133 unit of time) can be changed manually or in the config file. The user can toggle between the
 134 default histogram and a plot showing cumulative licks.
 135

136 A microstructural analysis is, in essence, a division of individual licks into groups of licks. To
 137 perform this grouping, the user must set several parameters. One of these is the ILI threshold,
 138 which is the minimum amount of time licks must be separated by in order to be considered
 139 separate bursts. Early studies identified ILIs of 251–500 ms as separating “bursts” of licking,
 140 and pauses of >500 ms as separating “clusters” of licking (i.e., a cluster of licks is made up
 141 of several bursts of licking). Others have argued that an ILI threshold of 1 s provides better

separation of lick bursts ([Spector & St. John, 1998](#)). In `lickcalc`, the user can set the ILI to any value (values between 250 ms and 3 s provided by default but can be adjusted using the config file). Another parameter that needs to be decided prior to the lick analysis is the minimum number of licks per burst. `lickcalc` allows between 1 and 5 licks by default. The appropriate number of minimum licks per burst may vary depending on experimental set up, and the likelihood that a single lick represents a lick rather than, for example, a paw touching the spout. Finally, in `lickcalc`, the user must set a “long-lick threshold” between 0.1 and 1 s. This parameter is only available when lick offset is included. Licks that are longer than the set threshold are counted as “long” and may indicate a problem (e.g., fluid bridges or a mouse holding the spout with its paws) rather than a true lick. The user can decide whether to remove “long licks” or not. All of these parameters can be set manually or through the config file. Four plots are generated (see *Key Features* section above), and tables are displayed showing values of several properties: total licks, intraburst frequency, number of long licks, maximum duration of long licks, number of bursts, mean licks per burst, Weibull α , Weibull β , and Weibull r^2 .

To save these data, the user has two options. The first is to export a single Excel file in which the user sets the animal ID and chooses which data to export. These data allow the user to recreate the plots displayed in `lickcalc` or perform further analyses. The second option for saving the data is to add the loaded data to the *Results Summary* table. The results in this table remain even as new data files are loaded, so the data from many sessions (and/or individual animals) can be exported into a single Excel file. In addition to the data from the whole session, the user can choose to divide the session into epochs, or to examine only the first n bursts, or perform a trial-based analysis (e.g., for Davis rig experiments). Each of these analysis epochs can be added to the table. The table contains the data and the analysis parameters (e.g., minimum burst size) used to generate them. Finally, a batch process feature is available allowing multiple files to be analysed using the same parameters.

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References

- 176 Aja, S., Schwartz, G. J., Kuhar, M. J., & Moran, T. H. (2001). Intracerebroventricular CART
177 peptide reduces rat ingestive behavior and alters licking microstructure. *American Journal*
178 of Physiology-Regulatory, Integrative and Comparative Physiology, 280(6), R1613–R1619.
<https://doi.org/10.1152/ajpregu.2001.280.6.R1613>
- 180 Chometton, S., Jung, A.-H., Mai, L., Dal Bon, T., Ramirez, A. O., Pittman, D. W., & Schier, L.
181 A. (2022). A glucokinase-linked sensor in the taste system contributes to glucose appetite.
182 *Molecular Metabolism*, 64, 101554. <https://doi.org/10.1016/j.molmet.2022.101554>
- 183 Davis, J. D. (1996). Deterministic and probabilistic control of the behavior of rats ingesting
184 liquid diets. *American Journal of Physiology-Regulatory, Integrative and Comparative*
185 *Physiology*, 270(4), R793–R800. <https://doi.org/10.1152/ajpregu.1996.270.4.R793>
- 186 Davis, J. D., & Smith, G. P. (1992). Analysis of the microstructure of the rhythmic tongue
187 movements of rats ingesting maltose and sucrose solutions. *Behavioral Neuroscience*,
188 106(1), 217–228. <https://doi.org/10.1037/0735-7044.106.1.217>

- 189 Frie, J. A., & Khokhar, J. Y. (2024). FARESHARE: An open-source apparatus for assessing
190 drinking microstructure in socially housed rats. *NPP—Digital Psychiatry and Neuroscience*,
191 2(1), 1. <https://doi.org/10.1038/s44277-024-00002-z>
- 192 Johnson, A. W. (2012). Dietary manipulations influence sucrose acceptance in diet induced
193 obese mice. *Appetite*, 58(1), 215–221. <https://doi.org/10.1016/j.appet.2011.09.015>
- 194 Johnson, A. W. (2018). Characterizing ingestive behavior through licking microstructure:
195 Underlying neurobiology and its use in the study of obesity in animal models. *International
196 Journal of Developmental Neuroscience: The Official Journal of the International Society for
197 Developmental Neuroscience*, 64, 38–47. <https://doi.org/10.1016/j.ijdevneu.2017.06.012>
- 198 Johnson, A. W., Sherwood, A., Smith, D. R., Wosiski-Kuhn, M., Gallagher, M., & Holland, P.
199 C. (2010). An analysis of licking microstructure in three strains of mice. *Appetite*, 54(2),
200 320–330. <https://doi.org/10.1016/j.appet.2009.12.007>
- 201 Lin, J.-Y., Amodeo, L. R., Arthurs, J., & Reilly, S. (2012). Taste neophobia and palatability:
202 The pleasure of drinking. *Physiology & Behavior*, 106(4), 515–519. <https://doi.org/10.1016/j.physbeh.2012.03.029>
- 204 McKay, N. J., & Daniels, D. (2013). Glucagon-like peptide-1 receptor agonist administration
205 suppresses both water and saline intake in rats. *Journal of Neuroendocrinology*, 25(10),
206 929–938. <https://doi.org/10.1111/jne.12086>
- 207 Monfared, M., Mascret, Q., Marroquin-Rivera, A., Blanc-Árabe, L., Lebouleux, Q., Lévesque,
208 J., Gosselin, B., & Labonté, B. (2024). High-throughput low-cost digital lickometer system
209 for the assessment of licking behaviours in mice. *Journal of Neuroscience Methods*, 410,
210 110221. <https://doi.org/10.1016/j.jneumeth.2024.110221>
- 211 Moran, T. H., Katz, L. F., Plata-Salaman, C. R., & Schwartz, G. J. (1998). Disordered
212 food intake and obesity in rats lacking cholecystokinin A receptors. *American Journal
213 of Physiology-Regulatory, Integrative and Comparative Physiology*, 274(3), R618–R625.
214 <https://doi.org/10.1152/ajpregu.1998.274.3.R618>
- 215 Naneix, F., Peters, K. Z., & McCutcheon, J. E. (2020). Investigating the Effect of Physiological
216 Need States on Palatability and Motivation Using Microstructural Analysis of Licking.
217 *Neuroscience*, 447, 155–166. <https://doi.org/10.1016/j.neuroscience.2019.10.036>
- 218 Patwell, R., Yang, H., Pandey, S. C., & Glover, E. J. (2021). An operant ethanol self-
219 administration paradigm that discriminates between appetitive and consummatory behaviors
220 reveals distinct behavioral phenotypes in commonly used rat strains. *Neuropharmacology*,
221 201, 108836. <https://doi.org/10.1016/j.neuropharm.2021.108836>
- 222 Petersen, N., Adank, D. N., Quan, Y., Edwards, C. M., Hallal, S. D., Taylor, A., Winder, D. G.,
223 & Doyle, M. A. (2024). A Novel Mouse Home Cage Lickometer System Reveals Sex- and
224 Housing-Based Influences on Alcohol Drinking. *eNeuro*, 11(10), ENEURO.0234–24.2024.
225 <https://doi.org/10.1523/ENEURO.0234-24.2024>
- 226 Raymond, M. A., Mast, T. G., & Breza, J. M. (2018). An open-source lickometer and
227 microstructure analysis program. *HardwareX*, 4, e00035. <https://doi.org/10.1016/j.ohx.2018.e00035>
- 229 Santollo, J., Edwards, A. A., Howell, J. A., & Myers, K. E. (2021). Bidirectional effects
230 of estradiol on the control of water intake in female rats. *Hormones and Behavior*, 133,
231 104996. <https://doi.org/10.1016/j.yhbeh.2021.104996>
- 232 Silva, A., Carriço, P., Fernandes, A. B., Saraiva, T., Oliveira-Maia, A. J., & Da
233 Silva, J. A. (2024). High-Precision Optical Fiber-Based Lickometer. *Eneuro*, 11(7),
234 ENEURO.0189–24.2024. <https://doi.org/10.1523/ENEURO.0189-24.2024>
- 235 Smith, G. P. (2001). John Davis and the meanings of licking. *Appetite*, 36(1), 84–92.
236 <https://doi.org/10.1006/appet.2000.0371>

- 237 Spector, A. C., & St. John, S. J. (1998). Role of taste in the microstructure of quinine
238 ingestion by rats. *American Journal of Physiology-Regulatory, Integrative and Comparative*
239 *Physiology*, 274(6), R1687–R1703. <https://doi.org/10.1152/ajpregu.1998.274.6.R1687>
- 240 St. John, S. J., Lu, L., Williams, R. W., Saputra, J., & Boughter, J. D. (2017). Genetic control
241 of oromotor phenotypes: A survey of licking and ingestive behaviors in highly diverse strains
242 of mice. *Physiology & Behavior*, 177, 34–43. <https://doi.org/10.1016/j.physbeh.2017.04.007>
- 244 Travers, J. B., Dinardo, L. A., & Karimnamazi, H. (1997). Motor and Premotor Mechanisms
245 of Licking. *Neuroscience & Biobehavioral Reviews*, 21(5), 631–647. [https://doi.org/10.1016/S0149-7634\(96\)00045-0](https://doi.org/10.1016/S0149-7634(96)00045-0)
- 247 Verharen, J. P. H., Roelofs, T. J. M., Menting-Henry, S., Luijendijk, M. C. M., Vanderschuren,
248 L. J. M. J., & Adan, R. A. H. (2019). Limbic control over the homeostatic need for sodium.
249 *Scientific Reports*, 9(1), 1050. <https://doi.org/10.1038/s41598-018-37405-w>
- 250 Volcko, K. L., Brakey, D. J., Przybysz, J. T., & Daniels, D. (2020). Exclusively drinking
251 sucrose or saline early in life alters adult drinking behavior by laboratory rats. *Appetite*,
252 149, 104616. <https://doi.org/10.1016/j.appet.2020.104616>