

EMGFlow: A Python package for preprocessing and feature extraction of electromyographic signals

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Software

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Summary

Surface electromyography (sEMG) is increasingly used to study human physiology and behaviour, spurred by advances in deep learning and wearable sensors. Here, we introduce EMGFlow, an open-source Python package that streamlines preprocessing and feature extraction for sEMG signals. Tailored for batch processing, EMGFlow handles large datasets typical in machine learning, extracting a comprehensive set of 33 statistical features across time and frequency domains. The package supports flexible file selection with regular expressions and uses Pandas DataFrames end-to-end to facilitate interoperability. An interactive dashboard visualises signals at each preprocessing stage to aid user decisions. EMGFlow is distributed under the GNU General Public License v3.0 (GPL-3.0) and is available on PyPI. Documentation with guides, API references, and runnable examples is available at https://wiiison.github.io/EMGFlow-Python-Package/.

Statement of Need

Although several packages process physiological and neurological signals, support for sEMG has remained limited. Many lack a comprehensive feature set for sEMG, forcing researchers to use a patchwork of tools. Others focus on event detection with GUI-centric workflows that suit continuous recordings of a single participant, but complicate batch feature extraction common in machine learning (Abadi et al., 2015; Chen et al., 2022; Koelstra et al., 2012; Schmidt et al., 2018; Sharma et al., 2019; Zhang et al., 2016). EMGFlow, a portmanteau of EMG and Workflow, fills this gap by providing a flexible pipeline for extracting a wide range of sEMG features, with a scalable design suited for large datasets. An overview of package metadata is presented in Table 1.

Table with 2 columns: Metadata, Description. Rows include License (GPLv3), Implementation (Python >= 3.9), Code repository (https://github.com/Willson/EMGFlow-Python-Package), Documentation (https://wiiison.github.io/EMGFlow-Python-Package), and PyPI installation (pip install EMGFlow).

Table 1: EMGFlow package metadata.

Comparison to Other Packages

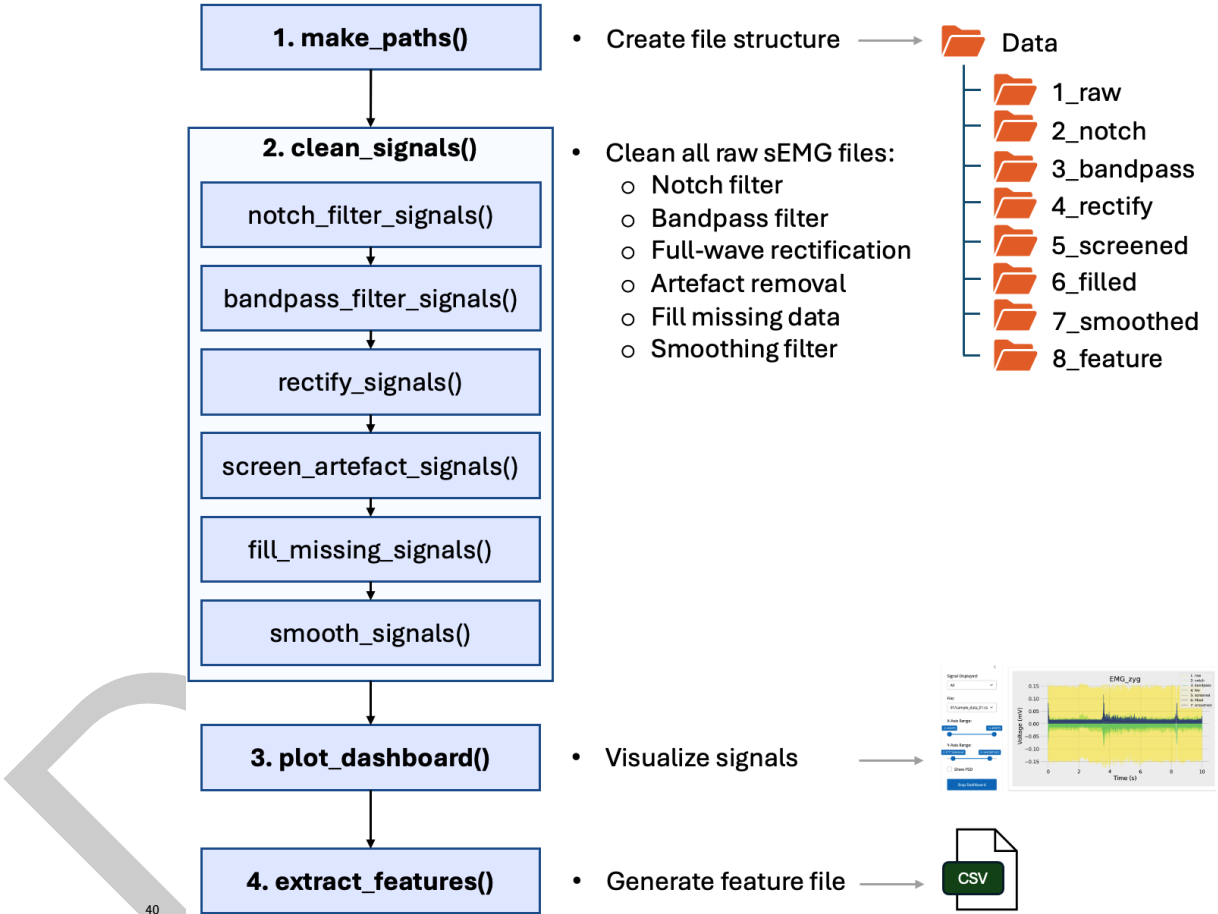
Compared to existing toolkits, EMGFlow provides a broader, sEMG-specific library of 33 features (Bizzego et al., 2019; Bota et al., 2024; Makowski et al., 2021; Sjak-Shie, n.d.;

31 [Soleymani et al., 2017](#)). Its dashboard visualises batch-processed files rather than single
32 recordings, enabling inspection of preprocessing effects across datasets ([Gabrieli et al., 2020](#)).
33 Adjustable filters and smoothing support international mains standards (50 vs 60 Hz), a subtle
34 detail some packages omit.

35 **Features**

36 **A Simplified Workflow**

37 Extracting features from large datasets is fundamental in machine learning and quantitative
38 analysis. *EMGFlow* supports batch-processing, enabling fully or semi-automated treatment of
39 sEMG recordings. Figure 1 outlines the pipeline.



41 **Figure 1:** An overview of the processing pipeline.

42 Example 1 demonstrates end-to-end preprocessing and feature extraction. We create project
43 paths with `make_paths()` and load bundled sample data with `make_sample_data()` (adapted
44 from *PeakAffectDS* ([Greene et al., 2022](#))). Next, we run automated preprocessing via
45 `clean_signals()` using sensible, literature-based defaults, and then write a plaintext CSV of
46 33 features per file with `extract_features()`.

```
# %% Example 1: Quick start (full pipeline)
import EMGFlow

# Create project paths
path_names = EMGFlow.make_paths()

# Load sample data
EMGFlow.make_sample_data(path_names)

# Preprocess signals
EMGFlow.clean_signals(path_names, sampling_rate=2000, notch_f0=50)

# Extract features to disk "Features.csv"
EMGFlow.extract_features(path_names, sampling_rate=2000)
```

47 Tailored Preprocessing

48 Example 2 shows how advanced users can tailor low-level preprocessing. After setup, Step 1
 49 applies a notch filter to remove AC mains interference. Most functions use common sense
 50 defaults, which can be modified task-wide or for select cases. For instance, the sample data
 51 were recorded in New Zealand (200-240 VAC 50Hz), so we set the notch frequency and quality
 52 factor accordingly.

```
# %% Example 2: Tailored preprocessing
import EMGFlow

# Setup workspace
path_names = EMGFlow.make_paths()
EMGFlow.make_sample_data(path_names)

# Data sampling rate
sampling_rate = 2000

# Notch filter for mains hum (Hz, Q-score)
notch_main = [(50, 5)]

# Columns names containing sEMG (Zygomaticus major, Corrugator supercilii)
muscles = ['EMG_zyg', 'EMG_cor']

# Step 1. Apply notch filter to all files in 1_raw, writing output to 2_notch
EMGFlow.notch_filter_signals(path_names['raw'], path_names['notch'],
                             muscles, sampling_rate, notch_main)

53 EMGFlow preserves the raw directory structure and mirrors it at each pipeline stage. All
54 preprocessing functions accept an optional regular expression to target specific files. In Step 1b,
55 we apply an additional notch filter at 150 Hz (the 3rd harmonic) only to files in subfolder /01.

# Custom notch settings
notch_custom = [(150, 25)]
path_pattern = '^01/'

# Step 1b. Apply custom notch filter all to files in subfolder "/01"
EMGFlow.notch_filter_signals(path_names['notch'], path_names['notch'],
                             muscles, sampling_rate, notch_custom,
                             expression=path_pattern)
```

56 Interference Attenuation

57 Surface EMG is susceptible to multiple sources of interference that affect the signal with
 58 distinct spectral signatures (Boyer et al., 2023). Band-pass filtering is typically performed
 59 in Step 2 to isolate the frequency spectrum of human muscle activity. Common passbands
 60 are 10-500 Hz (Livingstone et al., 2016; McManus et al., 2020; Sato et al., 2021; Tamietto
 61 et al., 2009), though precise edges vary by domain (Abadi et al., 2015). Step 3 performs
 62 full-wave rectification, converting negative values to positive (Dakin et al., 2014; Rutkowska et
 63 al., 2024).

```
# Passband edges (low, high)
passband_edges = [20, 450]
```

```
# Step 2. Apply band-pass filter
```

```
EMGFlow.bandpass_filter_signals(path_names['notch'], path_names['bandpass'],
                                muscles, sampling_rate, passband_edges)
```

```
# Step 3. Apply full-wave rectifier
```

```
EMGFlow.rectify_signals(path_names['bandpass'], path_names['fwr'], muscles)
```

64 Signal artefacts are another source of contamination and span a diverse range of phenomenon
 65 including thermal noise, eyeblinks, and random noise bursts (Boyer et al., 2023). These can
 66 be mitigated with screen_artefacts(), which applies a Hampel filter (default), or Wiener
 67 filter, both reported as robust denoisers (Allen, 2009; Bhowmik et al., 2017; Jarrah et al.,
 68 2022). Because artefact profiles vary across projects, we recommend visual inspection
 69 with the interactive dashboard to tune n_sigma (Hampel) and window_ms (Bhowmik et al.,
 70 2017; Pearson et al., 2016). In Step 4 we target /02/sample_data_04.csv which contains an
 71 artificial, band-limited noise pulse, and copy other files forward untouched.

```
screen_pattern = r'^02/sample_data_04\.csv$'
```

```
# Step 4. Apply Hampel artefact filter to 02/sample_data_04.csv
```

```
EMGFlow.screen_artefact_signals(path_names['fwr'], path_names['screened'],
                                muscles, sampling_rate,
                                expression=screen_pattern, copy_unmatched=True)
```

72 Missing data consisting of brief gaps or NaNs can be filled with fill_missing_signals(),
 73 which defaults to Piecewise Cubic Hermite Interpolating Polynomial (method=pchip). PCHIP
 74 is shape-preserving, monotonicity-respecting, and avoids overshoot - properties desirable for
 75 sEMG (SciPy Community, 2025). Cubic spline is also available (Shin et al., 2021). In Step 5,
 76 we address artificially injected gaps with PCHIP.

77 In Step 6, optional smoothing removes residual high-frequency noise before feature extraction.
 78 The default smoother RMS, equal to the square root of the total power, estimates signal
 79 amplitude and is commonly used in sEMG (McManus et al., 2020). Boxcar, Gaussian, and
 80 LOESS alternatives are also provided.

```
# Step 5. Fill missing data
```

```
EMGFlow.fill_missing_signals(path_names['screened'], path_names['filled'],
                              muscles, sampling_rate)
```

```
# Step 6. Apply smoothing filter
```

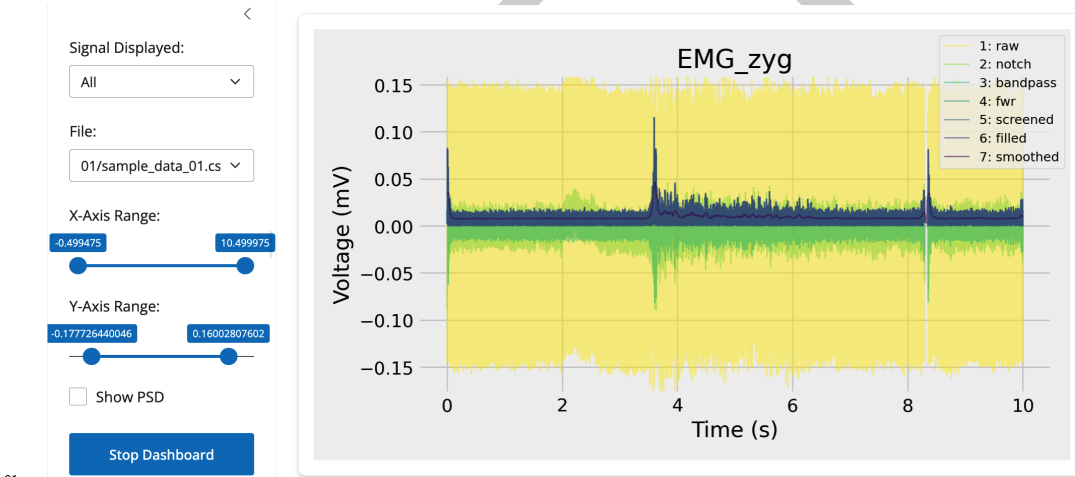
```
EMGFlow.smooth_signals(path_names['filled'], path_names['smooth'],
                        muscles, sampling_rate)
```

81 **An Interactive Dashboard**

82 *EMGFlow* includes a Shiny dashboard for visualising preprocessing effects. Pipeline steps can
83 be overlaid or shown individually, and files are selected from a drop-down menu. A checkbox
84 toggles between a time-domain amplitude view and a spectral view that displays the Power
85 Spectral Density (PSD). The amplitude view exposes transients and drift, guiding selection of
86 passband edges and confirming that filtering preserves waveform shape. The PSD highlights
87 mains peaks and harmonics, guiding the choice of notch parameters (f_0 , Q). Below we generate
88 a dashboard for the *Zygomaticus* major channel. When we have finished inspecting the signals,
89 we click 'Stop Dashboard' to shut down the dashboard server and end the interactive session
90 so that the analysis pipeline can proceed.

```
# Column and measurement units to plot
show_muscle = 'EMG_zyg'
units = 'mV'

# Plot data for the "EMG_zyg" column
EMGFlow.plot_dashboard(path_names, show_muscle, sampling_rate, units)
```



91
92 **Figure 2:** *EMGFlow*'s interactive dashboard visualizing effects of different preprocessing steps
93 on batch processed files.

94 **An Extensive Feature Library**

95 After preprocessing, files are ready for feature extraction. Surface EMG records voltage
96 differences at the skin arising from the summed motor-unit action potentials (Fridlund &
97 Cacioppo, 1986), yielding an interference signal whose amplitude (time domain) and spectrum
98 (frequency domain) reflect motor-unit recruitment, discharge rates, and muscle-fiber conduction
99 velocity (De Luca, 2008; McManus et al., 2020). *EMGFlow* extracts 33 features across time
100 and frequency domains, as listed in Table 2.

Domain	Feature
Temporal	minV, maxV, meanV, stdV, skewV, kurtosisV, maxF, IEMG, MAV, MMAV1, MMAV2, SSI, VAR, VOrder, RMS, WL, WAMP, LOG
Spectral	MFL, AP, SpecFlux, MDF, MNF, TwitchRatio, TwitchIndex, TwitchSlope, SC, SF, SS, SDec, SEntropy, SRoll, SBW

101 **Table 2:** Features extracted from sEMG signals.

102 We conclude Example 2 by extracting features, previewing the first rows, and outputting
103 package metadata.

```
# Step 7. Extract features and save results in "Features.csv"
df = EMGFlow.extract_features(path_names, muscles, sampling_rate)

# Inspect features
df.round(4).head()

"""
      File_Path  EMG_zyg_Min  ...  EMG_cor_SB  EMG_cor_Spectral_PCT_Missing
0  01/sample_data_01.csv    0.0031  ...    543.1803                0.0050
1  01/sample_data_02.csv    0.0050  ...    346.9988                0.0002
2  02/sample_data_03.csv    0.0001  ...    2183.3999                0.0153
3  02/sample_data_04.csv    0.0024  ...    1051.9444                0.0000

[4 rows x 71 columns]
"""

# Get package version
EMGFlow.package_version()

"""
EMGFlow 1.1.2
"""

# Get package citation
# EMGFlow.package_citation()
```

104 Temporal Feature Extraction

105 The set of 18 time-domain features include statistical moments (mean, variance, skew, kurtosis)
106 and sEMG-specific measures. Examples include Willison amplitude, a proxy for motor unit
107 firing that counts threshold crossings, and log-detector, an estimator of muscle force (Tkach
108 et al., 2010). Time-domain features can be computed after the first three preprocessing steps
109 (notch, band-pass, rectify); Steps 4-6 are optional.

110 Spectral Feature Extraction

111 The 15 frequency-domain features characterise power-spectrum shape and distribution. Median
112 frequency (Phinyomark et al., 2009) tracks changes in conduction velocity and is used in
113 muscle fatigue assessments (Boxtel et al., 1983; Lindstrom et al., 1977; McManus et al., 2020).
114 Standard measures include spectral centroid, flatness, entropy, and roll-off. We also introduce
115 Twitch Ratio, adapted from speech analysis (Eyben et al., 2016), defined as the ratio of upper-
116 to lower-band energy with a 60 Hz boundary between slow- and fast-twitch muscles fibres
117 (Hegedus et al., 2020).

118 Spectral features are computed by converting the Step 2 band-limited signal into a PSD. To
119 avoid discarding otherwise valid Welch frames due to isolated dropouts, we perform constrained
120 interpolation for micro-gaps <5 samples (2.5–5 ms at 1–2 kHz) and leave longer gaps as
121 NaN so affected frames are rejected (Jas et al., 2017). This limits interpolation bias, which
122 increases with gap size and density (Clifford & Tarassenko, 2005; Munteanu et al., 2016). We
123 do not apply Steps 3–6 before PSD: rectification is non-linear and distorts spectra (Farina et
124 al., 2013; McClelland et al., 2014; Neto & Christou, 2010); artefact-replacement filters can
125 violate stationarity assumptions for FFT-based PSD; and smoothing suppresses high-frequency
126 content. We estimate PSD with Welch's method using Hann windows, 50% overlap, and

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