



ATCG...



IN ADVANCE

Grow liquid overnight cultures of the 10 strains to be tested** Don't forget to add ampicillin and IPTG to growth media!

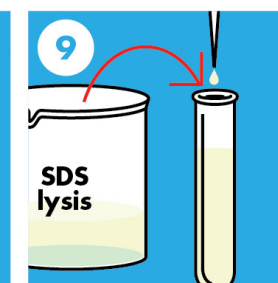
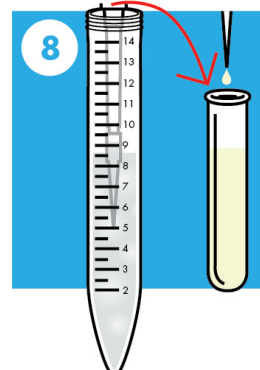
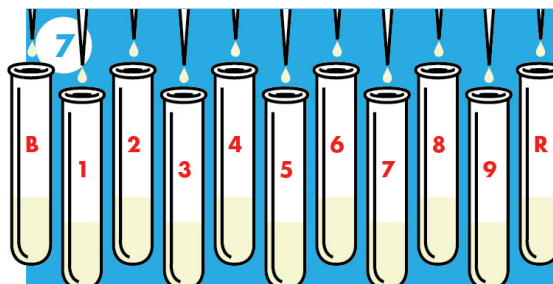
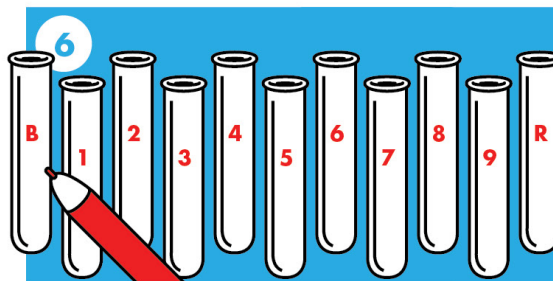
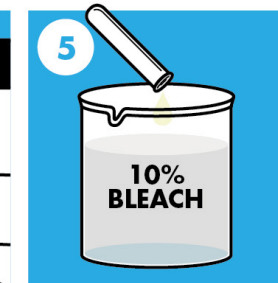
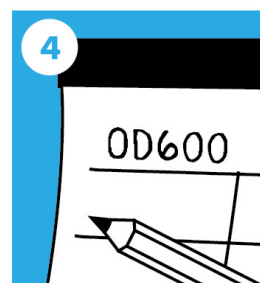
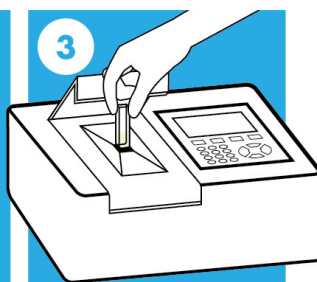
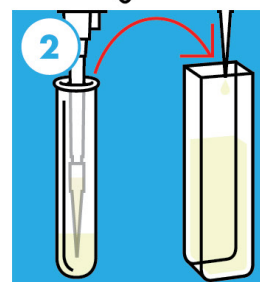
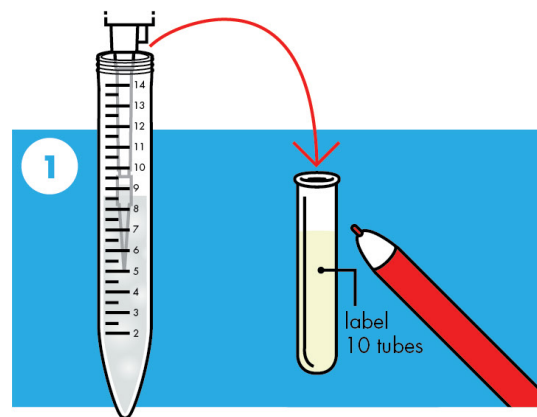
DAY OF LAB

Cell Density Measurements:

1. Label 10 glass tubes "1" through "9" and "R" for reference. Make a 1:10 dilution of each overnight culture by mixing 300 μ l of cells with 2.7 mL of bicarbonate buffer.
2. Transfer each sample to a cuvette, filling the cuvettes about three-quarters full.
3. Read and record the absorbance value for each sample. Start by zero-ing the spectrophotometer set at **600** nm using bicarbonate buffer or water.
4. Record the value x 10 in your data table under the **OD600** heading.
5. Discard all dilutions in 10% bleach.

Enzyme Reactions:

6. Label 11 glass tubes "1" through "9" as well as "B" for blank and "R" for the reference.
7. Add 1.0 mL bicarbonate buffer to each tube.
8. Transfer 100 μ l of cells from the UNDILUTED overnight cultures to the appropriate tube. Transfer 100 μ l of bicarbonate to the blank.
9. Add 100 μ l of SDS lysis solution to each tube, including the blank.



11. Add 50 μ l of competent solution to each tube. Pipet up and down to resuspend the cells. Change pipet tips between tubes.

12. Add 5 μ l of sterile water to the tube labelled (-). This is your negative control. Add 5 μ l of pRS414 DNA to the tube labelled (+). This is your positive control. Add 5 μ l of pRS414+crfYB¹ DNA to the tube labelled **Exp**. Flick all tubes to mix contents.

13. Add 500 μ l of transformation solution to each microfuge tube. Solution will be goopy. Pipet up and down to mix, changing tips between tubes.

14. Incubate tubes at 30° Celcius (C) for one hour along with 3 SC-trp petri dishes with lids slightly ajar to evaporate the moisture from the media surface. Periodically flick the tubes to mix during the incubation.

15. Label the media-side of the petri dishes as (-), (+) or **Exp**.

16. Pipet 250 μ l of each sample onto the media of the appropriate petri dish. Spread the sample evenly across the dish with a sterile spreader. ** Discard spreader and remainder of transformation mix in 10% bleach solution.

17. Incubate petri dishes, media side up, for 2 days at 30°C.

After the petri dishes have incubated for 2 days, count the colonies of each color in every dish.

** VIDEO OF PROCEDURE AVAILABLE ONLINE

