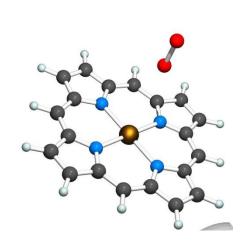
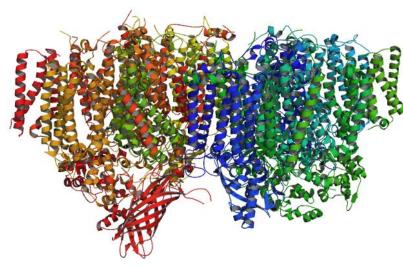
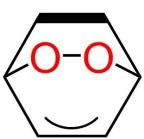
# Bioinorganic Chemistry (BIC) III. Metal-Dependent Hydrolase and Lyase Enzymes







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#### **Review on Part II**

Biological ligands for coordination to metals: Generally, heteroatoms of biological molecules (e.g. proteins, DNA/RNA, cofactor) as potential ligands can coordinate to metal: coordination modes depends on ligands, metal(s), biological molecules.

Coordination to metal can sometimes maintain the overall (folded) structure, but sometimes only stabilize its binding site in some biological molecules.

**Storage & Transport** of small molecules (e.g.  $O_2$  and metal ions) by some bioinorganic systems can keep **sufficient amounts/distribution** (not too many or few) of the molecules for various biological functionals.

## **Classification of Enzymes**

Enzyma Function

Enzyma Class

	Elizyllie Class	Elizyme Function
EC1	Oxidoreductase	Oxidation-reduction
EC2	Transferase	Transfer of a group from one compound to another
EC3	Hydrolase	Hydrolysis
EC4	Lyase	Nonhydrolytic addition or removal of groups
EC5	Isomerase	Conversion of a substance into an isomeric form
EC6	Ligase	Synthesis of a large molecule from two smaller ones

**EC**, Enzyme Commission number: Classification scheme for enzymes based on the **chemical reactions** involved.

## Hydrolases

A family of enzymes catalyze hydrolysis of a chemical bond, in which the metal (cofactor) acts as a Lewis acid generally.

#### $A-B + H_2O \rightarrow A-OH + B-H$

It can be further classified based on the bond involved:

EC 3.1: Ester bonds

EC 3.2: Sugars (DNA glycosylases, glycoside hydrolase)

EC 3.3: Ether bonds EC 3.4: Peptide bonds (proteases/peptidases)

EC 3.5: C-N bonds other than peptide bonds

EC 3.6: Acid anhydrides

EC 3.7: C-C bonds

EC 3.8: Halide bonds

EC 3.9: P-N bonds

EC 3.10: S-N bonds

#### Lyases

A family of enzymes catalyze non-hydrolytic addition or removal of groups (non-H<sub>2</sub>O).

It can be further classified based on the bond involved:

EC 4.1: C-C Lyases, e.g. decarboxylases, aldehyde

Iyases & others C-C Iyases

EC 4.2: C-O Lyases

EC 4.3: C-N Lyases

EC 4.4: C-S Lyases

EC 4.5: C-Halide Lyases

EC 4.6: P-O Lyases

EC 4.99: Other lyases

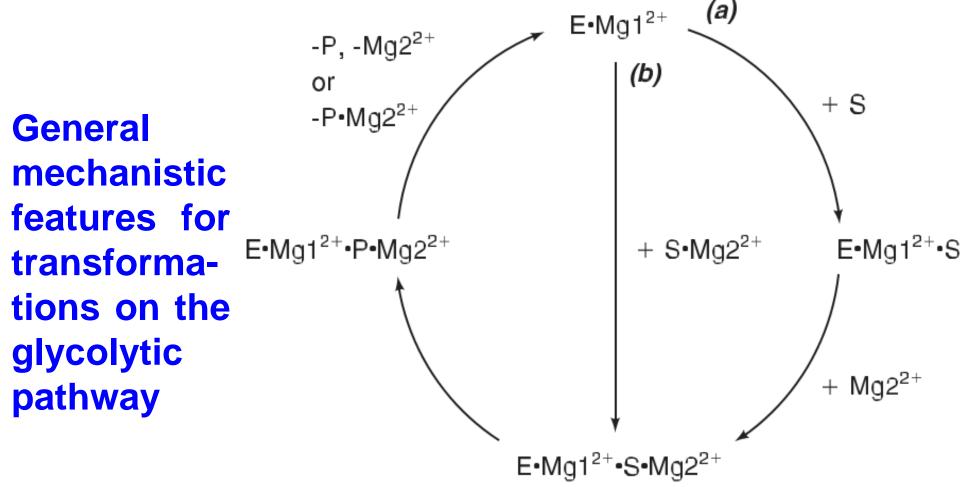
- For metal-dependent hydrolases & lyases, a metal cation, or sometimes cations, (as the cofactor) with a high Lewis acidity is important for many hydrolysis & condensation reactions under physiological conditions.
- The general roles of the metal(s):
- (1) **activate** a metal-bound **nucleophile** (e.g. ionization of M-OH<sub>2</sub> to give a **more reactive** M-OH form),
- (2) stabilize a metal-bound intermediate/product, or (3) sometimes bring the reactant species in the active site (structural role).

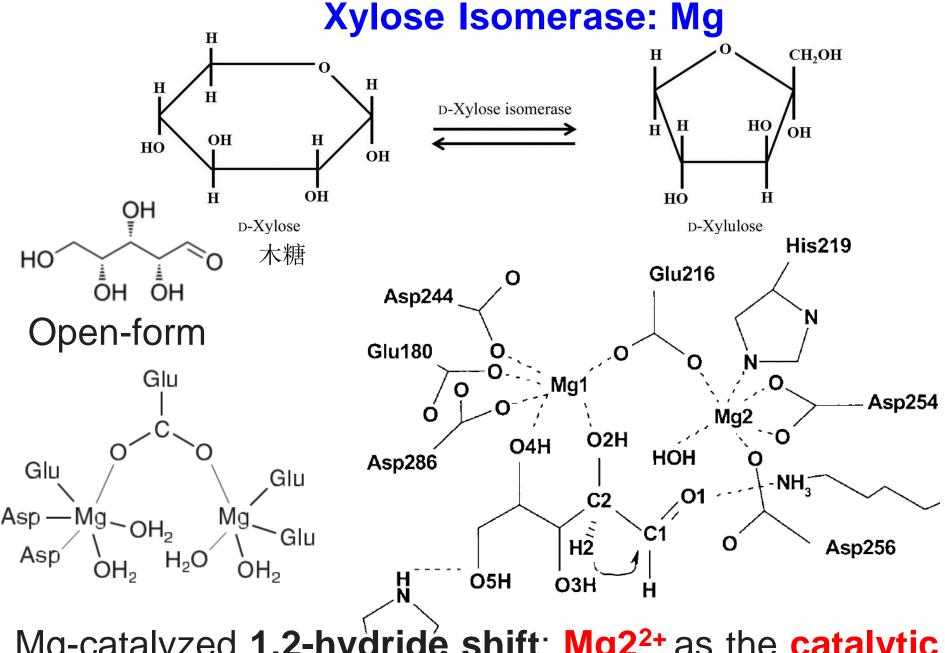
M + H<sub>2</sub>O → M-OH + H<sup>+</sup>
Zn(II), p
$$K_a$$
: ~8.8; Mg(II), p $K_a$ : ~11.4
p $K_W$ : 14.0 (25 deg.)

- <u>Non-redox</u> metals are widely used in these enzymes to protect sensitive functional groups in proteins and nucleic acids from oxidative damage. Namely, Zn<sup>2+</sup> & Mg<sup>2+</sup> (sometimes Ca<sup>2+</sup>) are common metals.
- "Intermediate" Lewis acid Zn<sup>2+</sup> ion center in these enzymes is usually tetrahedral & coordinates to "Intermediate" base histidine (His) & sometimes to soft cysteine (Cys) or hard carboxylate residues or H<sub>2</sub>O.
- Whereas, harder Mg<sup>2+</sup> ion center in these enzymes typically coordinates to hard carboxylate residues or water molecules (usually octahedral).

#### Magnesium (Mg)

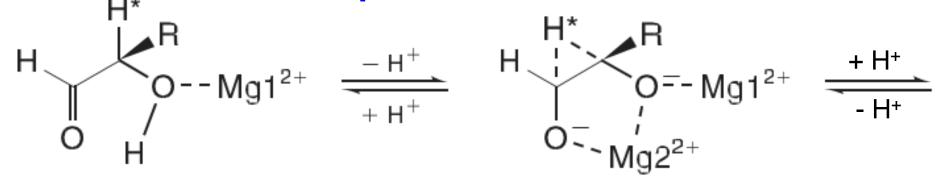
Some Mg-dependent enzymes require at least two metal-binding sites: (1) an allosteric regulatory site modulating either structure or binding (structural role), (2) a catalytic site.





Mg-catalyzed 1,2-hydridé shift: Mg2<sup>2+</sup> as the catalytic ion, while Mg1<sup>2+</sup> likely plays a structural role.

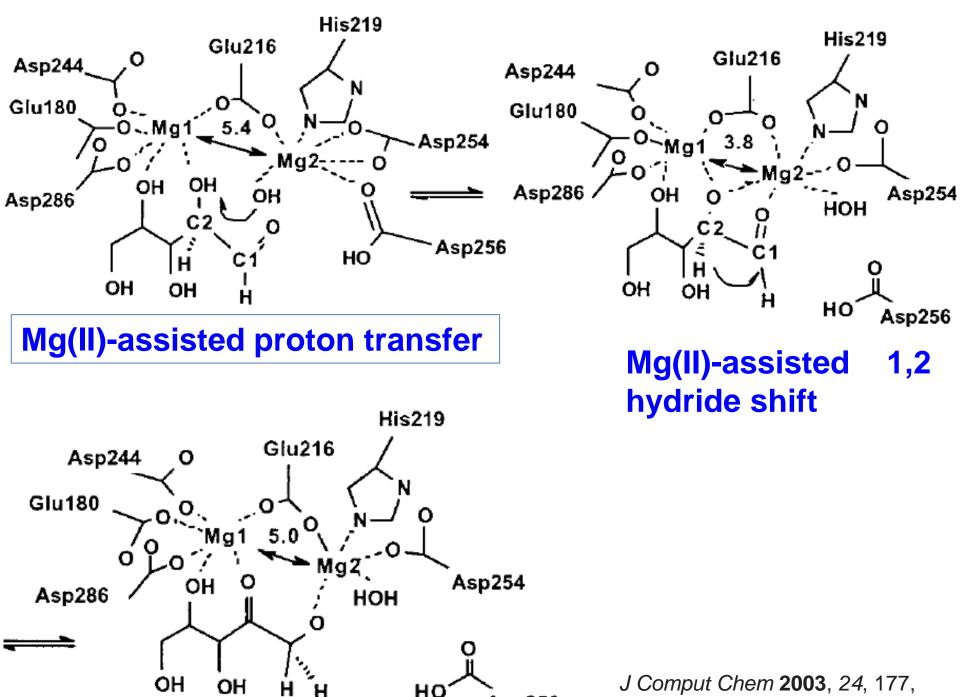
#### **Proposed Mechanism**



 $Mg2^{2+}$ 

Kinetic results suggests:

- Mg<sup>2+</sup> first binds to site 1 (higher affinity), facilitating substrate binding & proper orientation and stabilizing intermediates.
- The **second Mg**<sup>2+</sup> then binds, & **initiates the reaction**.
- A Mg<sup>2+</sup>-bound water molecule likely assists proton transfer between the hydroxyl and carbonyl oxygens.

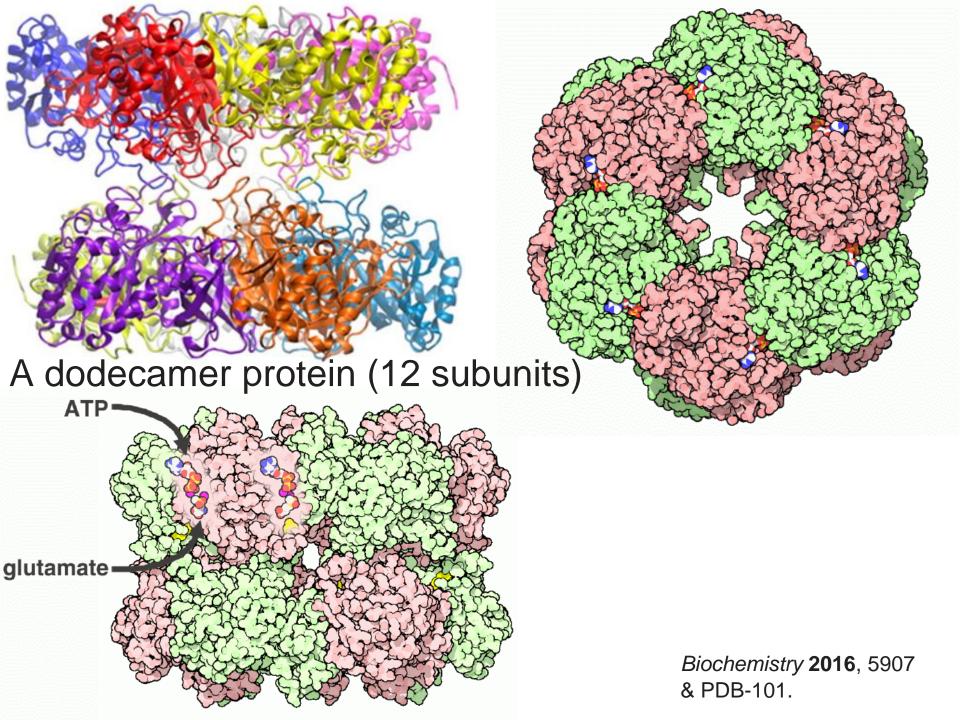


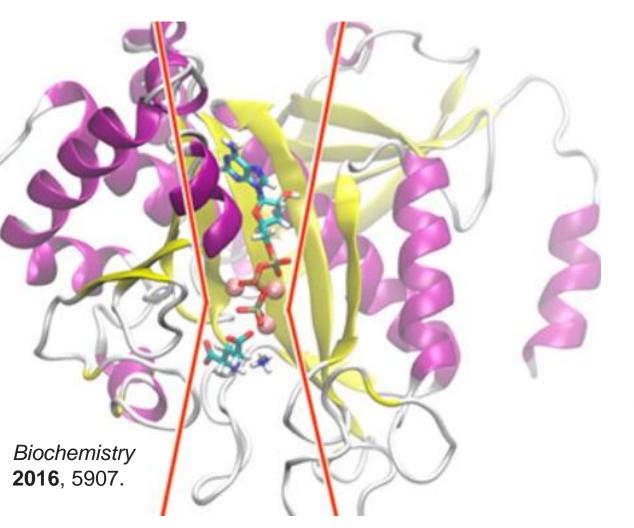
HO

Asp256

#### **Glutamine Synthetase: Mg**

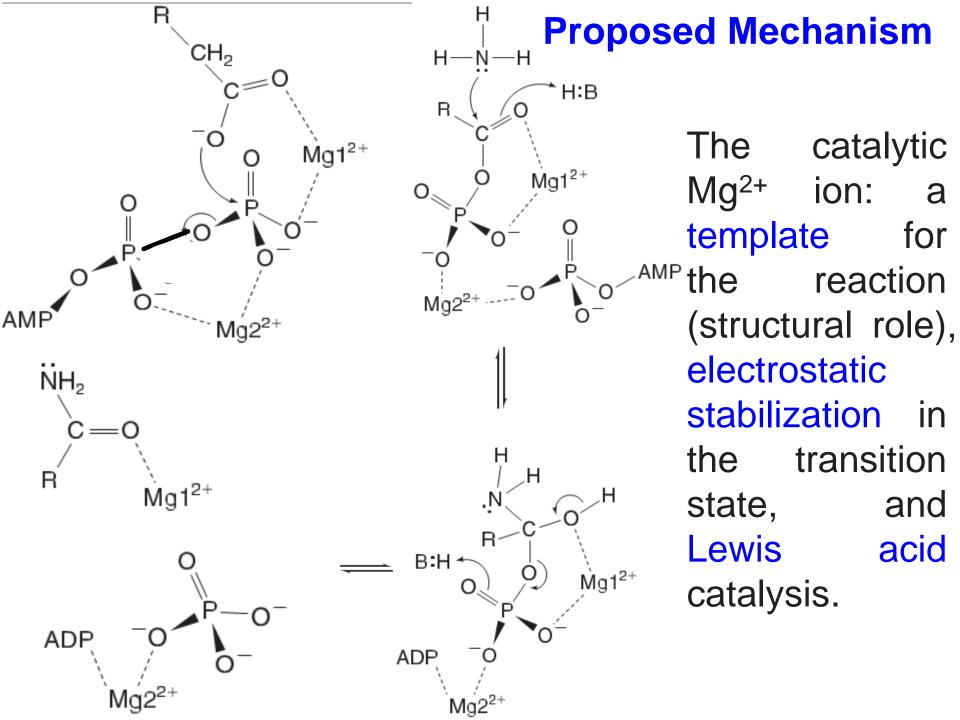
Catalyzes the formation of **glutamine** (**GIn**) from **glutamate** (**Glu**) & **NH**<sub>3</sub> with hydrolysis of ATP.





Crystal structure
& kinetic studies:
metal-binding
sites (high & low
affinity) with a Mg-Mg distance (5.8 Å).

• The high-affinity site (catalytic cofactor), while the weakly bound ion (possible binding of an Mg-ATP chelate) in the low-affinity site.

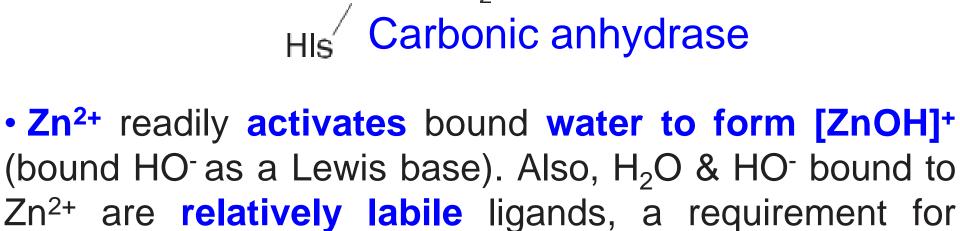


#### Zinc (Zn)

• pK<sub>a</sub> for  $Zn^{2+}$ (aq) is ~9-10, & can drop ~7 in an enzyme environment with the **more hydrophobic** environment (favoring reduction in the overall charge).

HIS  $\neg$   $^{2+}$  HIS - Zn $^{II}$  - OH $_2$ 

hydrolysis reactions.



• While, Zn<sup>2+</sup> forms kinetically inert bonds to His residues.

#### TABLE 12.1 Coordination Motifs in Catalytic Sites of Some Typical Mononuclear Zinc Enzymes

Carbonic anhydrase His-X-His-X<sub>22</sub>-His

β-lactamase His-X-His-X<sub>121</sub>-His

Thermolysin His-X<sub>3</sub>-His-X<sub>19</sub>-Glu

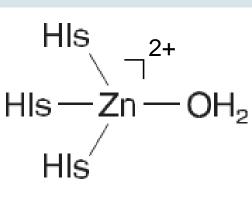
Carboxypeptidase His-X<sub>2</sub>-Glu-X<sub>123</sub>-His

Alcohol dehydrogenase Cys-X<sub>20</sub>-His-X<sub>106</sub>-Cys

Alkaline phosphatase Asp-X<sub>3</sub>-His-X<sub>80</sub>-His

Adenosine deaminase His-X-His-X<sub>196</sub>-His

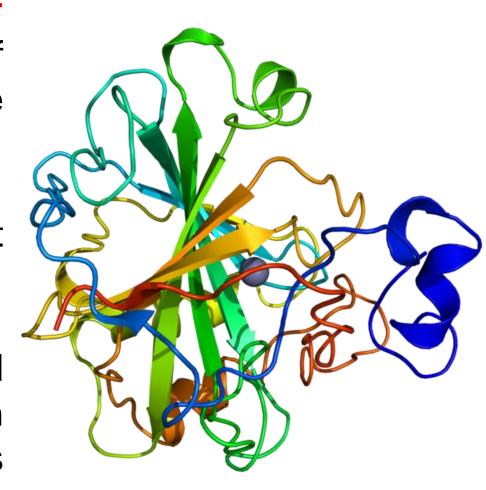
Carbonic anhydrase

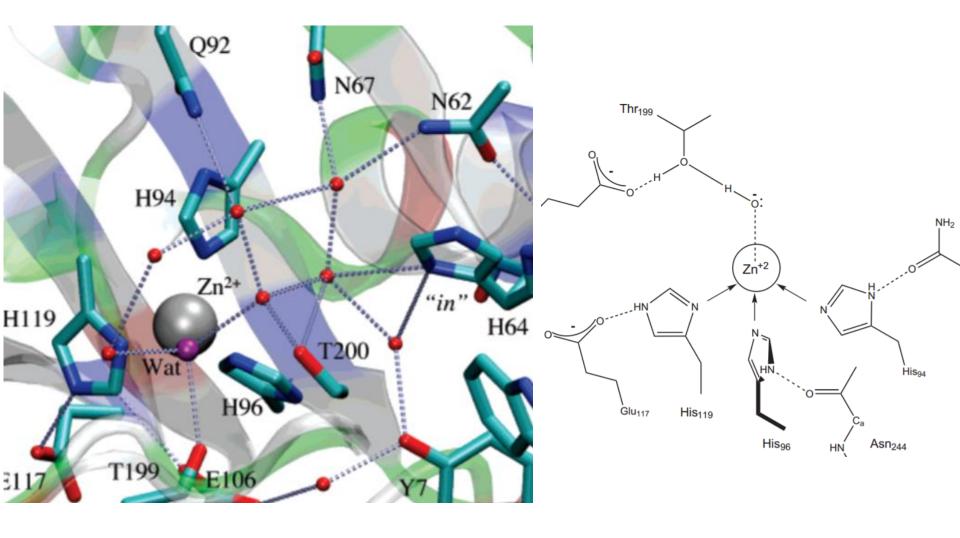


#### Carbonic Anhydrase (CA): Zn

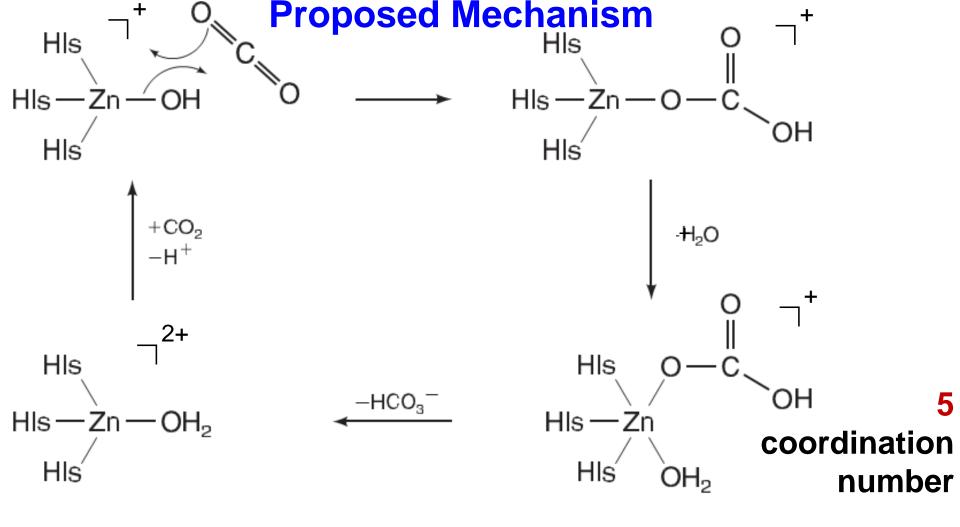
$$CO_2 + H_2O \rightarrow HCO_3^- + H^+$$

- Catalyzes hydration of CO<sub>2</sub>, with acceleration of ~10<sup>7</sup> relative to the uncatalyzed reaction.
- The first Zn-dependent enzyme to be discovered.
- Prevalent in red blood cells & key in respiration and pH buffering: CA helps transport of CO<sub>2</sub>.





• Zn<sup>2+</sup> with **3 His** and **1 labile** H<sub>2</sub>**O** (distorted tetrahedron coordination). Water molecules in the active site play key structural & functional roles (proton transfer).

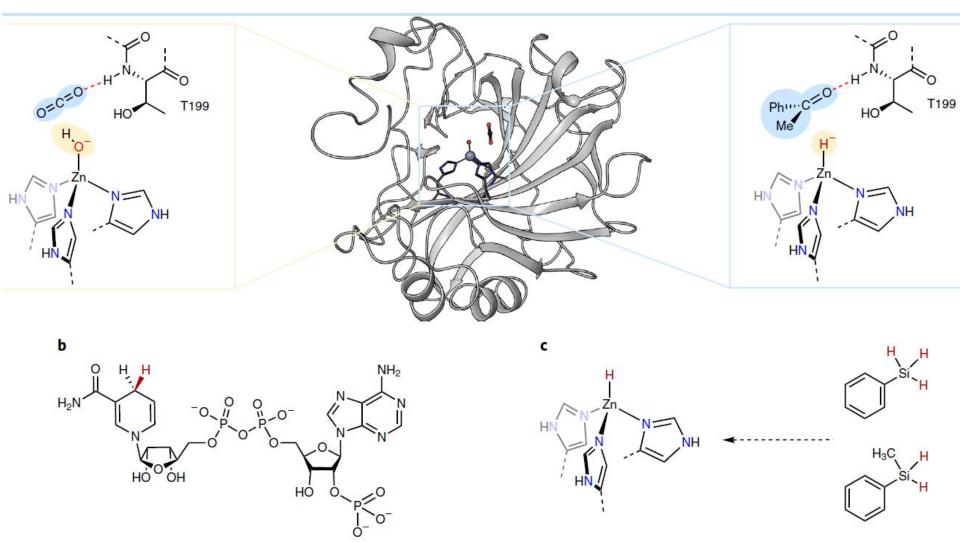


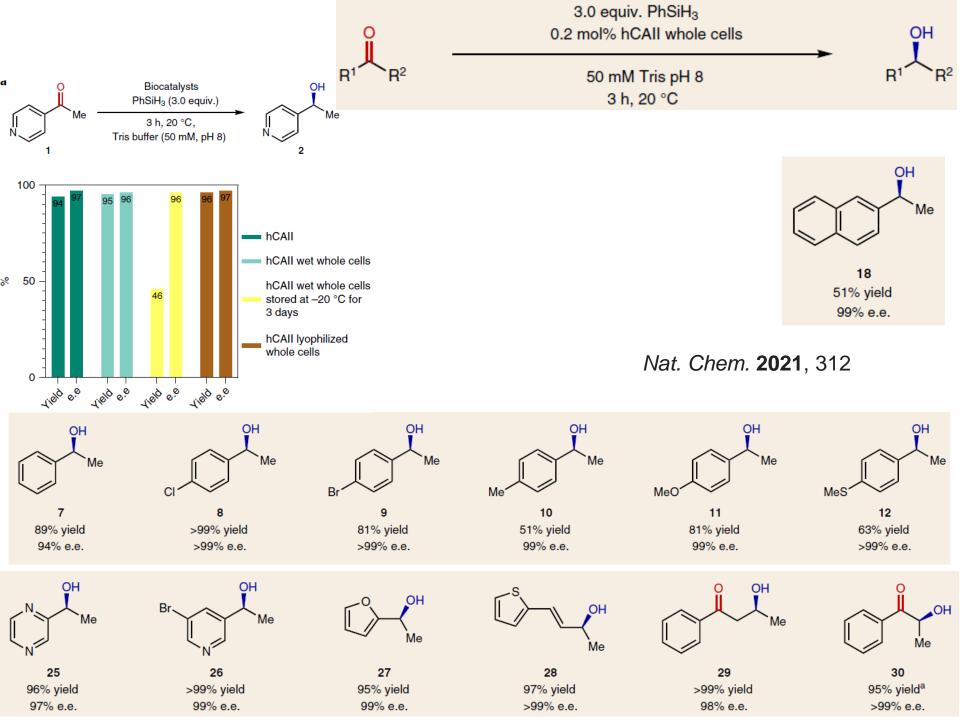
- A **Zn-bound hydroxide** (from **deprotonation**) attacks  $CO_2$ . The  $Zn^{2+}$  ion also **polarizes the O=C bond** and stabilizes the developing negative charge (Lewis acid).
- Subsequent uptake of a H<sub>2</sub>O & release of HCO<sub>3</sub><sup>-</sup>.

# Abiotic reduction of ketones with silanes catalysed by carbonic anhydrase through an enzymatic zinc hydride

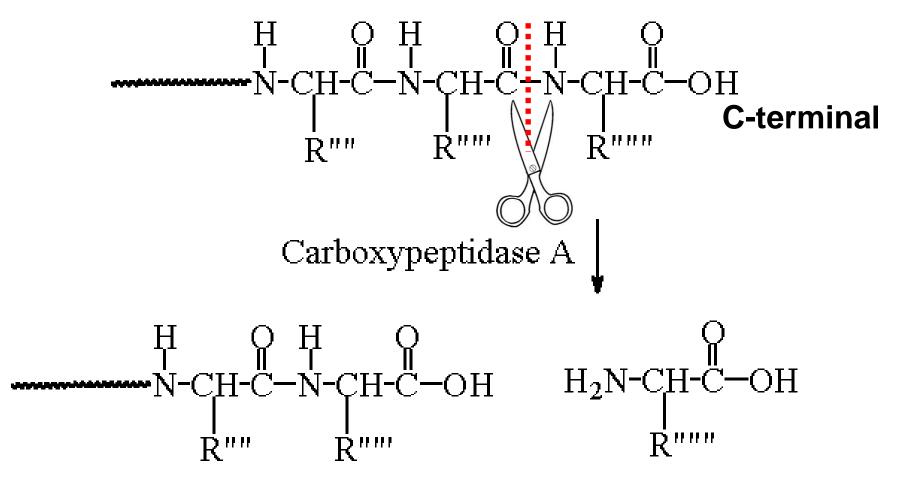
Nat. Chem. 2021, 312

Pengfei Ji<sup>1</sup>, Jeeyoung Park<sup>1</sup>, Yang Gu<sup>1</sup>, Douglas S. Clark<sup>1,2</sup> and John F. Hartwig<sup>1</sup>



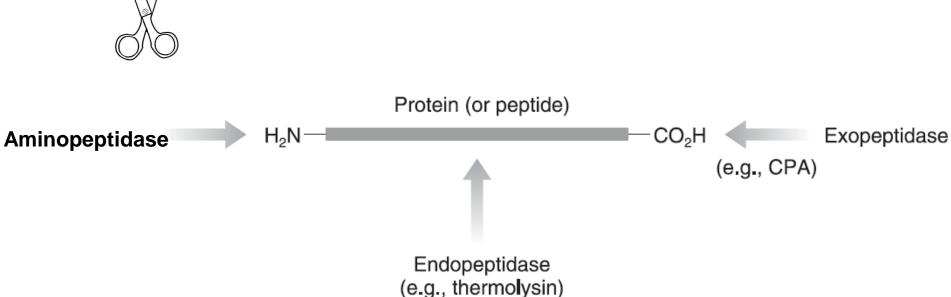


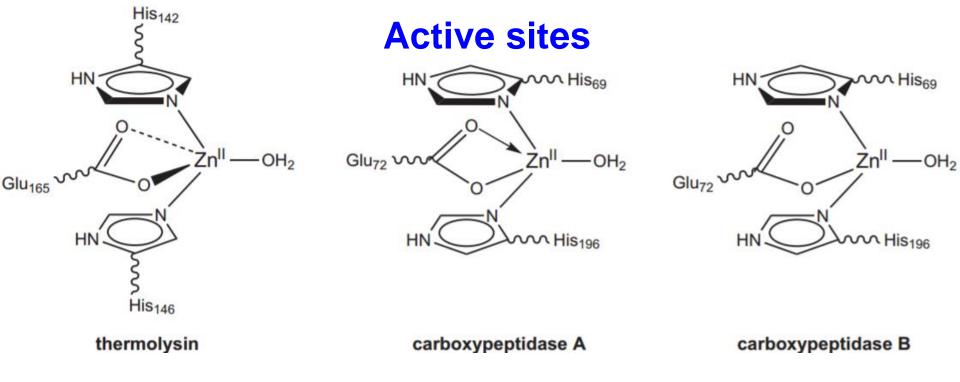
#### Carboxypeptidase: Zn



• Carboxypeptidase A (CPA): a digestive enzyme that hydrolyzes/cleaves a peptide bond of residues with aromatic or aliphatic side-chains at the carboxyterminal (C-terminal) of a protein.

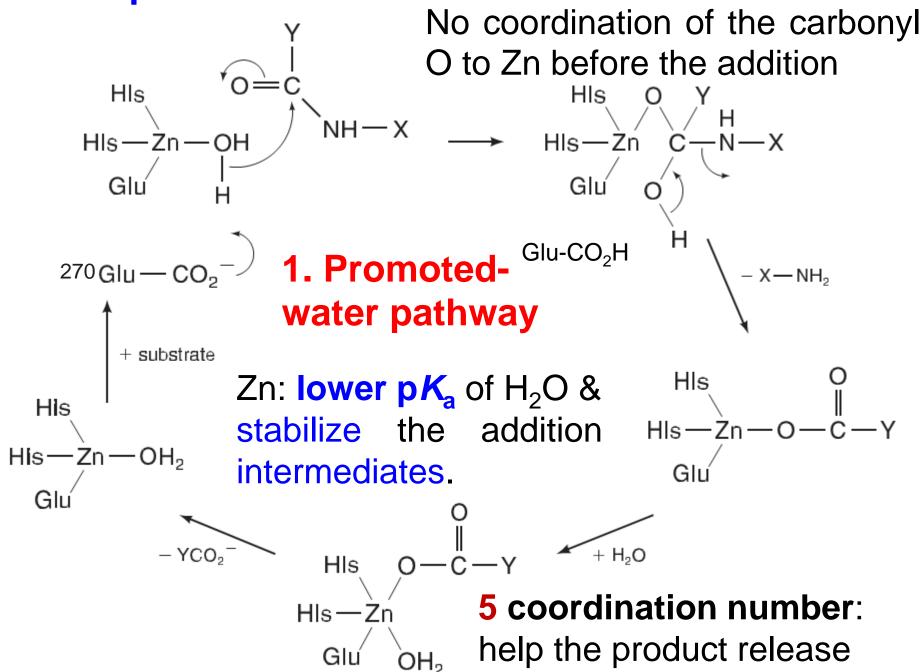
 Carboxypeptidase B (CPB) is similar to CPA, but it favors to cleave a peptide bond of residues with basic residues (Arg, Lys).



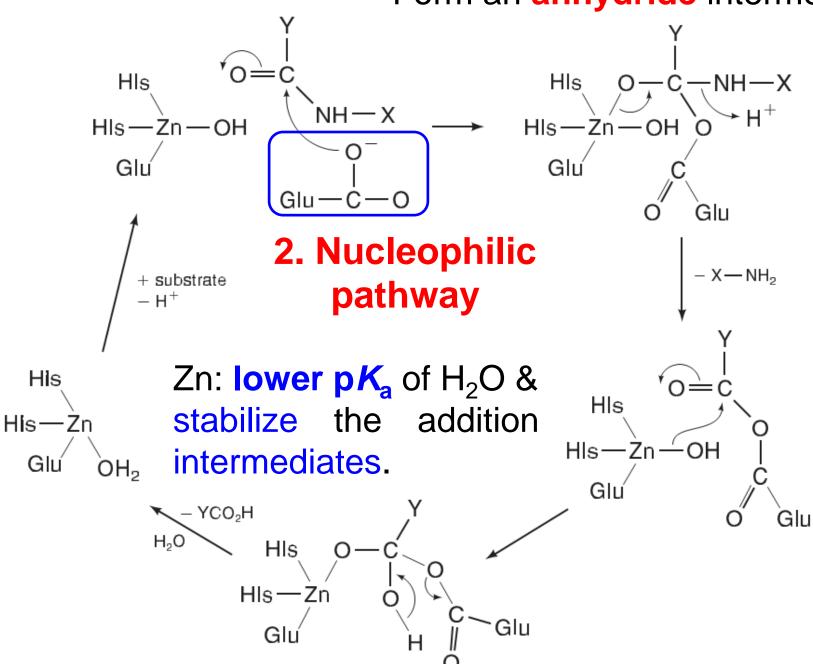


- The Zn ion in CPA: tetrahedral with 2 **His** ligands, 1 bidentate **Glu** & 1 **H<sub>2</sub>O**, which is hydrogen bonded to Ser197 and Glu270.
- In the presence of the substrate, the bound Glu shifts to monodentate coordination.
- The enzyme can also hydrolyze esters.

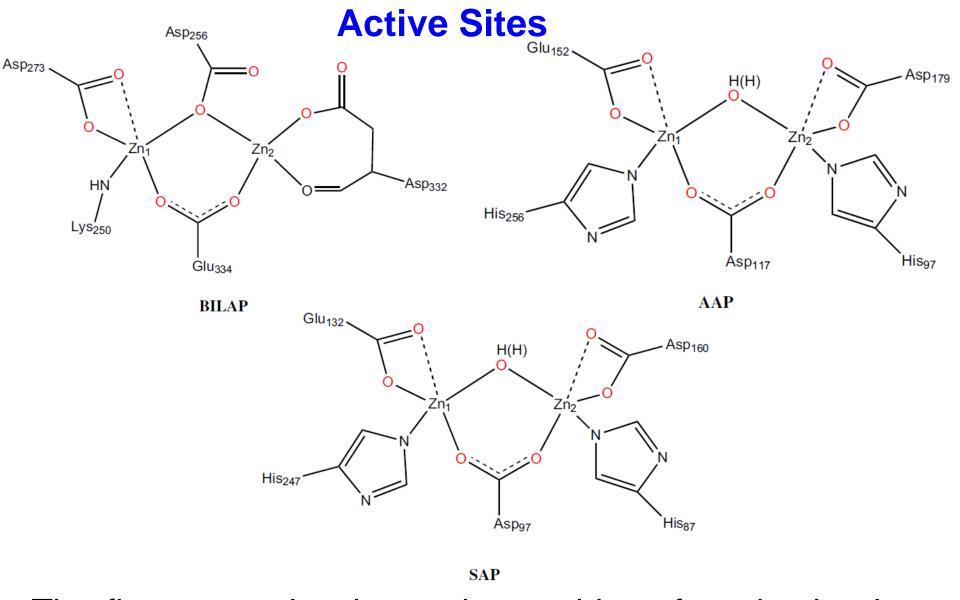
#### **Two Proposed Mechanisms**



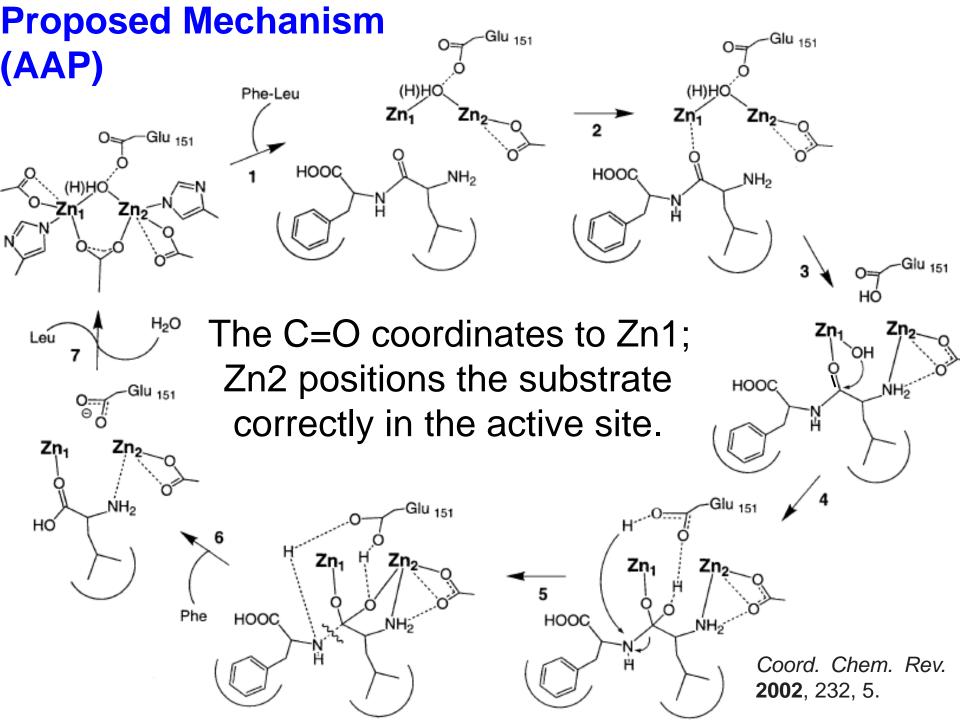
#### Form an anhydride intermediate



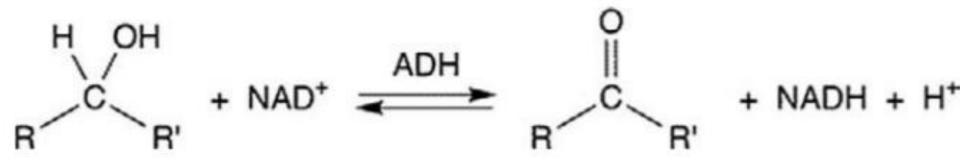
 Hydrolyzes/cleaves a peptide bond of residues at the amino-terminal (N-terminal) of a protein. A dinuclear Zn active site was found & both Zn ions are required with different roles.



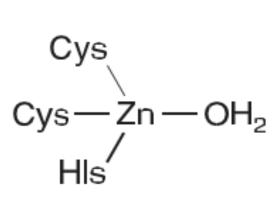
• The first group: leucine aminopeptidase from bovine lens; the second group: leucine aminopeptidases AAP from *Aeromonas proteolytica* and SAP from *Streptomyces griseus*.



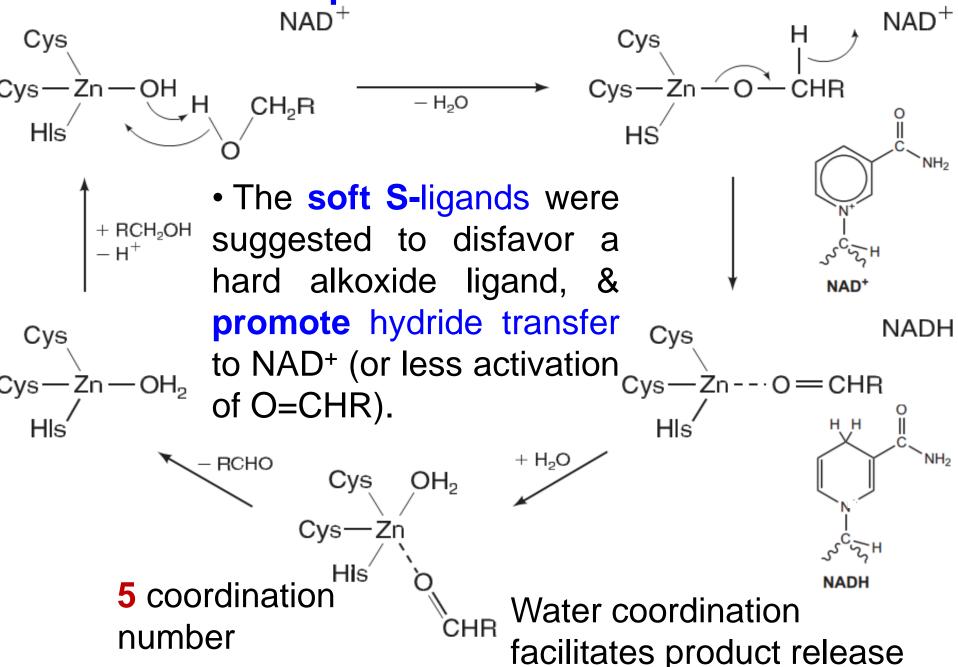
#### Liver Alcohol Dehydrogenase (LADH): Zn



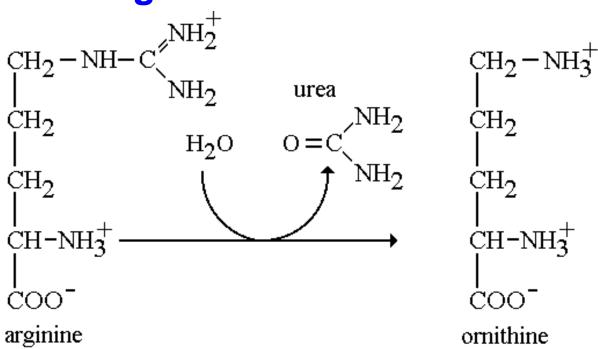
- Formally, LADH catalyzes an oxidation of a primary or secondary **alcohol** to give an **aldehyde** or **ketone**, respectively, through **hydride transfer to NAD**+.
- LADH in mammalian liver are dimeric proteins with 2 Zn<sup>2+</sup> ions in each subunit: one is catalytically active (coordinating with 1 His, 2 (soft) Cys & 1 H<sub>2</sub>O) & the other Zn plays a structural role (with 4 Cys).



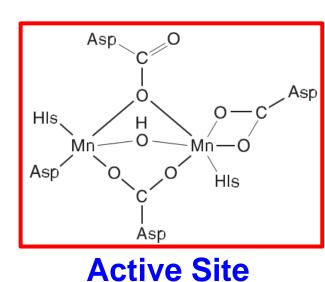
#### **Proposed Mechanism**



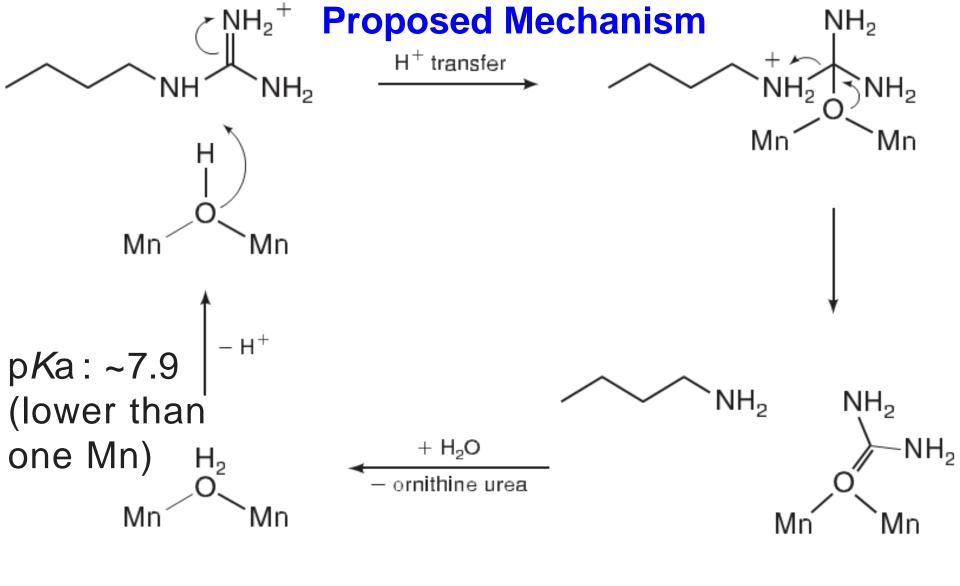
#### **Arginase: Mn**



- Catalyzes hydrolysis of arginine to give urea & ornithine (the final step in the urea cycle).
- A trimer protein; 2 Mn ions has 3 bridging ligands (a bidentate Asp, a monodentate Asp and a hydroxide) in the active site.



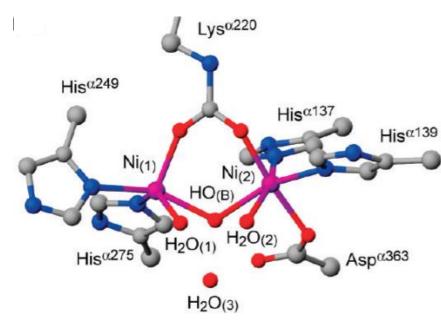
(Mn-Mn: ~3.3 Å)



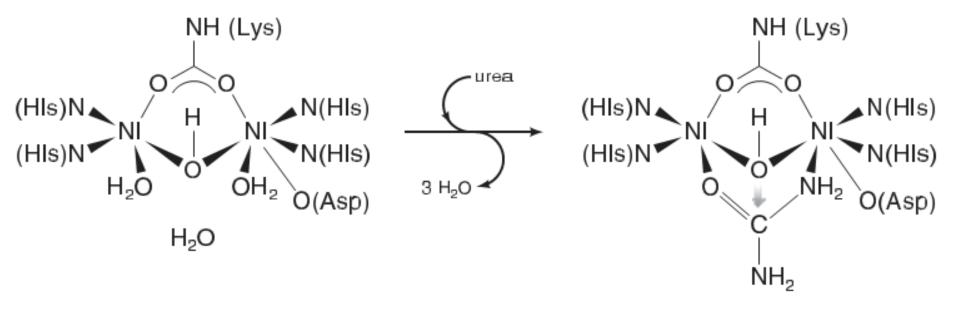
• The substrate does not bind directly to the metal cofactors. Why? Why not proton transfer to the terminal N atom?

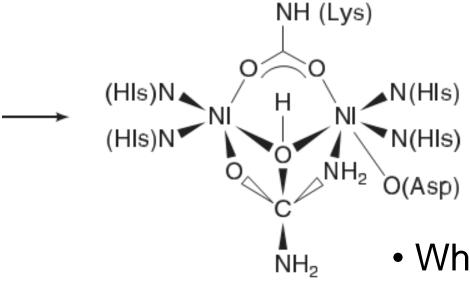
# 

- Hydrolyzes urea to give NH<sub>3</sub> & carbamate, which decomposes to give another NH<sub>3</sub> & HCO<sub>3</sub><sup>-</sup> (key role in nitrogen metabolism in plants & microbes).
- It was the **first Ni enzyme** & contains **two Ni ions**.



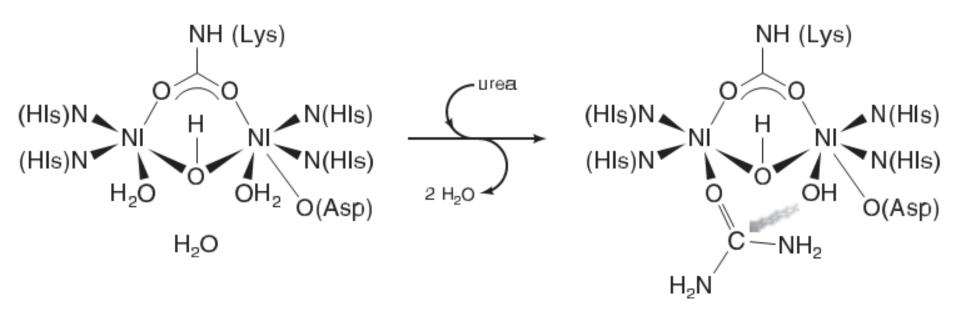
#### **Two Proposed Mechanisms**

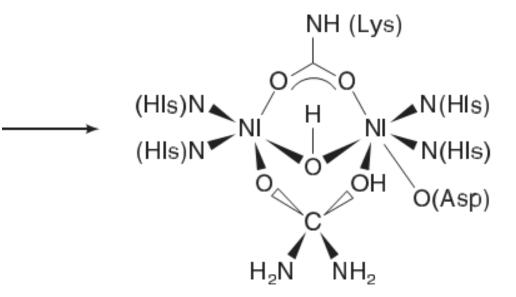




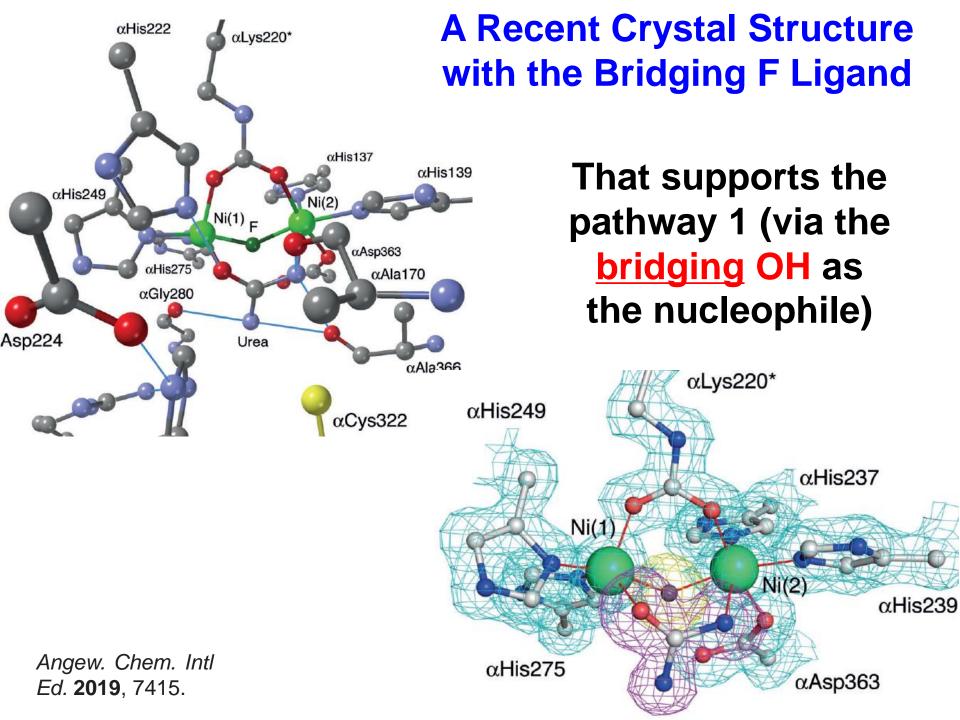
1. A pathway via the bridging hydroxide as the nucleophile

 Why the urea O preferentially coordinates to the left Ni center?



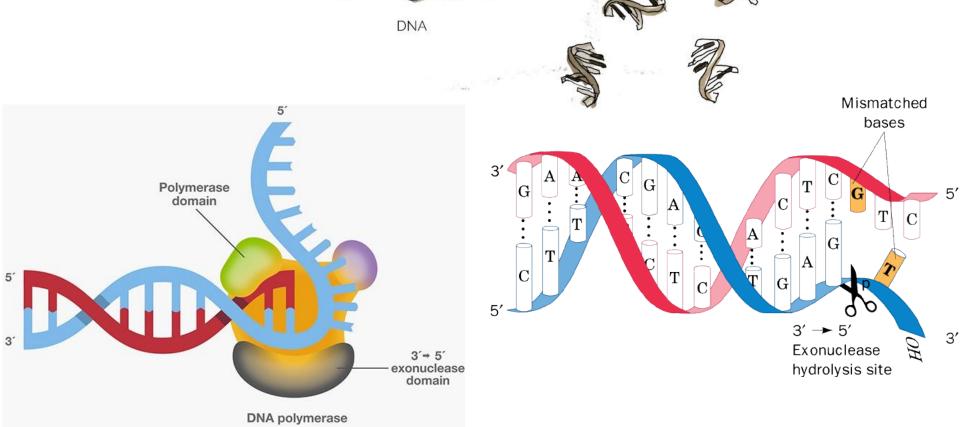


2. A pathway via the terminal hydroxide as the nucleophile



# Hydrolysis or Condensation Reactions with Nucleic Acids

**NUCLEASE** 



### Hydrolysis or Condensation Reactions with **Nucleic Acids**

 Almost all these enzymes require divalent metal ion cofactor to promote activity. Mg ion (relatively labile) is typically used, while Ca & Zn ions are also used in some cases.

Ion	Coordination Numbers	Geometry a	Radius (Å)	Ligand P
$Mg^{2+b}$	6	oct	0.65	$O; (H_2O)$
$Mn^{2+}$	5, 6	dist oct, sq pyr	0.85	O, N,

tet, sq pyr

dist oct, tet, sq pyr

0.81

0.79

N, S

O, N, S

4, 5, 6

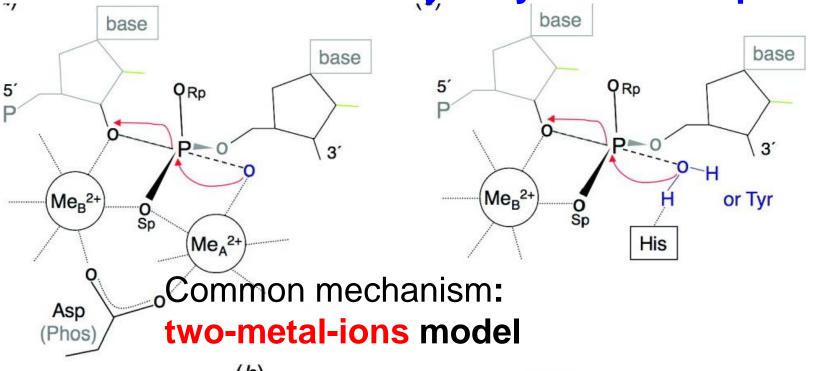
4, 5

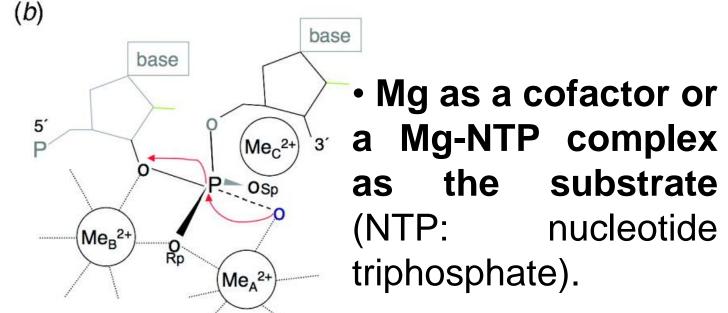
 $Zn^{2+}$ 

<sup>&</sup>lt;sup>a</sup> Octahedral = oct, distorted octahedral = dist oct, square pyramidal = sq pyr, tetrahedral = tet.

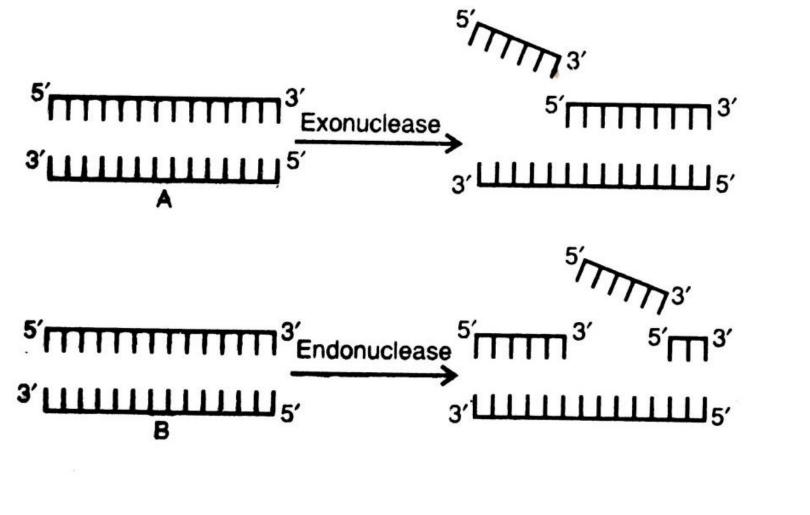
<sup>&</sup>lt;sup>b</sup> Maguire, M. E. and Cowan, J. A., "Magnesium Chemistry and Biochemistry", *Biometals* 15, 203–210 (2002).

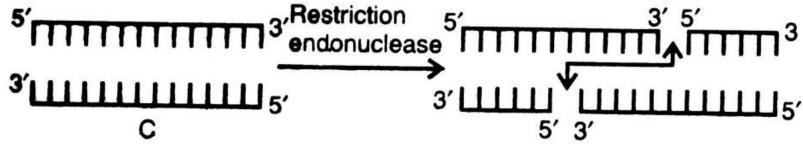
3 Generic Models for Hydrolysis of Phosphodiester





### **Nucleases** (cleavage of poly-nucleotide chain)





#### **Exonucleases**

 Cleave fragments from **end** of a polynucleotide chain (DNA or RNA) by **hydrolysis** of the terminal phosphodiester linkage.

dн<sub>z О</sub>

5' CH2 0

ÓΗ

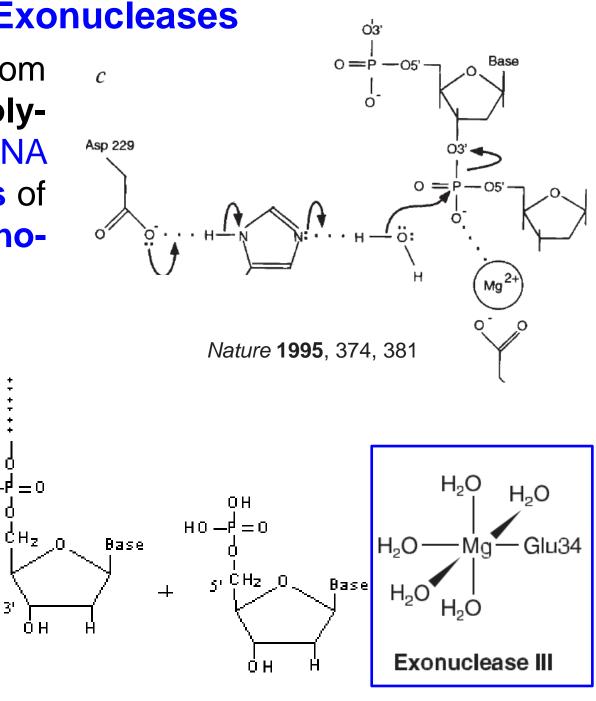
Base

Base

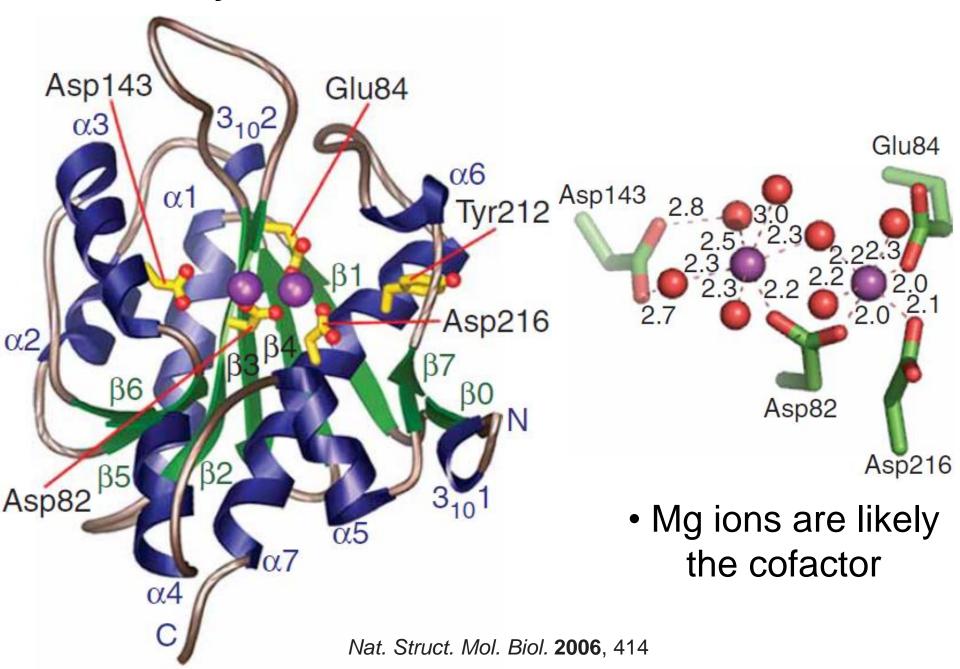
Exonuclease III

 $HO - \neq = 0$ 

ÓΗ



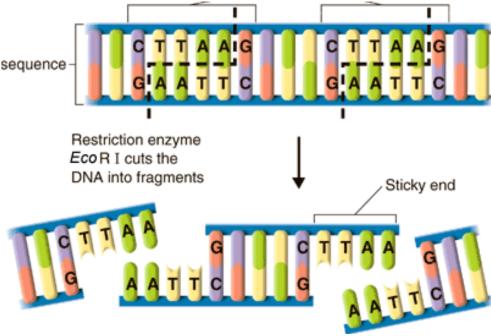
### **Crystal Structure of WRN Exonuclease**

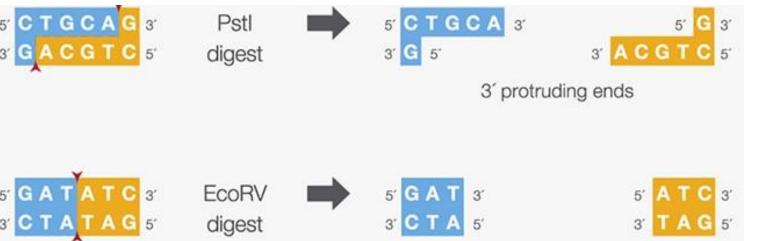


### **Endonucleases**

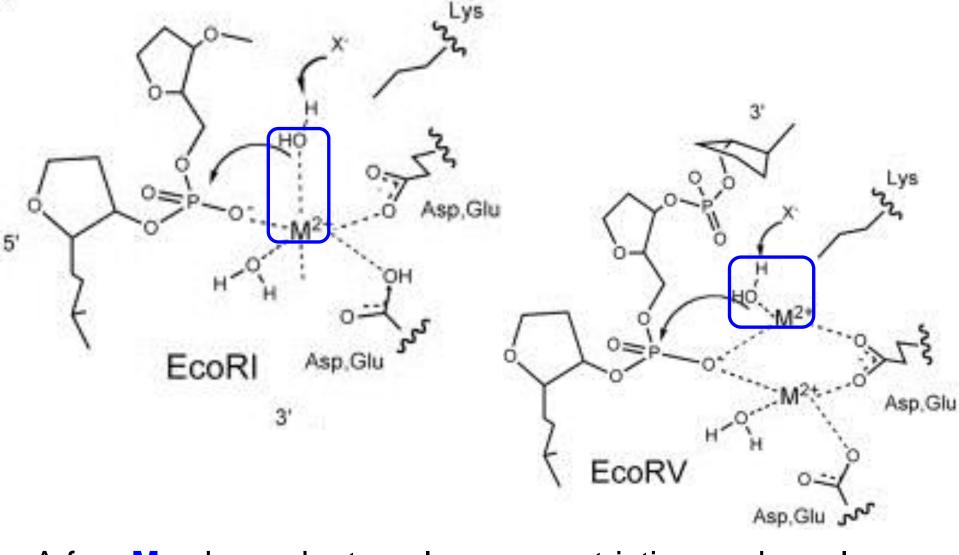
• Endonucleases cleave the phosphodiester bond within a polynucleotide chain.

• Restriction endonucleases cleave only at very specific nucleotide sequences (Most restriction enzymes are endonucleases).



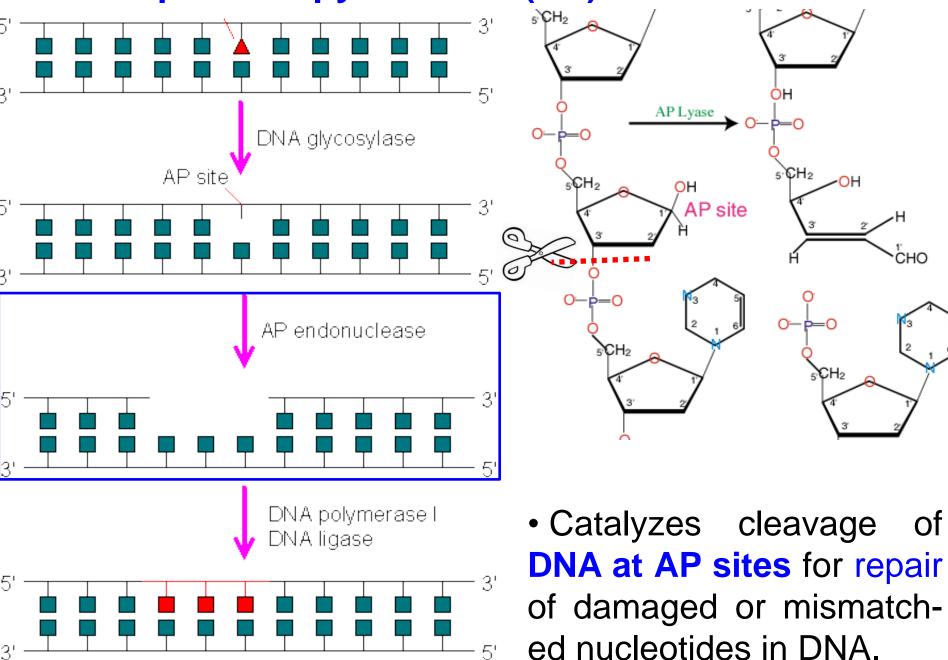


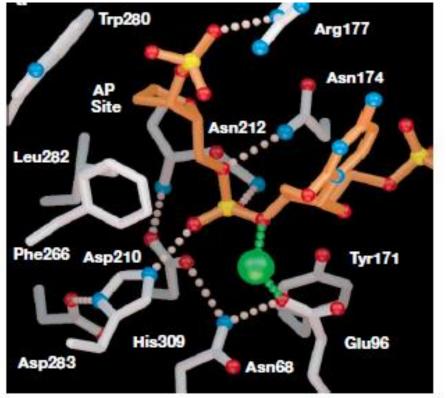
### **Restriction Endonucleases**



• A few Mg-dependent nucleases: restriction endonucleases. Some uncertainty about the metal cofactor stoichiometry, although an inner-sphere pathway seems to be likely.

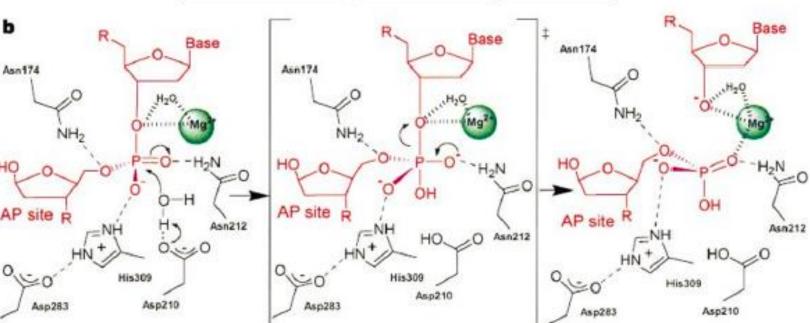
### Apurinic/apyrimidinic (AP) Endonuclease



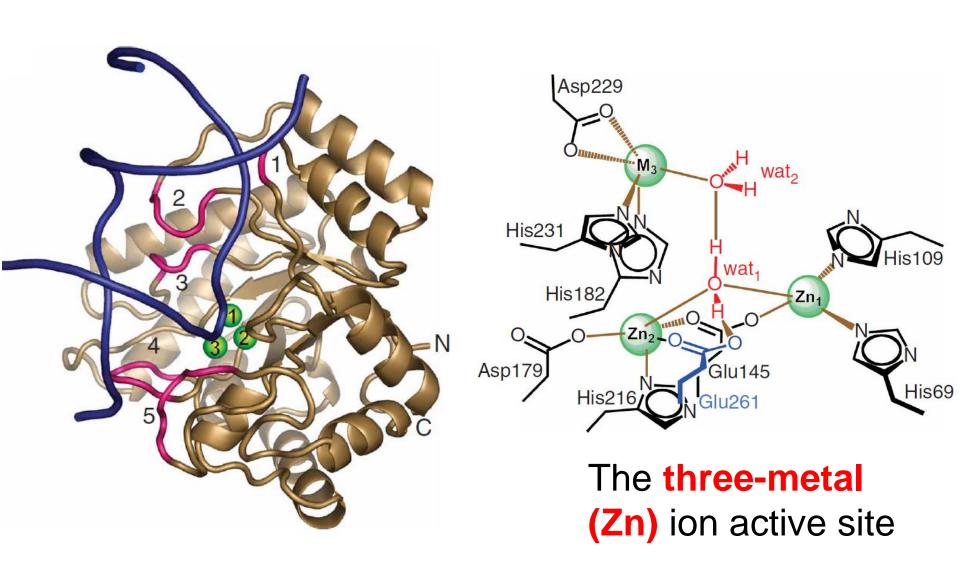


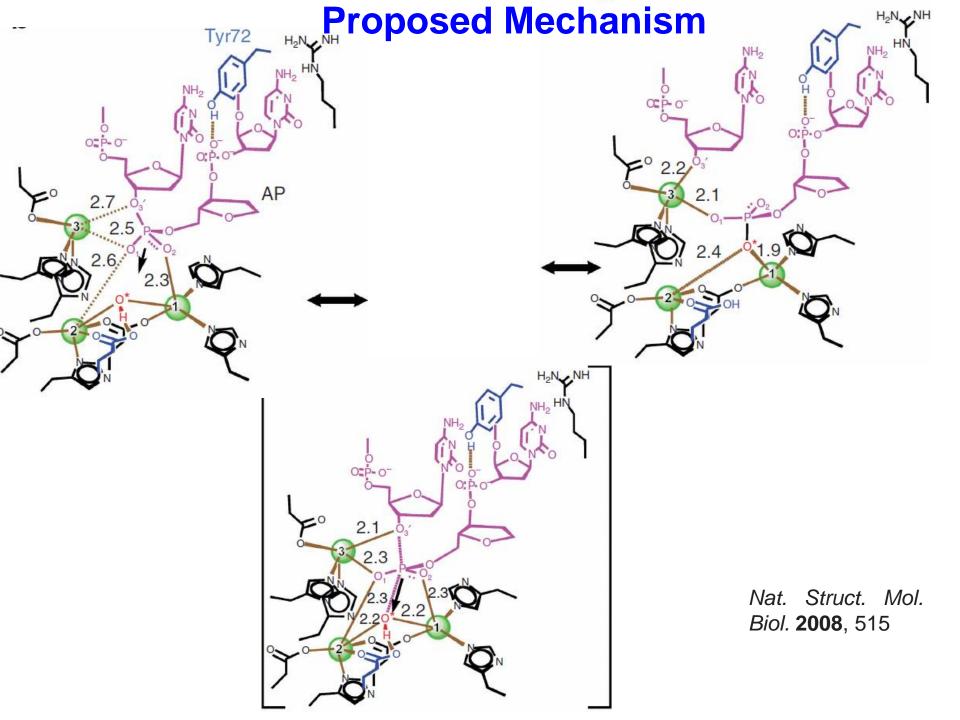
## Human base excision repair enzyme APE1

*Nature* **2000**, 403. 451

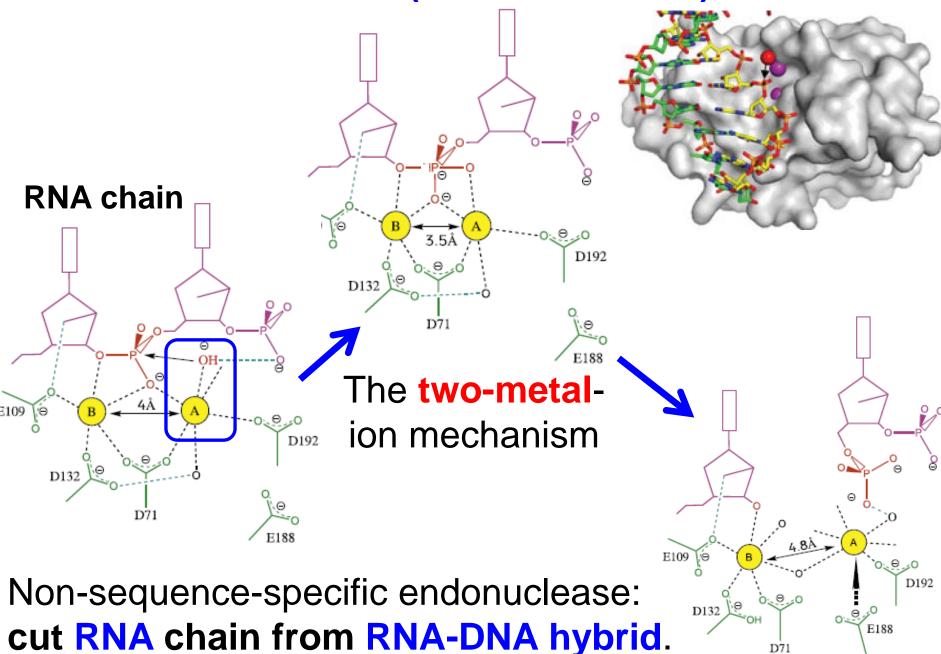


### E. Coli Endonuclease IV





### RNase H (Ribonuclease H)

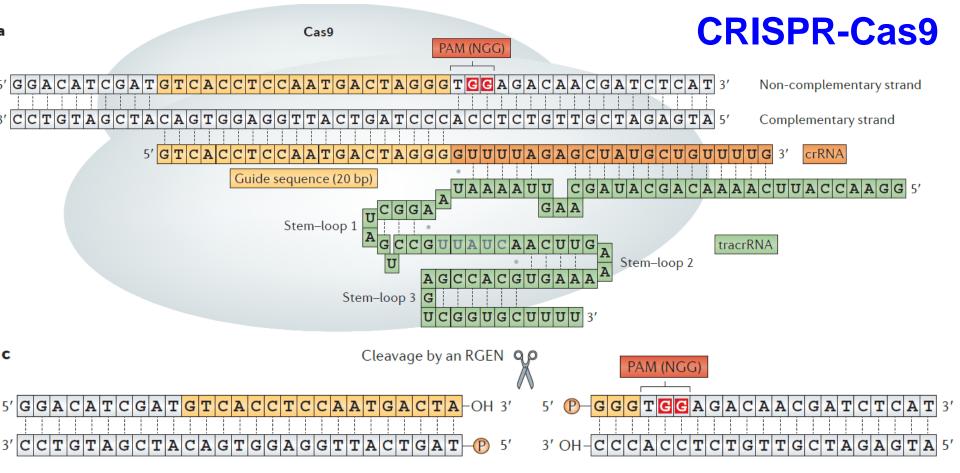


Staphylococcal Nuclease Arg87 HN N—Н CH<sub>3</sub>  $H_2N$ NH Arg35 Asp21 Ca<sup>2+</sup> Asp40 Glu43 Thr41

The active site with an inhibitor thymidine-3',5'-bisphosphate

enzyme hydrolyzes DNA or RNA.

• Ca<sup>2+</sup> binds the to phosphodiester stabilizes the leaving group after hydrolysis.



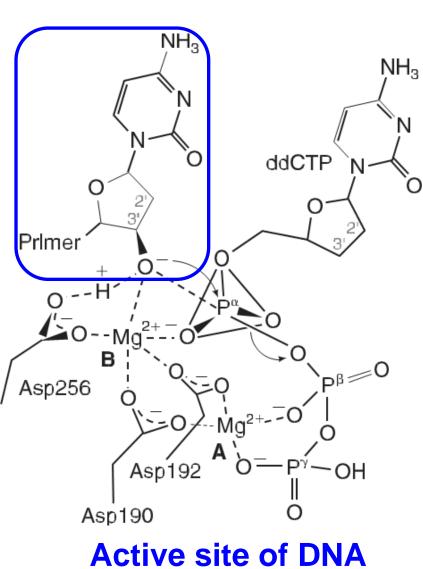
- CRISPR (clustered regularly interspaced short palindromic repeat)-associated protein 9 (Cas9)
- RGENs (RNA-guided engineered nucleases); crRNA (CRISPR RNA); tracrRNA (a transactivating crRNA)

  Nat. Rev. Genet. 2014, 321; Annu. Rev. Biochem. 2013, 237

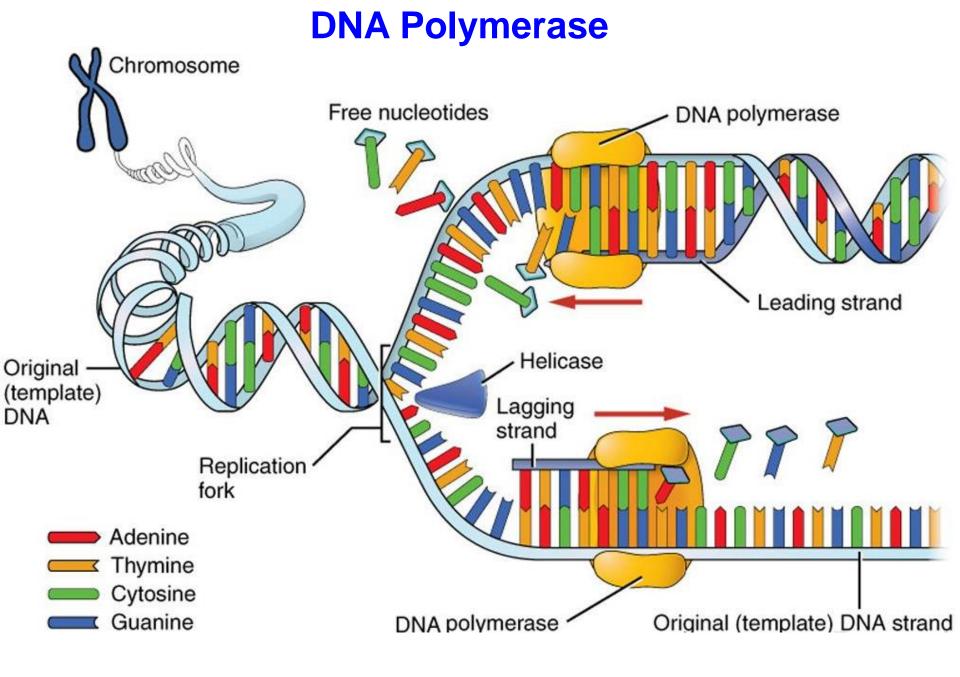
RNase III

### **Polymerases**

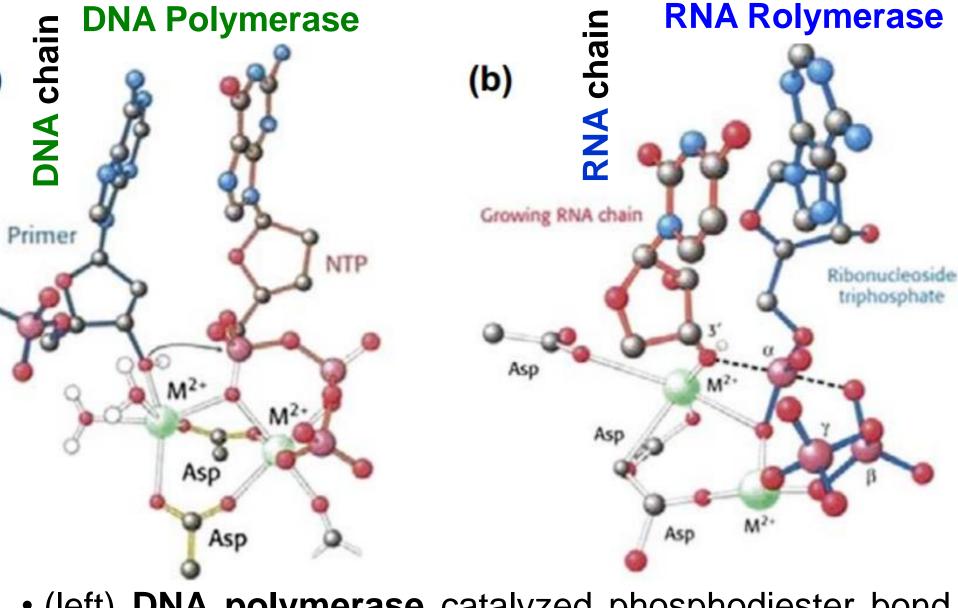
- Catalyzes replication & synthesis of DNA or RNA strands.
- Typically require 2 metals (usually Mg<sup>2+</sup>), & 2 bridging Asp groups.
- Acidic active-site residues can interact with the Mg-phosphate chelate to promote nucleophilic attack at the phosphate.
- Both metals help stabilize the negative charge on the 5-coordinate transition state & the leaving group during phosphoryl or nucleotidyl transfer reactions.



**polymerase** β



Catalyzes synthesis of DNA from deoxyribonucleotides

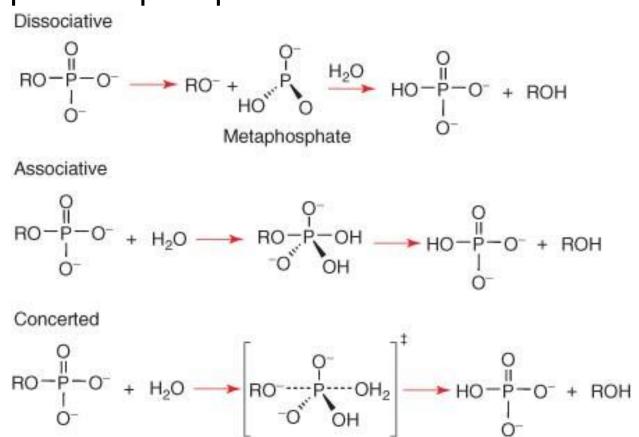


• (left) **DNA polymerase** catalyzed phosphodiester bond formation (Mg<sup>2+</sup>); (right) Transition-state model for phosphodiester bond formation in **RNA polymerase**.

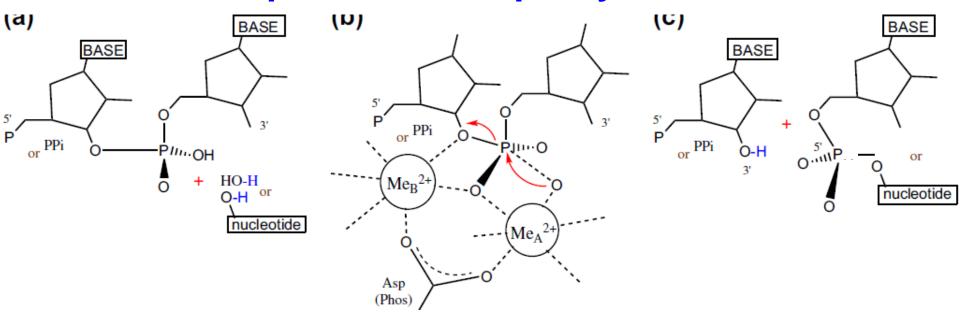
### **Phosphoryl Transfers**

- Enzymes that catalyze **phosphorylation** of substrates (usually Ser, Thr, or Tyr residues) typically use Mg<sup>2+</sup>-chelates to facilitate nucleophilic attack at the phosphate.
- Hydrolysis of simple phosphate esters is often catalyzed by enzymes with transition-metal cofactors, e.g. alkaline phosphatase & purple acid phosphatase.

### 3 possible mechanisms

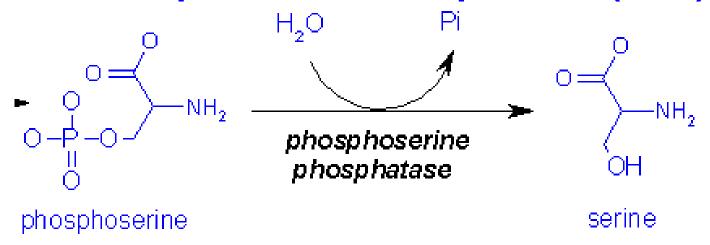


### 2-Metal-ion-dependent Phosphoryl Transfer Reaction

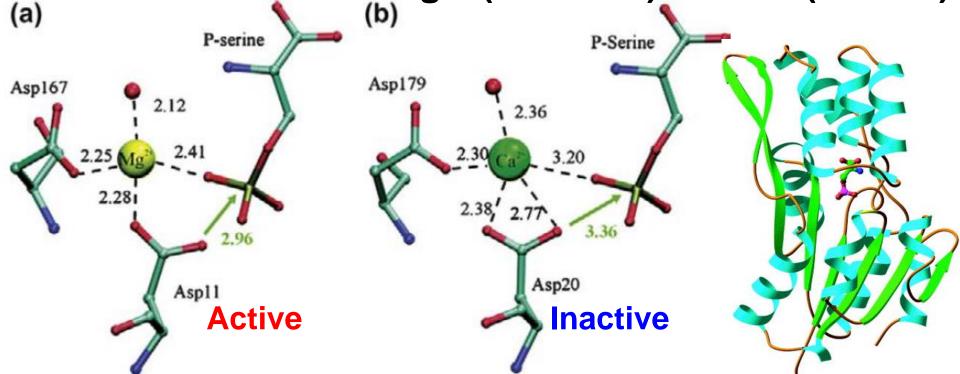


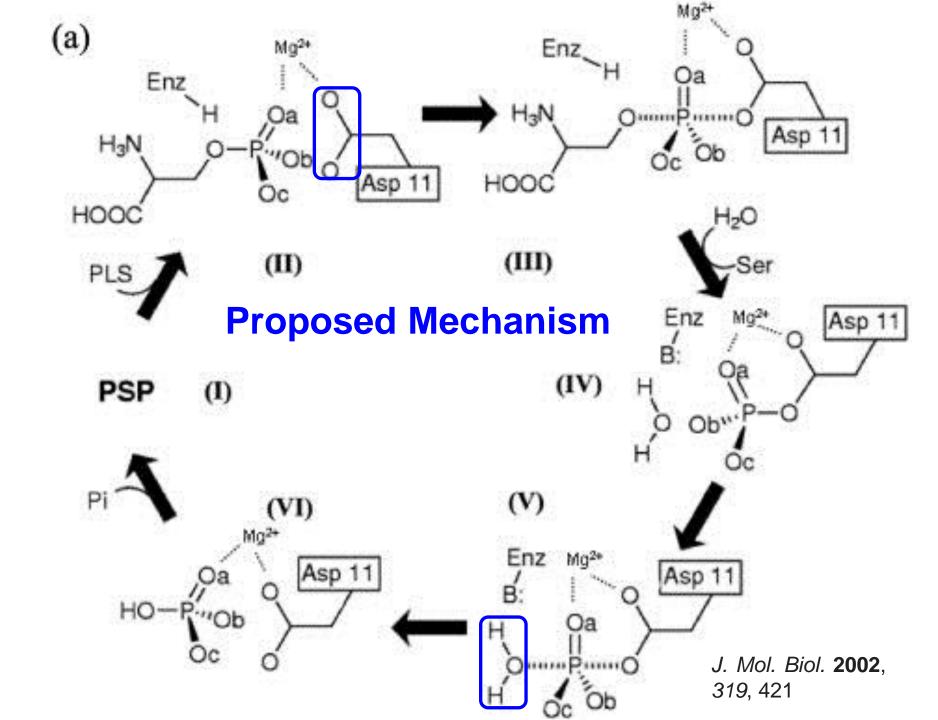
- (a) Substrates: phosphate & water (or RO-H).
- **(b) Intermediate**: the **2 metals** are always coordinated by **oxygen of the phosphate** and a conserved Asp.
- (c) Products: a new phosphoryl bond is formed between the nucleophile and phosphate.

### **Phosphoserine Phosphatase (PSP)**



Active site with one Mg<sup>2+</sup> (bacterial) or Ca<sup>2+</sup> (human)





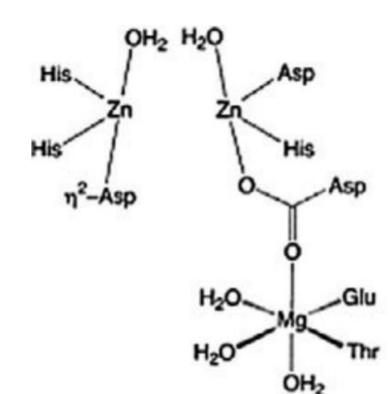
### E. coli Alkaline Phosphatase

Phosphate monoester

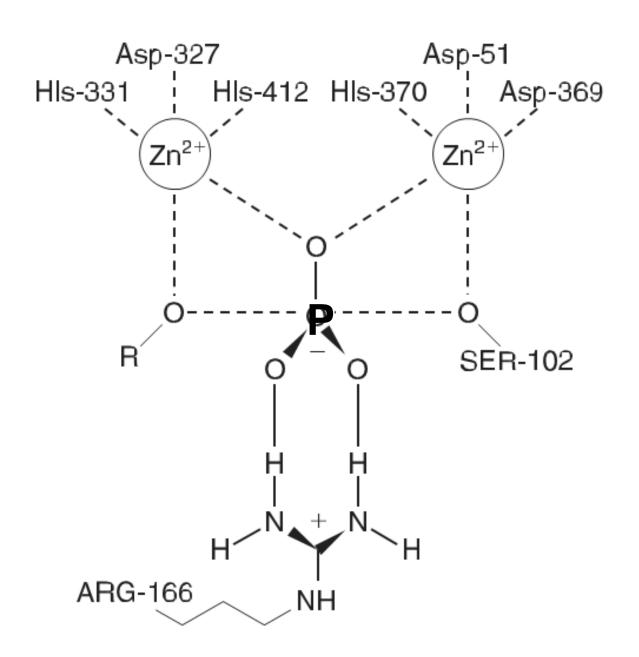
or R'OH (instead of H<sub>2</sub>O)

Alcohol Phosphate  $O=P(OH)_2(OR')$  phosphotransferase

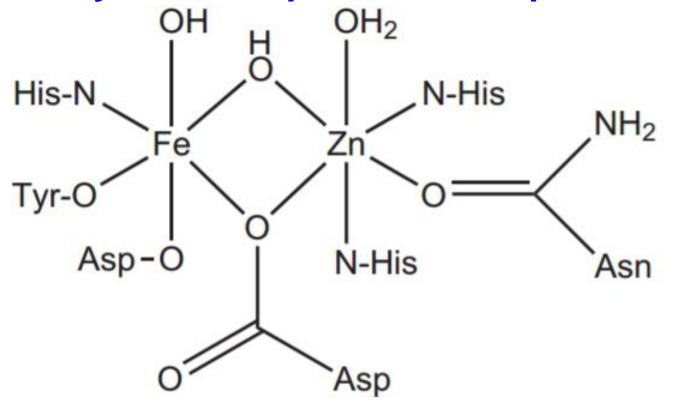
- Catalyzes removal of the phosphate groups.
- The optimal pH for activity:
   ~8 (basic) → a low pKa values for the Zn-bound water.



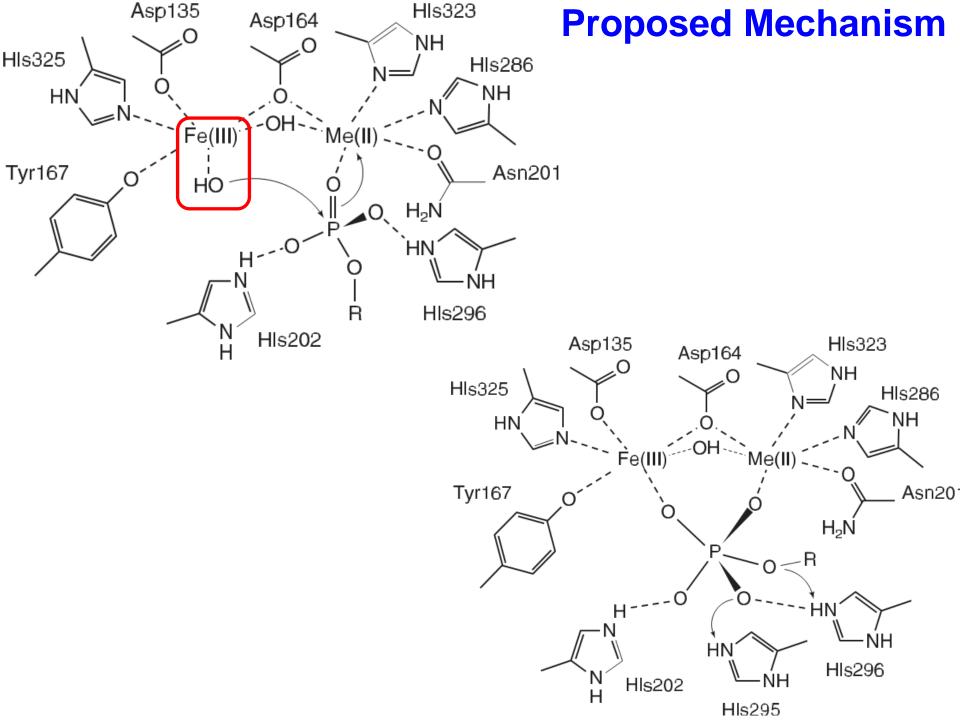
### **Proposed Mechanism**

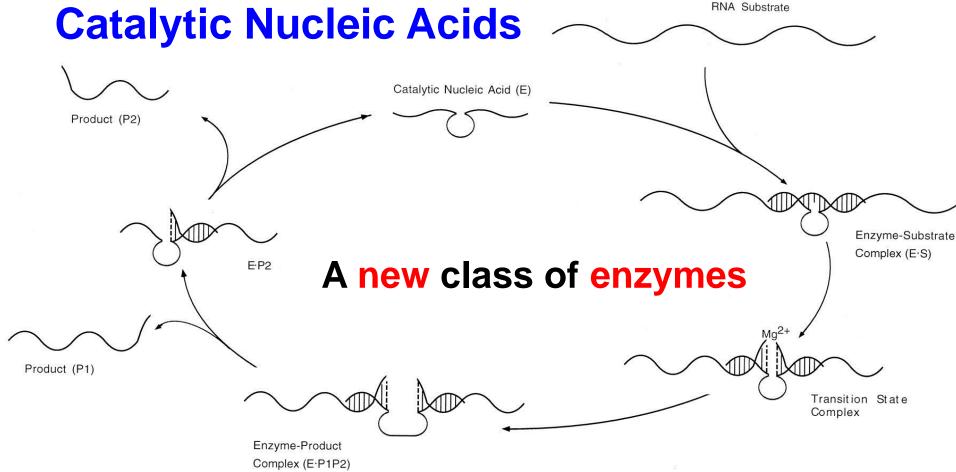


### **Kidney Bean Purple Acid Phosphatase**



- The optimal pH for activity: acidic → the high pKa values for the bound water.
- The substrates in acid phosphatases are **possibly** less reactive than alkaline phosphatase → require stronger Lewis acid (Fe(III)).



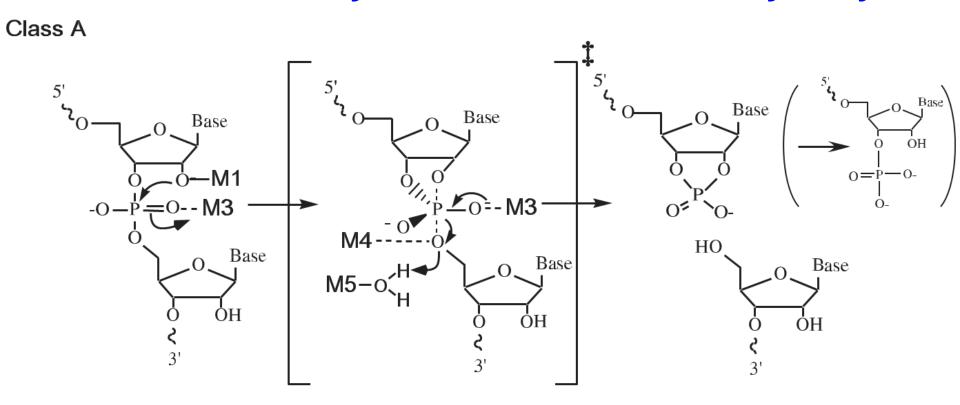


- Some RNA molecules were found to have enzymatic activities in 1982 (so-called catalytic RNA, ribozymes, RNA enzymes, or RNAzymes).
- Single-stranded DNA molecules were also found to catalyze RNA cleavage in 1994 (so-called deoxyribozymes, DNA enzymes or DNAzymes).

Reactions Catalyzed by Catalytic Nucleic Acids<sup>a</sup>

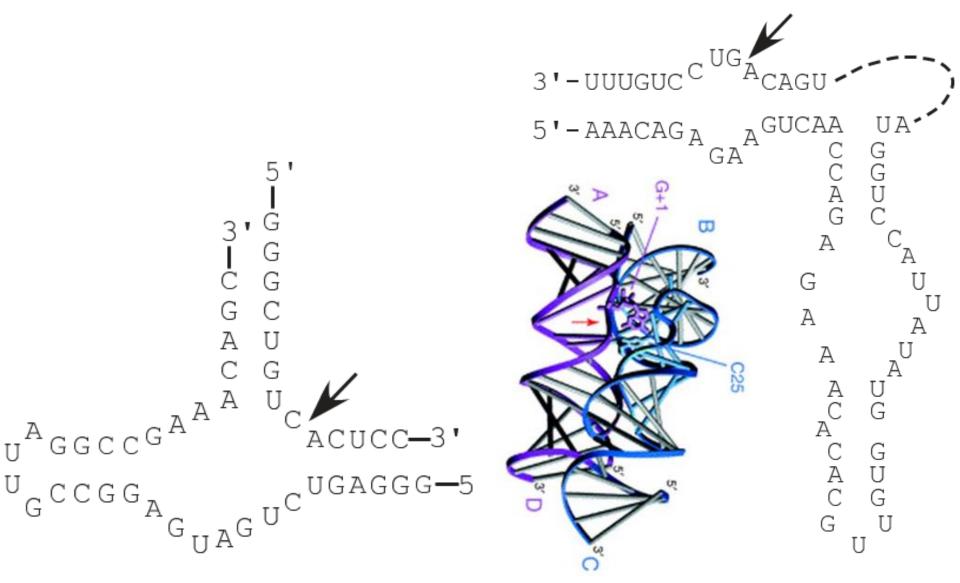
	Catalytic Activity				_	
Reaction <sup>b</sup>	Enzyme <sup>c</sup>	$k_{\rm cat}~({\rm min^{-1}})$	$K_{\rm m}~(\mu M)$	$k_{\rm cat}/k_{\rm u}$	<u>u</u>	
Phosphoester transfer	R-nat	0.1	$1 \times 10^{-3}$	1011	(R-nat):	
•	R-lab	0.3	0.02	$10^{13}$	catalytic	RNA
Phosphoester cleavage	R-nat	1	0.05	$10^{6}$	derived	from
	R-lab	0.1	0.03	$10^{5}$		ПОШ
	D-lab	3	$8 \times 10^{-4}$	$10^{6}$	naturally	
Polynucleotide ligation	R-nat	4	3	$10^{6}$	occurring	
	R-lab	100	9	$10^{9}$	•	
	D-lab	0.04	100	$10^{4}$	sources.	
Polynucleotide phosphorylation	R-lab	0.3	40	$> 10^{5}$		
Mononucleotide aminoacylation	R-lab	0.3	$5 \times 10^3$	$> 10^{7}$	(R-lab):	
Polynucleotide aminoacylation	R-lab	1	$9 \times 10^3$	$10^{6}$	,	
Aminoacyl ester hydrolysis	R-lab	0.02	0.5	10	catalytic	RNA
Aminoacyl transfer	R-lab	0.2	0.05	$10^{3}$	obtained	by in
Amide bond cleavage	R-lab			$10^{2}$	vitro selec	rtion
Amide bond formation	R-lab	0.04	2	$10^{5}$	villo selection.	
Peptide bond formation	R-lab	0.05	200	$10^{6}$		
N-Alkylation	R-lab	0.6	$1 \times 10^3$	$10^{7}$	(D-lab):	
S-Alkylation	R-lab			$10^{3}$	,	DNIV
Oxidative DNA cleavage	R-lab			$> 10^{6}$	catalytic	DNA
Biphenyl rotation	R-lab	$3 \times 10^{-5}$	500	$10^{2}$	obtained	by in
Porphyrin metallation	R-lab	0.9	10	$10^{3}$	vitro selec	ction
	D-lab	0.2	$3 \times 10^3$	$10^{3}$		
Diels-Alder cycloaddition	R-lab	> 0.1	> 500	$10^{3}$		

### 2 Classes of Catalytic Nucleic Acids for Hydrolysis



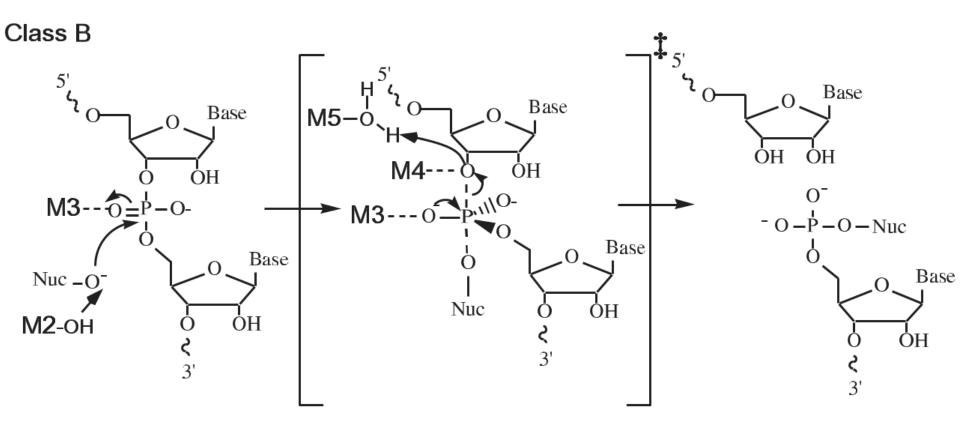
- They are typically small (< 200 bases).</li>
- M1 metal activates an internal 2'-OH for nucleophilic attack at the phosphorus of a phosphodiester bond & form 2',3'-cyclic phosphate & 5'-OH termini (e.g. in hammerhead, hairpin, hepatitis delta virus (HDV)).

### 2 Examples of Sequence of Catalytic RNA



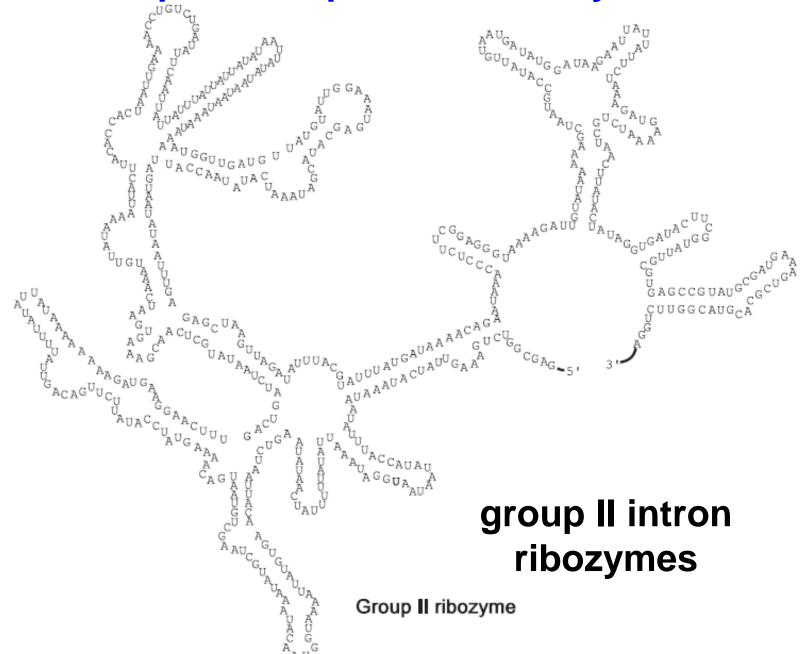
Hammerhead Ribozyme

Hairpin Ribozyme



- These catalytic nucleic acids are larger (> 400 bases, e.g. ribonuclease P, group I and group II intron ribozymes).
- They use an **external nucleophile**, e.g. an activated nucleotide or water, to attack the adjacent phosphodiester to form 3'-OH and 5'-phosphate termini.

### One Example of Sequence of Catalytic RNA



Catalytic Nucleic Acids	Functional	Nonfunctional					
Hammerhead <sup>b</sup>	$Mg^{2+}, Mn^{2+}, Ca^{2+}, \\ Cd^{2+}, Co^{2+}$	$\begin{array}{c} Ba^{2+},Sr^{2+},[Cr(NH_3)_6]^{3+},Pb^{2+},\\ Zn^{2+},Tb^{2+},Eu^{2+} \end{array}$					
Hairpin <sup>b</sup>	All tested, including $[Cr(NH_3)_6]^{3+}$						
Hepatitis $\delta$ virus <sup>b</sup>	$Mg^{2+}, Mn^{2+}, Ca^{2+}, Sr^{2+}$	$Cd^{2+}$ , $Ba^{2+}$ , $Co^{2+}$ , $Pb^{2+}$ , $Zn^{2+}$					
Neurospora VS <sup>b</sup>	Mg <sup>2+</sup> , Mn <sup>2+</sup> , Ca <sup>2+</sup>						
RNase $P^b$	$Mg^{2+}, Mn^{2+}, Ca^{2+}$	$Sr^{2+}$ , $Ba^{2+}$ , $Zn^{2+}$ , $Co^{2+}$ , $Cu^{2+}$ , $Fe^{2+}$ , $Ni^{2+}$					
Tetrahymena Group I <sup>b</sup>	$Mg^{2+}, Mn^{2+}$	$Ca^{2+}, Sr^{2+}, Ba^{2+}, Zn^{2+}, Co^{2+}, Cu^{2+}$					
Tetrahymena Group II <sup>b</sup>	$\mathrm{Mg}^{2+}$	$Ca^{2+}, Mn^{2+}$					
■ Mg²+, Mn²+, or Ca²+: generally essential for the catalytic							
function of most catalytic nucleic acids.							
● Structural role: facilitate folding to stable tertiary							
structures by charge neutralization with the anionic							
phosphodiester backbones.							
• Catalytic role: increase nucleophilicity of the							
nucleophile or stabilize the negative charge in transition							
states & products.							

### **Key Summary**

**Hydrolase:** enzymes catalyze **hydrolysis** of a chemical bond.

### General roles of the metal(s):

- 1. Structural role: bring the substrate into the active site & orient the substrate properly for the reactions.
- 2. Catalytic roles: as a Lewis acid to activate a metal-bound nucleophile (forming reactive M-OH or M-OR), stabilize the negative charge in transition states & leaving group/products.
- 3. Non-redox Zn<sup>2+</sup> & Mg<sup>2+</sup> are commonly used. The two metal ions are often required for the reactions.

Catalytic nucleic acids: a new class of enzymes.

# Thank You for Your Attention! Any Questions?