

# **GATK Best Practices for Variant Discovery**

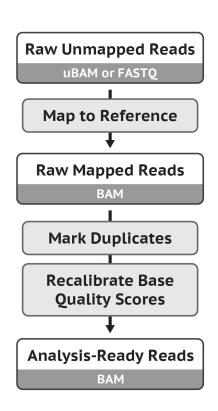
# Mapping

Finding where reads belong in the genome

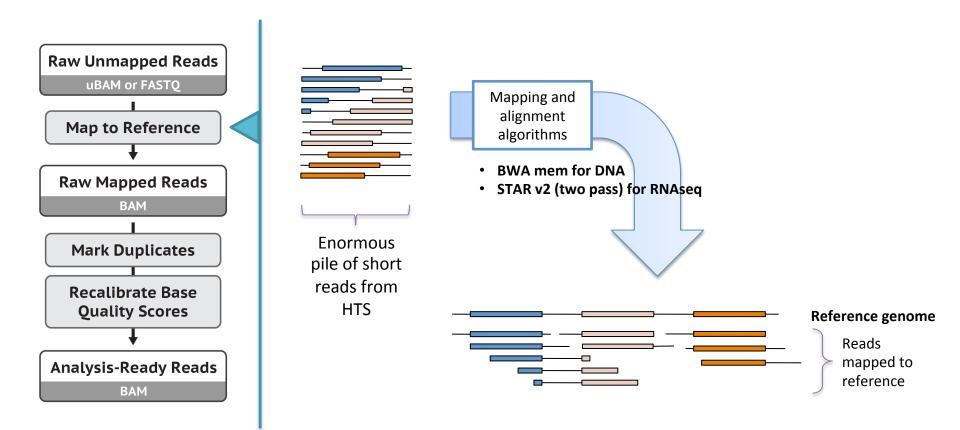




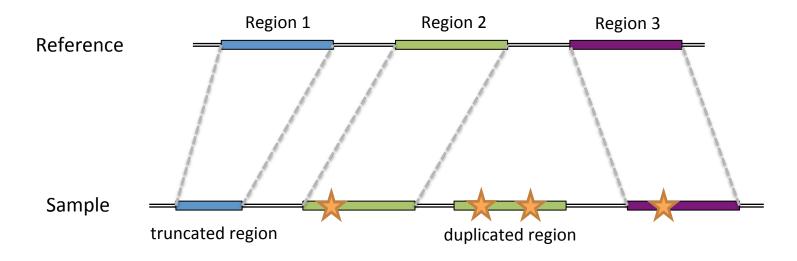
# **Data Pre-processing for Variant Discovery**



### **Step 1:** Map the reads produced by the sequencer to the reference



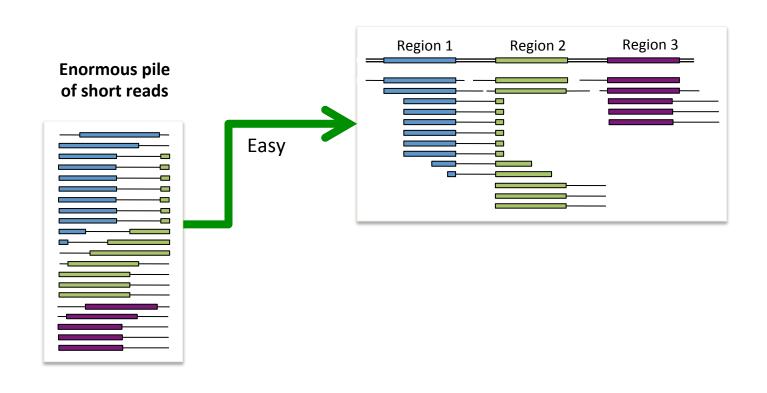
### Goal: align the sample genome to the reference genome



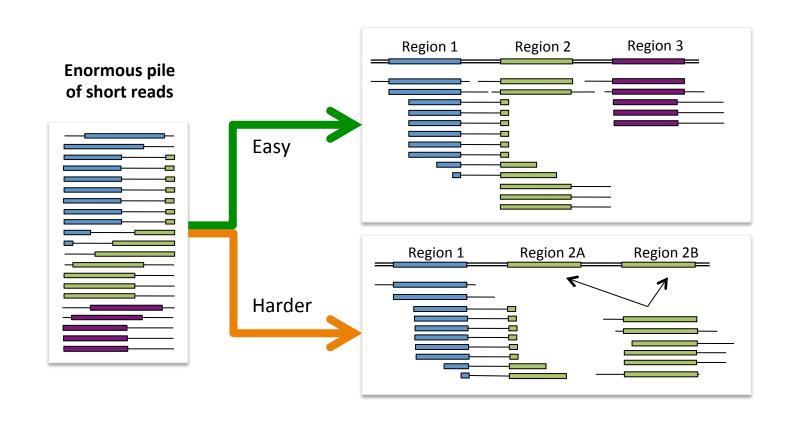


...But we don't have the whole sample in one piece.

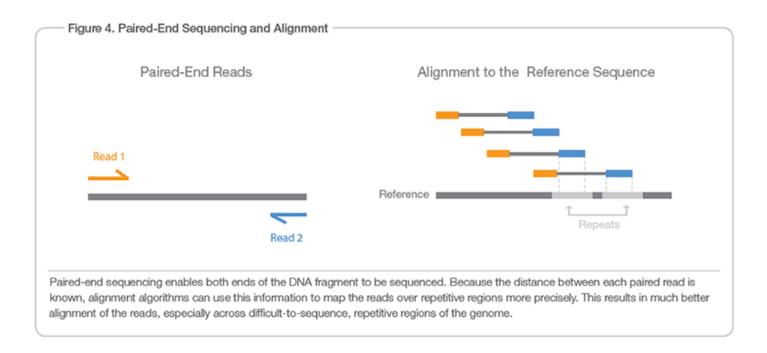
# So we have to map each little bit one by one



## **Complication:** mismatches, indels, duplicated regions...

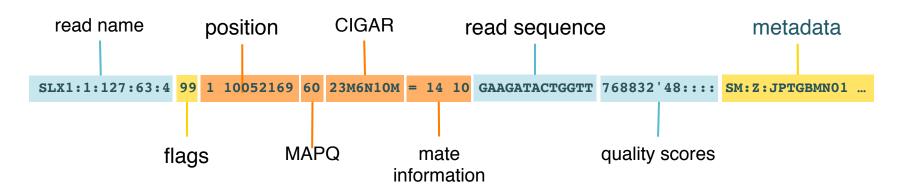


#### Paired-end sequencing helps a lot



# Output format: Sequence/Binary Alignment Map (SAM/BAM)

**HEADER** containing metadata (sequence dictionary, read group definitions etc) **RECORDS** containing structured read information (1 line per read record)

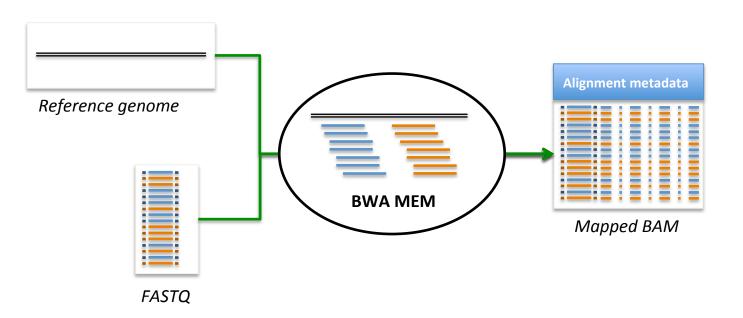


Added mapping info summarizes position, quality, and structure for each read

Special Note #1

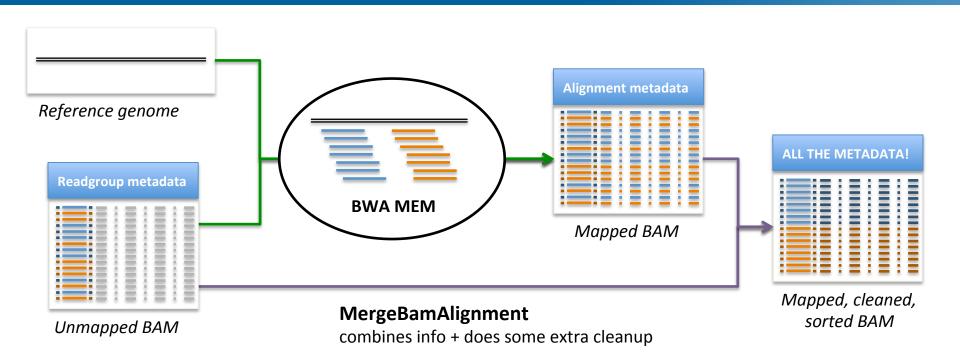
# THE UNMAPPED BAM WORKFLOW

### Regular FASTQ -> BAM workflow



! Adding Readgroup metadata requires additional step or injection of metadata into BWA command

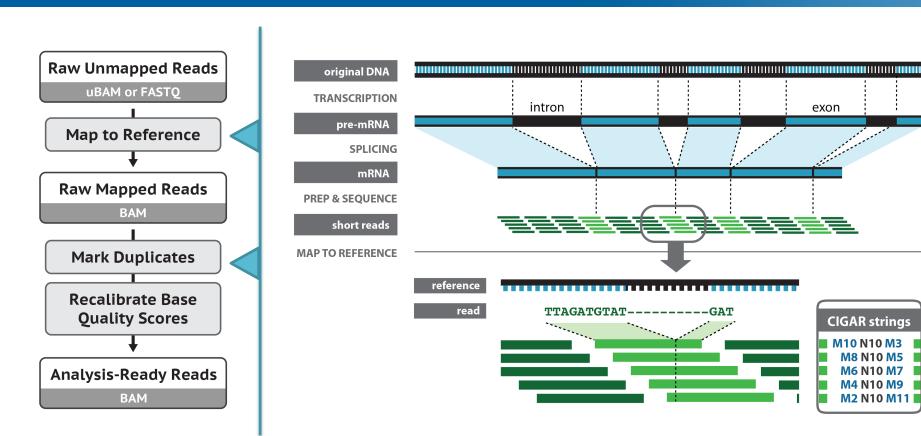
### Unmapped BAM -> BAM workflow



Special Note #2

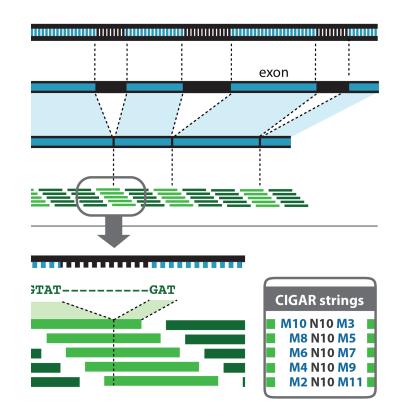
# **RNASEQ MAPPING**

## Special handling for RNAseq splice junctions



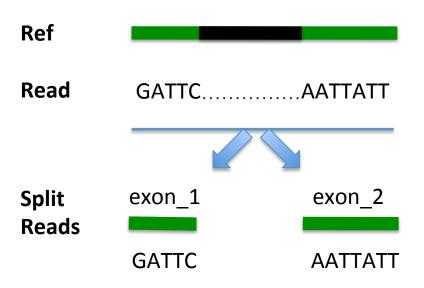
### Mapping RNAseq data with STAR v2

- Highest sensitivity for both SNPs and indels among all programs tested
- 2-pass approach described in
  - Pär G Engström et al. "Systematic evaluation of spliced alignment programs for RNA-seq data". *Nature Methods, 2013* (see Suppl. text p. 43 for detailed protocol)
  - First pass identifies splice junctions (SJ)
  - Use the SJ to guide the second round of alignment



# Split'N'Trim

1. Split reads with Ns in the CIGAR string



#### 2. Trim overhangs



# **Data Pre-processing for Variant Discovery**

