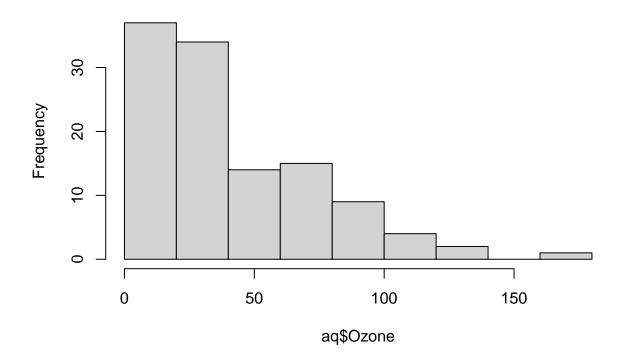
$Session\text{-}14_15.R$

SumanPaudel

2024-04-12

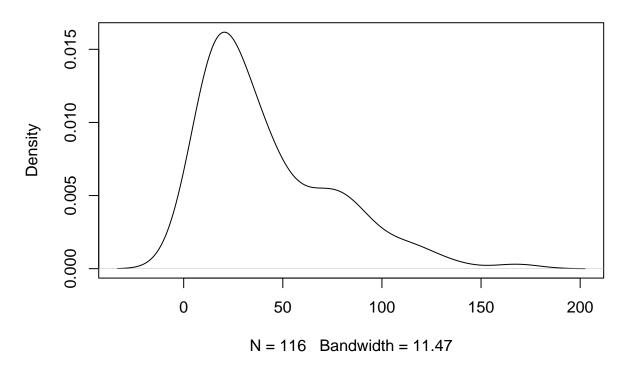
aq <- airquality
hist(aq\$0zone)</pre>

Histogram of aq\$Ozone

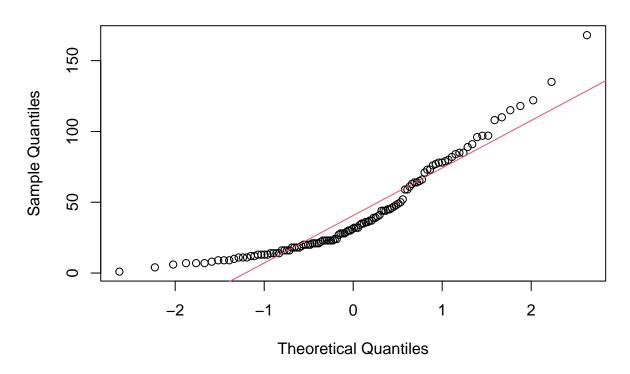


plot(density(aq\$0zone,na.rm=T))

density(x = aq\$Ozone, na.rm = T)



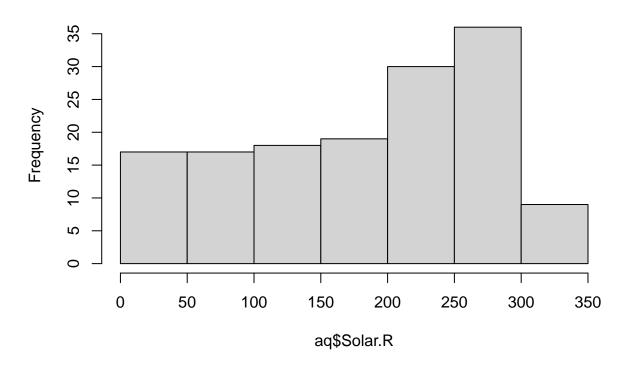
```
qqnorm(aq$0zone)
qqline(aq$0zone, col=2)
```



shapiro.test(aq\$0zone)

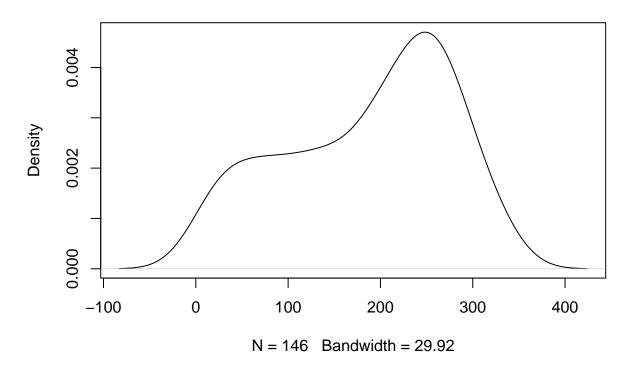
```
##
## Shapiro-Wilk normality test
##
## data: aq$0zone
## W = 0.87867, p-value = 2.79e-08
hist(aq$Solar.R)
```

Histogram of aq\$Solar.R

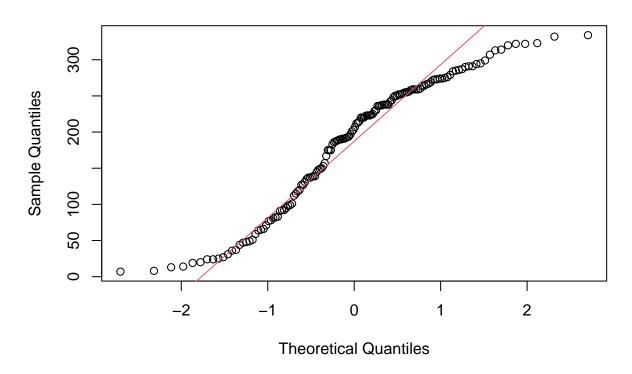


plot(density(aq\$Solar.R,na.rm=T))

density(x = aq\$Solar.R, na.rm = T)



```
qqnorm(aq$Solar.R)
qqline(aq$Solar.R, col=2)
```

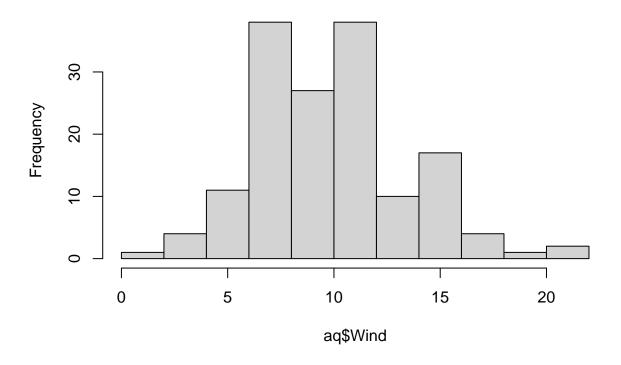


shapiro.test(aq\$Solar.R)

hist(aq\$Wind)

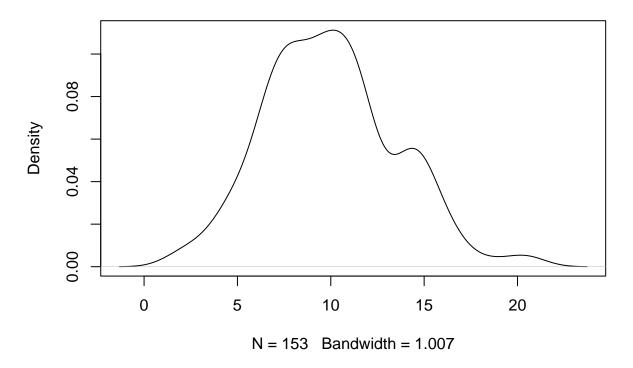
```
##
## Shapiro-Wilk normality test
##
## data: aq$Solar.R
## W = 0.94183, p-value = 9.492e-06
# testing normality for wind
```

Histogram of aq\$Wind

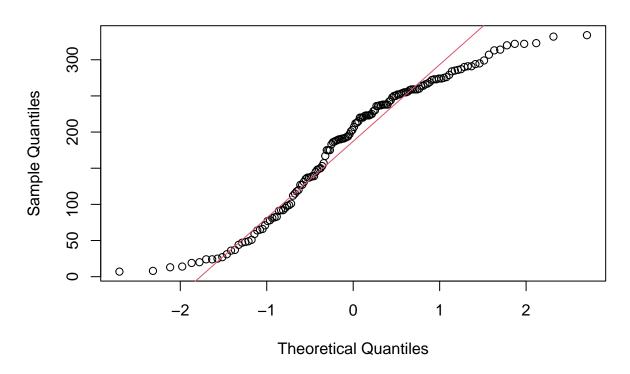


plot(density(aq\$Wind,na.rm=T))

density(x = aq\$Wind, na.rm = T)



```
qqnorm(aq$Solar.R)
qqline(aq$Solar.R, col=2)
```

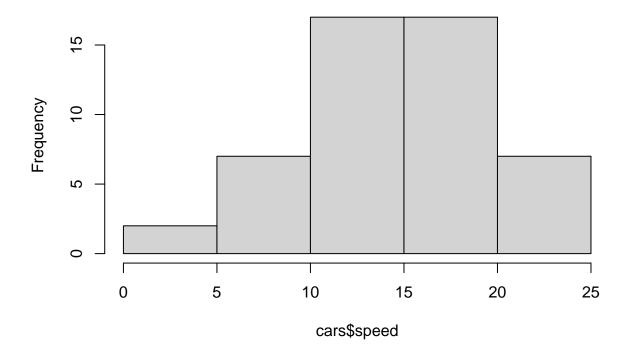


shapiro.test(aq\$Wind)

```
##
## Shapiro-Wilk normality test
##
## data: aq$Wind
## W = 0.98575, p-value = 0.1178

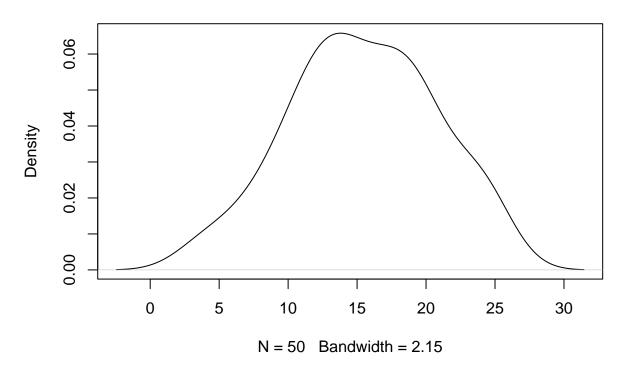
# testing normality for wind
# goodness of fit
hist(cars$speed)
```

Histogram of cars\$speed

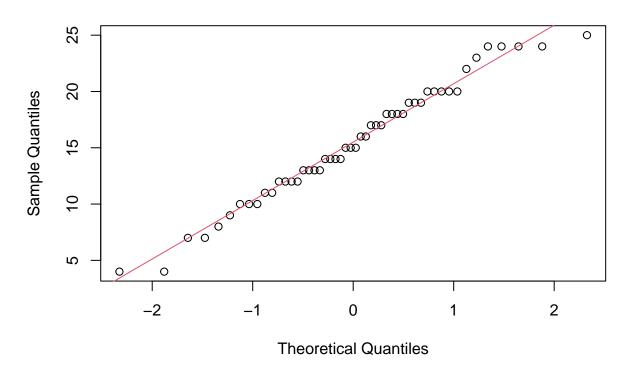


plot(density(cars\$speed,na.rm=T))

density(x = cars\$speed, na.rm = T)



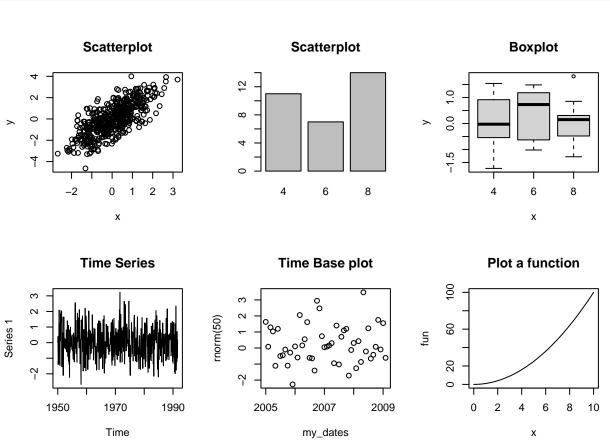
```
qqnorm(cars$speed)
qqline(cars$speed,col=2)
```



shapiro.test(cars\$speed)

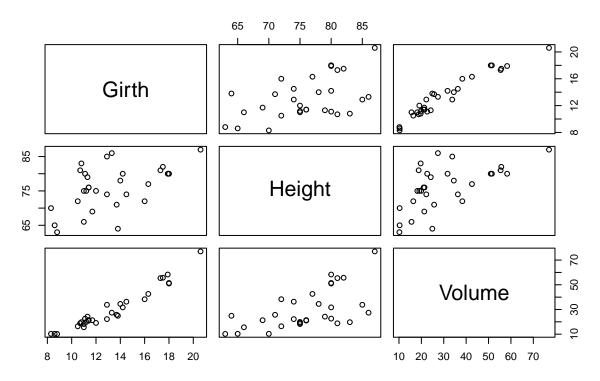
```
##
##
    Shapiro-Wilk normality test
##
## data: cars$speed
## W = 0.97765, p-value = 0.4576
# test of normality: types
# sekwness and kurtosis based
# # jarque-bera test of normality
# larges sample based
# kolmogorav-smirnov test of normality
# ks.test(cars$speed)
# small sample based
# shapiro wil test of normality
# multiple graph in single wind
x <- rnorm(500)
y \leftarrow x + rnorm(500)
```

```
my_ts <-
  ts(
    matrix(x,
    nrow = 500,
    ncol = 1),
    start = c(1950, 1),
    frequency = 12
  )
my_dates <- seq(as.Date("2005/1/1"),by="month",length=50)</pre>
my_factor <- factor(mtcars$cyl)</pre>
fun <- function(x) x^2</pre>
par(mfrow=c(2,3))
plot(x,y,main="Scatterplot")
plot(my_factor,main="Scatterplot")
plot(my_factor, rnorm(32), main="Boxplot")
plot(my_ts, main="Time Series")
plot(my_dates, rnorm(50), main= "Time Base plot")
plot(fun, 0, 10, main="Plot a function")
```



```
par(mfrow=c(1,1))
plot(trees[,1:3], main="Correlation Plot")
```

Correlation Plot



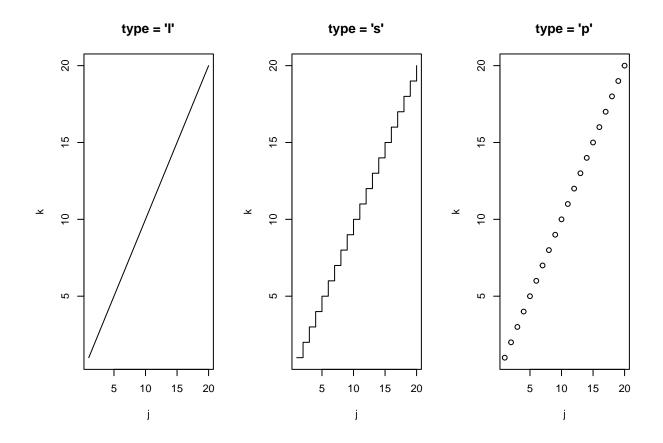
```
# if linear use pearson coefficeent
# if not linear use spearman coefficeent

# bi serial
# for testing both categorical and continuous

j <- 1:20
k <- j
par(mfrow=c(1,3))

plot(j,k,type="l",main="type = 'I'")

plot(j,k,type="s",main="type = 's'")
plot(j,k,type="p",main="type = 'p'")</pre>
```

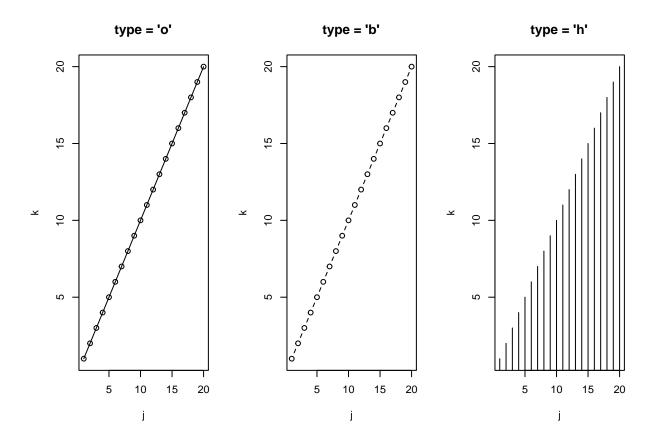


```
par(mfrow=c(1,3))

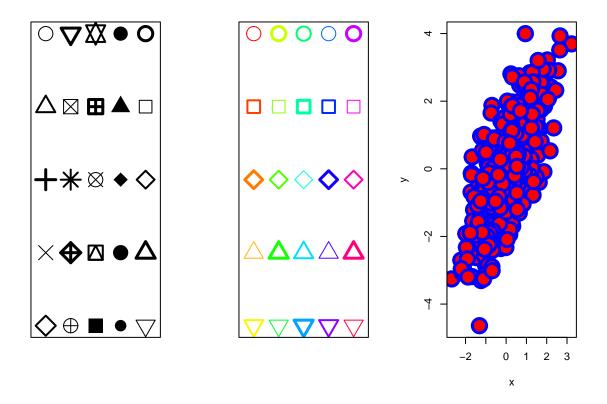
plot(j,k,type="o",main="type = 'o'")

plot(j,k,type="b",main="type = 'b'")

plot(j,k,type="h",main="type = 'h'")
```

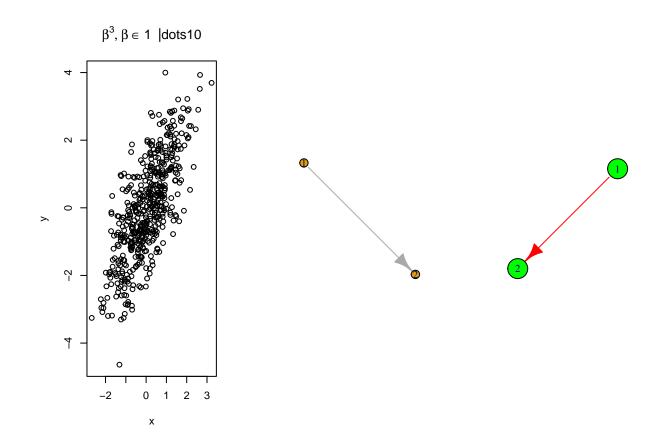


```
r <- c(sapply(seq(5, 25, 5), function(i) rep(i,5)))</pre>
t \leftarrow rep(seq(25, 5, -5), 5)
plot(r, t, pch=1:25, cex=3, yaxt="n", xaxt="n", ann=F,xlim=c(3,27),lwd=1:3)
plot(
 r,
  t,
  pch = 21:25,
  cex = 3,
  yaxt = "n",
  xaxt = "n",
  ann = F,
  xlim = c(3, 27),
  lwd = 1:3,
  col = rainbow(25)
)
plot(x,y, pch=21, bg="red",col="blue",cex=3,lwd=3)
```



```
# why values in the scatterplot lie in the range of -4 to 4
\# x < -rnom(5)
plot(x,y, main=latex2exp::TeX('$\\beta^3,\\beta\\in 1 \\ | dots 10$'))
# networks
# graph analysis
# Session 15
# SNA Basics
library(graph)
## Loading required package: BiocGenerics
##
## Attaching package: 'BiocGenerics'
## The following objects are masked from 'package:stats':
##
##
       IQR, mad, sd, var, xtabs
## The following objects are masked from 'package:base':
##
```

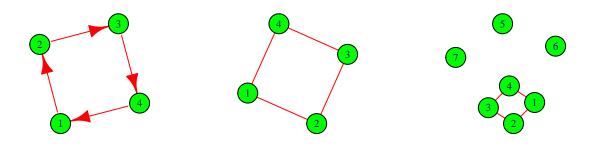
```
anyDuplicated, aperm, append, as.data.frame, basename, cbind,
##
##
       colnames, dirname, do.call, duplicated, eval, evalq, Filter, Find,
       get, grep, grepl, intersect, is.unsorted, lapply, Map, mapply,
##
##
       match, mget, order, paste, pmax, pmax.int, pmin, pmin.int,
       Position, rank, rbind, Reduce, rownames, sapply, setdiff, sort,
##
##
       table, tapply, union, unique, unsplit, which.max, which.min
library(igraph)
## Warning: package 'igraph' was built under R version 4.3.3
##
## Attaching package: 'igraph'
## The following objects are masked from 'package:graph':
##
##
       degree, edges, intersection, union
## The following objects are masked from 'package:BiocGenerics':
##
##
       normalize, path, union
## The following objects are masked from 'package:stats':
##
##
       decompose, spectrum
## The following object is masked from 'package:base':
##
       union
library(Rgraphviz)
## Loading required package: grid
g <- graph((c(1,2)))</pre>
plot(g)
plot(g, vertex.color='green',vertex.size=40,edge.color='red', edge.size=20)
```



```
g <- graph(c(1,2,2,3,3,4,4,1))
plot(g, vertex.color='green',vertex.size=40,edge.color='red', edge.size=20)

g <- graph(c(1,2,2,3,3,4,4,1),directed = F)
plot(g, vertex.color='green',vertex.size=40,edge.color='red', edge.size=20)

g <- graph(c(1, 2, 2, 3, 3, 4, 4, 1), directed = F, n = 7)
plot(g, vertex.color='green',vertex.size=40,edge.color='red', edge.size=20)</pre>
```



```
# defining nodes with text data
g1 <- graph(c("Sita", "Ram", "Ram", "Rita", "Rita", "Sita", "Sita", "Rita", "Anju", "Ram"))</pre>
plot(g1, vertex.color='green',vertex.size=40,edge.color='red', edge.size=20)
igraph::degree(g1)
## Sita Ram Rita Anju
##
     3
           3
               3
igraph::degree(g1, mode="in")
## Sita Ram Rita Anju
## 1
        2
               2 0
diameter(g1, directed = F)
## [1] 2
edge_density(g1, loops = F)
```

[1] 0.4166667

```
# edge density
ecount(g1) / (vcount(g1)*(vcount(g1-1)))

## [1] 0.4166667

reciprocity(g1)

## [1] 0.4

closeness(g1, mode='all', weights=NA)

## Sita Ram Rita Anju
## 0.2500000 0.3333333 0.2500000 0.2000000

betweenness(g1, directed = T, weights = NA)

## Sita Ram Rita Anju
## 1 2 2 0
```

