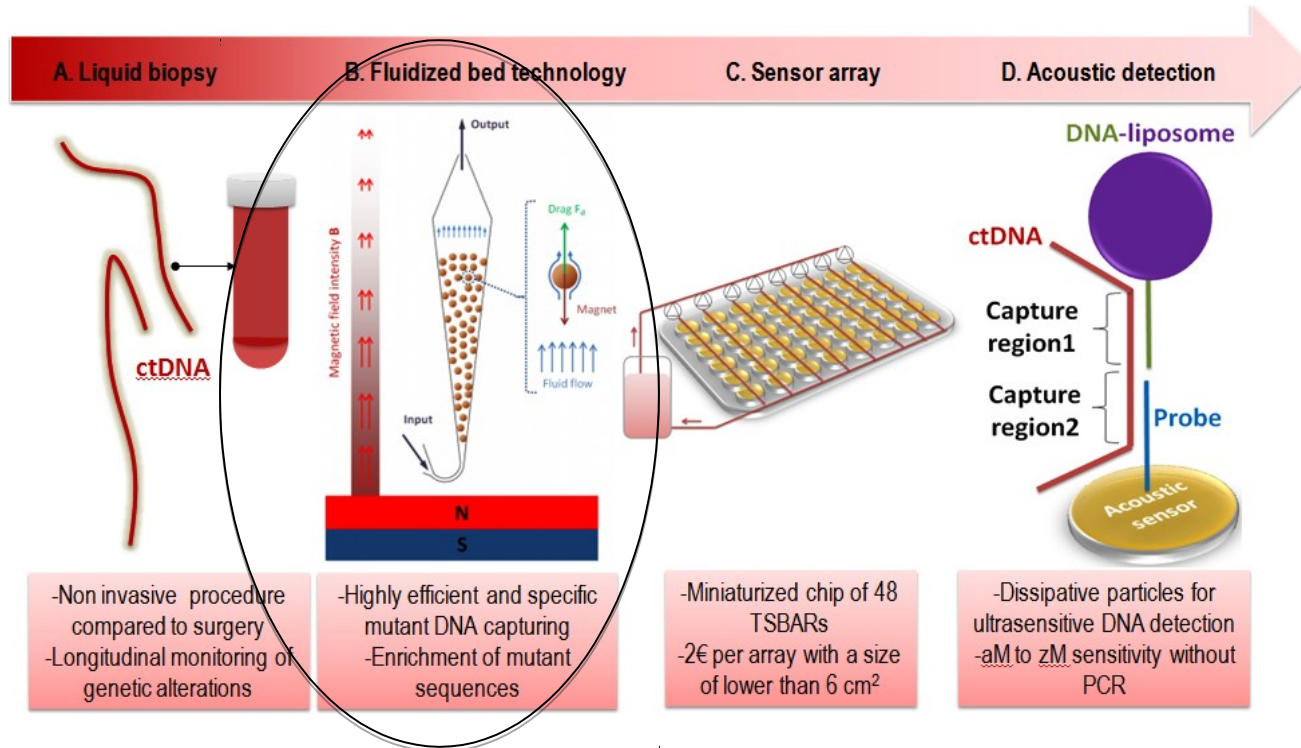


WP3: ctDNA enrichment strategies

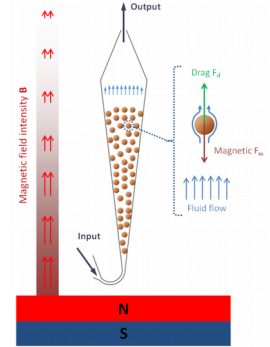


This WP is focused on innovative microfluidic-based strategy for processing of patient samples.

Our approach is to combine microfluidic technology with molecular biology to select and enrich ctDNA materials

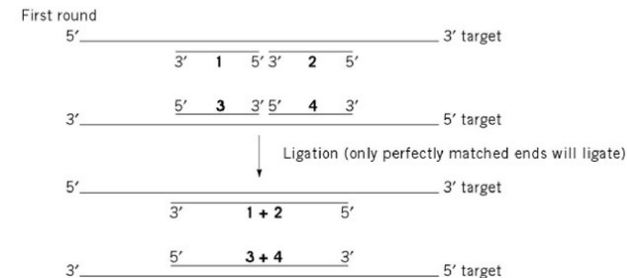
Task T3.1: Optimization of ctDNA selective capturing on magnetic beads in serum (Curie, M1-M24): (D3.1)

- Streptavidin-biotin system for capture oligo functionalization
- Capture through DNA-DNA interaction
- Improvement of pretreatment throughput and capture efficiency
- Control of hybridization $T^{\circ}C$



Task T3.2 : Optimization of a ligase-based assay for the selective enrichment of ctDNA targets (Curie, FORTH, M1-M30)

- Oligonucleotides sequence composition and length
- Number of cycles and temperatures
- Use of single-base 3' overhangs
- 5' phosphorylation of oligos
- Introduction of non-complementary tails



Task T3.3 : Optimized protocol for ctDNA isolation and enrichment (Curie, FORTH, M1-M36)

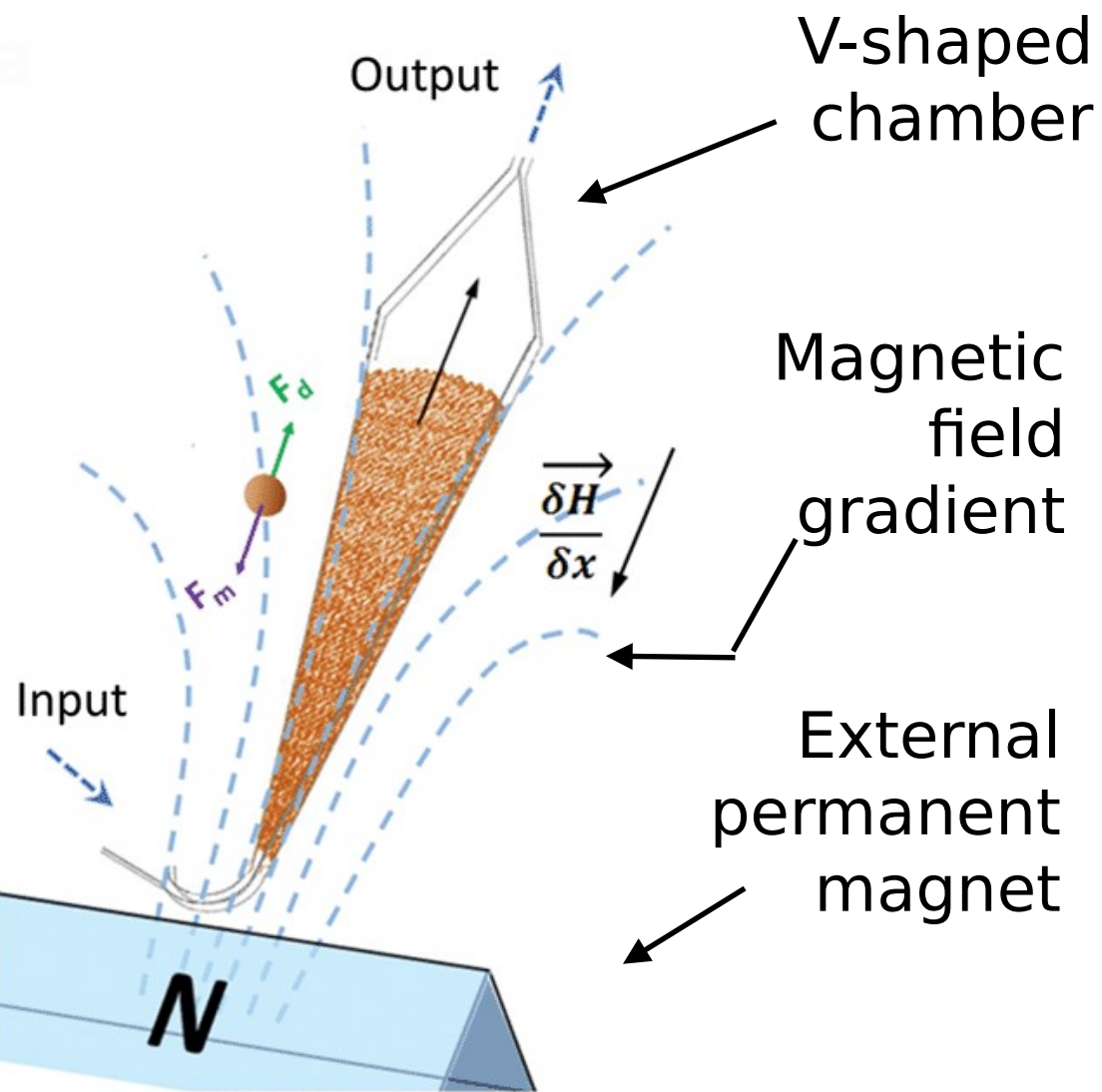
- Combination of tasks 3.2 and 3.3 outcomes
- Two strategy direct LCR or DNA release + LCR

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Task T3.1: Optimization of ctDNA selective capturing on magnetic beads in serum (Curie, M1-M24): (D3.1)

Monica Araya-Farias, Lucile Alexandre, Manh Louis Nguyen, Laura Trapiella-Alfonso, Jean-Louis Viovy and Stéphanie Descroix

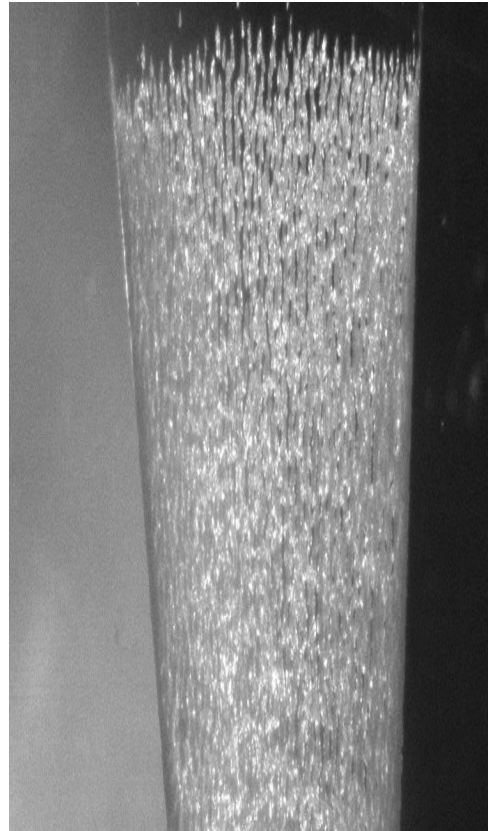
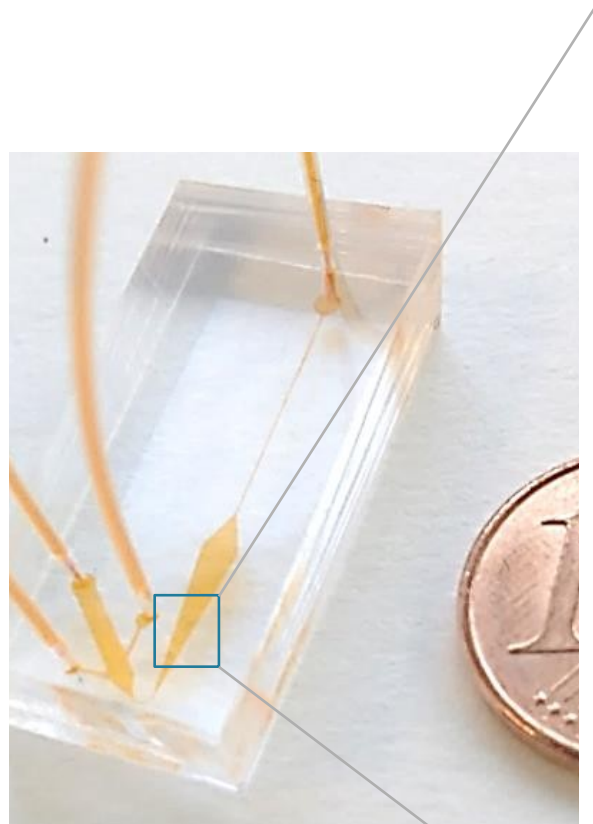
CATCH-U TECHNOLOGY : MICROFLUIDIC FLUIDIZED



Simplicity of fabrication and operation

THE MICROFLUIDIC FLUIDIZED BED

- Simplicity of fabrication and operation
- Well controlled particle recirculation and bead density



For Catch-U the beads will be functionalized to extract DNA from patients samples

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Task T3.1: ctDNA CAPTURE WITH THE FLUIDIZED BED TECHNOLOGY

Main objectives

1/ Improvement of the fluidized bed technology

† 2nd generation : Achieved
Bimodal beads composition +vibration

2/ ctDNA capture : two strategies

✓ Non specific capture - Achieved
(K. Perez-Toralla Sensors and actuators 2019)

✓ Specific hybridization : ongoing
(Carchi et al - Review Meeting 28th June 2019)

Second generation of fluidized bed

Similar design except : $h \times 5 : 250 \mu\text{m}$
Flow rate increase up to $15 \mu\text{L/min}$

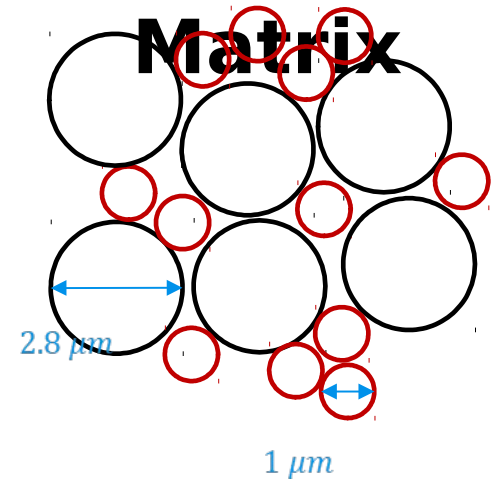
To avoid non uniformity in the bed of beads



External vibration



Bimodal Matrix



Model DNA capture with the second generation of fluidized bed

DNA WT

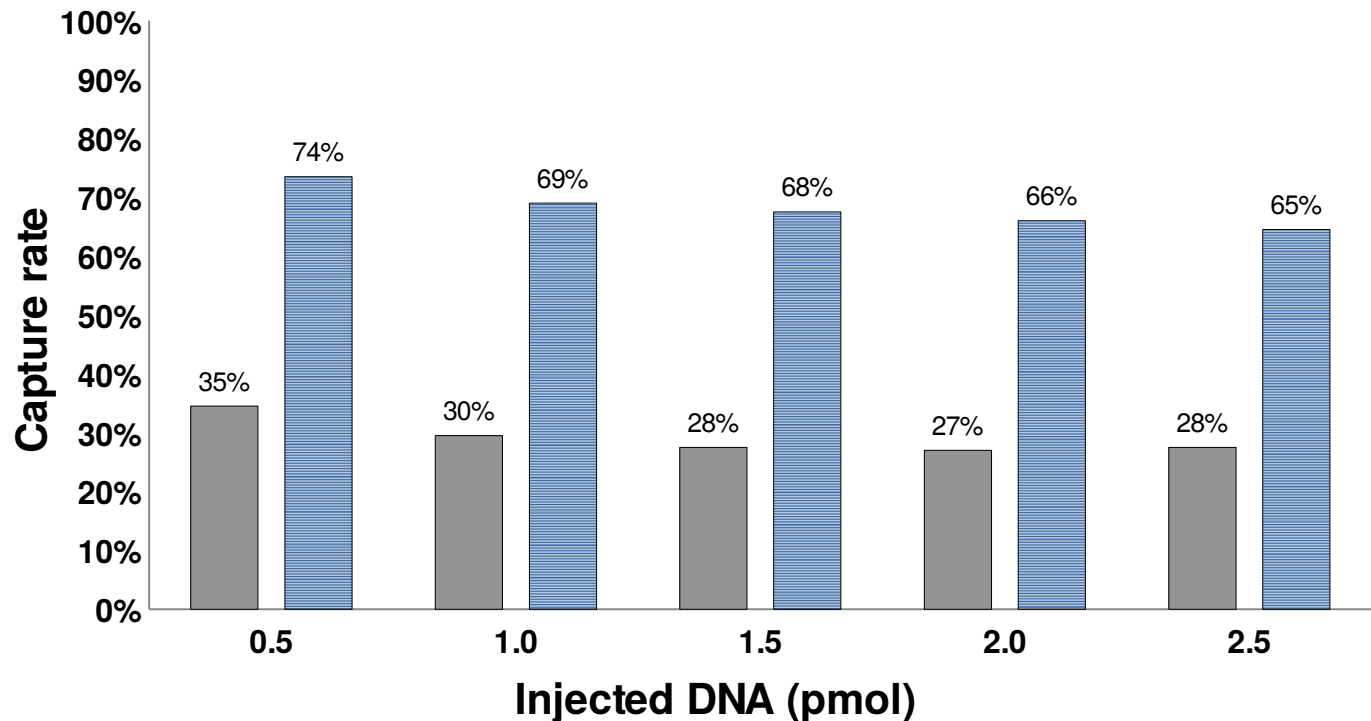


WT80bp

15
 $\mu\text{l}/\text{min}$

■ C Standard

■ Vib+bimodal



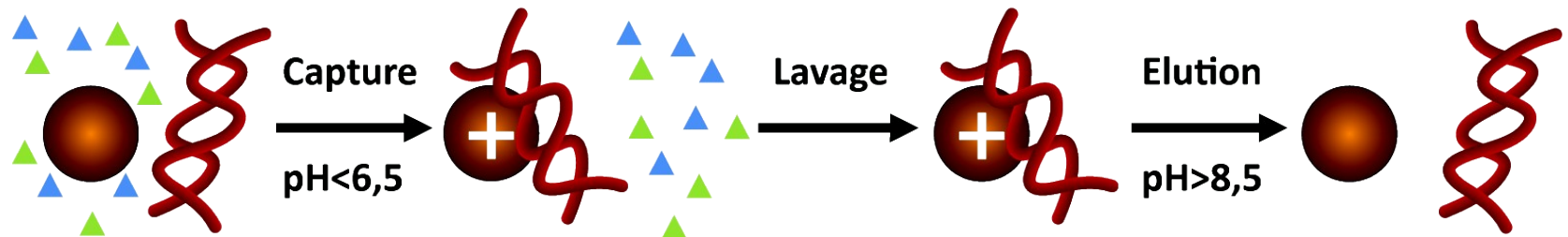
Objective 2 : ctDNA CAPTURE WITH THE FLUIDIZED BED TECHNOLOGY

- ctDNA capture : two strategies
 - ✓ non specific capture : achieved
 - ✓ specific hybridization : ongoing

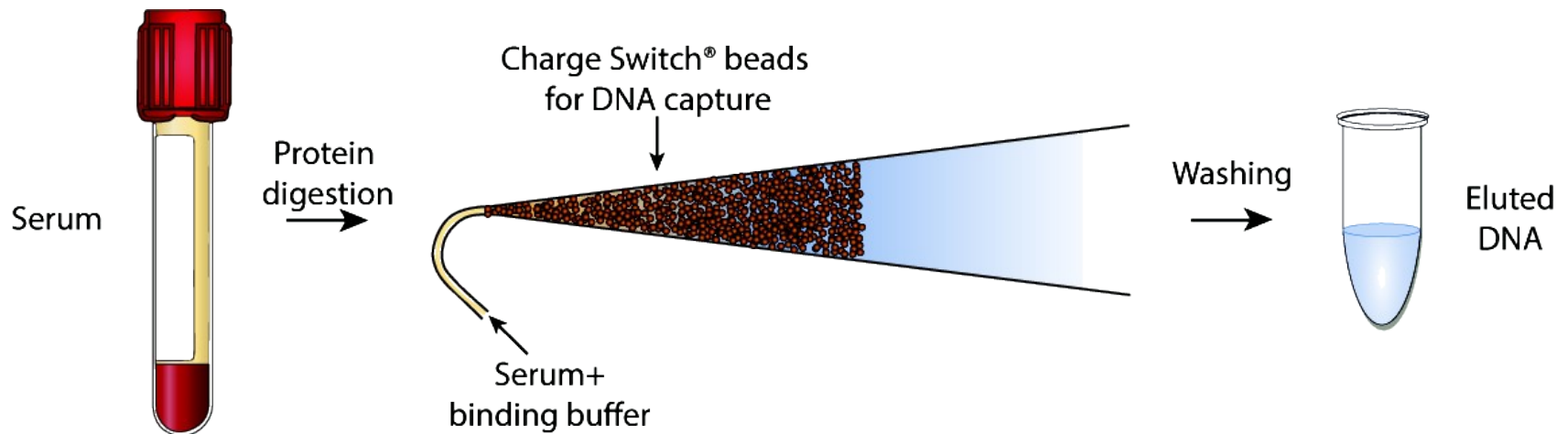


NON SPECIFIC DNA EXTRACTION

● ChargeSwitch® Technology



DNA capture based on electrostatic interactions

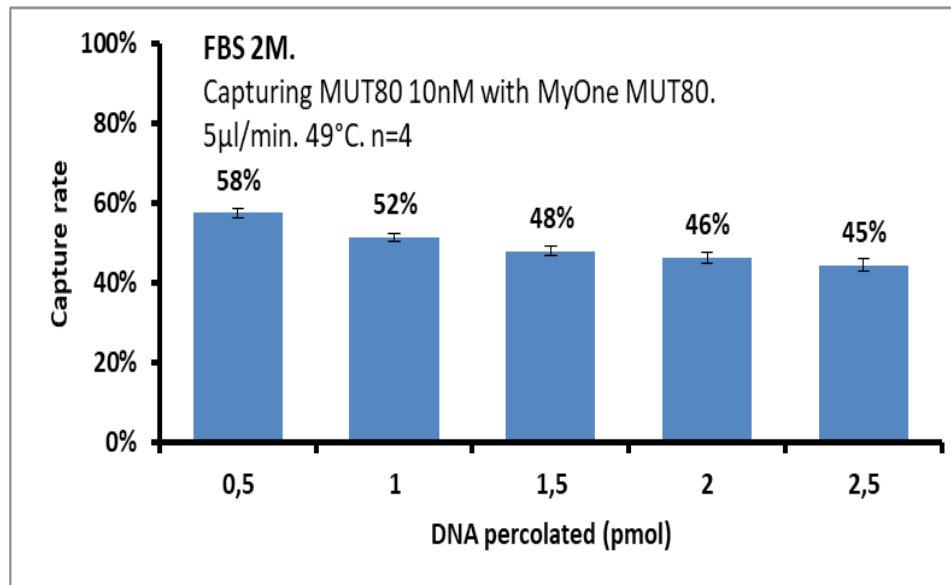


SPECIFIC DNA EXTRACTION WITH THE

First period : optimization and demonstration of single strand DNA capture

- Buffer salinity optimization
- Optimal temperature
- Optimization of the size of the capture oligo
- Study of the capture efficiency and

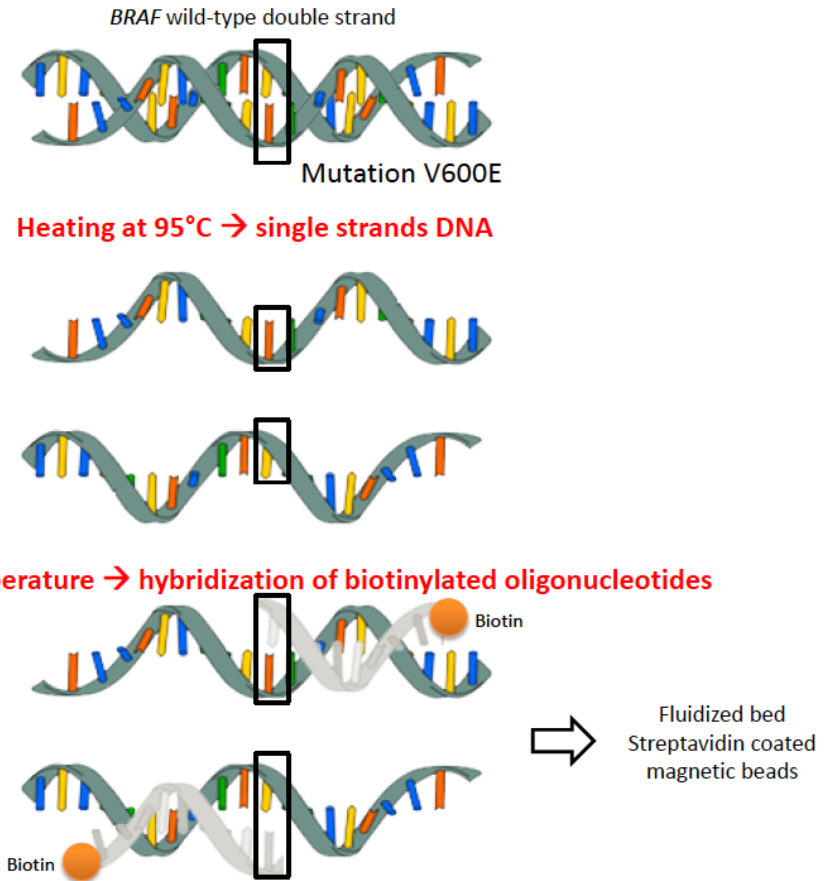
sp



SPECIFIC DNA EXTRACTION WITH THE FLUIDIZED BED

Second period : optimization and demonstration of double strand DNA capture

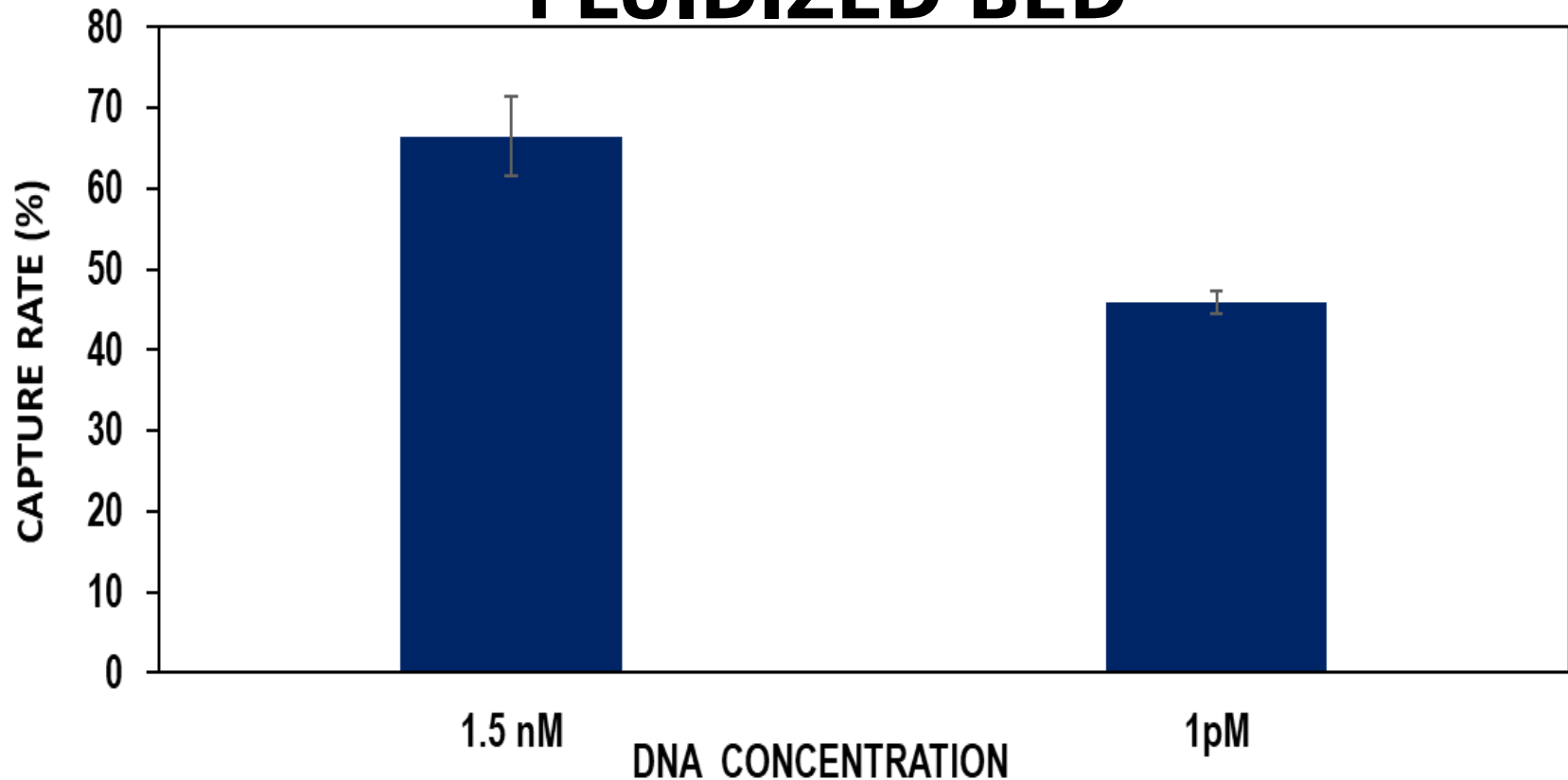
1. Heating to separate double stranded DNA
2. Introduction of biotinylated oligos
3. Capture on streptavidin beads



SPECIFIC DNA EXTRACTION WITH THE FLUIDIZED BED

- Optimization of the sample pretreatment :
Proteinase K
 - Incubation 2h at 37°C with PROTEINASE K (2.75 mg/mL)
- Inactivation of the enzyme to avoid ligands digestion
 - Heating at 95°C is a 'Win-Win' condition : DNA and PK denaturation
- Evaluation of DNA capture in buffer and in serum

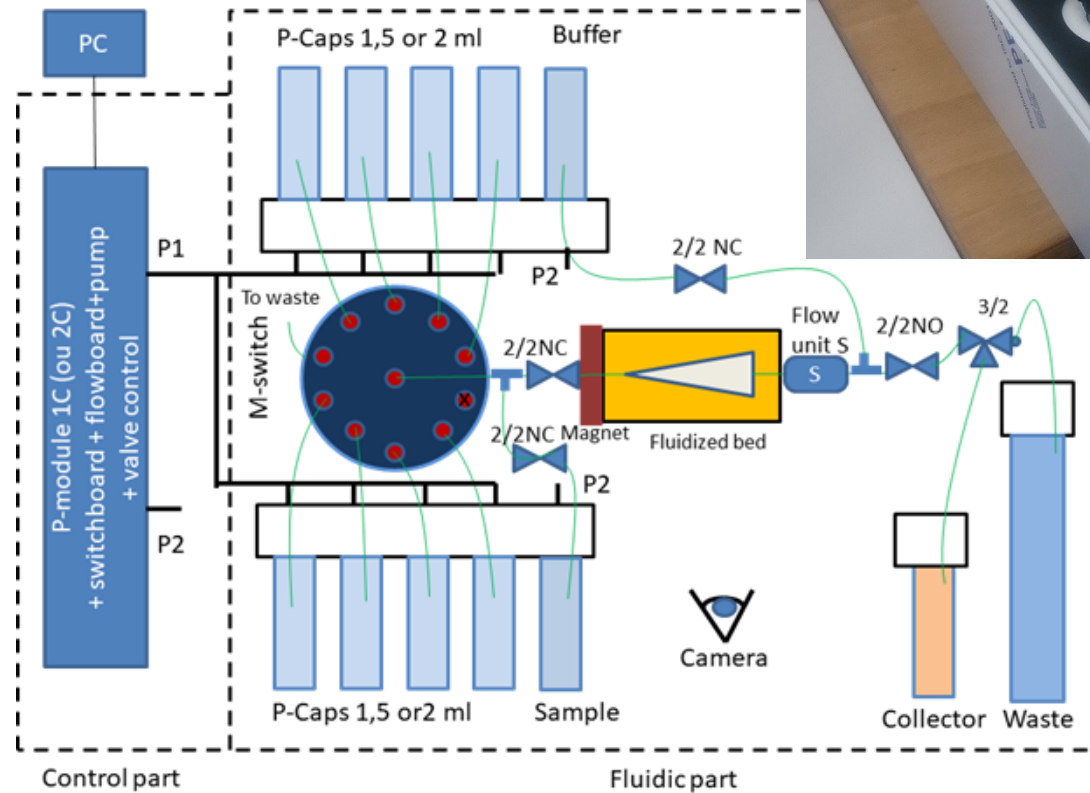
SPECIFIC DNA EXTRACTION WITH THE FLUIDIZED BED



Capture Rate (%) of dsDNA 277 BRAF MUT in human serum by using FB generation 2.0.
Error bars represent mean \pm SD of duplicate.

Capture was performed with streptavidin beads at Room Temperature, 5 μ L/min. Standard matrix + vibration. Probes (0,4 μ M). Human serum was pretreated with Proteinase K (2.75 mg/mL) 2h at 37°C following by a denaturation at 95°C (5 min) and an incubation at room temperature (15 min) before performing the capture on FB.

Fluidized Bed platform



**First Experiments to combine fluidized + LCR here
in Heraklion**

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Conclusion and Perspectives

Technology

- Development of fluidized 2.0 able to accommodate large volume
- Demonstration of the use of this second generation for DNA capture

DNA capture (D3.1 Delivered)

- Demonstration of non specific DNA capture on chip with patients samples
- New approaches for DNA specific capture from serum samples

DNA capture combined with LCR